Metal Ions Determined Self-Assembly Using Terpyridine Building Blocks

Qixia Bai,^{‡a} Ying liu, ^{‡a} Tun Wu,^a Haoyue Su,^c Gang Chen,^a Yuming Guan,^a Ming Wang,^c Ting-Zheng Xie,^a Zhe Zhang,^{*a,b} Pingshan Wang^{*a}

^a Institute of Environmental Research at Greater Bay Area; Key Laboratory for Water Quality and Conservation of the Pearl River Delta, Ministry of Education; ^bGuangzhou Key Laboratory for Clean Energy and Materials; Guangzhou University, Guangzhou 510006, China.

^b Guangdong Provincial Key Laboratory of Functional Supramolecular Coordination Materials and Applications, Jinan University, Guangzhou 510632, China

^c State Key Laboratory of Supramolecular Structure and Materials, College of Chemistry, Jilin University, Changchun, Jilin 130012, China

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1. Schemes of preparation of ligand L



Scheme S1. Synthesis of ligand L.

2. Experimental section

General procedures. Chemicals were purchased from Sigma/Aldrich, Fisher Scientific, Energy Chemical, Alfa Aesar and used without further purification. Thinlayer chromatography (TLC) was conducted on flexible sheets (Baker-flex) precoated with Al₂O₃ (IB-F) or SiO₂ (IB2-F). Column chromatography was conducted using basic Al₂O₃ Brockman Activity I (60-325 mesh) or SiO₂ (60-200 mesh) from Fisher Scientific. NMR spectra were recorded on Bruker NMR 400 or 500 MHz spectrometers, using CDCl₃ and CD₃OD for ligands, CD₃CN for metal products. ESI mass spectrometry (MS) experiments were performed on a Waters Synapt HDMS G2-Si quadrupole/time-of-flight (Q/TOF) tandem mass spectrometer. This instrument contains a triwave device between the Q and TOF analyzers, consisting of three collision cells in the order trap cell, ion mobility cell, and transfer cell. Trap and transfer cells are pressurized with Ar, and the ion mobility cell is pressurized with N₂ flowing in a direction opposite to that of the entering ions.

Mass Spectrometry and Ion Mobility. ESI-MS and TWIM-MS were recorded with a Waters Synapt G2-Si tandem mass spectrometer, using solutions of 0.01 mg sample in 1 mL of CHCl₃/CH₃OH (1:3, v/v) for ligands or 0.5 mg sample in 1 mL of CH₃CN/CH₃OH (3:1, v/v) for complexes. All samples were infused into the ESI source at a flow rate of 6 μ L/min by a syringe pump (KDS-100, KD Scientific). The TWIM-MS experiments were performed under the following conditions: ESI capillary voltage, 2 kV; sample cone voltage, 35 V; source offset, 42 V; source temperature 150 °C; desolvation temperature, 250 °C; cone gas flow, 10 L/h; desolvation gas flow, 700 L/h (N₂); source gas control, 0 mL/min; trap gas control, 3 mL/min; helium cell gas control, 120 mL/min; ion mobility (IM) cell gas control, 30 mL/min; sample flow rate, 8 μ L/min; IM traveling wave height, 25 V; and IM traveling wave velocity, 1200 m/s. Data were collected and analyzed by using Mass Lynx 4.2 and Drift Scope 2.9.

gMS². Gradient tandem mass spectrometry was performed under the following conditions: 13+ charged ions of complexes were isolated by quadrupole for the following collision-induced dissociation (CID), in which collision energy was

gradually increased by changing the voltage of trap cell depended on different complexes.

TEM. Transmission electron microscopy tests were performed on the JEOL JEM-2100F equipment. The sample solutions were diluted to a concentration of 10^{-6} M using CH₃CN, drop-casted onto a lacey carbon-covered Cu grid (300 mesh, purchased from Ted Pella Inc.) and the extra solution was absorbed by filter paper to avoid aggregation. And the size statistics were obtained by using the software image J.

AFM. AFM imaging was performed on a Bruker Dimension Icon AFM system with ScanAsyst and the data was processed by NanoScope Analysis version 2.0 (Bruker Software, Inc.). The sample was diluted to a concentration of 5×10^{-6} M using CH₃CN, dropped on freshly cleaved mica surface, and then dried in the air. Silicon cantilevers tip with a spring constant of around 0.1 N/m was used for the experiments.

UV-vis absorption and fluorescence properties. UV-vis absorption spectra were recorded on a Persee TU-1900 UV-visible spectrophotometer at room temperature (10^{-6} M in CH₃CN) and were corrected with the background spectrum of the solvent. Fluorescence properties were performed on Edinburgh-FS5 Fluorescence spectrometer at 73K (10^{-6} M in CH₃CN).

Cyclic Voltammetry. Cyclic voltammetry measurements were performed on a Princeton VersaSTAT 3F potentiostat with a standard three-electrode configuration using a glass-carbon working electrode, a platinum-rod auxiliary electrode, and a reference Ag/AgCl electrode. The electrochemical properties of these two complexes in CN_3CN (10⁻⁶ M) were studied in a three-electrode electrochemical cell with Bu₄NPF₆ (0.1 M) as an electrolyte.

3. Synthesis of the compounds and supramolecules



Compound 1, 2,8-Dibromodibenzofuran (1.00 g, 3.07 mmol), 4'-(4-boronatophenyl)-[2,2':6',2"]terpyridine (1.19 g, 3.37 mmol), Pd(PPh₃)₂Cl₂ (108 mg, 0.154 mmol) and sodium carbonate (976 mg, 9.21 mmol) were added into a 200 mL three-necked flask. 50 mL toluene, 30 mL H₂O and 15 mL tert-butyl alcohol were added under N₂. The mixture was stirred at 85 °C for 8 h. After cooling to room temperature, the mixture was extracted with CH₂Cl₂. The combined organic layer was washed with brine, dried over anhydrous Na₂SO₄, and then concentrated in vacuo, the residue was purified by flash column chromatography (silica gel), eluting with CH₂Cl₂: CH₃OH (100:0.75, ν/ν) to give the product as 1.27 g white solid (yield 75%). ¹H NMR (600 MHz, CDCl₃, 300 K) δ 8.82 (s, 2H, *Tpy-H*^{3',5'}), 8.76 (d, *J* = 5.5 Hz, 2H, *Tpy-*^{6,6''}), 8.70 (d, *J* = 7.9 Hz, 2H, *Tpy-H*^{3,3"}), 8.17 (d, J = 6.6 Hz, 2H, *Ph-H*^b), 8.05 (d, J = 8.3 Hz, 2H, *Ph-H*^{f,g}), 7.90 (t, J = 6.6 Hz, 2H, *Ph-H*^b), 8.05 (d, J = 8.3 Hz, 2H, *Ph-H*^{f,g}), 7.90 (t, J = 6.6 Hz, 2H, *Ph-H*^b), 8.05 (d, J = 8.3 Hz, 2H, *Ph-H*^{f,g}), 7.90 (t, J = 6.6 Hz, 2H, *Ph-H*^b), 8.05 (d, J = 8.3 Hz, 2H, *Ph-H*^{f,g}), 7.90 (t, J = 6.6 Hz, 2H, *Ph-H*^b), 8.05 (d, J = 8.3 Hz, 2H, *Ph-H*^{f,g}), 7.90 (t, J = 6.6 Hz, 2H, *Ph-H*^b), 8.05 (d, J = 8.3 Hz, 2H, *Ph-H*^{f,g}), 7.90 (t, J = 6.6 Hz, 2H, *Ph-H* $= 7.7 \text{ Hz}, 2\text{H}, Tpy-H^{4,4''}, 7.80 \text{ (d}, J = 8.4 \text{ Hz}, 2\text{H}, Ph-H^{a}), 7.79 - 7.77 \text{ (m, 1H, Ph-H^{d})},$ 7.65 (d, J = 8.5 Hz, 1H, *Ph-H^e*), 7.58 (d, J = 8.7 Hz, 1H, *Ph-H^h*), 7.48 (d, J = 8.7 Hz, 1H, *Ph-H^c*), 7.40 – 7.35 (m, 2H, *Tpy-H^{5,5}"*); ¹³C NMR (126 MHz, CDCl₃, 300 K) δ 156.32, 156.03, 149.81, 149.21, 137.13, 129.07, 128.14 - 127.66, 127.4, 127.27, 123.92, 123.69 - 123.63, 121.58, 119.12, 118.85;ESI-TOF (m/z): Calcd. for [C₃₃H₂₀BrN₃O+H] ⁺: 554.45. Found: 554.45.



Compound 2, Compound **1** (415.5 mg, 0.75 mmol) and RuCl₃•3H₂O (504.5 mg, 2.25 mmol) were mixed in 150 mL CHCl₃/CH₃OH (ν/ν , 1:1). The mixture was stirred at 65 °C for 24 h. After cooling to ambient temperature, the precipitates were filtered and washed with CH₃OH and H₂O to afford 485 mg compound **2** (yield 85%).



Compound 3, Compound 3 was synthesized according to the reported literature.¹



Compound 4, Compound **3** (5.20 g, 18.4 mmol) was added to a 100 mL single bottomed flask, 50 mL of acetone was poured, and then 1,8-diazabicycloundec-7-ene (13.8 mL, 92.2 mmol) was added dropwise to the flask and stirred at room temperature for 30 min. After the reaction was completed, 2 mol/L of HCl solution was added to neutralize the reaction, and when the solution became clarified, the organic phase was extracted with ether. The combined organic layer was washed with brine, dried over anhydrous Na₂SO₄, and then concentrated in vacuo, the residue was purified by flash column chromatography (silica gel), eluting with petroleum ether to give the product as 1.27 g white crystals (yield 75%). ¹H NMR (400 MHz, CDCl₃, 300 K) δ 7.05 (s, 2H, *Ph-H^e*), 3.83 (s, 3H, *-OCH₃-H^a*), 3.76 (s, 3H, *-OCH₃-H^b*); ¹³C NMR (101 MHz, CDCl₃, 300 K) δ 156.42, 148.11, 118.26, 117.92, 60.82, 56.03.



Compound 5, Compound 4 (440.8 mg, 1.5 mmol), 4'-(4-boronatophenyl)-[2,2':6',2"]terpyridine (1.59 g, 4.5 mmol), Pd(PPh₃)₂Cl₂ (105 mg, 0.15 mmol) and sodium carbonate (954 mg, 9 mmol) were added into a 250 mL three-necked flask. 60 mL toluene, 10 mL H₂O and 25 mL tert-butyl alcohol were added under N₂. The mixture was stirred at 85 °C for 48 h. After cooling to room temperature, the mixture was extracted with CH₂Cl₂. The combined organic layer was washed with brine, dried over anhydrous Na₂SO₄, and then concentrated in vacuo, the residue was purified by flash column chromatography (silica gel), eluting with CH_2Cl_2 : CH_3OH (100:1, v/v) to give the product as 880 mg white solid (yield 78%). ¹H NMR (400 MHz, CDCl₃, 300 K) δ 8.82 (s, 4H, *Tpy-H*^{3',5'}), 8.75 (d, J = 5.6 Hz, 4H, *Tpy-H*^{6,6"}), 8.70 (d, J = 7.9 Hz, 4H, *Tpy-H*^{3,3"}), 8.01 (d, J = 8.4 Hz, 4H, *Ph-H*^J), 7.89 (t, J = 7.7 Hz, 4H, *Tpy-H*^{4,4"}), 7.81 (d, J = 8.4 Hz, 4H, $Ph-H^{k}$), 7.39 – 7.33 (t, 4H, $Tpy-H^{5,5"}$), 6.99 (s, 2H, $Ph-H^{c}$), 3.91 (s, 3H, -OCH₃-H^a), 3.19 (s, 3H, -OCH₃-H^b); ¹³C NMR (101 MHz, CDCl₃, 300 K) δ 156.42, 155.86, 150.08, 149.06, 139.41, 137.41, 137.05, 136.02, 129.92, 127.28, 123.93, 121.48, 118.97, 115.50, 60.83, 55.87; ESI-TOF (m/z): Calcd. for $[C_{50}H_{36}N_6O_2+H]^+$: 753.88. Found: 753.88.



Compound 6, To a stirred solution of compound 5 (400 mg, 0.53 mmol) in CHCl₃ (30

mL) at room temperature, a solution of Br₂ (0.64 g, 4 mmol) in CHCl₃ (15 mL) was added drop-wise. After stirring at 90 °C for 48 h, the reaction mixture was washed with saturated Na₂SO₃ solution until colorless, dried over anhydrous MgSO₄, and then concentrated in vacuo to give the product as 310 mg light purple solid (yield 64%). ¹H NMR (400 MHz, CDCl₃) δ 8.81 (s, 4H, *Tpy-H^{3',5'}*), 8.74 (d, *J* = 5.6 Hz, 4H, *Tpy-H^{6,6''}*), 8.69 (d, *J* = 7.9 Hz, 4H, *Tpy-H^{3',3''}*), 8.00 (d, *J* = 8.3 Hz, 4H, *Ph-H^b*), 7.89 (t, *J* = 7.7 Hz, 4H, *Tpy-H^{4,4''}*), 7.53 (d, *J* = 8.3 Hz, 4H, *Ph-H^a*), 7.39 – 7.34 (t, 4H, *Tpy-H^{5,5''}*), 3.99 (s, 3H, *-OCH₃-H^d*), 3.06 (s, 3H, *-OCH₃-H^e*); ¹³C NMR (101 MHz, CDCl₃) δ 156.22, 155.99, 150.08, 149.18, 138.19, 137.61, 137.07, 130.59, 127.19, 123.94, 121.43, 119.50, 119.12, 77.33, 77.07, 76.75, 61.00, 60.60; ESI-TOF (*m/z*): Calcd. for [C₅₀H₃₆Br₂N₆O₂]: 910.67.



Compound 7: To a mixture of compound **6** (200 mg, 0.22 mmol) and compound **2** (502 mg, 0.66 mmol) in CHCl₃/MeOH (120 mL, v/v, 1:1), 500 µl of N-ethylmorpholine were added as reductant. After refluxing for 48 h, the solution was concentrated in vacuo to give a red solid that was chromatographed (Al₂O₃) using a mixed solvent of CH₂Cl₂/CH₃OH (100:1.75, v/v) to produce the product as 320 mg red powder (yield 62%). ESI-TOF (m/z): 555.37 [M-4Cl⁻]⁴⁺ (calcd m/z: 555.37), 752.33 [M-3Cl⁻]³⁺ (calcd m/z: 752.33), 1146.25 [M-2Cl⁻]²⁺ (calcd m/z: 1146.25); ¹H NMR (400 MHz, CD₃OD, 300 K) δ 9.40 (d, J = 6.5 Hz, 8H, Tpy-H^{4,B3'5'}), 8.95 (dd, J = 7.8, 4.1 Hz, 8H, Tpy-H^{4,B3,3''}), 8.54 (s, 2H, Ph-H^e), 8.47 (d, J = 8.1 Hz, 8H, Ph-H^{4,Ba}), 8.40 (d, J = 1.9 Hz, 2H, Ph-H^d), 8.16 (d, J = 8.3 Hz, 4H, Ph-H^{4b}), 8.05 (dd, J = 13.8, 5.9 Hz, 10H, Tpy-H^{4,B4,4''}, Ph-H^e), 7.84 – 7.78 (m, 6H, Ph-H^{Bb}, Ph-H^f), 7.69 (dd, J = 8.7, 2.0 Hz, 2H, Ph-

 H^{h}), 7.64 – 7.59 (m, 10H, *Tpy-H*^{4,B6,6"}, *Ph-H*^g), 7.33 – 7.29 (m, 8H, *Tpy-H*^{4,B5,5"}), 4.06 (d, J = 19.9 Hz, 6H, -*OCH*₃-*H*^{m,n}).



Ligand L: To a solution of 4'-(4-boronatophenyl)-[2,2':6',2"]terpyridine (179 mg, 0.508 mmol) and compound 7 (100 mg, 0.042 mmol) in CH₃CN/CH₃OH (40 mL, v/v, 1:1), aqueous K₂CO₃ (70.2 mg, 0.508 mmol, 0.5 mL) was added. The system was degassed for 10 minutes, and then Pd(PPh₃)₄ (38 mg, 0.034 mmol) was added. After refluxing for 4 days under N₂, the solvent was removed in vacuo to give a residue that was purified by flash column chromatography (Al₂O₃) with CH₂Cl₂/MeOH (100:4.5, v/v) to give L. Then LiNTf₂ was added to generate red precipitates, which were filtered and washed with H₂O and CH₃OH to obtain 75 mg reddish product (yield 43%). ESI-TOF (m/z): 783.88 [M-4 NTf₂]⁴⁺ (calcd m/z: 783.88), 1138.56 [M-3 NTf₂]³⁺ (calcd m/z: 1138.56); ¹H NMR (400 MHz, CD₃CN, 300 K) δ 9.11 (s, 4H, *Tpy-H*^{43'5'}), 9.03 (s, 4H, $Tpv-H^{B3'5'}$), 8.87 (s, 4H, $Tpv-H^{C3'5'}$), 8.80 (s, 4H, $Tpv-H^{D3'5'}$), 8.79 – 8.75 (m, 8H, $Tpv-H^{D3'5'}$), 8.75 (m, 8H, Tpv-H^{D3'5'}), 8.75 (m, 8H, T $H^{A,B3,3"}$), 8.69 (d, J = 8.4 Hz, 12H, $Tpy-H^{C,D3,3"}$, $Ph-H^{c.d}$), 8.63 (d, J = 7.7 Hz, 8H, $Tpy-H^{C,D3,3"}$) $H^{C,D6,6''}$), 8.39 (d, J = 8.4 Hz, 4H, $Tpy-H^{Aa}$), 8.21 (d, J = 3.6 Hz, 8H, $Tpy-H^{Ba,Ab}$), 8.11 $(d, J = 8.4 \text{ Hz}, 2H, Ph-H^e)$, 8.02 $(d, J = 8.1 \text{ Hz}, 8H, Tpy-H^{Ca,Da})$, 7.98 $(d, J = 8.0 \text{ Hz}, 2H, Ph-H^e)$ 8H, $Tpy-H^{A,B3,3"}$), 7.93 (dd, J = 8.5, 5.4 Hz, 16H, $Tpy-H^{C,D4,4",C,D6,6"}$), 7.87 – 7.82 (m, 4H, $Ph-H^{f,h}$), 7.77 (d, J = 8.4 Hz, 4H, $Tpv-H^{Ba}$), 7.65 (s, 2H, H^{g}), 7.51 – 7.47 (m, 8H, *Tpy-H*^{Cb,Db}), 7.44 – 7.41 (m, 8H, *Tpy-H*^{C,D5,5}"), 7.19 – 7.13 (m, 9H, *Tpy-H*^{4,B5,5}"), 3.22 (s, 3H, -*OCH*₃-*H*^m), 3.16 (s, 3H, -*OCH*₃-*H*ⁿ).



Zn₈L₄: To a solution of L (10 mg, 2.34 µmol) in CH₃CN (12 mL), Zn(NTf₂)₂ (2.94 mg, 4.68 µmol) in CH₃OH was added; then the mixture was stirred at 60 °C for 8 hours. Subsequently, LiNTf₂ was added to generate red precipitates, which were filtered and washed with H₂O and CH₃OH to obtain 11.3 mg reddish product (yield 98%). ESI-TOF (m/z): 1555.65 $[M-12NTf_2^{-1}]^{12+}$ (calcd m/z: 1555.65), 1414.44 $[M-13NTf_2^{-1}]^{13+}$ (calcd 1414.44), 1293.40 $[M-14NTf_2^{-1}]^{14+}$ (calcd m/z: 1293.40), 1188.49 m/z: $[M-15NTf_2^{-1}]^{15+}$ (calcd *m/z*: 1188.49), 1096.71 $[M-16NTf_2^{-1}]^{16+}$ (calcd *m/z*: 1096.71), 1015.72 $[M-17NTf_2^-]^{17+}$ (calcd *m/z*: 1015.72), 943.73 $[M-18NTf_2^-]^{18+}$ (calcd *m/z*: 943.73), 879.31 [M-19NTf₂]¹⁹⁺ (calcd *m/z*: 879.31);. ¹H NMR (400 MHz, CD₃CN, 300 K) δ 9.17 – 9.06 (br, 8H, *Tpy-H*^{*A,B,C,D3'5'*), 8.80 – 8.74 (br, *J* = 22.4 Hz, 10H, *Tpy-*} *H*^{*A*,*B*,*C*,*D*3,3^{*°*}, *Ph*-*H*^{*c.d*}), 8.47 – 8.39 (br, 4H, *Tpy*-*H*^{*A*,*B*4,4^{*°*}), 8.35 – 8.30 (br, 4H, *Tpy*-*H*^{*Ca*,*Da*}),}} 8.26 - 8.18 (br, 8H, Tpy-H^{A,B6,6"}, Tpy-H^{Cb,Db,Aa,Ba}), 8.12 - 8.09 (br, 2H, Ph-H^e), 8.01 -7.85 (br, J = 33.7 Hz, 14H, $Tpv-H^{A,B6,6",C,D4,4"}$, $Ph-H^h$), 7.52 – 7.38 (br, J = 4.2 Hz, 10H, $Tpy-H^{A,B5,5",C,D6,6"}$, $Ph-H^{g}$), 7.25 – 7.16 (br, 4H, $Tpy-H^{C,D5,5"}$), 3.45 (d, J = 7.0 Hz, 3H, $-OCH_{3}-H^{m,n}$).



Co₈L₄: To a solution of L (8 mg, 1.87 μ mol) in CH₃CN (2 mL), CoCl₂•6H₂O (0.89 mg, 3.76 μ mol) in CH₃OH was added; then the mixture was stirred at 60 °C for 8 hours. Subsequently, LiNTf₂ was added to generate red precipitates, which were filtered and washed with H₂O and CH₃OH to obtain 7.8 mg reddish product (98% yield). ESI-TOF

(*m/z*): 1989.75 [M–11 NTf₂⁻]¹¹⁺ (calcd *m/z*: 1989.75)1551.36 [M–12NTf₂⁻]¹²⁺ (calcd *m/z*: 1551.36), 1410.47 [M–13NTf₂⁻]¹³⁺ (calcd *m/z*: 1410.47), 1289.71 [M–14NTf₂⁻]¹⁴⁺ (calcd *m/z*: 1289.71), 1185.06 [M–15NTf₂⁻]¹⁵⁺ (calcd *m/z*: 1185.06), 1093.48 [M–16NTf₂⁻]¹⁶⁺ (calcd *m/z*: 1093.48), 1012.68 [M–17NTf₂⁻]¹⁷⁺ (calcd *m/z*: 1012.68), 940.86 [M–18NTf₂⁻]¹⁸⁺ (calcd *m/z*: 940.86), 876.60 [M–19NTf₂⁻]¹⁹⁺ (calcd *m/z*: 876.60).



4. ESI-MS spectra data of compounds (Cl⁻ or NTf₂⁻ as counterion)



Figure S1. ESI-MS of compound **7** (Cl^{-} as counterion).



Figure S2. ESI-MS of ligand L (Cl⁻ as counterion).



Figure S3. ESI-MS of ligand L (NTf_2^- as counterion).

5. ESI-MS spectra data of supramolecules (NTf2⁻ as counterion)



Figure S4. ESI-MS of L with $Cu(OTf)_2$ (NTf₂⁻ as counterion).



Figure S5. ESI-MS of L with $MnClO_4 \cdot 6H_2O$ (NTf_2^- as counterion).



Figure S6. Measured (bottom) and calculated (top) isotope patterns for different charge states observed from $[Co_8L_4]$ (NTf₂⁻ as counterion).



Figure S7. Measured (bottom) and calculated (top) isotope patterns for different charge states observed from $[Zn_8L_4]$ (NTf₂⁻ as counterion).



Figure S8. Measured (bottom) and calculated (top) isotope patterns for different charge states observed from $[Cd_6L_3]$ (NTf₂⁻ as counterion).



Figure S9. gMS² of (a) Co₈L₄ at m/z 1410.6 with different collision energies, (b) Zn₈L₄ at m/z 1414.6 with different collision energies, (c) Cd₆L₃ at m/z 1013.1 with different collision energies.

6. ¹H NMR, ¹³C NMR, 2D COSY NMR, 2D NOESY NMR, 2D DOSY NMR



Figure S10. ¹H NMR (600 MHz, CDCl₃, 300 K) spectrum of compound 1.

156.491 156.323 156.323 155.543 155.543 155.543 155.543 155.543 155.543 155.543 155.543 155.543 149.904 149.907 149.206 137.132 137.13	1305,201 (129,162) (129,162) (129,162) (129,005) (129,005) (129,007) (127,979) (127,97
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Figure S11. ¹³C NMR (126 MHz, CDCl₃, 300 K) spectrum of compound 1.



Figure S12. ¹H NMR (400 MHz, CDCl₃, 300 K) spectrum of compound 4.



Figure S13. ¹³C NMR (126 MHz, CDCl₃, 300 K) spectrum of compound 4.



Figure S15. ¹³C NMR (126 MHz, CDCl₃, 300 K) spectrum of compound 5.



170 160 150 140 130 120 110 100 90 80 70 60 50 40 Chemical shift (ppm)

Figure S17. ¹³C NMR (126 MHz, CDCl₃, 300 K) spectrum of compound 6.



Figure S18. ¹H NMR (400 MHz, CD₃OD, 300 K) spectrum of compound 7.



Figure **S19.** 2D COSY NMR (400 MHz, CD₃OD, 300 K) spectrum of compound 7 (aromatic region).



Figure S20. 2D NOESY NMR (400 MHz, CD₃OD, 300 K) spectrum of compound 7 (aromatic region).



Figure S21. ¹H NMR (400 MHz, CD₃CN, 300 K) spectrum of ligand L.



Figure S22. 2D COSY NMR (400 MHz, CD₃OD, 300 K) spectrum of ligand L (aromatic region).



Figure S23. 2D NOESY NMR (400 MHz, CD₃OD, 300 K) spectrum of ligand L (aromatic region).





Figure S24. ¹H NMR (500 MHz, CD₃CN, 300 K) spectrum of Zn₈L₄.



Figure S25. 2D COSY NMR (500 MHz, CD₃CN, 300 K) spectrum of **Zn₈L**₄ (all cross-peaks of Tpy-Ru-Tpy are illustrated by a blue line, all cross-peaks of Tpy-Zn-Tpy are illustrated by pink line).



Figure S26. 2D NOESY NMR (500 MHz, CD₃CN, 300 K) spectrum of **ZnsL**₄ (all cross-peaks of Tpy-Ru-Tpy are illustrated by a blue line, all cross-peaks of Tpy-Zn-Tpy are illustrated by pink line).



Figure S27. ¹H NMR (500 MHz, CD₃CN, 300 K) spectrum of Cd₆L₃.



Figure S28. 2D NOESY NMR (500 MHz, CD₃CN, 300 K) spectrum of Cd_6L_3 (all cross-peaks of Tpy-Ru-Tpy are illustrated by a blue line, all cross-peaks of Tpy-Zn-Tpy are illustrated by pink line).



Figure S29. DOSY (500 MHz, CD₃CN, 300 K) spectra of Zn₈L₄.

Specific calculation process

The sphere's hydrodynamic radius was estimated according to the Stokes-Einstein Equation, where **D** is the diffusion constant, **k** is the Boltzmann's constant, **T** is the temperature, μ is the viscosity of solvents, and **R** is the radius:

$$D = \frac{kT}{6\pi\mu R}$$

$$D = 10^{-9.85} \text{ m}^2 \text{ s}^{-1}$$

$$k = 1.38 \times 10^{-23} \text{ N m K}^{-1}$$

$$T = 298 \text{ K}$$

$$\mu = 3.43 \times 10^{-4} \text{ N m}^{-2} \text{ s } (\text{CD}_3 \text{CN})^{2,3}$$

$$R = \frac{kT}{6\pi\mu D} \approx 2.11 \times 10^{-9} \text{ m} = 2.11 \text{ nm}$$

The radius of $\mathbb{Z}n_8L_4$ was determined to be 4.22 nm, which is consistent with the results of computer modeling.



Figure S30. DOSY (500 MHz, CD₃CN, 300 K) spectra of Cd₆L₃.

Specific calculation process

The sphere's hydrodynamic radius was estimated according to the Stokes-Einstein Equation, where **D** is the diffusion constant, **k** is the Boltzmann's constant, **T** is the temperature, μ is the viscosity of solvents, and **R** is the radius:

$$D = \frac{kT}{6\pi\mu R}$$

$$D = 10^{-9.70} \text{ m}^2 \text{ s}^{-1}$$

$$k = 1.38 \times 10^{-23} \text{ N m K}^{-1}$$

$$T = 298 \text{ K}$$

$$\mu = 3.43 \times 10^{-4} \text{ N m}^{-2} \text{ s (CD_3CN)}$$

$$R = \frac{kT}{6\pi\mu D} \approx 1.62 \times 10^{-9} \text{ m} = 1.62 \text{ nm}$$

The radius of Cd_6L_3 was determined to be 3.24 nm, which is consistent with the results of computer modeling.

7. TEM and AFM images of supramolecules (NTf₂⁻ as counterion)



Figure S31. TEM images of (a) Co_8L_4 , (b) Zn_8L_4 , and (a) Cd_6L_3 on the lacey carboncoated Cu grid (c=10⁻⁶ M in CH₃CN).



Figure S32. TEM size statistical histogram of (a) CosL4, (b) ZnsL4, and (c) Cd6L3.



Figure S33. AFM 2D images of (a) Co₈L₄, (b) Zn₈L₄, and (c) Cd₆L₃.⁴



Figure S34. AFM height statistical histogram for 100 particles of (a) Co₈L₄, (b) Zn₈L₄, and (c) Cd₆L₃.



8. UV-vis and emission of L and supramolecules (NTf₂⁻ as counterion)

Figure S35. UV-vis $(10^{-6} \text{ M in CH}_3 \text{CN}, \text{ room temperature})$ of L and supramolecules.



Figure S36. Emission (10⁻⁶ M in CH₃CN, 73K) of L and supramolecules.

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