ELECTRONIC SUPPLEMENTARY INFORMATION

Use of dielectrophoresis for directing T cells to microwells before nanostraw transfection: modelling and experiments.

Mercy Lard¹, Bao D. Ho¹, Jason P. Beech¹, Jonas O. Tegenfeldt¹, Christelle N. Prinz^{1*}

¹ Division of Solid State Physics and NanoLund, Lund University, 221 00 Lund, Sweden

* corresponding author: Christelle N. Prinz (christelle.prinz@ftf.lth.se)



Figure S1: COMSOL simulations of the Real part of the Clausius Mossotti factor. (parameters)



Figure S2: Representative images of cells trapped in the microwells using DEP and subsequently stained with CalceinAM for cell viability. (a) Bright field image. (b) Calcein fluorescence. A red dot was added to the left of each visible cell in the bright field image and the red dots were added to the fluorescence image in order to facilitate comparison. Scale bars $30 \,\mu\text{m}$.



Figure S3: Representative images of cells trapped in the microwells using DEP and subsequently injected with PI. (a) Bright field image. (b) PI fluorescence after PI injection. A red dot was added to the left of each visible cell in the bright field image and the red dots were added to the fluorescence image in order to facilitate comparison. Scale bars $30 \,\mu\text{m}$.



Figure S4: Representative images of cells trapped in the microwells using DEP and subsequently undergoing mock EP followed by staining with CalceinAM for cell viability. (a) Bright field image. (b) Calcein fluorescence. A red dot was added to the left of each visible cell in the bright field image and the red dots were added to the fluorescence image in order to facilitate comparison.Scale bars 30 μ m.

Notation	Value	Description
D _{well}	15 μm	Diameter of the well
D _{straw}	160 nm	Outer diameter of the straws
t _{Al203}	10 nm	Thickness of the straws
L _{straw_to_straw}	2.2 μm	Distance between two straws in the same row or
		column
W _{domain}	32 µm	Width of the simulated domain, i.e. the
		geometrical unit cell that contains one microwell,
		mentioned in the text
t _{top}	34 µm	Thickness of the top membrane after the etching
		step
t _{bottom}	24 μm	Thickness of the bottom membrane after the
		etching step
H _{tip}	1 μm	Height of the tips of the straws, extruding at the
		bottom of the well.
t _{medium_above}	500 μm	Thickness of the liquid medium above the
		membranes
t_{medium_below}	50 μm	Thickness of the liquid medium below the
		membranes
D _{cell}	10 μm	Diameter of the cell
t _{cell_membrane}	5 nm	Thickness of the cell membrane
z _{cell}	varied	z-coordinate of the cell, relative to the bottom of
		the well. This parameter is varied together with
		the $\boldsymbol{x}_{\text{cell}}$ below so that a map of electrical force on
		the cell can be obtained.
x _{cell}	varied	x-coordinate of the cell, relative to the center of
		the well. This parameter is varied together with
		the $\boldsymbol{z}_{\text{cell}}$ above so that a map of electrical force on
		the cell can be obtained.
V ₀	25 V	The magnitude of the applied voltage. This
		corresponds to a 50 $V_{\text{peak-to-peak}}$.
f_0	2.5 <i>MHz</i>	The frequency of the applied voltage.

Table S1: List of parameters used in the simulations

Movie S1: Available at https://youtu.be/tBqDs4s9Azo

Time lapse images of Jurkat T cells being pulled to wells with nanostraws at the bottom. The DEP force is applied on frame 106 (15,1 sec) in the movie.

Movie S2: Available at https://youtu.be/Zo9ok9eQ-Q0

Time lapse images of Jurkat T cells being pulled to wells with nanostraws at the bottom. The DEP force is applied on frame 30 (4,3 sec) in the movie.

Movie S3 and S4: Trajectories of a cell in a simulated experiment where the cell settles from 500 μ m above the top membrane in medium. Overview (Movie S3) and Zoom (Movie S4). a) Whole trajectory. The field is turned off at 180 s. b) The cell floats at a balanced position before the field is turned off. c) After the field is off, the cell continues to settle down and finally rests on the nano straws.