

SUPPORTING INFORMATION

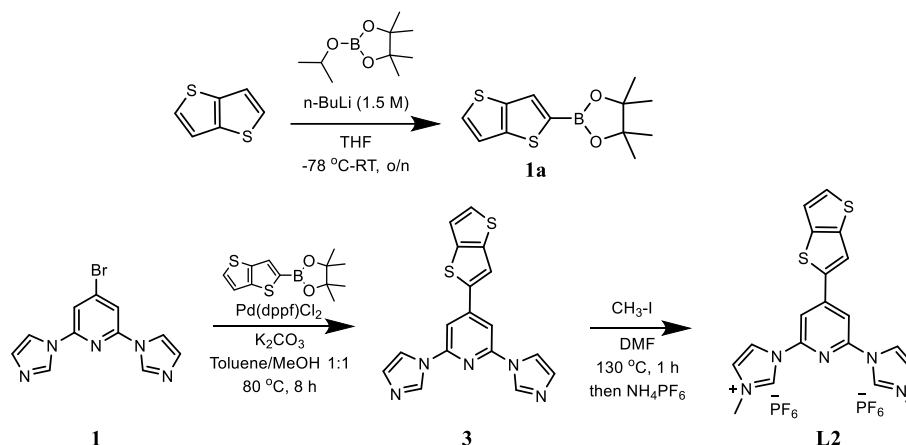
CONTENTS

Synthesis of ligands	S1
Synthesis of L2	S1
Synthesis of L3	S2
NMR spectra of ligands and complexes	S3
Cyclic Voltammograms	S13
Transient Absorption Spectroscopy	S14
Overview of fs-TAS results	S14
Global fit	S14
Computational details	S16
HOMO-LUMO Plots	S16
Computation of Geometry and Spin density of the ³ MCLT and ³ MC state	S17
Cartesian coordinates of the optimized structures	S18
References	S62

Synthesis of ligands

Ligand **L1** was prepared according to ref 1

Synthesis of **L2**

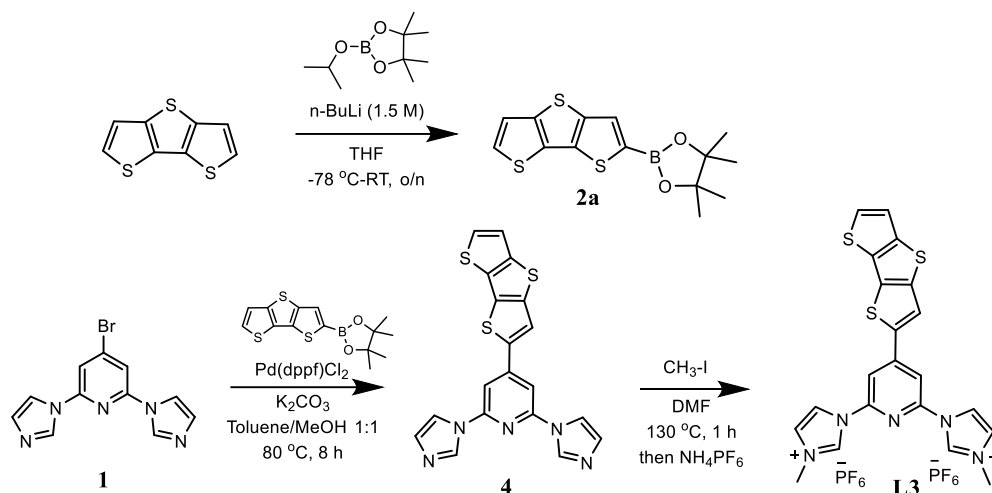


4,4,5,5-tetramethyl-2-(thieno[3,2-b]thiophen-2-yl)-1,3,2-dioxaborolane (1a) was prepared according to a literature procedure.²

2,6-di(1H-imidazol-1-yl)-4-(thieno[3,2-b]thiophen-2-yl)pyridine (3). To a solution of **1** (0.160 g, 0.551 mmol) in toluene (5 mL) was added a solution of compound **1a** (0.220 g, 0.827 mmol) and K_2CO_3 (0.228 g, 1.65 mmol) in methanol (5 mL). The mixture was degassed with Argon for 20 min then $PdCl_2(dppf)$ (0.040 g, 0.055 mmol) was added, and the solution was refluxed at 80 °C for 12 h. After completion of reaction, the reaction mixture was filtered through celite powder and washed with dichloromethane. After evaporation of solvents, the residue was purified by column chromatography over silica gel using 3-6% methanol in dichloromethane as the eluent to give the product as a white solid (0.12 g, 62% yield). 1H NMR (400 MHz, $DMSO-d_6$, δ ppm): 8.82(s, 2H), 8.43(s, 1H), 8.21(s, 2H), 7.93(s, 2H), 7.86(d, J = 5.20 Hz, 1H), 7.56(d, J = 5.19 Hz, 1H), 7.18(s, 2H). ^{13}C NMR (100 MHz, CD_3CN , δ ppm): 149.0, 148.3, 141.2, 141.2, 140.0, 136.2, 131.6, 130.7, 121.4, 120.8, 117.5, 105.8. ESI-HRMS calcd for $C_{17}H_{12}N_5S_2$ m/z = 350.0529. Found: 350.0485.

1,1'-(4-(thieno[3,2-b]thiophen-2-yl)pyridine-2,6-diyl)bis(3-methyl-1H-imidazol-3-ium)hexafluorophosphate(V) (L2). Compound **3** (0.07 g, 0.200 mmol) was charged in a 10 mL round bottomed flask and dissolved in 1 mL of DMF. Then added methyl iodide (0.05 mL, 0.801 mmol) and heated the mixture at 130 °C for 1 h. Desired compound was precipitated upon addition of diethyl ether. Solid was collected by vacuum filtration and washed thrice with diethyl ether and dried under vacuum. Then the solid was dissolved in minimum amount of water (ca. 2 mL) and added saturated NH_4PF_6 solution and stirred until the product precipitated. Compound **3** was collected by vacuum filtration and washed 3 times with water and then diethyl ether and dried under vacuum (0.128 g, 95.5% yield). 1H NMR (400 MHz, CD_3CN , δ ppm): 9.55(s, 2H), 8.29(s, 2H), 8.27(s, 1H), 8.06(s, 2H), 7.8(d, J = 5.3 Hz, 1H), 7.66(s, 2H), 7.52(d, J = 5.22 Hz, 1H), 4.05(s, 6H). ^{13}C NMR (100 MHz, $DMSO-d_6$, δ ppm): 149.7, 146.6, 142.4, 140.2, 139.8, 136.8, 132.99, 125.4, 122.7, 121.1, 119.7, 109.6, 37.1. ESI-HRMS calcd for $C_{19}H_{17}N_5S_2$ m/z = 189.5457. Found: 189.5486 [$M-2PF_6$].

Synthesis of L3



2-(dithieno[3,2-b:2',3'-d]thiophen-2-yl)-4,4,5,5-tetramethyl-1,3,2-dioxaborolane (2a) was prepared according to a literature procedure.³

4-(dithieno[3,2-b:2',3'-d]thiophen-2-yl)-2,6-di(1H-imidazol-1-yl)pyridine (4). To a solution of **1** (0.160 g, 0.551 mmol) in toluene (5 mL) was added a solution of compound **2a** (0.213 g, 0.661 mmol) and K_2CO_3 (0.228 g, 1.65 mmol) in methanol (5 mL). The mixture was degassed with Argon for 20 min then $PdCl_2(dppf)$ (0.040 g, 0.055 mmol) was added, and the solution was refluxed at 80 °C for 12 h. After completion of reaction, the reaction mixture was filtered through Celite powder and washed with dichloromethane. After evaporation of solvents, the residue was purified by column chromatography over silica gel using 5-7% methanol in dichloromethane as the eluent to give the product as yellow solid (0.157 g, 70 % yield). 1H NMR (400 MHz, $DMSO-d_6$, δ ppm): 8.84(s, 2H), 8.54(s, 1H), 8.24(s, 2H), 8.00(s, 2H), 7.86(d, J = 5.23 Hz, 1H), 7.62(d, J = 5.22 Hz, 1H), 7.19(s, 2H). ESI-HRMS calcd for $C_{19}H_{12}N_5S_3$ m/z = 406.0249. Found: 406.0248. The poor solubility prevented us from obtaining good quality ^{13}C spectra.

1,1'-(4-(dithieno[3,2-b:2',3'-d]thiophen-2-yl)pyridine-2,6-diyl)bis(3-methyl-1H-imidazol-3-ium) hexafluorophosphate(V) (L3). Compound **4** (0.100 g, 0.246 mmol) was charged in a 10 mL round bottomed flask and dissolved in 1 mL of DMF. Then added methyl iodide (0.077 mL, 1.23 mmol) and heated the mixture at 130 °C for 1 h. Desired compound was precipitated upon addition of diethyl ether. Solid was collected by vacuum filtration and washed thrice with diethyl ether and dried under vacuum. Then the solid was dissolved in minimum amount of water (ca. 3 mL) and added saturated NH_4PF_6 solution and stirred until the product precipitated. Compound **3** was collected by vacuum filtration and washed 3 times with water and then diethyl ether and dried under vacuum (0.160 g, 89.5% yield). 1H NMR (400 MHz, $DMSO-d_6$, δ ppm): 10.36(s, 2H), 8.83(s, 2H), 8.59(s, 1H), 8.45(s, 2H), 8.09(s, 2H), 7.92(d, 1H), 7.65(d, 1H), 4.06(s, 6H). ESI-HRMS calcd for $C_{21}H_{17}N_5S_3$ m/z = 217.5318. Found: 217.5324 [$M-2PF_6$]. The poor solubility prevented us from obtaining good quality ^{13}C spectra.

NMR spectra of ligands and complexes

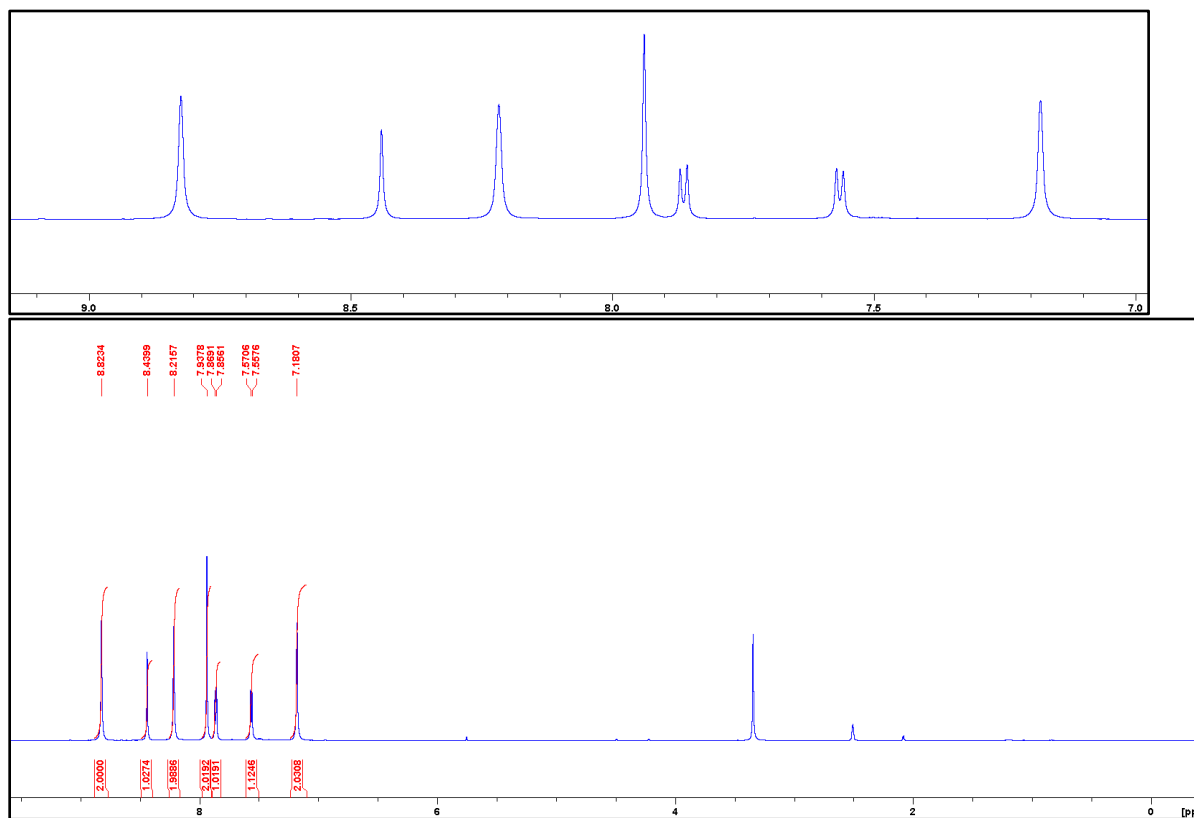


Figure S1. ^1H NMR spectra of compound 3

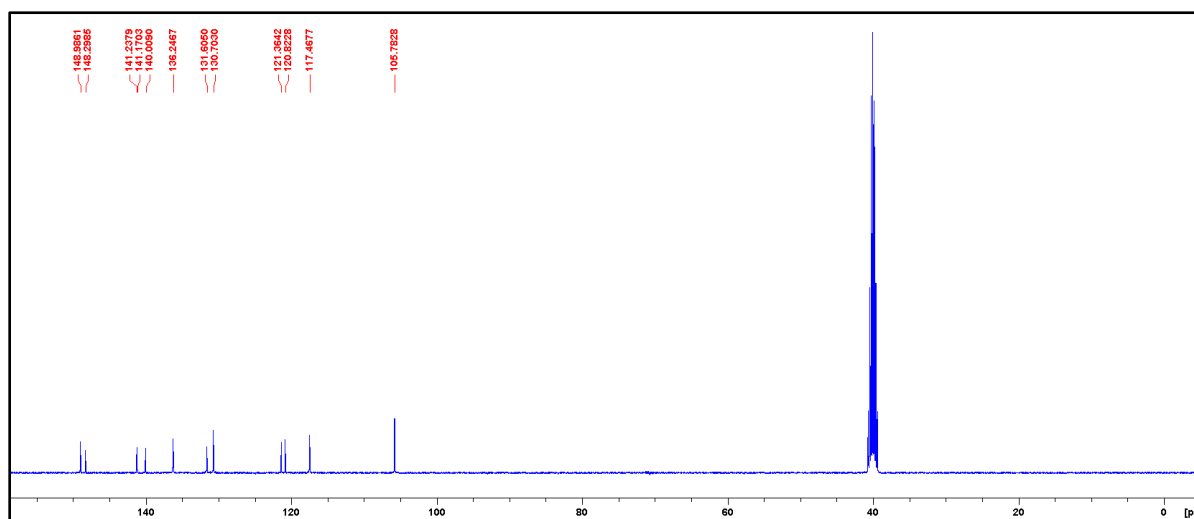


Figure S2. ^{13}C NMR spectra of compound 3.

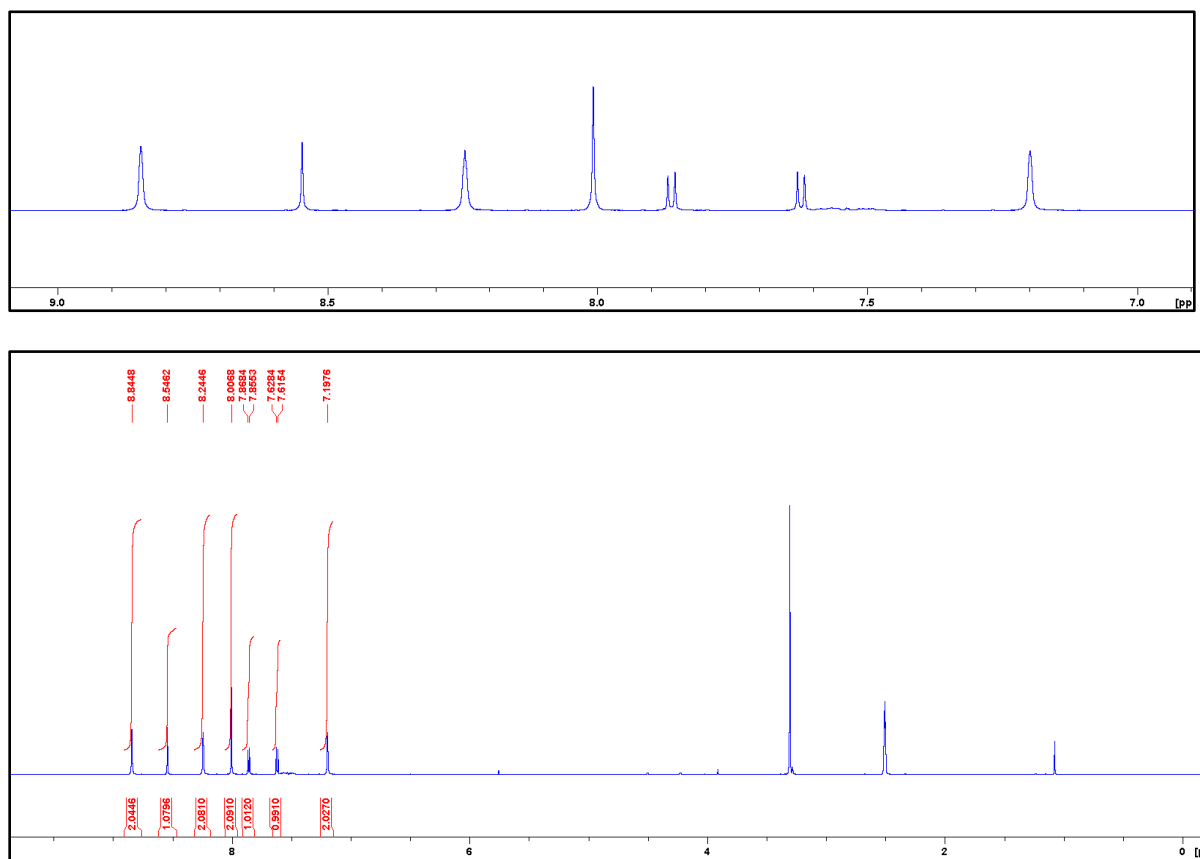


Figure S3. ^1H NMR spectra of compound 4.

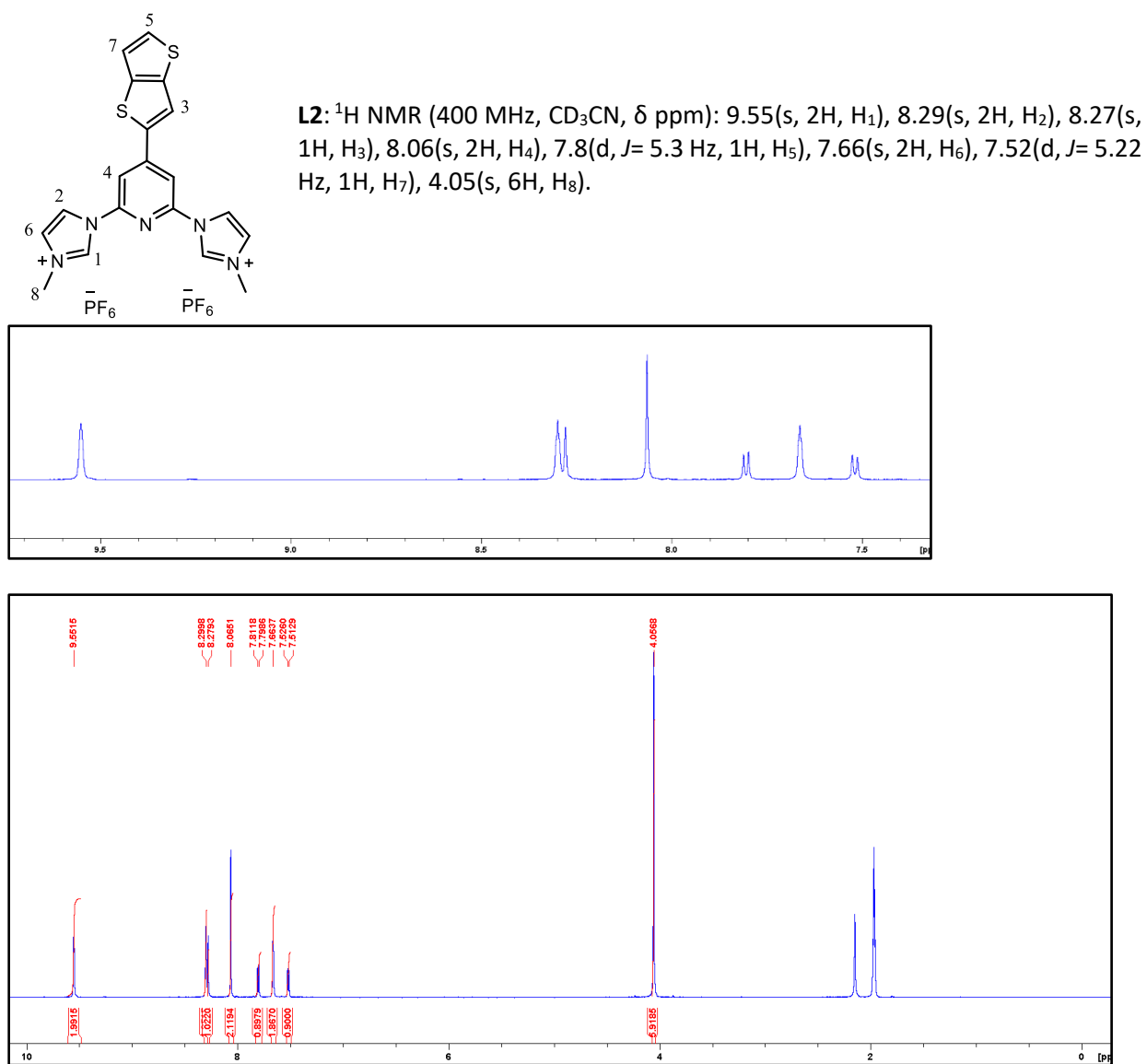


Figure S4. ^1H NMR spectra of **L2**.

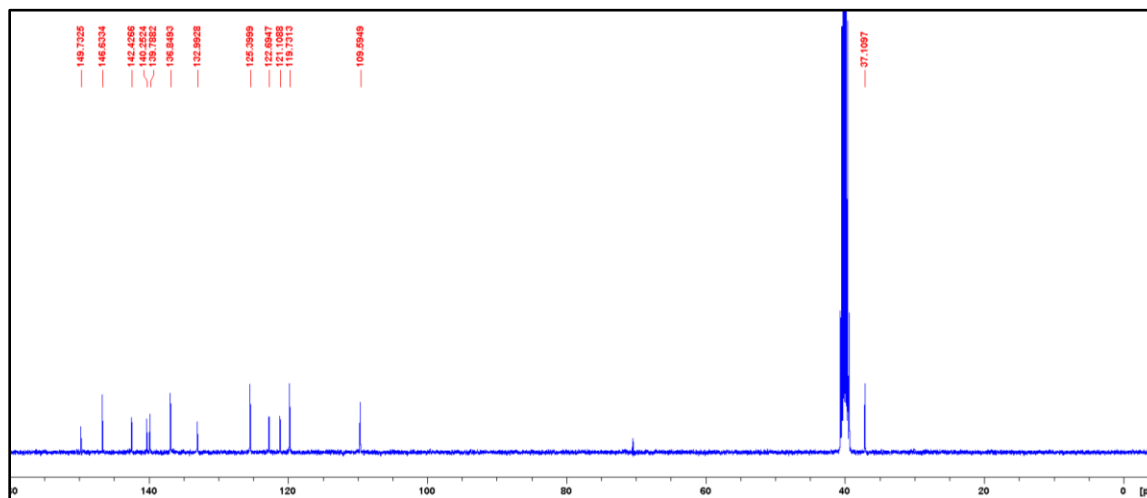
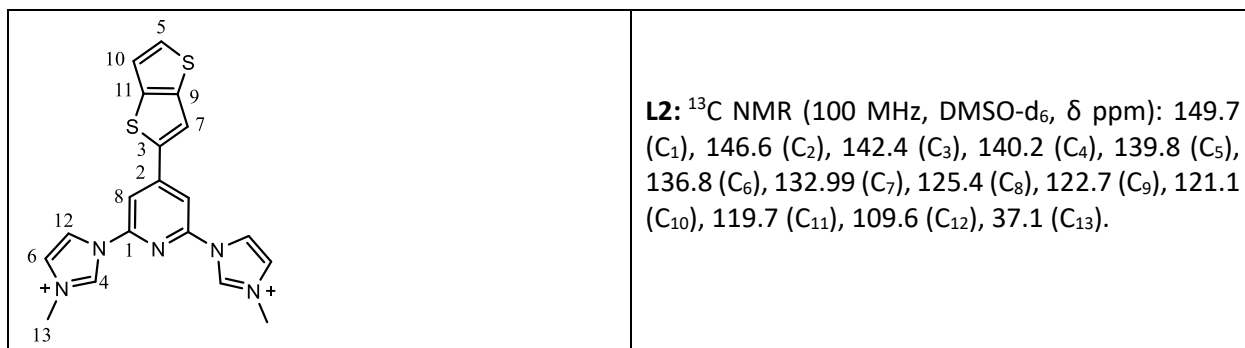
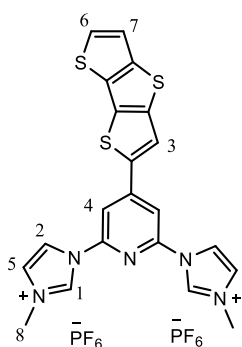


Figure S5. ^{13}C NMR spectra of **L2**.



L3: ^1H NMR (400 MHz, DMSO- d_6 , δ ppm): 10.36(s, 2H, H₁), 8.83(s, 2H, H₂), 8.59(s, 1H, H₃), 8.45(s, 2H, H₄), 8.09(s, 2H, H₅), 7.92(d, 1H, H₆), 7.65(d, 1H, H₇), 4.06(s, 6H, H₈).

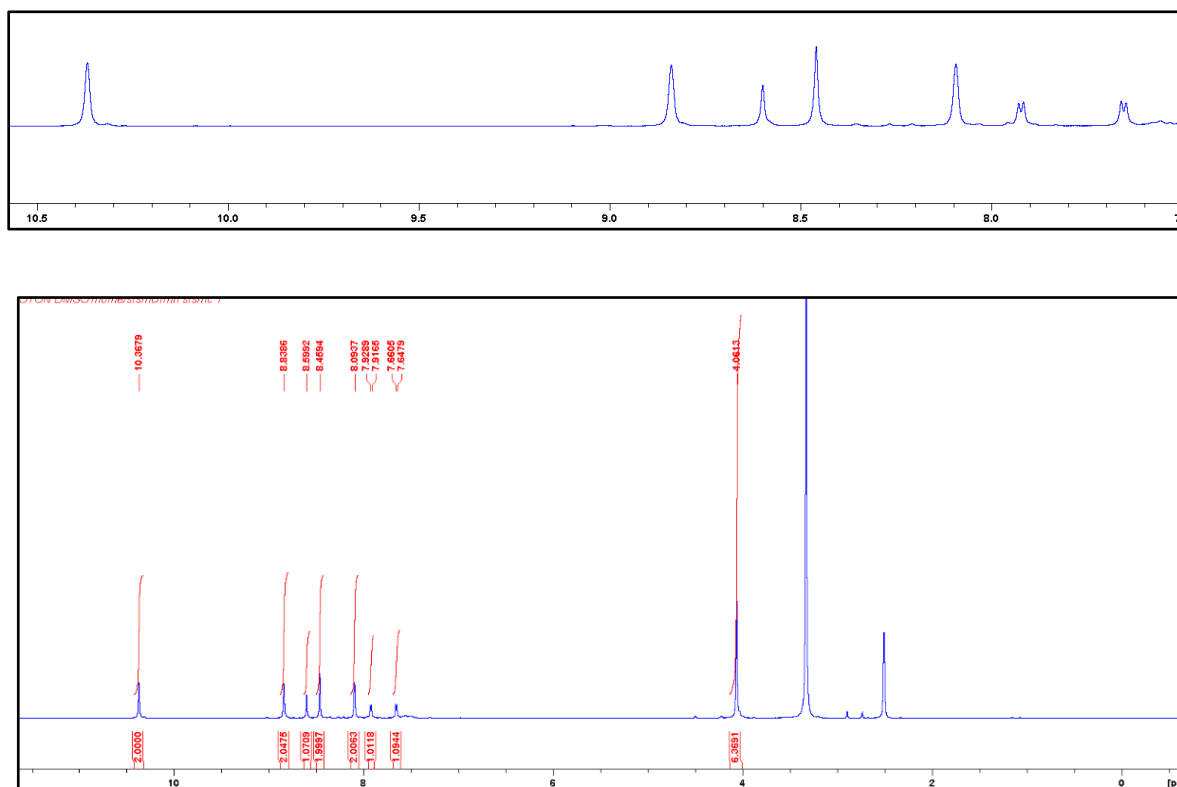
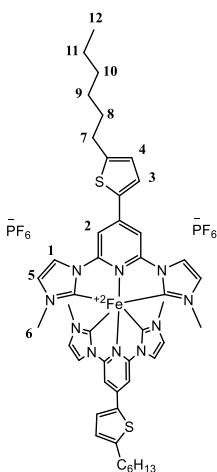


Figure S6. ^1H NMR spectra of compound **L3**.



CTh: ^1H NMR (400 MHz, CD_3CN , δ ppm): 8.12 (d, J = 2.19 Hz, 2H, H_1), 7.95 (s, 2H, H_2), 7.84 (d, J = 3.72 Hz, 1H, H_3), 7.06 (s, J = 3.72 Hz, 1H, H_4), 7.04 (d, J = 2.19 Hz, 2H, H_5), 2.99 (t, J = 7.51 Hz, 2H, H_6), 2.62 (s, 6H, H_7), 1.79 (q, 2H, H_8), 1.48-1.35 (m, 6H, $\text{H}_{9,10,11}$), 0.95 (t, J = 7.09 Hz, 3H, H_{12}).

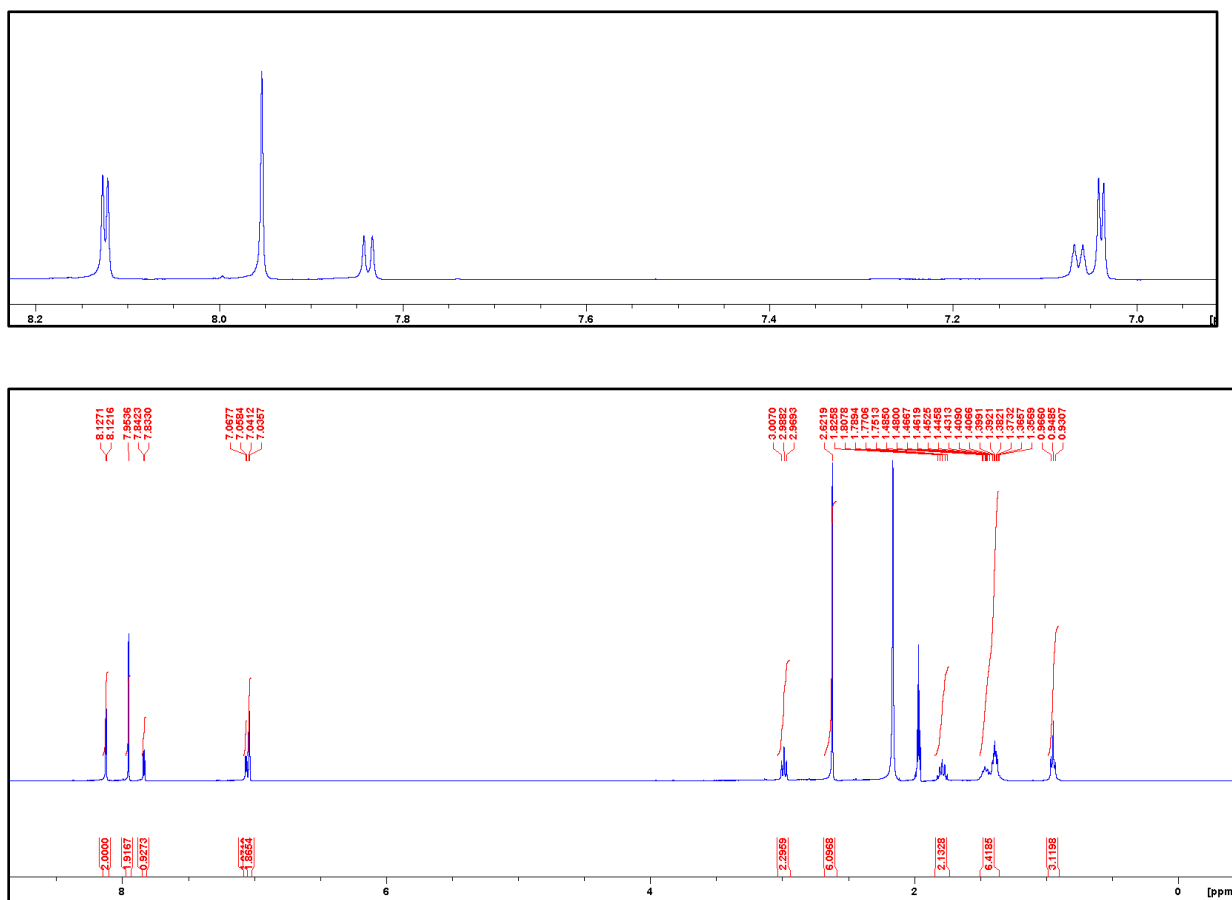
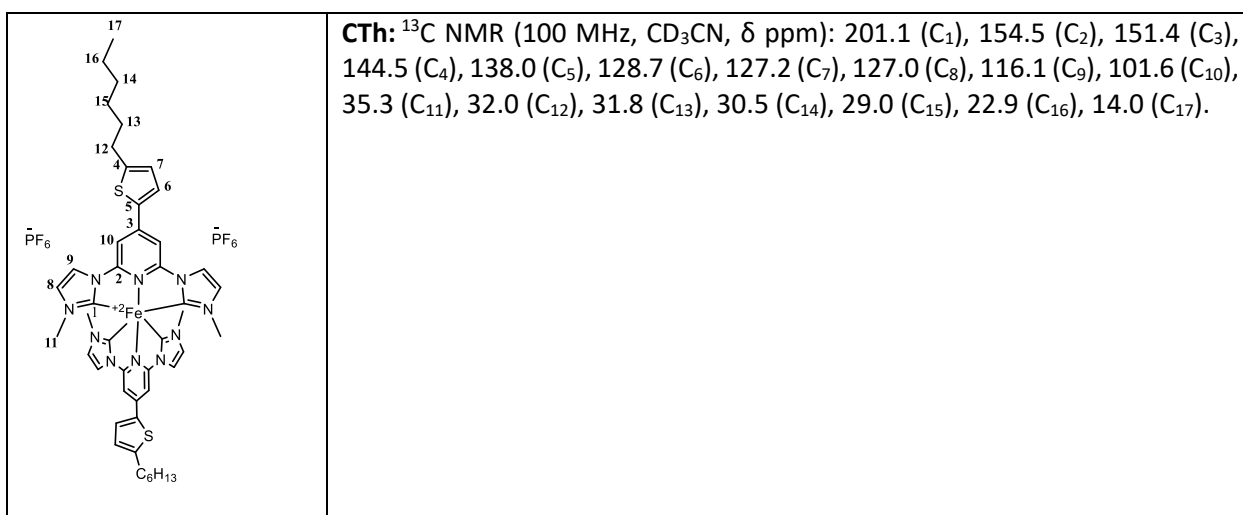


Figure S7. ^1H NMR spectra of **CTh**



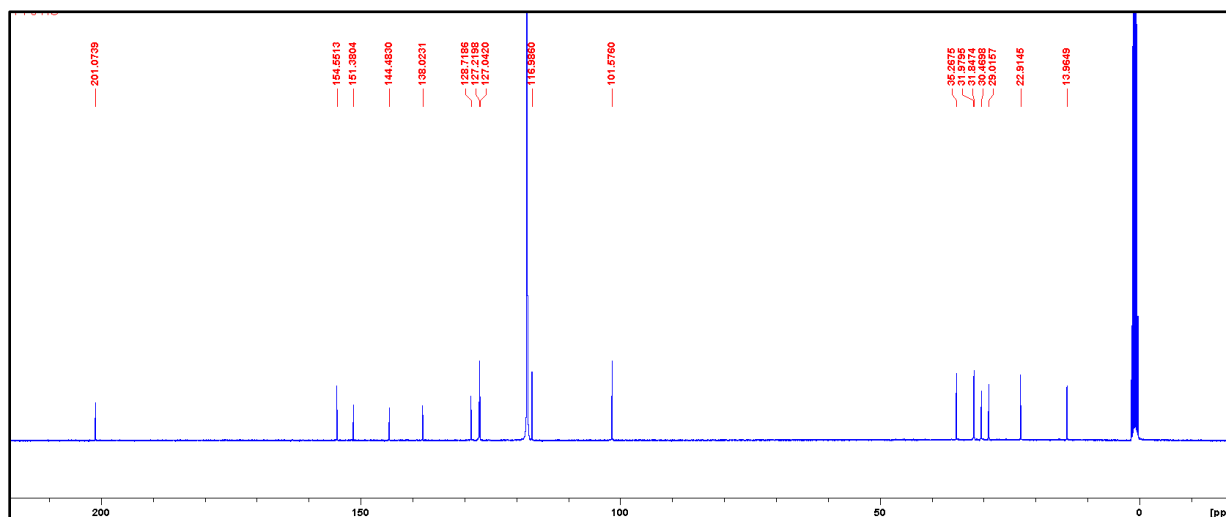
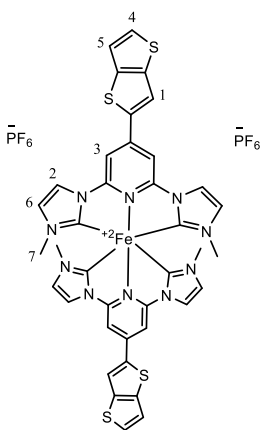
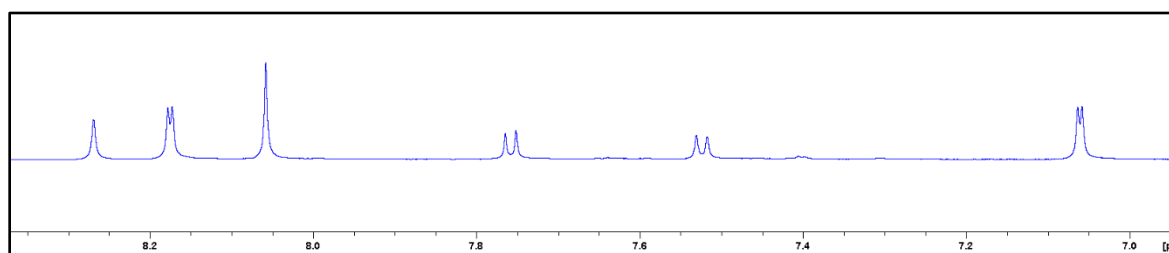


Figure S8. ^{13}C NMR spectra of **CTh**



CTh2: ^1H NMR (400 MHz, CD_3CN , δ ppm): 8.26 (s, 1H, H_1), 8.17(d, $J = 2.10$ Hz, 2H, H_2), 8.05(s, 2H, H_3), 7.76(d, $J = 5.30$ Hz, 1H, H_4), 7.52(d, $J = 5.28$ Hz, 1H, H_5), 7.05(d, $J = 2.10$ Hz, 2H, H_6), 2.64(s, 6H, H_7).



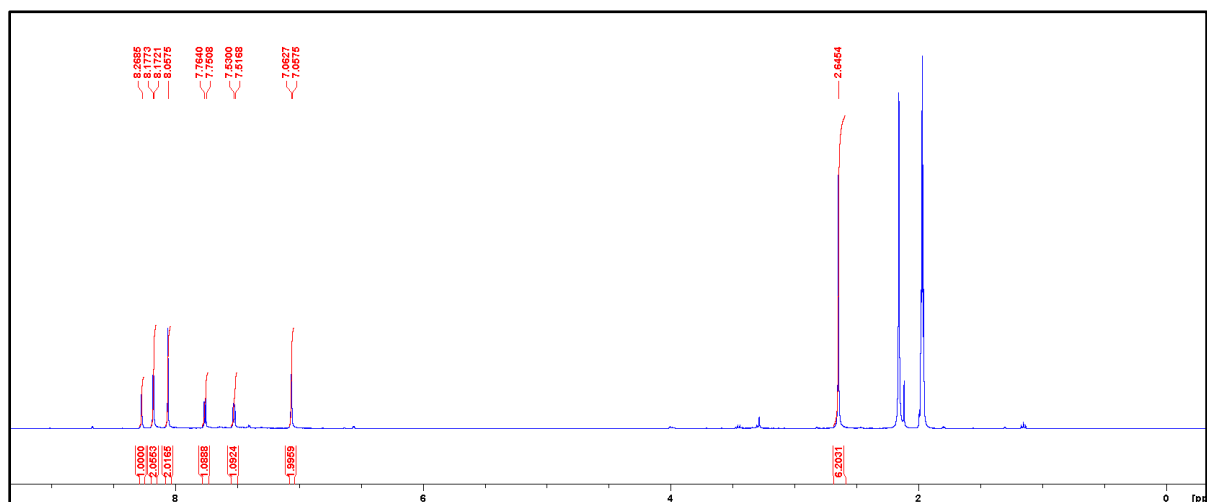
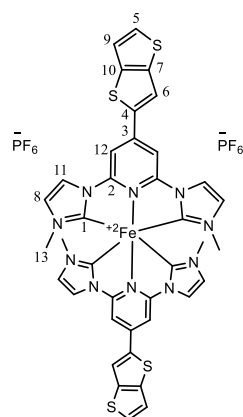


Figure S9. ¹H NMR spectra of CTh2



CTh2: ¹³C NMR (100 MHz, CD₃CN, δ ppm): 200.8 (C₁), 154.6 (C₂), 144.6 (C₃), 142.2 (C₄), 141.7 (C₅), 141.0 (C₆), 131.7 (C₇), 127.1 (C₈), 120.9 (C₉), 120.7 (C₁₀), 117.1 (C₁₁), 101.9 (C₁₂), 35.3 (C₁₃).

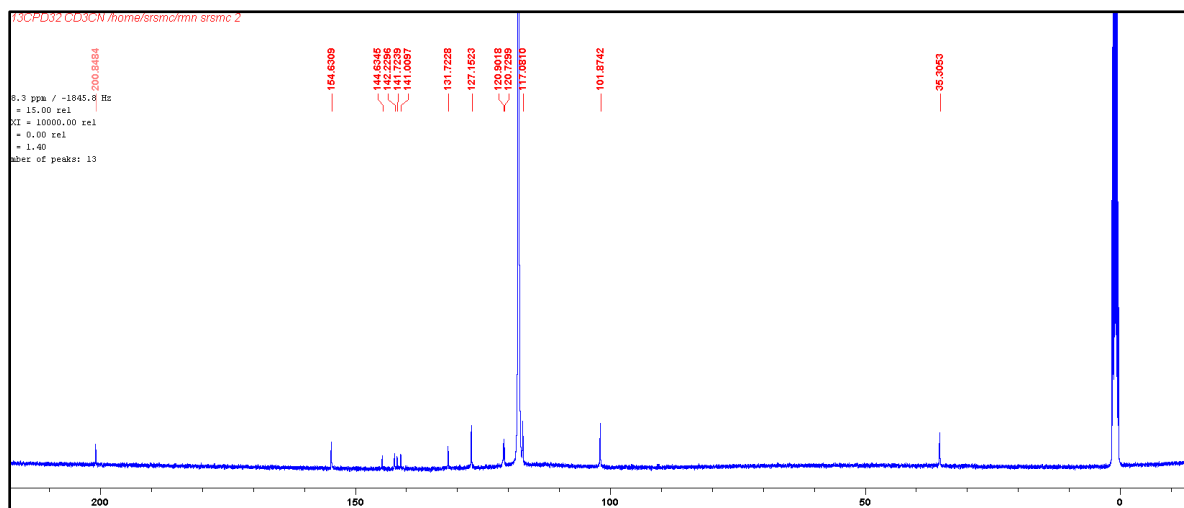
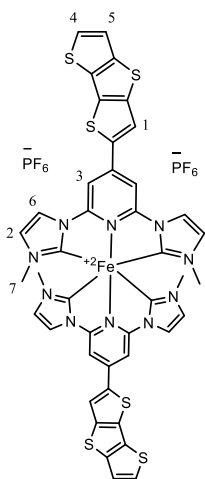


Figure S10. ¹³C NMR spectra of CTh2



CTh3: ^1H NMR (400 MHz, CD_3CN , δ ppm): 8.30 (s, 1H, H_1), 8.17 (d, $J = 2.12$ Hz, 2H, H_2), 8.06 (s, 2H, H_3), 7.71 (d, $J = 5.29$ Hz, 1H, H_4), 7.53 (d, $J = 5.30$ Hz, 1H, H_5), 7.06 (d, $J = 2.09$ Hz, 2H, H_6), 2.65 (s, 6H, H_7).

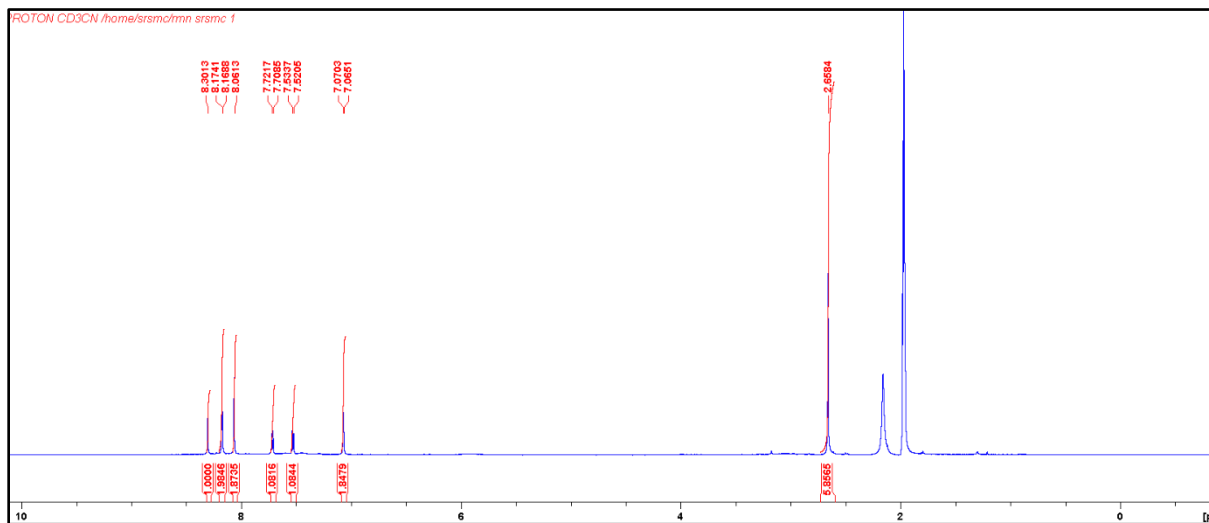
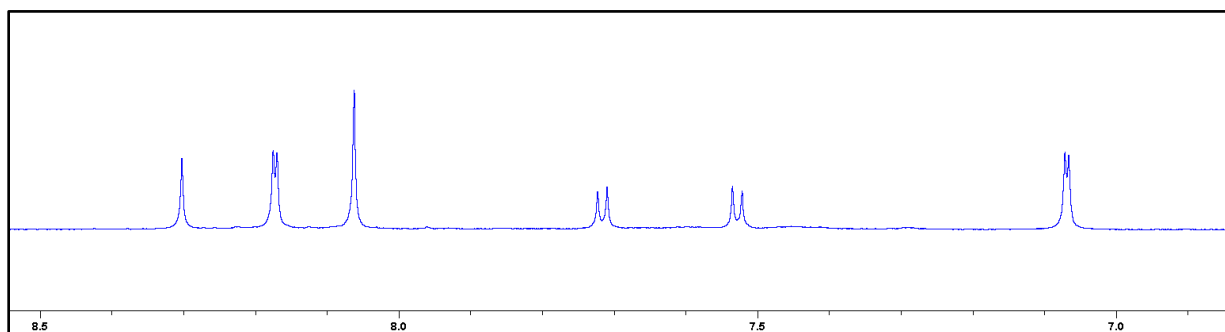


Figure S11. ^1H NMR spectra of CTh3

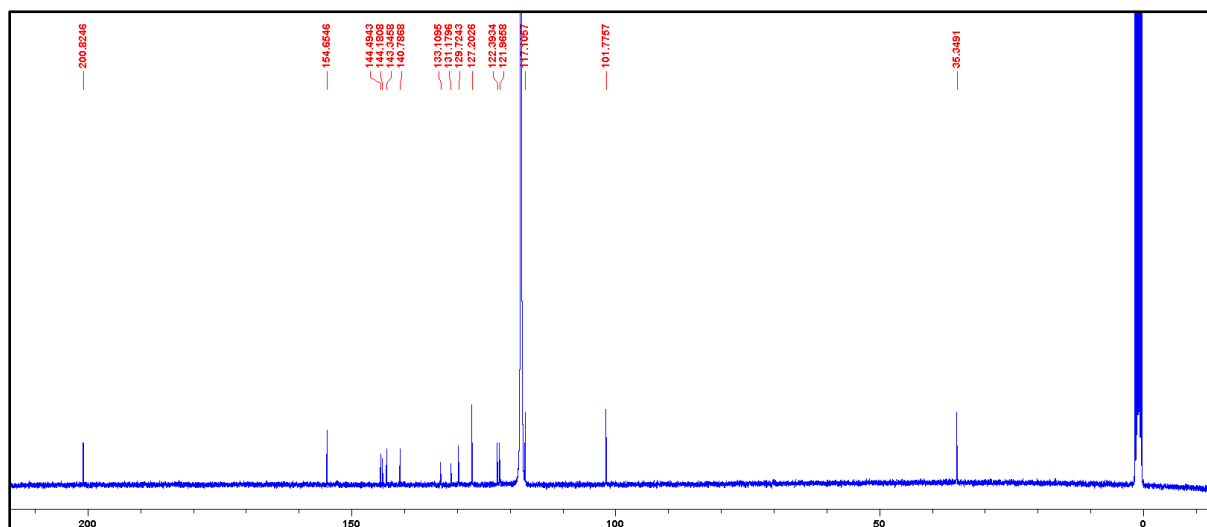
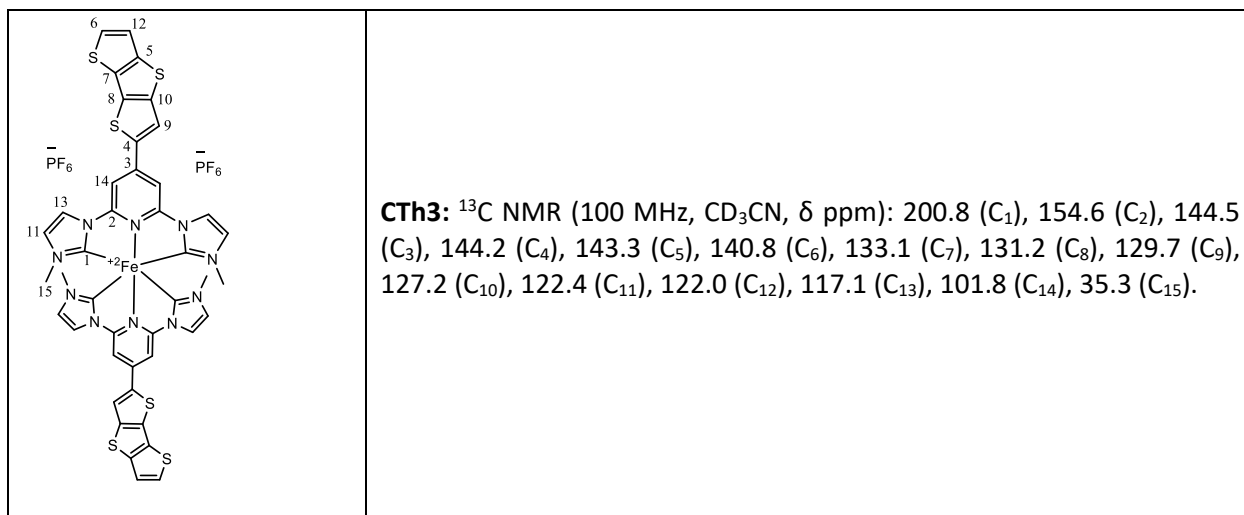


Figure S12. ^{13}C NMR spectra of **CTh3**.

Cyclic Voltammograms

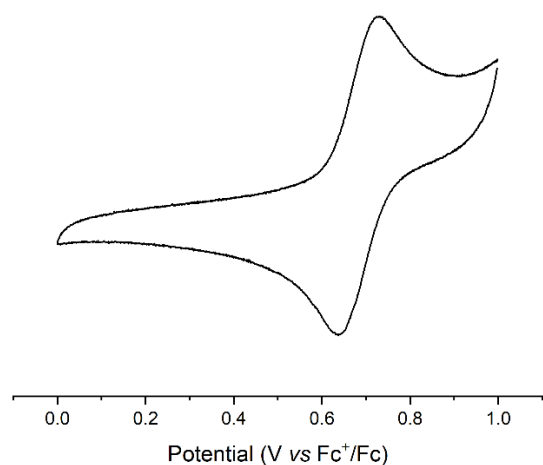


Figure S13. Cyclic Voltammetry of complex CTh1

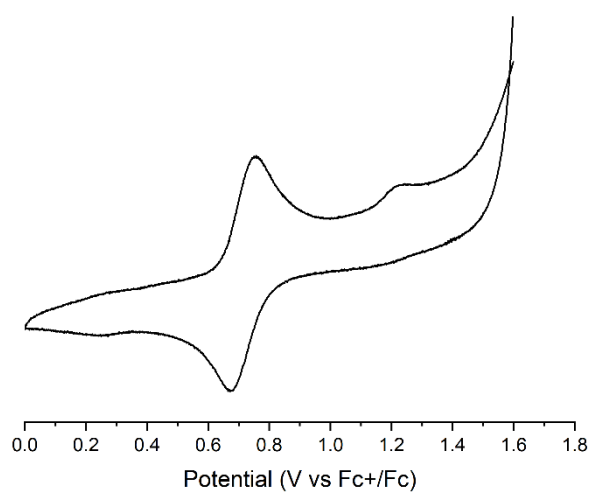
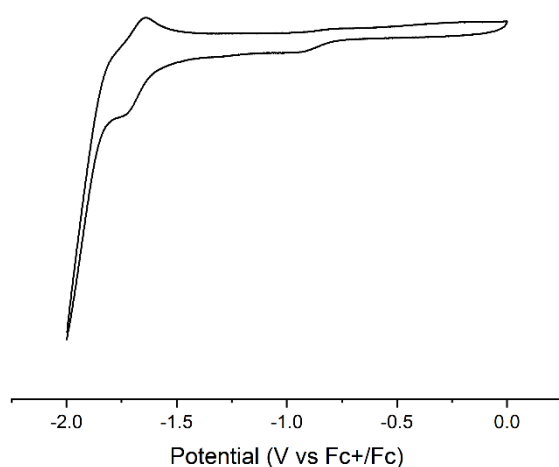


Figure S14. Cyclic Voltammetry of complex CTh2

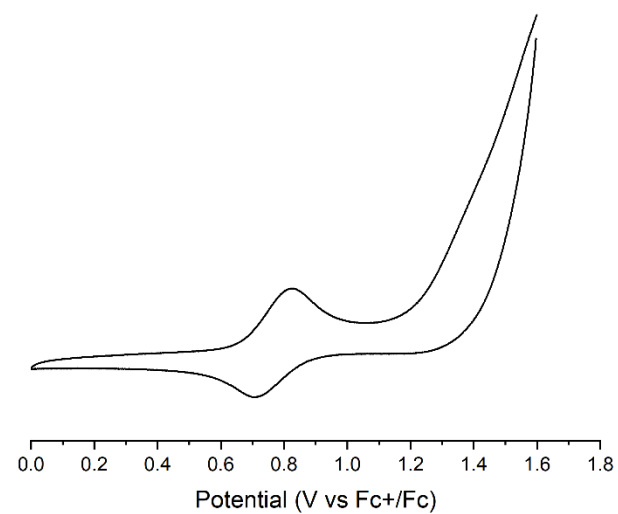
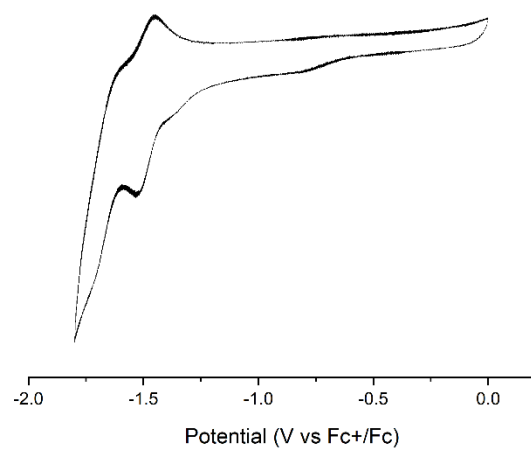
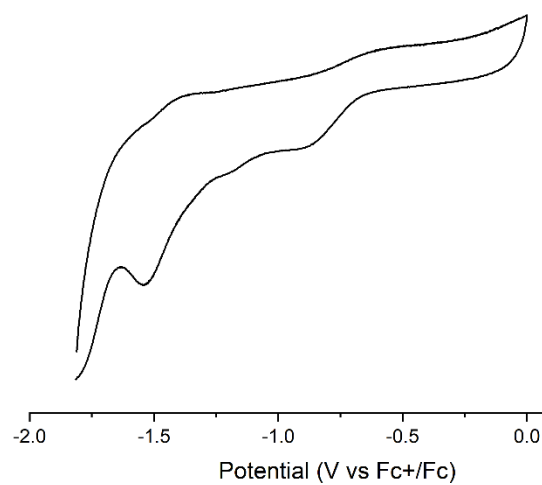


Figure S15. Cyclic Voltammetry of complex CTh3



Transient Absorption Spectroscopy

Overview of fs-TAS results

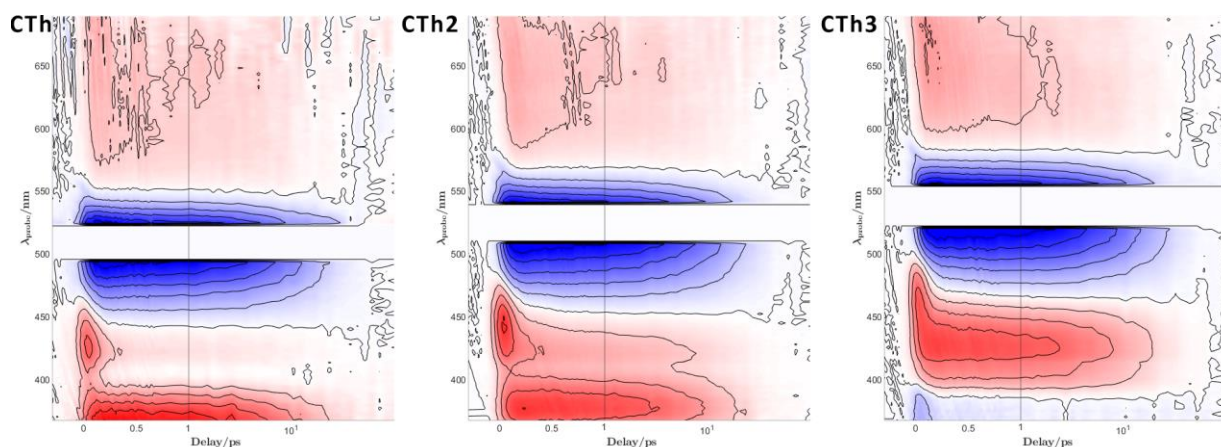


Figure S16. Two-dimensional contour plots of the fs-TAS data of the **CTh** series of molecules. Negative (bleach) signals are in shades of blue, whereas positive ones (induced absorption) are in red. Contour lines are drawn at 10 % increments of maximum absolute signal. Note the delay axis has a break at 1 ps and the scaling becomes logarithmic beyond that delay. The data around bleach maximum were omitted (white gap in 500-550 nm range) due to large noise of the scattered pump light.

Global fit

As described in the main text, all three datasets were subject to a global analysis employing a sequential relaxation scenario, which corresponds to a unidirectional first-order kinetic scheme in conjunction with a convolution with a Gaussian instrument response function (IRF, $\sigma=50$ -60 fs). The reported time constants correspond to the respective inverse rate coefficients. The minimal number of species/relaxation steps needed to adequately describe the experimental data was decided based on (1) the inspection of the singular values of the data matrix (Fig. S16) and (2) by ensuring that the residuals do not present any systematic features indicative of a missing component. Indeed, as can be seen in Fig. S17, the singular values appear to be almost constant as a function of the singular value index after three (**CTh**) and four (**CTh2** and **CTh3**). This suggests the first hint on the maximum number of components to include in the model.

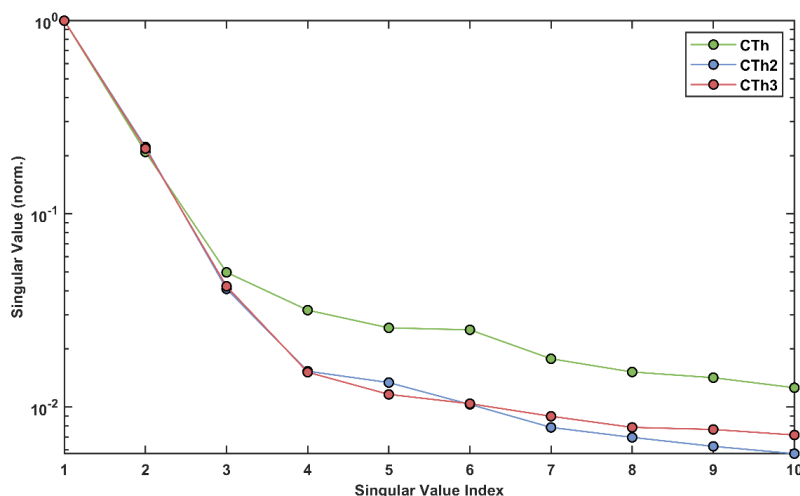


Figure S17. Normalized sorted singular values obtained by the singular value decomposition of the data matrices

We have tried the global sequential analysis with both three and four relaxation steps (+ a so-called coherent artifact contribution having the IRF temporal profile) and it turned out that actually 4 components are necessary for all three datasets **CTh**, **CTh2** and **CTh3**.

Figure S18 shows typical examples that highlight that the global fit with three exponentials does not capture properly the dynamics observed in the data in the 1-10 ps range. The additional time constant introduced by the four-step relaxation model falls in this range (3-5 ps). Note, however, that when fitted with 4 components, the CTh data yield an unphysically short time value of < 10fs for τ_1 .

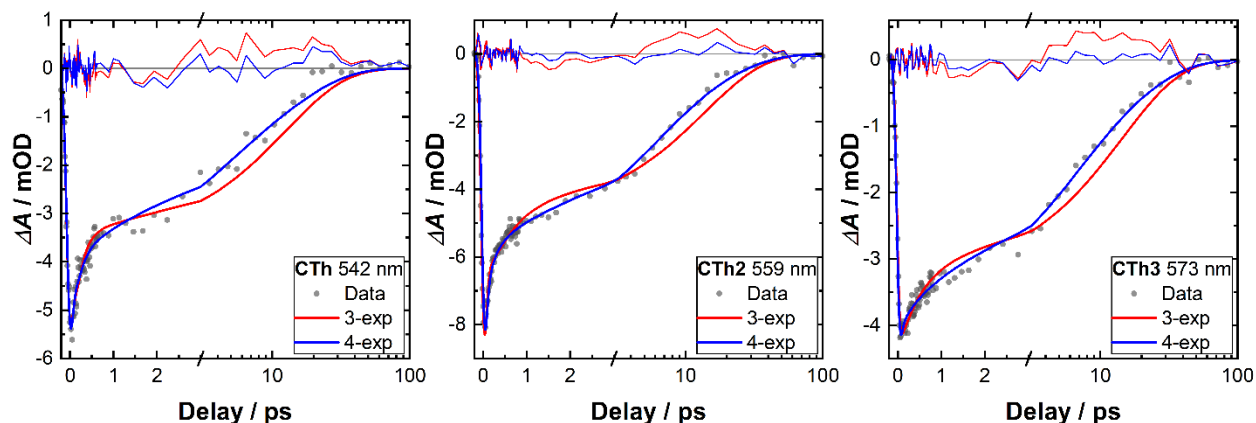


Figure S18. Examples of the results of the global sequential fit with three (red) and four (blue) exponentials at selected wavelengths. Residuals are drawn with thin lines and data are shown as dots.

As mentioned in the main text, given the similarity of the three datasets the evolution associated difference spectra (EADS) also show high resemblance (Fig. S19). Namely, for **CTh2** and **CTh3**, the first one accounts for the ultrafast redshift of the ESA overlapping the main bleach feature and the delayed rise of the latter. This component is attributed to the initial MLCT singlet to triplet conversion. The second sub-picosecond component captures the residual red shift of the ESA features (see e.g. the zero-crossing line for the red and blue curves in the 450-500 nm range) and intensity decrease. This feature can be assigned

to vibrational relaxation of a non-thermalized MLCT triplet state. The third and fourth species (blue and green traces in Figure S19) show almost identical spectral shapes and differ mostly in intensities. We tentatively assign the third step (~ 5 ps relaxation time constant) to residual slower thermalization process which involves vibrational modes that are less strongly coupled to the optical transitions and which therefore leads to little spectral variation. The last, ~ 15 ps, step corresponds to the return to the ground state in accordance with the relaxation cascade observed for the parent pyridylNHC Fe(II) complexes.⁴

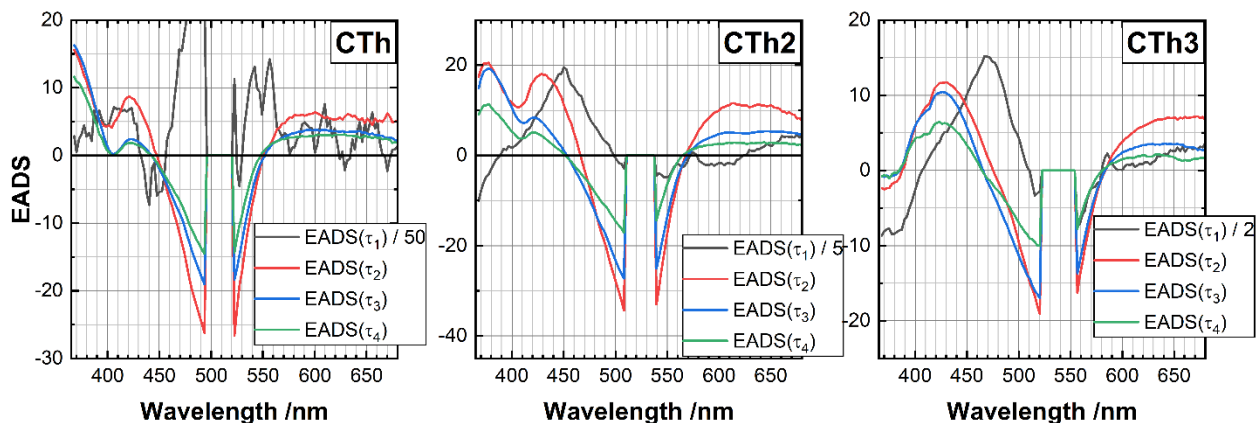
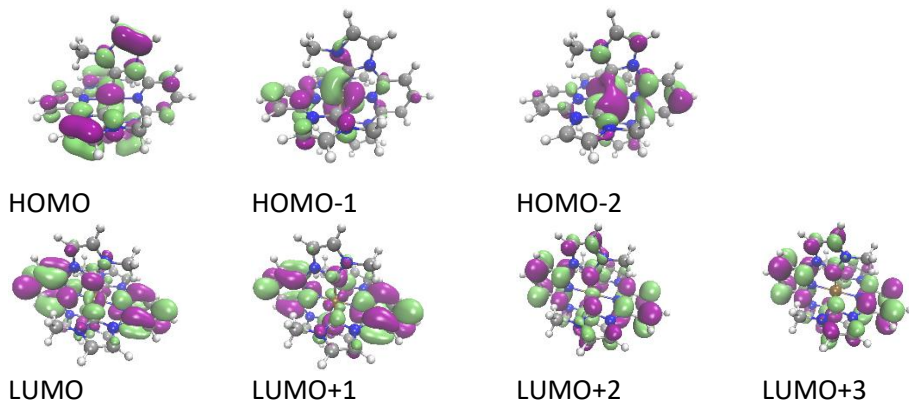


Figure S19. Evolution associated difference spectra obtained by the global sequential analysis with four kinetic components

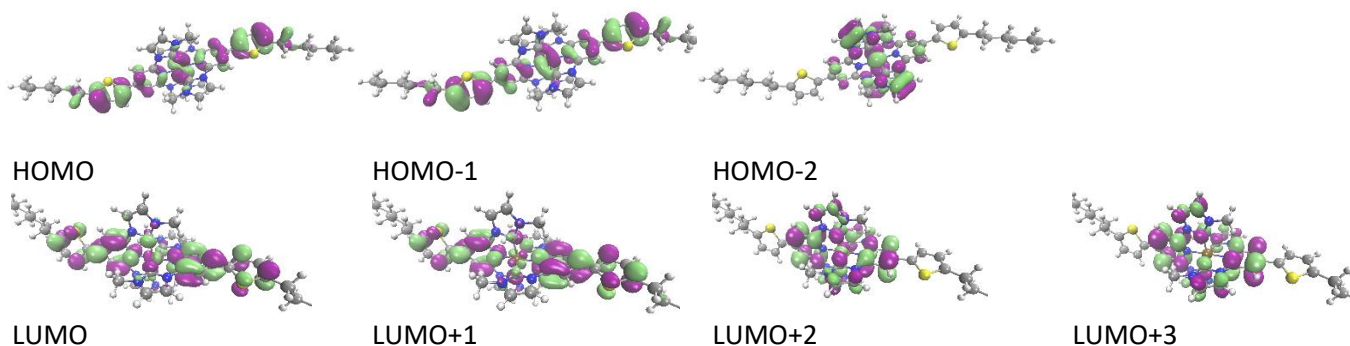
Computational details

HOMO-LUMO Plots

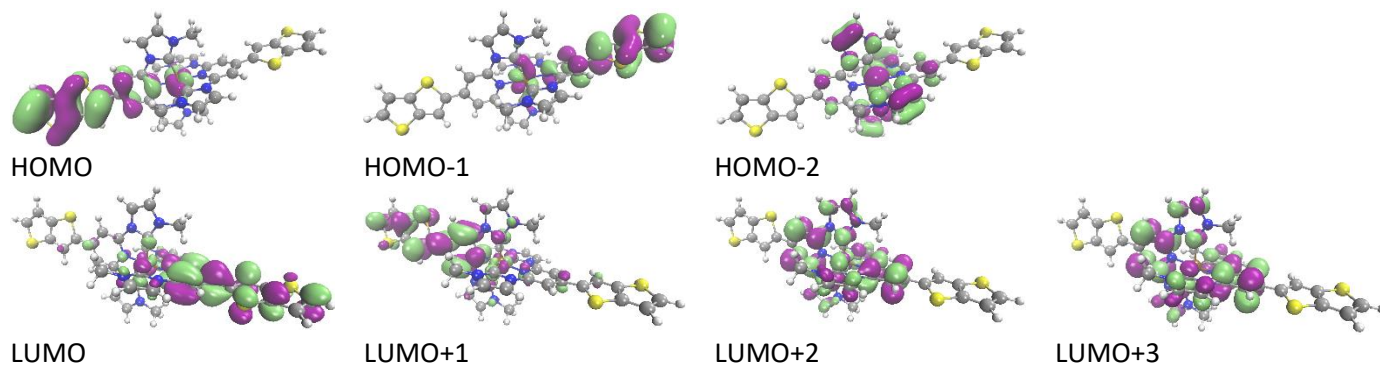
C0



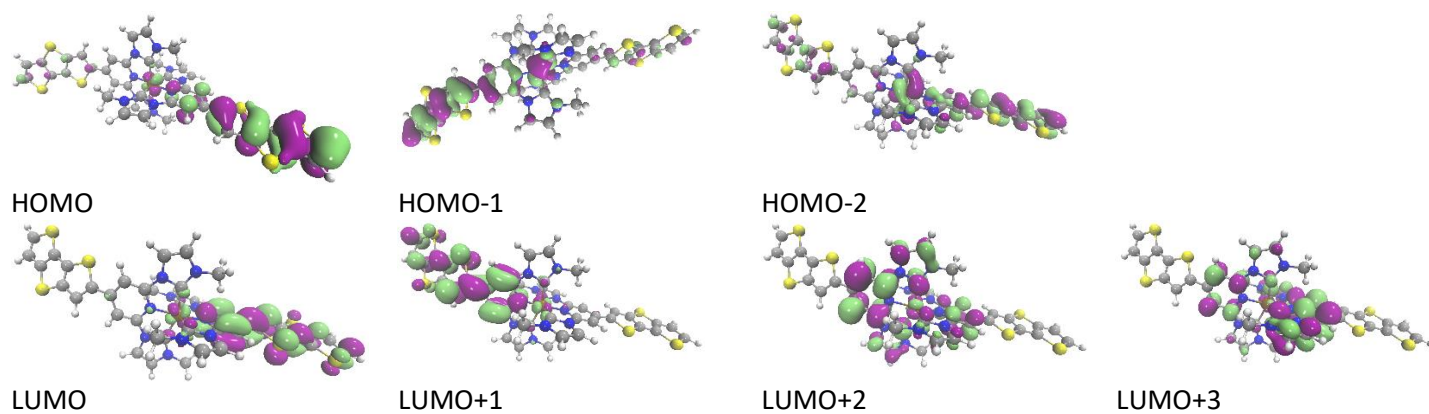
CTh



CTh2



CTh3

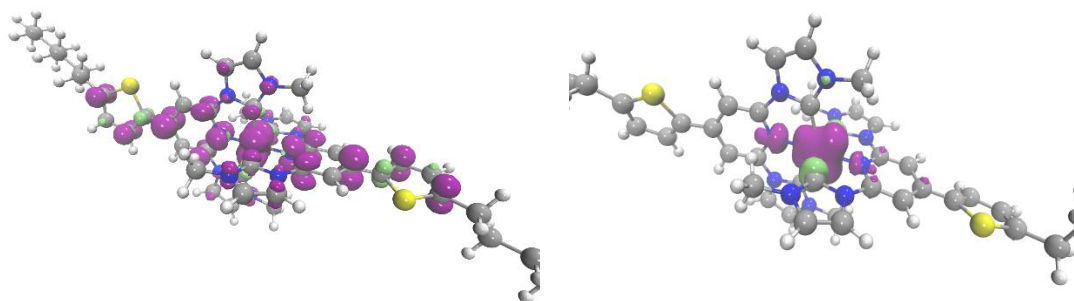


Computation of Geometry and Spin density of the $^3\text{MCLT}$ and ^3MC state

Complex
CTh

$^3\text{MLCT}$

^3MC



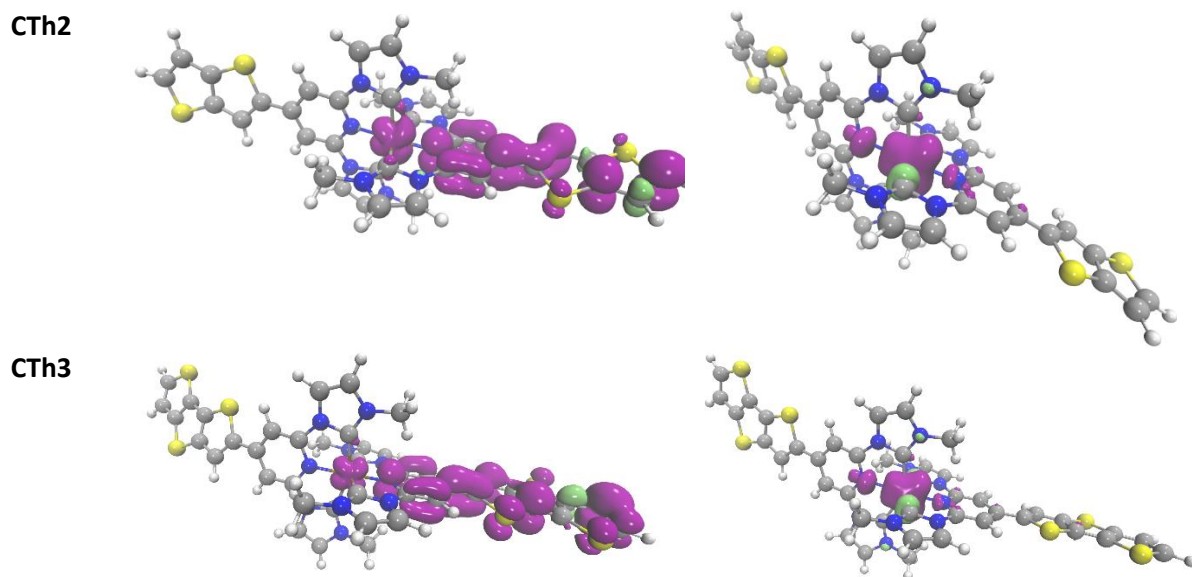


Figure S20. Geometry and Spin density of the $^3\text{MLCT}$ and ^3MC states of **CTh**, **CTh2** and **CTh3**.

Cartesian coordinates of the optimized structures

Ground state

C0

H	34.74433487615443	29.31776025755730	29.98286176114617
H	38.13051714840139	30.23438856687641	36.56875980411194
C	34.45576208538451	30.07176363702653	30.71389387142452
H	35.62211238883432	29.41617865641120	37.40638086471000
C	37.14060549232064	30.55246919856928	36.24471481930389
C	35.90952882590647	30.15094299996903	36.65557848850518
H	39.73055321502411	31.57498529074572	35.22034389451099
C	39.19538826283620	32.23753965918849	34.53692540818298
H	40.98374909720335	33.09218374293785	33.67346833911478
C	33.22408468214005	30.49023192960887	31.10563091349656
H	32.23475859095891	30.17065774772610	30.78126747339493
C	39.89044614040615	33.09030142123482	33.66827768547441
C	37.79995816801343	32.27152125733563	34.49249136337930

C	39.20086922254798	33.94052109424448	32.79291108756505
N	37.12661972248434	33.08526816101215	33.65476438481037
H	39.74022419040217	34.60490444369594	32.11457829292281
H	30.63241791379465	31.52982529949935	32.10855274384785
C	37.80520372819571	33.90130177065905	32.82347703272950
C	32.56196797481957	32.24772068665145	32.81825791500496
C	31.16658533018128	32.21042197545115	32.77480540285875
N	33.23404252545831	33.08191897854764	33.63664569304039
H	38.14959834353760	35.93640237516838	30.74796334369868
C	30.47021308872404	33.08212789059878	33.62338219872841
C	37.15749233696585	35.61587172355247	31.06273611011013
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N	36.176254	31.471384	32.651701
N	33.870942	35.403995	37.078932
C	41.103209	35.138669	34.798629
C	41.795588	35.990449	33.934139
C	43.194200	35.893789	34.085080

C	43.591429	34.960750	35.068615
S	42.224923	34.211034	35.813229
H	41.306611	36.653997	33.217096
S	44.575526	36.664162	33.312886
C	29.365399	30.427038	34.596878
C	28.916376	29.469489	33.679855
C	27.619467	29.013352	33.968370
C	27.055956	29.600425	35.126094
S	28.143395	30.737007	35.846924
H	29.506851	29.123806	32.827436
S	26.531062	27.862116	33.206294
C	45.669685	35.720928	34.312967
C	44.994983	34.862354	35.197912
S	46.115102	33.945204	36.160541
C	25.333476	28.168139	34.452884
C	25.758486	29.116918	35.404586
C	24.029317	27.661952	34.702194
C	47.086958	35.638564	34.413172
C	47.460200	34.720099	35.369081
H	48.475355	34.443695	35.665279
H	47.794777	36.219166	33.817837
S	24.537957	29.378137	36.619055
C	23.490655	28.228707	35.836001

H 22.507235 28.030903 36.270125

H 23.500991 26.933767 34.083845

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