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Stereoselective Peptide Catalysis in Complex Environments – From River Water to Cell Lysates

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1. General Aspects and Materials

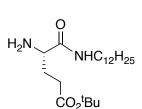
Reagents and materials were of the highest commercially available grade and used without further purification. Reactions were monitored by thin layer chromatography using Merck silica gel 60 F254 aluminium sheets. Visualization of the compounds was achieved by UV-Vis or KMnO₄. Flash chromatography and plug filtrations were performed using silica gel 60 (particle size 0.040 – 0.063 mm, 200 – 400 mesh) manufactured by Fluka. ¹H and ¹³C NMR spectra were recorded on a Bruker DRX 400, a Bruker AV III 400 (400 MHz/100 MHz) or a Bruker AV III 600 (600 MHz/150 MHz). All spectra were recorded at 25 °C, unless stated otherwise. Chemical shifts (δ) are reported in parts per million (ppm) relative to the signal of tetramethylsilane (TMS) using the residual solvent signals. SFC analyses were performed on an analytical SFC with a diode array detector ACQUITY-UPLC-PDA from Waters using chiral stationary phase columns (Trefoil, AS, AD, IA, Whelk, IC, OD, OJ) (150 mm x 30 mm) from Daicel or Waters under the reported conditions. HPLC analyses were performed on an analytical Ultimate 3000 HPLC system from Dionex with a diode array detector and chiral stationary phase columns (Daicel AD-H, Daicel AS-H, AY-H, OD-H or Daicel OJ-H). Highresolution electron ionization (HR-EI) mass spectra were measured on a Waters Micromass AutoSpec Ultima spectrometer. High-resolution MALDI spectra were acquired on a Bruker solariX 94 (ESI/MALDI-FT-ICR) and a Bruker Ultra-Flex II (MALDI-TOF) spectrometer.

2. Synthesis and Analytical Data of the Peptides

Peptide Synthesis

Peptides 1 and 2 were prepared by solution peptide synthesis, peptide 1a and 2b by solid phase peptide synthesis. The syntheses of 1^1 and $1a^2$ have been previously described.

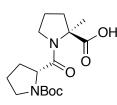
Procedure for the synthesis of H-Glu(O'Bu)-NHC₁₂H₂₅: Z-Glu(O'Bu)-OH(5.0 g, 14.8 mmol,



1 equiv.), dodecylamine (2.75 g, 14.82 mmol, 1.0 equiv.) and EDC•HCl (3.4 g, 17.8 mmol, 1.2 equiv.) were suspended in EtOAc (75 mL) and stirred at room temperature overnight. The mixture was diluted with EtOAc (350 mL) and washed with 0.1 M HCl (3x 100 mL), aq. 5% w/w Na₂CO₃ (2x 100 mL) and brine (150 mL). The organic layer was dried over MgSO₄ and the solvent was removed under reduced pressure. The

resulting colorless solid was purified by flash column chromatography on silica gel eluting with 1-5% MeOH in CH₂Cl₂. The colorless solid was then dissolved in MeOH (20 mL). Pd/C (10 % w/w) was added and the mixture was stirred under a hydrogen atmosphere (balloon) at room temperature overnight. The reaction mixture was filtered over a pad of celite, which was washed with MeOH (3x 10 mL). Removal of all volatiles yielded the desired product as colorless solid (3.95 g, 72%). ¹H NMR (400 MHz, CDCl₃) δ = 7.17 (s, *J* = 3.1 Hz, 1H), 3.37 (dd, *J* = 7.5, 5.0 Hz, 1H), 3.23 (td, *J* = 7.2, 5.9 Hz, 2H), 2.36 (m, 1H), 2.08 (dtd, *J* = 14.1, 7.5, 5.0 Hz, 1H), (dtd, *J* = 14.3, 7.7, 6.8 Hz, 1H), 1.55 – 1.45 (m, 2H), 1.43 (s, 9H), 1.36 – 1.17 (m, 20H), 0.86 (t, *J* = 6.8 Hz, 3H). ¹³C NMR (126 MHz, CDCl₃) δ = 174.3, 173.0, 80.7, 54.8, 39.3, 32.2, 32.05, 30.55, 29.9, 29.8, 29.73, 29.69, 29.5, 29.4, 28.2, 27.1, 22.8, 14.2. The spectroscopic data are in agreement with published data.¹

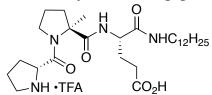
Procedure for the synthesis of Boc-DPro-MePro-OH: Boc-DPro-OH (9.5 g, 44.1 mmol,



synthesis of *Boc-Dirichlet ro-Ori*. Boc-Dirio-Ori (9.3 g, 44.1 million, 1.0 equiv.) and EDC•HCl (10.2 g, 53.0 mmol, 1.2 equiv.) were dissolved in EtOAc (110 mL). ${}^{i}Pr_{2}NEt$ (15.0 mL 88.3 mmol, 2.2 equiv.) was added and the mixture was stirred for 5 min before addition of H-MePro-OMe•HCl (6.3 g, 44.1 mmol, 1.0 equiv.). The reaction mixture was stirred at room temperature overnight. The reaction mixture was diluted with EtOAc and subsequently washed with 1 M aq. HCl (100 mL), H₂O (100

mL), sat. aq. NaHCO₃ solution (100 mL) and brine (100 mL). The combined aqueous layers were then reextracted with EtOAc (10x). The combined organic layers were dried over MgSO₄ and the solvent removed under reduced pressure. The crude product (colorless solid, 5.7 g) was purified by flash column chromatography on silica gel eluting with 1-5% MeOH in CH₂Cl₂. The resulting colorless solid was dissolved in THF/MeOH 1:1 (20 mL) and NaOH (1.33 g, 2 equiv.) and water (1 mL) was added. The reaction mixture was stirred for 1.5-2 h, acidified with concentrated HCl (pH \approx 2) and extracted with CH₂Cl₂ (3x). The combined organic layers were dried over MgSO₄. Removal of all volatiles yielded the desired product as colorless solid (5.20 g, 37%). ¹H NMR (400 MHz, CDCl₃) δ = 4.44 (m, 1H), 4.05 – 3.77 (m, 1H), 3.67 – 3.39 (m, 3H), 2.85 – 2.58 (m, 1H), 2.32 – 1.78 (m, 3H), 1.67 and 1.65 (2 s, 3H), 1.44 and 1.40 (2 s, 9H). ¹³C NMR (126 MHz, CDCl₃) δ = 176.6, 174.3, 173.8, 173.4, 80.8, 80.6, 69.9, 68.9, 58.5, 58.0, 49.6, 49.4, 47.1, 46.9, 38.2, 37.6, 30.6, 29.5, 28.6, 28.3, 24.8, 23.9, 23.7, 23.5, 22.4, 22.2. [M + H]⁺ C₁₆H₂₇N₂O₅⁺: 327.1923; found: 327.1830.

Procedure for solution peptide synthesis of H-DPro-MePro-Glu-NHC12H25 2: Boc-DPro-



MePro-OH (0.88 g, 2.40 mmol, 1.0 equiv) and EDC•HCl (0.57 g, 2.96 mmol, 1.2 equiv.) were suspended in EtOAc (30 mL) and ${}^{i}Pr_{2}NEt$ (0.92 mL, 2.2 equiv.). After stirring for 5 min at room temperature H-Glu-NHC₁₂H₂₅ (1.00 g, 2.70 mmol, 1.0 equiv.) was added and the suspension was

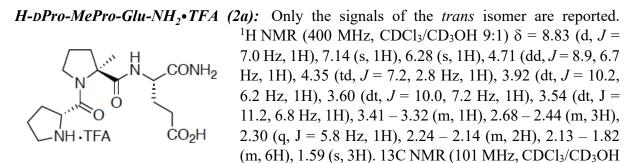
stirred at room temperature overnight. The reaction mixture was diluted with EtOAc (20 mL), washed with 0.1 M HCl (2x 5 mL), 5% w/w aq. Na₂CO₃ (5 mL) and brine (5 mL). The solution was dried over MgSO₄ and the solvent was removed under reduced pressure to give a yellowish oil which was purified by flash column chromatography on silica gel eluting with 1-5% MeOH in CH₂Cl₂. The protected peptide was dissolved in TFA/CH₂Cl₂ 2:1 (14 mL) and the mixture was stirred at room temperature for 30 min. Removal of all volatiles and precipitation from cold Et₂O yielded the desired product as a colorless solid (0.91 mg, 56 %). ¹H NMR (400 MHz, CDCl₃) δ = 8.84 (d, *J* = 7.1 Hz, 1H), 6.82 (t, *J* = 5.6 Hz, 1H), 4.80 – 4.71 (m, 1H), 4.44 – 4.35 (m, 1H), 3.95 – 3.85 (m, 1H), 3.81 – 3.70 (m, 1H), 3.65 – 3.54 (m, 1H), 3.42 – 3.31 (m, 1H), 3.30 – 3.14 (m, 1H), 2.64 – 2.21 (m, 6H), 2.15 – 1.86 (m, 4H), 1.83 – 1.72 (m, 1H), 1.64 (s, 3H), 1.53 – 1.42 (m, 2H), 1.33 – 1.19 (brs, 20H), 0.87 (td, *J* = 7.0, 2.3 Hz, 3H) (Mixture of *trans/cis* conformers in a ratio of > 30:1). ¹³C NMR (101 MHz, CDCl₃) δ = 179.9, 171.2, 170.1, 168.7, 77.5, 77.2, 76.8, 69.7, 59.8, 53.6, 49.1, 47.0, 39.7, 39.6, 32.1, 30.2, 30.1, 29.8, 29.8, 29.6, 29.5, 29.1, 27.1, 25.7, 25.0, 23.6, 22.8, 21.9, 14.3. HRMS (ESI): m/z calcd. for [M + H]⁺ C₂₈H₅₁N₄O₅⁺: 523.3854; found: 523.3850.

Peptide 2a was prepared on solid phase using Rink Amide resin. The general protocol for Fmoc/*t*Bu peptide synthesis was followed according to the general procedures below:

General procedure for peptide couplings: ${}^{i}Pr_{2}NEt$ (6 equiv.) was added to a solution of Fmoc-Xaa-OH (3 equiv.) and HATU (3 equiv.) in DMF. The solution of the activated amino acid ($\approx 100 \text{ mM}$) was added to the amino-functionalized resin (preswollen in CH₂Cl₂) and the mixture was agitated for 1 h before washing with DMF (3x) and CH₂Cl₂ (3x).

General procedure for Fmoc-deprotections: A solution of 20% piperidine in DMF was added to the resin (preswollen in CH_2Cl_2) and the reaction mixture was agitated for 10 min, drained and the piperidine treatment was repeated for 10 min. Finally, the resin was washed with DMF (3x) and CH_2Cl_2 (3x).

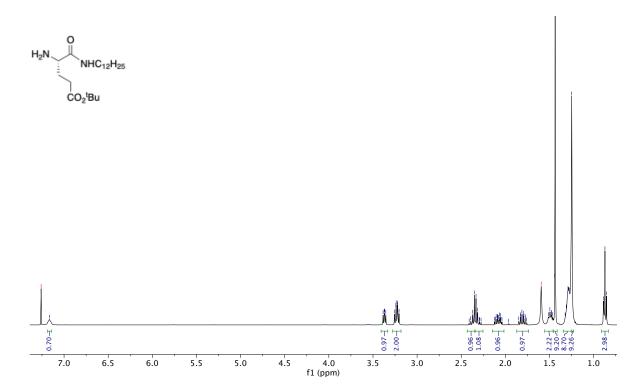
General procedure for side chain deprotection and cleavage of the peptides from the solid support: The peptides were side-chain deprotected and cleaved from the resin by shaking in a mixture of TFA/TIS/H₂O (95:2.5:2.5) for 1 h and a second time for 30 min. Pooling of the filtrates and removal of all volatiles under reduced pressure followed by precipitation and thorough washing with Et₂O afforded the peptides as their TFA-salts. The peptides were redissolved in MeCN/H₂O 1:1, dried by lyophilisation and used without further purification.

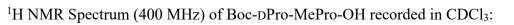


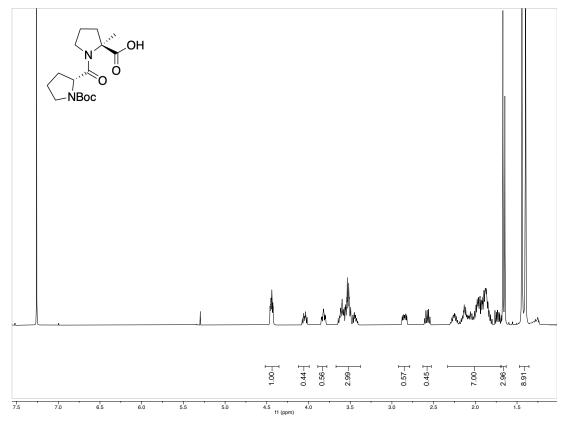
9:1) δ = 178.7, 174.8, 173.0, 168.5, 68.8, 59.4, 53.6, 48.7, 46.8, 39.4, 30.2, 29.1, 25.0, 24.8, 23.7, 21.2. HRMS (MALDI): m/z calcd. for [M + H]⁺ C₁₆H₂₇N₄O₅⁺: 355.1976; found: 355.1976.

¹H and ¹³C NMR Spectra

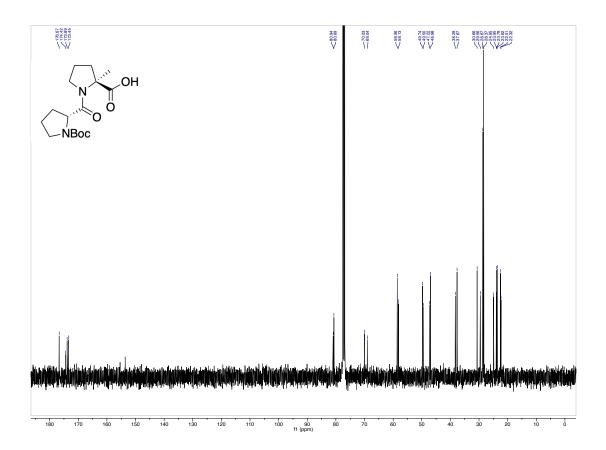
¹H NMR Spectrum (400 MHz) of H-Glu(O'Bu)-NHC₁₂H₂₅ recorded in CDCl₃:



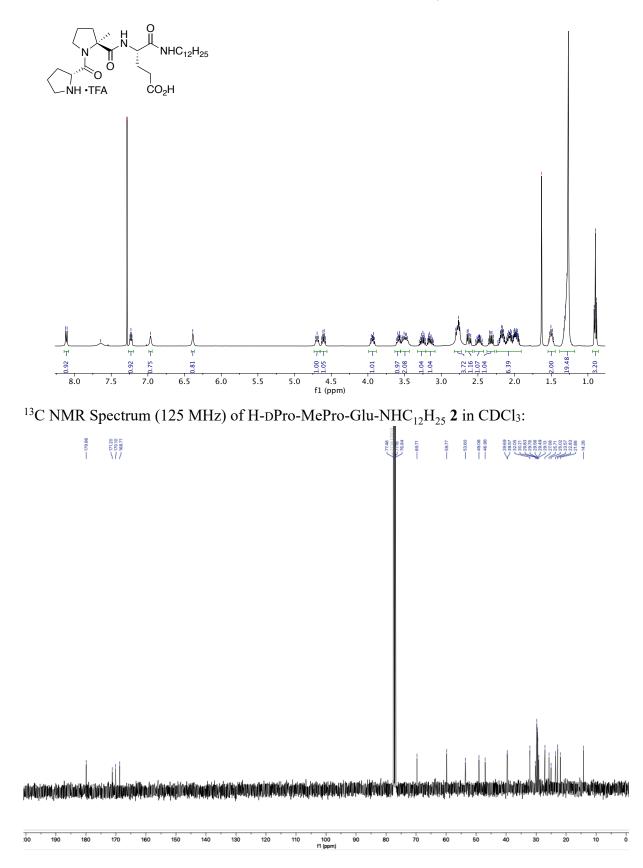




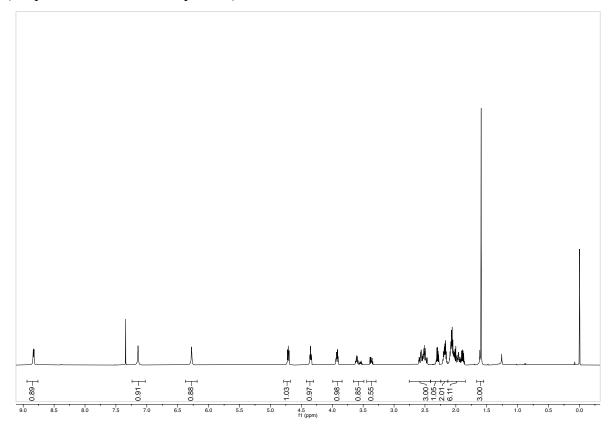
¹³C NMR Spectrum (126 MHz) of Boc-DPro-MePro-OH recorded in CDCl₃:



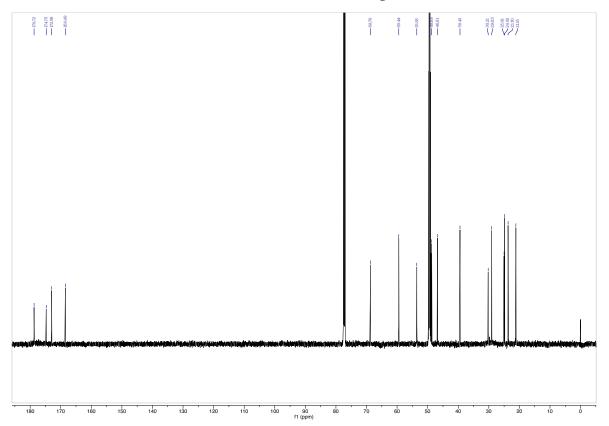
¹H NMR Spectrum (400 MHz) of H-DPro-MePro-Glu-NHC₁₂H₂₅ **2** in CDCl₃:



¹H NMR Spectrum (400 MHz) of H-DPro-MePro-Glu-NH₂ recorded in CDCl₃/CD₃OH 9:1: (the peak of CD₃O<u>H</u> was supressed)

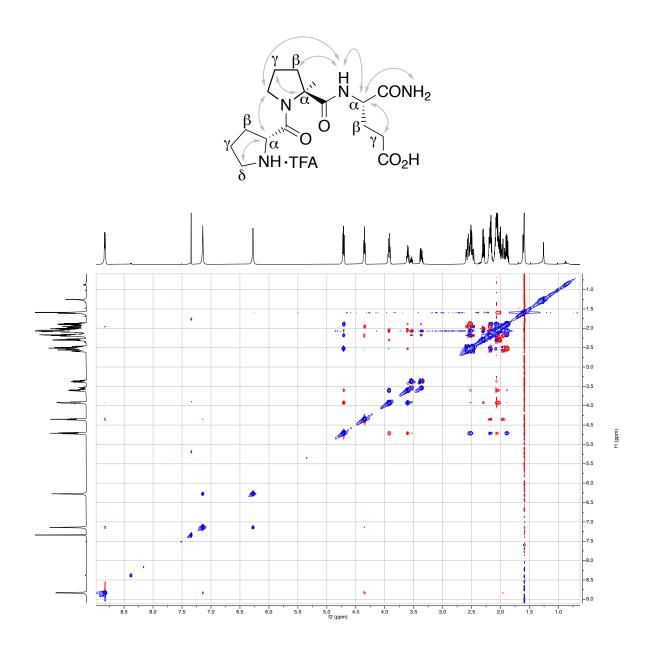


¹³C NMR Spectrum (150 MHz) of H-DPro-MePro-Glu-NH₂ recorded in CDCl₃/CD₃OH 9:1:



3. ROESY spectrum of 2a

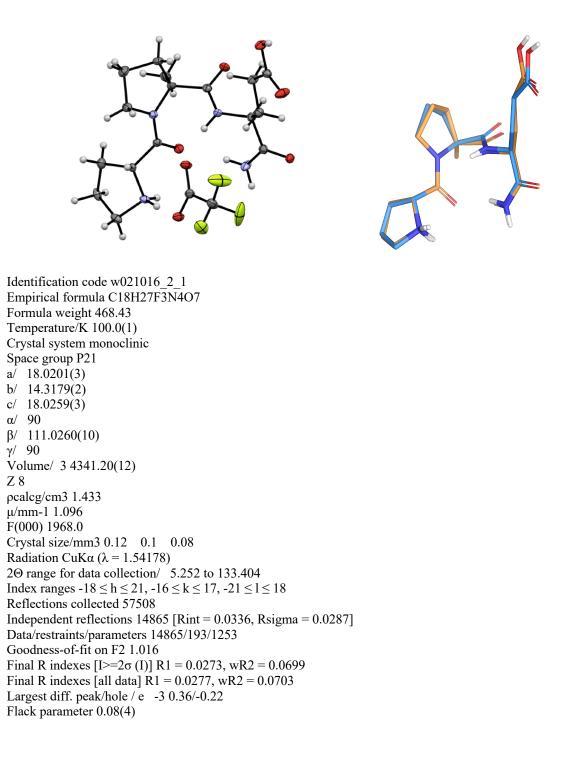
The ROESY NMR spectrum of peptide 2a was recorded in CDCl₃/CD₃OH 9:1, a solvent in which the population of the minor (*cis*) conformer is low and in which only the characteristic cross peaks for the major (*trans*) conformer are clearly visible. Note, 1a has been intensively studied in this solvent mixture,³ allowing for comparison of the conformation of 1a and 2a.



4. Crystal Structure of H-DPro-MePro-Glu-NH₂ (2a)

Single crystals of **2a** were obtained by vapor diffusion with MeOH as the solvent and THF as the antisolvent. A suitable crystal was selected and measured on a Bruker/Nonius ApexII diffractometer. The crystal was kept at 100.0(2) K during data collection. Using Olex2,⁴ the structure was solved with the XT⁵ structure solution program using Charge Flipping and refined with the XL⁶ refinement package using Least Squares minimization. Deposit number in CCSD: 2152664.

Left: ORTEP representation of the crystal structure of 2a. Right: Overlay of the crystal structures of $1a^7$ and 2a.



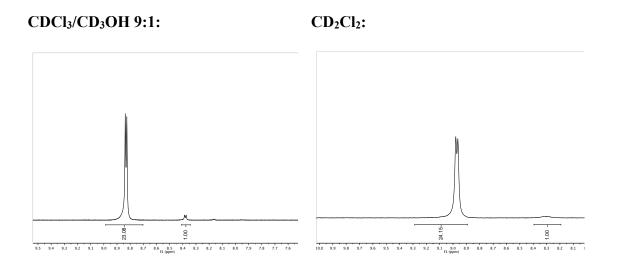
5. **Conjugate Addition Reaction with Peptides 1a and 2a in Organic Solvents**

Reaction procedure: The peptide-TFA salt (1 mol%, 2.5 µmol) was added to a solution of Nmethylmorpholine (1 mol%, 2.5 µmol), (E)-nitrostyrene (1 equiv., 250 µmol, 37.3 mg) and butanal (1.5 equiv., 375 µmol, 33.8 µl) in the respective solvent (0.5 ml). The reaction mixture was stirred for 2 h. The conversion and diastereoisomeric ratio were determined by ¹H NMR spectroscopy of the crude mixture by comparison of the aldehyde R-CHO signals. The enantiomeric excess was determined by chiral stationary phase SFC (AD-3, 5% MeOH, 2.0 ml/min, 40° C, 214 nm, 1.00 min (syn, minor), 1.21 min (syn, major)).

H Et	+ Ph	NO_2 1	mol% Peptide · TFA mol% NMM olvent, 0 °C, 2 h	H Et	Ph NO ₂
Solvent	Peptide	<i>trans</i> population ^s	a d.r. ^b	<i>ee</i> (%) ^c	conv. (%) ^b
CH ₂ Cl ₂	1a	91	18:1	89	23
	2a	95	34:1	98	27
THF	1a	88	31:1	84	6
1111	2a	96	36:1	95	18
Dioxane	1a	89	17:1	85	20
Dioxalle	2a	95	29:1	96	20 33 36
MeOH	1a	94	17:1	70	36
меон	2a	95	29:1	85	71
MeCN	1a	95	19:1	80	36
IVIECIN	2a	96	22:1	87	64
DMF	1a	78	15:1	63	20
DIVIT	2a	97	22:1	74	42
DMSO	1a	78	8:1	40	60
DIVISO	2a	96	14:1	77	8
CHCl ₃ / ^{<i>i</i>} PrOH 9:1	1a	98	35:1	97	quant.
	2a	96	54:1	98	quant.

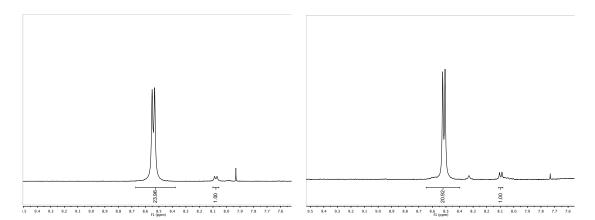
^a Determined by ¹H NMR spectroscopic analysis in the corresponding anhydrous deuterated solvent. Note: The *trans/cis* ratio depends on the water content of the deuterated solvent. ^b Determined after 2 hours by ¹H NMR spectroscopic analysis of the crude product and comparison of the aldehyde signals. ^c Determined by chiral stationary phase SFC analysis. The trans/cis ratios of 1a have been previously reported in ref 7.

¹H NMR spectroscopic analysis of *cis/trans* ratios: ¹H NMR spectra of peptides 2a were recorded at a concentration of \sim 5 mM, the concentration used in the conjugate addition reactions, or, in case of low solubility of the peptide in a saturated solution. No evidence for peptide aggregation was observed. The samples were equilibrated for at least 2 h and no changes of the *trans/cis* ratios over time were observed. The *trans/cis* ratios were determined by manual integration of baseline separated signals of the two spin systems.



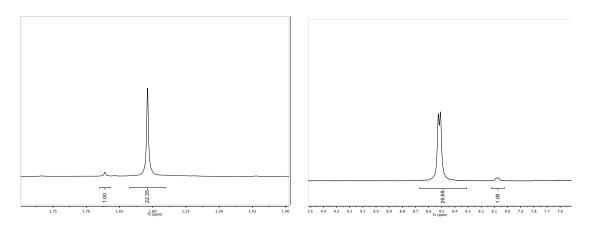
d⁸-THF:

d⁸-dioxane:



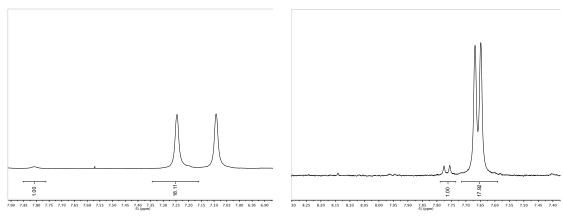
CD₃OH:

CD₃CN:



d⁷-DMF:

d⁶-DMSO



6. Conjugate Addition Reaction in the Presence of Additives

The peptide TFA salt (3 mol%) was added to a solution of *N*-methylmorpholine (3 mol%, 1.45 μ L), (*E*)-nitrostyrene (2 equiv., 0.88 mmol, 65.6 mg) and butanal (1.0 equiv., 0.44 mmol, 99.1 μ L) in the respective solutions (1.0 mL, complex mixture or additive solution in water). The reaction mixture was sonicated until no more solid (nitroolefin) was observed and a stable emulsion was obtained (approximately 2 min). The resulting emulsion was vigorously stirred at room temperature. The conversion and diastereoisomeric ratio were determined by ¹H NMR spectroscopic analysis of the crude mixture by comparison of the aldehyde R-C<u>H</u>O signals. The enantiomeric excess was determined by chiral stationary phase SFC (AD-3, 5% MeOH, 2.0 mL/min, 40 °C, 214 nm, 1.00 min (*syn*, minor), 1.21 min (*syn*, major)).

Additives	Conversion ^a (%) Amount of additive (mol%)			Major diastereomer ^a (%) Amount of additive (mol%)			ee ^b (%) Amount of additive (mol%)					
	3	10	50	100	3	10	50	100	3	10	50	100
No additive	>95	-	-	-	98	-	-	-	91	-	-	-
Α	>95	>95	>95	>95	96	96	95	95	91	89	90	91
В	>95	>95	>95	>95	95	91	68	66	91	90	88	88
С	>95	>95	>95	>95	97	96	96	96	91	91	91	90
D	>95	>95	>95	>95	97	96	97	96	90	90	91	91
Ε	>95	>95	>95	>95	96	97	96	96	91	91	90	91
F	>95	>95	>95	>95	96	96	95	96	90	91	91	91
G	>95	>95	>95	>95	97	96	97	97	91	91	91	91
Н	>95	>95	>95	>95	96	95	93	94	91	89	88	87
I	>95	>95	>95	>95	96	96	96	94	90	90	90	90
J	>95	>95	>95	>95	96	96	96	96	90	90	89	88
K	>95	>95	>95	>95	96	97	97	97	90	92	91	91
L	>95	>95	>95	>95	97	97	98	98	90	90	91	91
Μ	>95	>95	>95	>95	96	97	97	96	92	91	89	88
Ν	>95	93	56	36	97	97	98	98	90	90	90	90
0	81	84	37	24	98	98	98	98	91	91	92	92
Р	>95	>95	>95	>95	94	92	85	78	89	90	90	83

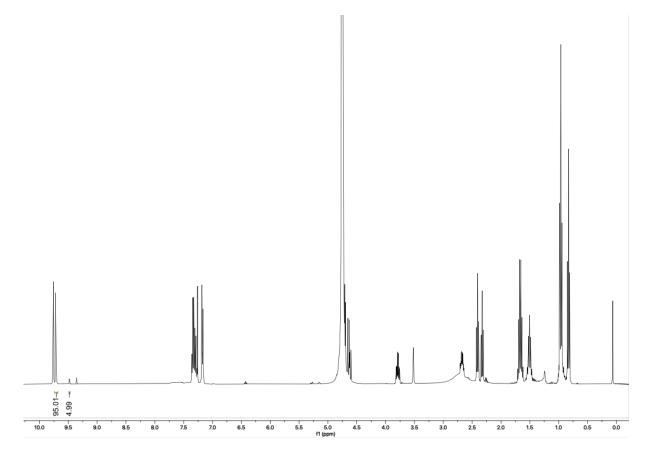
Results of the reactions in the presence of additives:

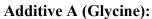
^aDetermined by ¹H NMR spectroscopic analysis of the crude mixture. ^bDetermined by chiral stationary phase SFC analysis.

¹H NMR spectra:

A sample of 50 μ L from the crude reaction mixture was added to an NMR tube that contained 600 μ L CDCl₃. Vigorous shaking / vortexing ensured extraction of the reaction products from the mixture. Conversion was determined by comparing the integrals of the olefinic signal of nitrostyrene and the RC<u>H</u>O signals of the γ -nitroaldehyde. The diastereomeric ratio was determined by integration of the RC<u>H</u>O signals of the γ -nitroaldehyde. Below an exemplary spectrum of the reaction with glycine is listed, for all other additives only a section.

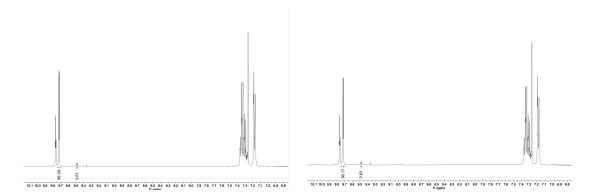
Exemplary spectrum of the reaction with 100 mol% glycine:

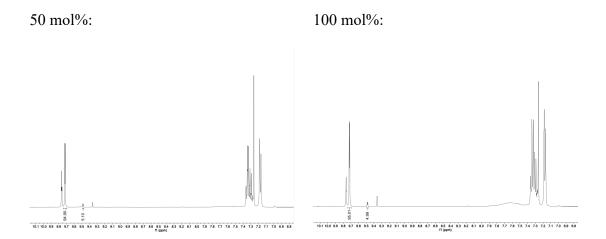




3 mol%:

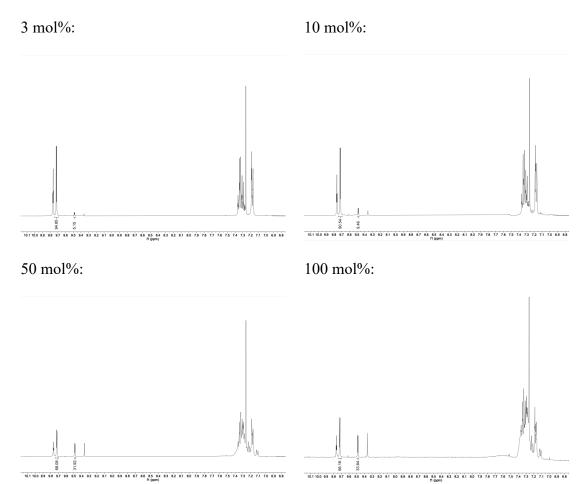




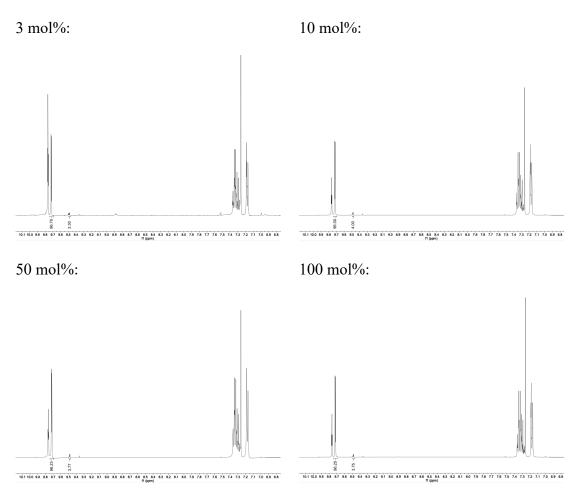


Additive B (Phenylalanine):

වි ව 10.1 10.0 19 98 97 96 95 94 93 92 91 90 89 88 87 86 85 84 83 82 81 80 79 78 77 78 75 74 73 72 71 70 69 68 ffigem)



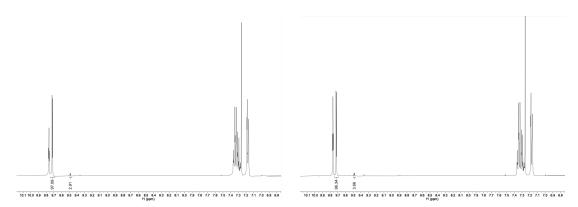
Additive C (Glucose):

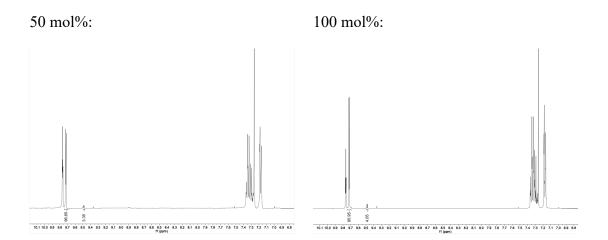


Additive D (Fructose):

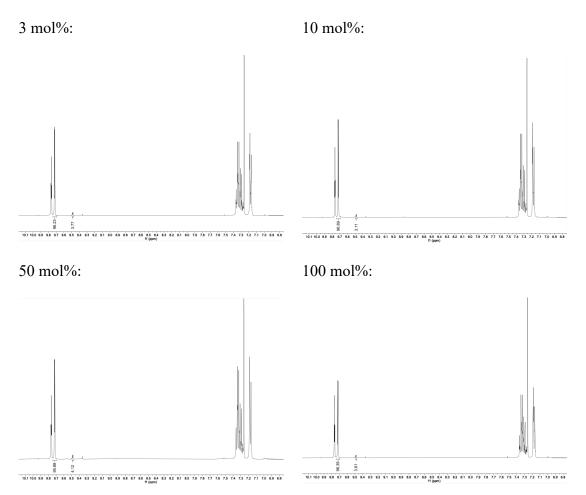
3 mol%:

10 mol%:

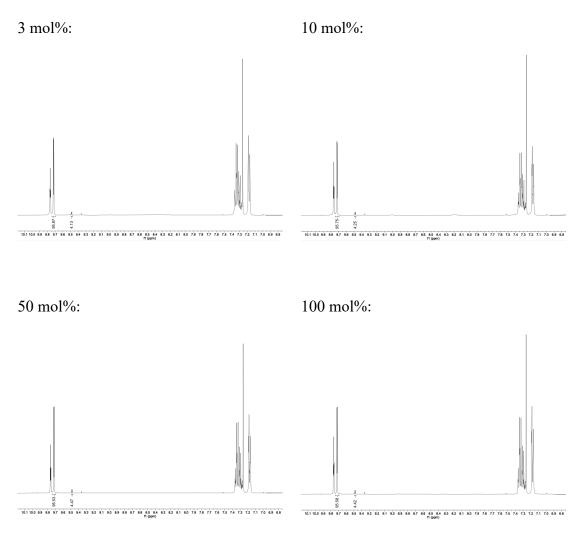




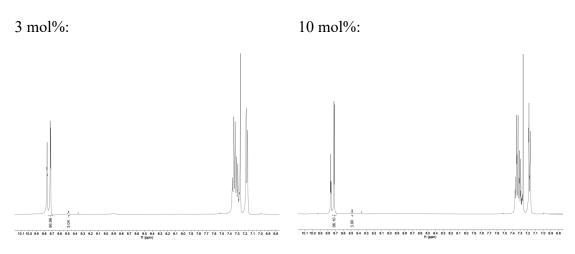
Additive E (Lactose):



Additive F (Adenine):

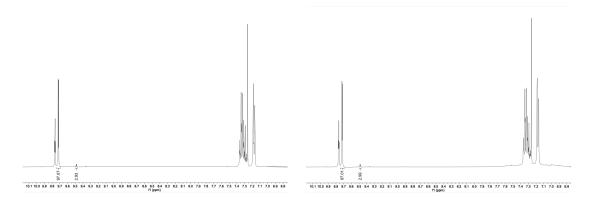


Additive G (Guanosine):

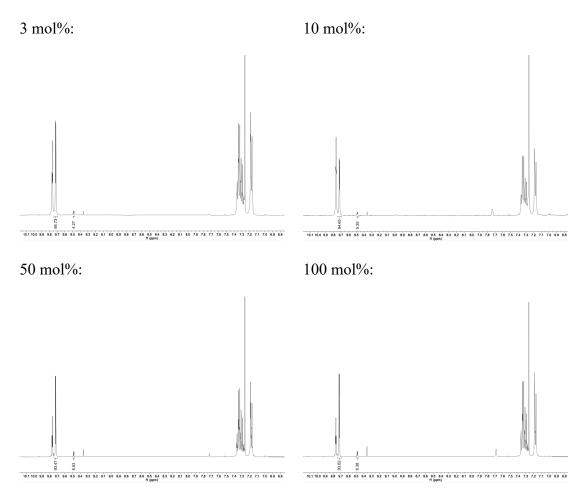




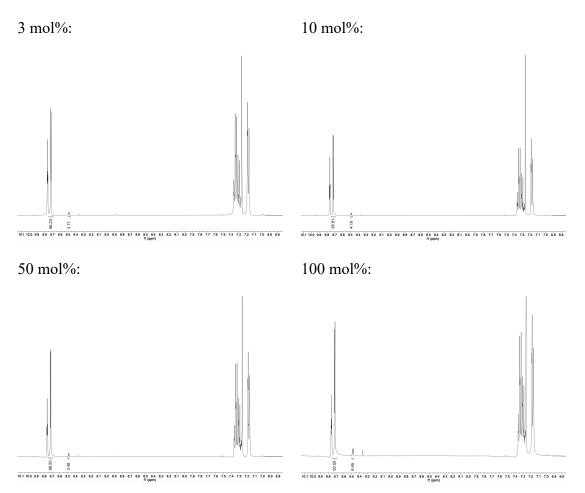
100 mol%:



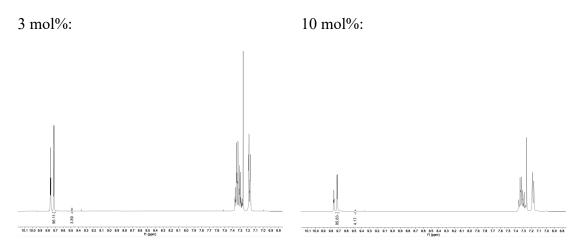
Additive H (Thymidine-5`-monophosphoric acid):

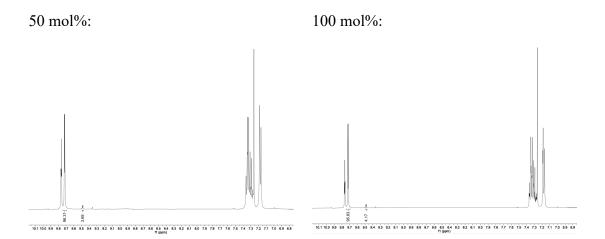


Additive I (Tristearine):

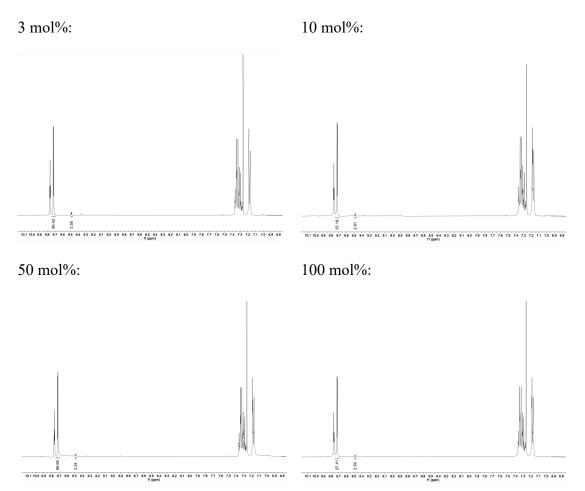


Additive J (Squalene):

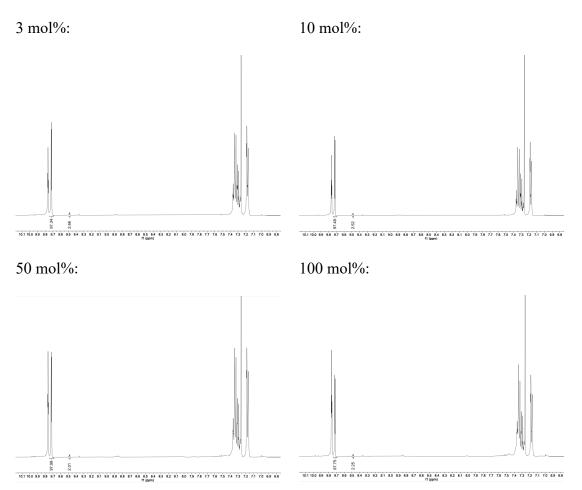




Additive K (Cholesterol):



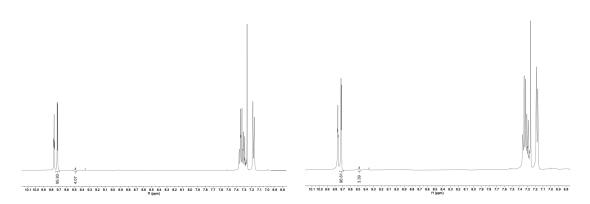
Additive L (Ascorbic acid):

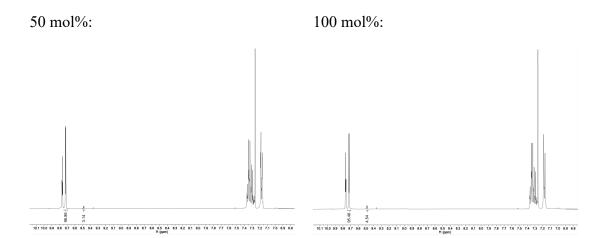


Additive M (Vitamine B₁₂):

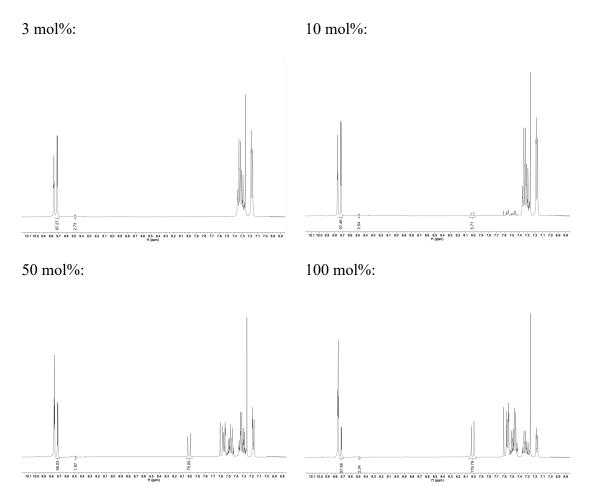
3 mol%:

10 mol%:

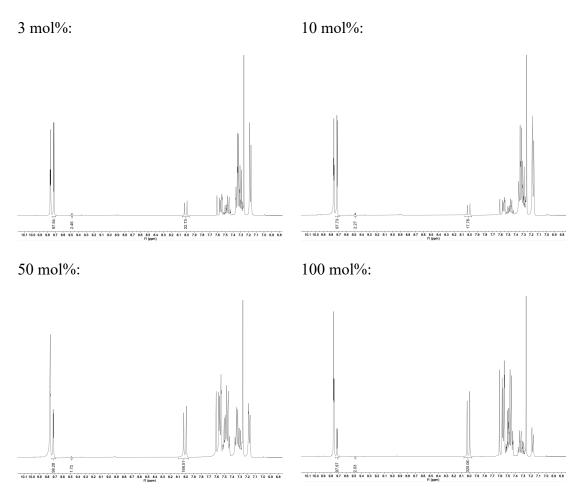




Additive N (Citric Acid):



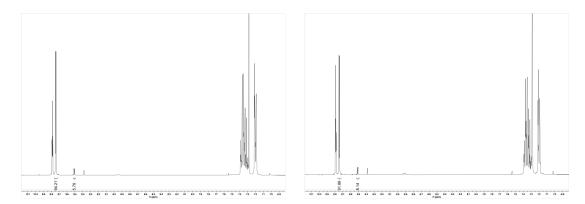
Additive O (Pyruvic acid):



Additive P (Glutathione):

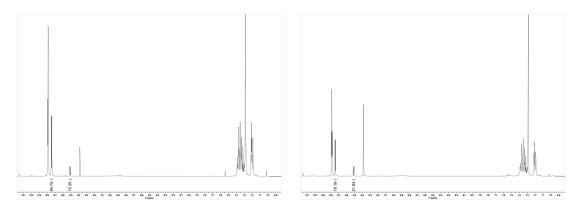
3 mol%:

10 mol%:



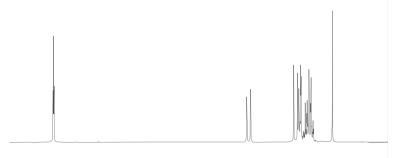


100 mol%:



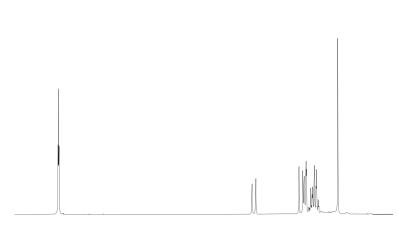
Control Experiments in the Absence of Catalyst (100 mol% additive)

Additive A (Glycine):



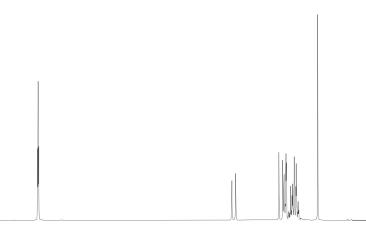
10.1 10.0 99 98 97 96 95 94 93 92 91 90 89 88 87 88 85 84 83 82 81 80 7.9 78 77 76 75 74 73 72 71 7.0 69 68 ff (ppm)

Additive B (Phenylalanine):



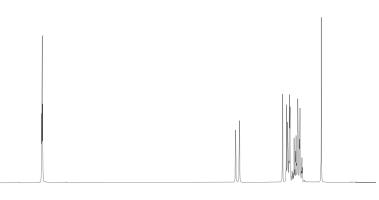
10.1 10.0 9.9 9.8 9.7 9.6 9.5 9.4 9.3 9.2 9.1 9.0 8.9 8.8 8.7 8.8 8.5 8.4 8.3 8.2 8.1 8.0 7.9 7.8 7.7 7.6 7.5 7.4 7.3 7.2 7.1 7.0 6.9 6.8 ff (ppm)

Additive C (Glucose):



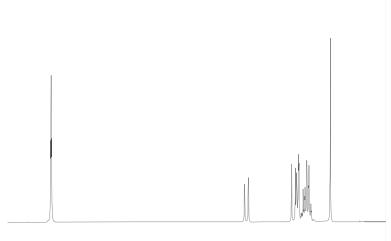
10.1 10.0 9.9 9.8 9.7 9.8 9.5 9.4 9.3 9.2 9.1 9.0 8.9 8.8 8.7 8.8 8.5 8.4 6.3 8.2 8.1 8.0 7.9 7.8 7.7 7.6 7.5 7.4 7.3 7.2 7.1 7.0 6.9 6.8 ft (gen)

Additive D (Fructose):



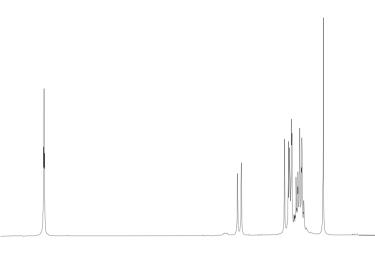
10.1 10.0 9.9 9.8 9.7 9.6 9.5 9.4 9.3 9.2 9.1 9.0 8.9 8.8 8.7 8.6 8.5 8.4 8.3 8.2 8.1 8.0 7.9 7.8 7.7 7.6 7.5 7.4 7.3 7.2 7.1 7.0 6.9 6.8 ff (pen)

Additive E (Lactose):



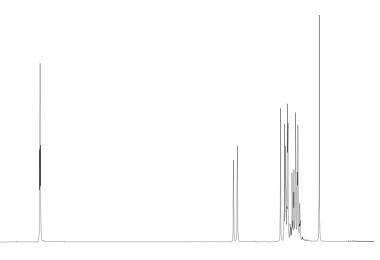
10.1 10.0 99 98 9.7 96 95 94 93 92 91 90 89 88 87 86 85 84 83 82 81 80 79 78 7.7 76 7.5 7.4 7.3 7.2 7.1 7.0 69 68 ff (ppm)

Additive F (Adenine):



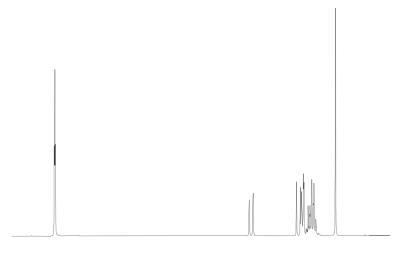
10.1 10.0 9.9 9.8 9.7 9.6 9.5 9.4 9.3 9.2 9.1 9.0 8.9 8.8 6.7 8.6 8.5 8.4 8.3 8.2 8.1 8.0 7.9 7.8 7.7 7.6 7.5 7.4 7.3 7.2 7.1 7.0 6.9 6.8 ff (ppm)

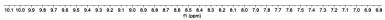
Additive G (Guanosine):



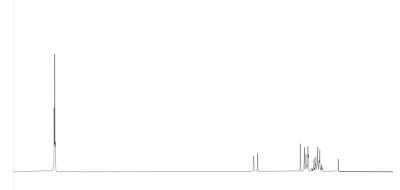
10.1 10.0 99 9.8 9.7 9.6 9.5 9.4 9.3 9.2 9.1 9.0 8.9 8.8 8.7 8.8 8.5 8.4 8.3 8.2 8.1 8.0 7.9 7.8 7.7 7.6 7.5 7.4 7.3 7.2 7.1 7.0 6.9 6.8 ff (ppm)

Additive H (Thymidine-5`-monophosphoric acid):



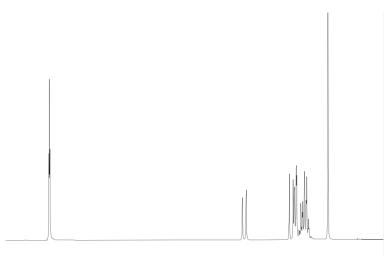


Additive I (Tristearine):



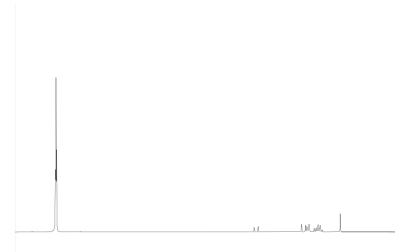
0.1 10.0 9.9 9.8 9.7 9.6 9.5 9.4 9.3 9.2 9.1 9.0 8.9 8.8 8.7 8.6 8.5 8.4 8.3 8.2 8.1 8.0 7.9 7.8 7.7 7.6 7.5 7.4 7.3 7.2 7.1 7.0 6.9 6 ft[bom]

Additive J (Squalene):



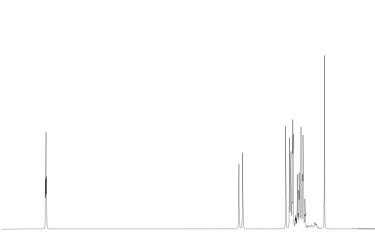
10.1 10.0 99 98 9.7 9.6 9.5 9.4 9.3 9.2 9.1 9.0 8.9 8.8 8.7 8.6 8.5 8.4 8.3 8.2 8.1 8.0 7.9 7.8 7.7 7.6 7.5 7.4 7.3 7.2 7.1 7.0 6.9 6.8 ff (ppm)

Additive K (Cholesterol):



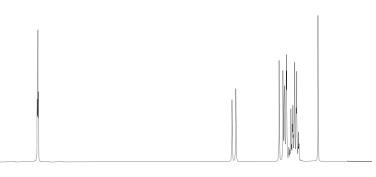
0.1 10.0 9.9 9.8 9.7 9.6 9.5 9.4 9.3 9.2 9.1 9.0 8.9 8.8 8.7 8.6 8.5 8.4 8.3 8.2 8.1 8.0 7.9 7.8 7.7 7.6 7.5 7.4 7.3 7.2 7.1 7.0 6.9 6 f1 (porm)

Additive L (Ascorbic acid):



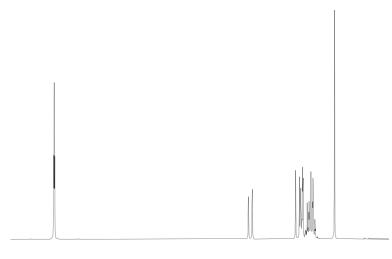
10.1 10.0 9.9 9.8 9.7 9.6 9.5 9.4 9.3 9.2 9.1 9.0 8.9 8.8 8.7 8.8 8.5 8.4 8.3 8.2 8.1 8.0 7.9 7.8 7.7 7.6 7.5 7.4 7.3 7.2 7.1 7.0 6.9 6.8 ff (ppm)

Additive M (Vitamine B₁₂):



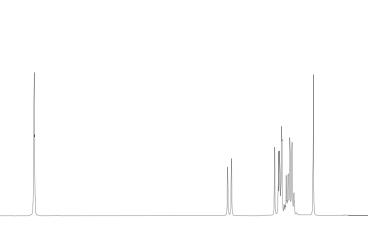
10.1 10.0 9.9 9.8 9.7 9.6 9.5 9.4 9.3 9.2 9.1 9.0 8.9 8.8 6.7 8.6 8.5 8.4 8.3 8.2 8.1 8.0 7.9 7.8 7.7 7.6 7.5 7.4 7.3 7.2 7.1 7.0 6.9 6.8 ff (ppm)

Additive N (Citric Acid):



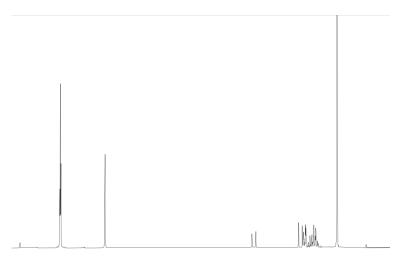
10.1 10.0 9.9 9.8 9.7 9.6 9.5 9.4 9.3 9.2 9.1 9.0 8.9 8.8 6.7 8.6 8.5 8.4 6.3 6.2 8.1 8.0 7.9 7.8 7.7 7.6 7.5 7.4 7.3 7.2 7.1 7.0 6.9 6.8 ff (ppm)

Additive O (Pyruvic acid):



10.1 10.0 99 98 97 96 95 94 93 92 91 90 89 88 87 88 85 84 83 82 81 80 7.9 78 7.7 76 7.5 7.4 7.3 7.2 7.1 7.0 69 68 ff (ppm)

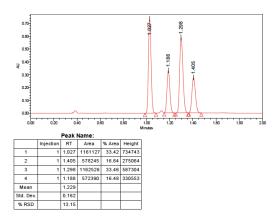
Additive P (Glutathione):



12 501 100 99 98 97 96 95 94 93 92 51 90 69 88 87 66 65 64 83 82 61 80 79 78 77 76 75 74 73 72 71 70 69 68 Filiperij

SFC chromatograms:

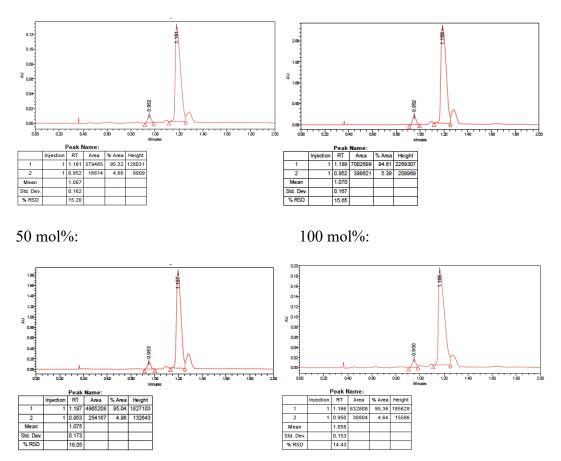
Racemate:



Additive A (Glycine):



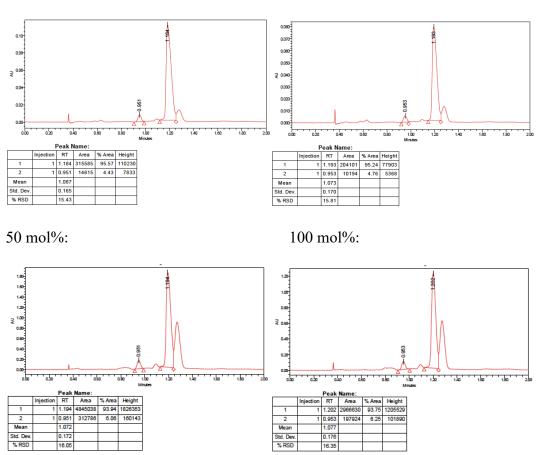
10 mol%:



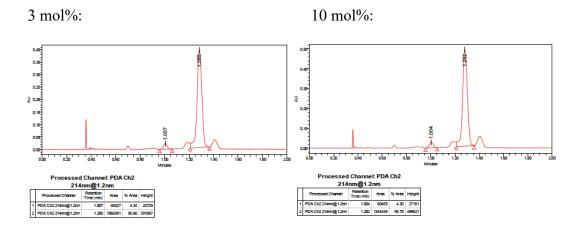
Additive B (Phenylalanine):





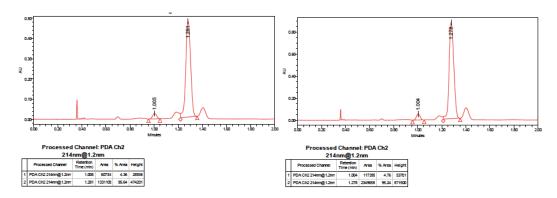


Additive C (Glucose):



S33

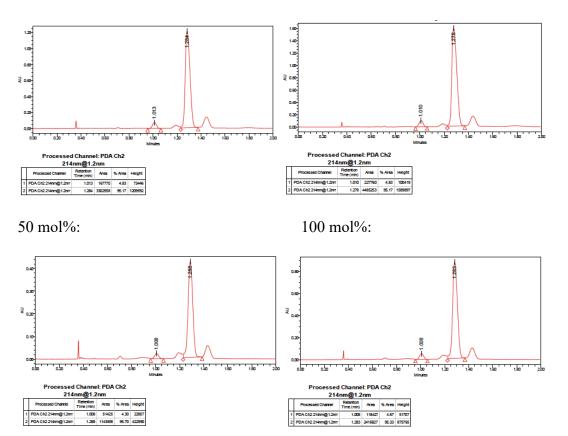
100 mol%:



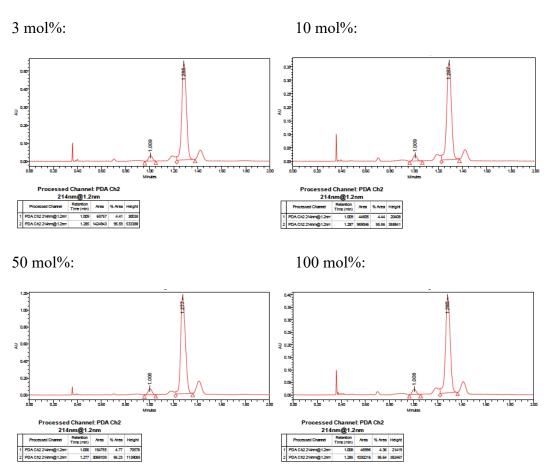
Additive D (Fructose):

3 mol%:

10 mol%:



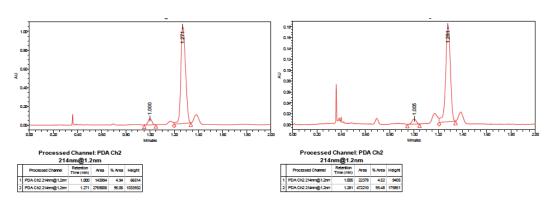
Additive E (Lactose):



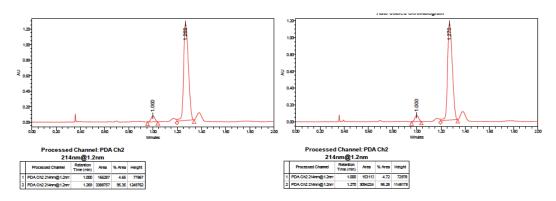
Additive F (Adenine):

3 mol%:

10 mol%:



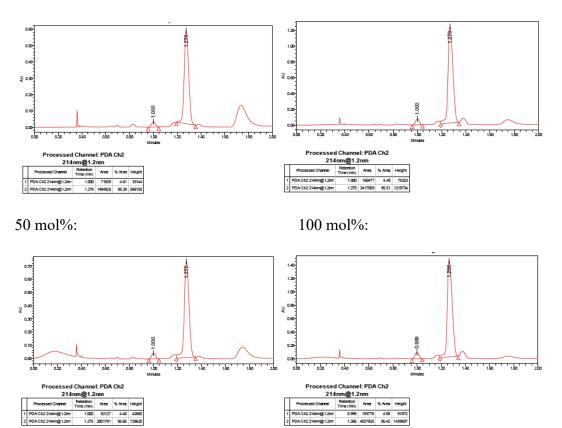
100 mol%:



Additive G (Guanosine):

3 mol%:

10 mol%:

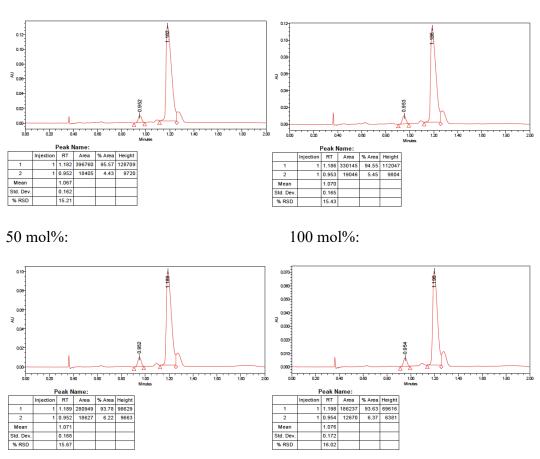


2 PDA Ch2 214nm@1.2nm

Additive H (Thymidine-5`-monophosphoric acid):

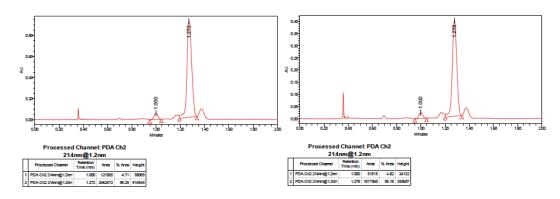


10 mol%:

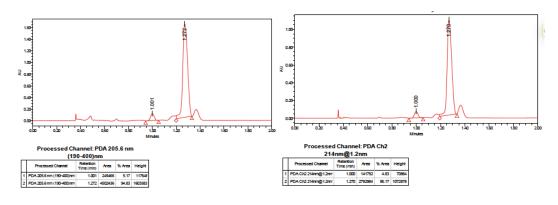


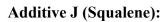
Additive I (Tristearine):

3 mol%:



100 mol%:

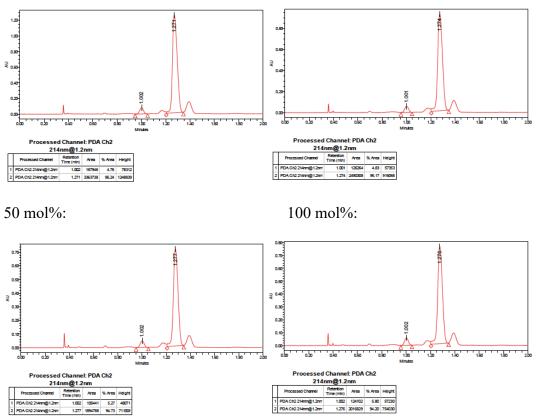




1.277 1894788 94.72

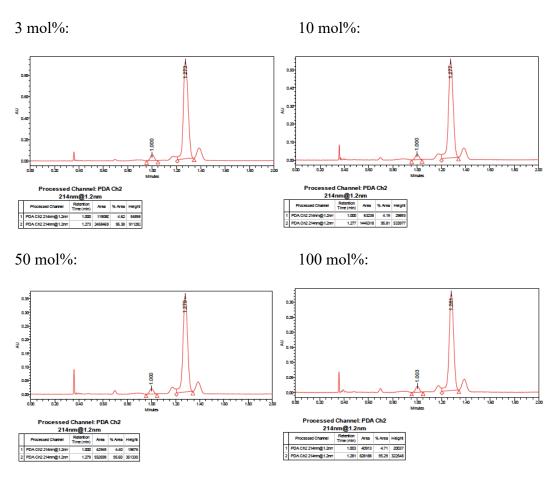


10 mol%:



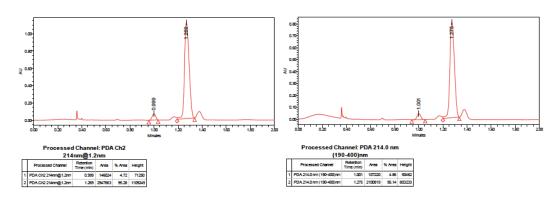
2 PDA Ch2 214

Additive K (Cholesterol):



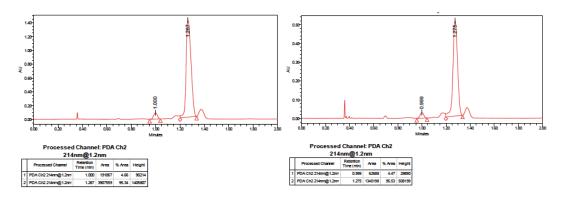
Additive L (Ascorbic acid):

10 mol%:



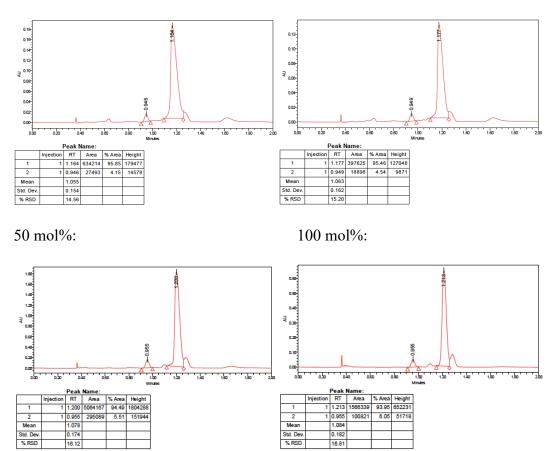
50 mol%:

100 mol%:

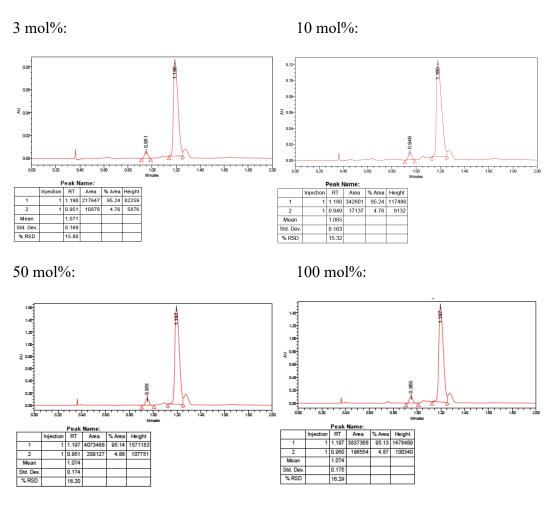


Additive M (Vitamine B₁₂):

10 mol%:

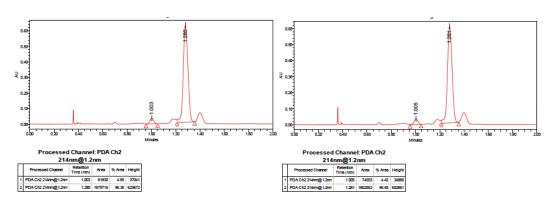


Additive N (Citric Acid):



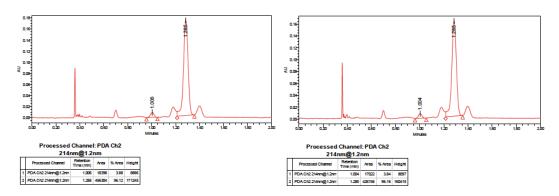
Additive O (Pyruvic acid):

3 mol%:

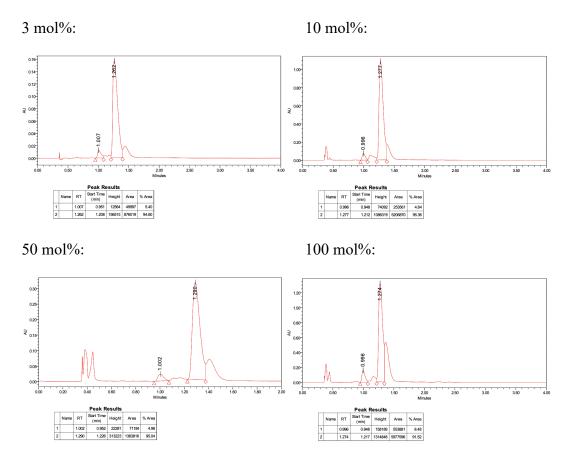


50 mol%:

100 mol%:



Additive P (Glutathione):



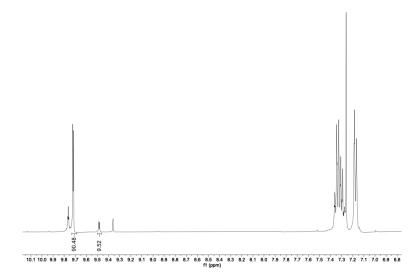
7. Conjugate Addition Reaction in Complex Mixtures

Solvent	Conversion ^a (%)		Major diastereomer ^a (%)		ee ^b (%)		Solvent	Conversion ^a (%)		Major diastereomer ^a (%)		ee ^b (%)	
Cat:	1	2	1	2	1	2		1	2	1	2	1	2
S1	>95	>95	90	90	85	96	S19	>95	>95	90	94	86	96
S2	>95	>95	90	90	84	96	S20	>95	>95	91	91	86	96
S3	>95	>95	90	89	85	96	S21	>95	>95	90	89	71	96
S4	>95	>95	89	90	84	96	S22	>95	>95	90	91	75	96
S5	>95	>95	91	87	88	96	S23	>95	>95	92	91	86	96
S6	>95	>95	91	87	88	96	S24	>95	>95	96	90	84	95
S7	>95	>95	90	89	87	96	S25	>95	>95	91	93	84	96
S8	>95	>95	90	89	86	96	S26	>95	>95	88	89	87	97
S9	>95	>95	91	91	86	96	S27	>95	>95	92	94	90	96
S10	>95	>95	92	93	89	96	S28	>95	>95	88	91	87	95
S11	>95	>95	92	95	80	96	S29	>95	>95	90	89	82	95
S12	>95	>95	92	94	89	96	S30	>95	>95	90	95	83	96
S13	>95	>95	88	89	88	96	S31	>95	>95	90	95	84	95
S14	>95	>95	89	91	87	96	S32	>95	>95	90	95	83	95
S15	>95	>95	87	91	86	96	S33	>95	>95	90	94	86	96
S16	>95	>95	92	90	88	96	S34	>95	>95	96	94	84	95
S17	>95	>95	90	91	79	96	S35	>95	>95	96	94	90	96
S18	>95	>95	90	90	83	96	S36	>95	>95	88	88	89	96
	ned by	¹ H NM					the crude						

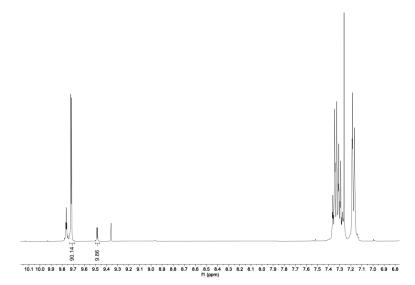
¹H NMR spectra of the reactions in the presence of H-DPro-Pro-Glu-NH₂ 1:

A sample of 50 μ L from the crude reaction mixture was added to an NMR tube that contained 600 μ L CDCl₃. Vigorous shaking or vortexing ensured extraction of the reaction products from the mixture. Conversion was determined by comparing the integrals of the olefinic signal of nitrostyrene and the RC<u>H</u>O signals of the γ -nitroaldehyde. The diastereomeric ratio was determined by integration of the RC<u>H</u>O signals of the γ -nitroaldehyde.

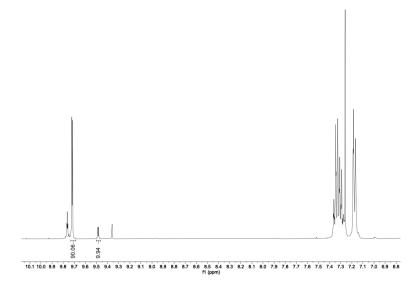
MilliQ water (S1):



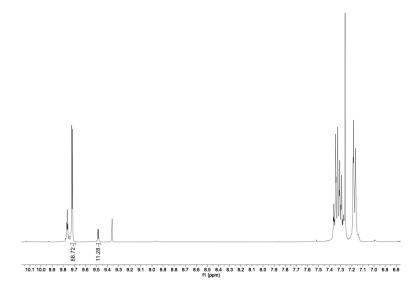




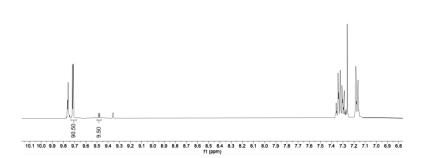
Lake Zürich water (S3):



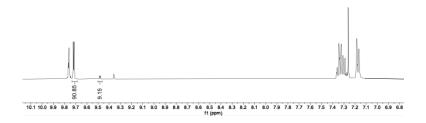




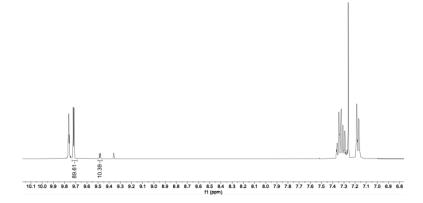




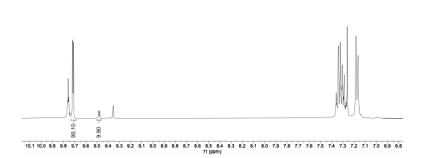
Low fat milk (S6):



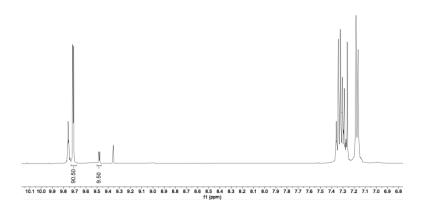


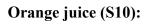


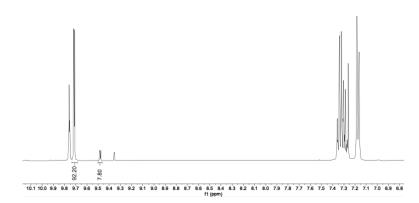




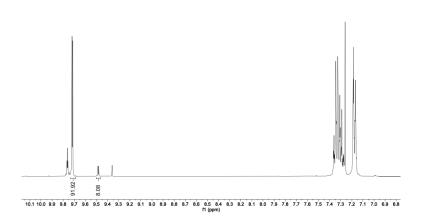
Rice milk (S9):



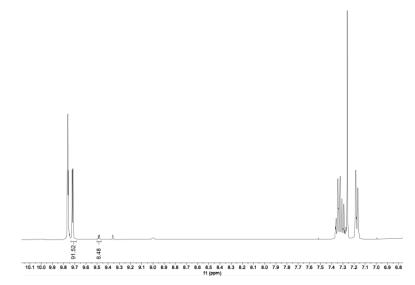


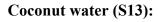


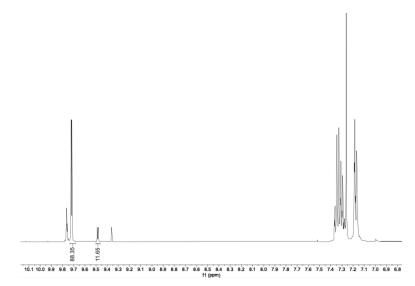
Blood orange juice (S11):

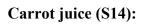


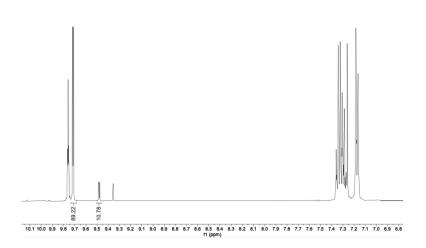
Apple juice (S12):



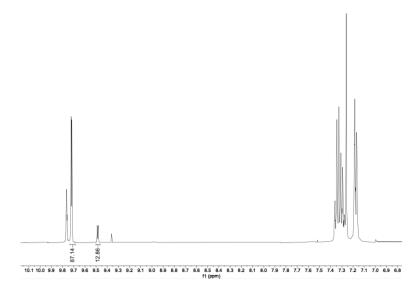




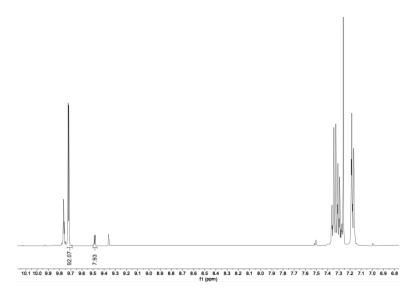


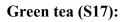


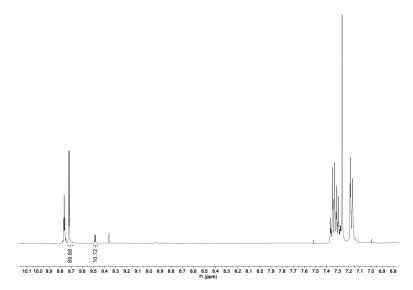
Beetroot juice (S15):



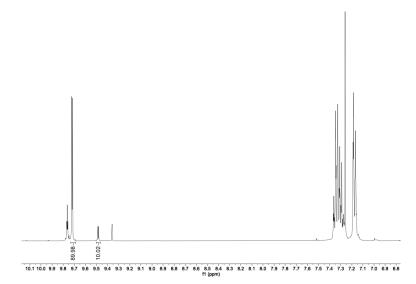


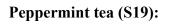


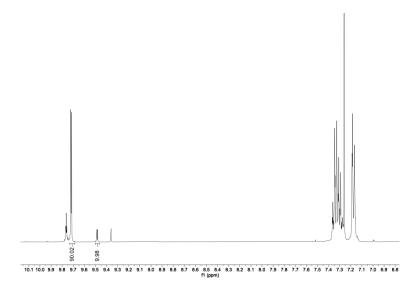


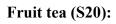


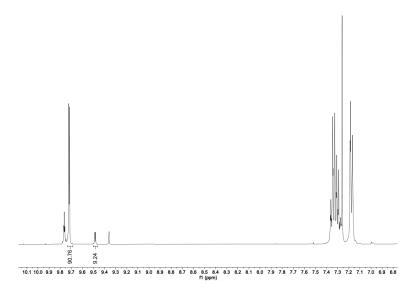
Black tea (S18):



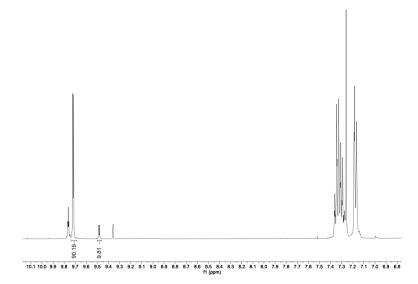




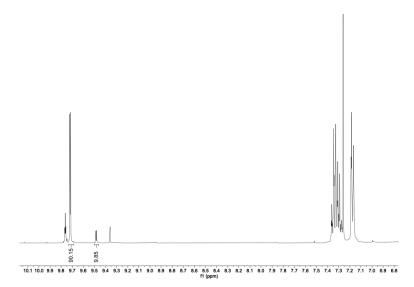


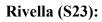


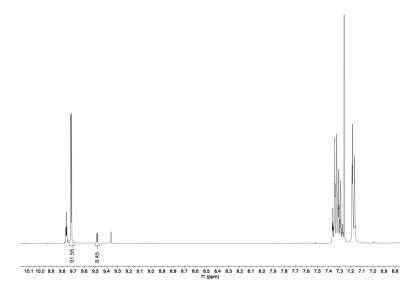
Honey solution 5 wt% (S21):



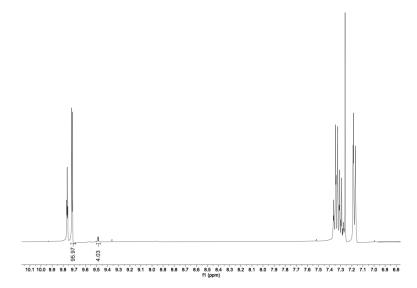




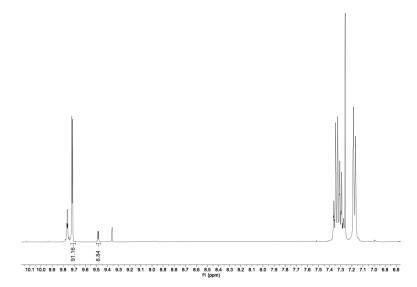




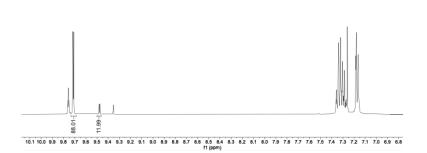
Diet Rivella (S24):



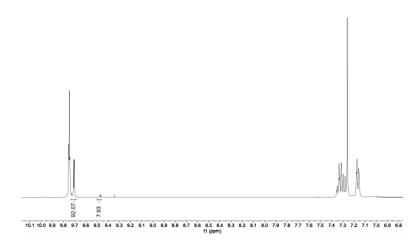




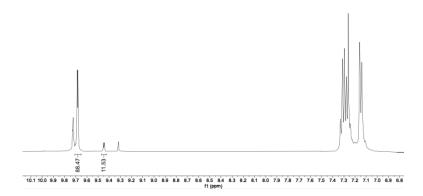




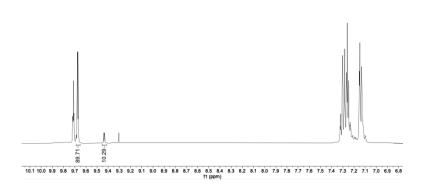
Red wine (S27):



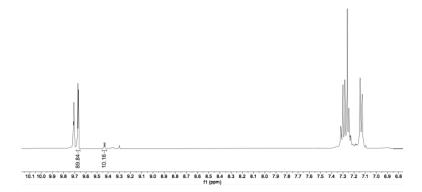




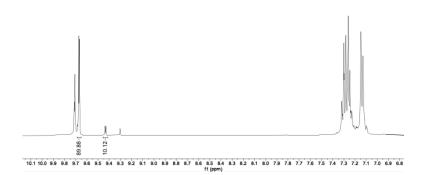
Herb Liqueur – "Jägermeister" (S29):



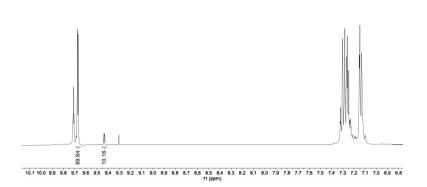
Scotch Whisky – Aberlour 18 y (S30):



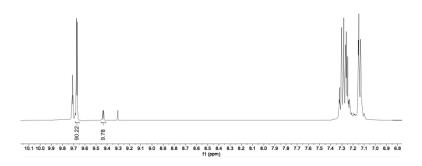




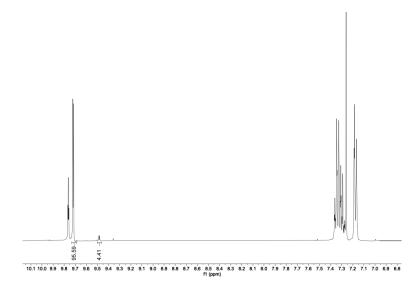


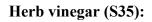


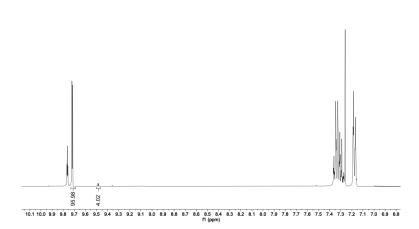
Rum (S33):



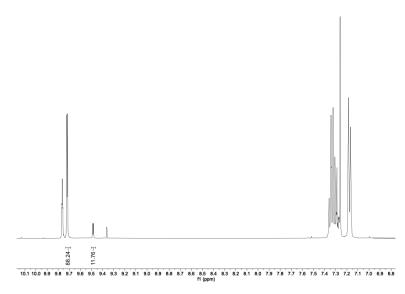






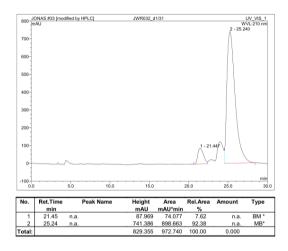


Olive oil (S36):

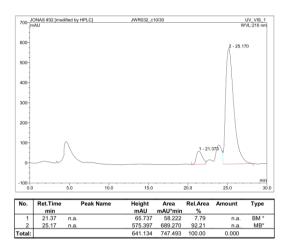


HPLC chromatograms of the reactions in presence of H-DPro-Pro-Glu-NH₂ 1:

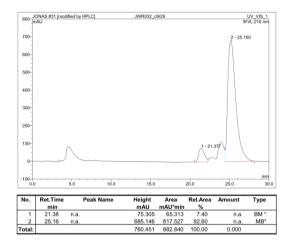
MilliQ water (S1):



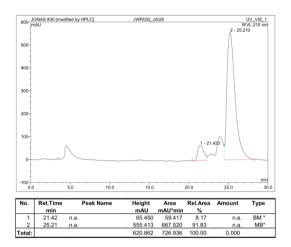
Tab water (S2):



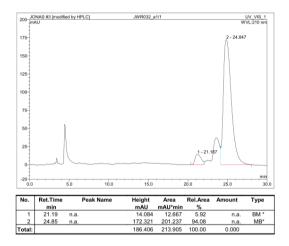
Lake Zürich water (S3):



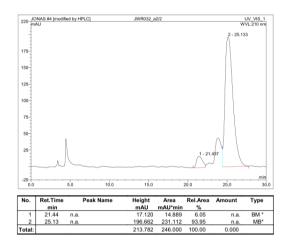
River Limmat water (S4):



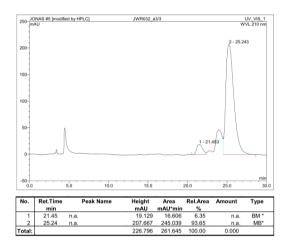
Milk (S5):



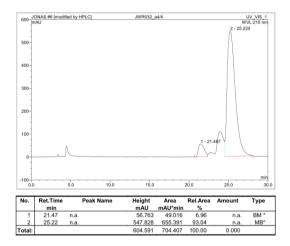
Low fat milk (S6):



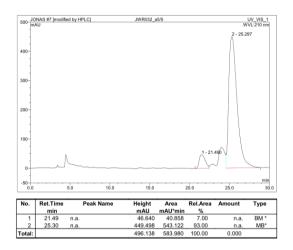
Soya milk (S7):



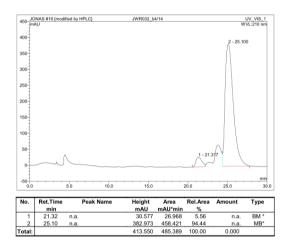
Almond milk (S8):



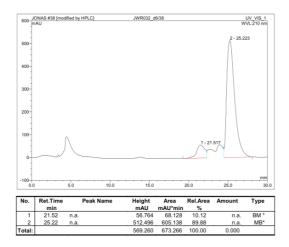
Rice milk (S9):



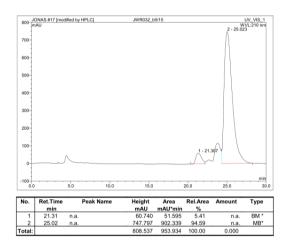
Orange juice (S10):



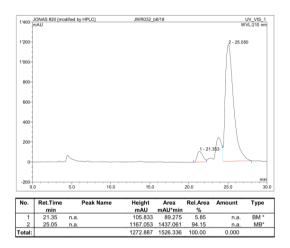
Blood orange juice (S11):



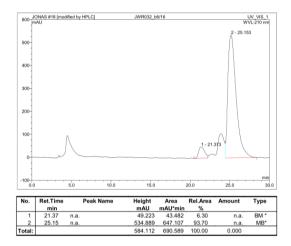
Apple juice (S12):



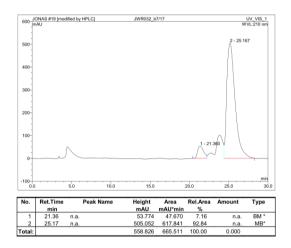
Coconut water (S13):



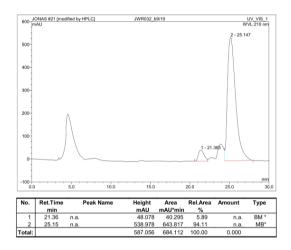
Carrot juice (S14):



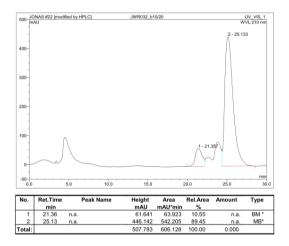
Beetroot juice (S15):



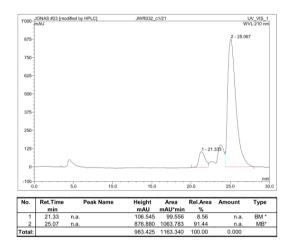
Coffee (S16):



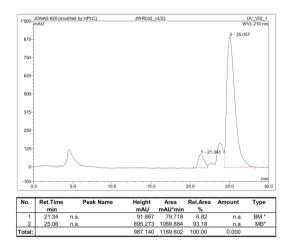
Green tea (S17):



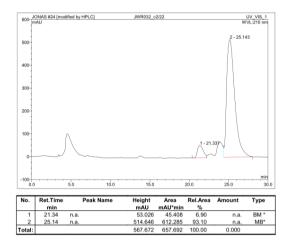
Black tea (S18):



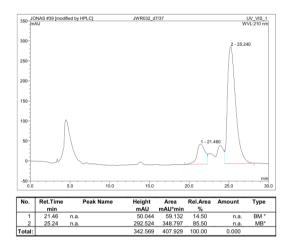
Peppermint tea (S19):



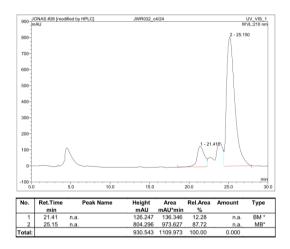
Fruit tea (S20):



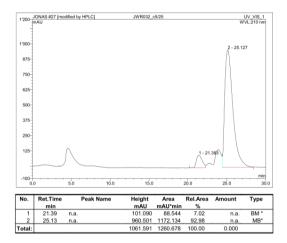
Honey solution 5 wt% (S21):



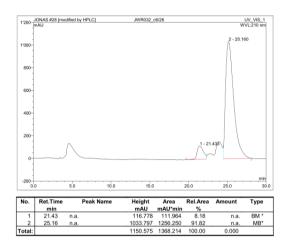
Coke (S22):



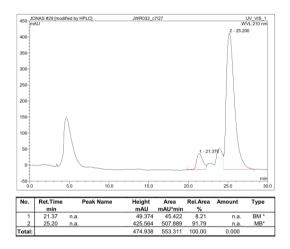
Rivella (S23):



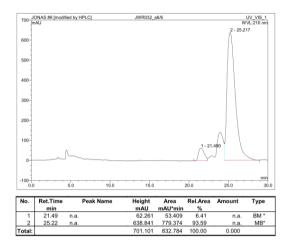
Diet Rivella (S24):



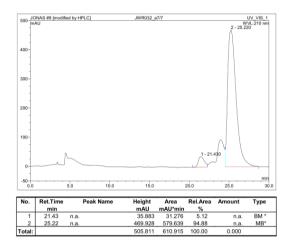
Ice tea (S25):



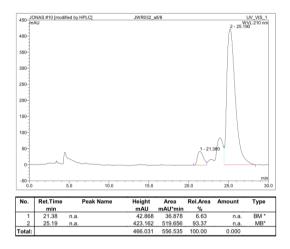
Beer (S26):



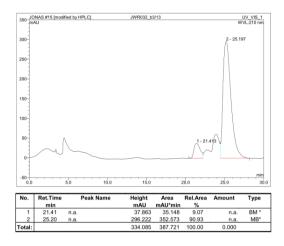
Red wine (S27):



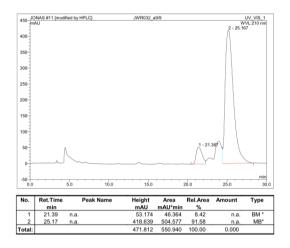
Amaretto (S28):



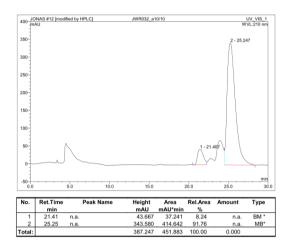
Herb Liqueur – "Jägermeister" (S29):



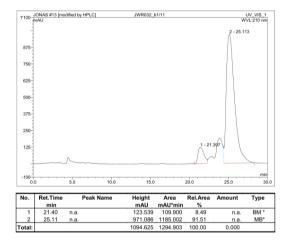
Scotch Whisky – Aberlour 18 y (S30):



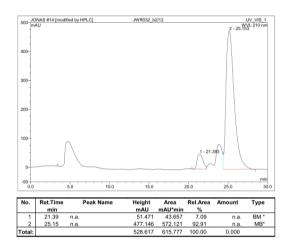
Vodka (S31):



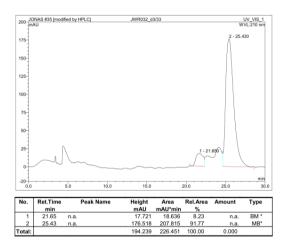
Gin (S32):



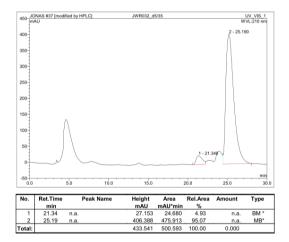
Rum (S33):



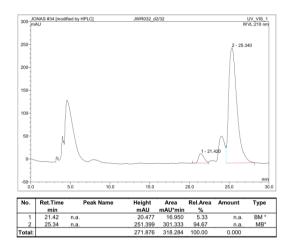
Aceto balsamico (S34):



Herb vinegar (S35):



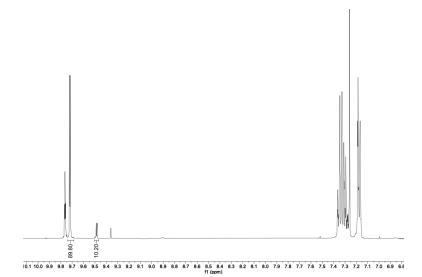
Olive oil (S36):



¹H NMR spectra of the reactions in the presence of H-DPro-MePro-Glu-NH₂ 2:

A sample of 50 μ L from the crude reaction mixture was added to an NMR tube that contained 600 μ L CDCl₃. Vigorous shaking or vortexing ensured extraction of the reaction products from the mixture. Conversion was determined by comparing the integrals of the olefinic signal of nitrostyrene and the RC<u>H</u>O signals of the γ -nitroaldehyde. The diastereomeric ratio was determined by integration of the RC<u>H</u>O signals of the γ -nitroaldehyde.

MilliQ water (S1):

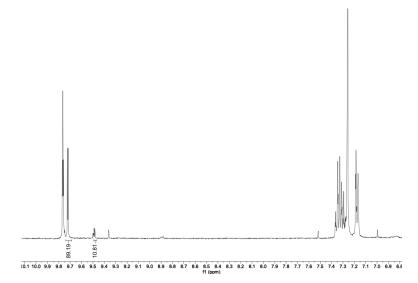




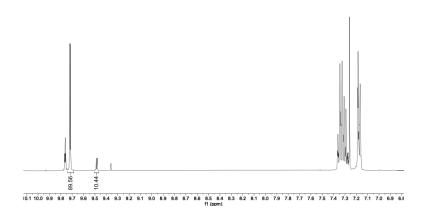


10.1 10.0 9.9 9.8 9.7 9.6 9.5 9.4 9.3 9.2 9.1 9.0 8.9 8.8 8.7 8.6 8.5 8.4 8.3 8.2 8.1 8.0 7.9 7.8 7.7 7.6 7.5 7.4 7.3 7.2 7.1 7.0 6.9 6.8

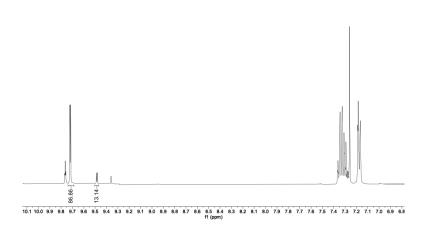
Lake Zürich water (S3):



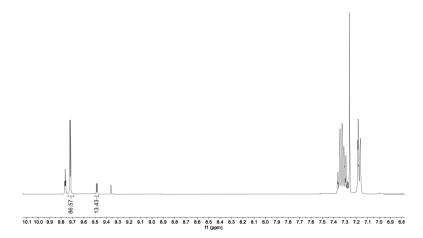




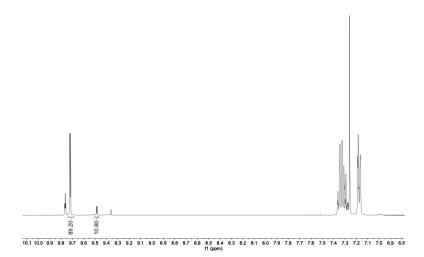


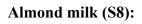


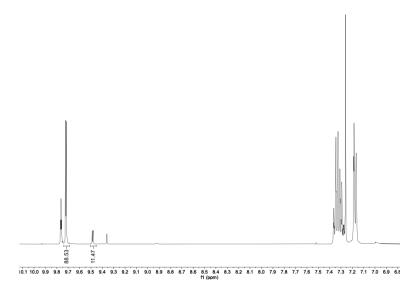
Low fat milk (S6):



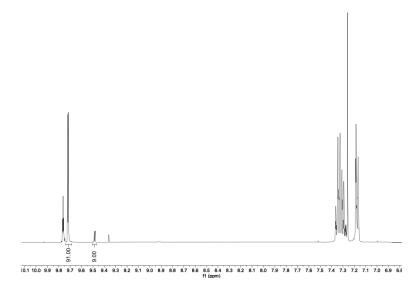


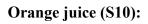


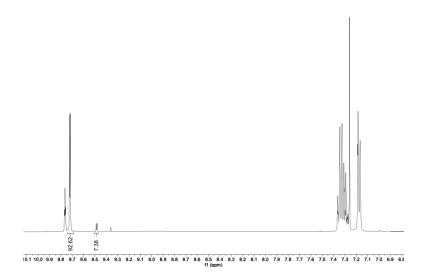




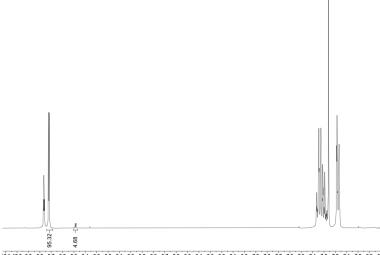
Rice milk (S9):

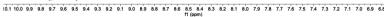




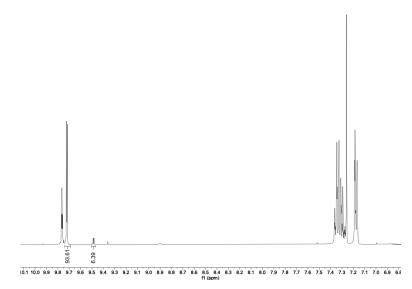


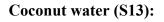
Blood orange juice (S11):

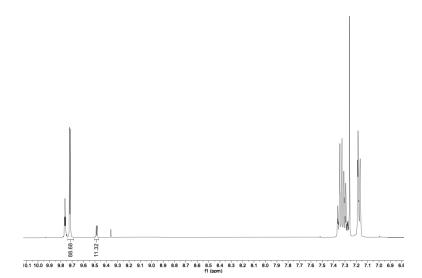




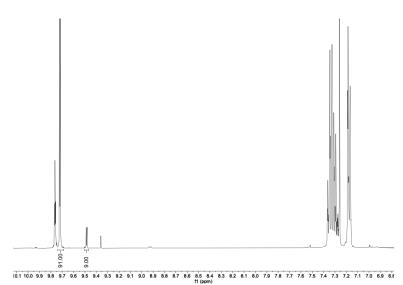
Apple juice (S12):





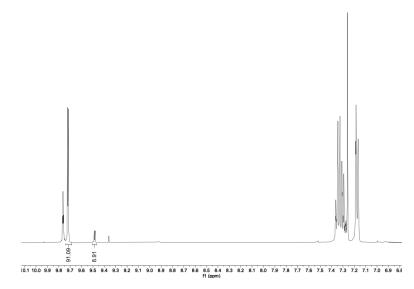


Carrot juice (S14):

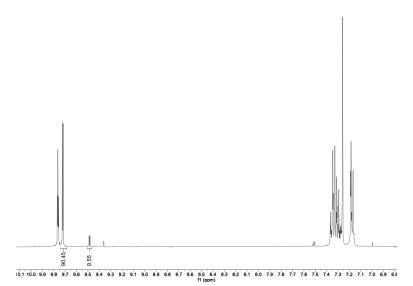


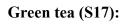
S73

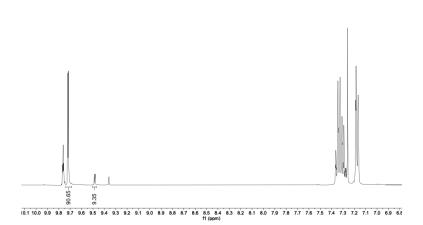
Beetroot juice (S15):



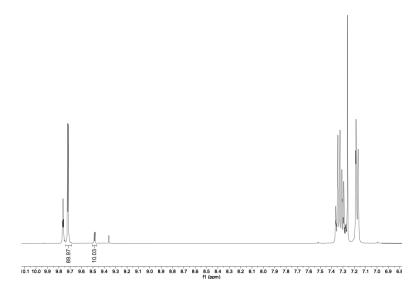




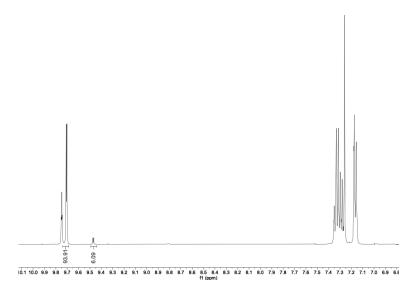


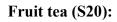


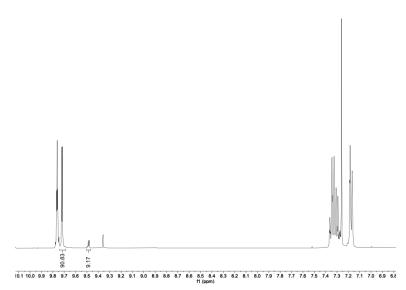
Black tea (S18):



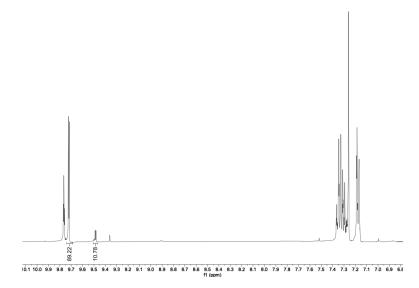




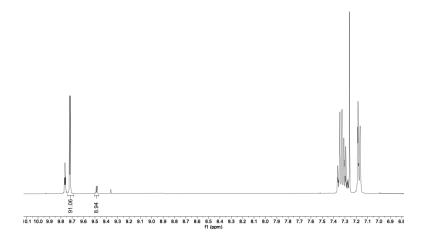




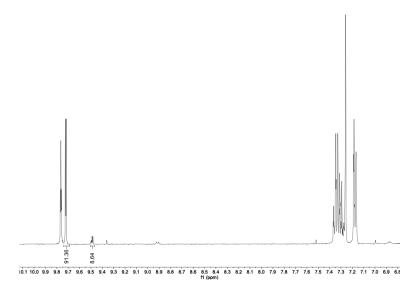
Honey solution 5 wt% (S21):





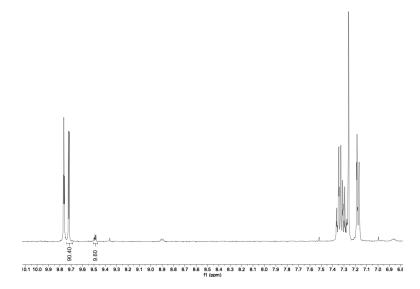




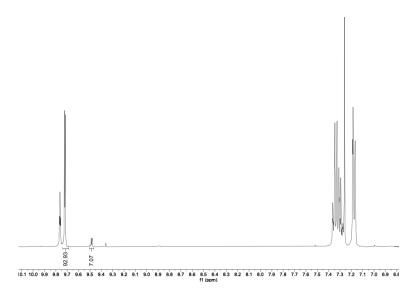


S76

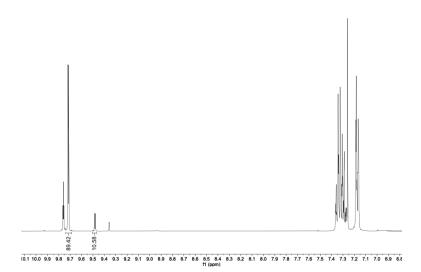
Diet Rivella (S24):





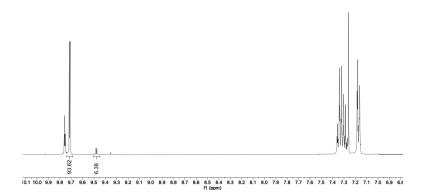


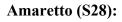


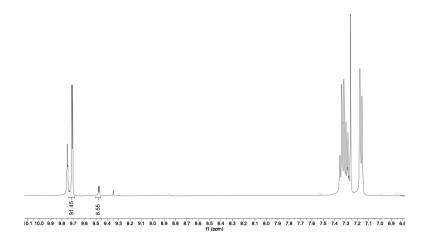


S77

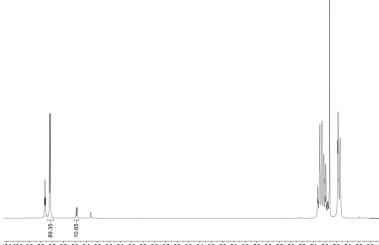
Red wine (S27):





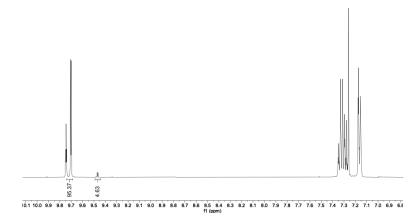


Herb Liqueur – "Jägermeister" (829):

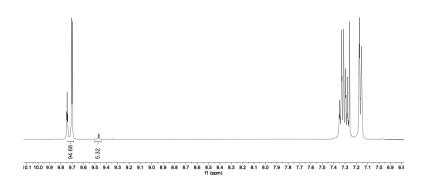


10.1 10.0 9.9 9.8 9.7 9.6 9.5 9.4 9.3 9.2 9.1 9.0 8.9 8.8 8.7 8.6 8.5 8.4 8.3 8.2 8.1 8.0 7.9 7.8 7.7 7.6 7.5 7.4 7.3 7.2 7.1 7.0 6.9 6.8 ff (porm)

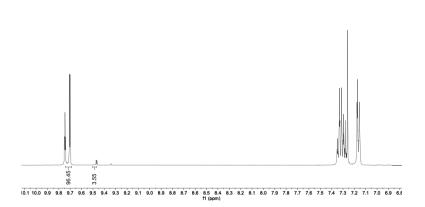
Scotch Whisky – Aberlour 18 y (S30):



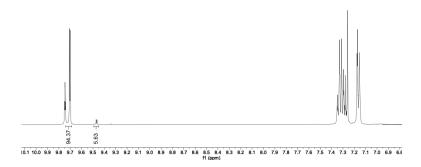




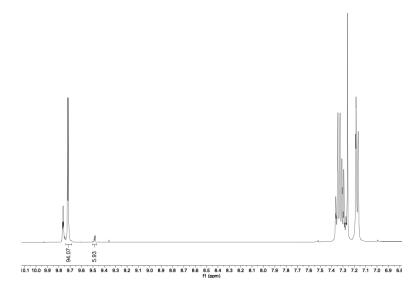




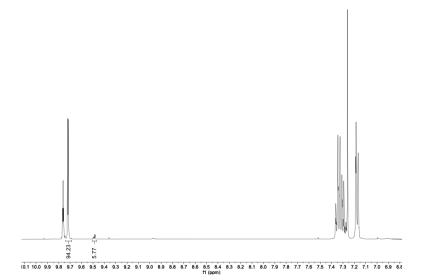
Rum (S33):





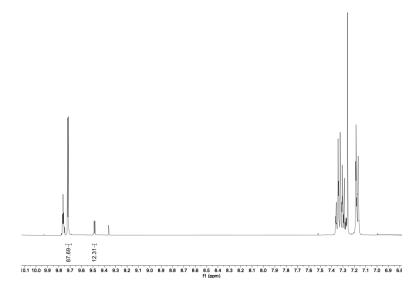


Herb vinegar (S35):



S80

Olive oil (S36):

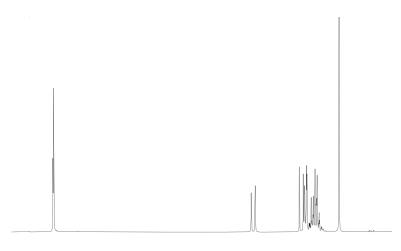


Control Experiments (Reaction without Catalyst in complex mixtures):

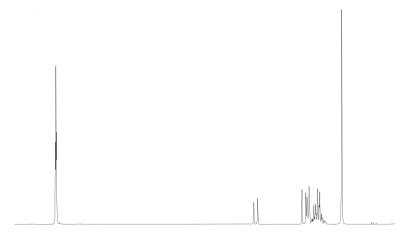
MilliQ water (S1):

See ref 1.

Tab water (S2):

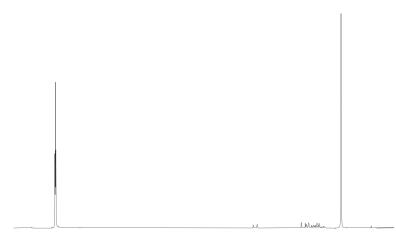


Lake Zürich water (S3):



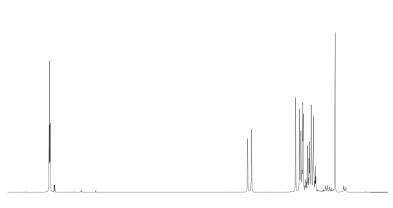
0.1 10.0 9.9 9.8 9.7 9.8 9.5 9.4 9.3 9.2 9.1 9.0 8.9 8.8 8.7 8.6 8.5 8.4 8.3 8.2 8.1 8.0 7.9 7.8 7.7 7.8 7.5 7.4 7.3 7.2 7.1 7.0 6.9 6. ff (pm)

River Limmat water (S4):

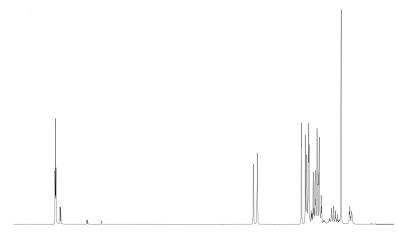


10.1 10.0 9.9 9.8 9.7 9.6 9.5 9.4 9.3 9.2 9.1 9.0 8.9 8.8 8.7 6.8 8.5 8.4 8.3 8.2 8.1 8.0 7.9 7.8 7.7 7.6 7.5 7.4 7.3 7.2 7.1 7.0 6.9 6. ft (ppm)

Milk (S5):

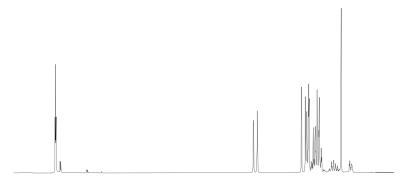


Low fat milk (S6):



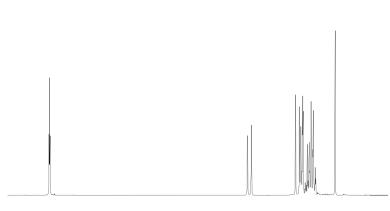
10.1 10.0 9.9 9.8 9.7 9.8 9.5 9.4 9.3 9.2 9.1 9.0 8.9 8.8 8.7 8.8 9.5 8.4 8.3 8.2 8.1 8.0 7.9 7.8 7.7 7.8 7.5 7.4 7.3 7.2 7.1 7.0 6.9 6. If (gen)



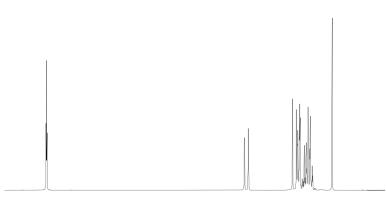


10.1 10.0 9.9 9.8 9.7 9.6 9.5 9.4 9.3 9.2 9.1 9.0 8.9 8.8 8.7 6.8 8.5 8.4 8.3 8.2 8.1 8.0 7.9 7.8 7.7 7.6 7.5 7.4 7.3 7.2 7.1 7.0 6.9 6. ft (ppm)

Almond milk (S8):

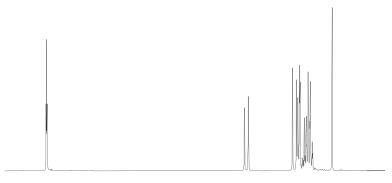


Rice milk (S9):



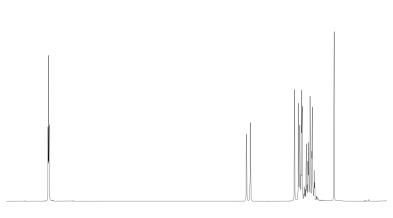
10.1 10.0 9.9 9.8 9.7 9.6 9.5 9.4 9.3 9.2 9.1 9.0 8.9 8.8 8.7 8.6 8.5 8.4 8.3 8.2 8.1 8.0 7.9 7.8 7.7 7.6 7.5 7.4 7.3 7.2 7.1 7.0 6.9 6. ft(ppm)

Orange juice (S10):

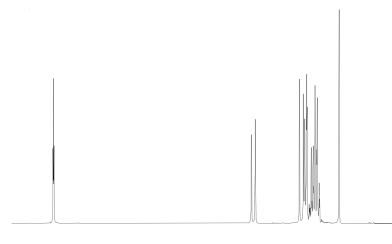


10.1 10.0 9.9 9.8 9.7 9.6 9.5 9.4 9.3 9.2 9.1 9.0 8.9 8.8 8.7 6.8 8.5 8.4 8.3 8.2 8.1 8.0 7.9 7.8 7.7 7.6 7.5 7.4 7.3 7.2 7.1 7.0 6.9 6. ft (ppm)

Blood orange juice (S11):

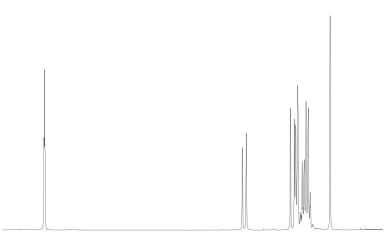


Apple juice (S12):



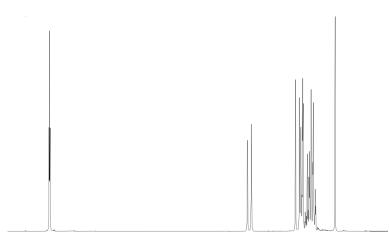
10.1 10.0 9.9 9.8 9.7 9.8 9.5 9.4 9.3 9.2 9.1 9.0 8.9 8.8 8.7 8.6 8.5 8.4 8.3 8.2 8.1 8.0 7.9 7.8 7.7 7.6 7.5 7.4 7.3 7.2 7.1 7.0 6.9 6. ft (pm)

Coconut water (S13):

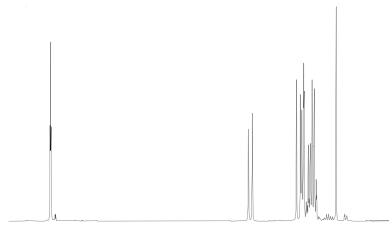


10.1 10.0 9.9 9.8 9.7 9.6 9.5 9.4 9.3 9.2 9.1 9.0 8.9 8.8 8.7 6.8 8.5 8.4 8.3 8.2 8.1 8.0 7.9 7.8 7.7 7.6 7.5 7.4 7.3 7.2 7.1 7.0 6.9 6. ft (ppm)

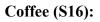
Carrot juice (S14):

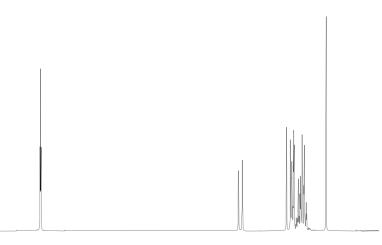


Beetroot juice (S15):



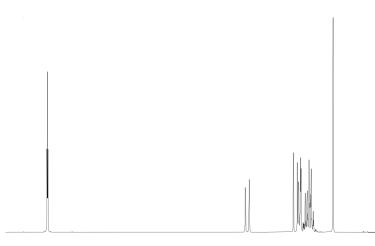
10.1 10.0 9.9 9.8 9.7 9.6 9.5 9.4 9.3 9.2 9.1 9.0 8.9 8.8 8.7 8.6 8.5 8.4 8.3 8.2 8.1 8.0 7.9 7.8 7.7 7.6 7.5 7.4 7.3 7.2 7.1 7.0 6.9 6. ft(ppm)



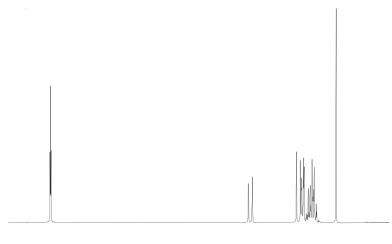


10.1 10.0 9.9 9.8 9.7 9.6 9.5 9.4 9.3 9.2 9.1 9.0 8.9 8.8 8.7 8.8 8.5 8.4 8.3 8.2 8.1 8.0 7.9 7.8 7.7 7.6 7.5 7.4 7.3 7.2 7.1 7.0 6.9 6. ft (gem)

Green tea (S17):

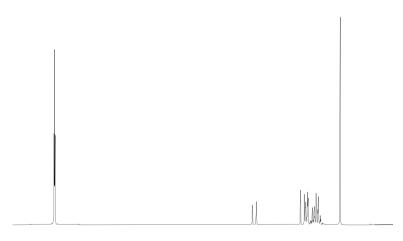


Black tea (S18):



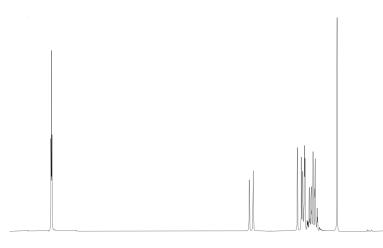
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Peppermint tea (S19):

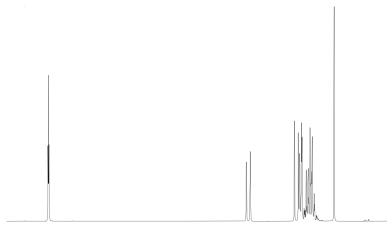


10.1 10.0 9.9 9.8 9.7 9.6 9.5 9.4 9.3 9.2 9.1 9.0 6.9 8.8 6.7 8.8 8.5 8.4 8.3 6.2 8.1 8.0 7.9 7.8 7.7 7.6 7.5 7.4 7.3 7.2 7.1 7.0 6.9 6. ff (ppm)

Fruit tea (S20):

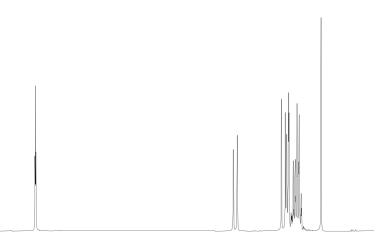


Honey solution 5 wt% (S21):



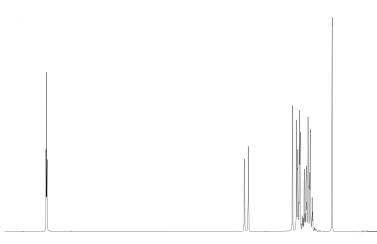
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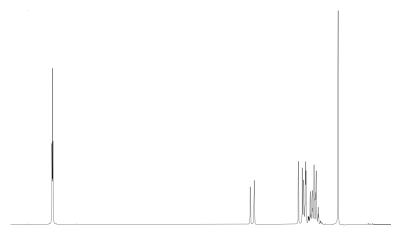


10.1 10.0 9.9 9.8 9.7 9.6 9.5 9.4 9.3 9.2 9.1 9.0 8.9 8.8 8.7 8.8 8.5 8.4 8.3 8.2 8.1 8.0 7.9 7.8 7.7 7.6 7.5 7.4 7.3 7.2 7.1 7.0 6.9 6. ft (gem)

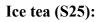
Rivella (S23):

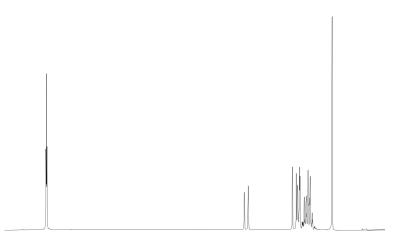


Diet Rivella (S24):



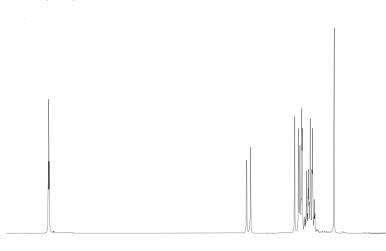
10.1 10.0 9.9 9.8 9.7 9.6 9.5 9.4 9.3 9.2 9.1 9.0 8.9 8.8 8.7 8.6 8.5 8.4 8.3 8.2 8.1 8.0 7.9 7.8 7.7 7.6 7.5 7.4 7.3 7.2 7.1 7.0 6.9 6. ft(ppm)



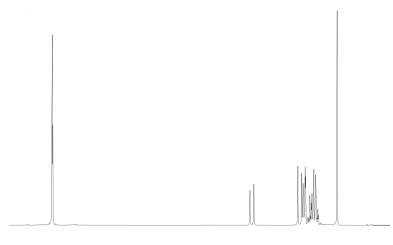


10.1 10.0 9.9 9.8 9.7 9.6 9.5 9.4 9.3 9.2 9.1 9.0 8.9 8.8 8.7 6.8 8.5 8.4 8.3 8.2 8.1 8.0 7.9 7.8 7.7 7.6 7.5 7.4 7.3 7.2 7.1 7.0 6.9 6. ft (ppm)

Beer (S26):



Red wine (S27):



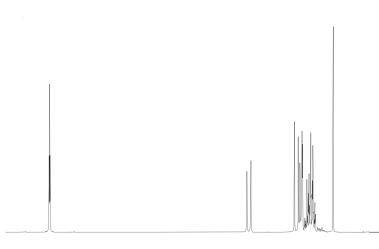
10.1 10.0 9.9 9.8 9.7 9.6 9.5 9.4 9.3 9.2 9.1 9.0 8.9 8.8 8.7 8.6 8.5 8.4 8.3 8.2 8.1 8.0 7.9 7.8 7.7 7.6 7.5 7.4 7.3 7.2 7.1 7.0 6.9 6. ft(ppm)

Amaretto (S28):

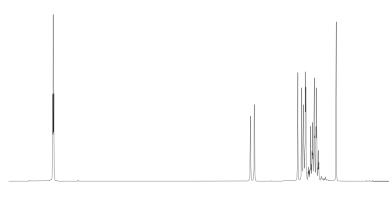


10.1 10.0 9.9 9.8 9.7 9.6 9.5 9.4 9.3 9.2 9.1 9.0 8.9 8.8 8.7 6.8 8.5 8.4 8.3 8.2 8.1 8.0 7.9 7.8 7.7 7.6 7.5 7.4 7.3 7.2 7.1 7.0 6.9 6. ft (ppm)

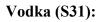
Herb Liqueur – "Jägermeister" (829):

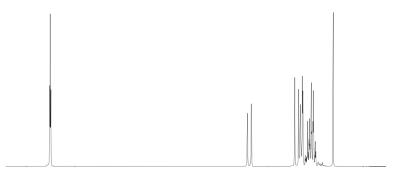


Scotch Whisky – Aberlour 18 y (830):



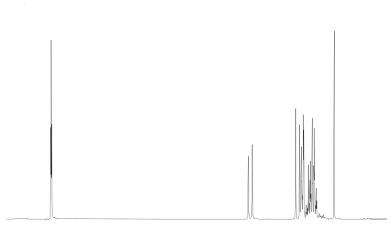
10.1 10.0 9.9 9.8 9.7 9.6 9.5 9.4 9.3 9.2 9.1 9.0 8.9 8.8 8.7 8.6 8.5 8.4 8.3 8.2 8.1 8.0 7.9 7.8 7.7 7.6 7.5 7.4 7.3 7.2 7.1 7.0 6.9 6. ft(ppm)



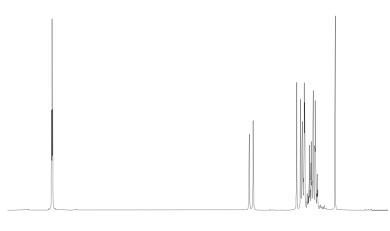


10.1 10.0 9.9 9.8 9.7 9.6 9.5 9.4 9.3 9.2 9.1 9.0 8.9 8.8 8.7 6.8 8.5 8.4 8.3 8.2 8.1 8.0 7.9 7.8 7.7 7.6 7.5 7.4 7.3 7.2 7.1 7.0 6.9 6. ft (ppm)

Gin (S32):

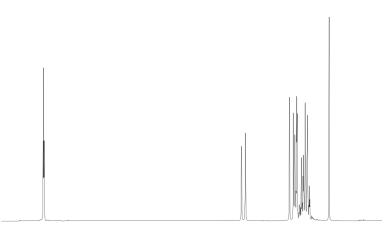


Rum (S33):



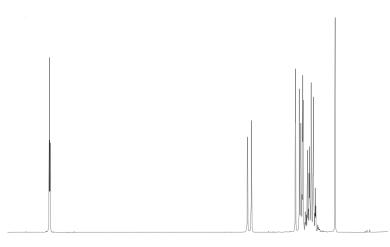
10.1 10.0 9.9 9.8 9.7 9.6 9.5 9.4 9.3 9.2 9.1 9.0 8.9 8.8 8.7 8.6 8.5 8.4 8.3 8.2 8.1 8.0 7.9 7.8 7.7 7.6 7.5 7.4 7.3 7.2 7.1 7.0 6.9 6. ft(ppm)

Aceto balsamico (S34):

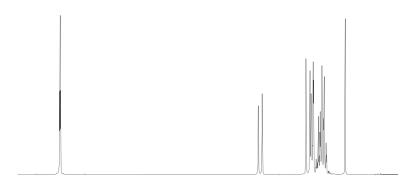


10.1 10.0 9.9 9.8 9.7 9.6 9.5 9.4 9.3 9.2 9.1 9.0 8.9 8.8 8.7 6.8 8.5 8.4 8.3 8.2 8.1 8.0 7.9 7.8 7.7 7.6 7.5 7.4 7.3 7.2 7.1 7.0 6.9 6. ft (ppm)

Herb vinegar (S35):

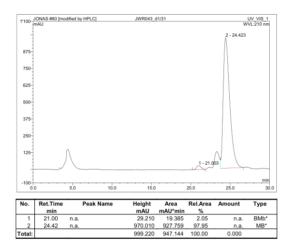


Olive oil (S36):

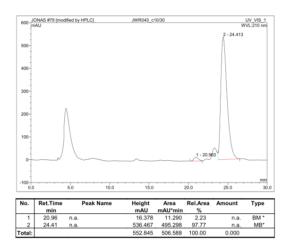


HPLC chromatograms of the reactions in the presence of H-DPro-MePro-Glu-NH₂ 2:

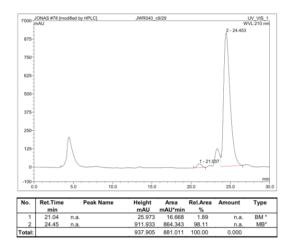
MilliQ water (S1):



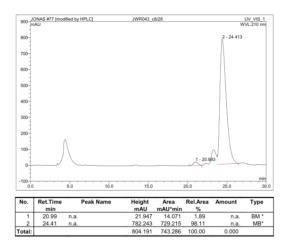
Tab water (S2):



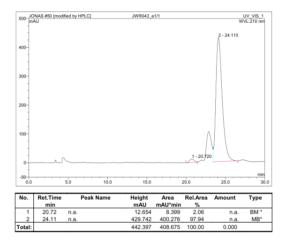
Lake Zürich water (S3):



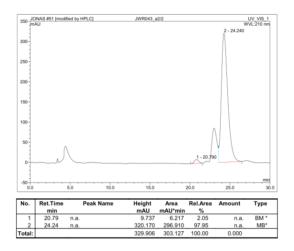
River Limmat water (S4):



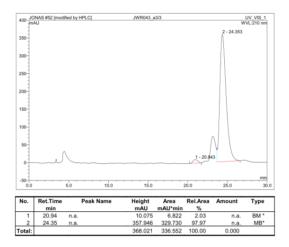
Milk (S5):



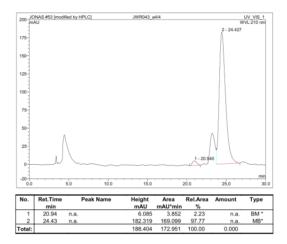
Low fat milk (S6):



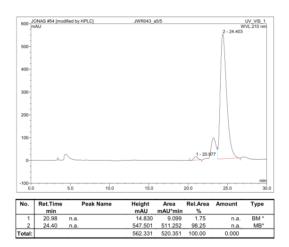
Soya milk (S7):



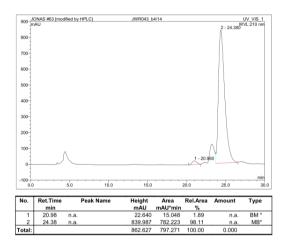
Almond milk (S8):



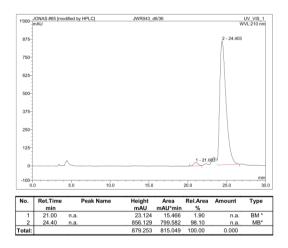
Rice milk (S9):



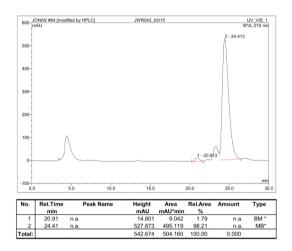
Orange juice (S10):



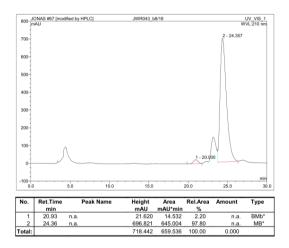
Blood orange juice (S11):



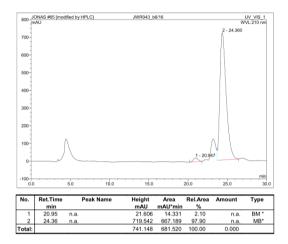
Apple juice (S12):



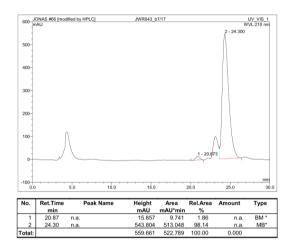
Coconut water (S13):



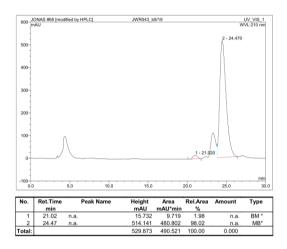
Carrot juice (S14):



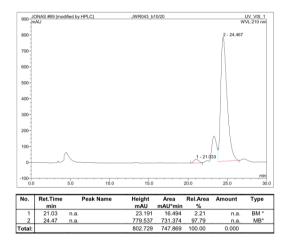
Beetroot juice (S15):



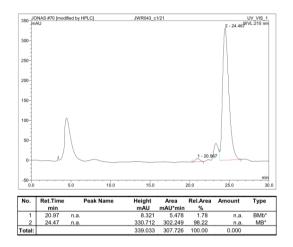
Coffee (S16):



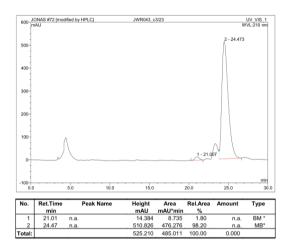
Green tea (S17):



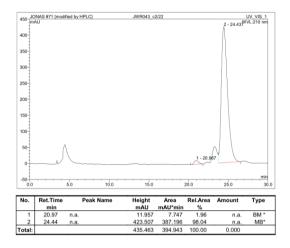
Black tea (S18):



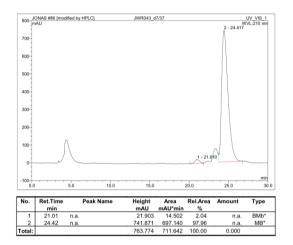
Peppermint tea (S19):



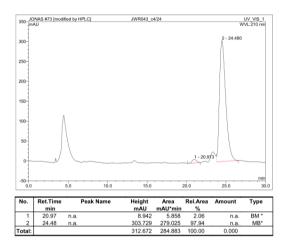
Fruit tea (S20):



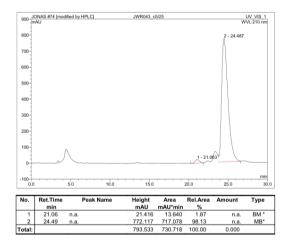
Honey solution 5 wt% (S21):



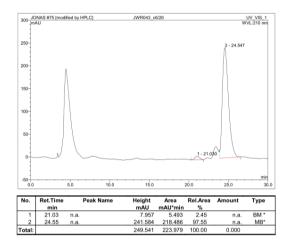
Coke (S22):



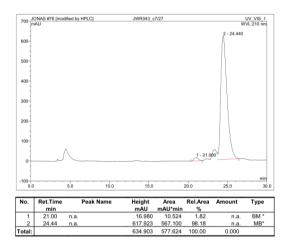
Rivella (S23):



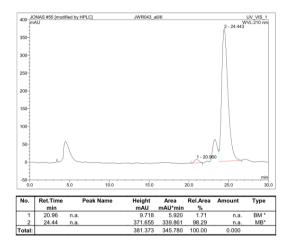
Diet Rivella (S24):



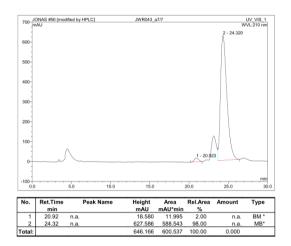
Ice tea (S25):



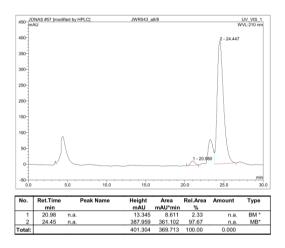
Beer (S26):



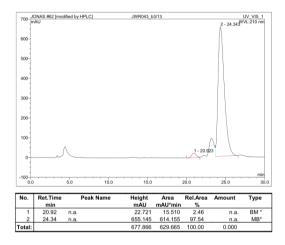
Red wine (S27):



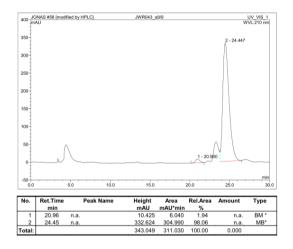
Amaretto (S28):



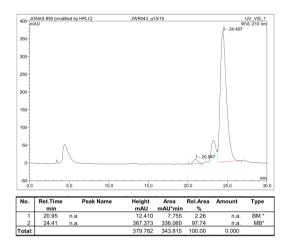
Herb Liqueur – "Jägermeister" (829):



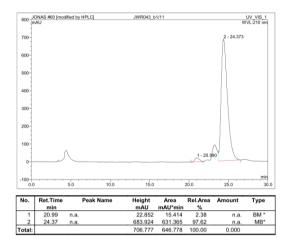
Scotch Whisky – Aberlour 18 y (S30):



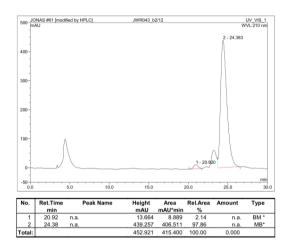
Vodka (S31):



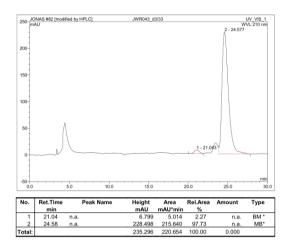
Gin (S32):



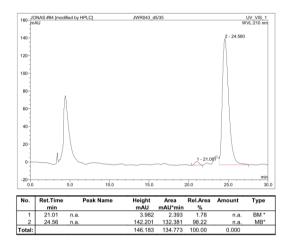
Rum (S33):



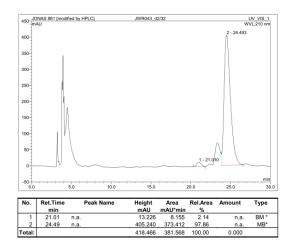
Aceto balsamico (S34):



Herb vinegar (S35):



Olive oil (S36):



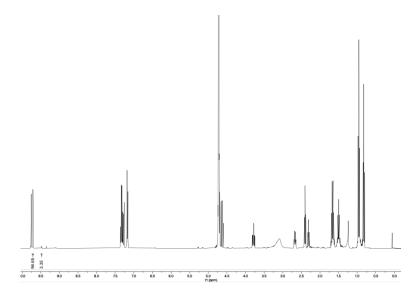
8. Conjugate Addition Reaction in Cell Lysate:

Preparation of the cell lysate:

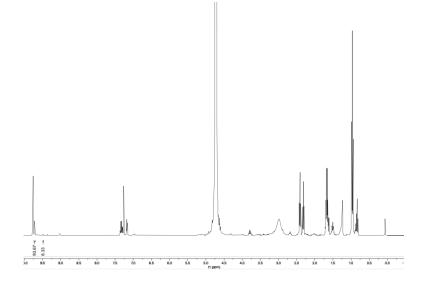
Hep G2 cells were seeded in a *TPP*® *Tissue Culture Flask 300* culture plate and grown to approximately 80% confluence in a cell culture incubator at 37 °C and 5% CO₂. The cells were washed three times with phosphate-buffered saline (PBS), then harvested off the bottom of the flask by cell scrapers into PBS buffer. The solution was transferred into conical *Falcon*® tubes. The collected cells were sonicated using a *Branson SFX 250 Sonifier*® while kept in an ice bath through the whole process. The cell lysate protein concentration was quantified at $\lambda = 280$ nm using a *NanoDrop*TM spectrophotometer.

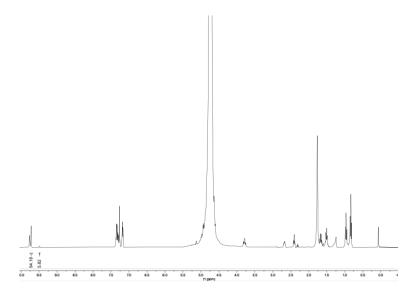
¹H NMR spectra:

Reaction with H-DPro-Pro-Glu-NH₂ 1 (440 mM nitrostyrene)

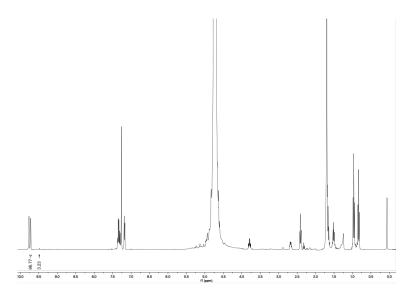


Reaction with H-DPro-MePro-Glu-NH₂ 2 (440 mM nitrostyrene)

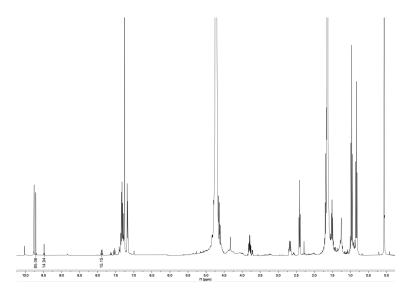




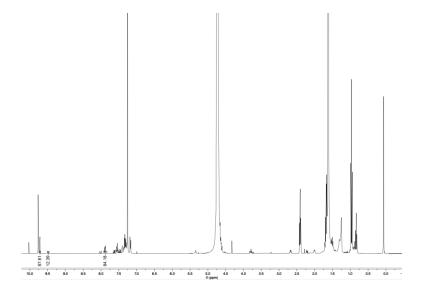
Reaction with H-DPro-MePro-Glu-NH₂ 2 (44 mM nitrostyrene)



Reaction with H-DPro-Pro-Glu-NH₂ 1 (4 mM nitrostyrene)

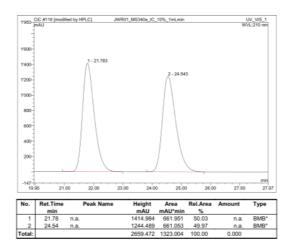


Reaction with H-DPro-MePro-Glu-NH₂ 2 (4 mM nitrostyrene)

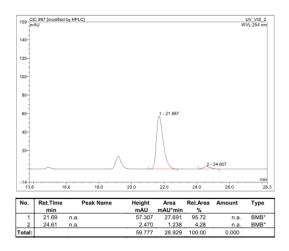


HPLC chromatograms:

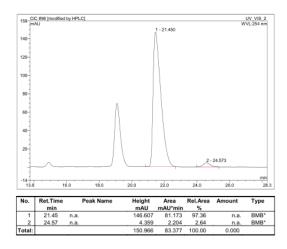
Racemic sample:



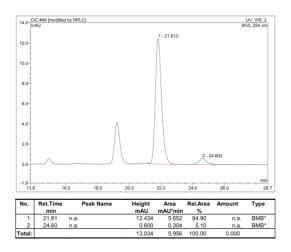
Reaction with H-DPro-Pro-Glu-NH₂ 1 (440 mM nitrostyrene)



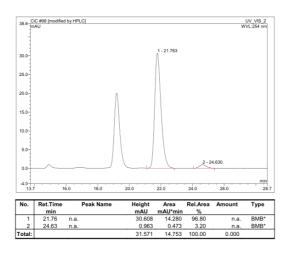
*Reaction with H-DPro-MePro-Glu-NH*₂ 2 (440 mM nitrostyrene)



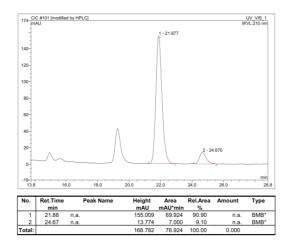
Reaction with H-DPro-Pro-Glu-NH₂ 1 (44 mM nitrostyrene)



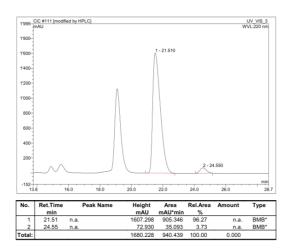
*Reaction with H-DPro-MePro-Glu-NH*₂ 2 (44 mM nitrostyrene)



Reaction with H-DPro-Pro-Glu-NH₂ 1 (4 mM nitrostyrene)



Reaction with H-DPro-MePro-Glu-NH₂ 2 (4 mM nitrostyrene)



9. References

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