

Supplementary Material

Identification of a novel ene reductase from *Pichia angusta* with potential application in (*R*)-levodione production

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Table and figure captions:

Fig. S1 Sequence alignment of PaER with the other nine OYEs. Substrate binding sites and catalytic residue are marked with black triangles.

Fig. S2 HPLC detection of PaER flavin species.

Fig. S3 (A) Standard curve of size-exclusion chromatography, (B) The size-exclusion chromatography of PaER.

Fig. S4 SDS-PAGE analysis of engineered *E. coli* cells. M: Molecular weight marker; T: whole cell lysate; S: supernatant; P: precipitate.

Fig. S5 GC-MS spectrum of (*R*)-levodione prepared by PaER. GC-EI-MS *m/z* (*M*+ 154 for C₉H₁₄O₂) 139, 111, 95, 83, 69, 56.

Fig. S6 NMR of product (*R*)-levodione prepared by PaER.

¹H NMR (400 MHz, Chloroform-*d*) δ 3.01 (dp, *J* = 13.2, 6.5 Hz, 1H), 2.81 – 2.68 (m, 2H), 2.52 (d, *J* = 15.4 Hz, 1H), 2.34 (dd, *J* = 17.7, 13.3 Hz, 1H), 1.29 – 1.10 (m, 8H).

Table S1 List of primers sequences.

Table S2 Comparison of PaER with other OYEs regarding half-lives.

Table S3 Kinetic parameters of PaER.

Table S4 Comparison of different ene-reductases for asymmetric reduction of ketoisophorone

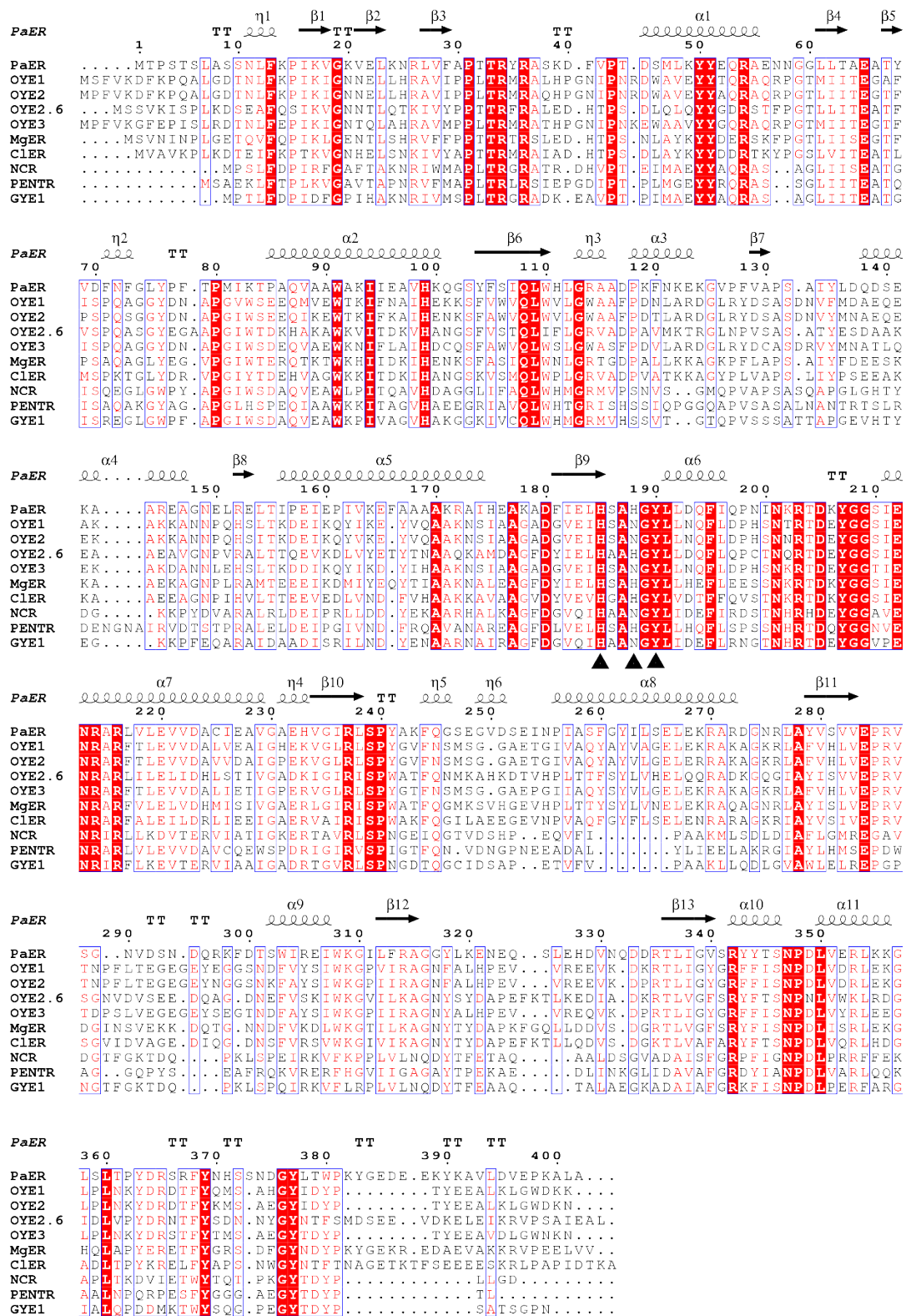


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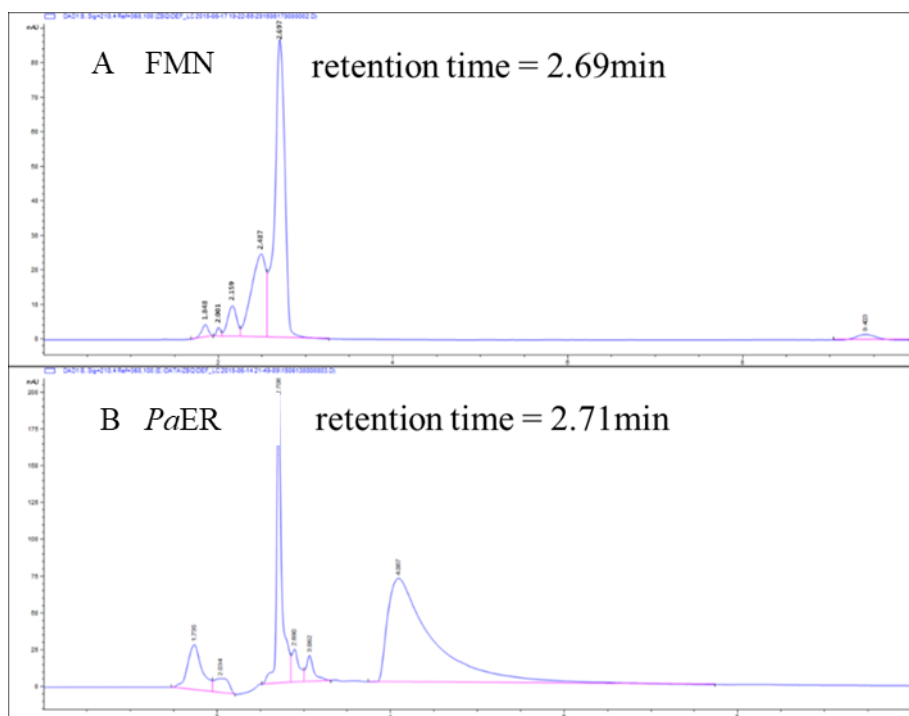


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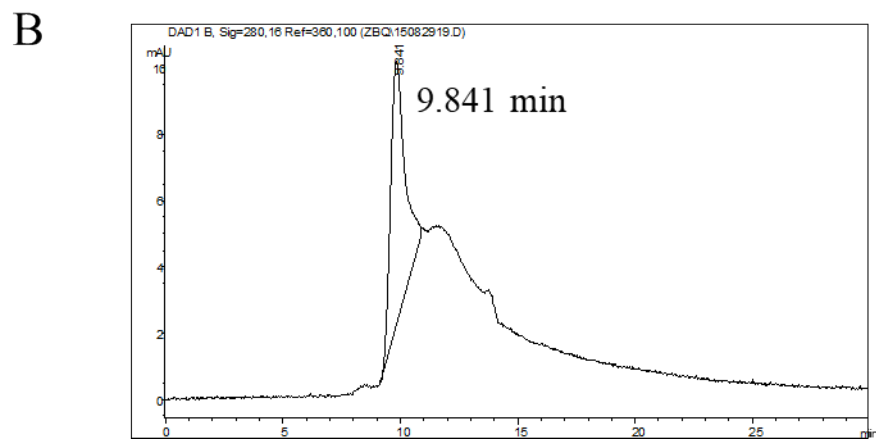
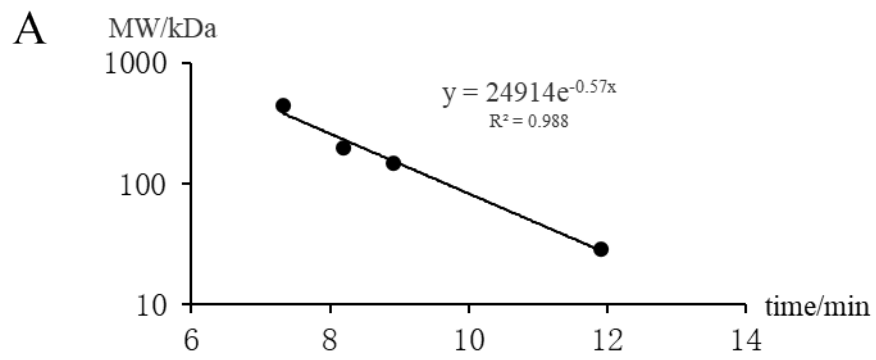


Fig. S3 (A) Standard curve of size-exclusion chromatography, (B) The size-exclusion chromatography of *PaER*.

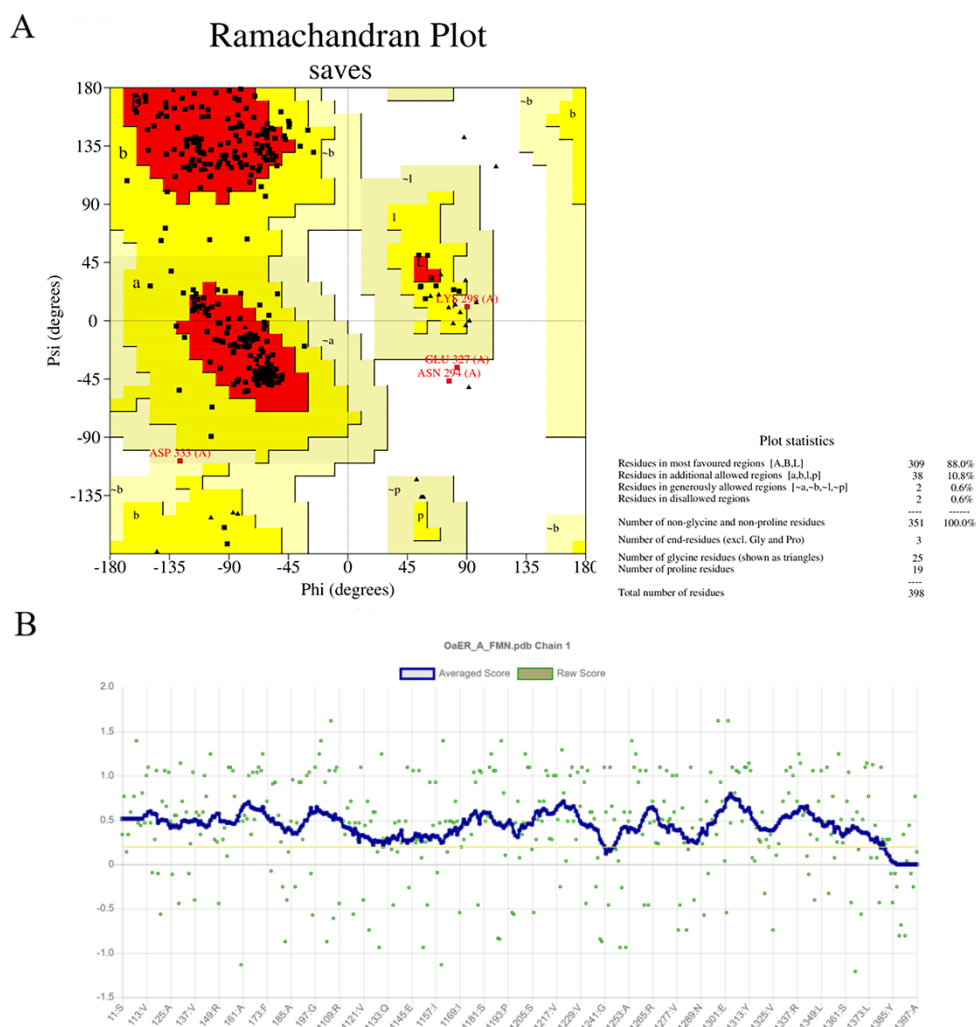


Fig. S4 (A) The Ramachandran plot of *PaER* model evaluation. (B) The Verify-3D analysis of *PaER* model evaluation.

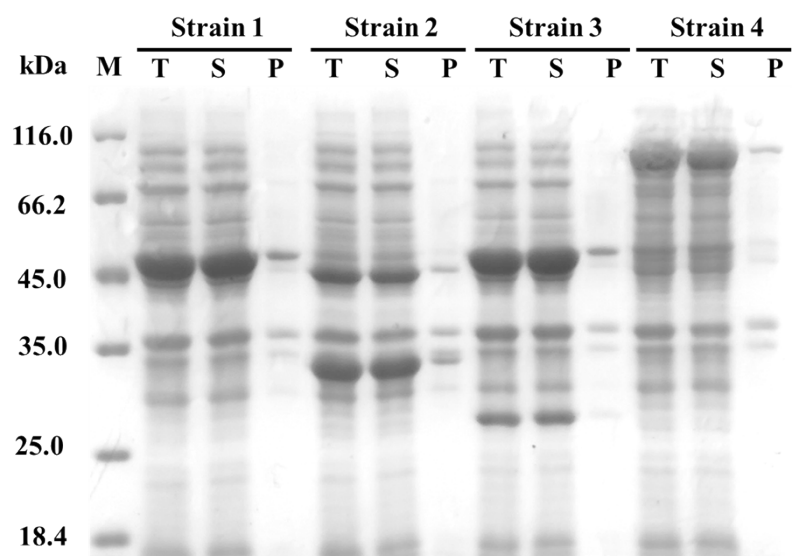


Fig. S5 SDS-PAGE analysis of engineered *E. coli* cells. M: Molecular weight marker; T: whole cell lysate; S: supernatant; P: precipitate.

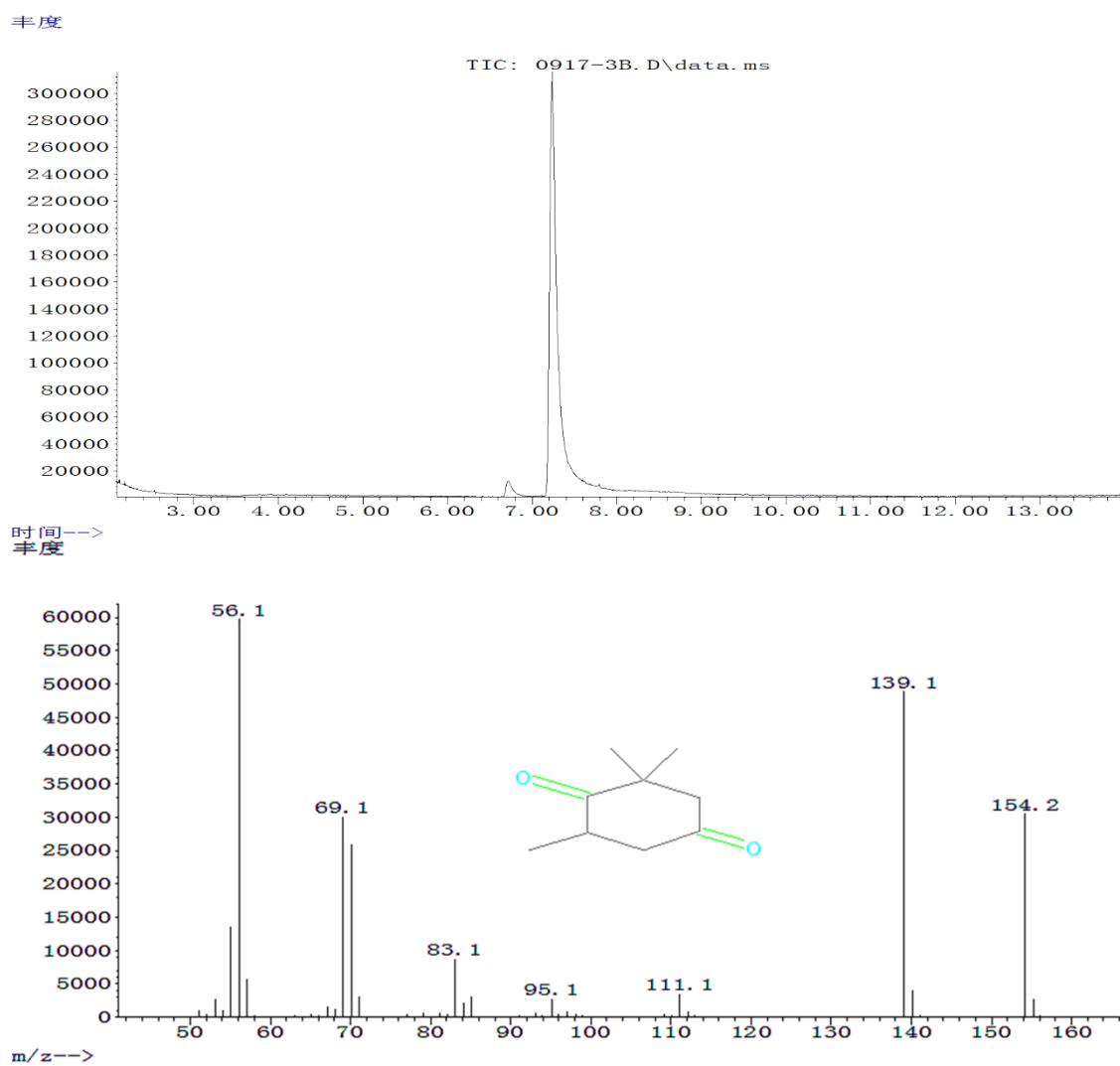


Fig. S6 GC-MS spectrum of (*R*)-levodione prepared by *PaER*. GC-EI-MS *m/z* (*M*+ 154 for C₉H₁₄O₂)

139, 111, 95, 83, 69, 56.

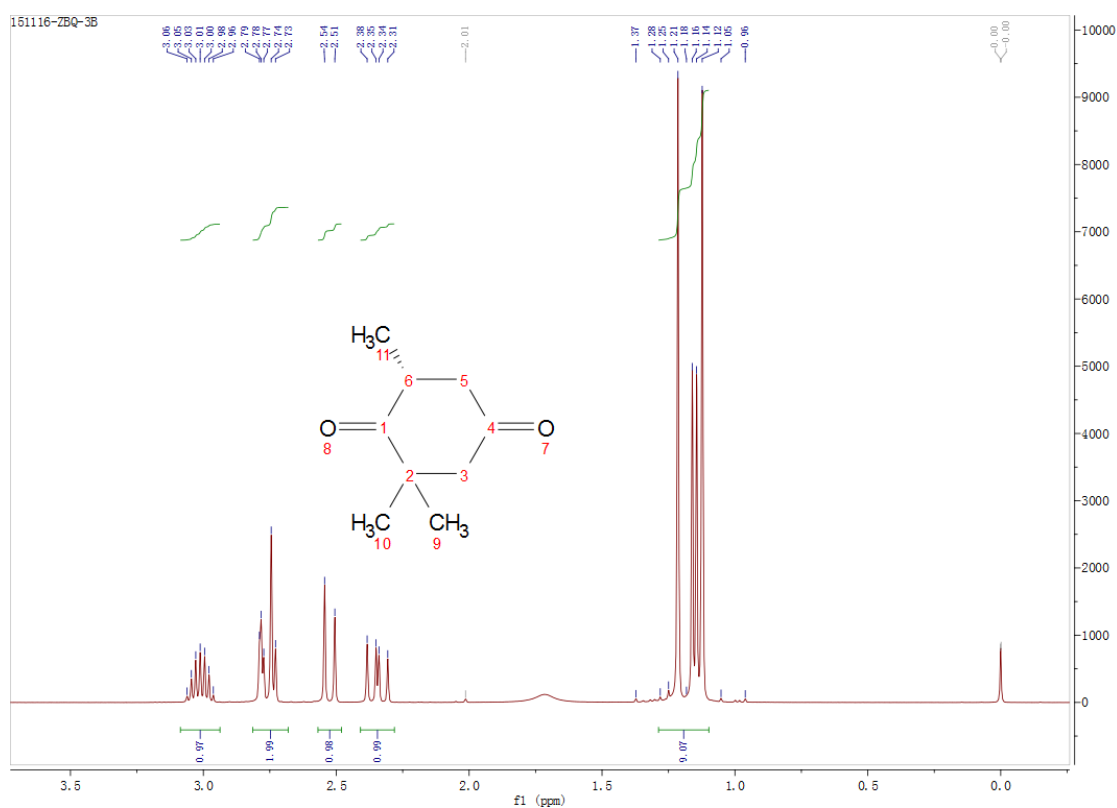


Fig. S7 NMR of product (*R*)-levodione prepared by *Pa*ER.

¹H NMR (400 MHz, Chloroform-d) δ 3.01 (dp, *J* = 13.2, 6.5 Hz, 1H), 2.81 – 2.68 (m, 2H), 2.52 (d, *J* = 15.4 Hz, 1H), 2.34 (dd, *J* = 17.7, 13.3 Hz, 1H), 1.29 – 1.10 (m, 8H).

Table S1 List of primers sequences.

Primer	Sequence (5'→3')
PaER-F	TGGACAGCAAATGGGT <u>CGCGGATCC</u> ATGACCCCAAGCACTTCTCT
PaER-R	GGTGCTCGAGTGCGGCCGCA <u>AAGCTT</u> CTACGCAAGGGCCTTTGGCT
rbs- GDH-F	AGCCAAAGGCCCTTGCGTAGA <u>AAGCTT</u> GAAAGGAGATATACCATGGGC
GDH-R	TGGTGCTCGAGTGCGGCCGCA <u>AAGCTT</u> TTAACCGCGGCCTGCCTG
MCS1- PaER-F	TTAAGAAGGAGATATAC <u>CCATGGT</u> TATGACCCCAAGCACTTCTCTT
MCS1- PaER-R	TGTCGACCTGCAGGCGCGCC <u>GAGCTC</u> CTACGCAAGGGCCTTTGGCT
MCS2- GDH-F	GTATAAGAAGGAGATATAC <u>CATATG</u> GCAGATCTCATGTATCCGGATTTAAAA GG
MCS2- GDH-R	CCGGCCGATATCCAATTG <u>AAGATCT</u> TTAACCGCGGCCTGCCTGGA
GDH- ERK-F	ATTCCAGGCAGGCCGCGGTGAAGAAGAGGAAAAAAGAAACAG
PaER- ERK-R	CAAGAGAAGTGCTTGGGGTCATGGATCCTTTTTTCTTACGTTTTTC

Table S2 Comparison of *PaER* with other OYEs regarding half-lives.

Enzyme	Species	Temperature (°C)	half-life	Reference
OYERo2	<i>Rhodococcus opacus</i>	32	28 min	1
OYERo2a	<i>Rhodococcus opacus</i>	32	87 min	1
FOYE-1	<i>Ferrovum</i> sp. JA12	50	5 h	2
OYE2p	<i>Saccharomyces cerevisiae</i>	40	11 h	3
ClER	<i>Clavispora lusitaniae</i>	40	36 h	4
MgER	<i>Meyerozyma guilliermondii</i>	40	60 h	5
CrOYE3	<i>Chlamydomonas reinhardtii</i>	40	46 h	6
CrOYE1	<i>Chlamydomonas reinhardtii</i>	40	137 h	6
CrOYE2	<i>Chlamydomonas reinhardtii</i>	40	134 h	6
<i>PaER</i>	<i>Pichia angusta</i>	40	89 h	This study

Table S3 Kinetic parameters of *PaER*.

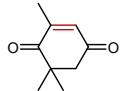
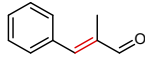
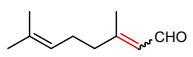
Substrate	K_m (mM)	k_{cat} (s ⁻¹)	k_{cat}/K_m (s ⁻¹ mM ⁻¹)
3a 	0.33	3.57	10.64
10a 	0.064	2.42	37.66
12a 	0.038	0.54	14.45

Table S4 Comparison of different ene-reductases for asymmetric reduction of ketoisophorone

Catalyst	Source	Concentration (mM)	Cofactor	Conversion (%)	<i>ee</i> (%)/config.	Reference
XenA	<i>Pseudomonas putida</i>	5	NADP ⁺ / 0.2 mM	98.9	2.25 (<i>S</i>)	7
PETNR	<i>Enterobacter cloacae</i> st. PB2	5	NADP ⁺ /0.01 mM	>99	57 (<i>R</i>)	8
Gox0502 ^a	<i>Gluconobacter. oxydans</i>	10	NADP ⁺ / 0.5 mM	>99	>99 (<i>R</i>)	9
SynER	<i>Synechococcus</i> sp. PCC 7942	10	NADP ⁺ /0.5 mM	93%	97 (<i>R</i>)	10
TsER C25D/I67T	<i>Thermus scotoductus</i> SA-01	125	NADP ⁺ /0.41 mM	>99	98 (<i>R</i>)	11
CIER	<i>Clavispora lusitaniae</i>	500	NADP ⁺ / 0.5 mM	>99	98 (<i>R</i>)	4
CYE	<i>Candida macedoniensis</i>	658	NADP ⁺ / 0.784 mM	95.4	n.d.	12
<i>Geobacillus</i> ene reductase	<i>Geobacillus</i> sp. 30	1000	NADH/1.0 mM	63.2	89.2 (<i>R</i>)	13
PaER	<i>Pichia angusta</i>	1000	NADP ⁺ /0.2 mM	>99%	>99 (<i>R</i>)	This study

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