Supporting Information

Synthesis, Molecular Docking, and Biological Evaluation of [3, 2-b] Indole Fused 18β-Glycyrrhetinic Acid Derivatives against Skin Melanoma

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Note: #Amit Kumar and Ragni Gupta are equal contributors to the study.

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1. Experimental section

- 1.1 Chemistry1.2 Biology
- NMR (1H, 13CNMR, 2D NMR) Spectra of [3, 2-b] Indole Fused 18β-Glycyrrhetinic Acid (GPD-1 to GPD-14)
- HRMS Spectra of of [3, 2-b] Indole Fused 18β-Glycyrrhetinic Acid (GPD-1 to GPD-14)
- 4. Molecular docking data
- 5. Raw images of western blot

1. Experimental section

1.1. Chemistry

All the required chemicals, reagents and solvents for modification were purchased from Sigma-Aldrich. Reactions were monitored through TLC on silica gel 60 F254 plates (E. Merck) by using ceric ammonium sulphate solution as spraying reagent for detection of spots. Purification of all synthesized derivatives was done through column chromatography using silica gel 60-120 mesh as stationary phase.

1.1.1. Isolation of 18β-Glycyrrhetinic acid (A01)

GA. was isolated in bulk quantity from MeOH extract of stem bark of *Glycyrrhiza glabra* and characterized by spectroscopic techniques.

GA-O. To a solution of compound **GA** (5 g, 11 mmol) in DCM was added PCC (3.54 g, 16 mmol) dissolved in DCM dropwise till dark colour appears and kept it at RT for 2 h. After completion, reaction mixture was passed through celite and filtrate was concentrated at rota vapour. Purification was done through column chromatography with EtoAc: Hexane (1: 13) as the eluent to afford product 7 colourless solid (3.5 g, 70% yield).

GPD-1¹H NMR (400 MHz, CDCl₃) δ 7.70 (s, 1H), 7.51 (d, J = 7.5 Hz, 1H), 7.29 (d, J = 7.8 Hz, 1H), 7.10 (dd, J = 11.0, 4.0 Hz, 1H), 7.06 (t, J = 6.9 Hz, 1H), 5.82 (s, 1H), 3.96 (d, J = 15.6 Hz, 1H), 2.68 (s, 1H),2.26 (d, J = 15.4 Hz, 2H), 2.13 – 2.05 (m, 2H),2.03 (d, J = 14.3 Hz, 2H), 1.95 (d, J = 16.4 Hz, 1H), 1.87 (dd, J = 21.4, 11.6 Hz, 1H), 1.78 (d, J = 9.2 Hz, 1H), 1.69 (d, J = 13.5 Hz, 2H), 1.62 (d, J = 15.0 Hz, 1H),1.53 (d, J = 12.4 Hz, 2H),1.46 (d, J = 12.4 Hz, 2H),1.43 (s, 3H), 1.32 (s, 3H), 1.25 (s, 6H), 1.23 (s, 3H), 1.19 (s, 3H), 1.07 (d, J = 13.5 Hz, 2H), 0.89 (s, 3H).¹³C NMR (101 MHz, CDCl₃) δ 199.35, 181.31, 169.12, 140.04, 136.22, 128.86, 128.30, 121.01, 118.93, 118.59, 110.16, 107.26, 60.55, 53.04, 48.21, 45.44, 43.82, 43.37, 41.02, 38.06, 37.77, 37.59, 34.05, 32.13, 31.97, 31.07, 30.97, 28.62, 28.45, 26.60, 26.52, 23.50, 23.34, 18.56, 18.37, 16.04.HRMS (ESI+): C₃₆H₄₈NO₃ m/z calcd for (M + H) +542.3634, found (M + H) +542.3634.

GPD-2

¹H NMR (400 MHz, CDCl₃) δ 7.42 (s, 1H), 7.35 – 7.28 (m, 1H), 7.23 – 7.16 (m, 1H), 7.16 – 7.11 (m, 1H), 6.90 (d, *J* = 7.9 Hz, 1H), 5.78 (s, 1H), 3.89 (d, *J* = 15.6 Hz, 1H), 2.67 (d, *J* = 7.4

Hz, 1H),2.23 (d, J= 15.8 Hz, 2H),2.03 (d, J = 14.2 Hz, 2H),1.95 (d, J = 18.1 Hz, 2H), 1.76 (d, J = 12.6 Hz, 2H), 1.68 – 1.59 (m, 4H), 1.53 (d, J = 14.3 Hz, 1H),1.42 (s, 3H), 1.39 – 1.36 (m, 2H), 1.31 (s, 3H), 1.26 (s, 3H), 1.24 (s, 3H), 1.21 (s, 3H), 1.16 (s, 3H), 1.07 (d, J = 9.3 Hz, 2H), 0.86 (s, 3H).¹³C NMR (101 MHz, CDCl₃) δ 200.37, 179.68, 170.19, 136.99, 128.54, 127.06, 122.93, 121.80, 119.52, 114.12, 113.16, 106.97, 60.47, 52.89, 49.75, 49.54, 49.32, 48.29, 45.42, 43.72, 43.38, 41.16, 37.99, 37.73, 37.40, 34.07, 31.89, 31.01, 30.81, 28.58, 28.43, 26.57, 23.25, 18.30, 15.95, 14.09. HRMS (ESI+): C₃₆H₄₆BrNO₃ m/z calcd for (M - H) 618.2587, found (M –H) 618.2583.

GPD-3 ¹H NMR (400 MHz, CDCl₃) δ 7.72 (s, 1H), 7.16 (ddd, J = 12.2, 9.2, 3.4 Hz, 2H), 6.84 (td, J = 9.2, 2.5 Hz, 1H), 5.82 (s, 1H), 3.88 (d, J = 15.5 Hz, 1H), 2.23 (d, J = 15.6 Hz, 2H), 2.04 (dd, J = 18.1, 9.2 Hz, 3H), 1.97 – 1.89 (m, 2H), 1.84 – 1.75 (m, 2H), 1.68 (d, J = 13.5 Hz, 2H), 1.63 (s, 1H), 1.53 (d, J = 9.8 Hz, 2H), 1.42 (s, 3H), 1.39 – 1.35 (m, 2H), 1.32 (s, 3H), 1.25 (d, J = 2.0 Hz, 6H), 1.22 (s, 3H), 1.18 (s, 3H), 1.08 – 1.02 (m, 2H), 0.88 (s, 3H).¹³C NMR (101 MHz, CDCl₃) δ 199.95, 181.83, 181.41, 169.39, 142.16, 132.63, 128.80, 110.60, 110.51, 109.05, 108.78, 107.60, 60.45, 52.92, 48.19, 45.42, 43.83, 43.36, 40.98, 38.01, 37.75, 37.50, 34.16, 32.05, 31.93, 31.05, 30.96, 28.62, 28.47, 26.58, 26.48, 23.53, 23.33, 18.53, 18.35, 16.02. HRMS (ESI+): C₃₆H₄₇FNO₃ m/z calcd for (M + H) +560.3540, found (M + H) +560.3547.

GPD-4 ¹H NMR (400 MHz, CDCl₃) δ 7.28 (d, *J* = 4.4 Hz, 2H), 7.21 (d, *J* = 8.7 Hz, 1H), 6.91 (d, *J* = 8.6 Hz, 1H), 5.73 (s, 1H), 3.84 (d, *J* = 15.5 Hz, 1H), 2.64 (s, 1H), 2.21 (d, *J* = 15.3 Hz, 2H), 2.03 (td, *J* = 13.4, 3.9 Hz, 1H), 1.98 – 1.84 (m, 3H), 1.75 (t, *J* = 12.8 Hz, 2H), 1.62 (d, *J* = 13.6 Hz, 3H), 1.54 – 1.44 (m, 2H), 1.39 (s, 3H), 1.37 – 1.32 (m, 1H), 1.29 (s, 3H), 1.25 (s, 1H), 1.22 (s, 3H), 1.18 (s, 3H), 1.16 (s, 3H), 1.14 (s, 3H), 1.05 (dd, *J* = 23.8, 11.6 Hz, 2H), 0.82 (s, 3H).¹³C NMR (101 MHz, CDCl₃) δ 200.66, 179.34, 170.69, 142.62, 142.30, 134.55, 128.34, 128.16, 122.09, 114.38, 110.66, 110.58, 107.02, 60.45, 52.92, 48.32, 45.42, 43.68, 43.38, 41.19, 37.95, 37.69, 37.40, 34.14, 31.97, 31.84, 30.98, 30.71, 28.52, 28.37, 26.53, 26.38, 23.17, 18.42, 18.24, 15.91. HRMS (ESI+):C₃₇H₄₆F₃NO₄m/z calcd for (M + H) +626.3464, found (M + H) +626.3457.

GPD-5 ¹H NMR (400 MHz, CDCl₃) δ 6.83 (dd, J = 20.7, 8.4 Hz, 1H), 6.57 (t, J = 10.3 Hz, 1H), 5.70 (d, J = 28.3 Hz, 1H), 3.79 (d, J = 15.5 Hz, 1H), 2.58 (s, 1H), 2.21 (t, J = 52.7 Hz, 4H), 1.93 (dd, J = 29.3, 11.7 Hz, 4H), 1.56 (dd, J = 25.5, 12.7 Hz, 5H), 1.35 (s, 3H), 1.31 (s, 3H), 1.26 (s, 4H), 1.18 (s, 9H), 1.14 (s, 3H), 1.09 (s, 3H), 0.78 (d, J = 9.5 Hz, 3H). ¹³C NMR (101 MHz, CDCl₃) δ 200.00, 176.27, 150.68, 128.34, 122.37, 114.10, 104.46, 103.27, 77.36,

77.05, 76.73, 60.38, 55.37, 52.87, 48.26, 48.21, 47.45, 45.29, 43.37, 41.08, 39.74, 36.69, 34.23, 31.94, 30.98, 29.72, 28.53, 26.39, 23.50, 23.32, 21.43, 18.52, 18.32, 16.00, 14.16, 13.65.

HRMS (ESI+): $C_{36}H_{45}F_2NO_3 m/z$ calcd for (M + H) +578.346, found (M + H) +578.3447.

GPD-6 ¹H NMR (400 MHz, CDCl₃) δ 7.37 (d, J = 12.4 Hz, 1H), 7.23 (d, J = 7.5 Hz, 1H), 6.94 (dd, J = 7.6, 4.7 Hz, 1H), 6.81 (d, J = 7.8 Hz, 1H), 5.76 (s, 1H), 3.90 (d, J = 15.3 Hz, 1H), 3.40 (d, J = 13.3 Hz, 1H), 2.69 (s, 1H), 2.26 (d, J = 14.9 Hz, 2H), 2.17 – 1.87 (m, 4H), 1.84 – 1.60 (m, 5H), 1.55 (d, J = 11.7 Hz, 2H), 1.44 (s, 6H), 1.36 (s, 3H), 1.28 (s, 3H), 1.23 (s, 3H), 1.20 (s, 3H), 1.18 (s, 2H), 1.13 – 1.02 (m, 2H), 0.87 (s, 3H).¹³C NMR (101 MHz, CDCl₃) δ 199.90, 180.86, 169.31, 148.07, 140.97, 128.82, 123.98, 119.15, 119.09, 114.36, 108.15, 106.20, 60.48, 52.98, 48.21, 45.42, 43.80, 43.37, 41.03, 38.05, 37.76, 37.68, 34.14, 32.07, 31.93, 31.04, 30.98, 29.26, 28.61, 28.43, 26.59, 26.49, 23.49, 23.33, 18.53, 18.35, 16.01. HRMS (ESI+): C₃₆H₄₆FNO₃ m/z calcd for 559.3462 found (M - H) 558.3383.

GPD-7 ¹H NMR (400 MHz, CDCl₃) δ 7.38 (d, J = 8.4 Hz, 1H), 7.27 (d, J = 1.8 Hz, 1H), 7.04 – 6.94 (m, 1H), 5.78 (s, 1H), 3.90 (d, J = 15.6 Hz, 1H), 2.66 (s, 2H), 2.24 (dd, J = 14.5, 6.7 Hz, 4H), 2.08 – 1.99 (m, 2H), 1.99 – 1.91 (m, 2H), 1.87 (dd, J = 13.5, 4.2 Hz, 1H), 1.80 – 1.74 (m, 1H), 1.66 (d, J = 13.4 Hz, 2H), 1.63 – 1.53 (m, 3H), 1.49 (d, J = 17.1 Hz, 1H), 1.42 (s, 3H), 1.31 (s, 3H), 1.25 (s, 3H), 1.24 (s, 2H), 1.21 (d, J = 1.8 Hz, 6H), 1.16 (s, 3H), 0.86 (s, 3H).¹³C NMR (101 MHz, CDCl₃) δ 200.32, 179.66, 170.09, 141.00, 136.62, 128.58, 126.57, 119.34, 119.18, 110.21, 107.09, 107.04, 60.46, 52.87, 48.28, 45.42, 43.74, 43.38, 41.15, 37.99, 37.75, 37.41, 34.09, 32.03, 31.91, 31.02, 30.89, 29.71, 28.60, 28.46, 26.58, 26.45, 23.27, 18.48, 18.32, 16.00. HRMS (ESI+):C₃₆H₄₇F₃ClNO₃ m/z calcd for (M + H) +576.3244, found (M + H) +576.3246.

GPD-8¹H NMR (400 MHz, CDCl₃) δ 7.57 (s, 1H), 7.37 (d, J = 7.7 Hz, 1H), 7.04 – 6.90 (m, 3H), 5.81 (s, 1H), 3.94 (d, J = 15.6 Hz, 1H), 3.38 (s, 1H), 2.67 (s, 2H), 2.48 (s, 3H), 2.24 (d, J = 3.6 Hz, 2H), 2.18 (s, 1H), 2.10 (d, J = 6.8 Hz, 1H), 2.05 (s, 2H), 1.96 (s, 2H), 1.88 (d, J = 13.2 Hz, 3H), 1.77 (d, J = 11.7 Hz, 2H), 1.66 (s, 2H), 1.52 (d, J = 5.8 Hz, 2H), 1.43 (s, 7H), 1.34 (s, 4H), 1.27 (s, 4H), 1.24 (s, 4H), 1.22 (s, 5H), 1.19 (s, 5H), 0.88 (s, 3H). ¹³C NMR (101 MHz, CDCl₃) δ 200.02, 181.04, 169.24, 149.01, 148.91, 146.42, 135.61, 128.85, 127.78, 119.26, 119.17, 116.36, 60.54, 53.07, 48.17, 45.43, 43.80, 43.36, 38.03, 37.72, 34.09,

32.11, 31.93, 31.11, 30.98, 28.62, 28.47, 26.58, 26.49, 23.55, 23.35, 23.23, 20.55, 18.56, 18.36, 16.78, 16.04. HRMS (ESI+):C₃₆H₄₇F₃ClNO₃ m/z calcd for (M + H) +556.3791, found (M + H) +556.3785.

GPD-9¹H NMR (400 MHz, MeOD) δ 8.38 (s, 1H), 8.20 (d, J = 8.9 Hz, 1H), 7.98 (d, J = 8.9 Hz, 1H), 7.80 (d, J = 8.6 Hz, 1H), 7.34 (d, J = 9.0 Hz, 1H), 3.94 (d, J = 15.7 Hz, 1H), 3.34 (d, J = 1.5 Hz, 1H), 2.94 – 2.85 (m, 1H), 2.75 (s, 1H), 2.62 (dd, J = 15.9, 9.6 Hz, 1H), 2.55 (s, 1H), 2.42 (dd, J = 13.2, 8.4 Hz, 1H), 2.26 (s, 1H), 2.20 (s, 1H), 2.10 (t, J = 12.6 Hz, 1H), 1.91 (d, J = 13.2 Hz, 2H), 1.68 (d, J = 14.0 Hz, 1H), 1.61 (d, J = 13.0 Hz, 2H), 1.48 (s, 2H), 1.43 (s, 4H), 1.26 (s, 4H), 1.19 (s, 5H), 1.12 (s, 3H), 1.09 (s, 3H), 0.86 (s, 3H).¹³C NMR (101 MHz, MeOD) δ 200.01, 178.81, 171.40, 144.15, 139.76, 139.65, 127.21, 126.87, 124.11, 60.44, 59.85, 54.43, 52.40, 44.88, 44.72, 43.10, 42.86, 40.58, 39.04, 37.34, 37.10, 36.03, 33.66, 33.44, 31.29, 31.18, 30.29, 27.75, 27.44, 25.85, 25.62, 22.42, 20.33, 18.11, 17.58, 14.92. HRMS (ESI+):C₃₆H₄₆N₂O₅ m/z calcd for (M + H) +587.3485, found (M + H) +587.3484

GPD-10¹H NMR (400 MHz, CDCl₃:MEOD) δ 7.53 (s, 1H), 7.30 (s, 1H), 7.05 (s, 1H), 5.76 (s, 1H), 3.82 (d, *J* = 15.3 Hz, 1H), 3.35 (s, 2H), 2.70 (s, 1H), 2.25 (t, *J* = 15.3 Hz, 3H), 2.11 (t, *J* = 11.8 Hz, 3H), 2.03 – 1.91 (m, 3H), 1.77 (dd, *J* = 27.9, 15.3 Hz, 3H), 1.63 (dd, *J* = 25.5, 12.3 Hz, 2H), 1.46 (s, 3H), 1.42 (s, 3H), 1.39 (s, 1H), 1.31 (s, 3H), 1.27 (s, 3H), 1.23 (s, 3H), 1.21 (s, 3H), 1.16 (s, 3H), 1.08 (d, *J* = 14.2 Hz, 1H), 0.88 (s, 3H). ¹³C NMR (101 MHz, CDCl₃: MEOD) δ 204.94, 183.26, 175.42, 147.57, 136.07, 135.04, 134.13, 132.08, 127.67, 123.76, 120.05, 111.17, 64.39, 57.10, 49.37, 47.61, 47.39, 45.17, 41.82, 41.63, 41.40, 38.27, 35.86, 35.74, 34.87, 34.06, 33.47, 32.34, 32.10, 30.43, 30.25, 27.01, 26.46, 22.32, 22.04, 19.62.

HRMS (ESI+):C₃₆H₄₅Cl₂NO₃m/z calcd for (M - H)⁺ 608.2698, found (M - H) ⁺ -608.2701

GPD-11¹H NMR (400 MHz, CDCl₃) δ 7.86 (s, 1H), 7.40 (d, J = 7.7 Hz, 1H), 7.11 (dd, J = 7.6, 0.8 Hz, 1H), 7.00 (t, J = 7.7 Hz, 1H), 5.82 (s, 1H), 3.94 (d, J = 15.6 Hz, 1H), 2.67 (s, 1H), 2.26 (d, J = 15.7 Hz, 2H), 2.14 – 1.98 (m, 2H), 1.98 – 1.85 (m, 2H), 1.84 – 1.72 (m, 2H), 1.69 (d, J = 13.5 Hz, 3H), 1.62 (d, J = 12.9 Hz, 2H), 1.54 (d, J = 12.6 Hz, 2H), 1.46 (d, J = 9.1 Hz, 2H), 1.43 (s, 3H), 1.42 – 1.37 (m, 1H), 1.35 (s, 3H), 1.29 (s, 3H), 1.25 (s, 3H), 1.23 (s, 3H), 1.18 (s, 3H), 1.07 (d, J = 13.4 Hz, 1H), 0.88 (s, 3H). ¹³C NMR (101 MHz, CDCl₃) δ

200.03, 181.50, 169.50, 141.00, 133.29, 129.79, 128.78, 120.44, 119.76, 117.21, 115.74, 108.46, 60.46, 52.95, 48.20, 45.43, 43.84, 43.37, 40.98, 38.00, 37.76, 37.63, 34.15, 32.04, 31.93, 31.03, 30.94, 28.63, 28.49, 26.58, 26.46, 23.48, 23.35, 18.52, 18.34, 16.02. HRMS (ESI+): $C_{36}H_{46}CINO_{3}m/z$ calcd for (M + H) +576.3244, found (M + H) +576.7248.

GPD-12¹H NMR (400 MHz, MEOD: CDCl₃) δ 7.81 (d, J = 2.8 Hz, 1H), 7.39 (ddd, J = 23.4, 15.3, 8.6 Hz, 3H), 5.79 (s, 1H), 3.91 (d, J = 15.7 Hz, 1H), 3.37 (s, 1H), 2.73 (s, 1H), 2.28 (d, J = 14.0 Hz, 2H), 2.22 (s, 3H), 2.21 (d, J = 0.6 Hz, 2H), 2.20 (s, 1H), 2.09 (d, J = 19.5 Hz, 1H), 1.97 (d, J = 14.0 Hz, 3H), 1.66 (dd, J = 27.9, 22.1 Hz, 4H), 1.46 (s, 3H), 1.43 (s, 1H), 1.36 (s, 3H), 1.29 (d, J = 7.6 Hz, 6H), 1.23 (s, 2H), 1.18 (s, 3H), 1.13 (d, J = 5.0 Hz, 1H), 1.10 (d, J = 5.2 Hz, 2H), 0.89 (d, J = 5.1 Hz, 3H). ¹³C NMR (101 MHz, MEOD: CDCl₃) δ 204.79, 183.30, 147.44, 132.13, 131.79, 127.43, 127.36, 125.49, 123.42, 115.13, 110.93, 104.02, 64.34, 56.86, 49.38, 47.62, 47.37, 45.14, 41.85, 41.61, 41.14, 38.08, 35.86, 35.77, 34.86, 34.59, 34.27, 33.53, 32.41, 32.21, 30.26, 27.08, 26.69, 22.33, 22.11, 19.73, 17.83. C₃₇H₄₇N₂O₃m/z calcd for (M + H) +567.3587, found (M + H) +567.3584.

GPD-13¹H NMR (400 MHz, CDCl₃) δ 7.58 (s, 1H), 7.33 (d, J = 3.4 Hz, 1H), 7.16 (d, J = 1.7 Hz, 2H), 5.76 (s, 1H), 3.85 (d, J = 15.5 Hz, 1H), 3.41 – 3.33 (m, 1H), 2.67 (s, 1H), 2.23 (t, J = 11.8 Hz, 2H), 2.12 – 1.82 (m, 5H), 1.80 – 1.49 (m, 6H), 1.43 (s, 3H), 1.40 – 1.36 (m, 1H), 1.32 (s, 3H), 1.29 (s, 1H), 1.25 (s, 3H), 1.21 (d, J = 5.2 Hz, 6H), 1.15 (s, 3H), 1.06 (d, J = 13.8 Hz, 1H), 0.86 (s, 3H). ¹³C NMR (101 MHz, CDCl₃) δ 200.03, 181.50, 169.50, 141.00, 133.29, 129.79, 128.78, 120.44, 119.76, 117.21, 115.74, 108.46, 60.46, 52.95, 48.20, 45.43, 43.84, 43.37, 40.98, 38.00, 37.76, 37.63, 34.15, 32.04, 31.93, 31.03, 30.94, 28.63, 28.49, 26.58, 26.46, 23.48, 23.35, 18.52, 18.34, 16.02. C₃₆H₄₇BrNO₃ m/z calcd for (M+ H) +620.2739, found (M+ H) +620.2733

GPD-14 ¹H NMR (400 MHz, CDCl₃) δ 6.88 – 6.82 (m, 1H), 6.64 (d, *J* = 8.3 Hz, 1H), 6.56 (d, *J* = 7.9 Hz, 1H), 5.31 (s, 1H), 3.73 (d, *J* = 5.3 Hz, 3H), 3.69 (d, *J* = 8.7 Hz, 2H), 3.63 (s, 2H), 2.61 (d, *J* = 15.6 Hz, 2H), 2.34 (d, *J* = 7.7 Hz, 2H), 2.28 – 2.02 (m, 2H), 1.95 (t, *J* = 8.1 Hz, 2H), 1.84 – 1.61 (m, 2H), 1.53 (d, *J* = 10.9 Hz, 2H), 1.42 (d, *J* = 11.6 Hz, 3H), 1.32 (s, 3H), 1.27 (d, *J* = 7.3 Hz, 3H), 1.18 (d, *J* = 7.6 Hz, 6H), 1.13 (d, *J* = 8.3 Hz, 6H), 0.94 (d, *J* = 10.6 Hz, 2H), 0.77 (d, *J* = 15.0 Hz, 3H). ¹³C NMR (101 MHz, CDCl₃) δ 199.27, 179.93, 152.00, 138.29, 125.14, 118.94, 115.84, 114.08, 113.73, 113.72, 113.50, 113.49, 59.44,

54.65, 51.98, 47.18, 44.40, 42.80, 42.33, 40.11, 37.01, 36.78, 33.11, 30.85, 29.95, 28.66, 27.57, 27.45, 25.42, 22.43, 22.28, 21.66, 20.02, 17.29, 15.03, 13.15. C₃₇H₄₉NO₄ m/z calcd for (M+ H) +572.3740, found (M+ H) +572.3743.

1.2. Biology

1.2.1. Chemicals and reagents

Cell culture media DMEM (Dulbecco's Modified Eagle Medium), Dulbecco's phosphate buffer saline (DPBS). Trypsin-EDTA (Ethylenediaminetetraacetic acid). 3-(4,5dimetylthiazolyl)- diphenyl tetrazolium bromide (MTT), DMSO (Dimethyl sulphoxide), Penicillin G, Streptomycin, Sodium bicarbonate Sodium pyruvate, RIPA lysis buffer, 1% protease and phosphatase inhibitor cocktail, Bradford reagent and Beta Actin antibody used as loading control were purchased from Sigma Aldrich. FBS (Fetal Bovine Serum) was purchased from GIBCO USA. Dichlorofluorescein diacetate (H2DCF-DA) was procured from Thermofisher. DAPI (4', 6-diamidino-2-phenylindole hydrochloride) was supplied by Invitrogen, Thermo Fischer Scientific. Antibodies against Bcl2, BAX and Caspase 9 were purchased from Cell Signalling Technology.

1.2.2. Cell culture

Human melanoma cell line, A375 and Murine melanoma cell line, B16F10 were purchased from American Type Culture Collection (Rockville, MD, USA). Primary Human Dermal Fibroblasts, HDF were purchased from HiMedia, India. Cell lines were cultured in DMEM media supplemented with 10%FBS, Penicillin G (120mg/L), Streptomycin (270mg/L), Sodium bicarbonate (1.2g/L) and Sodium pyruvate (220mg/L) and maintained in a humidified chamber (37°C, 5% CO₂).

1.2.3. Cell survival assay

MTT assay was performed for cell survival analysis ¹⁶. Briefly, A375 cells (5×10^3), B16F10 cells (5×10^3) and HDF cells (10^4) were seeded in 96 well plates and incubated for 24 hours. Cells were treated with different concentrations of GPD-12 and were incubated for 24 hours. Cells were again incubated with MTT solution (100μ L/well) at a concentration of 250µg/ml PBS for 3 hours. Formazan crystals formed were dissolved in DMSO (100μ L/well). The

absorbance was measured at 570nm using plate reader, Multiskan spectrum; Thermo Electron Corporations, USA.

1.2.4. Colony formation assay

A375 cells were seeded at a density of 10³ cells per well of 6 well plate and kept overnight. The media was replaced with fresh media containing different concentrations of GPD-12. After 24 hours the media containing GPD-12 was removed and cells were further cultured for 6 days in media without any effectors. On day 6, cells were washed with DPBS, fixed with methanol and stained with crystal violet dye. Cells were washed with distilled water and air dried for some time. Stained colonies were captured by EVOS- FL Cell Imaging System (Thermo Fisher).

1.2.5. DAPI Staining

A375 cells were seeded in 6 well plate (10^4 cells per well). After 24 hours, cells were subjected to treatment with different concentrations of GPD-12. Treatment was terminated after 24 hours. Cells were fixed with 4% Para formaldehyde, permeabilized with 0.1% Triton X-100 and stained with DAPI (5μ g/ml). Imaging was done by EVOS- FL Cell Imaging System (thermo fisher).

1.2.6. Reactive Oxygen Species measurement using Fluorescence microscopy

A375 cells were seeded in 35mm dishes. After 24 hours, cells were subjected to treatment with different concentrations of GPD-12 and incubated for 6 hours. Cells were washed with DPBS and stained with Dichlorodihydrofluorescein diacetate (H2DCF-DA) dye (5μ g/ml) for 30 minutes. Cells were again washed with DPBS and imaging was done under a thin layer of DPBS (EVOS- FL Cell Imaging System, thermo fisher).

1.2.7. Cell lysate preparation and western blotting

After respective treatments, cells were harvested and lysis was performed using RIPA lysis buffer containing 1% protease and phosphatase inhibitor cocktail. Protein estimation was done by Bradford Protein Estimation Method using Bovine Serum Albumin as standard. Protein samples were denatured at 100°C for 3 min in 5x Laemmli buffer and equal amounts of protein samples (40µg) were separated by SDS-PAGE (10-12%) using Miniprotein Tetra

System (Bio-Rad). Proteins were transferred onto PVDF membranes and membranes were blocked using 5% skimmed milk. Membranes were incubated overnight at 4°C with primary antibodies. Membranes were washed with Tris Buffered Saline containing Tween 20 and reincubated with secondary antibody for 2 hours at room temperature. Membranes were developed using immobilon Western Chemiluminescent HRP substrate. Blots were quantified using Image labTM software (BioRad).

1.2.8. Statistical analysis

Data is expressed as mean \pm Standard Deviation (SD) from three independent experiments. Results were considered significant at *p<0.05, **p<0.01. Data was analyzed by using GraphPad Prism software 9.



2. NMR (1H, 13CNMR, 2D NMR) Spectra of [3, 2-b] Indole Fused 18β-Glycyrrhetinic Acid (GPD-1 to GPD-14)



























 HRMS Spectra of of [3, 2-b] Indole Fused 18β-Glycyrrhetinic Acid (GPD-1 to GPD-14)











Monoisotopic Mass, Even Electron Ions 22 formula(e) evaluated with 1 results within limits (up to 3 closest results for each mass) Elements Used: C: 0-36 H: 0-100 N: 0-2 O: 0-5 GPD-9 QMI DIVISION, CSIR-IIIM JAMMU Xevo G2-XS OTOE YEC2015

050123_05	8 (0.172)					V¢.	VU UZ-NO	alor inc	2010				1: T(OF MS ES+ 1.67e+007
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Minimum: Maximum:			2.0	50.	0 51	1.5								
Mass 587.3484	Calc. 587.34	Mass 85	mDa -0.1	PPM -0.1	2 14	BE 4.5	i-FIT 812.0	Norm n/a	Conf(% n/a) Formu C36 H	ila 147 N2 05			

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Elemental Composition Report											
Single Mass Analysis Tolerance = 4.0 PPM / DBE: min = -1.5, max = 50.0 Element prediction: Off Number of isotope peaks used for i-FIT = 3											
Monoisotopic Mass, Even Electron Ions 167 formula(e) evaluated with 1 results within limits (up to 3 best isotopic matches for each mass) Elements Used: C: 0-36 H: 0-100 N: 0-1 O: 0-3 CI: 0-2 Br: 0-1											
GPD-11 QMI DIVISION, CSIR-IIIM JAMMU Xevo G2-XS QTOF YFC2015 300721_28 14 (0.293) Cm (14:15) Cm (14:15)											
576.3248											
%- 578.3237 579.3260											
141.9591 182.9868 301.1324 354.2844 438.4008 666.3003 850.7987 906.0811981.2759 100 200 300 400 500 600 700 800 900 1000	1153.6467 m/z 1100 1200										
Minimum: -1.5 Maximum: 2.0 4.0 50.0											
Mass Calc. Mass mDa PPM DBE i-FIT Norm Conf(%) Formula 576.3248 576.3244 0.4 0.7 13.5 28.4 n/a n/a C36 H47 N 03 Cl											





4. Molecular docking data

2D images of compound



5. Raw images of Western blot



Bax on SDS PAGE gel, Molecular Weight-23KDa

The molecular marker used is Precision Plus Protein[™] Kaleidoscope[™] Prestained Protein Standards by Bio-Rad Catalog # 161 0375



Bcl2 on SDS PAGE gel, Molecular Weight-26KDa

The molecular marker used is Precision Plus ProteinTM KaleidoscopeTM Prestained Protein Standards by Bio-Rad Catalog # 161 0375



 β -Actin (Bax and Bcl-2) on SDS PAGE gel, Molecular Weight-42KDa

The molecular marker used is Precision Plus ProteinTM KaleidoscopeTM Prestained Protein Standards by Bio-Rad Catalog # 161 0375



Caspase 3 on SDS PAGE gel, Molecular Weight-32KDa

The molecular marker used is Precision Plus ProteinTM KaleidoscopeTM Prestained Protein Standards by Bio-Rad Catalog # 161 0375



Caspase 9 on SDS PAGE gel, Molecular Weight-46KDa

The molecular marker used is Precision Plus ProteinTM KaleidoscopeTM Prestained Protein Standards by Bio-Rad Catalog # 161 0375



β-Actin (Caspase-3 and Caspase-9) on SDS PAGE gel, Molecular Weight-42KDa

> The molecular marker used is Precision Plus Protein™ Kaleidoscope™ Prestained Protein Standards by Bio-Rad Catalog # 161 0375