

## Supporting Information

### Aggregation Induced Emission Based Fluorenes as Dual–Channel Fluorescent Probes for Rapid Detection of Cyanide: Applications of Smartphone and Logic Gate

Shumaila Majeed<sup>a</sup>, Muhammad Tahir Waseem<sup>a</sup>, Hafiz Muhammad Junaid<sup>a</sup>, Gul Shahzada Khan<sup>b</sup>, Shamyla Nawazish<sup>c</sup>, Tariq Mahmood<sup>a,b</sup>, Asad Muhammad Khan<sup>a</sup>, Sohail Anjum Shahzad<sup>a,\*</sup>

<sup>a</sup> Department of Chemistry, COMSATS University Islamabad, Abbottabad Campus, University Road, Abbottabad 22060, Pakistan

<sup>b</sup> Department of Chemistry, College of Science, University of Bahrain, P.O. Box 32038, Bahrain

<sup>c</sup> Department of Environmental Sciences, COMSATS University Islamabad, Abbottabad Campus, Abbottabad 22060, Pakistan

\* Corresponding Author: Department of Chemistry, COMSATS University Islamabad, Abbottabad Campus, University Road, Abbottabad 22060, Pakistan (S.A. Shahzad). E-mail address: [sashahzad@cuiatd.edu.pk](mailto:sashahzad@cuiatd.edu.pk) (S.A. Shahzad).

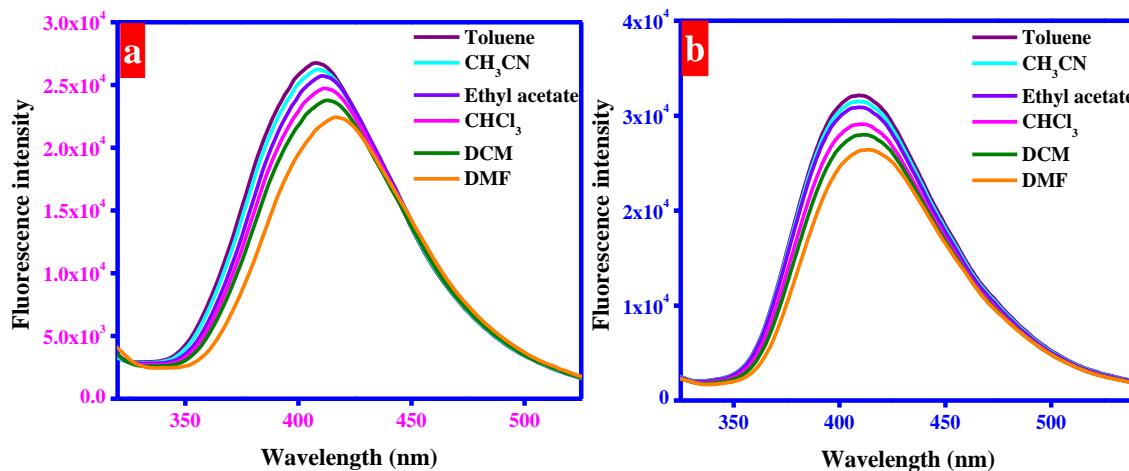
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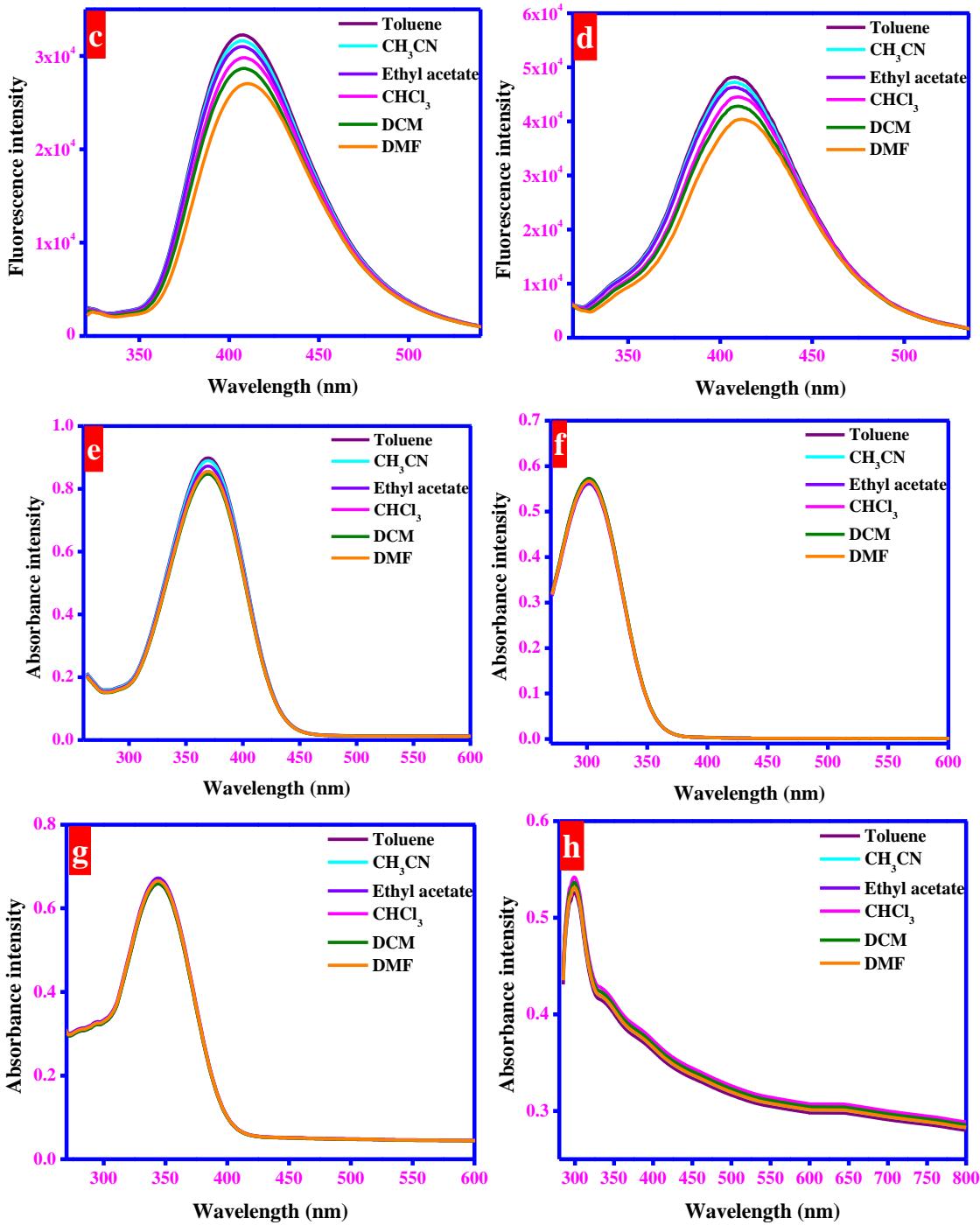
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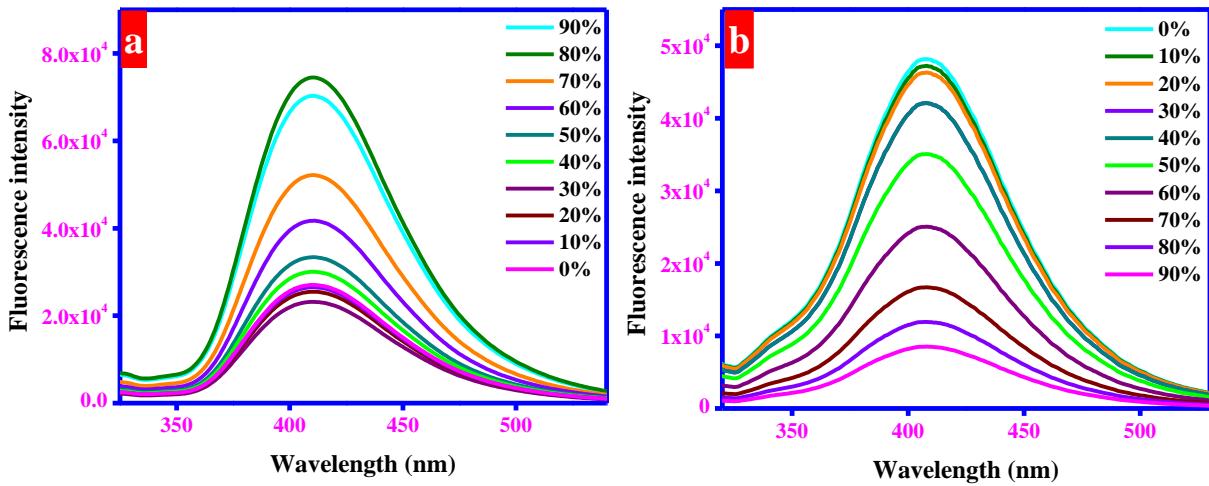
## SI-1. Instruments and reagents

The NMR ( $^1\text{H}$ ,  $^{13}\text{C}$ , and DEPT–135) spectra of all newly synthesized compounds (**1a**, **1b**, **1c**, and **1d**) were recorded at Bruker Avance 400 MHz nuclear magnetic resonance (NMR) spectrometer using  $\text{CDCl}_3$  as a solvent and tetramethylsilane (TMS) as a reference. Further, spectrofluorometer (model FluoroMax-Plus-P-C, Horiba Jobin Yvon Technology, USA) was used to record fluorescence emission spectra. The average particle size of compounds **1a** and **1b** was measured through Dynamic light scattering (DLS, model ZSP Malvern Analytical technology, UK). UV–visible studies were carried out at UV-Vis spectrophotometer (SPECORD 200 PLUS-223E2003C by Analytik Jena, Germany). All reagents and used chemicals were purchased from Alfa Aesar (UK), Daejung Chemicals & Metals (Korea), and Sigma Aldrich (USA) and were used without further purification. Solvents and chemicals utilized in this study include 9*H*-fluorene, ethanol, potassium tertiary butoxide, *N,N*-dimethylbenzaldehyde, 4-ethoxybenzaldehyde, 4-methoxybenzaldehyde, and 3-nitrobenzaldehyde, DCM, brine, distilled water, anhydrous  $\text{MgSO}_4$ , *n*-hexane, ethyl acetate, tetrahydrofuran (THF), ethanol, methanol, acetonitrile ( $\text{CH}_3\text{CN}$ ), dichloromethane (DCM), chloroform ( $\text{CHCl}_3$ ), toluene,  $\text{CN}^-$ ,  $\text{F}^-$ ,  $\text{Cl}^-$ ,  $\text{Br}^-$ ,  $\text{I}^-$ ,  $\text{ClO}_4^-$ ,  $\text{N}_3^-$ ,  $\text{NO}_3^-$ ,  $\text{H}_2\text{PO}_4^-$ ,  $\text{NO}_2^-$ , and  $\text{CH}_3\text{COO}^-$ .

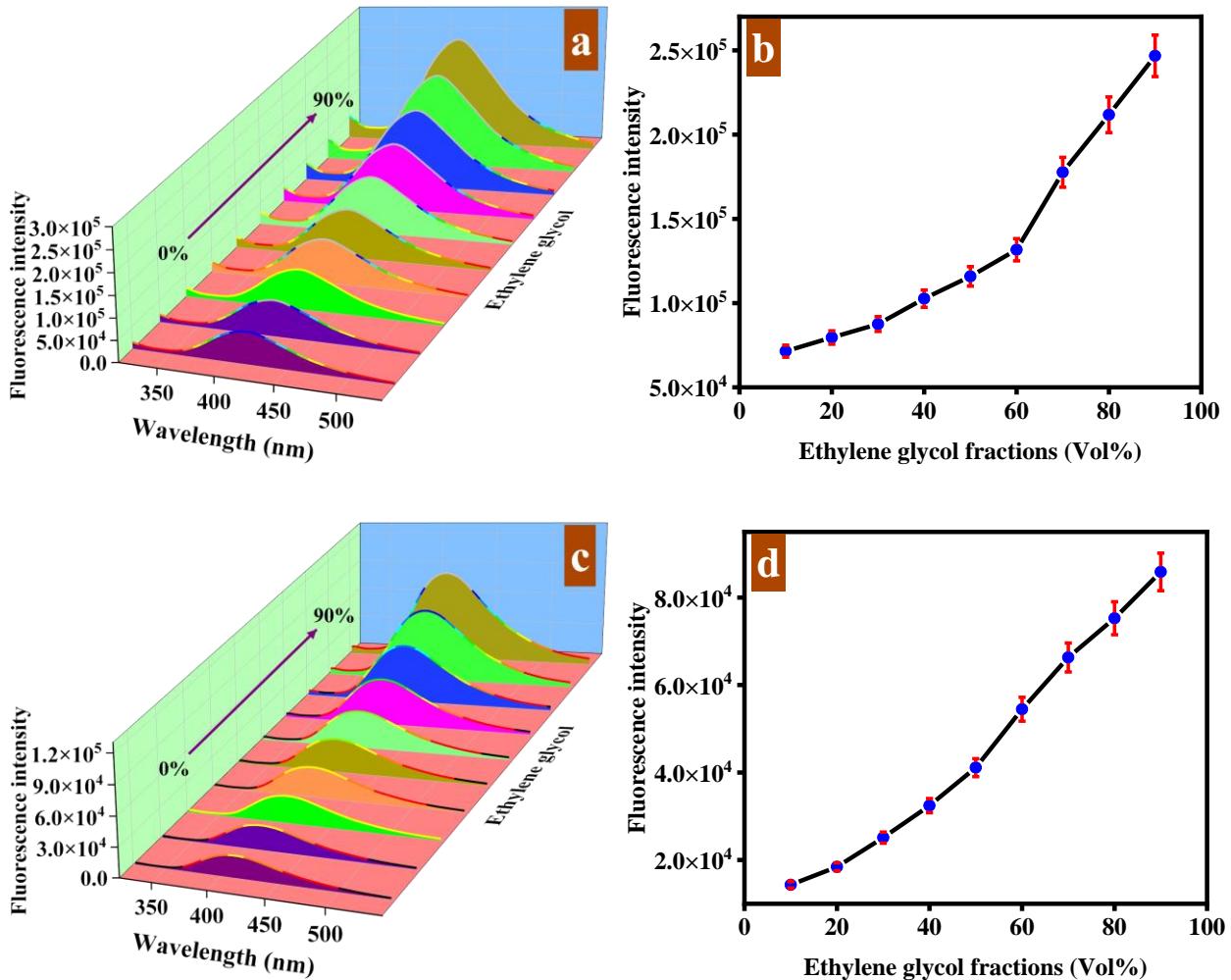


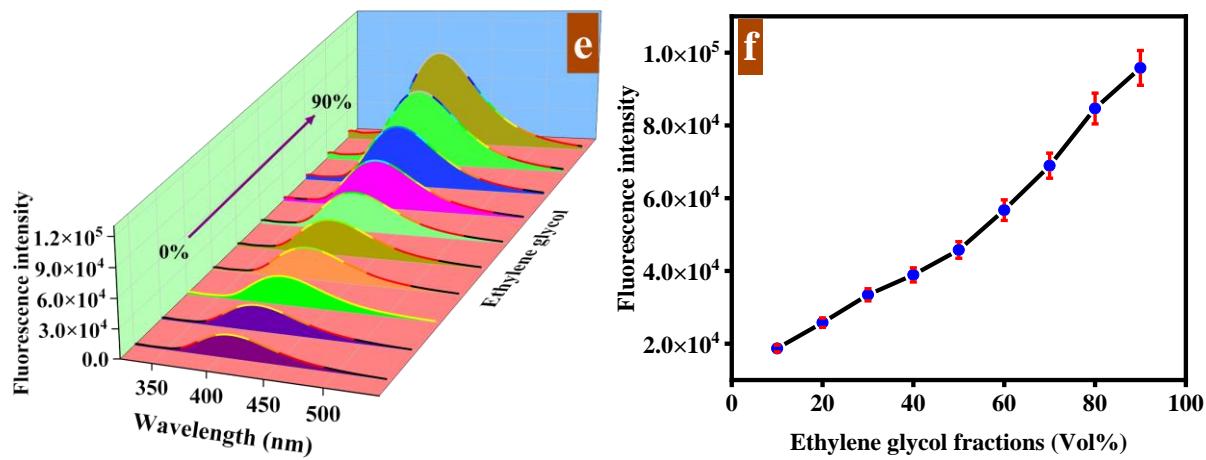


**Fig. S1.** Fluorescence and absorption spectra of probes **1a** (a and e), **1b** (b and f), **1c** (c and g), and **1d** (d and h) in different solvents (conc. = 20 µM (**1a**), 50 µM (**1b**), 10 µM (**1c**), and 20 µM (**1d**); for fluorescence,  $\lambda_{\text{ex}} = 390$  nm; slit width 2/2).

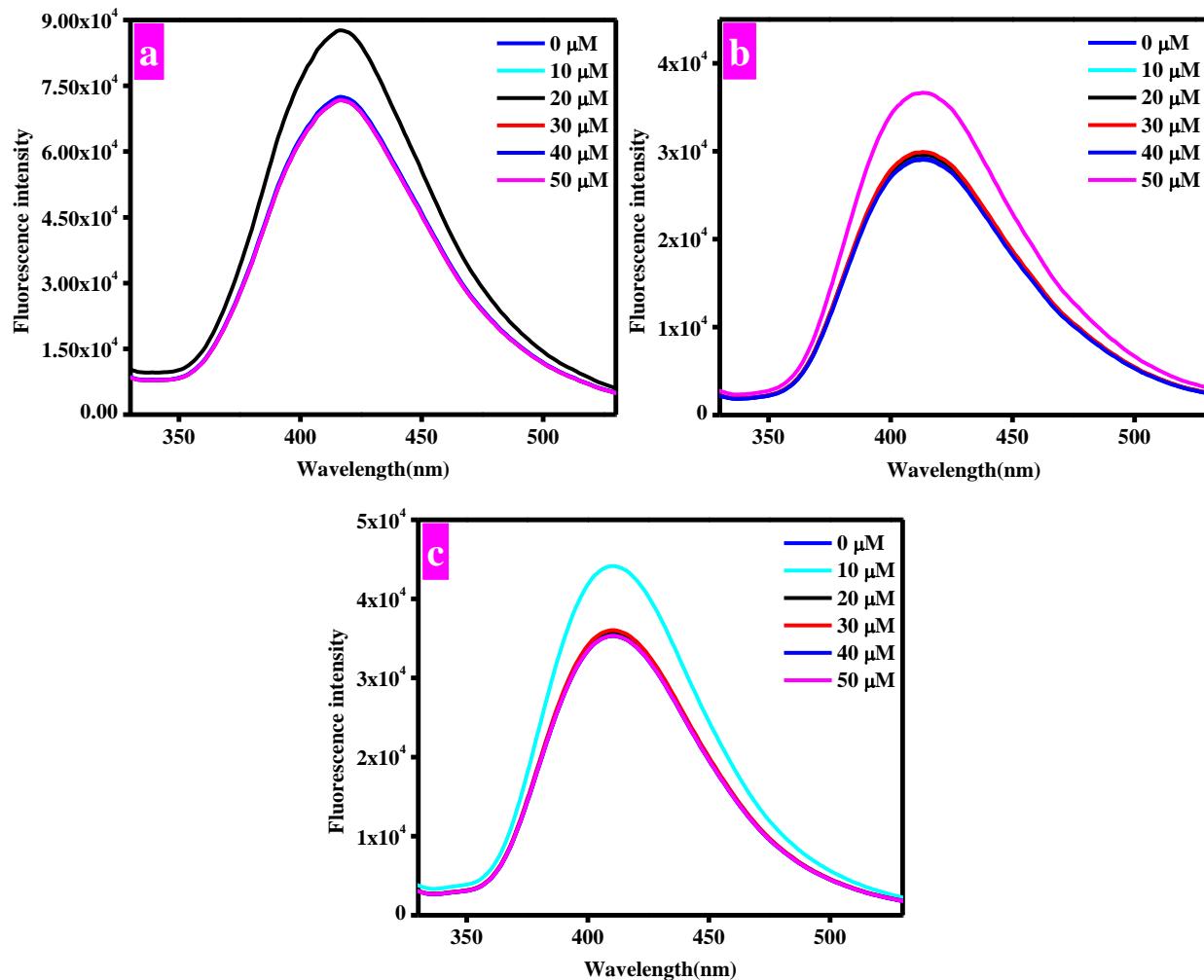


**Fig. S2.** Fluorescence spectra of probes **1c** (a) and **1d** (b) in DMF with increasing water contents from 0 to 90% (conc. = 10 µM (**1c**), and 20 µM (**1d**);  $\lambda_{\text{ex}}=390$  nm; slit width 2/2).

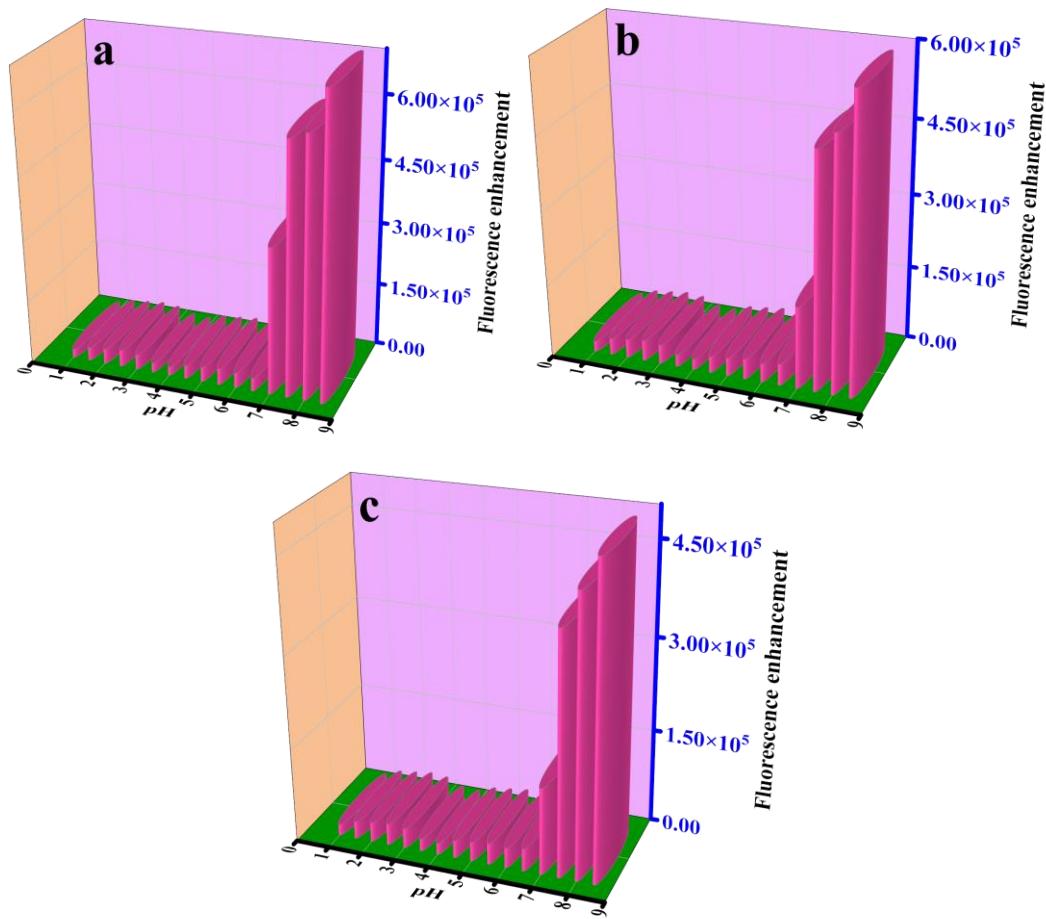




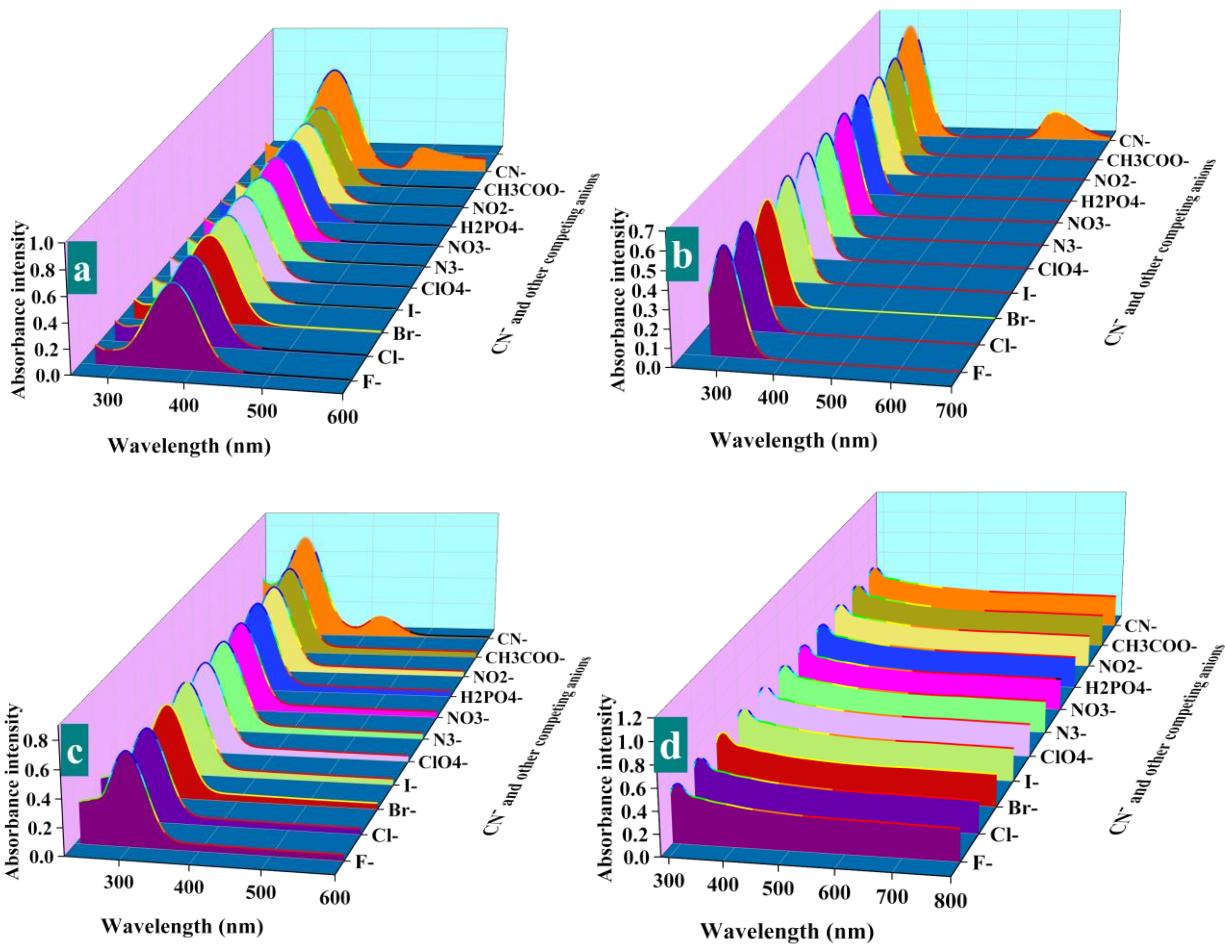
**Fig. S3.** Fluorescence spectra and linear fit plots of probes **1a** (a,b), **1b** (c,d), and **1c** (e,f) illustrating the constant increase in the fluorescence intensity upon regular increase of ethylene glycol contents from 0 to 90% (conc. = 20  $\mu\text{M}$  (**1a**), 50  $\mu\text{M}$  (**1b**), and 10  $\mu\text{M}$  (**1c**);  $\lambda_{\text{ex}}=390$  nm; slit width 2/2).



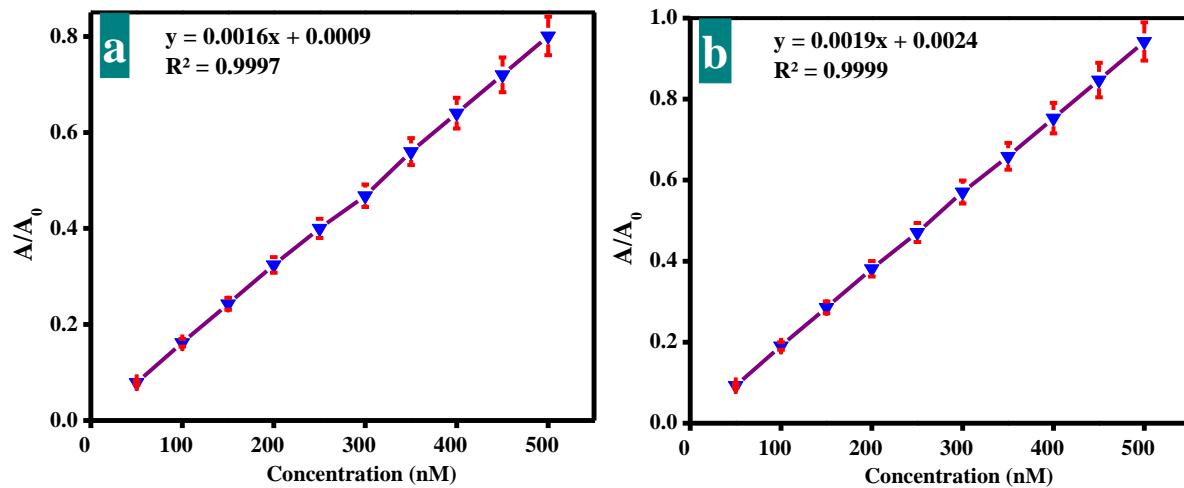
**Fig. S4.** Fluorescence spectra of probes **1a** (a), **1b** (b), and **1c** (c) in H<sub>2</sub>O/DMF (4:1, v/v) with increasing concentrations from 0 to 50  $\mu$ M ( $\lambda_{\text{ex}}=390$  nm; slit width 2/2).

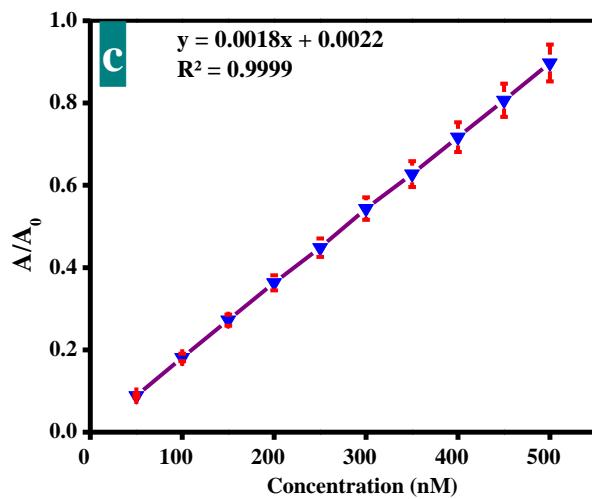


**Fig. S5.** Fluorescence spectra of probes **1a** (a), **1b** (b), and **1c** (c) before and after the addition of CN<sup>-</sup> (500 nM) over a wide pH range (2.0–12.0) (conc. = 20  $\mu$ M (**1a**), 50  $\mu$ M (**1b**), and 10  $\mu$ M (**1c**);  $\lambda_{\text{ex}} = 390$  nm; slit width 2/2).

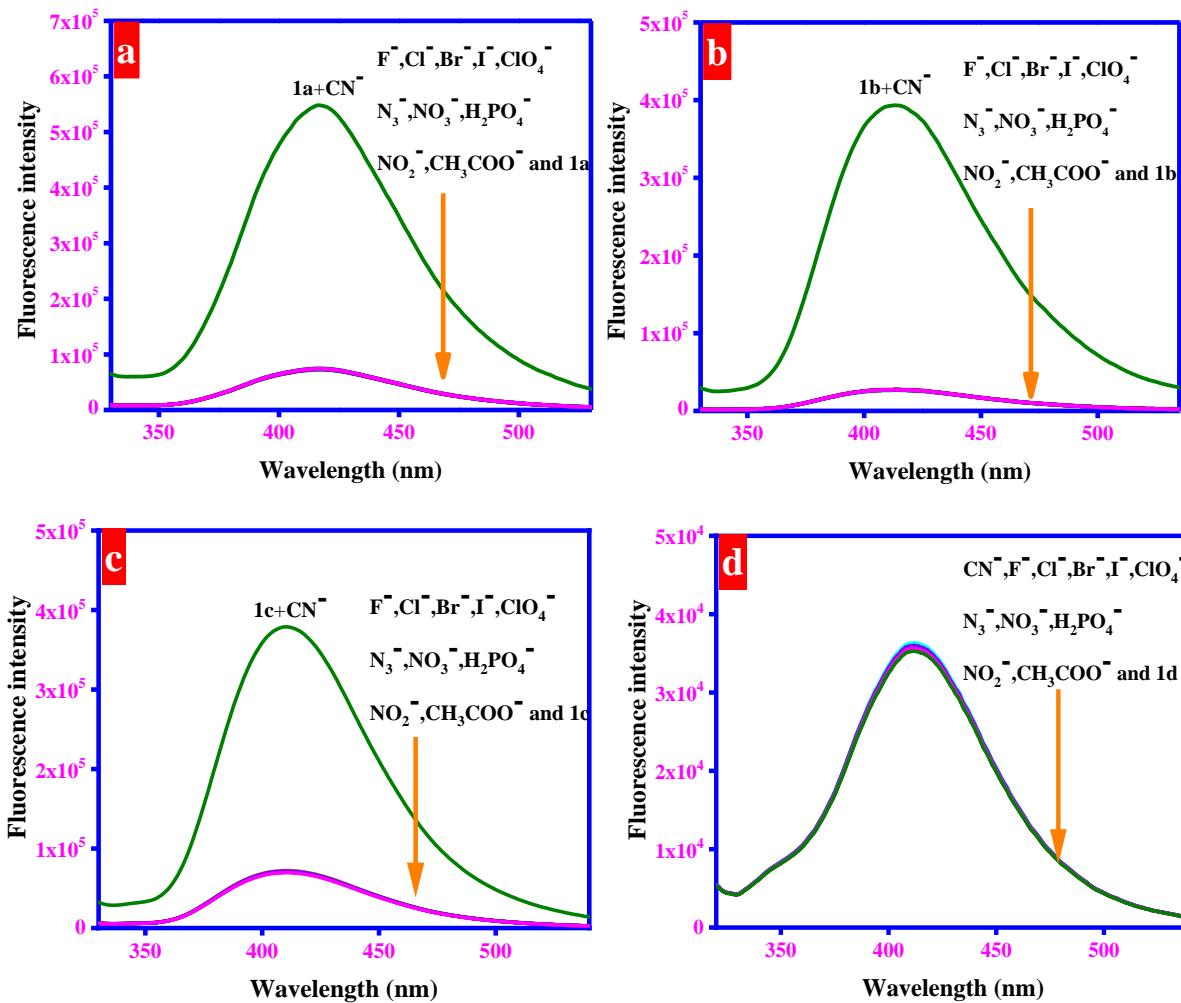


**Fig. S6.** Absorption spectra of probes **1a** (a), **1b** (b), **1c** (c), and **1d** (d) upon the addition of 500 nM of different anions (conc. = 20  $\mu$ M (**1a**), 50  $\mu$ M (**1b**), 10  $\mu$ M (**1c**), and 20  $\mu$ M (**1d**)).

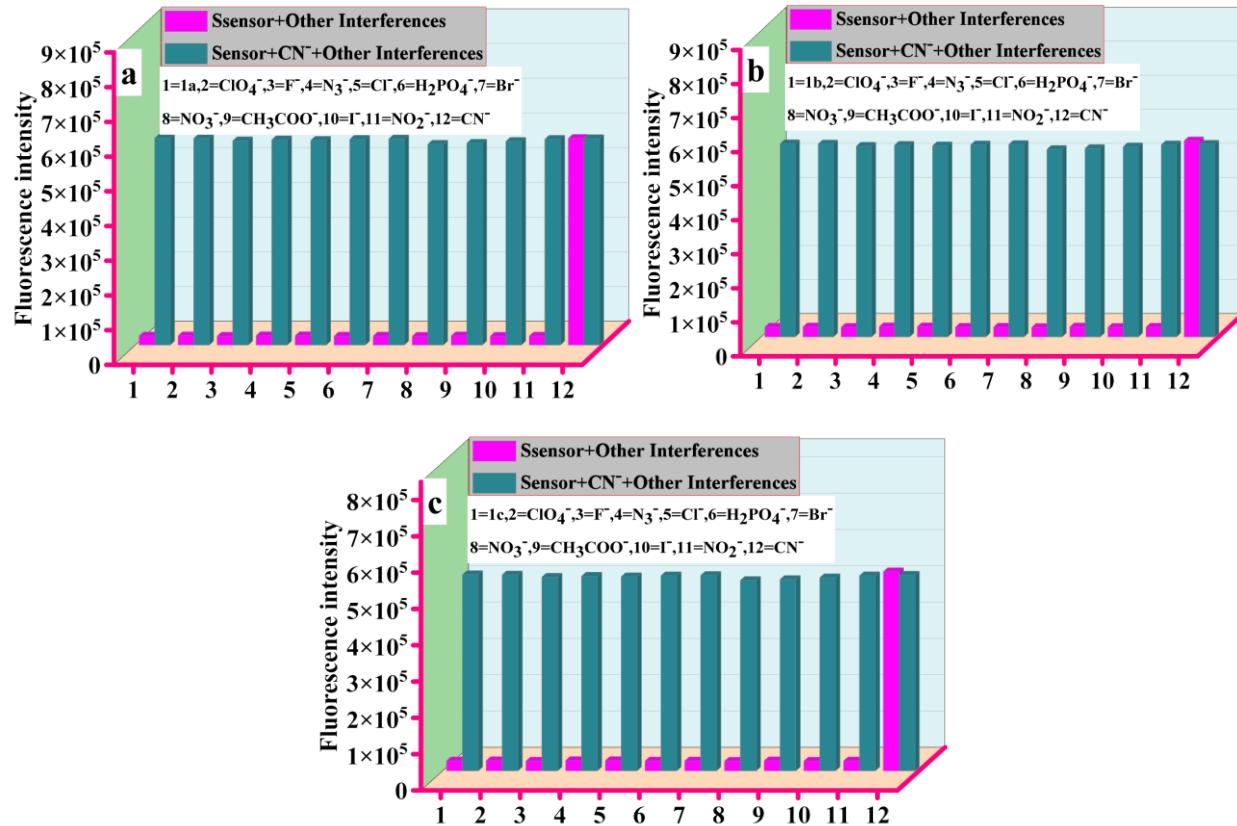




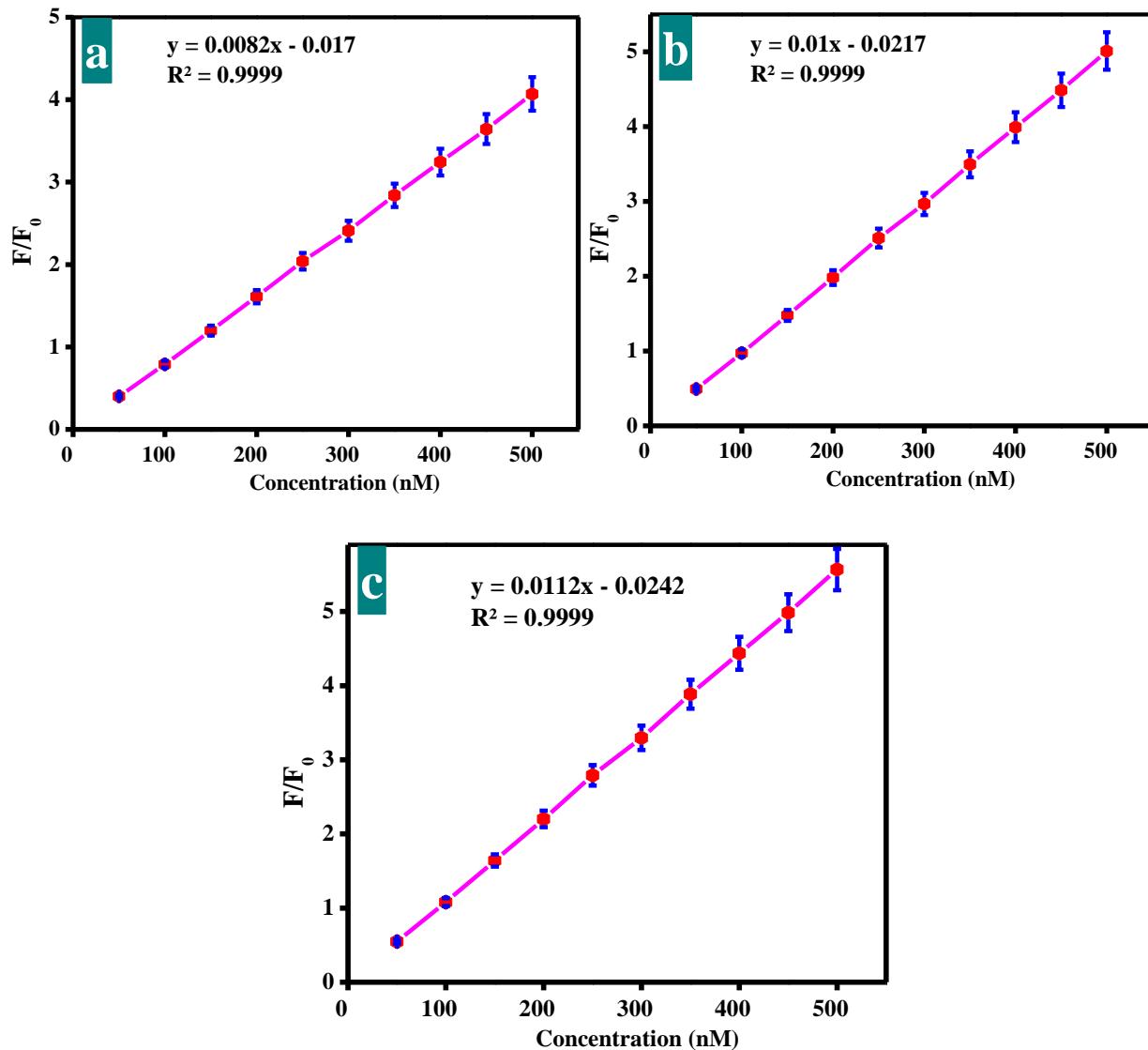
**Fig. S7.** Calibration curves demonstrating the steady elevation in the absorption intensity of probes **1a** (a), **1b** (b) and **1c** (c) upon constant addition (0–500 nM) of  $\text{CN}^-$ .



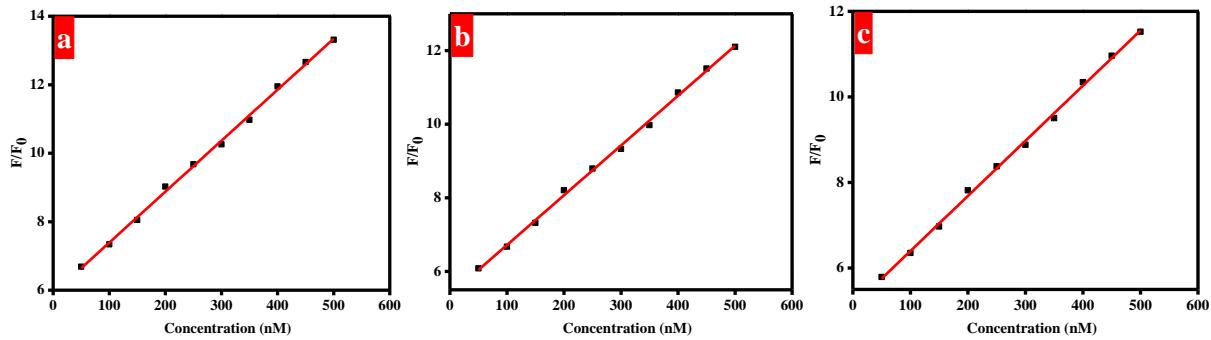
**Fig. S8.** Fluorescence spectra of probes **1a** (a), **1b** (b), **1c** (c), and **1d** (d) upon the addition of 500 nM of different anions (conc. = 20  $\mu$ M (**1a**), 50  $\mu$ M (**1b**), 10  $\mu$ M (**1c**), and 20  $\mu$ M (**1d**);  $\lambda_{\text{ex}} = 390$  nm; slit width 2/2).



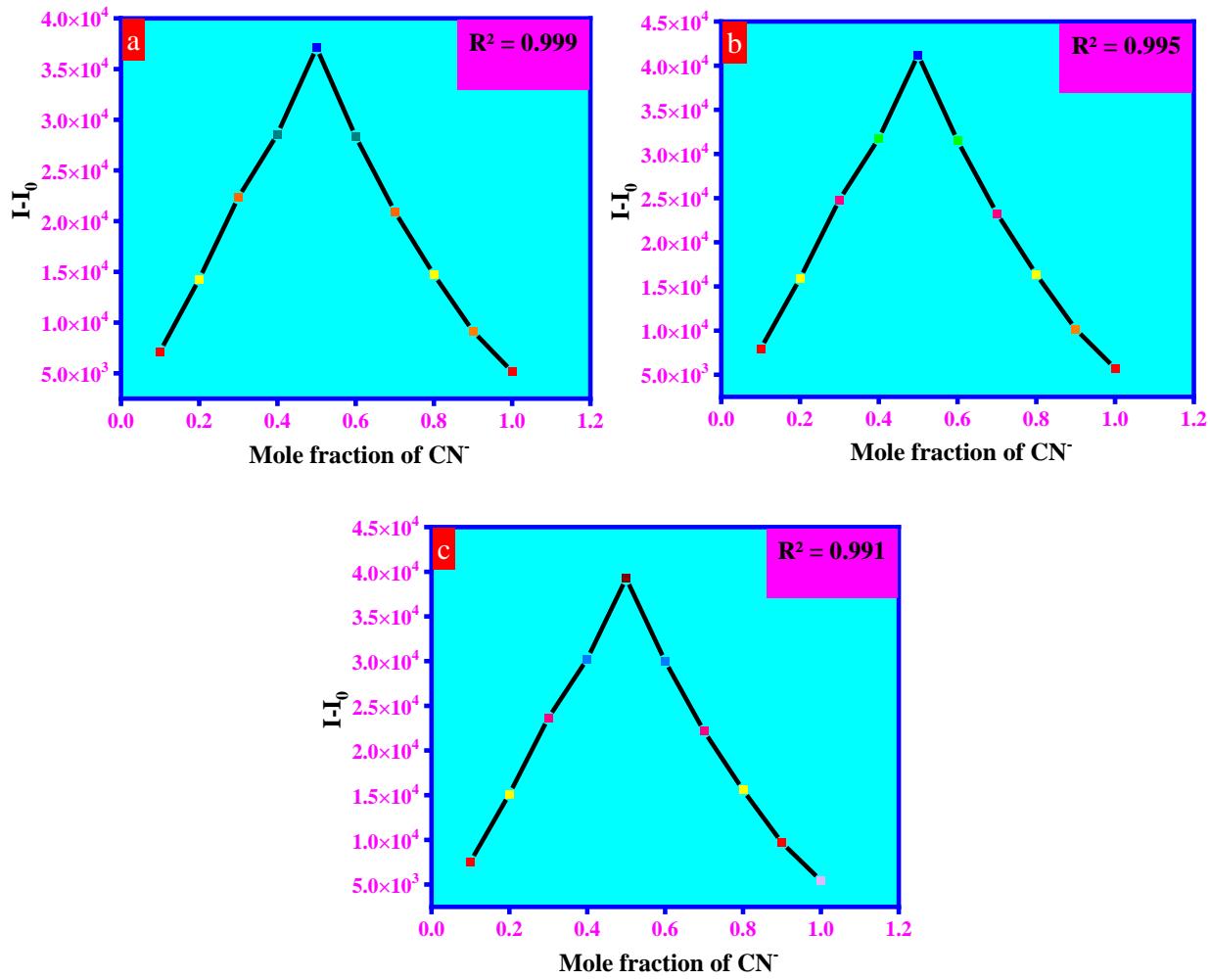
**Fig. S9.** Fluorescence spectra of probes **1a** (a), **1b** (b), and **1c** (c) upon the addition of CN<sup>-</sup> (500 nM) in the presence of other interferences (20 equiv) (conc. = 20  $\mu$ M (**1a**), 50  $\mu$ M (**1b**), and 10  $\mu$ M (**1c**);  $\lambda_{\text{ex}} = 390$  nm; slit width 2/2).



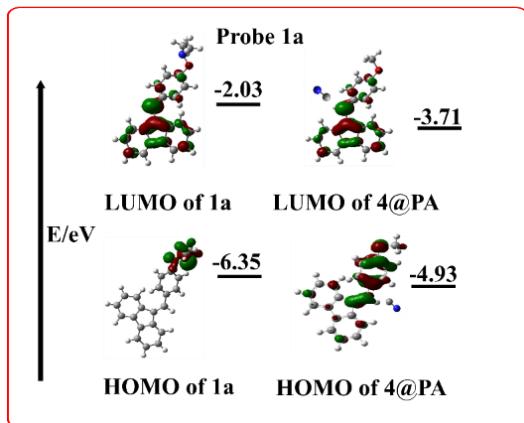
**Fig. S10.** Calibration curves demonstrating the linear fluorescence enhancement response probes **1a** (a), **1b** (b), and **1c** (c) towards  $\text{CN}^-$  ions in the range of 0–500 nM.



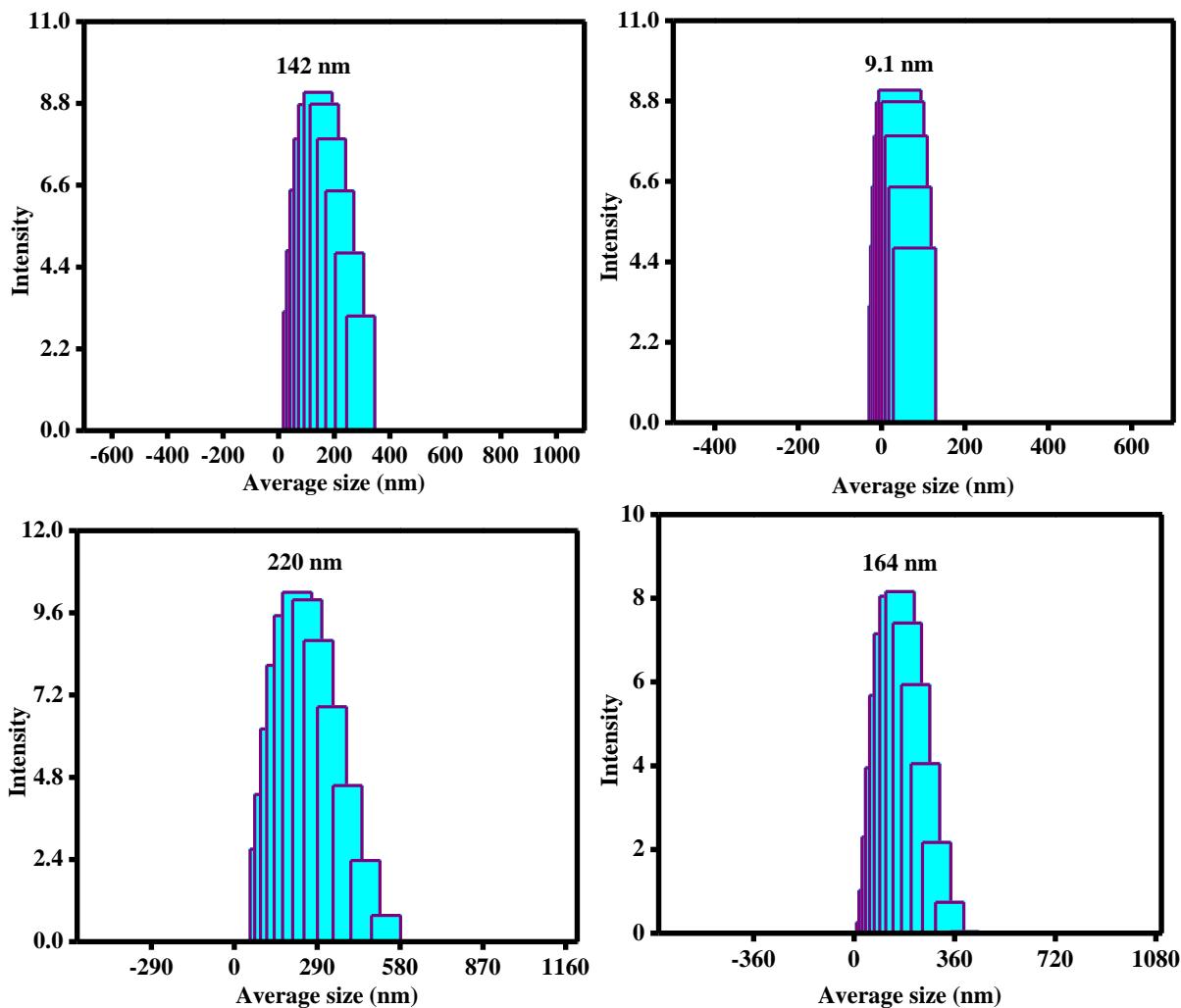
**Fig. S11.** Illustration of linear response of probes **1a** (a), **1b** (b), and **1c** (c) towards cyanide through the Benesi-Hildebrand plots.



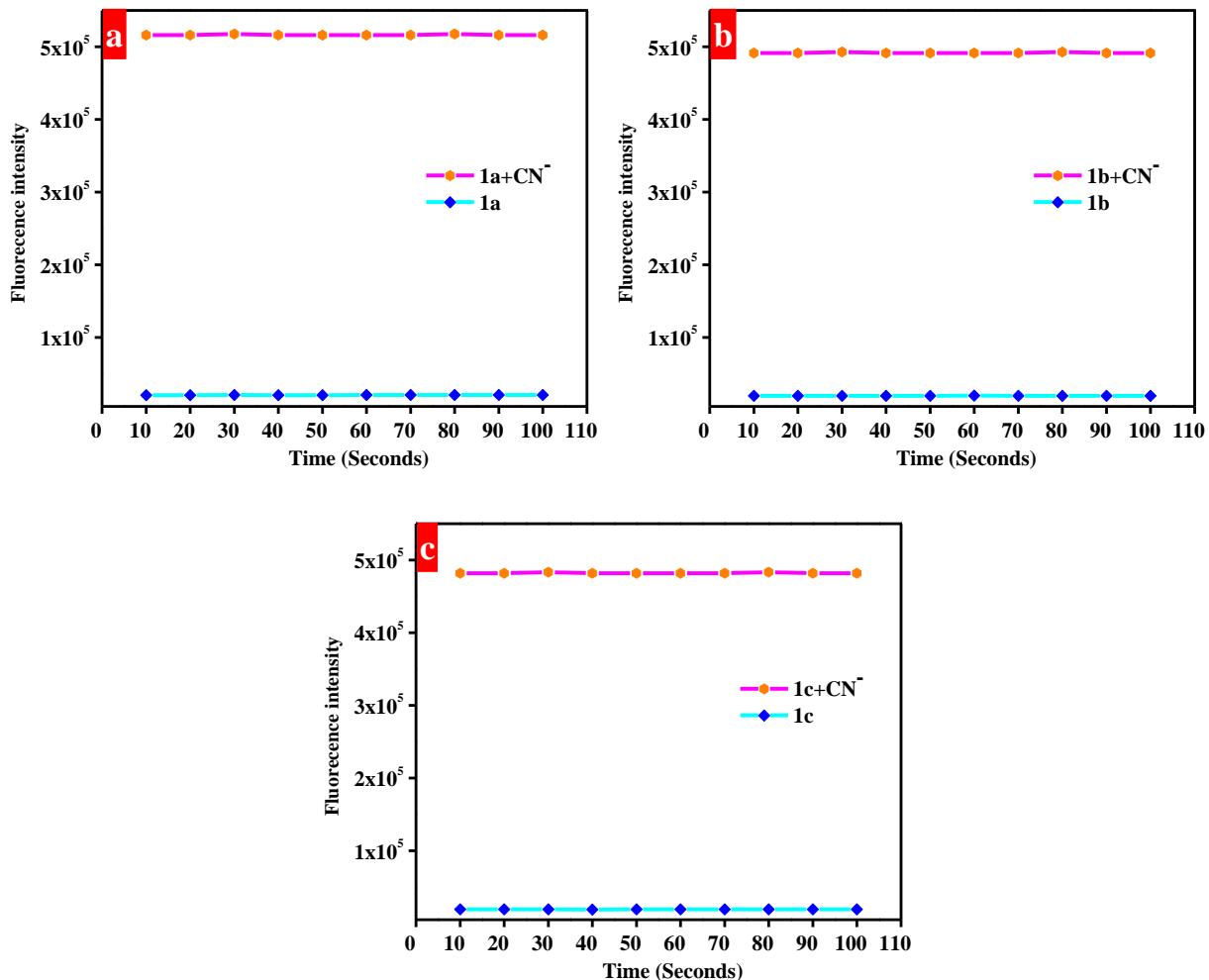
**Fig. S12.** Job's plots of probes **1a** (a), **1b** (b), and **1c** (c) upon the addition of 0–500 nM of  $\text{CN}^-$ .



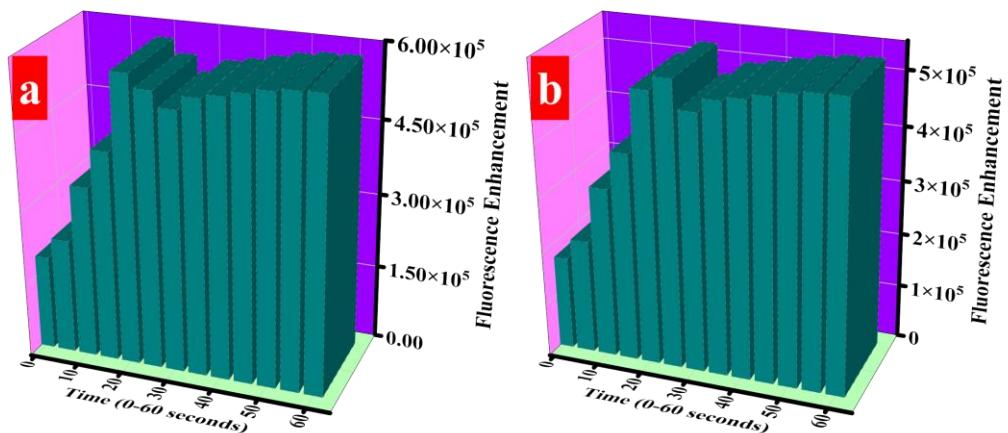
**Fig. S13.** The HOMO–LUMO energy level diagram for probe **1a** and their  $\text{CN}^-$  complexes.

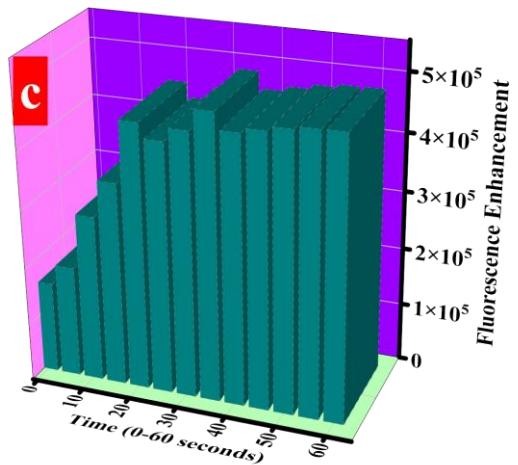


**Fig. S14.** Dynamic light scattering (DLS) measurements of probes **1a** and **1b** before and after the addition of 500 nM of  $\text{CN}^-$  (conc. = 20  $\mu\text{M}$  (**1a**) and 50  $\mu\text{M}$  (**1b**); solvent =  $\text{H}_2\text{O}/\text{DMF}$  (4:1, v/v).

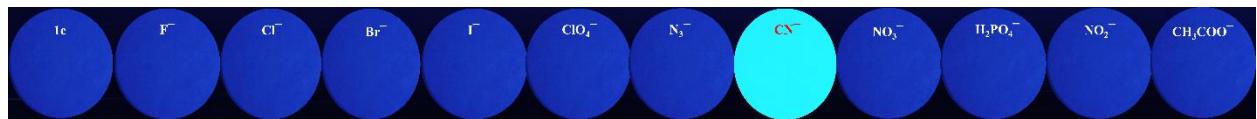


**Fig. S15.** Photostability test of probes **1a** (a), **1b** (b), and **1c** (c) and their probe–CN<sup>−</sup> adducts.

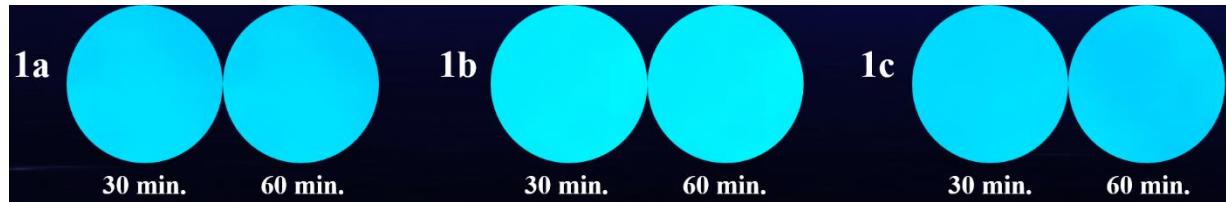




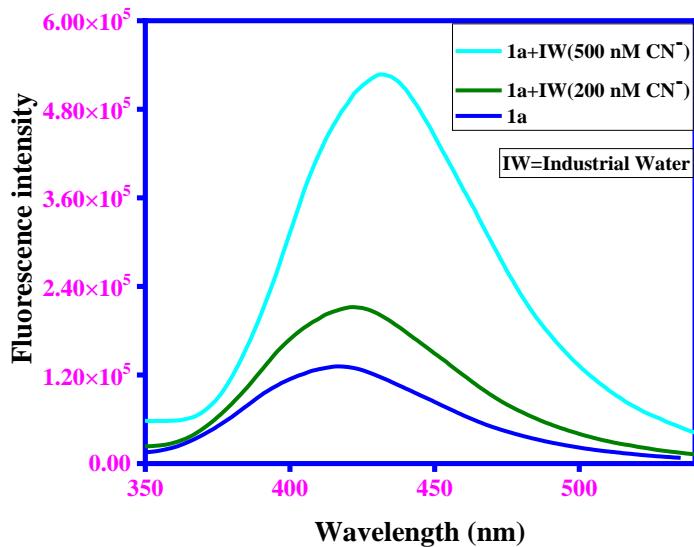
**Fig. S16.** Fluorescence spectra of probes **1a** (a), **1b** (b), and **1c** (c) as a function of time upon the addition of 500 nM of  $\text{CN}^-$  (conc. = 20  $\mu\text{M}$  (**1a**), 50  $\mu\text{M}$  (**1b**) and 10  $\mu\text{M}$  (**1c**);  $\lambda_{\text{ex}} = 390$  nm; slit width 2/2).



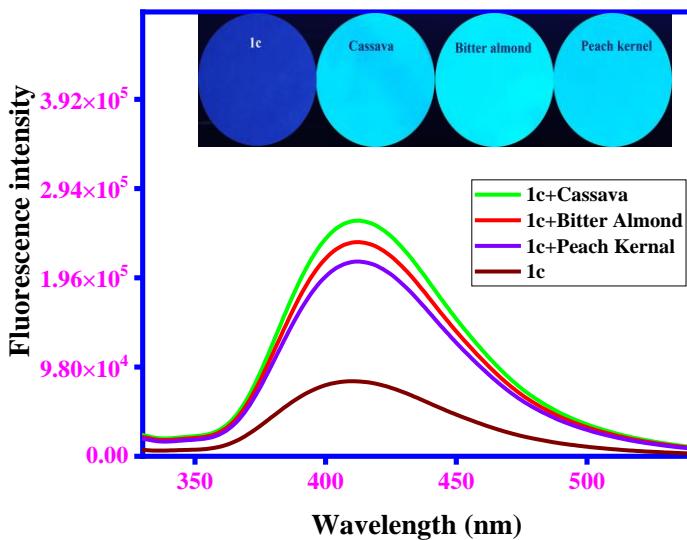
**Fig. S17.** Color change of probes' **1c** coated test strips upon dipping in THF solution of  $\text{CN}^-$  and other anions (500 nM).



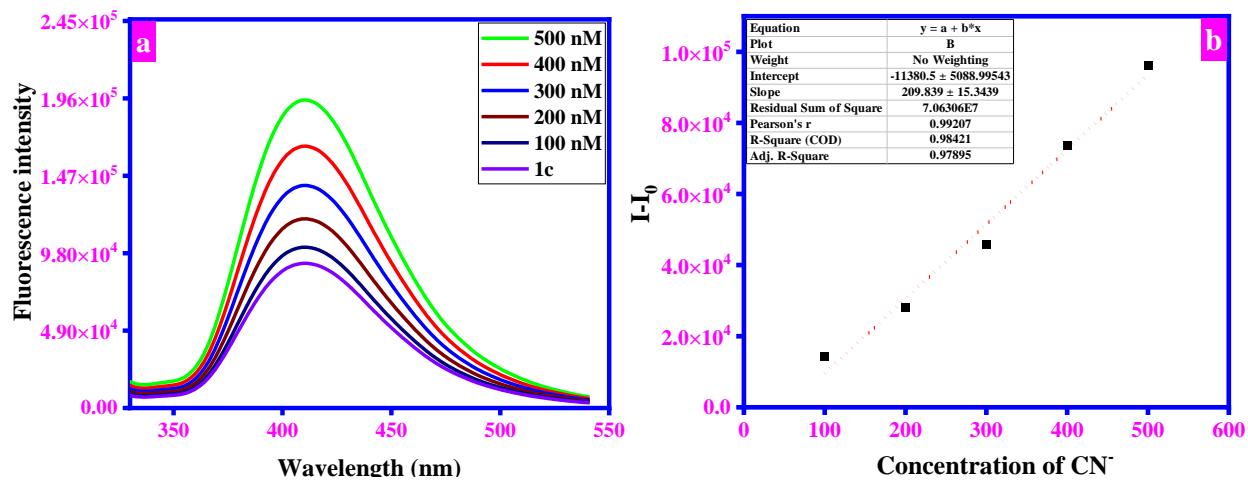
**Fig. S18.** Response of probes' coated tests strips when exposed to  $\text{CN}^-$  after regular interval of time.



**Fig. S19.** Fluorescence spectra of probes **1a** upon the addition of industrial wastewater containing  $\text{CN}^-$  (200 and 500 nM) (conc. = 20  $\mu\text{M}$  (**1a**);  $\lambda_{\text{ex}} = 390$  nm; slit width 2/2).



**Fig. S20.** Fluorescence spectra of probes **1c** upon the addition of  $\text{H}_2\text{O}/\text{DMF}$  (4:1, v/v) solution of peach kernel, bitter almond, and cassava powder (conc. = 20  $\mu\text{M}$  (**1a**), 50  $\mu\text{M}$  (**1b**) and 10  $\mu\text{M}$  (**1c**);  $\lambda_{\text{ex}} = 390$  nm; slit width 2/2).



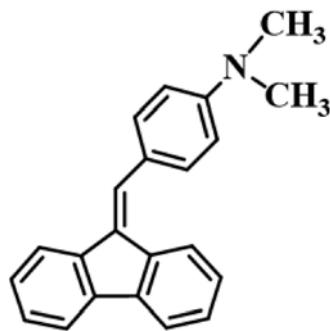
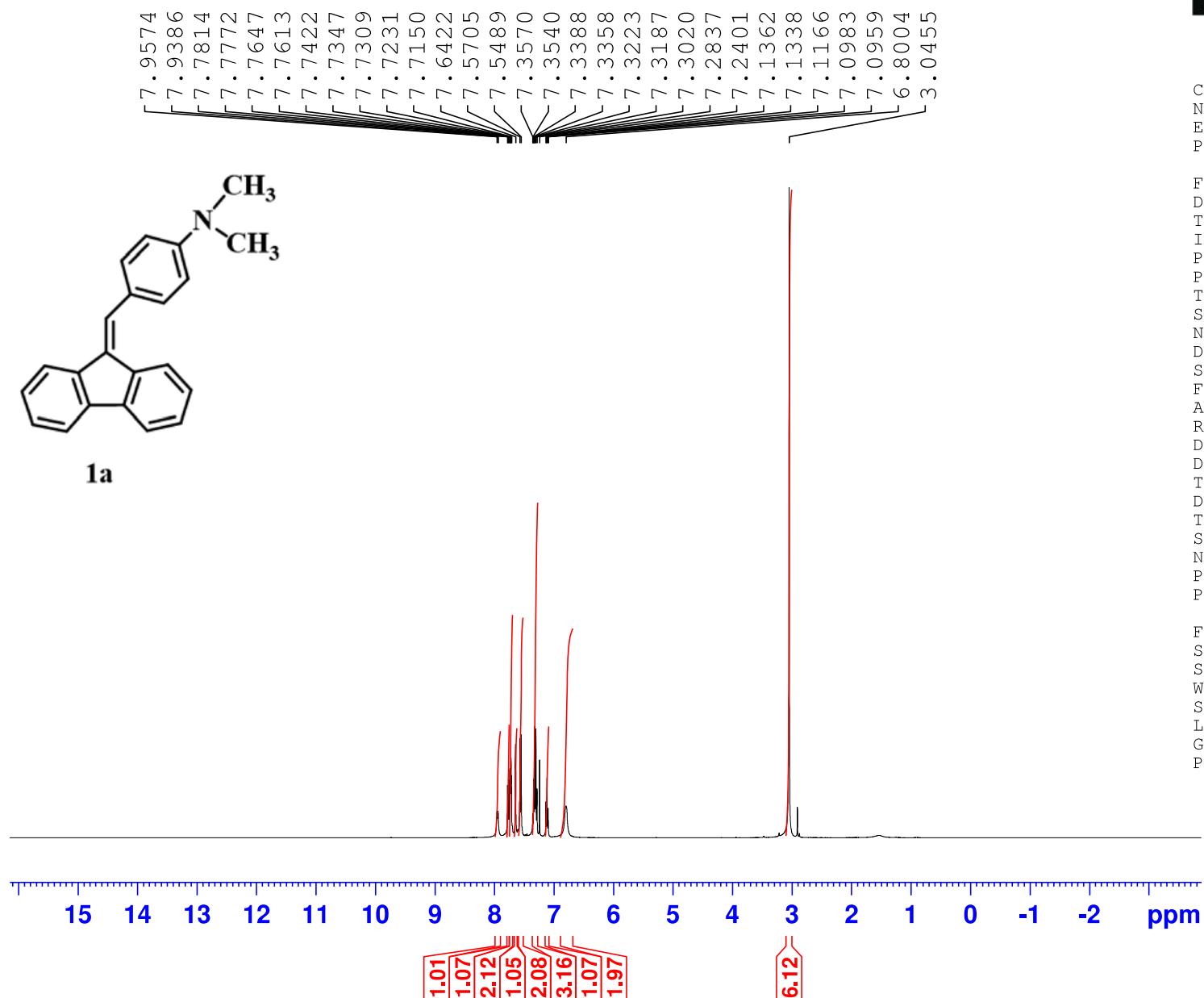
**Fig. S21.** Fluorescence spectra (a) and calibration curve (b) for the quantitation of  $\text{CN}^-$  ions in cassava food samples.

**Table S1.** Analysis of variance (ANOVA) for the linear fit model calculated for the method.

Analyte	Parameter	<sup>a</sup> SS	<sup>b</sup> DF	<sup>c</sup> MS	<sup>d</sup> F
CN <sup>-</sup>	Model	2340304.0	1	2452300.0	4860.9
	Residue	8244.35	12	8162.75	
	Lack of fit	2142.33	3	857.75	0.8955
	Pure error	6755.27	11	773.41	

<sup>a</sup>SS: sum of squares // <sup>b</sup>DF: Degrees of freedom // <sup>c</sup>MQ: mean of the squares // <sup>d</sup>F<sub>critic</sub> = 3.631// <sup>d</sup>F<sub>critic</sub> = 4.098

**Compound 1a, 1H NMR (400 MHz, CDCl<sub>3</sub>)**

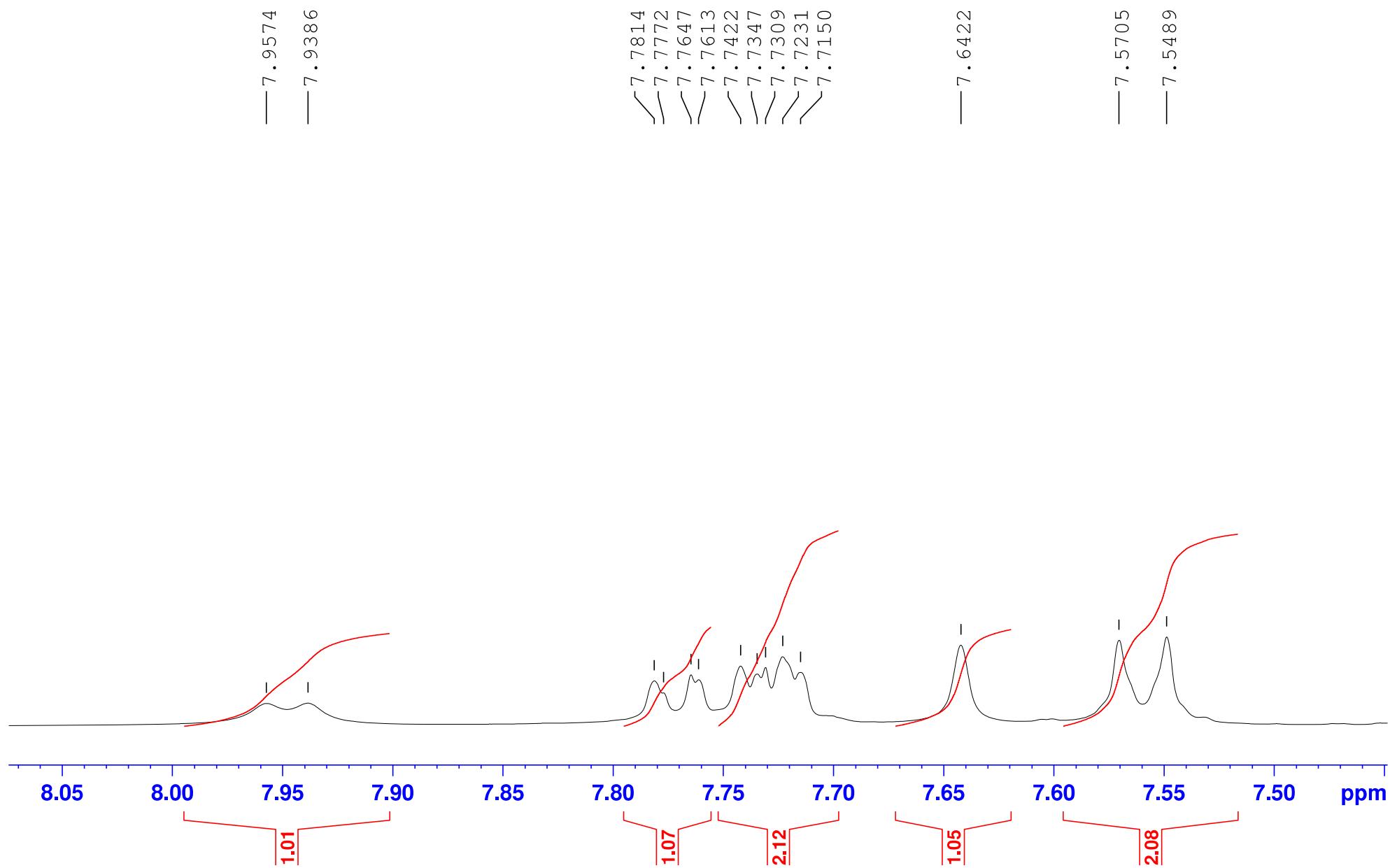


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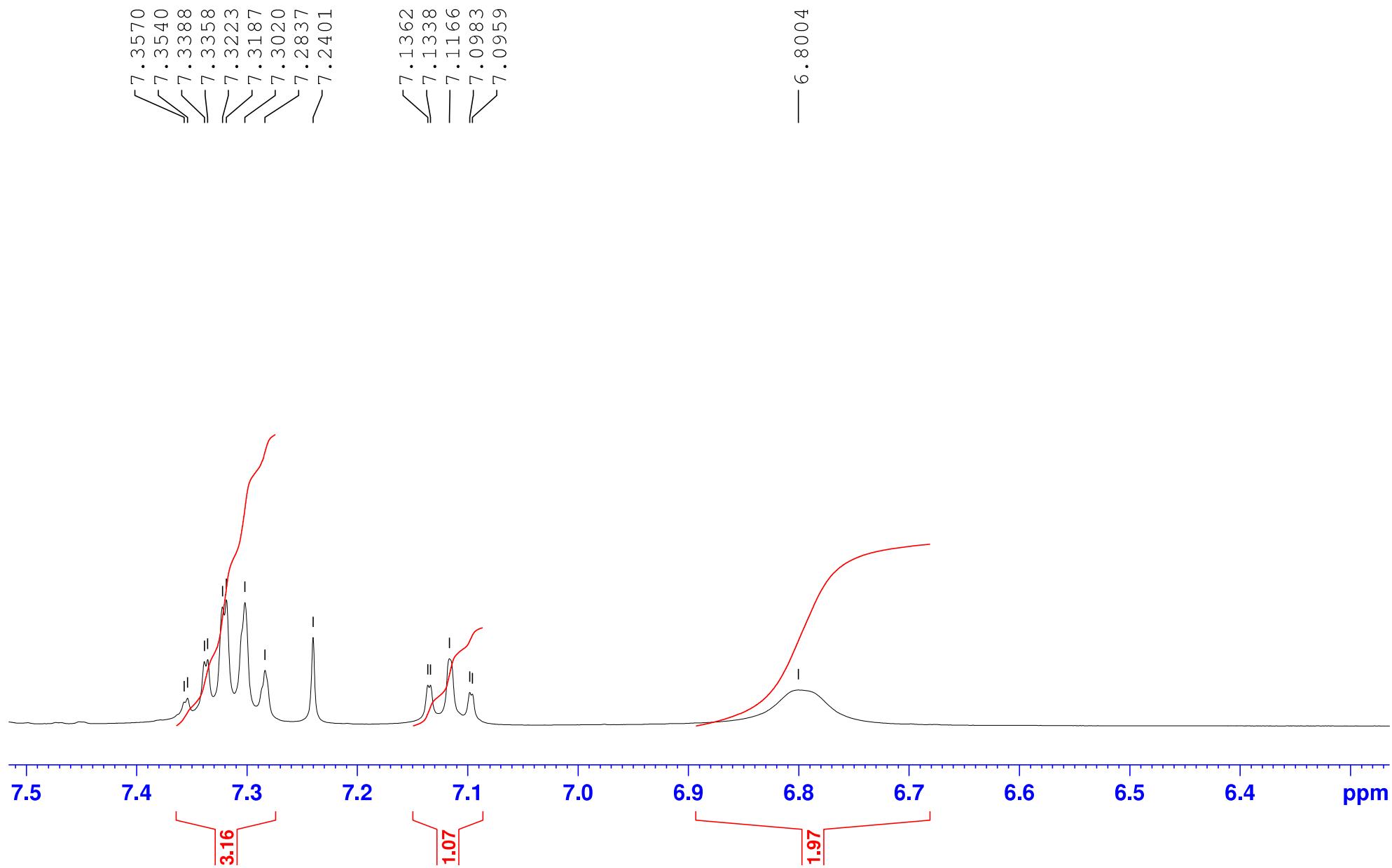
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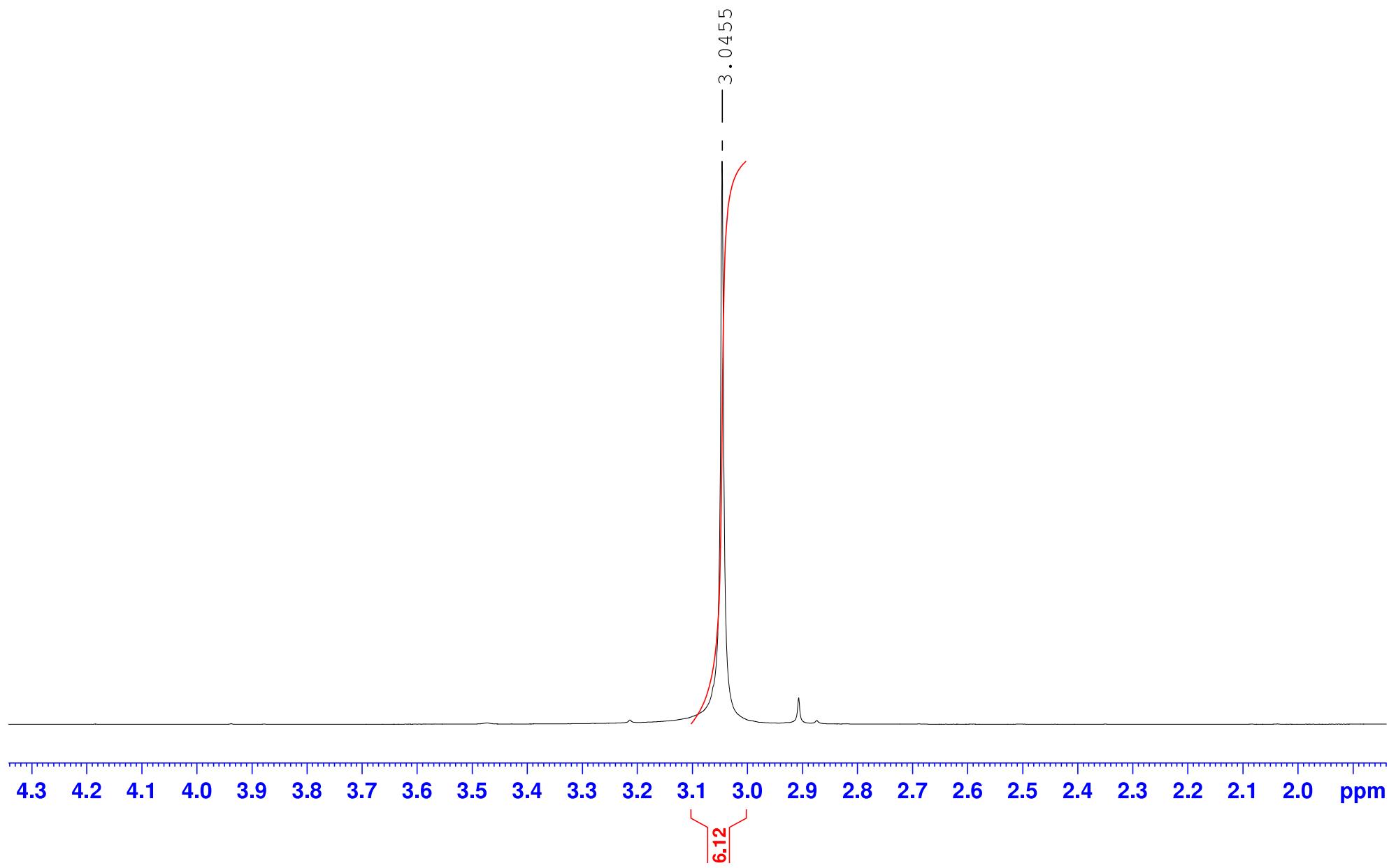
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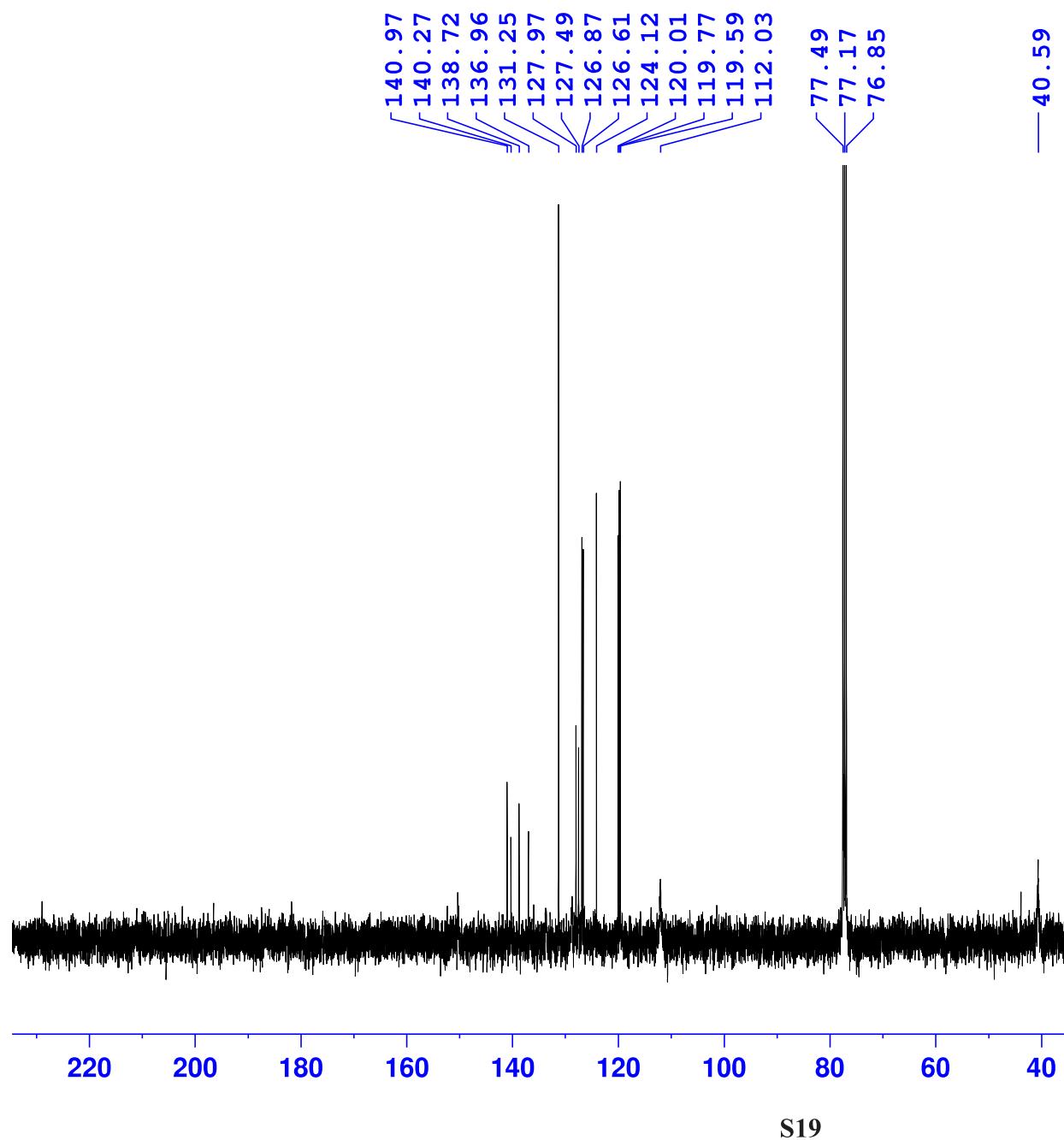
**Compound 1a,  $^1\text{H}$  NMR (400 MHz,  $\text{CDCl}_3$ )**



**Compound 1a,  $^1\text{H}$  NMR (400 MHz,  $\text{CDCl}_3$ )**



Compound 1a,  $^{13}\text{C}$  NMR (100 MHz,  $\text{CDCl}_3$ )

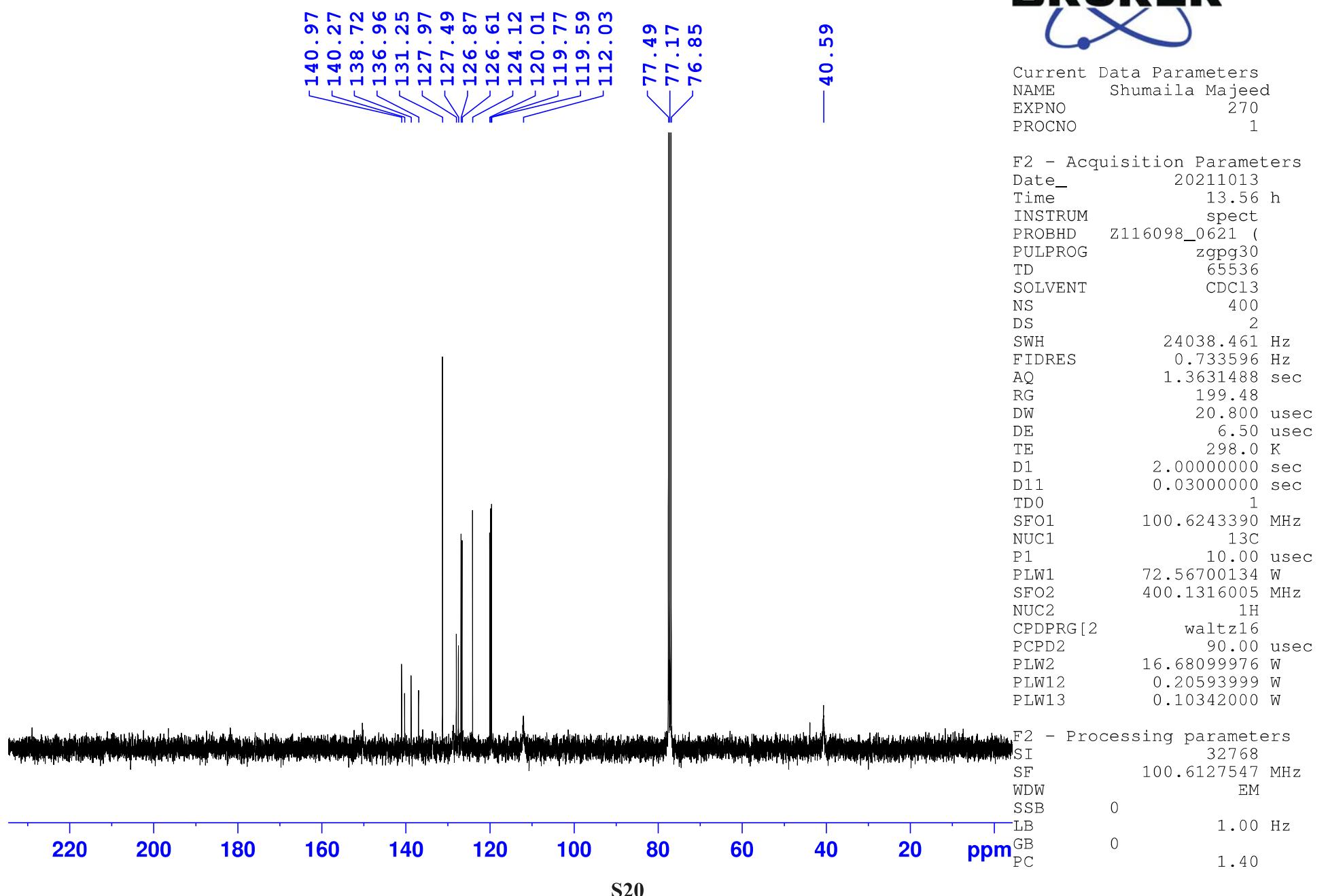


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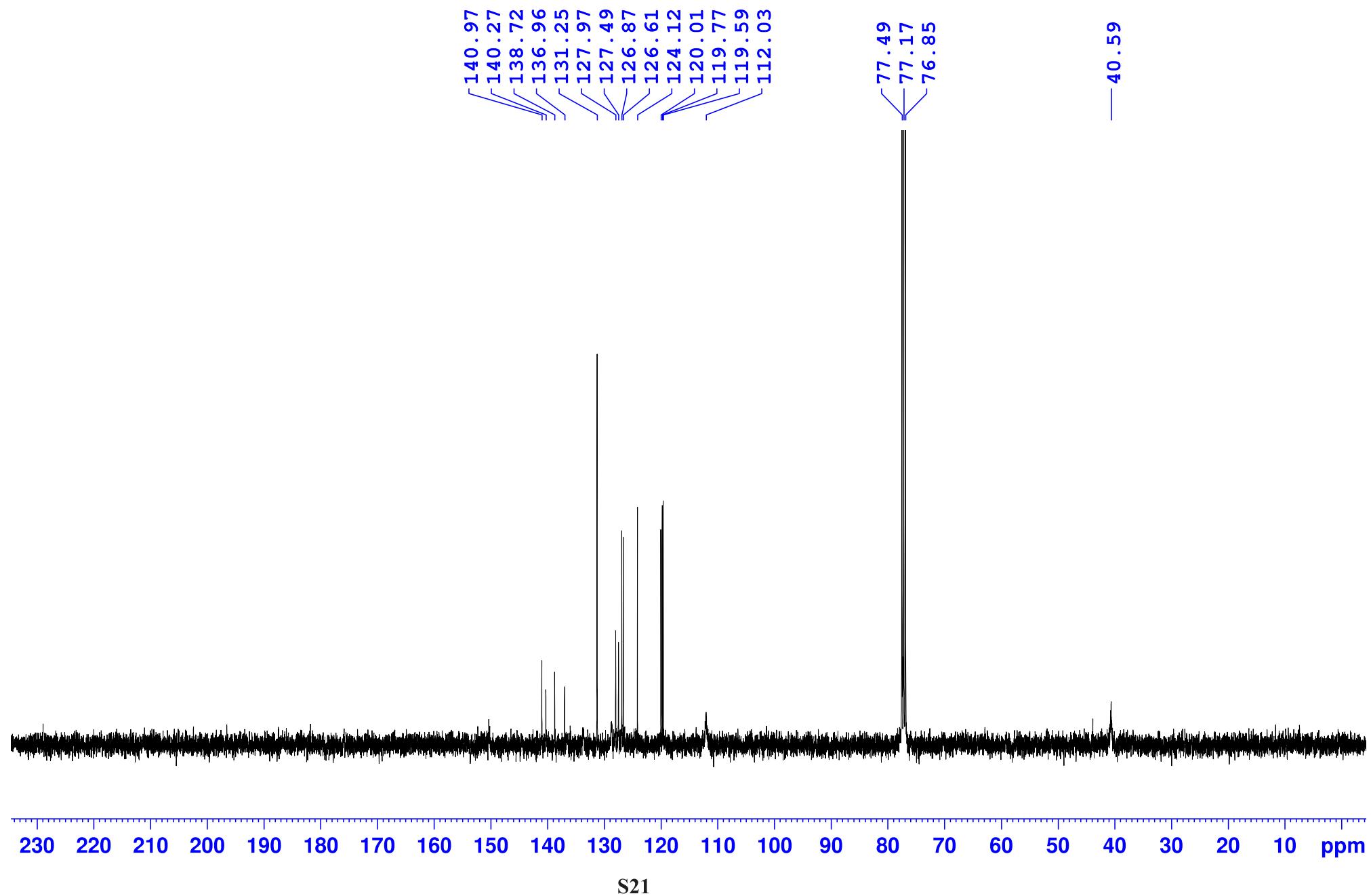
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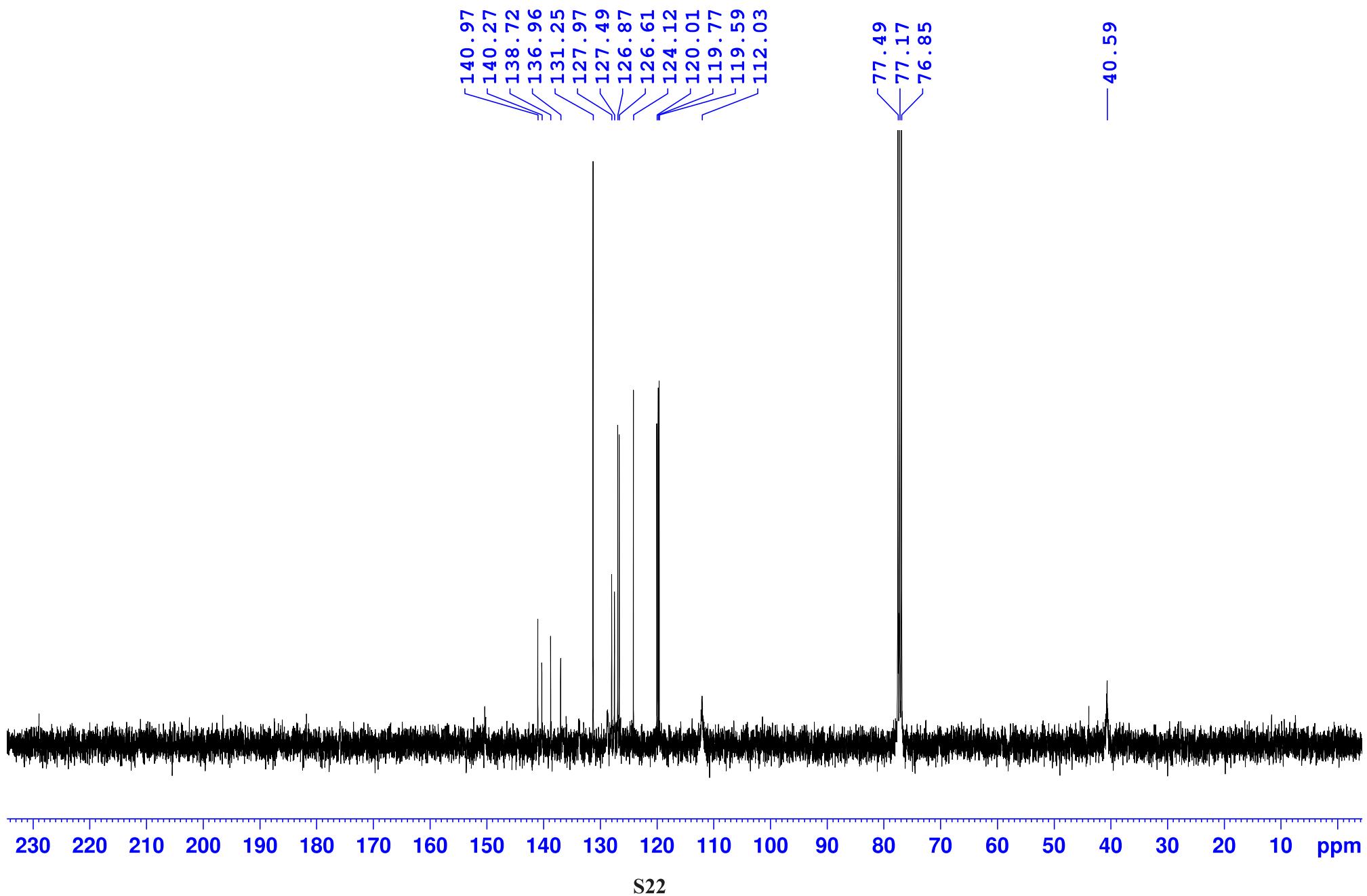
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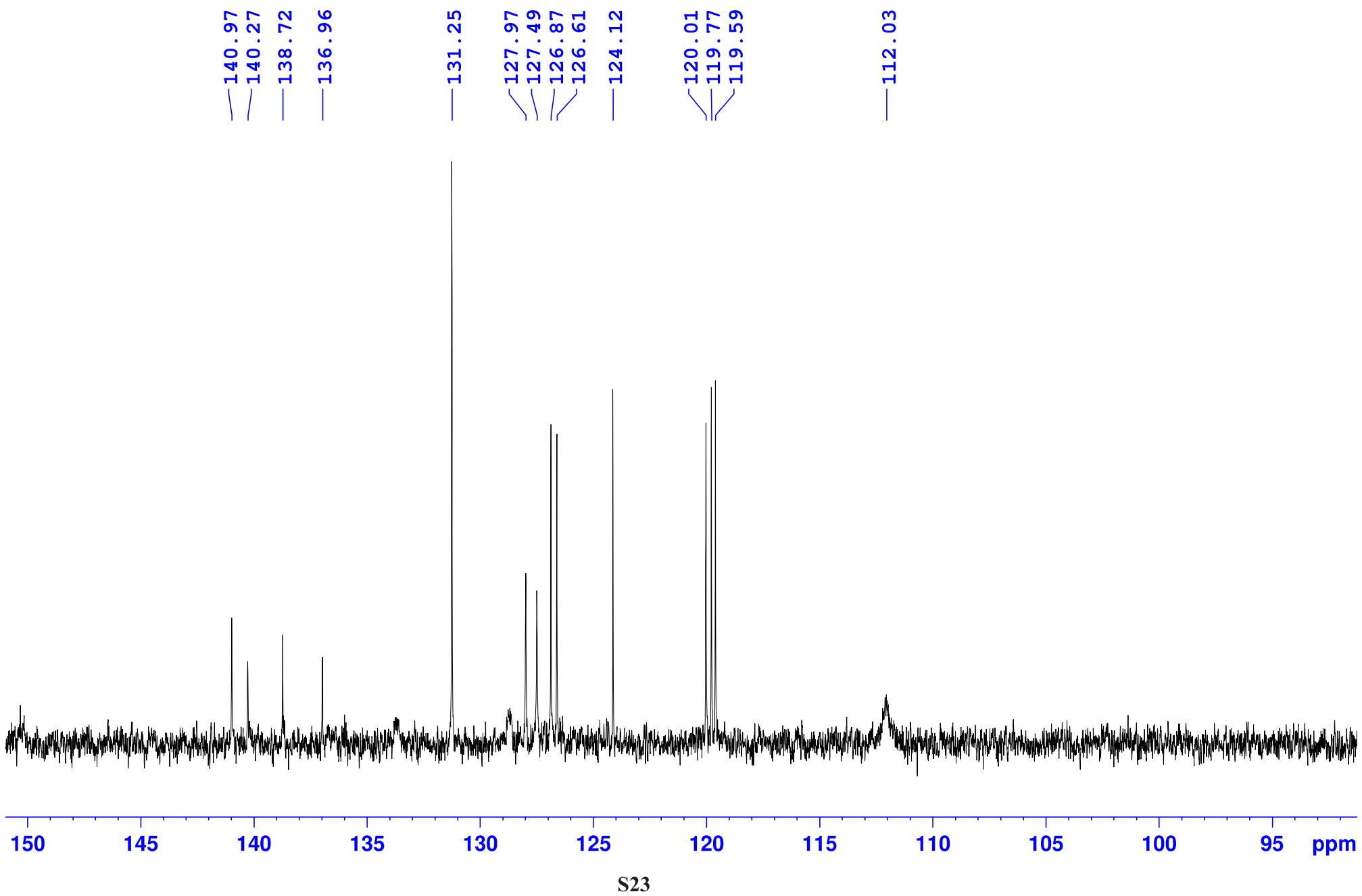
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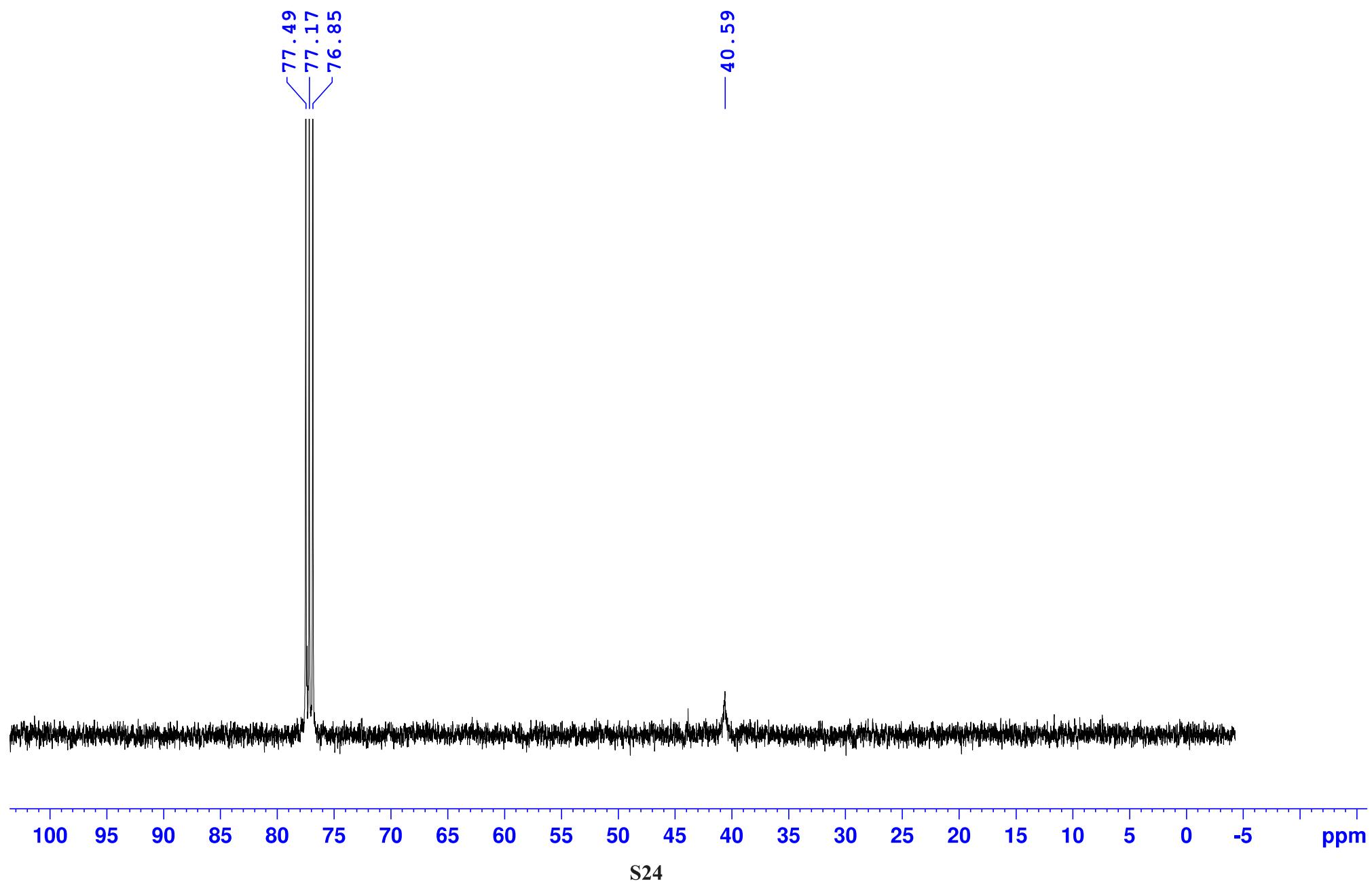
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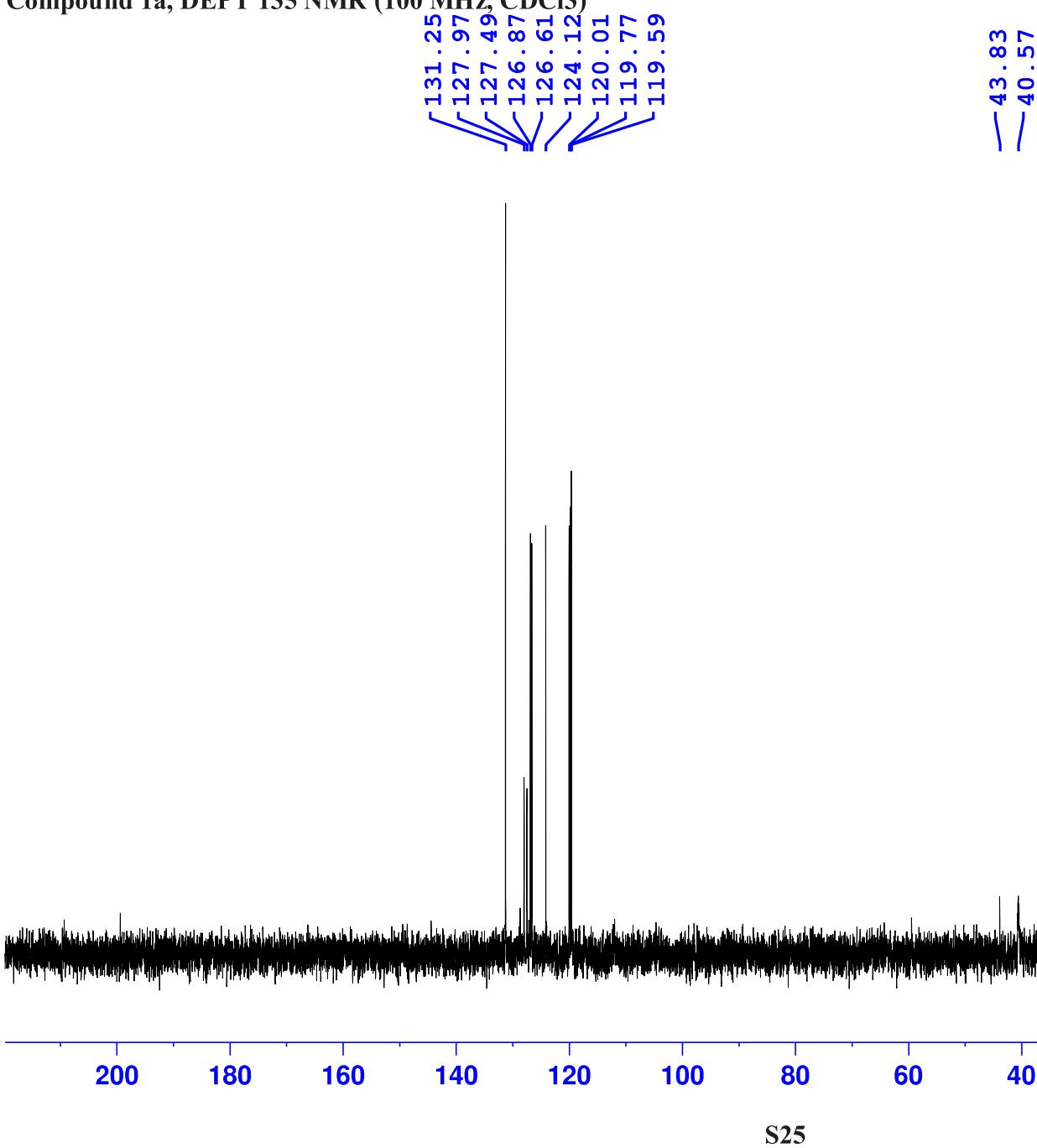
Compound 1a,  $^{13}\text{C}$  NMR (100 MHz,  $\text{CDCl}_3$ )



**Compound 1a,  $^{13}\text{C}$  NMR (100 MHz,  $\text{CDCl}_3$ )**



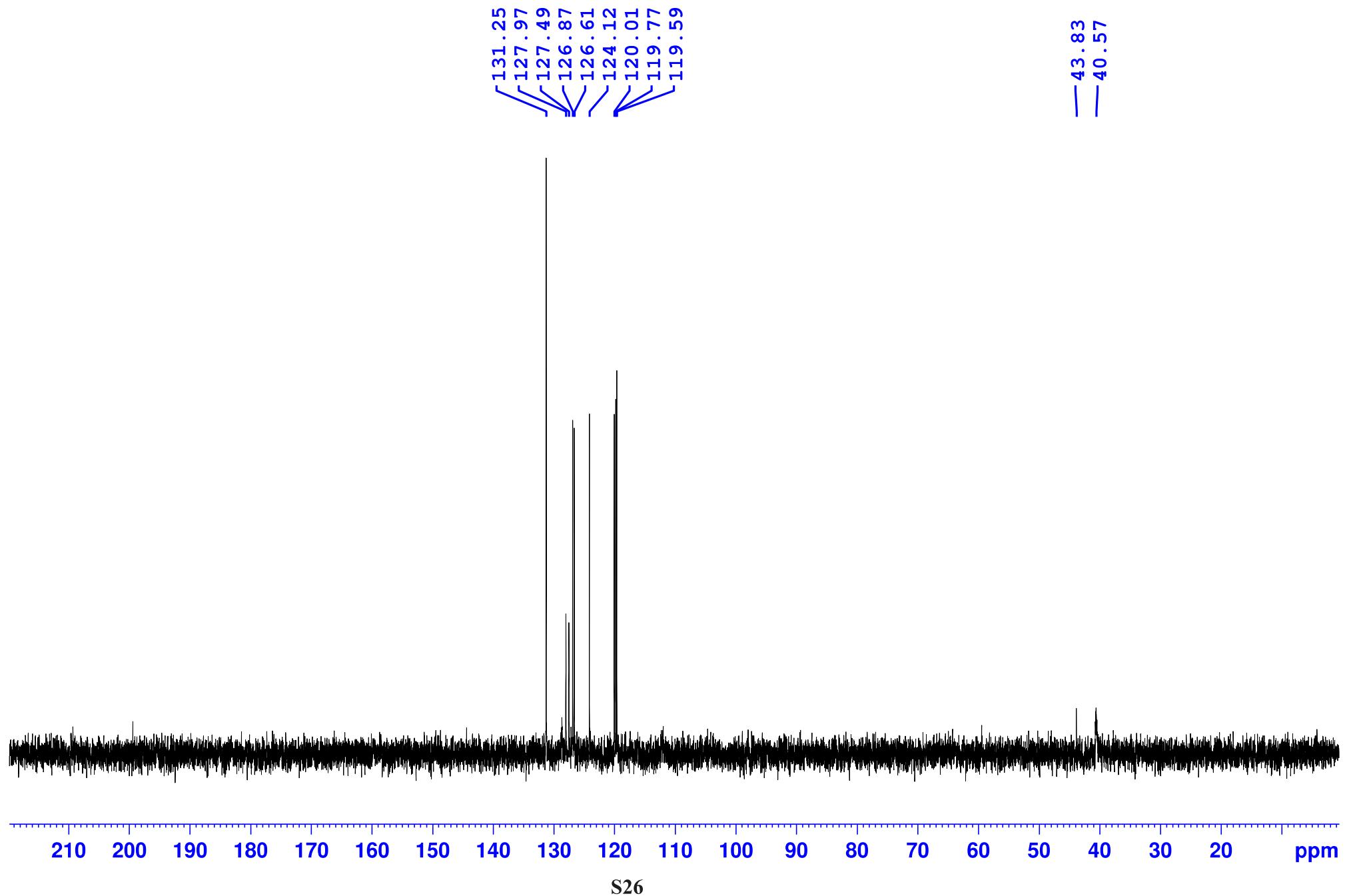
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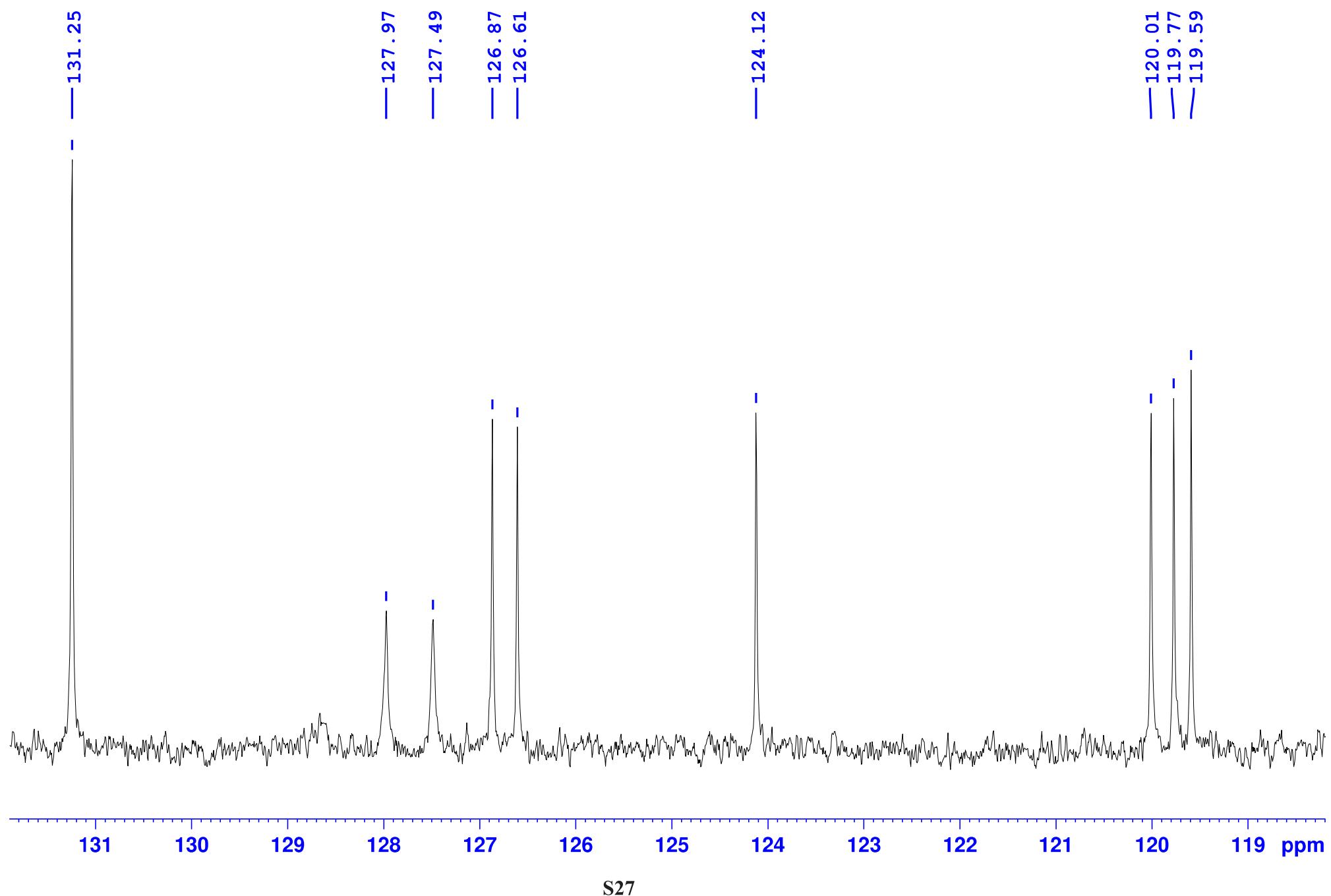
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 SPOAL5 0.500  
 SPOFFS5 0 Hz  
 SPW5 11.08699989 W  
 SFO2 400.1316005 MHz  
 NUC2 1H  
 CPDPRG[2] waltz16  
 P3 10.00 usec  
 P4 20.00 usec  
 PCPD2 90.00 usec  
 PLW2 16.68099976 W  
 PLW12 0.20593999 W

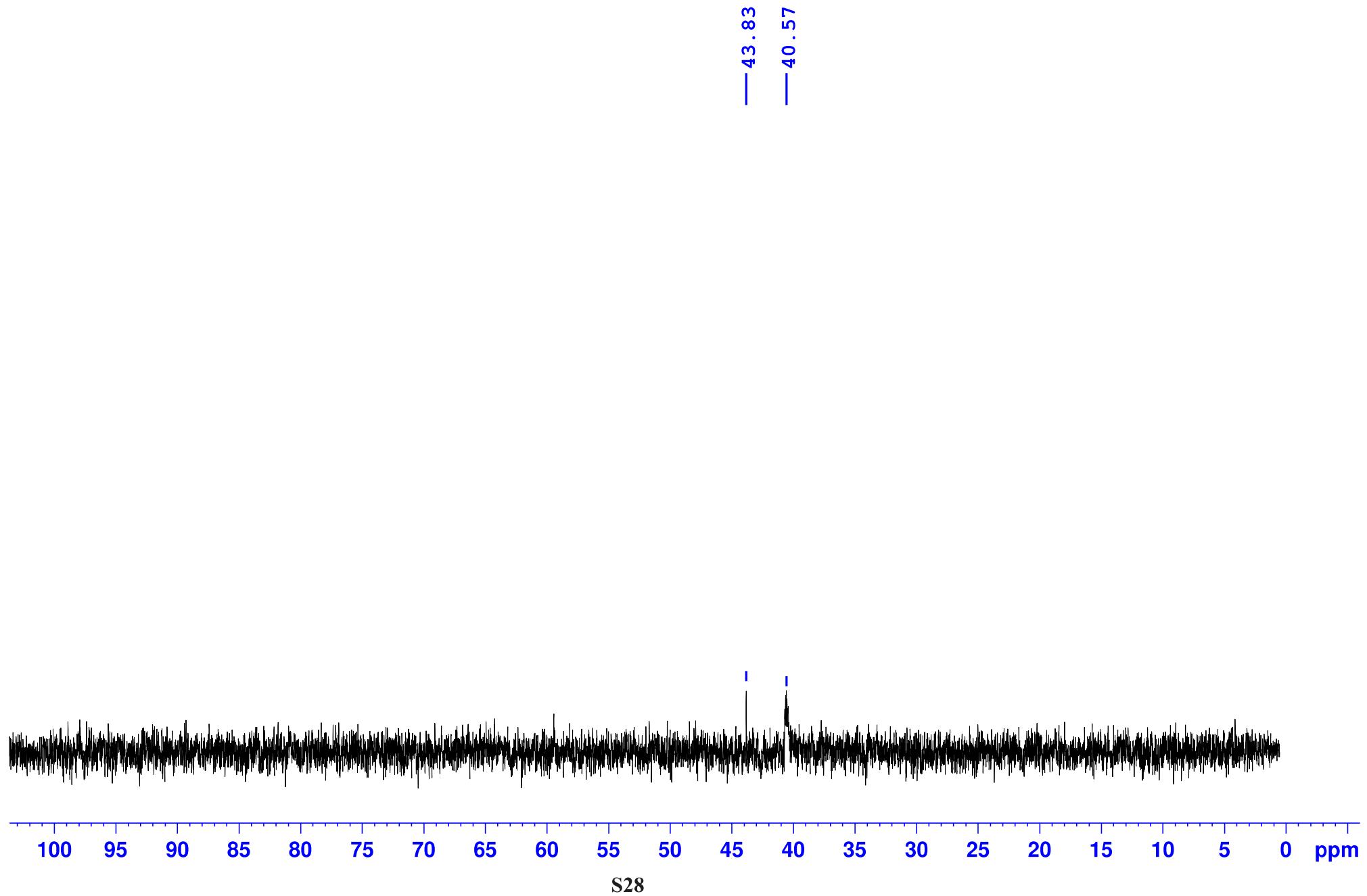
Compound 1a, DEPT 135 NMR (100 MHz, CDCl<sub>3</sub>)



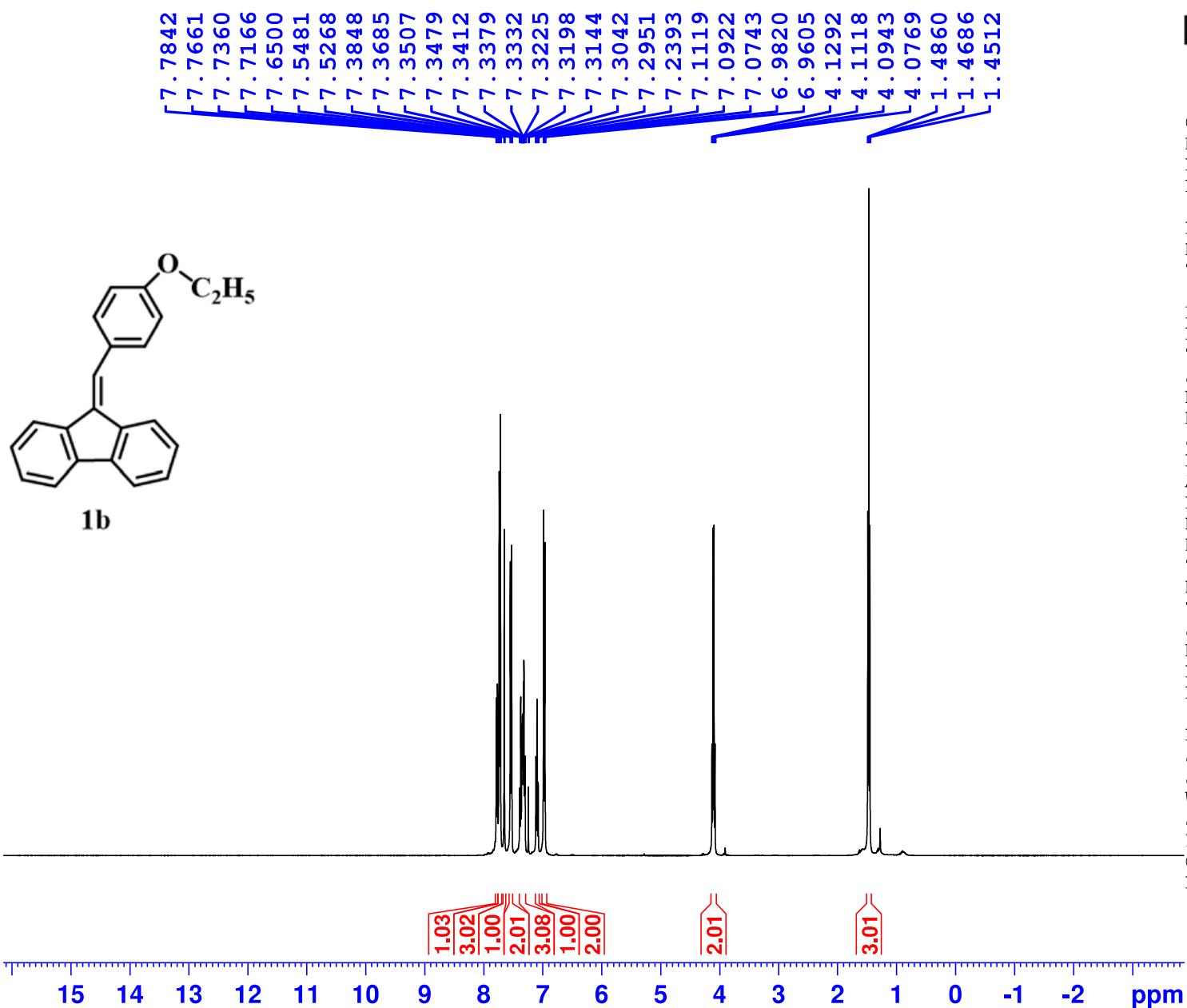
**Compound 1a, DEPT 135 NMR (100 MHz, CDCl<sub>3</sub>)**



Compound 1a, DEPT 135 NMR (100 MHz, CDCl<sub>3</sub>)



Compound 1b,  $^1\text{H}$  NMR (400 MHz,  $\text{CDCl}_3$ )

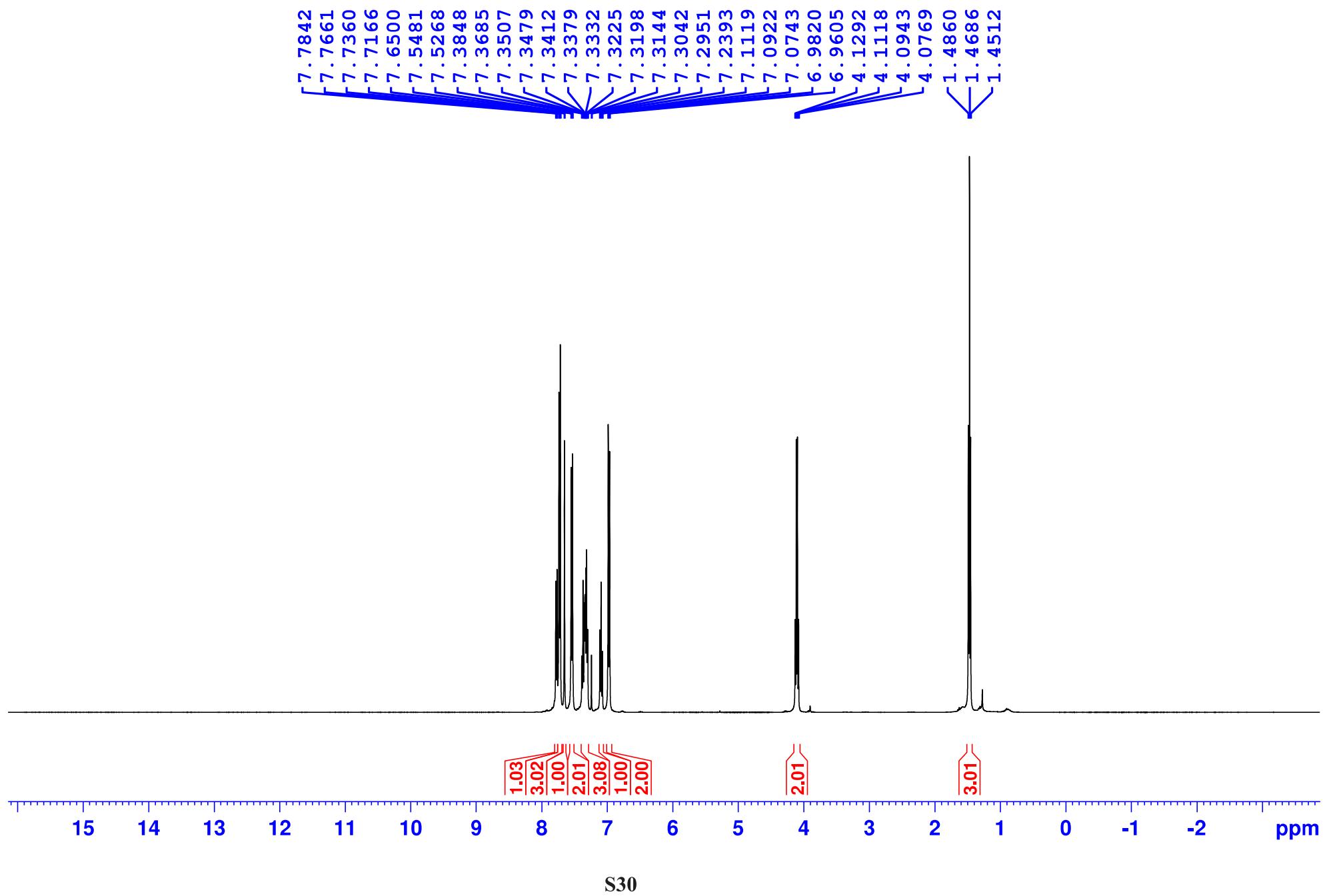


Current Data Parameters  
 NAME SMT-7  
 EXPNO 156  
 PROCNO 1

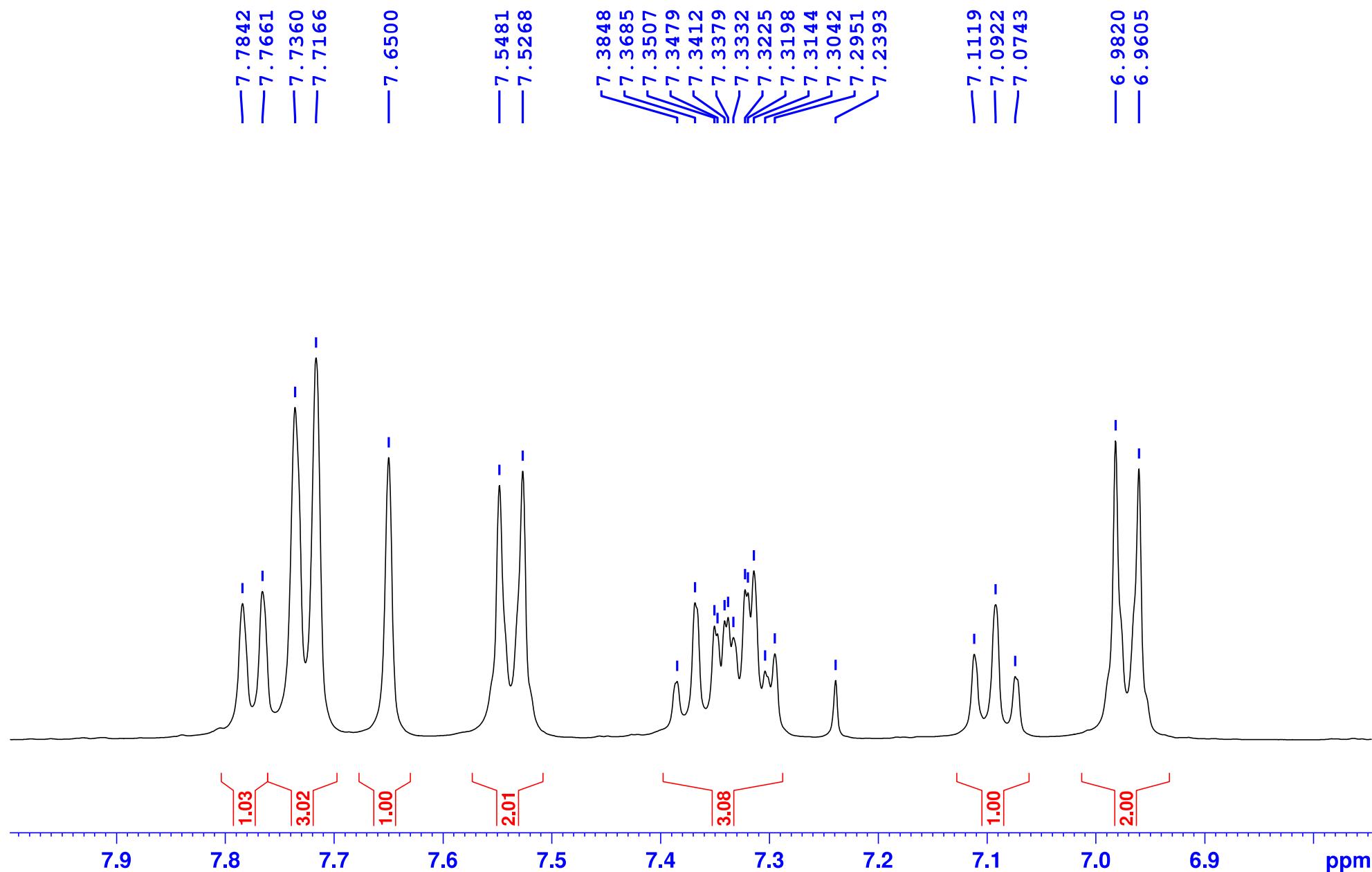
F2 - Acquisition Parameters  
 Date\_ 20210305  
 Time 15.45 h  
 INSTRUM spect  
 PROBHD Z116098\_0621 (zg30  
 PULPROG zg30  
 TD 65536  
 SOLVENT  $\text{CDCl}_3$   
 NS 16  
 DS 2  
 SWH 8012.820 Hz  
 FIDRES 0.244532 Hz  
 AQ 4.0894465 sec  
 RG 31.9  
 DW 62.400 usec  
 DE 6.50 usec  
 TE 298.0 K  
 D1 1.0000000 sec  
 TD0 1  
 SFO1 400.1324708 MHz  
 NUC1 1H  
 P1 10.00 usec  
 PLW1 16.68099976 W

F2 - Processing parameters  
 SI 65536  
 SF 400.1300172 MHz  
 WDW EM  
 SSB 0  
 LB 0.30 Hz  
 GB 0  
 PC 1.00

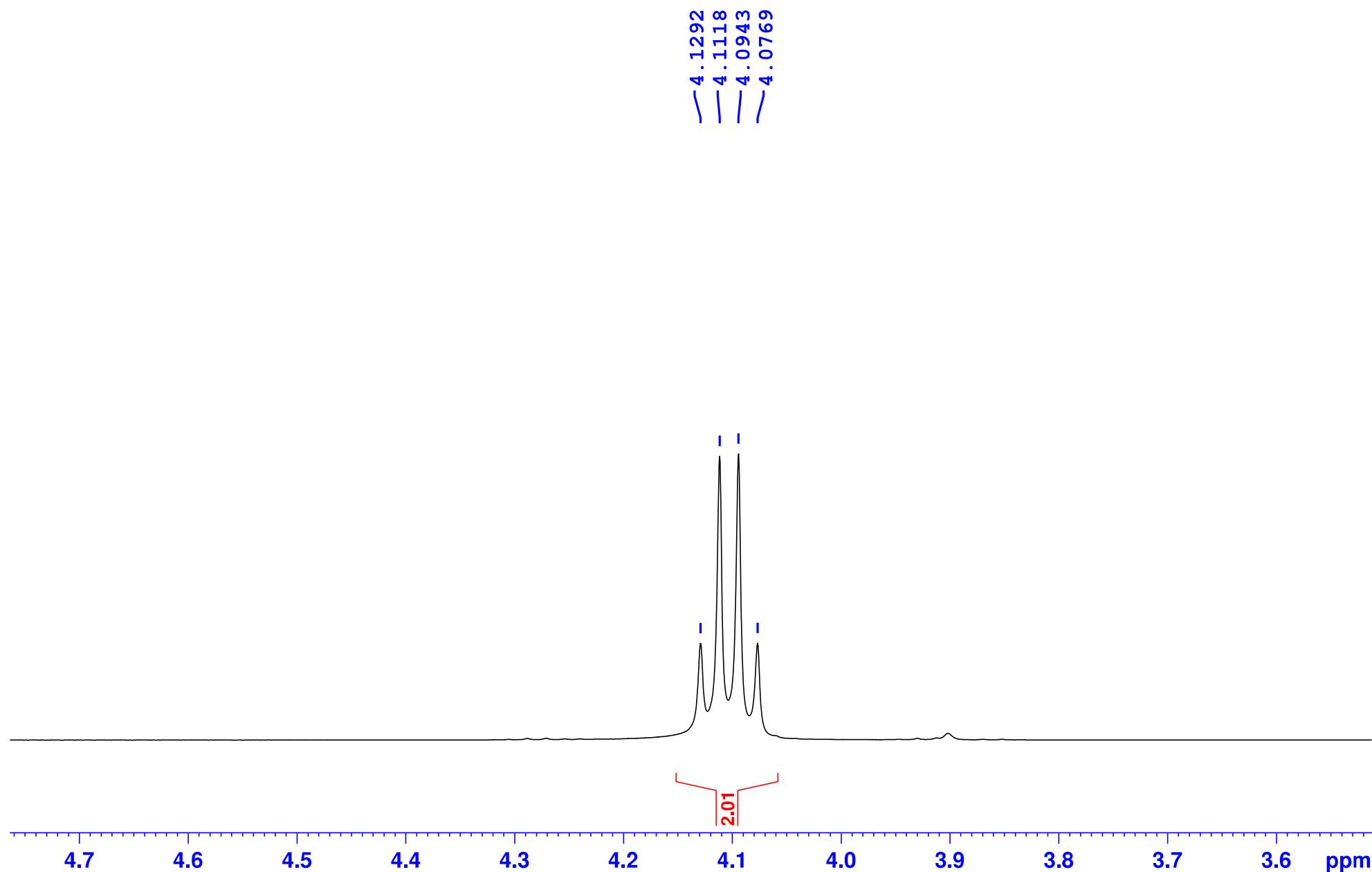
Compound 1b,  $^1\text{H}$  NMR (400 MHz,  $\text{CDCl}_3$ )



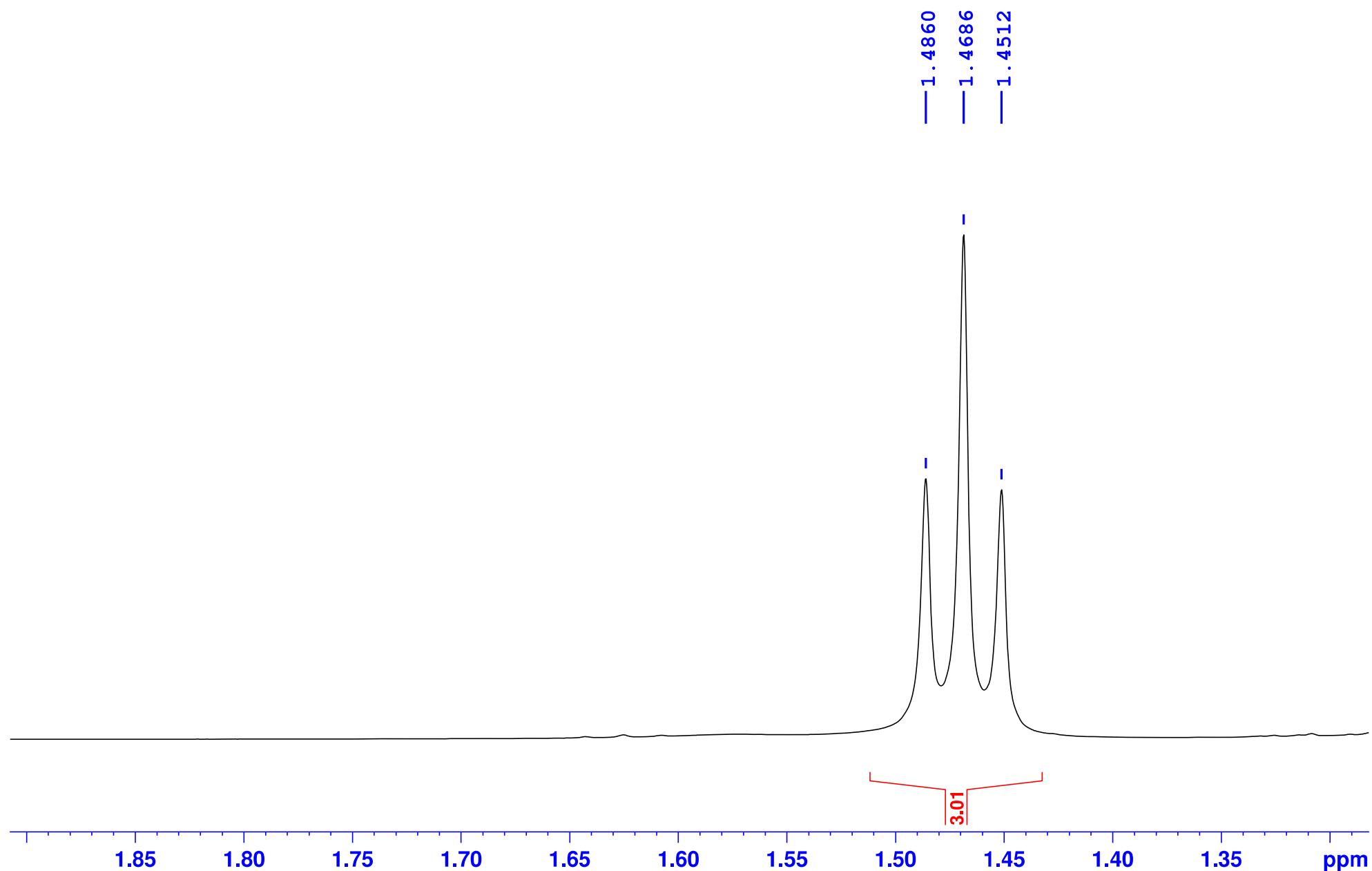
Compound 1b,  $^1\text{H}$  NMR (400 MHz,  $\text{CDCl}_3$ )



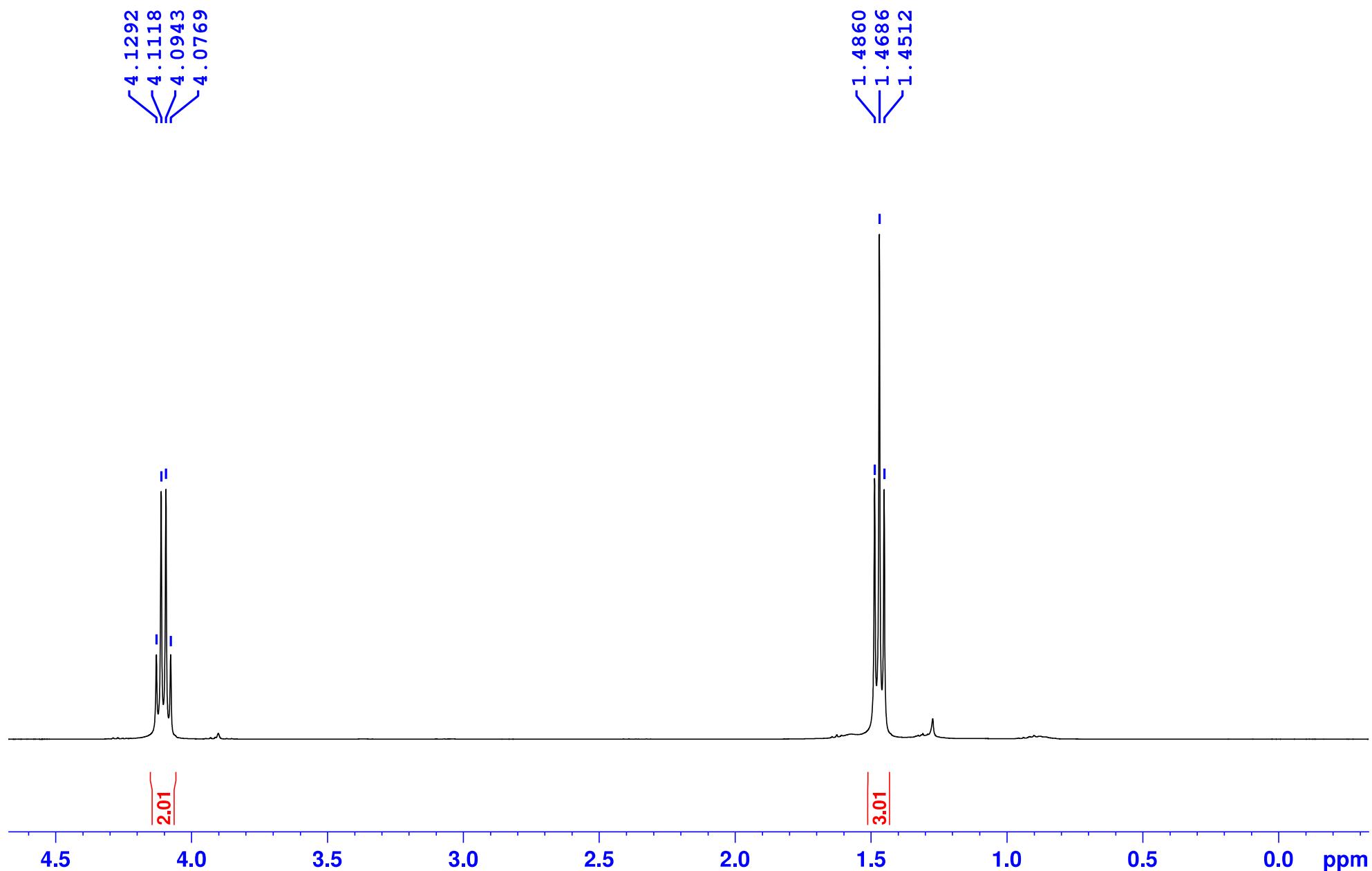
Compound 1b,  $^1\text{H}$  NMR (400 MHz,  $\text{CDCl}_3$ )



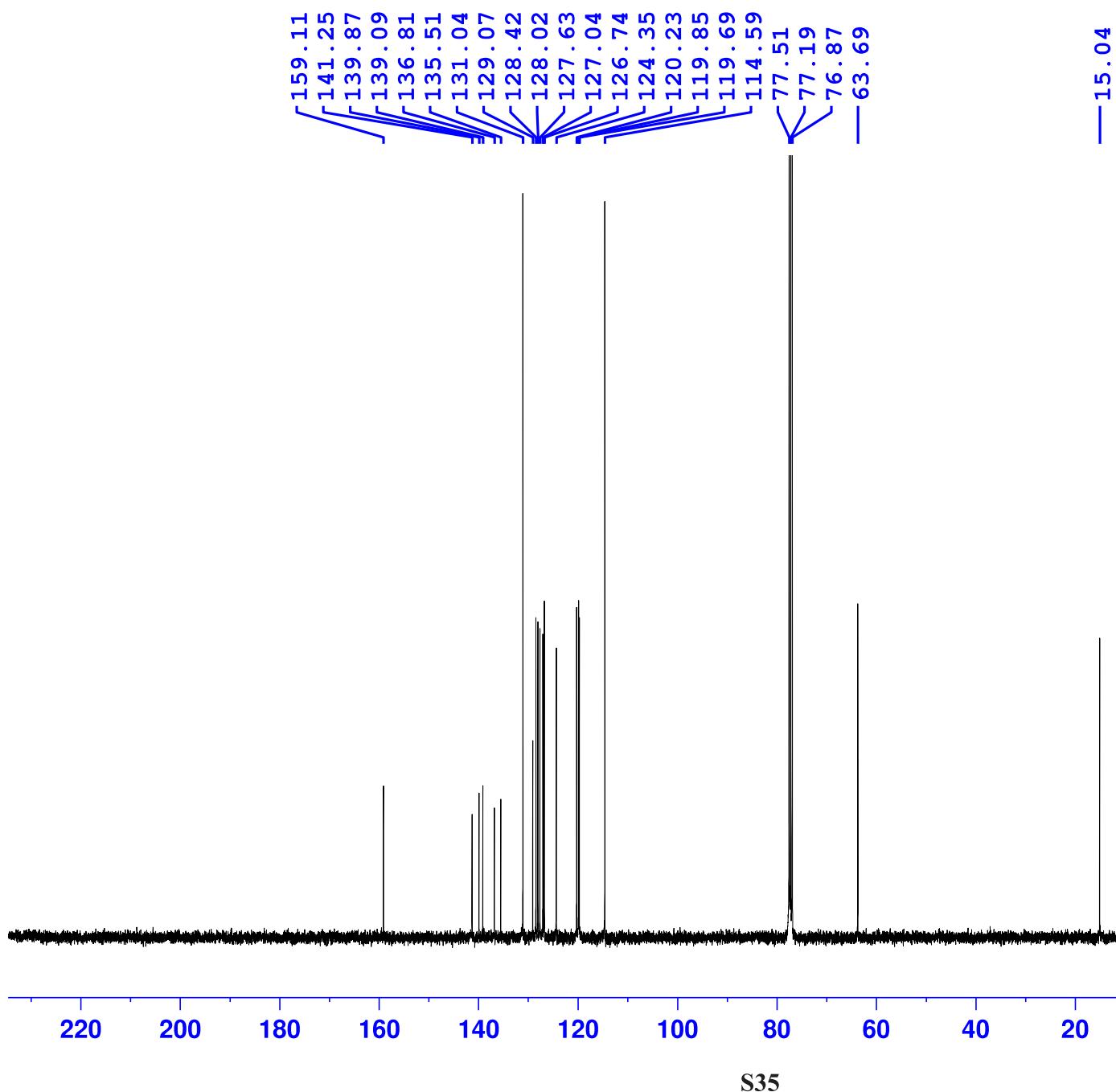
Compound 1b,  $^1\text{H}$  NMR (400 MHz,  $\text{CDCl}_3$ )



Compound 1b,  $^1\text{H}$  NMR (400 MHz,  $\text{CDCl}_3$ )



Compound 1b,  $^{13}\text{C}$  NMR (100 MHz,  $\text{CDCl}_3$ )

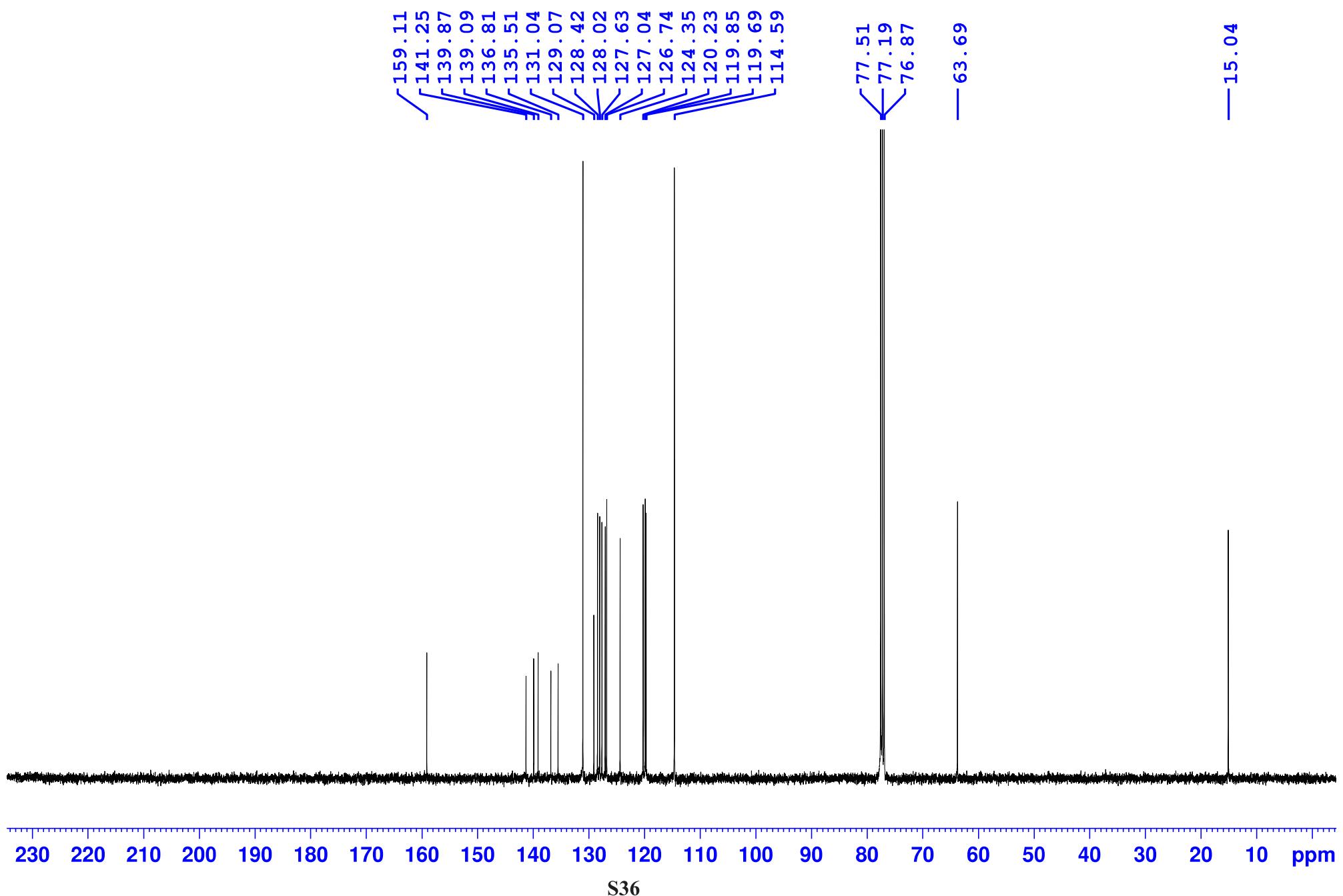


Current Data Parameters  
NAME Shumaila Majeed  
EXPNO 272  
PROCNO 1

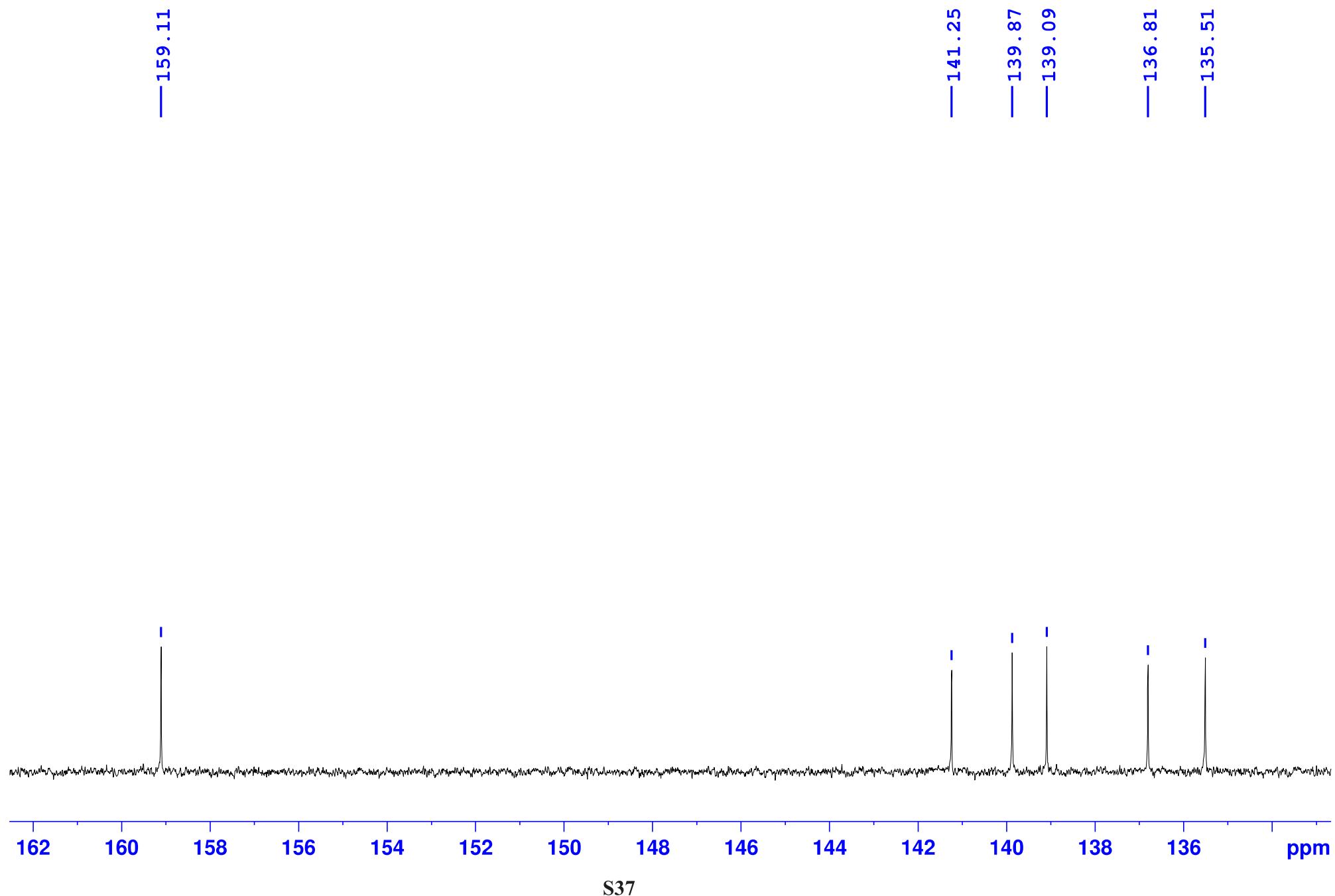
F2 - Acquisition Parameters  
Date\_ 20211013  
Time 14.39 h  
INSTRUM spect  
PROBHD Z116098\_0621 (zgpg30  
PULPROG 65536  
TD 400  
DS 2  
SWH 24038.461 Hz  
FIDRES 0.733596 Hz  
AQ 1.3631488 sec  
RG 199.48  
DW 20.800 usec  
DE 6.50 usec  
TE 298.0 K  
D1 2.0000000 sec  
D11 0.03000000 sec  
TD0 1  
SFO1 100.6243390 MHz  
NUC1  $^{13}\text{C}$   
P1 10.00 usec  
PLW1 72.56700134 W  
SFO2 400.1316005 MHz  
NUC2 1H  
CPDPRG[2] waltz16  
PCPD2 90.00 usec  
PLW2 16.68099976 W  
PLW12 0.20593999 W  
PLW13 0.10342000 W

F2 - Processing parameters  
SI 32768  
SF 100.6127547 MHz  
WDW EM  
SSB 0  
LB 1.00 Hz  
PC 0  
GB 1.40

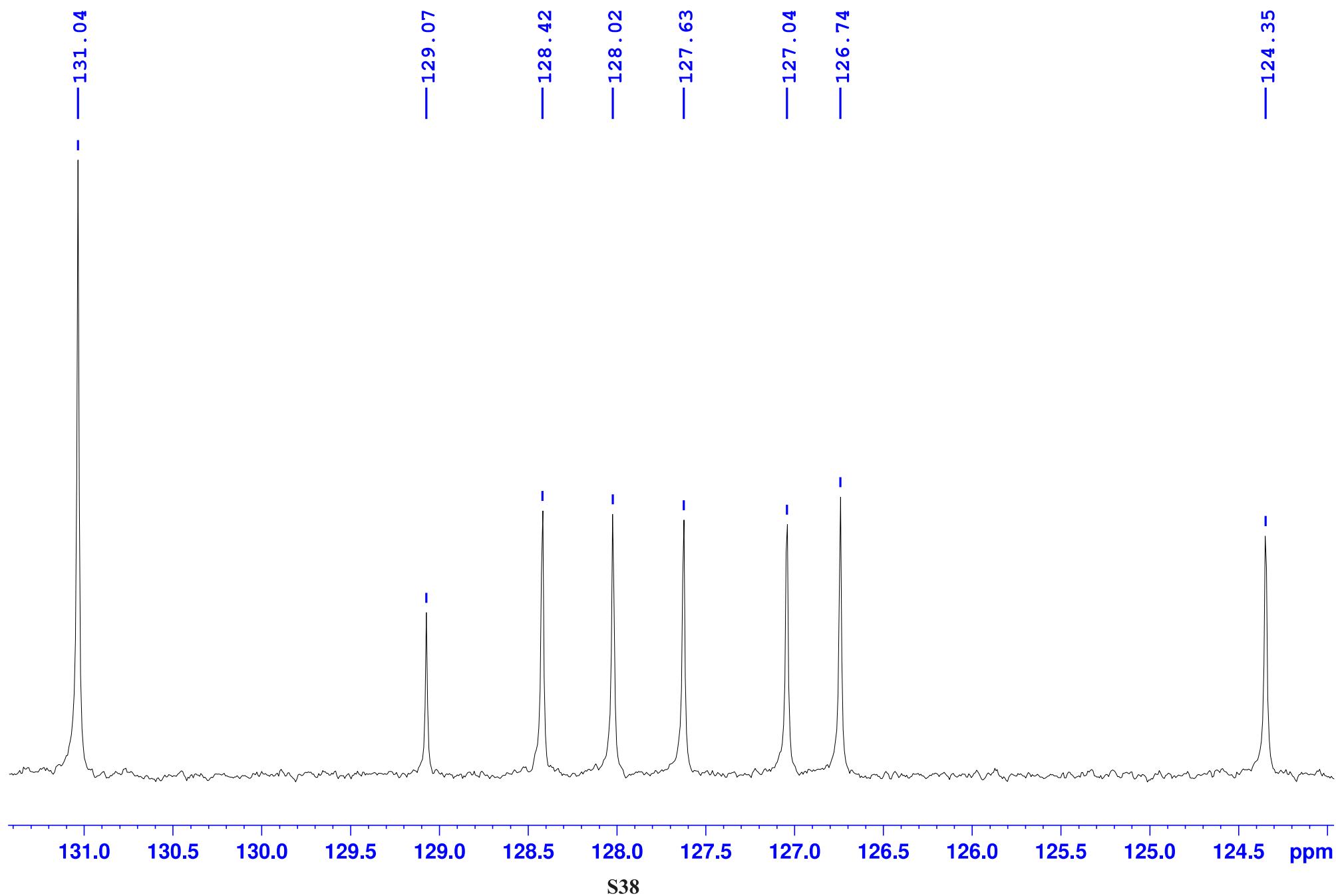
Compound 1b,  $^{13}\text{C}$  NMR (100 MHz,  $\text{CDCl}_3$ )



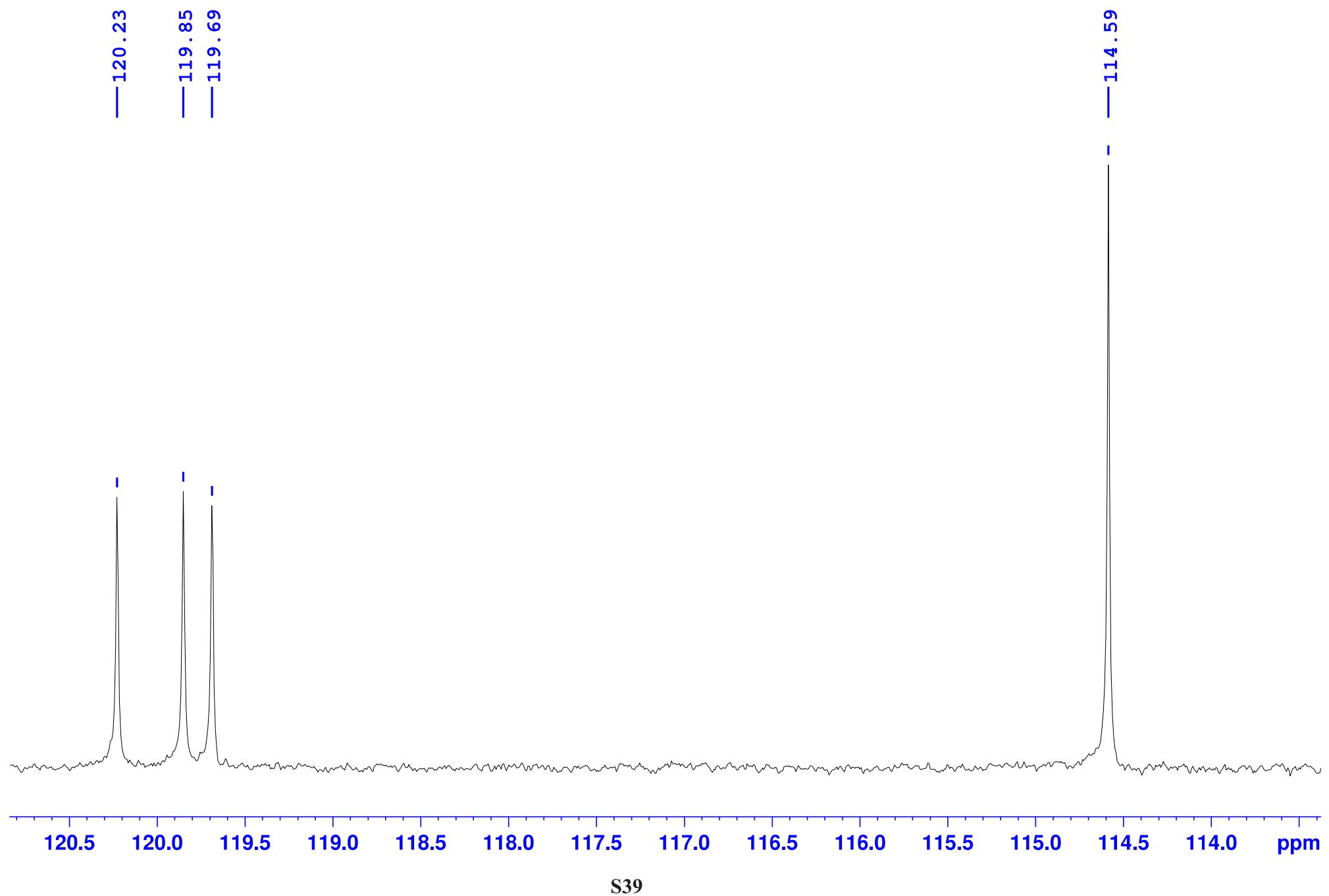
Compound 1b,  $^{13}\text{C}$  NMR (100 MHz,  $\text{CDCl}_3$ )



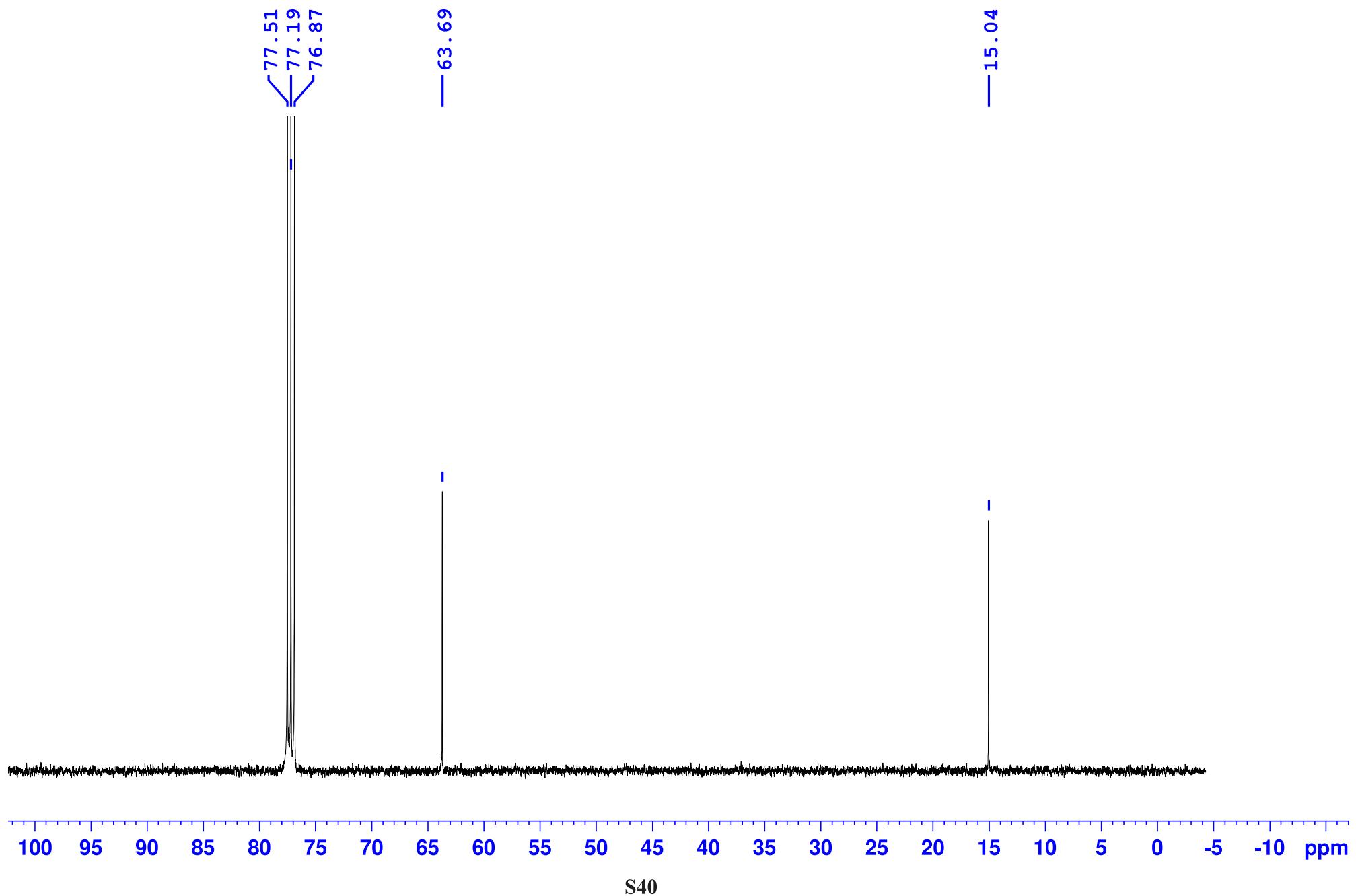
**Compound 1b,  $^{13}\text{C}$  NMR (100 MHz,  $\text{CDCl}_3$ )**



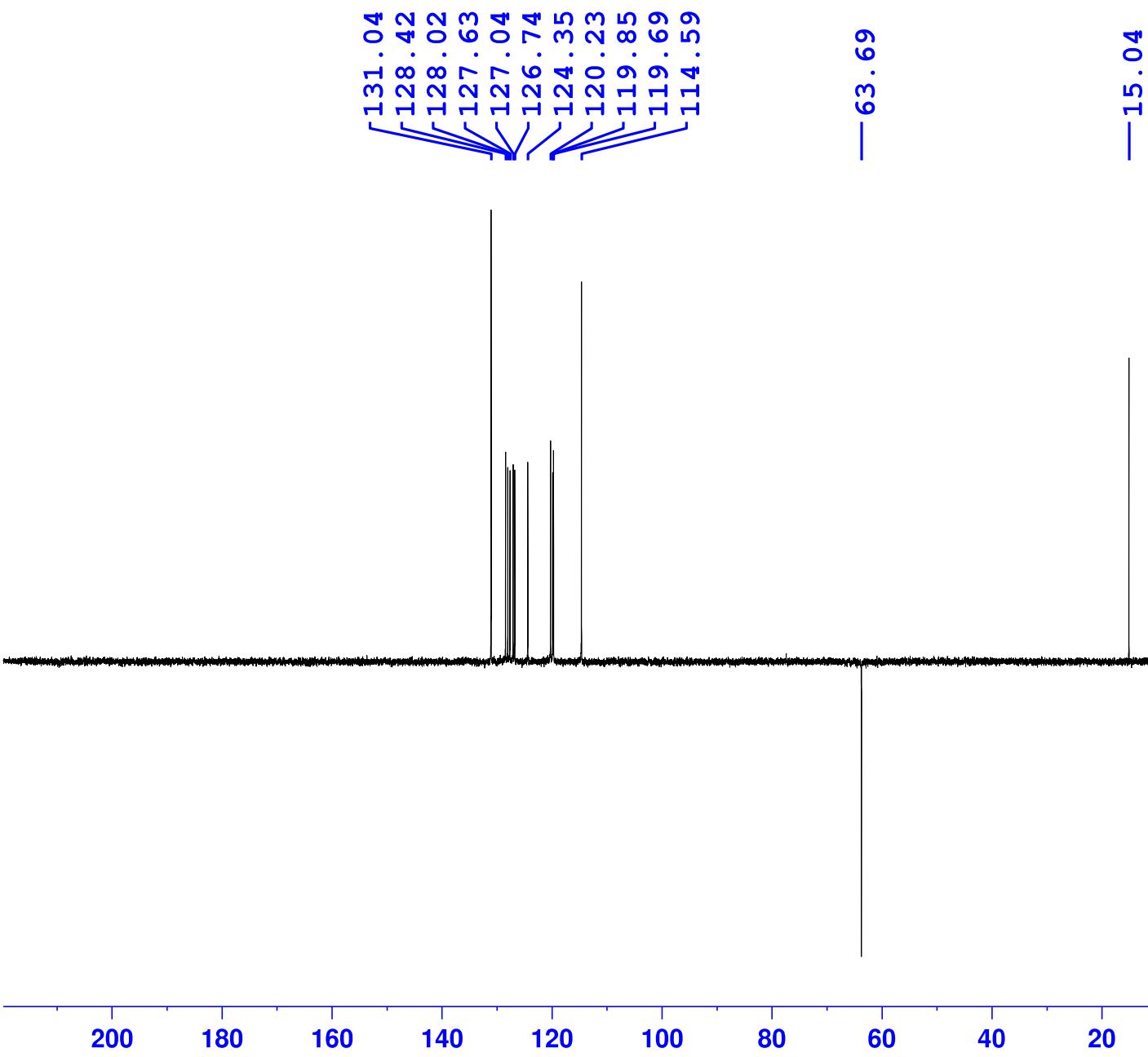
Compound 1b,  $^{13}\text{C}$  NMR (100 MHz,  $\text{CDCl}_3$ )



Compound 1b,  $^{13}\text{C}$  NMR (100 MHz,  $\text{CDCl}_3$ )



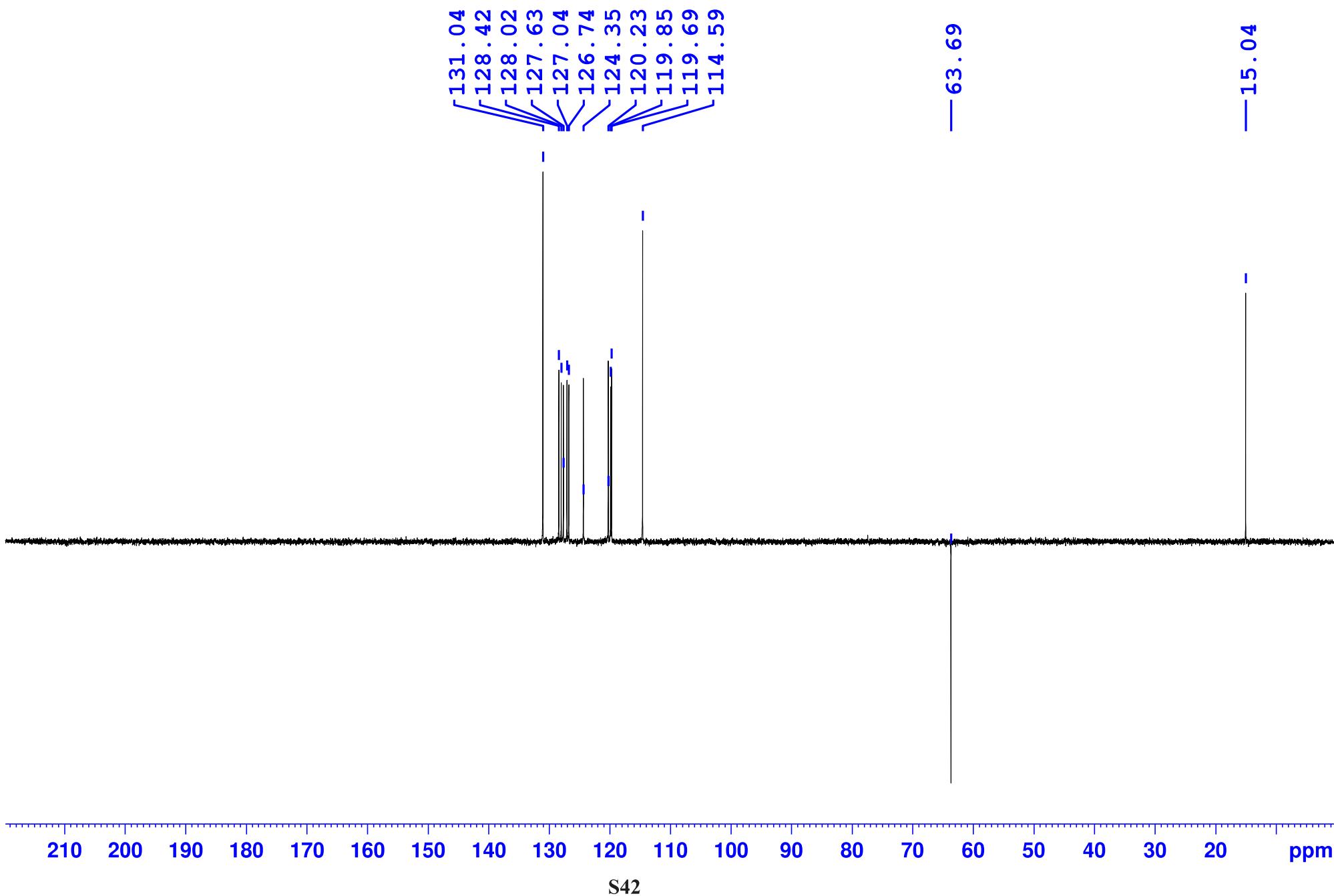
Compound 1b, DEPT 135 NMR (100 MHz, CDCl<sub>3</sub>)



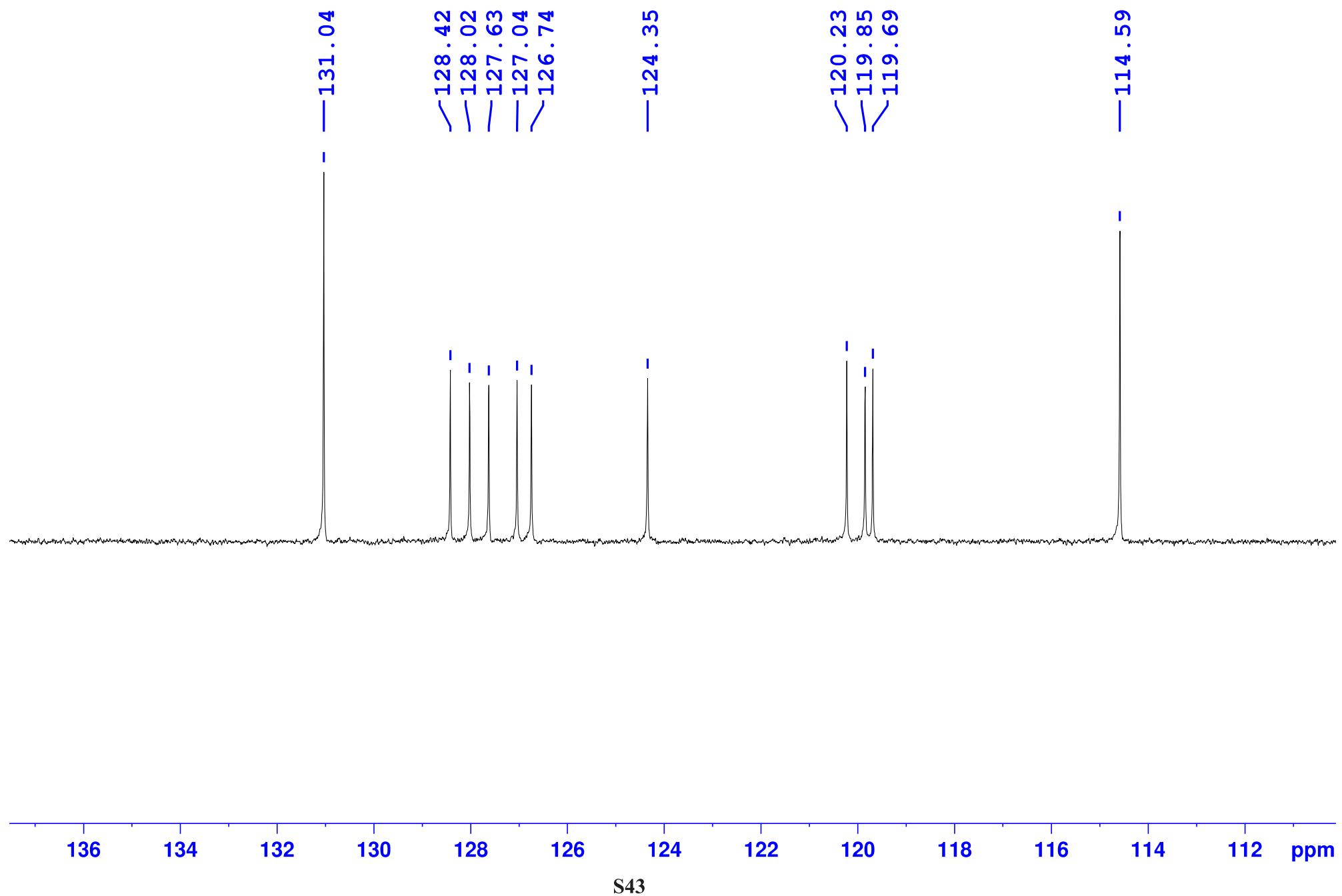
Current Data Parameters  
NAME Shumaila Majeed  
EXPNO 273  
PROCNO 1

F2 - Acquisition Parameters  
Date\_ 20211013  
Time 14.55 h  
INSTRUM spect  
PROBHD Z116098\_0621 (deptsp135  
PULPROG 65536  
TD 256  
SOLVENT CDC13  
NS 4  
DS 22058.824 Hz  
FIDRES 0.673182 Hz  
AQ 1.4854827 sec  
RG 199.48  
DW 22.667 usec  
DE 6.50 usec  
TE 298.0 K  
CNST2 145.0000000  
D1 2.00000000 sec  
D2 0.00344828 sec  
D12 0.00002000 sec  
TD0 1  
SF01 100.6238359 MHz  
NUC1 13C  
P1 10.00 usec  
P13 2000.00 usec  
PLW0 0 W  
PLW1 72.56700134 W  
SPNAM[5] Crp60comp.4  
SPOAL5 0.500  
SPOFFS5 0 Hz  
SPW5 11.08699989 W  
SF02 400.1316005 MHz  
NUC2 1H  
CPDPRG[2] waltz16  
P3 10.00 usec  
P4 20.00 usec  
PCPD2 90.00 usec  
PLW2 16.68099976 W  
PLW12 0.20593999 W

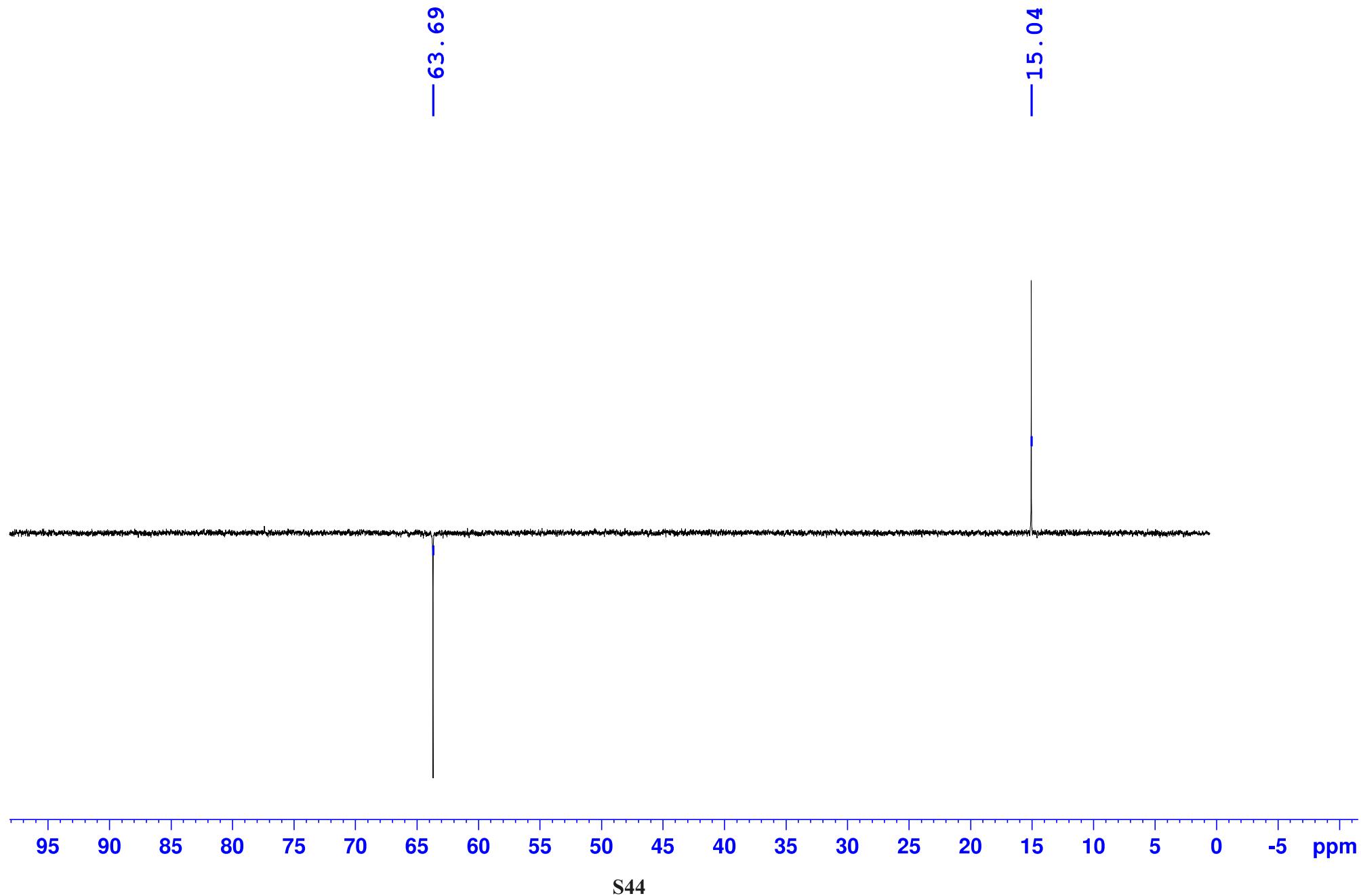
Compound 1b, DEPT 135 NMR (100 MHz, CDCl<sub>3</sub>)



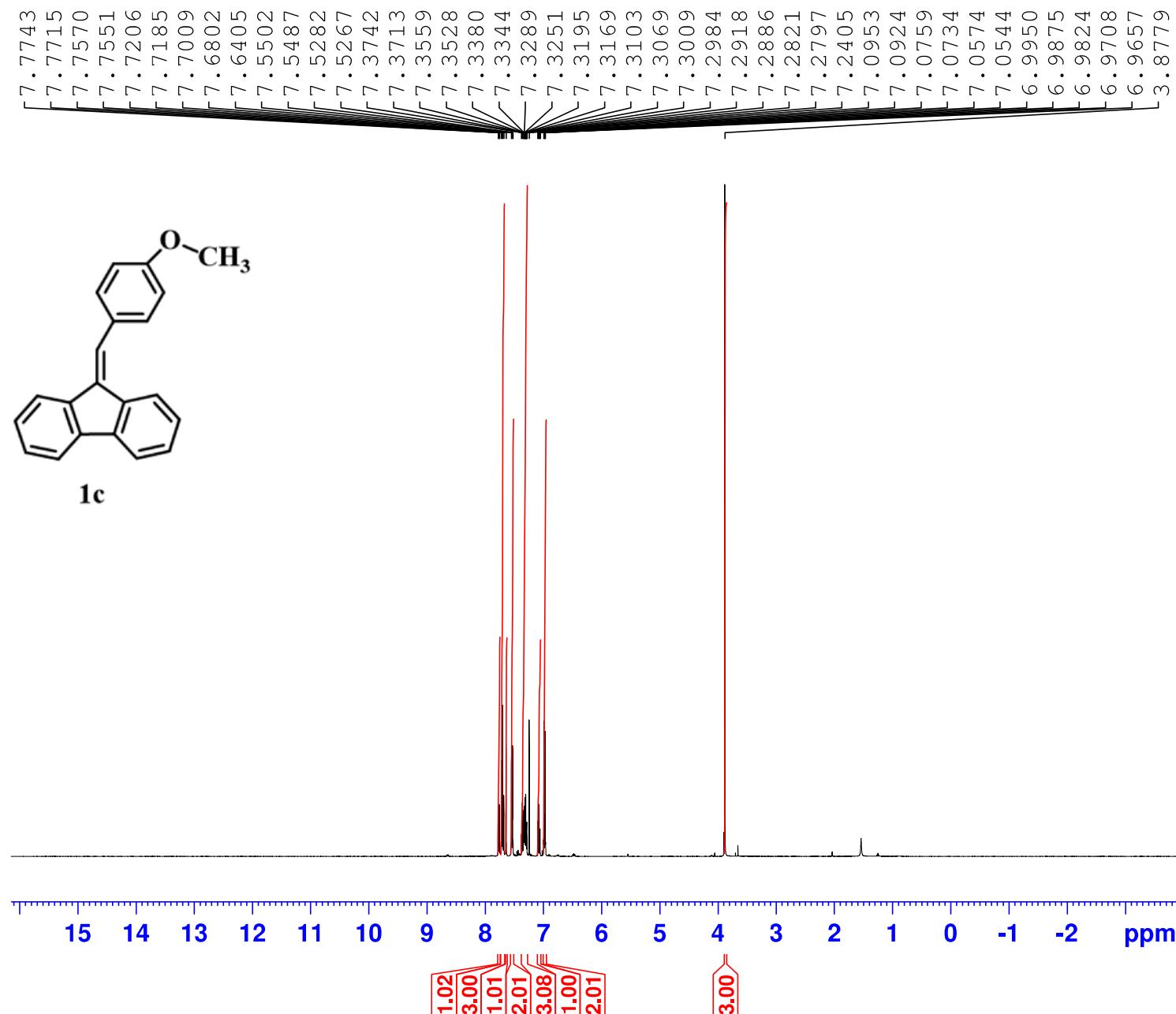
Compound 1b, DEPT 135 NMR (100 MHz, CDCl<sub>3</sub>)



**Compound 1b, DEPT 135 NMR (100 MHz, CDCl<sub>3</sub>)**



**Compound 1c, 1H NMR (400 MHz, CDCl<sub>3</sub>)**

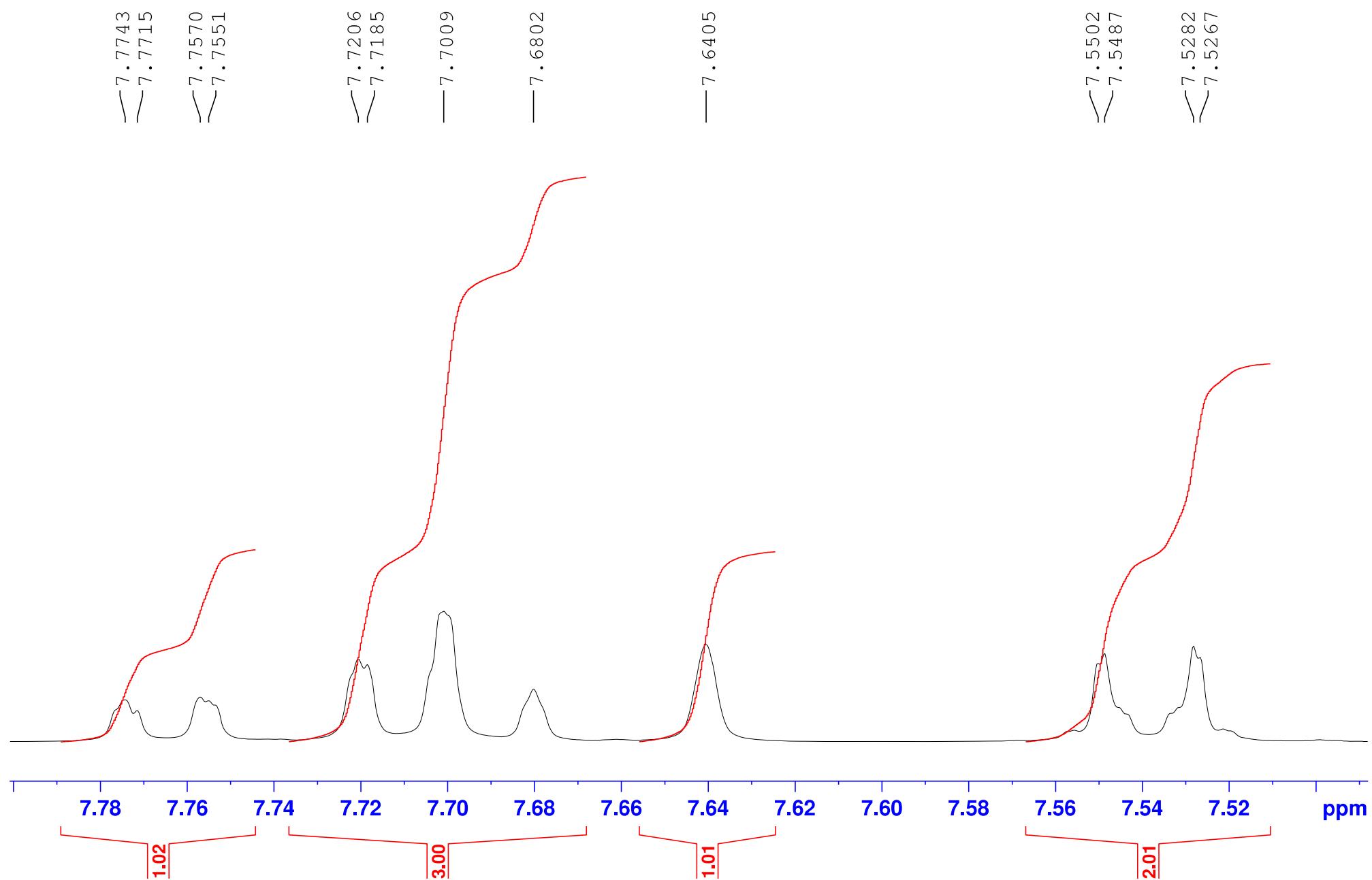


Current Data Parameters  
 NAME FID NMR  
 EXPNO 73  
 PROCNO 1

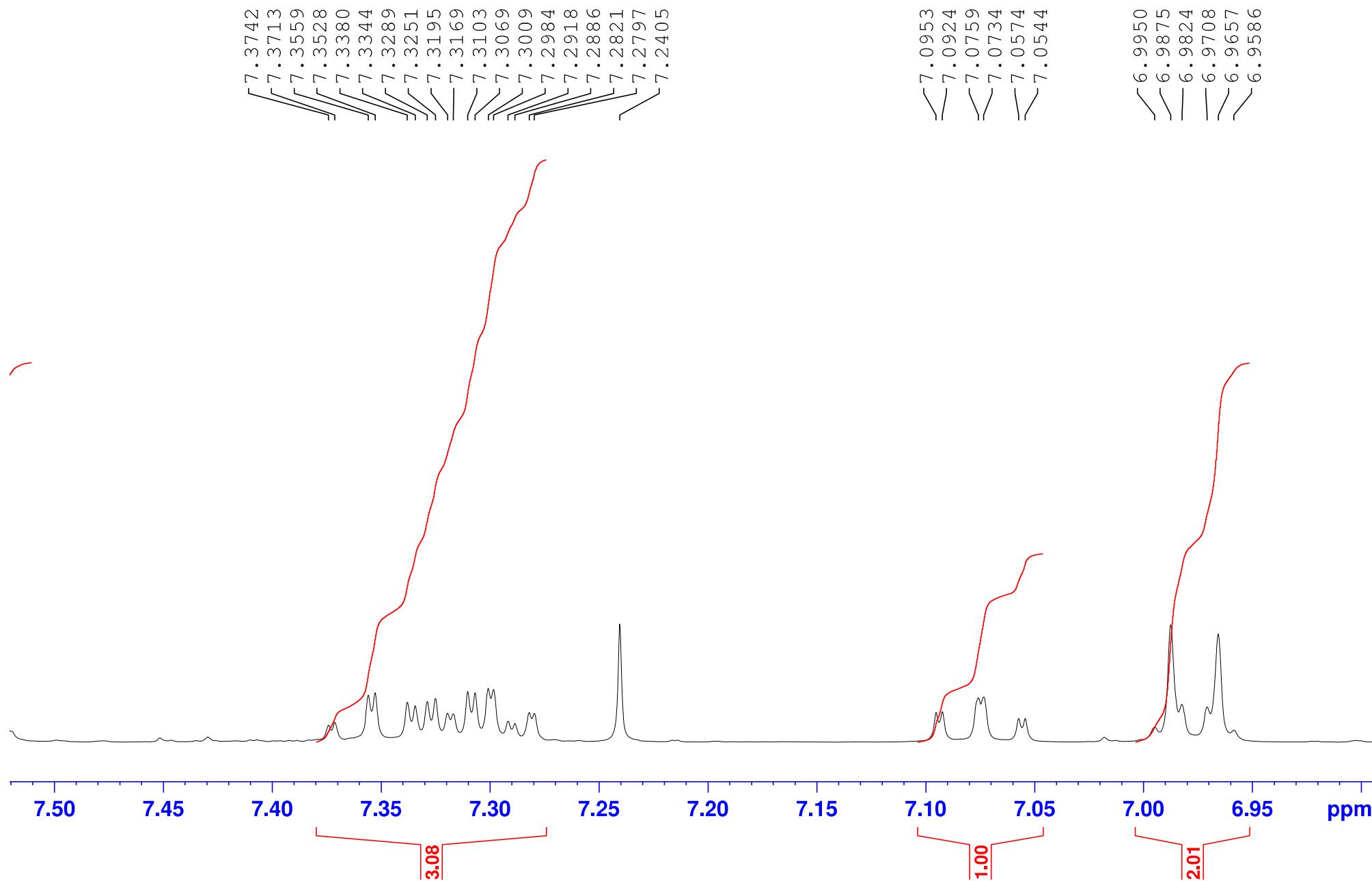
F2 - Acquisition Parameters  
 Date 20201103  
 Time 21.15 h  
 INSTRUM spect  
 PROBHD Z116098\_0621 (zg30  
 PULPROG zg30  
 TD 65536  
 SOLVENT CDCl<sub>3</sub>  
 NS 16  
 DS 2  
 SWH 8012.820 Hz  
 FIDRES 0.244532 Hz  
 AQ 4.0894465 sec  
 RG 98.53  
 DW 62.400 usec  
 DE 6.50 usec  
 TE 298.0 K  
 D1 1.00000000 sec  
 TDO 1  
 SFO1 400.1324708 MHz  
 NUC1 1H  
 P1 10.00 usec  
 PLW1 16.68099976 W

F2 - Processing parameters  
 SI 65536  
 SF 400.1300172 MHz  
 WDW EM  
 SSB 0  
 LB 0.30 Hz  
 GB 0  
 PC 1.00

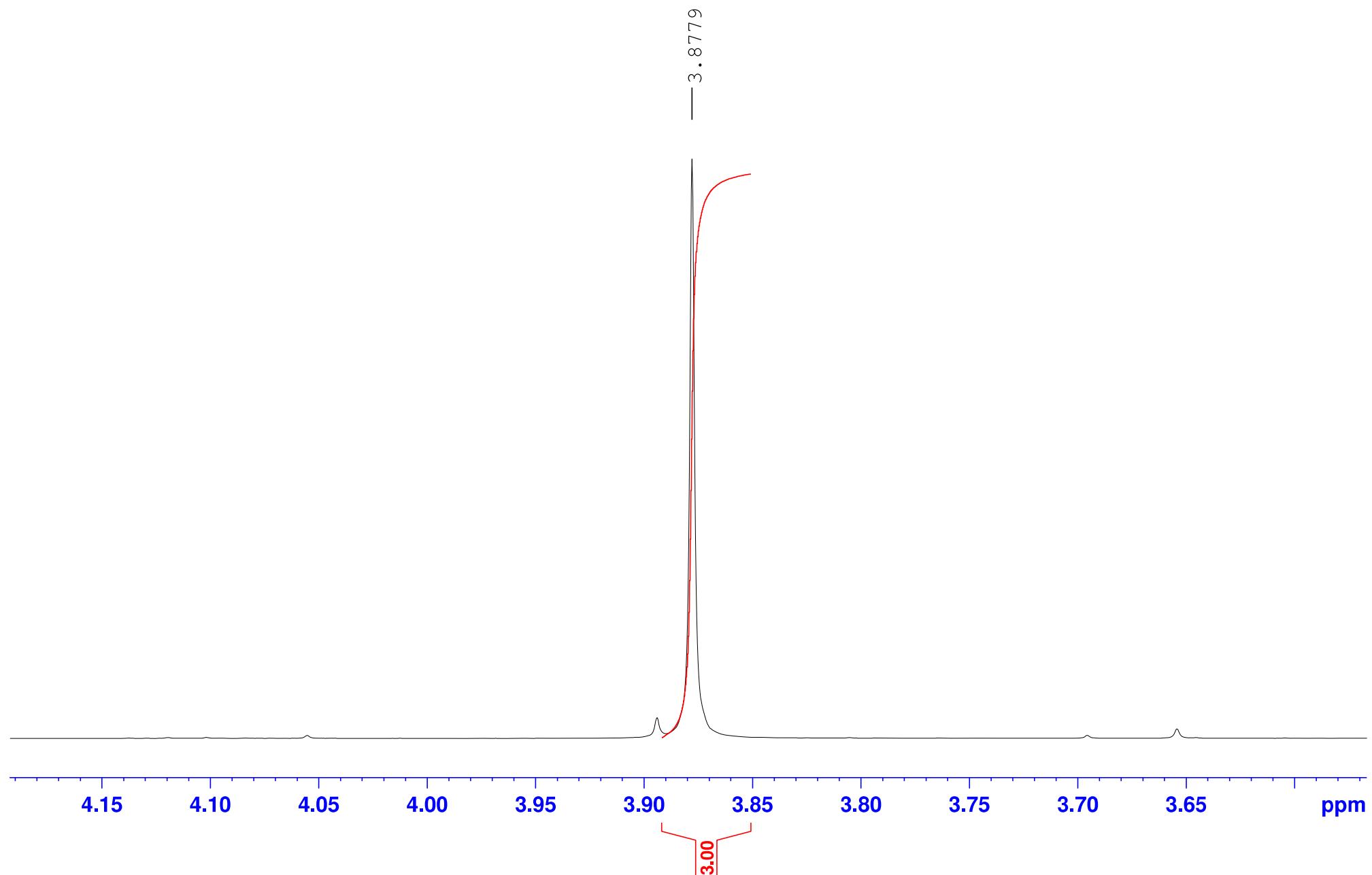
**Compound 1c,  $^1\text{H}$  NMR (400 MHz,  $\text{CDCl}_3$ )**



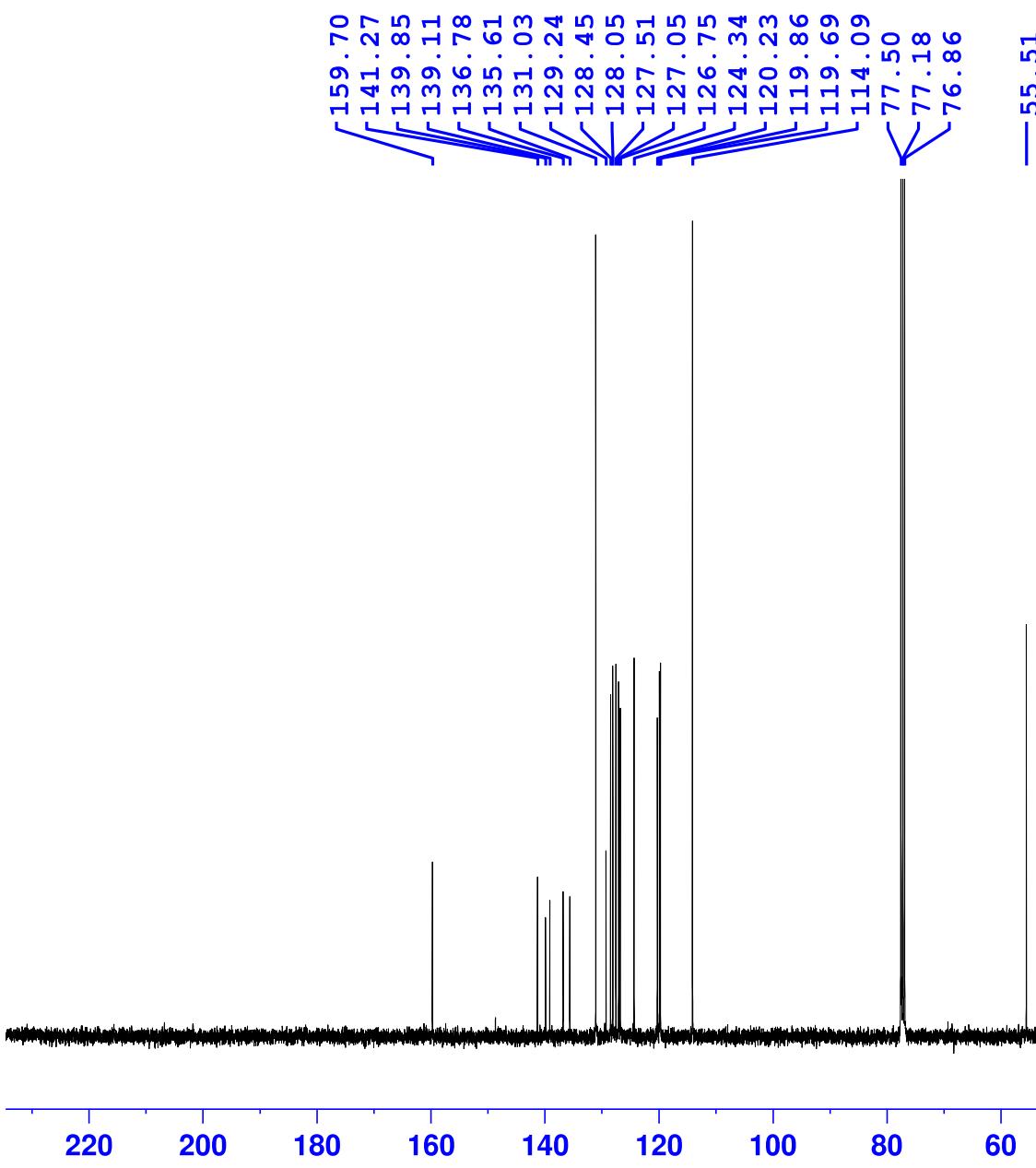
**Compound 1c, 1H NMR (400 MHz, CDCl<sub>3</sub>)**



**Compound 1c,  $^1\text{H}$  NMR (400 MHz,  $\text{CDCl}_3$ )**



Compound 1c,  $^{13}\text{C}$  NMR (100 MHz,  $\text{CDCl}_3$ )

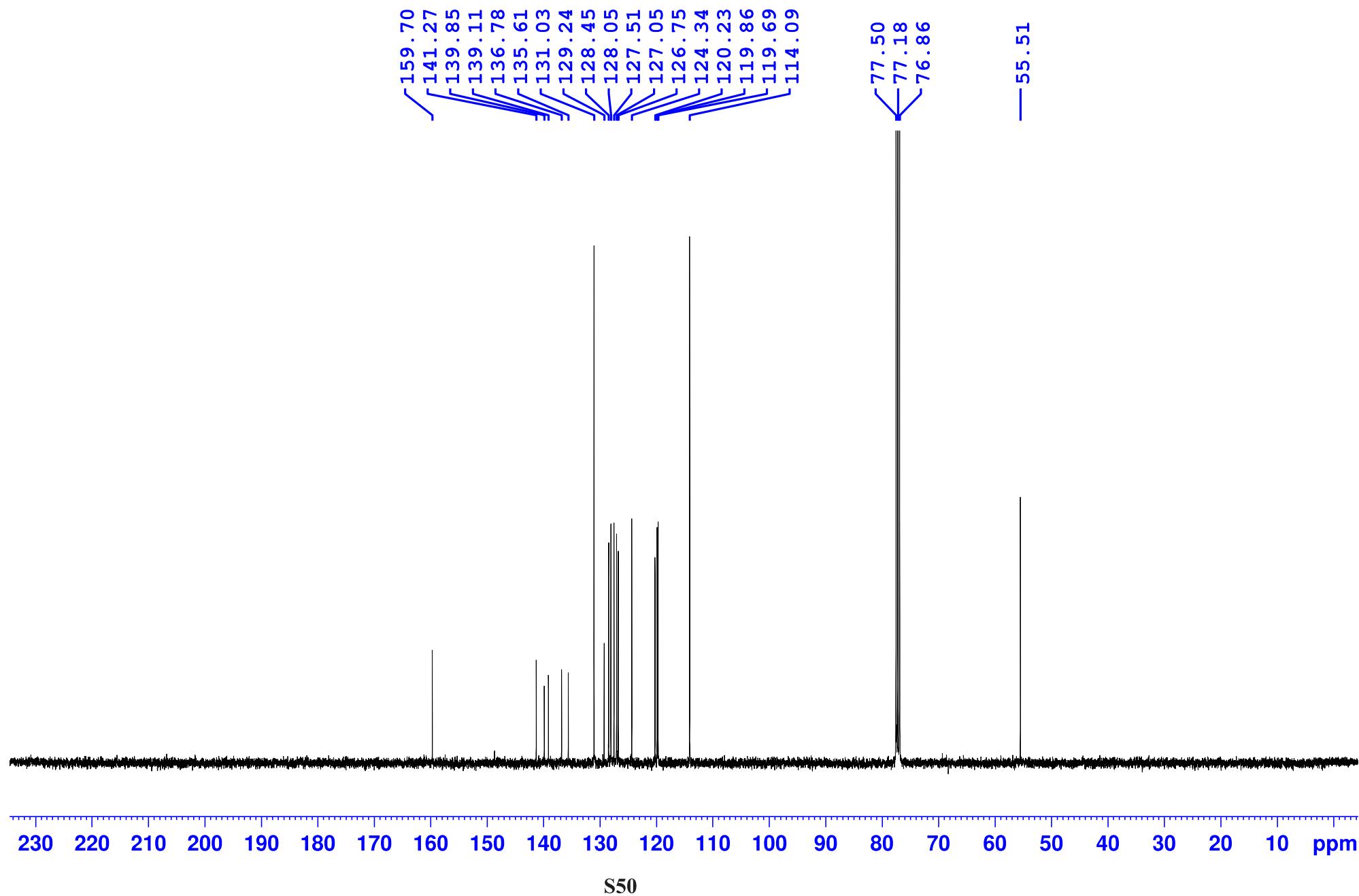


Current Data Parameters  
 NAME Shumaila Majeed  
 EXPNO 268  
 PROCNO 1

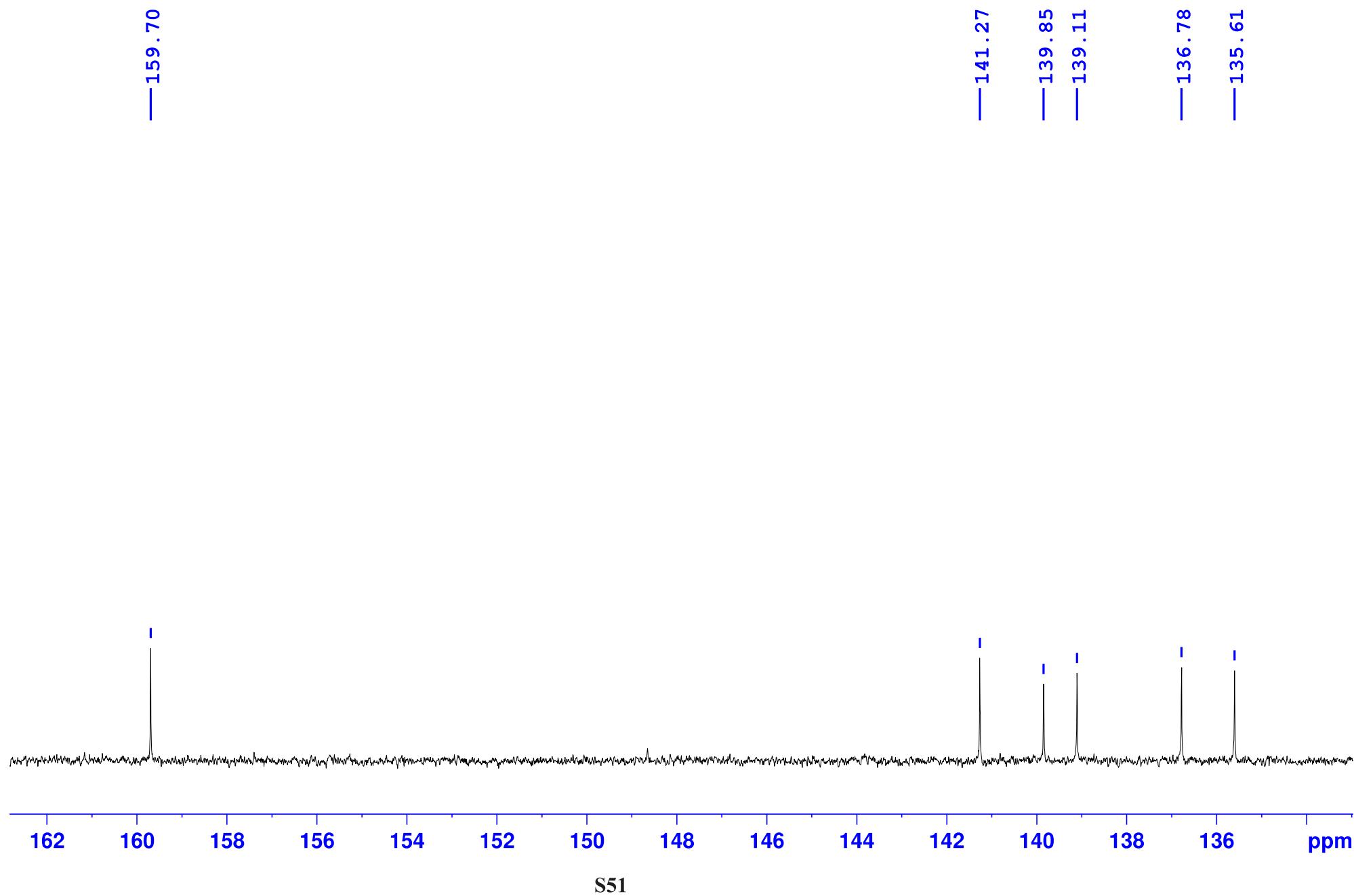
F2 - Acquisition Parameters  
 Date\_ 20211013  
 Time 13.13 h  
 INSTRUM spect  
 PROBHD Z116098\_0621 (zgpg30  
 PULPROG 65536  
 TD 400  
 SOLVENT CDC13  
 NS 2  
 DS 24038.461 Hz  
 SWH 0.733596 Hz  
 FIDRES 1.3631488 sec  
 AQ 199.48  
 RG 20.800 usec  
 DE 6.50 usec  
 TE 298.0 K  
 D1 2.0000000 sec  
 D11 0.0300000 sec  
 TDO 1  
 SFO1 100.6243390 MHz  
 NUC1  $^{13}\text{C}$   
 P1 10.00 usec  
 PLW1 72.56700134 W  
 SFO2 400.1316005 MHz  
 NUC2 1H  
 CPDPRG[2] waltz16  
 PCPD2 90.00 usec  
 PLW2 16.68099976 W  
 PLW12 0.20593999 W  
 PLW13 0.10342000 W

F2 - Processing parameters  
 SI 32768  
 SF 100.6127547 MHz  
 WDW EM  
 SSB 0  
 LB 1.00 Hz  
 GB 0  
 PC 1.40

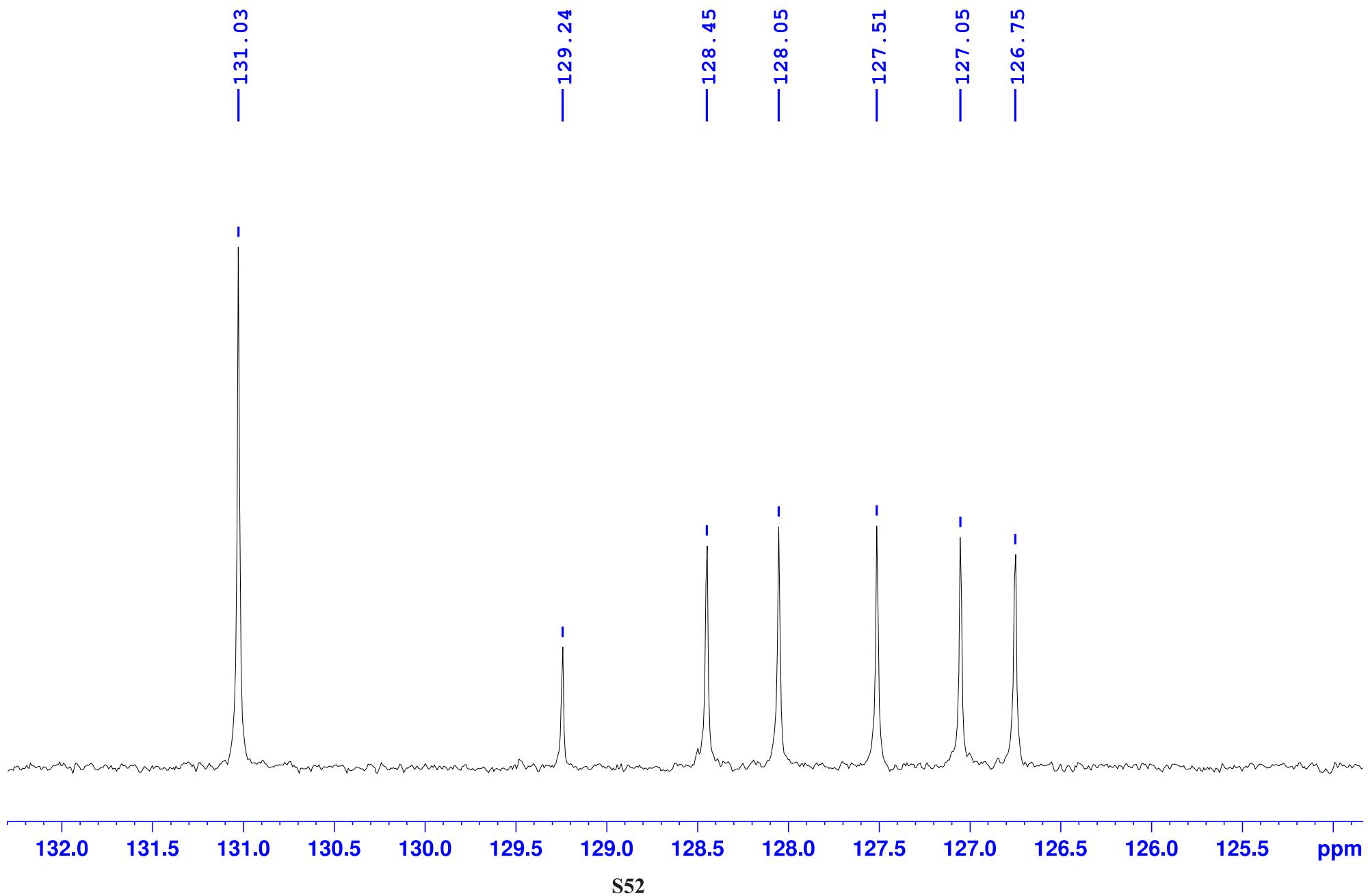
Compound 1c,  $^{13}\text{C}$  NMR (100 MHz,  $\text{CDCl}_3$ )



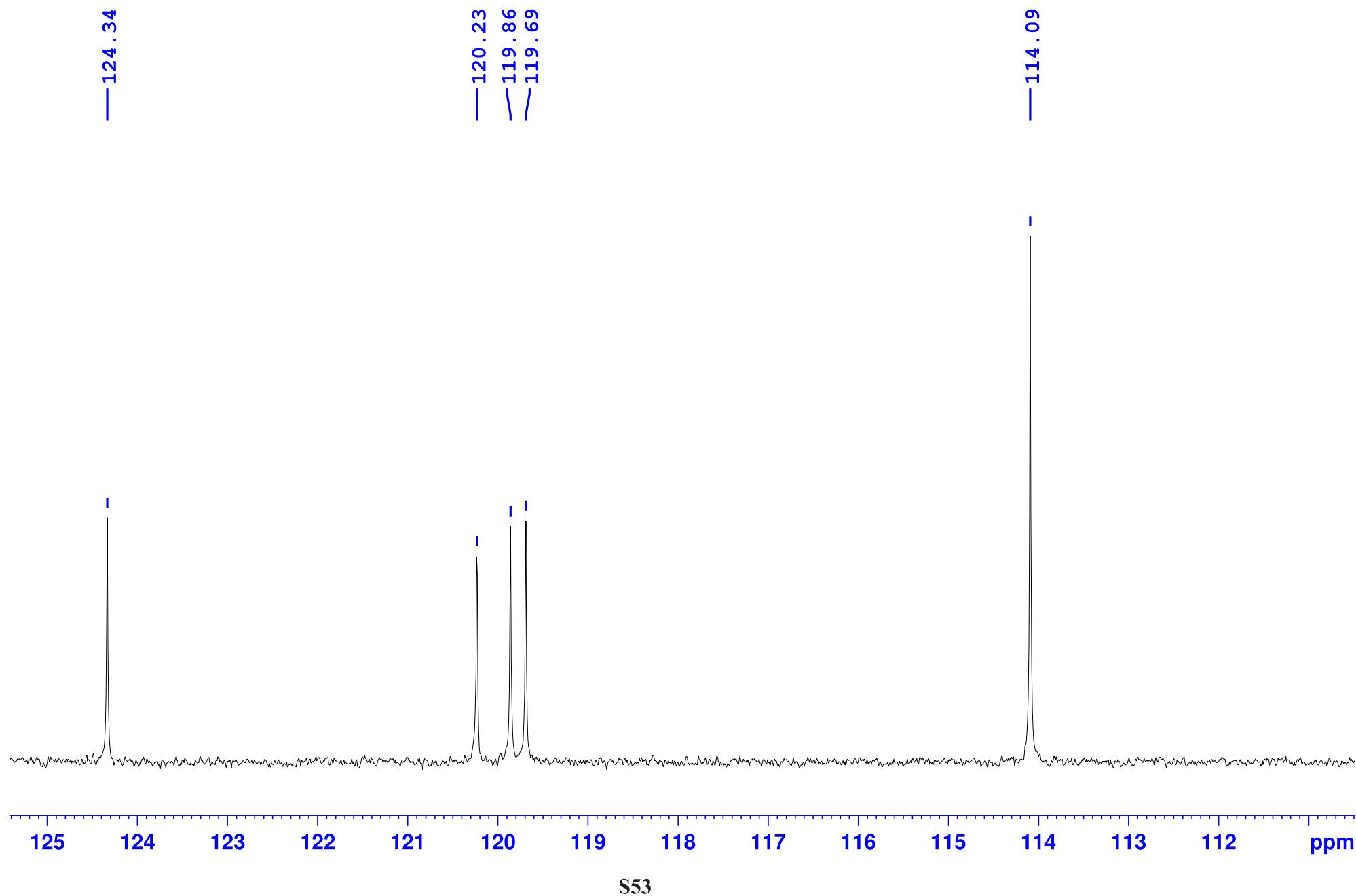
Compound 1c,  $^{13}\text{C}$  NMR (100 MHz,  $\text{CDCl}_3$ )



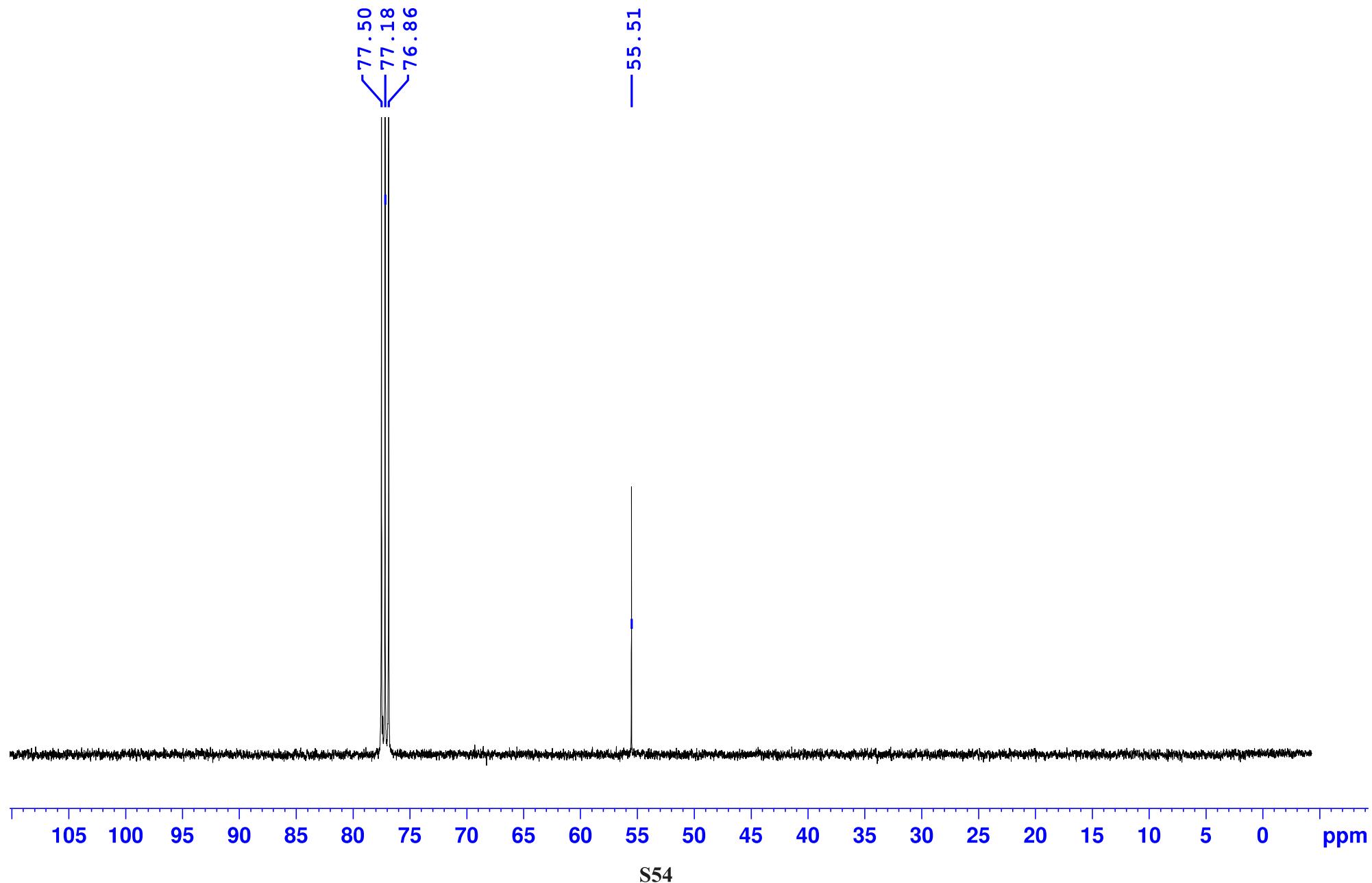
Compound 1c,  $^{13}\text{C}$  NMR (100 MHz,  $\text{CDCl}_3$ )



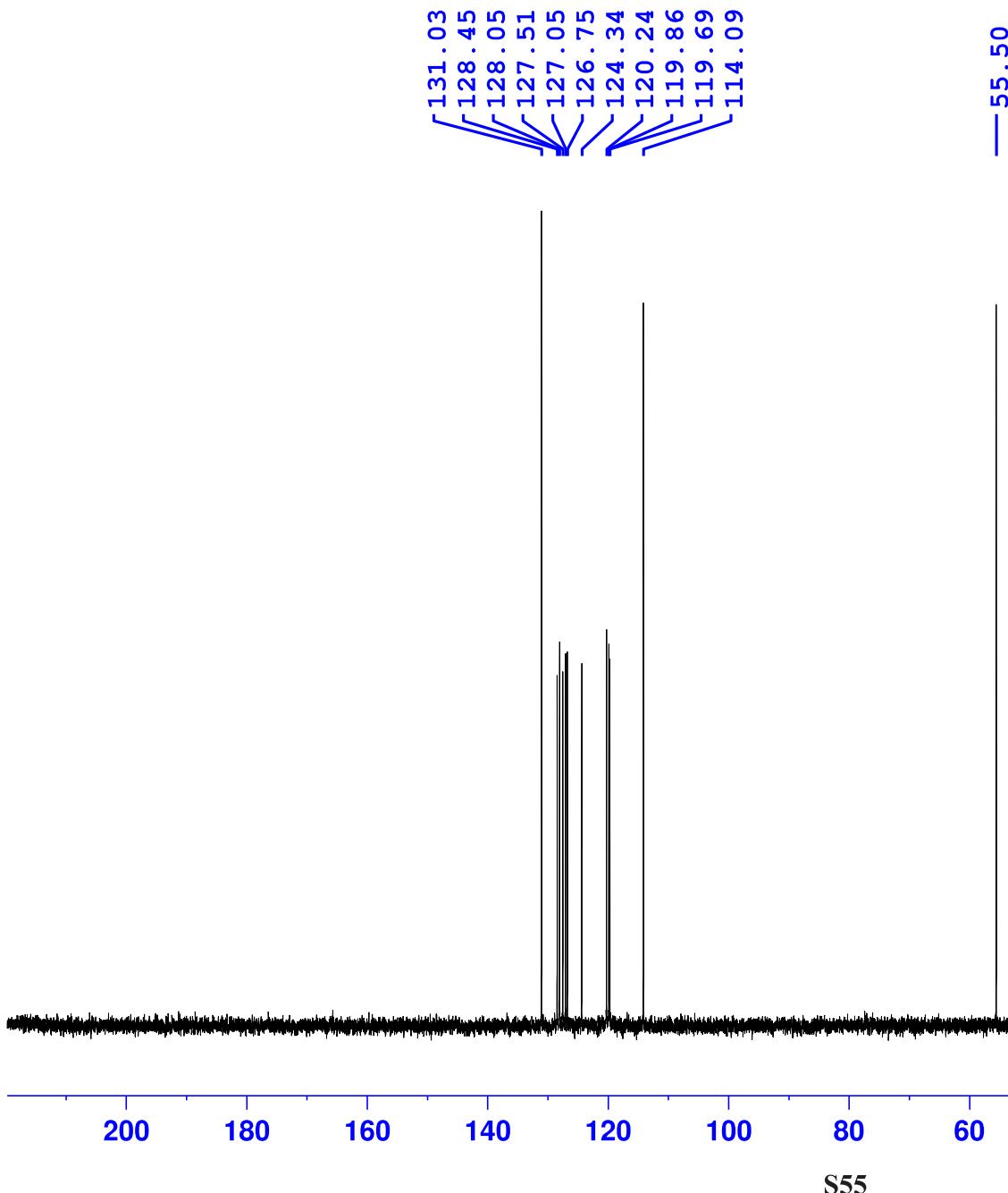
**Compound 1c,  $^{13}\text{C}$  NMR (100 MHz,  $\text{CDCl}_3$ )**



Compound 1c,  $^{13}\text{C}$  NMR (100 MHz,  $\text{CDCl}_3$ )



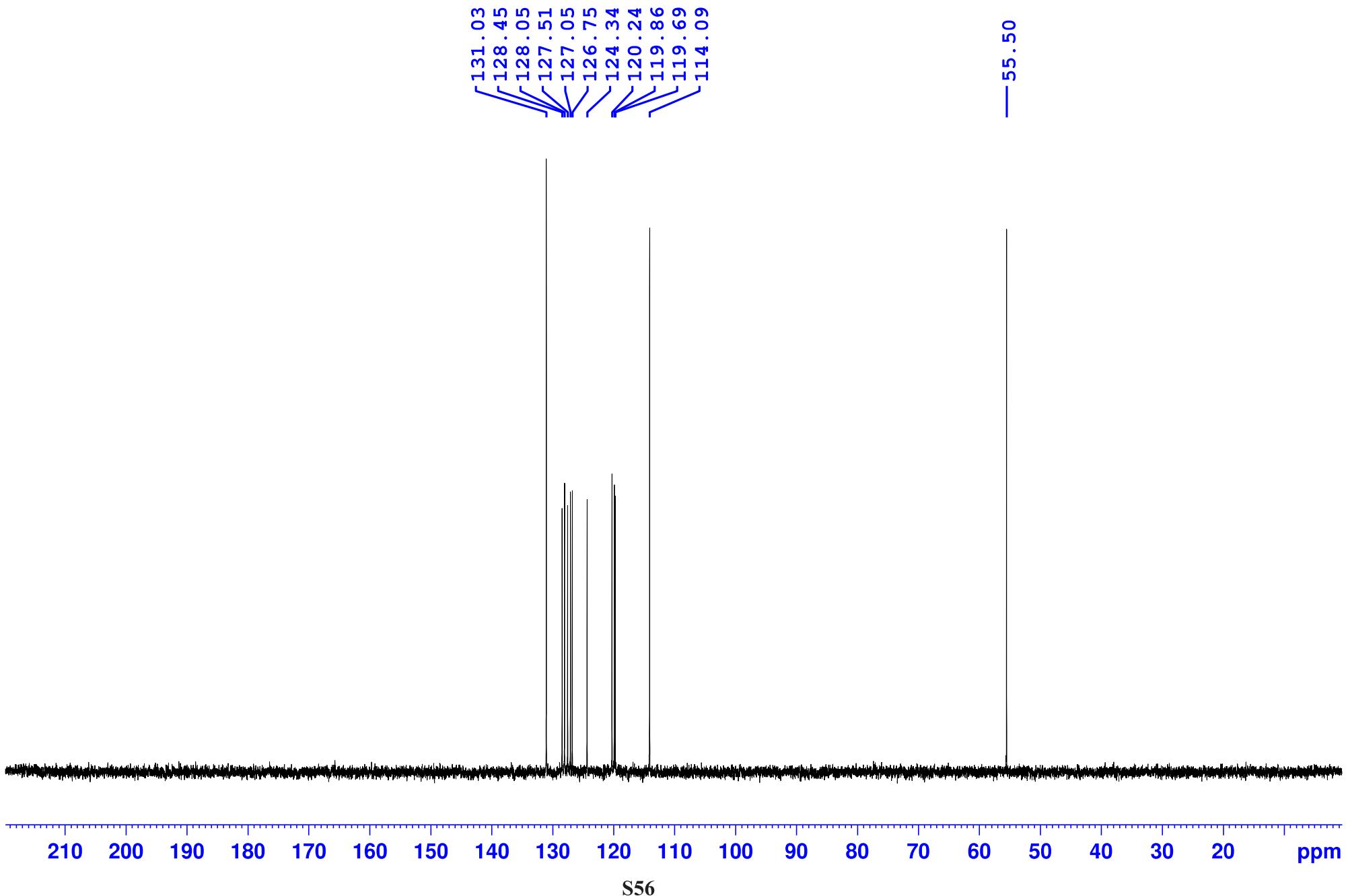
Compound 1c, DEPT 135 NMR (100 MHz, CDCl<sub>3</sub>)



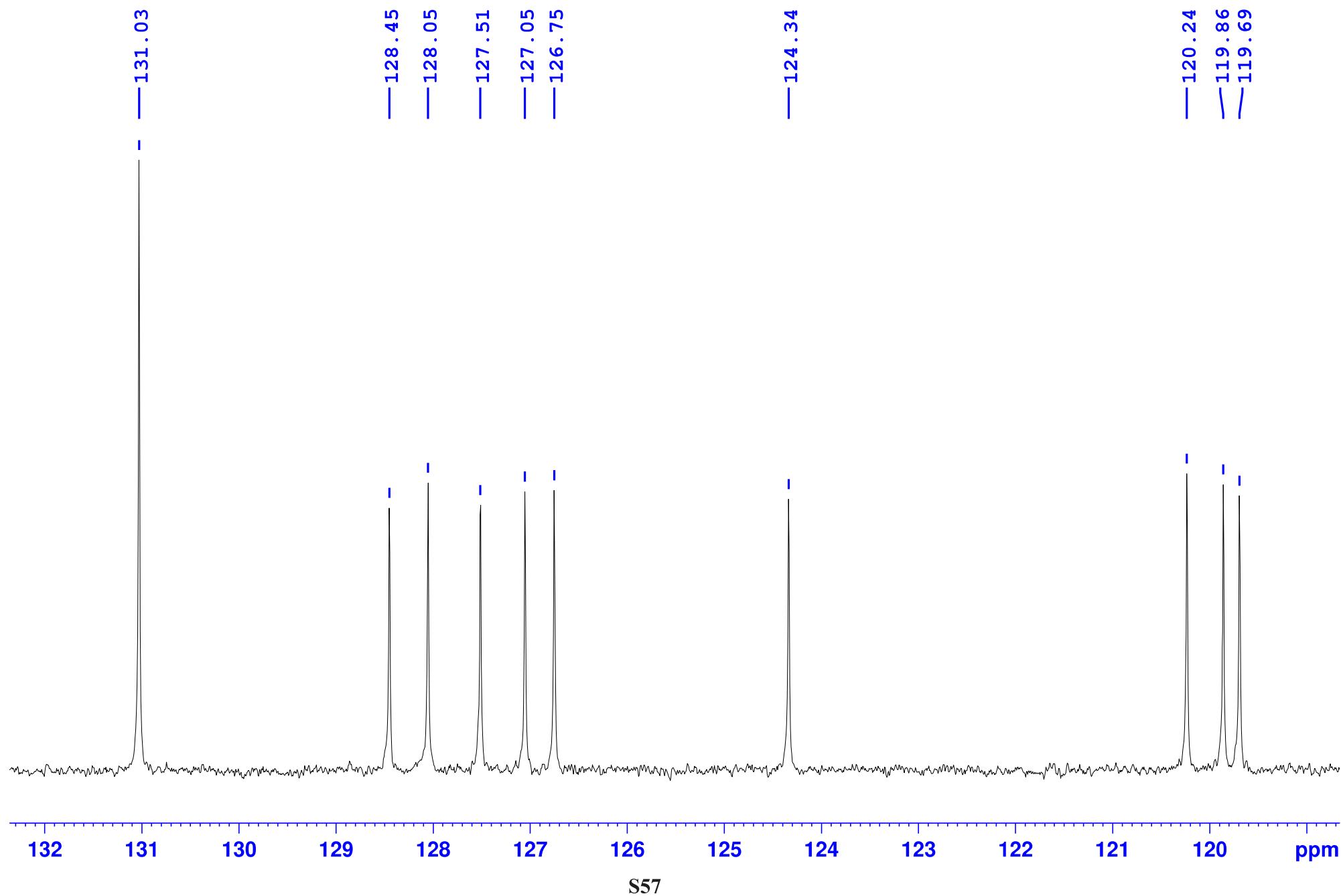
Current Data Parameters  
NAME Shumaila Majeed  
EXPNO 269  
PROCNO 1

F2 - Acquisition Parameters  
Date\_ 20211013  
Time 13.29 h  
INSTRUM spect  
PROBHD Z116098\_0621 (deptsp135  
PULPROG 65536  
TD 256  
SOLVENT CDC13  
NS 4  
DS 22058.824 Hz  
FIDRES 0.673182 Hz  
AQ 1.4854827 sec  
RG 199.48  
DW 22.667 usec  
DE 6.50 usec  
TE 298.0 K  
CNST2 145.000000  
D1 2.0000000 sec  
D2 0.00344828 sec  
D12 0.00002000 sec  
TD0 1  
SFO1 100.6238359 MHz  
NUC1 13C  
P1 10.00 usec  
P13 2000.00 usec  
PLW0 0 W  
PLW1 72.56700134 W  
SPNAM[5] Crp60comp.4  
SPOAL5 0.500  
SPOFFS5 0 Hz  
SPW5 11.08699989 W  
SFO2 400.1316005 MHz  
NUC2 1H  
CPDPKG[2] waltz16  
P3 10.00 usec  
P4 20.00 usec  
PCPD2 90.00 usec  
PLW2 16.68099976 W  
PLW12 0.20593999 W

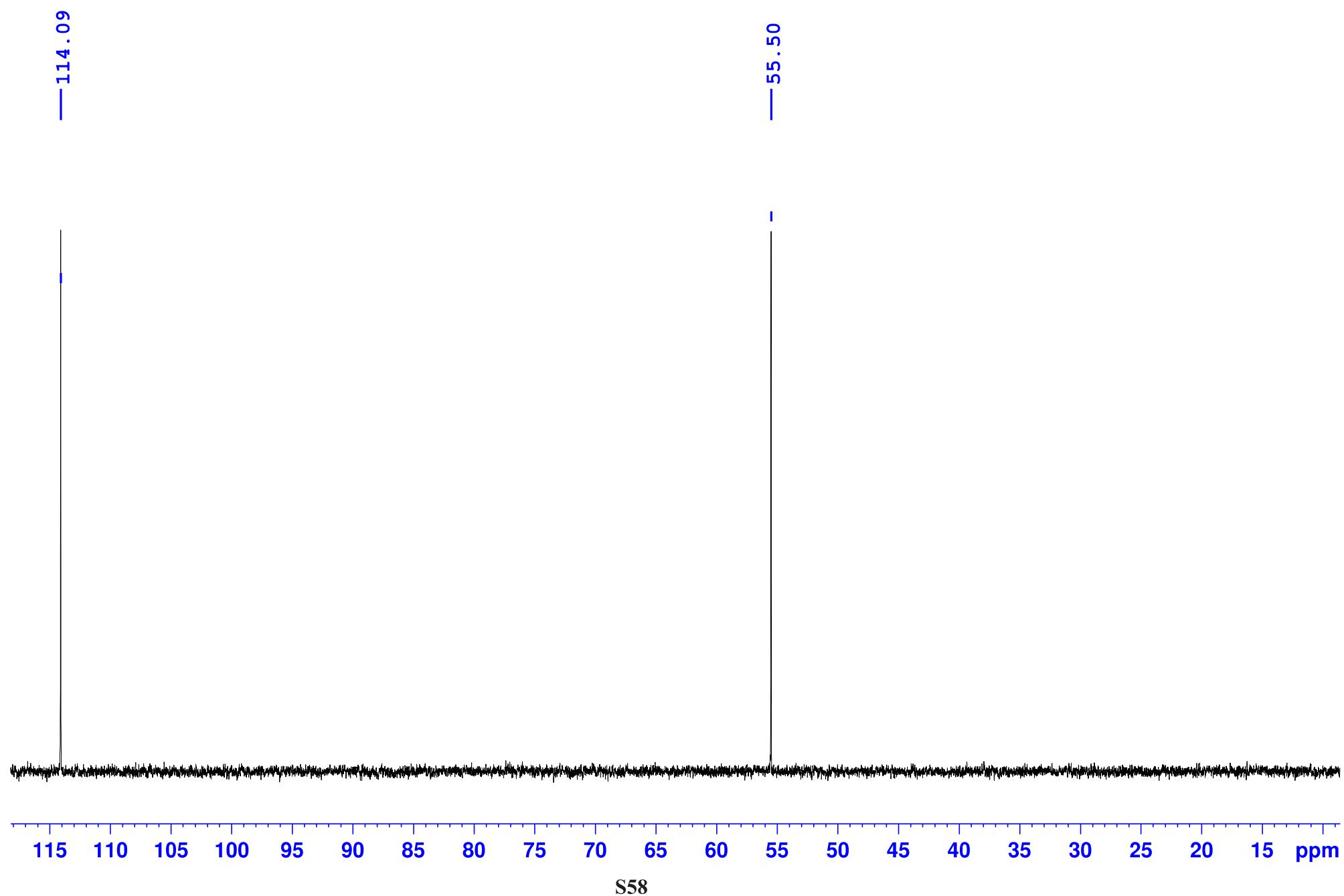
Compound 1c, DEPT 135 NMR (100 MHz, CDCl<sub>3</sub>)



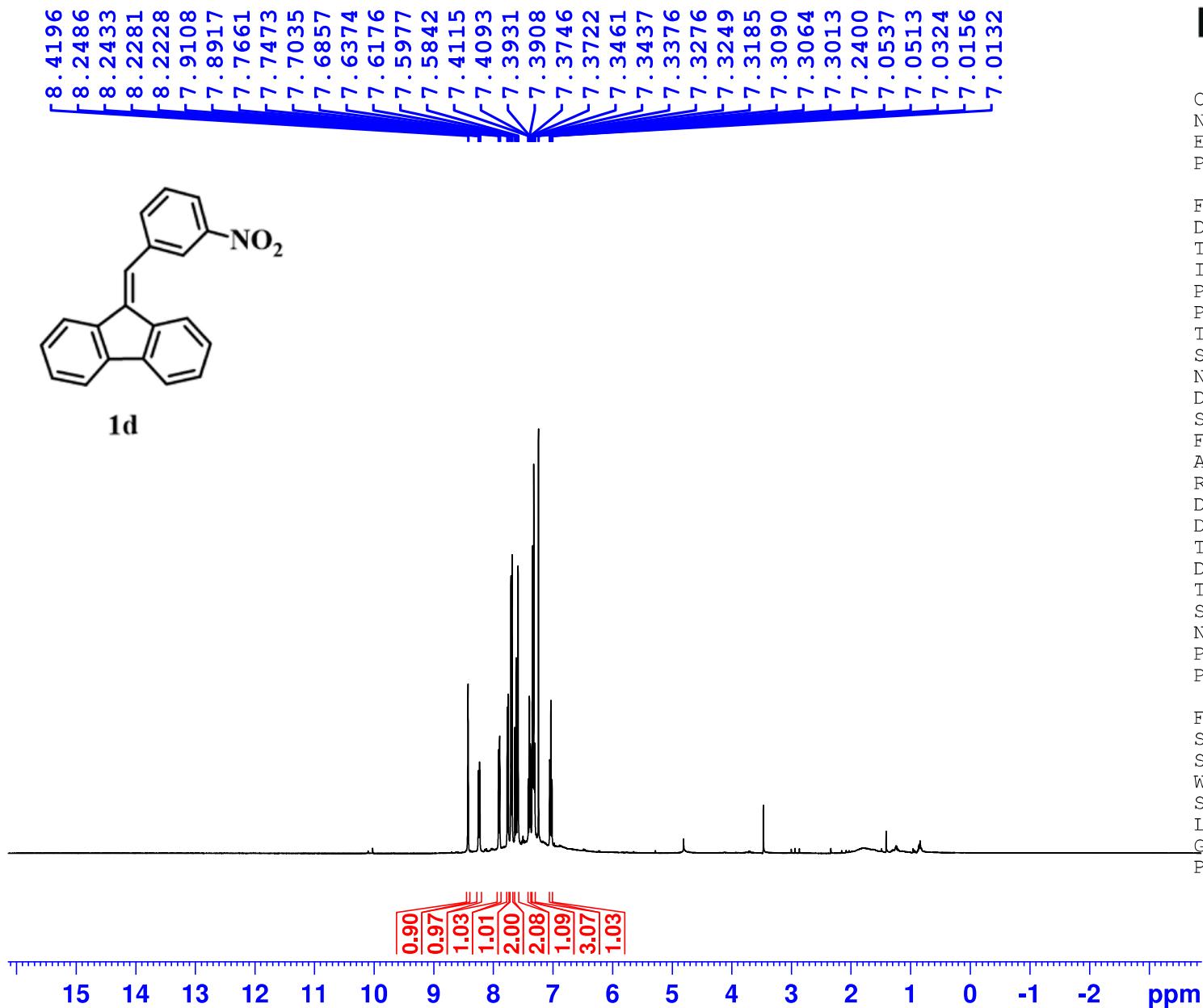
Compound 1c, DEPT 135 NMR (100 MHz, CDCl<sub>3</sub>)



Compound 1c, DEPT 135 NMR (100 MHz, CDCl<sub>3</sub>)



Compound 1d,  $^1\text{H}$  NMR (400 MHz,  $\text{CDCl}_3$ )



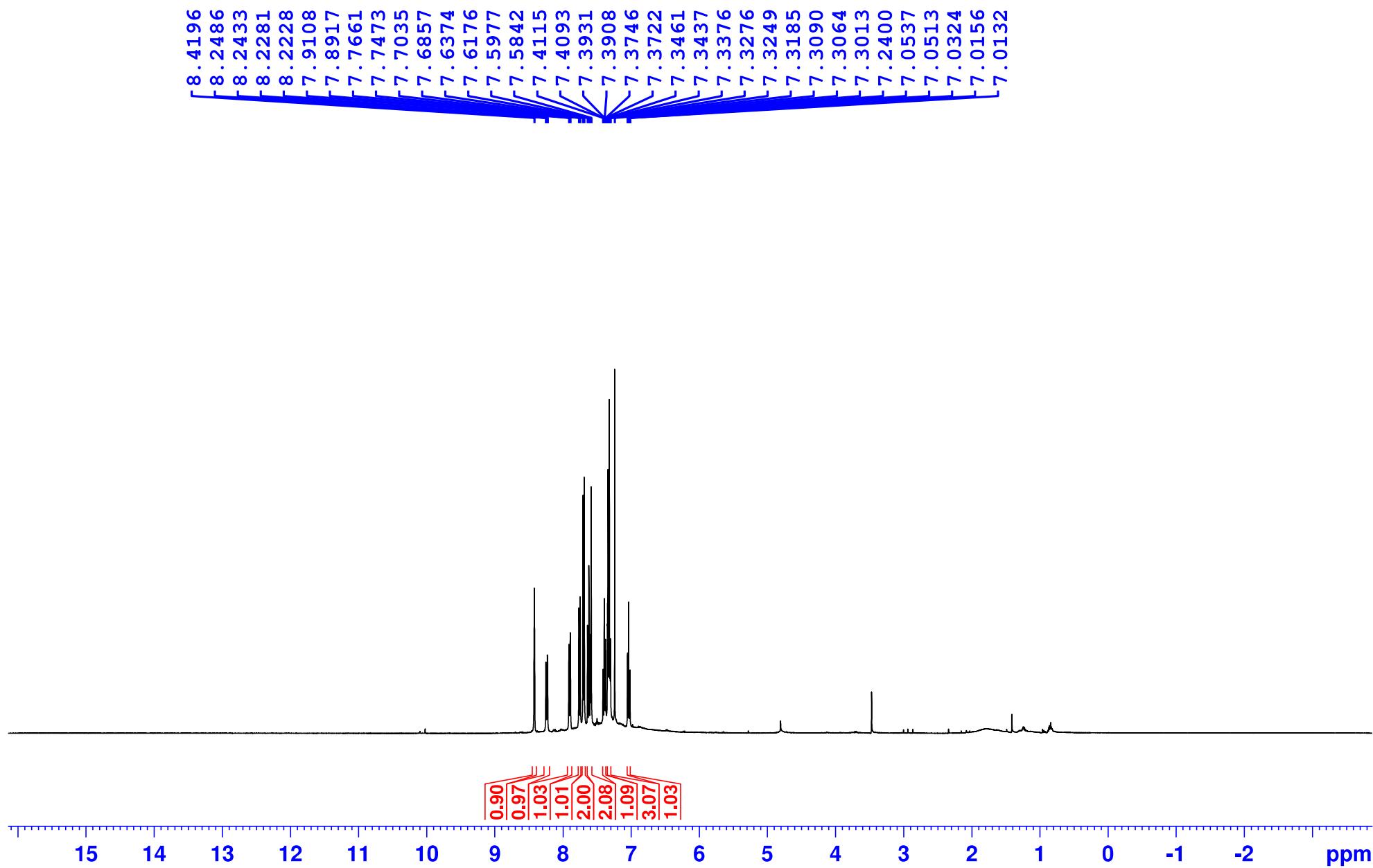
**BRUKER**

Current Data Parameters  
NAME SMT-8  
EXPNO 157  
PROCNO 1

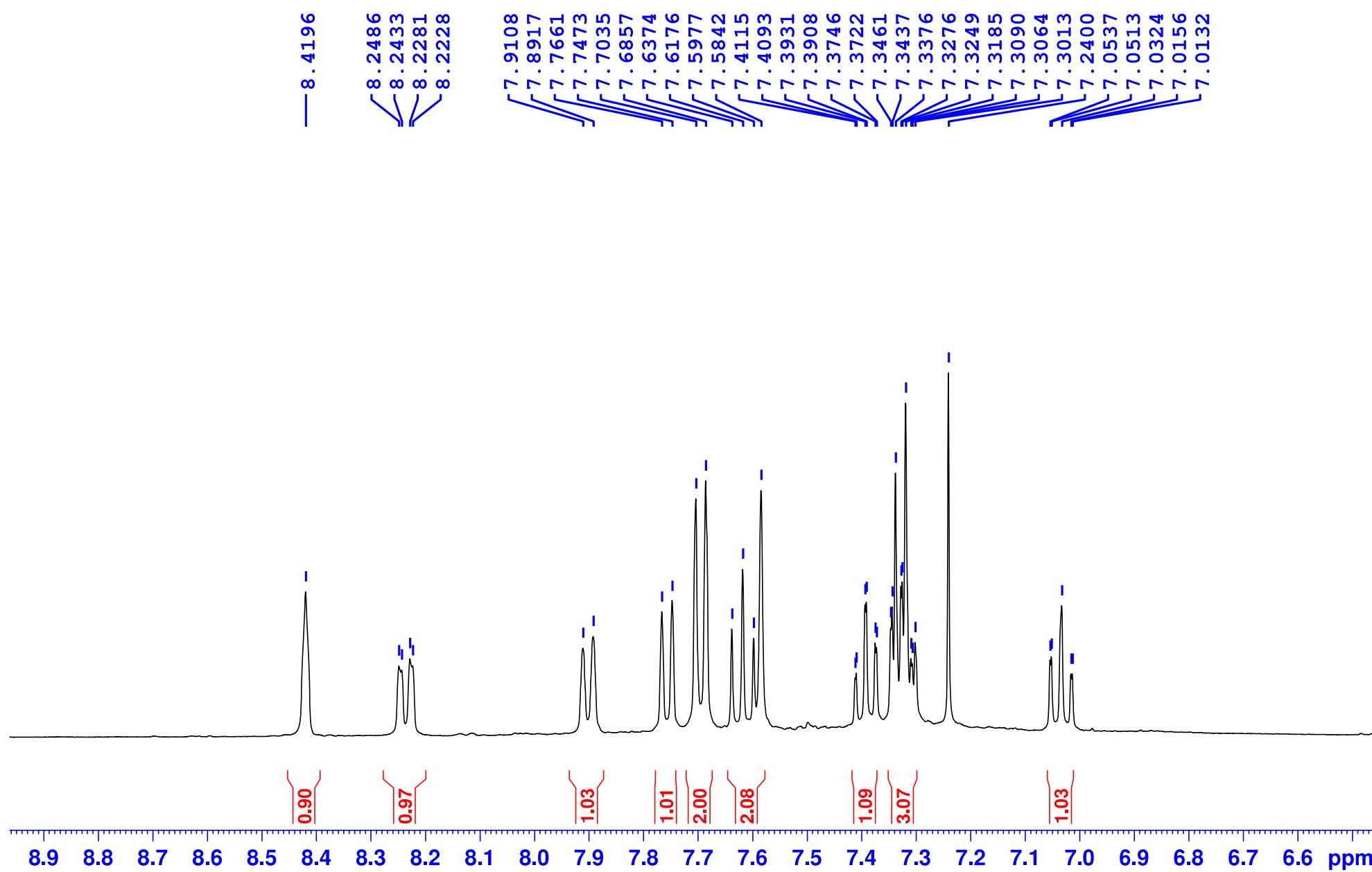
F2 - Acquisition Parameters  
Date\_ 20210305  
Time 15.50 h  
INSTRUM spect  
PROBHD Z116098\_0621 (   
PULPROG zg30  
TD 65536  
SOLVENT CDCl<sub>3</sub>  
NS 16  
DS 2  
SWH 8012.820 Hz  
FIDRES 0.244532 Hz  
AQ 4.0894465 sec  
RG 98.53  
DW 62.400 usec  
DE 6.50 usec  
TE 298.0 K  
D1 1.00000000 sec  
TD0 1  
SF01 400.1324708 MHz  
NUC1 1H  
P1 10.00 usec  
PLW1 16.68099976 W

F2 - Processing parameters  
SI 65536  
SF 400.1300172 MHz  
WDW EM  
SSB 0  
LB 0.30 Hz  
GB 0  
PC 1.00

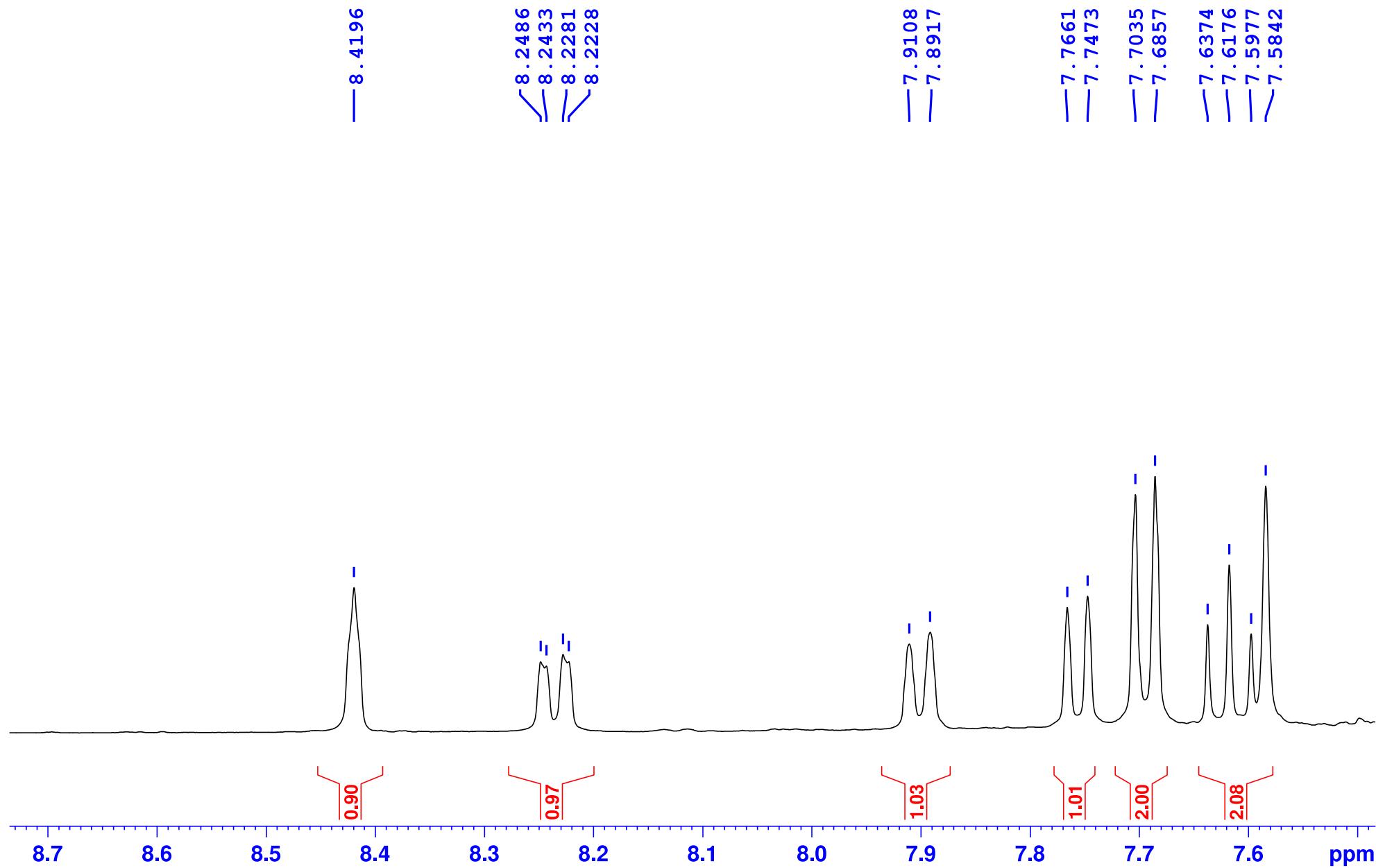
Compound 1d,  $^1\text{H}$  NMR (400 MHz,  $\text{CDCl}_3$ )



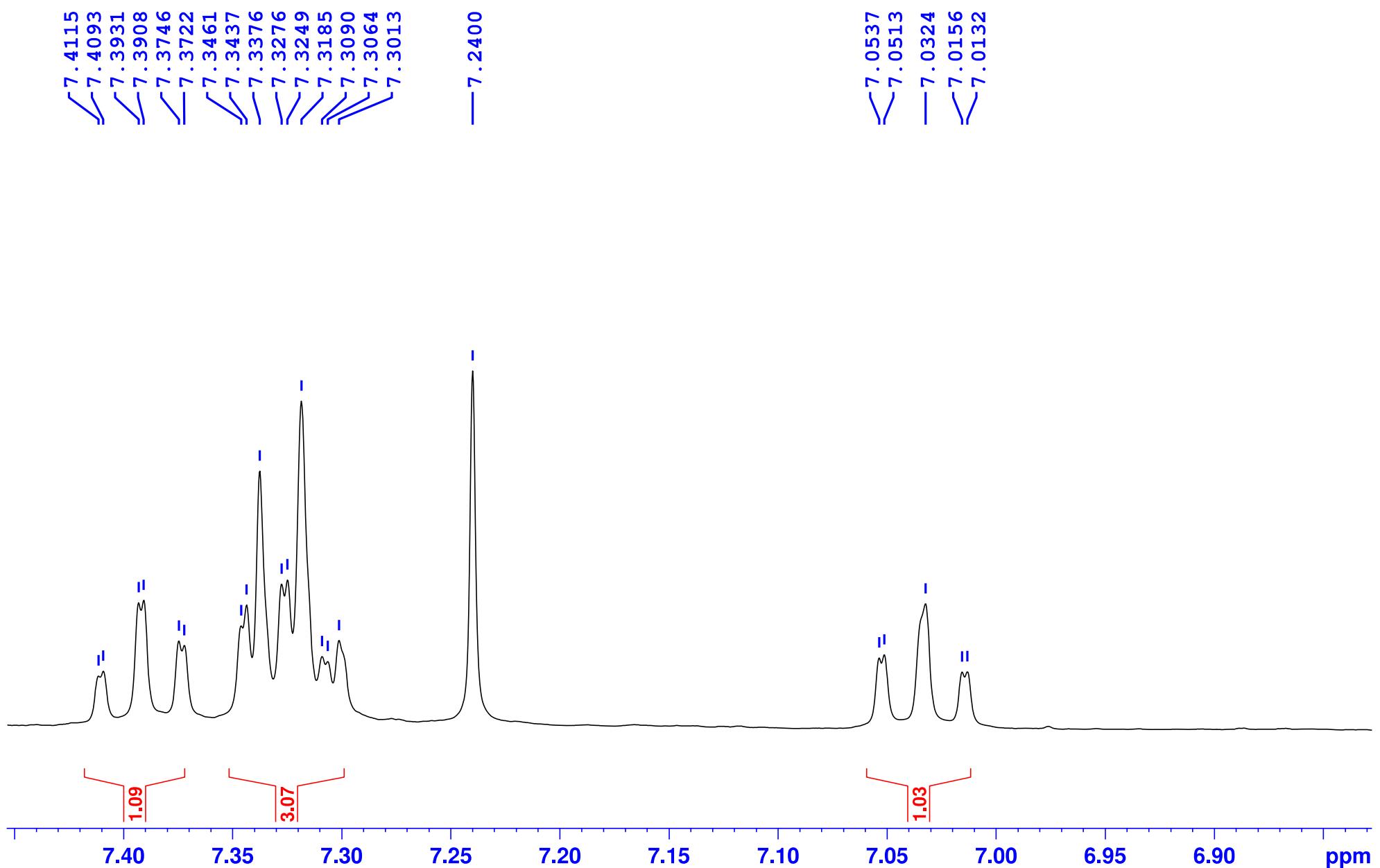
Compound 1d,  $^1\text{H}$  NMR (400 MHz,  $\text{CDCl}_3$ )



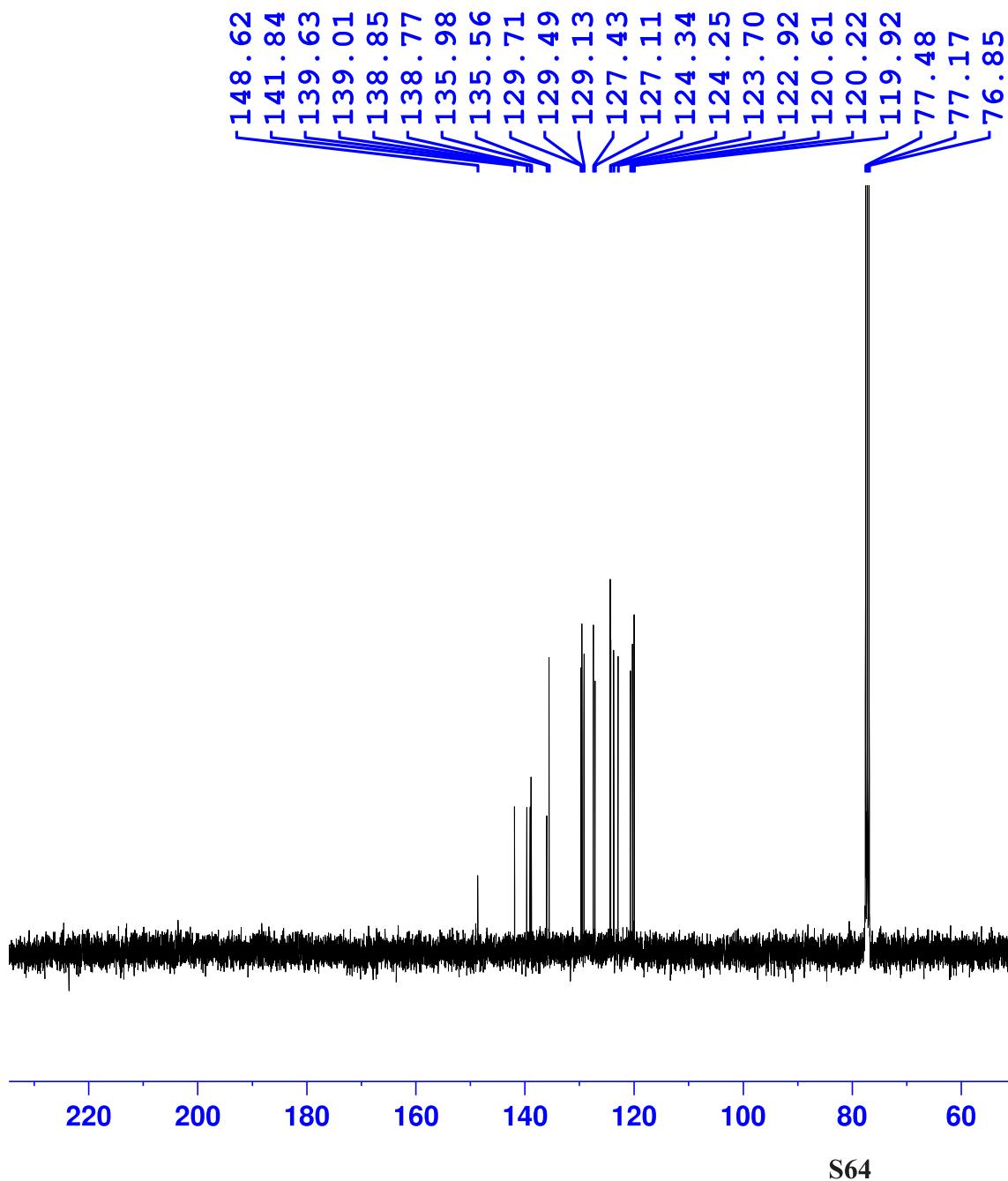
Compound 1d,  $^1\text{H}$  NMR (400 MHz,  $\text{CDCl}_3$ )



Compound 1d,  $^1\text{H}$  NMR (400 MHz,  $\text{CDCl}_3$ )



Compound 1d,  $^{13}\text{C}$  NMR (100 MHz,  $\text{CDCl}_3$ )

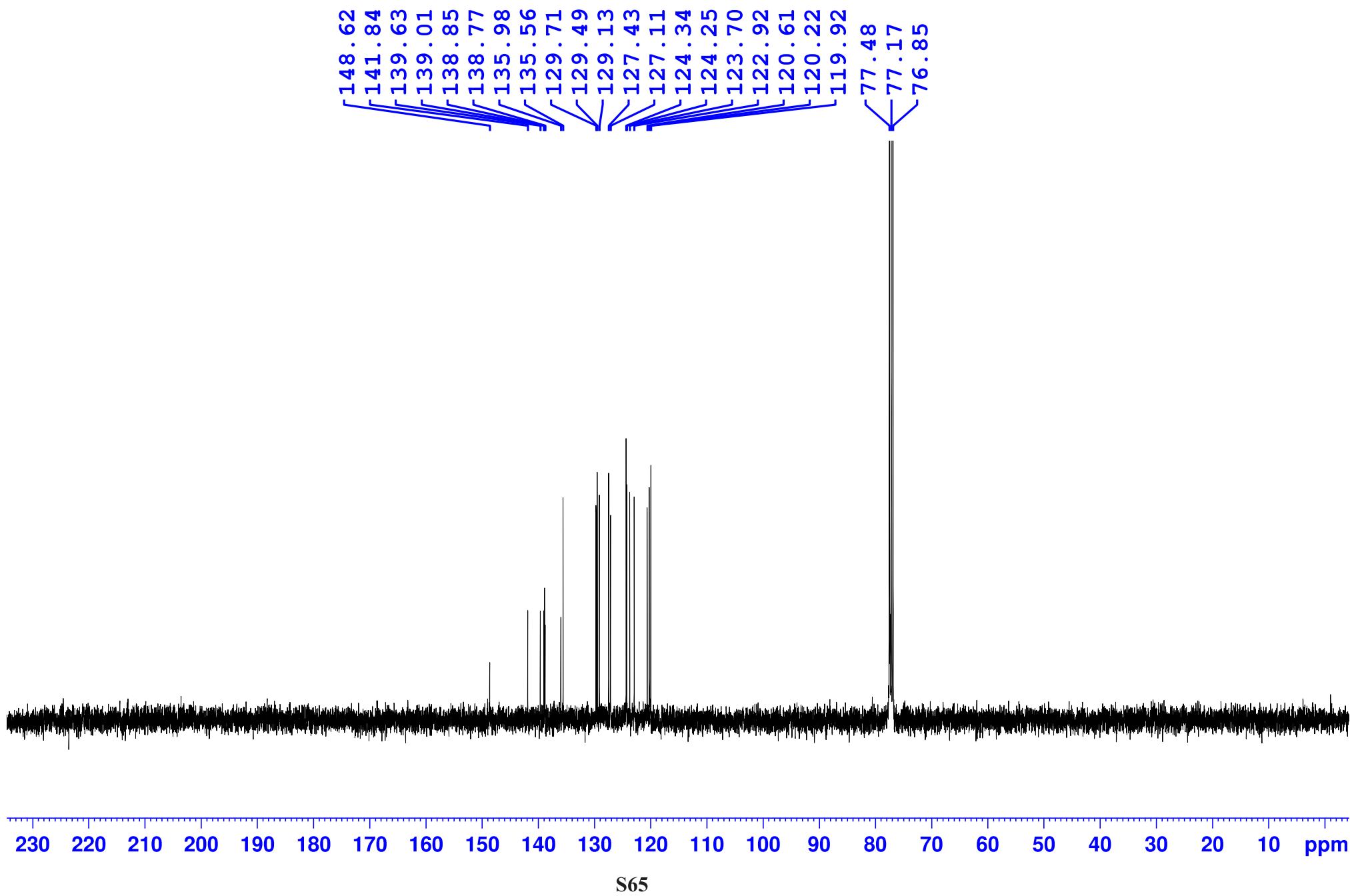


Current Data Parameters  
 NAME Shumaila Majeed  
 EXPNO 274  
 PROCNO 1

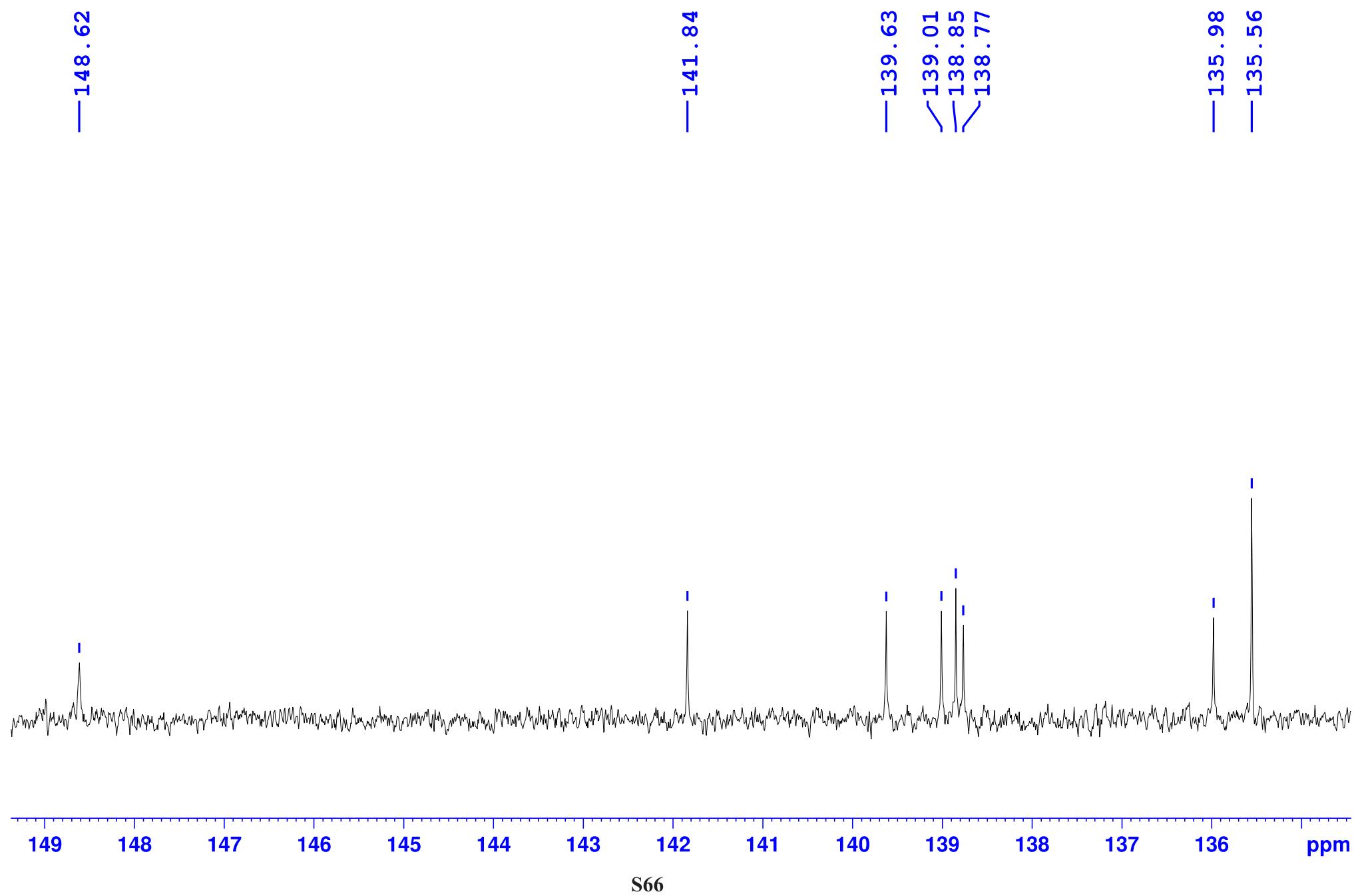
F2 - Acquisition Parameters  
 Date\_ 20211013  
 Time 15.22 h  
 INSTRUM spect  
 PROBHD Z116098\_0621 (zgpg30  
 PULPROG 65536  
 TD 400  
 SOLVENT CDC13  
 NS 2  
 DS 24038.461 Hz  
 SWH 0.733596 Hz  
 FIDRES 1.3631488 sec  
 AQ 199.48  
 RG 20.800 usec  
 DE 6.50 usec  
 TE 298.0 K  
 D1 2.0000000 sec  
 D11 0.0300000 sec  
 TDO 1  
 SFO1 100.6243390 MHz  
 NUC1  $^{13}\text{C}$   
 P1 10.00 usec  
 PLW1 72.56700134 W  
 SFO2 400.1316005 MHz  
 NUC2 1H  
 CPDPRG[2] waltz16  
 PCPD2 90.00 usec  
 PLW2 16.68099976 W  
 PLW12 0.20593999 W  
 PLW13 0.10342000 W

F2 - Processing parameters  
 SI 32768  
 SF 100.6127547 MHz  
 WDW EM  
 SSB 0  
 LB 1.00 Hz  
 GB 0  
 PC 1.40

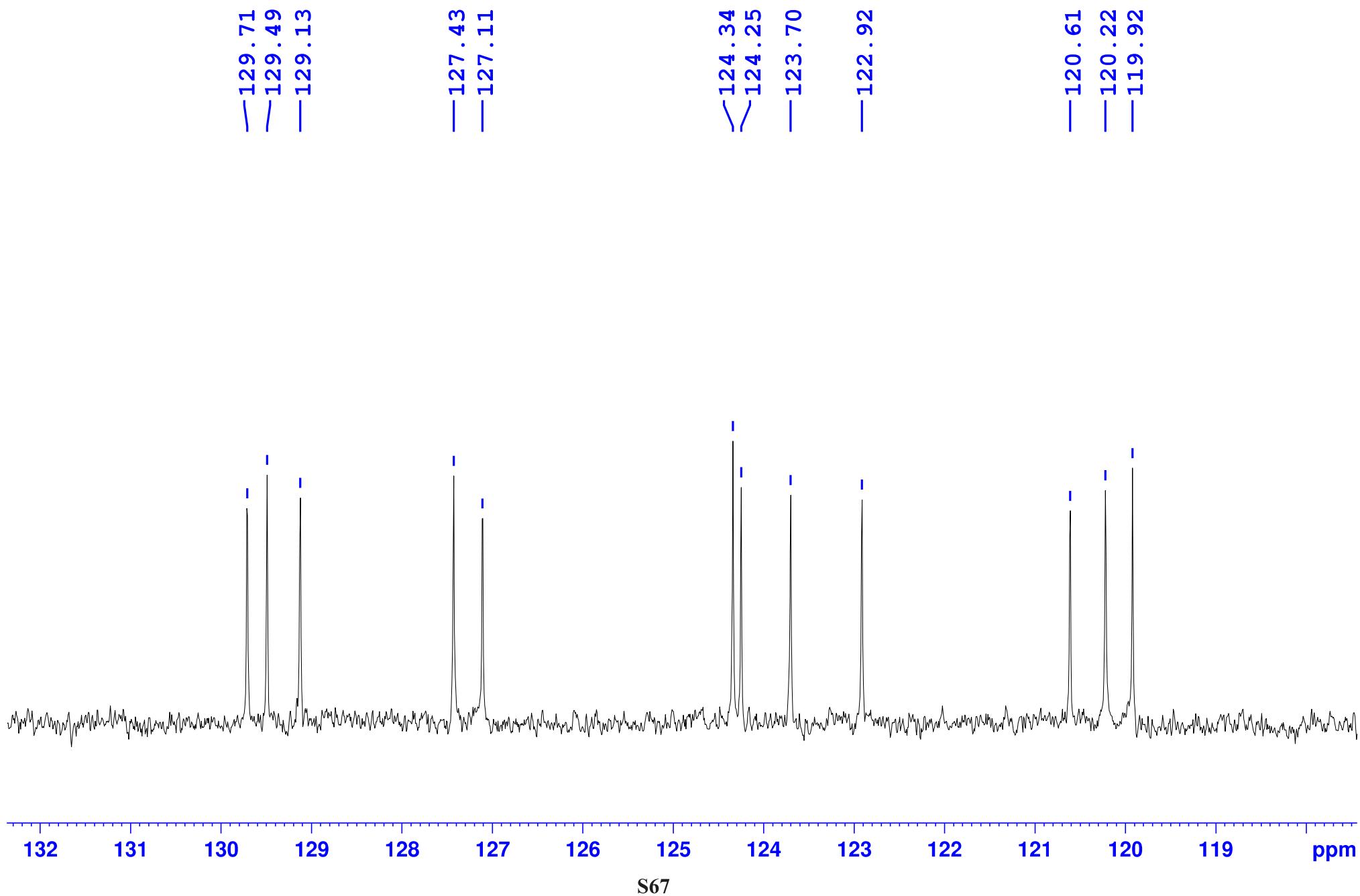
Compound 1d,  $^{13}\text{C}$  NMR (100 MHz,  $\text{CDCl}_3$ )



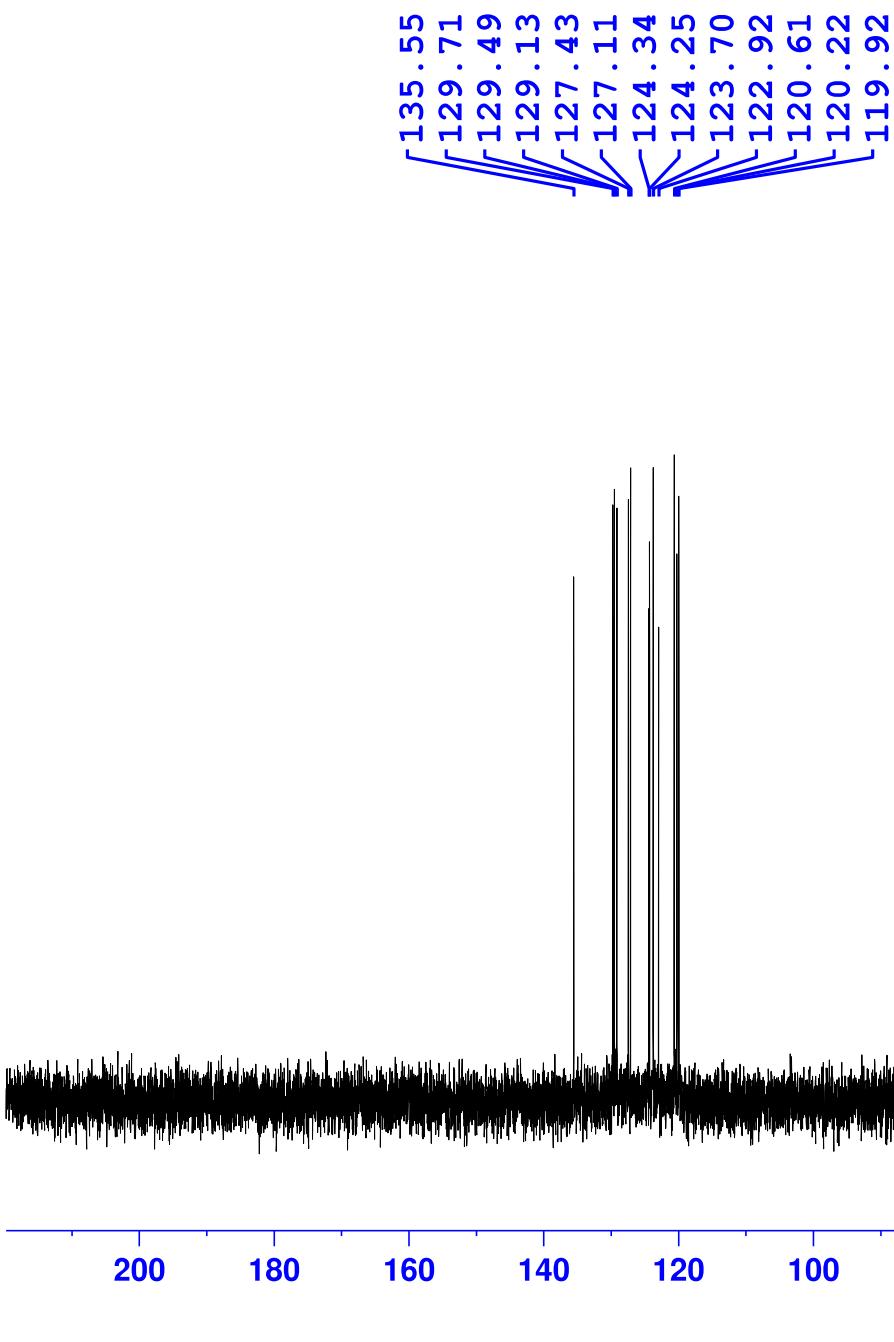
**Compound 1d,  $^{13}\text{C}$  NMR (100 MHz,  $\text{CDCl}_3$ )**



**Compound 1d,  $^{13}\text{C}$  NMR (100 MHz,  $\text{CDCl}_3$ )**



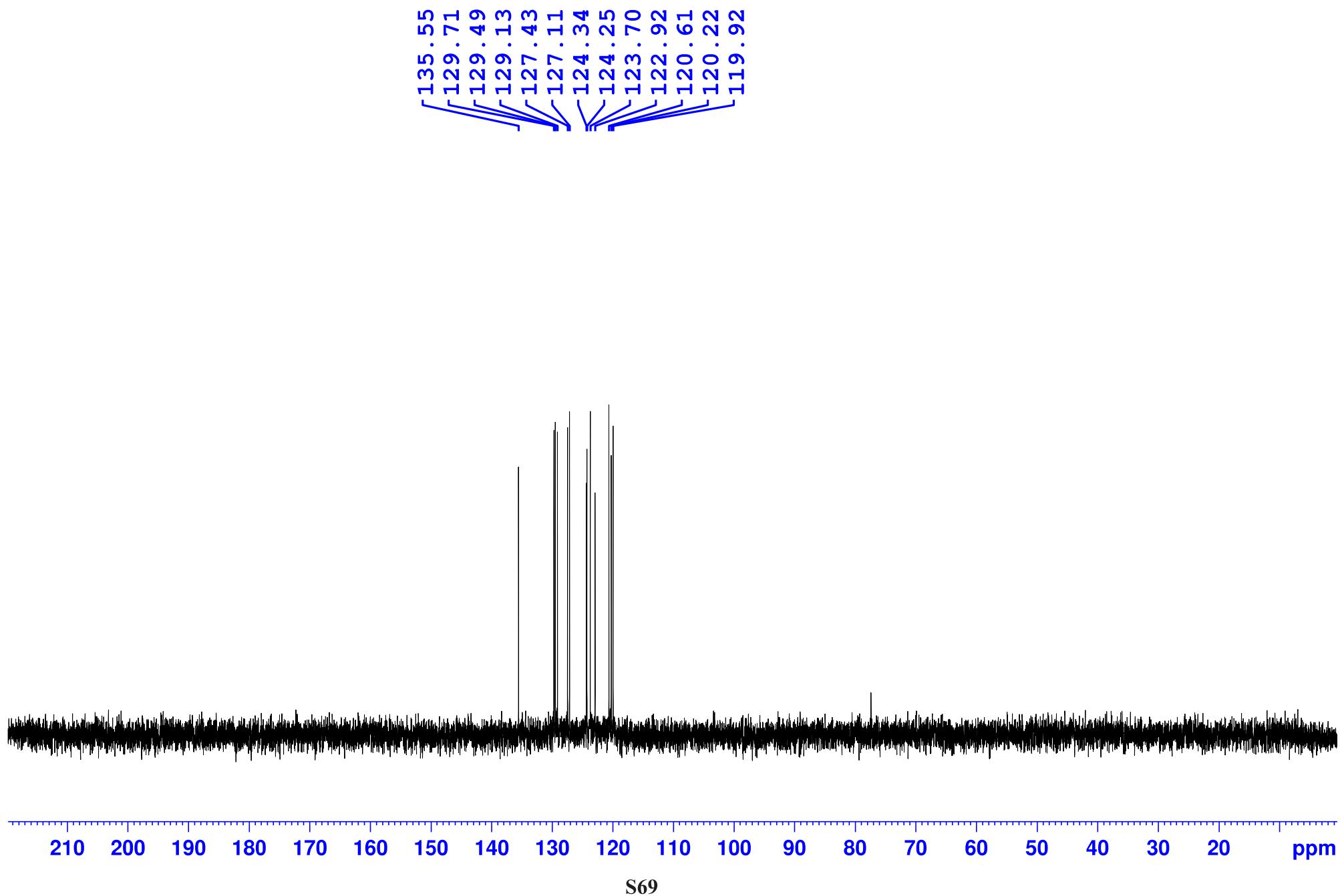
Compound 1d, DEPT 135 NMR (100 MHz, CDCl<sub>3</sub>)



Current Data Parameters  
 NAME Shumaila Majeed  
 EXPNO 275  
 PROCNO 1

F2 - Acquisition Parameters  
 Date\_ 20211013  
 Time 15.38 h  
 INSTRUM spect  
 PROBHD Z116098\_0621 (   
 PULPROG deptsp135  
 TD 65536  
 SOLVENT CDCl<sub>3</sub>  
 NS 256  
 DS 4  
 SWH 22058.824 Hz  
 FIDRES 0.673182 Hz  
 AQ 1.4854827 sec  
 RG 199.48  
 DW 22.667 usec  
 DE 6.50 usec  
 TE 298.0 K  
 CNST2 145.0000000  
 D1 2.00000000 sec  
 D2 0.00344828 sec  
 D12 0.00002000 sec  
 TD0 1  
 SFO1 100.6238359 MHz  
 NUC1 13C  
 P1 10.00 usec  
 P13 2000.00 usec  
 PLW0 0 W  
 PLW1 72.56700134 W  
 SPNAM[5] Crp60comp.4  
 SPOAL5 0.500  
 SPOFFS5 0 Hz  
 SPW5 11.08699989 W  
 SFO2 400.1316005 MHz  
 NUC2 1H  
 CPDPRG[2] waltz16  
 P3 10.00 usec  
 P4 20.00 usec  
 PCPD2 90.00 usec  
 PLW2 16.68099976 W  
 PLW12 0.20593999 W

Compound 1d, DEPT 135 NMR (100 MHz, CDCl<sub>3</sub>)



**Compound 1d, DEPT 135 NMR (100 MHz, CDCl<sub>3</sub>)**

