Anion Effect in Enantioselective Oxidative NHC Catalysis: Highly Efficient Kinetic Resolution of Tertiary alcohols

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Supporting Information

Contents	
1. General Information	S3
2. Synthesis of azolium chloride	S5
3. General procedure for the anion exchange reaction	S6
4. General procedure for the synthesis of N-benzyl Isatin derivations	S8
5. General procedure A for the synthesis of 3-Hydroxy-3-Substituted Oxindoles	S9
6. General Procedures for Kinetic Resolution of 3-Hydroxy-3-Substituted Oxindoles	s S10
7. General procedure B for the synthesis of 3-Hydroxy-3-Substituted Oxindoles	S11
8. Table 1. Screening different additives for kinetic resolution of substate 1e	S13
9. General Procedures B for Kinetic Resolution of 3-Hydroxy-3-Subst Oxindoles	ituted S14
10. Characterization of compounds.	S15
11. Reference	S 31
12. NMR Spectrum	S32

1. General Information

¹H and ¹³C NMR spectra were recorded on a Bruker ACF300 (300 MHz) or AMX500 (500 MHz) spectrometer. Chemical shifts were reported in parts per million (ppm), and the residual solvent peak was used as an internal reference: proton (chloroform δ 7.26, acetone δ 2.05, dimethyl sulfoxide δ 2.50), carbon (chloroform δ 77.0, acetone δ 29.84, dimethyl sulfoxide δ 39.6) or tetramethylsilane (TMS δ 0.00) was used as a reference. Multiplicity was indicated as follows: s (singlet), d (doublet), t (triplet), q (quartet), m (multiplet), dd (doublet of doublet), bs (broad singlet). Coupling constants were reported in Hertz (Hz). Low resolution mass spectra were obtained on a Finnigan/MAT LCQ spectrometer in ESI mode, and a Finnigan/MAT 95XL-T mass spectrometer in EI mode. All high resolution mass spectra (HRMS) were obtained on a Finnigan/MAT 95XL-T spectrometer. For thin layer chromatography (TLC), Merck pre-coated TLC plates (Merck 60 F254) were used, and compounds were visualized with a UV light at 254 nm. Further visualization was achieved by staining with iodine, or potassium permanganate solution followed by heating using a heat gun. Flash chromatography separations were performed on Merck 60 (0.040-0.063 mm) mesh silica gel. Optical rotations were measured using an Anton Paar MCP-150 digital polarimeter using a 1 cm glass cell. The enantiomeric excess (ee) of products were determined by chiral phase HPLC analysis on on ThermoFisher HPLC units, or Agilent HPLC units, including the following instruments: pump, LPG-3400SDN; detector, VWD-3400RS; column, Chiralcel OD-H, Chiralpak AS-H, IA, or IC.

All reactions were carried out under nitrogen atmosphere. All commercially available reagents were used as received for the reactions without any purification. All solvents were dried on alumina columns using a solvent dispensing system. Sodium hydride, benzyl bromide, cinnamaldehyde, indole-3-propionic acid, 5-methoxyisatin, isatin, tetrabutyl ammonium

bromide, triethyl phosphite, malonic acid, pyridine, thionyl chloride were purchased from Aldrich or Fluka and used directly.

2. Synthesis of the azolium chloride



The synthesis is an adaption of literature procedures^{1,2} from Rovis procedure. Methylation of (4aS,9aR)-4,4a,9,9a-tetrahydroindeno[2,1-b][1,4]oxazin-3(2H)-one with Meerwein's reagent, followed by liberation of the free base by treatment with sodium bicarbonate gave imino ether. Reaction of the imino ether with an equimolar amount of Pentafluorophenylhydrazine hydrochloride and a catalytic amount of anhydrous hydrochloric acid in methanol afforded amidrazone hydrochloride. Treatment of amidrazone with excess trimethylorthoformate in MeOH at 80 °C for 36 hours provided the crude product which subsequent precipitation from acetonitrile by the addition of diethyl ether. The azolium chloride was obtained as a white solid.

White solid. 45% yield, **MP:** 210 - 212 °C. ¹**H NMR** (500 MHz, DMSO) δ 11.89 (s, 1H), 7.65 (d, J = 7.5 Hz, 1H), 7.42 (dt, J = 14.7, 7.3 Hz, 2H), 7.35 (t, J = 7.2 Hz, 1H), 6.18 (d, J =3.6 Hz, 1H), 5.36 (d, J = 16.2 Hz, 1H), 5.10 (d, J = 16.2 Hz, 1H), 4.99 (t, J = 4.5 Hz, 1H), 3.49 (dd, J = 17.0, 4.9 Hz, 1H), 3.17 (d, J = 17.1 Hz, 1H). ¹³**C NMR** (126 MHz, DMSO) δ 151.3, 147.5, 143.7 (m), 141.6 (m), 141.1, 139.2 (m), 137.2 (m), 135.8, 129.9, 127.5, 125.9, 124.9, 77.3, 62.2, 60.0, 37.3. ¹⁹**F NMR** (471 MHz, DMSO) δ -145.35 (d, J = 20.0 Hz), -147.85, -159.99 (d, J = 25.0 Hz). **HRMS (ESI)** m/z Calcd for [C₁₈H₁₁F₅N₃O, M - Cl]⁺: 380.0817; Found: 380.0819.

3. General procedure for the anion exchange reaction.^{2,3}



To a suspension of azolium chloride (0.1 mmol) in CH_3CN (1 mL) was added AgSbF₆ or NaBPh₄ (0.1 mmol). After being stirred for 12 h at 24 °C, the reaction mixture was filtered through silica gel and the solvent was removed under reduced pressure. Recrystallization from Hexanes/Ethyl acetate gave the desired product as white solid.



White solid. 82% yield, **MP:** 221 - 223 °C. ¹**H NMR** (500 MHz, Acetone) δ 11.29 (s, 1H), 7.64 (d, J = 7.7 Hz, 1H), 7.48-7.43 (m, 2H), 7.37 (t, J = 7.4 Hz, 1H), 6.38 (d, J = 3.4 Hz, 1H), 5.44 (d, J = 16.4 Hz, 1H), 5.28 (d, J = 16.4 Hz, 1H), 5.21 (t, J = 4.5 Hz, 1H), 3.57 (dd, J = 17.1, 5.0 Hz, 1H), 3.33 (d, J = 17.1 Hz, 1H). ¹³**C NMR** (126 MHz, Acetone) δ 151.7, 146.1, 143.9 (m), 141.9 (m), 140.8, 139.4 (m), 137.3 (m), 135.1, 129.7, 127.2, 125.7, 124.2, 77.3, 62.7, 59.9, 37.0. ¹⁹**F NMR** (471 MHz, Acetone) δ -146.65 (d, J = 18.6 Hz), -149.45 (t, J = 22.2 Hz), -161.41 - -161.55 (m). **HRMS (ESI)** m/z Calcd for [C₁₈H₁₁F₅N₃O, M – SbF₆]⁺: 380.0817; Found: 380.0820.



White solid. 70% yield, **MP:** 216 - 218 °C. ¹**H NMR** (500 MHz, Acetone) δ 10.77 (s, 1H), 7.51 (d, *J* = 7.7 Hz, 1H), 7.46 - 7.40 (m, 2H), 7.40 - 7.29 (m, 9H), 6.95 (t, *J* = 7.4 Hz, 8H), 6.81 (t, J = 7.2 Hz, 4H), 6.05 (d, J = 4.1 Hz, 1H), 5.26 (t, J = 21.0 Hz, 1H), 5.10 – 4.98 (m, 1H), 4.91 (t, J = 4.5 Hz, 1H), 3.46 – 3.33 (m, 1H), 3.23 (d, J = 17.2 Hz, 1H). ¹³C NMR (126 MHz, Acetone) δ 164.0 (dd, J = 98.7, 49.4 Hz), 151.7, 145.8, 143.9 (m), 141.8 (m), 140.8, 139.4 (m), 137.4 (m), 136.1 (d, J = 1.4 Hz), 134.8, 129.7, 127.1, 125.7, 125.2 (m), 124.1, 121.4, 77.2, 62.5, 59.8, 36.9. ¹⁹F NMR (471 MHz, Acetone) δ -146.36 (d, J = 17.0 Hz), -148.74 (t, J = 20.0 Hz), -160.68 – -161.51 (m). HRMS (ESI) m/z Calcd for [C₁₈H₁₁F₅N₃O, M – BPh₄]⁺: 380.0817; Found: 380.0824.

4. General procedure for the synthesis of N-benzyl Isatin derivations



A solution of isatin (5.0 g, 34 mmol) in DMF (60 mL) was cooled to 0 °C (ice bath). NaH (60% dispersion in mineral oil, 1.4 g, 36 mmol) was added portion wise to the orange solution. The colour of the solution turned to deep purple. When the gas evolution stopped, benzyl bromide (6.7 g, 39 mmol) was added slowly, upon which the mixture turned red-brown. After the reaction mixture was stirred for 15 min at room temperature, ice water (300 mL) was introduced to precipitate the product. After suction filtration, the product was washed with hexane to afford 1-benzylindoline-2,3-dione after drying.

5. General procedure A for the synthesis of 3-Hydroxy-3-Substituted Oxindoles.⁴



The isatin derivative (20 mmol) was dissolved in anhydrous THF (60 mL) and cooled to -78 $^{\circ}$ C followed by dropwise addition of a 2.0 M solution of RMgBr in THF (12.0 ml, 24.0 mmol). Then, the reaction warmed to room temperature, and the reaction was stirred under N₂ for 30 min at which point TLC analysis indicated consumption of the starting material. The reaction mixture was quenched by the addition of saturated aqueous NH₄Cl (20 mL), and extracted with ethyl acetate (3 × 20 mL). The combined organic layers were washed with brine (20 mL), dried over anhydrous MgSO₄, filtered, concentrated, and the residue was purified by silica gel chromatography with EtOAc/Hexane to afford desired product.

6. General procedure for the kinetic resolution of 3-Hydroxy-3-Substituted Oxindoles through NHC-Catalyzed Oxidative Esterification



To a 4 mL vial was added alcohol (0.15 mmol), triazolium salt **3** (0.015 mmol), Manganese (IV) oxide activated (0.75 mmol), additive (0.075 mmol). In glovebox Magnesium trifluoromethanesulfonate (0.015 mmol), cinnamaldehyde (48 μ L, 0.375 mmol), 100 mg 4Å, anhydrous THF (0.3 mL) were added, and after 10 min DBU (23 μ L, 0.15 mmol) were added. The mixture was stirred at 23 °C for 24 h, and purified by column chromatography on silica gel with hexane/ethyl acetate (10:1 v/v) as eluent to afford the annulation product. Racemic products were synthesized *via* similar procedure using achiral triazolium salt.

7. General procedure B for the synthesis of 3-Hydroxy-3-Substituted Oxindoles.⁵



To a solution of *N*-protected isatin derivative (6 mmol) in pyridine (8 mL) and ethanol (24 mL), malonic acid (956 mg, 9.2 mmol) was added. The reaction mixture was refluxed for 20 h, after which TLC revealed the complete consumption of the starting material. The reaction mixture was quenched with 2M HCl (25 mL), extracted with dichloromethane (3 x 25 mL), and concentrated under reduced pressure to yield a solid residue. The crude product was then dissolved in 30 mL of the alcohol and cooled to 0 °C with an ice bath. Thionyl chloride (2.2 mL) was added dropwise to the solution and left to stir overnight at room temperature. The resulting tertiary alcohol was quenched with sodium carbonate to bring the solution to pH 8, extracted with DCM, dried over Na₂SO₄, evaporated in vacuo and purified by column chromatography with Hexane/Ethyl acetate as eluent.



To a solution of 3-(1H-indol-3-yl)propanoic acid (6 mmol) in IPA was added SOCl₂ (2.2 mL) at 0 °C and warmed to room temperature and stirred overnight. The reaction was quenched with sodium carbonate to bring the solution to pH 8, extracted with DCM, dried over Na₂SO₄, evaporated *in vacuo* and purified by column chromatography with hexanes/ethyl acetate as

eluent to yield the ester product. Benzylation of the product was performed as above procedures to yield isopropyl 3-(1-benzyl-1*H*-indol-3-yl)proponoate.

The benzylated ester product (2 g, 6.2 mmol) was dissolved in DMSO (10 mL) and to it was added dropwise conc. HCl (14 mL) at r.t. The resulting dark solution was stirred overnight, and poured into ice-water (120 mL). Aqueous NaHCO₃ was added slowly to bring the pH of the solution to \sim 7, and extracted with extracted with EtOAc, washed with brine, dried over Na₂SO₄, and concentrated *in vacuo*. The residue was purified by column chromatography with hexanes/ethyl acetate (8:1) as eluent.

The purified isopropyl 3-(1-benzyl-2-oxoindolin-3-yl)propanoate (900 mg, 2.7 mg) was dissolved in toluene (20 mL). Tetra-n-butylammonium bromide (TBAB) (150 mg, 0.53 mmol), P(OEt)₃ (0.9 mL, 5.3 mmol) and 50% aqueous KOH (6 mL) was added. Reaction was stirred at r.t. overnight in air, after which the reaction mixture was diluted with EtOAc and water. Aqueous layer was extracted EtOAc (2 x 15 mL) and combined organic extracts was dried over Na₂SO₄, concentrated under reduced pressure and residue was purified by column chromatography with hexane/ethyl acetate as eluent to yield the racemic version of **1**i.

MeO OH N Bn (±)-1e	Ph H -	3 (10 mol%) MnO₂ (5 equiv.) DBU (1 equiv.) additive (50 mol%) 4 Å MS, THF, 24 °C	MeO O N Bn 1e	OMe OMe Bn 4a
entry	Additive	ee _{ie} ^b	Conv. ^c	$k_{rel}{}^{c}$
1	none	49	45	6
2	LiBF ₄	48	43	7
3	NaBF ₄	45	40	8
4	KBF ₄	40	42	5
5	NaPF ₆	59	50	7
6	NaSbF ₆	72	48	17
7	NaBPh ₄	65	47	12
8 ^d	NaSbF ₆	84	50	30

8. Table 1. Screening different additives for kinetic resolution of substate 1e^a

^{*a*}The reaction was carried out at 0.15 mmol scale using **1e** (1.0 equiv.), cinnamaldehyde (2.5 equiv.), NHC precursor **3** (10 mol%), DBU (1.0 equiv.), MnO₂ (5 equiv.), 4 Å MS (100 mg) and one of the additives (50 mol%) in THF (0.5 M). ^{*b*}Determined by HPLC. ^{*c*}Conv. was indicated by ¹H NMR using 1,1,2,2-tetrachloroethane as the internal standard. $k_{rel} = \ln[(1-\text{Conv.})(1-\text{ee}_{1e})] /\ln[(1-\text{Conv.})(1+\text{ee}_{1e})]$. ^{*d*}Reaction was carried out with 10 mol% of Mg(OTf)₂.

9. General Procedures B for Kinetic Resolution of 3-Hydroxy-3-Substituted Oxindoles



To a 4 mL vial was added the (\pm)-**1e-1k** (0.15 mmol), **3** (0.015 mmol), activated manganese (IV) oxide (0.75 mmol), NaSbF₆ (0.075 mmol) and 4Å molecular sieves (100mg). The mixture was taken into the glovebox, where magnesium trifluoromethanesulfonate (0.015 mmol), cinnamaldehyde (50 μ L, 0.375 mmol) and anhydrous THF (300 μ L) were added. The reaction mixture was taken outside the glovebox, and DBU (23 μ L, 0.15 mmol) was added using a micropipette. The reaction vial was then sealed and allowed to stir at ambient temperature overnight, and filtered over a short column of silica gel with dichloromethane as eluent. The filtrate was concentrated under reduced pressure and the conversion was indicated by ¹H NMR using 1,1,2,2-tetrachloroethane as the internal standard. The crude reaction mixture was directly purified by silica gel column chromatography with hexanes/ethyl acetate (10:1 v/v) as eluent to afford the ester product and recovered the tertiary alcohol in pure form.

10. Characterization of compounds.

(S)-3-allyl-1-benzyl-3-hydroxyindolin-2-one (Scheme 3, 1a).



Recovered starting material: colorless syrup, 45 % yield. ¹H NMR (500 MHz, CDCl₃) δ 7.43 (d, *J* = 7.3 Hz, 1H), 7.37 – 7.26 (m, 5H), 7.23 (t, *J* = 7.5 Hz, 1H), 7.09 (t, *J* = 7.5 Hz, 1H), 6.73 (d, *J* = 7.8 Hz, 1H), 5.66 (dddd, *J* = 14.7, 10.1, 8.4, 6.3 Hz, 1H), 5.17 (d, *J* = 17.1 Hz, 1H), 5.12 (d, *J* = 10.1 Hz, 1H), 5.05 (d, *J* = 15.7 Hz, 1H), 4.76 (d, *J* = 15.7 Hz, 1H), 3.55 (br, 1H), 2.85 (dd, *J* = 13.2, 6.2 Hz, 1H), 2.74 (dd, *J* = 13.0, 8.4 Hz, 1H). ¹³C NMR (CDCl₃, 125 MHz): δ 178.0, 142.4, 135.3, 130.5, 129.7, 129.4, 128.6, 127.6, 127.2, 124.1, 123.0, 120.3, 109.4, 76.0, 43.7, 42.9. **IR** *v* (cm⁻¹): 2465, 2344, 2282, 2115, 2038, 1850, 1681, 1621, 799, 757. The NMR date were consistent with that reported in the literature.⁶ **Specific Rotation**: [α]²⁵_D -36.0 (*c* = 0.10, CHCl₃). The absolute configuration of **1a** was assigned by comparing its specific rotation with that of the same compound reported in the literature.⁴ 97% ee (HPLC condition: Chiralpak IC column, *n*-Hexane/*i*-PrOH = 80:20, flow rate = 1.0 mL/min, wave length = 254 nm, *t*_R = 8.24 min for minor isomer, *t*_R = 9.22 min for major isomer).



(R)-3-allyl-1-benzyl-2-oxoindolin-3-yl cinnamate (Scheme 3, 2a).



Product: colorless syrup, 48 % yield. ¹**H** NMR (500 MHz, CDCl₃) δ 7.71 (d, *J* = 16.0 Hz, 1H), 7.55 - 7.52 (m, 2H), 7.45 - 7.36 (m, 7H), 7.32 - 7.28 (m, 2H), 7.22 (td, *J* = 7.5, 1.0 Hz, 1H), 7.04 (t, *J* = 7.5 Hz, 1H), 6.72 (d, *J* = 7.5 Hz, 1H), 6.51 (d, *J* = 16.0 Hz, 1H), 5.74 - 5.65 (m, 1H), 5.21- 5.15 (m, 2H), 5.08 (d, *J* = 16.0 Hz, 1H), 4.97 (d, *J* = 16.0 Hz, 1H), 3.03 (dd, *J* = 13.5, 6.0 Hz, 1H), 2.79 (dd, *J* = 13.5, 8.5 Hz, 1H); ¹³C NMR (CDCl₃, 125 MHz): δ 174.3, 164.7, 146.4, 142.9, 135.6, 134.1, 130.5, 129.5, 129.5, 128.8, 128.6, 128.1, 127.4, 127.2, 127.1, 122.9, 122.5, 120.7, 116.6, 109.4, 79.4, 44.0, 41.2. HRMS (ESI) m/z Calcd for [C₂₇H₂₃NNaO₃, M + Na]⁺ : 432.1570; Found: 432.1572. IR ν (cm⁻¹): 2463, 2310, 2113, 1989, 1885, 1671, 1599, 806, 757. Specific Rotation: [α]²⁵D -35.0 (*c* = 0.10, CHCl₃). 94% ee (HPLC condition: Chiralcel OD-H column, *n*-Hexane/*i*-PrOH = 80:20, flow rate = 1.0 mL/min, wave length = 254 nm, *t*_R = 15.11 min for major isomer, *t*_R = 17.64 min for minor isomer).



(S)-1-benzyl-3-hydroxy-3-methylindolin-2-one (Scheme 3, 1b).



Recovered starting material: colorless syrup, 44 % yield.¹**H** NMR (500 MHz, CDCl₃) δ 7.45 (d, *J* = 7.2 Hz, 1H), 7.27 - 7.35 (m, 5H), 7.24 (td, *J* = 7.5, 1.2 Hz, 1H), 7.09 (t, *J* = 7.5 Hz, 1H), 6.75 (d, *J* = 7.8 Hz, 1H), 4.99 (d, *J* = 15.7 Hz, 1H), 4.84 (d, *J* = 15.7 Hz, 1H), 3.44 (br, 1H), 1.70 (s, 3H). ¹³**C** NMR (CDCl₃, 125 MHz): δ 178.9, 141.8, 141.7, 135.4, 131.6, 129.4, 129.3, 128.8, 127.6, 127.1, 123.4, 123.2, 109.5, 73.7, 43.6, 25.0. The NMR date were consistent with that reported in the literature.⁶ IR *v* (cm⁻¹): 2426, 2358, 2282, 2145, 1976, 1855, 1676, 1625, 1175, 734, 483. Specific Rotation: [α]²⁵_D -55.0 (*c* = 0.10, CHCl₃). The absolute configuration of **1b** was assigned by analogy to **1a**. 96% ee (HPLC condition: Chiralpak IC column, *n*-Hexane/*i*-PrOH = 70:30, flow rate = 1.0 mL/min, wave length = 254 nm, *t*_R = 5.32 min for minor isomer, *t*_R = 5.66 min for major isomer).



(R)-1-benzyl-3-methyl-2-oxoindolin-3-yl cinnamate (Scheme 3, 2b).



Product: colorless syrup, 47 % yield. ¹**H NMR** (CDCl₃, 500 MHz): δ 7.71 (d, *J* = 16.0 Hz, 1H), 7.54 - 7.52 (m, 2H), 7.45 - 7.37 (m, 7H), 7.32 - 7.28 (m, 2H), 7.21 (t, *J* = 7.5 Hz, 1H), 7.04 (t, *J* = 7.5 Hz, 1H), 6.72 (d, *J* = 8.0 Hz, 1H), 6.51 (d, *J* = 16.0 Hz, 1H), 5.04 (s, 2H), 1.80 (s, 3H); ¹³C NMR (CDCl₃, 125 MHz): δ 175.2, 164.8, 146.3, 142.3, 135.6, 134.1, 130.5, 129.4, 129.2, 128.8, 128.7, 128.1, 127.5, 127.1, 122.8, 122.0, 116.7, 109.5, 77.4, 44.0, 23.7. HRMS (ESI) m/z Calcd for [C₂₅H₂₁NNaO₃, M + Na]⁺: 406.1414; Found: 406.1391. IR *v* (cm⁻¹): 2467, 2412, 2291, 2115, 1978, 1876, 1676, 1588, 748, 671. Specific Rotation: [α]²⁵_D - 45.0 (*c* = 1.00, CHCl₃). 93% ee (HPLC condition: Chiralcel OD-H column, *n*-Hexane/*i*-PrOH = 70:30, flow rate = 1.0 mL/min, wave length = 254 nm, *t*_R = 8.67 min for major isomer, *t*_R = 11.63 min for minor isomer).



(S)-1-benzyl-3-hydroxy-3-phenylindolin-2-one (Scheme 3, 1c).



Recovered starting material: colorless syrup, 43 % yield. ¹H NMR (500 MHz, CDCl₃) δ 7.49 – 7.43 (m, 2H), 7.40 – 7.33 (m, 6H), 7.33 – 7.28 (m, 2H), 7.25 (td, *J* = 7.8, 1.1 Hz, 1H), 7.07 (dd, *J* = 10.9, 4.2 Hz, 1H), 6.81 (d, *J* = 7.8 Hz, 1H), 5.07 (d, *J* = 15.6 Hz, 1H), 4.85 (dd, *J* = 15.7, 1.8 Hz, 1H), 3.73 (br, 1H). ¹³C NMR (CDCl₃, 125 MHz): δ 177.7, 142.5, 140.2, 135.4, 131.8, 129.6, 128.8, 128.5, 128.1, 127.7, 127.2, 125.3, 124.9, 123.5, 109.7, 77.9, 43.9. The NMR date were consistent with that reported in the literature.⁶ **IR** v (cm⁻¹): 2486, 2430, 2277, 2076, 2082, 1743, 1985, 1616, 762. **Specific Rotation**: $[\alpha]^{25}_{D}$ -32.5 (c = 1.00, CHCl₃). The absolute configuration of **1c** was assigned by analogy to **1a**. 97% ee (HPLC condition: Chiralpak AS-H column, *n*-Hexane/*i*-PrOH = 70:30, flow rate = 1.0 mL/min, wave length = 254 nm, $t_{R} = 13.51$ min for minor isomer, $t_{R} = 18.68$ min for major isomer).



(R)-1-benzyl-2-oxo-3-phenylindolin-3-yl cinnamate (Scheme 3, 2c).



product: colorless syrup, 50 %. ¹**H** NMR (CDCl₃, 500 MHz): δ 7.79 (d, *J* = 16.0 Hz, 1H), 7.57 - 7.56 (m, 2H), 7.51 - 7.49 (m, 2H), 7.43-7.27 (m, 13H), 7.10 (t, *J* = 7.5 Hz, 1H), 6.79 (d, *J* = 8.0 Hz, 1H), 6.62 (d, *J* = 16.0 Hz, 1H), 5.05 (d, *J* = 16.0 Hz, 1H), 4.97 (d, *J* = 16.0 Hz, 1H); ¹³C NMR (CDCl₃, 125 MHz): δ 174.0, 165.0, 146.6, 143.6, 136.8, 135.6, 134.1, 130.6, 129.9, 128.9, 128.9, 128.7, 128.6, 128.2, 127.5, 127.2, 126.4, 124.1, 123.0, 116.8, 109.7, 81.3, 44.3. HRMS (ESI) m/z Calcd for [C₃₀H₂₃NNaO₃, M + Na]⁺: 468.1570; Found: 468.1557. IR *v* (cm⁻¹): 2488, 2319, 2122, 1999, 1857, 1676, 1607, 799, 752. Specific Rotation: [α]²⁵_D -12.2 (*c* = 0.10, CHCl₃). 85% ee (HPLC condition: Chiralcel OD-H column, *n*-Hexane/*i*-PrOH = 70:30, flow rate = 1.0 mL/min, wave length = 254 nm, t_R = 8.56 min for major isomer, t_R = 11.83 min for minor isomer).



(R)-1-benzyl-3-hydroxy-3-(pyridin-2-yl)indolin-2-one (Scheme 3, 1d).



Recovered starting material: colorless syrup, 51 % yield. ¹H NMR (500 MHz, CDCl₃) δ 8.66 (d, *J* = 4.6 Hz, 1H), 7.64 (td, *J* = 7.7, 1.3 Hz, 1H), 7.43 – 7.23 (m, 8H), 7.19 (d, *J* = 7.3 Hz, 1H), 7.05 (t, *J* = 7.5 Hz, 1H), 6.96 (d, *J* = 7.9 Hz, 1H), 6.83 (d, *J* = 7.9 Hz, 1H), 6.30 (s, 1H), 5.09 (d, *J* = 15.7 Hz, 1H), 4.84 (d, *J* = 15.7 Hz, 1H). ¹³C NMR (CDCl₃, 125 MHz): δ 176.6, 156.6, 148.1, 143.6, 137.4, 135.5, 130.9, 130.0, 128.8, 127.6, 127.3, 124.7, 123.4, 119.9, 109.6, 77.7, 44.0. The NMR date were consistent with that reported in the literature.⁶ IR *v* (cm⁻¹): 2484, 2323, 2117, 2001, 1676, 1595, 801, 755, 648. Specific Rotation: [α]²⁵_D-15.0 (*c* = 0.10, CHCl₃). The absolute configuration of 1d was assigned by analogy to 1a. 71% ee (HPLC condition: Chiralpak IC column, *n*-Hexane/*i*-PrOH = 70:30, flow rate = 1.0 mL/min, wave length = 254 nm, *t*_R = 8.85 min for minor isomer, *t*_R = 14.26 min for major isomer).



(S)-1-benzyl-2-oxo-3-(pyridin-2-yl)indolin-3-yl cinnamate (Scheme 3, 2d).



Product: colorless syrup, 41 % yield. ¹H NMR (CDCl₃, 500 MHz): δ 8.56 (d, *J* = 3.5 Hz, 1H), 7.95 (d, *J* = 8.5 Hz, 1H), 7.84 (td, *J* = 8.0, 1.5 Hz, 1H), 7.79 (d, *J* = 16.0 Hz, 1H), 7.58 - 7.54 (m, 4H), 7.43 - 7.37 (m, 5H), 7.32 - 7.26 (m, 2H), 7.23 - 7.20 (m, 2H), 7.00 (t, *J* = 7.5 Hz, 1H), 6.74 (t, *J* = 8.0 Hz, 1H), 6.66 (d, *J* = 16.0 Hz, 1H), 5.22 (d, *J* = 16.0 Hz, 1H), 5.02 (d, *J* = 16.0 Hz, 1H); ¹³C NMR (CDCl₃, 125 MHz): δ 173.7, 164.6, 156.9, 149.6, 146.9, 144.0, 137.1, 135. 6, 134.1, 130.7, 129.8, 128.9, 128.8, 128.7, 128.3, 127.4, 127.2, 123.5, 123.3, 122.8, 120.6, 116.6, 109.8, 82.4, 44.3. HRMS (ESI) m/z Calcd for [C₂₉H₂₂N₂NaO₃, M + Na]⁺: 469.1523; Found: 469.1511. IR v (cm⁻¹): 2470, 2351, 2291, 2113, 1982, 1857, 1669, 1607, 810, 750. Specific Rotation: [α]²⁵_D -13.6 (*c* = 1.00, CHCl₃). 86% ee (HPLC condition: Chiralpak AD-H column, *n*-Hexane/*i*-PrOH = 60:40, flow rate = 1.0 mL/min, wave length = 254 nm, *t*_R = 9.09 min for major isomer, *t*_R = 10.05 min for minor isomer).



methyl (S)-2-(1-benzyl-3-hydroxy-2-oxoindolin-3-yl)acetate, (Scheme 4, 1e)



Recovered starting material: 45% yield. ¹H NMR (400 Mhz) δ 7.31 (dd, J = 9.0 Hz, 1H), 7.23 – 7.19, (m, 5H), 7.14 (td, J = 7.8, 1.2 Hz, 1H), 6.97 (td, J = 7.6, 0.8 Hz, 1H), 6.64 (d, J =7.6 Hz, 1H), 4.86 (d, J = 15.6 Hz, 1H), 4.77 (d, J = 15.6 Hz, 1H), 4.03 (br, 1H), 3.59 (s, 3H), 2.92 (d, J = 2.4 Hz, 2H). ¹³C NMR (100 MHz) δ 176.2, 170.8, 142.6, 135.3, 130.1, 129.2, 128.8, 127.7, 127.3, 123.9, 123.2, 109.7, 73.5, 52.1, 43.9, 41.0. HRMS (ESI) m/z Calcd for [C₁₈H₁₇NNaO₄, M + Na]⁺: 334.1050; Found: 334.1049. IR v (cm⁻¹): 2460, 2347, 2268, 1987, 1859, 1597, 1175, 799, 757. Specific Rotation: [α]²⁵_D +10.70 (c = 0.1, CHCl₃). The absolute configuration of 1e was assigned by analogy to 1k. 84% ee (HPLC condition: Chiralpak IC column, *n*-Hexane/*i*-PrOH = 80:20, flow rate = 1.0 mL/min, wave length = 254 nm, $t_R = 18.77$ min for minor isomer, $t_R = 29.73$ min for major isomer).



Isopropyl 2-(1-benzyl-3-hydroxy-2-oxoindolin-3-yl)acetate, (Scheme 5, 1f)



Recovered starting material: 42% yield. ¹H NMR (300 MHz, CDCl₃): δ 7.40 (d, J = 7.5 Hz, 1H), 7.31 - 7.25 (m, 5H), 7.19 (dd, J = 8.0 Hz, 1H) 7.04 (dd, J = 8.0 Hz, 1H), 6.70 (d, J = 7.5Hz, 1H), 5.03 (h, J = 6.4, 6.2 Hz, 1H), 4.89 (d, J = 1.5 Hz, 2H), 2.93 (s, 2H), 1.73 (br, 1H), 1.23 (dd, J = 6.4, 6.2 Hz, 6 H). ¹³C NMR (300 MHz, CDCl₃): δ 176.2, 170.0, 142.6, 135.3, 129.9, 129.3, 128.8, 127.6, 127.2, 123.9, 123.1, 109.6, 73.6, 68.9, 43.8, 41.4, 21.6. HRMS (ESI) m/z Calcd for [C₂₀H₂₁NNaO₄, M + Na]⁺: 362.1363; Found: 362.1369. IR ν (cm⁻¹): 2486, 2410, 2317, 2293, 2117, 2003, 1783, 1671, 1599, 752, 666. Specific Rotation: [α]²⁵_D +12.00 (c = 0.1, CHCl₃). The absolute configuration of 1f was assigned by analogy to 1k. 99% ee (HPLC condition: Chiralcel OD-H column, *n*-Hexane/*i*-PrOH = 85:15, flow rate = 0.5 mL/min, wave length = 254 nm, $t_R = 13.97$ min for minor isomer, $t_R = 12.11$ min for major isomer).



2-(1-benzyl-3-hydroxy-2-oxoindolin-3-yl)acetonitrile, (Scheme 5, 1g)



Recovered starting material: 39% yield. ¹H NMR (500 MHz, CDCl₃): δ 7.65 (d, J = 7.5 Hz, 1 H), 7.34 – 7.26 (m, 6 H), 7.14, (dd, J = 7.5 Hz, 1 H), 6.78 (d, J = 7.5 Hz, 1 H), 4.90 (d, J =15.5 Hz, 1 H), 4.84 (d, J = 15.5 Hz, 1 H), 3.13 (d, J = 16.5 Hz, 1 H), 2.80 (d, J = 16.5 Hz, 1 H). ¹³C NMR (125 MHz, CDCl₃): δ 175.2, 142.1, 134.6, 130.9, 128.9, 128.0, 127.3, 127.2, 124.3, 123.9, 115.2, 110.1, 72.5, 44.1, 27.5. HRMS (ESI) m/z Calcd for [C₁₇H₁₄N₂NaO₂, M + Na]⁺: 301.0947; Found: 301.0946. IR v (cm⁻¹): 2486, 2347, 2314, 2117, 1996, 1848, 1674, 1597, 762, 715. Specific Rotation: [α]²⁵_D +18.60 (c = 0.1, CHCl₃). The absolute configuration of 1g was assigned by analogy to 1k. 85% ee (HPLC condition: Chiralcel OD-H column, *n*-Hexane/*i*-PrOH = 70:30, flow rate = 2.0 mL/min, wave length = 254 nm, $t_R = 6.90$ min for major isomer, $t_R = 8.22$ min for minor isomer).



Isopropyl 2-(1-benzyl-3-hydroxy-5-methoxy-2-oxoindolin-3-yl)acetate, (Scheme 5, 1h)



Recovered starting material: 36% yield. ¹**H NMR** (400 MHz, CDCl₃): δ 7.34 – 7.28 (m, 5 H), 7.06 (d, 2.4 Hz, 1 H), 6.74, (dd, J = 8.4, 2.4 Hz, 1 H), 6.61 (d, J = 8.4 Hz, 1 H), 5.08 (h, J = 6.0 Hz, 1H), 4.89 (s, 2 H), 4.51 (s, 1 H), 3.77 (s, 3 H), 2.93 (d, J = 0.4 Hz, 2 H), 1.23 (d, J = 6.0 Hz, 3H), 1.22 (d, J = 6.0 Hz, 3H). ¹³**C NMR** (100 MHz, CDCl₃): δ 175.9, 170.1, 156.3, 135.8, 135.4, 130.5, 128.8, 127.6, 127.2, 114.5, 111.1, 110.1, 76.6, 74.0, 69.0, 55.8, 43.9, 41.4, 21.6. **HRMS (ESI)** m/z Calcd for [C₂₁H₂₃NNaO₅, M + Na]⁺: 392.1468; Found: 392.1470. **IR** v (cm⁻¹): 2470, 2386, 2293, 2096, 1971, 1841, 1750, 1614, 1595, 1159, 752, 702. **Specific Rotation**: [α]²⁵_D +13.30 (c = 0.1, CHCl₃). The absolute configuration of **1h** was assigned by analogy to **1k**. 99% ee (HPLC condition: Chiralcel OD-H column, *n*-Hexane/*i*-PrOH = 85:15, flow rate = 1.0 mL/min, wave length = 254 nm, $t_R = 8.85$ min for minor isomer, $t_R = 10.34$ min for major isomer).



Isopropyl 3-(1-benzyl-3-hydroxy-2-oxoindolin-3-yl)propanoate, (Scheme 5, 1i)



Recovered starting material: 43%. ¹H NMR (400 MHz, CDCl₃): δ 7.32 (dd, *J* = 7.6, 0.8 Hz, 1 H), 7.25 – 7.20, (m, 5 H), 7.13 (dd, *J* = 8.0, 1.2 Hz, 1 H), 7.01 (dd, *J* = 7.6, 0.8 Hz, 1 H), 6.64 (d, *J* = 7.6 Hz, 1 H), 4.86 – 4.83 (m, 3 H), 2.57 – 2.53 (m, 2 H), 2.33 – 2.26 (m, 2H), 1.12 (d, *J* = 6.0 Hz, 3 H), 1.07 (d, *J* = 6.0 Hz, 3 H). ¹³C NMR (100 MHz, CDCl₃): δ 173.6, 171.3, 141.7, 135.0, 130.3, 128.9, 128.9, 127.8, 127.1, 124.4, 123.5, 109.8, 68.1, 64.0, 44.1, 34.0, 29.7, 21.7, 21.7. HRMS (ESI) m/z Calcd for [C₂₁H₂₃NNaO₄, M + Na]⁺: 376.1519; Found: 376.1515. IR v (cm⁻¹): 2484, 2275, 2029, 1836, 1614, 803, 752, 725. Specific Rotation: [α]²⁵_D +22.00 (*c* = 0.1, CHCl₃). The absolute configuration of **1i** was assigned by analogy to **1k**. 96% ee (HPLC condition: Chiralpak ID column, *n*-Hexane/*i*-PrOH = 80:20, flow rate = 1.0 mL/min, wave length = 254 nm, *t*_R = 13.04 min for minor isomer, *t*_R = 14.30 min for major isomer).



Isopropyl 2-(3-hydroxy-1-(4-methoxybenzyl)-2-oxoindolin-3-yl)acetate, (Scheme 5, 1j)



Recovered starting material: 40%, ¹H NMR (500 MHz, CDCl₃): δ 7.40 (dd, J = 7.5 Hz, 1 H), 7.26 – 7.19 (m, 3 H), 7.03 (dd, J = 7.5 Hz, 1 H), 6.84 (d, J = 8.5 Hz, 2 H), 6.72 (d, J = 7.5 Hz, 1 H), 5.04 – 4.99 (m, 1 H), 4.86 – 4.79 (m, 2 H), 3.76 (s, 3H), 2.94 (s, 2 H), 1.17 – 1.15 (m, 6 H),. ¹³C NMR (125 MHz): δ 176.2, 169.9, 159.0, 142.6, 129.9, 129.3, 128.6, 127.3, 123.8, 123.0, 114.1, 109.5, 73.6, 68.8, 55.2, 43.3, 41.5, 21.5, 21.5. HRMS (ESI) m/z Calcd for [C₂₁H₂₃NNaO₅, M + Na]⁺: 392.1468; Found: 392.1468. IR *v* (cm⁻¹): 2474, 2384, 2354, 2092, 1982, 1676, 1616, 1161, 736, 708. Specific Rotation: [α]²⁵_D +24.00 (*c* = 0.1, CHCl₃). The absolute configuration of 1j was assigned by analogy to 1k. 99% ee (HPLC condition: Chiralcel OD-H column, *n*-Hexane/*i*-PrOH = 85:15, flow rate = 0.5 mL/min, wave length = 254 nm, *t*_R = 18.03 min for major isomer, *t*_R = 20.60 min for minor isomer).



Isopropyl 2-(3-hydroxy-2-oxo-1-tritylindolin-3-yl)acetate, (Scheme 5, 1k)



Recovered starting material: 43% yield. ¹H NMR (400 MHz, CDCl₃): δ 7.40 – 7.38 (m, 5H), 7.27 – 7.25 (m, 1H), 7.20-7.10 (m, 10 H), 6.90 – 6.84 (m, 2H), 6.24 – 6.22 (m, 1H), 3.74 (br, 1h), 3.57 (s, 3H), 2.94 (d, *J* = 15.2 Hz, 1H), 2.88 (d, *J* = 15.2 Hz, 1H). ¹³C NMR (100 MHz, CDCl₃): δ 177.8, 170.2, 143.2, 141.8, 129.2, 128.5, 127.9, 127.6, 126.9, 123.2, 122.6, 116.3, 74.5, 73.3, 52.0, 42.1. HRMS (ESI) m/z Calcd for [C₃₀H₂₅NNaO₄, M + Na]⁺: 486.1676; Found: 486.1680. IR *v* (cm⁻¹): 2493, 2284, 2117, 2031, 1920, 1769, 1678, 1602, 750. Specific Rotation: [α]²⁵_D + 24.20 (*c* = 0.1, CH₂Cl₂). The absolute configuration of 1k was assigned by comparing its specific rotation with that of the enantiomer reported in the literature.⁷ 88% ee (HPLC condition: Chiralcel OD-H column, *n*-Hexane/*i*-PrOH = 85:15, flow rate = 1.0 mL/min, wave length = 254 nm, *t*_R = 7.45 min for minor isomer, *t*_R = 9.78 min for major isomer).



1-benzyl-3-(2-methoxy-2-oxoethyl)-2-oxoindolin-3-yl cinnamate, (Scheme 4b, 2e)



¹H NMR (500 MHz, CDCl₃) δ 7.70 (d, J = 16.0 Hz, 1H), 7.58 – 7.36 (m, 10H), 7.31 (dd, J = 14.6, 7.3 Hz, 1H), 7.23 (t, J = 7.6 Hz, 1H), 7.04 (t, J = 7.4 Hz, 1H), 6.72 (d, J = 7.8 Hz, 1H), 6.48 (d, J = 15.9 Hz, 1H), 5.07 (d, J = 15.8 Hz, 1H), 5.02 (d, J = 15.9 Hz, 1H), 3.58 (s, 3H), 3.37 (d, J = 15.4 Hz, 1H), 3.17 (d, J = 15.4 Hz, 1H). ¹³C NMR (126 MHz, CDCl₃) δ 173.7,

168.2, 164.4, 146.7, 143.8, 135.6, 134.1, 130.6, 130.2, 128.9, 128.8, 128.2, 127.5, 127.4, 126.0, 123.8, 122.7, 116.6, 109.6, 77.1, 51.8, 44.5, 41.2. **HRMS (ESI)** m/z Calcd for [C₂₇H₂₃NNaO₅, M + Na]⁺: 464.1468; Found: 464.1467. **IR** *v* (cm⁻¹): 2479, 2423, 2310, 1987, 1781, 1595, 1168, 813, 752

(HPLC condition: Chiralcel AD column, *n*-Hexane/*i*-PrOH = 70:30, flow rate = 1.0 mL/min, wave length = 254 nm, $t_{\rm R}$ = 15.89 min for one isomer, $t_{\rm R}$ = 19.15 min for the other isomer).



(Z)-methyl 2-(1-benzyl-2-oxoindolin-3-ylidene)acetate, (Scheme 4a, 4a)



¹H NMR (500 MHz, CDCl₃) δ 8.59 (d, *J* = 7.7 Hz, 1H), 7.37 – 7.26 (m, 6H), 7.06 (td, *J* = 7.7, 0.8 Hz, 1H), 7.01 (s, 1H), 6.72 (d, *J* = 7.8 Hz, 1H), 4.96 (s, 2H), 3.91 (s, 3H). ¹³C NMR (126 MHz, CDCl₃) δ 167.6, 166.1, 145.1, 137.9, 135.4, 132.4, 128.8, 127.7, 127.2, 122.9, 122.2, 119.9, 109.2, 52.2, 43.8. **IR** *v* (cm⁻¹): 2474, 2356. 2298, 1980, 1848, 1774, 1681, 1173, 796, 762

The NMR date were consistent with that reported in the literature.⁸

(R)-5-acetyl-6-methyl-4-phenyl-3,4-dihydro-2H-pyran-2-one, (Scheme 6b, 7)

42% yield, ¹**H** NMR (500 MHz, CDCl₃) δ 7.36 (dd, J = 10.8, 3.9 Hz, 2H), 7.32 – 7.26 (m, 1H), 7.17 (d, J = 7.6 Hz, 2H), 4.17 (d, J = 6.8 Hz, 1H), 2.99 (ddd, J = 15.6, 7.2, 1.4 Hz, 1H), 2.86 (dd, J = 15.7, 2.4 Hz, 1H), 2.45 (s, 3H), 2.14 (s, 3H). ¹³C NMR (126 MHz, CDCl₃) δ 197.9, 165.6, 160.2, 139.7, 129.4, 127.9, 126.6, 117.3, 38.8, 37.2, 29.7, 19.1. IR v (cm⁻¹): 2484, 2417, 2312, 2261, 1989, 1662, 1590, 1166, 810, 7552. The NMR date were consistent with that reported in the literature.⁸ Specific Rotation: $[\alpha]^{25}_{D}$ - 64.00 (c = 1.0, CHCl₃). The absolute configuration of **7** was assigned by comparing its specific rotation with that of the same compound reported in the literature.⁹ 55% ee (HPLC condition: Chiralpak AD-H column, *n*-Hexane/*i*-PrOH = 99:1, flow rate = 1 mL/min, wave length = 254 nm, $t_{R} = 33.10$ min for major isomer, $t_{R} = 42.06$ min for minor isomer).



11. Reference

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s36



100 -105 -110 -115 -120 -125 -130 -135 -140 -145 -150 -155 -160 -165 -170 -175 -180 -185 -190 -195 -2(f1 (ppm)



(S)-3-allyl-1-benzyl-3-hydroxyindolin-2-one (Scheme 3, 1a).







1H AMX500 3-methyl alcohol



(*R*)-1-benzyl-3-methyl-2-oxoindolin-3-yl cinnamate (Scheme 3, 2b).



s41

(S)-1-benzyl-3-hydroxy-3-phenylindolin-2-one (Scheme 3, 1c).

1H AMX500 phenyl alcohol



(*R*)-1-benzyl-2-oxo-3-phenylindolin-3-yl cinnamate (Scheme 3, 2c).





(*R*)-1-benzyl-3-hydroxy-3-(pyridin-2-yl)indolin-2-one (Scheme 3, 1d).

(S)-1-benzyl-2-oxo-3-(pyridin-2-yl)indolin-3-yl cinnamate (Scheme 3, 2d)







methyl (S)-2-(1-benzyl-3-hydroxy-2-oxoindolin-3-yl)acetate, (Scheme 4, 1e)



Isopropyl 2-(1-benzyl-3-hydroxy-2-oxoindolin-3-yl)acetate, (Scheme 5, 1f)



2-(1-benzyl-3-hydroxy-2-oxoindolin-3-yl)acetonitrile, (Scheme 5, 1g)



Isopropyl 2-(1-benzyl-3-hydroxy-5-methoxy-2-oxoindolin-3-yl)acetate, (Scheme 5, 1h)



Isopropyl 3-(1-benzyl-3-hydroxy-2-oxoindolin-3-yl)propanoate, (Scheme 5, 1i)



Isopropyl 2-(3-hydroxy-1-(4-methoxybenzyl)-2-oxoindolin-3-yl)acetate, (Scheme 5, 1j)



Isopropyl 2-(3-hydroxy-2-oxo-1-tritylindolin-3-yl)acetate, (Scheme 5, 1k)

s52



1-benzyl-3-(2-methoxy-2-oxoethyl)-2-oxoindolin-3-yl cinnamate, (Scheme 4b, 2e)



(Z)-methyl 2-(1-benzyl-2-oxoindolin-3-ylidene)acetate, (Scheme 4a, 4a)



(R)-5-acetyl-6-methyl-4-phenyl-3,4-dihydro-2H-pyran-2-one, (Scheme 6b, 7)