

## Supporting Information

### “Turn-on” Fluorescence Sensor for Organic Amines Fabricated via

#### Sustainable Processing

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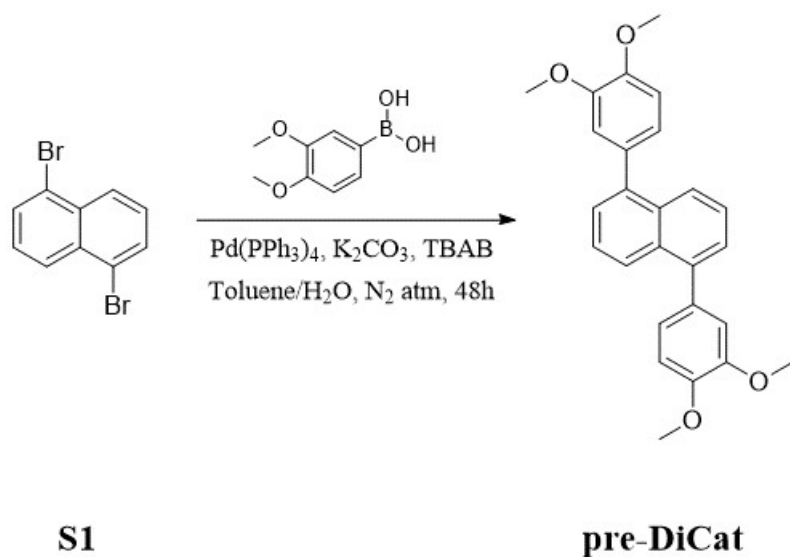
## **General information**

All the reagents were purchased and used directly without further purification. Thin layer chromatography (TLC) was performed on EMD preloaded plated (silica gel 60 F254) and visualized with hand-held UV lamp at 254 or 365 nm.

<sup>1</sup>H-NMR (400 MHz) and <sup>13</sup>C-NMR (101 MHz) spectra were measured on Bruker spectrometer. Chemical shifts were expressed in parts per million (ppm) with respect to internal standard of TMS. Coupling constants were reported as Hertz (Hz), signal shapes and splitting patterns were indicated as follows: s, singlet; d, doublet; t, triplet; q, quartet; m, multiplet. Mass spectra data were collected on MALDI-TOF MS Performance (Shimadzu, Japan).

IR spectra were recorded with Perkin Elmer (Spectrum 100) FT-IR Spectrometer with attenuated total reflectance (ATR) accessory. UV-visible spectra were collected on Perkin Elmer (Lambda 750) UV/Vis/NIR Spectrometer using THF or THF/H<sub>2</sub>O solution in quartz cuvette. Fluorescence spectra were recorded by SHIMADZU Spectro fluorophotometer RF-6000 using THF or THF/H<sub>2</sub>O solution in quartz cuvette at specific excitation wavelength.

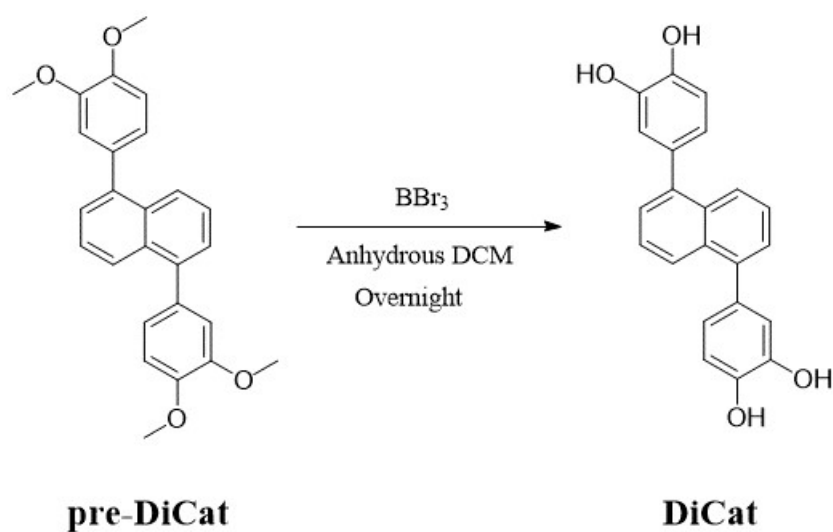
## Synthesis of pre-DiCat



**Figure S1** Synthetic procedure for **pre-DiCat**

Round-bottom flask equipped with a magnetic stirrer was charged with 1,5-dibromonaphthalene (2.86g, 10 mmol), (3,4-dimethoxyphenyl)boronic acid (4.00 g, 22 mmol), anhydrous  $K_2CO_3$  (6.91 g, 50 mmol), TBAB (665 mg, 2.0 mmol),  $Pd(PPh_3)_4$  (116 mg, 0.10 mmol). The flask was evacuated and filled with  $N_2$  for three times before the addition of 80 mL toluene and 25 mL DI  $H_2O$ . The reaction mixture was vigorously stirred at 90 cent. for 48h and then cooled to room temperature before post-treatment. Adding DI  $H_2O$  into reaction mixture gradually and extracting with EA for three times. The combined organic layer was washed with brine and dried with anhydrous  $Na_2SO_4$  before being concentrated under reduced pressure to afford the crude product. It was further purified by washing with EA, PE and DCM for several times to give 1,5-bis(3,4-dimethoxyphenyl)naphthalene (**pre-DiCat**) (3.60 g, 90.0%) as white powder.  $^1H-NMR$  (400 MHz,  $DMSO-d_6$ )  $\delta$  = 7.91 (d,  $J$  = 8.3 Hz, 2H), 7.56 (t,  $J$  = 7.6 Hz, 2H), 7.48 (d,  $J$  = 6.9 Hz, 2H), 7.16 (d,  $J$  = 8.2 Hz, 2H), 7.09 (s, 2H), 7.05 (d,  $J$  = 8.9 Hz, 2H), 3.89 (s, 6H), 3.84 (s, 6H).  $^{13}C-NMR$  (101 MHz,  $DMSO-d_6$ )  $\delta$  = 149.22, 148.91, 140.56, 133.44, 132.22, 127.34, 126.19, 125.59, 122.61, 114.30, 112.48, 56.26, 56.22. **MALDI-TOF-MS**  $m/z$  calculated for  $C_{26}H_{24}O_4$   $[M]^+$  400.2, found 400.2. **m.p.** 184-190 cent.

## Synthesis of DiCat



**Figure S2** Synthetic procedure for **DiCat**

A flame-dried round-bottom flask equipped with a magnetic stirrer was charged with **pre-DiCat** (4.00 g, 10 mmol). It was evacuated and filled with N<sub>2</sub> for three times before the addition of 100 mL extra dry DCM. BBr<sub>3</sub> (10.0 g, 40 mmol) was dissolved in extra dry DCM and added into the reaction mixture dropwise at -78 cent.. The solution was allowed to warm to R.T. and stirred overnight. DI H<sub>2</sub>O was gradually added to quench excess amount of BBr<sub>3</sub>. Reaction mixture was filtrated with Buchner funnel and washed with DI H<sub>2</sub>O several times to afford crude product. Transferring the filter residue into another flask and adding EA to disperse the impure sample. It was ultrasonicated for 5 minutes before centrifugation. This process was repeated for several times and white solid (2.84 g, 82.7%) was obtained after drying under vacuum oven for 24h. <sup>1</sup>H-NMR (400 MHz, acetone-*d*<sub>6</sub>) δ = 8.08 (s, 2H), 8.03 (s, 2H), 7.91 (d, J = 8.5 Hz, 2H), 7.49 – 7.40 (m, 2H), 7.36 (d, J = 6.7 Hz, 2H), 6.98 – 6.94 (m, 4H), 6.81 (dd, J = 8.0, 1.9 Hz, 2H). <sup>13</sup>C-NMR (101 MHz, acetone-*d*<sub>6</sub>) δ = 144.97, 144.74, 140.70, 132.87, 132.37, 126.58, 125.26, 121.62, 117.17, 115.27. **MALDI-TOF-MS** m/z calculated for C<sub>22</sub>H<sub>16</sub>O<sub>4</sub> [M]<sup>+</sup> 344.1, found 344.2. **m.p.** > 300 cent.

### NMR spectra

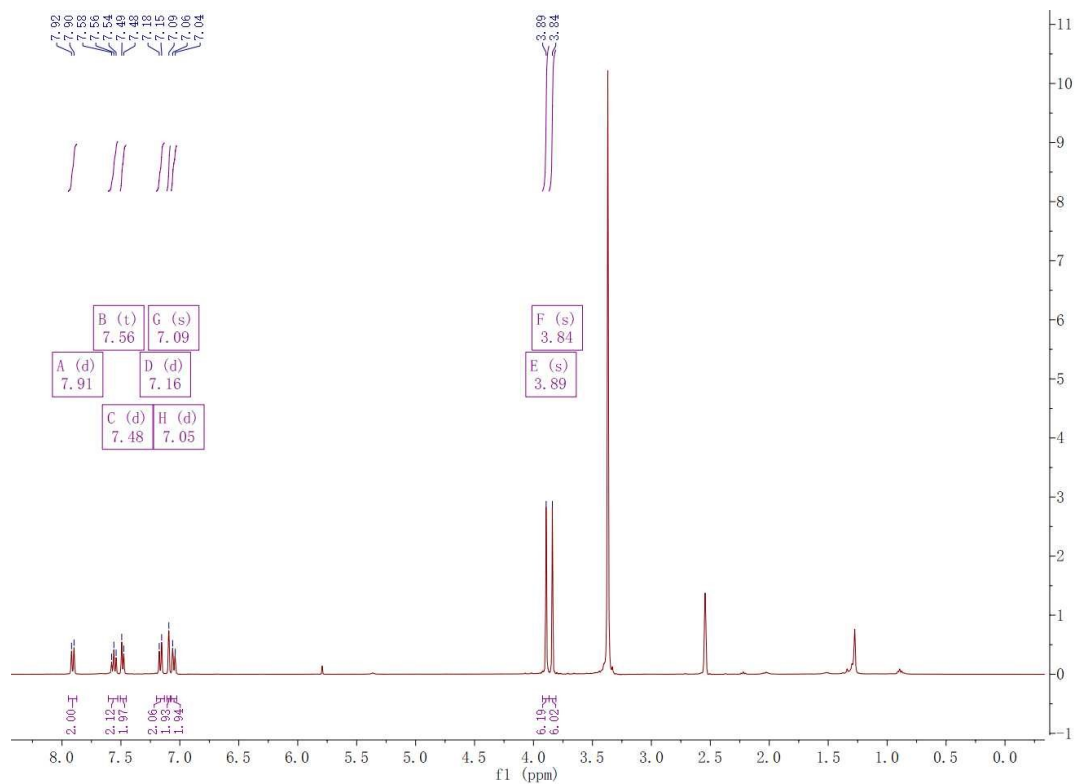


Figure S3 <sup>1</sup>H-NMR spectrum of pre-DiCat

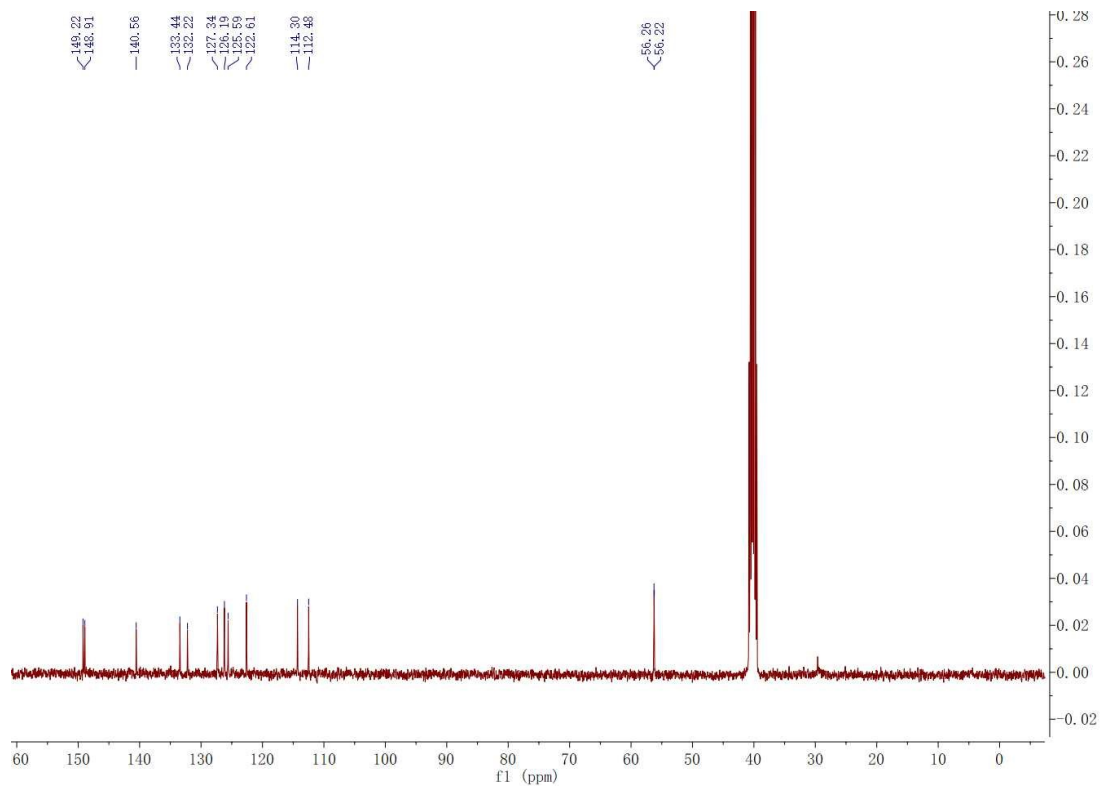
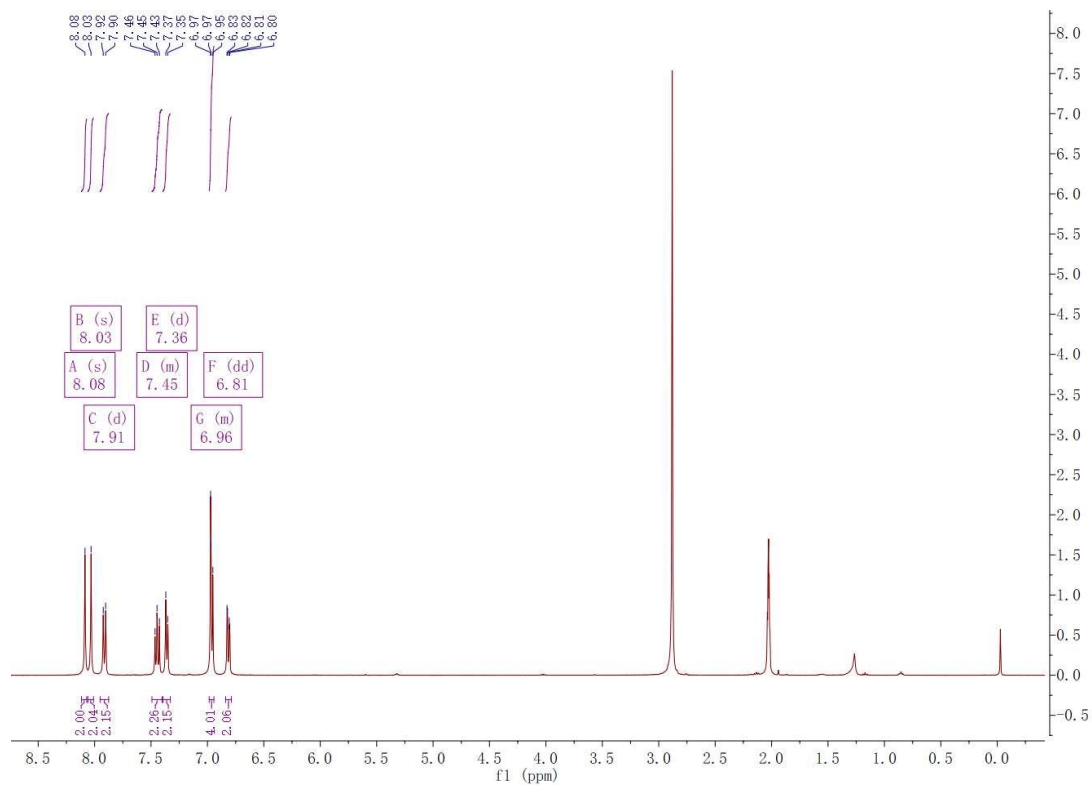
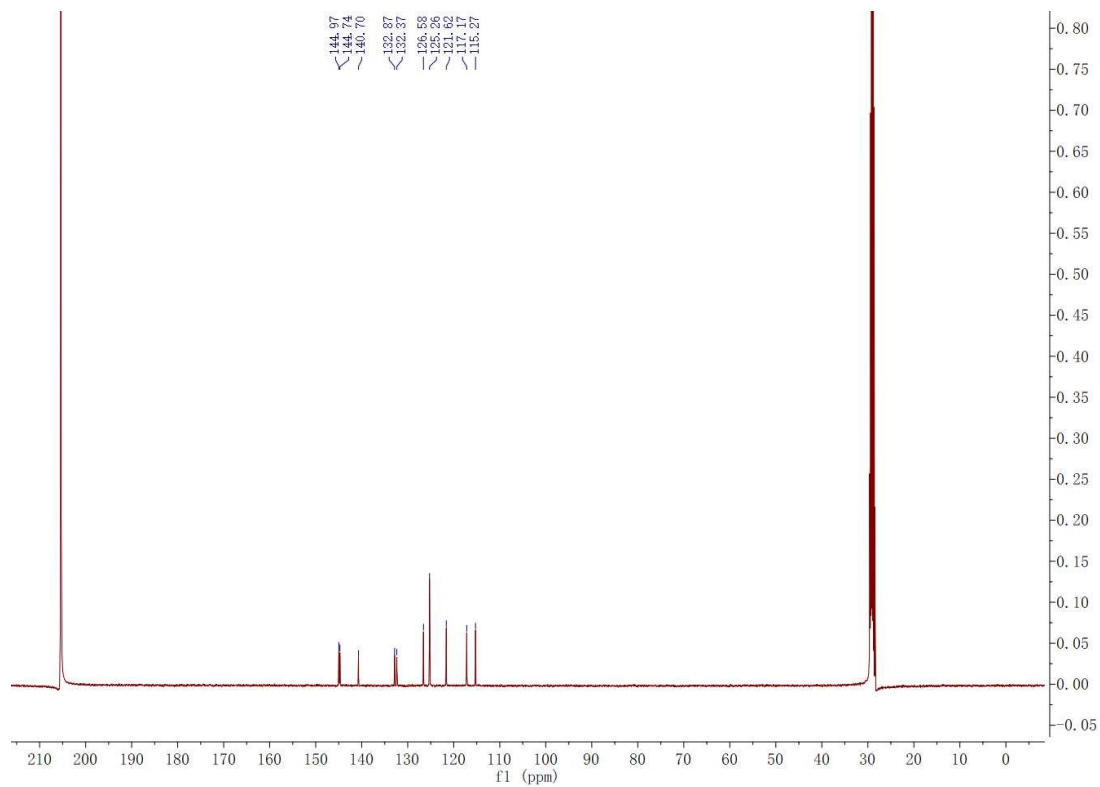


Figure S4 <sup>13</sup>C-NMR spectrum of pre-DiCat



**Figure S5  $^1\text{H-NMR}$  spectrum of DiCat**



**Figure S6  $^{13}\text{C-NMR}$  spectrum of DiCat**

## MALDI-TOF-MS

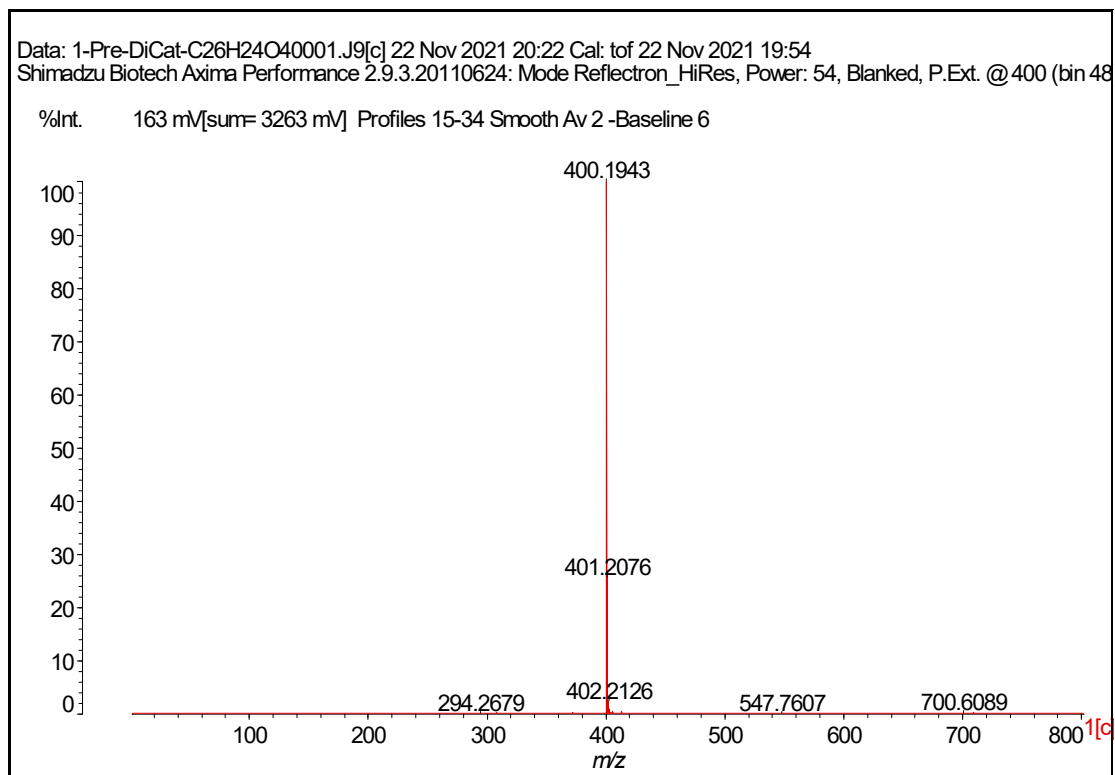


Figure S7 MALDI-TOF-MS record of pre-DiCat

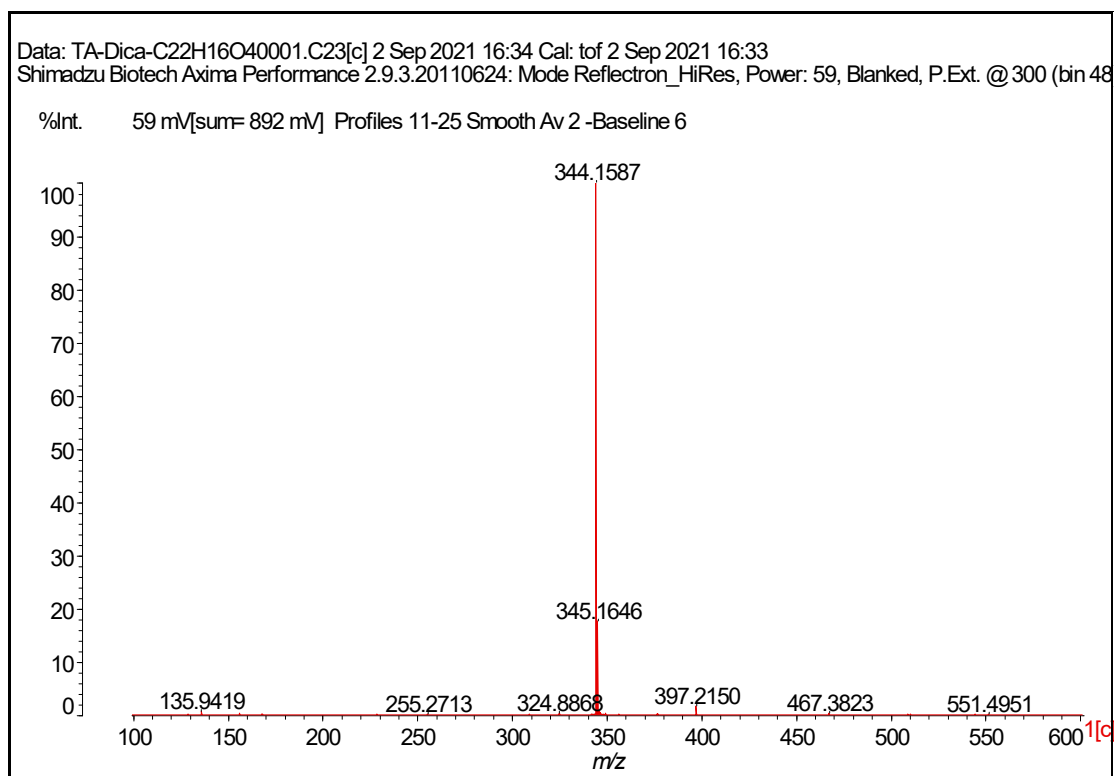
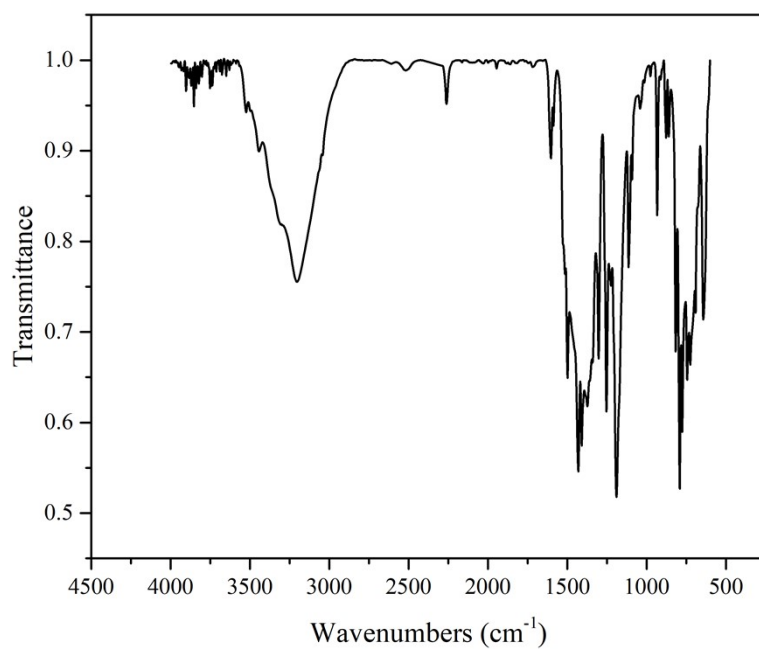


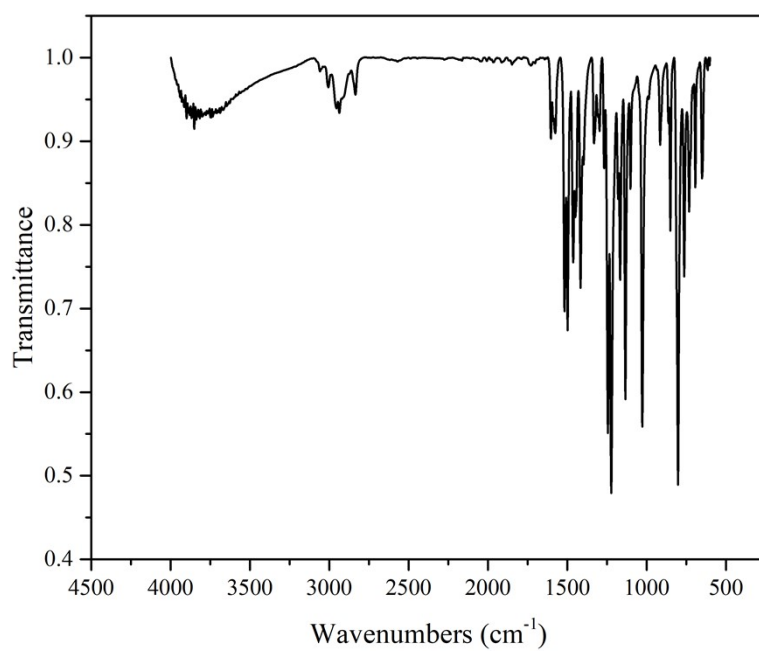
Figure S8 MALDI-TOF-MS record of DiCat



## FT-IR spectra

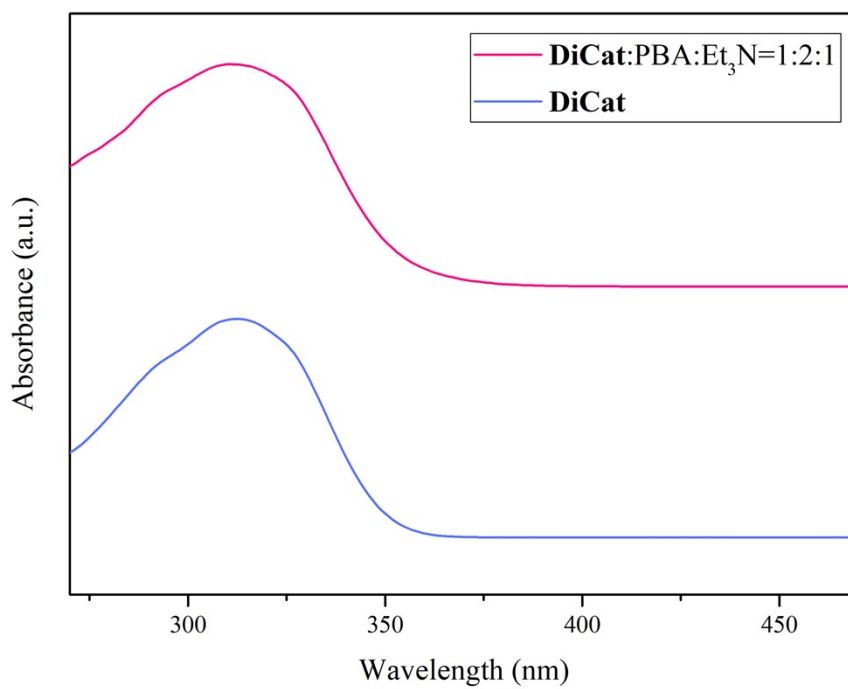


**Figure S9** FT-IR spectrum of **pre-DiCat**



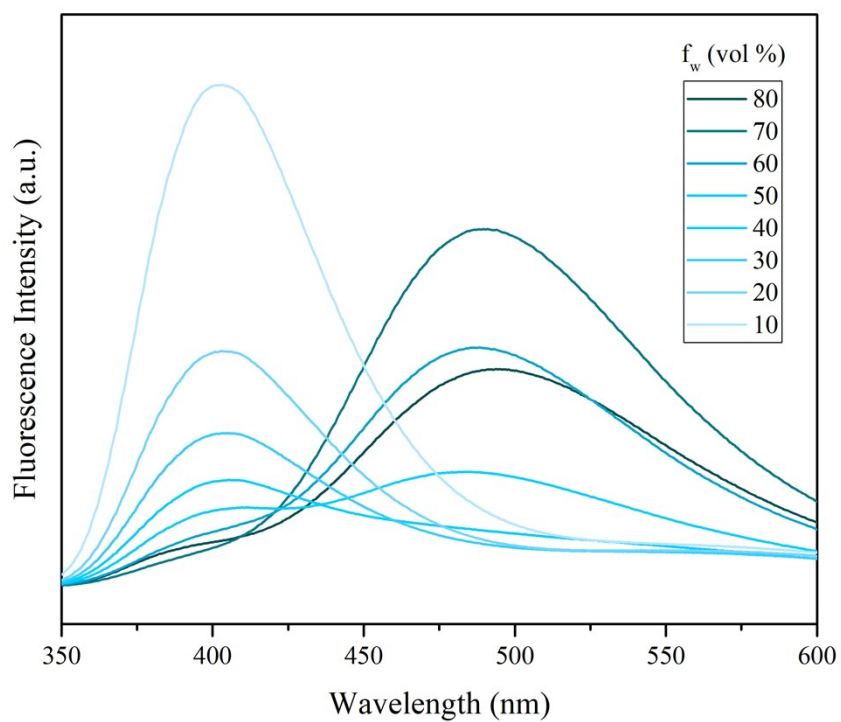
**Figure S10** FT-IR spectrum of **DiCat**

## UV-Vis spectra

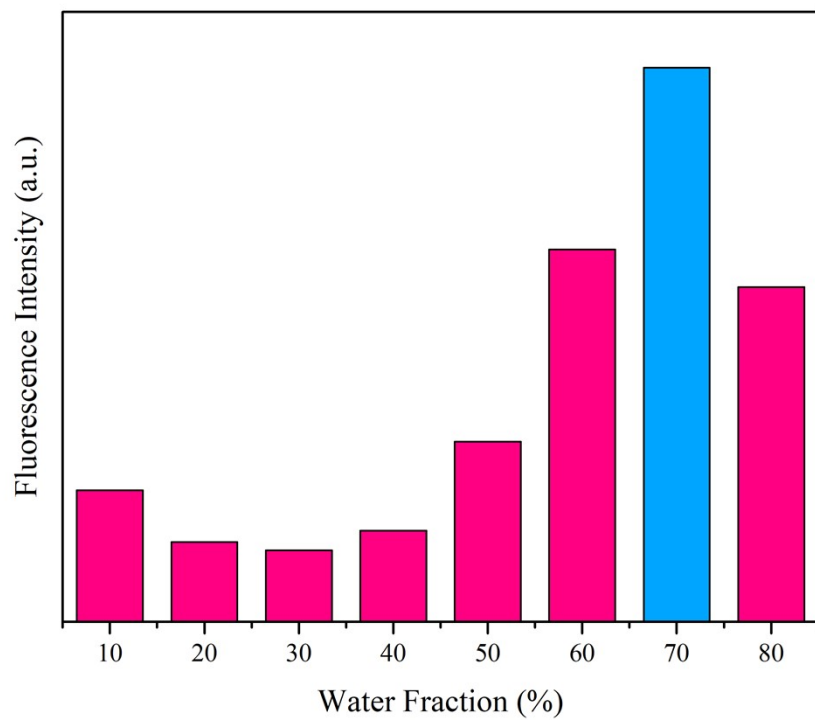


**Figure S11** UV-Vis spectrum of **DiCat** and sensing system under optimal detection condition (binary solvent of THF and DI H<sub>2</sub>O, water fraction of 70%).

## Optimization of the detection condition

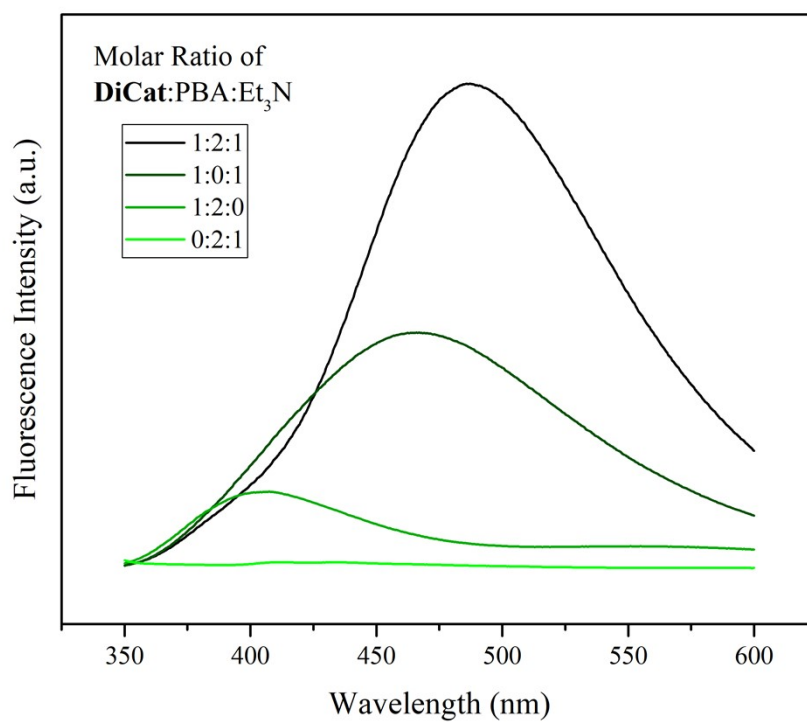


**Figure S12** Fluorescence spectrum recorded in THF/DI H<sub>2</sub>O solution of DiCat/PBA/Et<sub>3</sub>N (molar ratio 1:2:1) with varying water fraction.



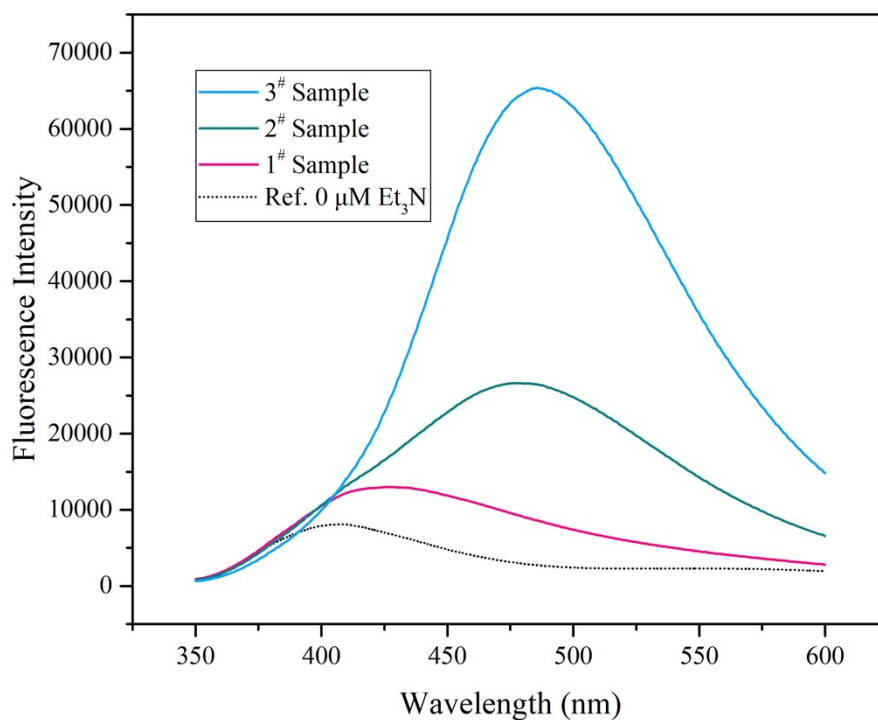
**Figure S13** Column chart of PL intensity versus water fraction which determines 70% water fraction as the optimal detection condition.

## Control experiments



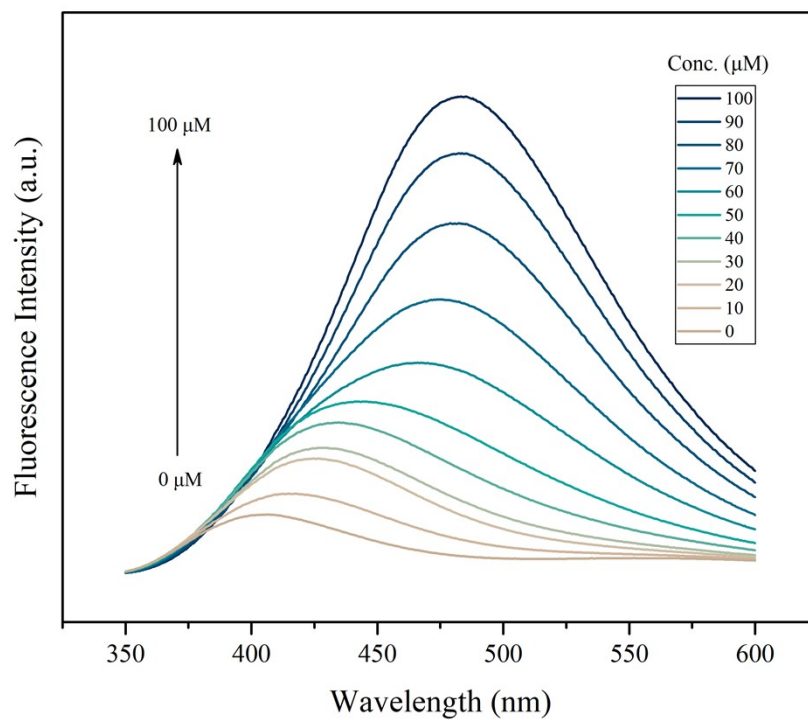
**Figure S14** Fluorescence spectrum of **DiCat/PBA/Et<sub>3</sub>N** system with different molar ratio in THF/DI H<sub>2</sub>O binary solvent ( $f_w = 70\%$ ).

## Fluorescence spectrum of unknown samples containing Et<sub>3</sub>N

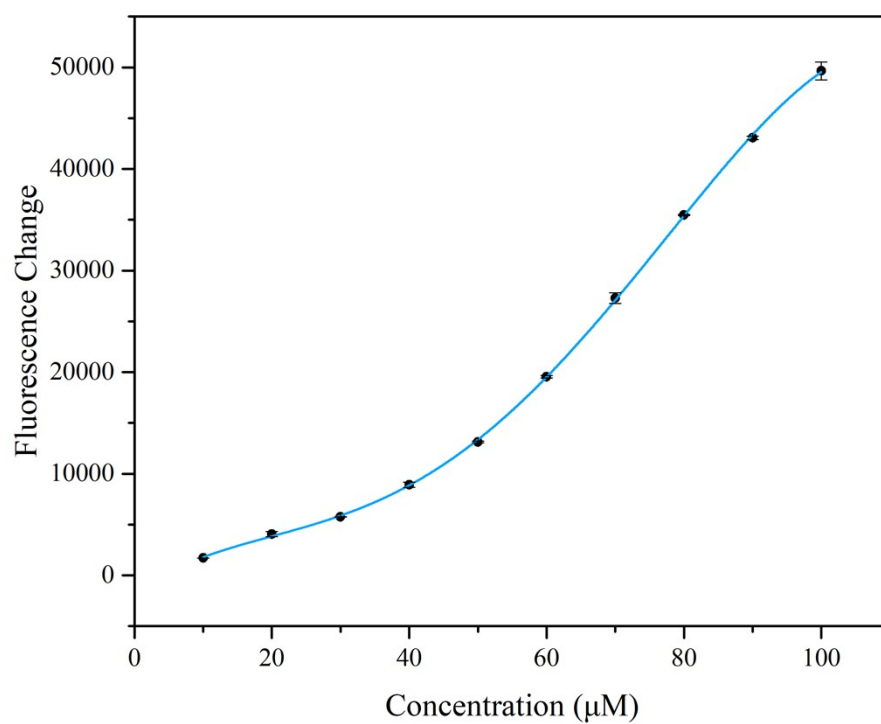


**Figure S15** Fluorescence spectrum of three unknown samples containing Et<sub>3</sub>N comparing with reference sensing system without Et<sub>3</sub>N (recorded under optimal detection condition with THF/DI H<sub>2</sub>O binary solvent,  $f_w = 70\%$ ; concentration of **DiCat** is 100 μM; concentration of PBA is 200 μM).

## Fluorescence spectrum and fitting curves of diverse organic amines

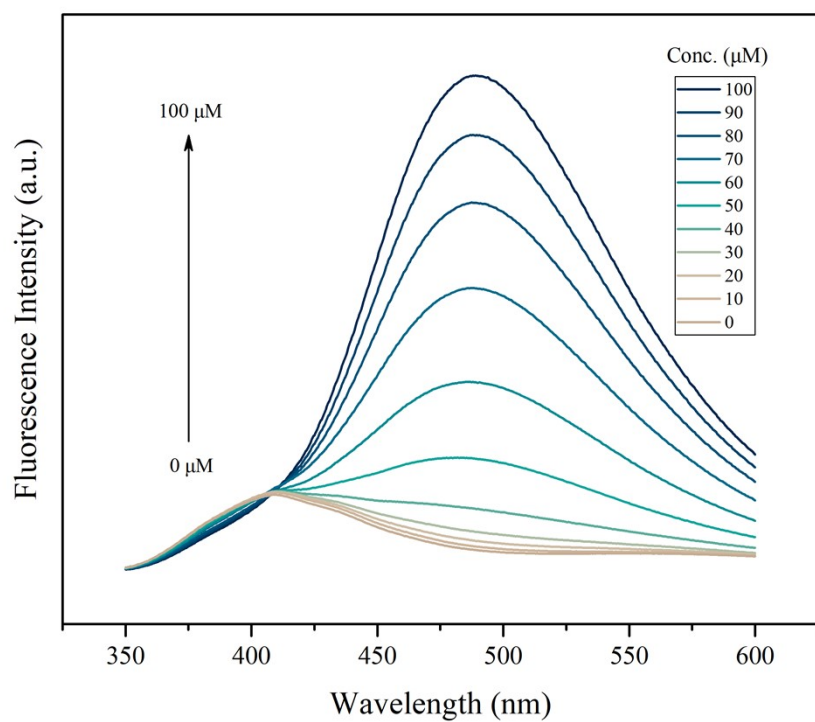


**Figure S16** Fluorescence spectrum of **DiCat/PBA** with increasing concentration of diisopropylamine under optimal detection condition. (concentration of **DiCat** and PBA is fixed to be 100  $\mu\text{M}$  and 200  $\mu\text{M}$  respectively)

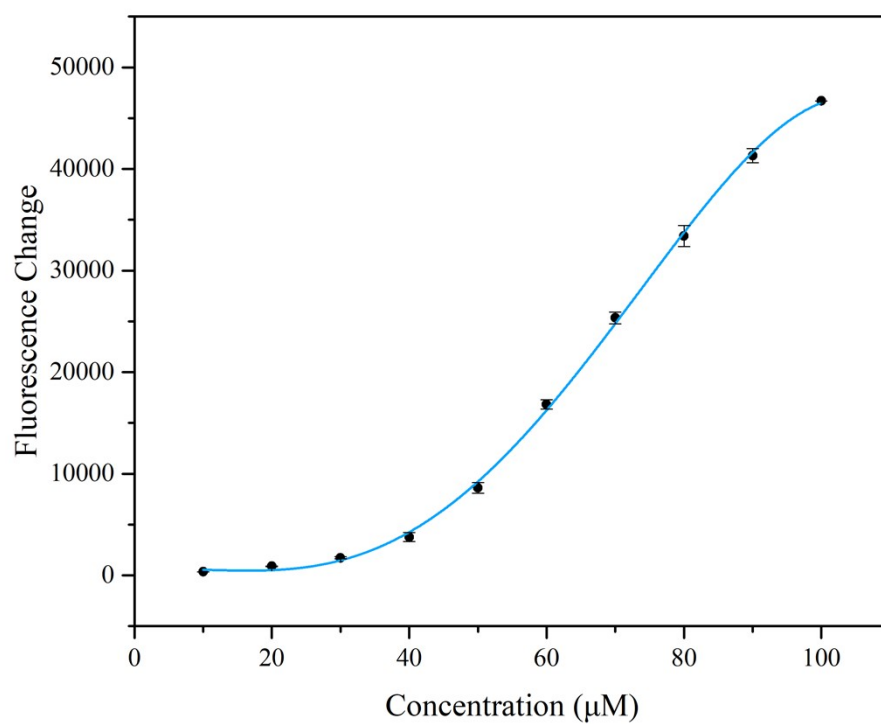


**Figure S17** Polynomial regression of fluorescence change versus diisopropylamine concentration.

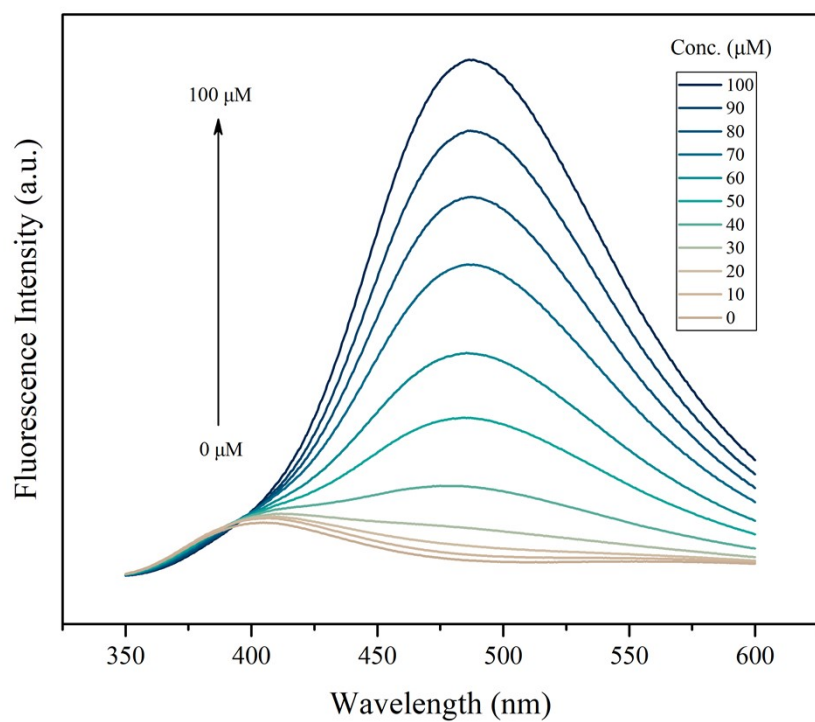




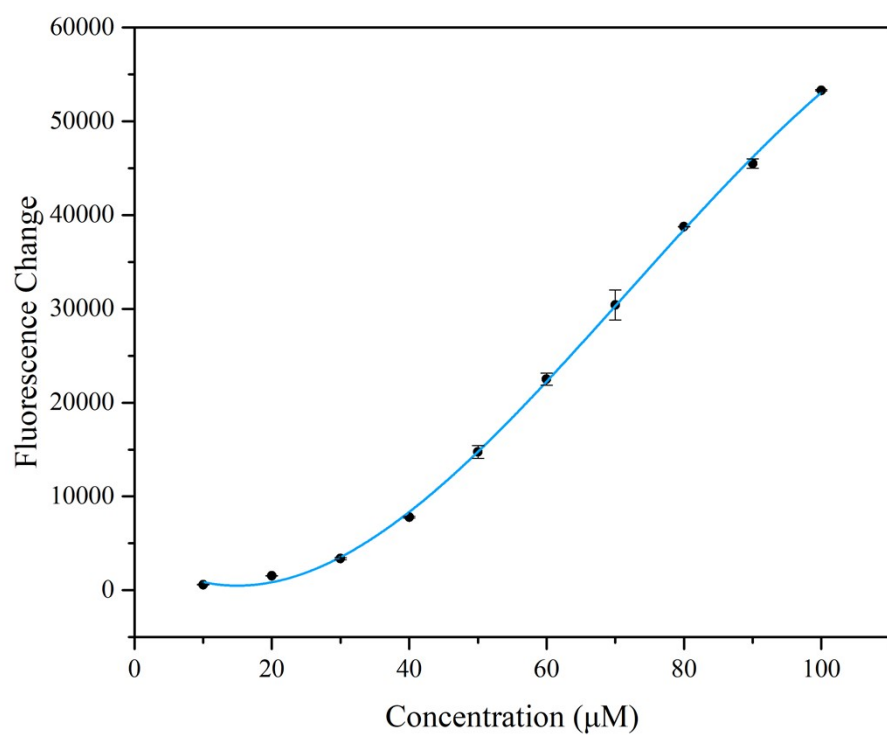
**Figure S18** Fluorescence spectrum of **DiCat**/PBA with increasing concentration of n-butylamine under optimal detection condition. (concentration of **DiCat** and PBA is fixed to be 100  $\mu\text{M}$  and 200  $\mu\text{M}$  respectively)



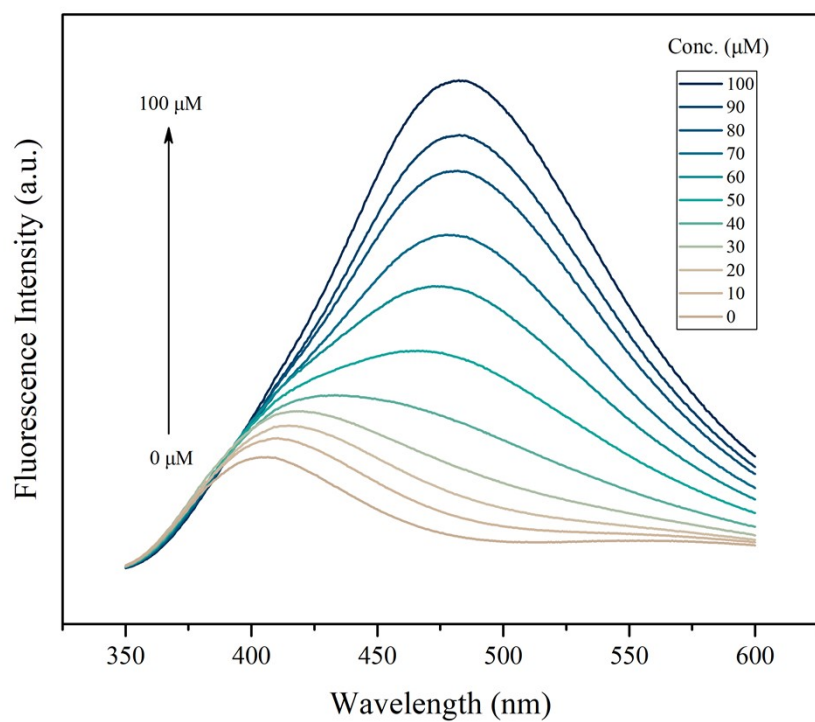
**Figure S19** Polynomial regression of fluorescence change versus n-butylamine concentration.



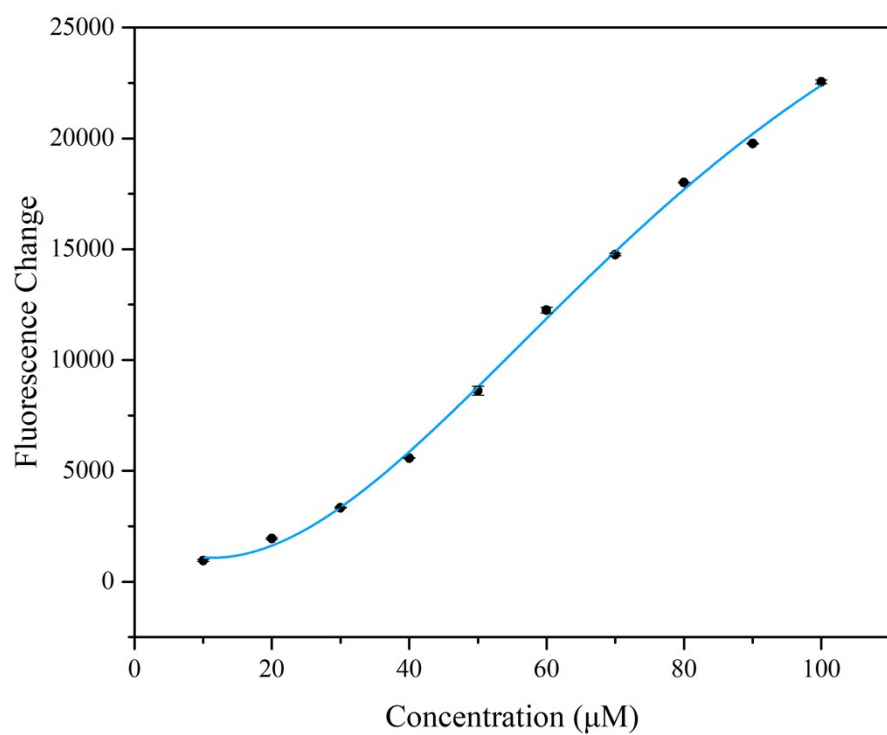
**Figure S20** Fluorescence spectrum of **DiCat/PBA** with increasing concentration of **DIPEA** under optimal detection condition. (concentration of **DiCat** and **PBA** is fixed to be 100  $\mu\text{M}$  and 200  $\mu\text{M}$  respectively)



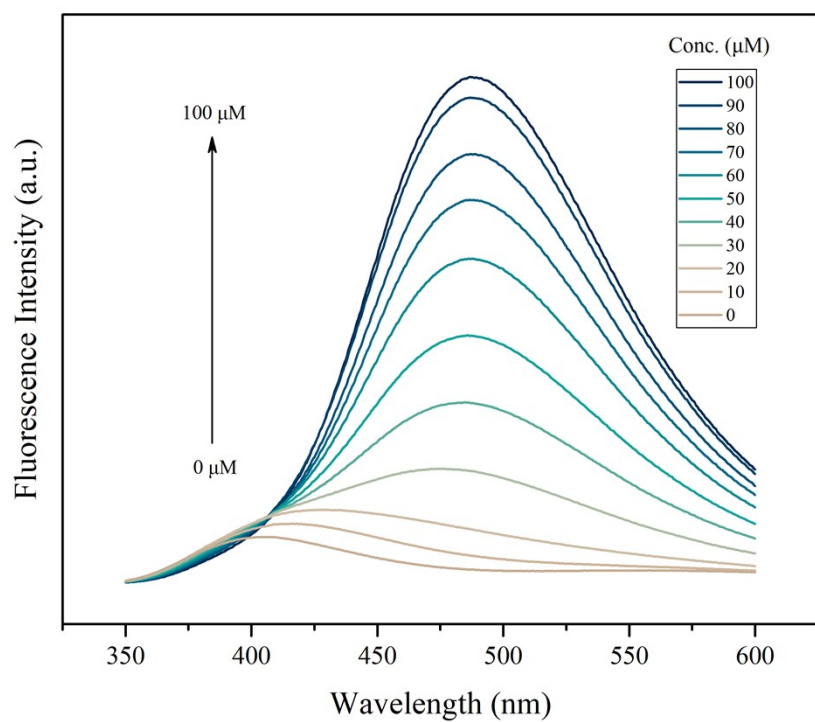
**Figure S21** Polynomial regression of fluorescence change versus DIPEA concentration.



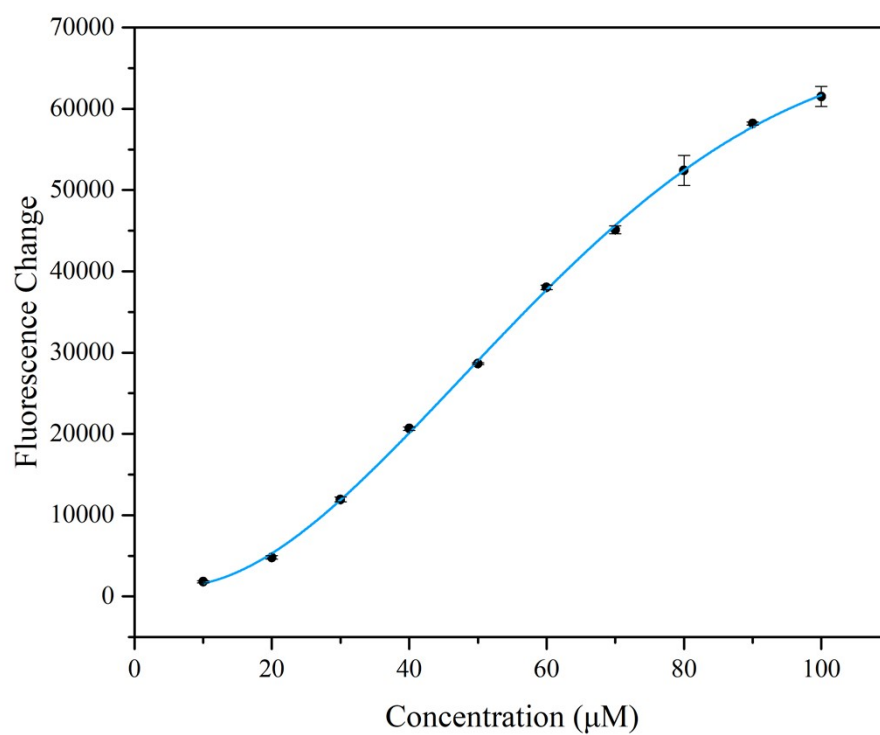
**Figure S22** Fluorescence spectrum of **DiCat/PBA** with increasing concentration of **DMAP** under optimal detection condition. (concentration of **DiCat** and **PBA** is fixed to be 100 μM and 200 μM respectively)



**Figure S23** Polynomial regression of fluorescence change versus DMAP concentration.

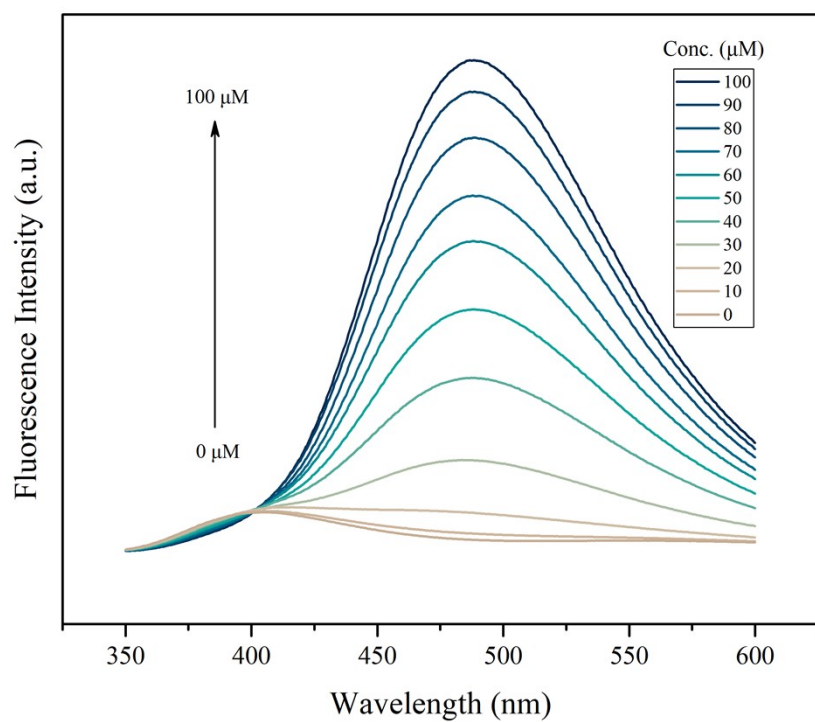


**Figure S24** Fluorescence spectrum of **DiCat/PBA** with increasing concentration of putrescine under optimal detection condition. (concentration of **DiCat** and PBA is fixed to be 100  $\mu\text{M}$  and 200  $\mu\text{M}$  respectively)

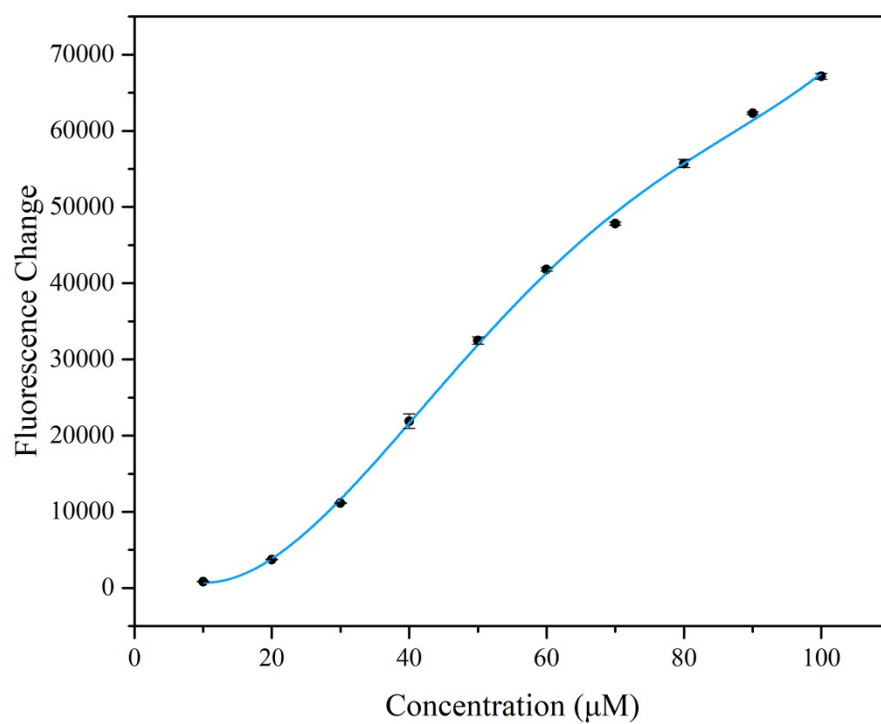


**Figure S25** Polynomial regression of fluorescence change versus putrescine concentration.

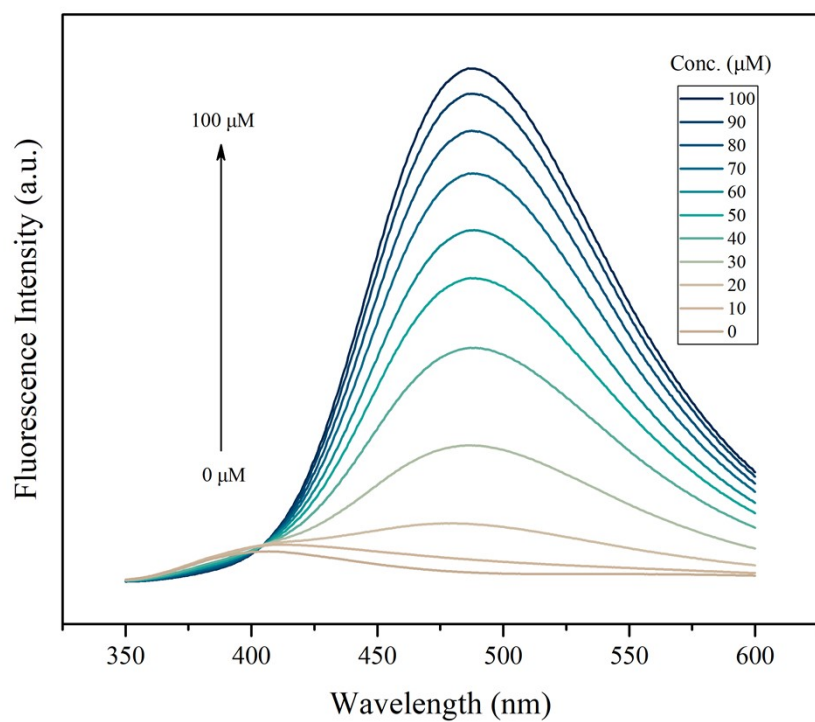




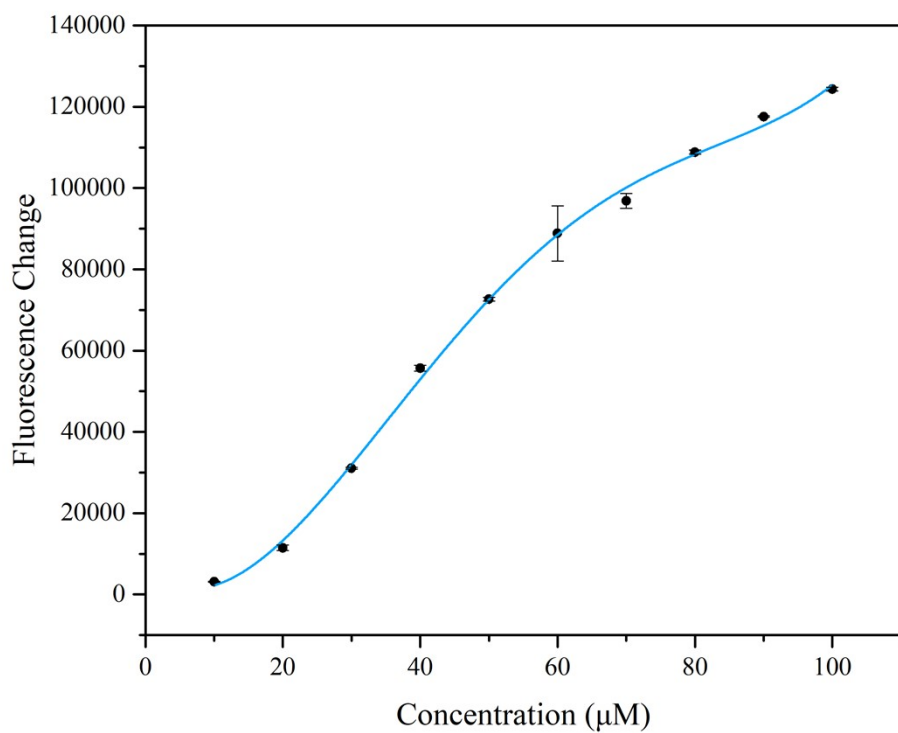
**Figure S26** Fluorescence spectrum of **DiCat/PBA** with increasing concentration of cadaverine under optimal detection condition. (concentration of **DiCat** and PBA is fixed to be 100  $\mu\text{M}$  and 200  $\mu\text{M}$  respectively)



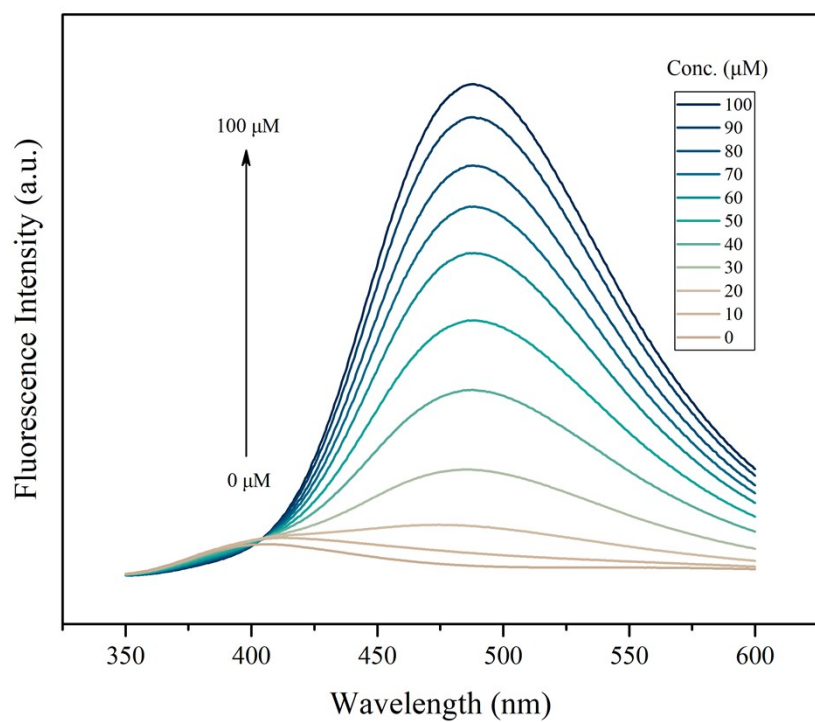
**Figure S27** Polynomial regression of fluorescence change versus cadaverine concentration.



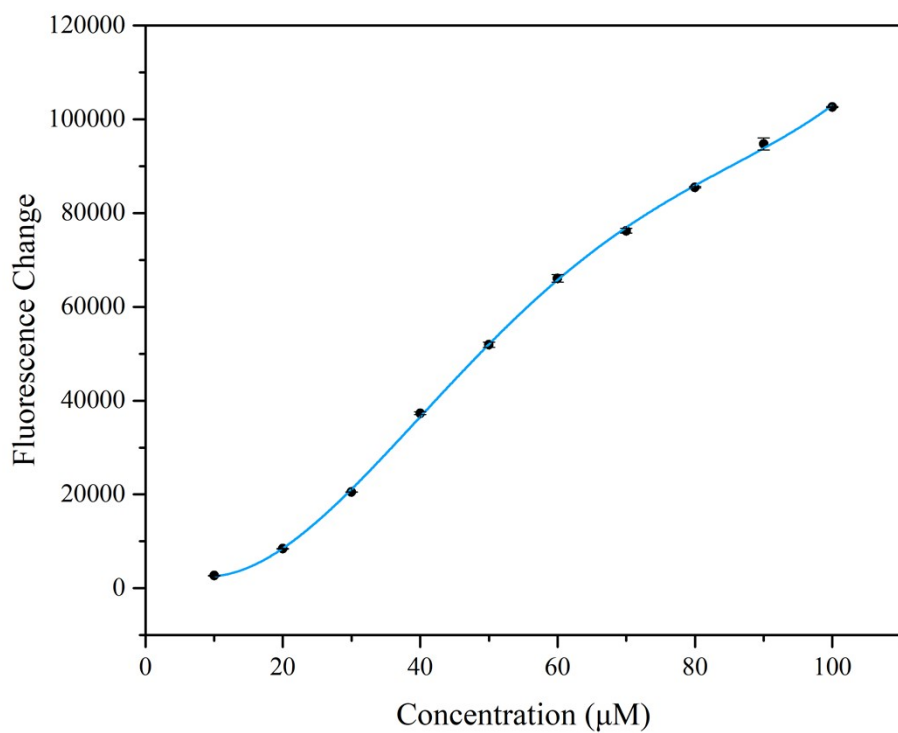
**Figure S28** Fluorescence spectrum of **DiCat/PBA** with increasing concentration of spermine under optimal detection condition. (concentration of **DiCat** and PBA is fixed to be 100 μM and 200 μM respectively)



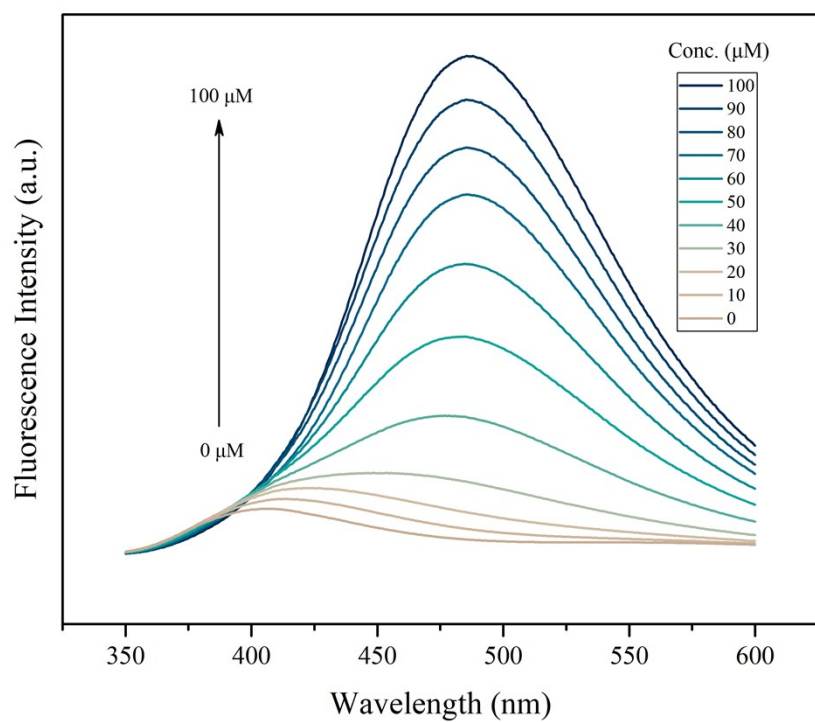
**Figure S29** Polynomial regression of fluorescence change versus spermine concentration.



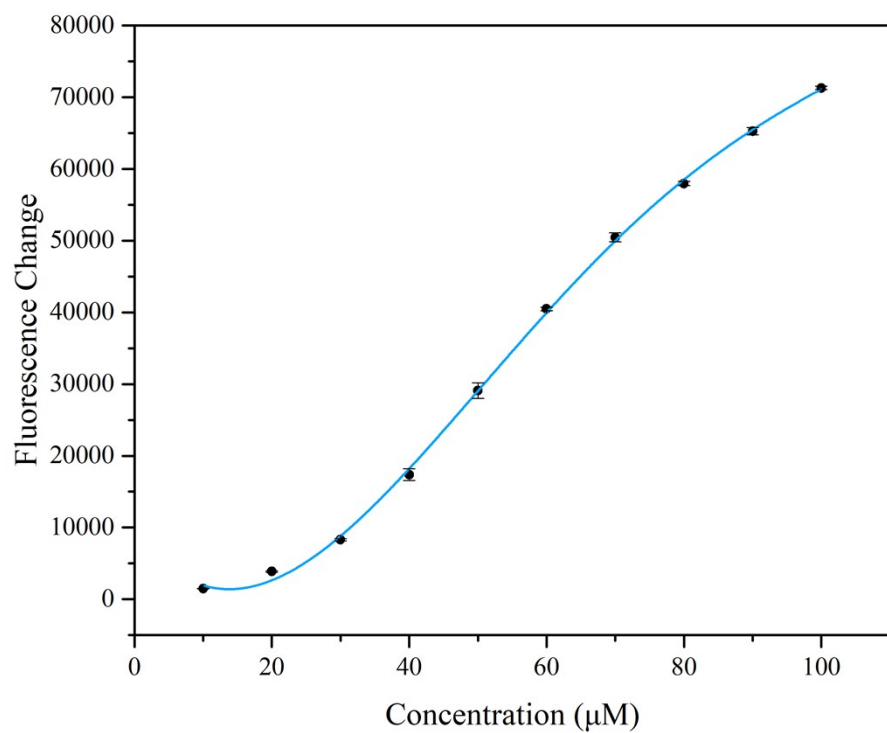
**Figure S30** Fluorescence spectrum of **DiCat/PBA** with increasing concentration of spermidine under optimal detection condition. (concentration of **DiCat** and PBA is fixed to be 100  $\mu\text{M}$  and 200  $\mu\text{M}$  respectively)



**Figure S31** Polynomial regression of fluorescence change versus spermidine concentration.

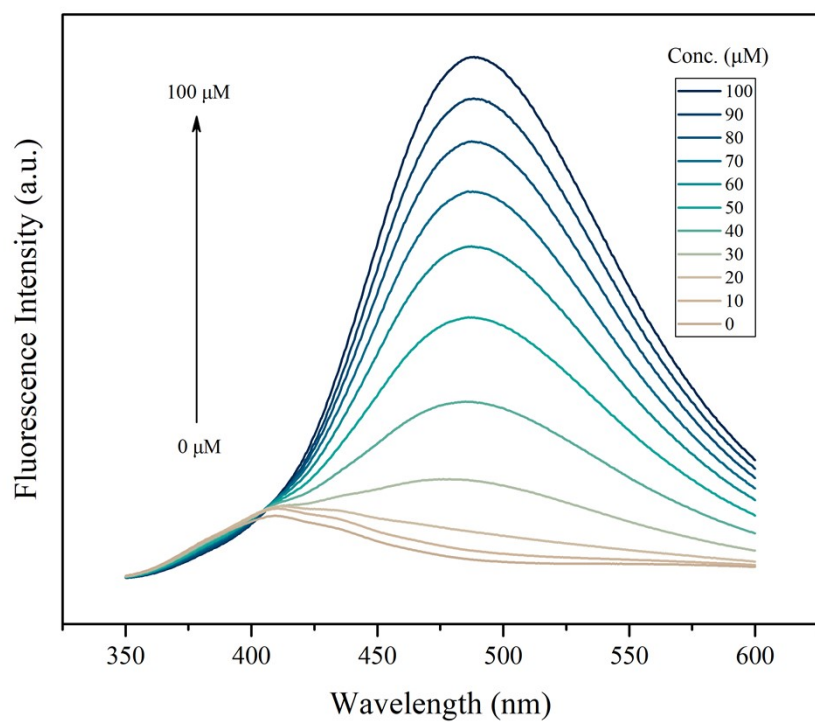


**Figure S32** Fluorescence spectrum of **DiCat/PBA** with increasing concentration of tyramine under optimal detection condition. (concentration of **DiCat** and PBA is fixed to be 100  $\mu\text{M}$  and 200  $\mu\text{M}$  respectively)

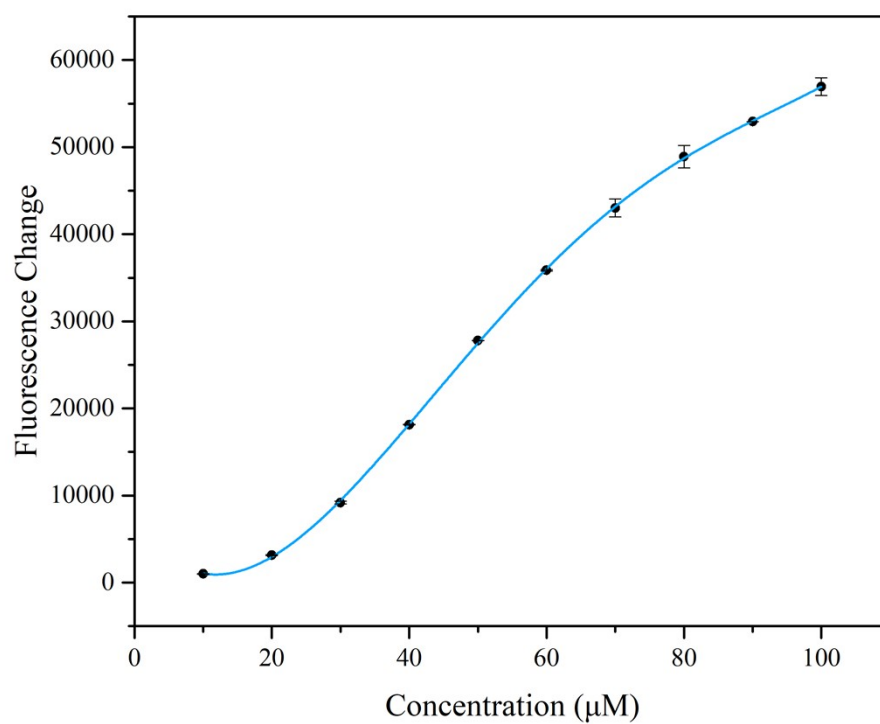


**Figure S33** Polynomial regression of fluorescence change versus tyramine concentration.

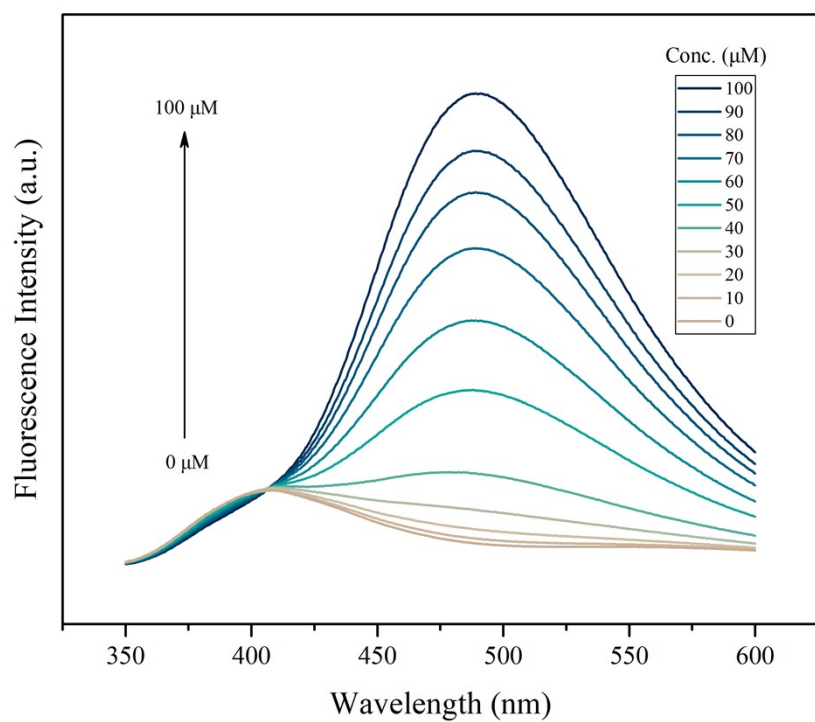




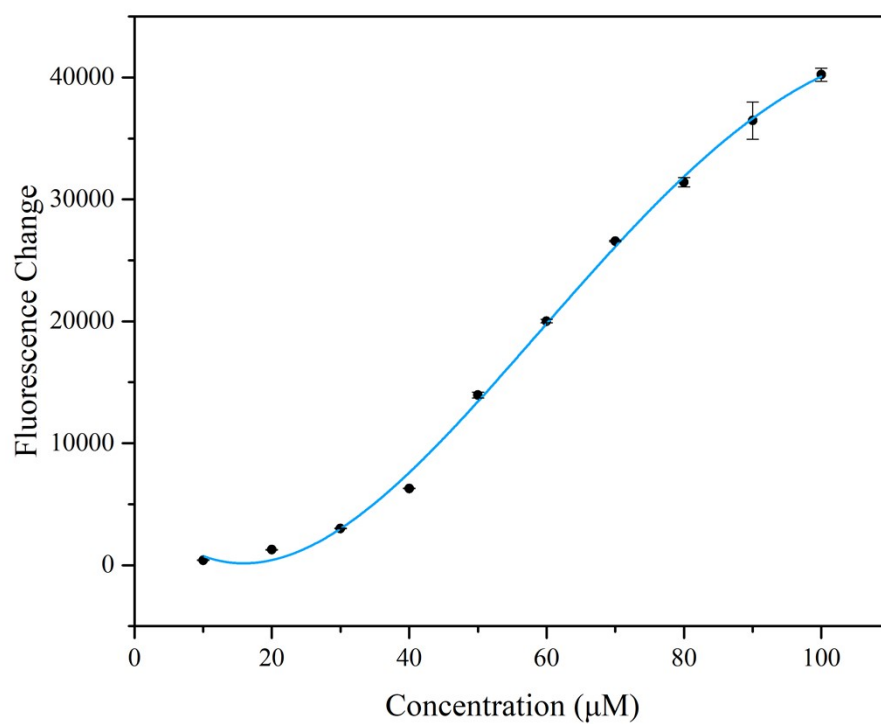
**Figure S34** Fluorescence spectrum of **DiCat/PBA** with increasing concentration of phenylethylamine under optimal detection condition. (concentration of **DiCat** and PBA is fixed to be 100  $\mu\text{M}$  and 200  $\mu\text{M}$  respectively)



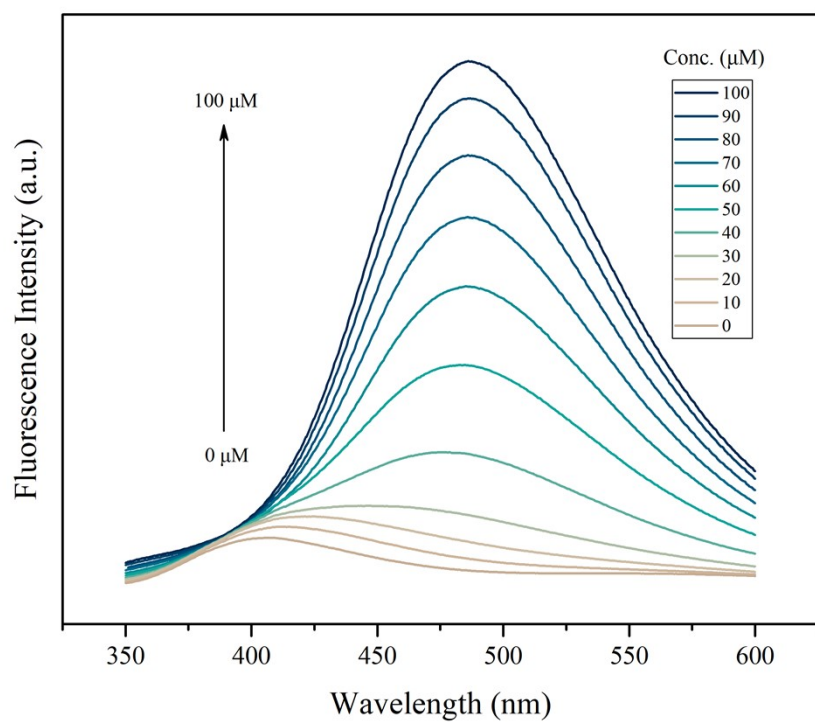
**Figure S35** Polynomial regression of fluorescence change versus phenylethylamine concentration.



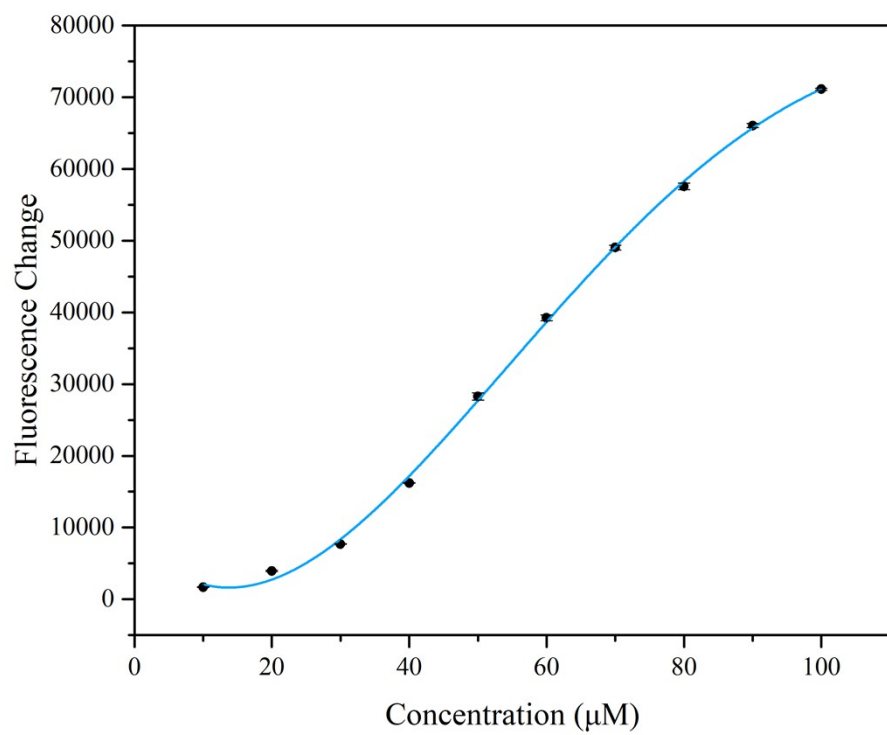
**Figure S36** Fluorescence spectrum of **DiCat/PBA** with increasing concentration of histamine under optimal detection condition. (concentration of **DiCat** and PBA is fixed to be 100  $\mu\text{M}$  and 200  $\mu\text{M}$  respectively)



**Figure S37** Polynomial regression of fluorescence change versus histamine concentration.

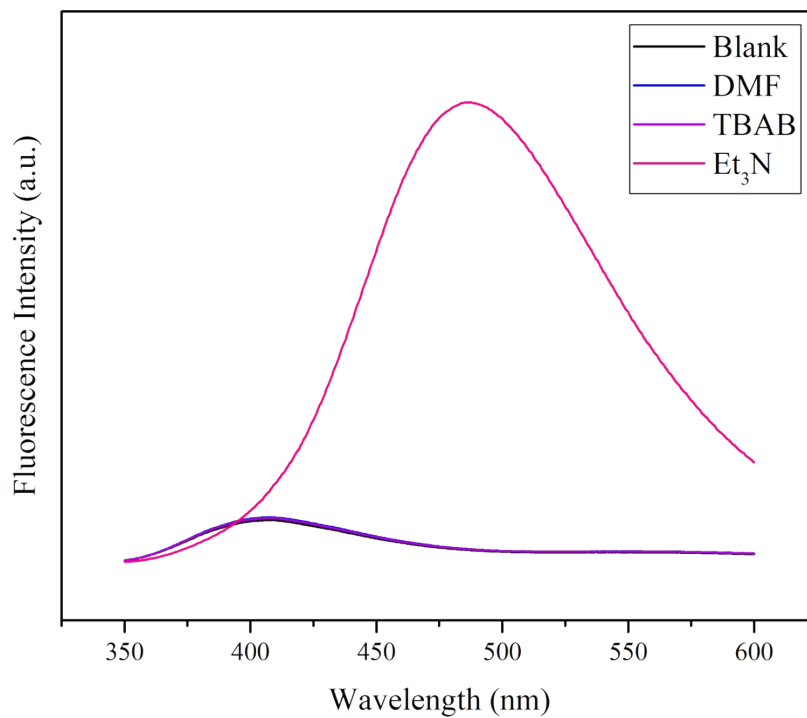


**Figure S38** Fluorescence spectrum of **DiCat/PBA** with increasing concentration of tryptamine under optimal detection condition. (concentration of **DiCat** and PBA is fixed to be 100  $\mu\text{M}$  and 200  $\mu\text{M}$  respectively)

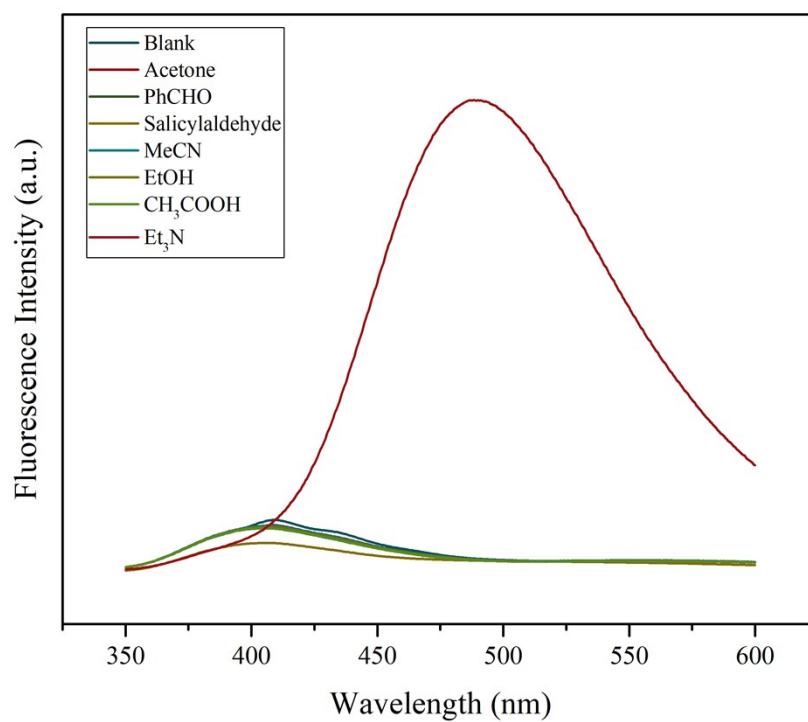


**Figure S39** Polynomial regression of fluorescence change versus tryptamine concentration.

## Selectivity of this detection protocol

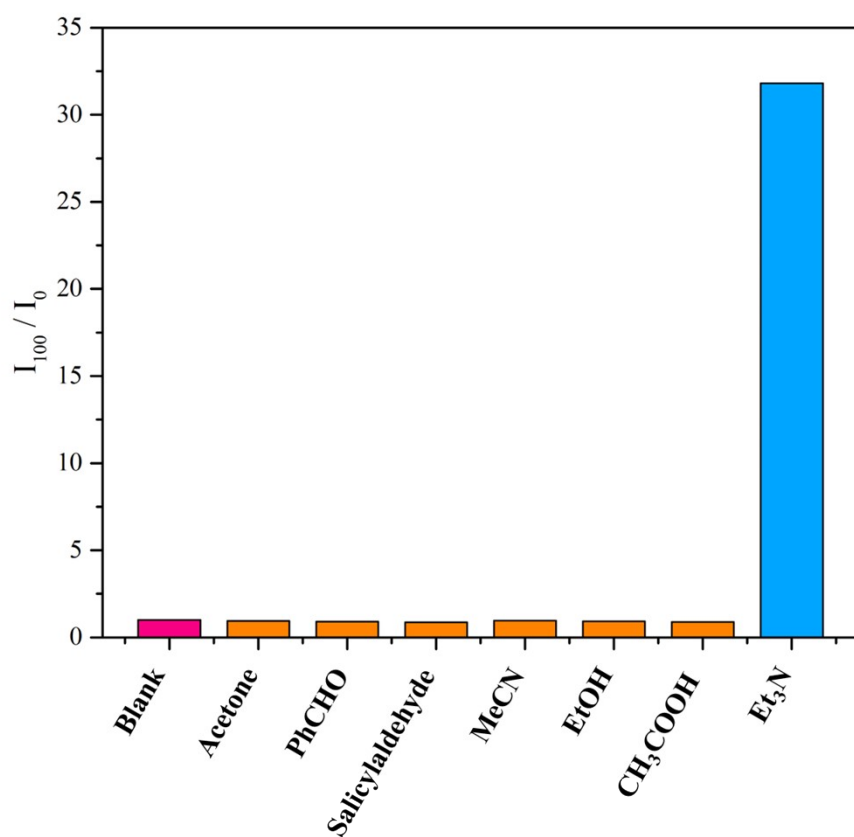


**Figure S40** Fluorescence spectrum of **DiCat/PBA** with DMF (100  $\mu\text{M}$ ), TBAB (100  $\mu\text{M}$ ), and Et<sub>3</sub>N (100  $\mu\text{M}$ ) under optimal detection condition. (concentration of **DiCat** and PBA is fixed to be 100  $\mu\text{M}$  and 200  $\mu\text{M}$  respectively)



**Figure S41** Fluorescence spectrum of **DiCat**/PBA with addition of other common water-soluble organic compounds compared with Et<sub>3</sub>N. (with the substance concentration of 100  $\mu$ M; optimal detection condition is used as concentration of **DiCat** and PBA is fixed to be 100  $\mu$ M and 200  $\mu$ M respectively)





**Figure S42** Relative response intensity of DiCat/PBA towards different water-soluble organic compounds. (detection condition is the same as shown in **Figure S41**, fluorescence intensity at 488 nm is taken for comparison)

## Computational details

Density functional theory (DFT) calculations were carried out with B3LYP functional and 6-31G\* basis set for structural optimization of **DiCat**. Detailed atomic coordinates were listed below.

**Table S1** Atomic coordinates within the optimal molecular structure of **DiCat**.

Atom Type	x	y	z
C	-1.35601	-1.73323	-1.00533
C	-0.00869	-2.08637	-1.22635
C	1.00394	-1.22858	-0.86479
C	0.71847	0.01318	-0.23317
C	-0.65749	0.3849	-0.02707
C	-1.69875	-0.51647	-0.44079
C	1.76005	0.9133	0.182
C	1.41766	2.13155	0.74292
C	0.07008	2.48716	0.95844
C	-0.94308	1.62892	0.59962
C	-3.14174	-0.1811	-0.29319
C	3.20324	0.57649	0.04128
C	4.07392	1.43814	-0.63851
C	5.43867	1.1588	-0.73262
C	5.96209	0.01168	-0.14392
C	5.09783	-0.85753	0.54573
C	3.74082	-0.57888	0.63805
C	-3.69949	0.96654	-0.87513
C	-5.06489	1.23511	-0.77207
C	-5.90348	0.3613	-0.08516
C	-5.3555	-0.79426	0.49792
C	-3.99604	-1.06112	0.39369
O	-7.23658	0.6307	0.01588

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O	-6.25906	-1.59772	1.1607
O	7.29547	-0.25908	-0.23789
O	5.7065	-1.96101	1.10573
H	-2.14707	-2.40602	-1.32539
H	0.22505	-3.03082	-1.71073
H	2.0348	-1.49011	-1.07416
H	2.20918	2.80512	1.05947
H	-0.16397	3.43575	1.43424
H	-1.97421	1.89526	0.80057
H	3.67406	2.33056	-1.1103
H	6.11454	1.82008	-1.26556
H	3.08915	-1.25381	1.189
H	-3.05984	1.64729	-1.42796
H	-5.50139	2.11859	-1.22706
H	-3.58419	-1.95438	0.85933
H	-7.64433	-0.08886	0.52703
H	-5.79917	-2.36119	1.53988
H	7.46092	-1.09099	0.23736
H	5.04608	-2.49671	1.56954

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We calculated the excited electronic structure of the proposed tetra-coordinated boronic intermediate while **DiCat**/PBA ensemble interacting with Et<sub>3</sub>N. TDDFT method under PBE0-D3/def2-TZVP level was taken to obtain the excited energy, oscillator strength and emission spectrum of the given structure. All the calculations were performed using Gaussian 16 program. The natural transition orbitals (NTOs) and the contributions of molecular orbital transitions were obtained by electron excitation analysis from the transition density matrix of TD-DFT calculation using Multiwfn program<sup>[s1]</sup>. The SMD (Solvation Model Based on Density) <sup>[s2]</sup> implicit solvent model was used to describe the solvation effect of tetrahydrofuran and water. The visualization of the frontier molecular orbitals and natural transition orbitals were also rendered using Visual Molecular Dynamic program (VMD)<sup>[s3]</sup>.

Ref.

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