

## SUPPORTING INFORMATION

### Sulfonylation of Aryl Boronic Acids by Sulfonyl Fluorides in water under Visible Light Irradiation

Jingsong Zhen<sup>a,b</sup> , Yihui Li<sup>a</sup> , Han Yuan<sup>a</sup> , Xiaohong Xu<sup>a</sup> , Xian Du<sup>a</sup> , Xin-Qing Li<sup>c</sup> \* , Yong Luo<sup>a,b</sup> \*

<sup>a</sup>School of Pharmaceutical Sciences (Shenzhen), Shenzhen Campus of Sun Yat-sen University, Shenzhen, 518107, China.

<sup>b</sup>Key Laboratory of Functional Molecular Solids, Ministry of Education, Anhui Normal University, Wuhu, 241002, China

<sup>c</sup>Department of Pharmacy, Ganzhou People's Hospital (The Affiliated Ganzhou Hospital of Nanchang University), Ganzhou, 341000, China.

---

\* Dr. Xin-Qing Li  
E-mail address: lixinqing16@163.com

\* Dr. Yong Luo  
E-mail address: luoyong5@mail.sysu.edu.cn

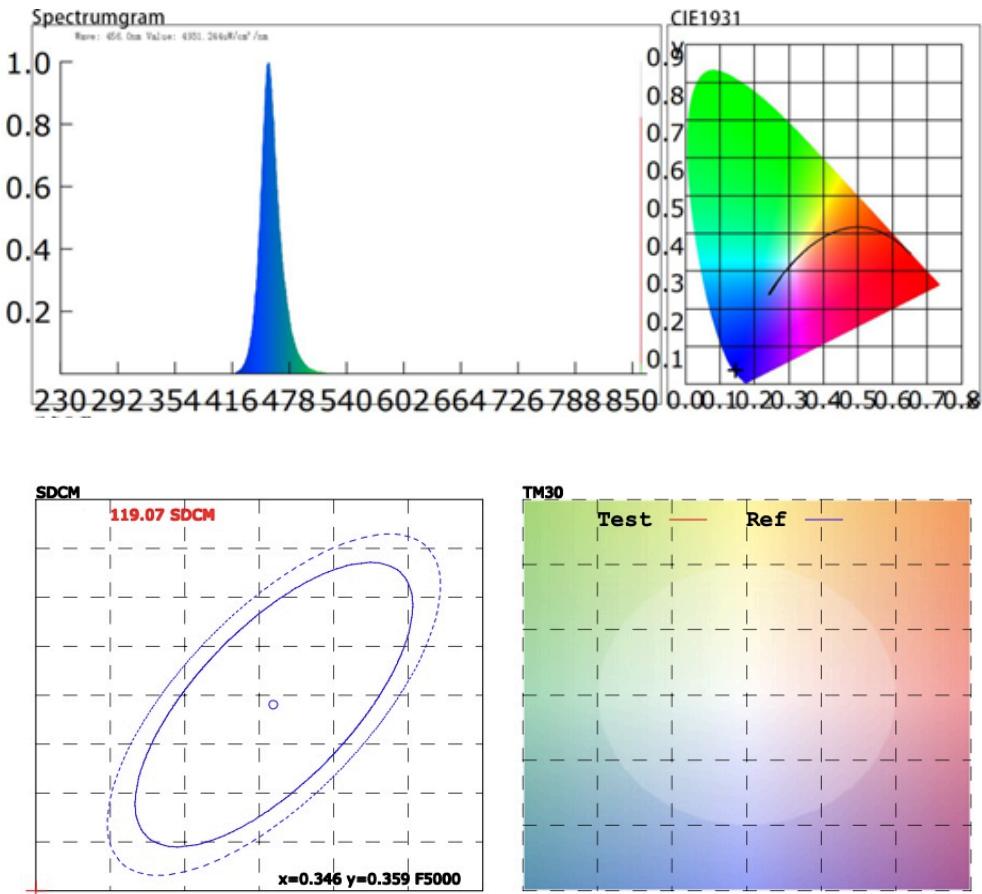
# Table of Contents

<b>1. General Information.....</b>	<b>S3</b>
<b>2. Optimization of Reaction Conditions .....</b>	<b>S4</b>
<b>3. General Procedures .....</b>	<b>S5</b>
3.1. Preparation of Sulfonyl Fluorides .....	S5
3.2. General Procedure A: Substrate Scope of Boronic Acids .....	S5
3.3. General Procedure B: Substrate Scope of Sulfonyl Fluorides .....	S5
<b>4. Derivative Experiments .....</b>	<b>S6</b>
4.1. Control Experiments: Sulfonyl Fluoride vs. Sulfonyl Chloride .....	S6
4.2. Control Experiments: Water vs. PBS .....	S6
4.3. Scale-up Experiments .....	S6
4.4. Tandem One-pot Reactions of Sulfonyl Fluorides .....	S7
<b>5. Mechanism Studies .....</b>	<b>S9</b>
5.1. UV-Vis Absorption Spectra Studies .....	S9
5.2. Stern-Volmer Quenching Experiments .....	S10
5.3. Control Experiments .....	S11
<b>6. Characterization of Products .....</b>	<b>S12</b>
<b>7. Spectra for Described Compounds .....</b>	<b>S20</b>
<b>8. References .....</b>	<b>S56</b>

## 1. General Information

Sulfonyl chlorides, boronic acids and solvents were purchased from commercial sources (Adamas-Beta Co., Bidepharm Co. or Energy Chemical Co.) and used as received.  $[\text{Ir}(\text{dF}(\text{CF}_3)\text{ppy})_2(\text{dtbbpy})]\text{PF}_6$ ,  $[\text{Ir}(\text{dF}(\text{CF}_3)\text{ppy})_2(\text{bpy})]\text{PF}_6$ ,  $[\text{Ir}(\text{ppy})_2(\text{dtbbpy})]\text{PF}_6$ ,  $[\text{Ir}(\text{ppy})_2(\text{bpy})]\text{PF}_6$ ,  $[\text{Ru}(\text{bpy})_3]\text{PF}_6$  and 4-CzIPN were prepared according to the literature procedures.<sup>[1-3]</sup> Thin-layer chromatography (TLC) was performed on glass plates coated with silica gel 60 F254, 0.2 mm thickness. The plates were visualized using a 254 nm ultraviolet lamp or iodine cylinder. Flash chromatography was performed using silica gel, 300-400 mesh. Products were identified using NMR analysis and comparison with authentic samples. NMR spectra were recorded in  $\text{CDCl}_3$  on Bruker spectrometers at 400 MHz. All chemical shifts are reported in parts per million (ppm) relative to residual  $\text{CDCl}_3$  peak (7.26 and 77.0 ppm,  $^1\text{H}$  NMR and  $^{13}\text{C}$  NMR, respectively). Data for NMR are reported as follows: chemical shift ( $\delta$  ppm), integration, multiplicity (s = singlet, d = doublet, t = triplet, q = quartet, m = multiplet), coupling constant (Hz), and assignment. High resolution mass spectra were obtained on a Waters Synapt G2-Si (ESI) or Thermo Fisher Q Exactive GC (EI). UV-Visible absorption spectra were recorded on a YOKE double beam UV-vis spectrophotometer-T3202S. UV-Visible emission spectra were recorded on a TECHCOMP fluorescence spectrometer-FL970. GC data were recorded on a FULI INSTRUMENT GC9790Plus.

Blue LEDs (30 W, 455 nm) were purchased from Shenzhen FENGSHENG Electronic Technology Co., and the parameter information was shown below.



## 2. Optimization of Reaction Conditions

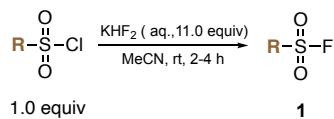
	<chem>Ts-F</chem>	<chem>+ MeOOCc1ccc(B(OH)2)cc1</chem>	<chem>PC, base, additive</chem>	<chem>MeOOCc1ccc(Ts)cc1</chem>
	<b>1a</b>	<b>2a</b>	<small>solvent, rt, 12h Blue LEDs</small>	<b>3a</b>
Entry	PC	base	additive	<sup>b</sup> Yield (%)
1	[Ir(ppy <sub>2</sub> )(dtbbpy)]PF <sub>6</sub>	K <sub>2</sub> CO <sub>3</sub>	-	1,4-dioxane trace
2	[Ru(bpy) <sub>3</sub> ]PF <sub>6</sub>	K <sub>2</sub> CO <sub>3</sub>	-	1,4-dioxane nd
3	Solvent Red	K <sub>2</sub> CO <sub>3</sub>	-	1,4-dioxane trace
4	Rhodamine B	K <sub>2</sub> CO <sub>3</sub>	-	1,4-dioxane 11
5	Fluorescin	K <sub>2</sub> CO <sub>3</sub>	-	1,4-dioxane trace
6	Mes-Arc	K <sub>2</sub> CO <sub>3</sub>	-	1,4-dioxane nd
7	4-CzIPN	K <sub>2</sub> CO <sub>3</sub>	-	1,4-dioxane 70
8	4-CzIPN	-	-	1,4-dioxane nd
9	4-CzIPN	CsF	-	1,4-dioxane 38
10	4-CzIPN	K <sub>2</sub> HPO <sub>4</sub>	-	1,4-dioxane 16
11	4-CzIPN	K <sub>3</sub> PO <sub>4</sub>	-	1,4-dioxane 14
12	4-CzIPN	Cs <sub>2</sub> CO <sub>3</sub>	-	1,4-dioxane 68
13	4-CzIPN	NET <sub>3</sub>	-	1,4-dioxane nd
14	4-CzIPN	DMAP	-	1,4-dioxane nd
15	4-CzIPN	DABCO	-	1,4-dioxane nd
16	4-CzIPN	Rb <sub>2</sub> CO <sub>3</sub>	-	1,4-dioxane 74
17	4-CzIPN	Rb <sub>2</sub> CO <sub>3</sub>	-	THF 46
18	4-CzIPN	Rb <sub>2</sub> CO <sub>3</sub>	-	DME 24
19	4-CzIPN	Rb <sub>2</sub> CO <sub>3</sub>	-	DMSO 13
20	4-CzIPN	Rb <sub>2</sub> CO <sub>3</sub>	-	DMA 15
21	4-CzIPN	Rb <sub>2</sub> CO <sub>3</sub>	-	MeCN trace
22	4-CzIPN	Rb <sub>2</sub> CO <sub>3</sub>	-	H <sub>2</sub> O nd
23	4-CzIPN	Rb <sub>2</sub> CO <sub>3</sub>	Tween20	H <sub>2</sub> O 80
24	4-CzIPN	Rb <sub>2</sub> CO <sub>3</sub>	Tween60	H <sub>2</sub> O 74
25	4-CzIPN	Rb <sub>2</sub> CO <sub>3</sub>	Span40	H <sub>2</sub> O nd
26	4-CzIPN	Rb <sub>2</sub> CO <sub>3</sub>	TBAB	H <sub>2</sub> O 23
27	4-CzIPN	Rb <sub>2</sub> CO <sub>3</sub>	BrijL23	H <sub>2</sub> O 43
28	4-CzIPN	Rb <sub>2</sub> CO <sub>3</sub>	DDBS	H <sub>2</sub> O 13
29	4-CzIPN	Rb <sub>2</sub> CO <sub>3</sub>	Betaine	H <sub>2</sub> O nd
30	-	Rb <sub>2</sub> CO <sub>3</sub>	Tween20	H <sub>2</sub> O nd
31	4-CzIPN	Rb <sub>2</sub> CO <sub>3</sub>	Tween20	H <sub>2</sub> O <sup>c</sup> nd
32	4-CzIPN	Rb <sub>2</sub> CO <sub>3</sub>	Tween20	H <sub>2</sub> O <sup>d</sup> nd

<sup>a</sup> Reaction conditions:: **1a** (0.3 mmol, 1.5 equiv), **2a** (0.2 mmol, 1.0 equiv), PC (1 mol%), base (0.4 mmol, 2.0 equiv), additive (3.2 equiv), solvent (1 mL), 30 W blue LED, N<sub>2</sub>, 12 h, room temperature. nd = not detected. TBAB = Tetrabutylammonium bromide. BrijL23 = Polyoxyethylene lauryl ether. DDBS = Sodium dodecylbenzenesulphonate.

<sup>b</sup> GC yield was calculated with Dodecane as internal standard. <sup>c</sup> Under air. <sup>d</sup> In dark.

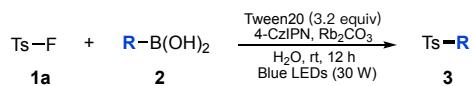
### 3. General Procedures

#### 3.1 Preparation of Sulfonyl Fluorides (Cl-F exchanging)



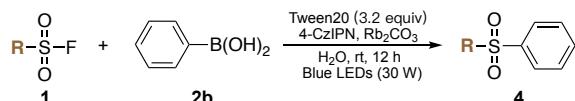
Sulfonyl chloride (5 mmol, 1.0 equiv) was dissolved in MeCN (10 mL), and aq. KHF<sub>2</sub> (10 mL, 11.0 equiv) was added. The suspension was stirred for 2-4 hours at room temperature. Then the reaction mixture was diluted with H<sub>2</sub>O (5 mL) and extracted with EtOAc (3 x 5 mL). The combined organic layer was dried over Na<sub>2</sub>SO<sub>4</sub>, and filtrate was concentrated in vacuo to directly give the desired sulfonyl fluoride.

#### 3.2. General Procedure A: Substrate Scope of Boronic Acids



An oven-dried Schlenk tube equipped with a stir bar was charged with sulfonyl fluoride **1a** (0.3 mmol, 1.5 equiv), boronic acid **2** (0.2 mmol, 1.0 equiv), Rb<sub>2</sub>CO<sub>3</sub> (0.4 mmol, 2.0 equiv) and 4-CzIPN (1 mol%). Then the tube was sealed with a cap, evacuated and backfilled with nitrogen for 3 times. Subsequently, Tween20 (0.2 mL, 3.2 equiv) and H<sub>2</sub>O (1 mL) was added. The tube was placed in a water bath with irradiation by 30 W blue LEDs (455 nm, distance: 1 cm, the reaction temperature reached room temperature) for 12 hours. Then the mixture was washed with sat. aq. NH<sub>4</sub>Cl (1 x 2 mL), diluted with EtOAc (1 x 4 mL), and concentrated. The crude product was purified by flash chromatography (EtOAc/PE: 1/4, unless otherwise noted) to give analytically pure product **3**.

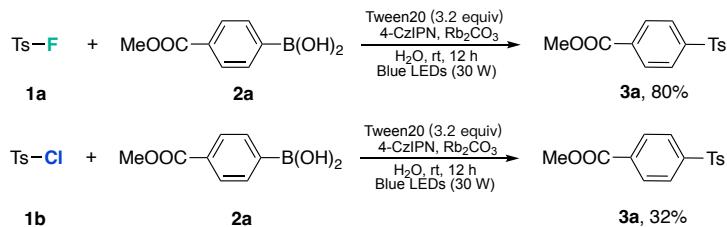
#### 3.3. General Procedure B: Substrate Scope of Sulfonyl Fluorides



An oven-dried Schlenk tube equipped with a stir bar was charged with sulfonyl fluoride **1** (0.3 mmol, 1.5 equiv), boronic acid **2b** (0.2 mmol, 1.0 equiv), Rb<sub>2</sub>CO<sub>3</sub> (0.4 mmol, 2.0 equiv) and 4-CzIPN (1 mol%). Then the tube was sealed with a cap, evacuated and backfilled with nitrogen for 3 times. Subsequently, Tween20 (0.2 mL, 3.2 equiv) and H<sub>2</sub>O (1 mL) was added. The tube was placed in a water bath with irradiation by 30 W blue LEDs (455 nm, distance: 1 cm, the reaction temperature reached room temperature) for 12 hours. Then the mixture was washed with sat. aq. NH<sub>4</sub>Cl (1 x 2 mL), diluted with EtOAc (1 x 4 mL), and concentrated. The crude product was purified by flash chromatography (EtOAc/PE: 1/4, unless otherwise noted) to give analytically pure product **4**.

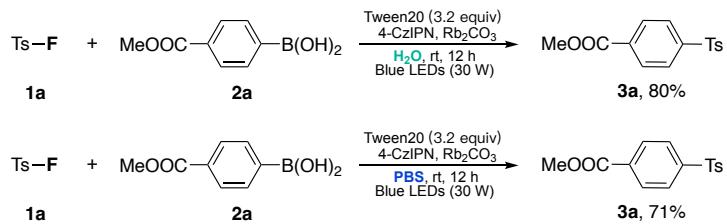
## 4. Derivative Experiments

### 4.1. Control Experiments: Sulfonyl Fluoride vs. Sulfonyl Chloride



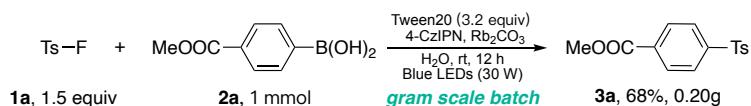
An oven-dried Schlenk tube equipped with a stir bar was charged with sulfonyl fluoride **1a** or **1b** (0.3 mmol, 1.5 equiv), boronic acid **2a** (0.2 mmol, 1.0 equiv), Rb<sub>2</sub>CO<sub>3</sub> (0.4 mmol, 2.0 equiv) and 4-CzIPN (1 mol%). Then the tube was sealed with a cap, evacuated and backfilled with nitrogen for 3 times. Subsequently, Tween20 (0.2 mL, 3.2 equiv) and H<sub>2</sub>O (1 mL) was added. The tube was placed in a water bath with irradiation by 30 W blue LEDs (455 nm, distance: 1 cm, the reaction temperature reached room temperature) for 12 hours. Then the mixture was washed with sat. aq. NH<sub>4</sub>Cl (1 x 2 mL), diluted with EtOAc (1 x 4 mL), and concentrated. The crude product was purified by flash chromatography (EtOAc/PE: 1/4) to give analytically pure product **3a**.

### 4.2. Control Experiments: Water vs. PBS



An oven-dried Schlenk tube equipped with a stir bar was charged with sulfonyl fluoride **1a** (0.3 mmol, 1.5 equiv), boronic acid **2a** (0.2 mmol, 1.0 equiv), Rb<sub>2</sub>CO<sub>3</sub> (0.4 mmol, 2.0 equiv) and 4-CzIPN (1 mol%). Then the tube was sealed with a cap, evacuated and backfilled with nitrogen for 3 times. Subsequently, Tween20 (0.2 mL, 3.2 equiv) and H<sub>2</sub>O or PBS (1 mL) was added. The tube was placed in a water bath with irradiation by 30 W blue LEDs (455 nm, distance: 1 cm, the reaction temperature reached room temperature) for 12 hours. Then the mixture was washed with sat. aq. NH<sub>4</sub>Cl (1 x 2 mL), diluted with EtOAc (1 x 4 mL), and concentrated. The crude product was purified by flash chromatography (EtOAc/PE: 1/4) to give analytically pure product **3a**.

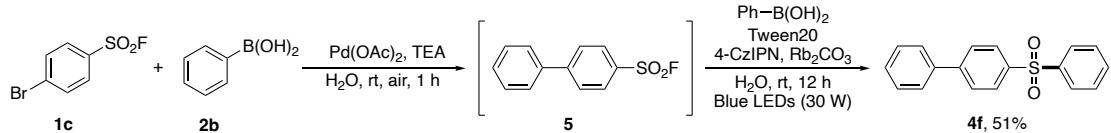
### 4.3. Scale-up Experiments



A scale-up experiment for synthesis of **3a** was carried out by following the **General Procedure A**. The whole process was carried out in an oven-dried 50 mL round bottom flask. Finally, **3a** was purified to afford as colorless solid, 0.20 g, 68% yield.

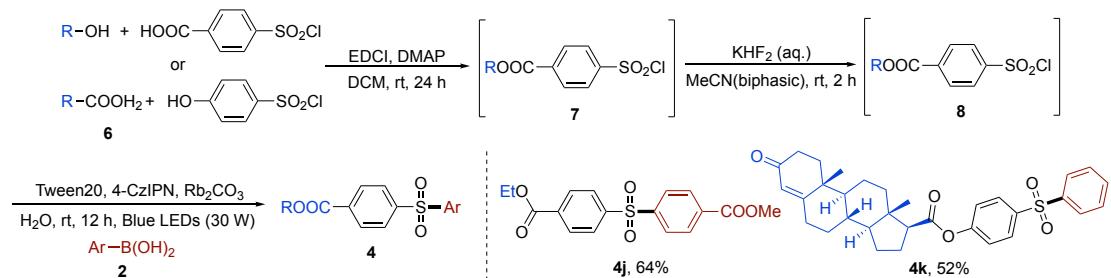
#### 4.4. Tandem One-pot Reactions of Sulfonyl Fluorides

##### 4.4.1. Cross-coupling



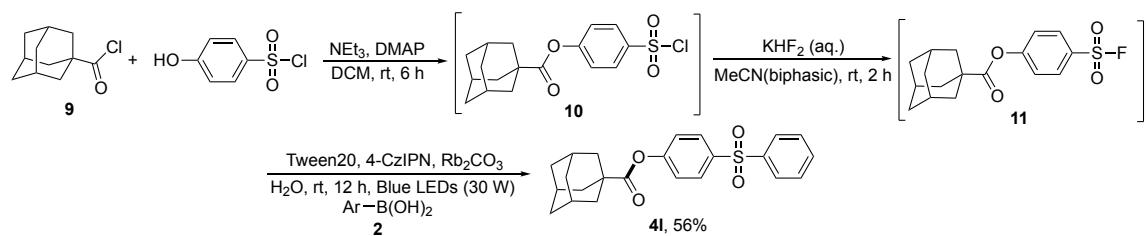
An oven-dried Schlenk tube equipped with a stir bar was charged with 4-bromobenzenesulfon fluoride **1c** (0.3 mmol, 1.5 equiv), phenylboronic acid **2b** (0.45 mmol, 2.25 equiv), TEA (0.9 mmol, 4.5 equiv), palladium (II) acetate (1 mol%) and  $\text{H}_2\text{O}$  (1 mL). The suspension was stirred under air at room temperature for 1 hour. After that, the suspension was washed with aq. HBr (1 mL, 1 M), and extracted with EtOAc (2 x 1 mL). The organic layer was placed in a new oven-dried Schlenk tube, and concentrated. Then phenylboronic acid (0.2 mmol, 1.0 equiv),  $\text{Rb}_2\text{CO}_3$  (0.4 mmol, 2.0 equiv) and 4-CzIPN (1 mol%) was added in  $\text{H}_2\text{O}$  (1 mL). Then the tube was sealed with a cap, evacuated and backfilled with nitrogen for 3 times. Subsequently, Tween20 (0.2 mL, 3.2 equiv) was added. The tube was placed in a water bath with irradiation by 30 W blue LEDs (455 nm, distance: 1 cm, the reaction temperature reached room temperature) for 12 hours. Then the mixture was washed with sat. aq.  $\text{NH}_4\text{Cl}$  (1 x 2 mL) and diluted with EtOAc (1 x 4 mL). The crude product was purified by flash chromatography (EtOAc/PE: 1/4) to give analytically pure product **4f**.

##### 4.4.2. Esterification



Carboxylic acid (1 mmol, 5.0 equiv.), hydroxyl substrate (1 mmol) and EDCI (1-ethyl-3-(3-(dimethylamino)propyl) carbodiimide hydrochloride) (1.1 mmol) was suspended in DCM (10 mL) and stirred at room temperature for 24 hours. The reaction mixture was diluted with DCM and quenched with sat. aq.  $\text{NaHCO}_3$  (10 mL). The aqueous layer was extracted with DCM (3 x 5 mL). The combined organic layer was dried over  $\text{Na}_2\text{SO}_4$ . The filtrate was concentrated in vacuo and the residue was then suspended in sat. aq. KHF<sub>2</sub> (1.1 mmol, 2 mL) and MeCN (2 mL), and stirred at room temperature for 2 hours. The reaction mixture was diluted with EtOAc (1 mL) and water (2 mL). The aqueous layer was extracted with EtOAc (3 x 1 mL). The combine organic layer was dried over  $\text{Na}_2\text{SO}_4$ . The filtrate was concentrated in vacuo. Then follow the **General Procedure B** to give analytically pure product **4j** and **4k** (EtOAc/PE: 1/1).

#### 4.4.3. Acyl Esterification

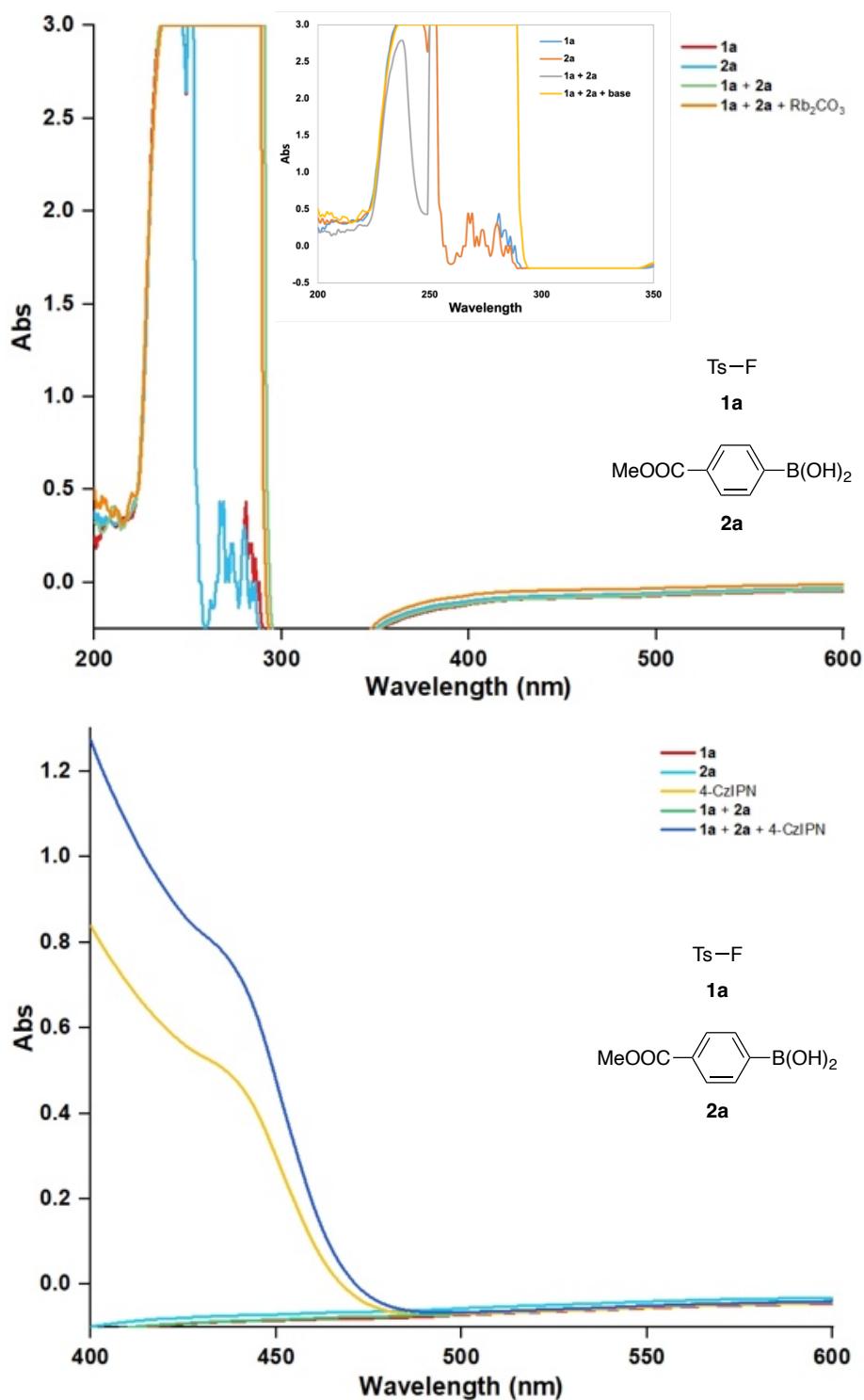


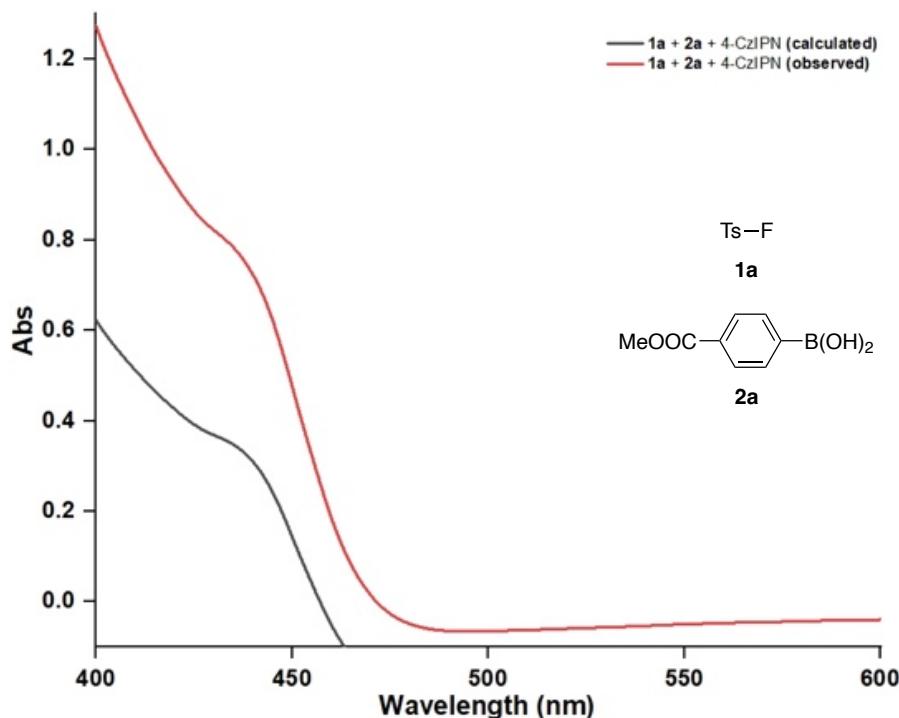
4-hydroxybenzenesulfonyl chloride (1.1 mmol, 5.5 equiv), DMAP (4-dimethylaminopyridine) (0.2 mmol) and  $\text{Et}_3\text{N}$  (2.2 mmol) in  $\text{DCM}$  (10 mL) was suspended and stirred for 10 minutes. Then 1-Adamantanecarbonyl chloride **9** (1 mmol, 5.0 equiv) was added dropwise at 0 °C. The reaction mixture was warmed to room temperature and stirred for 6 hours before quenching with  $\text{H}_2\text{O}$  (5 mL) and extracting with  $\text{DCM}$  (3 x 5 mL). The combined organic layer was dried over  $\text{Na}_2\text{SO}_4$ . The filtrate was concentrated in vacuo and the residue was then suspended in sat. aq.  $\text{KHF}_2$  (1.1 mmol, 2 mL) and  $\text{MeCN}$  (2 mL), and stirred at room temperature for 2 hours. The reaction mixture was diluted with  $\text{EtOAc}$  (1 mL) and water (2 mL). The aqueous layer was extracted with  $\text{EtOAc}$  (3 x 1 mL). The combine organic layer was dried over  $\text{Na}_2\text{SO}_4$ . The filtrate was concentrated in vacuo. Then follow the **General Procedure B** to give analytically pure product **4l**.

## 5. Mechanism Studies

### 5.1. UV-Vis Absorption Spectra Studies

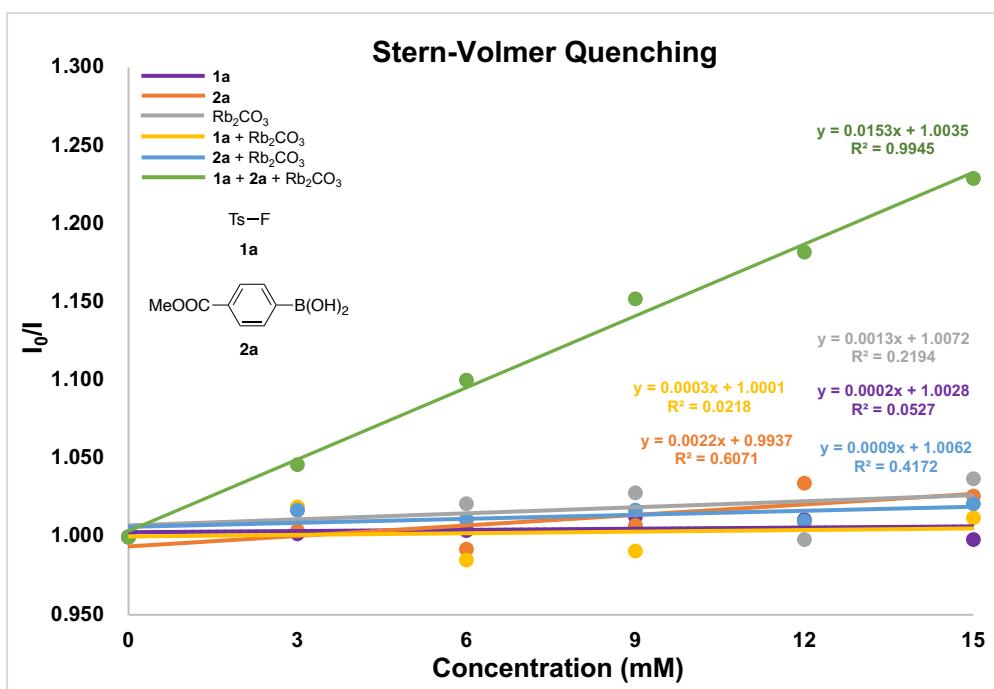
UV-visible spectrum was conducted with **1a** (0.075 M), **2a** (0.05 M), base ( $\text{Rb}_2\text{CO}_3$ , 0.1 M) and 4-CzIPN (1 mol%) in degassed THF.





### 5.2. Stern-Volmer Quenching Experiments

Stern-Volmer experiments were conducted with 0.3 mM  $[\text{Ir}(\text{ppy})_2(\text{dtbbpy})]\text{PF}_6$  and while probing the quenching ability of **1a**, **2a**, **1a + base**, **2a + base** and **1a + 2a + base** (in degassed THF). The solutions were irradiated at 361 nm and luminescence was measured at 524 nm.

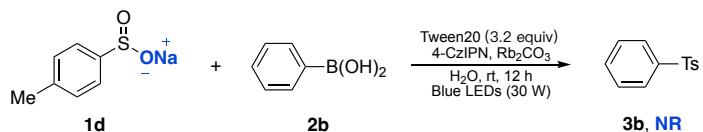


### 5.3. Control Experiments

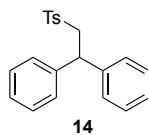
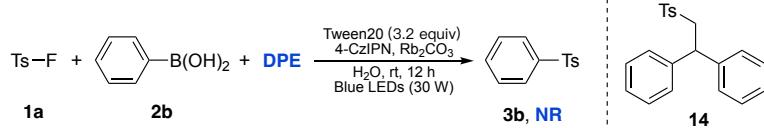
a) Radical Trapping with TEMPO (2,2,6,6-tetramethylpiperidinoxy)



b) Sodium 4-toluenesulfinate (1d) Instead



c) Radical Trapping with DPE (1,1-diphenylethylene)



#### a) Radical Trapping Experiments with TEMPO

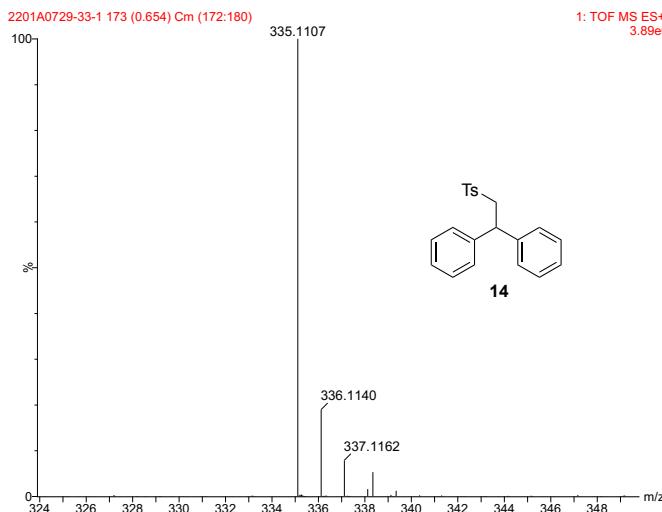
An oven-dried Schlenk tube equipped with a stir bar was charged with sulfonyl fluoride **1a** (0.3 mmol, 1.5 equiv), boronic acid **2b** (0.2 mmol, 1.0 equiv),  $\text{Rb}_2\text{CO}_3$  (0.4 mmol, 2.0 equiv), 4-CzIPN (1 mol%) and TEMPO (2,2,6,6-Tetramethylpiperidinoxy) (0.2 mmol, 1.0 equiv). Then follow the **General Procedure B**. Finally, the desired product was not detected.

#### b) Sodium 4-toluenesulfinate Instead

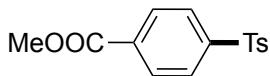
An oven-dried Schlenk tube equipped with a stir bar was charged with sodium 4-toluenesulfinate **1d** (0.3 mmol, 1.5 equiv), boronic acid **2b** (0.2 mmol, 1.0 equiv),  $\text{Rb}_2\text{CO}_3$  (0.4 mmol, 2.0 equiv) and 4-CzIPN (1 mol%). Then follow the **General Procedure B**. Finally, the desired product was not detected.

#### c) Radical Trapping Experiments with DPE

An oven-dried Schlenk tube equipped with a stir bar was charged with sulfonyl fluoride **1a** (0.3 mmol, 1.5 equiv), boronic acid **2b** (0.2 mmol, 1.0 equiv),  $\text{Rb}_2\text{CO}_3$  (0.4 mmol, 2.0 equiv), 4-CzIPN (1 mol%) and DPE (1,1-diphenylethylene) (0.2 mmol, 1.0 equiv). Then follow the **General Procedure B**. Finally, the desired product was not detected yet DPE adduct **14** was detected by HRMS (ESI/[M-H]<sup>+</sup>): Calculated for  $\text{C}_{21}\text{H}_{20}\text{O}_2\text{S}$ : 335.1106, Found: 335.1107.

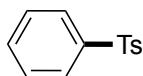


## 6. Characterization of Products



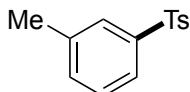
Following the **General Procedure A**, **3a** was isolated as white solid (46.5 mg, 80%).

**<sup>1</sup>H NMR** (400 MHz, CDCl<sub>3</sub>) δ 8.13 (d, *J* = 8.0 Hz, 2H), 7.98 (d, *J* = 8.0 Hz, 2H), 7.83 (d, *J* = 8.0 Hz, 2H), 7.31 (d, *J* = 8.0 Hz, 2H), 3.92 (s, 3H), 2.40 (s, 3H). **<sup>13</sup>C NMR** (101 MHz, CDCl<sub>3</sub>) δ 165.5, 145.9, 144.8, 137.8, 134.0, 130.4, 130.1, 127.9, 127.5, 52.7, 21.6. **HRMS** (ESI/[M+H]<sup>+</sup>) calculated for C<sub>15</sub>H<sub>15</sub>O<sub>4</sub>S: 291.0686, found: 291.0682.



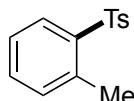
Following the **General Procedure A**, **3b** was isolated as white solid (23.7 mg, 51%).

**<sup>1</sup>H NMR** (400 MHz, CDCl<sub>3</sub>) δ 7.93 (d, *J* = 8.0 Hz, 2H), 7.83 (d, *J* = 8.0 Hz, 2H), 7.56–7.47 (m, 3H), 7.29 (d, *J* = 8.0 Hz, 2H), 2.39 (s, 3H). **<sup>13</sup>C NMR** (101 MHz, CDCl<sub>3</sub>) δ 144.3, 142.1, 138.7, 133.1, 130.0, 129.3, 127.8, 127.6, 21.6. **HRMS** (ESI/[M+H]<sup>+</sup>) calculated for C<sub>14</sub>H<sub>15</sub>O<sub>2</sub>S: 247.0787, found: 247.0788.



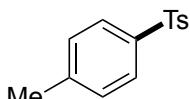
Following the **General Procedure A**, **3c** was isolated as white solid (26.1 mg, 53%).

**<sup>1</sup>H NMR** (400 MHz, CDCl<sub>3</sub>) δ 7.82 (d, *J* = 8.0 Hz, 2H), 7.72 (d, *J* = 8.0 Hz, 2H), 7.36 (d, *J* = 8.0 Hz, 2H), 7.29 (d, *J* = 8.0 Hz, 2H), 2.39 (s, 6H). **<sup>13</sup>C NMR** (101 MHz, CDCl<sub>3</sub>) δ 144.1, 141.8, 139.5, 138.8, 133.8, 129.9, 129.1, 127.8, 127.7, 124.7, 21.6, 21.4. **HRMS** (ESI/[M+H]<sup>+</sup>) calculated for C<sub>14</sub>H<sub>15</sub>O<sub>2</sub>S: 247.0787, found: 247.0788.



Following the **General Procedure A**, **3d** was isolated as white solid (27.1 mg, 55%).

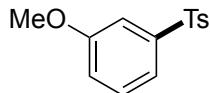
**<sup>1</sup>H NMR** (400 MHz, CDCl<sub>3</sub>) δ 8.04–7.99 (m, 4H), 7.82 (d, *J* = 8.0 Hz, 2H), 7.31 (d, *J* = 8.0 Hz, 2H), 2.60 (s, 3H), 2.39 (s, 3H). **<sup>13</sup>C NMR** (101 MHz, CDCl<sub>3</sub>) δ 144.0, 139.2, 138.3, 137.9, 133.7, 132.6, 129.7, 129.3, 127.8, 126.4, 21.6, 20.2. **HRMS** (ESI/[M+H]<sup>+</sup>) calculated for C<sub>14</sub>H<sub>15</sub>O<sub>2</sub>S: 247.0787, found: 247.0787.



Following the **General Procedure A**, **3e** was isolated as white solid (32.5 mg, 66%).

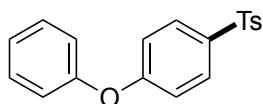
**<sup>1</sup>H NMR** (400 MHz, CDCl<sub>3</sub>) δ 7.83 (d, *J* = 8.0 Hz, 4H), 7.30 (d, *J* = 8.0 Hz, 4H), 2.41 (s, 6H). **<sup>13</sup>C**

**NMR** (101 MHz, CDCl<sub>3</sub>) δ 144.0, 139.1, 129.9, 127.6, 21.6. **HRMS** (ESI/[M+H]<sup>+</sup>) calculated for C<sub>14</sub>H<sub>15</sub>O<sub>2</sub>S: 247.0787, found: 247.0785.



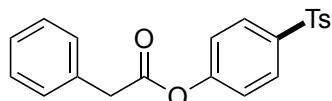
Following the **General Procedure A**, **3f** was isolated as white solid (31.5 mg, 60%).

**<sup>1</sup>H NMR** (400 MHz, CDCl<sub>3</sub>) δ 7.82 (d, *J* = 8.0 Hz, 2H), 7.50–7.36 (m, 3H), 7.29 (d, *J* = 8.0 Hz, 2H), 7.05 (d, *J* = 8.0 Hz, 1H), 3.83 (s, 3H), 2.39 (s, 3H). **<sup>13</sup>C NMR** (101 MHz, CDCl<sub>3</sub>) δ 160.1, 144.3, 143.2, 138.6, 130.4, 130.0, 127.8, 119.8, 119.4, 112.2, 55.8, 21.7. **HRMS** (ESI/[M+H]<sup>+</sup>) calculated for C<sub>14</sub>H<sub>15</sub>O<sub>3</sub>S: 263.0736, found: 263.0734.



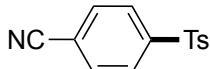
Following the **General Procedure A**, **3g** was isolated as light yellow solid (44.8 mg, 69%).

**<sup>1</sup>H NMR** (400 MHz, CDCl<sub>3</sub>) δ 7.86 (d, *J* = 8.0 Hz, 2H), 7.81 (d, *J* = 8.0 Hz, 2H), 7.41–7.37 (m, 2H), 7.29 (d, *J* = 8.0 Hz, 2H), 7.23–7.19 (m, 1H), 7.04–6.99 (m, 4H), 2.40 (s, 3H). **<sup>13</sup>C NMR** (101 MHz, CDCl<sub>3</sub>) δ 162.0, 155.0, 144.0, 139.1, 135.4, 130.2, 129.9, 129.8, 127.5, 125.0, 120.4, 117.7, 21.6. **HRMS** data are in accordance with the literature report.<sup>[10]</sup>



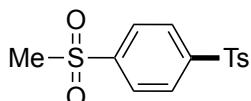
Following the **General Procedure A**, **3h** was isolated as white solid (52.0 mg, 71%).

**<sup>1</sup>H NMR** (400 MHz, CDCl<sub>3</sub>) δ 8.16 (d, *J* = 8.0 Hz, 2H), 7.98 (d, *J* = 8.0 Hz, 2H), 7.82 (d, *J* = 8.0 Hz, 2H), 7.42–7.34 (m, 5H), 7.30 (d, *J* = 8.0 Hz, 2H), 5.36 (s, 2H), 2.40 (s, 3H). **<sup>13</sup>C NMR** (101 MHz, CDCl<sub>3</sub>) δ 165.0, 146.1, 144.8, 137.9, 135.5, 134.2, 130.6, 130.2, 128.8, 128.6, 128.4, 128.0, 127.6, 67.5, 21.7. **HRMS** (ESI/[M+H]<sup>+</sup>) calculated for C<sub>21</sub>H<sub>18</sub>O<sub>4</sub>S: 367.0999, found: 367.1027.



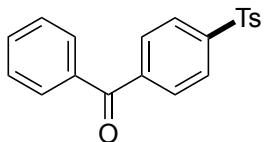
Following the **General Procedure A**, **3i** was isolated (EtOAc/PE: 1/2 to 1/1) as white solid (40.7 mg, 79%).

**<sup>1</sup>H NMR** (400 MHz, CDCl<sub>3</sub>) δ 8.01 (d, *J* = 8.0 Hz, 2H), 7.83–7.76 (m, 4H), 7.33 (d, *J* = 8.0 Hz, 2H), 2.41 (s, 3H). **<sup>13</sup>C NMR** (101 MHz, CDCl<sub>3</sub>) δ 146.3, 145.3, 137.1, 133.1, 130.3, 128.2, 128.1, 117.3, 116.7, 21.7; **HRMS** (ESI/[M+H]<sup>+</sup>) calculated for C<sub>14</sub>H<sub>12</sub>O<sub>2</sub>S: 258.0583, found: 258.0582.



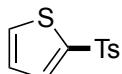
Following the **General Procedure A**, **3j** was isolated as white solid (45.9 mg, 74%).

**<sup>1</sup>H NMR** (400 MHz, CDCl<sub>3</sub>) δ 8.12–8.04 (m, 4H), 7.83 (d, *J* = 8.0 Hz, 2H), 7.33 (d, *J* = 8.0 Hz, 2H), 3.05 (s, 3H), 2.41 (s, 3H). **<sup>13</sup>C NMR** (101 MHz, CDCl<sub>3</sub>) δ 147.3, 145.4, 144.6, 137.2, 130.4, 128.64, 128.56, 128.2, 44.3, 21.7. **HRMS** (ESI/[M+H]<sup>+</sup>) calculated for C<sub>14</sub>H<sub>14</sub>O<sub>4</sub>S<sub>2</sub>: 311.0406, found: 311.0419.



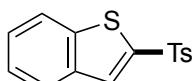
Following the **General Procedure A**, **3k** was isolated as white solid (37.7 mg, 56%).

**<sup>1</sup>H NMR** (400 MHz, CDCl<sub>3</sub>) δ 8.03 (d, *J* = 12.0 Hz, 2H), 7.86 (d, *J* = 8.0 Hz, 4H), 7.76 (d, *J* = 8.0 Hz, 2H), 7.64–7.60 (m, 1H), 7.51–7.47 (m, 2H), 7.33 (d, *J* = 8.0 Hz, 2H), 2.42 (s, 3H). **<sup>13</sup>C NMR** (101 MHz, CDCl<sub>3</sub>) δ 195.3, 145.1, 144.8, 141.5, 137.9, 136.4, 133.3, 130.5, 130.1, 128.6, 128.0, 127.5, 21.7. **HRMS** data are in accordance with the literature report.<sup>[7]</sup>



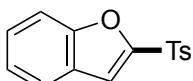
Following the **General Procedure A**, **3l** was isolated as white solid (37.2 mg, 78%).

**<sup>1</sup>H NMR** (400 MHz, CDCl<sub>3</sub>) δ 7.87 (d, *J* = 8.0 Hz, 2H), 7.66 (d, *J* = 4.0 Hz, 1H), 7.62 (d, *J* = 4.0 Hz, 1H), 7.31 (d, *J* = 8.0 Hz, 2H), 7.07–7.05 (m, 1H), 2.40 (s, 3H). **<sup>13</sup>C NMR** (101 MHz, CDCl<sub>3</sub>) δ 144.4, 143.5, 139.1, 133.6, 133.1, 130.0, 127.8, 127.4, 21.6. **HRMS** data are in accordance with the literature report.<sup>[5]</sup>



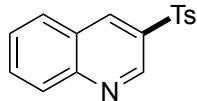
Following the **General Procedure A**, **3m** was isolated as white solid (46.1 mg, 80%).

**<sup>1</sup>H NMR** (400 MHz, CDCl<sub>3</sub>) δ 7.95–7.91 (m, 3H), 7.86 (d, *J* = 8.0 Hz, 1H), 7.81 (d, *J* = 8.0 Hz, 1H), 7.45–7.40 (m, 2H), 7.32 (d, *J* = 8.0 Hz, 2H), 2.40 (s, 3H). **<sup>13</sup>C NMR** (101 MHz, CDCl<sub>3</sub>) δ 144.7, 143.4, 142.7, 138.4, 137.8, 130.0, 127.7, 127.4, 125.9, 125.5, 122.8, 21.7. **HRMS** data are in accordance with the literature report.<sup>[5]</sup>



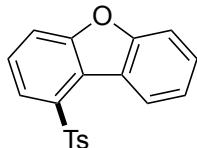
Following the **General Procedure A**, **3n** was isolated as white solid (28.9 mg, 53%).

**<sup>1</sup>H NMR** (400 MHz, CDCl<sub>3</sub>) δ 7.96 (d, *J* = 8.0 Hz, 2H), 7.67 (d, *J* = 8.0 Hz, 1H), 7.54–7.49 (m, 2H), 7.45–7.41 (m, 1H), 7.36–7.29 (m, 3H), 2.42 (s, 3H). **<sup>13</sup>C NMR** (101 MHz, CDCl<sub>3</sub>) δ 156.3, 151.9, 145.3, 136.4, 130.1, 128.3, 127.9, 126.0, 124.3, 123.1, 112.9, 112.5, 21.7. **HRMS** (ESI/[M+H]<sup>+</sup>) calculated for C<sub>15</sub>H<sub>13</sub>O<sub>3</sub>S: 273.0580, found: 273.0577.



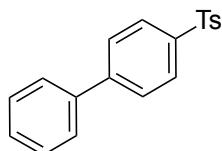
Following the **General Procedure A**, **3o** was isolated as yellow solid (23.2 mg, 41%).

**<sup>1</sup>H NMR** (400 MHz, CDCl<sub>3</sub>) δ 9.28 (s, 1H), 8.83 (s, 1H), 8.18 (d, *J* = 8.0 Hz, 1H), 7.99–7.87 (m, 4H), 7.72–7.68 (m, 1H), 7.35 (d, *J* = 8.0, 2H), 2.42 (s, 3H). **<sup>13</sup>C NMR** (101 MHz, CDCl<sub>3</sub>) δ 149.3, 147.1, 144.9, 138.0, 136.7, 135.1, 132.7, 130.2, 129.6, 129.2, 128.3, 127.9, 126.4, 21.6. **HRMS** (ESI/[M+H]<sup>+</sup>) calculated for C<sub>16</sub>H<sub>14</sub>NO<sub>2</sub>S: 284.0745, found: 284.0747.



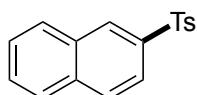
Following the **General Procedure A**, **3p** was isolated as white solid (32.9 mg, 51%).

**<sup>1</sup>H NMR** (400 MHz, CDCl<sub>3</sub>) δ 8.81 (d, *J* = 8.0 Hz, 1H), 8.08 (d, *J* = 8.0 Hz, 1H), 7.89–7.83 (m, 3H), 7.64–7.54 (m, 3H), 7.45–7.41 (m, 1H), 7.27 (d, *J* = 8.0 Hz, 2H), 2.37 (s, 3H). **<sup>13</sup>C NMR** (101 MHz, CDCl<sub>3</sub>) δ 156.8, 144.3, 138.1, 134.6, 129.8, 128.9, 127.3, 126.6, 125.9, 124.5, 123.5, 121.6, 121.1, 117.0, 111.6, 21.6. **HRMS** (ESI/[M+H]<sup>+</sup>) calculated for C<sub>19</sub>H<sub>15</sub>O<sub>3</sub>S: 323.0736, found: 323.0731.



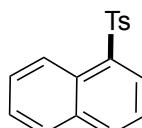
Following the **General Procedure A**, **3q** was isolated as white solid (55.5 mg, 90%).

**<sup>1</sup>H NMR** (400 MHz, CDCl<sub>3</sub>) δ 7.99 (d, *J* = 8.0 Hz, 2H), 7.87 (d, *J* = 8.0 Hz, 2H), 7.69 (d, *J* = 8.0 Hz, 2H), 7.56 (d, *J* = 8.0 Hz, 2H), 7.47–7.40 (m, 3H), 7.31 (d, *J* = 8.0 Hz, 2H), 2.40 (s, 3H). **<sup>13</sup>C NMR** (101 MHz, CDCl<sub>3</sub>) δ 146.0, 144.2, 140.5, 139.2, 138.8, 130.0, 129.07, 128.6, 128.1, 127.9, 127.7, 127.4, 21.6. **HRMS** data are in accordance with the literature report.<sup>[7]</sup>



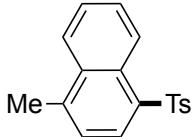
Following the **General Procedure A**, **3r** was isolated as white solid (33.9 mg, 60%).

**<sup>1</sup>H NMR** (400 MHz, CDCl<sub>3</sub>) δ 8.56 (s, 1H), 7.97 (d, *J* = 8.0 Hz, 1H), 7.92–7.83 (m, 5H), 7.64–7.57 (m, 2H), 7.29 (d, *J* = 8.0 Hz, 2H), 2.38 (s, 3H). **<sup>13</sup>C NMR** (101 MHz, CDCl<sub>3</sub>) δ 144.2, 138.8, 138.7, 134.9, 132.2, 130.0, 129.6, 129.4, 129.1, 128.8, 127.9, 127.8, 127.6, 122.6, 21.6. **HRMS** data are in accordance with the literature report.<sup>[6]</sup>



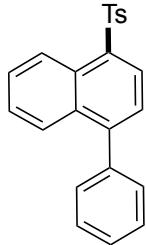
Following the **General Procedure A**, **3s** was isolated as white solid (32.2 mg, 57%).

**<sup>1</sup>H NMR** (400 MHz, CDCl<sub>3</sub>) δ 8.66 (d, *J* = 8.0 Hz, 1H), 8.51 (d, *J* = 8.0 Hz, 1H), 8.09 (d, *J* = 8.0 Hz, 1H), 7.91–7.86 (m, 3H), 7.64–7.54 (m, 4H), 7.27 (d, *J* = 8.0 Hz, 2H), 2.37 (s, 3H). **<sup>13</sup>C NMR** (126 MHz, CDCl<sub>3</sub>) δ 144.0, 138.8, 136.2, 135.0, 134.2, 129.8, 129.0, 128.4, 128.3, 127.5, 126.9, 124.4, 124.4, 21.6. **HRMS** data are in accordance with the literature report.<sup>[6]</sup>



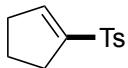
Following the **General Procedure A**, **3t** was isolated as white solid (43.4 mg, 73%).

**<sup>1</sup>H NMR** (400 MHz, CDCl<sub>3</sub>) δ 8.69–8.66 (m, 1H), 8.42 (d, *J* = 8.0 Hz, 1H), 8.07–8.04 (m, 1H), 7.85 (d, *J* = 8.0 Hz, 2H), 7.58–7.56 (m, 2H), 7.47 (d, *J* = 8.0 Hz, 1H), 7.25 (d, *J* = 8.0 Hz, 2H), 2.76 (s, 3H), 2.35 (s, 3H). **<sup>13</sup>C NMR** (126 MHz, CDCl<sub>3</sub>) δ 143.8, 142.4, 139.1, 134.2, 133.3, 129.7, 128.5, 127.8, 127.4, 126.7, 125.3, 125.04, 125.0, 21.5, 20.2. **HRMS** data are in accordance with the literature report.<sup>[7]</sup>



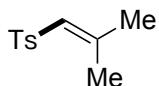
Following the **General Procedure A**, **3u** was isolated as white solid (47.3 mg, 66%).

**<sup>1</sup>H NMR** (400 MHz, CDCl<sub>3</sub>) δ 8.74 (d, *J* = 8.0 Hz, 1H), 8.55 (d, *J* = 8.0 Hz, 1H), 7.91 (d, *J* = 8.0 Hz, 3H), 7.58–7.43 (m, 8H), 7.29 (d, *J* = 8.0 Hz, 2H), 2.37 (s, 3H). **<sup>13</sup>C NMR** (101 MHz, CDCl<sub>3</sub>) δ 147.3, 144.0, 139.4, 138.9, 135.3, 132.6, 129.8, 129.3, 128.9, 128.5, 128.2, 128.0, 127.6, 127.4, 126.8, 125.5, 124.6, 21.6. **HRMS** data are in accordance with the literature report.<sup>[7]</sup>



Following the **General Procedure A**, **3v** was isolated as white solid (17.78 mg, 40%).

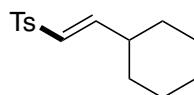
**<sup>1</sup>H NMR** (400 MHz, CDCl<sub>3</sub>) δ 7.78 (d, *J* = 8.0 Hz, 2H), 7.33 (d, *J* = 8.0 Hz, 2H), 6.71(s, 1H), 2.53 (t, *J* = 8.0 Hz, 4H), 2.44 (s, 3H), 2.05–1.98 (m, 2H). **<sup>13</sup>C NMR** (101 MHz, CDCl<sub>3</sub>) δ 144.9, 144.3, 142.8, 136.7, 129.8, 128.0, 32.9, 30.8, 23.7, 21.6. **HRMS** data are in accordance with the literature report.<sup>[4]</sup>



Following the **General Procedure A**, **3w** was isolated as light yellow solid (14.9 mg, 38%).

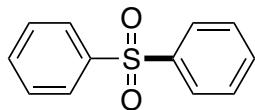
**<sup>1</sup>H NMR** (400 MHz, CDCl<sub>3</sub>) δ 7.78 (d, *J* = 8.0 Hz, 2H), 7.32 (d, *J* = 8.0 Hz, 2H), 6.17(s, 1H), 2.43 (s, 3H), 2.14 (s, 3H), 1.87 (s, 3H). **<sup>13</sup>C NMR** (101 MHz, CDCl<sub>3</sub>) δ 153.7, 143.9, 139.5, 129.8, 127.2,

126.6, 27.1, 21.6, 19.2. **HRMS** data are in accordance with the literature report.<sup>[8]</sup>



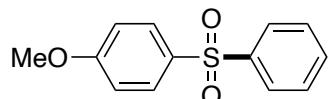
Following the **General Procedure A**, **3x** was isolated as white solid (30.1 mg, 57%).

**<sup>1</sup>H NMR** (400 MHz, CDCl<sub>3</sub>) δ 7.74 (d, *J* = 8.0 Hz, 2H), 7.31 (d, *J* = 8.0 Hz, 2H), 6.93-6.88 (m 1H), 6.22 (d, *J* = 16.0 Hz, 1H), 2.42 (s, 3H), 1.76-1.63 (m 6H), 1.25-1.10 (m, 5H). **<sup>13</sup>C NMR** (101 MHz, CDCl<sub>3</sub>) δ 151.2, 144.1, 137.9, 129.9, 128.6, 127.6, 39.9, 31.3, 25.7, 25.6, 21.6. **HRMS** data are in accordance with the literature report.<sup>[9]</sup>



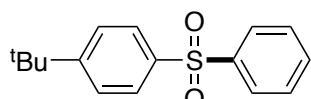
Following the **General Procedure B**, **4a** was isolated as white solid (26.2 mg, 60%).

**<sup>1</sup>H NMR** (400 MHz, CDCl<sub>3</sub>) δ 7.96–7.94 (m, 4H), 7.58–7.49 (m, 6H). **<sup>13</sup>C NMR** (101 MHz, CDCl<sub>3</sub>) δ 141.6, 133.2, 129.3, 127.7. **HRMS** (ESI/[M+H]<sup>+</sup>) calculated for C<sub>12</sub>H<sub>10</sub>O<sub>2</sub>S: 219.0474, found: 219.0457.



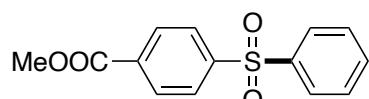
Following the **General Procedure B**, **4b** was isolated as white solid (27.3 mg, 55%).

**<sup>1</sup>H NMR** (400 MHz, CDCl<sub>3</sub>) δ 7.92–7.86 (m, 4H), 7.54–7.46 (m, 3H), 6.97–6.94 (m, 2H), 3.83 (s, 3H). **<sup>13</sup>C NMR** (101 MHz, CDCl<sub>3</sub>) δ 163.4, 142.4, 133.1, 132.8, 129.9, 129.2, 127.3, 114.5, 55.7. **HRMS** (ESI/[M+H]<sup>+</sup>) calculated for C<sub>13</sub>H<sub>12</sub>O<sub>3</sub>S: 249.0580, found: 249.0590.



Following the **General Procedure B**, **4c** was isolated as white solid (28.1 mg, 51%).

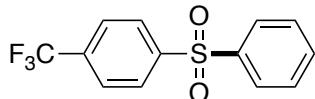
**<sup>1</sup>H NMR** (400 MHz, CDCl<sub>3</sub>) δ 7.95 (d, *J* = 8.0 Hz, 2H), 7.86 (d, *J* = 8.0 Hz, 2H), 7.57–7.53 (m, 1H), 7.51–7.47 (m, 4H), 1.30 (s, 9H). **<sup>13</sup>C NMR** (101 MHz, CDCl<sub>3</sub>) δ 157.1, 141.9, 138.5, 133.0, 129.2, 127.6, 127.5, 126.3, 35.2, 31.0. **HRMS** (ESI/[M+H]<sup>+</sup>) calculated for C<sub>16</sub>H<sub>18</sub>O<sub>2</sub>S: 275.1100, found: 275.1076.



Following the **General Procedure B**, **4d** was isolated as white solid (35.4 mg, 64%).

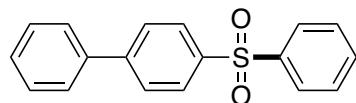
**<sup>1</sup>H NMR** (400 MHz, CDCl<sub>3</sub>) δ 8.17 (d, *J* = 8.0 Hz, 2H), 8.03 (d, *J* = 8.0 Hz, 2H), 7.97 (d, *J* = 8.0 Hz, 2H), 7.63–7.60 (m, 1H), 7.56–7.52 (m, 2H), 3.95 (s, 3H). **<sup>13</sup>C NMR** (101 MHz, CDCl<sub>3</sub>) δ 165.5,

145.5, 140.8, 134.3, 133.7, 130.5, 129.5, 127.9, 127.7, 52.7. **HRMS** (ESI/[M+H]<sup>+</sup>) calculated for C<sub>14</sub>H<sub>13</sub>O<sub>4</sub>S: 277.0529, found: 277.0525.



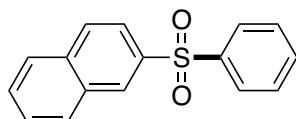
Following the **General Procedure B**, **4e** was isolated as white solid (40.8 mg, 71%).

**<sup>1</sup>H NMR** (400 MHz, CDCl<sub>3</sub>) δ 8.07 (d, *J* = 8.0 Hz, 2H), 7.96 (d, *J* = 8.0 Hz, 2H), 7.76 (d, *J* = 8.0 Hz, 2H), 7.63–7.52 (m, 3H). **<sup>13</sup>C NMR** (101 MHz, CDCl<sub>3</sub>) δ 145.2, 140.6, 134.9 (q, *J* = 32.8 Hz), 133.8, 129.6, 128.2, 127.9, 126.5, 122.0 (q, *J* = 274.7 Hz). **<sup>19</sup>F NMR** (377 MHz, CDCl<sub>3</sub>) δ 63.20. **HRMS** (ESI/[M+Na]<sup>+</sup>) calculated for C<sub>13</sub>H<sub>9</sub>O<sub>2</sub>F<sub>3</sub>S: 309.0168, found: 309.0153.



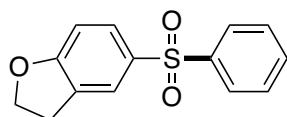
Following the **General Procedure B**, **4f** was isolated as white solid (40.0 mg, 68%).

**<sup>1</sup>H NMR** (400 MHz, CDCl<sub>3</sub>) δ 8.05–8.01 (m, 4H), 7.72 (d, *J* = 12.0 Hz, 2H), 7.59–7.53 (m, 5H), 7.50–7.41 (m, 3H). **<sup>13</sup>C NMR** (101 MHz, CDCl<sub>3</sub>) δ 146.2, 141.7, 140.1, 139.2, 133.2, 129.4, 129.1, 128.6, 128.2, 128.0, 127.7, 127.4. **HRMS** data are in accordance with the literature report.<sup>[10]</sup>



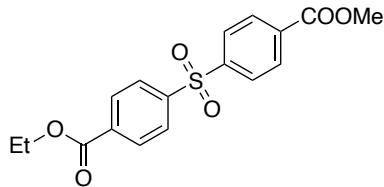
Following the **General Procedure B**, **4g** was isolated as white solid (38.6 mg, 72%).

**<sup>1</sup>H NMR** (400 MHz, CDCl<sub>3</sub>) δ 8.59 (s, 1H), 8.02–7.85 (m, 6H), 7.65–7.58 (m, 2H), 7.55–7.48 (m, 3H). **<sup>13</sup>C NMR** (101 MHz, CDCl<sub>3</sub>) δ 141.6, 138.4, 135.0, 133.2, 132.2, 129.7, 129.4, 129.3, 129.2, 129.1, 127.9, 127.7, 127.7, 122.7. **HRMS** data are in accordance with the literature report.<sup>[11]</sup>



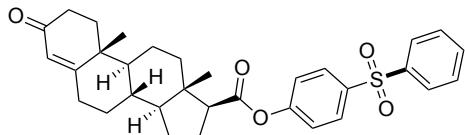
Following the **General Procedure B**, **4h** was isolated (EtOAc/PE: 1/2 to 1/1) as white solid (38.5 mg, 74%).

**<sup>1</sup>H NMR** (400 MHz, CDCl<sub>3</sub>) δ 7.93 (d, *J* = 8.0 Hz, 2H), 7.76 (d, *J* = 4.0 Hz, 2H), 7.57–7.48 (m, 3H), 6.84 (d, *J* = 8.0 Hz, 1H), 4.66 (t, *J* = 8.0 Hz, 2H), 3.25 (t, *J* = 8.0 Hz, 2H). **<sup>13</sup>C NMR** (101 MHz, CDCl<sub>3</sub>) δ 164.4, 142.5, 133.0, 132.8, 129.5, 129.2, 128.7, 127.2, 125.0, 109.8, 72.4, 29.0. **HRMS** data are in accordance with the literature report.<sup>[11]</sup>



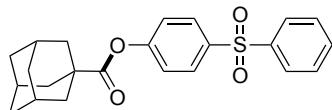
According to tandem one-pot reaction, **4j** was isolated as white solid (44.6 mg, 64%).

**<sup>1</sup>H NMR** (400 MHz, CDCl<sub>3</sub>) δ 8.19–8.16 (m, 4H), 8.03 (d, *J* = 8.0 Hz, 4H), 4.40 (q, *J* = 8.0 Hz, 2H), 3.95 (s, 3H), 1.40 (t, *J* = 8.0 Hz, 3H). **<sup>13</sup>C NMR** (101 MHz, CDCl<sub>3</sub>) δ 165.3, 164.8, 144.8, 144.6, 135.1, 134.7, 130.6, 130.5, 127.9, 61.8, 52.7, 14.2. **HRMS** (ESI/[M+H]<sup>+</sup>) calculated for C<sub>17</sub>H<sub>16</sub>O<sub>6</sub>S: 349.0740, found: 349.0734.



According to tandem one-pot reaction, **4k** was isolated (EtOAc/PE: 1/1 to 2/1) as colorless liquid (55.3 mg, 52%).

**<sup>1</sup>H NMR** (400 MHz, CDCl<sub>3</sub>) δ 7.96–7.91 (m, 4H), 7.56–7.47 (m, 3H), 7.22 (d, *J* = 8.0 Hz, 2H), 5.73 (s, 1H), 3.86 (s, 1H), 2.63–2.58 (m, 1H), 2.42–2.36 (m, 2H), 2.30–2.26 (m, 1H), 2.21–2.12 (m, 2H), 2.04–2.01 (m, 1H), 1.95–1.85 (m, 2H), 1.79–1.71 (m, 2H), 1.64–1.54 (m, 2H), 1.47–1.39 (m, 2H), 1.27–1.23 (m, 2H), 1.18 (s, 3H), 1.08–0.96 (m, 2H), 0.82 (s, 3H). **<sup>13</sup>C NMR** (101 MHz, CDCl<sub>3</sub>) δ 199.7, 171.8, 171.1, 154.5, 141.4, 138.7, 133.9, 133.3, 129.4, 127.6, 124.0, 122.6, 55.4, 55.1, 53.6, 44.6, 38.6, 38.2, 35.7, 33.9, 32.8, 31.9, 29.7, 24.4, 23.6, 20.9, 17.4, 13.7. **HRMS** data are in accordance with the literature report.<sup>[6]</sup>



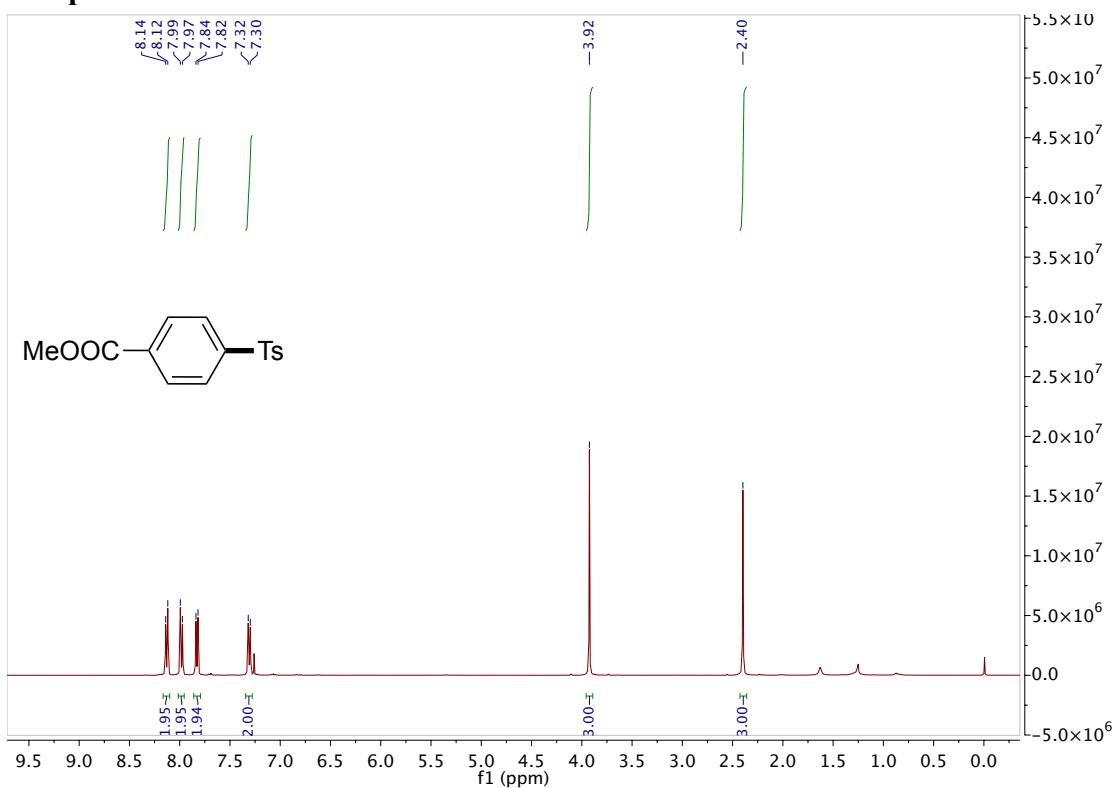
According to tandem one-pot reactions, **4l** was isolated as white solid (44.4 mg, 56%).

**<sup>1</sup>H NMR** (400 MHz, CDCl<sub>3</sub>) δ 7.96–7.92 (m, 4H), 7.58–7.48 (m, 3H), 7.19 (d, *J* = 8.0 Hz, 2H), 2.08–2.02 (m, 9H), 1.79–1.72 (m, 6H). **<sup>13</sup>C NMR** (101 MHz, CDCl<sub>3</sub>) δ 175.4, 154.9, 141.5, 138.5, 133.3, 129.4, 127.6, 122.6, 41.2, 38.6, 36.3, 27.8. **HRMS** data are in accordance with the literature report.<sup>[6]</sup>

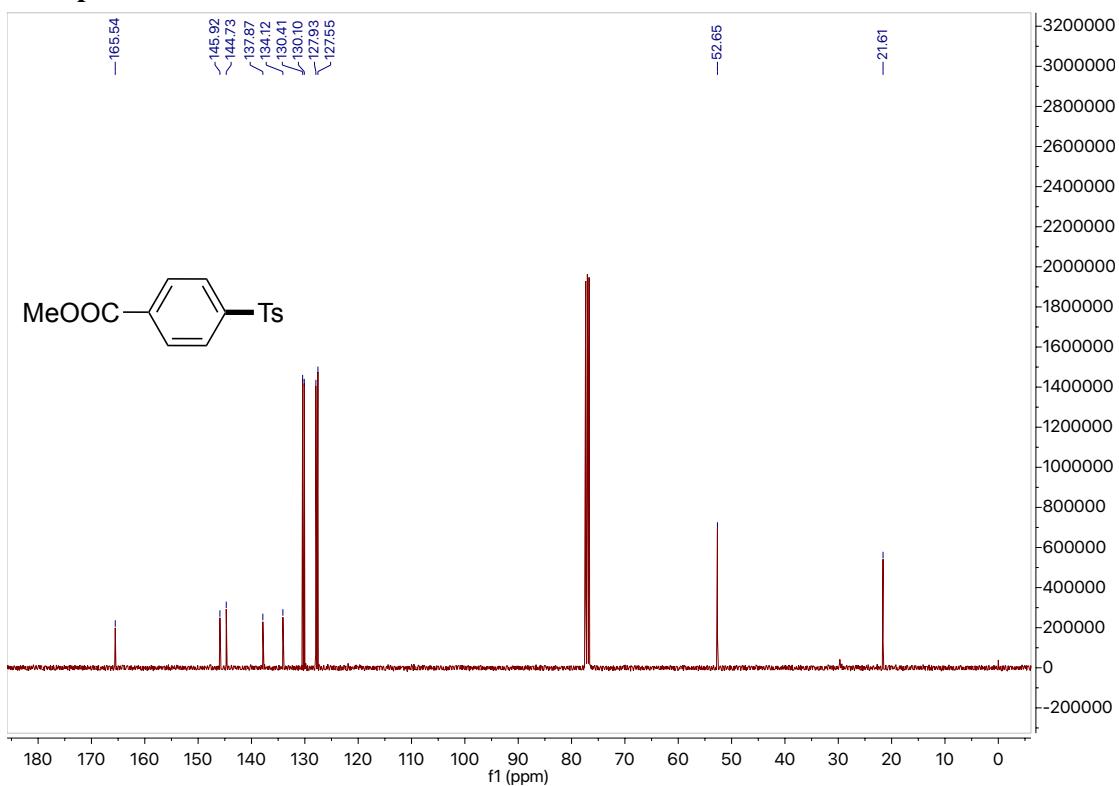
## 7. Spectra for Described Compounds

(3a)

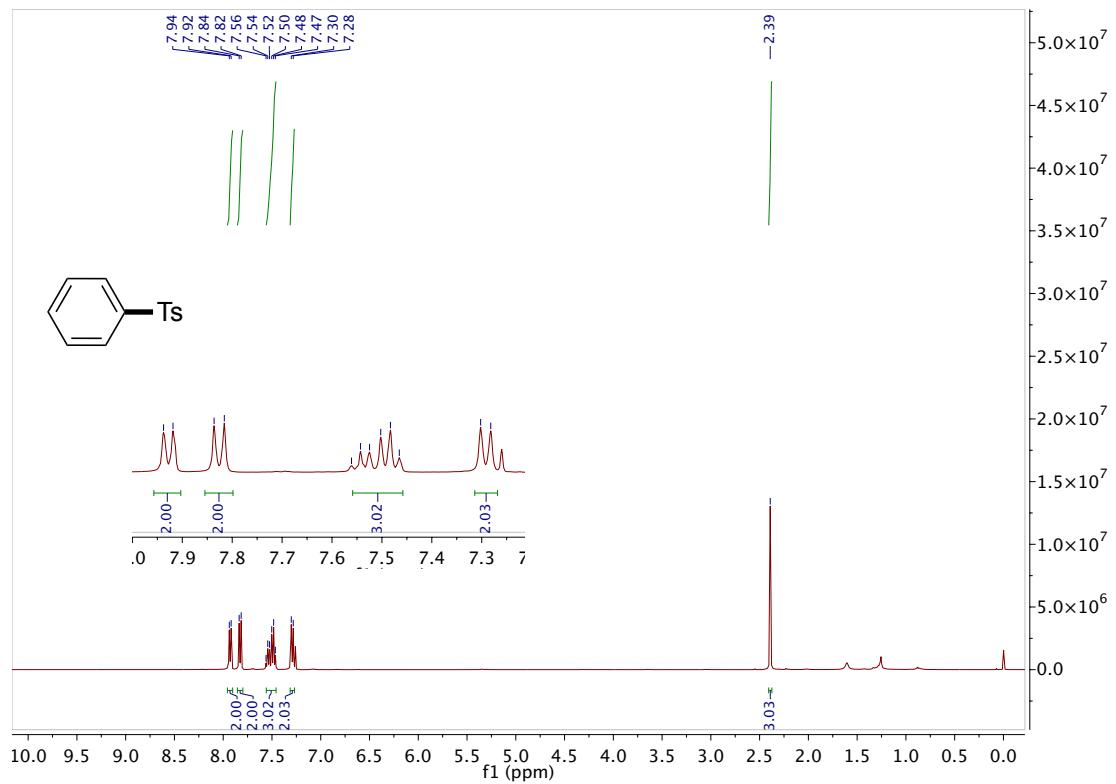
<sup>1</sup>H Spectra



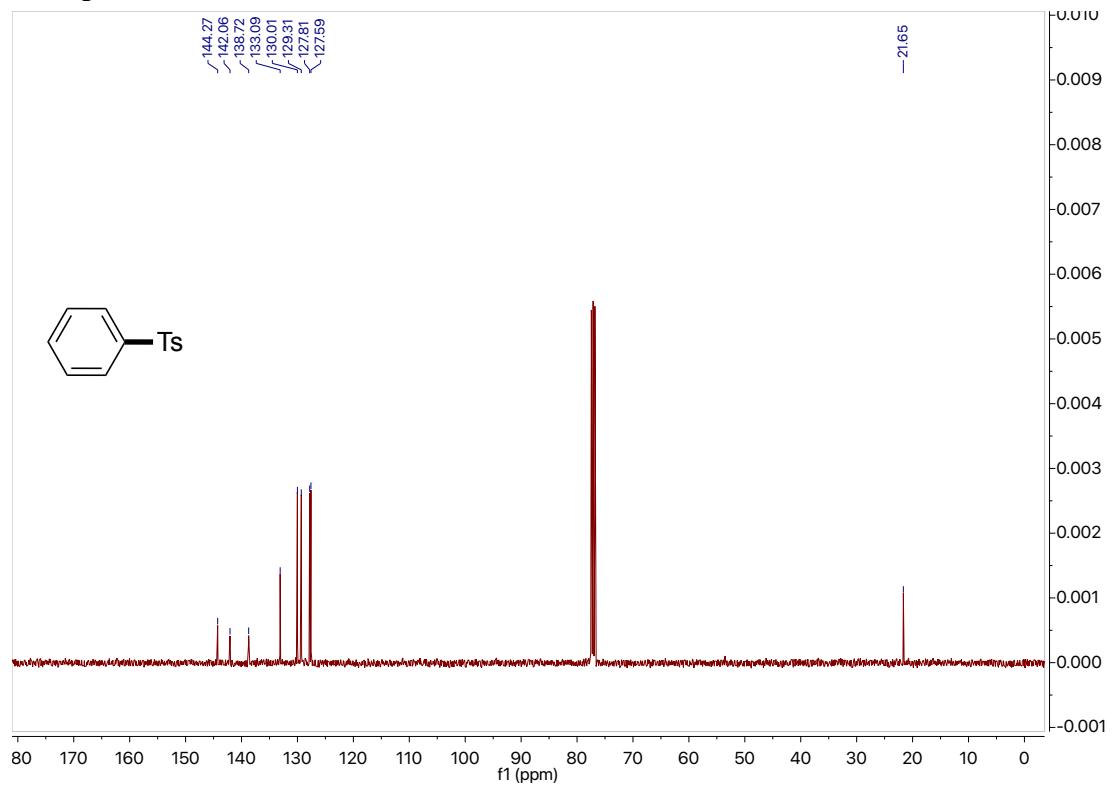
<sup>13</sup>C Spectra



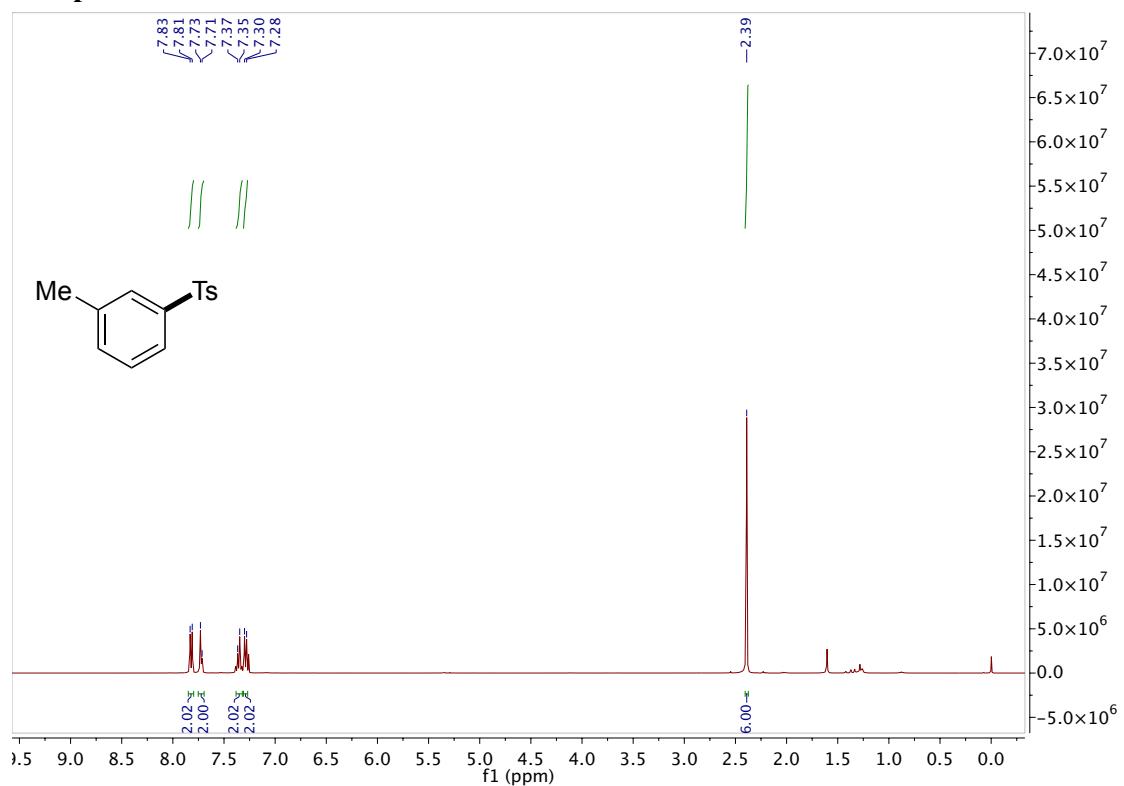
**(3b)**  
 **$^1\text{H}$  Spectra**



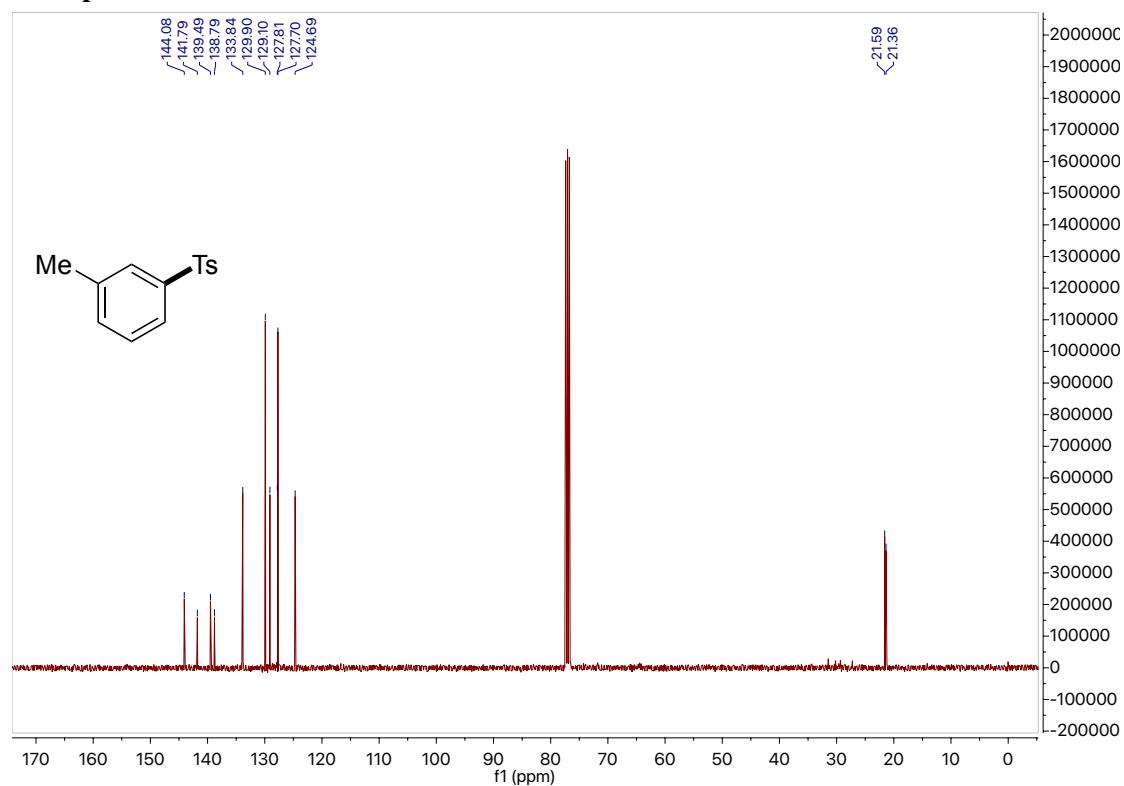
**$^{13}\text{C}$  Spectra**



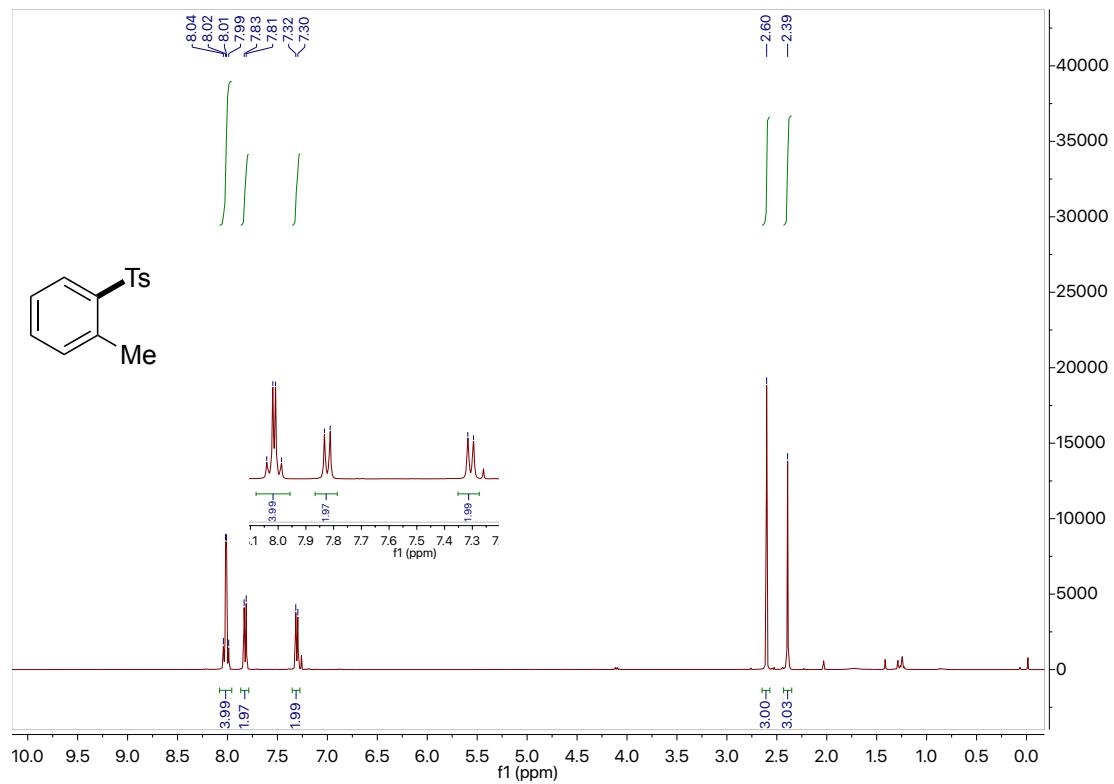
(3c)  
<sup>1</sup>H Spectra



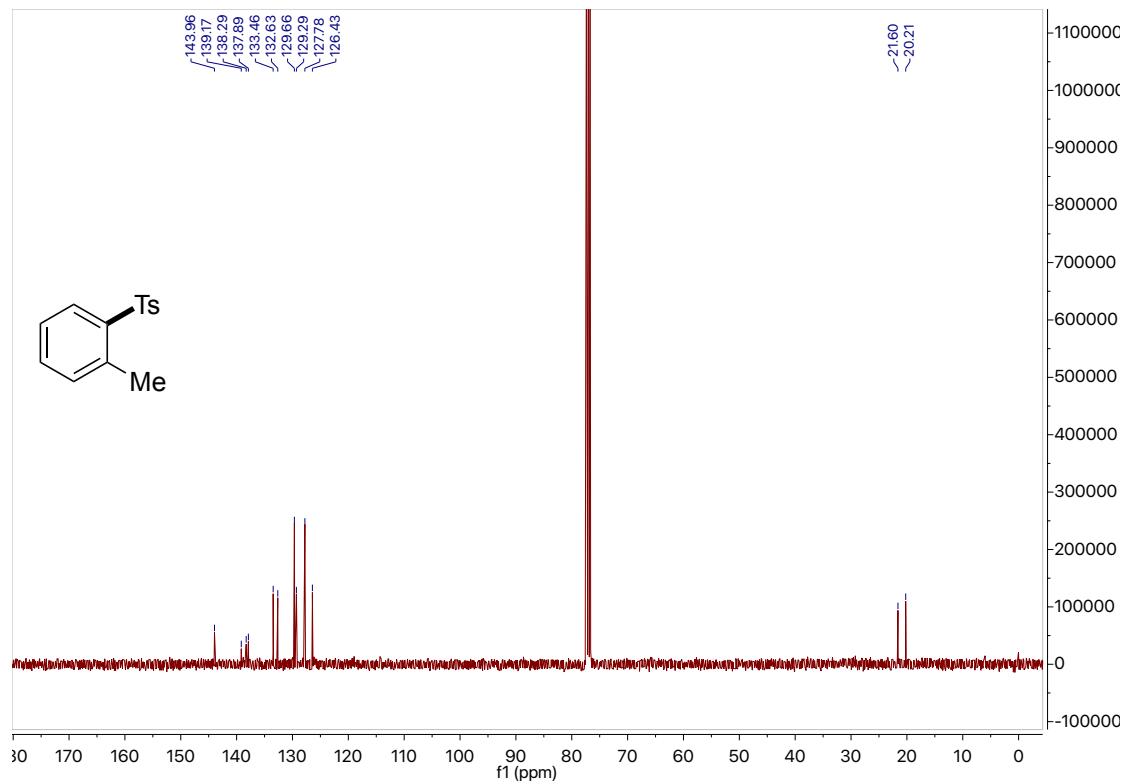
<sup>13</sup>C Spectra



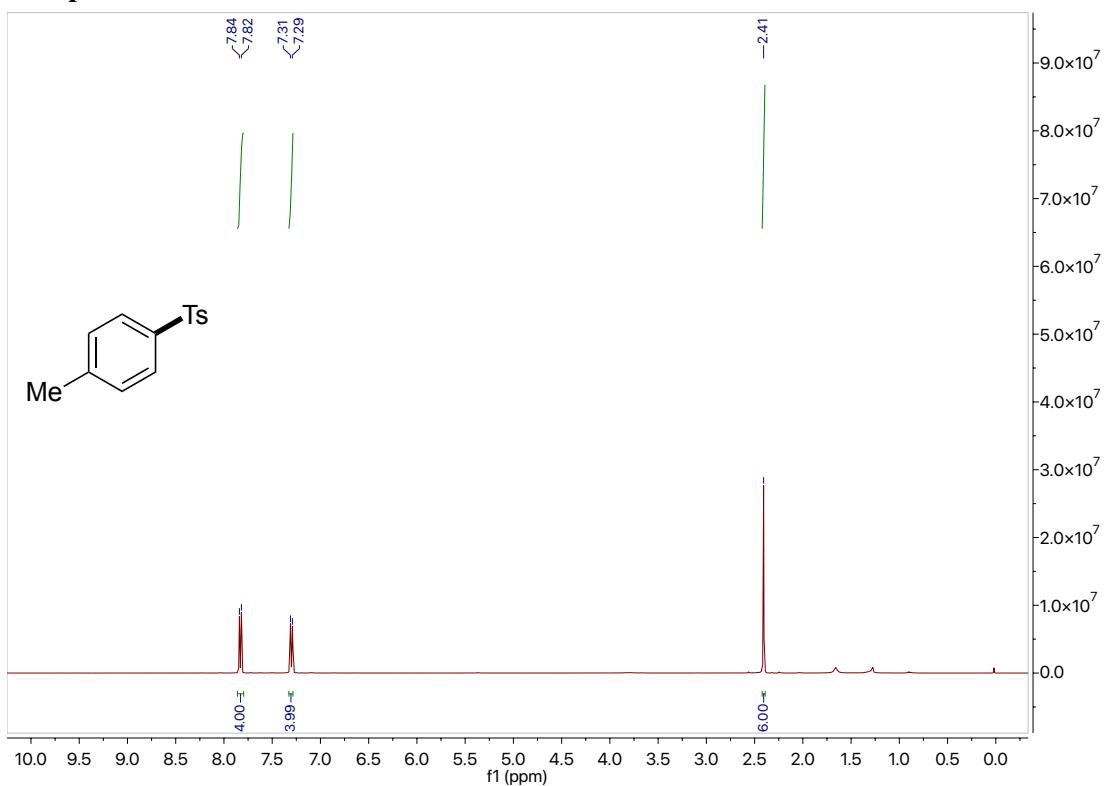
(3d)  
<sup>1</sup>H Spectra



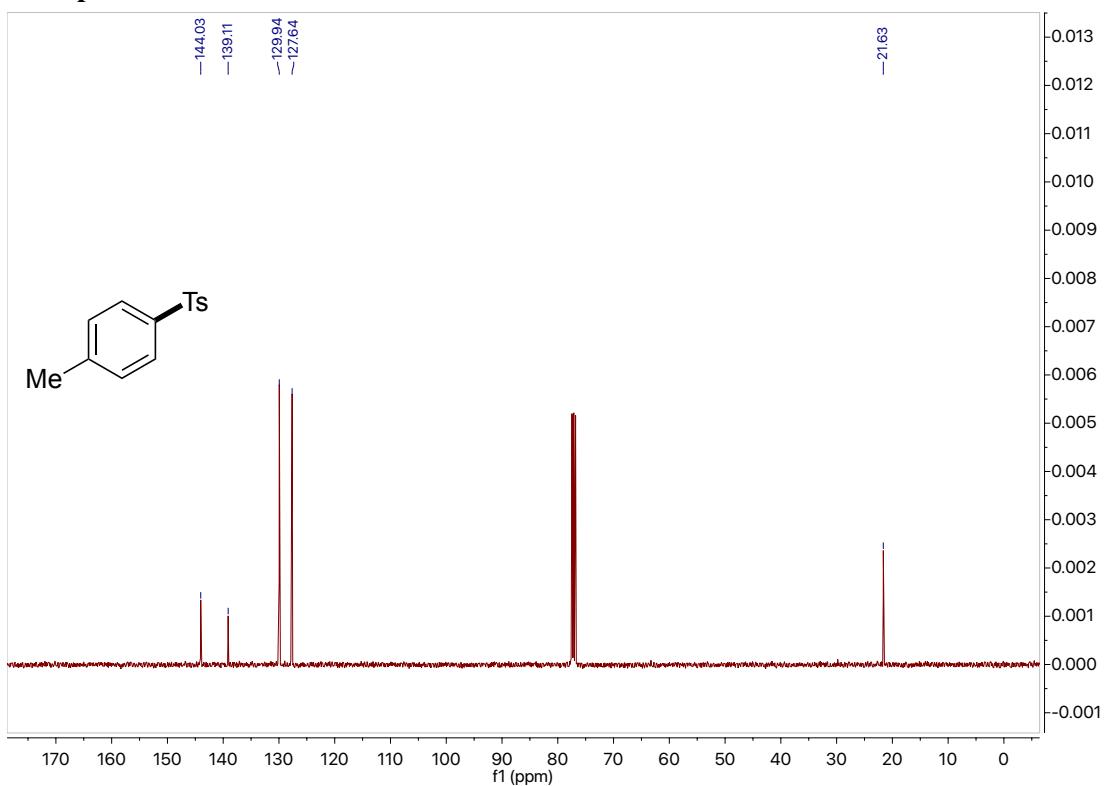
<sup>13</sup>C Spectra



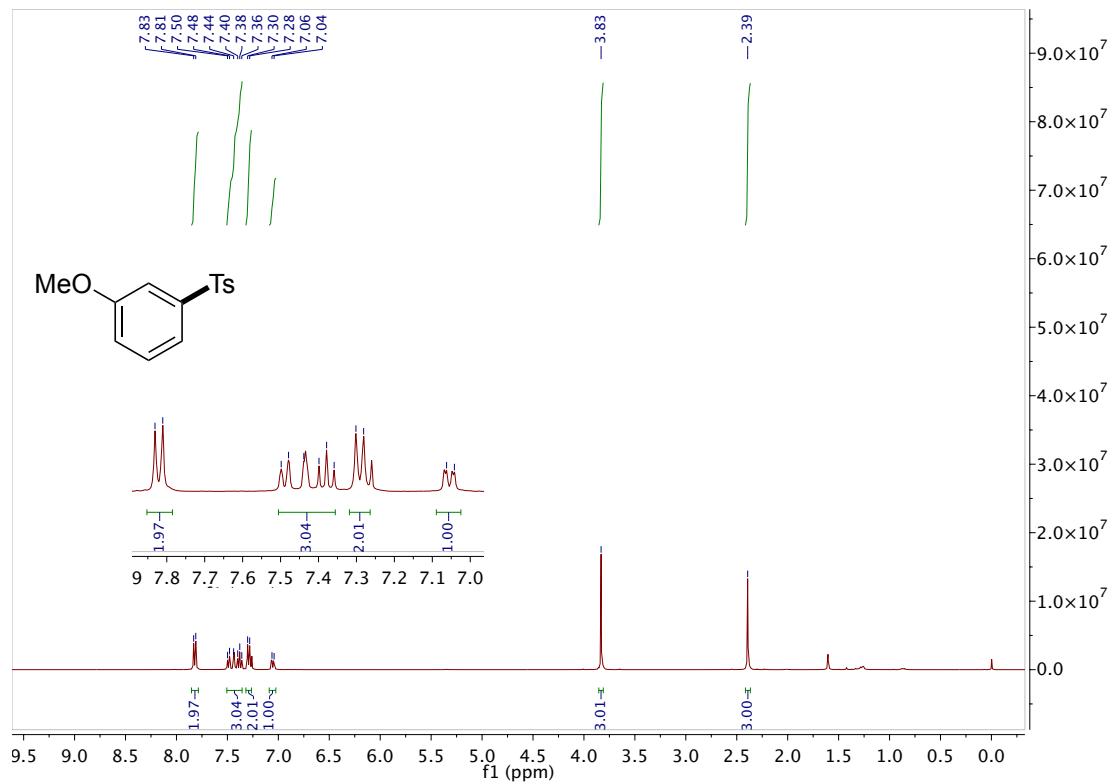
(3e)  
<sup>1</sup>H Spectra



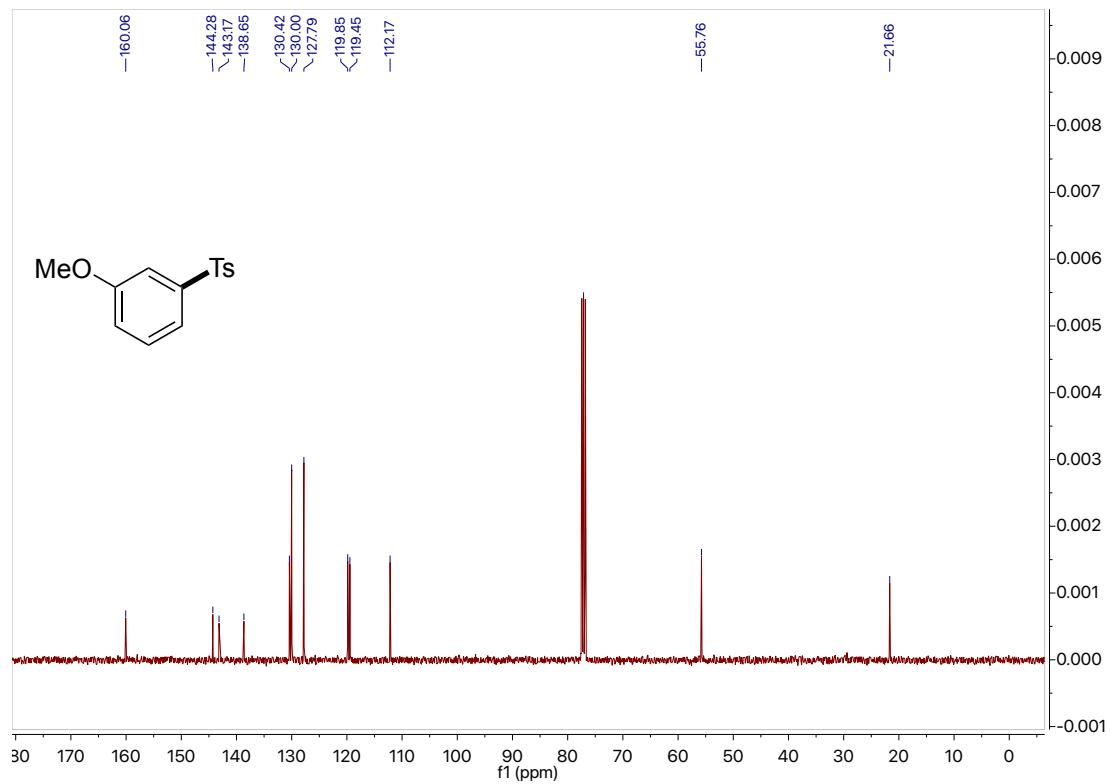
<sup>13</sup>C Spectra



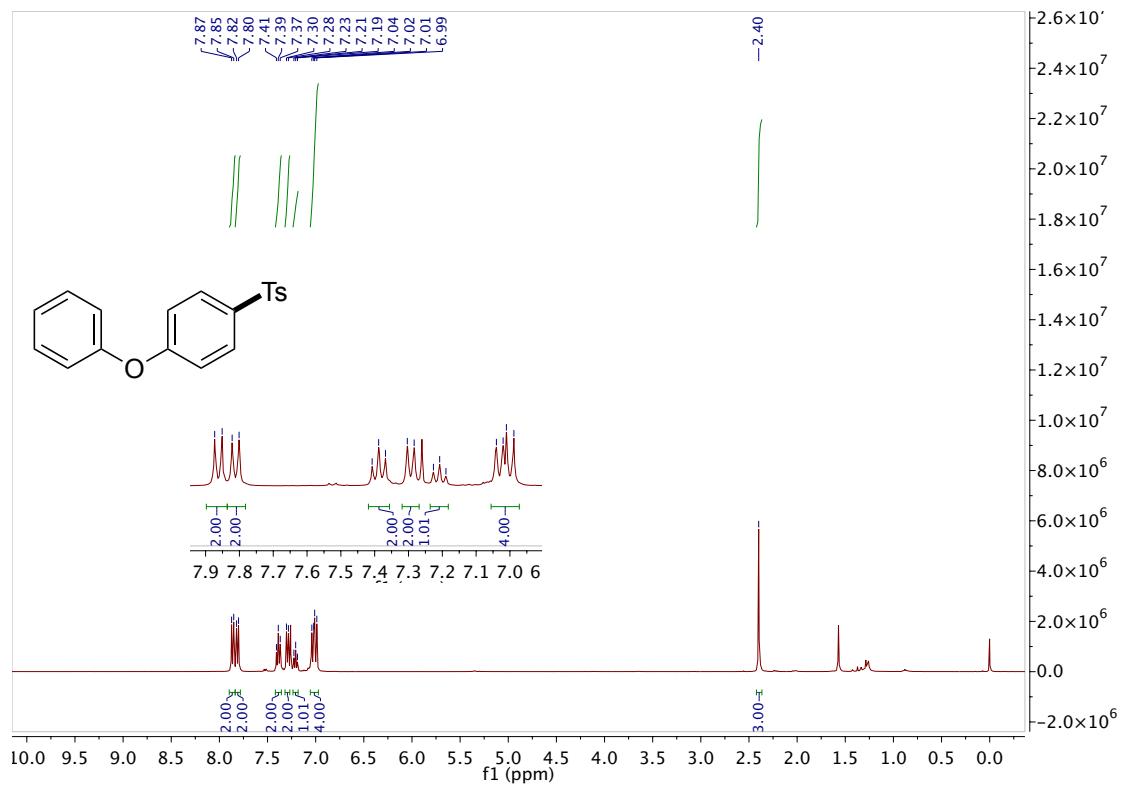
**(3f)**  
 **$^1\text{H}$  Spectra**



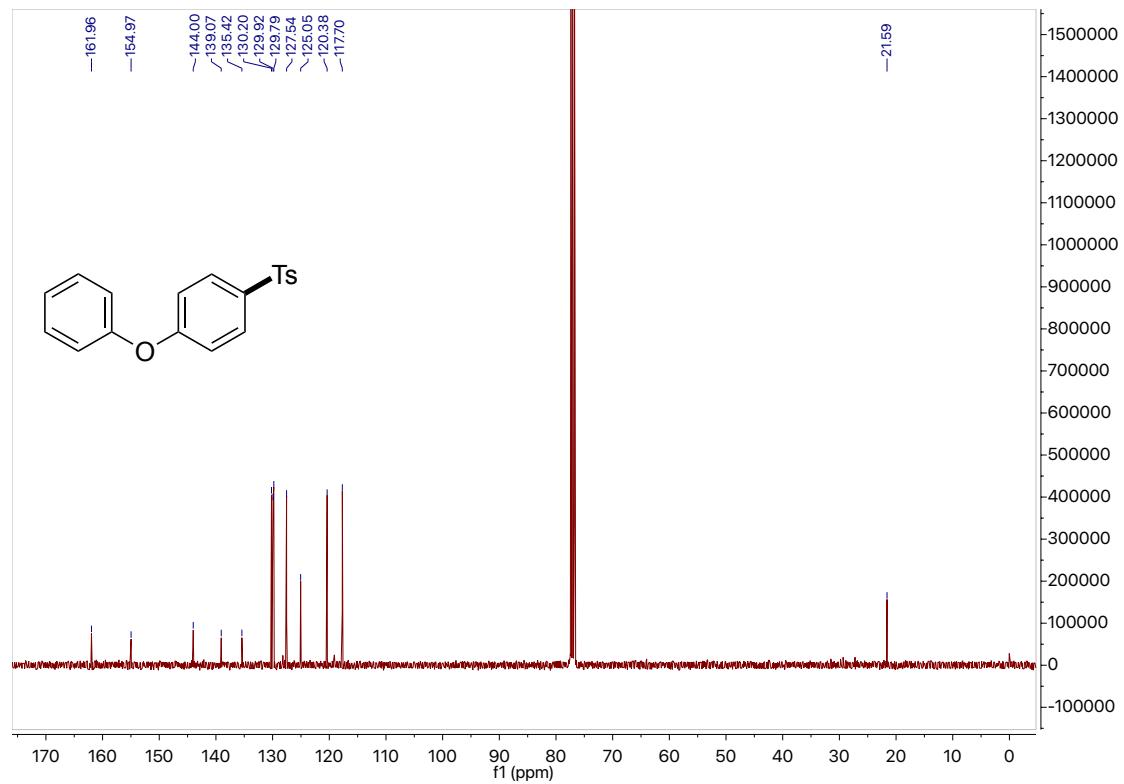
**$^{13}\text{C}$  Spectra**



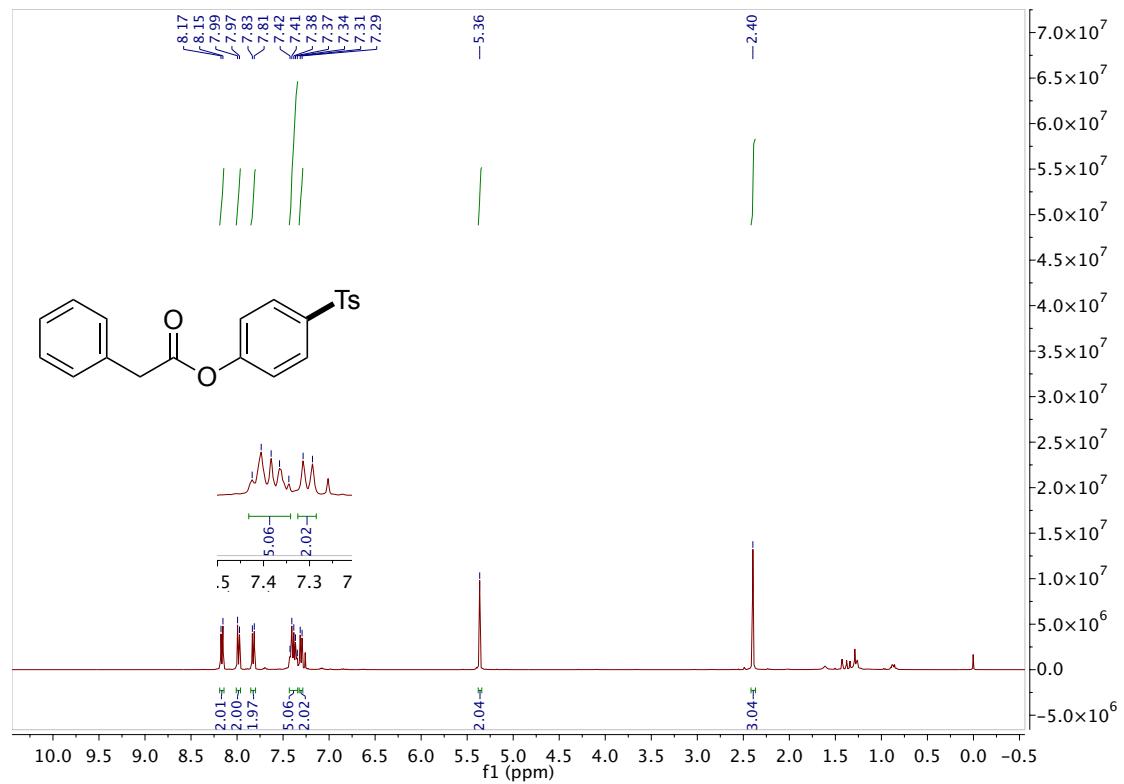
(3g)  
<sup>1</sup>H Spectra



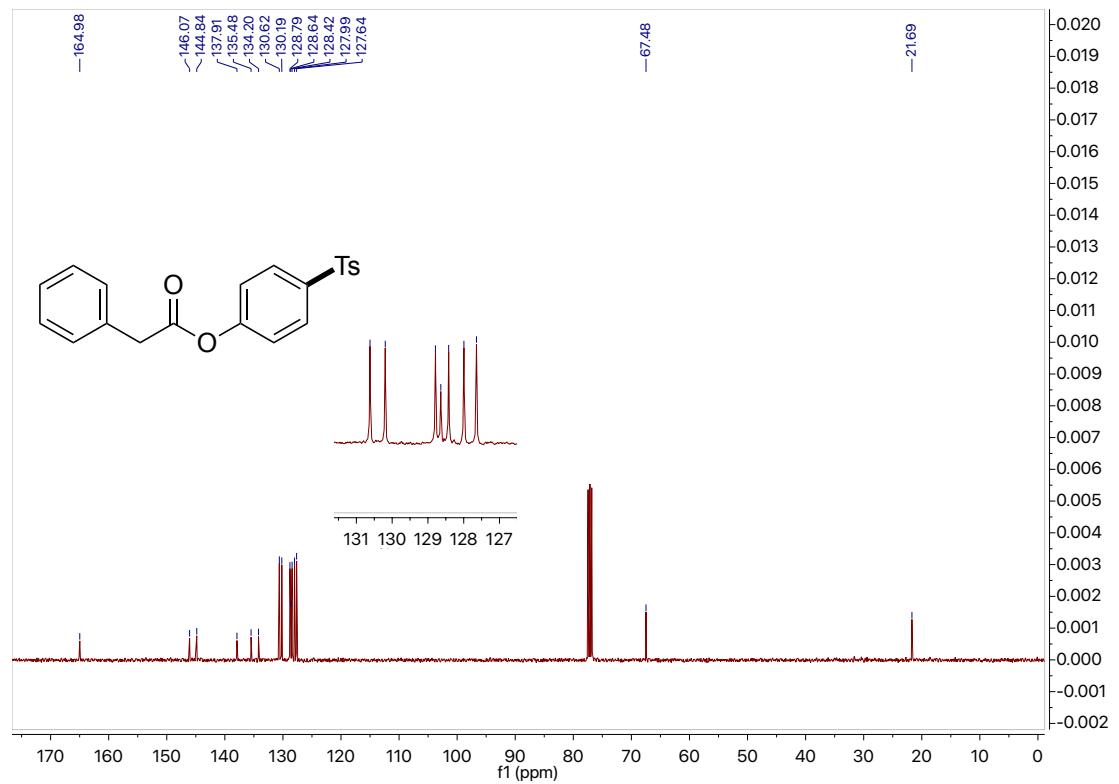
<sup>13</sup>C Spectra



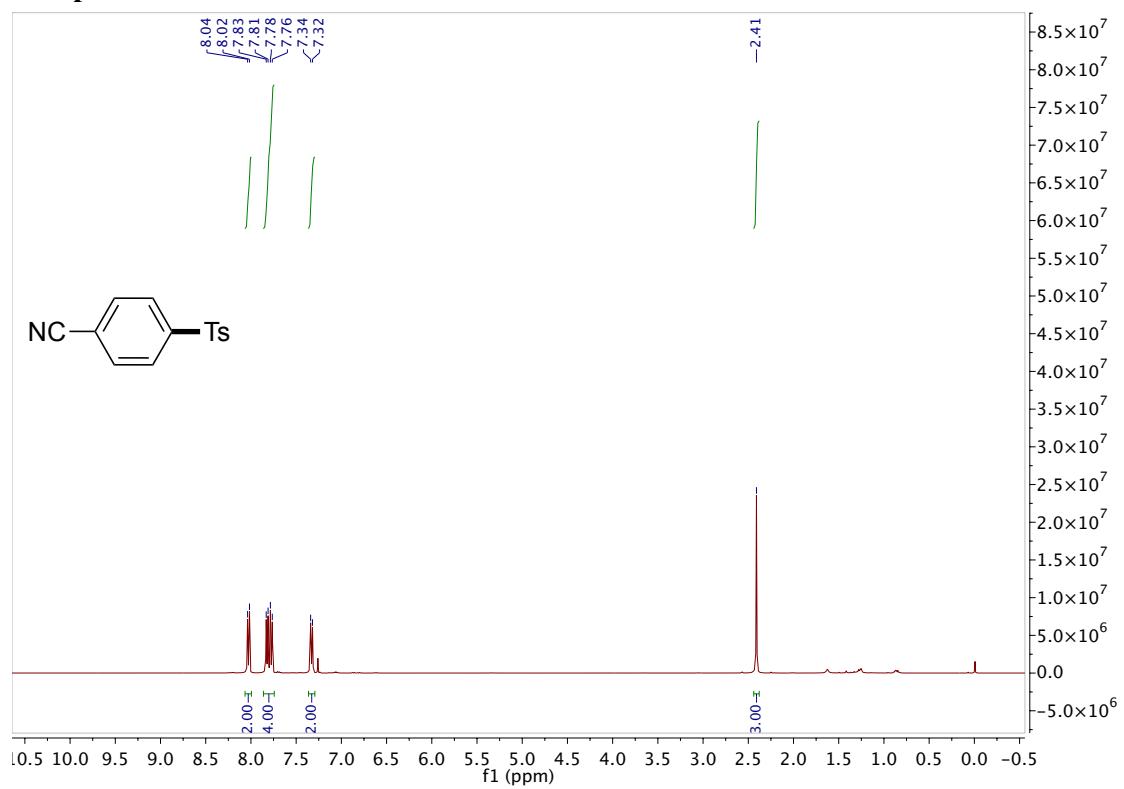
**(3h)**  
 **$^1\text{H}$  Spectra**



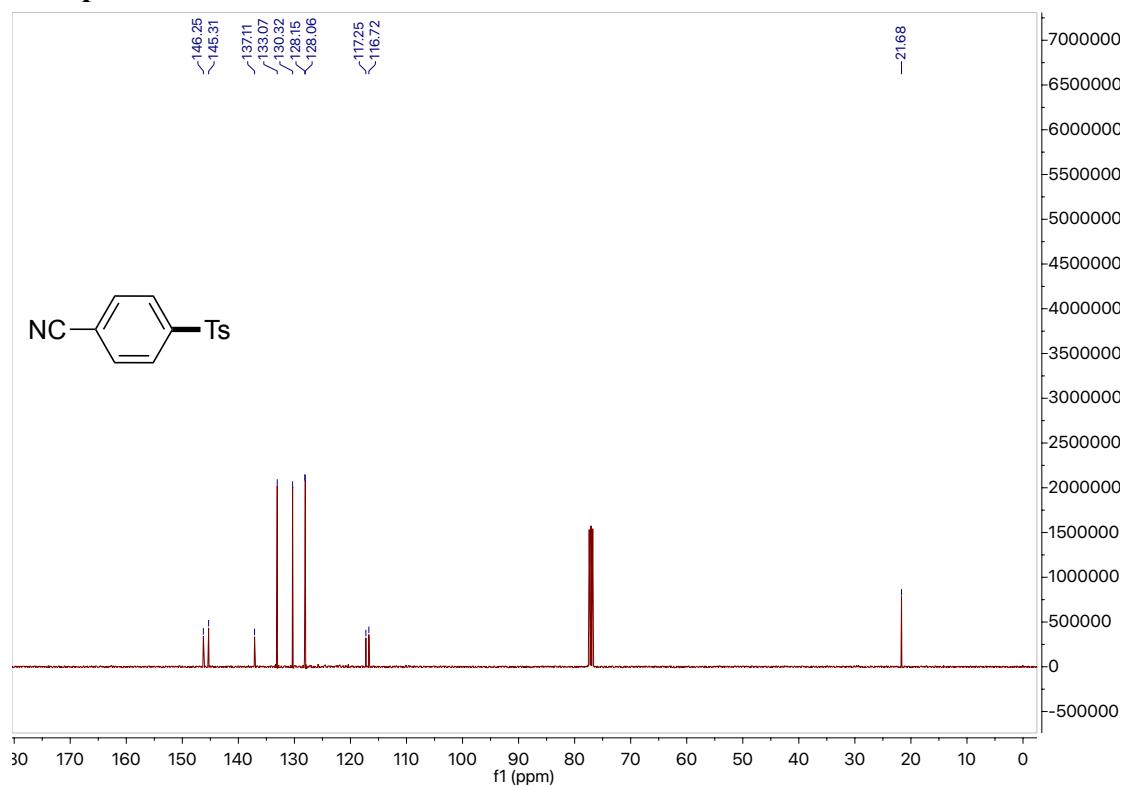
**$^{13}\text{C}$  Spectra**



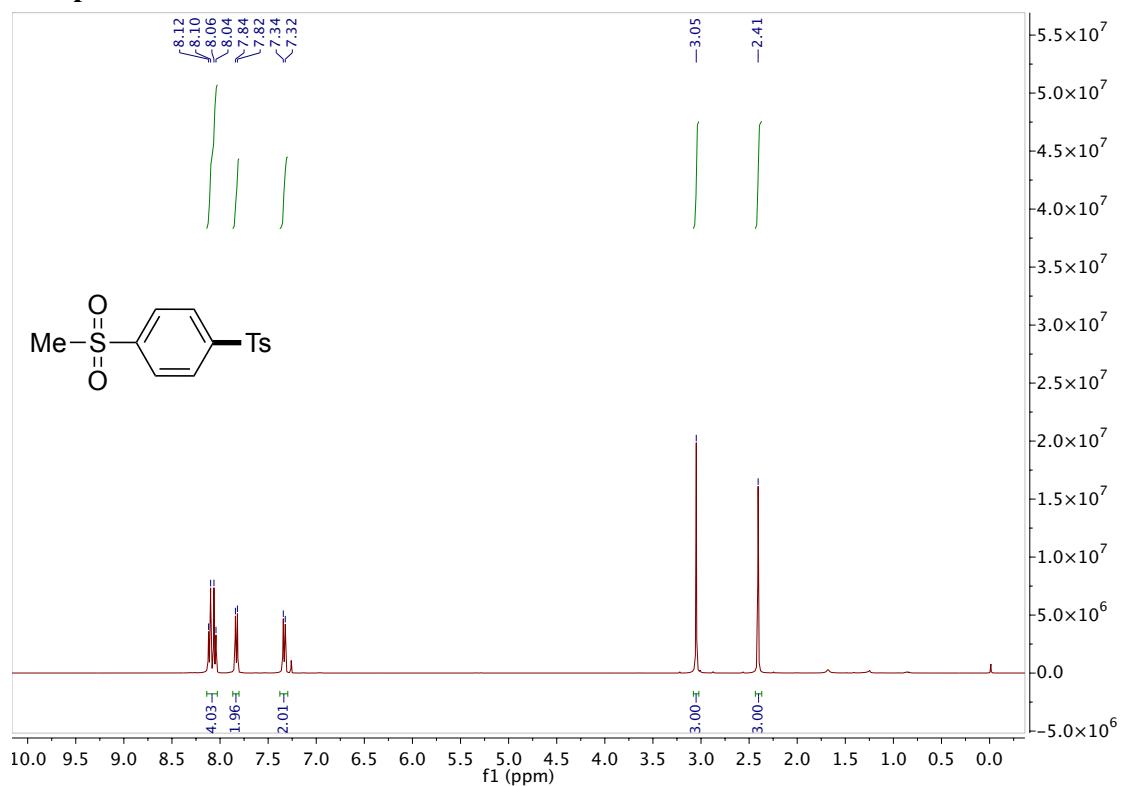
(3i)  
<sup>1</sup>H Spectra



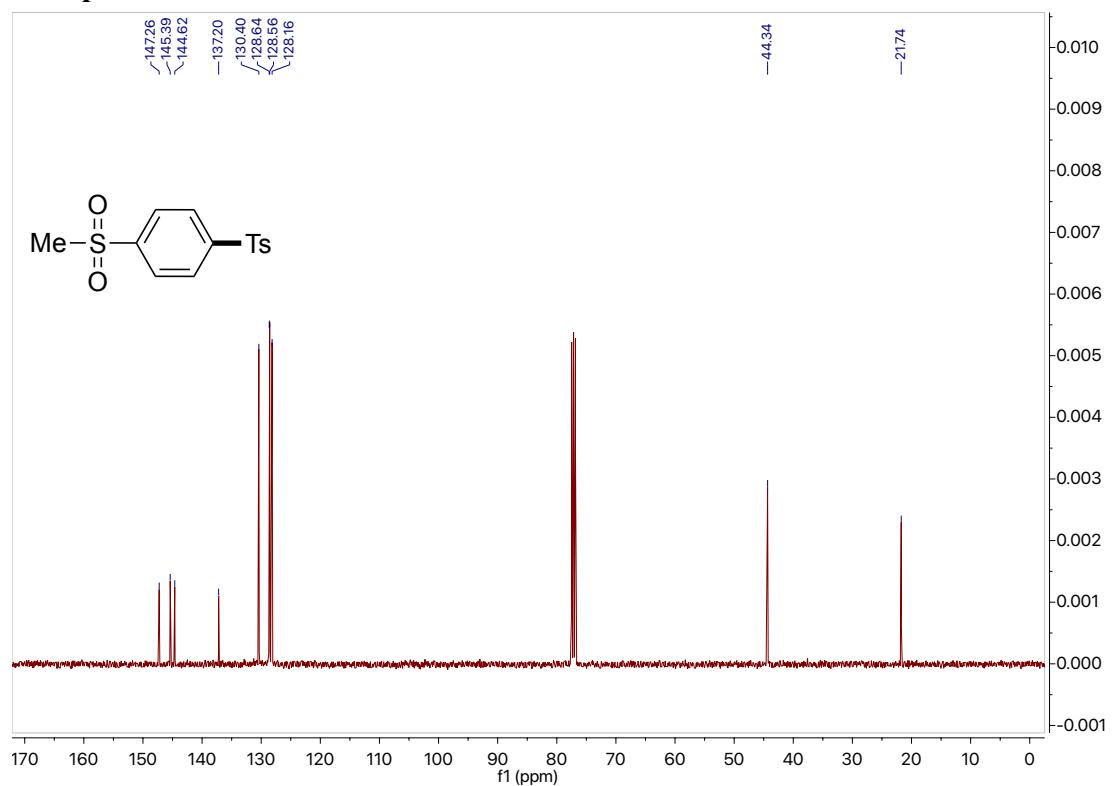
<sup>13</sup>C Spectra



**(3j)**  
 **$^1\text{H}$  Spectra**

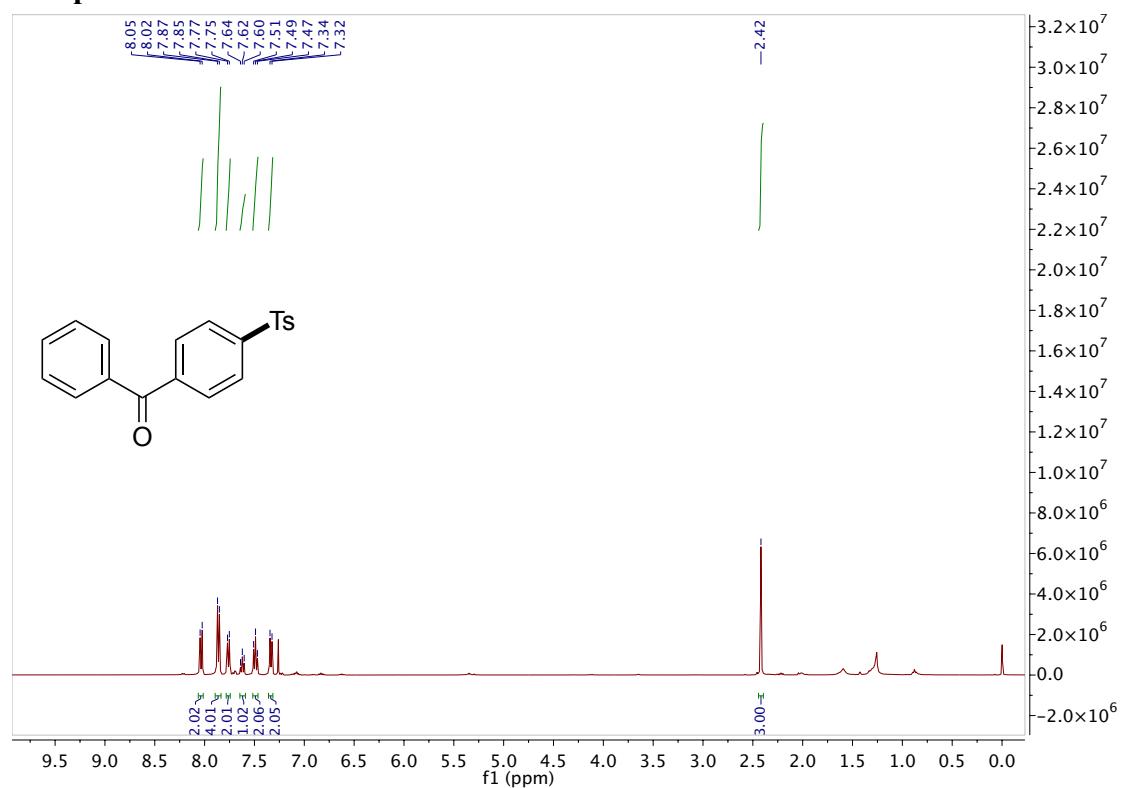


**$^{13}\text{C}$  Spectra**

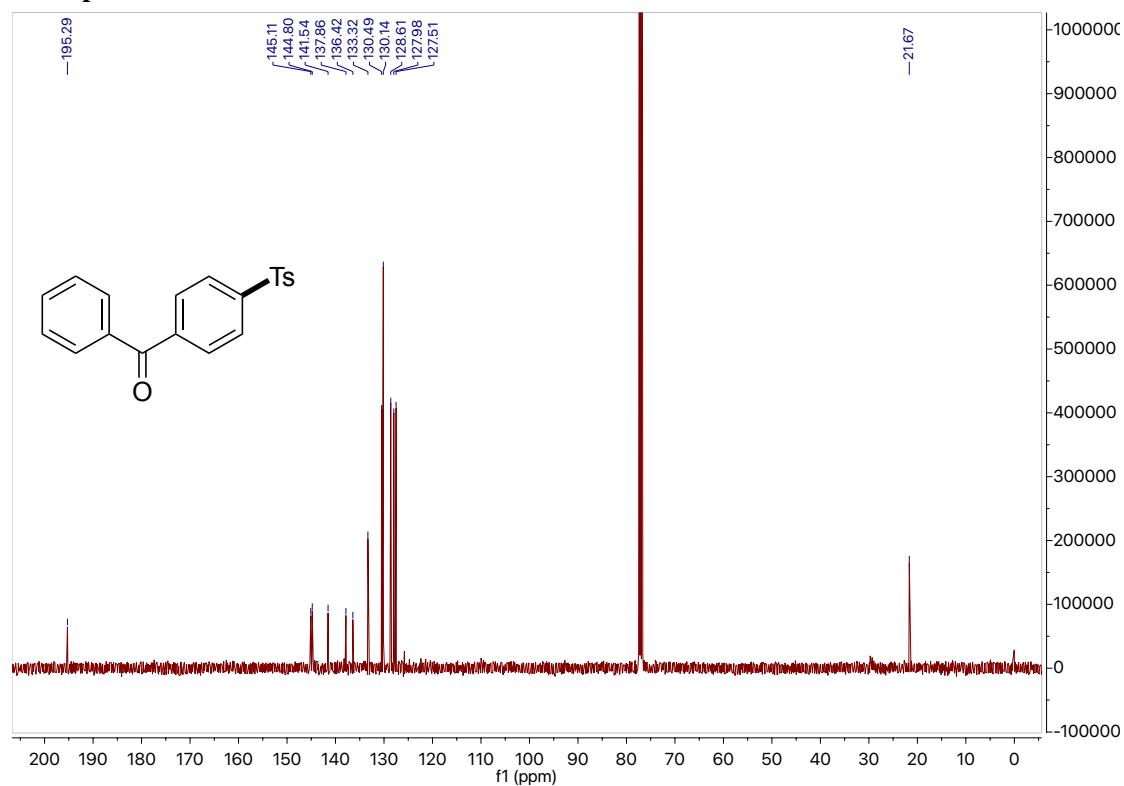


(3k)

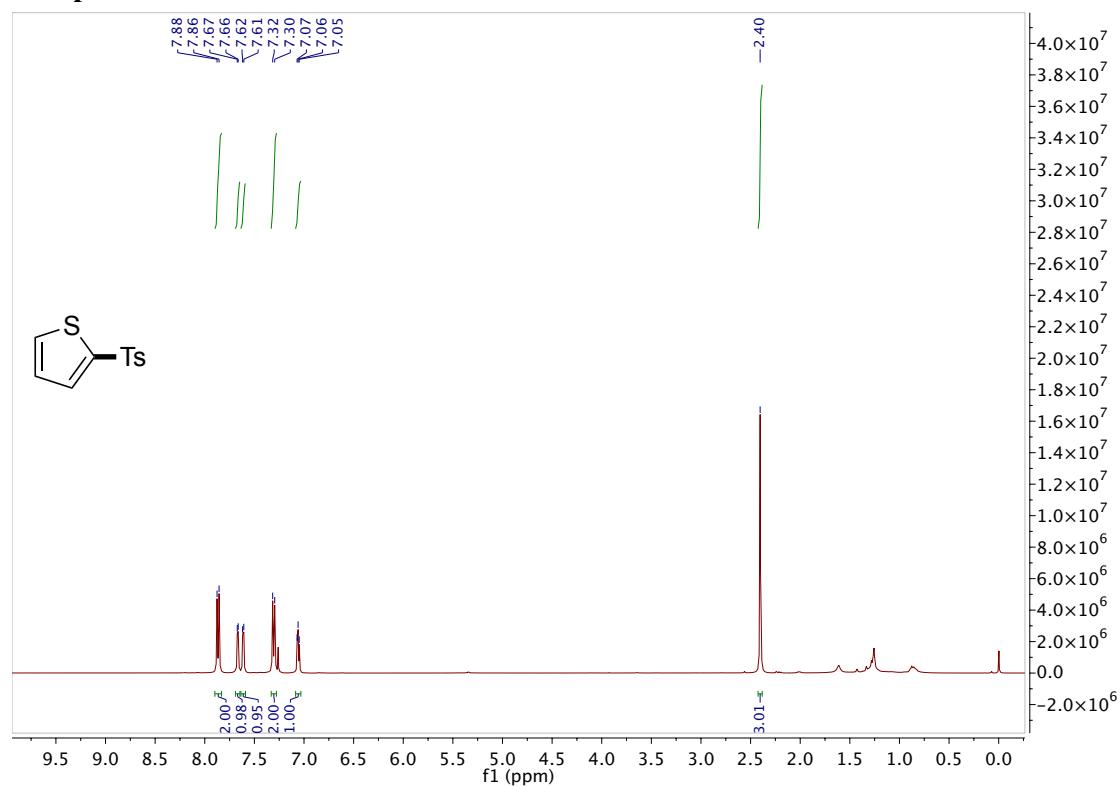
<sup>1</sup>H Spectra



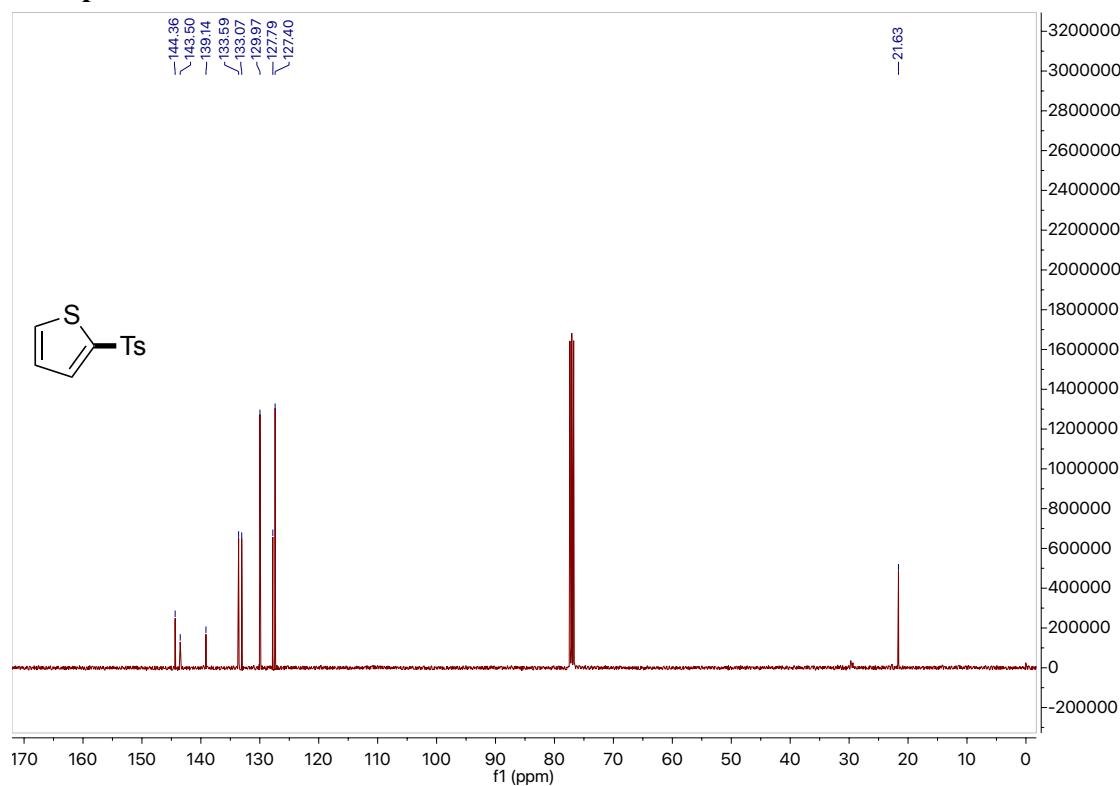
<sup>13</sup>C Spectra



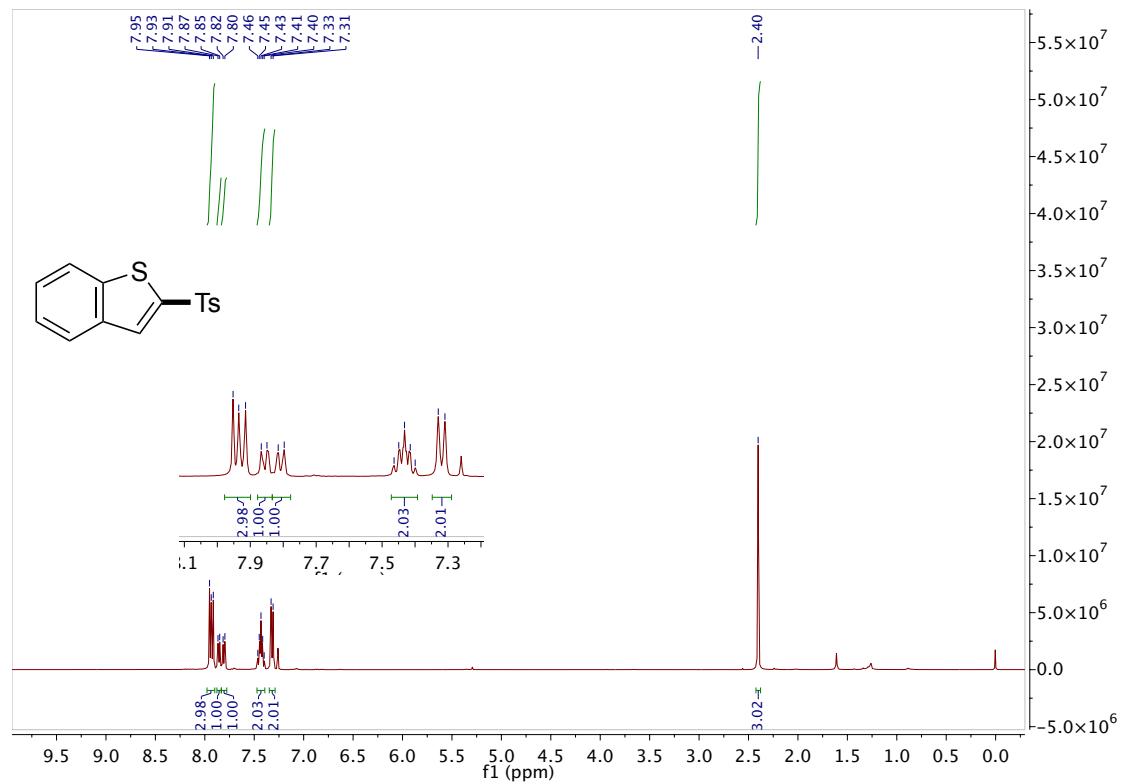
**(3l)**  
 **$^1\text{H}$  Spectra**



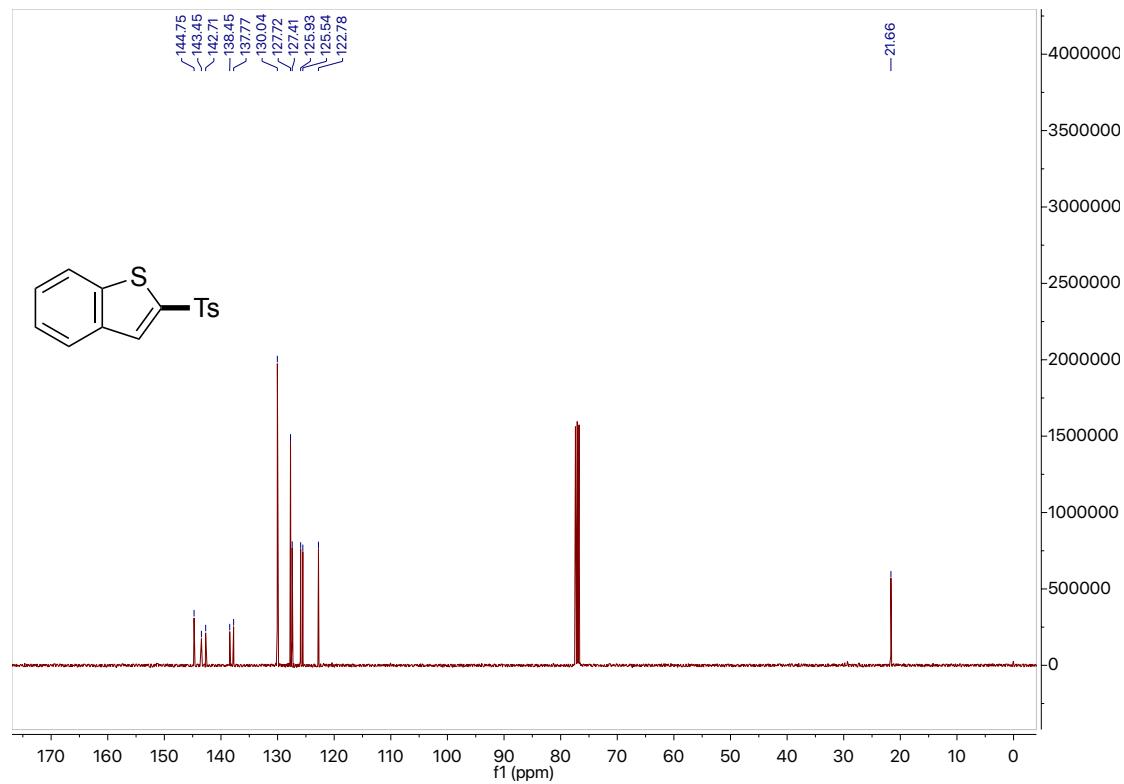
**$^{13}\text{C}$  Spectra**



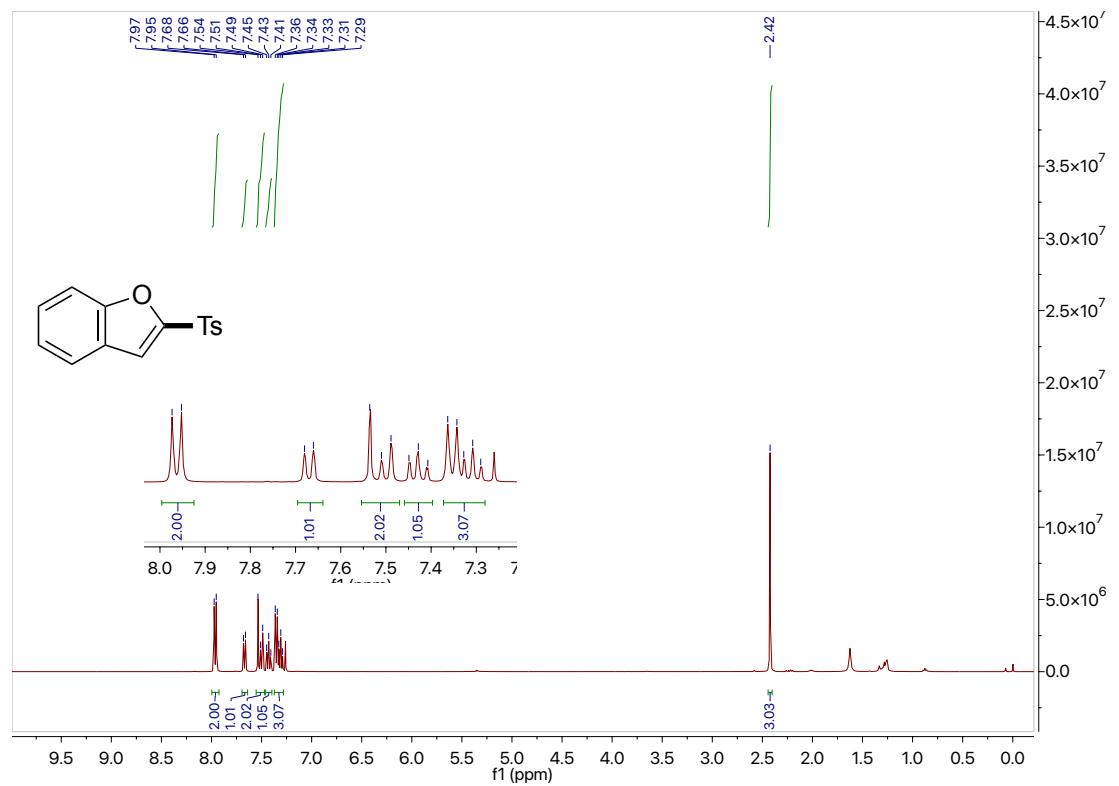
**(3m)**  
 **$^1\text{H}$  Spectra**



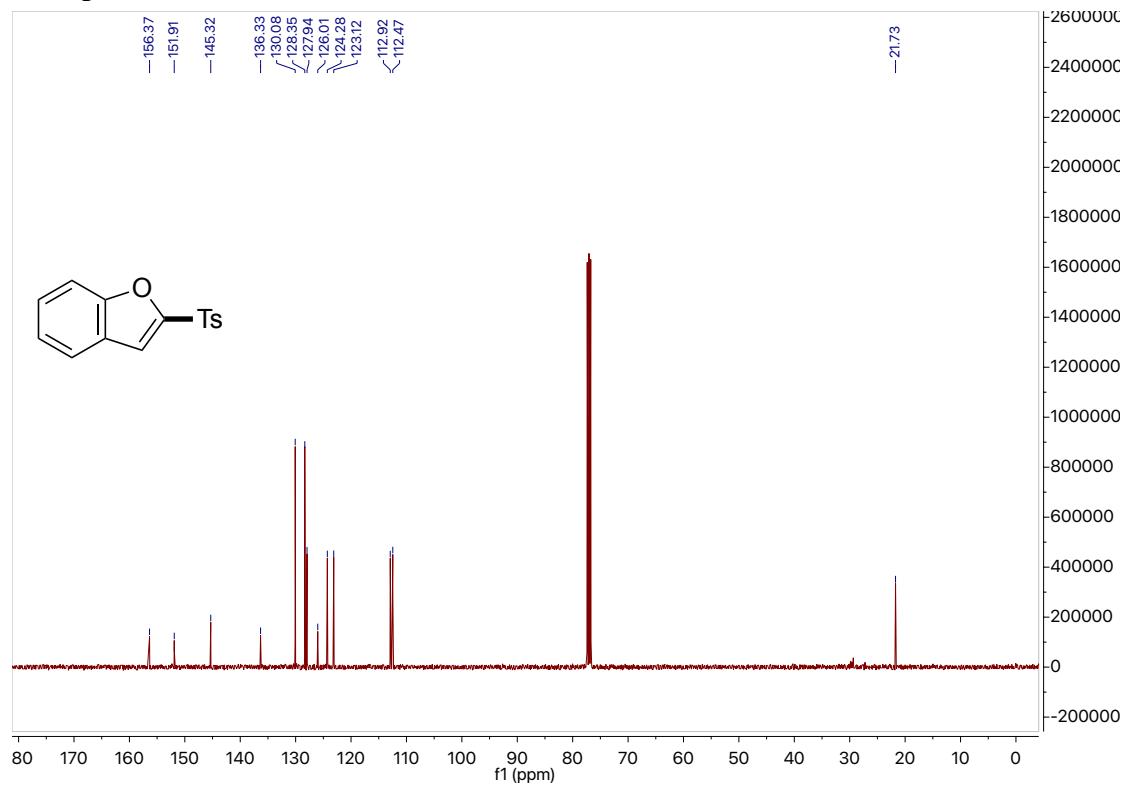
**$^{13}\text{C}$  Spectra**



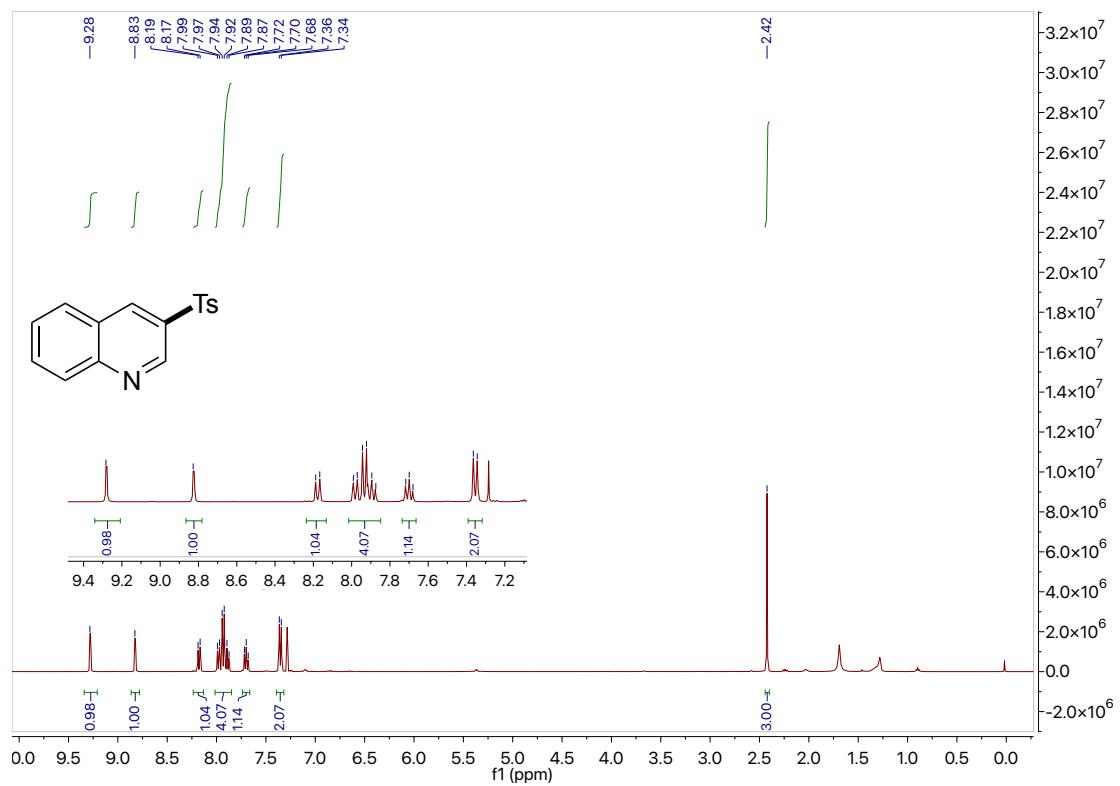
**(3n)**  
 **$^1\text{H}$  Spectra**



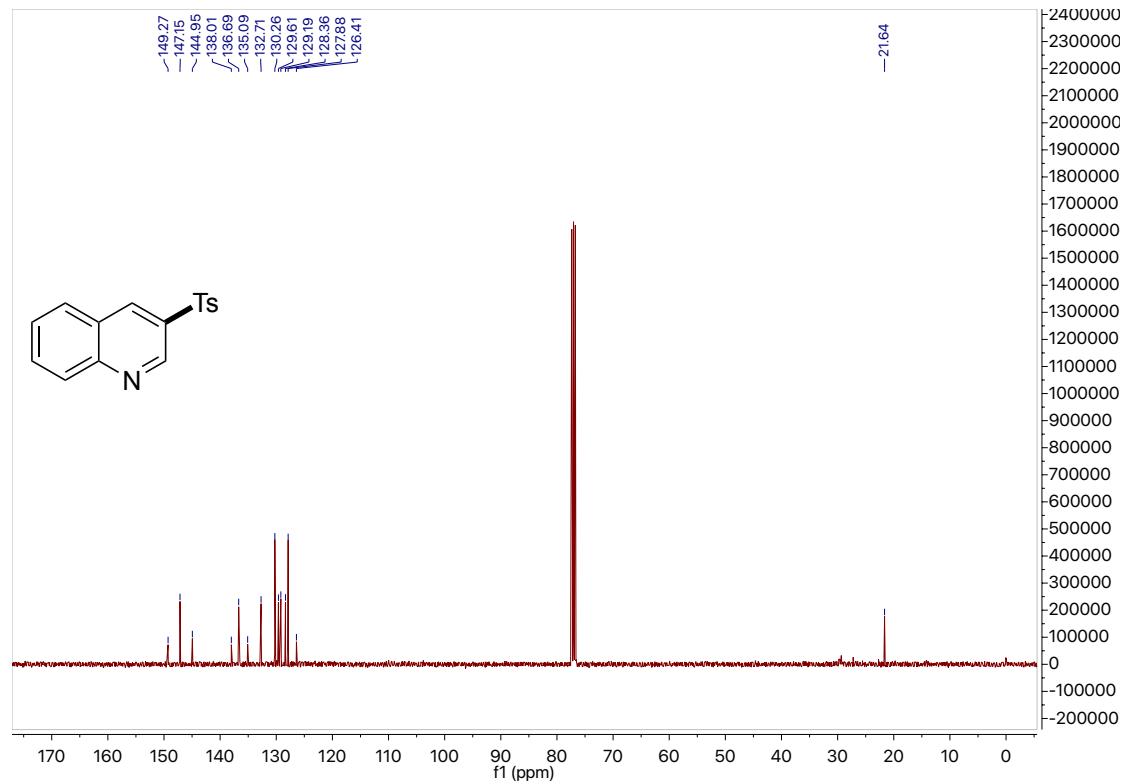
**$^{13}\text{C}$  Spectra**



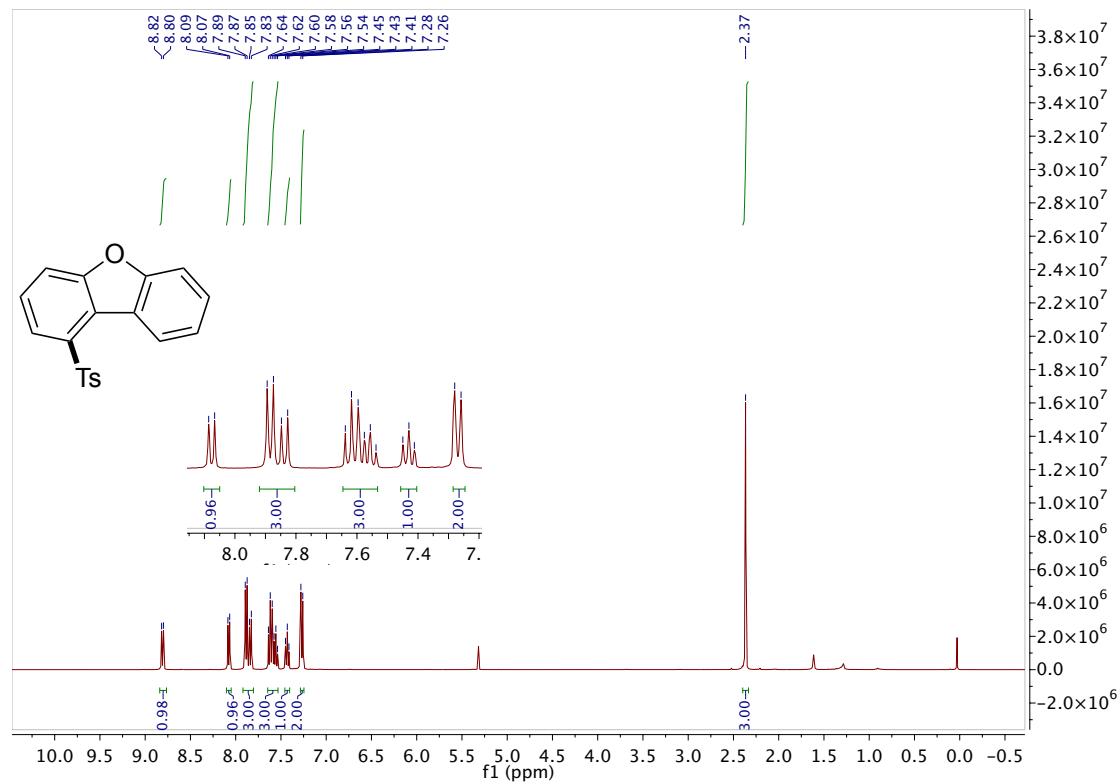
(3o)  
<sup>1</sup>H Spectra



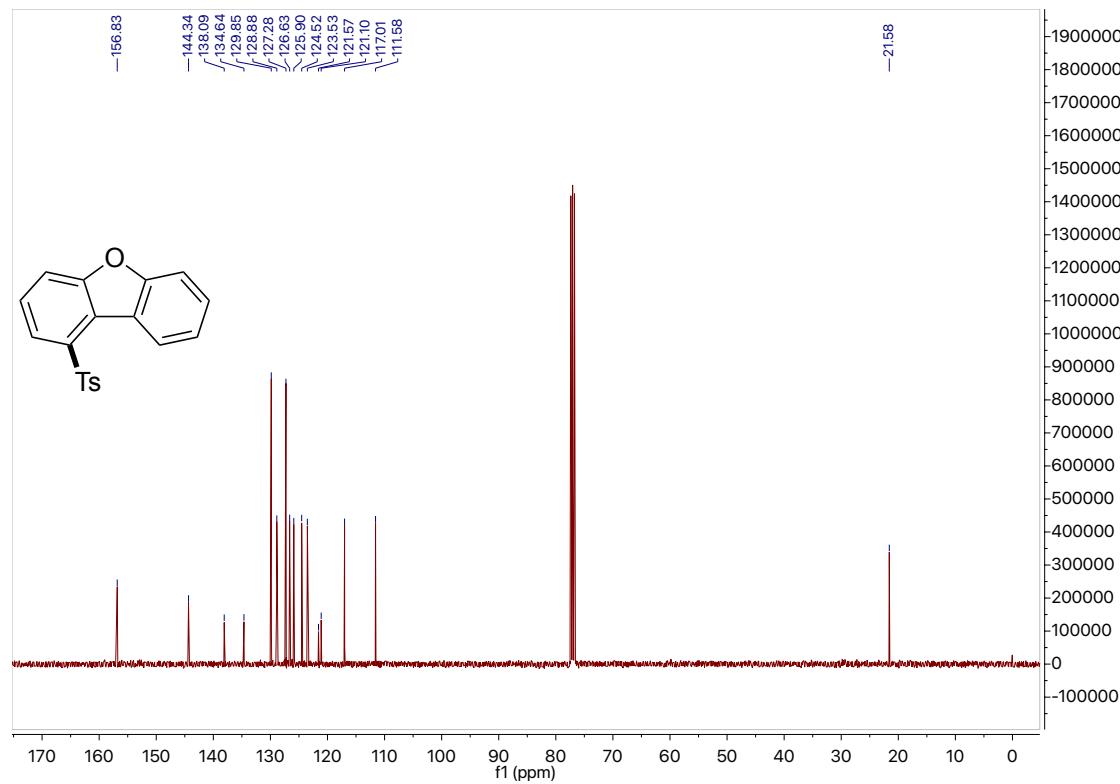
<sup>13</sup>C Spectra



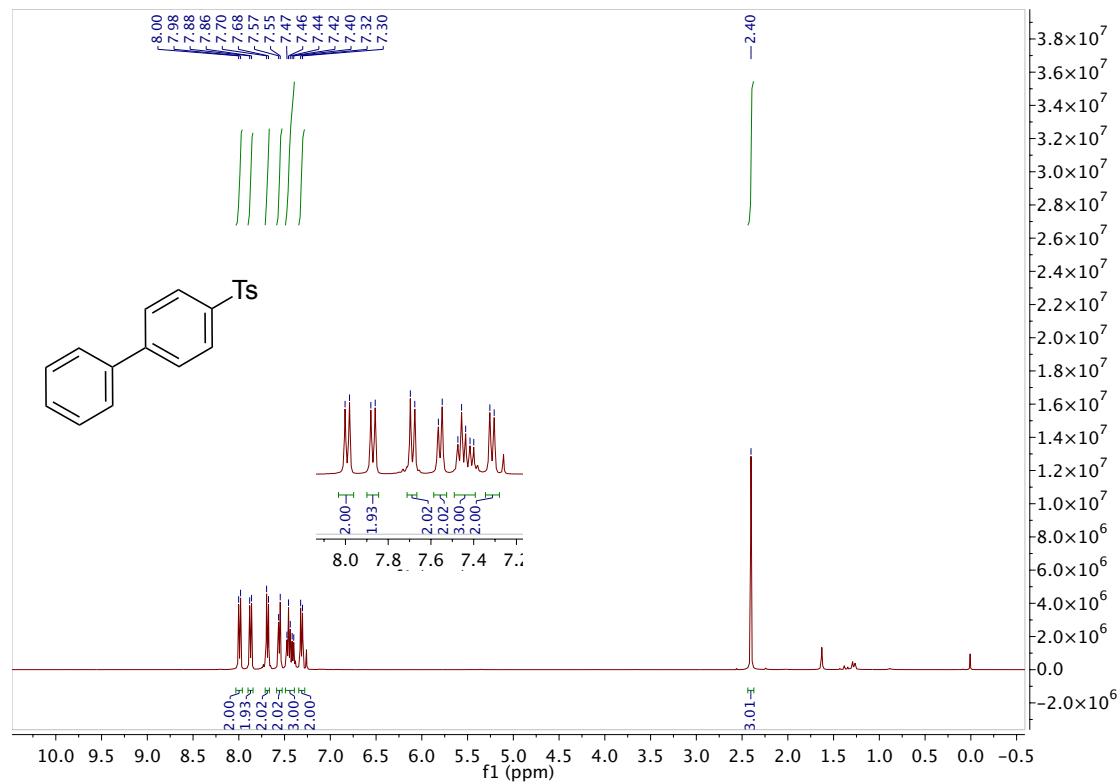
**(3p)**  
 **$^1\text{H}$  Spectra**



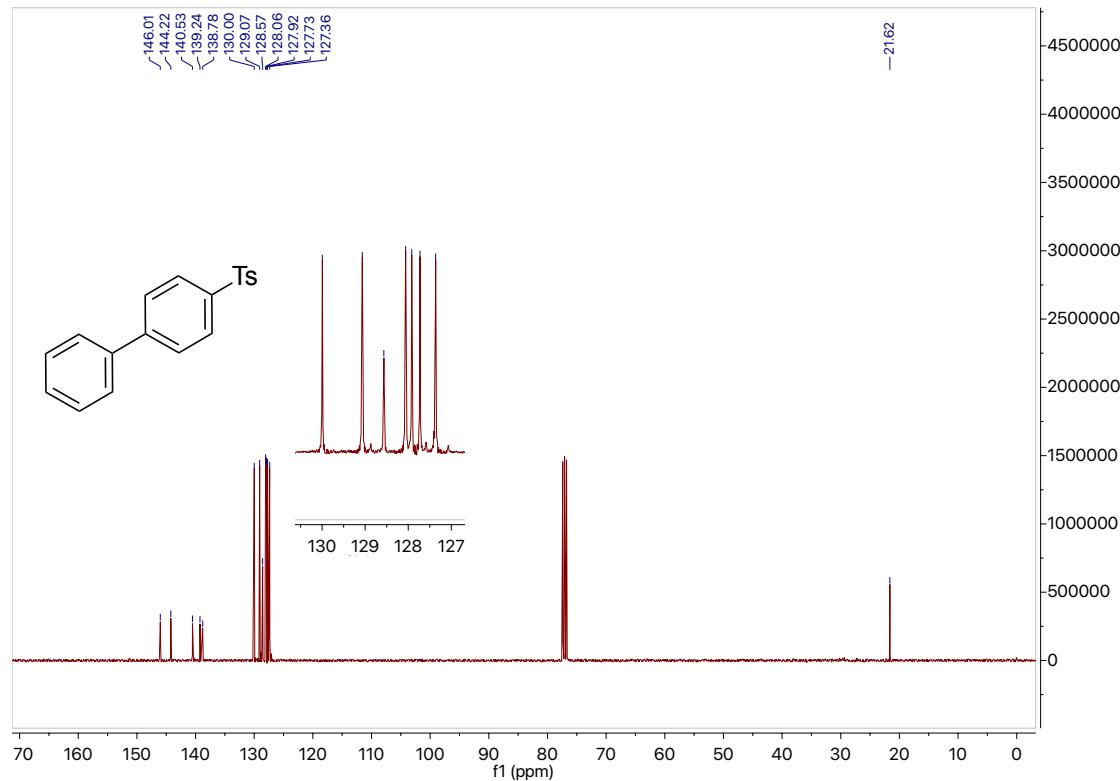
**$^{13}\text{C}$  Spectra**



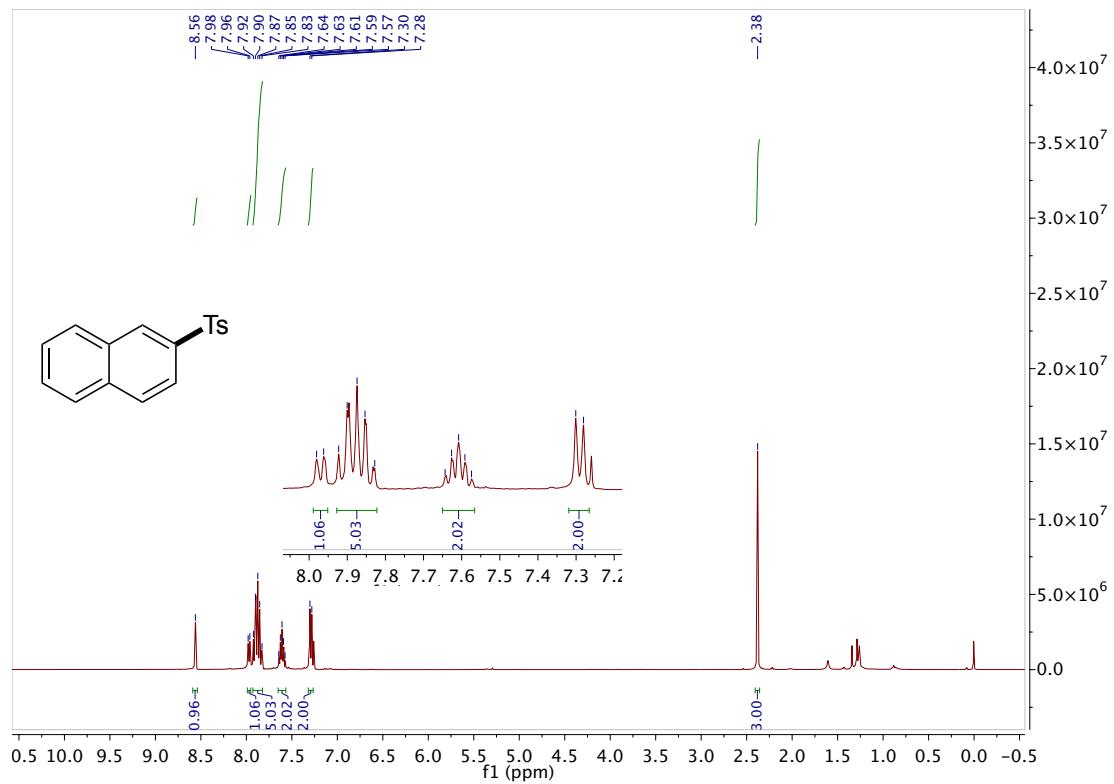
**(3q)**  
 **$^1\text{H}$  Spectra**



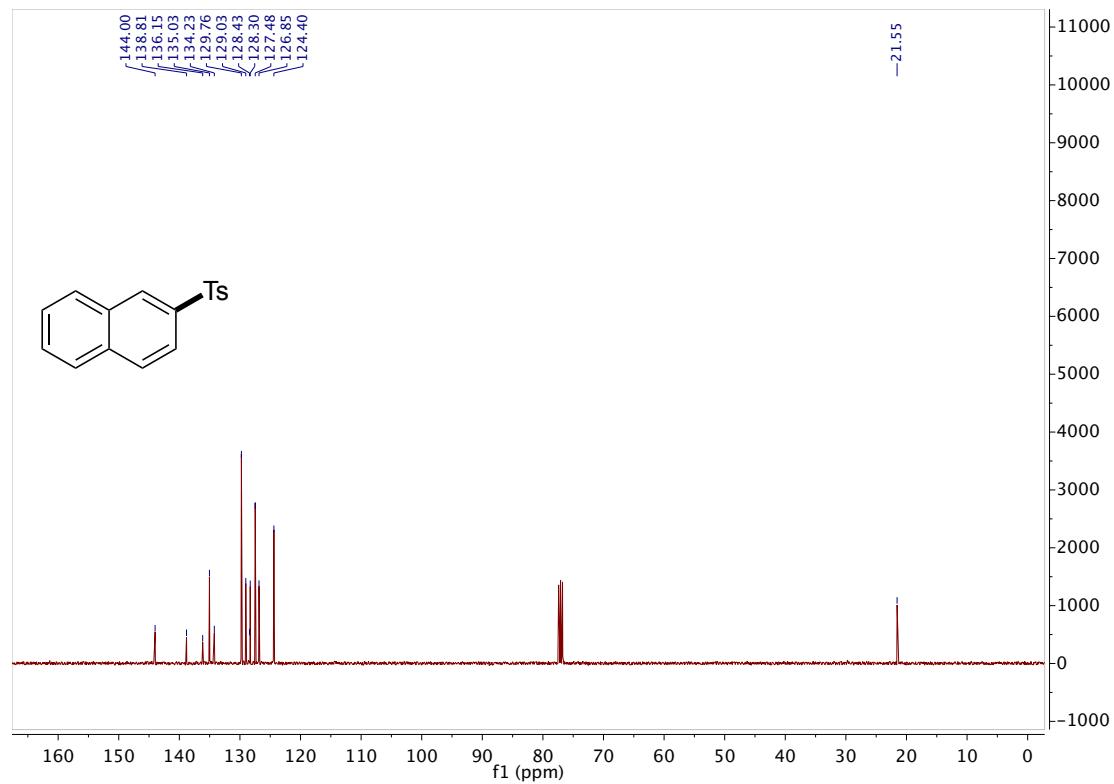
**$^{13}\text{C}$  Spectra**



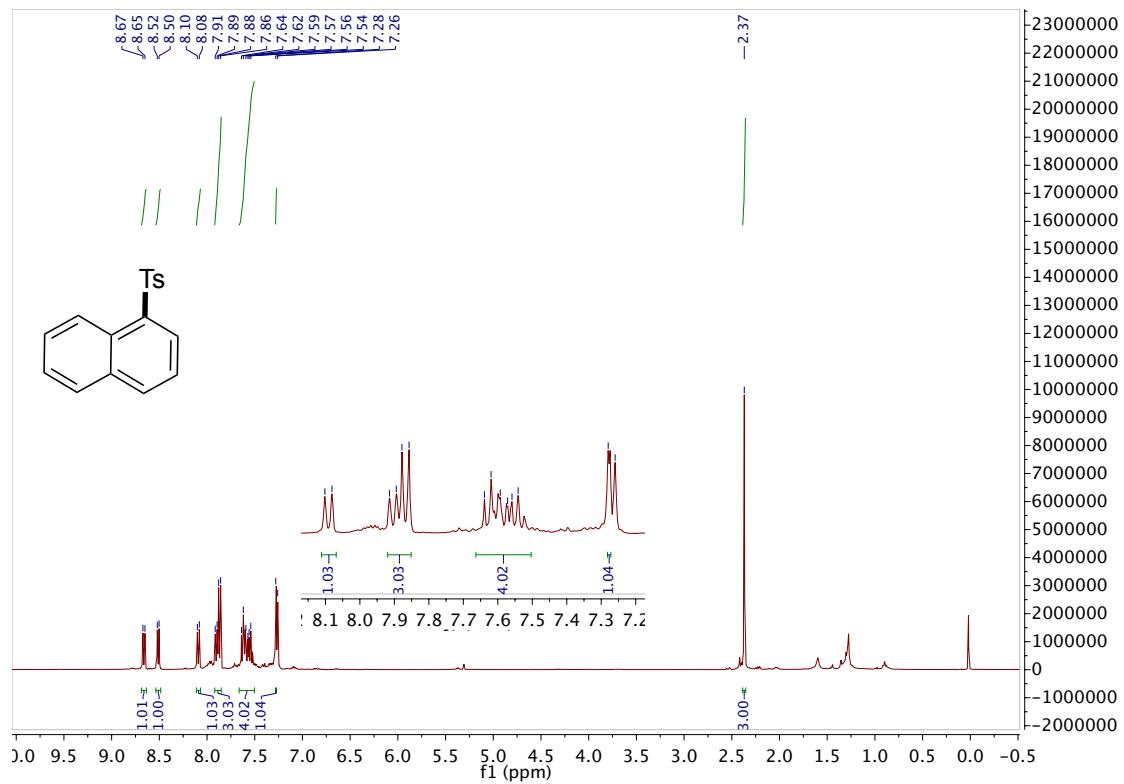
**(3r)**  
 **$^1\text{H}$  Spectra**



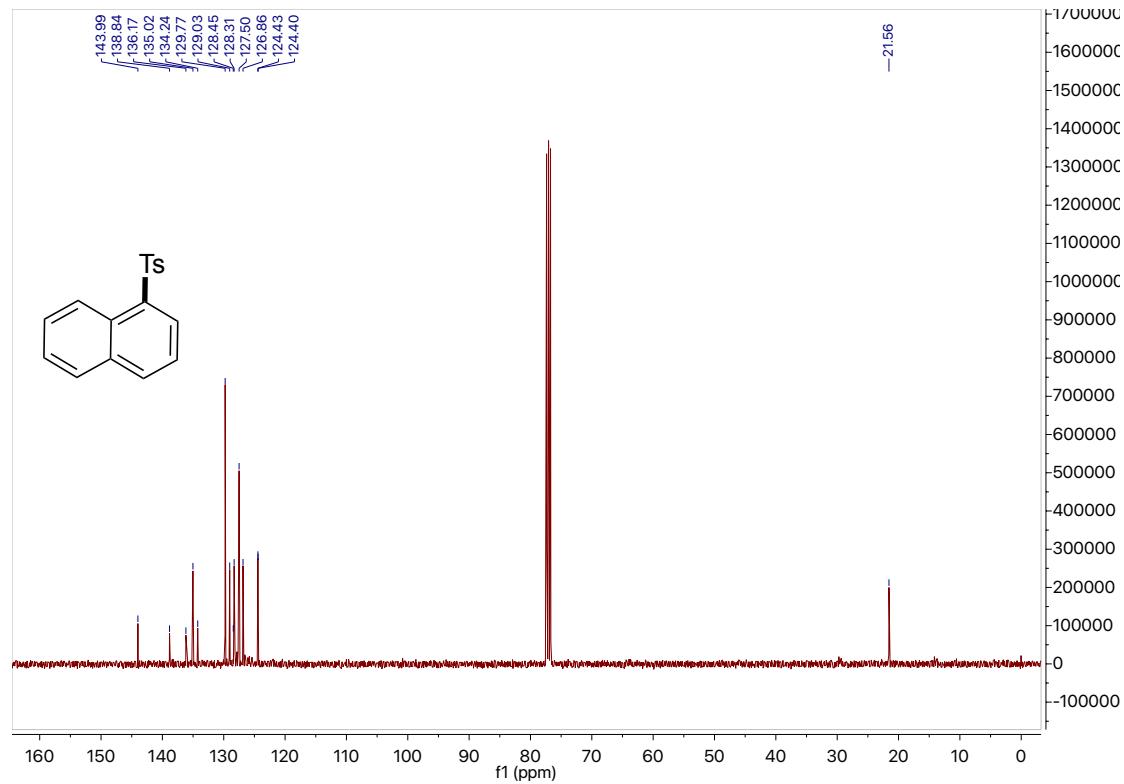
**$^{13}\text{C}$  Spectra**



**(3s)  
<sup>1</sup>H Spectra**

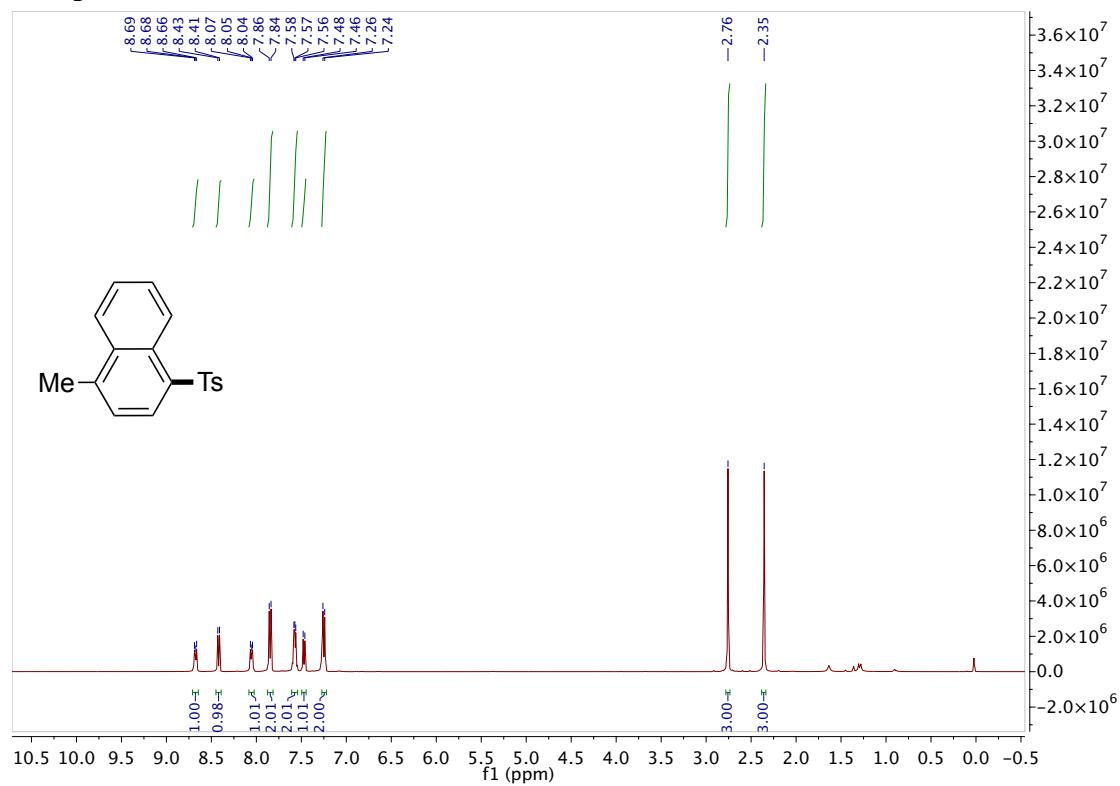


**<sup>13</sup>C Spectra**

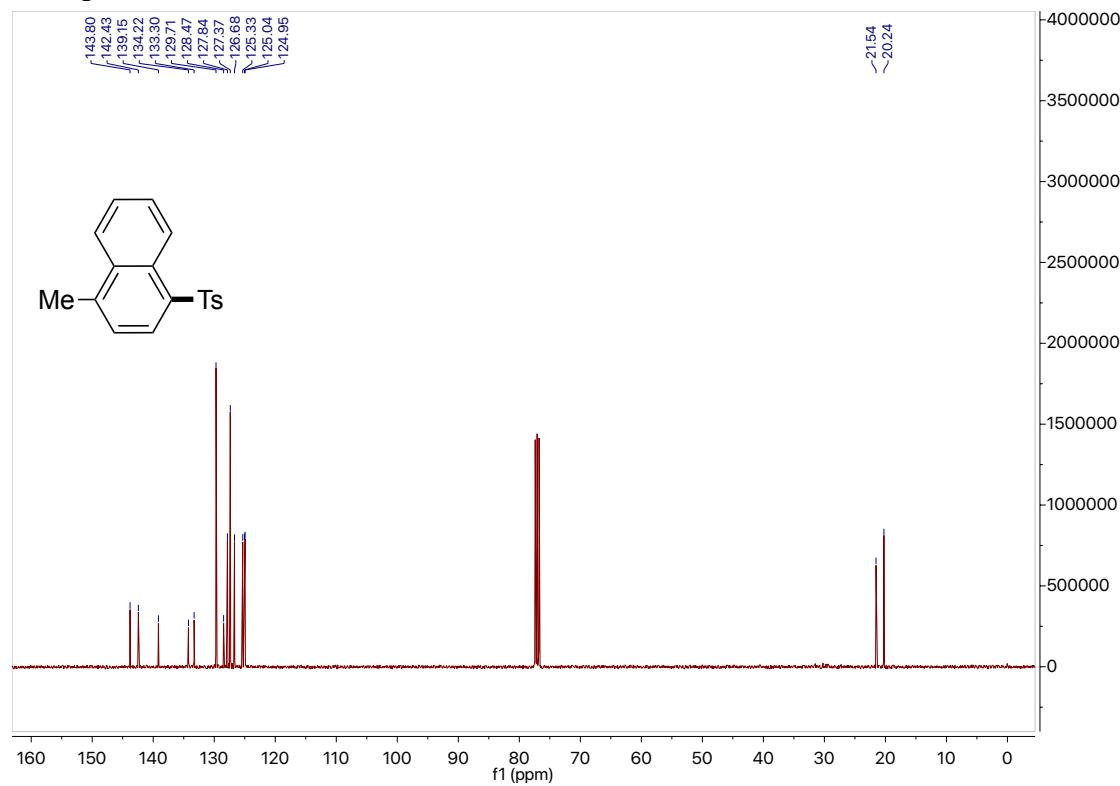


(3t)

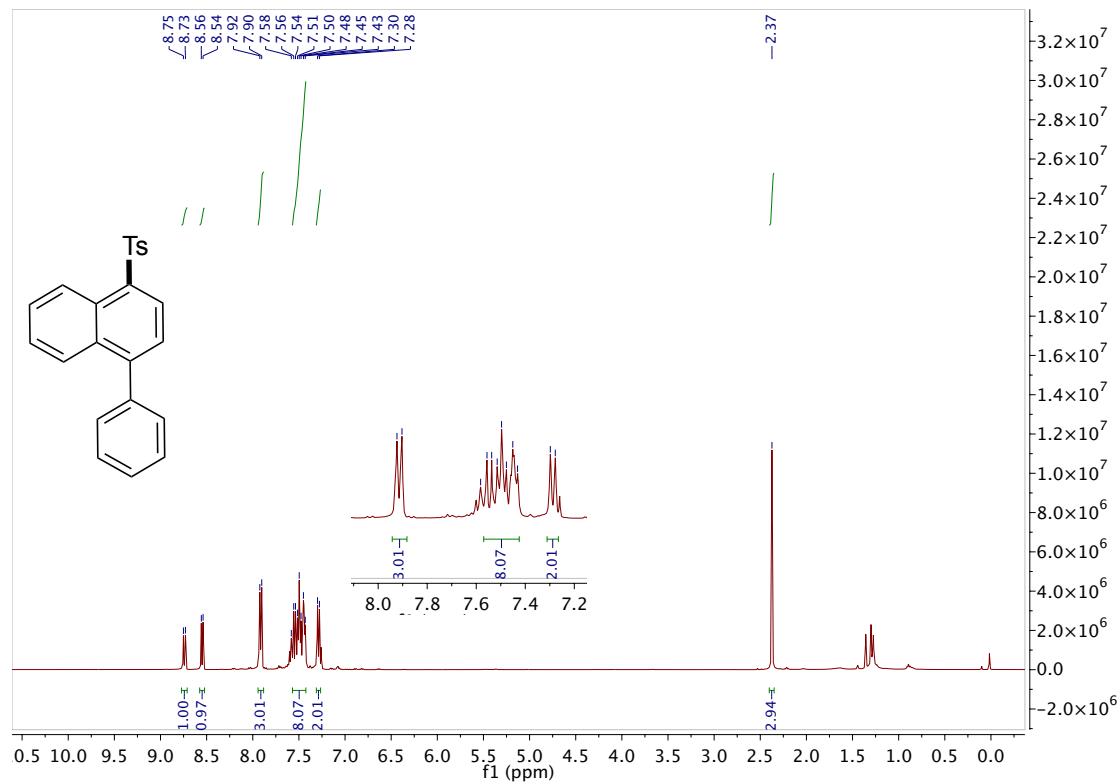
<sup>1</sup>H Spectra



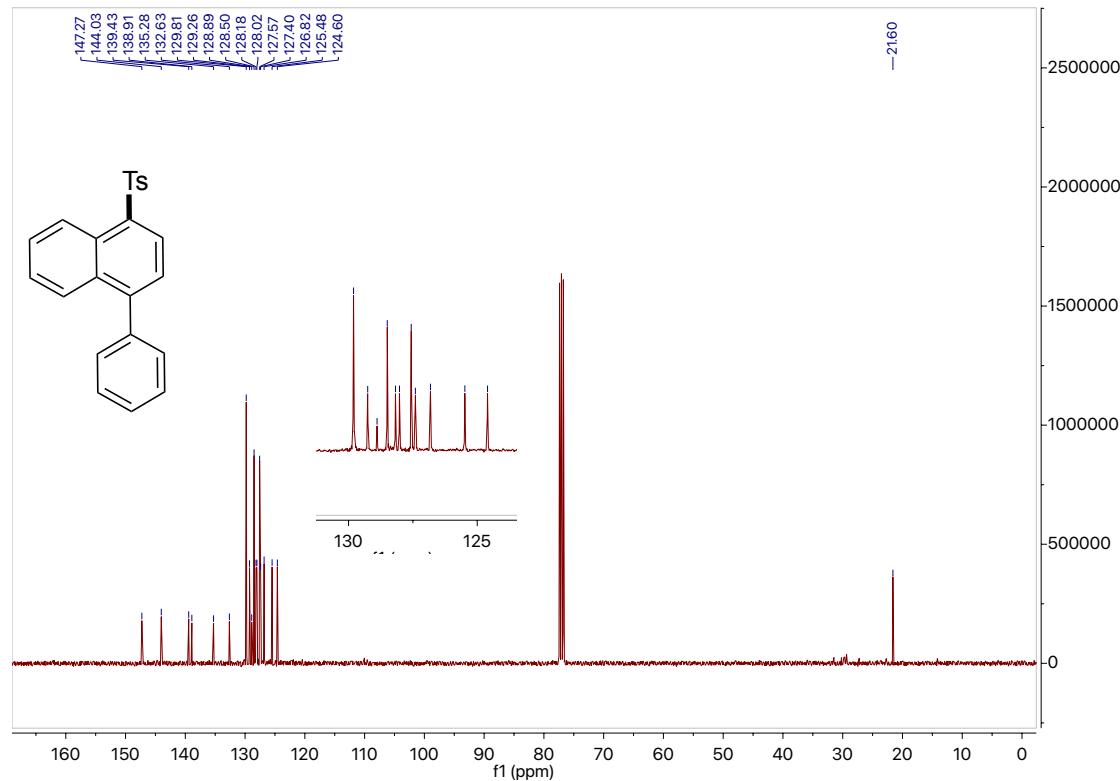
<sup>13</sup>C Spectra



**(3u)**  
 **$^1\text{H}$  Spectra**

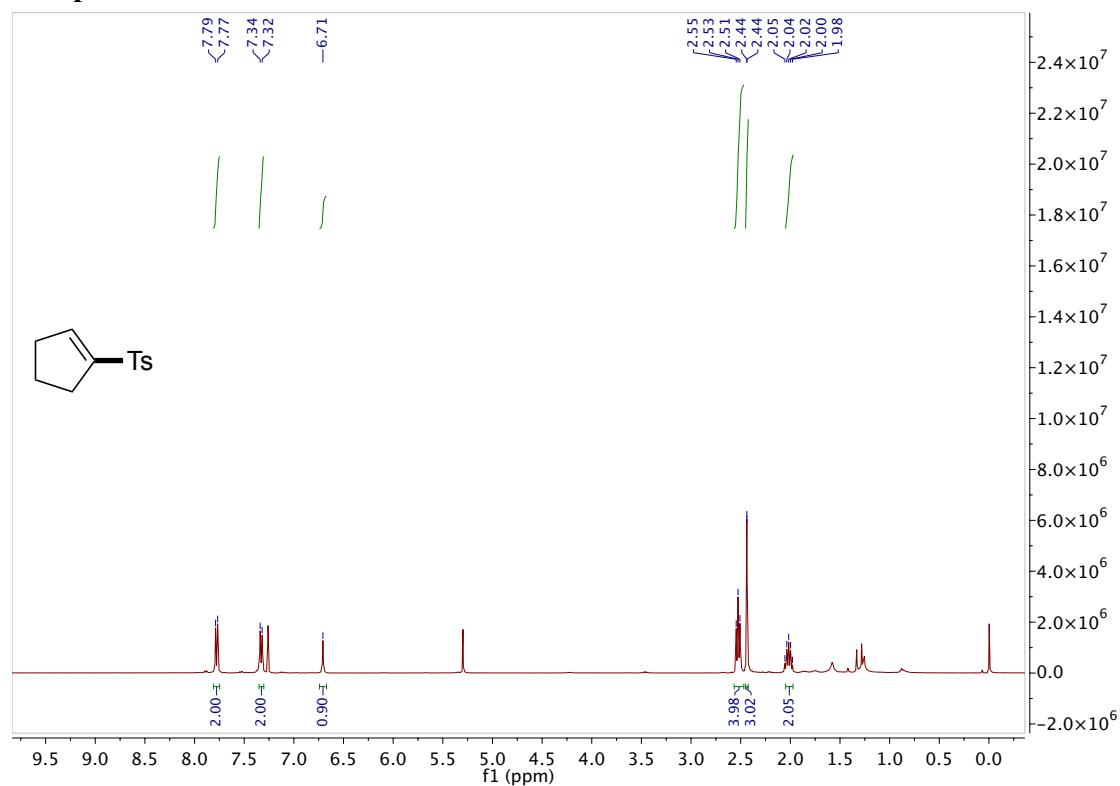


**$^{13}\text{C}$  Spectra**

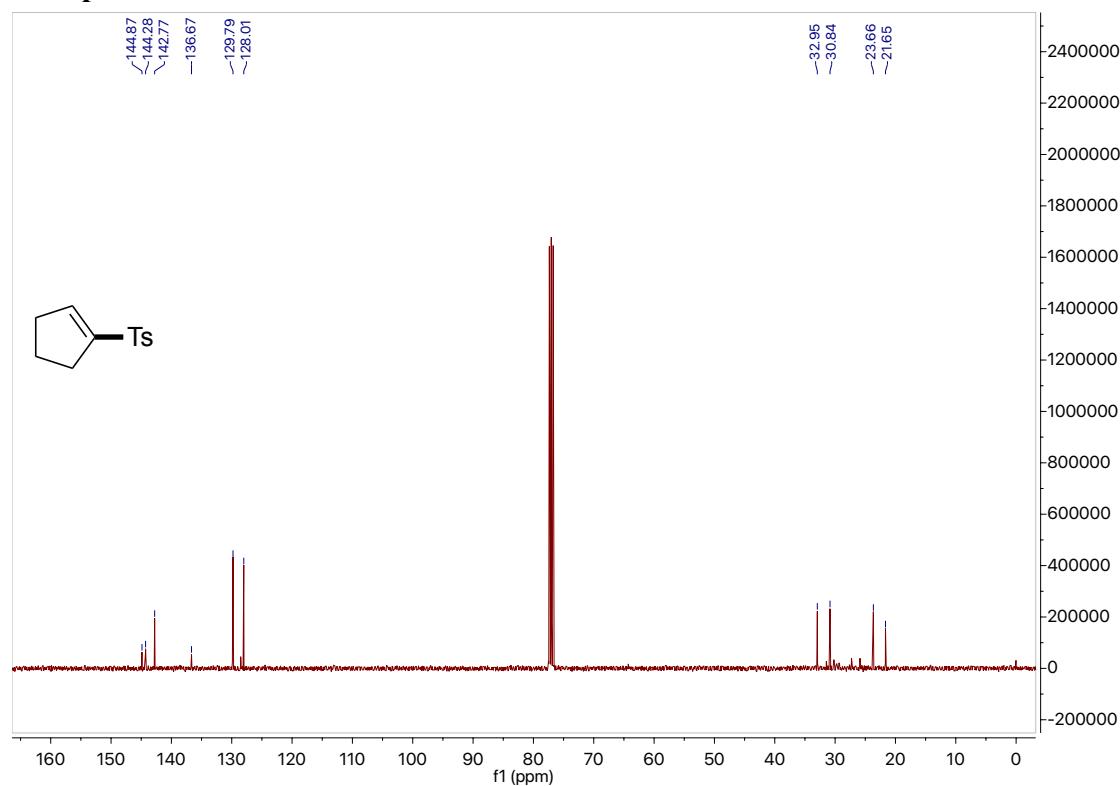


(3v)

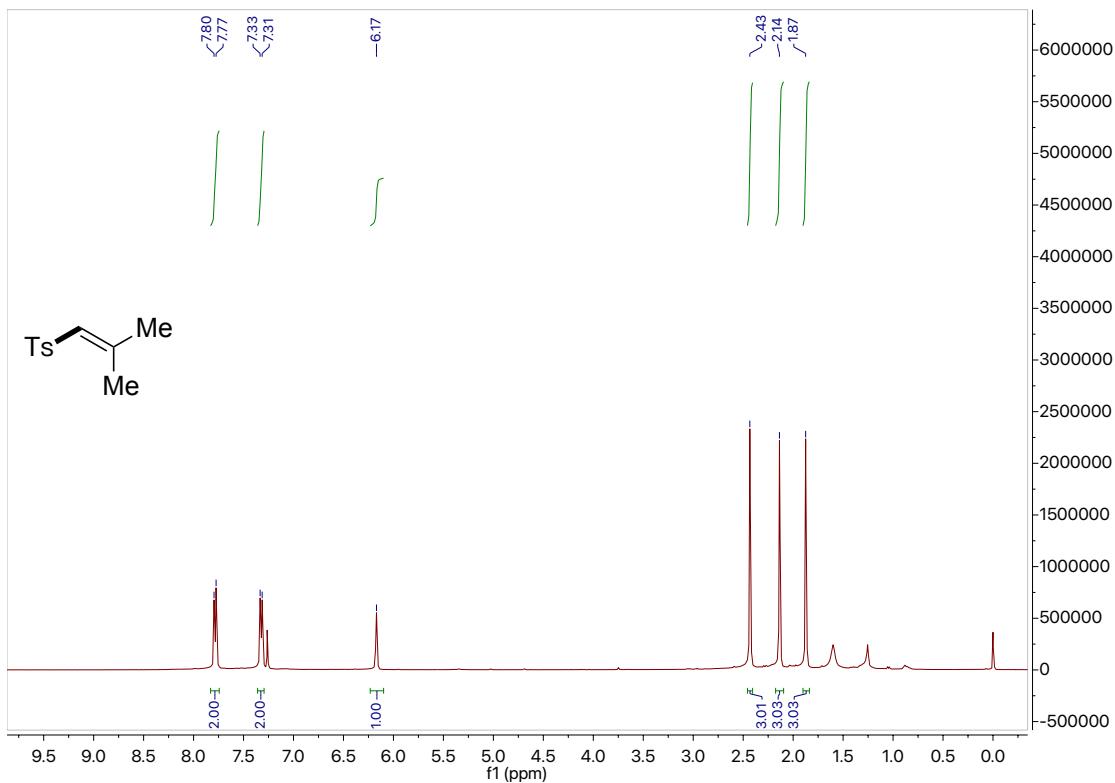
<sup>1</sup>H Spectra



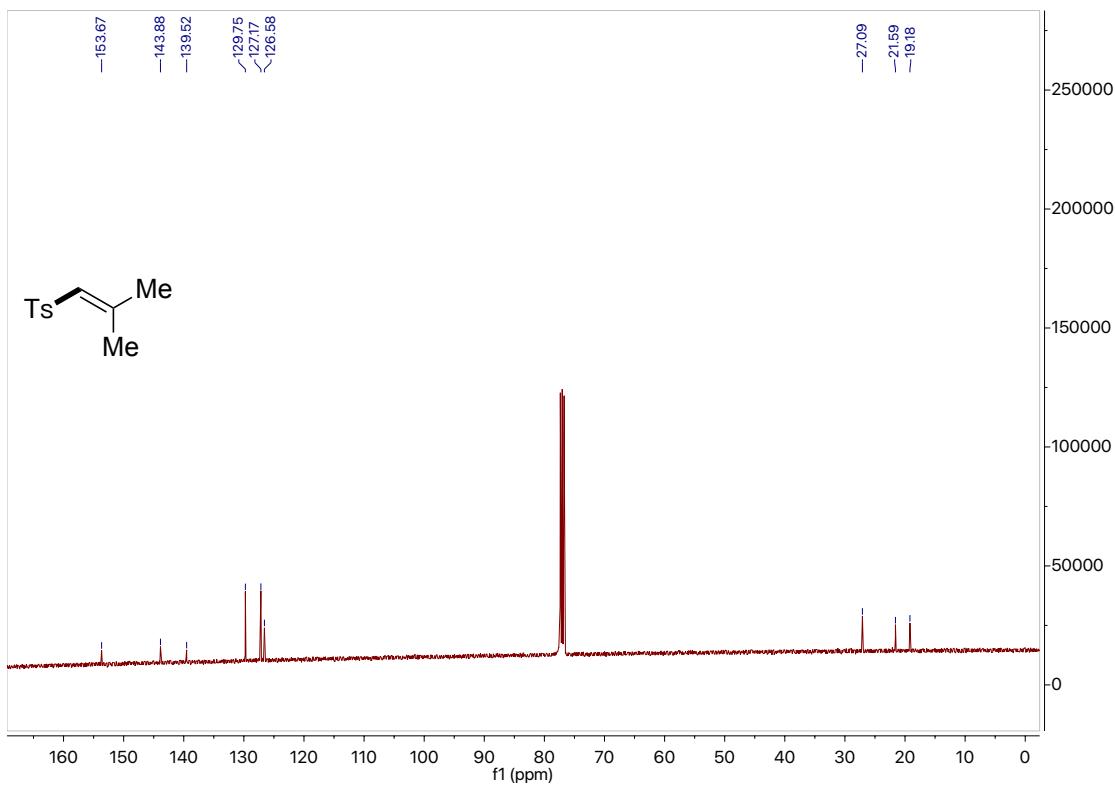
<sup>13</sup>C Spectra



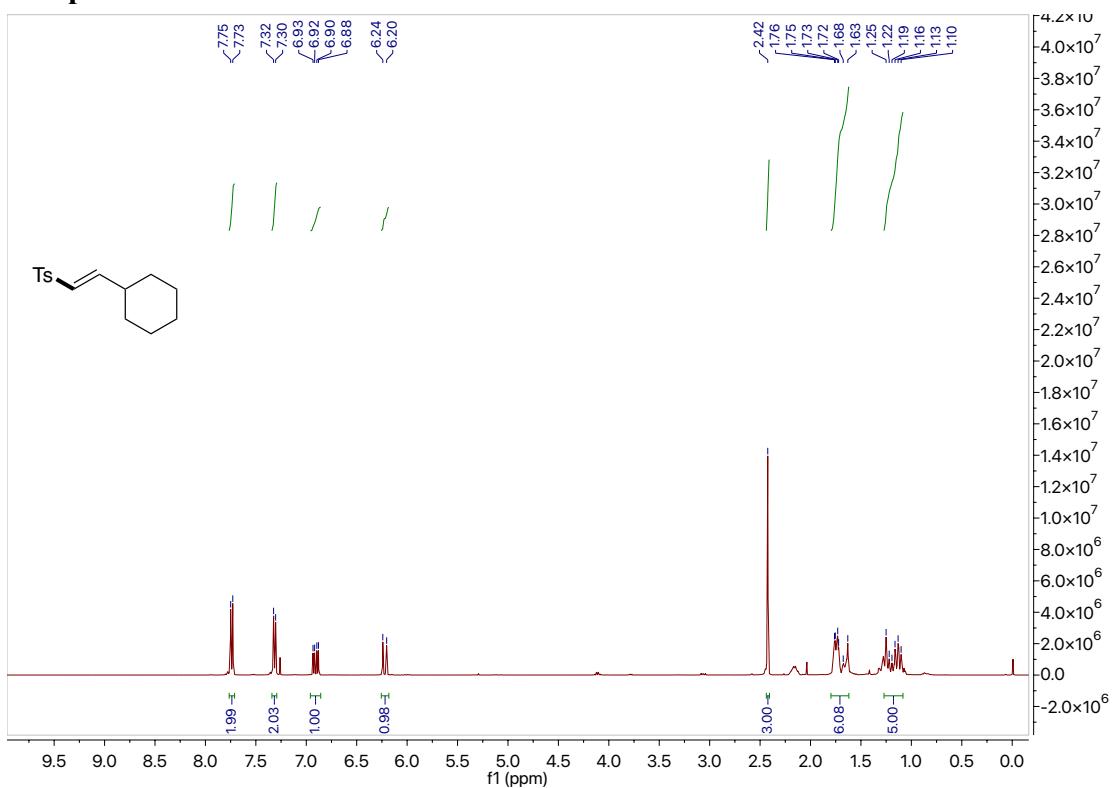
(3w)  
<sup>1</sup>H Spectra



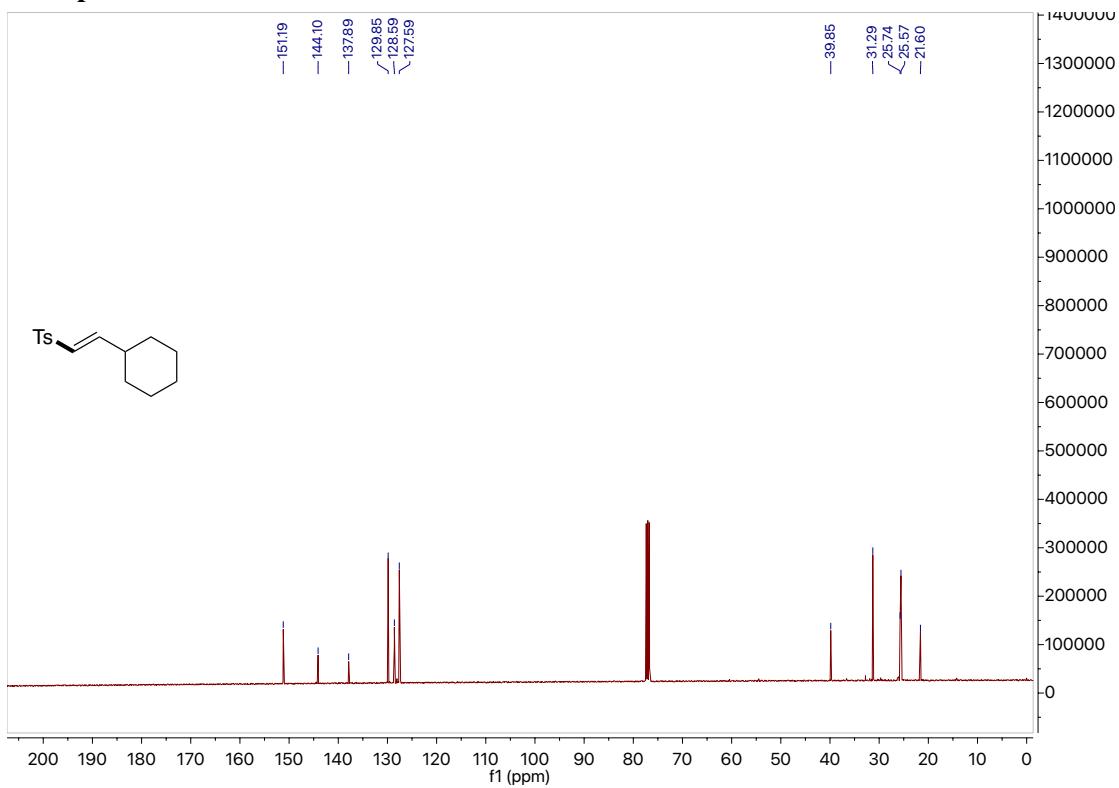
<sup>13</sup>C Spectra



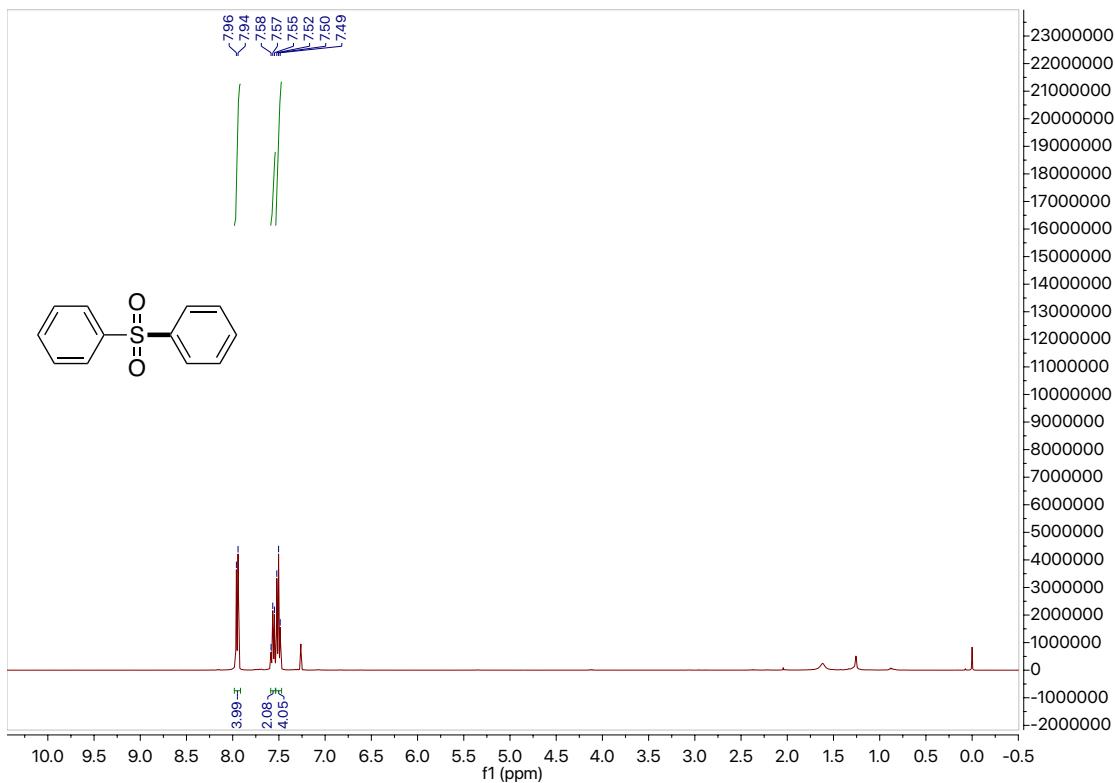
(3x)  
<sup>1</sup>H Spectra



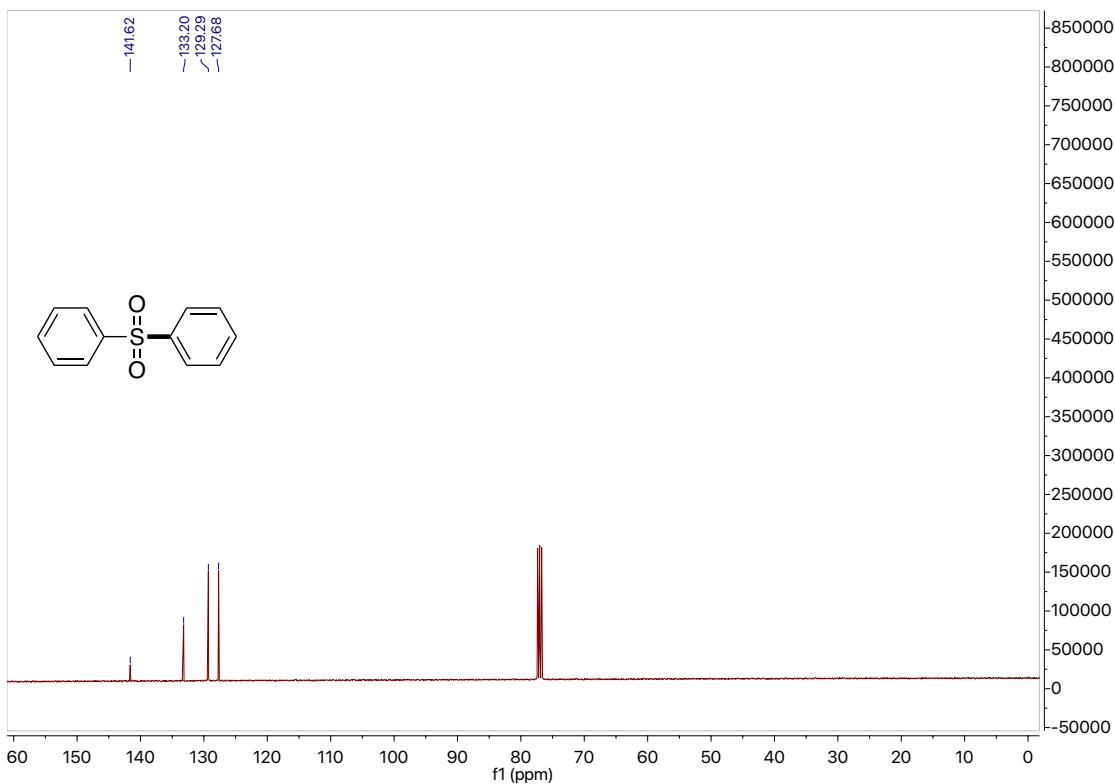
<sup>13</sup>C Spectra



(4a)  
<sup>1</sup>H Spectra

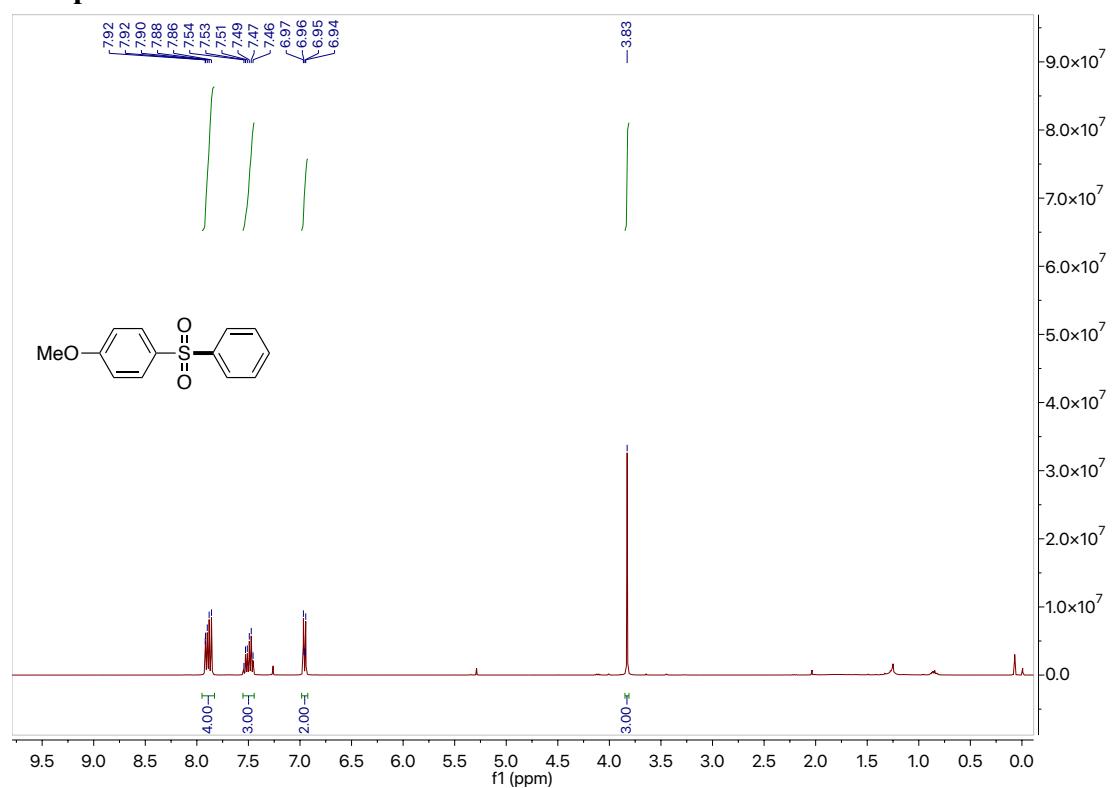


<sup>13</sup>C Spectra

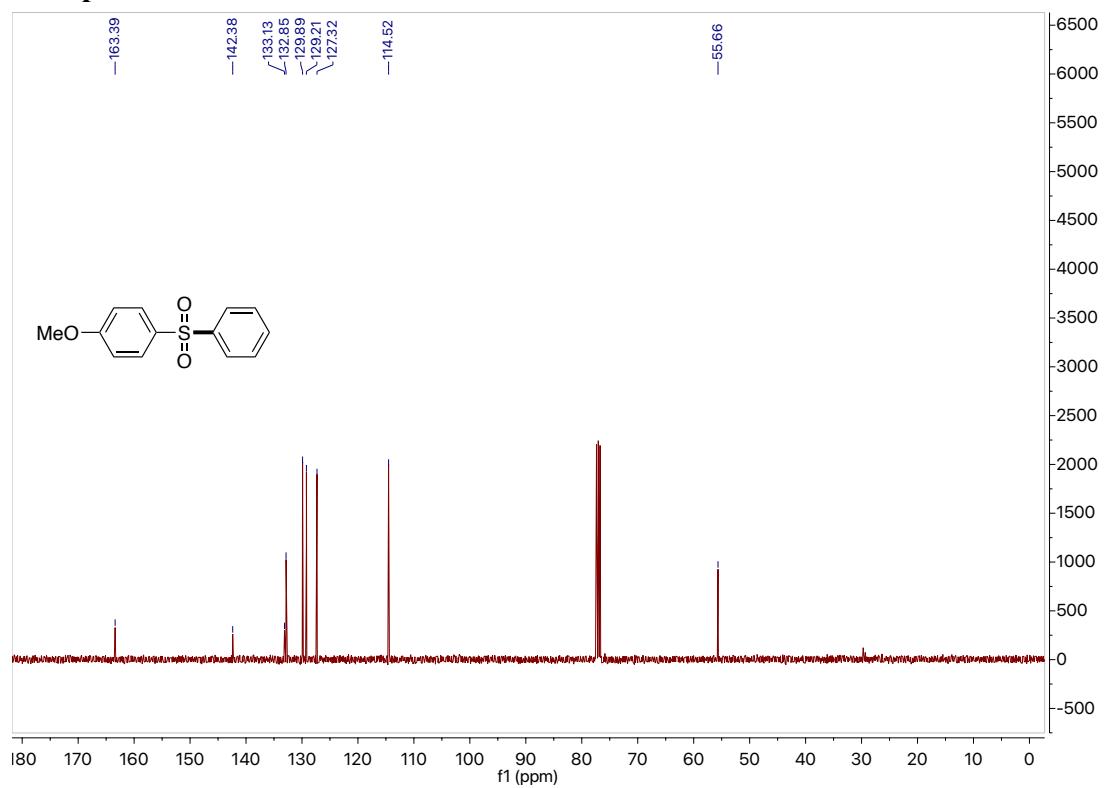


(4b)

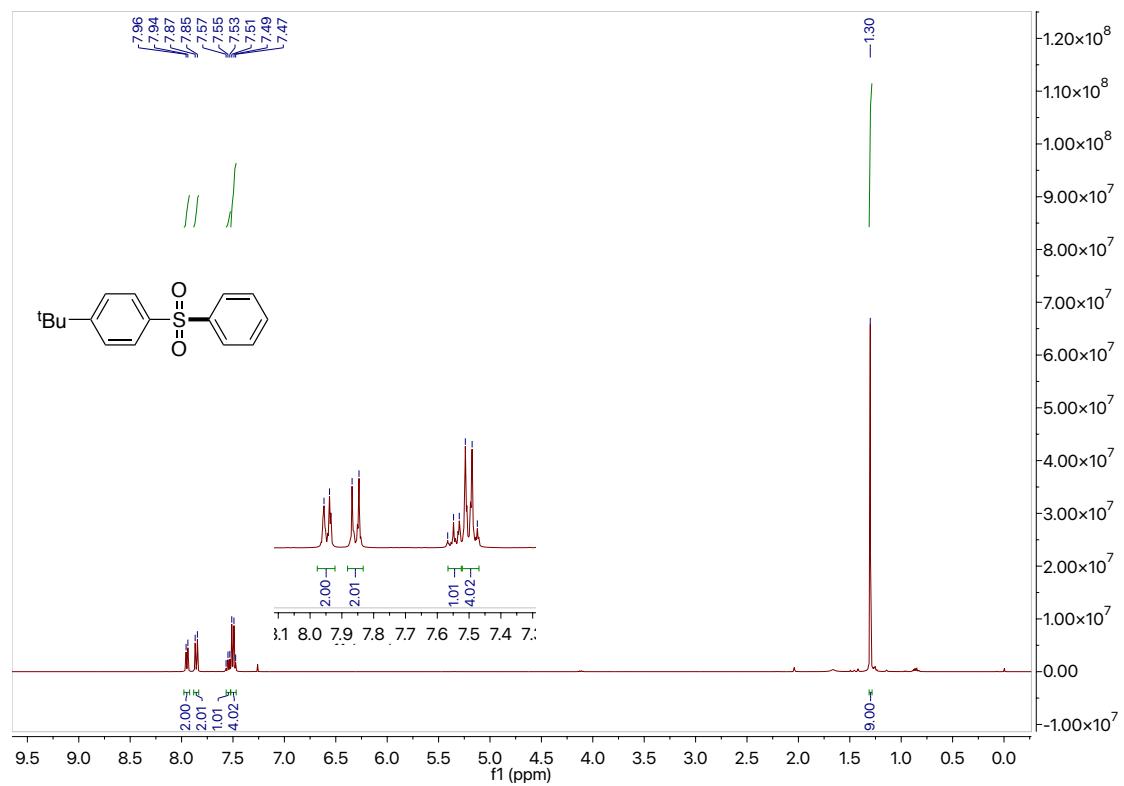
<sup>1</sup>H Spectra



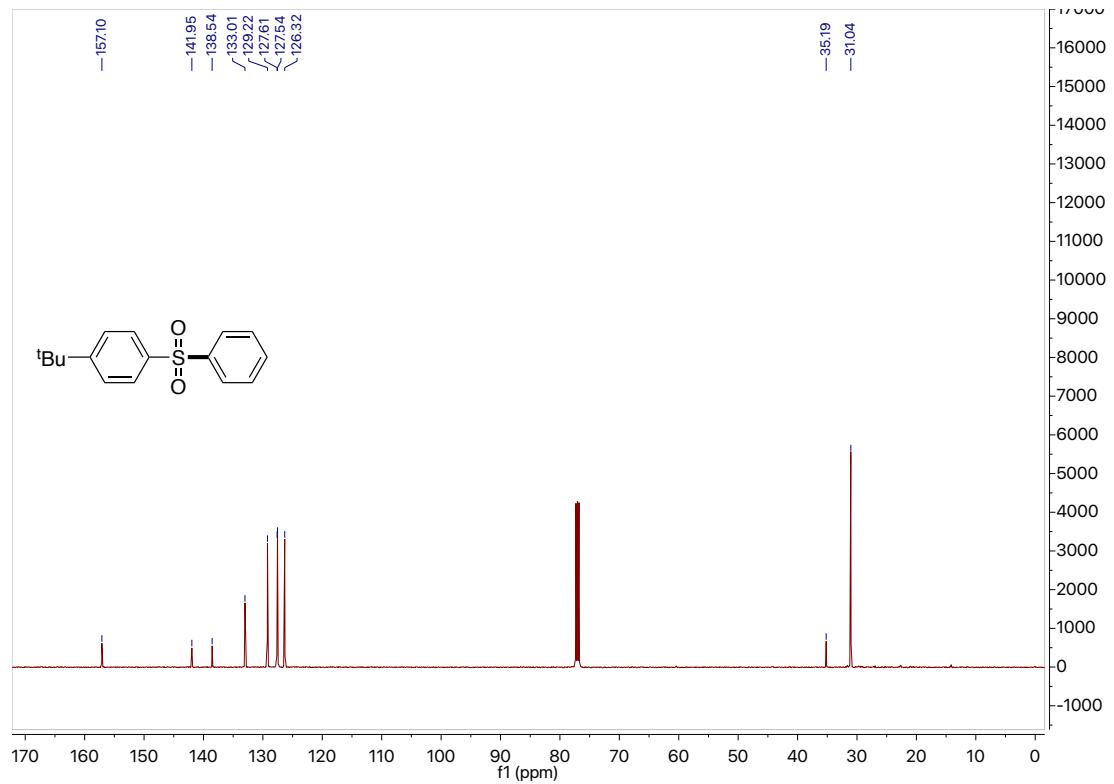
<sup>13</sup>C Spectra



**(4c)**  
 **$^1\text{H}$  Spectra**

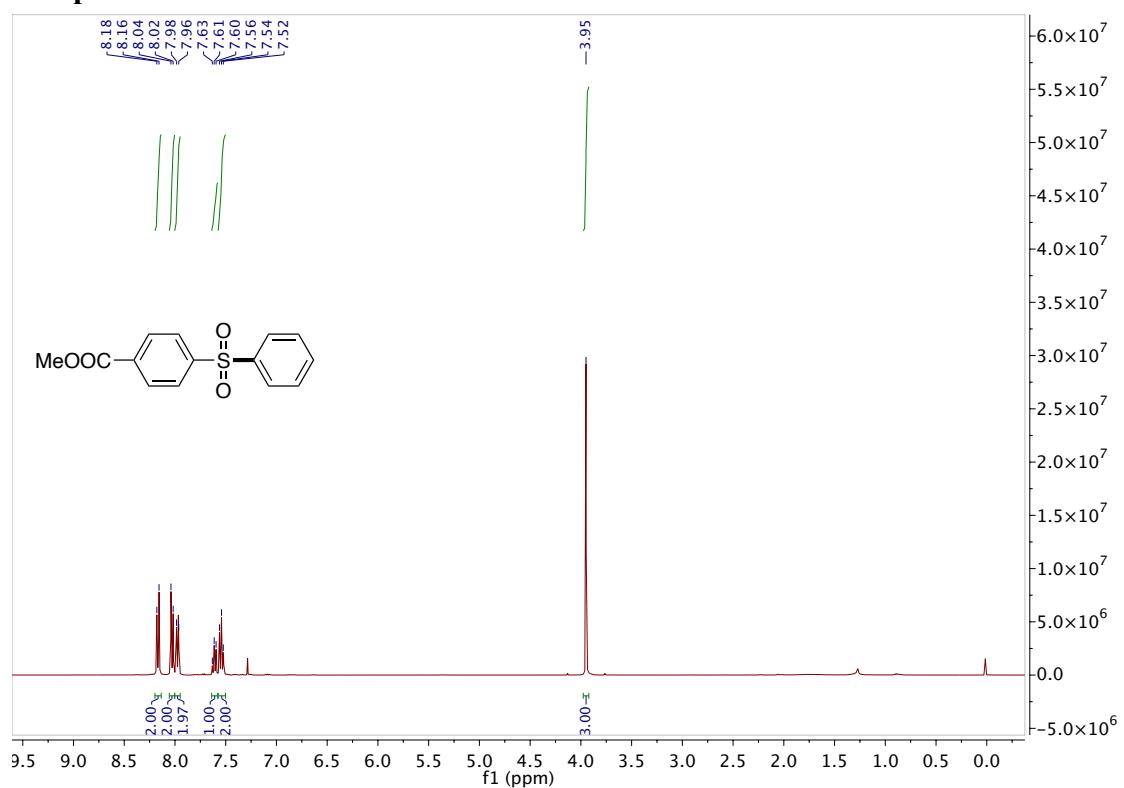


**$^{13}\text{C}$  Spectra**

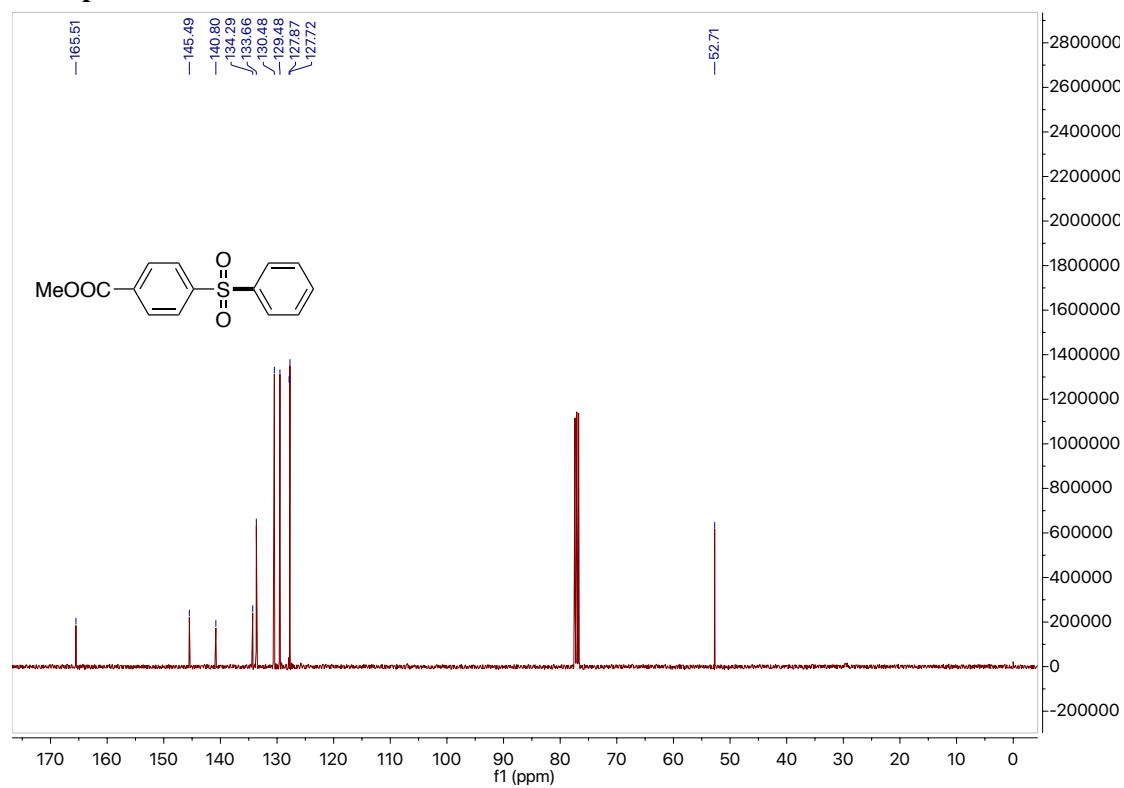


(4d)

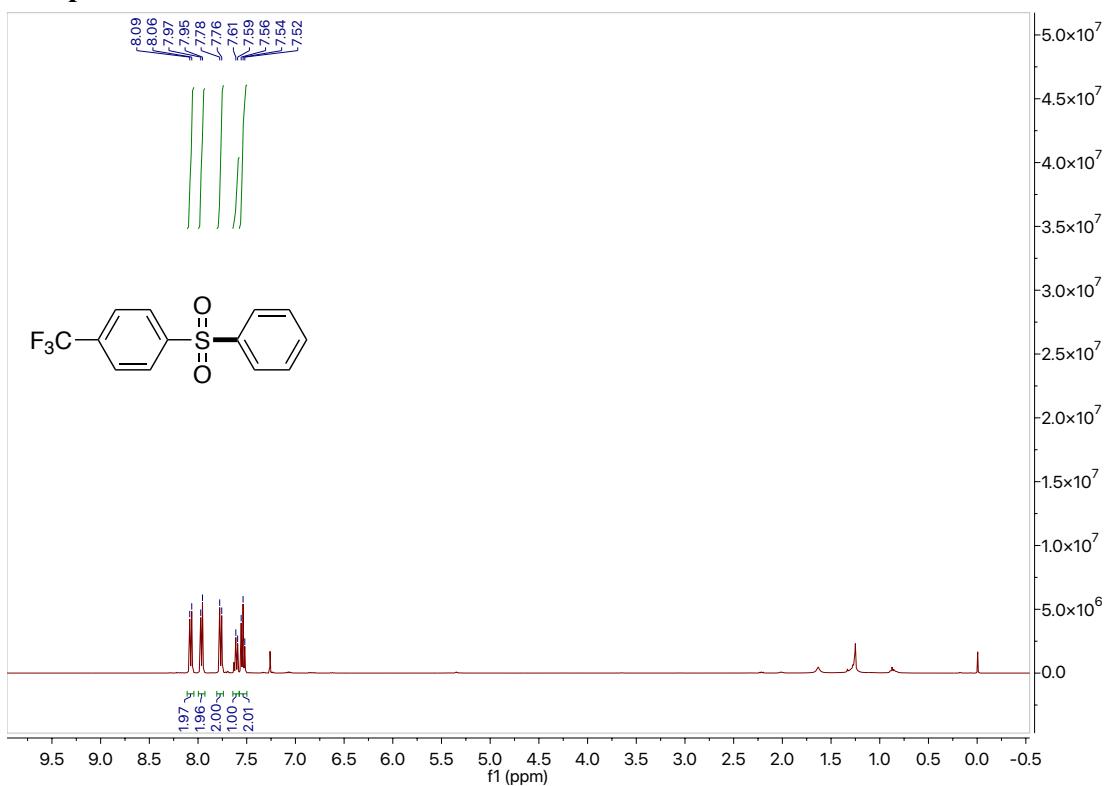
<sup>1</sup>H Spectra



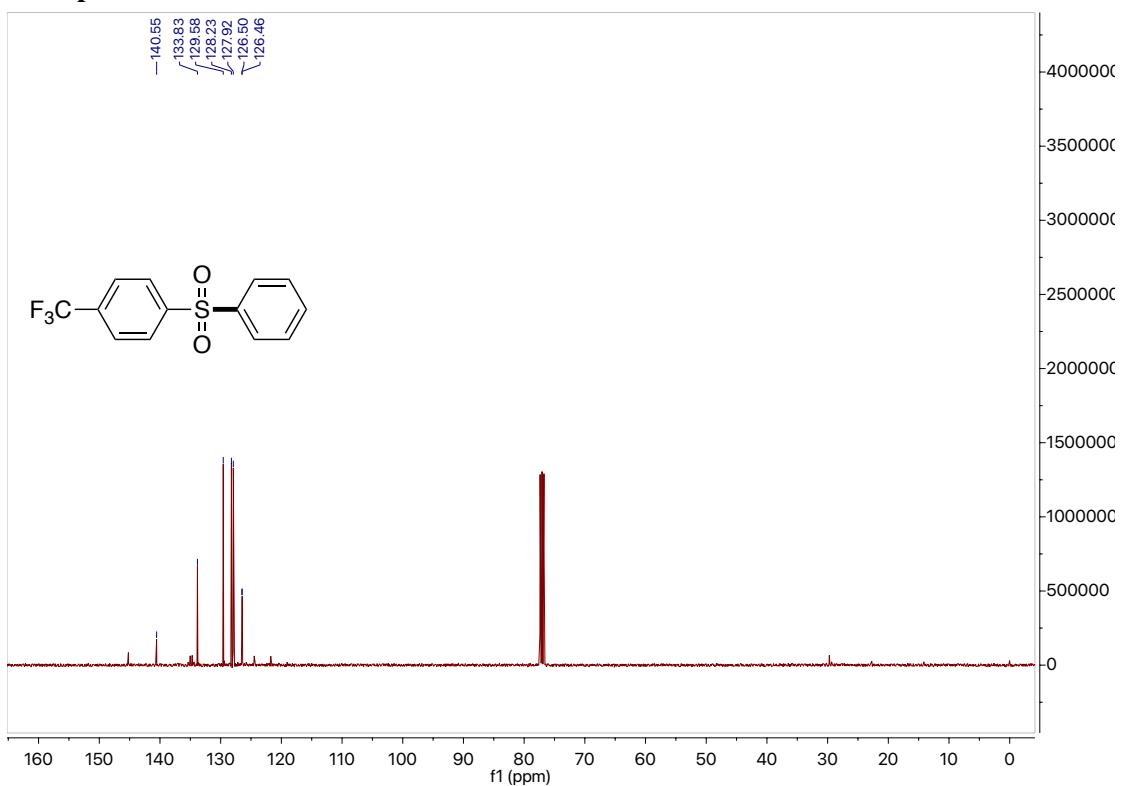
<sup>13</sup>C Spectra



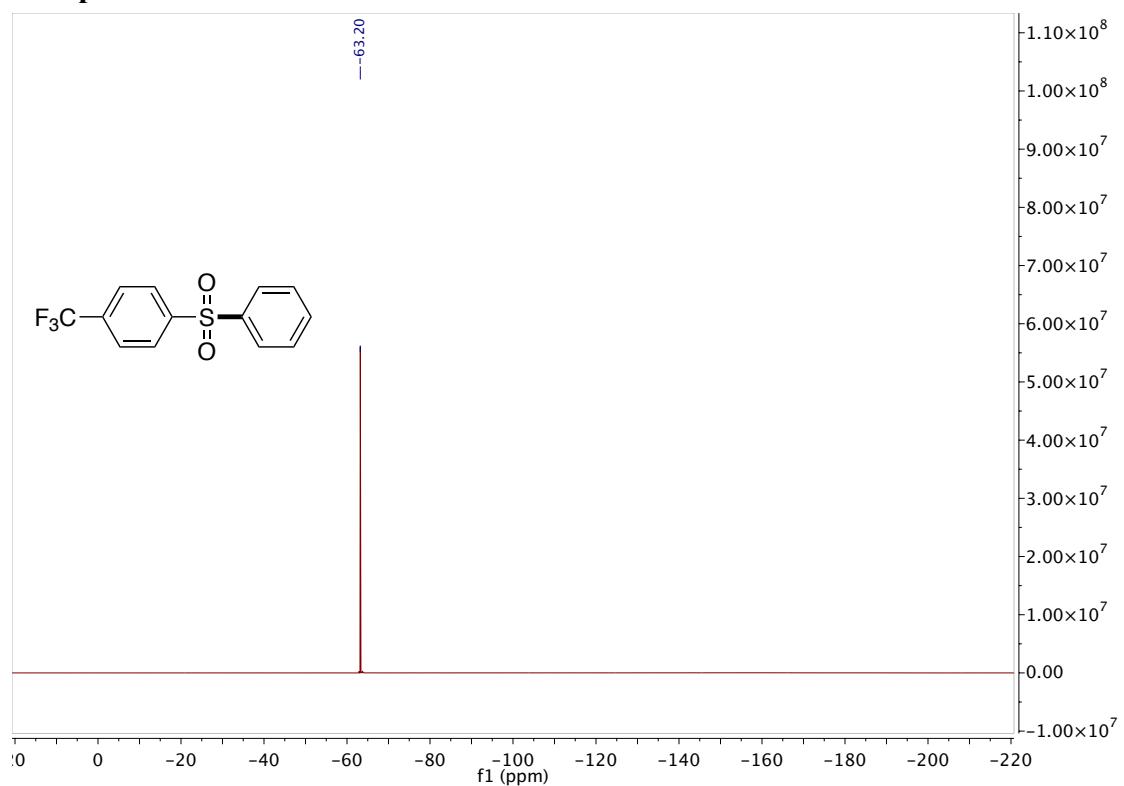
**(4e)**  
 **$^1\text{H}$  Spectra**



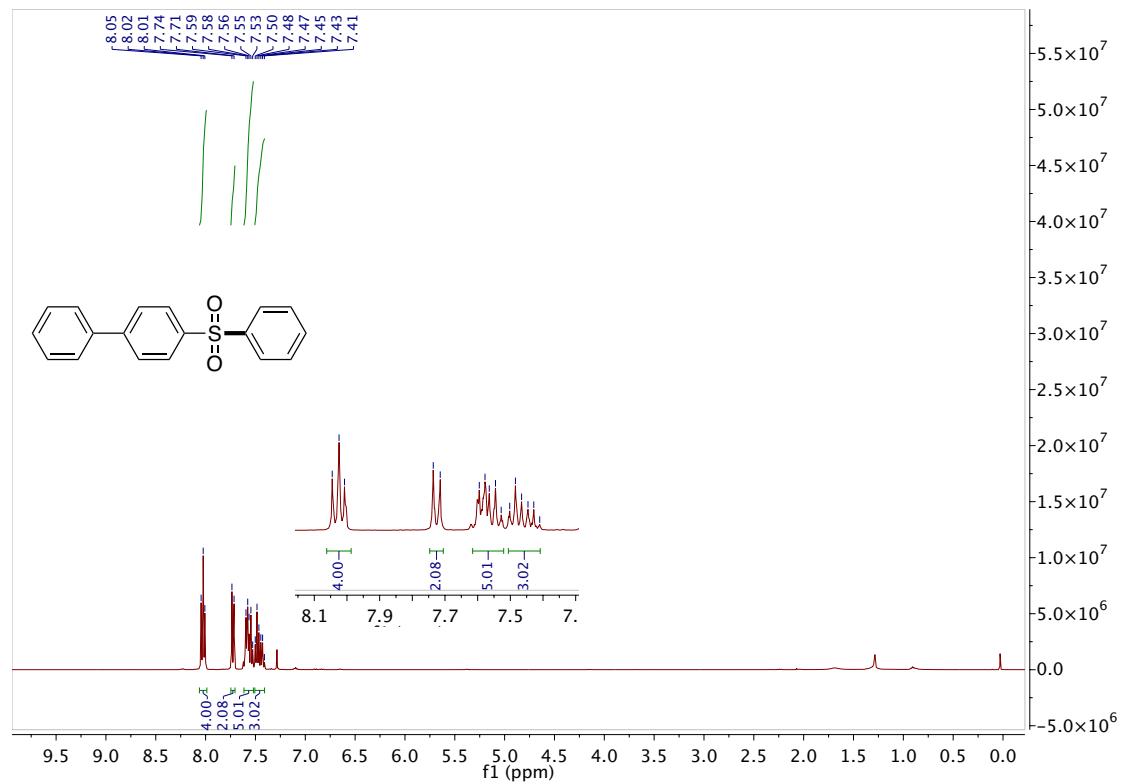
**$^{13}\text{C}$  Spectra**



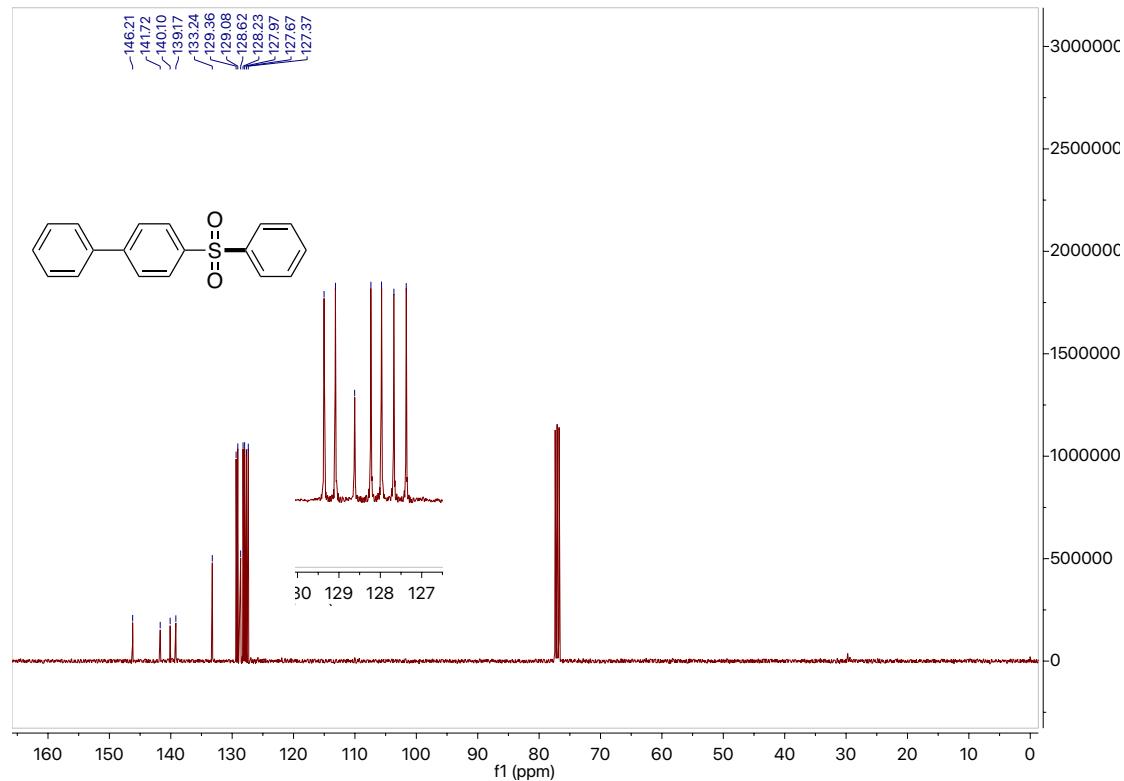
<sup>19</sup>F Spectra



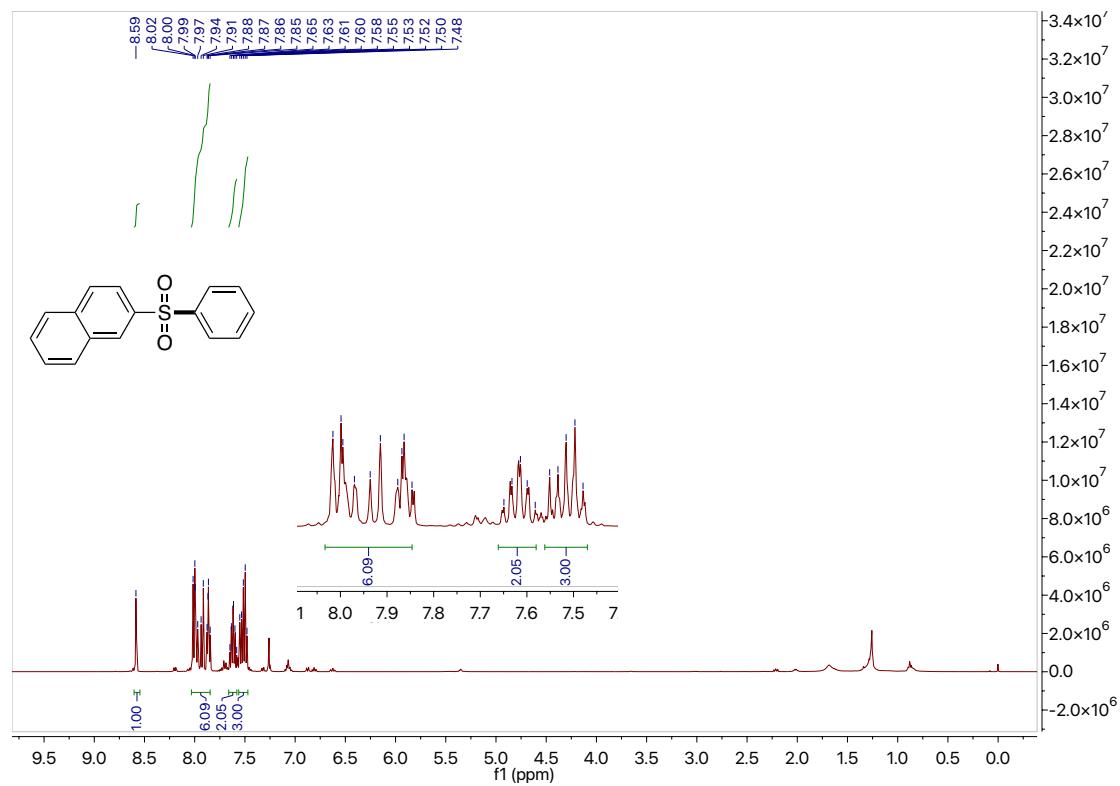
**(4f)**  
 **$^1\text{H}$  Spectra**



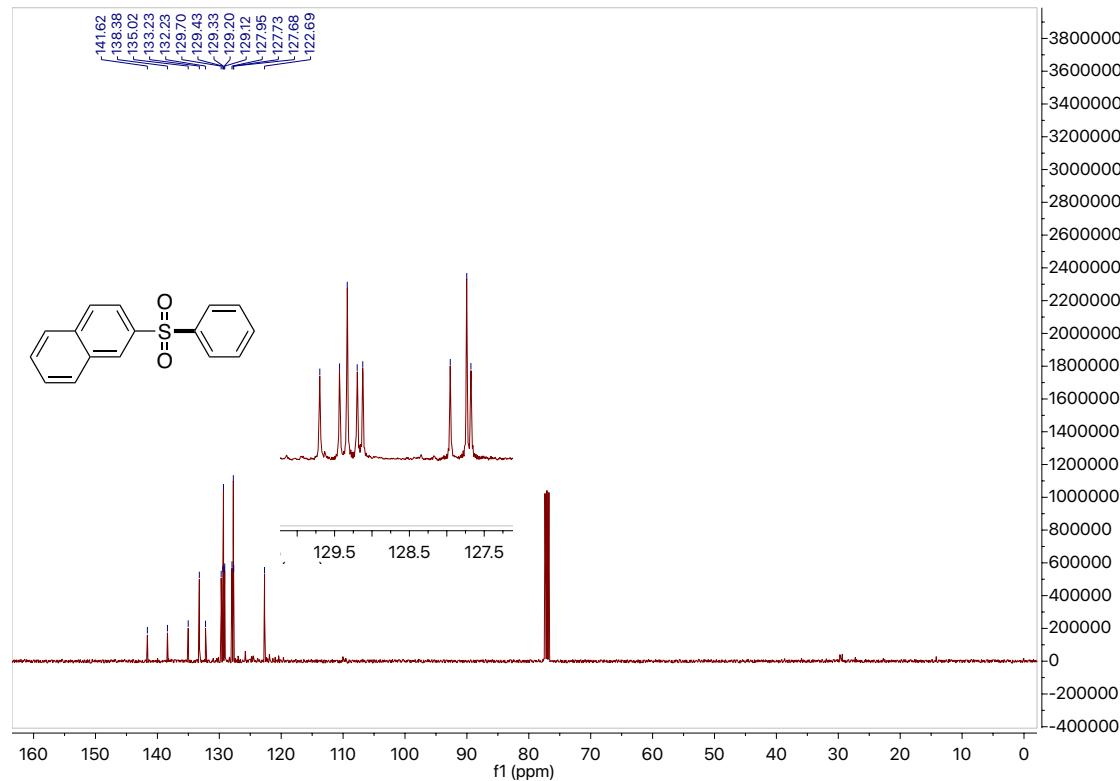
**$^{13}\text{C}$  Spectra**



**(4g)**  
 **$^1\text{H}$  Spectra**

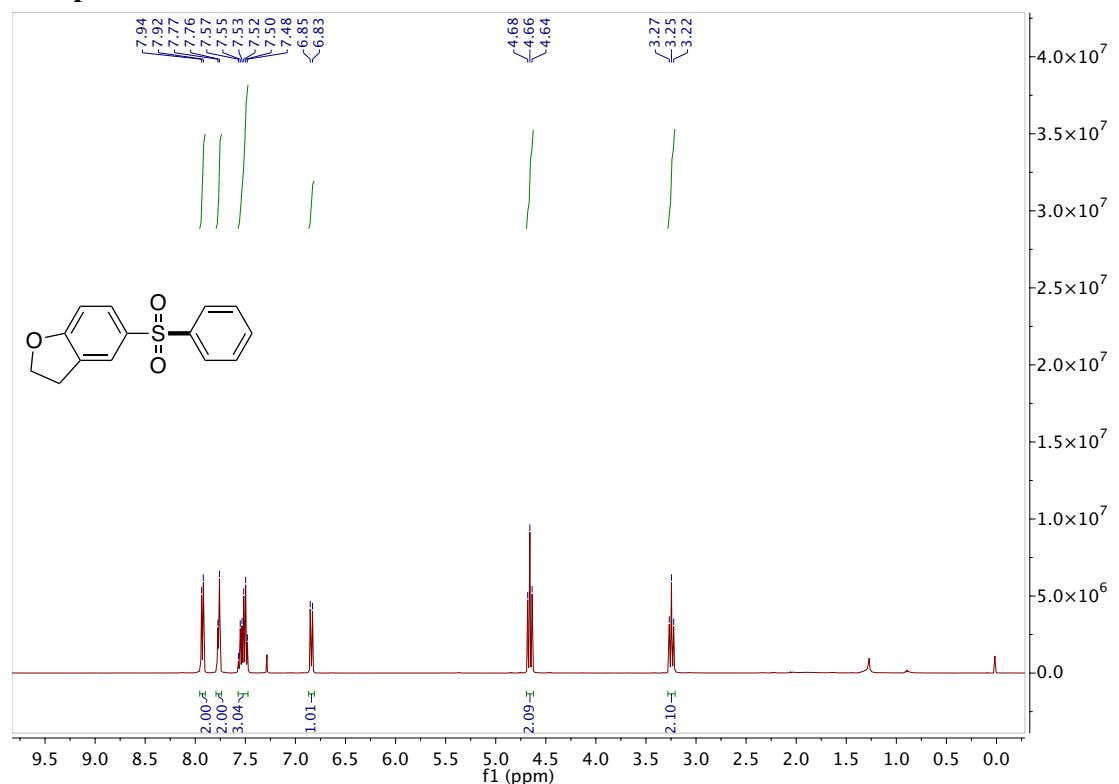


**$^{13}\text{C}$  Spectra**

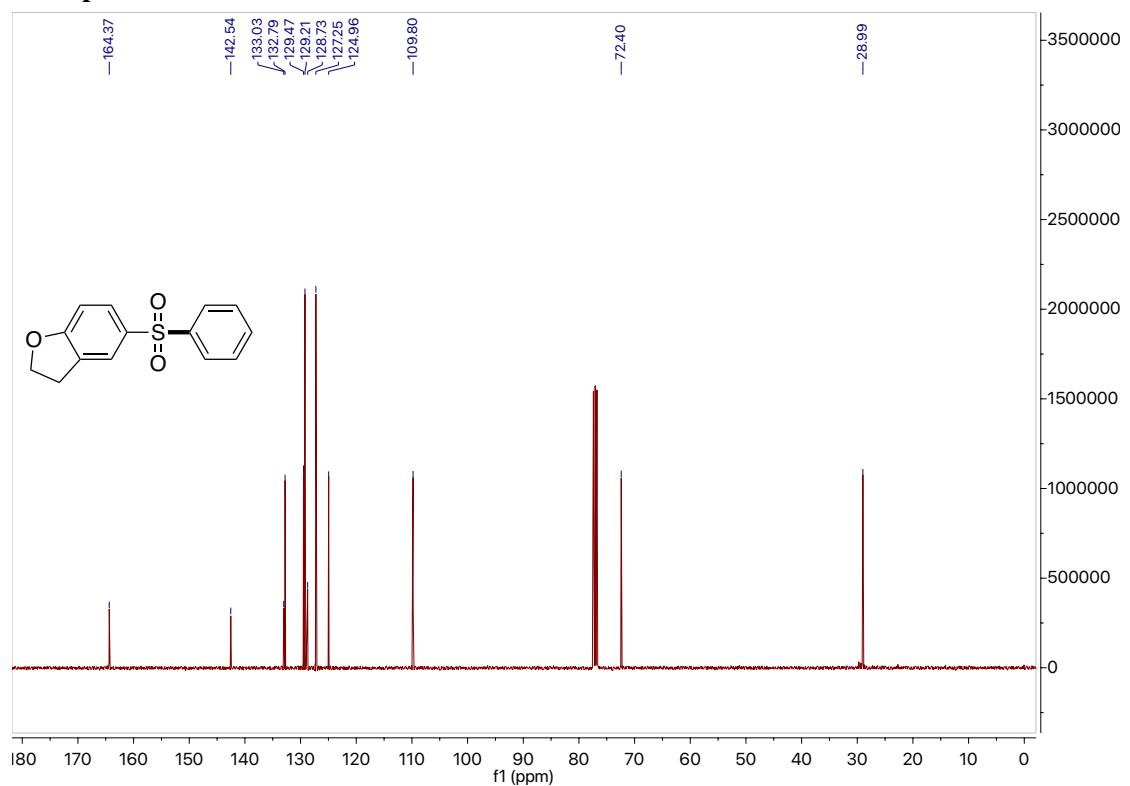


**(4h)**

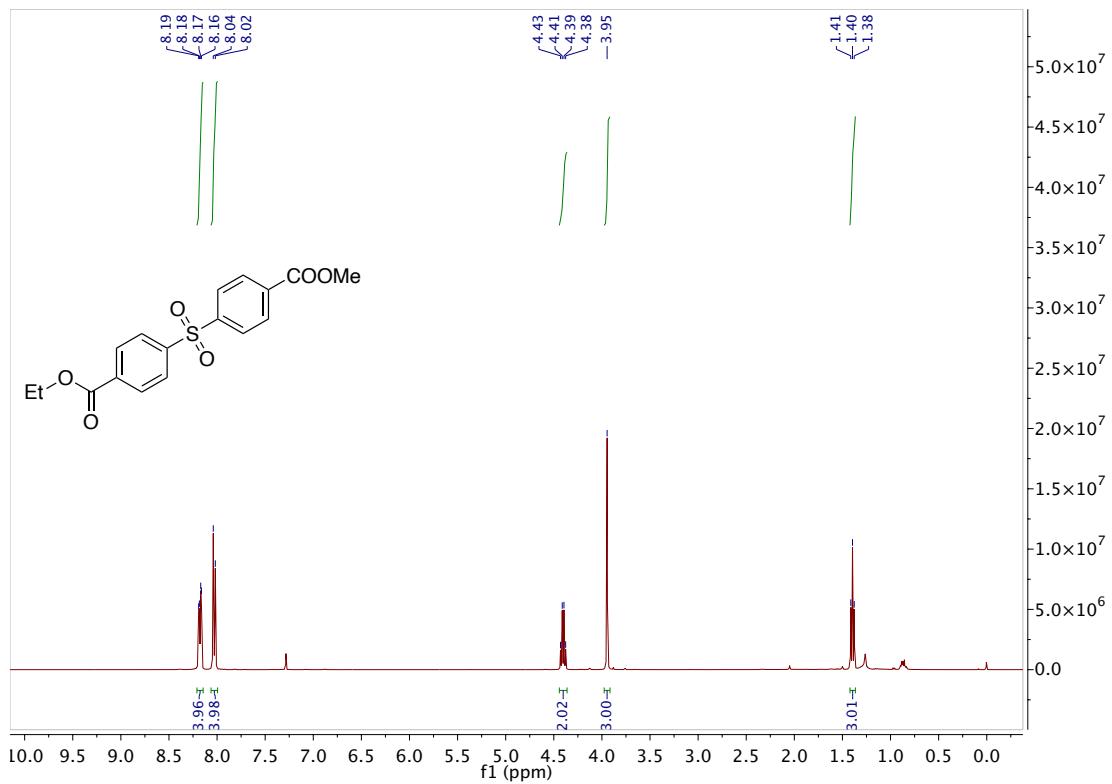
**<sup>1</sup>H Spectra**



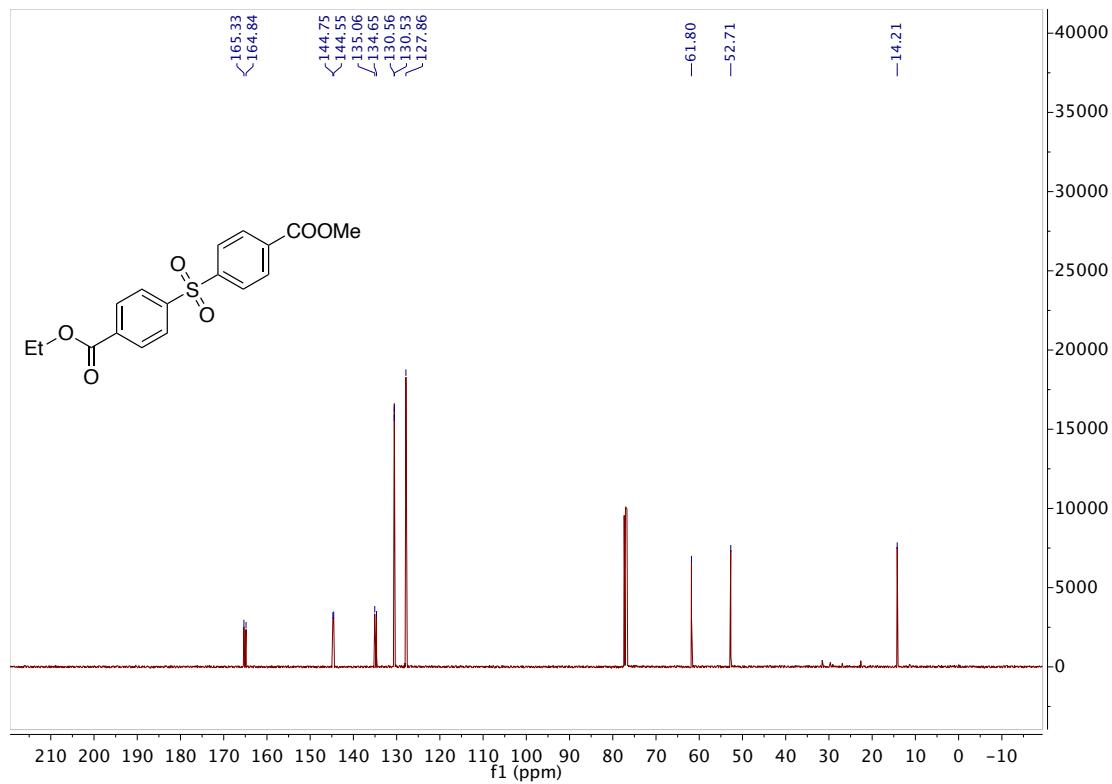
**<sup>13</sup>C Spectra**



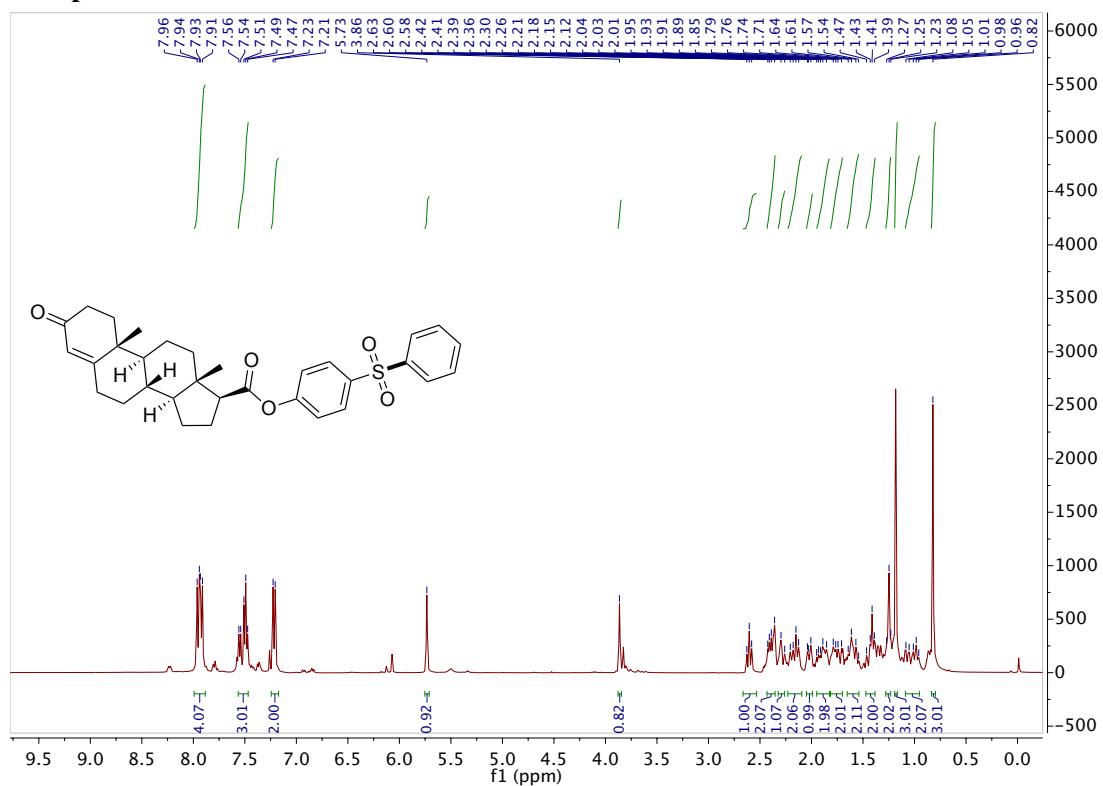
## (4j) <sup>1</sup>H Spectra



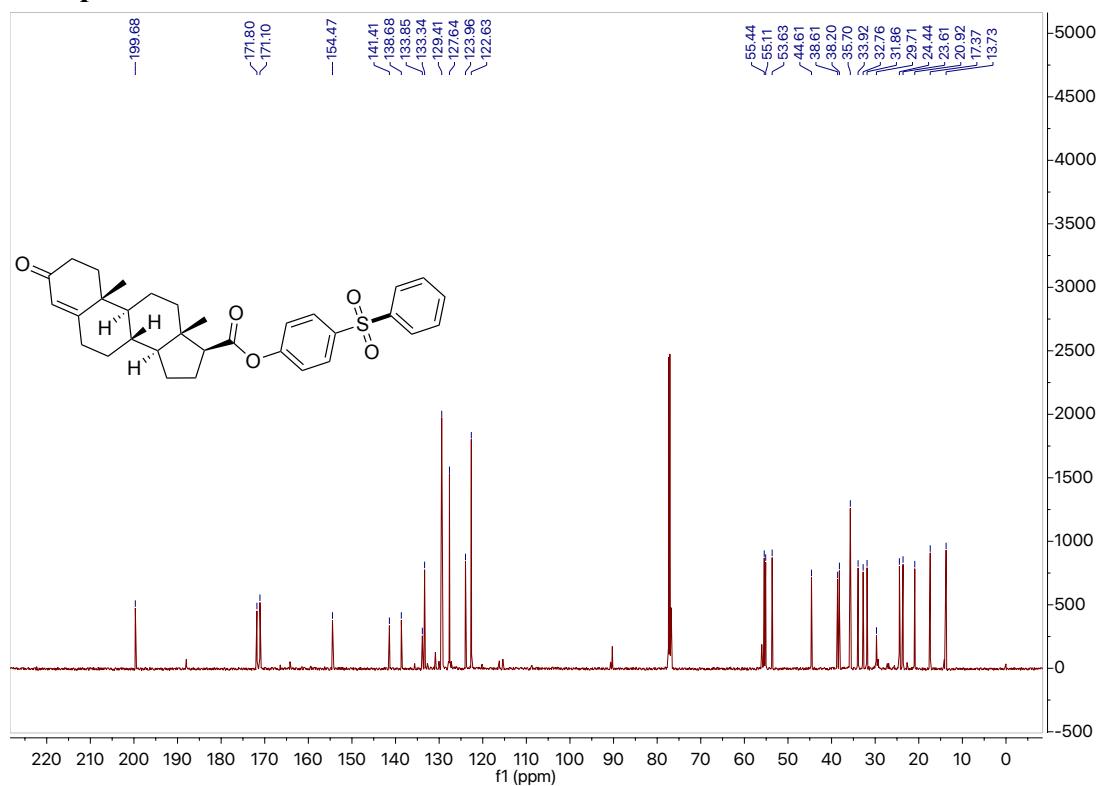
## **<sup>13</sup>C Spectra**



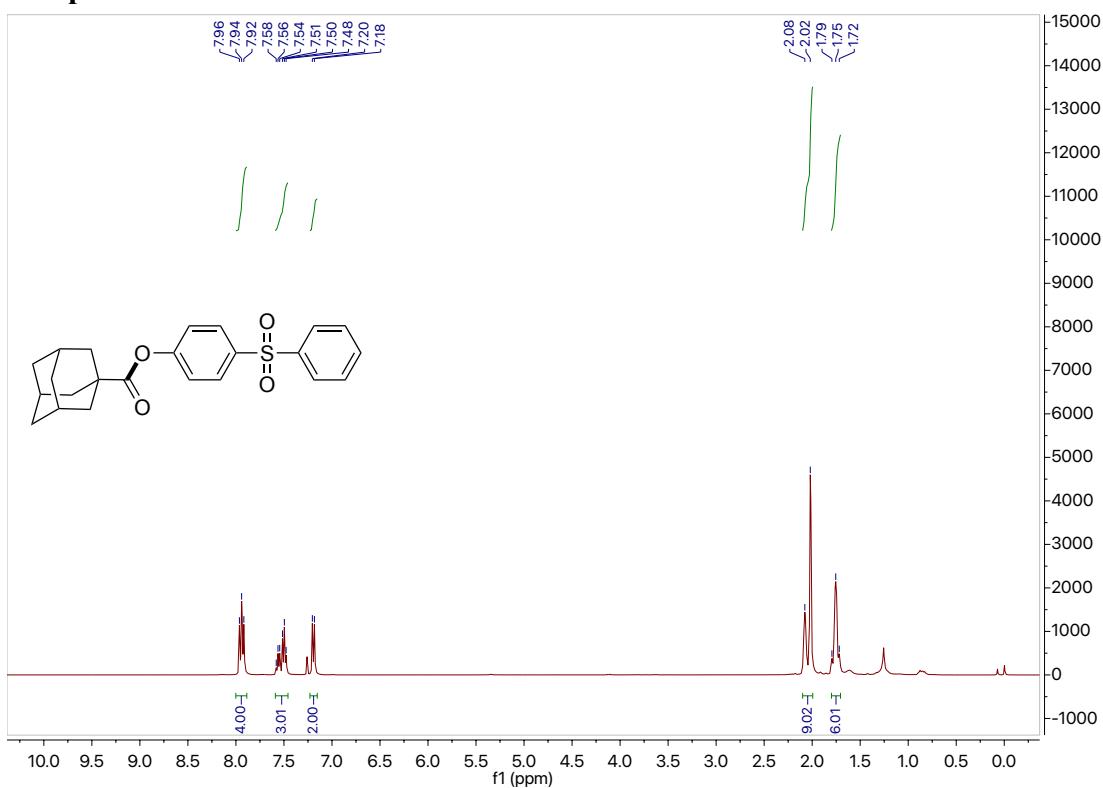
## (4k) <sup>1</sup>H Spectra



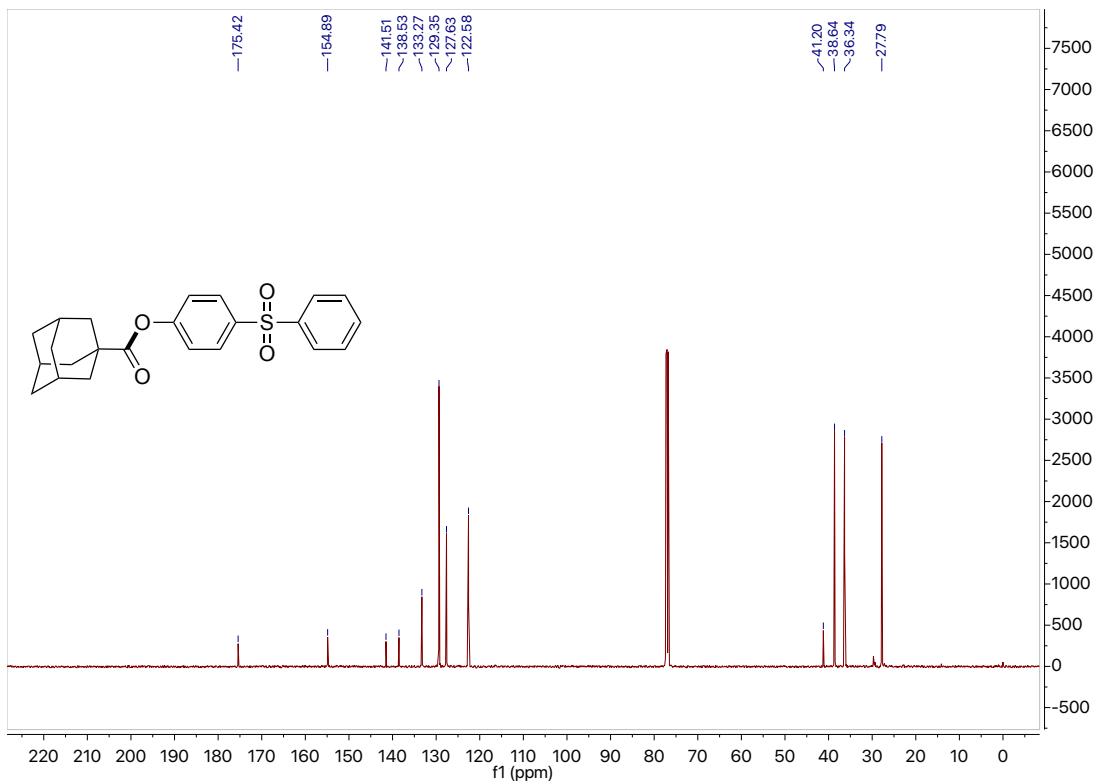
## **<sup>13</sup>C Spectra**



(4I)  
<sup>1</sup>H Spectra



<sup>13</sup>C Spectra



## 8. References

---

- [1] M.S. Lowry, J. I. Goldsmith, J. D. Slinker, R. Rohl, R. A. Pascal, G. G. Malliaras, and S. Bernhard, *Chem. Mater.*, 2005, **17**, 5712.
- [2] S. Ladouceur, D. Fortin, and E. Zsymon-Colman, *Inorg. Chem.*, 2011, **50**, 11514.
- [3] T. Rossolini, J. A. Leitch, R. Grainger, and D. J. Dixon, *Org. Lett.*, 2018, **20**, **21**, 6794-6798.
- [4] D. C. Reeves, S. Rodriguez, H. Lee, N. Haddad, D. Krishnamurthy, and C. H. Senanayake, *Tetrahedron Lett.*, 2009, **50**, 2870-2873.
- [5] K. Aboutayab, S. Caddick, K. Jenkins, S. Joshi, and S. Khan, *Tetrahedron*, 1996, **52**, 11329-11340.
- [6] J. S. Zhen, X. Du, X. H. Xu, Y. H. Li, H. Yuan, D. Xu, C. Xue and Y. Luo, *ACS Catal.*, 2022, **12**, 1986-1991.
- [7] Y. Luo, H. Ding, J. S. Zhen, X. Du, X. H. Xu, H. Yuan, Y. H. Li, W. Y. Qi, B. Z. Liu, S. M. Lu, C. Xue and Q. Ding, *Chem. Sci.*, 2021, **12**, 9556-9560.
- [8] a) G. Zhang, J. G. Fu, Q. Zhao, G. S. Zhang, M. Y. Li, C. G. Feng and G. Q. Lin, *Chem. Commun.*, 2020, **56**, 4688. b) M. Ishigaki, M. Inumaru and T. Satoh, *Tetrahedron Lett.*, 2011, **52**, 5563–5566.
- [9] R. A. Doohan, J. J. Hannan and Niall W. A. Geraghty, *Org. Biomol. Chem.*, 2006, **4**, 942–952.
- [10] X. Liang, Y. F. Li, Q. Xia, L. Cheng, J. B. Guo, P. Zhang, W. H. Zhang, and Q. M. Wang, *Green Chem.*, 2021, **23**, 8865.
- [11] S. Jiang, Z. T. Zhang, D. J. Young, L. L Chai, Q. Wu, and H. X Li, *Org. Chem. Front.*, 2022, **9**, 1437-1444.