

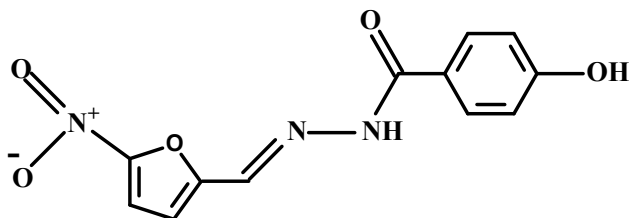
## Electrochemical stripping voltametrical sensor based on polypyrrole exfoliated polyetheramine-montmorillonite nanocomposite for nanomolar detection of nifuroxazide

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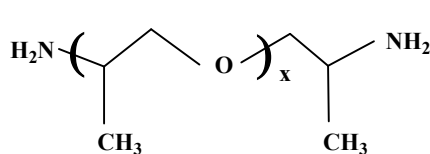
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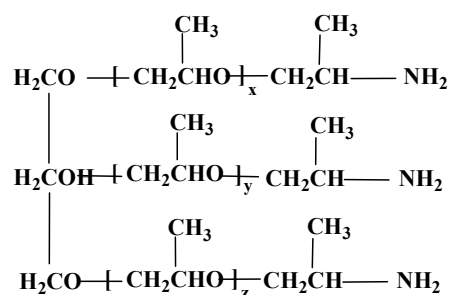
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**Scheme. S<sub>1</sub>.** Structure of nifuroxazide (NF).



$\underline{D}_{400}$ :  $x \approx 6.1$  &  $\underline{D}_{2000}$ :  $x \approx 33$



$\underline{T}_{403}$ :  $x+y+z = 5.3$  &  $\underline{T}_{5000}$ :  $x+y+z = 81$

**Scheme. S<sub>2</sub>.** Structure of Polyoxyalkylene amine  $\underline{D}_{400}$ ,  $\underline{D}_{2000}$ ,  $\underline{T}_{403}$  and  $\underline{T}_{5000}$ .

## 2. Experimental

### 2.1. Materials, equipment, electrochemical liquids, and fabrication of unmodified and modified CPSs

All reagents were in pure form and used as purchased. Pyrrole monomer (Py, reagent grade, 98%), and Iron (III) chloride ( $\text{FeCl}_3$ , reagent grade, 97%) were obtained from Sigma-Aldrich company. Na-Montmorillonite (Na-MMT) was acquired from southern clay products (colloid BP), Inc (Gonzales, Texas, USA) with (114.8 meq/100g) cation exchange capacity. The Na-Montmorillonite clay was dried in a vacuum oven at  $100^\circ\text{C}$  for 24 h, yielding the interlayer spacing ( $d_{001}$ ) was 9.6 Å. Four kinds of polyoxypropylene diamine and triamine (Jeffamine) were ordered from (Huntsman Corporation, TX, USA) including Jeffamine D<sub>400</sub>, D<sub>2000</sub>, T<sub>403</sub>, and T<sub>5000</sub> with an average molecular mass of 430, 2000, 440, 5000 and the primary amine contents (PACs) are 4.3, 0.97, 6.1, and 0.52 meq/ g, respectively. The commercial capsules of NF (*Antinal*® 200 mg, and Drotazide® 200 mg), and ultra-pure active powder were ordered from Amoun Pharmaceutical Co., El-Obour City, Egypt.

A Perkin Elmer spectrophotometer and a Phillips Powder-Diffractometer equipped with a Ni-filtered Cu- $K_\alpha$  ( $\lambda=1.5418$  Å) were used to obtain Fourier transform infrared (FT-IR) spectra and Wide-angle X-ray diffraction (WAXD) patterns of prepared samples, respectively. JEOL JSM 5400 Scanning electron microscopy and a JEOL JEM-100CX electron microscope were used to investigate the morphological structure of MMT and the prepared nanocomposites. **In addition, the ultra-thin films of nanocomposites were fabricated for transmission electron microscopy (TEM) analysis by cutting capsules of samples using an ultra-microtome, LKB 8800 ultra-microtome III equipped with a glass knife.** The conductivity of MMT and prepared nanocomposites were measured via two probe system of direct current circuit (DC circuit). PC-controlled potentiostat (PAR) Model 273 A and PAR-394 (Princeton Applied Research, Oak Ridge, TN, USA) with the software 270/250-PAR were utilized for the analytical voltammetry measurements.

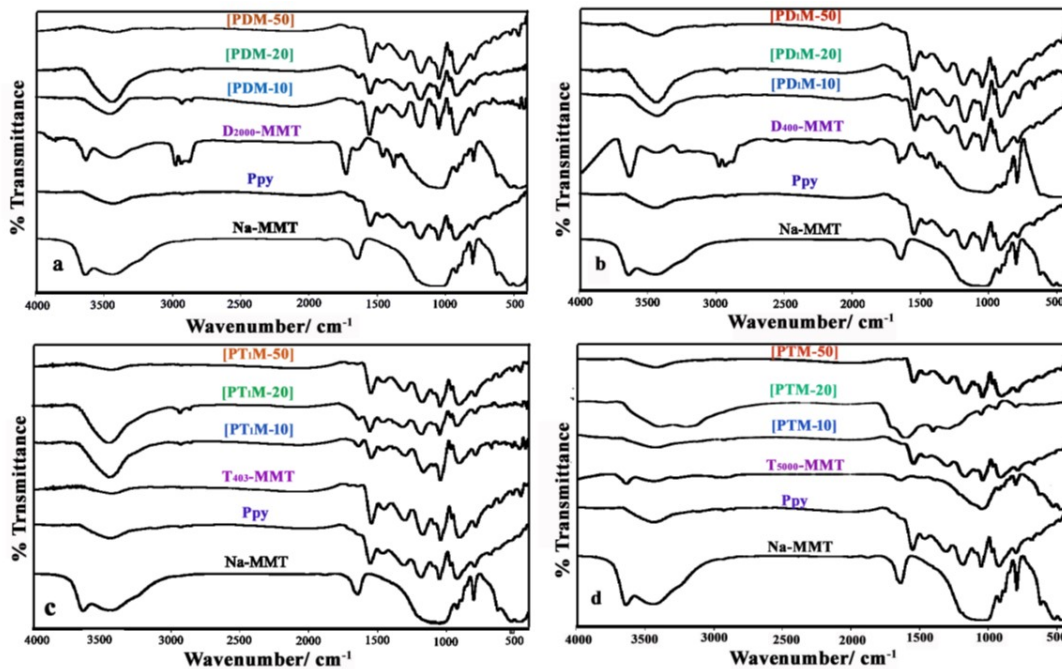
NF stock ( $10^{-3}\text{M}$ ) and diluted (1.0 to 1000 nM) solutions were prepared in methanol. Urine samples of 3 healthy volunteers were added to a micro-electrolysis cell containing a B-R buffer of pH 5.0 (9: 1, buffer: urine). Britton–Robinson (B–R) universal buffer series (pH

2-11) were prepared by mixing various proportions of boric, phosphoric, and acetic (0.04 M acids) with the NaOH (0.2 M).

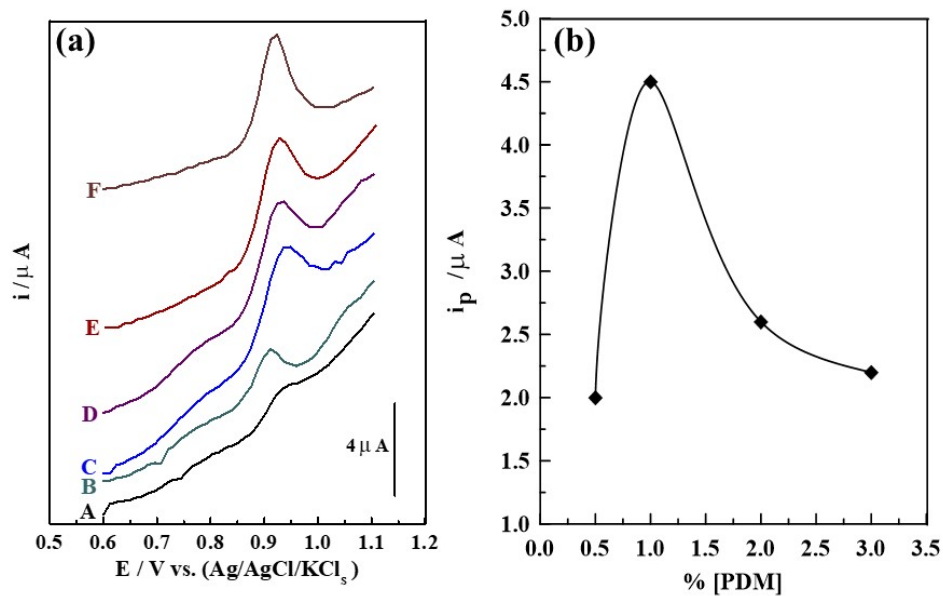
5.0 g of carbon powder (CP) was pasted with 1.8 mL of paraffin oil for 1 h to acquire bare carbon paste [B] CP. Whereas, 1.0 % [Ppy / (D<sub>2000</sub>, D<sub>400</sub>, T<sub>5000</sub>, and T<sub>403</sub>)-MMT-50] nanocomposites modulated CP sensors (MCPSs) were constructed by blending 4.95 g of graphite powder, and 0.05 g of modulation material with 1.8 mL paraffin oil to acquire a uniform MCP. The same method was carried out by changing the weight percent of the nanocomposite at the range of 0.5% - 3.0% of [PDM-50]. The [B] CP, and fabricated MCPs were inserted into the cavity of the sensor (Inner diameter (ID): 3mm), the superficial of the sensor was smoothed until glossy.

**Table. S<sub>1</sub>.** Composition data of Ppy/ polyetheramine - MMT nanocomposites.

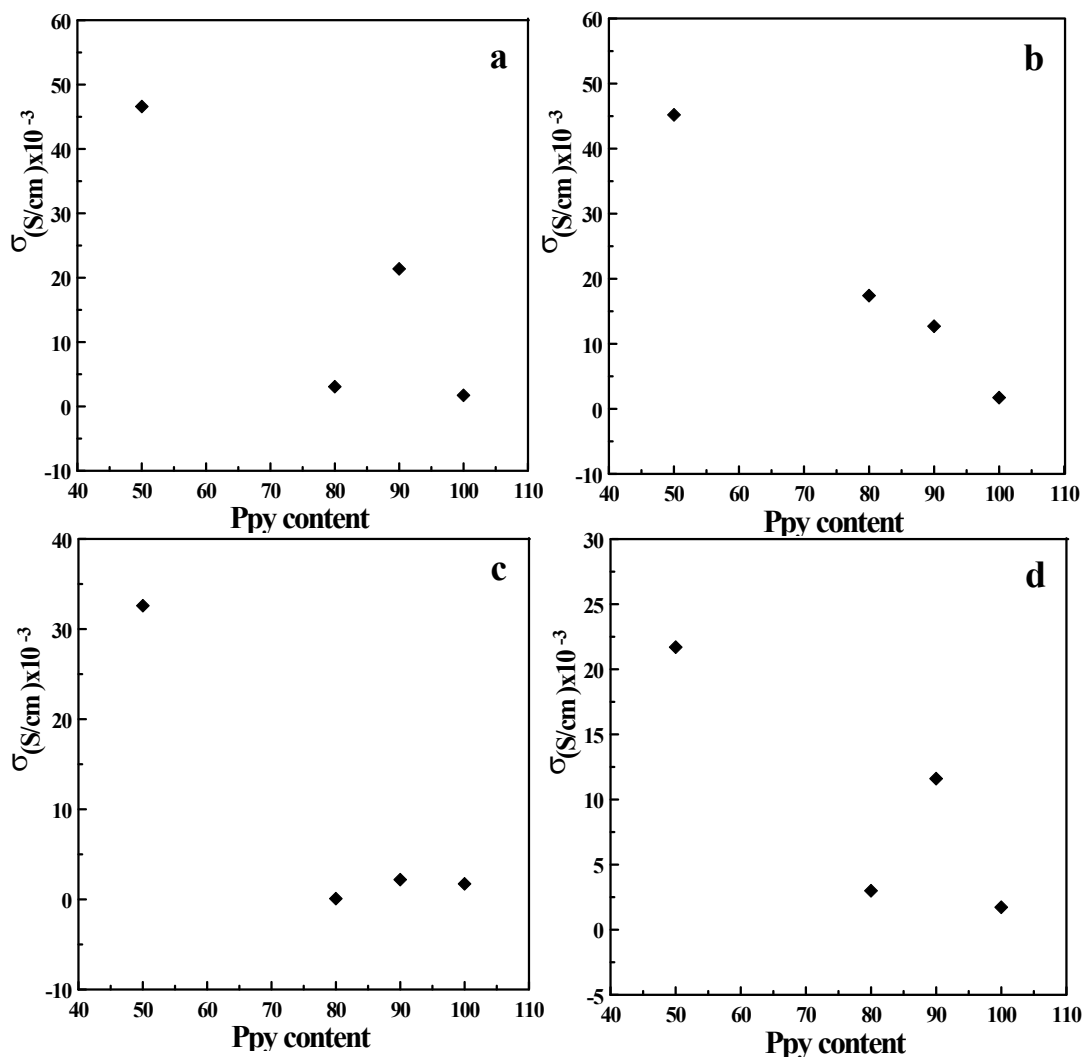
<i>Code</i>	<i>Polyetheramine-montmorillonite</i>			<i>Ppy/ polyetheramine - MMT</i>
	<i>Type</i>	<i>Wt / g</i>	<i>Wt %</i>	<i>Wt %</i>
[PDM-10]	D <sub>2000</sub> -MMT	0.1	10	93.23
[PDM-20]	D <sub>2000</sub> -MMT	0.2	20	90.22
[PDM-50]	D <sub>2000</sub> -MMT	0.5	50	66.22
[PD <sub>1</sub> M-10]	D <sub>400</sub> -MMT	0.1	10	88.90
[PD <sub>1</sub> M -20]	D <sub>400</sub> -MMT	0.2	20	85.84
[PD <sub>1</sub> M-50]	D <sub>400</sub> -MMT	0.5	50	83.08
[PTM-10]	T <sub>5000</sub> -MMT	0.1	10	90.20
[PTM-20]	T <sub>5000</sub> -MMT	0.2	20	95.94
[PTM-50]	T <sub>5000</sub> -MMT	0.5	50	76.97
[PT <sub>1</sub> M-10]	T <sub>403</sub> -MMT	0.1	10	92.74
[PT <sub>1</sub> M-20]	T <sub>403</sub> -MMT	0.2	20	86.37
[PT <sub>1</sub> M-50]	T <sub>403</sub> -MMT	0.5	50	70.29



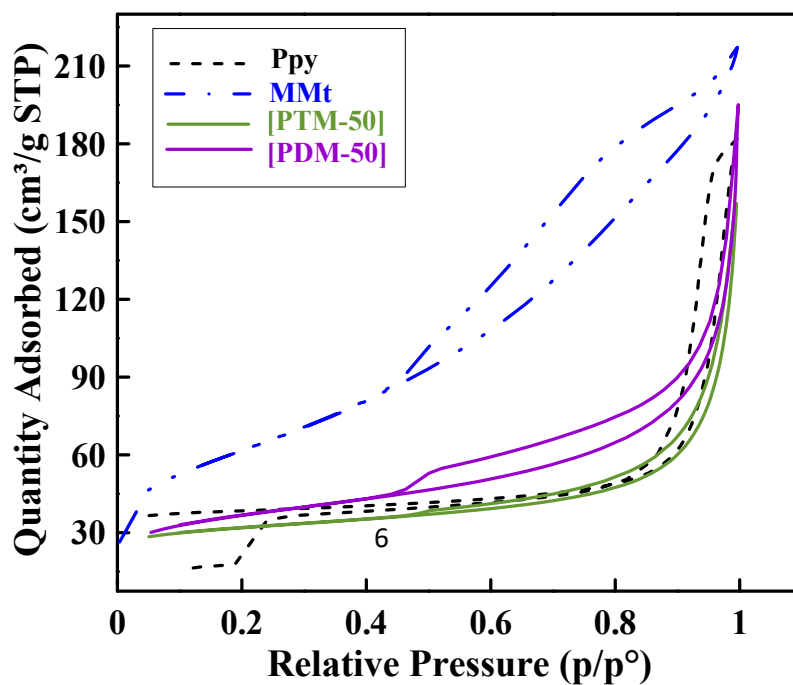
**Fig.S1.** FT-IR spectra of Na-MMT, Ppy, organoclay, and Ppy/ organoclay nanocomposites at different % of organoclay in the region 4000-400  $\text{cm}^{-1}$ .



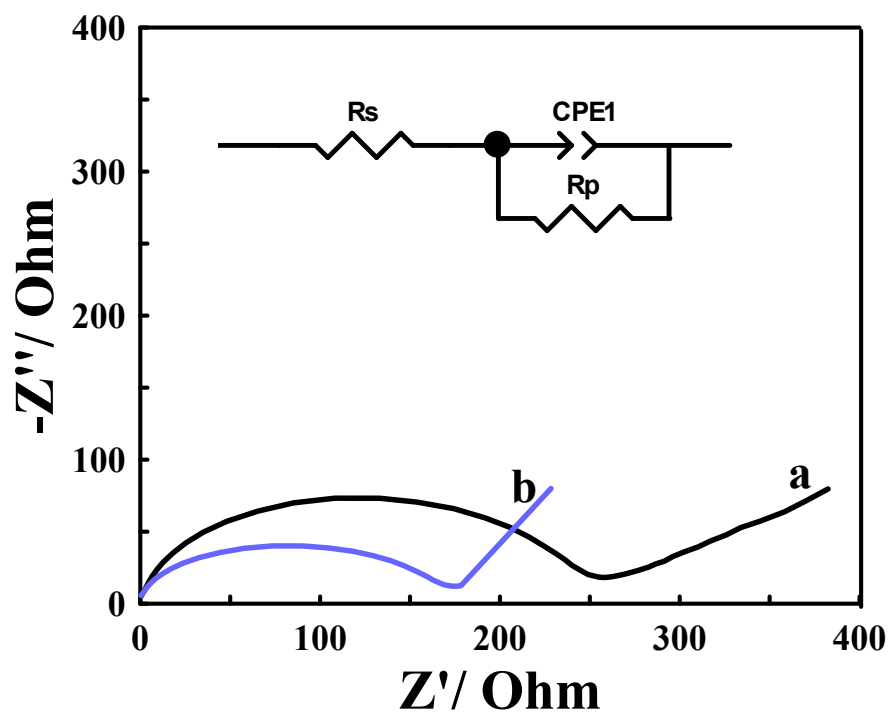
**Fig.S2. (a):** SW-AdAS voltammograms of 30.0 nM *NF* in pH 5 (B-R buffer) at  $E_{acc} = 0.60$  V for 10 s at A) [B] CPS, B) 7.0% [Na-MMT, C) 1.0 % [PT<sub>1</sub>M-50], D) 1.0 % [PTM-50], E) 1.0 % [PD<sub>1</sub>M-50], and F) 1.0 % [PDM-50] MCPSs ( $f = 100$  Hz,  $a = 25$  mV and  $\Delta E_s = 10$  mV). (b) Plot of 30.0 nM *NF* in (pH 5) at  $E_{acc} = 0.60$  V for 10 s of different % of [PDM-50].



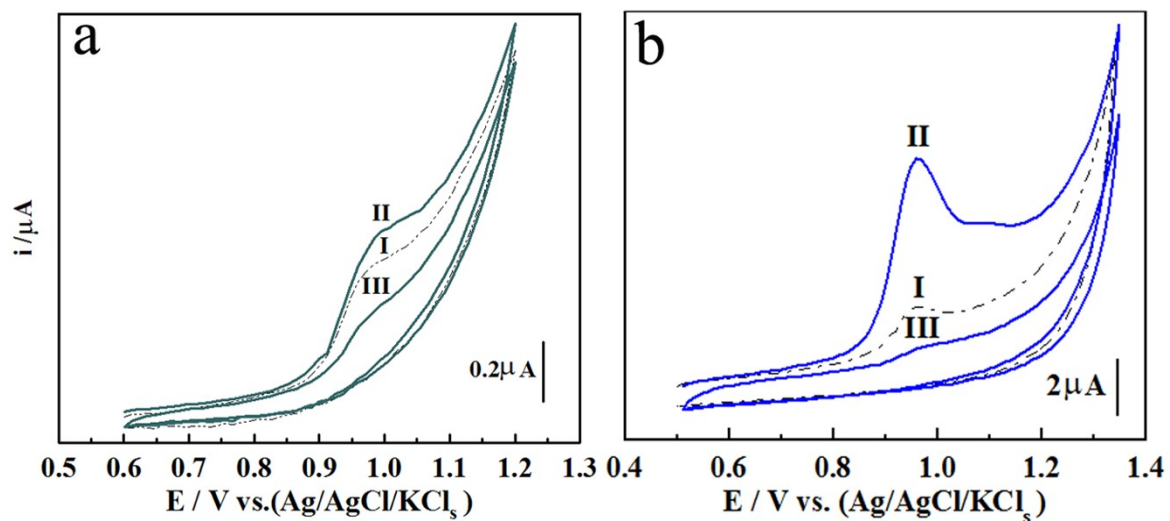
**Fig.S3.** Conductivity plots of (a) [PDM], (b) [PTM], (c) [PD<sub>1</sub>M], and (d) [PT<sub>1</sub>M].



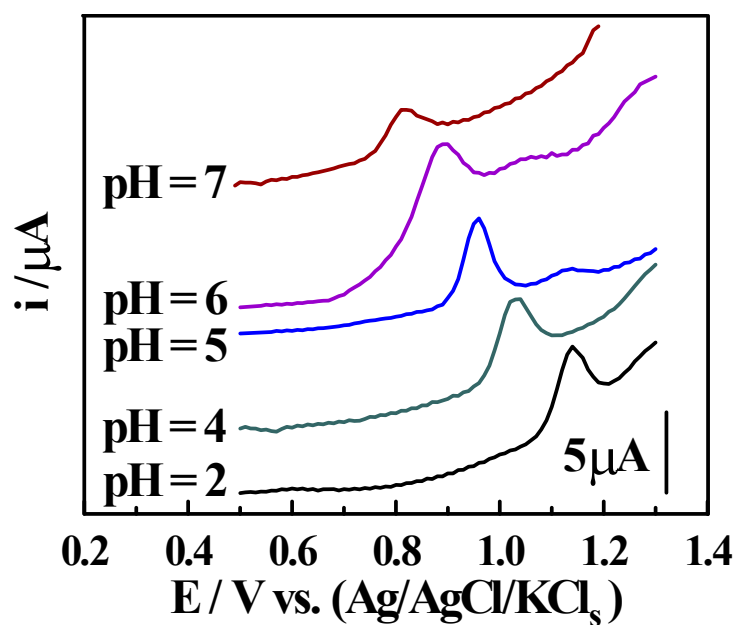
**Fig.S4.** N<sub>2</sub> adsorption-desorption isotherm of some of prepared materials



**Fig.S5.** EIS of (a) [B] CPS, and (b) 1.0% [PDM-50] MCPS.

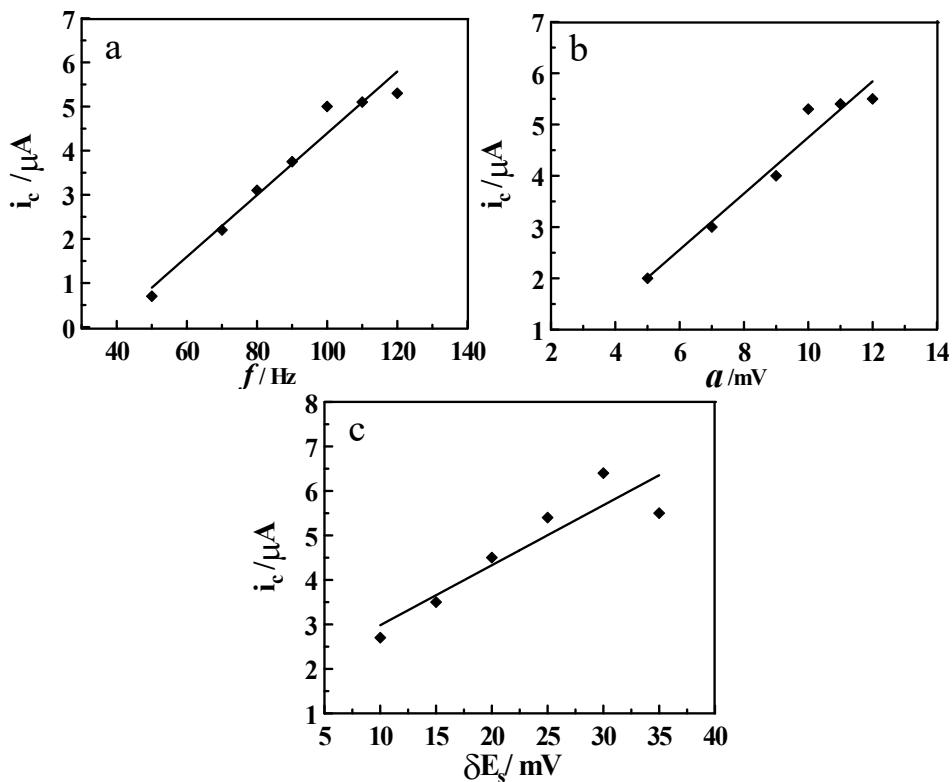


**Fig. S<sub>6</sub>.** CVs of 50.0 nM NF (pH 5) utilizing [B] CPS (a), and 1.0% [Ppy/ D<sub>2000</sub>-MMT 50] MCPS(b) at 0.0s (Cycle I) for 10.0 s (Cycles; 1<sup>st</sup> (II) and 2<sup>nd</sup> (III)) at  $v=100\text{ mV}\cdot\text{s}^{-1}$ .

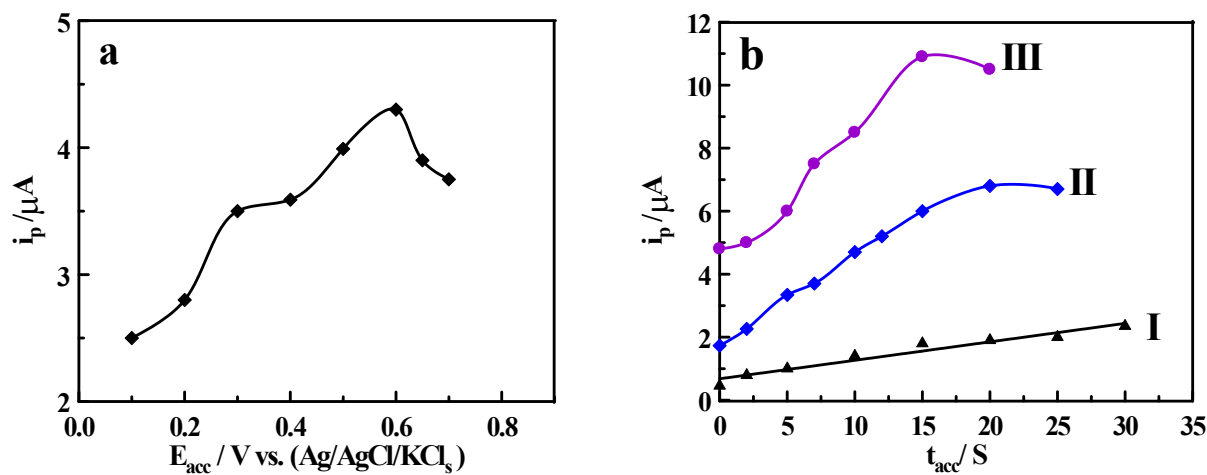


**Fig. S<sub>7</sub>.** The effect of using of 30.0 nM NF in series of the B-R buffer upon 1.0 % [PDM-50] MCPS ( $f = 120\text{ Hz}$ ,  $\Delta E_s = 12\text{ mV}$  and,  $a = 30\text{ mV}$ ).

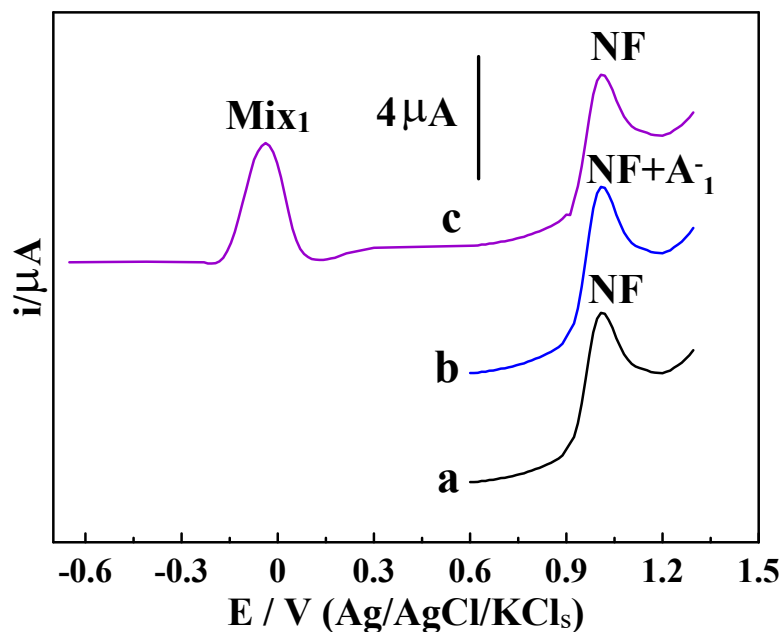




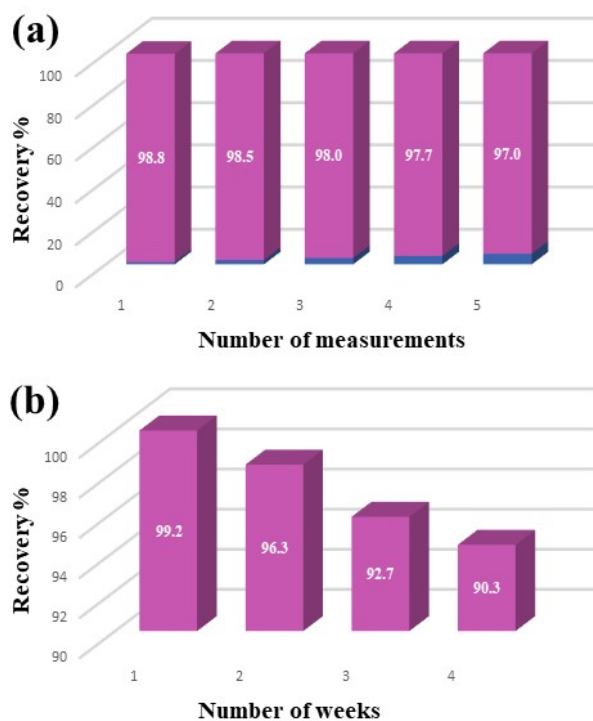
**Fig. S8.** Effect of pulse parameters of 30.0 nM NF in the B-R universal buffer pH 5 upon 1.0 % [PDM-50] MCPS at  $E_{\text{acc}} = 0.5$  V for 20 s.



**Fig. S9.** (a) The effect of  $E_{\text{acc}}$  on SW-AdsASVs peaks of 0.1  $\mu\text{M}$  NF in pH 5 upon 1.0 % [PDM-50] MCPS for 20s ( $\Delta E_s = 12$  mV,  $f = 120$  Hz, and  $a = 30$  mV), and (b) The effect of the  $t_{\text{acc}}$  of (I) 0.01, (II) 0.05, and (III) 0.1  $\mu\text{M}$  at  $E_{\text{acc}} = 0.6$  V.



**Fig. S<sub>10</sub>.** SW-AdAS voltammetry peaks of (a) 20.0 nM NF, (b) 20.0 nM NF mixed with 2.0  $\mu M$  ( $\sim 100$ -fold) of  $A_1^-$ , and (c) 20.0 nM NF mixed with 20.0 nM of  $Mix_1$  upon the surface of 1.0 % [PDM-50] MCPS.



**Fig. S<sub>11</sub>.** The reusability (a), and stability (b) histogram plots of the 1.0 % [PDM-50] MCPS.