

Supporting Information

Fe^{III}/TBHP mediated remote C-O bond construction of 8-aminoquinolines: access to methoxylation and cyanomethoxylation

*Mengfei Zhao, Kaixin Zhang, Jianxiong Xu, Jizhen Li ^{*a,b}*

*^a Department of Organic Chemistry, College of Chemistry, Jilin University, 2519 Jiefang Road, Changchun,
130021, P. R. China*

*^b State Key Laboratory of Rare Earth Resource Utilization, Changchun Institute of Applied Chemistry,
Chinese Academy of Sciences, Changchun, 130022, P.R. China*

Corresponding author E-mail: ljz@jlu.edu.cn

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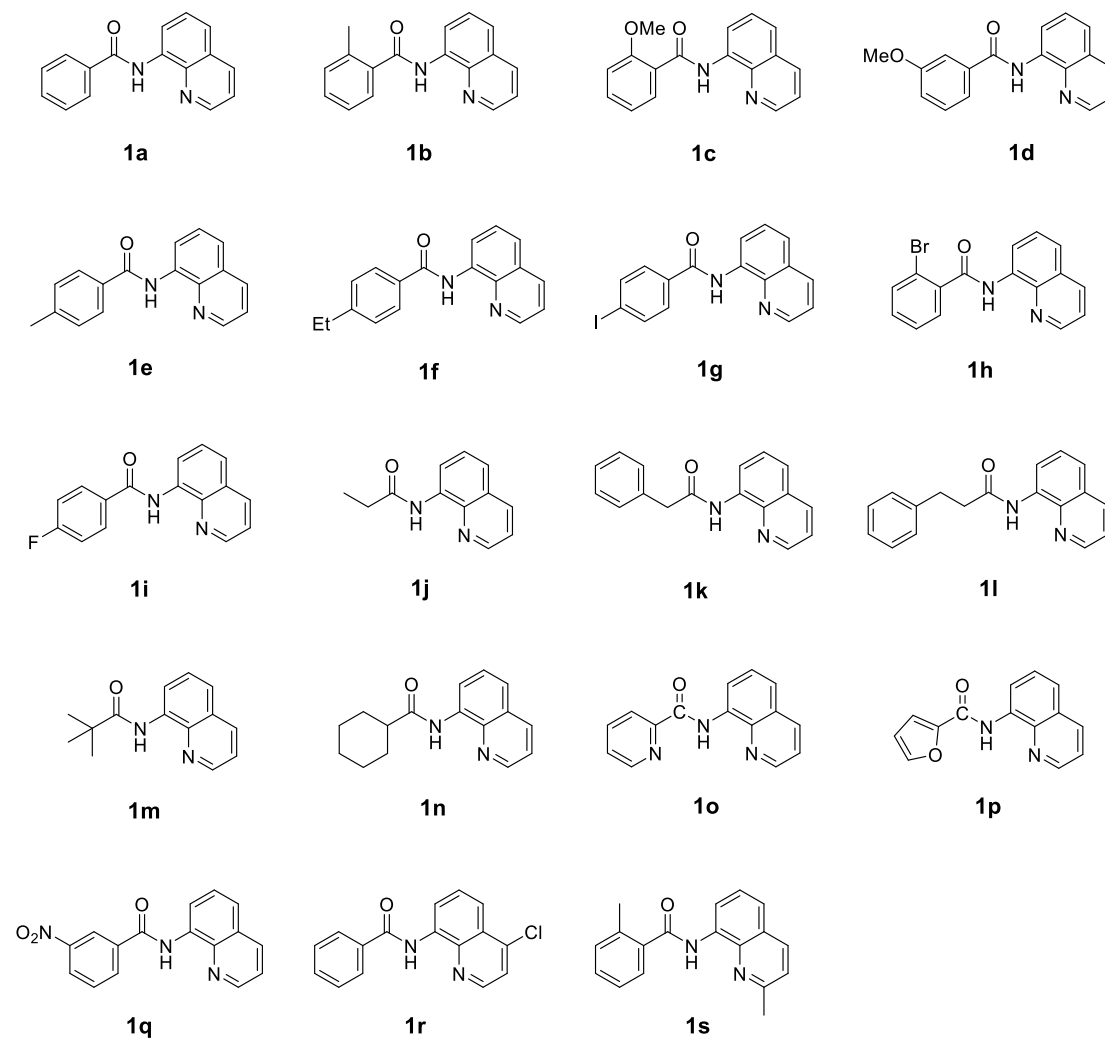
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1. General information¹

All reagents, starting materials, and solvents were purchased from commercial sources and used without treatment unless otherwise indicated. All the solvents were dried and newly distilled. NMR spectra were obtained on a Bruker AMX 400 system using chloroform-d as deuterated solvents. The ¹H-NMR spectra were recorded at 400 MHz in CDCl₃, and the ¹³C-NMR spectra were recorded at 101 MHz in CDCl₃. All shifts were given in ppm. All coupling constants (*J* values) were reported in Hertz (Hz). High-Resolution Liquid Chromatography-Mass Spectrometry was recorded on the Bruker MicrOTOF QII. Column chromatography was performed on silica gel 100-200 mesh or 200-300 mesh. Ethyl acetate and petroleum ether were used for column chromatography.

2. Preparation of starting materials¹

Preparation of starting materials: Aromatic amine (5.0 mmol, 1.0 equiv) was dissolved in 10 mL of dichloromethane and cooled to 0 °C using an ice bath. NEt₃ (6.0 mmol, 1.2 equiv) was added to the aniline solution followed by the corresponding acid chloride (6.0 mmol, 1.2 equiv) dropwise. The mixture was stirred for 10 h at room temperature. Then, the mixture was washed with sat. NaHCO₃ (50 mL), and was extracted with dichloromethane three times (3 x 40 mL). The organic layer was dried over Na₂SO₄. After filtration and evaporation, the amides were purified by column chromatography through silica gel.

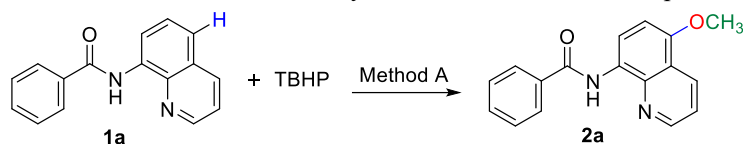


^a indolin-2-one **1r** and **1s** were purchased from *Energy Chemical*.

3. Experimental section

3.1 Optimization of reaction conditions

Table S1 Optimization for selective methoxylation reaction of 8-aminoquinolines^a



Entry	T (°C)	catalyst	solvent	t (h)	additive	yield ^b
1	140	dppf(NiCl ₂)	PhCF ₃	4		27
2	80	dppf(NiCl ₂)	PhCF ₃	4		trace
3	100	dppf(NiCl ₂)	PhCF ₃	4		6
4	120	dppf(NiCl ₂)	PhCF ₃	4		17
5	150	dppf(NiCl ₂)	PhCF ₃	4		25
6	140	NiCl ₂	PhCF ₃	4		n.r.
7	140	FeCl ₃	PhCF ₃	4		36
8	140	K ₃ Fe(CN) ₆	PhCF ₃	4		n.r.
9	140	FeF ₂	PhCF ₃	4		trace
10	140	FeCl ₂	PhCF ₃	4		22
11	140	Ni(OTf) ₂	PhCF ₃	4		n.r.
12	140	NiSO ₄	PhCF ₃	4		n.r.
13	140	Sc(OTf) ₃	PhCF ₃	4		n.r.
14	140	CeCl ₃	PhCF ₃	4		n.r.
15	140	FeCl ₃	DMSO	4		trace
16	140	FeCl ₃	DCM	4		6
17	140	FeCl ₃	CH ₃ CN	4		26/24 ^c
18	140	FeCl ₃	EtOH	4		n.r.
19	140	FeCl ₃	DMF	4		n.r.
20	140	FeCl ₃	DCE	4		n.r.
21	140	FeCl ₃	toluene	4		n.r.
22	140	FeCl ₃	TFE	4		45
23	140	FeCl ₃	TFE	8		47
24	140	FeCl ₃	TFE	12		46
25 ^d	140	FeCl ₃	TFE	8		39
26 ^e	140	FeCl ₃	TFE	8		46
27	140	FeCl ₃	TFE	8	PPh ₃ (2.0 eq)	48
28	140	FeCl ₃	TFE	8	PivOH (0.2 eq)	64
29	140	FeCl ₃	TFE	8	PivOH (0.5 eq)	67
30	140	FeCl₃	TFE	8	PivOH (1.0 eq)	69
31	140	FeCl ₃	TFE	8	PivOH (2.0 eq)	61
32 ^f	140	FeCl ₃	TFE	8	PivOH (2.0 eq)	61
33 ^g	140	FeCl ₃	TFE	8	PivOH (2.0 eq)	61
34	140	FeCl ₃	TFE	8	NaHCO ₃ (2.0 eq)	34
35 ^h	140	FeCl ₃	TFE	8	PivOH (1.0 eq)	16
36	130	FeCl ₃	TFE	8	PivOH (1.0 eq)	64
37	120	FeCl ₃	TFE	8	PivOH (1.0 eq)	52

^a Reaction conditions: **1a** (0.2 mmol, 1.0 equiv), TBHP (70% in water, 4.0 equiv), catalyst (5 mol%), solvent (1.5 mL), in sealed tube (10 mL). ^b Isolated yield. ^c 24% yield of C-5 cyanomethylation by-product. ^d TBHP (5.0–6.0 mol/L in decane, 4.0 eq). ^e FeCl₃ (10 mol%). ^f under N₂. ^g under O₂. ^h DTBP (4.0 eq) instead of TBHP.

Table S2 Optimization for cyanomethoxylation reaction of quinoline amides^a

c1ccc(cc1)C(=O)Nc2ccc3c(c2)ncn3 + TBHP $\xrightarrow{\text{Method B}}$ c1ccc(cc1)C(=O)Nc2ccc3c(c2)ncn3COCC#N

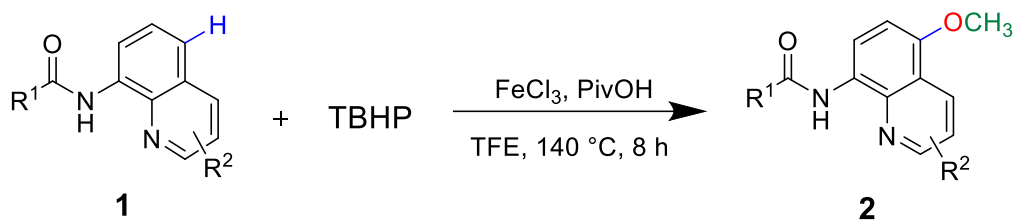
1a **3a**

Entry	T(°C)	catalyst	solvent	additive	t (h)	yield ^b
1	70	FeCl ₂	CH ₃ CN		12	6
2	70	FeCl ₃	CH ₃ CN		12	8
3	80	FeCl ₃	CH ₃ CN		12	17
3	85	FeCl ₃	CH ₃ CN		12	18
4	90	FeCl ₃	CH ₃ CN		12	18/6 ^c
5	100	FeCl ₃	CH ₃ CN		12	24/16 ^c
6	80	FeF ₂	CH ₃ CN		12	n.r.
7	80	NiCl ₂	CH ₃ CN		12	n.r.
8	80	NiSO ₄	CH ₃ CN		12	n.r.
9	80	CoCl ₂	CH ₃ CN		12	n.r.
10	80	FeCl ₃	CH ₃ CN/TFE (1/1)		12	6
11	80	FeCl ₃	CH ₃ CN /PhCF ₃ (1/1)		12	8
12	80	FeCl ₃	CH ₃ CN		8	14
13	80	FeCl ₃	CH ₃ CN		24	33
14	80	FeCl ₃	CH ₃ CN		48	32
15 ^d	80	FeCl ₃	CH ₃ CN		24	42
16 ^d	80	FeCl ₃ (5 mol%)	CH ₃ CN		24	16
17 ^d	80	FeCl ₃	CH ₃ CN	PivOH (2.0 eq)	24	35
18 ^d	80	FeCl ₃	CH ₃ CN	NaHCO ₃ (2.0 eq)	24	trace
19 ^d	80	FeCl ₃	CH ₃ CN	N ₂	24	37
20 ^d	80	FeCl ₃	CH ₃ CN	O ₂	24	38
21 ^d	80	FeCl ₃ /FeCl ₂ (5/5 mol%)	CH ₃ CN		24	41
22 ^d	80	FeCl ₃ (20 mol%)	CH ₃ CN		24	39
23 ^{d, e}	80	FeCl ₃	CH ₃ CN		24	trace
24 ^{d, f}	80	FeCl ₃	CH ₃ CN		24	41
24 ^g	80	FeCl ₃	CH ₃ CN		24	25
24 ^h	80	FeCl ₃	CH ₃ CN		24	39
24^g	85	FeCl₃	CH₃CN (3 mL)		24	50

^a Reaction conditions: **1a** (0.2 mmol, 1.0 equiv), TBHP (70% in water, 0.4 mmol, 4.0 equiv), catalyst (10 mol%), solvent (1.5 mL), in sealed tube (10 mL). ^b Isolated yield. ^c yield of C-5 methoxylation by-product. ^d TBHP (5.0-6.0 mol/L in decane, 4.0 eq). ^e DTBP (4.0 eq) instead of TBHP. ^f **1a** (0.5 mmol, 1.0 equiv). ^g TBHP (5.0-6.0 mol/L in decane, 8.0 eq). ^h TBHP (5.0-6.0 mol/L in decane, 2.0 eq).

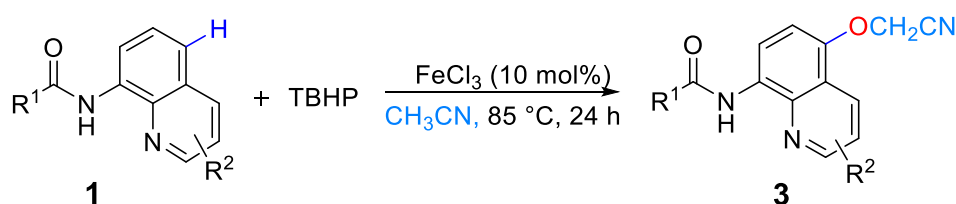
3.2 General procedure

3.2.1 General procedure for C-5 methoxylation



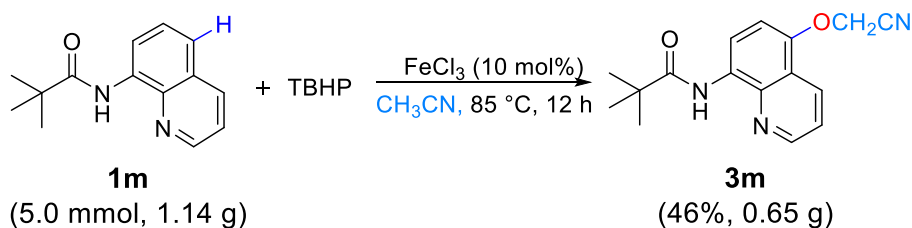
1 (0.20 mmol, 1.0 equiv), TBHP (4.0 equiv, 70% in water), FeCl₃ (5 mol%), PivOH (1.0 eq) were mixed in TFE (1.5 mL) and stirred in a dried sealed tube (10 mL) at 140 °C for 8 h. After completion of the reaction (TLC monitored), it was cooled to room temperature and quenched with 1 mol/L Na₂SO₃ solution (10 mL, for removal of excess TBHP). Then the mixture was extracted with ethyl acetate (3 x 5 mL) and the organic layer was transferred to a round bottom flask after being dried over anhydrous Na₂SO₄. The solvent was concentrated under reduced pressure and further purified by flash chromatography (SiO₂, petroleum ether/ethyl acetate gradient), yielding the target products **2**.

3.2.2 General procedure for C-5 cyanomethoxylation



1 (0.20 mmol, 1.0 equiv), TBHP (8.0 equiv, 5-6 mol/L in decane), FeCl₃ (10 mol%) were mixed in CH₃CN (3.0 mL) and stirred in a dried sealed tube (10 mL) at 85 °C for 24 h. After completion of the reaction (TLC monitored), it was cooled to room temperature and quenched with 1 mol/L Na₂SO₃ solution (10 mL, for removal of excess TBHP). Then the mixture was extracted with ethyl acetate (3 x 5 mL) and the organic layer was transferred to a round bottom flask after being dried over anhydrous Na₂SO₄. The solvent was concentrated under reduced pressure and further purified by flash column chromatography (SiO₂, petroleum ether/ethyl acetate gradient), yielding the target products **2**.

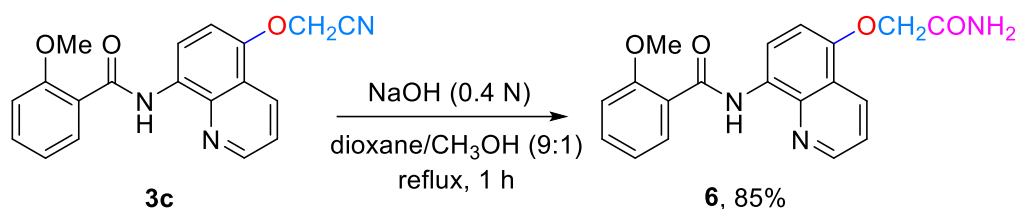
3.2.3 General procedure for gram-scale reaction of cyanomethoxylation



1m (5.0 mmol, 1.0 equiv), TBHP (5-6 mol/L in decane, 2.0 mL), FeCl₃ (10 mol%) were mixed in CH₃CN (10.0 mL) and stirred in a dried sealed tube at 85 °C for 12 h. After completion of the

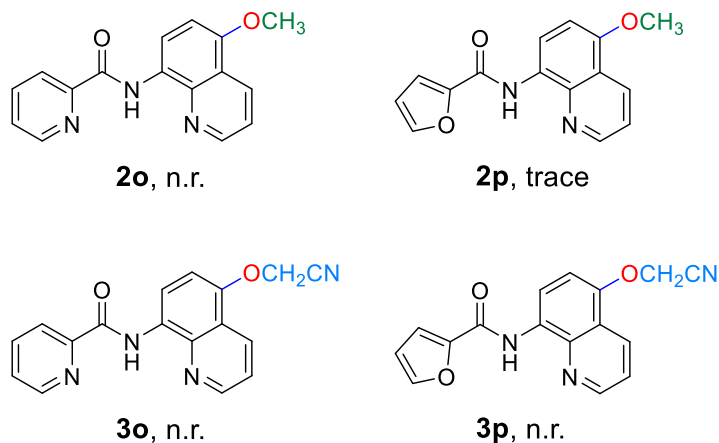
reaction (TLC monitored), it was cooled to room temperature and quenched with 1 mol/L Na_2SO_3 solution (20 mL, for removal of excess TBHP). Then the mixture was extracted with ethyl acetate (3 x 20 mL) and the organic layer was transferred to a round bottom flask after being dried over anhydrous Na_2SO_4 . The solvent was concentrated under reduced pressure and further purified by flash column chromatography (SiO_2 , petroleum ether/ethyl acetate gradient), yielding the target product **3m** 0.65g.

3.2.4 General procedure for the transformation of cyanomethoxylation compound



3c (0.5 mmol), NaOH (2 mmol, dissolved in 0.5 mL CH_3OH) were mixed in dioxane (4.5 mL) and the mixture was refluxed at 80 °C for 1 h. The course of the reaction was monitored by TLC analysis. Then the reaction mixture was cooled and evaporated to dryness, diluted with ethyl acetate and washed with water. The organic phase was concentrated after being dried over anhydrous Na_2SO_4 and purified by column chromatography chromatography (SiO_2 , petroleum ether/ethyl acetate gradient), yielding amide **6** with 149 mg.

4 Tolerance of this alkoxylation strategy to heteroaryl carboxamides substrates



5. The single crystal X-ray diffraction data

5.1 Methoxylation product N-[5-(methoxyl)-8-quinolinyl]-2- phenyl-propionamide **2l**

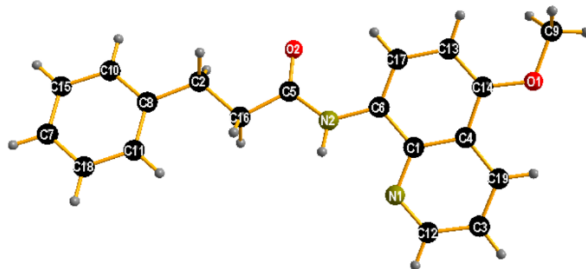


Table S3 Crystal data and structure refinement for **2l**

Bond precision: C-C = 0.0051 Å Wavelength = 26.474(3)

Cell: a = 5.3192(6) b = 10.9600(14) c = 26.474(3)

alpha = 90 beta = 90 gamma = 90

Temperature: 180 K

	Calculated	Reported
Volume	1543.4(3)	1543.4(3)
Space group	P 21/c	P 1 21/c 1
Hall group	-P 2ybc	-P 2ybc
Moiety formula	C ₁₉ H ₁₈ N ₂ O ₂	C ₁₉ H ₁₈ N ₂ O ₂
Sum formula	C ₁₉ H ₁₈ N ₂ O ₂	C ₁₉ H ₁₈ N ₂ O ₂
Mr	306.35	306.35
Dx (g · cm ⁻³)	1.318	1.318
Z	4	4
Mu (mm ⁻¹)	0.087	0.087
F000	648.0	648.0
F000'	648.27	

h, k, lmax 6,13,31 6,13,31

Nref 2798 2204

Tmin, Tmax 0.978,0.989 0.542,0.745

Tmin' 0.978

Correction method = # Reported T Limits: Tmin = 0.542, Tmax = 0.745,

AbsCorr = MULTI-SCAN

Data completeness = 0.788 Theta(max) = 25.234

R(reflections) = 0.0705(1575) wR2(reflections)= 0.1949(2204)

S = 1.025 Npar = 209

5.2 Cyanomethoxylation product N-[5-(cyanomethoxyl)-8-quinoliny]-2,2-dimethyl- propionamide 3m

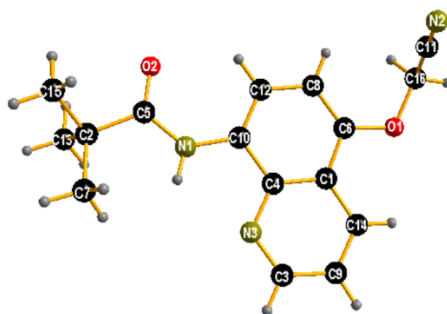


Table S4 Crystal data and structure refinement for **3m**

Bond precision: C-C = 0.0024 Å Wavelength = 0.71073

Cell: a=7.8041(6) b=21.7738(13) c=9.3578(8)

alpha=90 beta=112.022(3) gamma=90

Temperature: 180 K

	Calculated	Reported
Volume	1474.11(19)	1474.11(19)
Space group	P 21/c	P 1 21/c 1

Hall group	-P 2ybc	-P 2ybc
Moiety formula	C ₁₆ H ₁₇ N ₃ O ₂	C ₁₆ H ₁₇ N ₃ O ₂
Sum formula	C ₁₆ H ₁₇ N ₃ O ₂	C ₁₆ H ₁₇ N ₃ O ₂
Mr	283.33	283.32
Dx (g • cm ⁻³)	1.277	1.277
Z	4	4
Mu (mm ⁻¹)	0.086	0.086
F000	600.0	600.0
F000'	600.25	
h,k,lmax	9,25,11	9,25,11
Nref	2621	2607
Tmin,Tmax	0.983,0.987	0.638,0.745
Tmin'	0.981	

Correction method= # Reported T Limits: Tmin=0.638, Tmax=0.745

AbsCorr = MULTI-SCAN

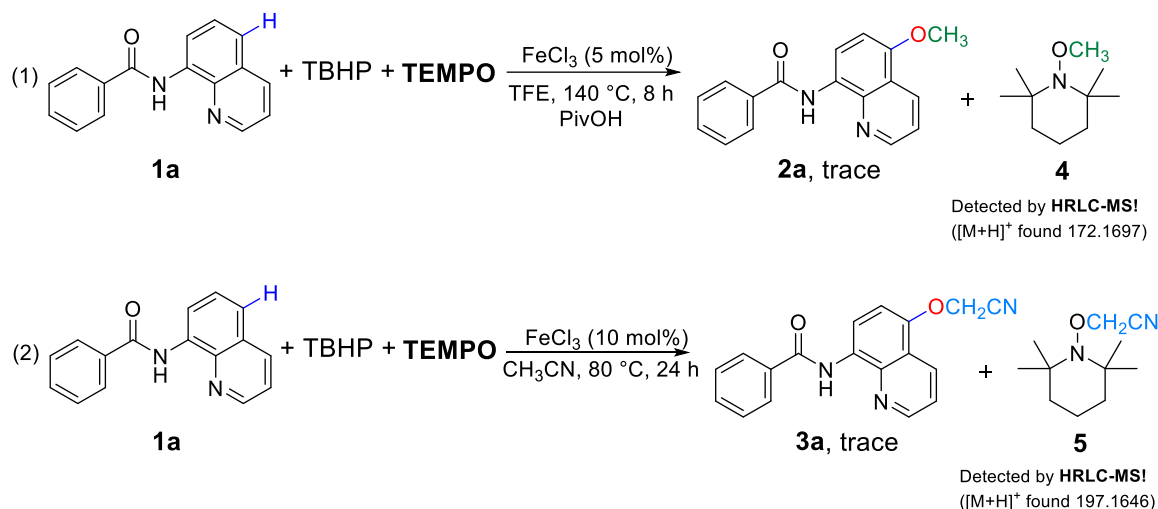
Data completeness= 0.995 Theta(max)= 25.068

R(reflections)= 0.0405(2005) wR2(reflections)= 0.1197(2607)

S = 1.079 Npar= 193

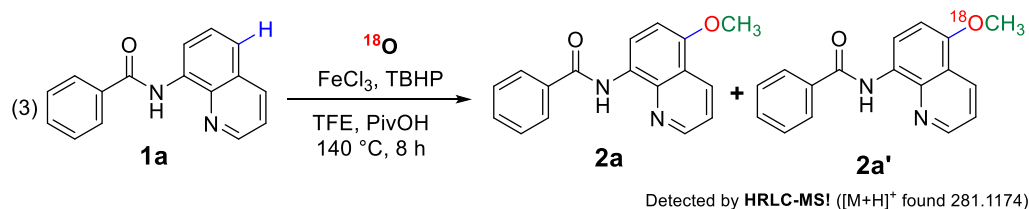
6. Mechanism investigation

6.1 Control experiments



- (1) **1a** (0.10 mmol, 1.0 equiv), TBHP (4.0 equiv, 70% in water), FeCl₃ (5 mol%), PivOH (2.0 equiv), Tempo (3.0 equiv) were mixed in TFE (1.5 mL) and stirred in a dried sealed tube (10 mL) at 140 °C for 8 h. After completion of the reaction (TLC monitored), it was cooled to room temperature and quenched with a saturated solution of Na₂SO₃ (10 mL, for removal of excess TBHP). Then the mixture was extracted with ethyl acetate (3 x 5 mL) and the organic layer was transferred to a round bottom flask after being dried over anhydrous Na₂SO₄. The mixture was then filtered by flash chromatography (SiO₂, petroleum ethyl acetate gradient) and detected by LC-HRMS.
- (2) **1a** (0.10 mmol, 1.0 equiv), TBHP (4.0 equiv, 5-6 mol/L in decane), FeCl₃ (10 mol%), Tempo (3.0 equiv) were mixed in CH₃CN (1.5 mL) and stirred in a dried sealed tube (10 mL) at 80 °C for 24 h. After completion of the reaction (TLC monitored), it was cooled to room temperature and quenched with a saturated solution of Na₂SO₃ (10 mL, for removal of excess TBHP). Then the mixture was extracted with ethyl acetate (3 x 5 mL) and the organic layer was transferred to a round bottom flask after being dried over anhydrous Na₂SO₄. The mixture was then filtered by flash chromatography (SiO₂, petroleum ethyl acetate gradient) and detected by LC-HRMS.

6.2 ¹⁸O₂ Isotope labeling experiments



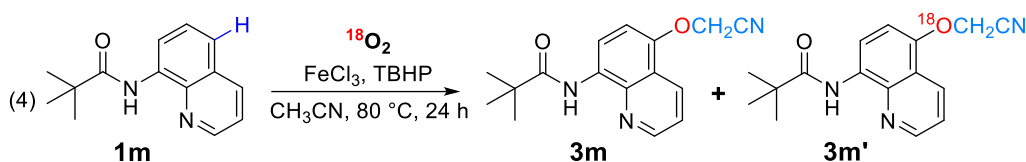
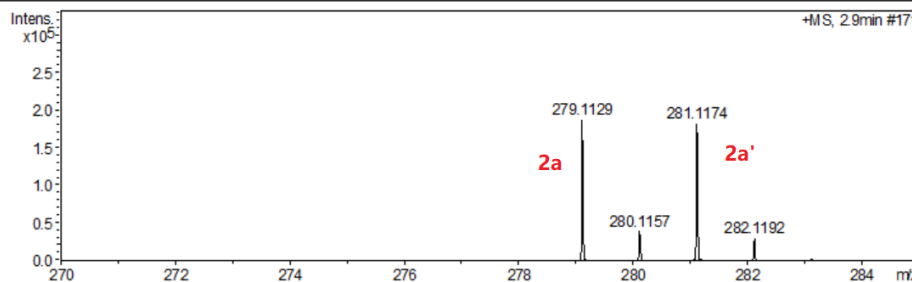
Mass Spectrum SmartFormula Report

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Method: lc-ms-hr-low.m	Operator: zlwei
Sample Name: Sample	Instrument / Ser#: microTOF-Q II 10351
Comment:	

Acquisition Parameter

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Detected by HR/LC-MS! ([M+H]⁺ found 286.1432)

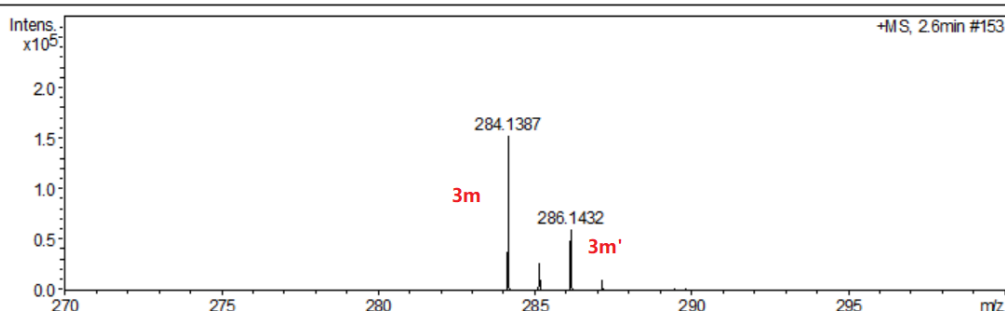
Mass Spectrum SmartFormula Report

Analysis Info

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Method: lc-ms-hr-low.m	Operator: zlwei
Sample Name: Sample	Instrument / Ser#: microTOF-Q II 10351
Comment:	

Acquisition Parameter

Source Type: ESI	Ion Polarity: Positive	Set Nebulizer: 0.6 Bar	Set Dry Gas: 8.0 l/min
Focus: Active	Set Capillary: 4500 V	Set Dry Heater: 200 °C	
Scan Begin: 50 m/z	Set End Plate Offset: -500 V	Set Dry Gas: 8.0 l/min	
Scan End: 1200 m/z	Set Collision Cell RF: 100.0 Vpp	Set Divert Valve: Waste	



Scheme S1 Investigation of the radical pathway

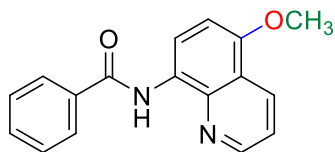
- (3) **1a** (0.10 mmol, 1.0 equiv), TBHP (4.0 equiv, 70% in water), FeCl₃ (5 mol%), were mixed in TFE (1.5 mL) under ¹⁸O₂ and stirred in a dried schlenk tube (10 mL) at 140 °C for 8 h. After completion of the reaction (TLC monitored), it was cooled to room temperature and quenched with a saturated solution of Na₂SO₃ (10 mL, for removal of excess TBHP). Then the mixture was **extracted** with ethyl acetate (3 x 5 mL) and the organic layer was transferred to a round bottom flask after being dried over anhydrous Na₂SO₄. The mixture was then filtered by flash

chromatography (SiO₂, petroleum ethyl acetate gradient) and detected by LC-HRMS (**2a**: **2a'** = 1:1).

- (4) **1a** (0.10 mmol, 1.0 equiv), TBHP (4.0 equiv, 5-6 mol/L in decane), FeCl₃ (10 mol%), Tempo (3.0 equiv) were mixed in CH₃CN (1.5 mL) under ¹⁸O₂ and stirred in a dried schlenk tube (10 mL) at 80 °C for 24 h. After completion of the reaction (TLC monitored), it was cooled to room temperature and quenched with a saturated solution of Na₂SO₃ (10 mL, for removal of excess TBHP). Then the mixture was extracted with ethyl acetate (3 x 5 mL) and the organic layer was transferred to a round bottom flask after being dried over anhydrous Na₂SO₄. The mixture was then filtered by flash chromatography (SiO₂, petroleum ethyl acetate gradient) and detected by LC-HRMS (**3m**: **3m'** = 2.5:1).

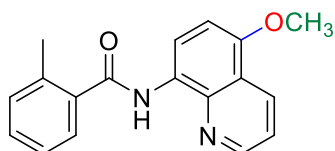
7. Characterization data of products

2a



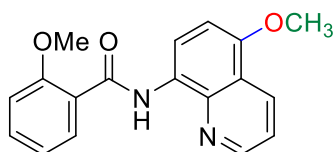
White solid, isolated yield: 69%; ^1H NMR (400 MHz, CDCl_3) δ 10.51 (s, 1H), 8.87 (dd, $J = 4.9$, 3.7 Hz, 2H), 8.61 (dd, $J = 8.4$, 1.2 Hz, 1H), 8.10 – 8.05 (m, 2H), 7.58 – 7.51 (m, 3H), 7.47 (dd, $J = 8.4$, 4.2 Hz, 1H), 6.90 (d, $J = 8.6$ Hz, 1H), 4.02 (s, 3H). ^{13}C NMR (101 MHz, CDCl_3) δ 165.13 (s), 150.44 (s), 148.75 (s), 139.47 (s), 135.37 (s), 131.61 (s), 131.32 (s), 128.75 (s), 128.01 (s), 127.20 (s), 120.80 (s), 120.52 (s), 116.72 (s), 104.39 (s), 55.80 (s). HRMS (ESI): m/z : calcd for $[\text{M}+\text{H}]^+$ $\text{C}_{17}\text{H}_{15}\text{N}_2\text{O}_2$: 279.1128, found: 279.1134.

2b



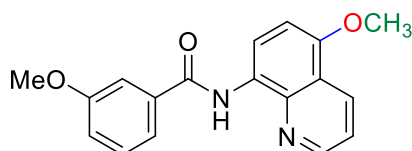
White solid, isolated yield: 71%; ^1H NMR (400 MHz, CDCl_3) δ 9.96 (s, 1H), 8.86 (d, $J = 8.5$ Hz, 1H), 8.78 (dd, $J = 4.2$, 1.7 Hz, 1H), 8.59 (dd, $J = 8.4$, 1.7 Hz, 1H), 7.67 (d, $J = 7.6$ Hz, 1H), 7.44 (dd, $J = 8.4$, 4.2 Hz, 1H), 7.41 – 7.37 (m, 1H), 7.31 (t, $J = 7.4$ Hz, 2H), 6.90 (d, $J = 8.6$ Hz, 1H), 4.02 (s, 3H), 2.60 (s, 3H). ^{13}C NMR (101 MHz, CDCl_3) δ 167.89 (s), 150.50 (s), 148.72 (s), 139.33 (s), 136.89 (s), 136.57 (s), 131.30 (d, $J = 1.5$ Hz), 130.13 (s), 128.19 (s), 127.25 (s), 125.97 (s), 120.78 (s), 120.52 (s), 116.70 (s), 104.33 (s), 55.84 (s), 20.21 (s). HRMS (ESI): m/z : calcd for $[\text{M}+\text{H}]^+$ $\text{C}_{18}\text{H}_{17}\text{N}_2\text{O}_2$: 293.1285, found: 293.1284.

2c



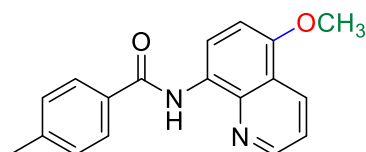
White solid, isolated yield: 70%; ^1H NMR (400 MHz, CDCl_3) δ 12.14 (s, 1H), 8.97 (d, $J = 8.6$ Hz, 1H), 8.89 (dd, $J = 4.1$, 1.5 Hz, 1H), 8.60 (dd, $J = 8.4$, 1.5 Hz, 1H), 8.36 (dd, $J = 7.8$, 1.7 Hz, 1H), 7.52 – 7.48 (m, 1H), 7.45 (dd, $J = 8.4$, 4.2 Hz, 1H), 7.14 (t, $J = 7.5$ Hz, 1H), 7.08 (d, $J = 8.3$ Hz, 1H), 6.90 (d, $J = 8.6$ Hz, 1H), 4.20 (s, 3H), 4.01 (s, 3H). ^{13}C NMR (101 MHz, CDCl_3) δ 157.69 (s), 150.27 (d, $J = 4.2$ Hz), 148.73 (s), 139.95 (s), 132.86 (s), 132.25 (s), 131.10 (s), 129.19 (s), 121.27 (s), 120.56 (s), 117.47 (s), 111.56 (s), 104.61 (s), 56.12 (s), 55.78 (s). HRMS (ESI): m/z : calcd for $[\text{M}+\text{H}]^+$ $\text{C}_{18}\text{H}_{17}\text{N}_2\text{O}_3$: 309.1234, found: 309.1243.

2d



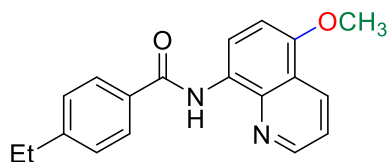
White solid, isolated yield: 71%; ^1H NMR (400 MHz, CDCl_3) δ 10.49 (s, 1H), 8.87 – 8.82 (m, 2H), 8.59 (dd, $J = 8.4, 1.7$ Hz, 1H), 7.62 (dd, $J = 4.3, 2.4$ Hz, 2H), 7.47 – 7.42 (m, 2H), 7.10 (dd, $J = 7.9, 2.5$ Hz, 1H), 6.89 (d, $J = 8.6$ Hz, 1H), 4.01 (s, 3H), 3.91 (s, 3H). ^{13}C NMR (101 MHz, CDCl_3) δ 164.94 (s), 159.95 (s), 150.46 (s), 148.77 (s), 139.46 (s), 136.86 (s), 131.30 (s), 129.73 (s), 127.98 (s), 120.81 (s), 120.51 (s), 118.99 (s), 117.80 (s), 116.71 (s), 112.56 (s), 104.36 (s), 55.80 (s), 55.51 (s). HRMS (ESI): m/z : calcd for $[\text{M}+\text{H}]^+$ $\text{C}_{18}\text{H}_{17}\text{N}_2\text{O}_3$: 309.1234, found: 309.1238.

2e



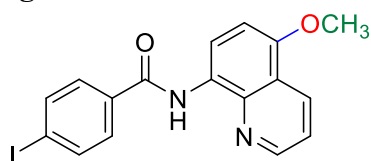
White solid, isolated yield: 73%; ^1H NMR (400 MHz, CDCl_3) δ 10.47 (s, 1H), 8.88 – 8.83 (m, 2H), 8.60 (dd, $J = 8.4, 1.7$ Hz, 1H), 7.97 (d, $J = 8.2$ Hz, 2H), 7.46 (dd, $J = 8.4, 4.2$ Hz, 1H), 7.34 (d, $J = 7.9$ Hz, 2H), 6.89 (d, $J = 8.6$ Hz, 1H), 4.01 (s, 3H), 2.45 (s, 3H). ^{13}C NMR (101 MHz, CDCl_3) δ 165.14 (s), 150.33 (s), 148.71 (s), 142.05 (s), 139.47 (s), 132.55 (s), 131.31 (s), 129.41 (s), 128.13 (s), 127.21 (s), 120.77 (s), 120.53 (s), 116.65 (s), 104.44 (s), 55.80 (s), 21.55 (s). HRMS (ESI): m/z : calcd for $[\text{M}+\text{H}]^+$ $\text{C}_{18}\text{H}_{17}\text{N}_2\text{O}_2$: 293.1285, found: 293.1284.

2f



White solid, isolated yield: 70%; ^1H NMR (400 MHz, CDCl_3) δ 10.48 (s, 1H), 8.86 (dd, $J = 5.0, 3.5$ Hz, 2H), 8.60 (dd, $J = 8.4, 1.5$ Hz, 1H), 8.00 (d, $J = 8.1$ Hz, 2H), 7.46 (dd, $J = 8.4, 4.2$ Hz, 1H), 7.36 (d, $J = 8.1$ Hz, 2H), 6.90 (d, $J = 8.6$ Hz, 1H), 4.01 (d, $J = 7.6$ Hz, 3H), 2.75 (q, $J = 7.6$ Hz, 2H), 1.29 (t, $J = 7.6$ Hz, 3H). ^{13}C NMR (101 MHz, CDCl_3) δ 165.17 (s), 150.33 (s), 150.11 (s), 148.69 (s), 148.26 (s), 139.49 (s), 132.83 (s), 131.30 (s), 128.20 (d, $J = 4.9$ Hz), 127.30 (s), 120.75 (s), 116.64 (s), 104.46 (s), 55.81 (s), 28.86 (s), 15.36 (s). HRMS (ESI): m/z : calcd for $[\text{M}+\text{H}]^+$ $\text{C}_{19}\text{H}_{19}\text{N}_2\text{O}_2$: 307.1442, found: 307.1441.

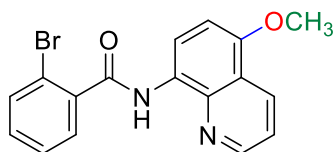
2g



White solid, isolated yield: 53%; ^1H NMR (400 MHz, CDCl_3) δ 10.49 (s, 1H), 8.90 – 8.83 (m, 2H), 8.63 (dd, $J = 8.4, 1.2$ Hz, 1H), 7.91 (d, $J = 8.3$ Hz, 2H), 7.81 (d, $J = 8.3$ Hz, 2H), 7.50 (dd, $J = 8.4, 4.2$ Hz, 1H), 6.92 (d, $J = 8.6$ Hz, 1H), 4.04 (s, 3H). ^{13}C NMR (101 MHz, CDCl_3) δ 148.80 (s), 139.42 (s),

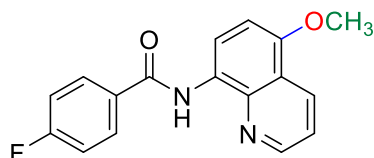
137.96 (s), 134.81 (s), 131.40 (s), 128.79 (s), 120.86 (s), 116.84 (s), 104.36 (s), 55.81 (s). HRMS (ESI): m/z : calcd for $[M+H]^+$ $C_{17}H_{14}N_2O_2$: 405.0094, found: 405.0091.

2h



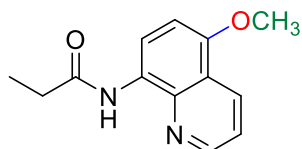
White solid, isolated yield: 57%; 1H NMR (400 MHz, $CDCl_3$) δ 10.05 (s, 1H), 8.87 (d, $J = 8.5$ Hz, 1H), 8.79 (dd, $J = 4.2, 1.7$ Hz, 1H), 8.59 (dd, $J = 8.4, 1.7$ Hz, 1H), 7.69 (ddd, $J = 10.1, 7.8, 1.4$ Hz, 2H), 7.46 – 7.41 (m, 2H), 7.36 – 7.31 (m, 1H), 6.91 – 6.88 (m, 1H), 4.02 (s, 3H). ^{13}C NMR (101 MHz, $CDCl_3$) δ 150.77 (s), 148.81 (s), 133.63 (s), 131.32 (d, $J = 2.5$ Hz), 129.53 (s), 127.69 (d, $J = 14.8$ Hz), 120.84 (s), 120.51 (s), 119.77 (s), 117.12 (s), 104.29 (s), 55.85 (s). HRMS (ESI): m/z : calcd for $[M+H]^+$ $C_{17}H_{14}BrN_2O_2$: 357.0233, 359.0213, found: 357.0238, 359.0217.

2i



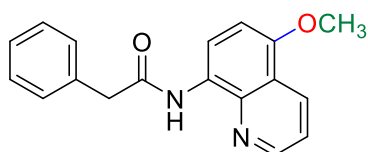
White solid, isolated yield: 81%; 1H NMR (400 MHz, $CDCl_3$) δ 10.43 (s, 1H), 8.83 (ddd, $J = 13.6, 6.8, 5.1$ Hz, 2H), 8.59 (dt, $J = 8.4, 1.8$ Hz, 1H), 8.07 (dd, $J = 8.3, 5.5$ Hz, 2H), 7.46 (ddd, $J = 8.4, 4.2, 1.8$ Hz, 1H), 7.21 (dd, $J = 12.7, 4.6$ Hz, 2H), 6.88 (dd, $J = 8.6, 1.8$ Hz, 1H), 4.00 (d, $J = 1.4$ Hz, 3H). ^{13}C NMR (101 MHz, $CDCl_3$) δ 164.85 (d, $J = 252.9$ Hz), 164.00 (s), 150.51 (s), 148.76 (s), 139.40 (s), 131.53 (d, $J = 3.2$ Hz), 131.38 (s), 129.53 (d, $J = 9.0$ Hz), 127.83 (s), 120.84 (s), 120.53 (s), 116.73 (s), 115.78 (d, $J = 21.9$ Hz), 104.35 (s), 55.80 (s). HRMS (ESI): m/z : calcd for $[M+H]^+$ $C_{17}H_{14}FN_2O_2$: 297.1034, found: 297.1035.

2j



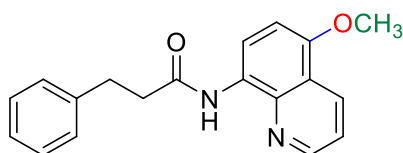
White solid, isolated yield: 48%; 1H NMR (400 MHz, $CDCl_3$) δ 9.60 (s, 1H), 8.84 (dd, $J = 4.2, 1.6$ Hz, 1H), 8.73 (d, $J = 8.5$ Hz, 1H), 8.61 (dd, $J = 8.4, 1.6$ Hz, 1H), 7.47 (dd, $J = 8.4, 4.2$ Hz, 1H), 6.87 (d, $J = 8.6$ Hz, 1H), 4.02 (s, 3H), 2.60 (q, $J = 7.6$ Hz, 2H), 1.36 (t, $J = 7.6$ Hz, 3H). ^{13}C NMR (101 MHz, $CDCl_3$) δ 172.12 (s), 148.54 (s), 131.30 (s), 128.00 (s), 120.67 (s), 116.56 (s), 104.38 (s), 55.76 (s), 31.17 (s), 9.88 (s). HRMS (ESI): m/z : calcd for $[M+H]^+$ $C_{13}H_{15}N_2O_2$: 231.1129, found: 231.1126.

2k



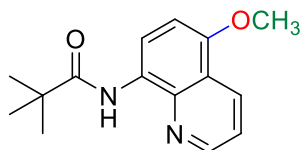
White solid, isolated yield: 64%; ^1H NMR (400 MHz, CDCl_3) δ 9.66 (s, 1H), 8.70 (dd, $J = 4.3, 1.7$ Hz, 1H), 8.67 (d, $J = 8.5$ Hz, 1H), 8.53 (dd, $J = 8.4, 1.7$ Hz, 1H), 7.45 – 7.37 (m, 5H), 7.35 – 7.30 (m, 1H), 6.81 (d, $J = 8.6$ Hz, 1H), 3.97 (s, 3H), 3.87 (s, 2H). ^{13}C NMR (101 MHz, CDCl_3) δ 169.10 (s), 150.32 (s), 148.61 (s), 134.89 (s), 131.17 (s), 129.55 (s), 128.93 (s), 127.83 (s), 127.25 (s), 120.64 (s), 120.35 (s), 116.53 (s), 104.25 (s), 55.75 (s), 45.26 (s). HRMS (ESI): m/z : calcd for $[\text{M}+\text{H}]^+$ $\text{C}_{18}\text{H}_{17}\text{N}_2\text{O}_2$: 293.1285, found: 293.1283.

2l



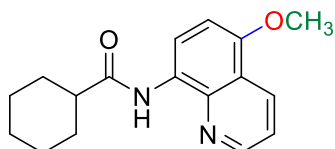
White solid, isolated yield: 62%; ^1H NMR (400 MHz, CDCl_3) δ 9.57 (s, 1H), 8.81 (dd, $J = 4.2, 1.5$ Hz, 1H), 8.73 (d, $J = 8.5$ Hz, 1H), 8.60 (dd, $J = 8.4, 1.6$ Hz, 1H), 7.46 (dd, $J = 8.4, 4.2$ Hz, 1H), 7.32 (d, $J = 4.4$ Hz, 4H), 7.23 (dq, $J = 8.8, 4.2$ Hz, 1H), 6.87 (d, $J = 8.6$ Hz, 1H), 4.02 (s, 3H), 3.20 – 3.14 (m, 2H), 2.91 – 2.86 (m, 2H). ^{13}C NMR (101 MHz, CDCl_3) δ 170.34 (s), 150.23 (s), 148.55 (s), 144.90 (s), 140.90 (s), 131.27 (s), 128.47 (d, $J = 12.8$ Hz), 127.89 (s), 126.19 (s), 120.68 (s), 120.43 (s), 116.68 (s), 104.37 (s), 55.78 (s), 39.68 (s), 31.61 (s). HRMS (ESI): m/z : calcd for $[\text{M}+\text{H}]^+$ $\text{C}_{19}\text{H}_{19}\text{N}_2\text{O}_2$: 307.1442, found: 307.1442.

2m



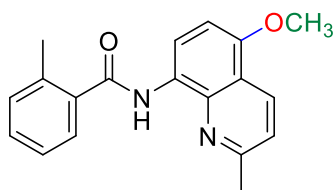
White solid, isolated yield: 74%; ^1H NMR (400 MHz, CDCl_3) δ 10.03 (s, 1H), 8.82 (dd, $J = 4.2, 1.7$ Hz, 1H), 8.71 (d, $J = 8.5$ Hz, 1H), 8.56 (dd, $J = 8.4, 1.7$ Hz, 1H), 7.43 (dd, $J = 8.4, 4.3$ Hz, 1H), 6.83 (d, $J = 8.6$ Hz, 1H), 3.98 (s, 3H), 1.42 (s, 9H). ^{13}C NMR (101 MHz, CDCl_3) δ 176.85 (s), 150.06 (s), 148.67 (s), 131.19 (s), 120.64 (s), 116.28 (s), 104.36 (s), 55.76 (s), 40.18 (s), 27.79 (s). HRMS (ESI): m/z : calcd for $[\text{M}+\text{H}]^+$ $\text{C}_{15}\text{H}_{19}\text{N}_2\text{O}_2$: 259.1441, found: 259.1440.

2n



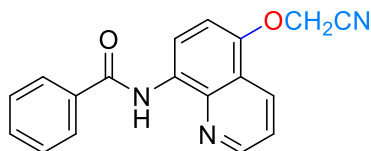
White solid, isolated yield: 67%; ^1H NMR (400 MHz, CDCl_3) δ 9.64 (s, 1H), 8.82 (dd, $J = 4.2, 1.7$ Hz, 1H), 8.72 (d, $J = 8.5$ Hz, 1H), 8.57 (dd, $J = 8.4, 1.6$ Hz, 1H), 7.44 (dd, $J = 8.4, 4.2$ Hz, 1H), 6.84 (d, $J = 8.6$ Hz, 1H), 3.98 (d, $J = 8.0$ Hz, 3H), 2.45 (tt, $J = 11.7, 3.5$ Hz, 1H), 2.07 (d, $J = 13.3$ Hz, 2H), 1.91 – 1.84 (m, 2H), 1.76 – 1.60 (m, 3H), 1.34 (ddd, $J = 22.0, 12.5, 9.5$ Hz, 3H). ^{13}C NMR (101 MHz, CDCl_3) δ 174.51 (s), 150.05 (s), 148.56 (s), 139.17 (s), 131.26 (s), 128.09 (s), 120.66 (s), 120.42 (s), 116.52 (s), 104.38 (s), 55.77 (s), 46.85 (s, 2C), 29.82 (s), 25.82 (s). HRMS (ESI): m/z : calcd for $[\text{M}+\text{H}]^+$ $\text{C}_{17}\text{H}_{21}\text{N}_2\text{O}_2$: 285.1598, found: 285.1605.

2s



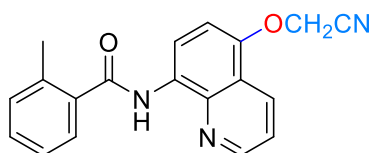
yellow solid, isolated yield: 54%; ^1H NMR (400 MHz, CDCl_3) δ 10.02 (s, 1H), 8.82 (d, $J = 8.5$ Hz, 1H), 8.46 (d, $J = 8.5$ Hz, 1H), 7.69 (d, $J = 7.4$ Hz, 1H), 7.39 (d, $J = 7.5$ Hz, 1H), 7.35 – 7.29 (m, 3H), 6.83 (d, $J = 8.5$ Hz, 1H), 4.00 (s, 3H), 2.68 (s, 3H), 2.62 (s, 3H). ^{13}C NMR (101 MHz, CDCl_3) δ 167.70 (s), 157.79 (s), 150.61 (s), 138.82 (s), 136.55 (s), 131.33 (s), 131.30 (s), 130.10 (s), 127.67 (s), 127.43 (s), 126.22 (s), 126.00 (s), 121.51 (s), 118.51 (s), 116.64 (s), 103.50 (s), 55.75 (s), 25.24 (s), 20.34 (s). HRMS (ESI): m/z : calcd for $[\text{M}+\text{H}]^+$ $\text{C}_{19}\text{H}_{19}\text{N}_2\text{O}_2$: 307.1442, found: 307.1440.

3a



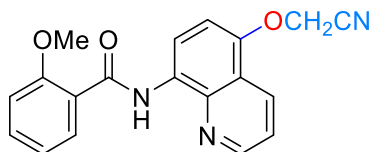
White solid, isolated yield: 50%; ^1H NMR (400 MHz, CDCl_3) δ 10.56 (s, 1H), 8.91 (dd, $J = 5.0, 3.4$ Hz, 2H), 8.58 (dd, $J = 8.5, 1.6$ Hz, 1H), 8.08 (dd, $J = 7.8, 1.5$ Hz, 2H), 7.60 – 7.53 (m, 4H), 7.03 (d, $J = 8.6$ Hz, 1H), 5.00 (s, 2H). ^{13}C NMR (101 MHz, CDCl_3) δ 163.72 (s), 149.26 (s), 146.98 (s), 140.26 (s), 131.86 (s), 130.91 (s), 130.30 (s), 129.55 (s), 128.83 (s), 127.24 (s), 121.84 (s), 121.60 (s), 120.48 (s), 115.84 (s), 106.75 (s), 54.13 (s). HRMS (ESI): m/z : calcd for $[\text{M}+\text{H}]^+$ $\text{C}_{18}\text{H}_{14}\text{N}_3\text{O}_2$: 304.1081, found: 304.1076.

3b



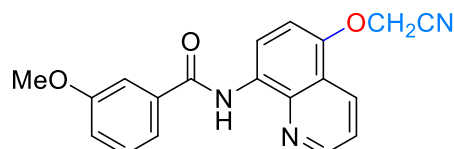
White solid, isolated yield: 46%; ^1H NMR (400 MHz, CDCl_3) δ 10.02 (s, 1H), 8.91 (d, $J = 8.5$ Hz, 1H), 8.83 (dd, $J = 4.2, 1.4$ Hz, 1H), 8.56 (dd, $J = 8.4, 1.4$ Hz, 1H), 7.67 (d, $J = 7.4$ Hz, 1H), 7.52 (dd, $J = 8.5, 4.3$ Hz, 1H), 7.41 (t, $J = 6.8$ Hz, 1H), 7.32 (t, $J = 7.4$ Hz, 2H), 7.02 (d, $J = 8.6$ Hz, 1H), 5.00 (s, 2H), 2.60 (s, 3H). ^{13}C NMR (101 MHz, CDCl_3) δ 168.07 (s), 149.23 (s), 140.05 (s), 139.27 (s), 136.69 (s), 136.54 (s), 135.71 (s), 134.74 (s), 131.39 (s), 130.85 (s), 130.44 (s), 130.36 (s), 127.25 (s), 126.03 (s), 121.57 (s), 115.80 (s), 106.68 (s), 54.13 (s), 20.23 (s). HRMS (ESI): m/z : calcd for $[\text{M}+\text{H}]^+$ $\text{C}_{19}\text{H}_{16}\text{N}_3\text{O}_2$: 318.1237, found: 318.1232.

3c



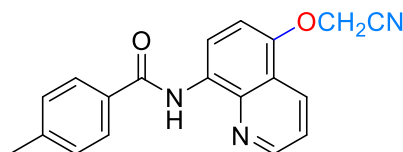
White solid, isolated yield: 46%; ^1H NMR (400 MHz, CDCl_3) δ 12.25 (s, 1H), 9.02 (d, $J = 8.6$ Hz, 1H), 8.96 (dd, $J = 4.1, 1.5$ Hz, 1H), 8.59 (dd, $J = 8.5, 1.6$ Hz, 1H), 8.38 (dd, $J = 7.8, 1.6$ Hz, 1H), 7.57 – 7.52 (m, 2H), 7.18 (t, $J = 7.6$ Hz, 1H), 7.11 (d, $J = 8.4$ Hz, 1H), 7.03 (d, $J = 8.6$ Hz, 1H), 5.01 (s, 2H), 4.24 (s, 3H). ^{13}C NMR (101 MHz, CDCl_3) δ 163.45 (s), 157.71 (s), 149.21 (s), 146.81 (s), 139.87 (s), 133.16 (s), 132.33 (s), 131.43 (s), 130.69 (s), 122.21 (s), 121.35 (s), 120.48 (s), 120.29 (s), 116.55 (s), 115.39 (s), 111.59 (s), 106.91 (s), 56.15 (s), 54.15 (s). HRMS (ESI): m/z : calcd for $[\text{M}+\text{H}]^+$ $\text{C}_{19}\text{H}_{16}\text{N}_3\text{O}_3$: 334.1186, found: 334.1189.

3d



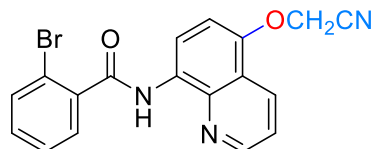
White solid, isolated yield: 44%; ^1H NMR (400 MHz, CDCl_3) δ 10.54 (s, 1H), 8.89 (d, $J = 8.3$ Hz, 2H), 8.57 (dd, $J = 8.5, 1.4$ Hz, 1H), 7.63 (d, $J = 6.3$ Hz, 2H), 7.54 (dd, $J = 8.5, 4.2$ Hz, 1H), 7.46 (t, $J = 8.1$ Hz, 1H), 7.13 (dd, $J = 8.2, 2.2$ Hz, 1H), 7.02 (d, $J = 8.6$ Hz, 1H), 5.00 (s, 2H), 3.92 (s, 3H). ^{13}C NMR (101 MHz, CDCl_3) δ 165.17 (s), 156.45 (s), 153.60 (s), 149.28 (s), 136.54 (s), 132.51 (s), 130.89 (s), 130.26 (s), 129.81 (s), 128.58 (s), 121.61 (s), 118.89 (d, $J = 24.3$ Hz), 118.67 – 118.22 (m), 118.02 (s), 115.83 (s), 112.62 (s), 106.70 (s), 55.54 (s), 54.11 (s). HRMS (ESI): m/z : calcd for $[\text{M}+\text{H}]^+$ $\text{C}_{19}\text{H}_{16}\text{N}_3\text{O}_3$: 334.1186, found: 334.1191.

3e



White solid, isolated yield: 44%; ^1H NMR (400 MHz, CDCl_3) δ 10.52 (s, 1H), 8.90 (dd, $J = 7.0, 4.7$ Hz, 2H), 8.57 (dd, $J = 8.5, 1.5$ Hz, 1H), 7.97 (d, $J = 8.1$ Hz, 2H), 7.54 (dd, $J = 8.4, 4.1$ Hz, 1H), 7.35 (d, $J = 8.0$ Hz, 2H), 7.02 (d, $J = 8.6$ Hz, 1H), 4.99 (s, 2H), 2.45 (s, 3H). ^{13}C NMR (101 MHz, CDCl_3) δ 165.35 (s), 149.21 (s), 146.87 (s), 142.37 (s), 139.40 (s), 130.89 (s), 129.48 (s), 127.76 (s), 127.29 (d, $J = 8.6$ Hz), 121.56 (s), 115.75 (s), 106.77 (s), 54.13 (s). HRMS (ESI): m/z : calcd for $[\text{M}+\text{H}]^+$ $\text{C}_{19}\text{H}_{16}\text{N}_3\text{O}_2$: 318.1237, found: 318.1220.

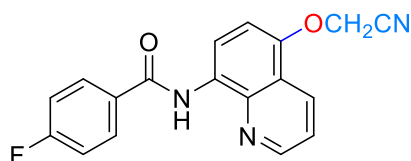
3h



White solid, isolated yield: 45%; ^1H NMR (400 MHz, CDCl_3) δ 10.12 (s, 1H), 8.92 (d, $J = 8.6$ Hz, 1H), 8.85 (d, $J = 2.7$ Hz, 1H), 8.57 (d, $J = 7.0$ Hz, 1H), 7.70 (t, $J = 6.8$ Hz, 2H), 7.53 (dd, $J = 8.5, 4.2$ Hz, 1H), 7.46 (t, $J = 7.5$ Hz, 1H), 7.36 (t, $J = 6.9$ Hz, 1H), 7.02 (d, $J = 8.6$ Hz, 1H), 5.00 (s, 2H). ^{13}C NMR (101 MHz, CDCl_3) δ 170.22 (s), 149.32 (s), 147.27 (s), 136.44 (s), 133.71 (s), 131.54 (s), 130.96 (d, $J = 19.3$ Hz), 130.18 – 130.11 (m), 129.80 (d, $J = 42.1$ Hz), 127.82 (d, $J = 26.3$ Hz), 121.63 (s), 118.98 (s),

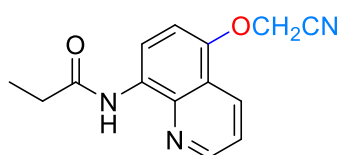
116.21 (s), 111.91 (s), 106.57 (s), 54.10 (s). HRMS (ESI): m/z : calcd for $[M+H]^+$ $C_{18}H_{13}BrN_3O_2$: 382.0186, 384.0165, found: 382.0188, 384.0173.

3i



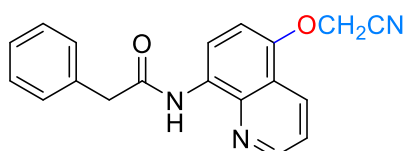
White solid, isolated yield: 46%; 1H NMR (400 MHz, $CDCl_3$) δ 10.50 (s, 1H), 8.90 (dd, $J = 14.8$, 6.4 Hz, 2H), 8.58 (d, $J = 8.5$ Hz, 1H), 8.09 (dd, $J = 7.4$, 5.3 Hz, 2H), 7.56 (dd, $J = 8.5$, 4.3 Hz, 1H), 7.22 (d, $J = 7.8$ Hz, 2H), 7.03 (d, $J = 8.5$ Hz, 1H), 5.00 (s, 2H). ^{13}C NMR (101 MHz, $CDCl_3$) δ 164.99 (d, $J = 252.5$ Hz), 164.23 (s), 149.28 (s), 147.05 (s), 139.35 (s), 131.21 (d, $J = 3.1$ Hz), 130.97 (s), 130.09 (s), 129.61 (d, $J = 9.0$ Hz), 121.64 (s), 120.48 (s), 115.89 (d, $J = 22.0$ Hz), 115.87 (s), 114.75 (s), 106.69 (s), 54.10 (s). HRMS (ESI): m/z : calcd for $[M+H]^+$ $C_{18}H_{13}FN_3O_2$: 322.0986, found: 322.0989.

3j



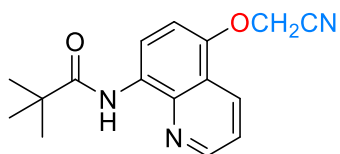
White solid, isolated yield: 43%; 1H NMR (400 MHz, $CDCl_3$) δ 9.63 (s, 1H), 8.86 (dd, $J = 4.2$, 1.7 Hz, 1H), 8.75 (d, $J = 8.6$ Hz, 1H), 8.54 (dd, $J = 8.5$, 1.7 Hz, 1H), 7.51 (dd, $J = 8.5$, 4.2 Hz, 1H), 6.96 (d, $J = 8.6$ Hz, 1H), 4.96 (s, 2H), 2.59 (q, $J = 7.6$ Hz, 2H), 1.34 (t, $J = 7.6$ Hz, 3H). ^{13}C NMR (101 MHz, $CDCl_3$) δ 172.34 (s), 149.05 (s), 146.67 (s), 130.84 (s), 130.28 (s), 126.08 (s), 121.46 (s), 120.39 (s), 115.64 (s), 114.78 (s), 106.78 (s), 54.13 (s), 31.17 (s), 9.78 (s). HRMS (ESI): m/z : calcd for $[M+H]^+$ $C_{14}H_{14}N_3O_2$: 256.1081, found: 256.1082.

3k



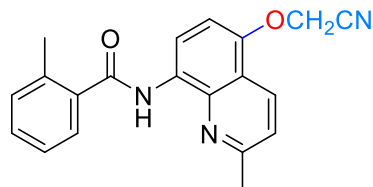
White solid, isolated yield: 36%; 1H NMR (400 MHz, $CDCl_3$) δ 9.71 (s, 1H), 8.74 (dd, $J = 4.2$, 1.4 Hz, 1H), 8.71 (d, $J = 8.6$ Hz, 1H), 8.49 (dd, $J = 8.5$, 1.4 Hz, 1H), 7.45 (dt, $J = 13.7$, 5.6 Hz, 5H), 7.34 (d, $J = 6.5$ Hz, 1H), 6.93 (d, $J = 8.6$ Hz, 1H), 4.94 (s, 2H), 3.88 (s, 2H). ^{13}C NMR (101 MHz, $CDCl_3$) δ 169.10 (s), 150.32 (s), 148.61 (s), 134.89 (s), 131.17 (s), 129.55 (s), 129.55 (s), 128.93 (s), 127.83 (s), 127.25 (s), 120.64 (s), 120.35 (s), 116.53 (s), 104.25 (s), 55.75 (s), 45.26 (s). HRMS (ESI): m/z : calcd for $[M+H]^+$ $C_{19}H_{16}N_3O_2$: 318.1237, found: 318.1235.

3m



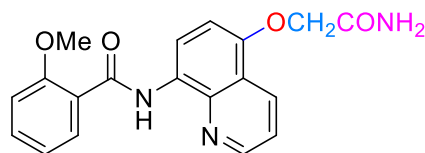
White solid, isolated yield: 59%; ^1H NMR (400 MHz, CDCl_3) δ 10.08 (s, 1H), 8.87 (dd, $J = 4.2, 1.7$ Hz, 1H), 8.75 (d, $J = 8.6$ Hz, 1H), 8.54 (dd, $J = 8.5, 1.7$ Hz, 1H), 7.51 (dd, $J = 8.5, 4.2$ Hz, 1H), 6.96 (d, $J = 8.6$ Hz, 1H), 4.96 (s, 2H), 1.42 (s, 9H). ^{13}C NMR (101 MHz, CDCl_3) δ 177.12 (s), 149.17 (s), 146.63 (s), 139.40 (s), 130.78 (s), 121.43 (s), 115.41 (s), 114.81 (s), 106.75 (s), 54.12 (s), 40.26 (s), 27.74 (s). HRMS (ESI): m/z : calcd for $[\text{M}+\text{H}]^+$ $\text{C}_{16}\text{H}_{18}\text{N}_3\text{O}_2$: 284.1394, found: 284.1392.

3s



White solid, isolated yield: 34%; ^1H NMR (400 MHz, CDCl_3) δ 10.07 (s, 1H), 8.86 (d, $J = 8.6$ Hz, 1H), 8.42 (d, $J = 8.6$ Hz, 1H), 7.69 (d, $J = 7.5$ Hz, 1H), 7.38 (ddd, $J = 18.1, 15.0, 7.1$ Hz, 4H), 6.94 (d, $J = 8.6$ Hz, 1H), 4.97 (s, 2H), 2.71 (s, 3H), 2.62 (s, 3H). ^{13}C NMR (101 MHz, CDCl_3) δ 165.38 (s), 158.49 (s), 147.17 (s), 138.79 (s), 136.66 (s), 131.42 (s), 130.86 (s), 130.24 (d, $J = 14.4$ Hz), 129.86 (s), 127.41 (s), 126.07 (s), 122.35 (s), 118.50 (s), 115.80 (s), 114.87 (s), 105.79 (s), 54.11 (s), 25.30 (s), 20.34 (s). HRMS (ESI): m/z : calcd for $[\text{M}+\text{H}]^+$ $\text{C}_{20}\text{H}_{18}\text{N}_3\text{O}_2$: 332.1394, found: 332.1392.

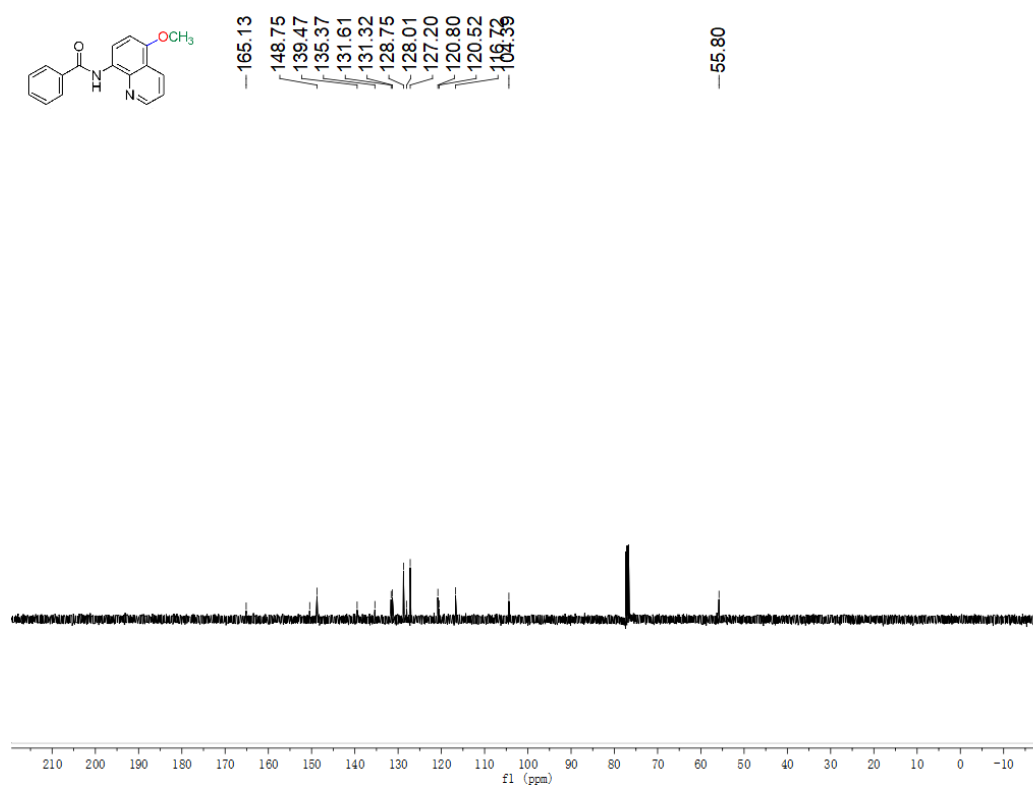
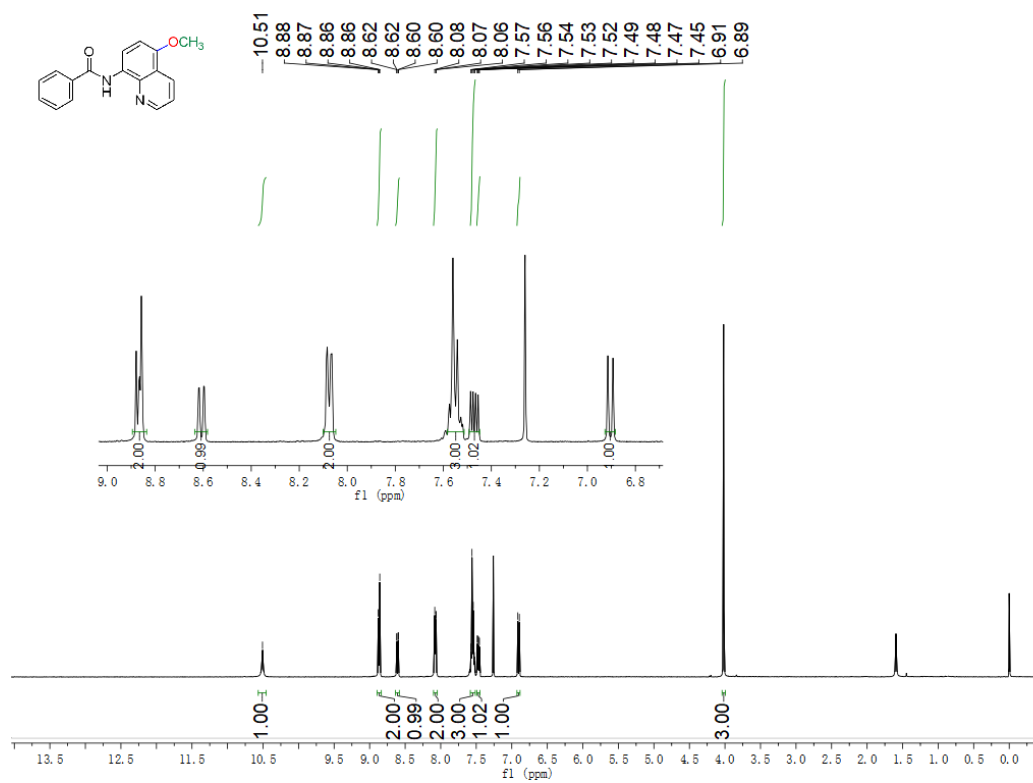
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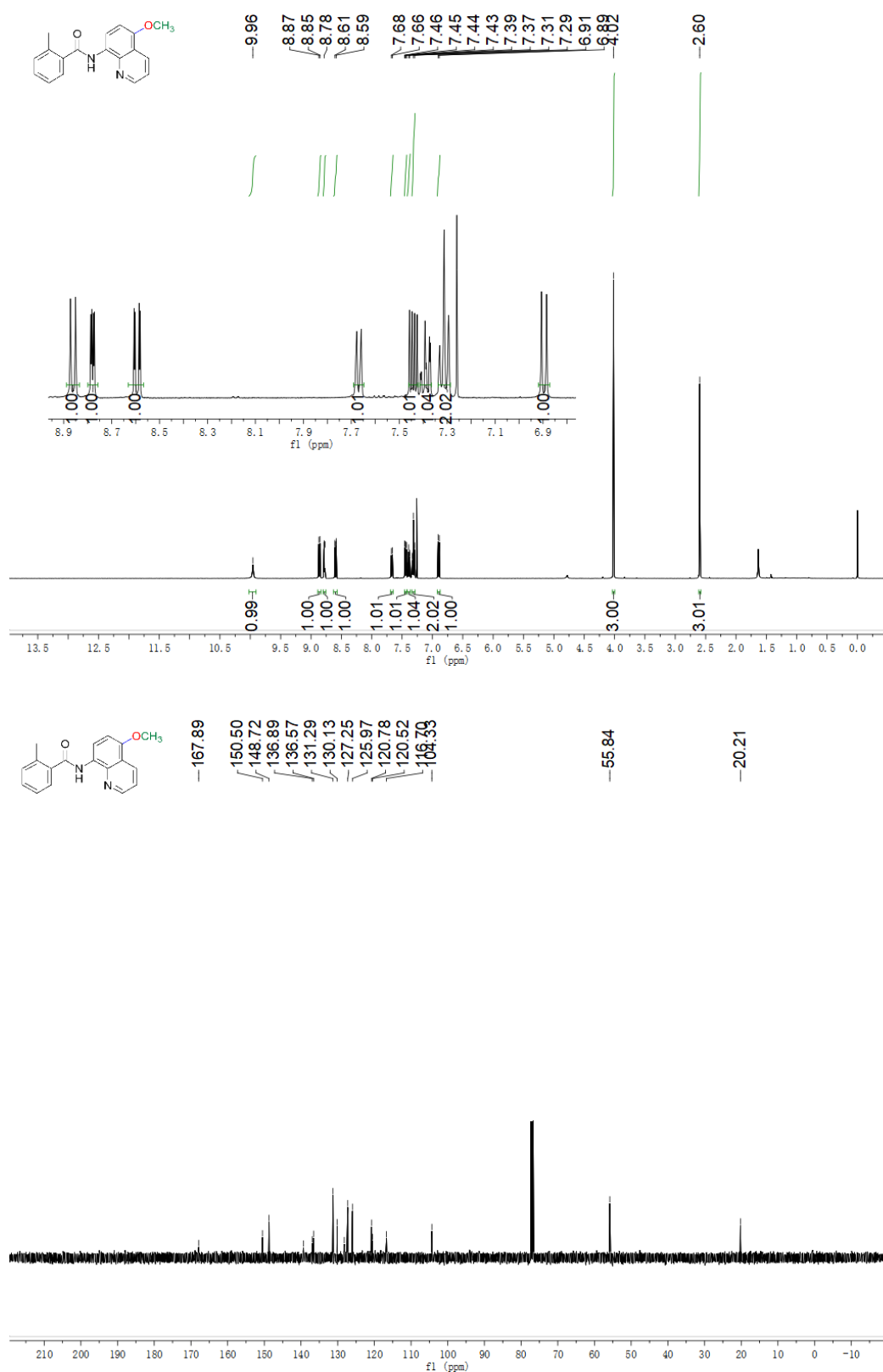
White solid, isolated yield: 85%; ^1H NMR (400 MHz, CDCl_3) δ 12.22 (s, 1H), 9.02 – 8.96 (m, 2H), 8.60 (d, $J = 8.2$ Hz, 1H), 8.38 (d, $J = 7.6$ Hz, 1H), 7.58 – 7.54 (m, 2H), 7.20 – 7.16 (m, 1H), 7.12 (d, $J = 8.4$ Hz, 1H), 6.97 (d, $J = 8.7$ Hz, 1H), 6.55 (s, 1H), 5.68 (s, 1H), 4.76 (s, 2H), 4.24 (s, 3H). ^{13}C NMR (101 MHz, CDCl_3) δ 177.21 (s), 163.42 (s), 157.62 (s), 149.36 (s), 146.89 (s), 139.81 (s), 133.16 (s), 132.27 (s), 131.30 (s), 130.81 (s), 122.21 (s), 121.35 (s), 120.48 (s), 120.29 (s), 116.40 (s), 111.41 (s), 106.21 (s), 67.95 (s), 54.15 (s). HRMS (ESI): m/z : calcd for $[\text{M}+\text{H}]^+$ $\text{C}_{19}\text{H}_{18}\text{N}_3\text{O}_4$: 352.1292, found: 352.1290.

7. ^1H and ^{13}C NMR spectra

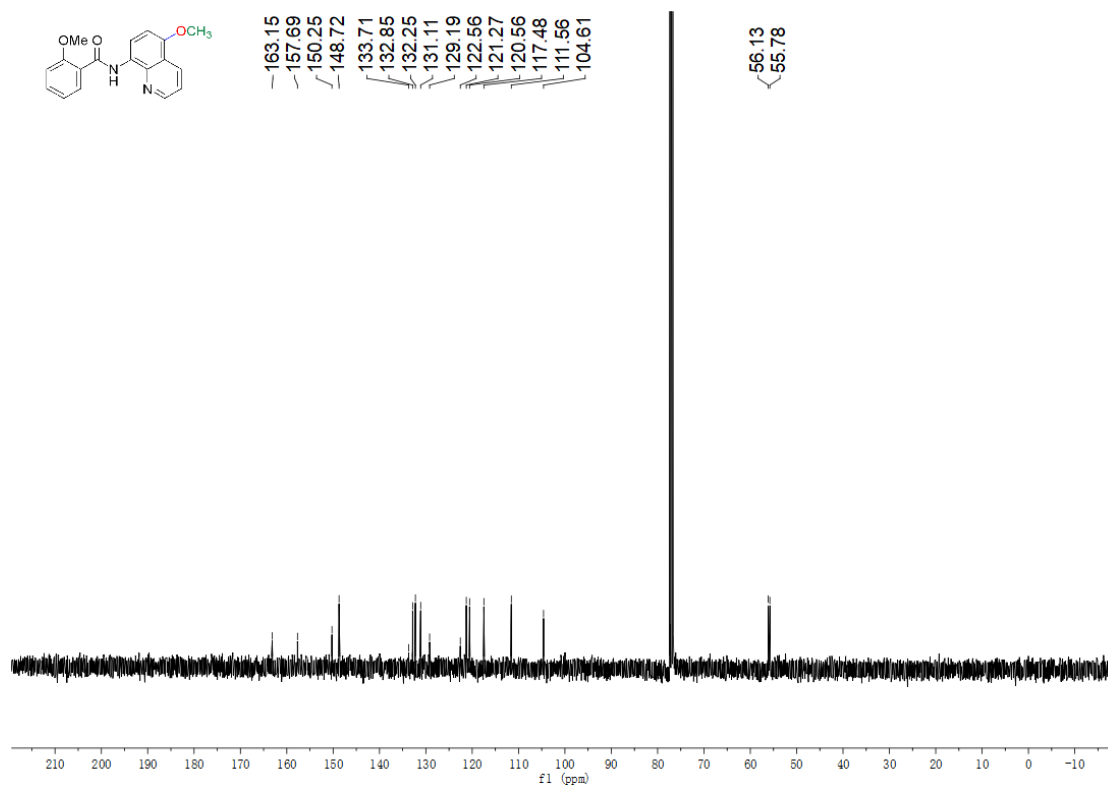
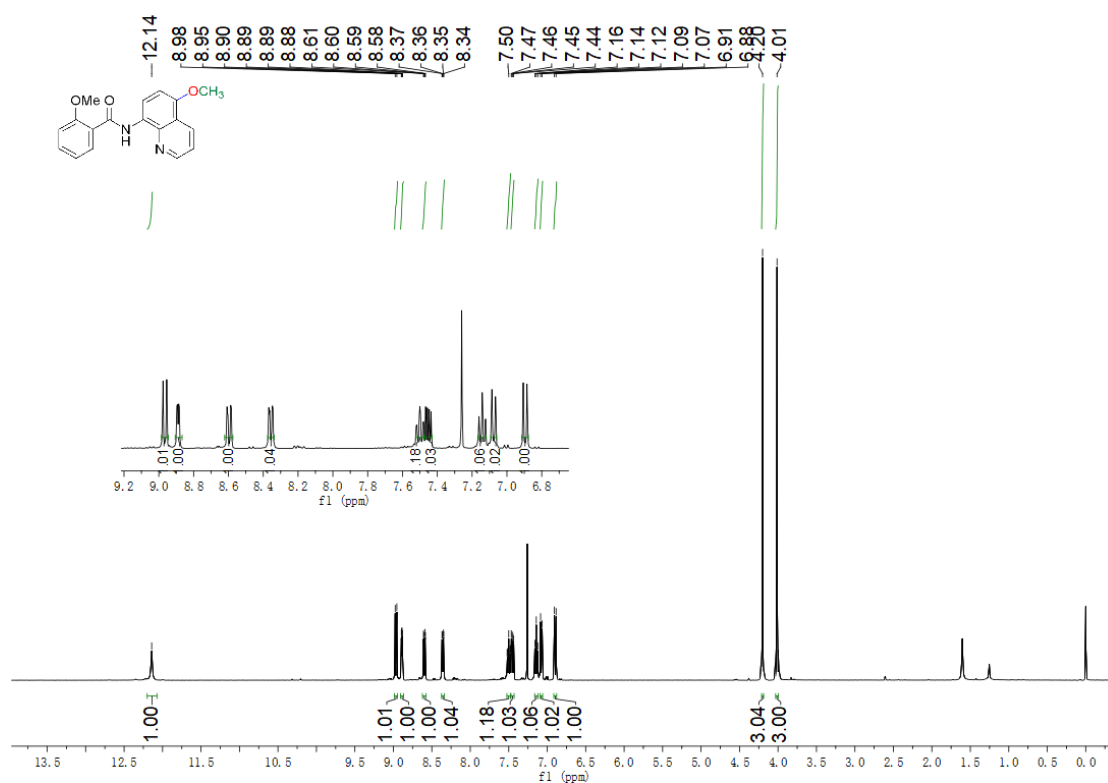
2a



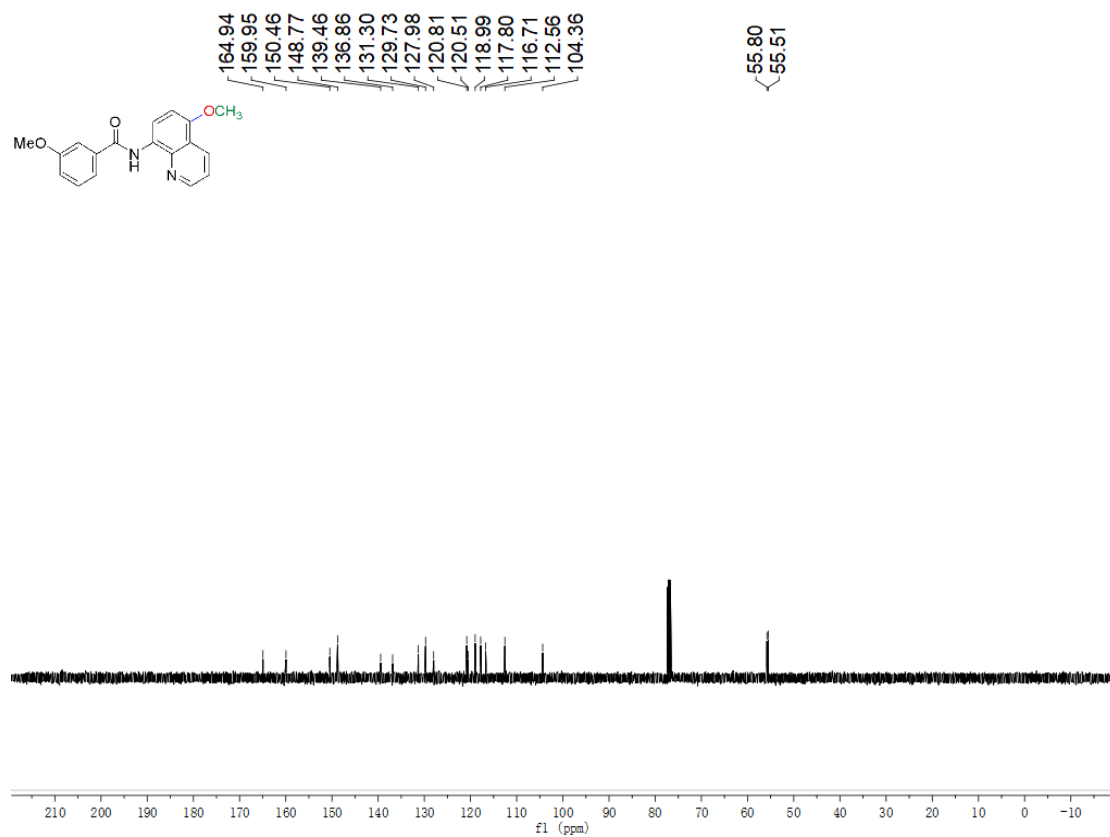
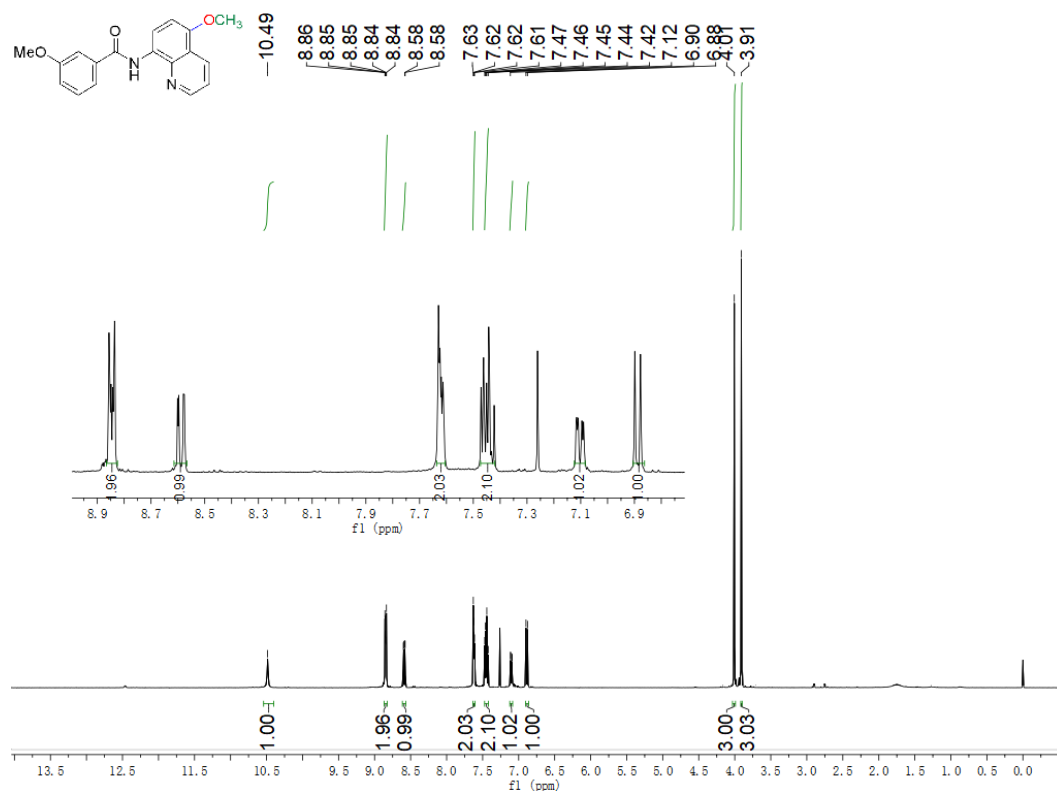
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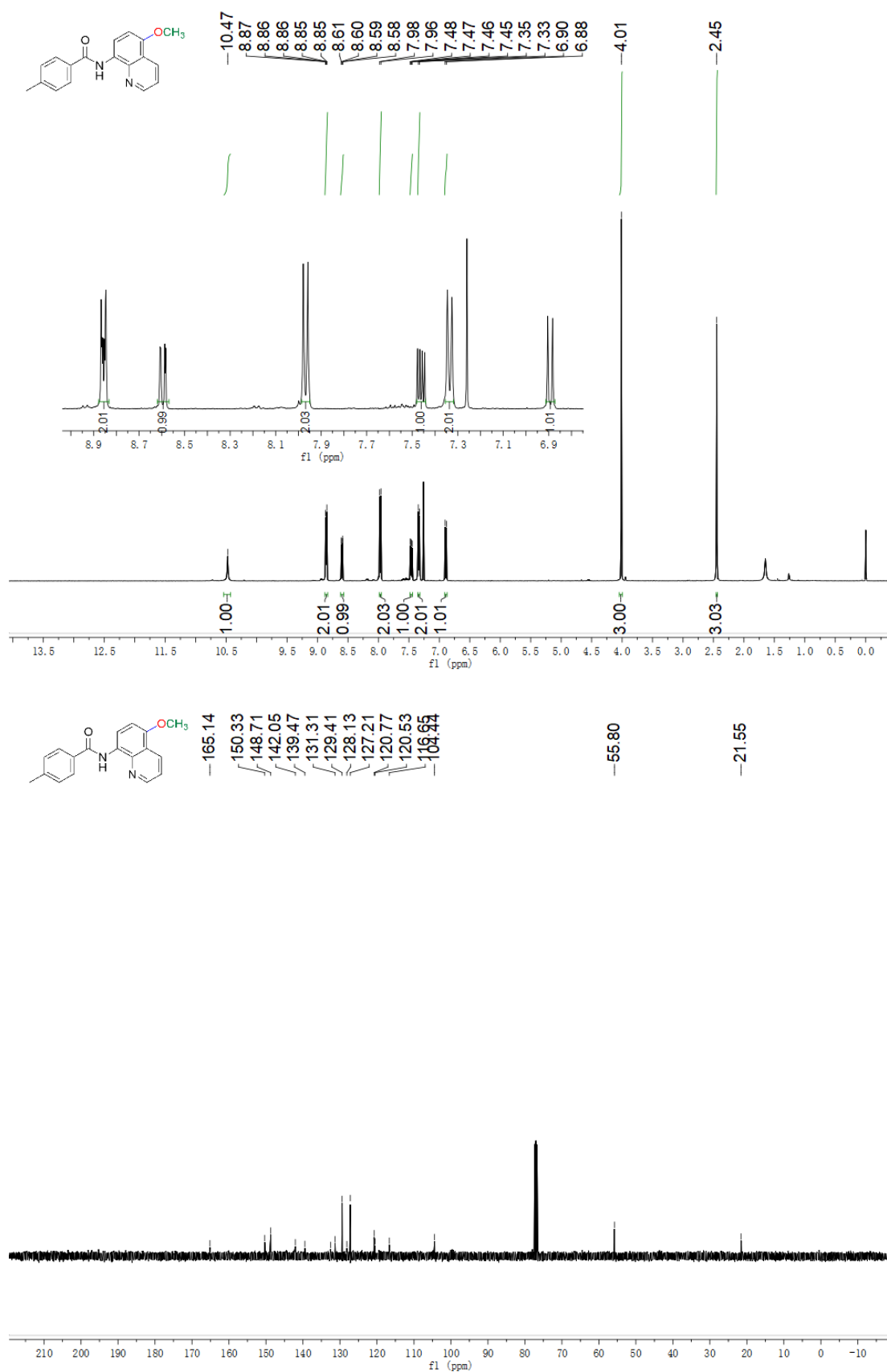
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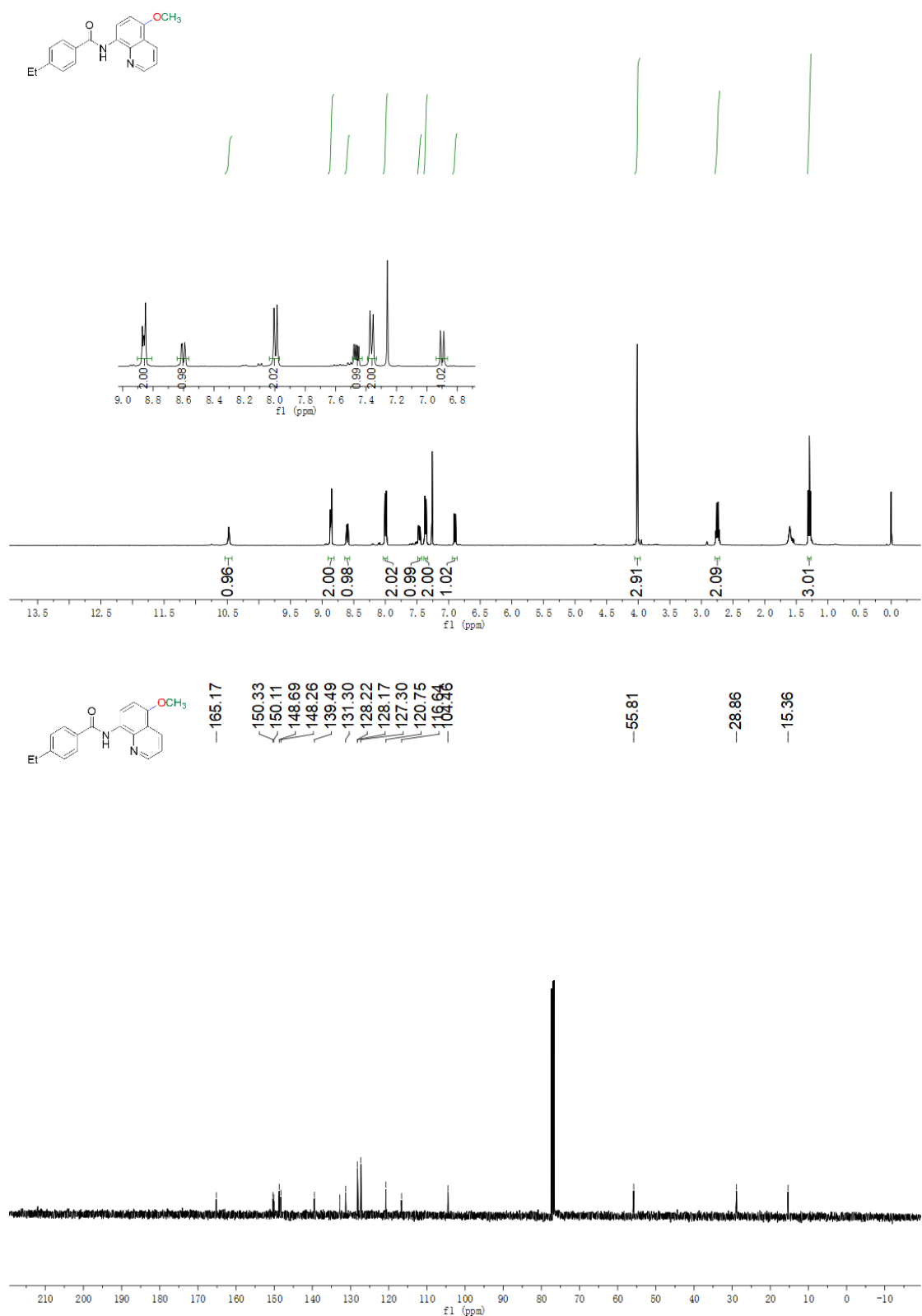
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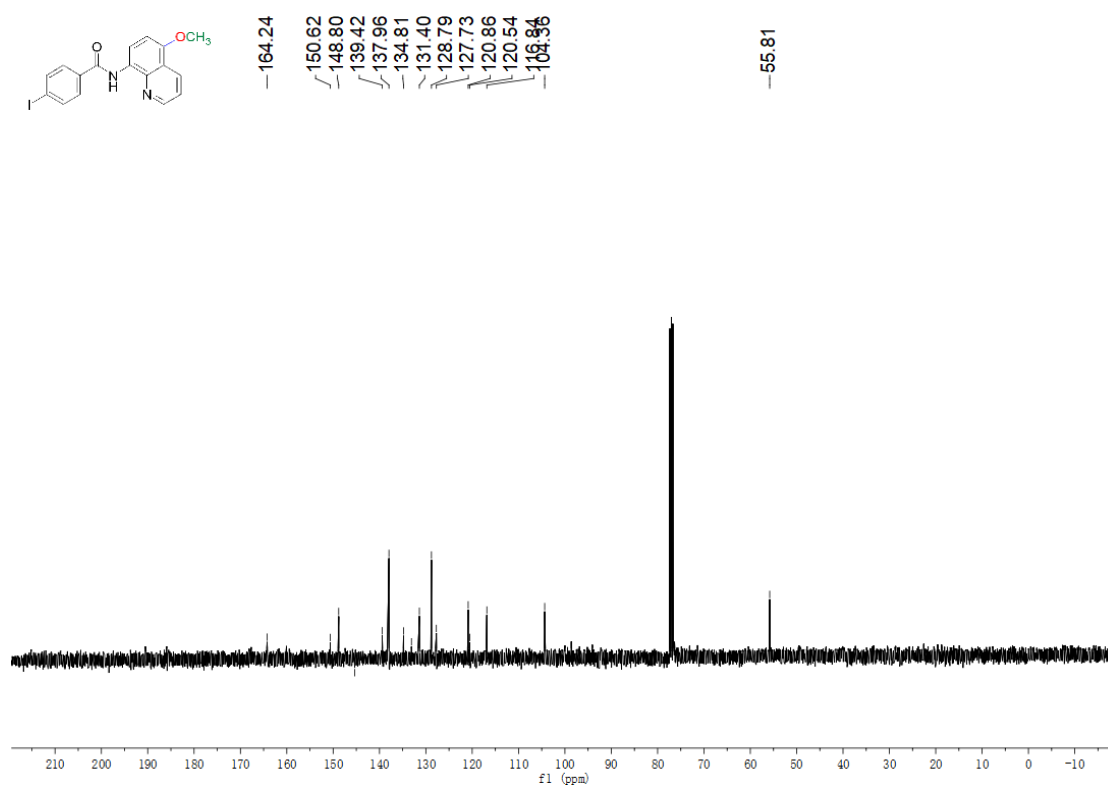
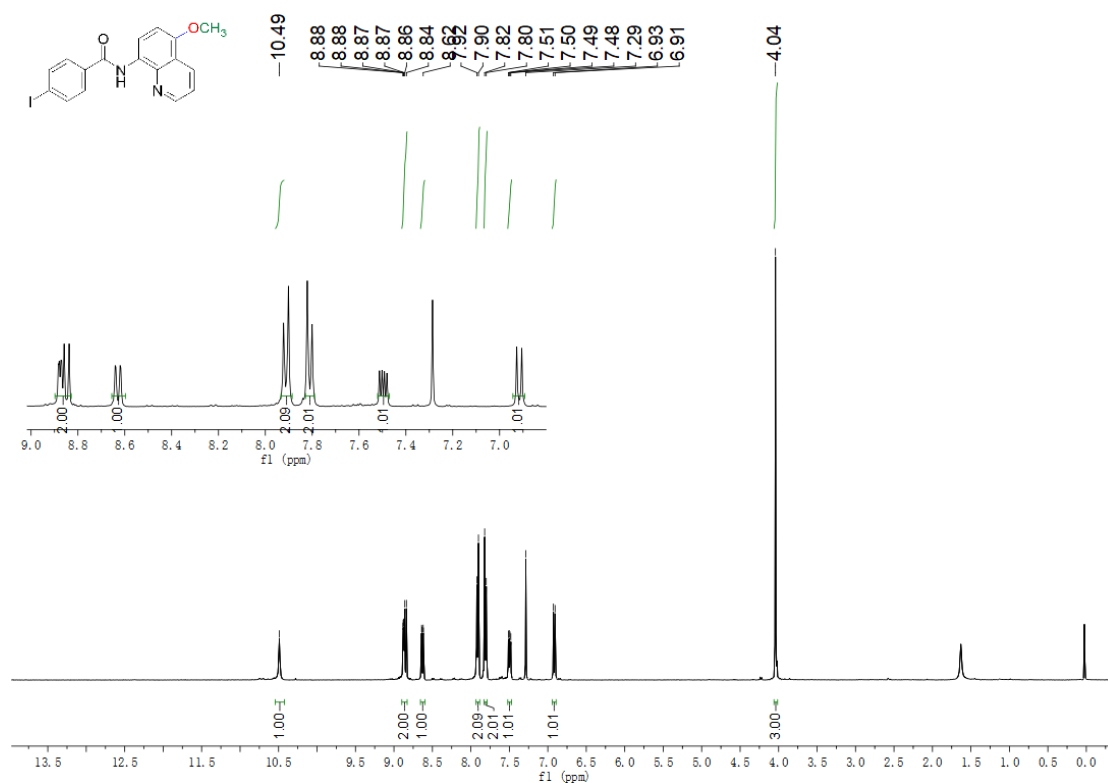
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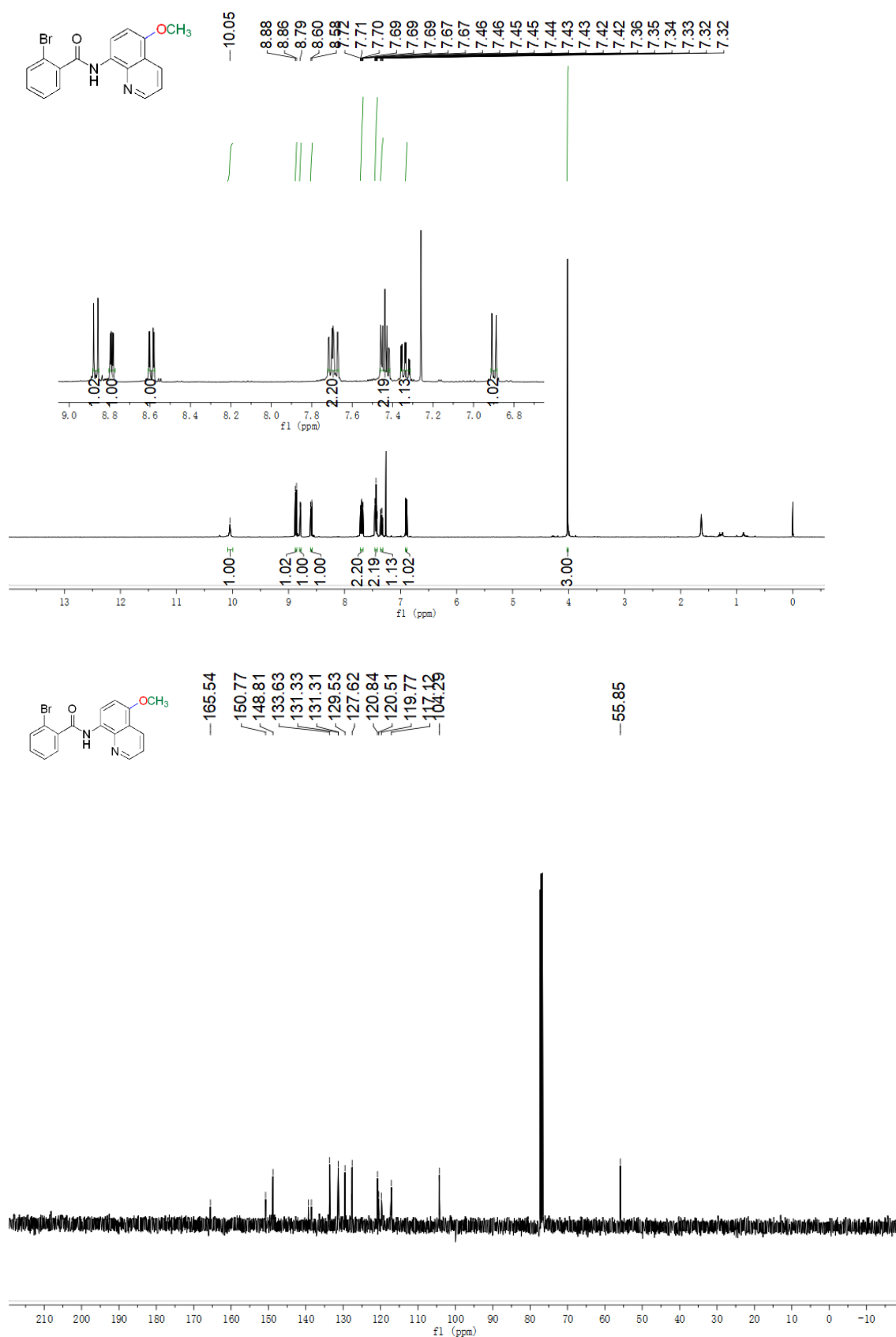
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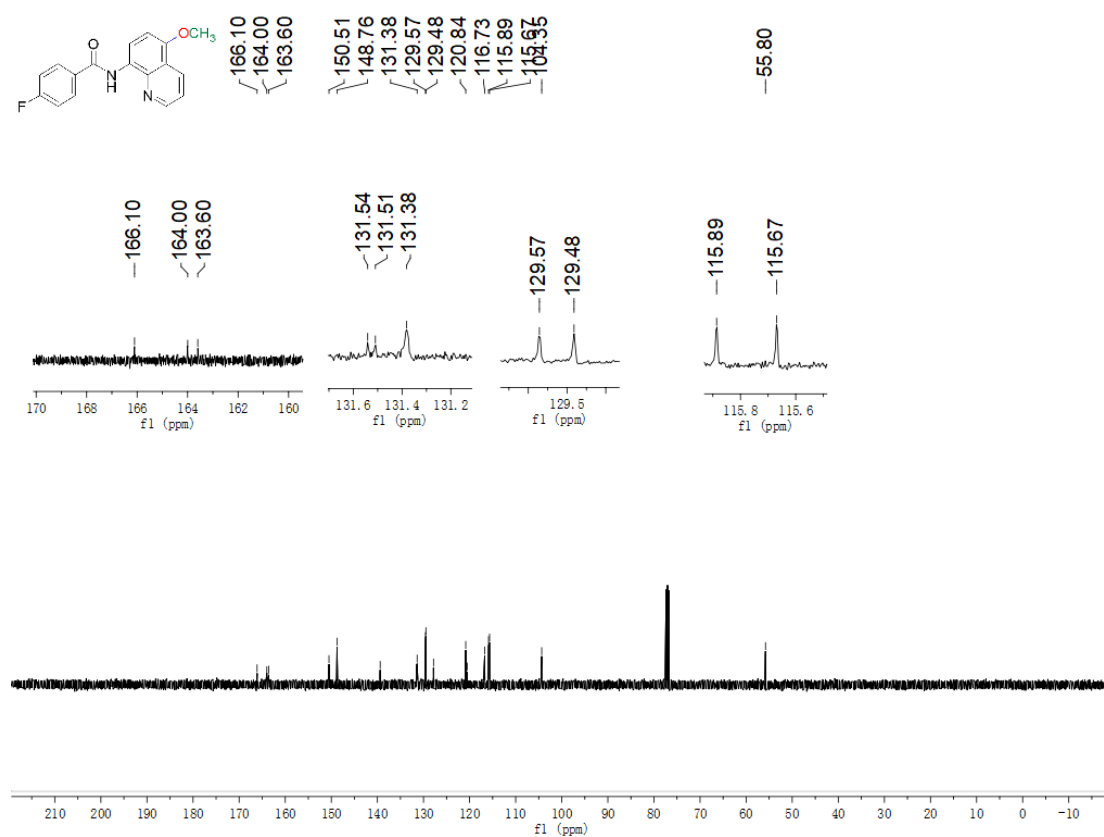
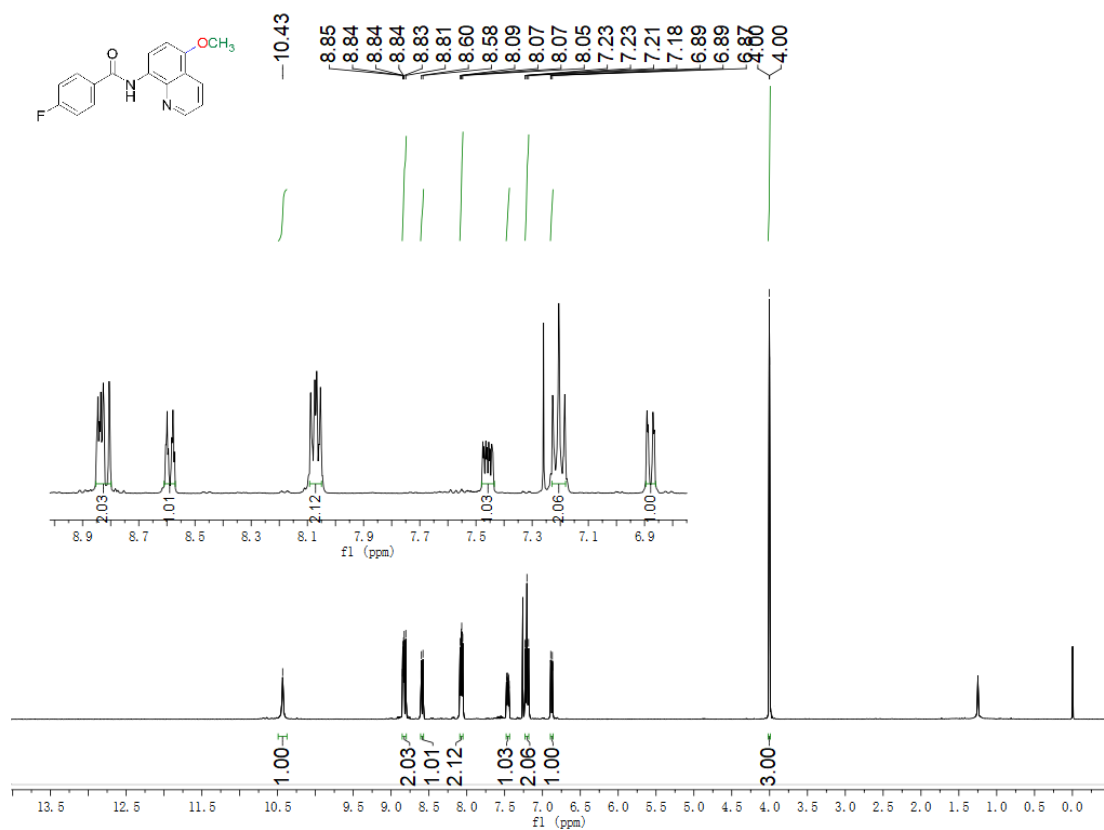
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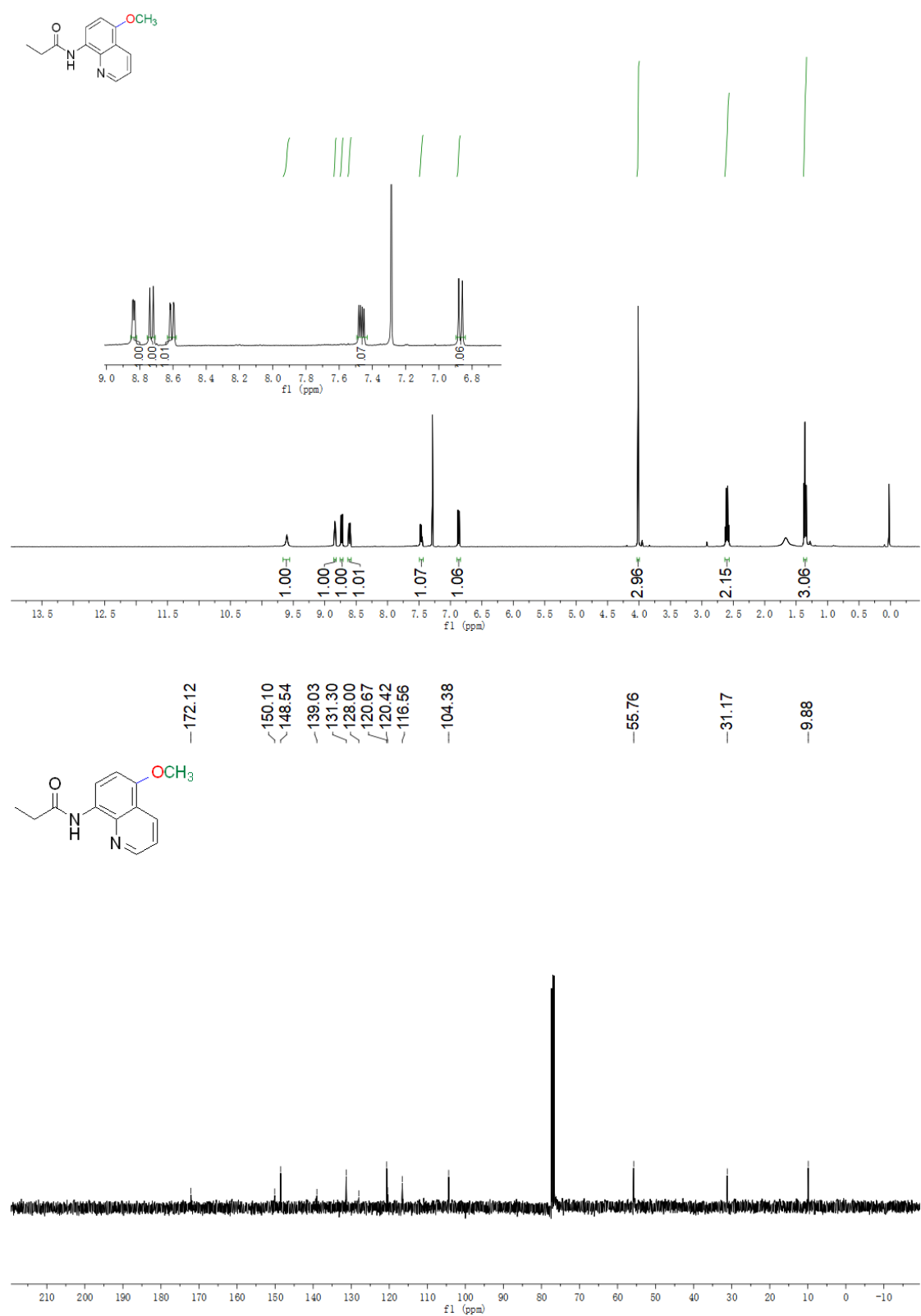
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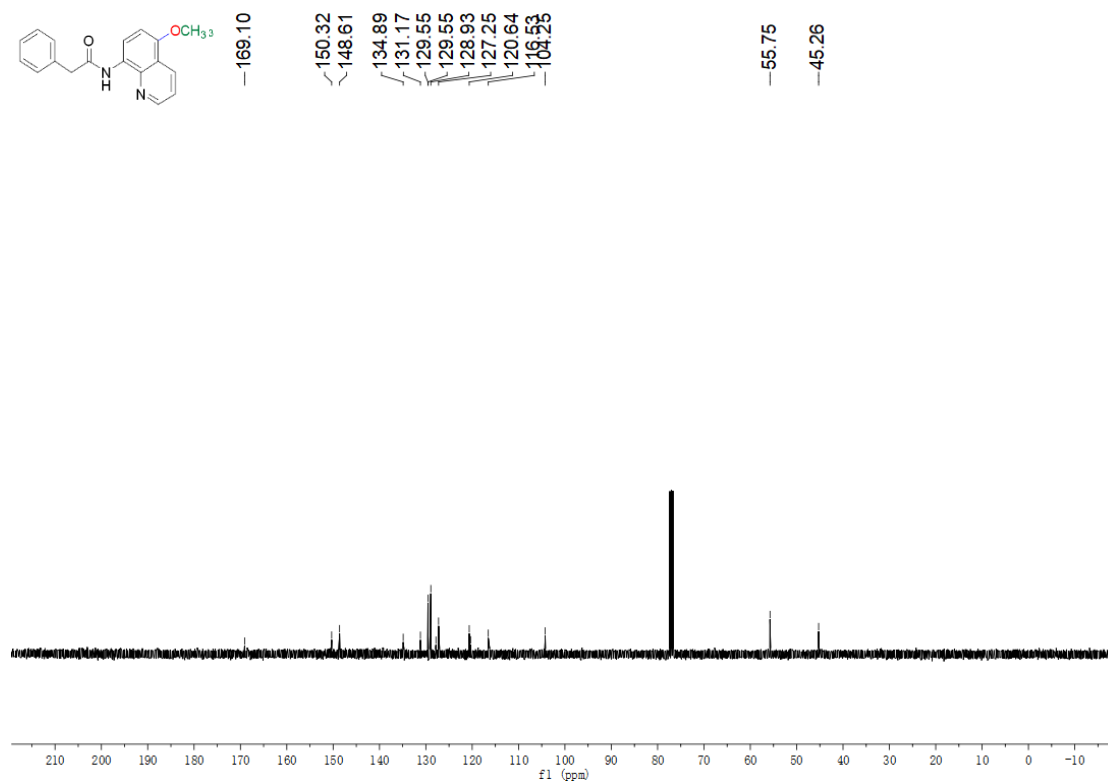
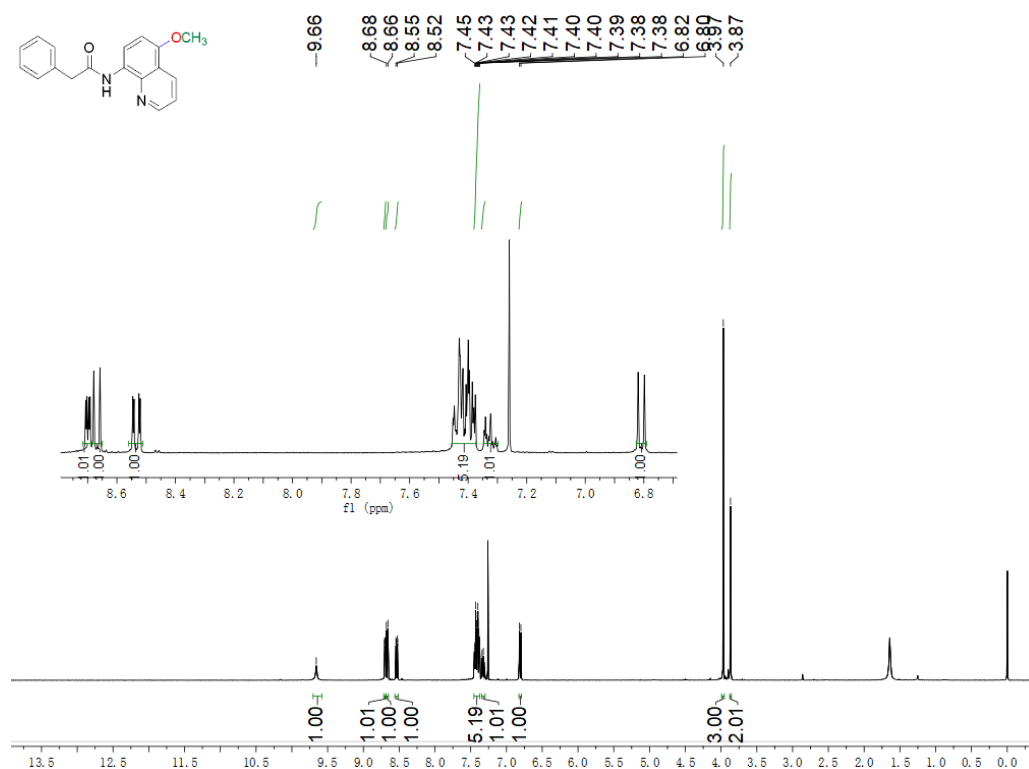
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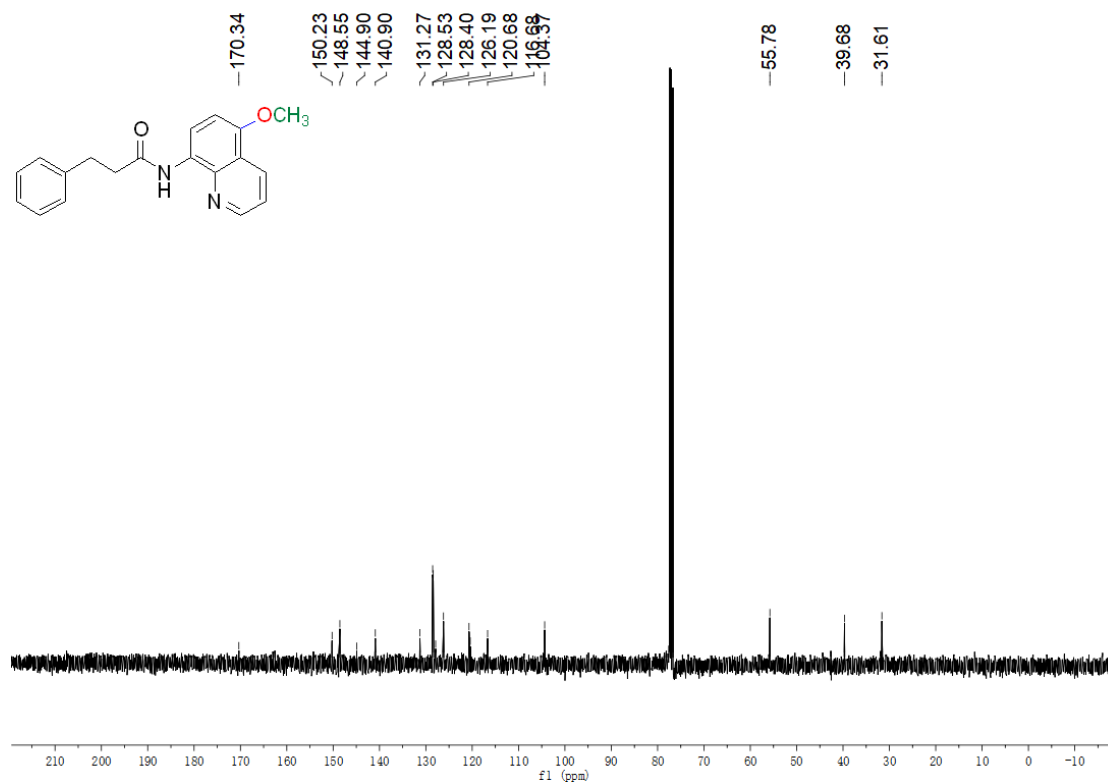
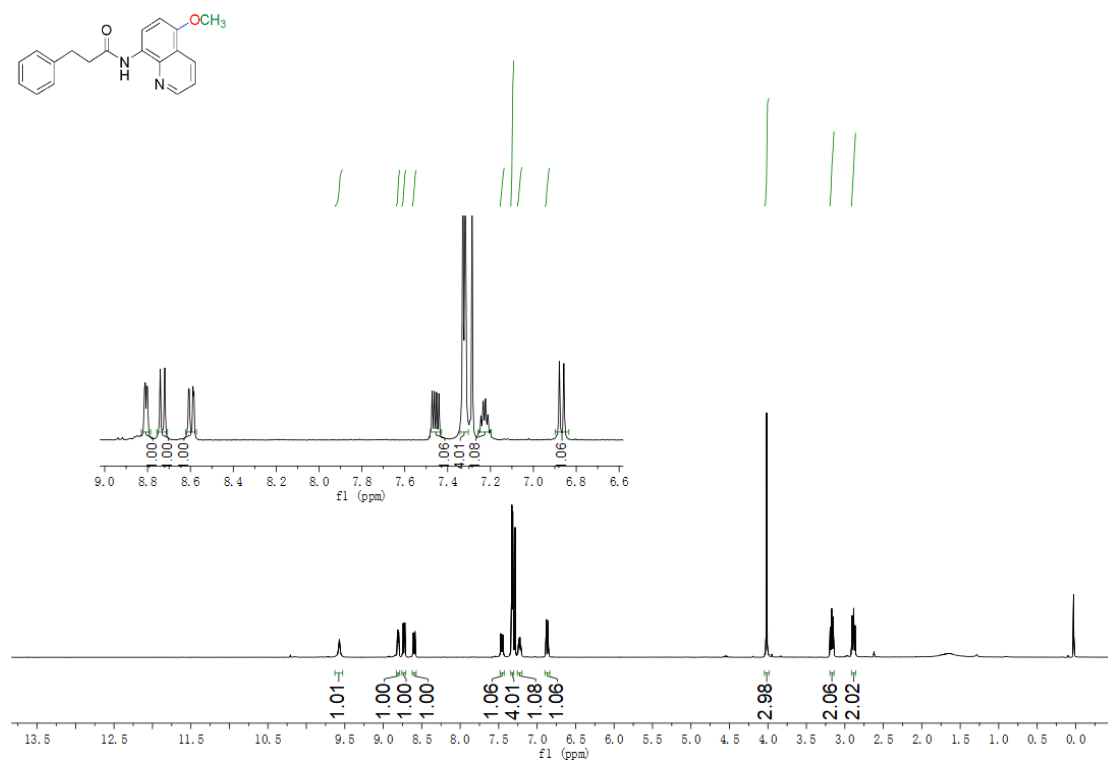
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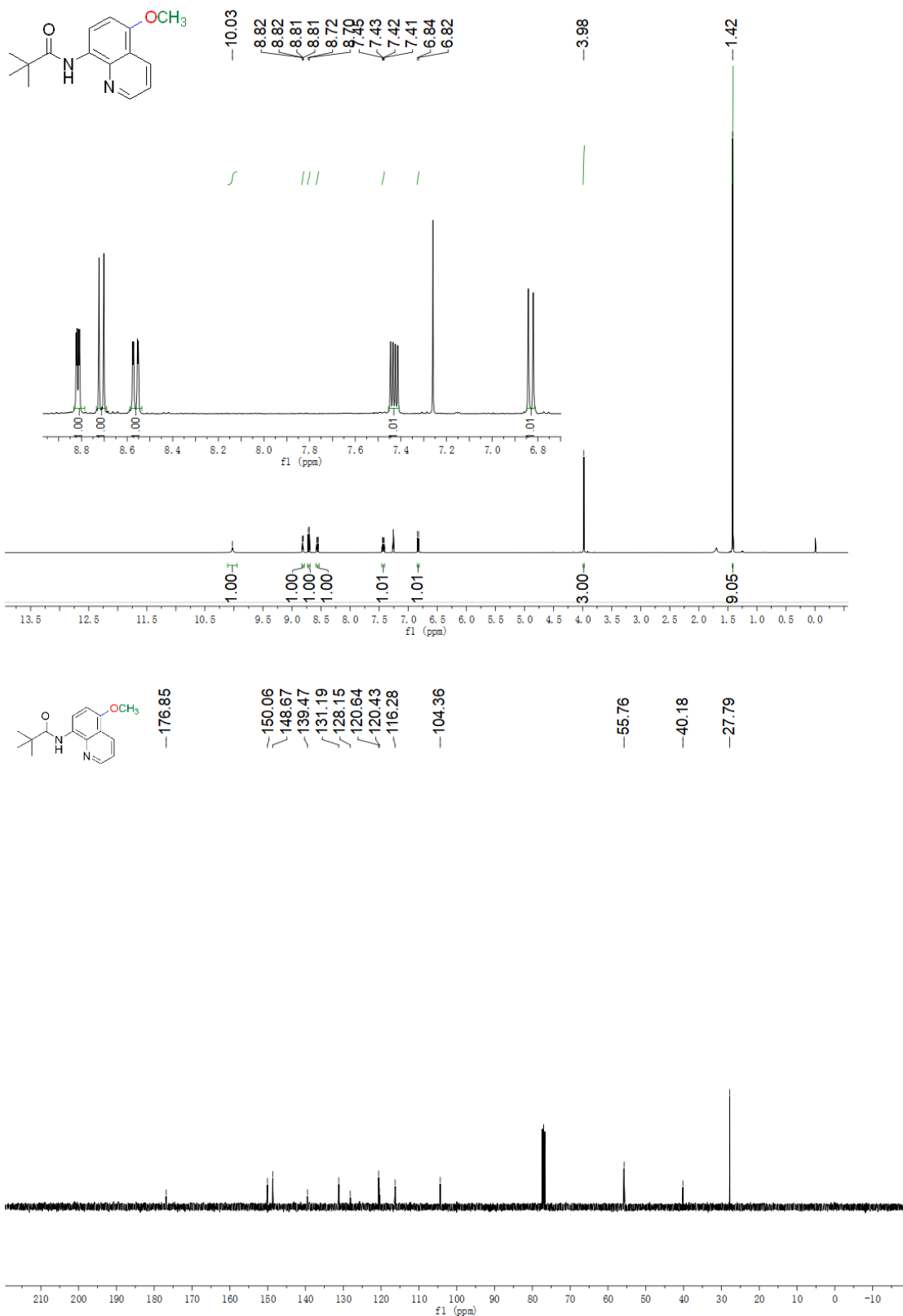
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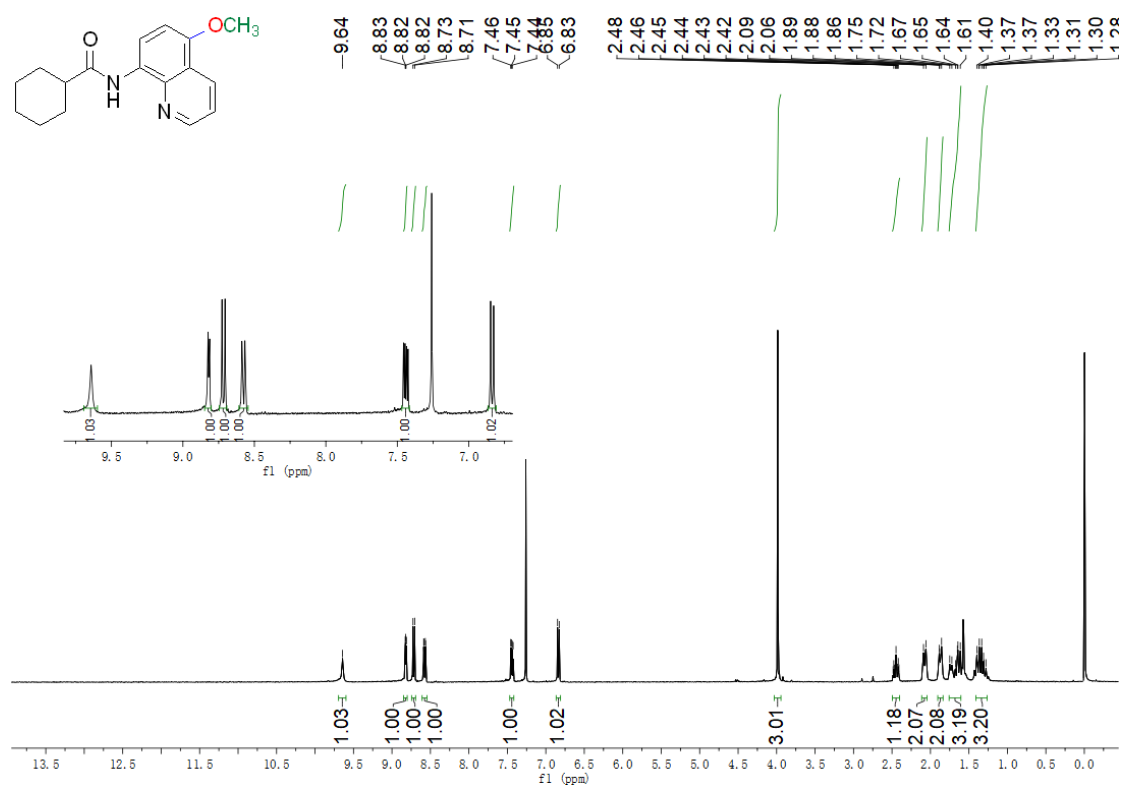
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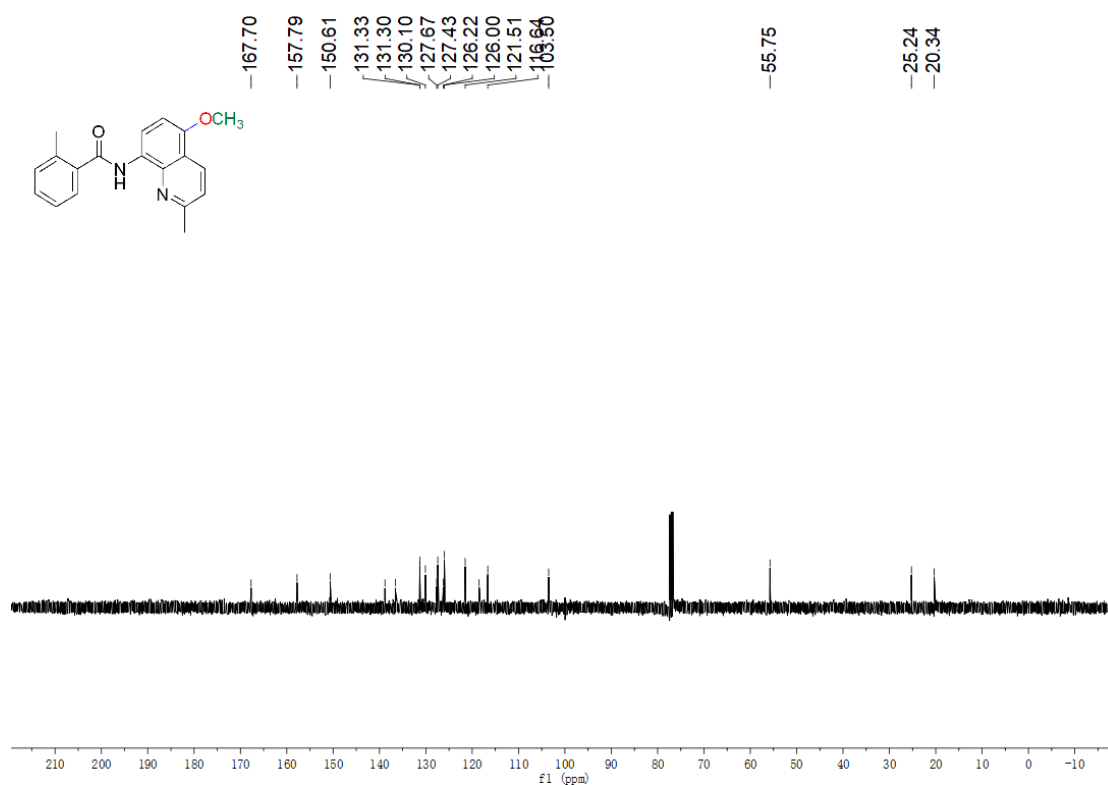
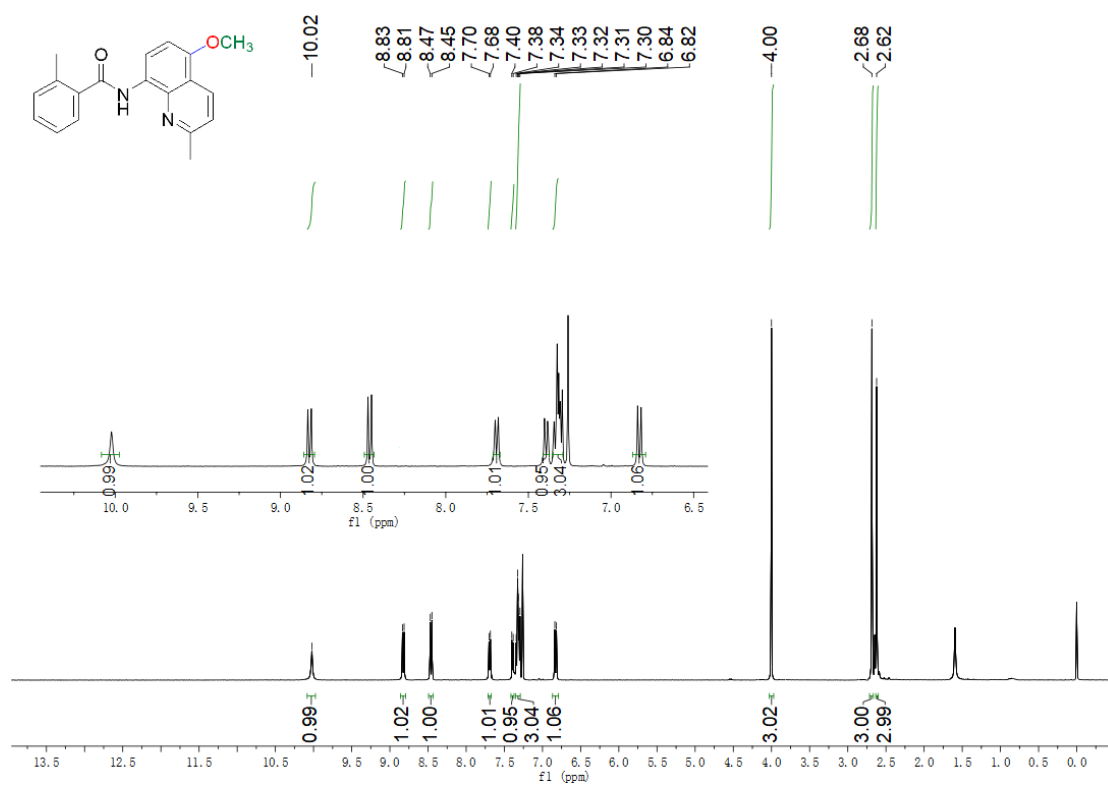
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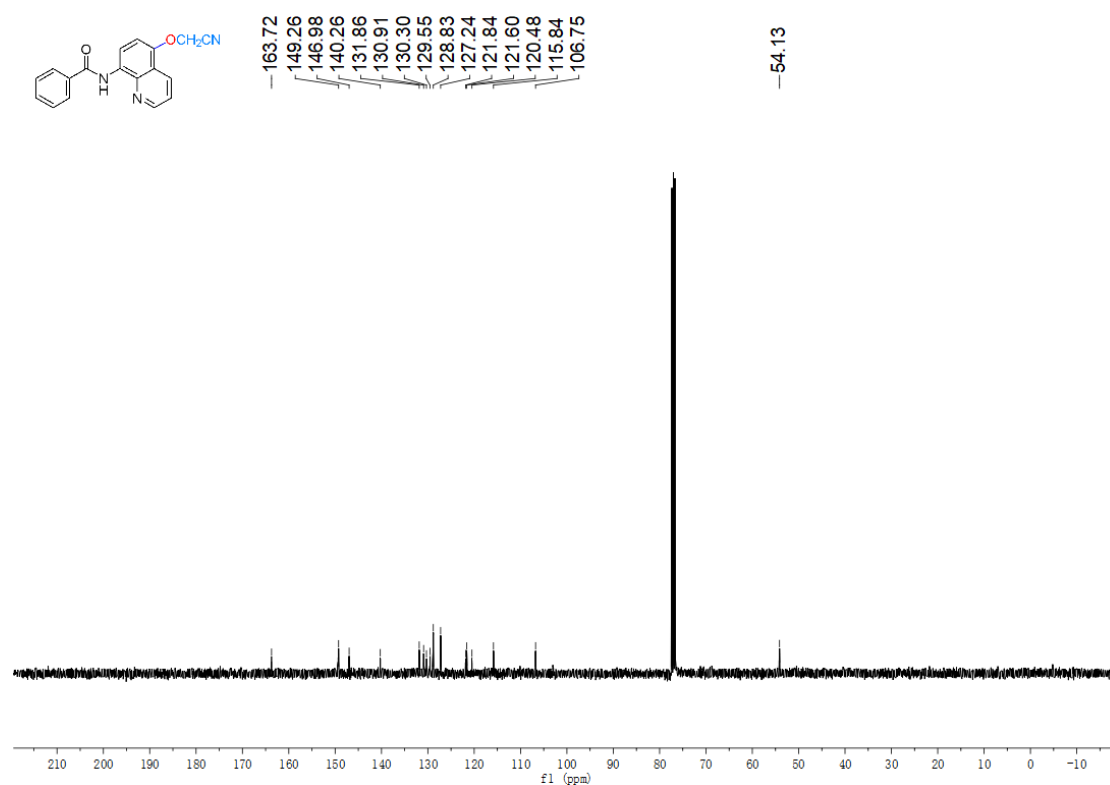
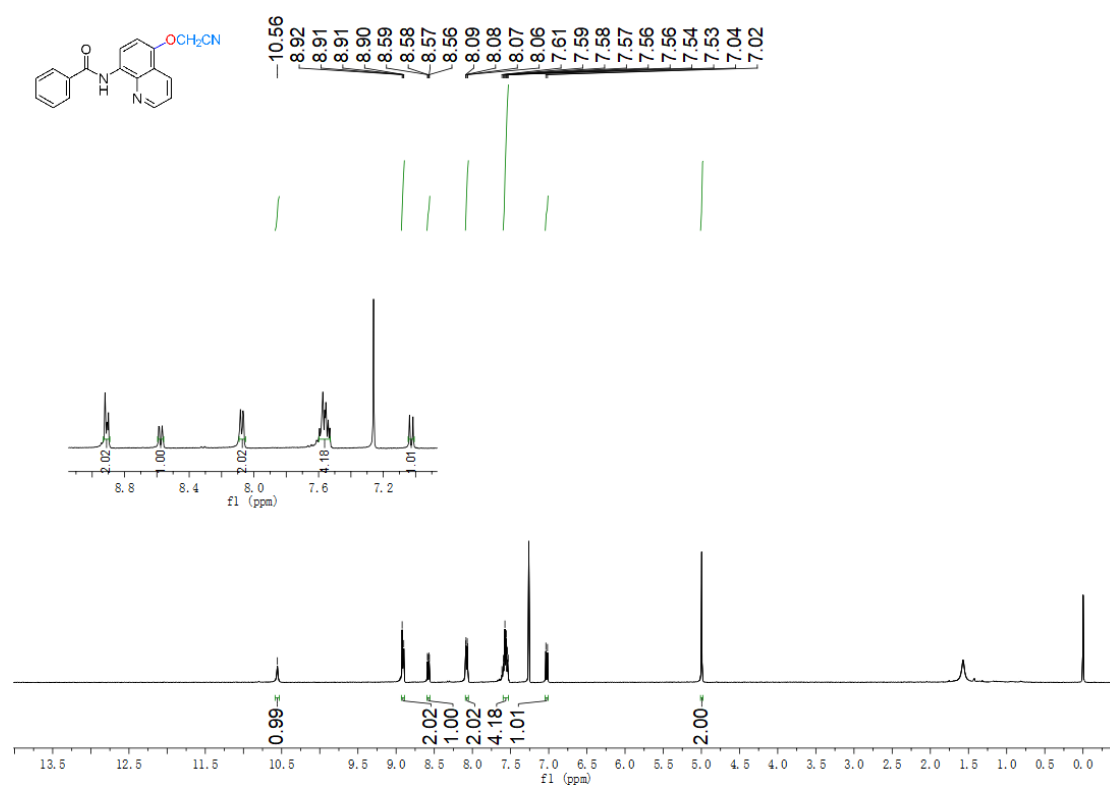
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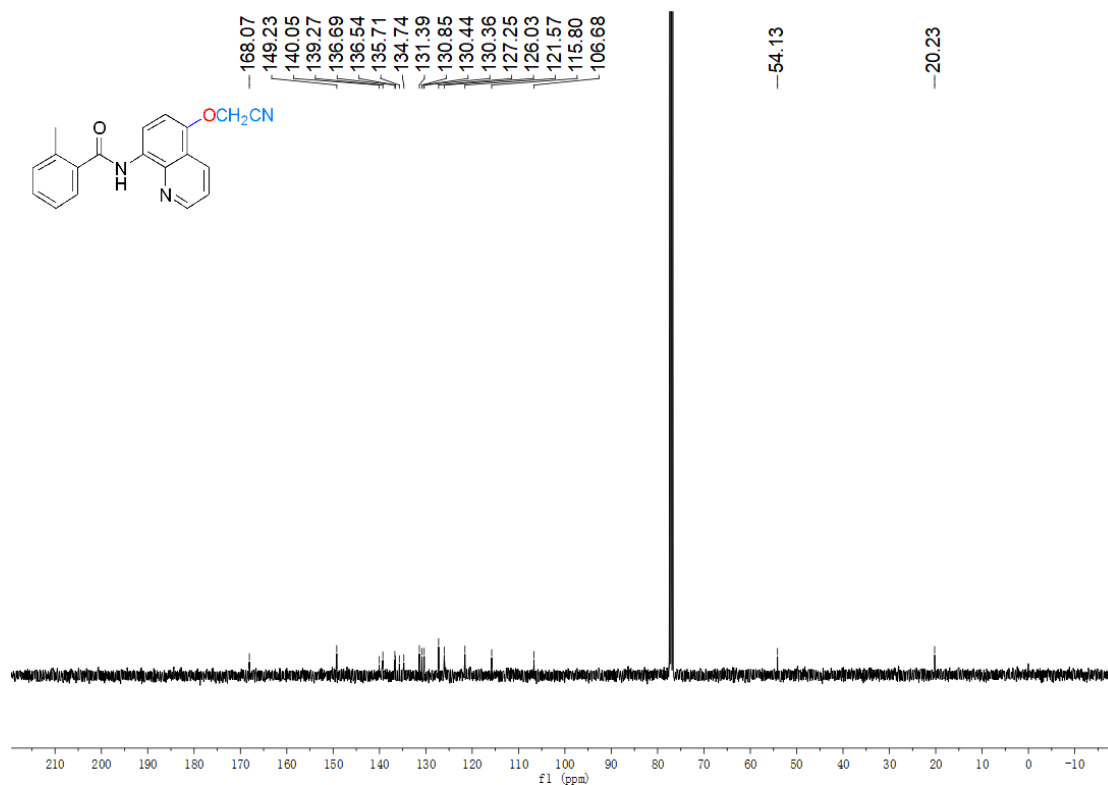
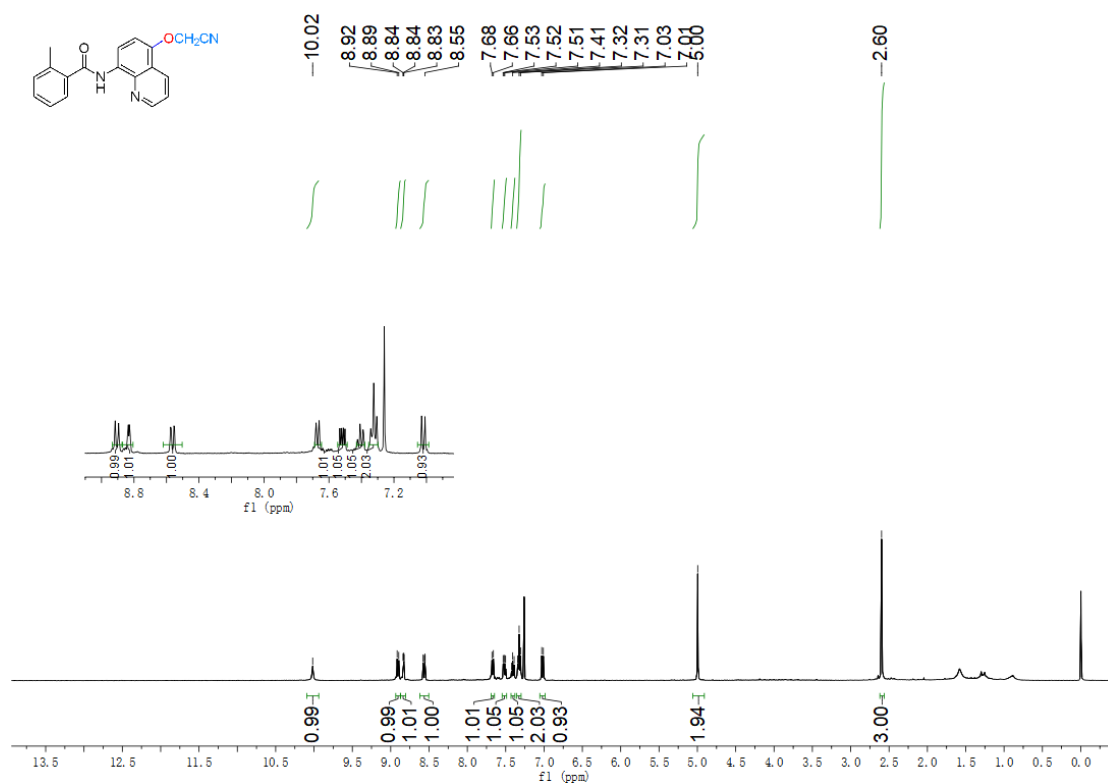
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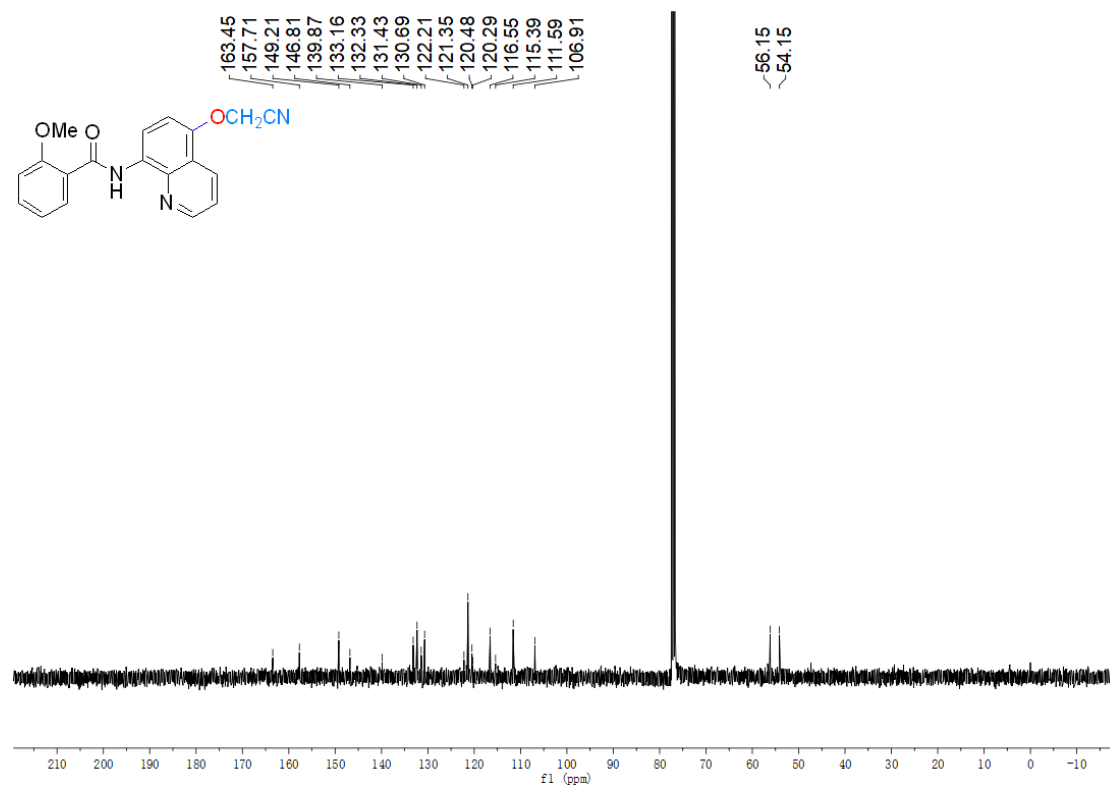
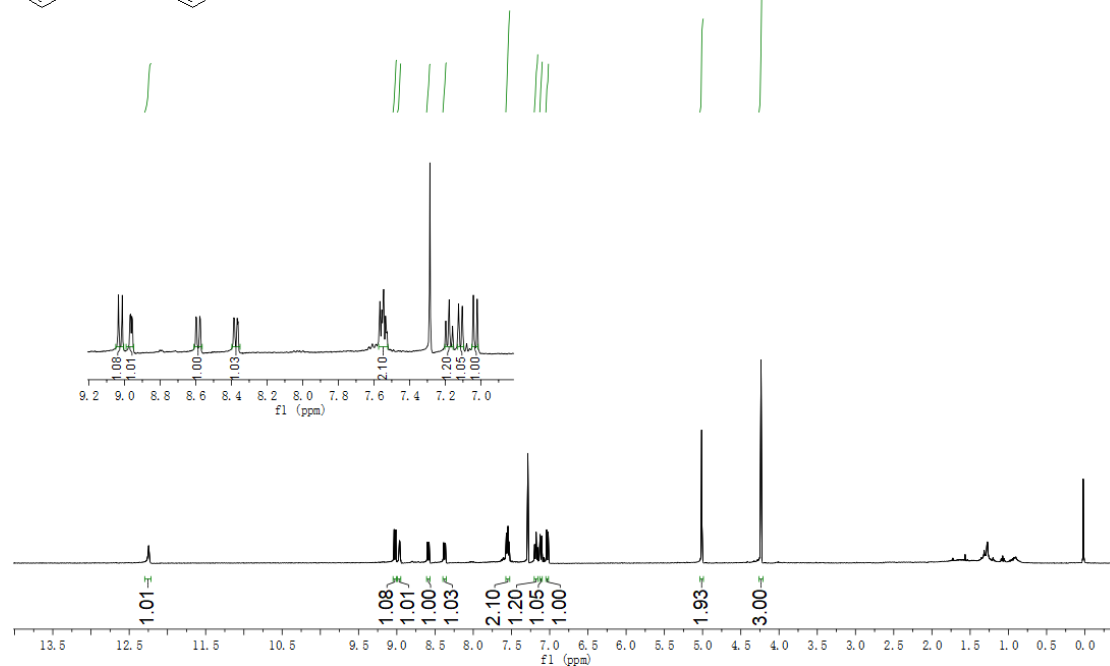
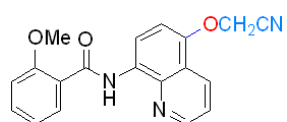
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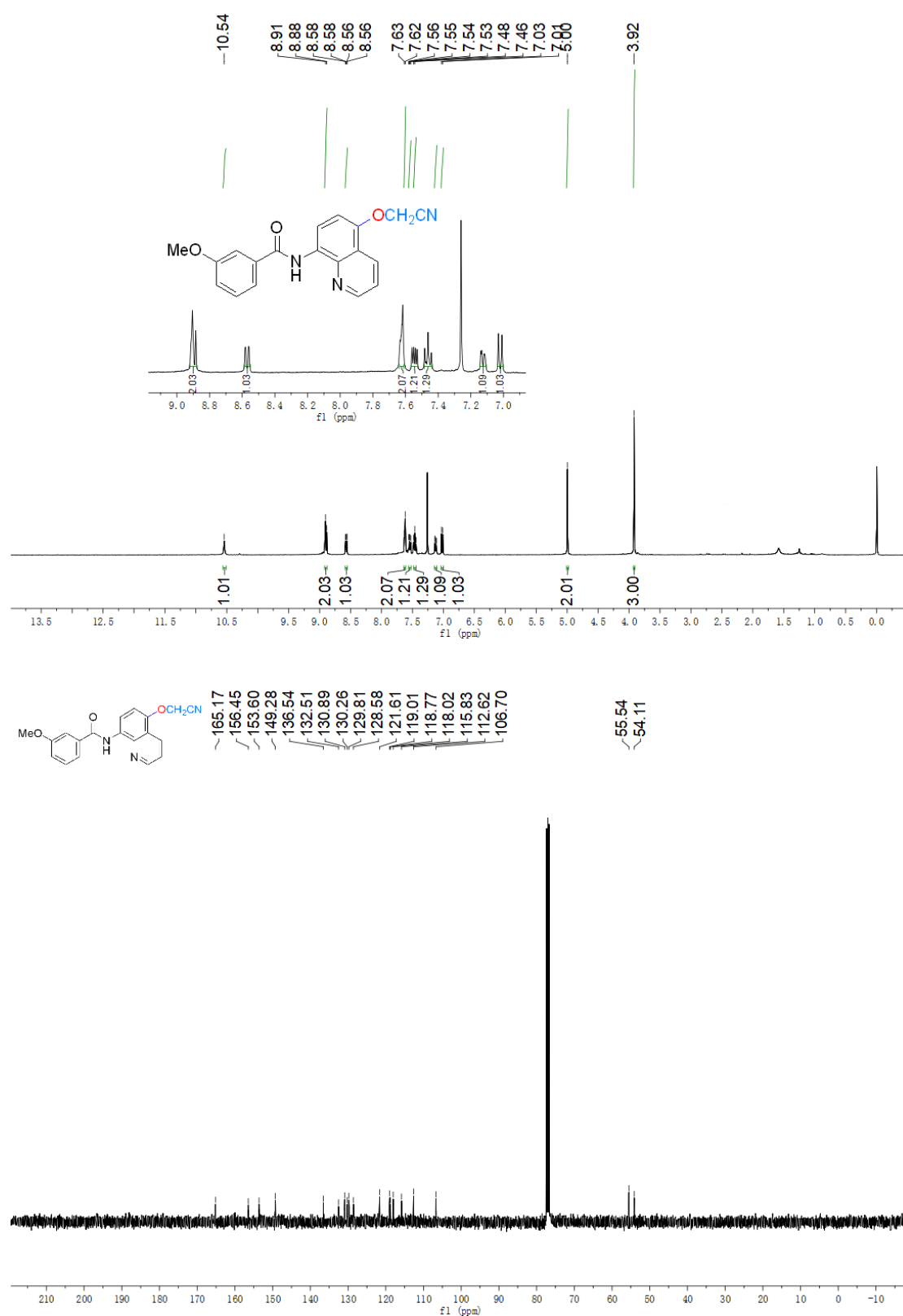
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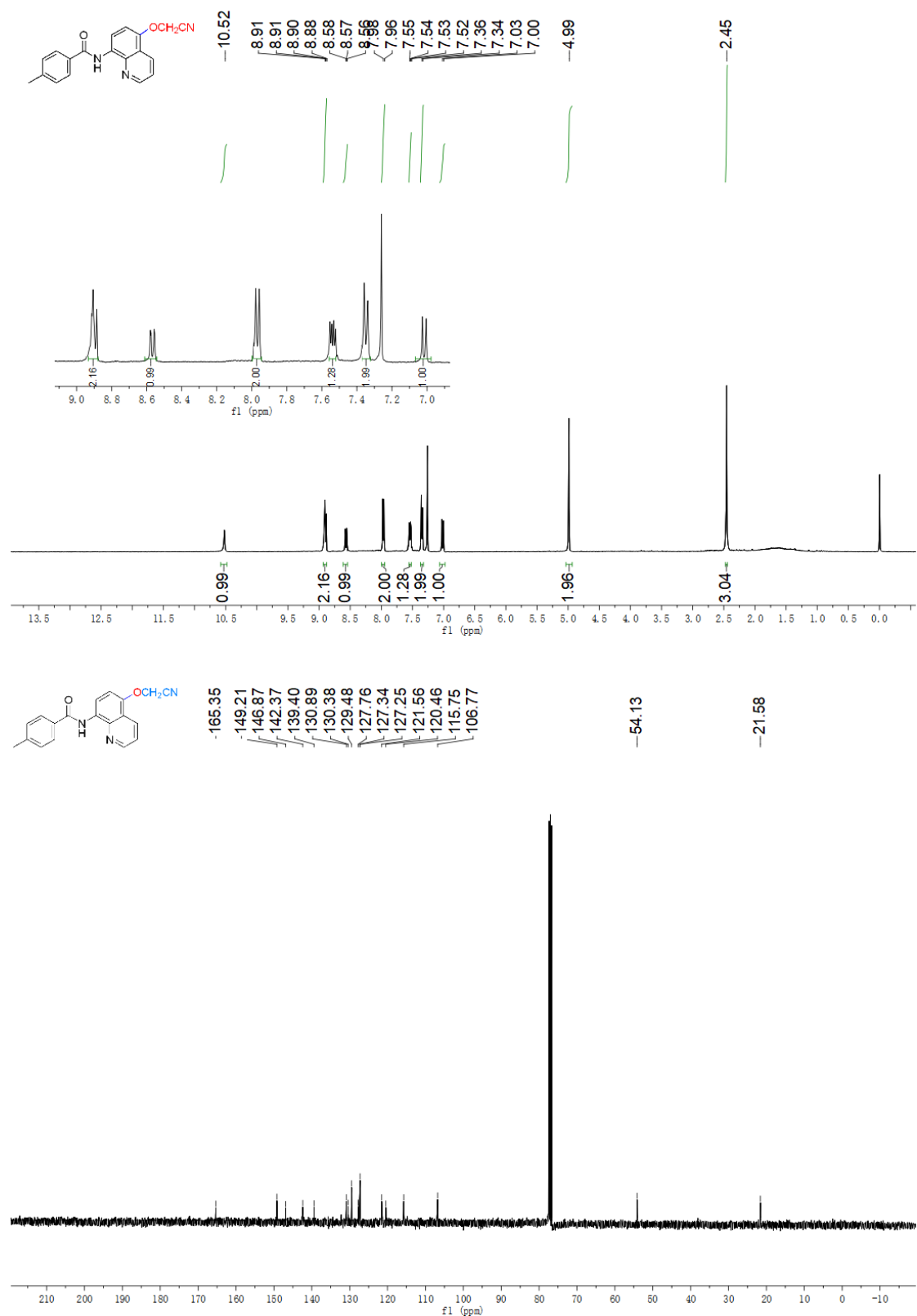
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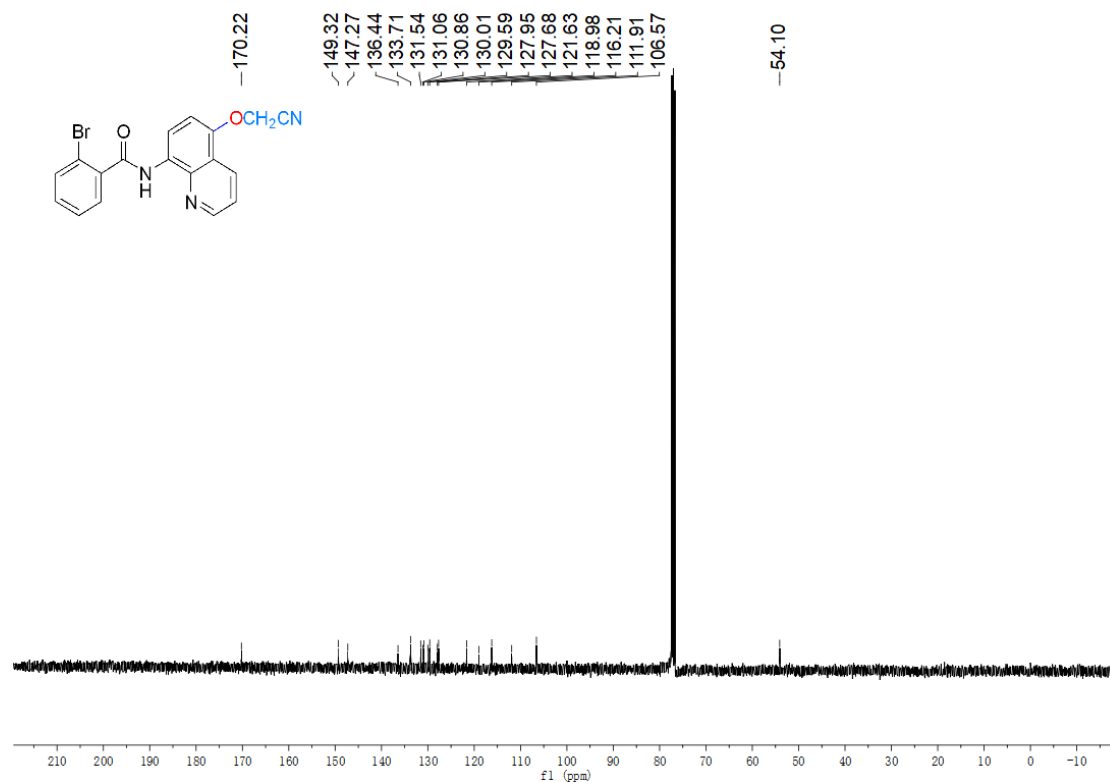
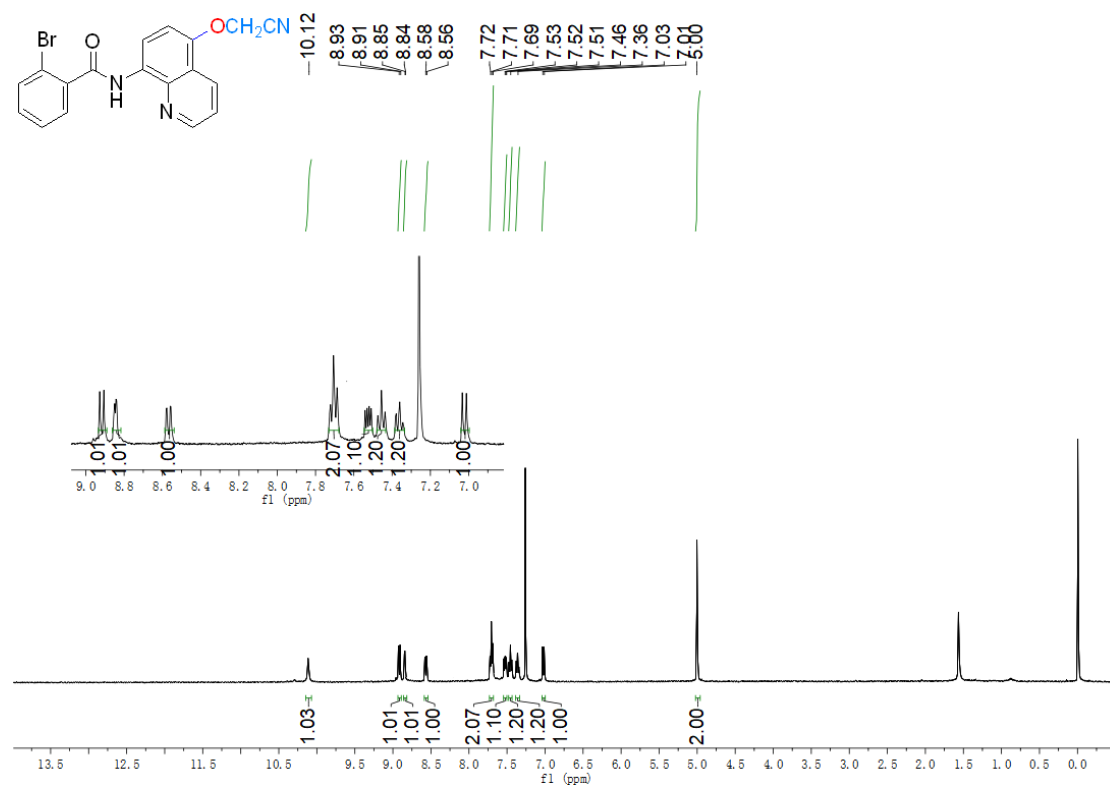
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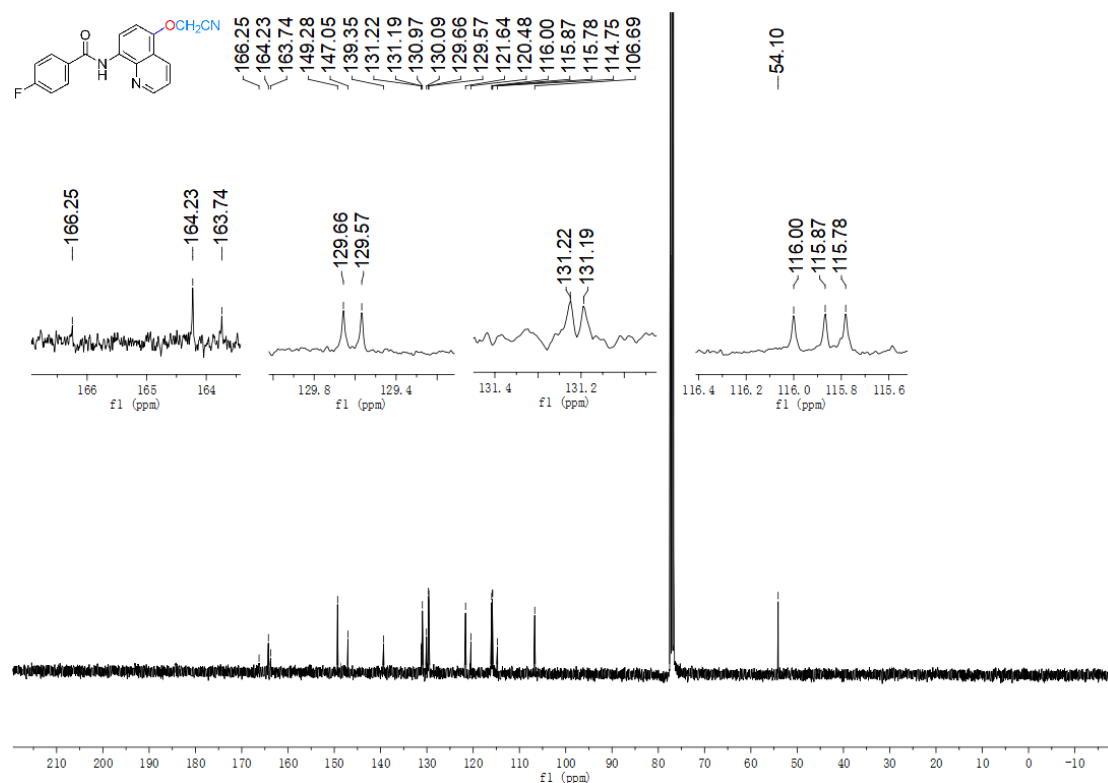
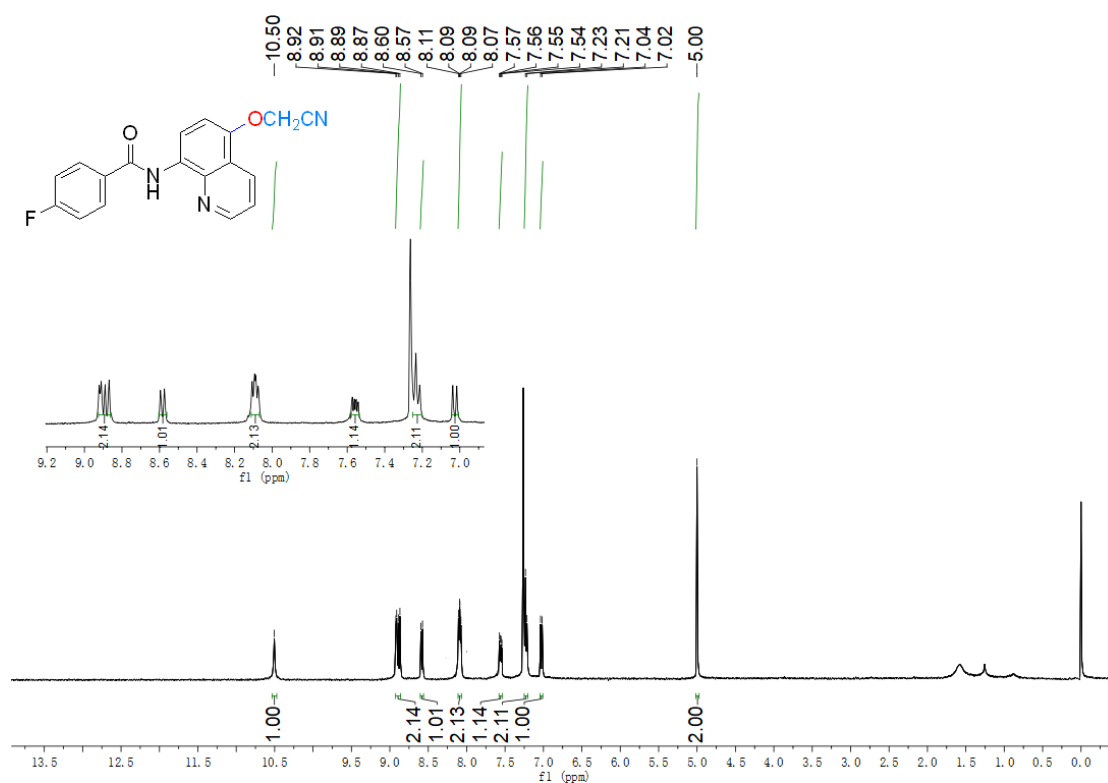
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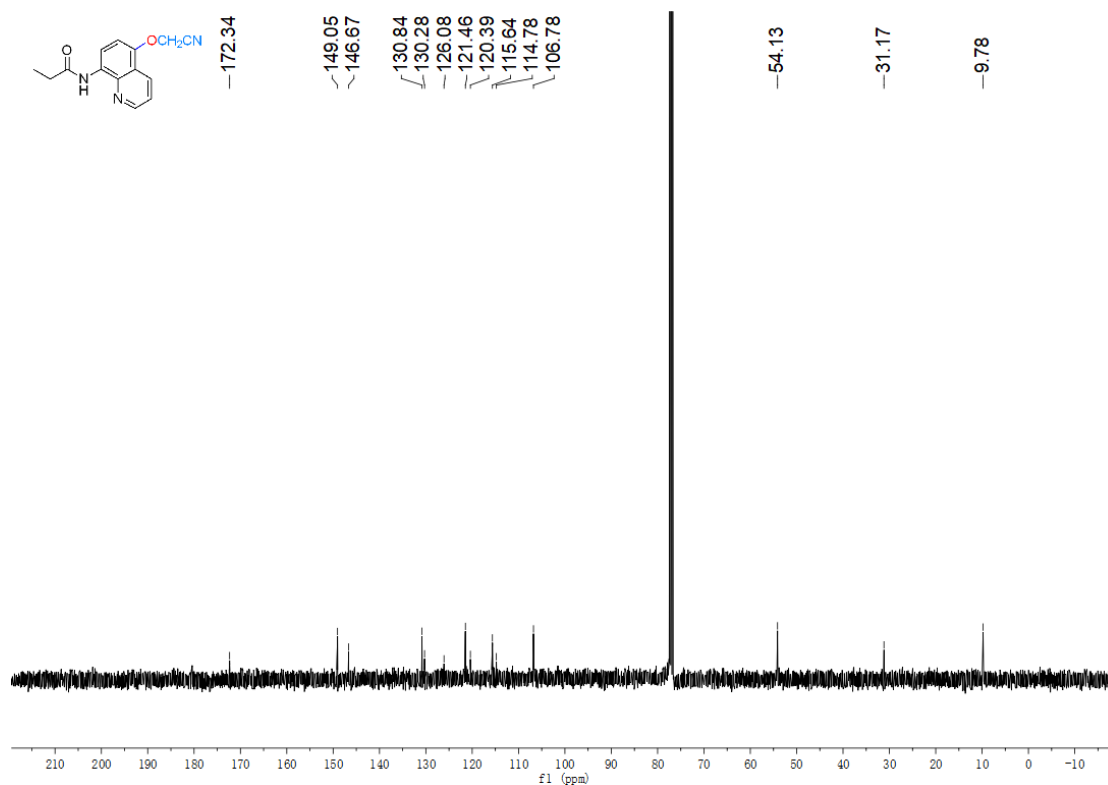
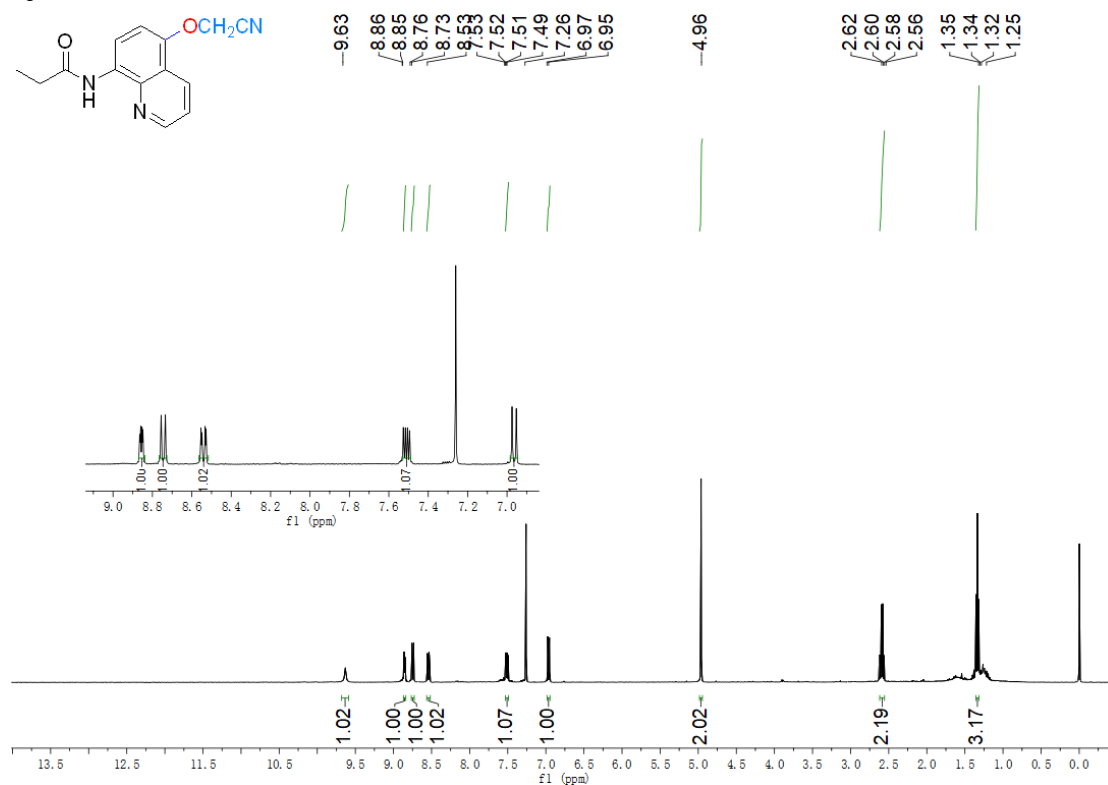
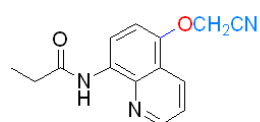
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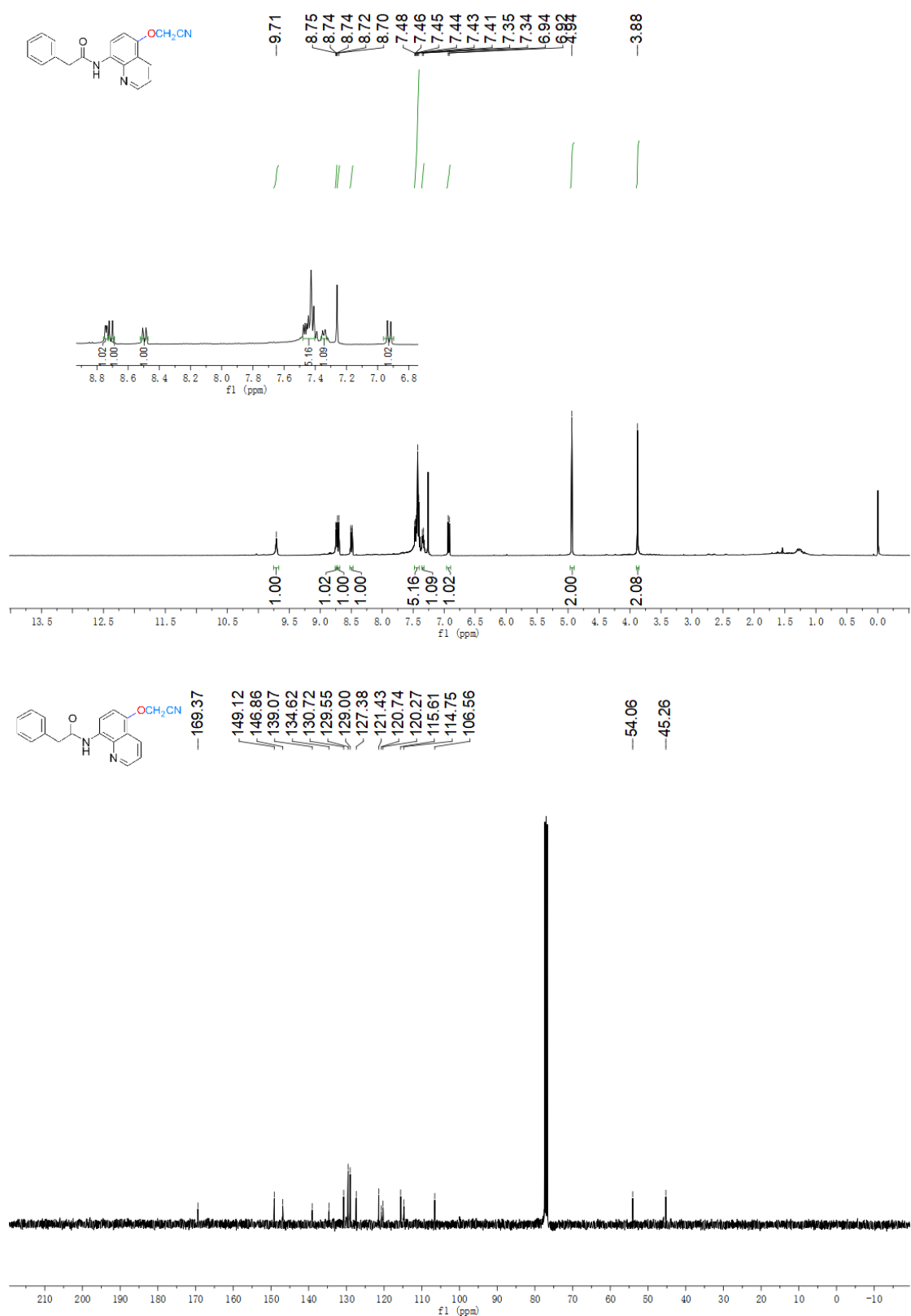
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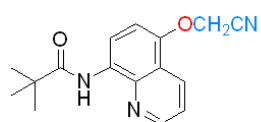
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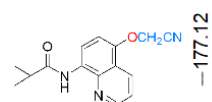
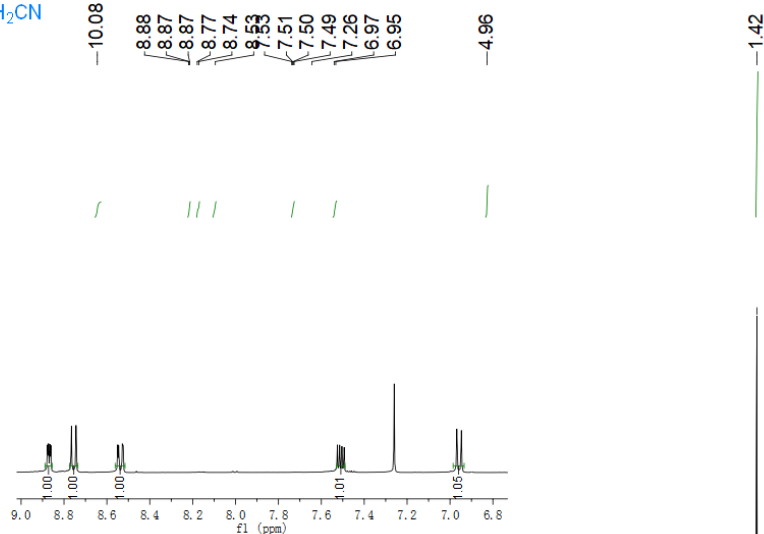
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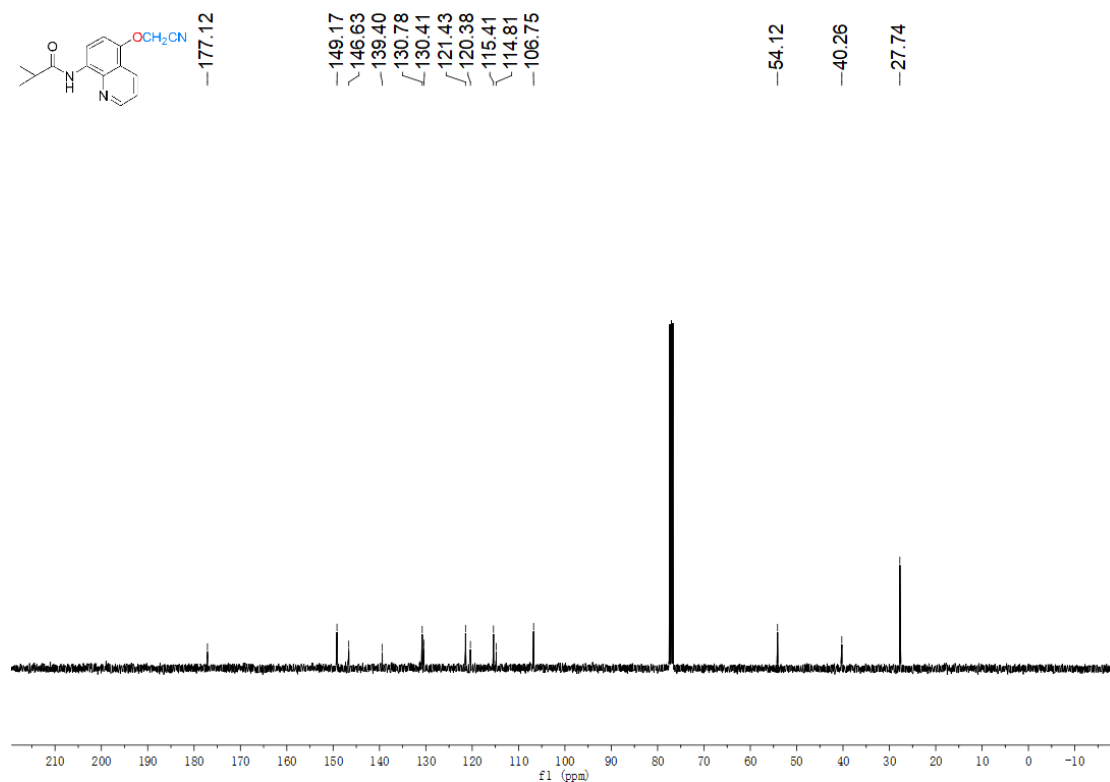
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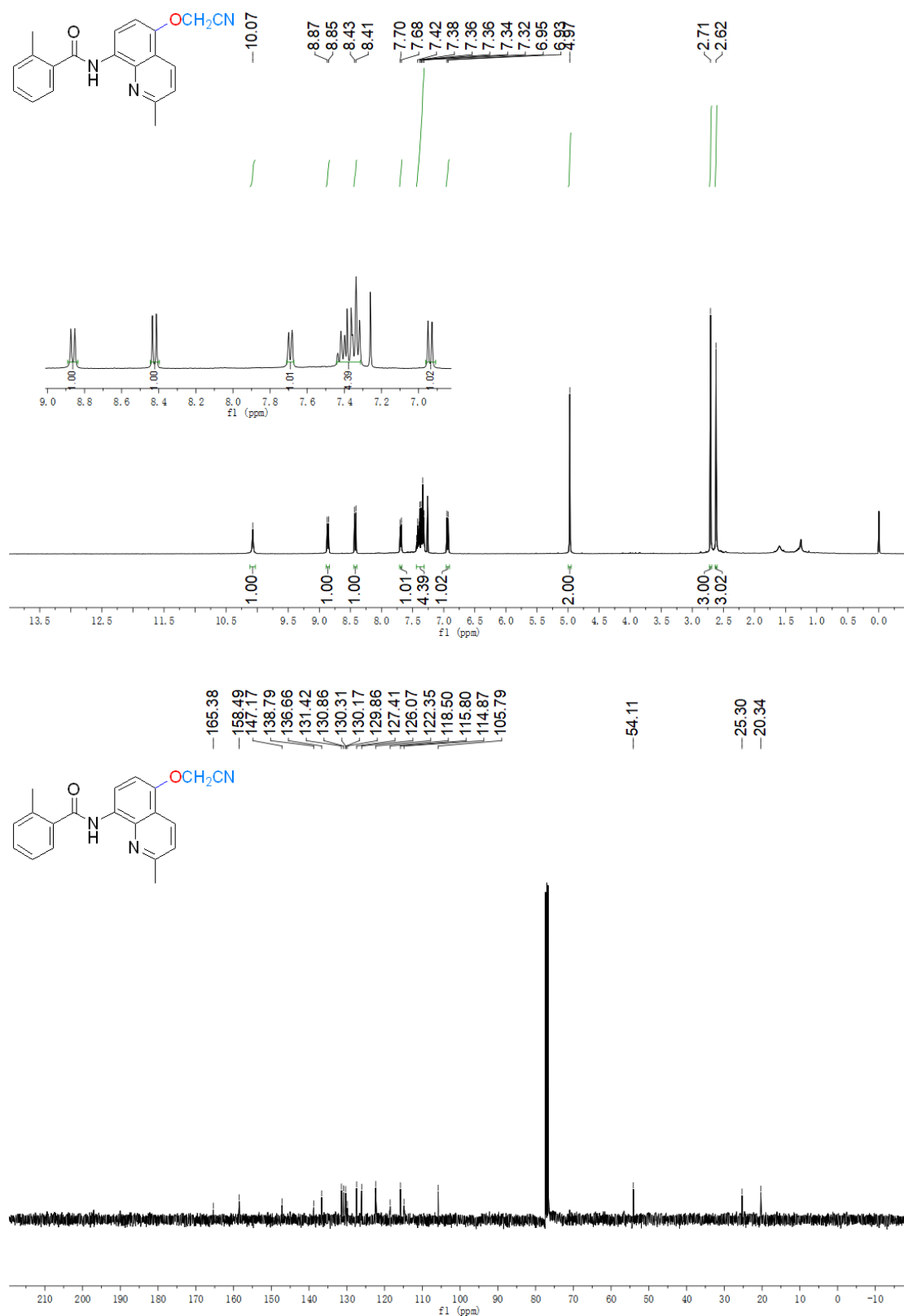
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4.96



177.12
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146.63
139.40
130.78
130.41
121.43
120.38
115.41
114.81
106.75
54.12
40.26
27.74



3s



6

