

Supplementary Materials for

Electrochemical Oxidative Rearrangement of Tetrahydro- β -carbolines in Zero-gap Flow Cell

Yiting Zheng^{#[a][c]}, Yuen Tsz Cheung^{#[b]}, Lixin Liang^[b], Huiying Qiu^[b], Lei Zhang^[c], Anson Tsang^[a],
Qing Chen^{*[a][b]} & Rongbiao Tong^{*[b]}

*Corresponding author. Email: chenqing@ust.hk; rtong@ust.hk;

Index of Supplementary Materials

| | |
|--|----|
| General Information..... | 2 |
| 1. Faradaic Efficiency and Productivity..... | 4 |
| 2. Flow cell set-up..... | 4 |
| 3. Other conditions screened..... | 6 |
| 4. Electro-chemical test (undivided cell)..... | 8 |
| 5. Electro-chemical test (flow cell)..... | 9 |
| 6. Comparison with literature data..... | 9 |
| 7. Preparation of indole substrates..... | 13 |
| 8. Electrochemical oxidative rearrangement of indoles to 2-oxindoles..... | 16 |
| 9. Mechanistic study..... | 22 |
| 10. References..... | 26 |
| 11. Copies of ¹ H- and ¹³ C-NMR spectra (S28-S85)..... | 28 |

General Information

All the electrochemical experiments and synthesis were carried on electrochemical workstation VMP-300 (as Figure S1A present). Reactions in a flow cell were carried on the two PEEK housing zero-gap membrane reactor with PTFE gasket and celgard 3501, the detail information could be found in section Flow cell set-up in this file. The inlet and outlet injectors were connected to PEEK housing with SMC stainless steel air-tight connector and PTFE tube. The reaction solutions were sealed in bottle with holes, as Figure S1B present, and bumped through the PTFE tube connected with PharMed BPT tube by LongerPump BT-600-2J with YZ1515x in 5-20 mL/min.

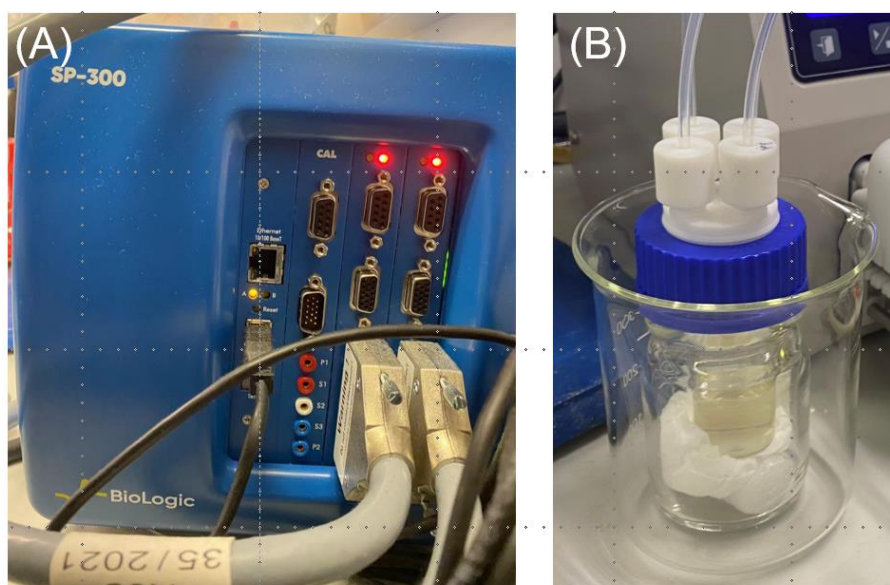


Figure S1. (A) Electrochemical workstation VMP-300; (B) Sealed bottle with holes connect with PTFE tubes connect with zero-gap flow cell.

Solvents used in workup, extraction and column chromatography were used as received from commercial suppliers without prior purification. Reactions for substrate preparation were magnetically stirred and monitored by thin layer chromatography (TLC, 0.25 mm) on pre-coated silica gel plates. Flash chromatography was performed with silica gel 60 (particle size 0.040 – 0.062 mm). ^1H and ^{13}C NMR spectra were recorded on a Bruker AV-400 spectrometer (400 MHz for ^1H , 100 MHz for ^{13}C). Chemical shifts are reported in parts per million (ppm) as values relative to the internal chloroform (7.26 ppm for ^1H NMR and 77.16 ppm for ^{13}C NMR). Abbreviations for signal coupling are as follows: s, singlet; d, doublet; t, triplet; q, quartet; m, multiplet; br, broad. High resolution mass spectra were measured at the Hong Kong University of Science and Technology Mass Spectrometry Service

Center on an Agilent GC/MS. Carbon paper (CP, SGL-39AA) and organic deposition on CP after the synthesis mediated by KBr were characterized on the scanning electron microscopy (SEM) (JEOL-7100, 10 kV).

General Procedure A (using flow cell, Figure S2A) of Electrochemical oxidative rearrangement of indoles to 2-oxindoles: Reactions were conducted in two 20 mL vials with anolyte and catholyte. The assemble and detail information were discussed in **Flow cell setup**. The reaction was performed in the two PEEK housing flow cell at constant voltage 1.2 V using potentiostatic method in VMP-300. Anolyte solution of **1a** (0.2 mmol) in MeCN/AcOH/H₂O (15:2.4:2, 10 mL) was added bromide salt (0.4 mmol, 2.0 equiv.). The anolyte was separated from the cathode electrolyte in H₂SO₄ (0.25M, 10 mL) by a proton-exchange membrane Nafion 117. The 20 mL/min was set for synthesis, and the cut-off charge was set 2 F/mol. Reaction was monitored by TLC. The reaction mixture was diluted with EtOAc (10 mL). Saturated aqueous Na₂CO₃ solution (10 mL) was added. The organic fraction was collected, and the aqueous fraction was extracted with EtOAc (2 x 10 mL). The combined organic fraction was washed with brine, dried over Na₂SO₄, filtered and concentrated under reduced pressure. The resulting residue was purified by flash column chromatography on silica gel using hexane/ethyl acetate eluents.

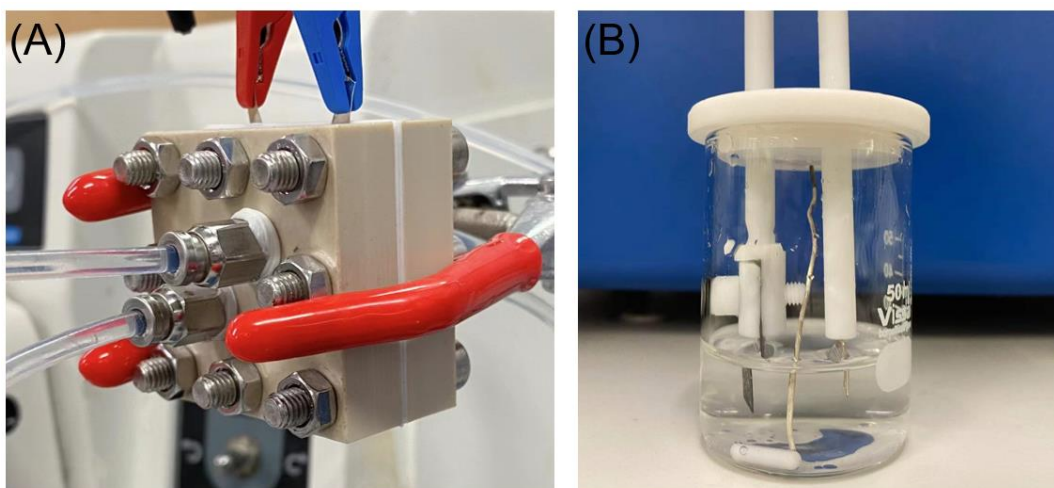
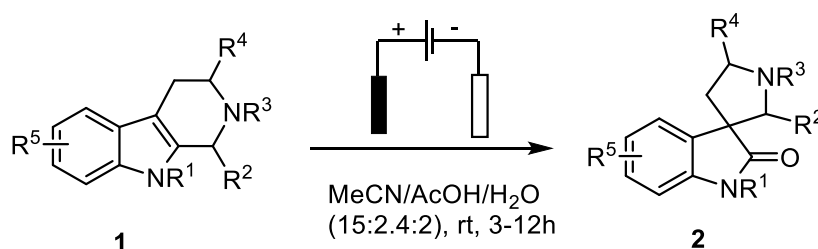


Figure S2. (A) Flow cell using in General Procedure A; (B) Set-up of undivided cell using in General Procedure B

General Procedure B (using undivided cell, Figure S2B) of Electrochemical oxidative rearrangement of indoles to 2-oxindoles: Reactions were conducted in a 20 mL vial with a stir bar and a carbon paper (10 mm*10 mm*0.6 mm) working electrode (anode), a platinum-plated (10 mm*10 mm*0.1 mm) counter-electrode (cathode) and Ag /AgBr reference electrode. Constant voltage mode with 2.0 V was applied by VMP-300. The Ag/AgBr electrode was made by Ag wire and HBr, the potential was calibrated with SCE and convert to -0.26 V vs. RHE. To a solution of **1** (0.1 mmol) in MeCN/AcOH/H₂O (15:2.4:2, 10 mL) was added KBr (0.05 mmol, 0.5 equiv.). The resistance in the set-up was measured 122 ohm by VMP-300. Reaction was monitored by TLC and stopped after complete conversion of **1a**. The reaction mixture was diluted with EtOAc (10 mL). Saturated Na₂CO₃ solution (10 mL) was added. The organic fraction was collected, and the aqueous fraction was extracted with EtOAc (2 x 10 mL). The combined organic fraction was washed with brine, dried over Na₂SO₄, filtered and concentrated under reduced pressure. The resulting residue was purified by flash column chromatography on silica gel using hexane/ethyl acetate eluents.

1. Faradaic Efficiency and Productivity

The faradaic efficiency (FE) of product formation and productivity were calculated using the following equation^[1,2]:

$$\text{Faradaic efficiency}(\%) = \frac{n \times F \times \text{mole of product formed}}{\text{total charge passed}} \times 100\% \quad \text{Equation (1)}$$

$$\text{productivity}(\mu\text{mol}/(\text{h} \times \text{cm}^2)) = \frac{\text{mole of product formed}}{\text{total time} \times \text{electrode area}} \quad \text{Equation (2)}$$

where n is the number of electron transfer for each product formation and F is the Faraday constant (96485 C mol⁻¹). Total time is the time achieved 100% conversion of substrate. Electrode area is the anode geometric area.

2. Flow cell set-up

2.1 Materials for zero-gap flow cell set-up using in General Procedure A:

2 x PEEK endplates (40 mm*40 mm*10 mm) with snake shape of flow channel, as Figure S3A&B shown

4 x PTFE Gasket (40 mm*40 mm*0.3 mm) with a hollow part (10 mm*10 mm) in the middle, as Figure S3C shown

2 x Celgard 3501 (12 mm* 12 mm), pretreatment with MeCN

1 x Proton-exchange membrane Nafion[®] 117 (12 mm* 12 mm, pre-treatment with MeCN)

Anode: 2 x Carbon papers (CP, SGL 39AA) (10 mm*10 mm*0.3 mm), heat treatment in 400 °C in air, 24 h

Cathode: 1 x 20% Pt/graphite carbon paper (10 mm*10 mm*0.3 mm), prepared by dripping 20% Pt/graphite (purchase from Sigma-Aldrich) Nafion[®] solution (5 wt% Nafion amount) on heat treated CP

Cuurent Collector: 2 x Platinum foil (3 mm*25 mm*0.1 mm)

8 x Screws and Screw caps

4 x SMC stainless steel air-tight connectors (tube: 1/8 to M5)

3 x PTFE tube (1/8)

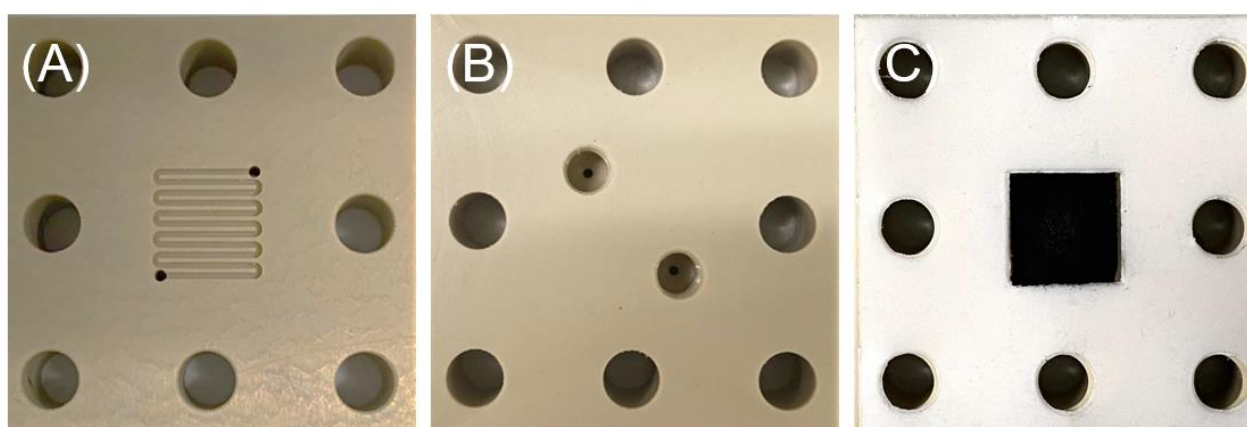


Figure S3. (A) PEEK endplates with flow channels, contact with PTFE gasket and electrode; (B) PEEK endplates with M5 holes connect with SMC air-tight connectors; (C) PTFE gasket with holes and CP in the hole.

2.2 Procedure for flow cell assembly:

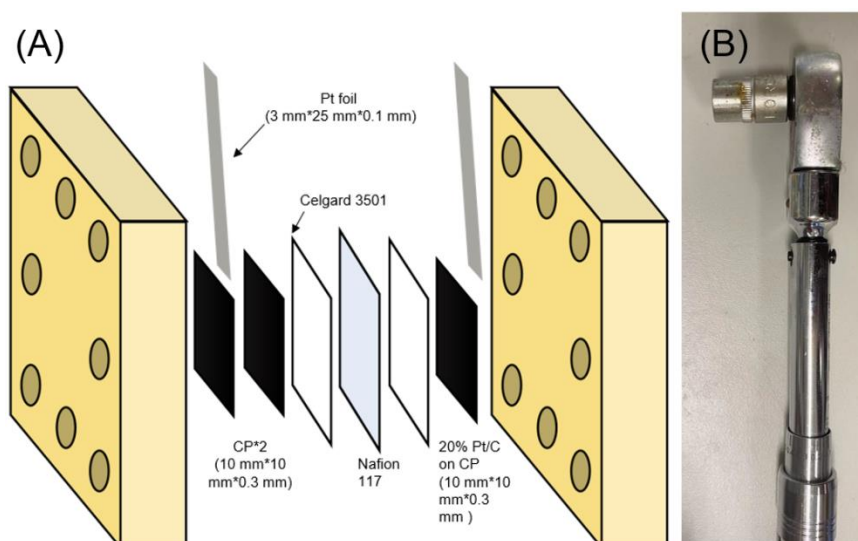
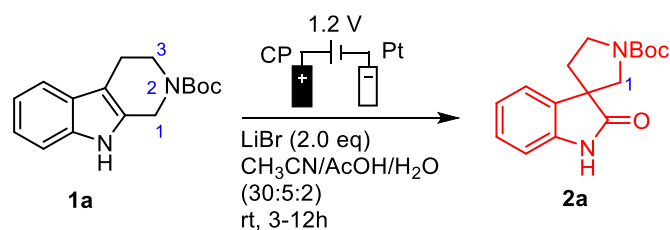


Figure S4. (A) Schematic of flow cell using in General Procedure A; (B) Torque wrench using to tight screw

A PEEK endplate with flow channel facing top was equipped with eight screws. The assemble schematic was presented in Figure S4A. A piece of PTFE gasket was placed on top of the endplate. Two pieces of carbon papers in the middle were fitted in the hollow part of the gasket. Current collector was placed beyond the edge of the set-up. A piece of celgard 3501 was placed on top covering the whole carbon paper electrode. Then additional two pieces of PTFE Gasket with a proton-exchange membrane Nafion 117 in the middle were placed on top. Another piece of celgard 3501 was placed to cover the hollow part. The 20% Pt/graphite CP and current collector were then placed, covered by the last PTFE Gasket and another PEEK endplates and tightened the screws with screw caps using torque wrench, as Figure S4B presented, with uniform force to prevent the deformation and leakage of using PTFE gasket. Finally, four SMC stainless steel air-tight connectors were screwed on two sides of the flow cell with Teflon tape to prevent corrosion and connected with four PTFE tube at the other end.

3. Other conditions screened

Table S1. Selected conditions for Br mediated electro-oxidative rearrangement^[a]



| Entry | Different from base case ^a | Current density (mA*cm ⁻²) | Conv. (%) | Yield. (%) | Faradaic Efficiency (%) | Productivity (μmol*h ⁻¹ *cm ⁻²) |
|-------|---------------------------------------|--|-----------|------------|-------------------------|--|
| S1 | NaBr 40 mL*min ⁻¹ | 5.2 | 100 | 82 | 82 | 78.7 |
| S2 | KBr 40 mL*min ⁻¹ | 3.5 | 100 | 75 | 75 | 48.5 |
| S3 | TBAB 40 mL*min ⁻¹ | 1.4 | 100 | 58 | 58 | 14.7 |
| S4 | LiBr 5 eq | 7.0 | 100 | 87 | 87 | 113.6 |
| S5 | LiBr 2.5 eq | 4.4 | 100 | 95 | 95 | 82.2 |
| S6 | LiBr 1.5 eq | 2.0 | 100 | 73 | 73 | 37.2 |
| S7 | 40 mL*min ⁻¹ | 6.4 | 100 | 96 | 96 | 114.7 |
| S8 | 5 mL*min ⁻¹ | 2.0 | 100 | 89 | 89 | 33.2 |

^[a] The reaction was carried out at room temperature with **1a** (0.2 mmol), LiBr (2.0 eq), solvents (20 mL) followed general procedure A. NMR yield was obtained

4. Electro-chemical test (undivided cell)

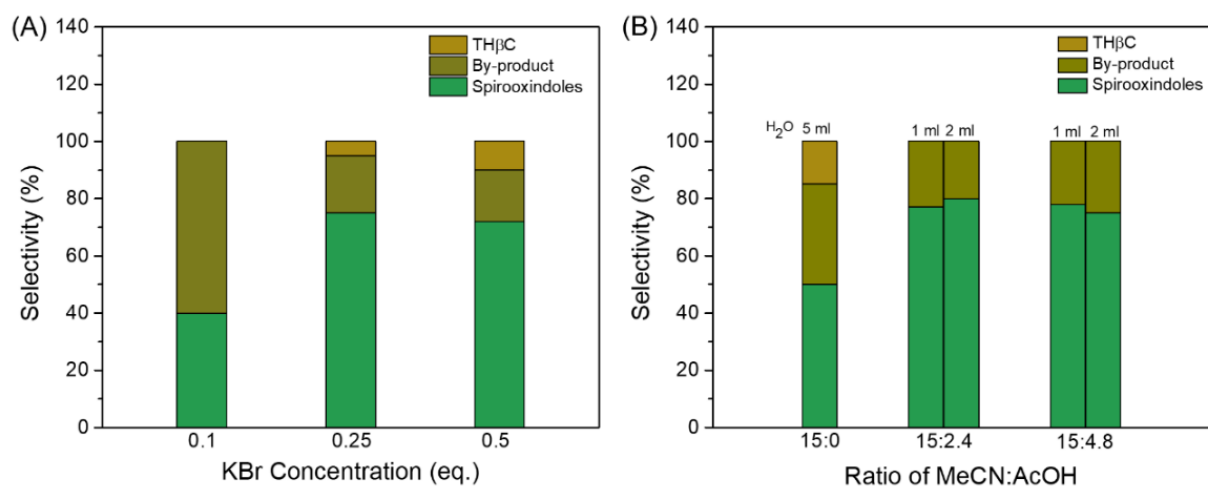


Figure S5. (A) Influence of KBr concentration in General Procedure B; (B) Influence of AcOH addition in General Procedure B

5. Electro-chemical test (flow cell)

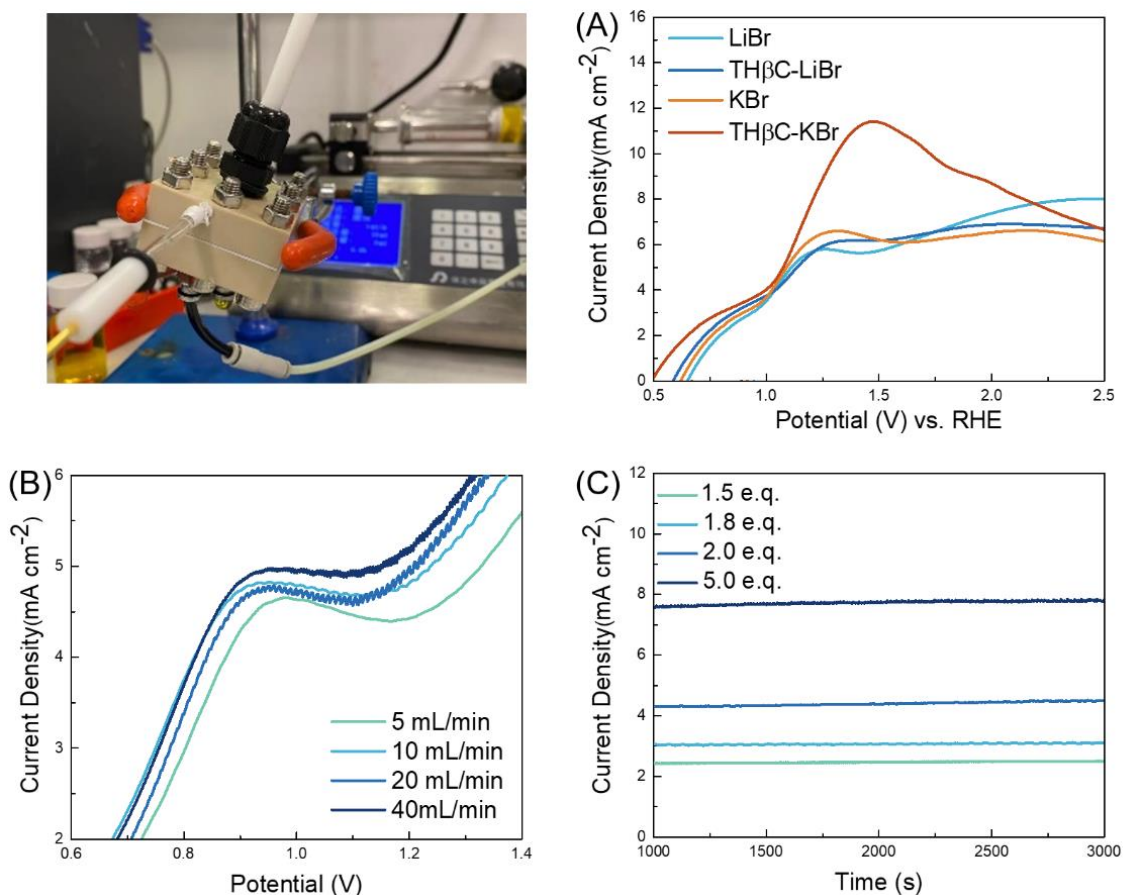


Figure S6. (A) LSV of half-flow cell (as photo present) with condition: Working electrode: CP; Counter electrode: Pt mesh; Reference electrode: Ag/AgCl, the $E_{RHE} = E + 0.0591 \cdot \text{pH} + 0.197$ ^[3]; Scan rate 50 mV/s with 0.8 M LiClO₄ as supporting electrolyte, 0.2 mmol THβC, 2 e.q. LiBr or KBr; (B) LSV of different flow rate under General Procedure A. (C) Current density in different equivalent of LiBr under General procedure A.

6. Comparison with literature data

We also compared our results with other electrochemical oxidation reactions, in particular, of indole or related N-heterocycle compounds^[4-9] in Tables S2-3, which including: (a) electrochemical 1,2-diarylation^[4] of alkenes with a CoCl₂ catalyst; (b) electrochemical diazidation^[5] reaction of alkenes with the MnBr₂ catalyst; (c) electro-oxidative [3+2] annulation^[6] of phenol and indole; (d) electrochemical radical cascade cyclization^[7] of N-methacryloyl-2-phenylbenzimidazole and alkyl

boronic acid; (e) electrochemical flow microreactor^[8] for efficient synthesis of isoquinoline-6(5H)-ones; and (f) flow Rhodaelectro-catalyzed alkyne annulations^[9]. The comparison was based on terms of the yield and productivity, i.e., the number of mole of the desired product per unit of reaction time and per area of the cell [in $\mu\text{mol}/(\text{h}\cdot\text{cm}^2)$]. The higher productivity, the shorter time and the smaller cell required, the more cost-effective. As shown in Figure S7, the yield (**2p**, 94%) of our reaction system was among the best, while the productivity (**2p**, $144 \mu\text{mol}/(\text{h}\cdot\text{cm}^2)$) (also see Table S3 in Supporting Information) was much higher than all other electrochemical reactions. We attributed our outstanding performance to the following two factors: 1) the zero-gap flow cell minimizes the overpotential and mitigates the trade-off between a high current and a high selectivity, which is typically encountered in a cell with less forced convection; and 2) the use of bifunctional mediator LiBr with a suitable oxidation potential leaves a sufficiently wide potential window for the high selectivity.

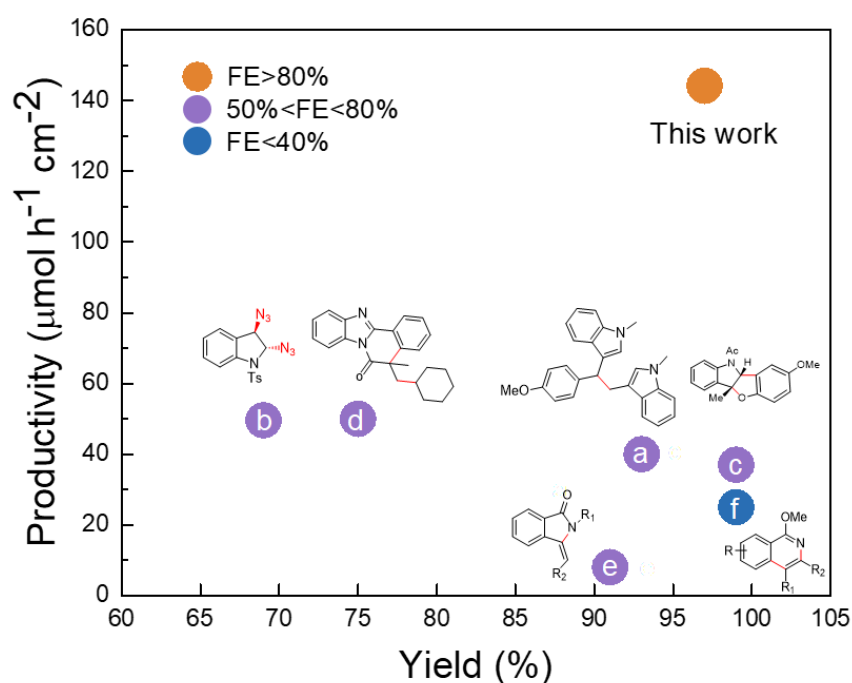


Figure S7. Comparison of productivity, Faradaic efficiencies and yield cycle life (Noted: all selected electro-oxidation of indole or N-heterocyclic compounds for a fair comparison, Table S3 summarizes the detail calculation; red bond represents new generated bond during the electrochemical reaction)^[4-9]

Table S2. Summary of the detailed reactions in Figure S7 in the article.

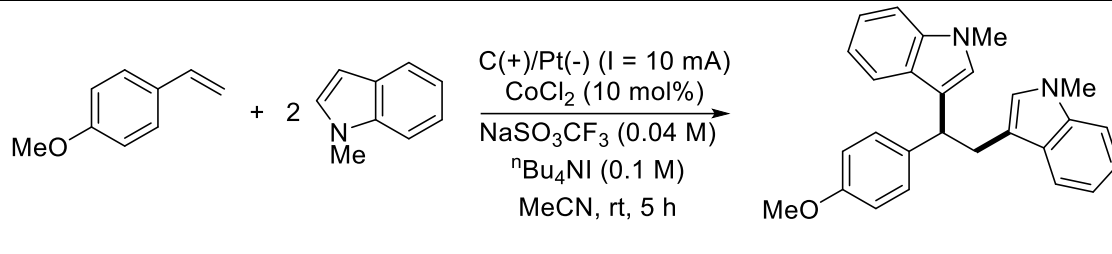
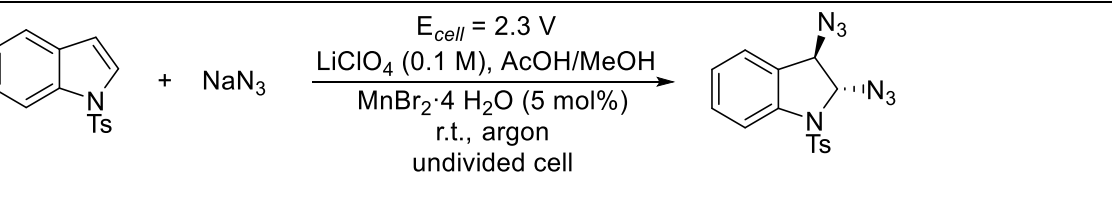
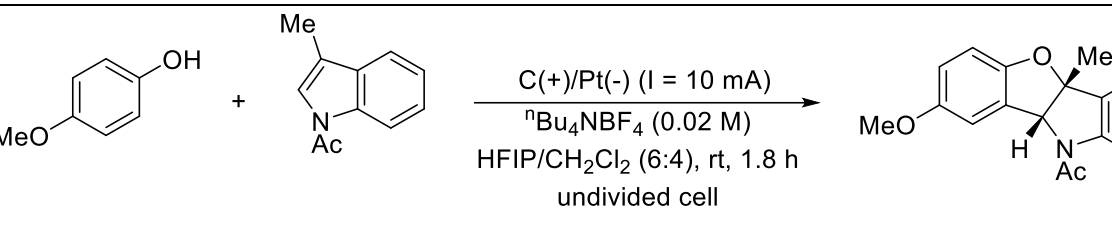
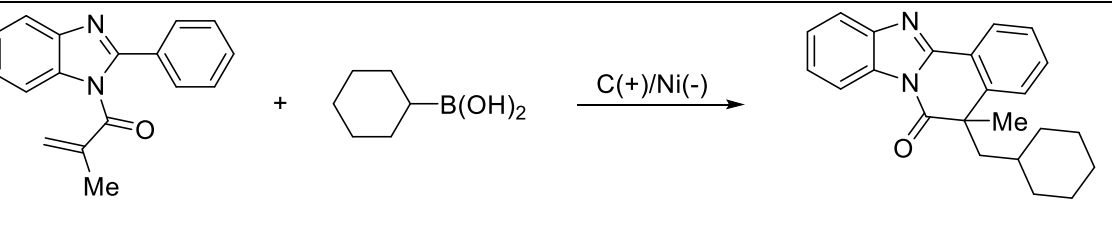
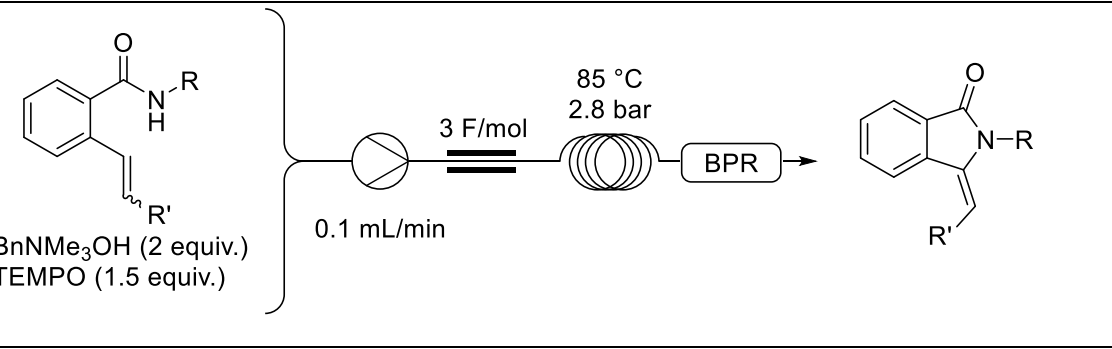
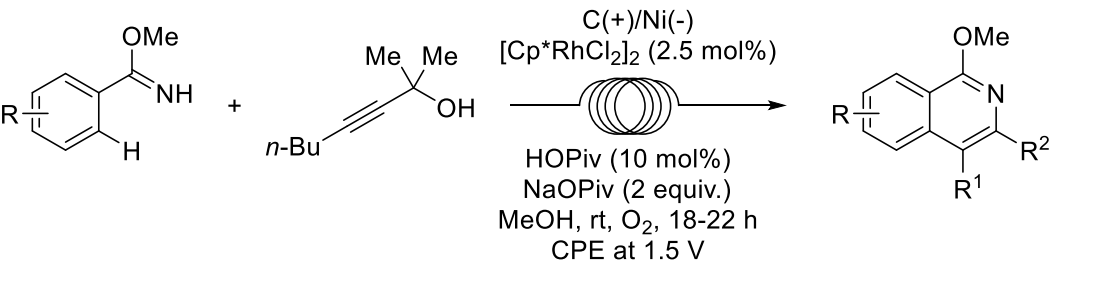
| | |
|------------------|---|
| a ^[4] |  <p> $\text{C}(+)/\text{Pt}(-)$ ($I = 10 \text{ mA}$) CoCl_2 (10 mol%) NaSO_3CF_3 (0.04 M) $n\text{Bu}_4\text{NI}$ (0.1 M) MeCN, rt, 5 h </p> |
| b ^[5] |  <p> $E_{\text{cell}} = 2.3 \text{ V}$ LiClO_4 (0.1 M), AcOH/MeOH $\text{MnBr}_2 \cdot 4 \text{ H}_2\text{O}$ (5 mol%) r.t., argon undivided cell </p> |
| c ^[6] |  <p> $\text{C}(+)/\text{Pt}(-)$ ($I = 10 \text{ mA}$) $n\text{Bu}_4\text{NBF}_4$ (0.02 M) HFIP/CH_2Cl_2 (6:4), rt, 1.8 h undivided cell </p> |
| d ^[7] |  <p> $\text{C}(+)/\text{Ni}(-)$ </p> |
| e ^[8] |  <p> $85 \text{ }^\circ\text{C}$ 2.8 bar 3 F/mol 0.1 mL/min BPR BnNMe_3OH (2 equiv.) TEMPO (1.5 equiv.) </p> |
| f ^[9] |  <p> $\text{C}(+)/\text{Ni}(-)$ $[\text{Cp}^*\text{RhCl}_2]_2$ (2.5 mol%) HOPiv (10 mol%) NaOPiv (2 equiv.) MeOH, rt, O_2, 18-22 h CPE at 1.5 V </p> |

Table S3. Summary of productivity, Faradaic efficiency and yield in Figure S7

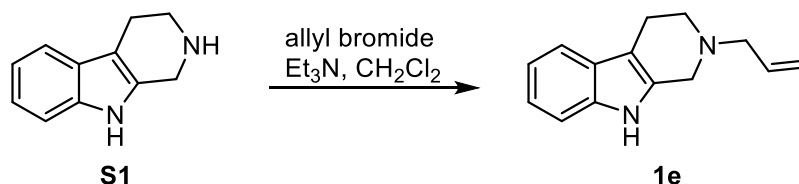
| Entry | Reaction cell | Electrode | Yield. (%) | Faradaic Efficiency (%) | Productivity ($\mu\text{mol}\cdot\text{h}^{-1}\cdot\text{cm}^{-2}$) |
|------------------|----------------------------|--------------------------|---------------|-------------------------------|--|
| Our work | Divided zero-gap flow cell | CP/Pt C | 94 | 93 | 144 |
| a ^[4] | Undivided beaker | CC ^a /Pt | 93 | 64 | 40 |
| b ^[5] | Undivided beaker | RVC ^b /Pt | 69 | 62 | 49 |
| c ^[6] | Undivided beaker | GR ^c /Pt | 99 | 60 | 37 |
| d ^[7] | Undivided beaker | CF ^d /Ni foam | 75 | 54 | 50 |
| e ^[8] | Single-pass flow | Pt/Pt | 91 | 67 | 8 |
| f ^[9] | Undivided flow cell | GF ^d /Ni foam | 99 | 27 | 25 |

^aCarbon Cloth; ^bReticulated Vitreous Carbon; ^cGraphite Rod; ^dCarbon Felt

7. Preparation of indole substrates

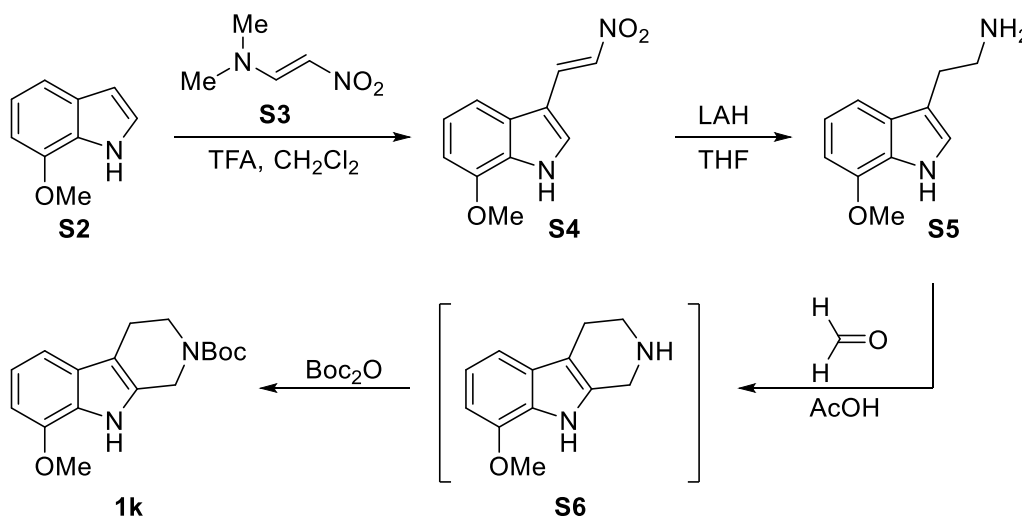
Substrates **1a**^[10], **1b**^[11], **1c**^[12], **1d**^[13], **1f-1h**^[14], **1i-1j**^[15], **1l**^[16], **1m**^[17], **1o**^[18], **1p**^[19], **1q**^[17], **1r**^[20], and **1t**^[17] were prepared according to the published procedures.

1. Preparation of **1e**



To a solution of **S1** (516 mg, 3.0 mmol) and Et_3N (1.25 mL, 9.0 mmol) in CH_2Cl_2 (3 mL) was added allyl bromide (0.31 mL, 3.6 mmol) at 0 °C. The reaction mixture was stirred at rt for 2 h. Saturated NaHCO_3 (10 mL) solution was added and extracted with CH_2Cl_2 (3 x 10 mL). The combined organic fraction was washed with brine, dried over Na_2SO_4 , filtered, and concentrated under reduced pressure. The resulting residue was purified by flash column chromatography (hexane : EtOAc = 4 : 1) to give compound **1e** as a yellow solid (334 mg, 52%). ^1H NMR (400 MHz, CDCl_3) δ 7.92 (s, 1H), 7.48 (d, J = 7.0 Hz, 1H), 7.27 (d, J = 6.8 Hz, 1H), 7.21 – 7.04 (m, 2H), 6.07 – 5.88 (m, 1H), 5.39 – 5.15 (m, 2H), 3.62 (s, 2H), 3.25 (d, J = 5.6 Hz, 2H), 2.97 – 2.74 (m, 4H). $^{13}\text{C}\{^1\text{H}\}$ NMR (100 MHz, CDCl_3) δ 136.2, 135.4, 131.8, 127.3, 121.4, 119.4, 118.3, 118.1, 110.8, 108.4, 60.9, 50.8, 50.2, 21.3. HRMS (ESI) m/z : Calcd for $\text{C}_{14}\text{H}_{17}\text{N}_2^+$ $[\text{M}+\text{H}]^+$ 213.1386; Found 213.1383.

2. Preparation of **1k**

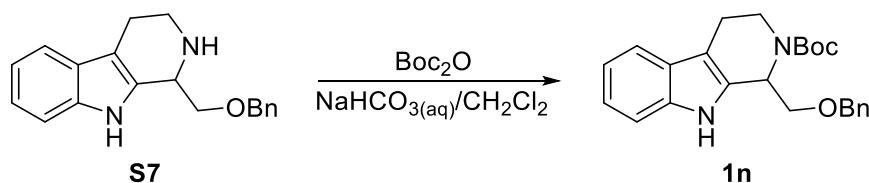


To a solution of **S3** (418 mg, 3.6 mmol) and TFA (0.53 mL, 6.9 mmol) in CH₂Cl₂ (10 mL) was added **S2** (0.39 mL, 3.0 mmol) at 0 °C and stirred at this temperature for 2 h. Diluted with water (10 mL) and extracted with CH₂Cl₂ (3 x 10 mL). The combined organic fraction was washed with saturated NaHCO₃ solution, brine, dried over Na₂SO₄, filtered, and concentrated under reduced pressure. The resulting residue was directly used in the next step without further purification.

To a solution of crude **S4** (269 mg) in THF (5 mL) at 0 °C was added LAH (280 mg, 7.4 mmol) portion wise and stirred for 18 h. Then, diluted with Et₂O (5 mL). Saturated sodium potassium tartrate solution (10mL) was added and stirred for another 2 h. The resulting mixture was extracted with EtOAc (3 x 10 mL). The combined organic fraction was acidified to pH 1-2 by addition of 1 M HCl. The collected aqueous fraction was then basified to pH 9-10 by addition of 3 N of NaOH. Extracted with EtOAc (3 x 10 mL). The combined organic fraction was washed with brine, dried over Na₂SO₄, filtered, and concentrated under reduced pressure. The resulting residue was directly used in the next step without further purification.

To a solution of crude **S5** (161mg) and paraformaldehyde (25 mg, 0.85 mmol) in AcOH (8.5 mL) was heated at at 80 °C for 1 h. After cooling down to rt, the reaction mixture was basified to pH 9-10 by addition of saturated Na₂CO₃ solution and diluted with CH₂Cl₂. Boc₂O (0.23 mL, 1.0 mmol) was added and stirred for another 3 h. The reaction mixture was extracted with CH₂Cl₂ (2 x 10mL). The combined organic fraction was washed with brine, dried over Na₂SO₄, filtered, and concentrated under reduced pressure. The resulting residue was purified by flash column chromatography (hexane : EtOAc = 12 : 1) to give compound **1k** as a white solid (105 mg, 12%). ¹H NMR (400 MHz, CDCl₃) δ 8.28 – 7.97 (m, 1H), 7.10 (d, J = 7.8 Hz, 1H), 7.03 (t, J = 7.8 Hz, 1H), 6.64 (d, J = 7.7 Hz, 1H), 4.64 (br, 2H), 3.95 (s, 3H), 3.76 (br, 2H), 2.79 (br, 2H), 1.51 (s, 9H). ¹³C{¹H} NMR (100 MHz, CDCl₃) δ 155.3, 146.0, 130.4, 128.4, 126.4, 120.1, 111.0, 109.7, 109.2, 102.2, 80.1, 55.5, 42.7, 41.8, 28.6, 21.6. HRMS (ESI) *m/z*: Calcd for C₁₇H₂₂N₂O₃Na⁺ [M+Na]⁺ 325.1523; Found 325.1525.

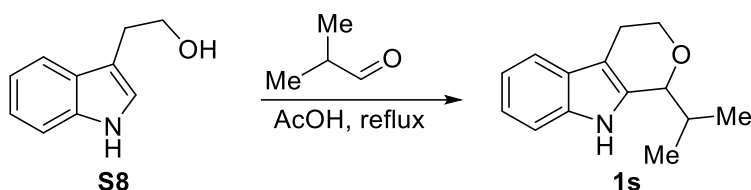
3. Preparation of **1n**



To a solution of **S7** (877 mg, 3.0 mmol) in a mixture of saturated NaHCO₃ solution/CH₂Cl₂ (1:4) (25 mL) was added Boc₂O (0.83 mL, 3.6 mmol) and stirred for 3 h. The reaction mixture was extracted with CH₂Cl₂ (2 x 30 mL). The combined organic fraction was washed with brine, dried over Na₂SO₄, filtered, and concentrated under reduced pressure. The resulting residue was purified by flash column

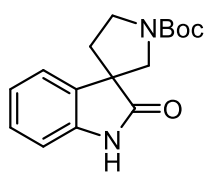
chromatography (hexane : EtOAc = 12 : 1) to give compound **1n** as pale-yellow oil (913mg, 78%). ¹H NMR (400 MHz, CDCl₃) δ 8.79 – 8.48 (m, 1H), 7.60 (d, J = 7.6 Hz, 1H), 7.54 – 7.30 (m, 6H), 7.30 – 7.23 (m, 1H), 7.20 (t, J = 7.3 Hz, 1H), 5.70 – 5.29 (m, 1H), 4.79 – 4.40 (m, 3H), 3.92 (s, 1H), 3.75 (s, 1H), 3.33 – 3.05 (m, 1H), 2.98 – 2.87 (m, 1H), 2.87 – 2.76 (m, 1H), 1.75 – 1.53 (m, 9H). ¹³C{¹H} NMR (100 MHz, CDCl₃) δ 154.7, 137.8, 136.1, 132.8, 128.5, 127.8, 126.9, 126.5, 121.7, 119.3, 118.1, 111.0, 108.8, 80.2, 73.5, 71.3, 50.7, 50.0, 40.4, 39.2, 28.5, 21.5. HRMS (ESI) *m/z*: Calcd for C₂₄H₂₈N₂O₃Na⁺ [M+Na]⁺ 415.1992; Found 415.1994.

4. Preparation of **1s**



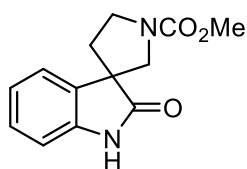
To a solution of **S8** (322 mg, 2.0 mmol) in AcOH (7 mL) was added isobutyraldehyde (0.18 mL, 2.0 mmol) and heated at reflux for 5 h. After cooling down to rt, the reaction mixture was basified to pH 9-10 by addition of saturated Na₂CO₃ solution and diluted with CH₂Cl₂. The reaction mixture was extracted with CH₂Cl₂ (2 x 20 mL). The combined organic fraction was washed with brine, dried over Na₂SO₄, filtered, and concentrated under reduced pressure. The resulting residue was purified by flash column chromatography (hexane : EtOAc = 15 : 1) to give compound **1s** as white solid (131 mg, 30%). ¹H NMR (400 MHz, CDCl₃) δ 7.77 (br, 1H), 7.55 (d, J = 7.6 Hz, 1H), 7.36 (d, J = 7.8 Hz, 1H), 7.21 (t, J = 7.1 Hz, 1H), 7.16 (t, J = 7.2 Hz, 1H), 4.78 – 4.70 (m, 1H), 4.33 (ddd, J = 11.0, 5.4, 1.5 Hz, 1H), 3.80 (td, J = 10.8, 3.7 Hz, 1H), 2.98 (dddd, J = 15.9, 10.6, 5.4, 2.1 Hz, 1H), 2.76 – 2.67 (m, 1H), 2.16 (dtd, J = 13.7, 6.9, 3.3 Hz, 1H), 1.19 (d, J = 6.9 Hz, 3H), 0.91 (d, J = 6.9 Hz, 3H). ¹³C{¹H} NMR (100 MHz, CDCl₃) δ 136.0, 134.4, 127.3, 121.8, 119.7, 118.2, 111.0, 109.4, 104.9, 77.6, 65.0, 32.6, 22.5, 19.1, 16.8, 16.5. HRMS (ESI) *m/z*: Calcd for C₁₄H₁₆NO⁻ [M-H]⁻ 214.1237; Found 214.1232

8. Electrochemical oxidative rearrangement of indoles to 2-oxindoles



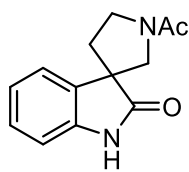
2a (95%)

2a was obtained by General Procedure A [54.7 mg, 95% yield, 89.55 $\mu\text{mol}/(\text{h}\cdot\text{cm}^{-2})$] and by General Procedure B (23.1 mg, 80% yield). Eluent solvents: $\text{CH}_2\text{Cl}_2/\text{MeOH} = 50:1$. ^1H NMR (400 MHz, CDCl_3) (presence of rotamers) δ 9.27 – 9.03 (m, 1H), 7.23 (d, $J = 7.7$ Hz, 1H), 7.17 (br, 1H), 7.04 (t, $J = 7.5$ Hz, 1H), 6.96 (d, $J = 7.7$ Hz, 1H), 3.92 – 3.67 (m, 3H), 3.67 – 3.50 (m, 1H), 2.41 (dt, $J = 12.6, 8.2$ Hz, 1H), 2.07 (br, 2H), 1.57 – 1.40 (m, 9H). $^{13}\text{C}\{^1\text{H}\}$ NMR (100 MHz, CDCl_3) δ 180.2, 154.6, 140.3, 133.1, 128.5, 123.0, 122.8, 110.3, 80.0, 54.5, 53.5, 45.4, 35.7, 28.6. HRMS (ESI) m/z : Calcd for $\text{C}_{16}\text{H}_{20}\text{N}_2\text{O}_3\text{Na}$ $[\text{M}+\text{Na}]^+$ 311.1366; Found 311.1375.



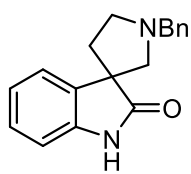
2b (84%)

2b was obtained by General Procedure A with flow rate of 40 mL/min [41.2 mg, 84% yield, 114.59 $\mu\text{mol}/(\text{h}\cdot\text{cm}^{-2})$]. Eluent solvents: $\text{CH}_2\text{Cl}_2/\text{MeOH} = 50:1$. ^1H NMR (400 MHz, CDCl_3) (presence of rotamers) δ 8.49 – 8.36 (m, 1H), 7.24 (t, $J = 7.7$ Hz, 1H), 7.17 (d, $J = 7.1$ Hz, 1H), 7.05 (t, $J = 7.5$ Hz, 1H), 6.93 (d, $J = 7.5$ Hz, 1H), 3.97 – 3.57 (m, 7H), 2.48 – 2.38 (m, 1H), 2.18 – 2.04 (m, 1H). $^{13}\text{C}\{^1\text{H}\}$ NMR (100 MHz, CDCl_3) δ 179.8, 155.6, 140.2, 132.8, 132.4, 128.7, 123.2, 122.9, 110.2, 54.5, 54.1, 53.4, 52.8, 52.4, 45.8, 45.3, 36.5, 35.6. HRMS (ESI) m/z : Calcd for $\text{C}_{13}\text{H}_{14}\text{N}_2\text{O}_3\text{Na}^+$ $[\text{M}+\text{Na}]^+$ 269.0897; Found 269.0905.



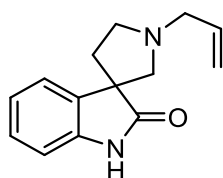
2c (79%)

2c was obtained by General Procedure A [25.5 mg, 79% yield, 0.1 mmol scale, 45.69 $\mu\text{mol}/(\text{h}\cdot\text{cm}^{-2})$]. Eluent solvents: $\text{CH}_2\text{Cl}_2/\text{MeOH} = 50:1$. ^1H NMR (400 MHz, CDCl_3) (presence of rotamers) δ 8.49 – 8.24 (m, 1H), 7.31 – 7.20 (m, 2H), 7.20 – 7.11 (m, 1H), 7.11 – 7.00 (m, 1H), 7.00 – 6.88 (m, 1H), 4.06 – 3.93 (m, 1H), 3.93 – 3.80 (m, 2H), 3.80 – 3.59 (m, 1H), 2.53 – 2.38 (m, 1H), 2.29 – 2.02 (m, 4H). $^{13}\text{C}\{^1\text{H}\}$ NMR (100 MHz, CDCl_3) (presence of rotamers) δ 180.3, 179.2, 169.7, 169.5, 140.5, 140.2, 132.7, 131.8, 128.9, 128.8, 123.3, 123.1, 122.8, 110.4, 110.3, 55.6, 53.7, 53.7, 51.9, 46.8, 45.3, 36.6, 35.3, 22.7, 22.6. HRMS (ESI) m/z : Calcd for $\text{C}_{13}\text{H}_{14}\text{N}_2\text{O}_3\text{Na}^+$ $[\text{M}+\text{Na}]^+$ 269.0897; Found 269.0905.



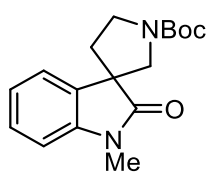
2d (71%)

2d was obtained by General Procedure A with 0.4 mmol LiBr [40.4 mg, 71% yield, 63.59 $\mu\text{mol}/(\text{h}\cdot\text{cm}^{-2})$] and by General Procedure B (11.5mg, 41% yield). Eluent solvents: $\text{CH}_2\text{Cl}_2/\text{MeOH} = 100:1-30:1$. ^1H NMR (400 MHz, CDCl_3) δ 7.70 (br, 1H), 7.49 (d, $J = 7.4$ Hz, 1H), 7.39 (d, $J = 7.2$ Hz, 2H), 7.31 (t, $J = 7.4$ Hz, 2H), 7.26 – 7.21 (m, 1H), 7.19 (td, $J = 7.7, 1.2$ Hz, 1H), 7.06 (dd, $J = 7.5, 0.8$ Hz, 1H), 6.84 (d, $J = 7.7$ Hz, 1H), 3.75 (ABq, 2H, $J = 13$ Hz), 3.15 (dt, $J = 8.4, 3.9$ Hz, 1H), 2.92 (d, $J = 9.1$ Hz, 1H), 2.81 (d, $J = 9.1$ Hz, 1H), 2.73 (q, $J = 8.3$ Hz, 1H), 2.42 (ddd, $J = 12.4, 8.3, 3.9$ Hz, 1H), 2.09 (dt, $J = 13.0, 7.8$ Hz, 1H). $^{13}\text{C}\{^1\text{H}\}$ NMR (100 MHz, CDCl_3) δ 182.0, 139.8, 139.3, 137.0, 128.7, 128.4, 127.8, 127.1, 123.7, 123.1, 109.3, 77.5, 77.2, 76.8, 64.3, 59.8, 54.4, 53.3, 37.3. HRMS (ESI) m/z : Calcd for $\text{C}_{18}\text{H}_{19}\text{N}_2\text{O}^+$ $[\text{M}+\text{H}]^+$ 279.1492; Found 279.1494.



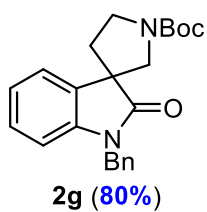
2e (73%)

2e was obtained by General Procedure A with 0.4 mmol LiBr [33.4 mg, 73% yield, 43.58 $\mu\text{mol}/(\text{h}\cdot\text{cm}^{-2})$]. Eluent solvents: $\text{CH}_2\text{Cl}_2/\text{MeOH} = 100:1-30:1$. ^1H NMR (400 MHz, CDCl_3) δ 9.16 (s, 1H), 7.41 (d, $J = 7.3$ Hz, 1H), 7.19 (td, $J = 7.7, 1.2$ Hz, 1H), 7.03 (td, $J = 7.6, 0.9$ Hz, 1H), 6.91 (d, $J = 7.7$ Hz, 1H), 5.95 (ddt, $J = 16.6, 10.2, 6.3$ Hz, 1H), 5.29 – 5.17 (m, 1H), 5.15 – 5.05 (m, 1H), 3.32 – 3.17 (m, 2H), 3.08 (ddd, $J = 8.9, 7.6, 4.8$ Hz, 1H), 2.96 – 2.86 (m, 2H), 2.81 (dd, $J = 16.5, 7.7$ Hz, 1H), 2.40 (ddd, $J = 12.7, 7.8, 4.8$ Hz, 1H), 2.09 (dt, $J = 12.8, 7.4$ Hz, 1H). $^{13}\text{C}\{^1\text{H}\}$ NMR (100 MHz, CDCl_3) δ 183.3, 140.3, 136.5, 135.8, 127.8, 123.4, 122.9, 117.1, 109.8, 64.1, 58.6, 54.5, 53.3, 37.4. HRMS (ESI) m/z : Calcd for $\text{C}_{14}\text{H}_{17}\text{N}_2\text{O}^+$ $[\text{M}+\text{H}]^+$ 229.1335; Found 229.1344.

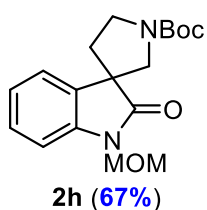


2f (89%)

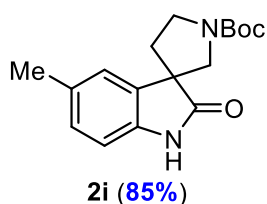
2f was obtained by General Procedure A [53.5 mg, 89% yield, 39.85 $\mu\text{mol}/(\text{h}\cdot\text{cm}^{-2})$] and by General Procedure B (18.9 mg, 63% yield). Eluent solvents: hexane/EtOAc = 5:1. ^1H NMR (400 MHz, CDCl_3) (presence of rotamers) δ 7.30 (t, $J = 7.6$ Hz, 1H), 7.18 (br, 1H), 7.06 (t, $J = 7.4$ Hz, 1H), 6.86 (d, $J = 7.8$ Hz, 1H), 3.88 – 3.63 (m, 3H), 3.62 – 3.47 (m, 1H), 3.22 (s, 3H), 2.46 – 2.33 (m, 1H), 2.00 (br, 1H), 1.55 – 1.38 (m, 9H). $^{13}\text{C}\{^1\text{H}\}$ NMR (100 MHz, CDCl_3) (presence of rotamers) δ 177.4, 154.5, 142.9, 132.9, 128.5, 123.1, 122.5, 108.3, 79.9, 54.5, 53.0, 45.3, 35.6, 28.6, 26.5. HRMS (ESI) m/z : Calcd for $\text{C}_{17}\text{H}_{22}\text{N}_2\text{O}_3\text{Na}^+$ $[\text{M}+\text{Na}]^+$ 325.1523; Found 325.1528.



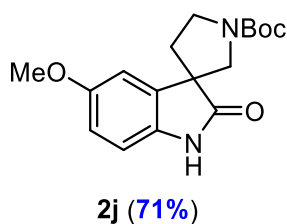
2g was obtained by General Procedure A [60.2 mg, 80% yield, 41.79 $\mu\text{mol}/(\text{h}\cdot\text{cm}^{-2})$] and by General Procedure B (22.7 mg, 60% yield). Eluent solvents: hexane/EtOAc = 5:1. ^1H NMR (400 MHz, CDCl_3) (presence of rotamers) δ 7.37 – 7.27 (m, 7H), 7.24 – 7.13 (m, 2H), 7.09 – 6.99 (m, 1H), 6.82 – 6.67 (m, 1H), 5.03 – 4.84 (m, 2H), 3.94 – 3.70 (m, 2H), 3.70 – 3.55 (m, 1H), 2.54 – 2.39 (m, 1H), 2.18 – 2.02 (m, 1H), 1.56 – 1.43 (m, 9H). $^{13}\text{C}\{^1\text{H}\}$ NMR (100 MHz, CDCl_3) (presence of rotamers) δ 177.6, 154.6, 142.0, 135.8, 132.9, 129.0, 128.4, 127.9, 127.4, 123.2, 122.7, 109.4, 80.0, 54.6, 54.2, 53.1, 52.1, 45.6, 45.4, 44.0, 36.6, 35.9, 28.7, 28.6. HRMS (ESI) m/z : Calcd for $\text{C}_{23}\text{H}_{26}\text{N}_2\text{O}_3\text{Na}^+$ [$\text{M}+\text{Na}$] $^+$ 401.1836; Found 401.1843.



2h was obtained by General Procedure A [44.6 mg, 67% yield, 25.01 $\mu\text{mol}/(\text{h}\cdot\text{cm}^{-2})$]. Eluent solvents: hexane/EtOAc = 5:1. ^1H NMR (400 MHz, CDCl_3) (presence of rotamers) δ 7.30 (t, $J = 7.6$ Hz, 1H), 7.21 (t, $J = 8.5$ Hz, 1H), 7.11 (t, $J = 7.5$ Hz, 1H), 7.06 (d, $J = 7.8$ Hz, 1H), 5.14 (s, 2H), 3.90 – 3.67 (m, 3H), 3.65 – 3.53 (m, 1H), 3.33 (s, 3H), 2.48 – 2.34 (m, 1H), 2.16 – 2.00 (m, 1H), 1.55 – 1.41 (m, 9H). $^{13}\text{C}\{^1\text{H}\}$ NMR (100 MHz, CDCl_3) (presence of rotamers) δ 178.7, 178.2, 154.5, 141.3, 141.2, 132.3, 131.6, 128.7, 123.7, 123.7, 122.7, 109.8, 80.0, 71.6, 56.4, 54.8, 54.3, 53.3, 52.3, 45.6, 45.3, 36.8, 36.0, 28.6, 28.6. HRMS (ESI) m/z : Calcd for $\text{C}_{18}\text{H}_{24}\text{N}_2\text{O}_4\text{Na}^+$ [$\text{M}+\text{Na}$] $^+$ 355.1628; Found 355.1632.

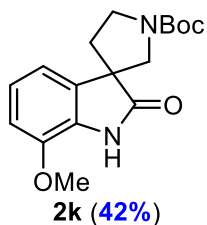


2i was obtained by General Procedure A [51.2 mg, 85% yield, 50.75 $\mu\text{mol}/(\text{h}\cdot\text{cm}^{-2})$] and by General Procedure B (17.0 mg, 56% yield). Eluent solvents: hexane/EtOAc = 4:1-2:1. ^1H NMR (400 MHz, CDCl_3) (presence of rotamers) δ 9.35 – 9.15 (m, 1H), 7.09 – 6.93 (m, 2H), 6.84 (d, $J = 7.7$ Hz, 1H), 3.91 – 3.67 (m, 3H), 3.66 – 3.51 (m, 1H), 2.46 – 2.35 (m, 1H), 2.31 (s, 3H), 2.06 (br, 1H), 1.58 – 1.39 (m, 9H). $^{13}\text{C}\{^1\text{H}\}$ NMR (100 MHz, CDCl_3) (presence of rotamers) δ 180.3, 154.6, 137.8, 133.4, 132.6, 128.8, 123.6, 110.0, 80.0, 54.5, 53.6, 45.4, 35.6, 28.6, 21.3. HRMS (ESI) m/z : Calcd for $\text{C}_{17}\text{H}_{22}\text{N}_2\text{O}_3\text{Na}^+$ [$\text{M}+\text{Na}$] $^+$ 325.1523; Found 325.1527.

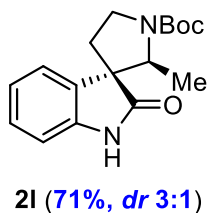


2j was obtained by General Procedure A [45.0 mg, 71% yield, 34.44 $\mu\text{mol}/(\text{h}\cdot\text{cm}^{-2})$]. Eluent solvents: hexane/EtOAc = 4:1-2:1. ^1H NMR (400 MHz, CDCl_3) (presence of rotamers) δ 8.56 – 8.35 (m, 1H), 6.87 – 6.81 (m, 1H), 6.81 – 6.72 (m, 2H), 3.90 – 3.65 (m, 6H), 3.65 – 3.52 (m, 1H), 2.47 – 2.35 (m, 1H), 2.12 – 2.00 (m, 1H), 1.54 – 1.41 (m, 9H). $^{13}\text{C}\{^1\text{H}\}$ NMR (100 MHz, CDCl_3) (presence of rotamers) δ 180.0, 179.8, 156.3, 154.6, 134.6, 133.4, 112.8, 112.6, 110.5,

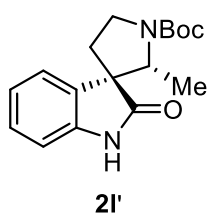
110.2, 80.0, 56.0, 54.5, 54.0, 53.9, 52.9, 45.5, 45.3, 36.5, 35.7, 28.7, 28.6. HRMS (ESI) m/z : Calcd for $C_{17}H_{22}N_2O_4Na^+$ $[M+Na]^+$ 341.1472; Found 341.1474.



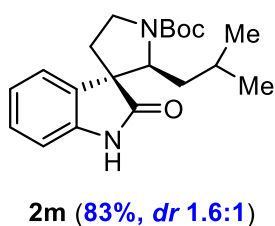
2k was obtained by General Procedure A [13.3 mg, 42% yield, 0.1 mmol scale, 19.59 $\mu\text{mol}/(\text{h}^*\text{cm}^{-2})$]. Eluent solvents: hexane/EtOAc = 4:1-2:1. ^1H NMR (400 MHz, CDCl_3) δ 7.66 – 7.55 (m, 1H), 7.06 – 6.98 (m, 1H), 6.87 – 6.76 (m, 2H), 3.88 (s, 3H), 3.86 – 3.66 (m, 3H), 3.66 – 3.51 (m, 1H), 2.47 – 2.35 (m, 1H), 2.14 – 2.00 (m, 1H), 1.54 – 1.41 (m, 9H). ^{13}C NMR (100 MHz, CDCl_3) δ 178.9, 178.5, 154.6, 144.0, 134.0, 133.5, 128.7, 123.8, 123.7, 115.2, 110.9, 80.0, 55.9, 54.5, 54.1, 54.0, 53.1, 45.5, 45.3, 36.5, 35.7, 28.7, 28.6. HRMS (ESI) m/z : Calcd for $C_{17}H_{22}N_2O_4Na^+$ $[M+Na]^+$ 341.1472; Found 341.1470.



2l was obtained by General Procedure A [32.5 mg, 53% yield, 35.60 $\mu\text{mol}/(\text{h}^*\text{cm}^{-2})$]. Eluent solvents: hexane/EtOAc = 6:1-2:1. ^1H NMR (400 MHz, CDCl_3) δ 9.17 – 8.92 (m, 1H), 7.22 (t, $J = 7.6$ Hz, 1H), 7.11 (d, $J = 7.4$ Hz, 1H), 7.03 (t, $J = 7.5$ Hz, 1H), 6.93 (d, $J = 7.7$ Hz, 1H), 4.14 – 3.90 (m, 1H), 3.91 – 3.68 (m, 2H), 2.42 (br, 1H), 2.17 – 1.99 (m, 1H), 1.47 (s, 9H), 1.30 (d, $J = 6.5$ Hz, 3H). $^{13}\text{C}\{^1\text{H}\}$ NMR (100 MHz, CDCl_3) δ 179.1, 154.5, 140.4, 133.3, 128.4, 123.0, 122.8, 110.0, 79.9, 61.0, 56.8, 44.6, 33.4, 28.6, 16.7. HRMS (ESI) m/z : Calcd for $C_{17}H_{22}N_2O_3Na^+$ $[M+Na]^+$ 325.1523; Found 325.1523.

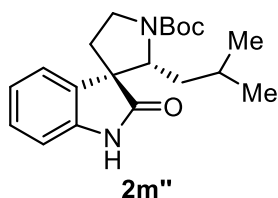


2l' was obtained by General Procedure A [10.6 mg, 18% yield, 12.09 $\mu\text{mol}/(\text{h}^*\text{cm}^{-2})$]. Eluent solvents: hexane/EtOAc = 6:1-2:1. ^1H NMR (400 MHz, CDCl_3) δ 8.13 (br, 1H), 7.26 – 7.20 (m, 2H), 7.05 (td, $J = 7.6, 0.9$ Hz, 1H), 6.92 (d, $J = 7.6$ Hz, 1H), 4.19 – 4.08 (m, 1H), 3.95 (br, 1H), 3.74 – 3.62 (m, 1H), 2.32 (dt, $J = 12.6, 8.1$ Hz, 1H), 2.07 (br, 1H), 1.49 (s, 9H). $^{13}\text{C}\{^1\text{H}\}$ NMR (100 MHz, CDCl_3) δ 180.0, 155.1, 140.6, 130.0, 128.5, 125.3, 122.5, 110.1, 59.1, 46.0, 34.0, 28.7, 17.8. HRMS (ESI) m/z : Calcd for $C_{17}H_{22}N_2O_3Na^+$ $[M+Na]^+$ 325.1523; Found 325.1525.

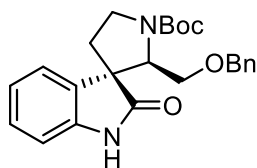


2m was obtained by General Procedure A with 20 mL solvent [29.3 mg, 56% yield, 17.76 $\mu\text{mol}/(\text{h}^*\text{cm}^{-2})$] and by General Procedure B (<40%). Eluent solvents: hexane/EtOAc = 6:1-2:1. ^1H NMR (400 MHz, CDCl_3) δ 9.33 – 9.12 (m, 1H), 7.22 (dd, $J = 10.7, 5.4$ Hz, 1H), 7.13 – 6.97 (m, 2H), 6.94 (d, $J = 7.6$ Hz, 1H), 4.19 – 3.61 (m, 3H), 2.62 – 2.34 (m, 1H), 2.08 – 1.97 (m, 1H), 1.86

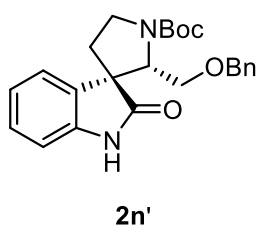
(br, 1H), 1.59 – 1.16 (m, 11H), 0.81 (br, 6H). $^{13}\text{C}\{^1\text{H}\}$ NMR (100 MHz, CDCl_3) δ 178.8, 154.8, 139.7, 134.8, 128.2, 123.0, 122.6, 110.0, 80.3, 63.7, 55.9, 44.0, 40.3, 34.0, 28.7, 25.2, 23.1, 22.8. HRMS (ESI) m/z : Calcd for $\text{C}_{20}\text{H}_{28}\text{N}_2\text{O}_3\text{Na}^+$ $[\text{M}+\text{Na}]^+$ 367.1992; Found 367.1998.



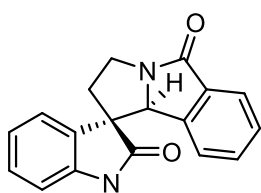
2m'' was obtained by General Procedure A with 20 mL solvent [18.4 mg, 27% yield, $8.57 \mu\text{mol}/(\text{h}^*\text{cm}^{-2})$]. Eluent solvents: hexane/EtOAc = 6:1-2:1. ^1H NMR (400 MHz, CDCl_3) δ 8.85 (s, 1H), 7.26 – 7.20 (m, 2H), 7.04 (t, $J = 7.5$ Hz, 1H), 6.94 (d, $J = 7.9$ Hz, 1H), 4.29 – 3.95 (m, 2H), 3.58 (dt, $J = 11.2, 7.4$ Hz, 1H), 2.31 (dt, $J = 12.4, 8.5$ Hz, 1H), 2.06 – 1.95 (m, 1H), 1.75 (br, 1H), 1.50 (s, 10H), 1.35 (br, 1H), 1.00 (br, 1H), 0.83 (d, $J = 6.4$ Hz, 3H), 0.58 (d, $J = 6.3$ Hz, 3H). $^{13}\text{C}\{^1\text{H}\}$ NMR (100 MHz, CDCl_3) δ 181.1, 155.4, 140.7, 130.1, 128.5, 125.3, 122.3, 110.3, 80.0, 61.2, 45.8, 41.0, 35.9, 31.1, 28.7, 24.7, 23.5, 22.0. HRMS (ESI) m/z : Calcd for $\text{C}_{20}\text{H}_{28}\text{N}_2\text{O}_3\text{Na}^+$ $[\text{M}+\text{Na}]^+$ 367.1992; Found 367.1997.



2n was obtained by General Procedure A [34.1 mg, 43% yield, $15.24 \mu\text{mol}/(\text{h}^*\text{cm}^{-2})$]. Eluent solvents: hexane/EtOAc = 6:1-2:1. ^1H NMR (400 MHz, CDCl_3) δ 8.66 – 8.40 (m, 1H), 7.25 – 7.06 (m, 6H), 7.06 – 6.93 (m, 2H), 6.83 (br, 1H), 4.52 – 3.69 (m, 7H), 2.45 (br, 1H), 2.07 (br, 1H), 1.64 – 1.32 (m, 9H). $^{13}\text{C}\{^1\text{H}\}$ NMR (100 MHz, CDCl_3) δ 178.7, 154.4, 140.0, 138.3, 134.3, 128.1, 127.4, 122.8, 122.4, 109.9, 80.3, 72.8, 68.6, 64.4, 55.1, 45.3, 34.7, 28.6. HRMS (ESI) m/z : Calcd for $\text{C}_{24}\text{H}_{28}\text{N}_2\text{O}_4\text{Na}^+$ $[\text{M}+\text{Na}]^+$ 431.1941; Found 431.1943.



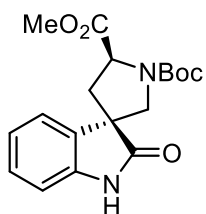
2n' was obtained by General Procedure A [16.9 mg, 21% yield, $7.44 \mu\text{mol}/(\text{h}^*\text{cm}^{-2})$]. Eluent solvents: hexane/EtOAc = 6:1-2:1. ^1H NMR (400 MHz, CDCl_3) δ 8.34 (br, 1H), 7.32 (d, $J = 7.4$ Hz, 1H), 7.29 – 7.16 (m, 4H), 7.11 – 6.95 (m, 3H), 6.88 (d, $J = 7.7$ Hz, 1H), 4.46 – 3.75 (m, 5H), 3.75 – 3.59 (m, 1H), 3.58 – 3.29 (m, 1H), 2.35 – 2.07 (m, 2H), 1.46 (s, 9H). ^{13}C NMR (100 MHz, CDCl_3) δ 180.7, 155.0, 141.3, 138.1, 129.9, 128.4, 128.3, 127.5, 127.3, 124.9, 122.3, 110.1, 80.2, 73.3, 69.5, 62.2, 46.5, 35.2, 28.6. HRMS (ESI) m/z : Calcd for $\text{C}_{24}\text{H}_{28}\text{N}_2\text{O}_4\text{Na}^+$ $[\text{M}+\text{Na}]^+$ 431.1941; Found 431.1944.



2o (81%, dr 5:1)

2o was obtained by General Procedure A with 20 mL solvent [23.4 mg, 81% yield, 0.1 mmol scale, 48.36 $\mu\text{mol}/(\text{h}\cdot\text{cm}^2)$]. Eluent solvents: $\text{CH}_2\text{Cl}_2/\text{MeOH} = 100:1$. Data of the major isomer: ^1H NMR (400 MHz, CDCl_3) δ 8.39 (s, 1H), 7.71 (d, $J = 7.4$ Hz, 1H), 7.30 (dt, $J = 7.5, 3.8$ Hz, 1H), 7.28 – 7.22 (m, 1H), 7.06 – 6.98 (m, 2H), 6.81 (d, $J = 7.7$ Hz, 1H), 6.68 (td, $J = 7.6, 1.0$ Hz, 1H), 6.49 (d, $J = 7.5$ Hz, 1H), 5.25 (s, 1H), 4.17 – 4.07 (m, 1H), 3.85 (ddd, $J = 11.7, 9.9, 1.6$

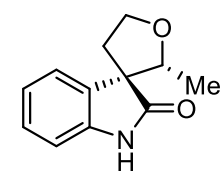
Hz, 1H), 3.11 (dt, $J = 13.0, 9.8$ Hz, 1H), 2.53 (ddd, $J = 13.0, 8.1, 1.9$ Hz, 1H). $^{13}\text{C}\{^1\text{H}\}$ NMR (100 MHz, CDCl_3) δ 178.0, 171.3, 142.1, 139.6, 134.4, 132.0, 129.2, 129.0, 128.4, 123.8, 123.5, 123.1, 122.0, 110.0, 71.0, 54.9, 41.6, 39.8. HRMS (ESI) m/z : Calcd for $\text{C}_{18}\text{H}_{14}\text{N}_2\text{O}_2\text{Na}^+$ $[\text{M}+\text{Na}]^+$ 313.0947; Found 313.0951.



2p (94%, dr 5:1)

2p was obtained by General Procedure A with flow rate of 40 mL/min [65.3 mg, 94% yield, 144.21 $\mu\text{mol}/(\text{h}\cdot\text{cm}^2)$] and by General Procedure B (25.3 mg, 73% yield). Eluent solvents: hexane/EtOAc = 10:1-3:1. Data of the major isomer: ^1H NMR (400 MHz, CDCl_3) (presence of rotamers) δ 8.93 – 8.82 (m, 1H), 7.29 – 7.20 (m, 1H), 7.13 – 7.08 (m, 1H), 7.08 – 7.01 (m, 1H), 6.99 – 6.92 (m, 1H), 4.79 – 4.57 (m, 1H), 3.89 – 3.66 (m, 1H), 2.63 – 2.50 (m, 1H), 2.44 – 2.34 (m, 1H), 1.51 – 1.40 (m, 1H).

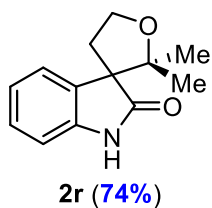
$^{13}\text{C}\{^1\text{H}\}$ NMR (100 MHz, CDCl_3) (presence of rotamers) δ 178.1, 178.0, 172.6, 172.3, 154.4, 153.5, 139.9, 139.9, 133.4, 133.2, 128.8, 123.3, 123.3, 122.6, 110.4, 81.0, 59.2, 58.8, 55.5, 54.9, 53.2, 52.6, 52.5, 52.3, 40.6, 39.7, 28.4. HRMS (ESI) m/z : Calcd for $\text{C}_{18}\text{H}_{22}\text{N}_2\text{O}_5\text{Na}^+$ $[\text{M}+\text{Na}]^+$ 369.1421; Found 369.1428.



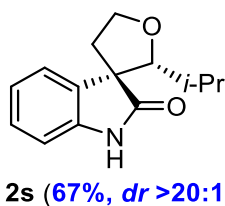
2q (63%, dr >20:1)

2q was obtained by General Procedure A [12.8 mg, 63% yield, 0.1 mmol scale, 65.68 $\mu\text{mol}/(\text{h}\cdot\text{cm}^2)$]. Eluent solvents: hexane/EtOAc = 3:1-1:1. ^1H NMR (400 MHz, CDCl_3) δ 8.73 (br, 1H), 7.28 – 7.20 (m, 2H), 7.06 (t, $J = 7.3$ Hz, 1H), 6.95 (d, $J = 7.6$ Hz, 1H), 4.30 – 4.12 (m, 3H), 2.70 (ddd, $J = 12.7, 9.4, 6.6$ Hz, 1H), 2.20 (ddd, $J = 10.0, 7.8, 3.7$ Hz, 1H), 0.91 (d, $J = 6.2$ Hz, 3H).

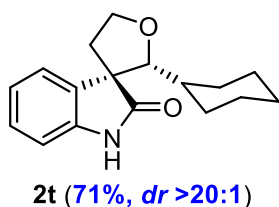
$^{13}\text{C}\{^1\text{H}\}$ NMR (100 MHz, CDCl_3) δ 180.0, 140.3, 131.8, 128.1, 124.8, 122.8, 110.0, 82.4, 67.3, 58.4, 38.2, 15.2. HRMS (ESI) m/z : Calcd for $\text{C}_{12}\text{H}_{13}\text{NO}_2\text{Na}^+$ $[\text{M}+\text{Na}]^+$ 226.0838; Found 226.0838.



2r was obtained by General Procedure A [32.2 mg, 74% yield, 80.08 $\mu\text{mol}/(\text{h}^*\text{cm}^{-2})$]. Eluent solvents: hexane/EtOAc = 3:1-1:1. ^1H NMR (400 MHz, CDCl_3) δ 8.21 (br, 1H), 7.26 (d, $J = 7.4$ Hz, 2H), 7.22 (t, $J = 7.6$ Hz, 1H), 7.03 (t, $J = 7.6$ Hz, 1H), 6.90 (d, $J = 7.7$ Hz, 1H), 4.27 (td, $J = 8.9, 5.6$ Hz, 1H), 4.18 (td, $J = 8.7, 6.0$ Hz, 1H), 2.67 (ddd, $J = 12.8, 9.3, 6.0$ Hz, 1H), 2.31 (ddd, $J = 12.9, 8.9, 5.4$ Hz, 1H), 1.34 (s, 3H), 1.10 (s, 3H). $^{13}\text{C}\{^1\text{H}\}$ NMR (100 MHz, CDCl_3) δ 179.9, 140.6, 131.8, 128.2, 125.1, 122.5, 109.7, 84.9, 64.5, 60.0, 36.1, 24.4, 23.4. HRMS (ESI) m/z : Calcd for $\text{C}_{13}\text{H}_{15}\text{NO}_2\text{Na}^+$ $[\text{M}+\text{Na}]^+$ 240.0995; Found 240.0998.



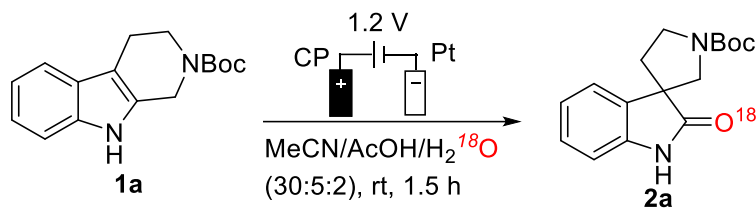
2s was obtained by General Procedure A [31.0 mg, 67% yield, 23.75 $\mu\text{mol}/(\text{h}^*\text{cm}^{-2})$]. Eluent solvents: hexane/EtOAc = 3:1-1:1. ^1H NMR (400 MHz, CDCl_3) δ 8.92 (s, 1H), 7.30 – 7.19 (m, 2H), 7.06 (t, $J = 7.5$ Hz, 1H), 6.97 (d, $J = 7.7$ Hz, 1H), 4.24 – 4.09 (m, 2H), 3.73 (d, $J = 9.9$ Hz, 1H), 2.68 (ddd, $J = 12.4, 9.1, 7.6$ Hz, 1H), 2.13 (ddd, $J = 12.7, 7.8, 5.2$ Hz, 1H), 1.59 (ddt, $J = 13.2, 10.0, 6.6$ Hz, 1H), 1.00 (d, $J = 6.5$ Hz, 3H), 0.43 (d, $J = 6.7$ Hz, 3H). $^{13}\text{C}\{^1\text{H}\}$ NMR (100 MHz, CDCl_3) δ 180.9, 140.2, 132.1, 128.1, 124.8, 122.7, 110.1, 91.6, 66.6, 56.9, 40.6, 30.3, 21.1, 17.6. HRMS (ESI) m/z : Calcd for $\text{C}_{14}\text{H}_{17}\text{NO}_2\text{Na}^+$ $[\text{M}+\text{Na}]^+$ 254.1151; Found 254.1148.



2t was obtained by General Procedure A [38.5 mg, 71% yield, 46.36 $\mu\text{mol}/(\text{h}^*\text{cm}^{-2})$]. Eluent solvents: hexane/EtOAc = 3:1-1:1. ^1H NMR (400 MHz, CDCl_3) δ 8.62 (s, 1H), 7.29 – 7.21 (m, 2H), 7.06 (td, $J = 7.6, 1.0$ Hz, 1H), 6.94 (d, $J = 7.7$ Hz, 1H), 4.23 – 4.08 (m, 2H), 3.81 (d, $J = 9.8$ Hz, 1H), 2.64 (ddd, $J = 12.4, 9.3, 7.5$ Hz, 1H), 2.09 (ddd, $J = 12.7, 7.8, 5.1$ Hz, 1H), 2.03 – 1.96 (m, 1H), 1.69 – 1.60 (m, 1H), 1.52 – 1.43 (m, 1H), 1.43 – 1.29 (m, 2H), 1.14 – 0.96 (m, 3H), 0.90 – 0.81 (m, 1H), 0.81 – 0.71 (m, 2H). $^{13}\text{C}\{^1\text{H}\}$ NMR (100 MHz, CDCl_3) δ 180.6, 139.9, 132.1, 128.0, 124.7, 122.8, 110.1, 90.1, 66.4, 56.6, 40.6, 39.4, 31.1, 27.4, 26.2, 25.5. HRMS (ESI) m/z : Calcd for $\text{C}_{17}\text{H}_{21}\text{NO}_2\text{Na}^+$ $[\text{M}+\text{Na}]^+$ 294.1464; Found 294.1465.

9. Mechanistic study

9.1 Isotopic labelling experiment with ^{18}O



Reactions were conducted in two 4 mL vials with anolyte and catholyte. The assemble and detail information were discussed in **Flow cell setup**. The reaction was performed in the two PEEK housing flow cell at constant voltage 1.2 V using potentiostatic method in VMP-300. Electrolyte was prepared by the addition of LiBr (0.2 mmol, 2.0 equiv.) in MeCN/AcOH/H₂¹⁸O (30:5:2, 3 mL). **1a** (27.2mg, 0.1 mmol) was added in the anolyte. The anolyte was separated from the cathode electrolyte (3mL) by a proton-exchange membrane Nafion 117. The 20 mL/min was set for synthesis, and the cut-off charge was set 2 F/mol. Reaction was monitored by TLC. The reaction mixture was diluted with EtOAc (10 mL). Saturated aqueous Na₂CO₃ solution (10 mL) was added. The organic fraction was collected, and the aqueous fraction was extracted with EtOAc (2 x 10 mL). The combined organic fraction was washed with brine, dried over Na₂SO₄, filtered and concentrated under reduced pressure. The resulting residue was purified by flash column chromatography on silica gel (CH₂Cl₂/MeOH = 50:1) to give compound **2a** (26.9mg, 92.6%). ¹H NMR (400 MHz, CDCl₃) (presence of rotamers) δ 8.69 – 8.60 (m, 1H), 7.26 - 7.20 (m, 1H), 7.18 (t, J = 9.7 Hz, 1H), 7.05 (t, J = 7.4 Hz, 1H), 6.97 - 6.90 (m, 1H), 3.92 - 3.68 (m, 3H), 3.65 – 3.55 (m, 1H), 2.47 - 2.36 (m, 1H), 2.15 - 2.01 (m, 1H), 1.48 (d, J = 24.1 Hz, 9H). ¹³C{¹H} NMR (100 MHz, CDCl₃) (presence of rotamers) δ 180.3, 180.0, 154.6, 140.2, 133.2, 128.5, 123.1, 122.9, 110.2, 80.0, 54.5, 54.0, 53.5, 52.5, 45.6, 45.3, 36.5, 35.7, 28.7, 28.6. HRMS (ESI) *m/z*: Calcd for C₁₆H₂₀N₂O₂¹⁸ONa⁺ [M+Na]⁺ 313.1409; Found 313.1413. *m/z*: Calcd for C₁₆H₂₀N₂O₃Na⁺ [M+Na]⁺ 311.1366; Found 311.1370.

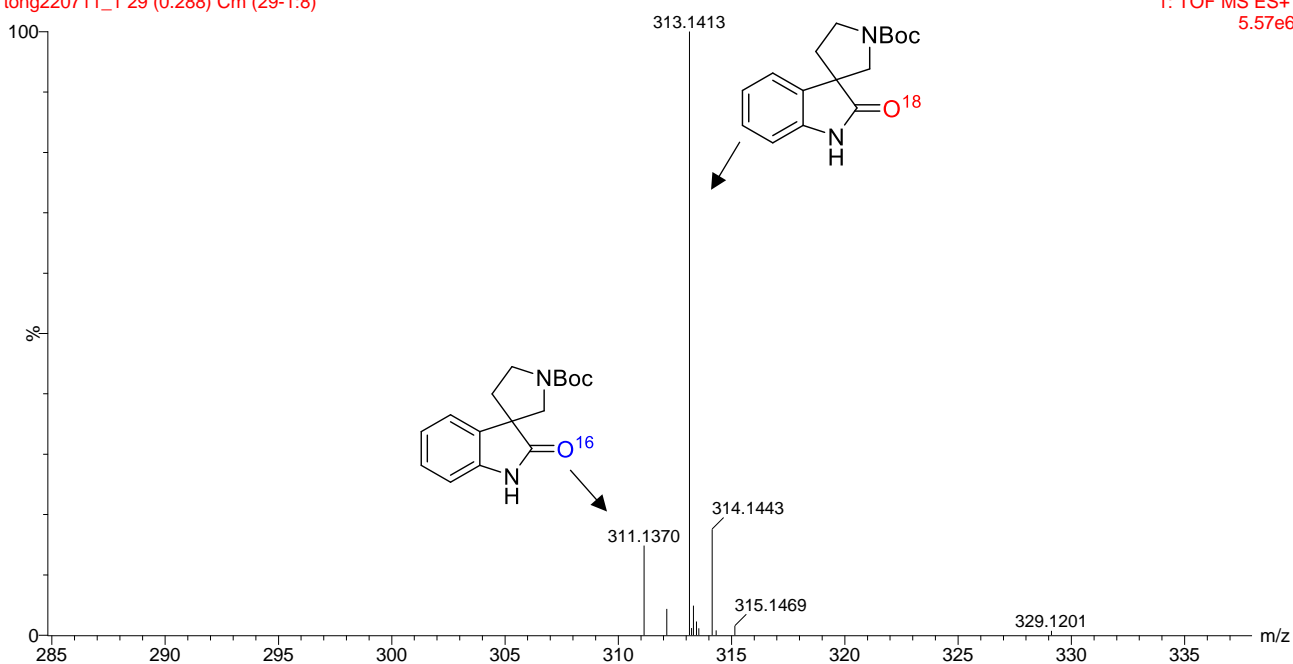
Intensity of peak *m/z* 313.1413 (C₁₆H₂₀N₂O₂¹⁸ONa⁺) : 5.572e6

Intensity of peak *m/z* 311.1370 (C₁₆H₂₀N₂O₃Na⁺) : 8.255e5

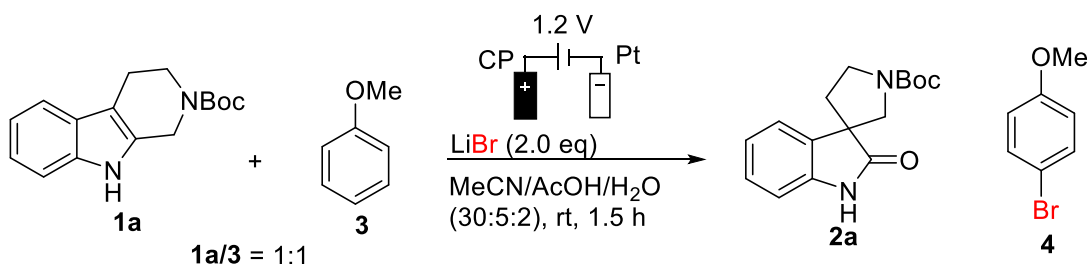
$$\text{Percentage of } ^{18}\text{O} \text{ substitution} = \frac{5.572e6}{5.572e6+8.255e5} = 87\%$$

cyt-YT-0044, MW=290, 288
tong220711_1 29 (0.288) Cm (29-1:8)

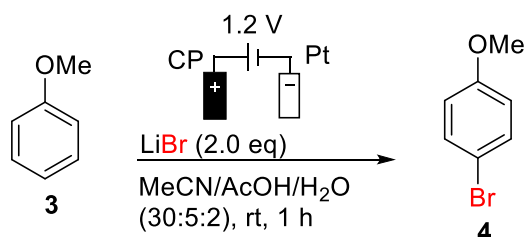
1: TOF MS ES+
5.57e6



9.2 RBS trapping



The reaction was conducted following General Procedure A in 0.1 mmol scale with 1.0 equiv. **3** (0.1 mmol) added in anolyte. After workup, the ratio of **2a** and **4** was determined by NMR of the crude product. It was found that $\mathbf{2a}/\mathbf{4} = 1:0$.

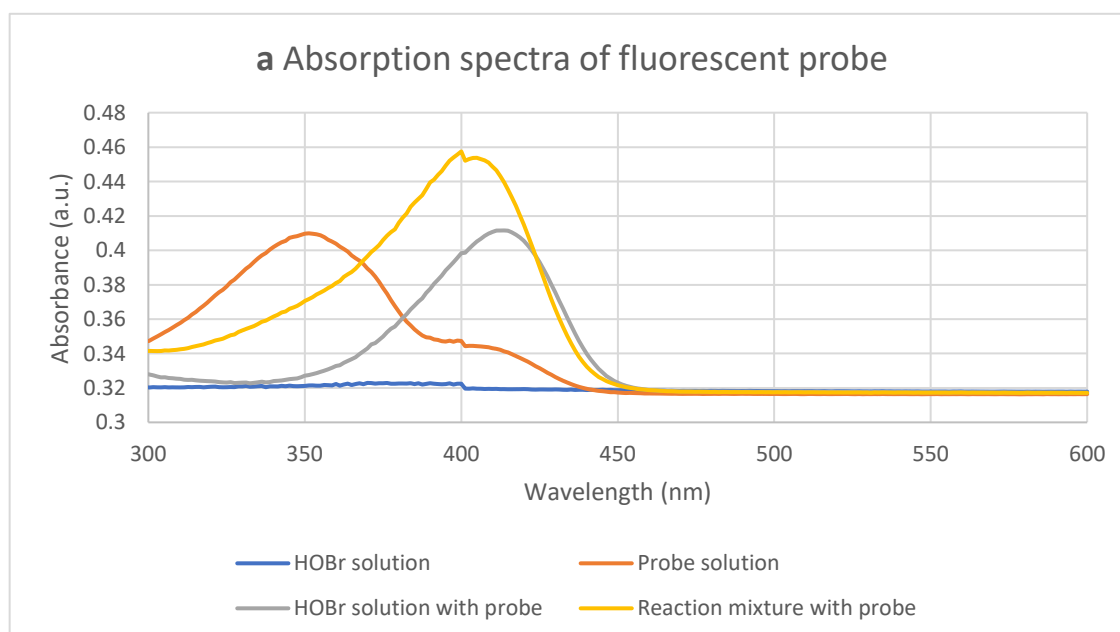
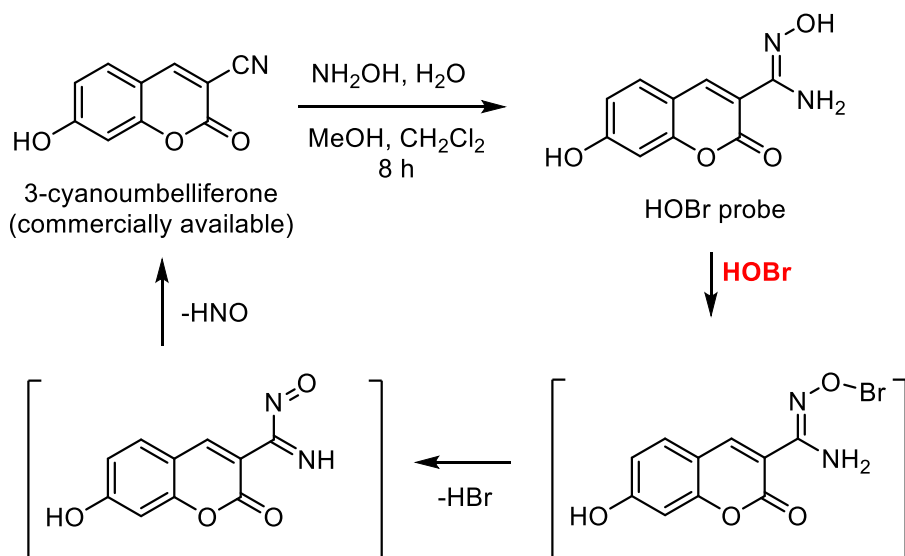


The reaction was conducted following General Procedure A in 0.1 mmol scale with **3** instead of **1a** added in anolyte. Compound **4** was obtained without further purification (16.9mg, 91%). ^1H NMR

(400 MHz, CDCl₃) δ 7.42 - 7.34 (m, 2H), 6.82 - 6.75 (m, 2H), 3.78 (s, 3H). ¹³C {¹H} NMR (100 MHz, CDCl₃) δ 158.8, 132.4, 115.9, 113.0, 55.6. HRMS (CI) *m/z*: Calcd for C₇H₇OBr⁺ [M]⁺ 185.9675; Found 185.9687.

9.3 Detection of RBS

The fluorescent probe was prepared according to the published literature^[21].



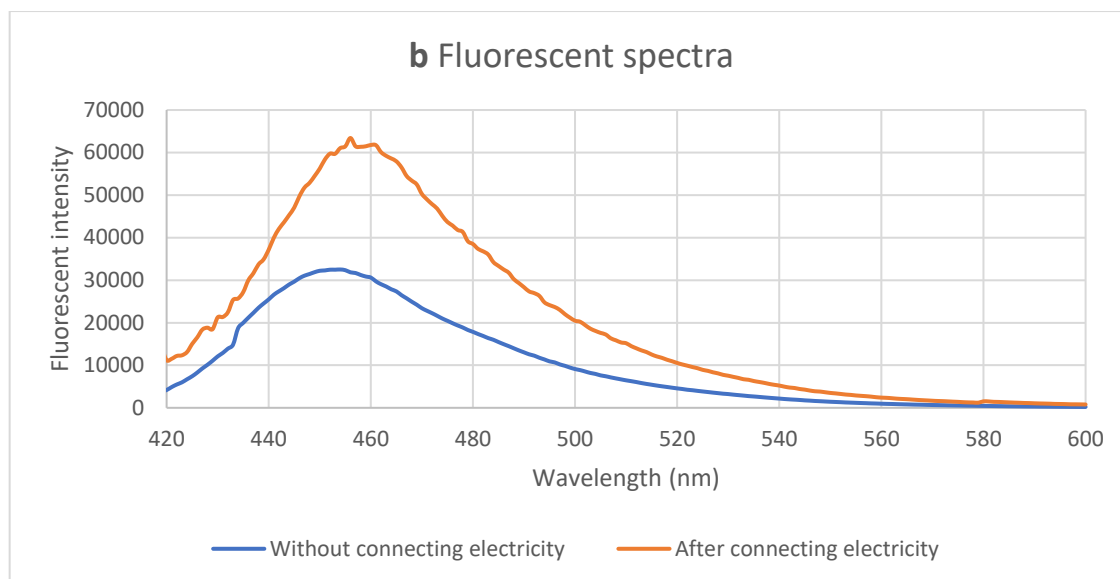


Figure S8. (a). Absorption spectra of fluorescent probe in different solution and HOBr solution. (b). Fluorescent spectra of reaction mixture before and during reaction.

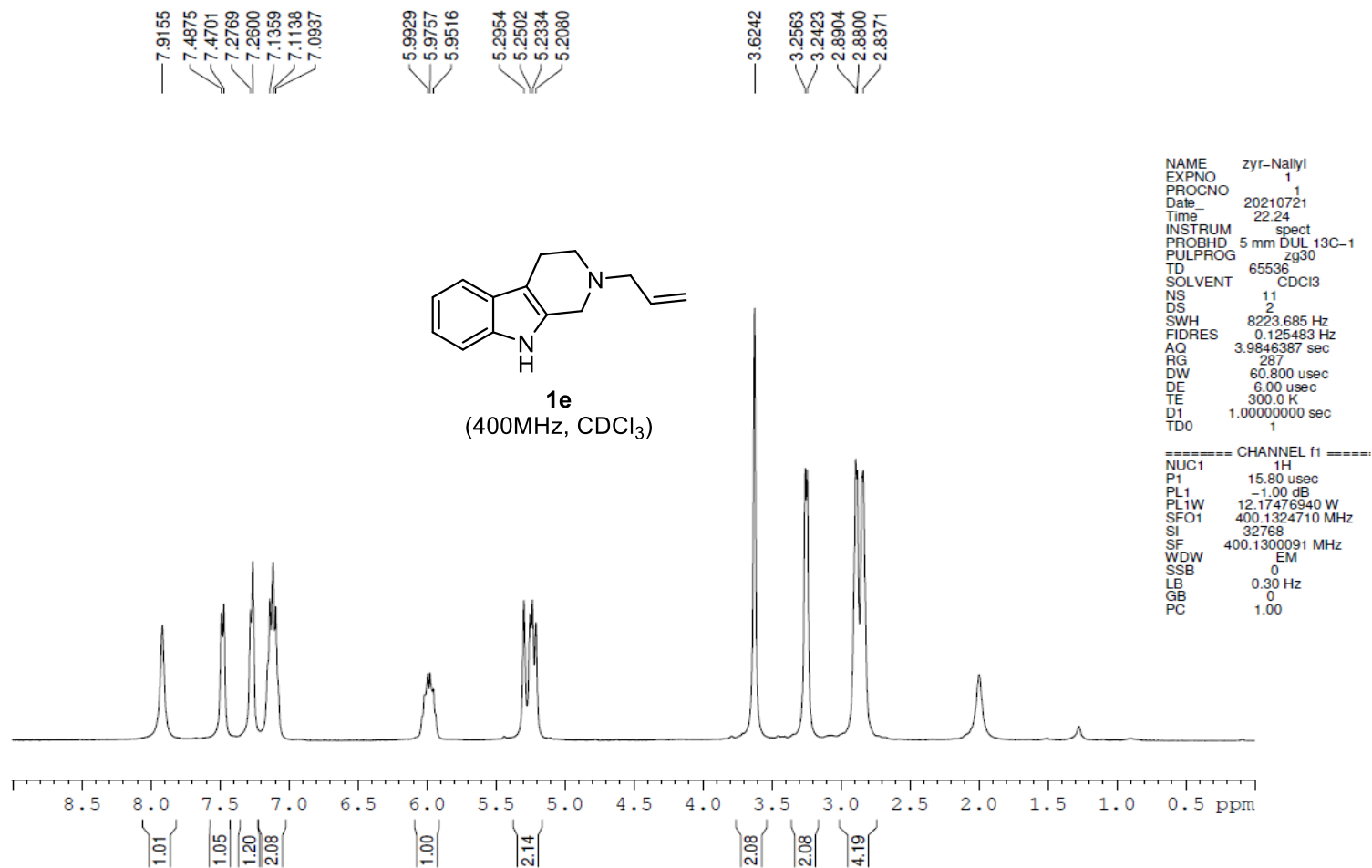
Based on the results from Zeng^[21], the shift in both UV absorption and fluorescent emission refer to the presence of HOBr (RBS) in the reaction mixture after connecting to electricity. This can support the *in situ* generation of HOBr after connecting electricity.

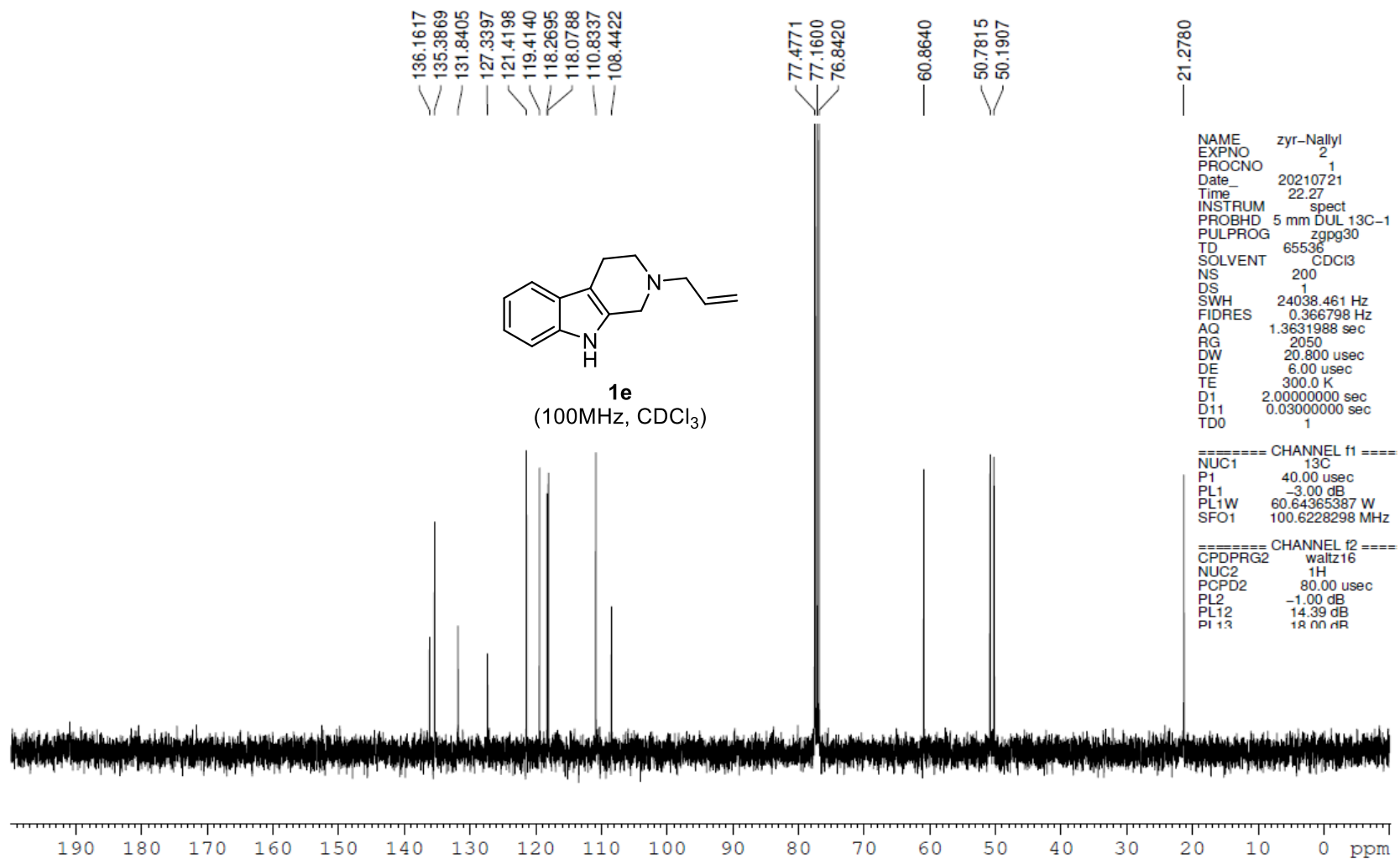
10. References

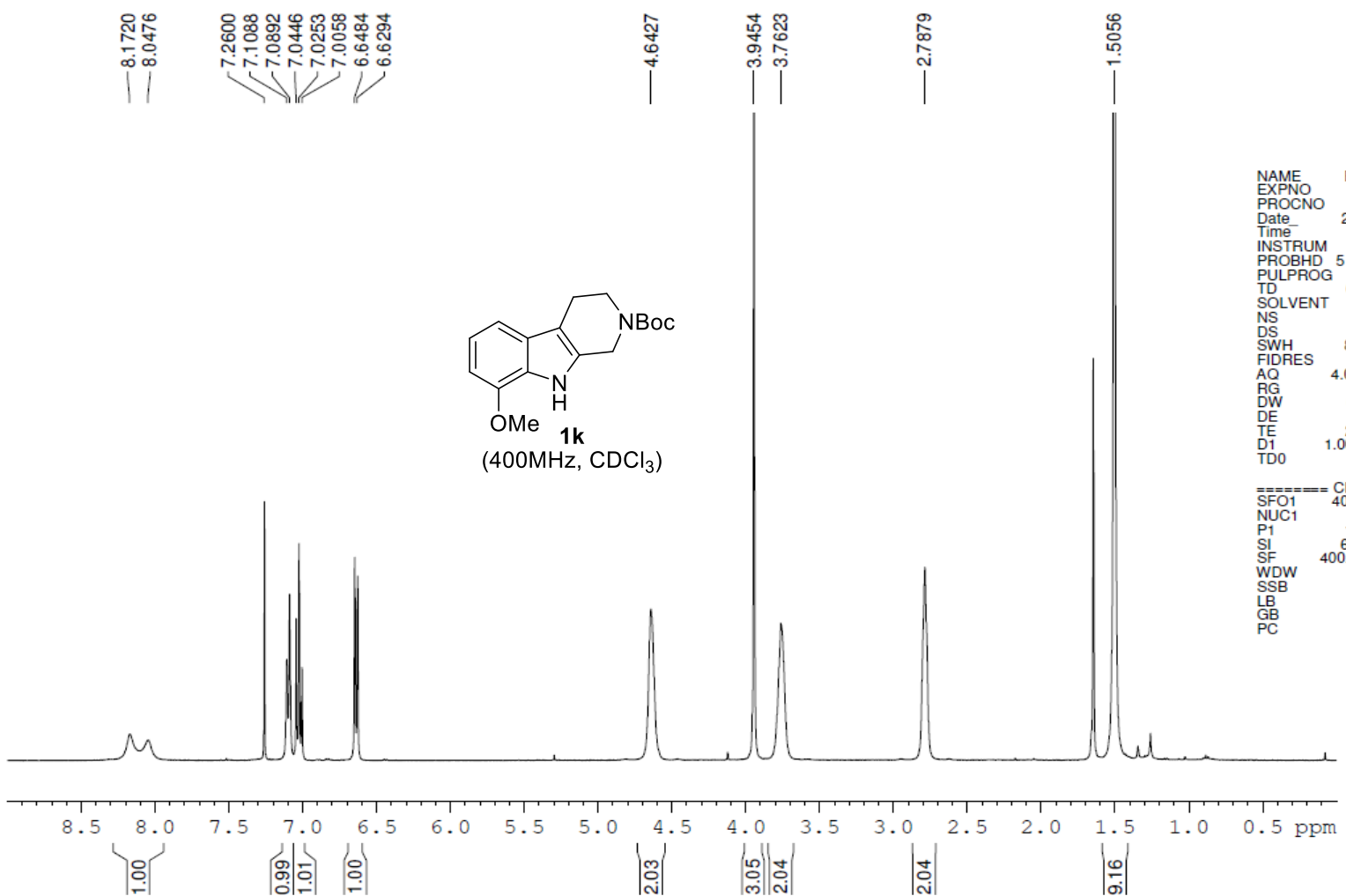
- [1] M. Wang, S. Liu, T. Qian, J. Liu, J. Zhou, H. Ji, J. Xiong, J. Zhong, C. Yan, *Nat Commun* **2019**, *10*, 341.
- [2] M. J. Kim, Y. Seo, M. A. Cruz, B. J. Wiley, *ACS Nano* **2019**, *13*, 6998–7009.
- [3] Keiichi. Tsuji, P. J. Elving, *Anal. Chem.* **1969**, *41*, 216–218.
- [4] J. Qin, M. Luo, D. An, J. Li, *Angew. Chem.* **2021**, *133*, 1889–1896.
- [5] N. Fu, G. S. Sauer, A. Saha, A. Loo, S. Lin, *Science* **2017**, *357*, 575–579.
- [6] K. Liu, S. Tang, P. Huang, A. Lei, *Nat Commun* **2017**, *8*, 775.
- [7] Y. Yuan, Y. Zheng, B. Xu, J. Liao, F. Bu, S. Wang, J.-G. Hu, A. Lei, *ACS Catal.* **2020**, *10*, 6676–6681.
- [8] A. A. Folgueiras-Amador, K. Philipps, S. Guilbaud, J. Poelakker, T. Wirth, *Angew. Chem. Int. Ed.* **2017**, *56*, 15446–15450.
- [9] W.-J. Kong, L. H. Finger, A. M. Messinis, R. Kuniyil, J. C. A. Oliveira, L. Ackermann, *J. Am. Chem. Soc.* **2019**, *141*, 17198–17206.
- [10] M. Géraldy, M. Morgen, P. Sehr, R. R. Steimbach, D. Moi, J. Ridinger, I. Oehme, O. Witt, M. Malz, M. S. Nogueira, O. Koch, N. Gunkel, A. K. Miller, *J. Med. Chem.* **2019**, *62*, 4426–4443.

- [11] J. Ye, Y. Lin, Q. Liu, D. Xu, F. Wu, B. Liu, Y. Gao, H. Chen, *Org. Lett.* **2018**, *20*, 5457–5460.
- [12] J. Ye, J. Wu, T. Lv, G. Wu, Y. Gao, H. Chen, *Angew. Chem. Int. Ed.* **2017**, *56*, 14968–14972.
- [13] K. Buaban, W. Phutdhawong, T. Taechowisan, W. S. Phutdhawong, *Molecules* **2021**, *26*, 207.
- [14] E. J. Cochrane, L. A. Hassall, I. Coldham, *J. Org. Chem.* **2015**, *80*, 5964–5969.
- [15] N. Jana, Q. Nguyen, T. G. Driver, *J. Org. Chem.* **2014**, *79*, 2781–2791.
- [16] G. Zhao, L. Liang, E. Wang, S. Lou, R. Qi, R. Tong, *Green Chem.* **2021**, *23*, 2300–2307.
- [17] J. Xu, L. Liang, H. Zheng, Y. R. Chi, R. Tong, *Nat Commun* **2019**, *10*, 4754.
- [18] Y.-N. Sun, C.-L. Wang, N. Zhang, Z. Wang, Z.-L. Liu, J.-L. Liu, *Chinese Chemical Letters* **2014**, *25*, 1503–1506.
- [19] L. Chen, J. Xie, H. Song, Y. Liu, Y. Gu, L. Wang, Q. Wang, *J. Agric. Food Chem.* **2016**, *64*, 6508–6516.
- [20] X. Zhang, X. Li, J. C. Lanter, Z. Sui, *Org. Lett.* **2005**, *7*, 2043–2046.
- [21] X. Huo, X. Wang, R. Yang, Z. Li, Y. Sun, L. Qu, H. Zeng. *Sensors and Actuators: B. Chemical.* **2020**, *315*, 128125.

11. Copies of ^1H - and ^{13}C -NMR spectra (S28-S85)







```

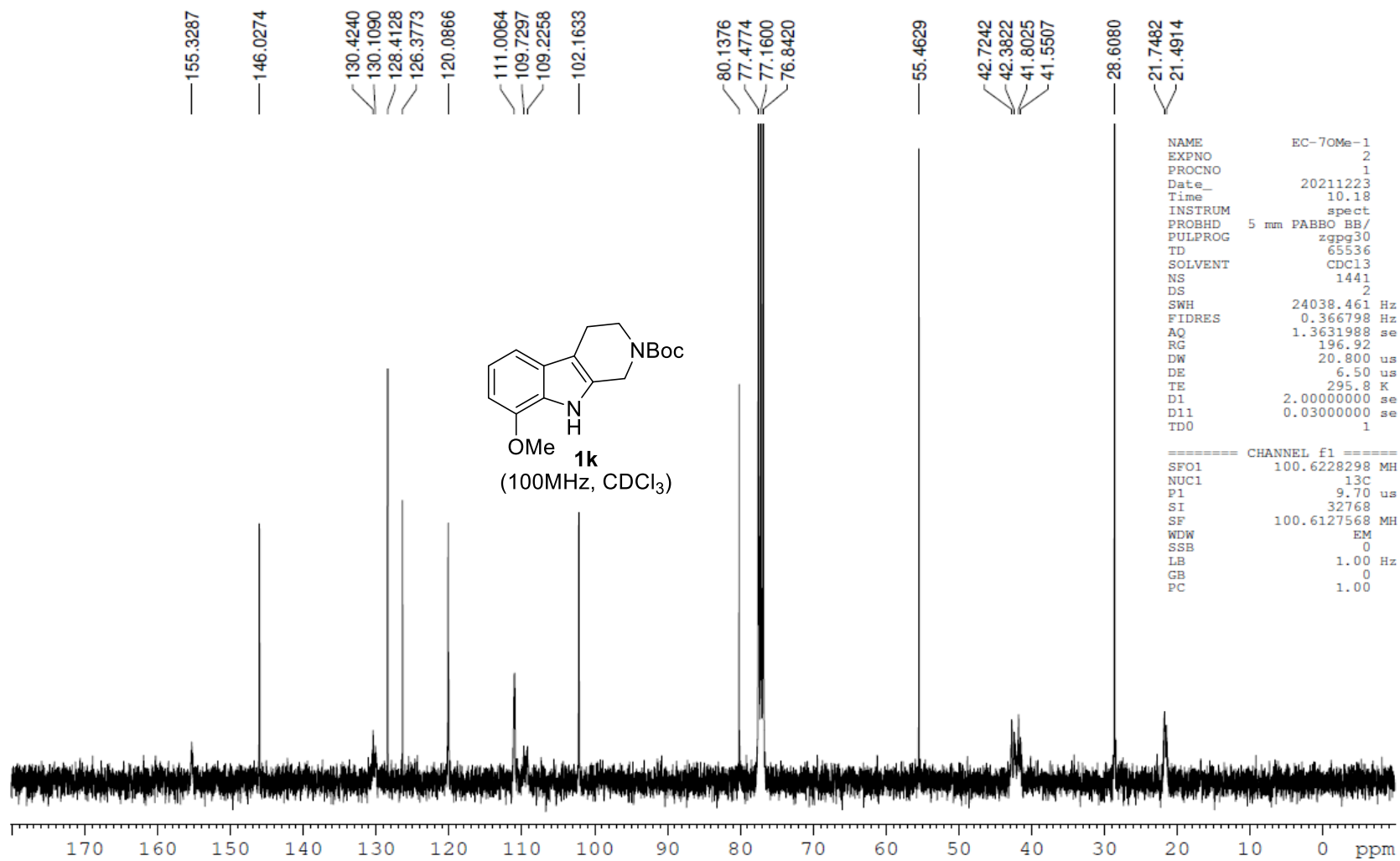
NAME      EC-7OMe-1
EXPNO    1
PROCNO   1
Date_    20211222
Time     22.18
INSTRUM  spect
PROBHD   5 mm PABBO BB/
PULPROG  zg30
TD       65536
SOLVENT  CDCl3
NS       16
DS       2
SWH      8012.820 Hz
FIDRES   0.122266 Hz
AQ       4.0894966 sec
RG       88.84
DW       62.400 usec
DE       6.50 usec
TE       296.1 K
D1       1.00000000 sec
TD0      1

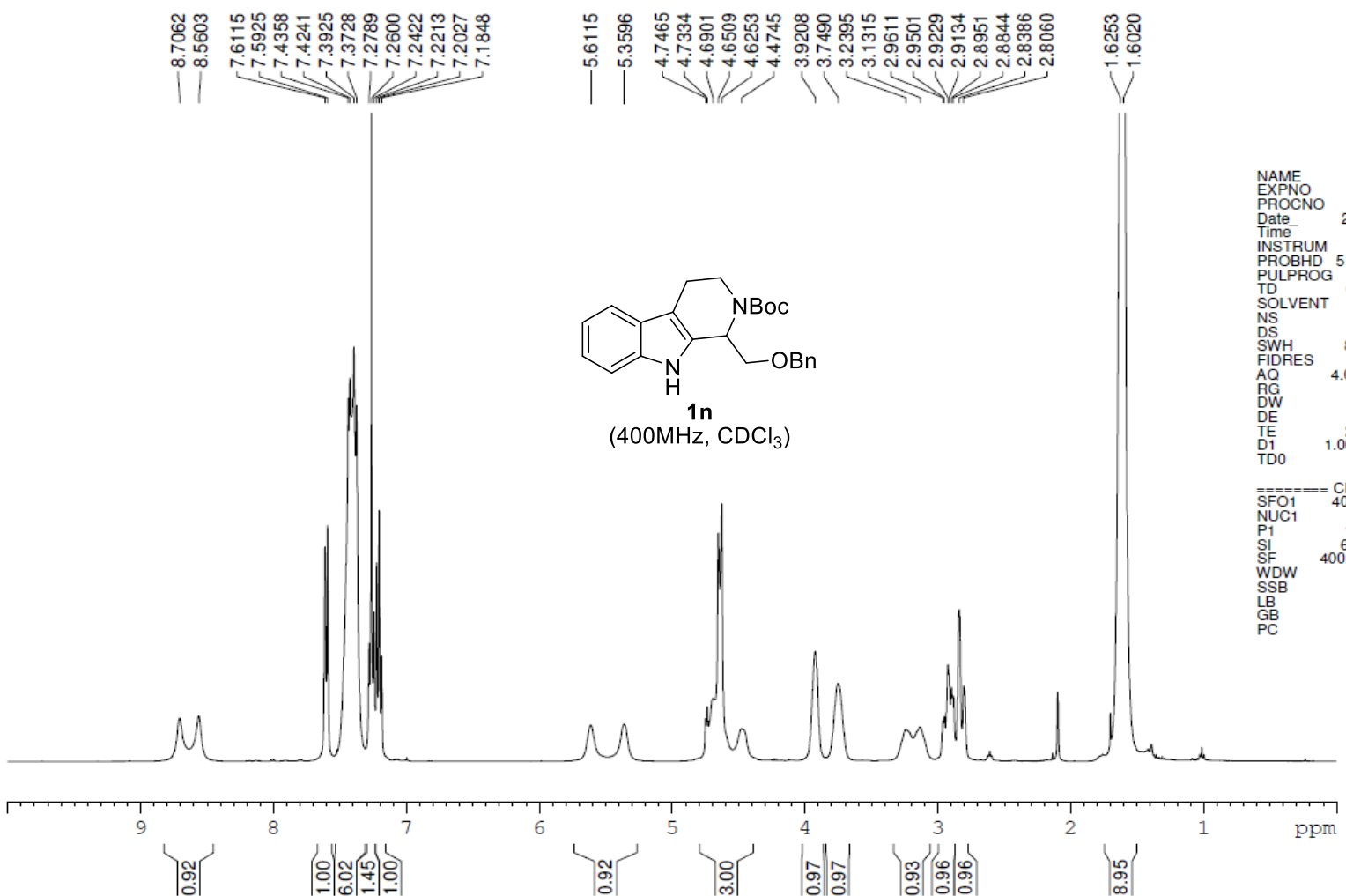
```

```

===== CHANNEL f1 =====
SFO1    400.1324710 MHz
NUC1     1H
P1      14.50 usec
SI      65536
SF      400.1300101 MHz
WDW     EM
SSB     0
LB      0.30 Hz
GB      0
PC      1.00

```



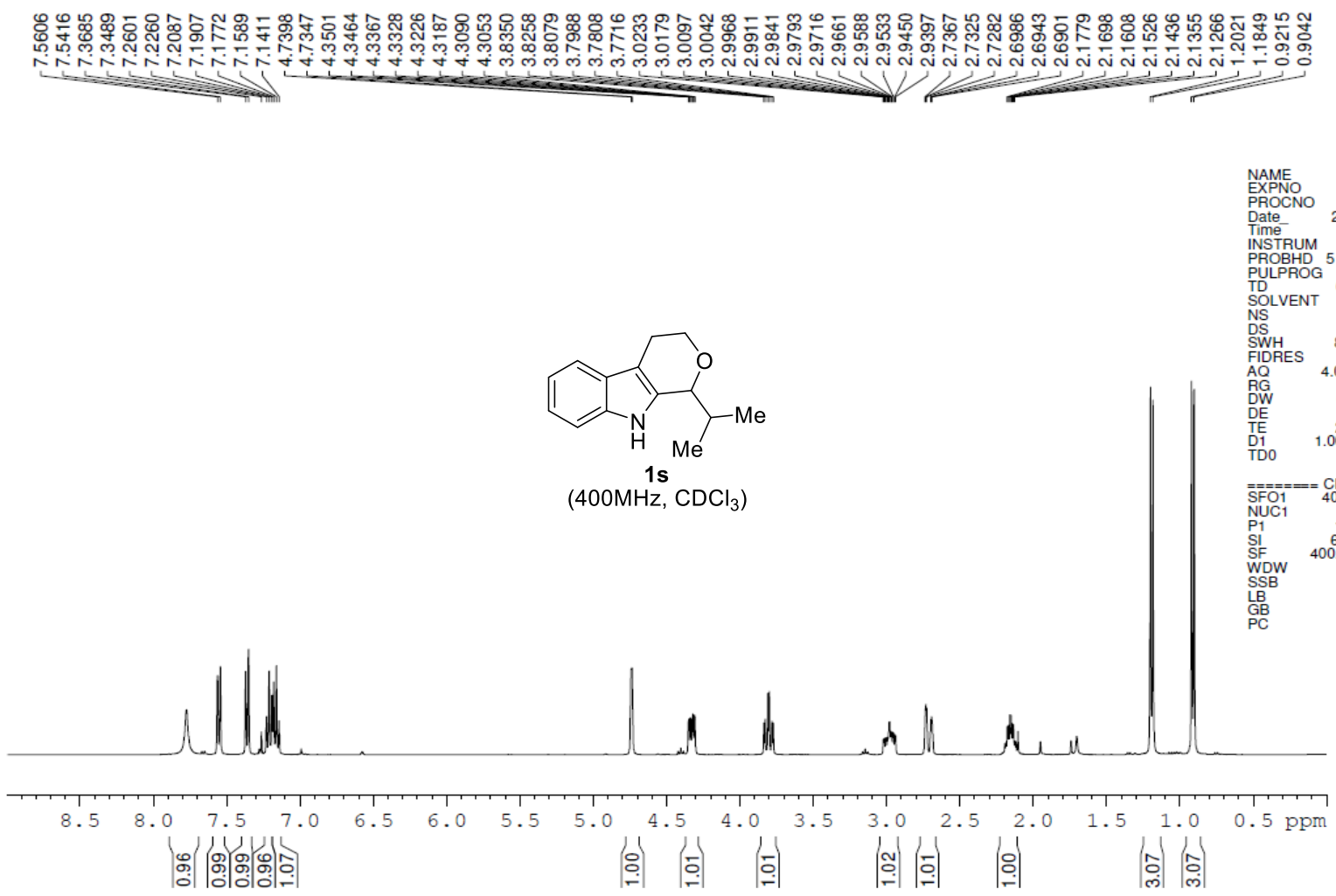


```

NAME      EC-OBn-1
EXPNO    1
PROCNO   1
Date_    20211222
Time     22.48
INSTRUM  spect
PROBHD   5 mm PABBO BB/
PULPROG  zg30
TD       65536
SOLVENT  CDCl3
NS       16
DS       2
SWH      8012.820 Hz
FIDRES   0.122266 Hz
AQ       4.0894966 sec
RG       12.56
DW       62.400 usec
DE       6.50 usec
TE       296.2 K
D1       1.00000000 sec
TD0      1
  
```

```

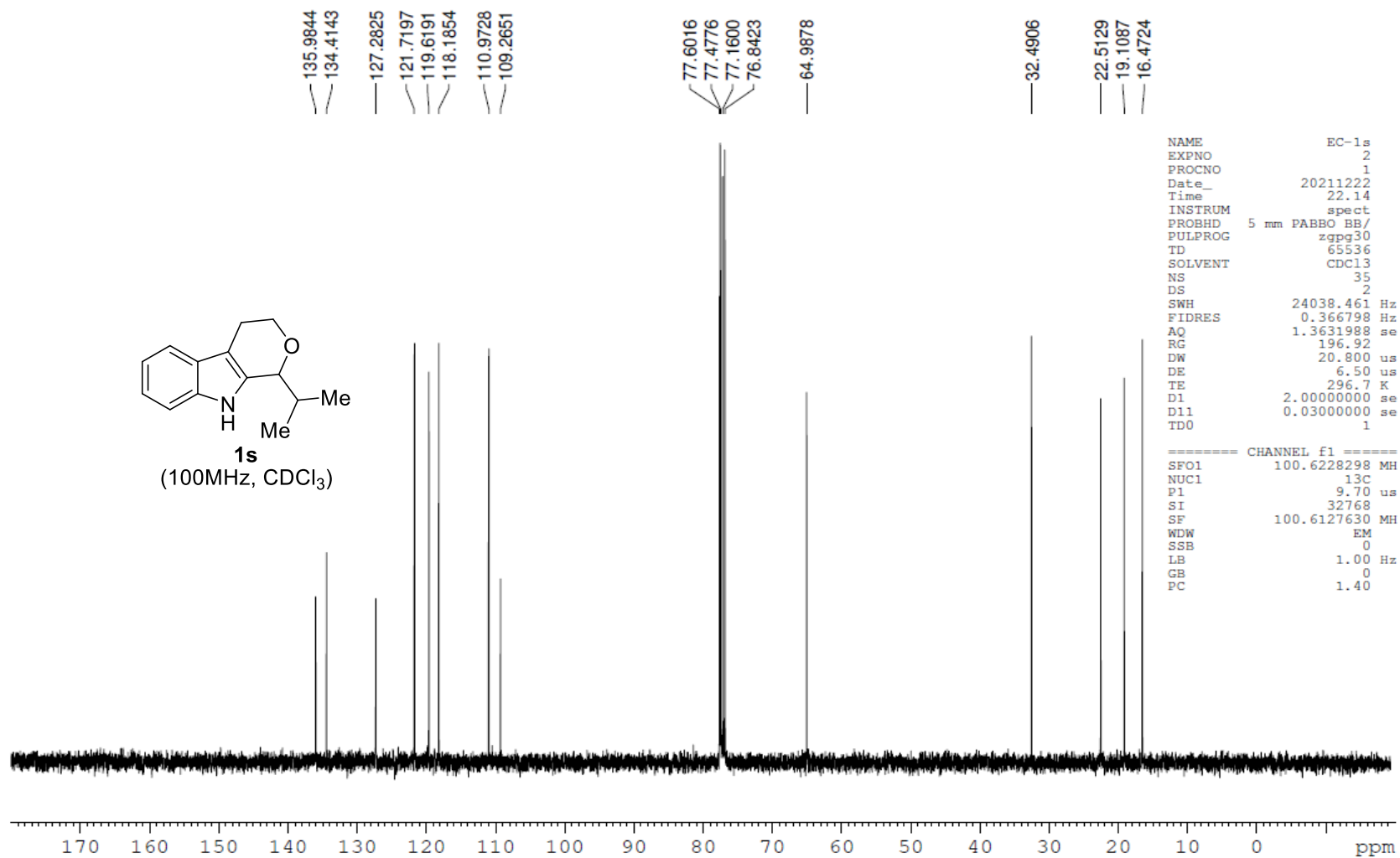
===== CHANNEL f1 =====
SFO1    400.1324710 MHz
NUC1    1H
P1      14.50 usec
SI      65536
SF      400.1300096 MHz
WDW     EM
SSB     0
LB      0.30 Hz
GB      0
PC      1.00
  
```

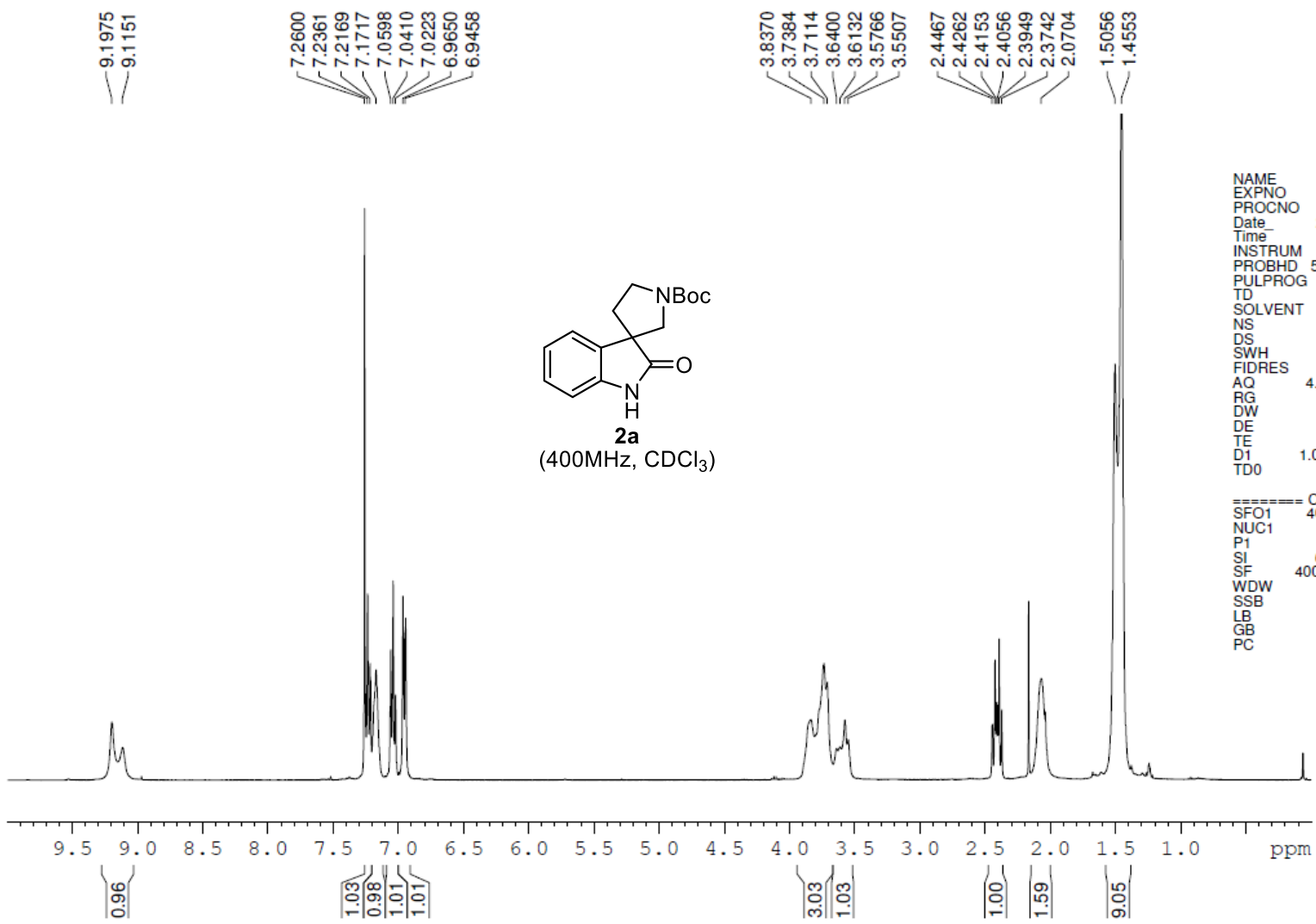



```

NAME      EC-1s
EXPNO     1
PROCNO    1
Date_     20211222
Time      22.10
INSTRUM   spect
PROBHD    5 mm PABBO BB/
PULPROG   zg30
TD         65536
SOLVENT   CDCl3
NS         16
DS         2
SWH        8012.820 Hz
FIDRES     0.122266 Hz
AQ         4.0894966 sec
RG         27.78
DW         62.400 usec
DE         6.50 usec
TE         296.0 K
D1         1.00000000 sec
TD0        1

===== CHANNEL f1 =====
SFO1      400.1324710 MHz
NUC1       1H
P1         14.50 usec
SI         65536
SF         400.1300099 MHz
WDW        EM
SSB        0
LB         0.30 Hz
GB         0
PC         1.00
  
```



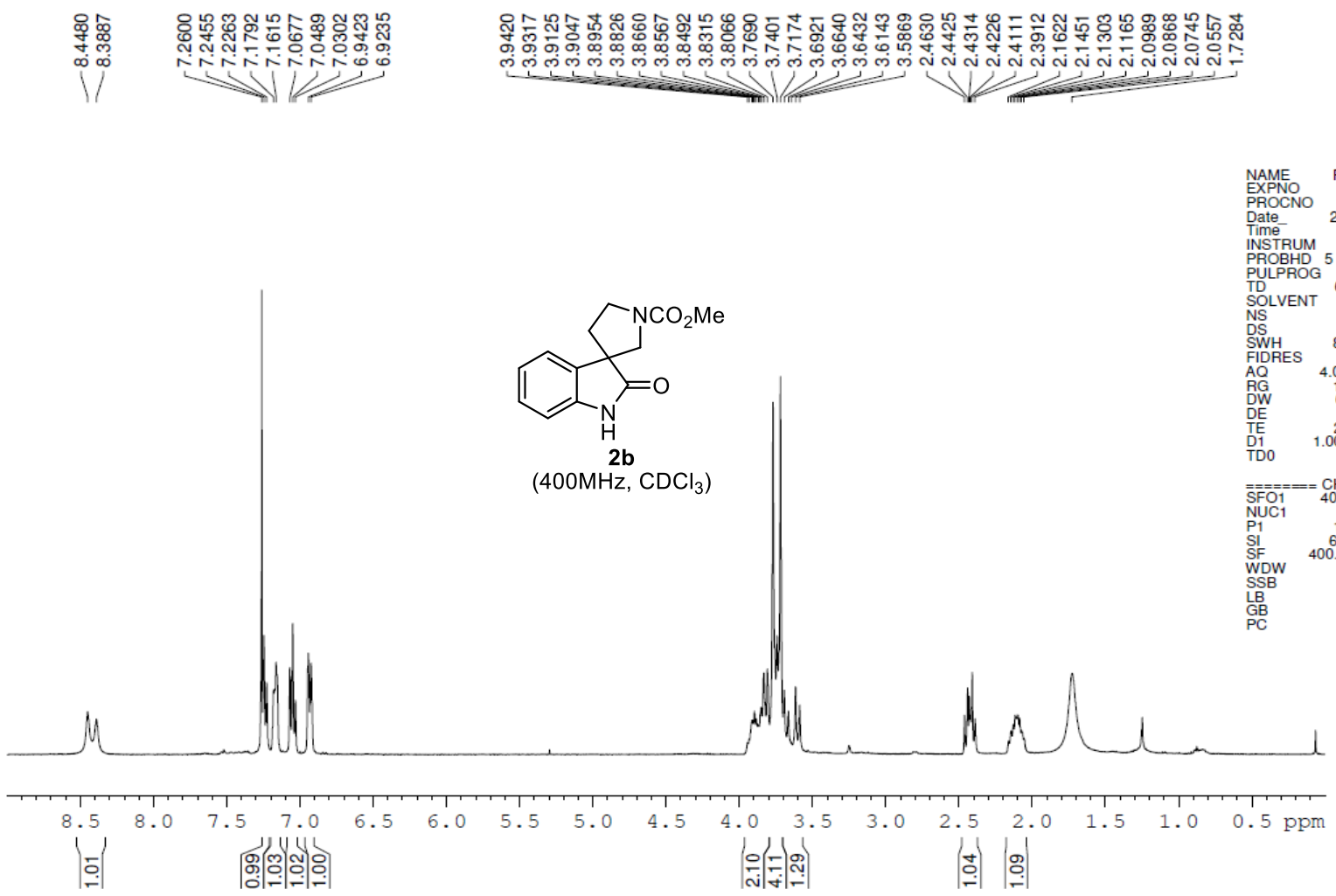


```

NAME      EC-Boc
EXPNO     3
PROCNO    1
Date_     20211211
Time      12.53
INSTRUM   spect
PROBHD    5 mm PABBO BB/
PULPROG   zg30
TD         65536
SOLVENT   CDCl3
NS         16
DS         2
SWH        8012.820 Hz
FIDRES     0.122266 Hz
AQ         4.0894966 sec
RG         34.77
DW         62.400 usec
DE         6.50 usec
TE         295.7 K
D1         1.00000000 sec
TD0        1

===== CHANNEL f1 =====
SFO1      400.1324710 MHz
NUC1       1H
P1         14.50 usec
SI         65536
SF         400.1300100 MHz
WDW        EM
SSB        0
LB         0.30 Hz
GB         0
PC         1.00

```

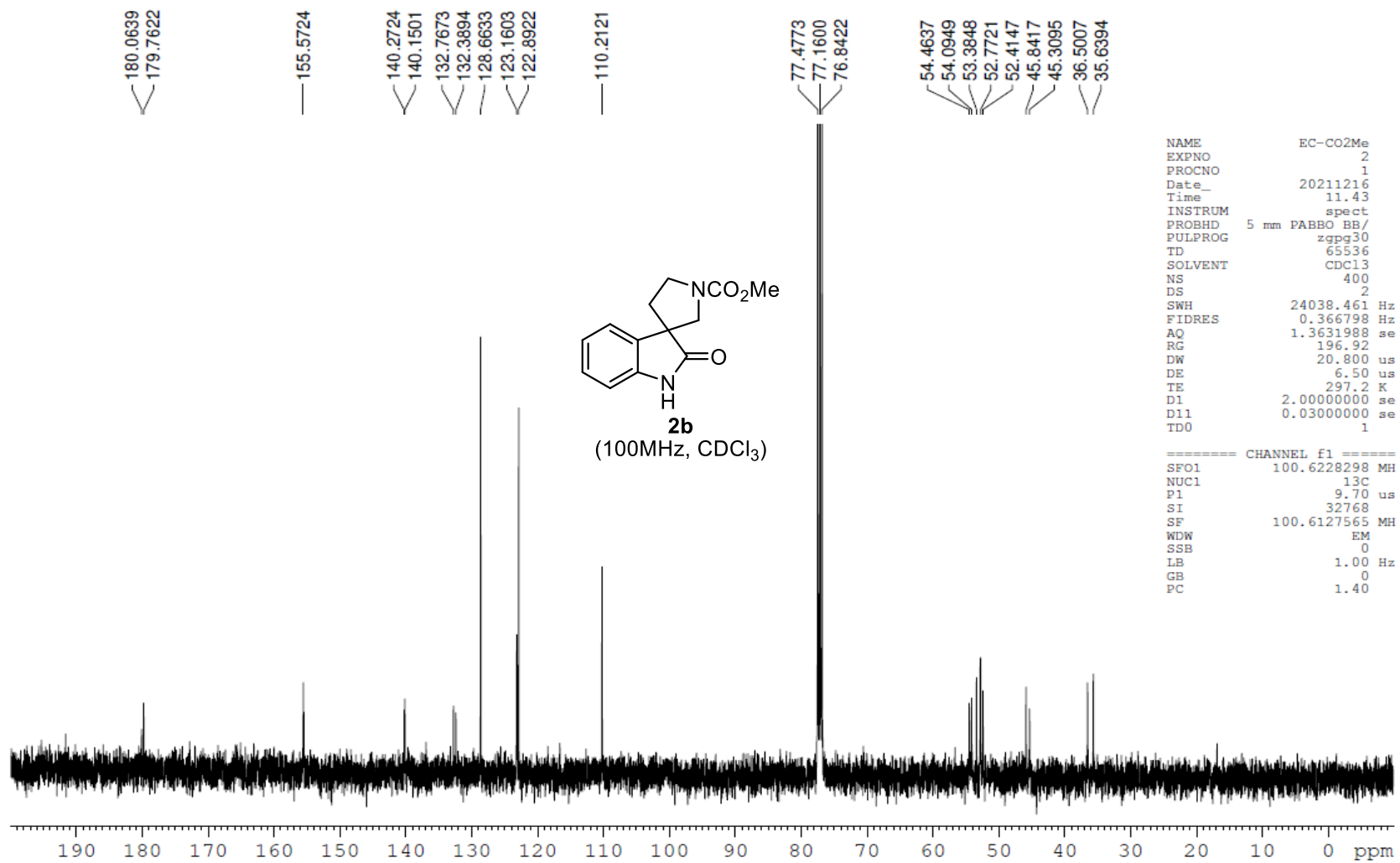



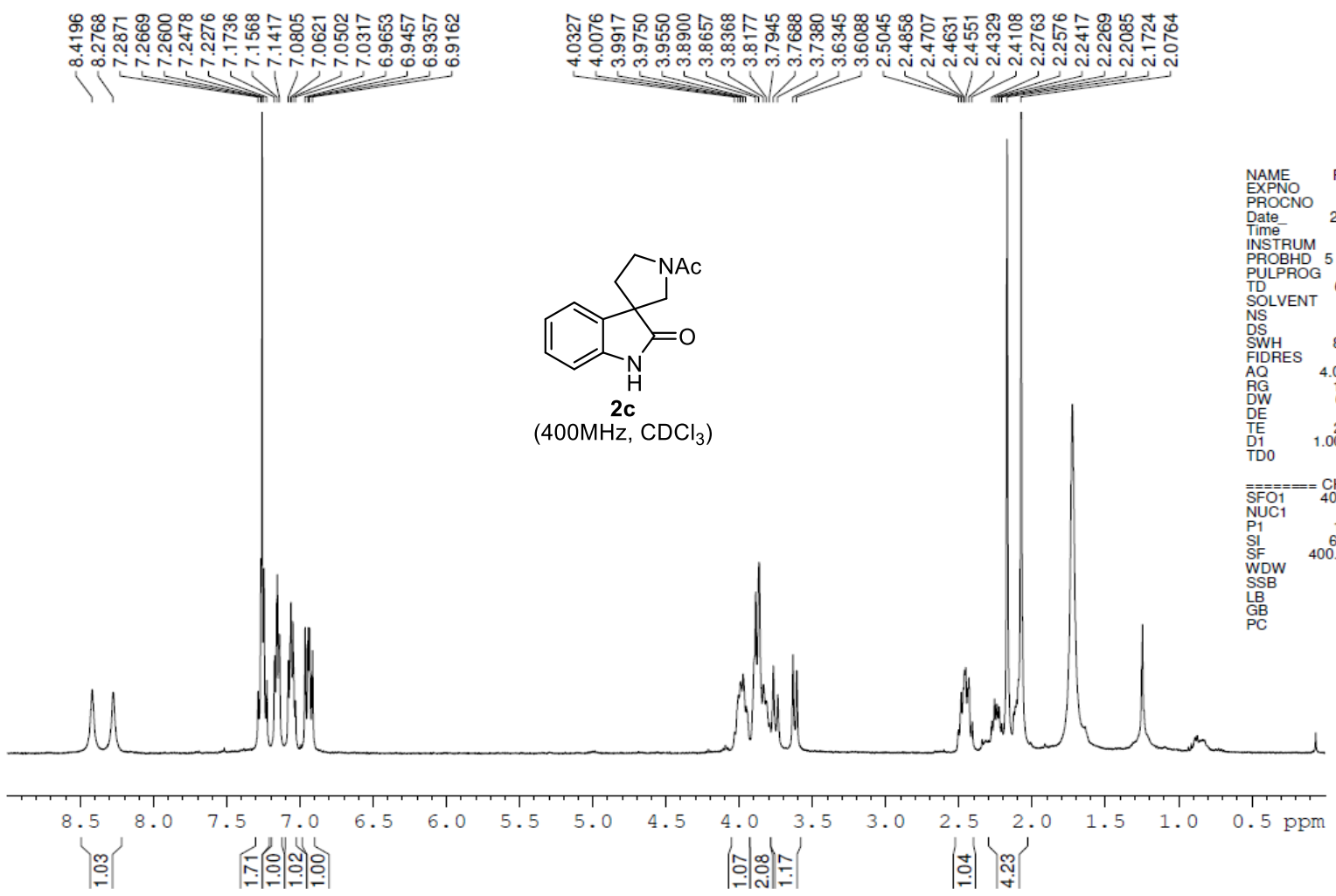
```

NAME      FZ-2307-1
EXPNO    2
PROCNO   1
Date_    20210525
Time     23.12
INSTRUM  spect
PROBHD   5 mm PABBO BB/
PULPROG  zg30
TD       65536
SOLVENT  CDCl3
NS       16
DS       2
SWH      8012.820 Hz
FIDRES   0.122266 Hz
AQ       4.0894966 sec
RG       126.97
DW       62.400 usec
DE       6.50 usec
TE       298.3 K
D1       1.00000000 sec
TD0      1

===== CHANNEL f1 =====
SFO1    400.1324710 MHz
NUC1    1H
P1      14.50 usec
SI      65536
SF      400.1300101 MHz
WDW     EM
SSB     0
LB      0.30 Hz
GB      0
PC      1.00

```



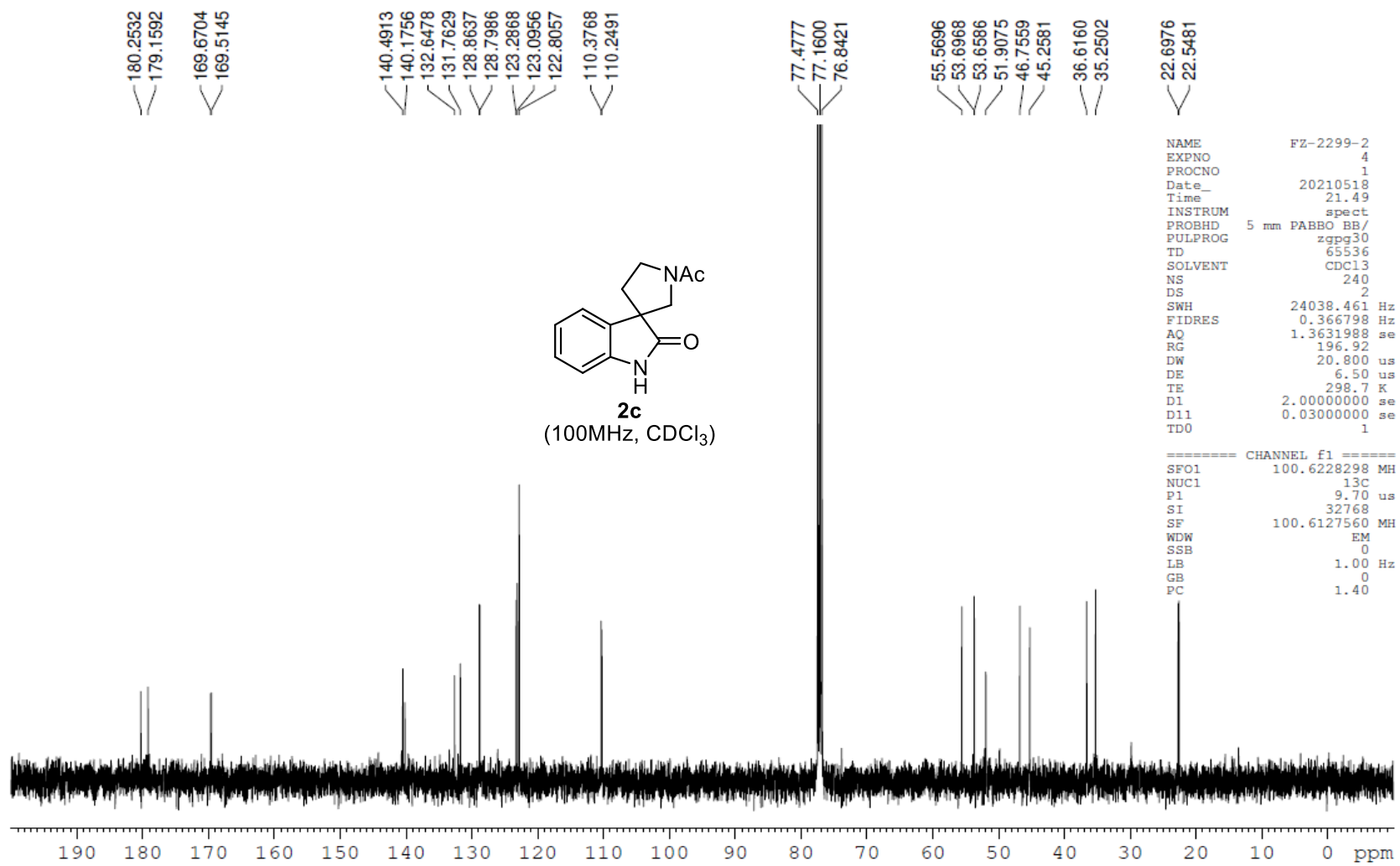


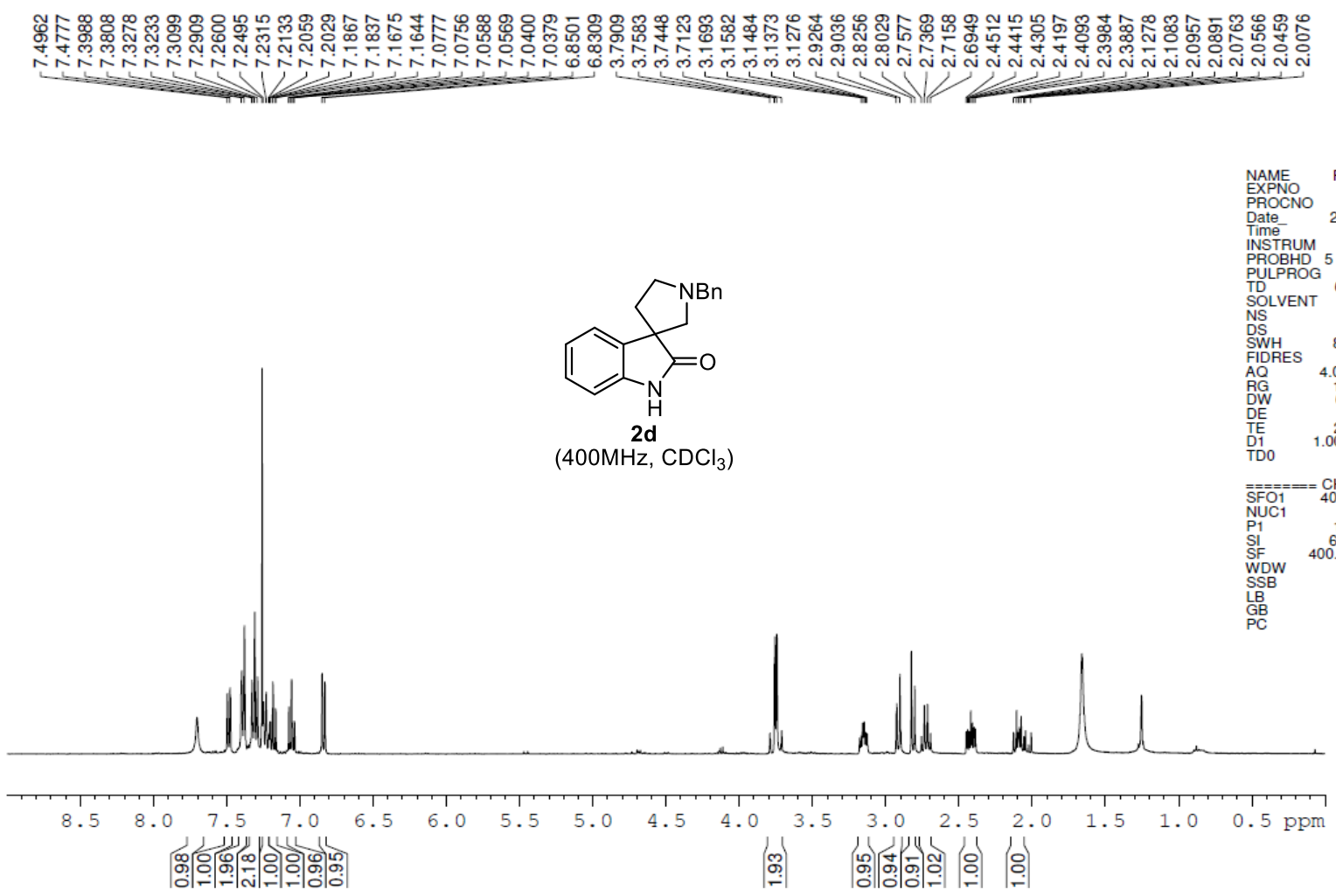
```

NAME      FZ-2299-2
EXPNO     6
PROCNO    1
Date_     20210527
Time      16.21
INSTRUM   spect
PROBHD    5 mm PABBO BB/
PULPROG   zg30
TD         65536
SOLVENT   CDCl3
NS         16
DS         2
SWH        8012.820 Hz
FIDRES     0.122266 Hz
AQ         4.0894966 sec
RG         142.88
DW         62.400 usec
DE         6.50 usec
TE         297.4 K
D1         1.00000000 sec
TD0        1

===== CHANNEL f1 =====
SFO1      400.1324710 MHz
NUC1       1H
P1         14.50 usec
SI         65536
SF         400.1300101 MHz
WDW        EM
SSB        0
LB         0.30 Hz
GB         0
PC         1.00

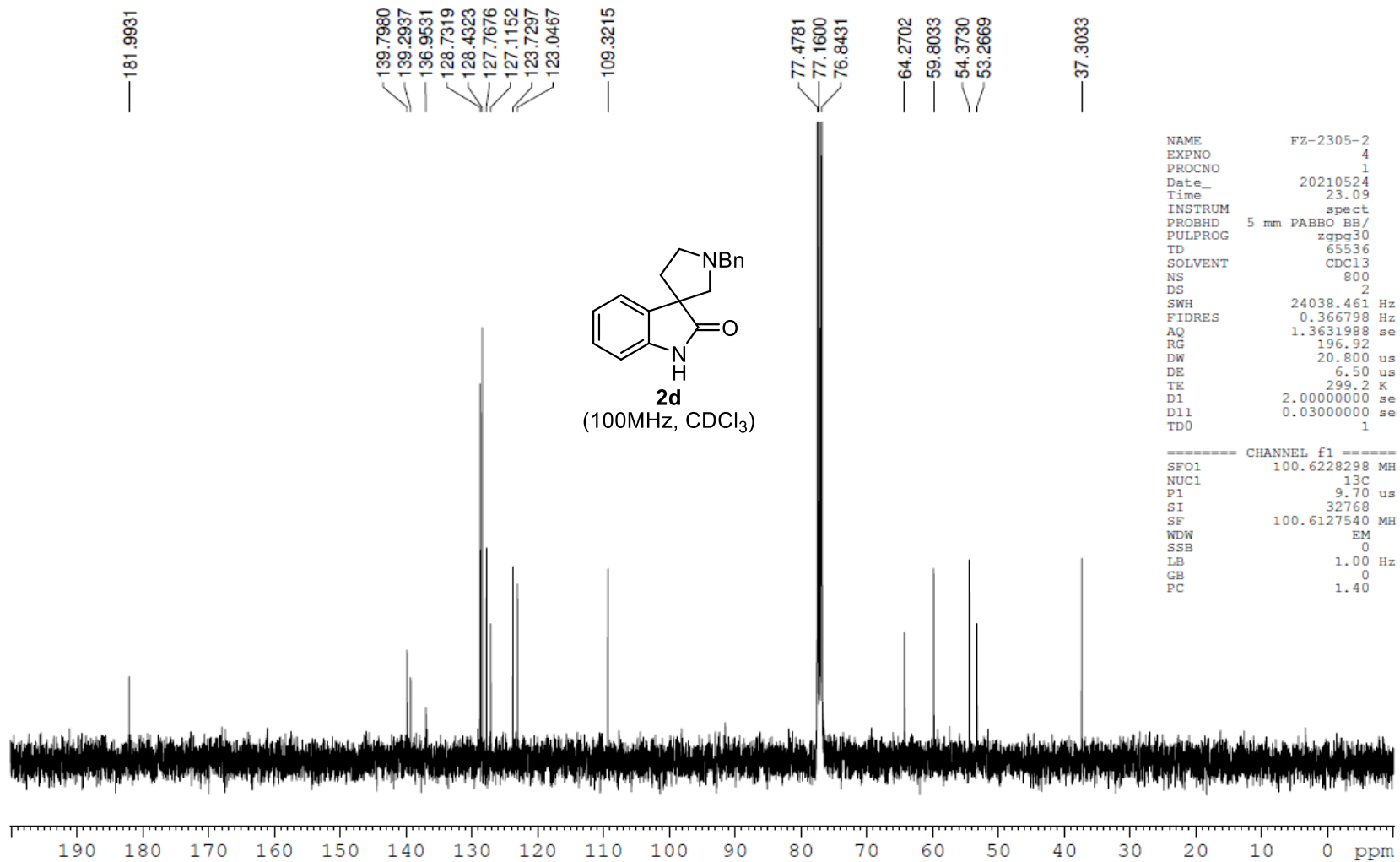
```

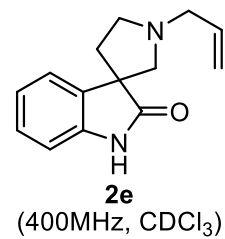
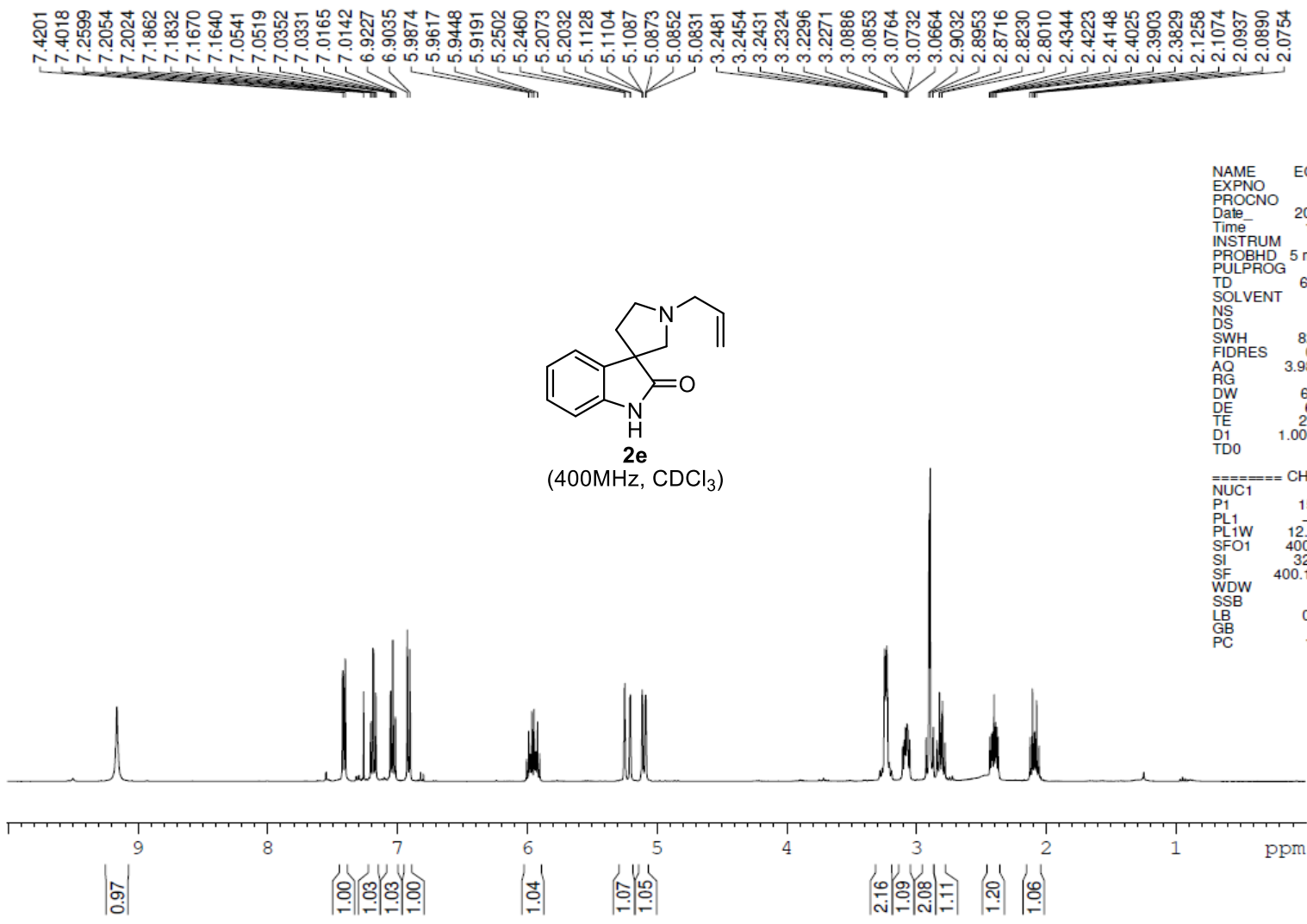





NAME FZ-2305-2
 EXPNO 1
 PROCNO 1
 Date_ 20210524
 Time 14.29
 INSTRUM spect
 PROBHD 5 mm PABBO BB/
 PULPROG zg30
 TD 65536
 SOLVENT CDCl3
 NS 16
 DS 2
 SWH 8012.820 Hz
 FIDRES 0.122266 Hz
 AQ 4.0894966 sec
 RG 164.33
 DW 62.400 usec
 DE 6.50 usec
 TE 297.8 K
 D1 1.00000000 sec
 TD0 1

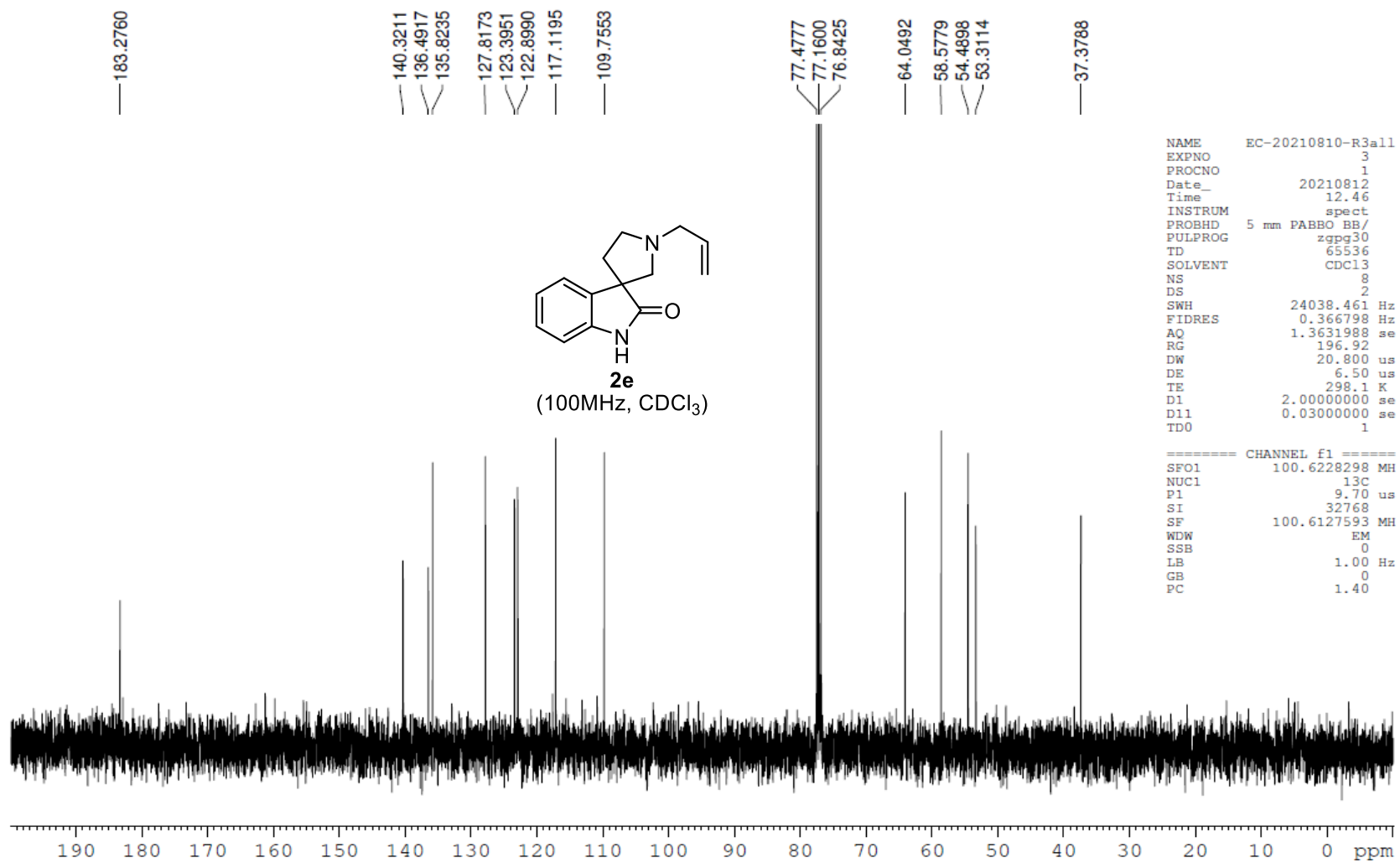
===== CHANNEL f1 =====
 SFO1 400.1324710 MHz
 NUC1 1H
 P1 14.50 usec
 SI 65536
 SF 400.1300100 MHz
 WDW EM
 SSB 0
 LB 0.30 Hz
 GB 0
 PC 1.00

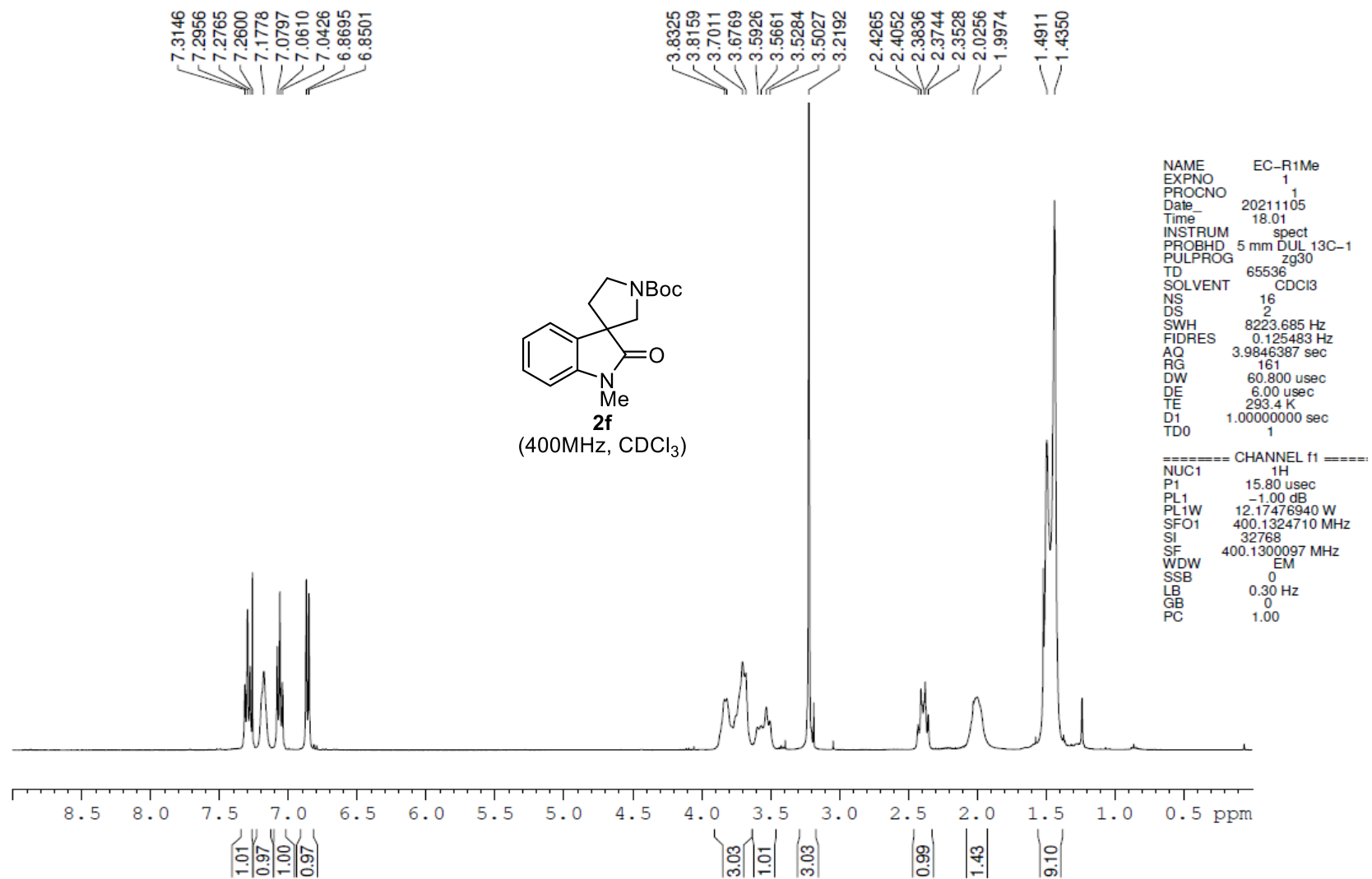


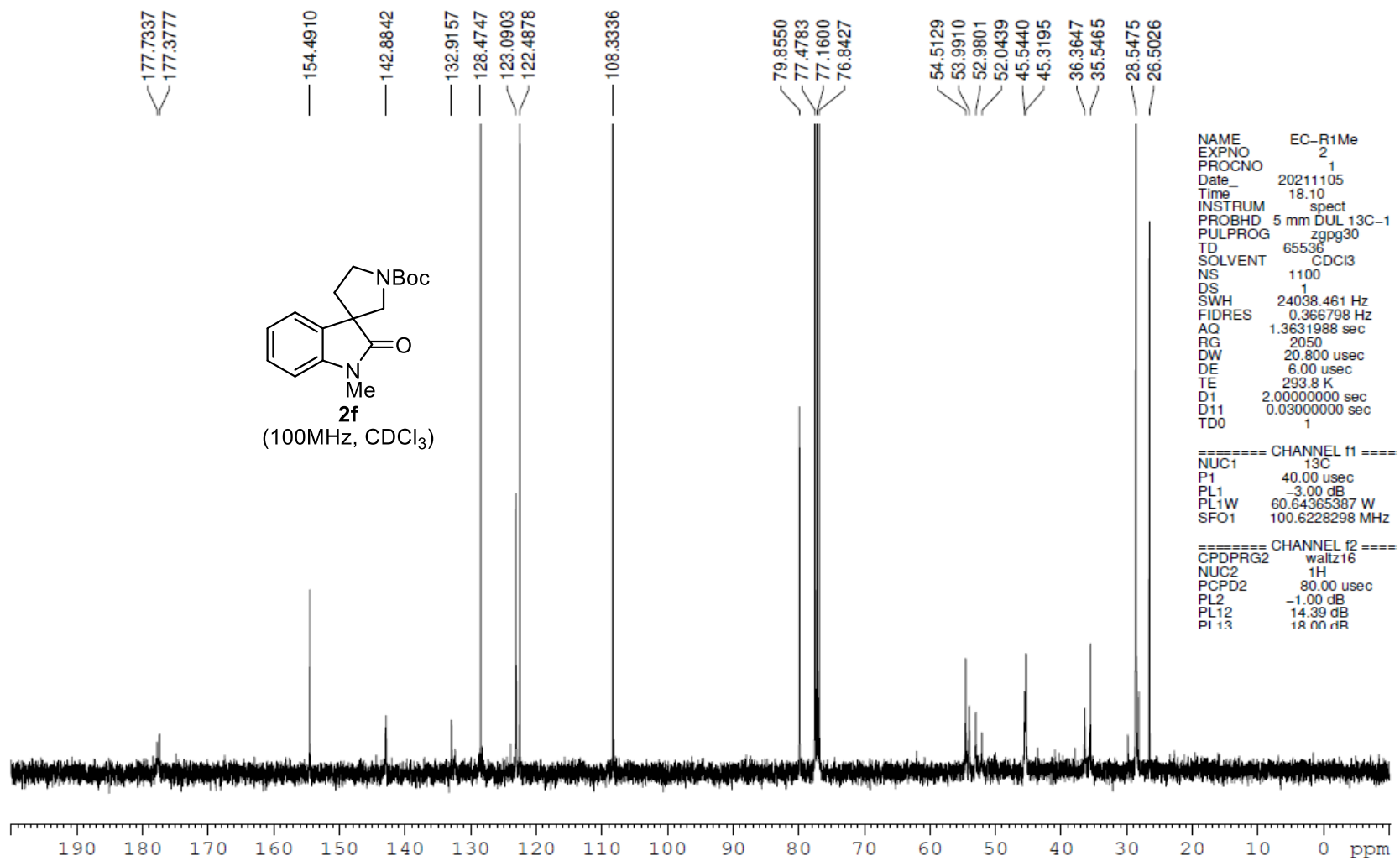


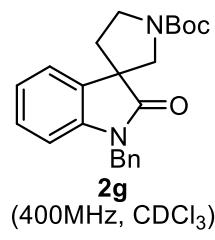
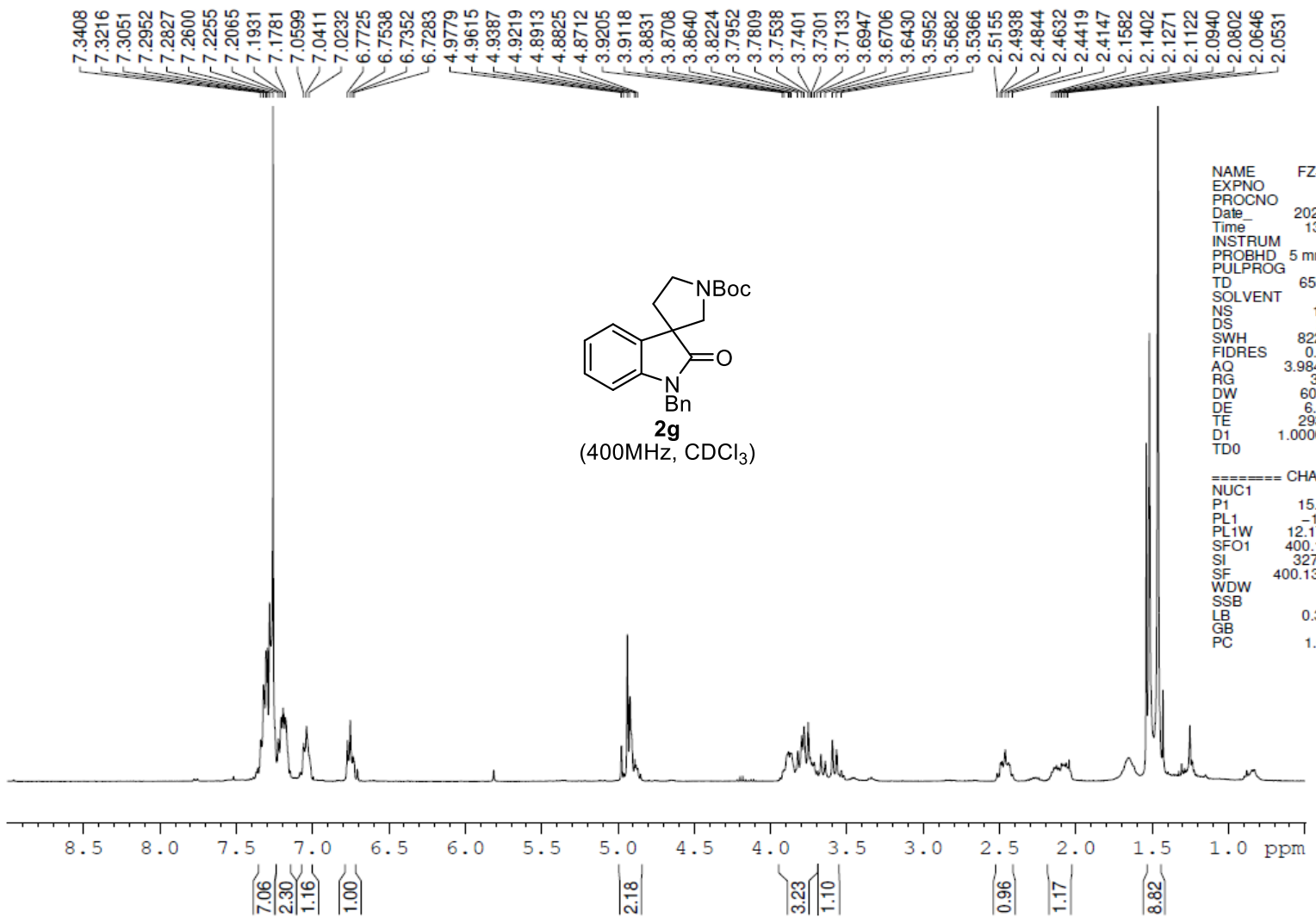
NAME EC-R3allyl
 EXPNO 1
 PROCNO 1
 Date 20211209
 Time 18.08
 INSTRUM spect
 PROBHD 5 mm DUL 13C-1
 PULPROG zg30
 TD 65536
 SOLVENT CDCl3
 NS 16
 DS 2
 SWH 8223.685 Hz
 FIDRES 0.125483 Hz
 AQ 3.9846387 sec
 RG 203
 DW 60.800 usec
 DE 6.00 usec
 TE 293.4 K
 D1 1.00000000 sec
 TD0 1

===== CHANNEL f1 =====
 NUC1 1H
 P1 15.80 usec
 PL1 -1.00 dB
 PL1W 12.17476940 W
 SFO1 400.1324710 MHz
 SI 32768
 SF 400.1300099 MHz
 WDW EM
 SSB 0
 LB 0.30 Hz
 GB 0
 PC 1.00







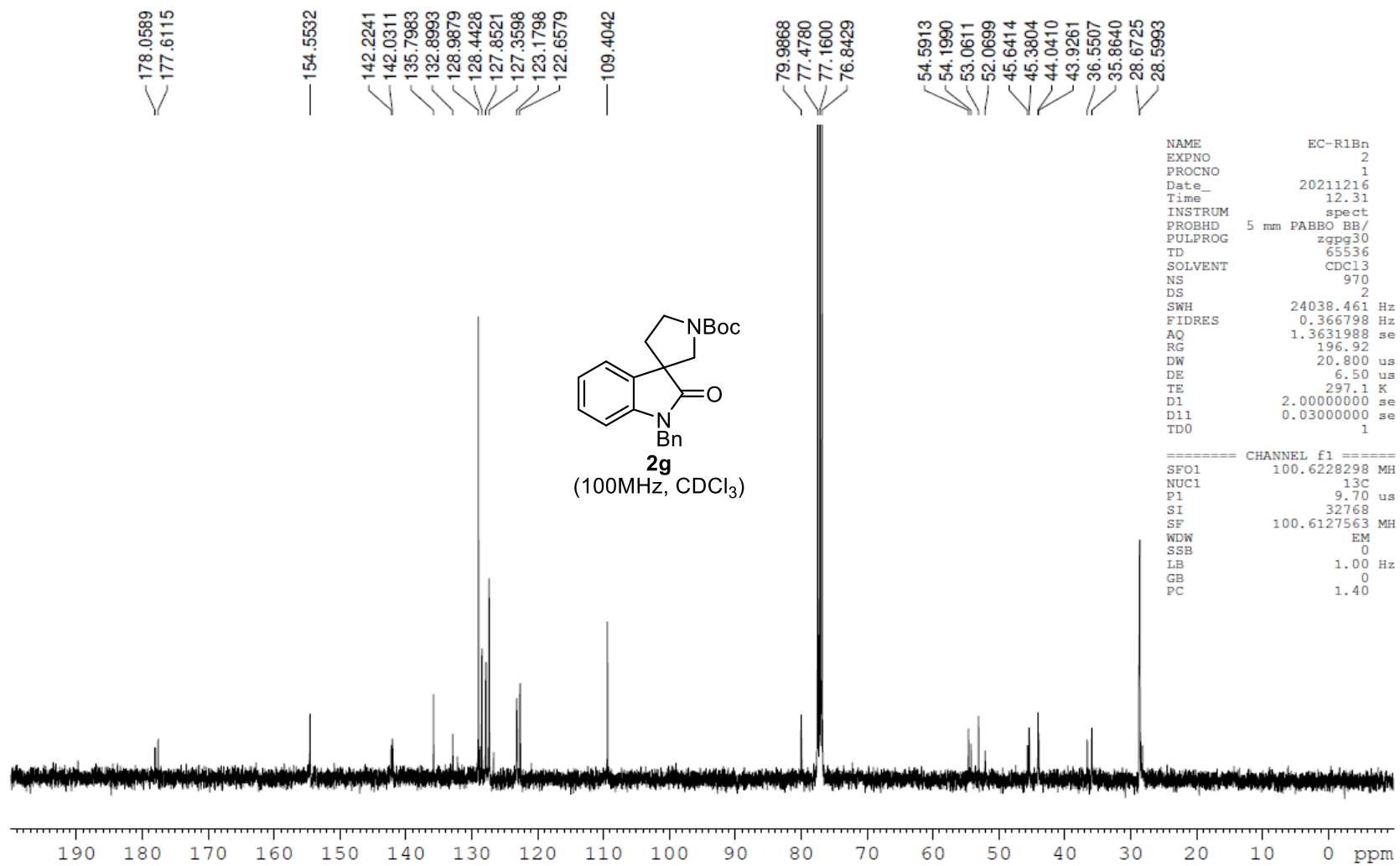


```

NAME      FZ-3007-3
EXPNO     1
PROCNO    1
Date_     20210604
Time      13.30
INSTRUM   spect
PROBHD    5 mm DUL 13C-1
PULPROG   zg30
TD         65536
SOLVENT   CDCl3
NS         16
DS         2
SWH       8223.685 Hz
FIDRES    0.125483 Hz
AQ        3.9846387 sec
RG         362
DW        60.800 usec
DE        6.00 usec
TE        293.5 K
D1        1.00000000 sec
TD0       1

===== CHANNEL f1 =====
NUC1      1H
P1        15.80 usec
PL1       -1.00 dB
PL1W      12.17476940 W
SFO1      400.1324710 MHz
SI        32768
SF        400.1300099 MHz
WDW       EM
SSB       0
LB        0.30 Hz
GB        0
PC        1.00

```

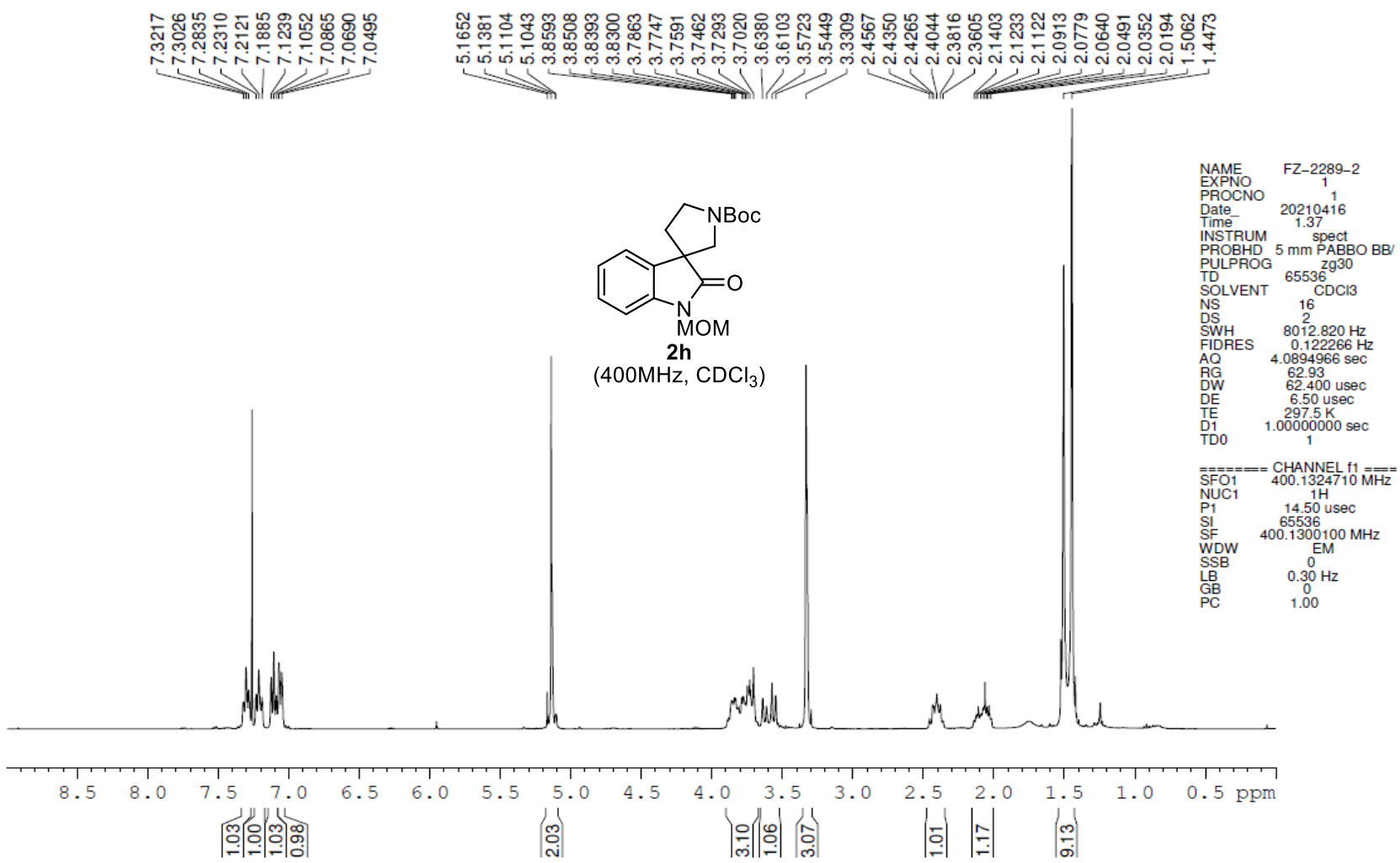



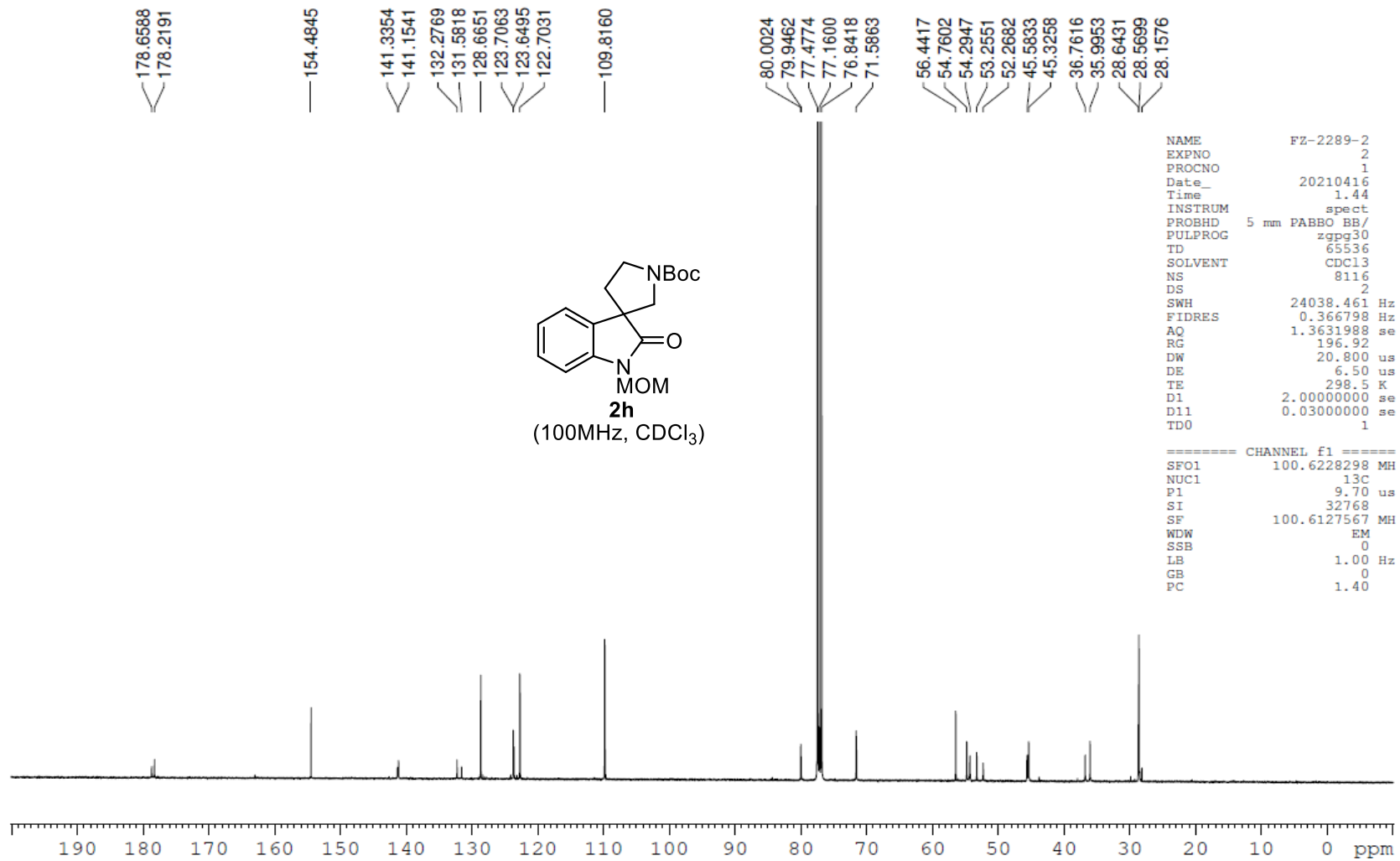
```

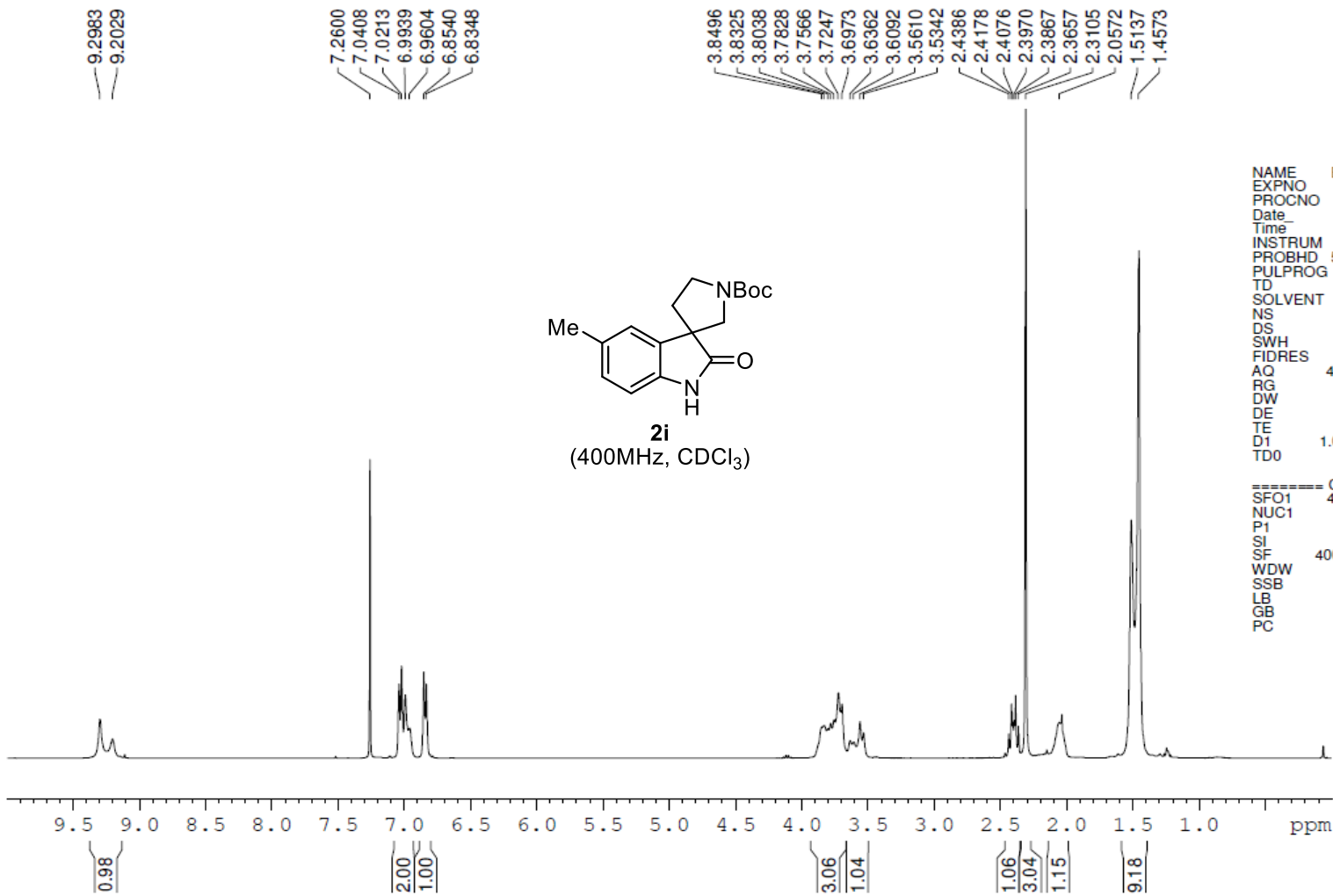
NAME          EC-R1Bn
EXPNO         2
PROCNO        1
Date_         20211216
Time_         12.31
INSTRUM       spect
PROBHD        5 mm PABBO BB/
PULPROG       zgpg30
TD            65536
SOLVENT       CDC13
NS            970
DS            2
SWH           24038.461 Hz
FIDRES        0.366798 Hz
AQ            1.3631988 sec
RG            196.92
RG            196.92
DW            20.800 us
DE            6.50 us
TE            297.1 K
D1            2.00000000 sec
D11           0.03000000 sec
TD0           1
  
```

```

===== CHANNEL f1 =====
SFO1         100.6228298 MH
NUC1          13C
P1            9.70 us
SI           32768
SF           100.6127563 MH
WDW           EM
SSB           0
LB            1.00 Hz
GB            0
PC            1.40
  
```





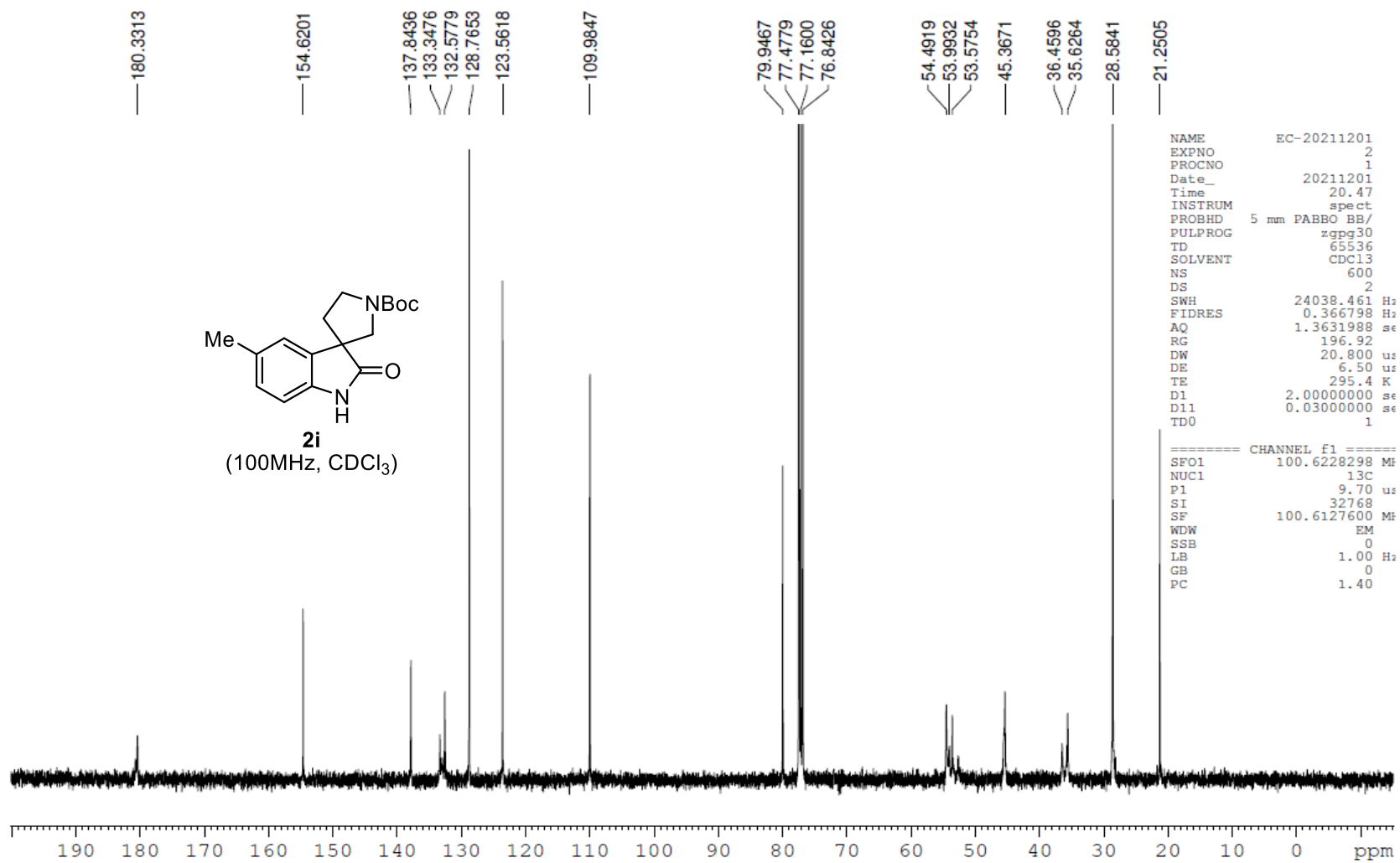


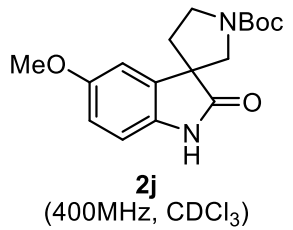
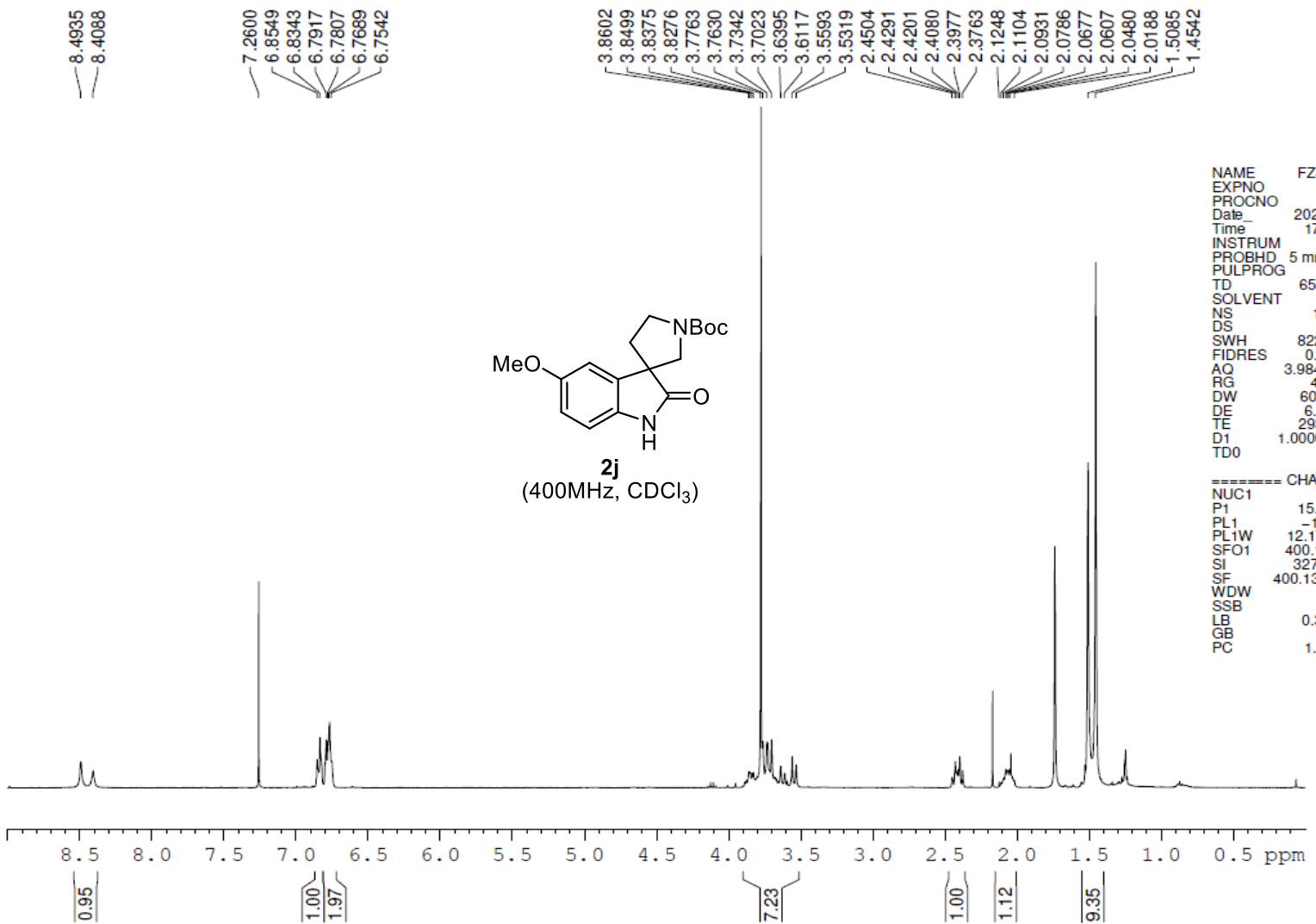
```

NAME      EC-20211201
EXPNO     1
PROCNO    1
Date_     20211201
Time      20.19
INSTRUM   spect
PROBHD    5 mm PABBO BB/
PULPROG   zg30
TD         65536
SOLVENT   CDCl3
NS         16
DS         2
SWH        8012.820 Hz
FIDRES     0.122266 Hz
AQ         4.0894966 sec
RG         27.78
DW         62.400 usec
DE         6.50 usec
TE         294.8 K
D1         1.00000000 sec
TD0        1

===== CHANNEL f1 =====
SFO1      400.1324710 MHz
NUC1       1H
P1         14.50 usec
SI         65536
SF         400.1300099 MHz
WDW        EM
SSB        0
LB         0.30 Hz
GB         0
PC         1.00

```

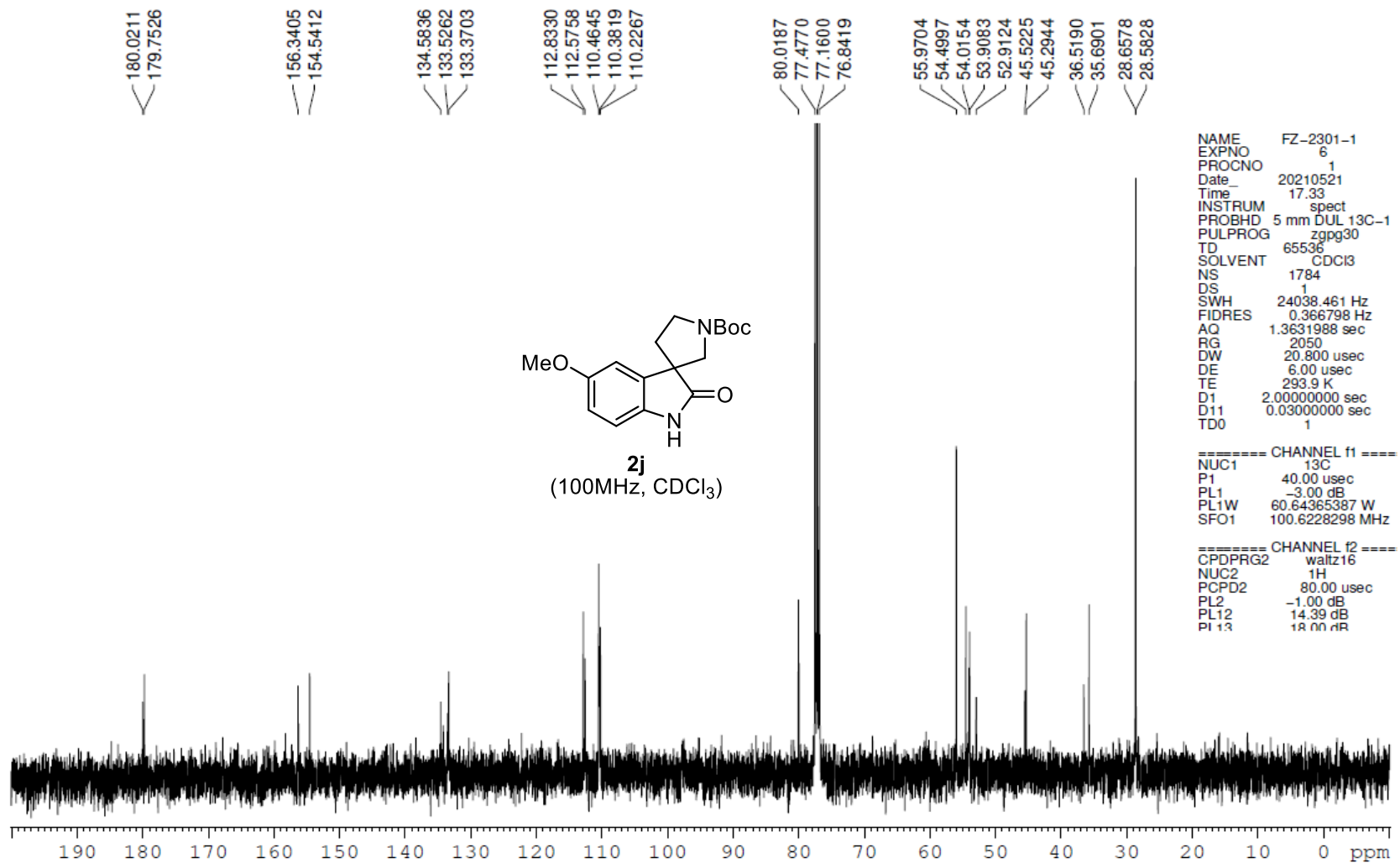


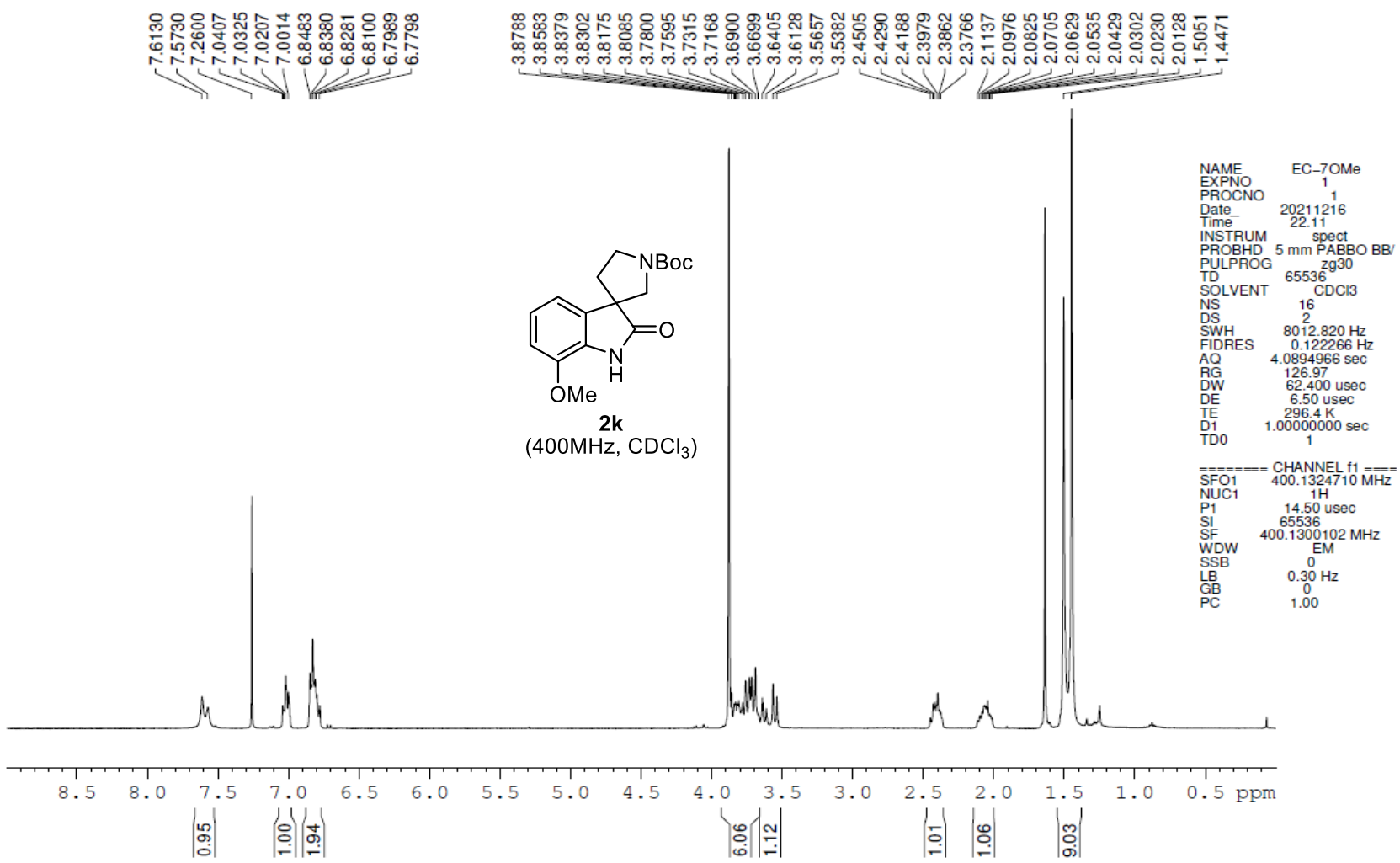


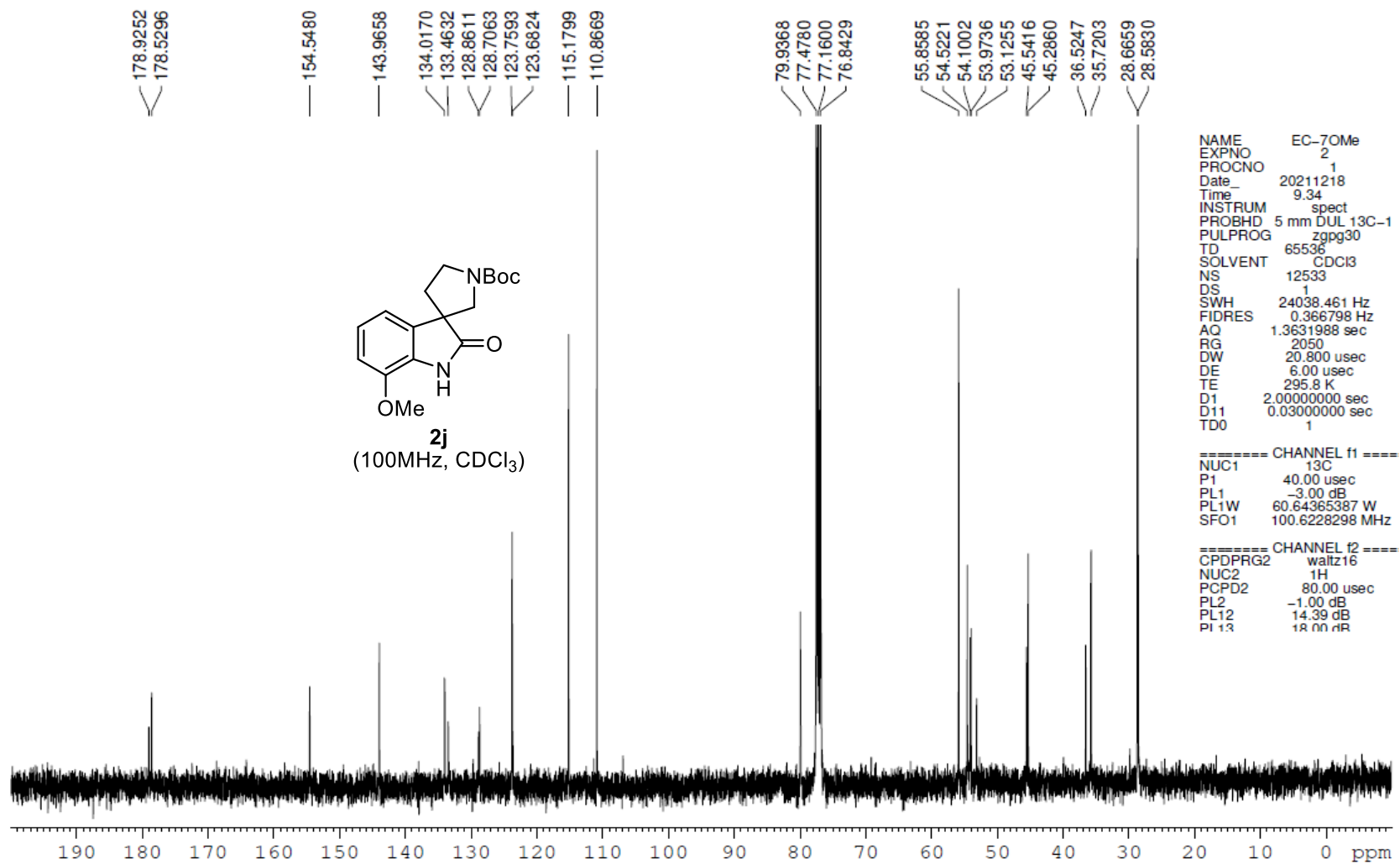
```

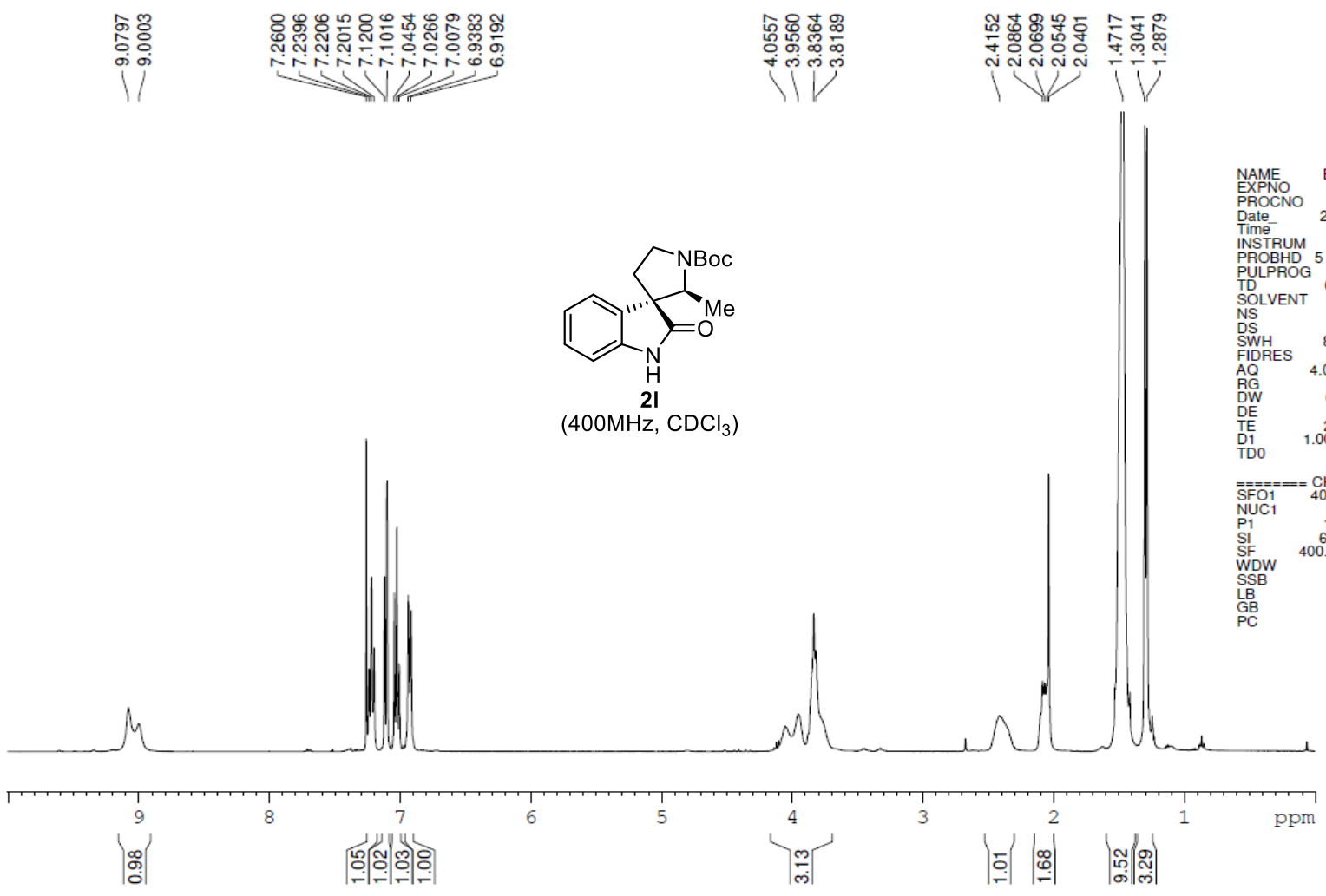
NAME      FZ-2301-1
EXPNO     5
PROCNO    1
Date_     20210521
Time      17.26
INSTRUM   spect
PROBHD    5 mm DUL 13C-1
PULPROG   zg30
TD         65536
SOLVENT   CDCl3
NS         16
DS         2
SWH        8223.685 Hz
FIDRES     0.125483 Hz
AQ         3.9846387 sec
RG         406
DW         60.800 usec
DE         6.00 usec
TE         293.5 K
D1         1.00000000 sec
TD0        1

===== CHANNEL f1 =====
NUC1       1H
P1         15.80 usec
PL1        -1.00 dB
PL1W       12.17476940 W
SFO1       400.1324710 MHz
SI         32768
SF         400.1300099 MHz
WDW        EM
SSB        0
LB         0.30 Hz
GB         0
PC         1.00
  
```







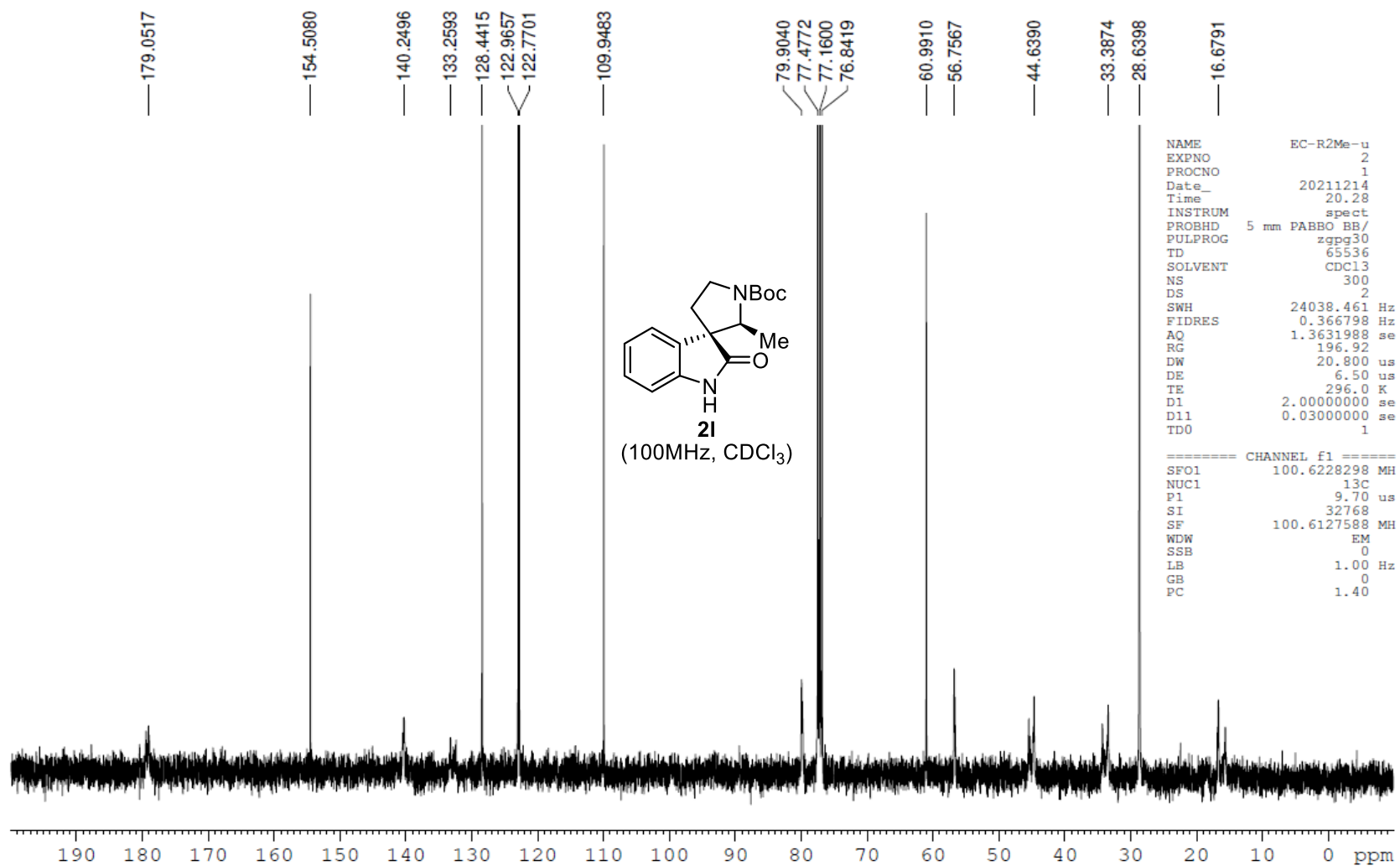


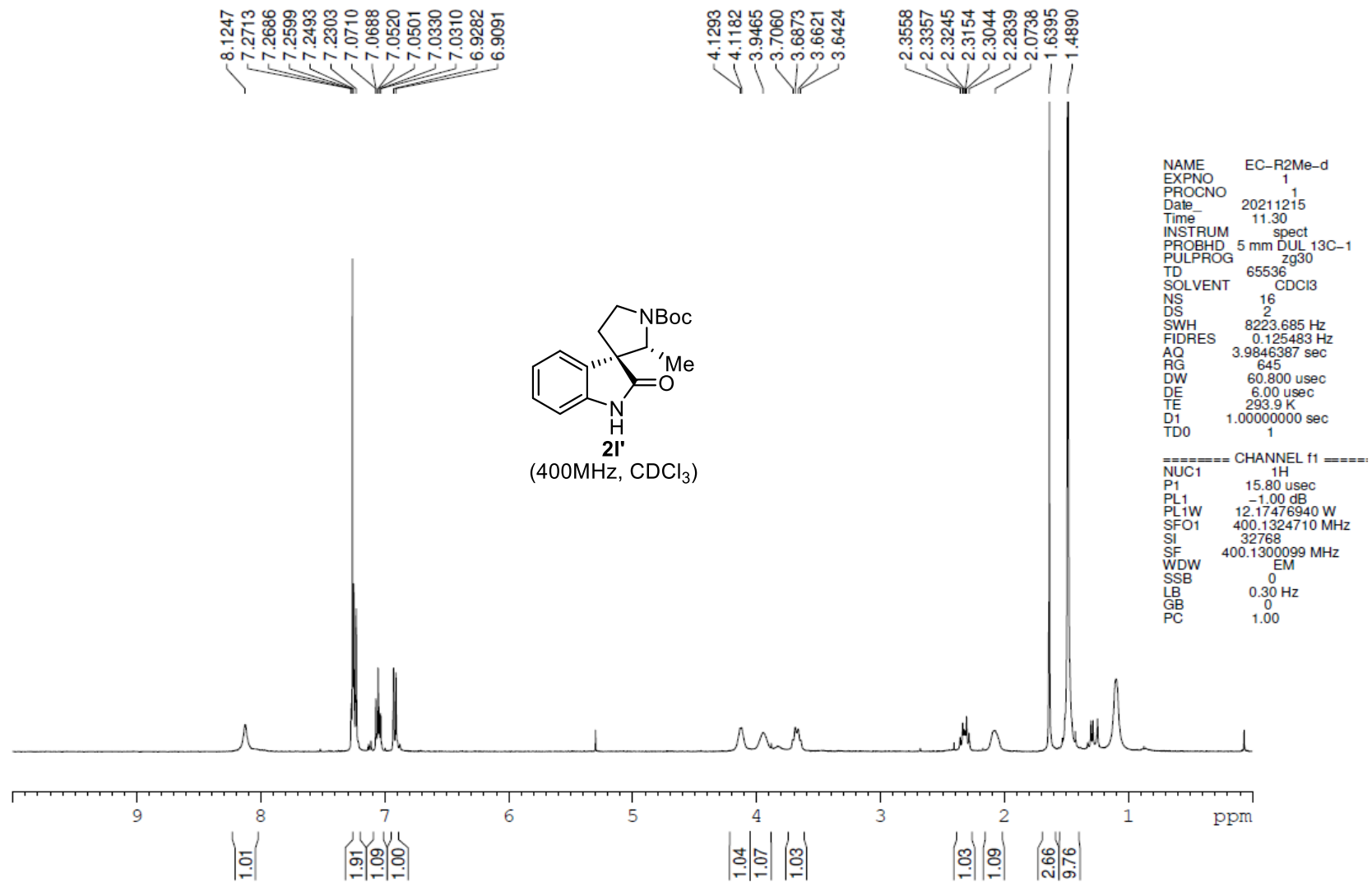
```

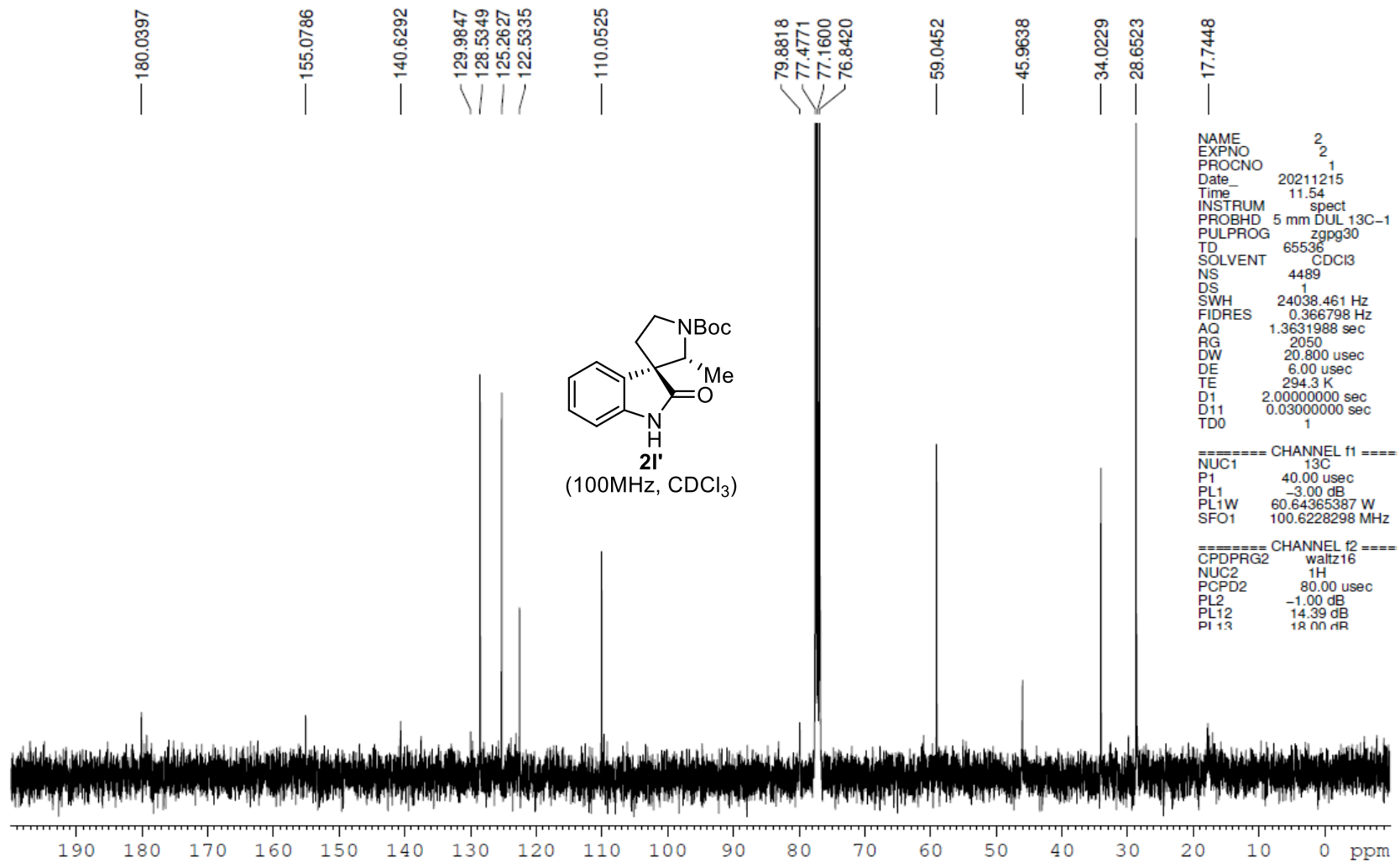
NAME      EC-R2Me-u
EXPNO    1
PROCNO   1
Date_    20211214
Time     20.23
INSTRUM  spect
PROBHD   5 mm PABBO BB/
PULPROG  zg30
TD       65536
SOLVENT  CDCl3
NS       16
DS       2
SWH      8012.820 Hz
FIDRES   0.122266 Hz
AQ       4.0894966 sec
RG       31.55
DW       62.400 usec
DE       6.50 usec
TE       295.1 K
D1       1.00000000 sec
TD0      1

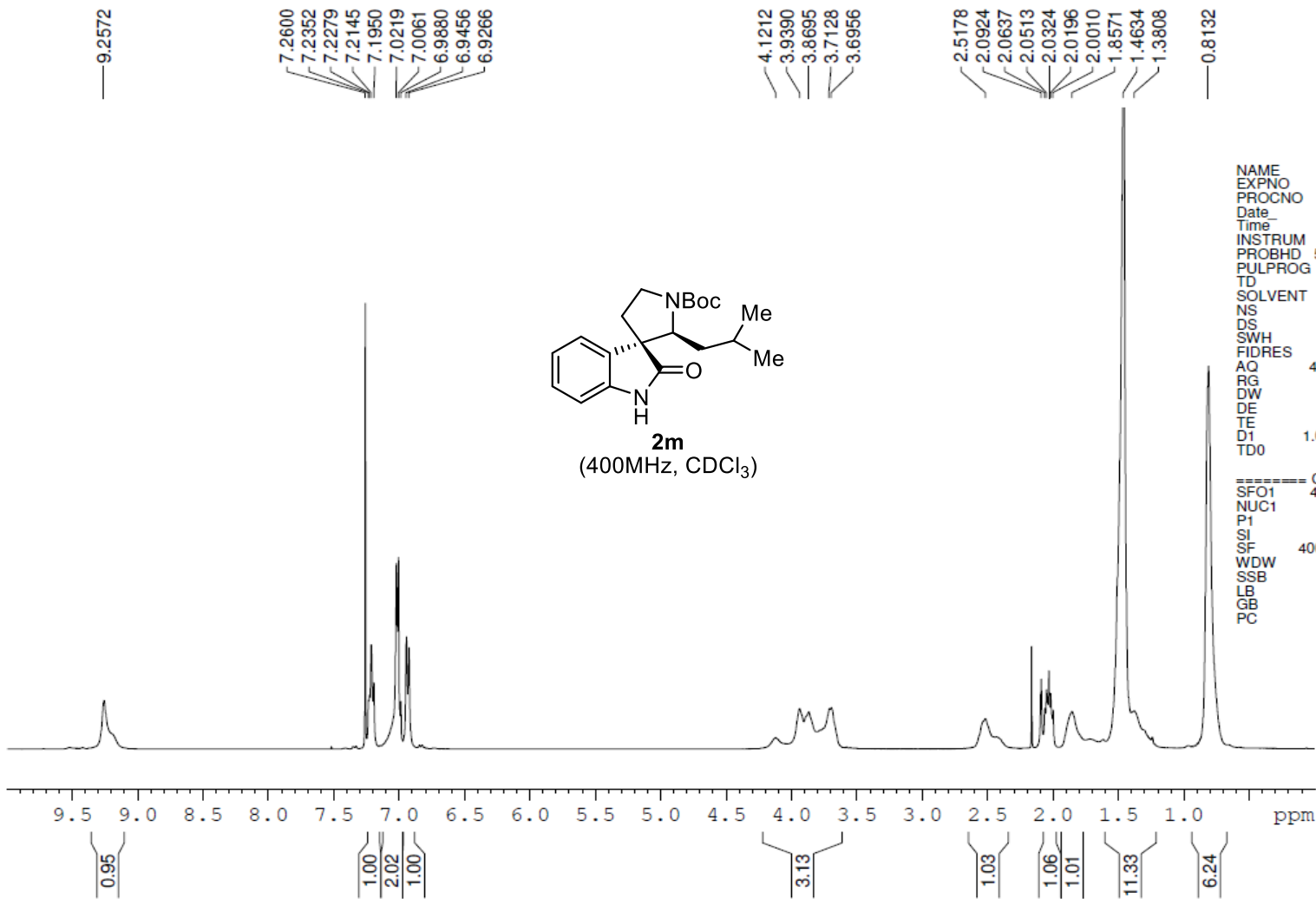
===== CHANNEL f1 =====
SFO1    400.1324710 MHz
NUC1     1H
P1      14.50 usec
SI      65536
SF      400.1300102 MHz
WDW     EM
SSB     0
LB      0.30 Hz
GB      0
PC      1.00

```







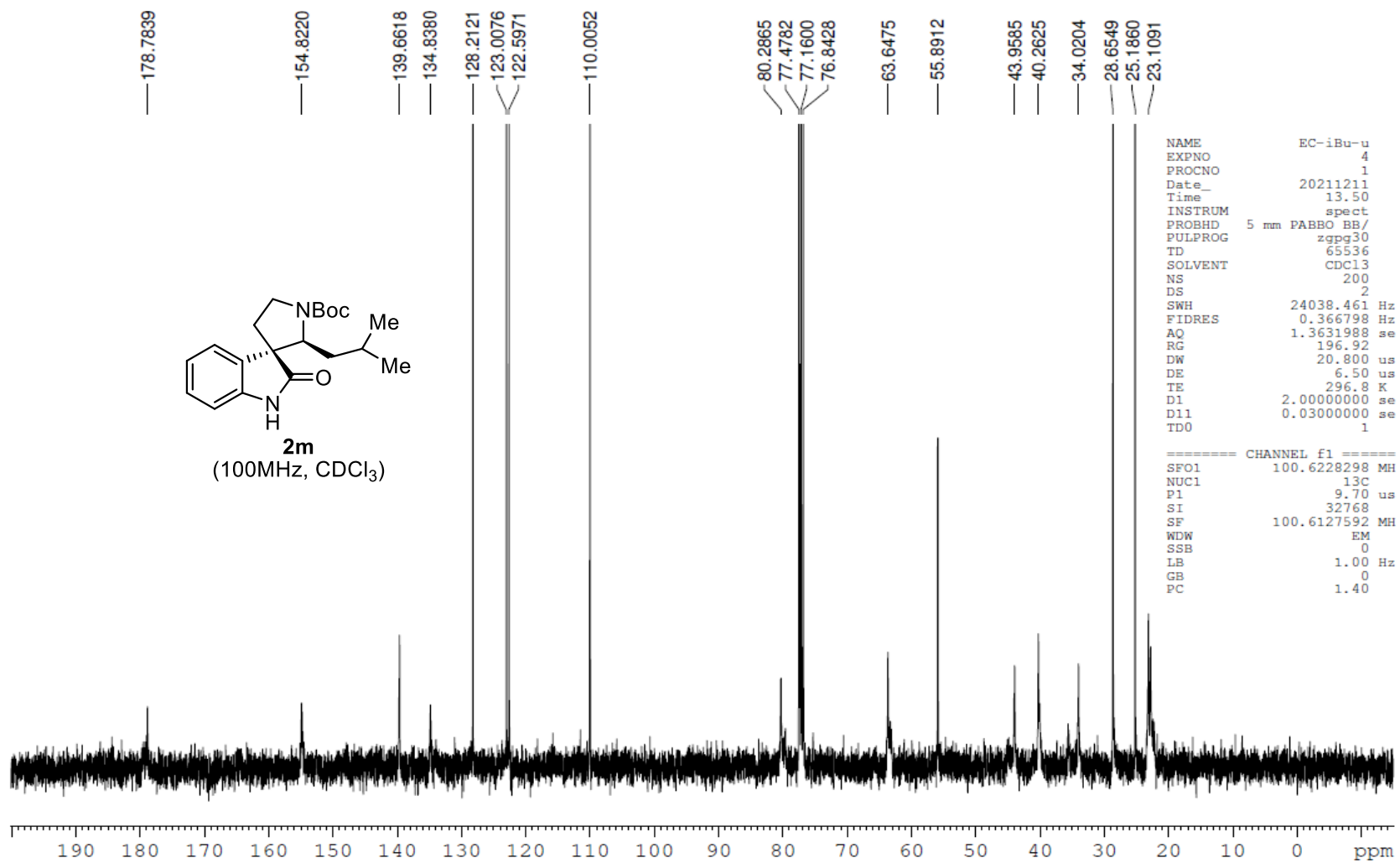


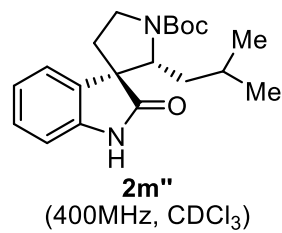
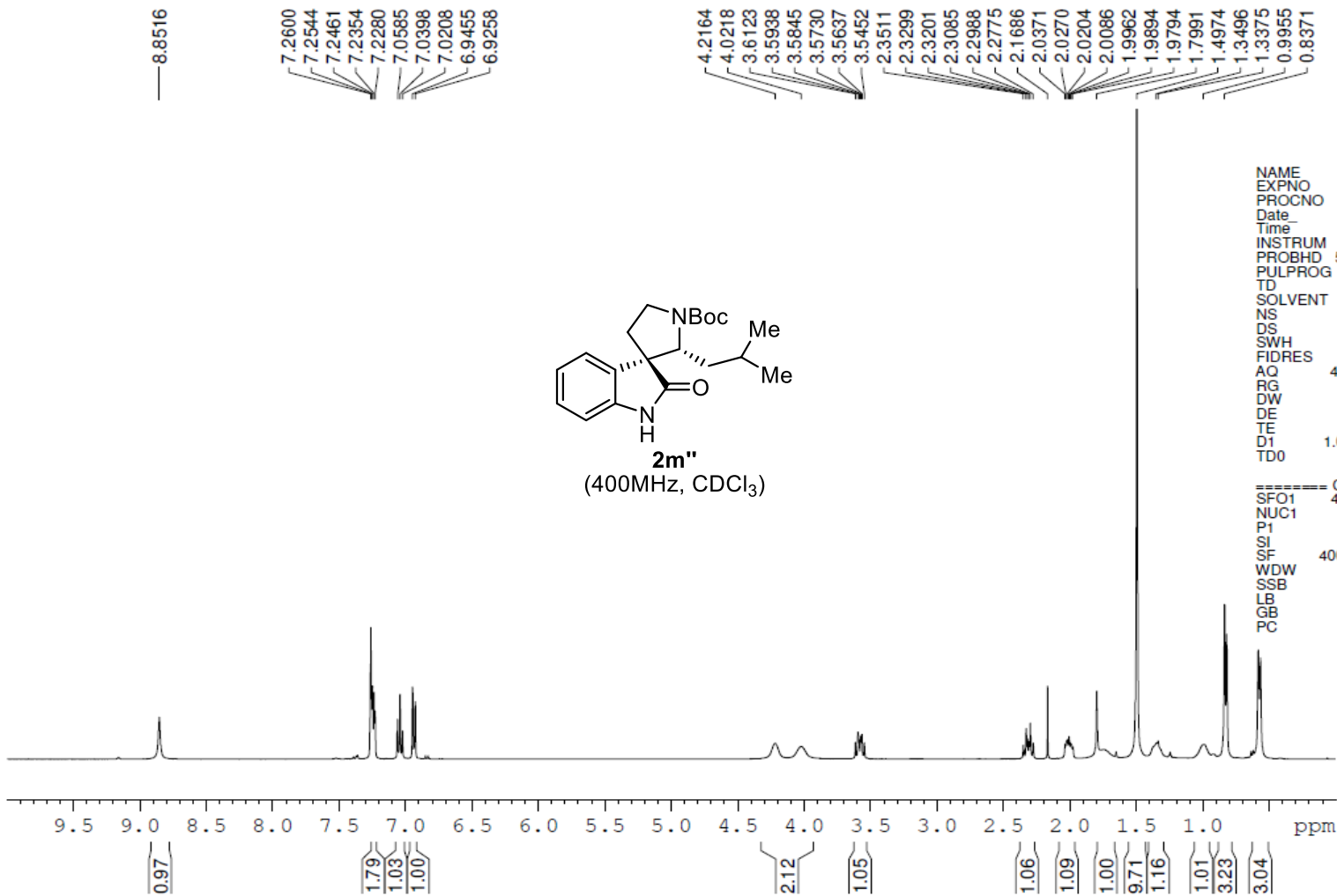
```

NAME      EC-IBu-u
EXPNO     3
PROCNO    1
Date_     20211211
Time      13.39
INSTRUM   spect
PROBHD    5 mm PABBO BB/
PULPROG   zg30
TD         65536
SOLVENT   CDCl3
NS         16
DS         2
SWH        8012.820 Hz
FIDRES     0.122266 Hz
AQ         4.0894966 sec
RG         25.32
DW         62.400 usec
DE         6.50 usec
TE         295.9 K
D1         1.00000000 sec
TD0        1

===== CHANNEL f1 =====
SFO1      400.1324710 MHz
NUC1       1H
P1         14.50 usec
SI         65536
SF         400.1300100 MHz
WDW        EM
SSB        0
LB         0.30 Hz
GB         0
PC         1.00

```

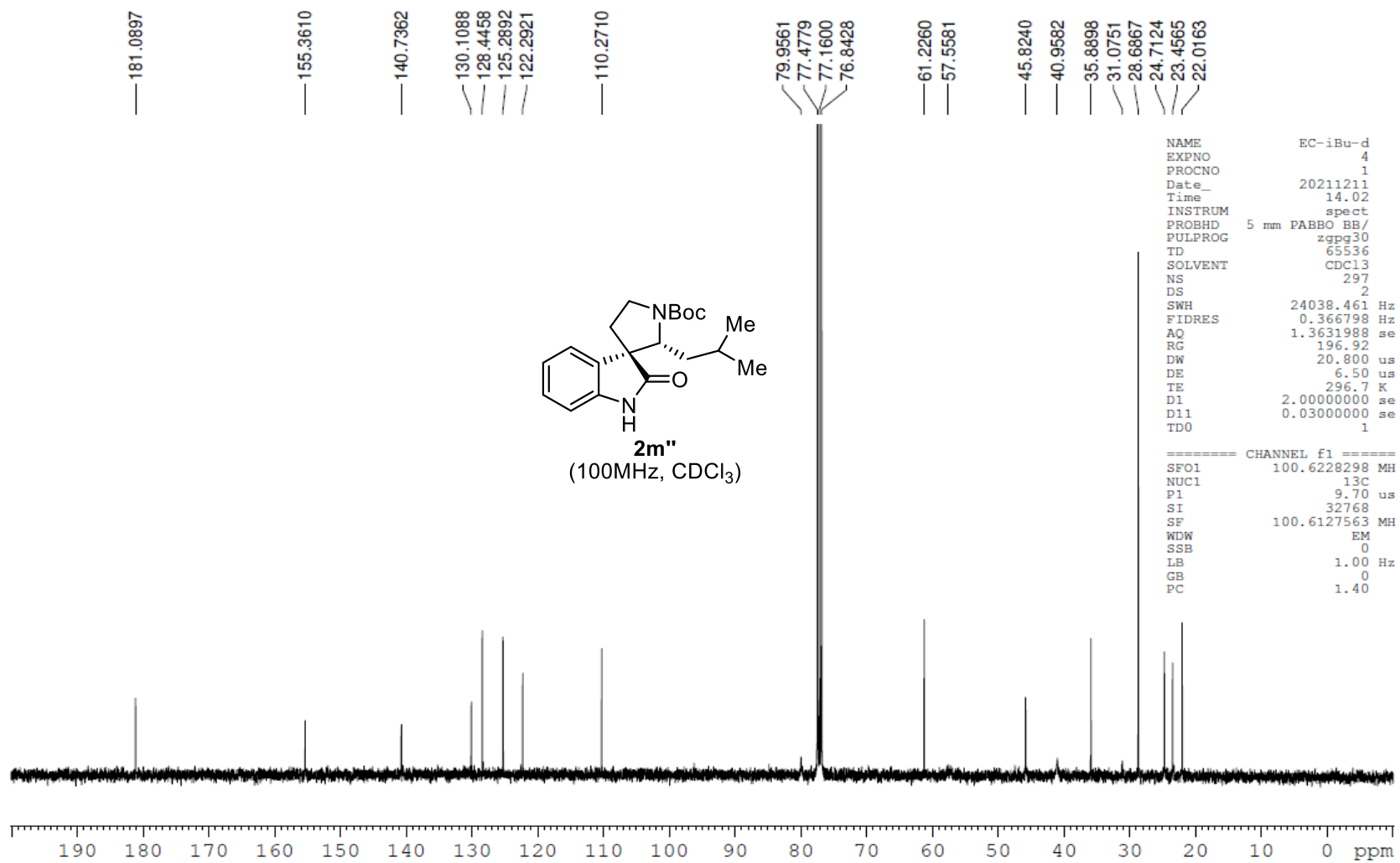


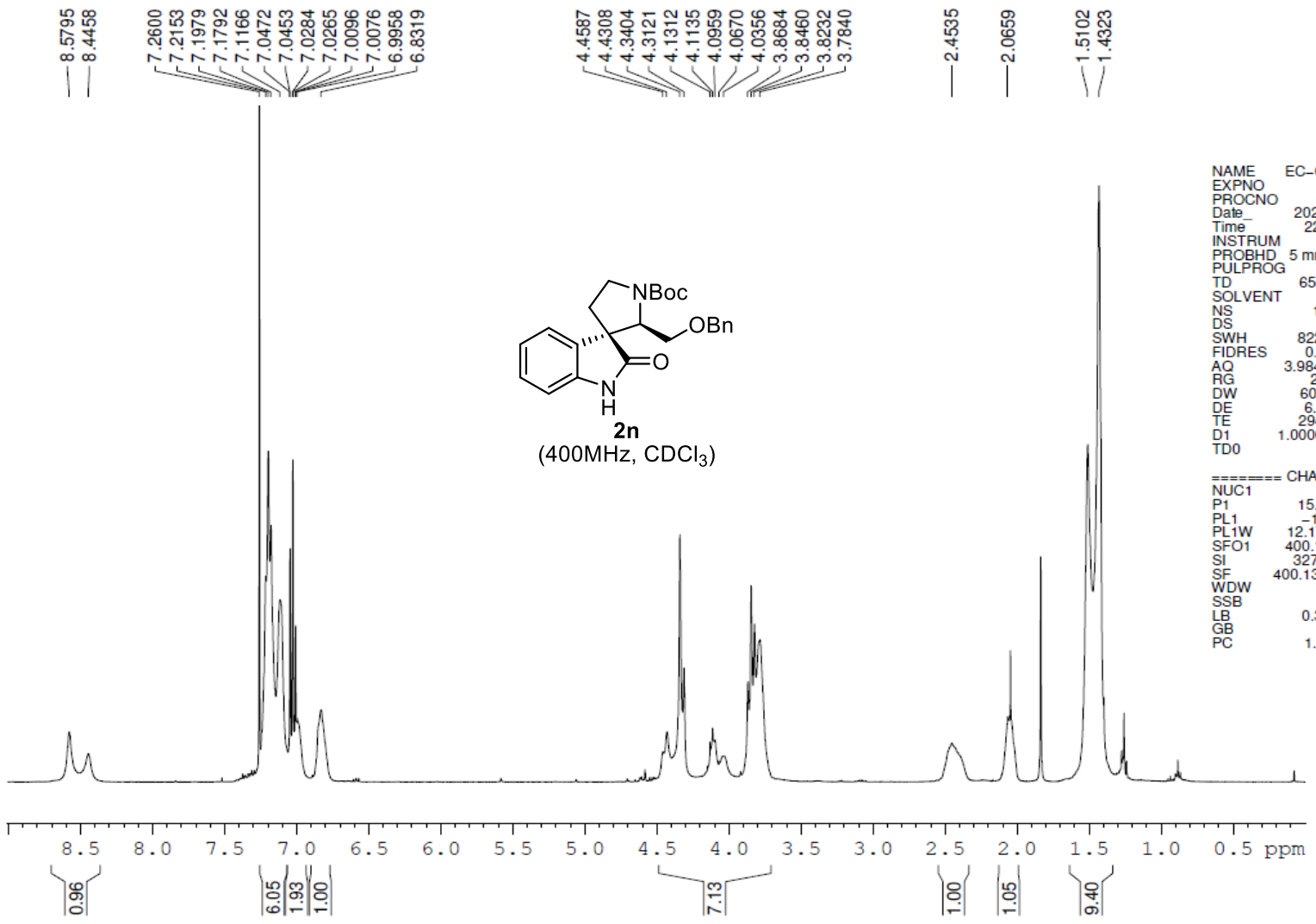


```

NAME      EC-IBu-d
EXPNO     3
PROCNO    1
Date_     20211211
Time      13.56
INSTRUM   spect
PROBHD    5 mm PABBO BB/
PULPROG   zg30
TD         65536
SOLVENT   CDCl3
NS         16
DS         2
SWH        8012.820 Hz
FIDRES     0.122266 Hz
AQ         4.0894966 sec
RG         70.97
DW         62.400 usec
DE         6.50 usec
TE         295.9 K
D1         1.00000000 sec
TD0        1

===== CHANNEL f1 =====
SFO1      400.1324710 MHz
NUC1       1H
P1         14.50 usec
SI         65536
SF         400.1300100 MHz
WDW        EM
SSB        0
LB         0.30 Hz
GB         0
PC         1.00
  
```

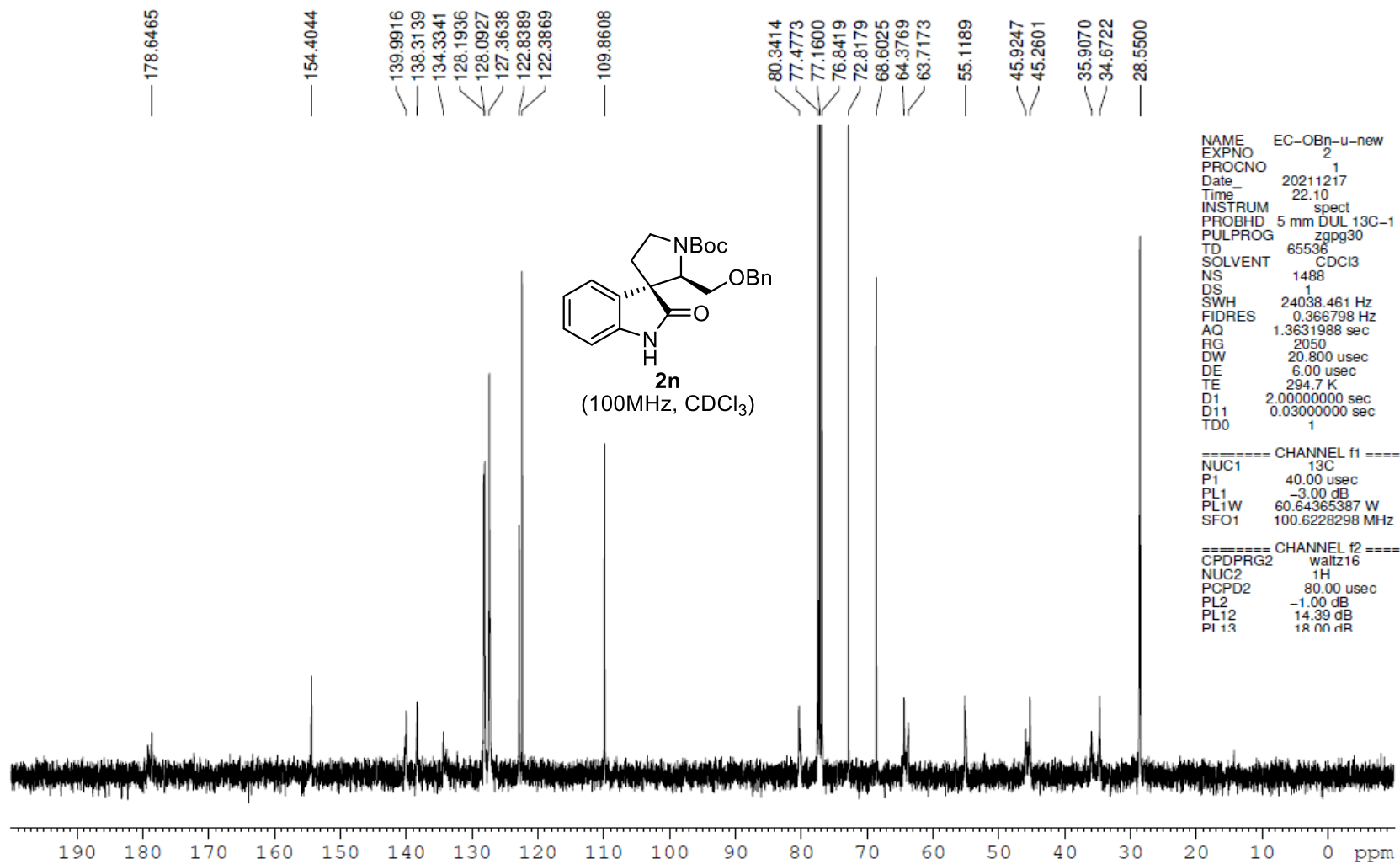


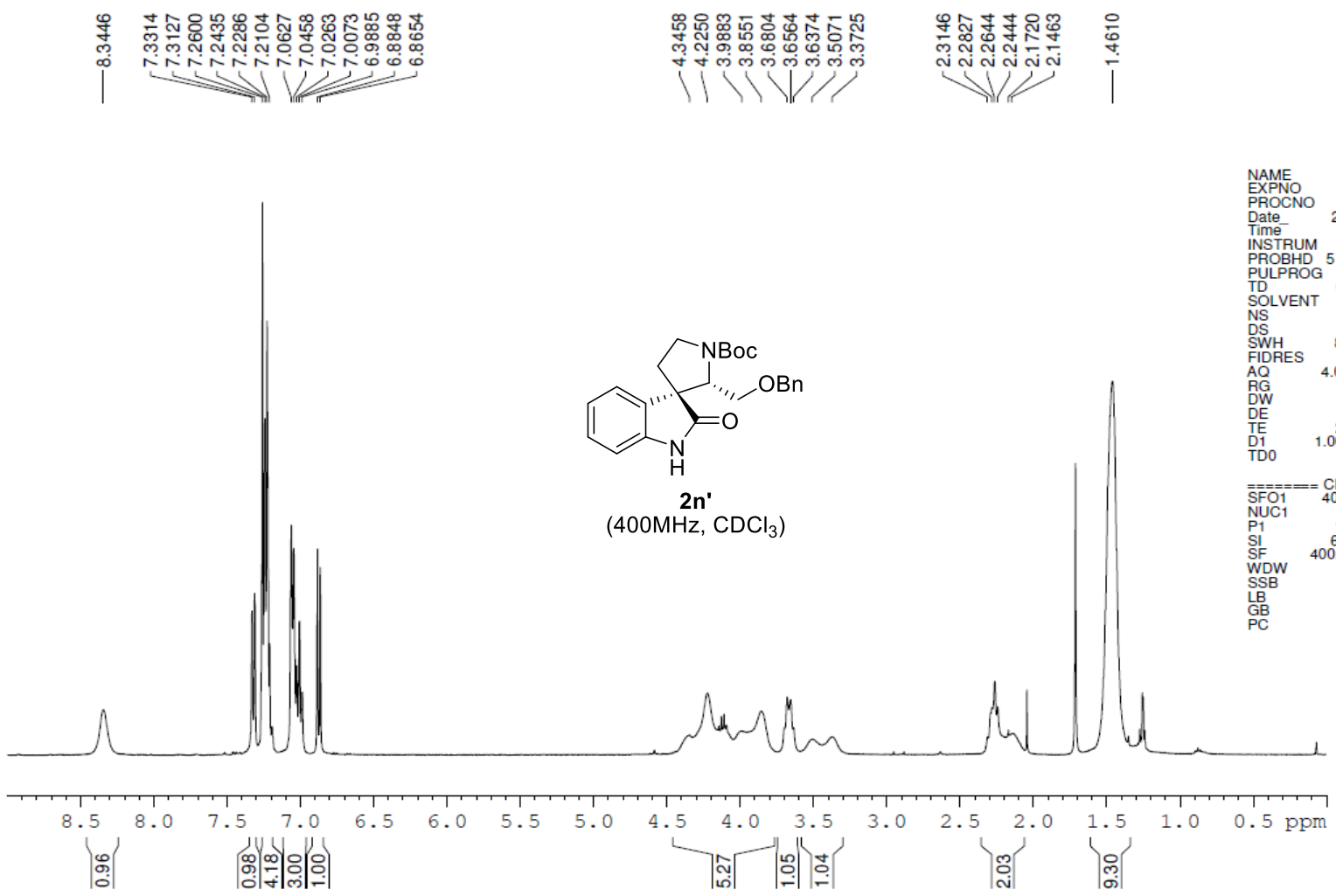
```

NAME EC-OBn-u-new
EXPNO 1
PROCNO 1
Date_ 20211217
Time 22.06
INSTRUM spect
PROBHD 5 mm DUL 13C-1
PULPROG zg30
TD 65536
SOLVENT CDCl3
NS 16
DS 2
SWH 8223.685 Hz
FIDRES 0.125483 Hz
AQ 3.9846387 sec
RG 203
DW 60.800 usec
DE 6.00 usec
TE 294.3 K
D1 1.00000000 sec
TD0 1

===== CHANNEL f1 =====
NUC1 1H
P1 15.80 usec
PL1 -1.00 dB
PL1W 12.17476940 W
SFO1 400.1324710 MHz
SI 32768
SF 400.1300097 MHz
WDW EM
SSB 0
LB 0.30 Hz
GB 0
PC 1.00

```



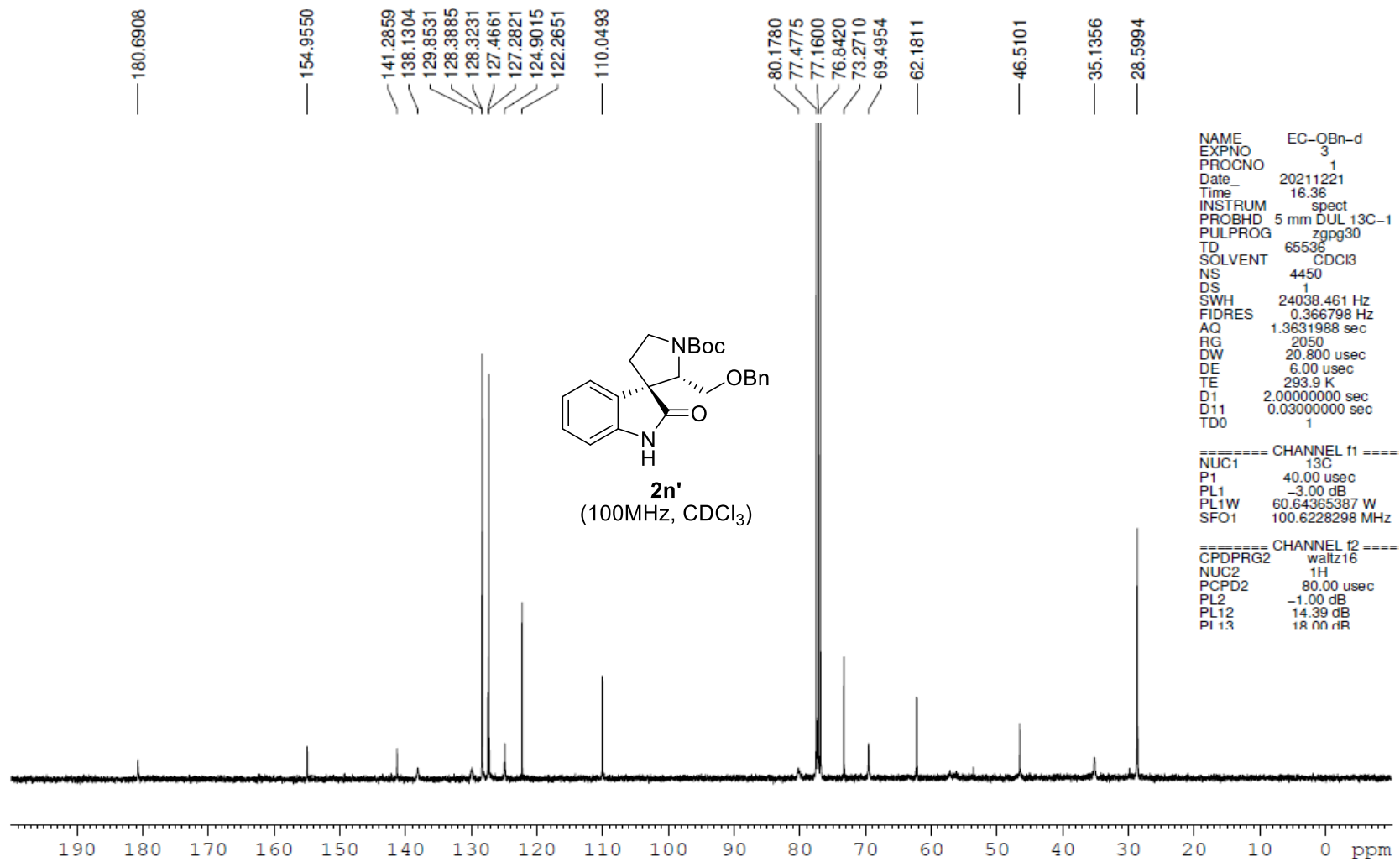


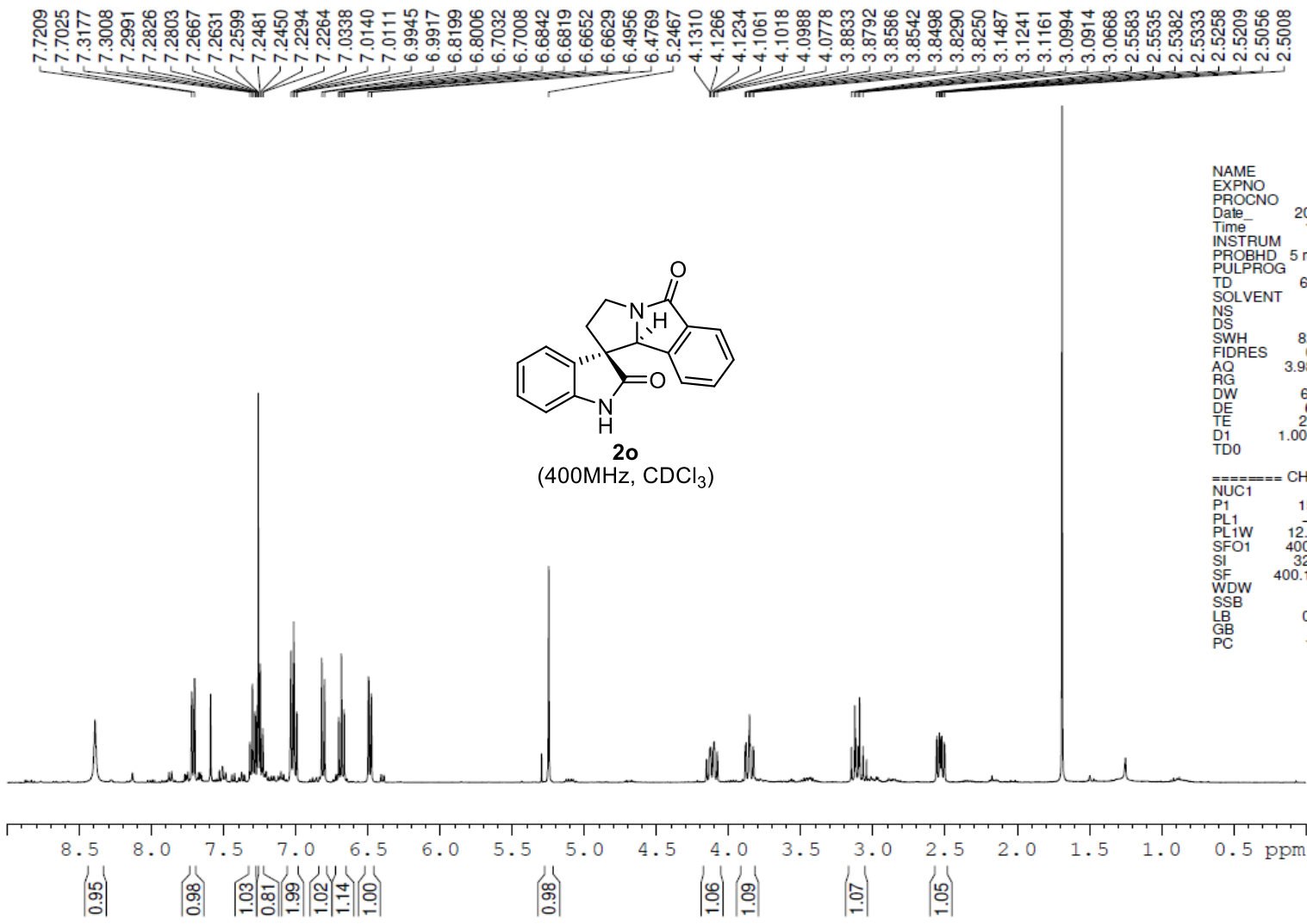
```

NAME      EC-OBn-d
EXPNO     4
PROCNO    1
Date_     20211223
Time      10.10
INSTRUM   spect
PROBHD    5 mm PABBO BB/
PULPROG   zg30
TD         65536
SOLVENT   CDCl3
NS         16
DS         2
SWH        8012.820 Hz
FIDRES     0.122266 Hz
AQ         4.0894966 sec
RG         88.84
DW         62.400 usec
DE         6.50 usec
TE         295.1 K
D1         1.00000000 sec
TD0        1

===== CHANNEL f1 =====
SFO1      400.1324710 MHz
NUC1       1H
P1         14.50 usec
SI         65536
SF         400.1300100 MHz
WDW        EM
SSB        0
LB         0.30 Hz
GB         0
PC         1.00

```



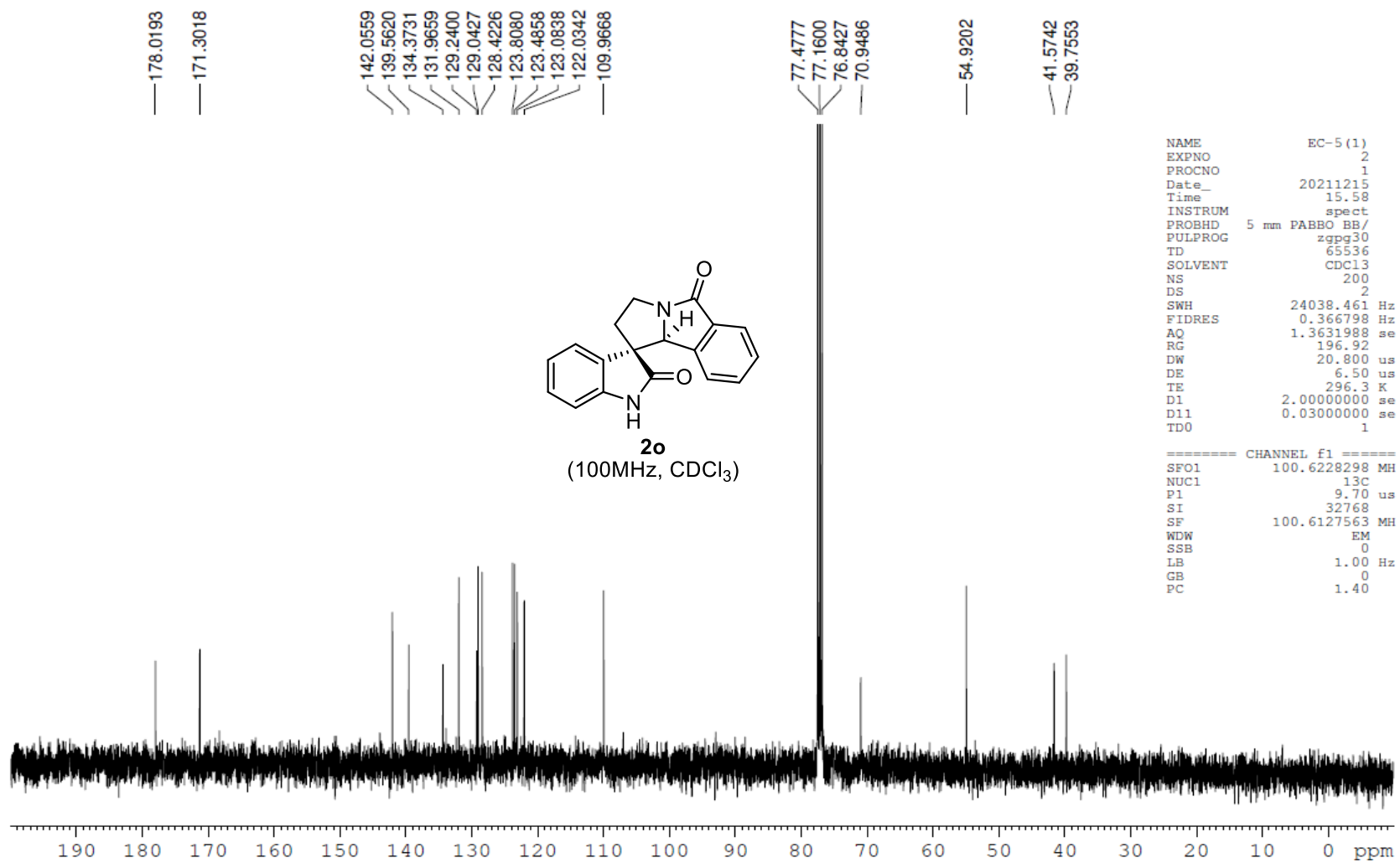


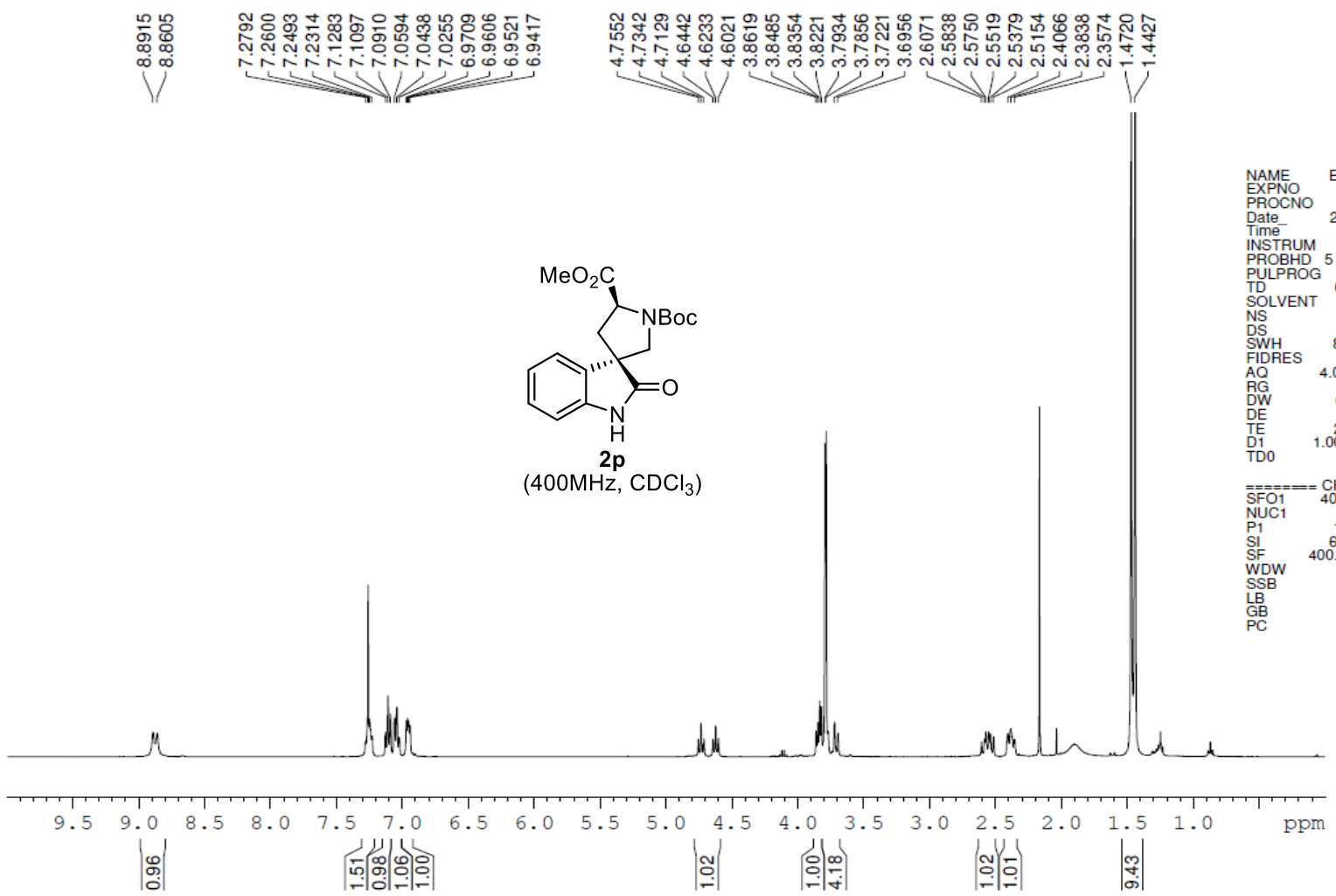
```

NAME          EC-5
EXPNO         1
PROCNO        1
Date_         20211215
Time          19.05
INSTRUM       spect
PROBHD        5 mm DUL 13C-1
PULPROG       zg30
TD            65536
SOLVENT       CDCl3
NS            16
DS            2
SWH           8223.685 Hz
FIDRES        0.125483 Hz
AQ            3.9846387 sec
RG            575
DW            60.800 usec
DE            6.00 usec
TE            293.4 K
D1            1.00000000 sec
TD0           1

===== CHANNEL f1 =====
NUC1          1H
P1            15.80 usec
PL1           -1.00 dB
PL1W          12.17476940 W
SFO1          400.1324710 MHz
SI            32768
SF            400.1300099 MHz
WDW           EM
SSB           0
LB            0.30 Hz
GB            0
PC            1.00

```



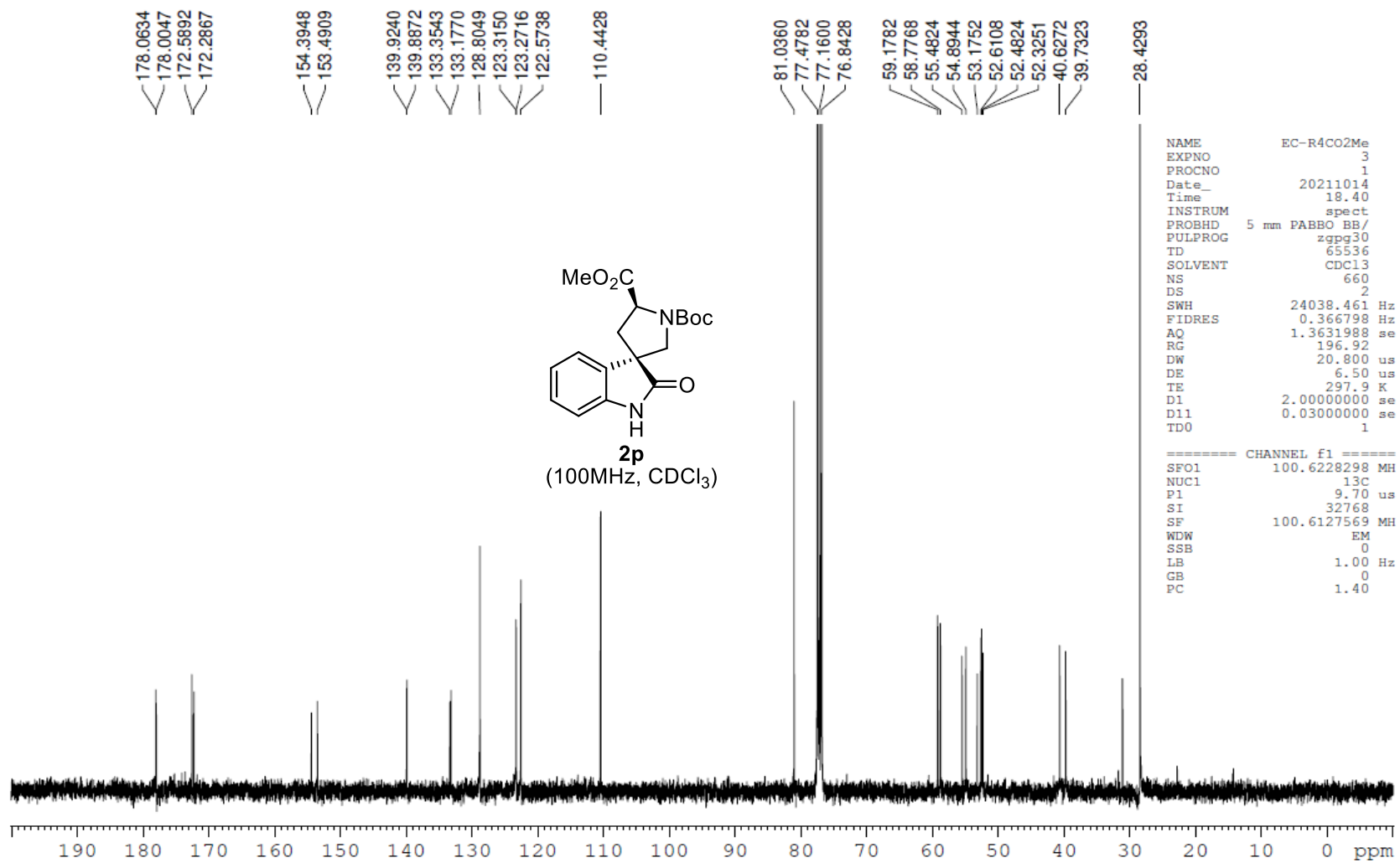


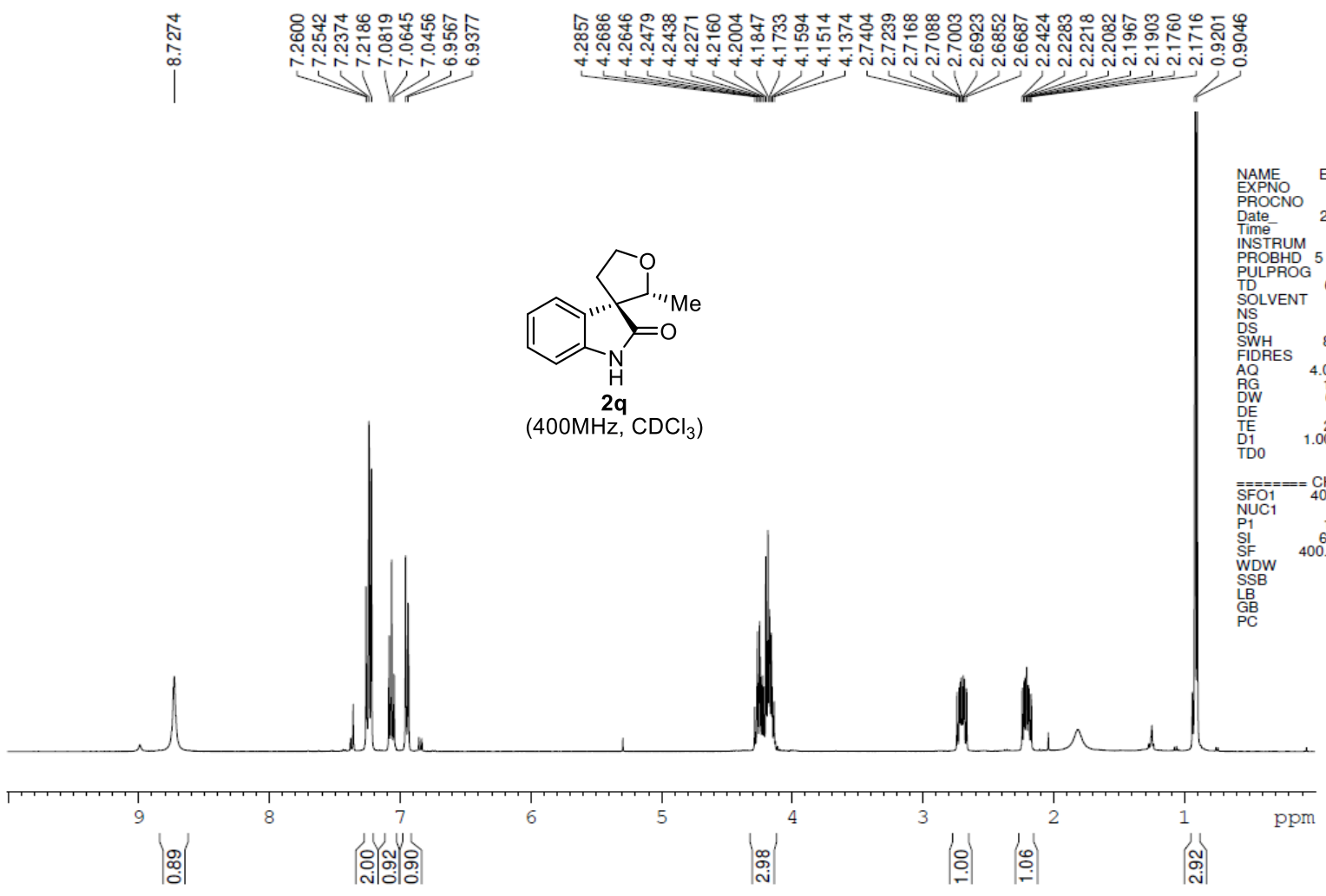
```

NAME      EC-R4CO2Me
EXPNO    1
PROCNO   1
Date_    20211014
Time     18.11
INSTRUM  spect
PROBHD   5 mm PABBO BB/
PULPROG  zg30
TD       65536
SOLVENT  CDCl3
NS       16
DS       2
SWH      8012.820 Hz
FIDRES   0.122266 Hz
AQ       4.0894966 sec
RG       70.97
DW       62.400 usec
DE       6.50 usec
TE       296.8 K
D1       1.00000000 sec
TD0      1

===== CHANNEL f1 =====
SFO1    400.1324710 MHz
NUC1    1H
P1      14.50 usec
SI      65536
SF      400.1300101 MHz
WDW     EM
SSB     0
LB      0.30 Hz
GB      0
PC      1.00

```

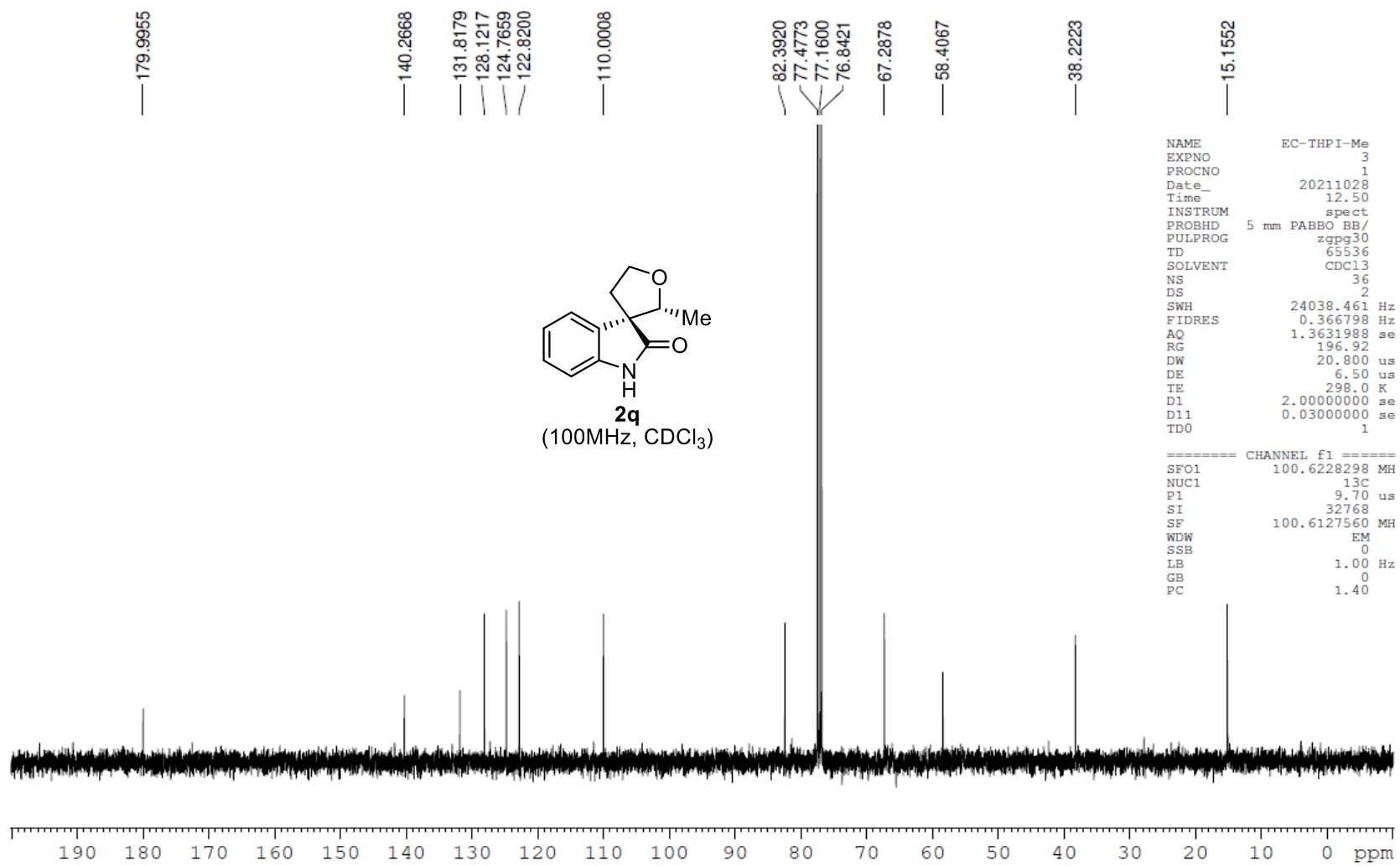


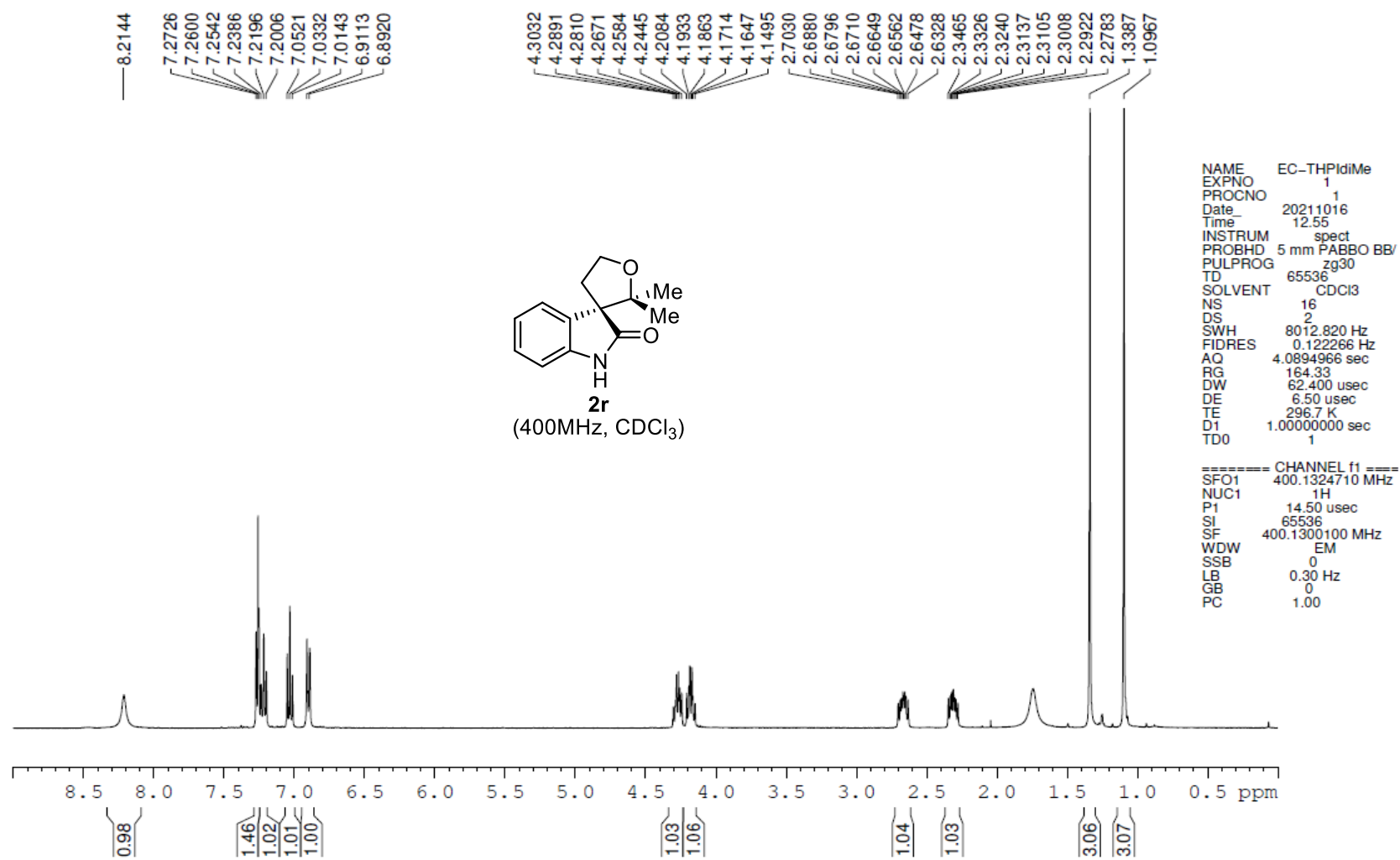
```

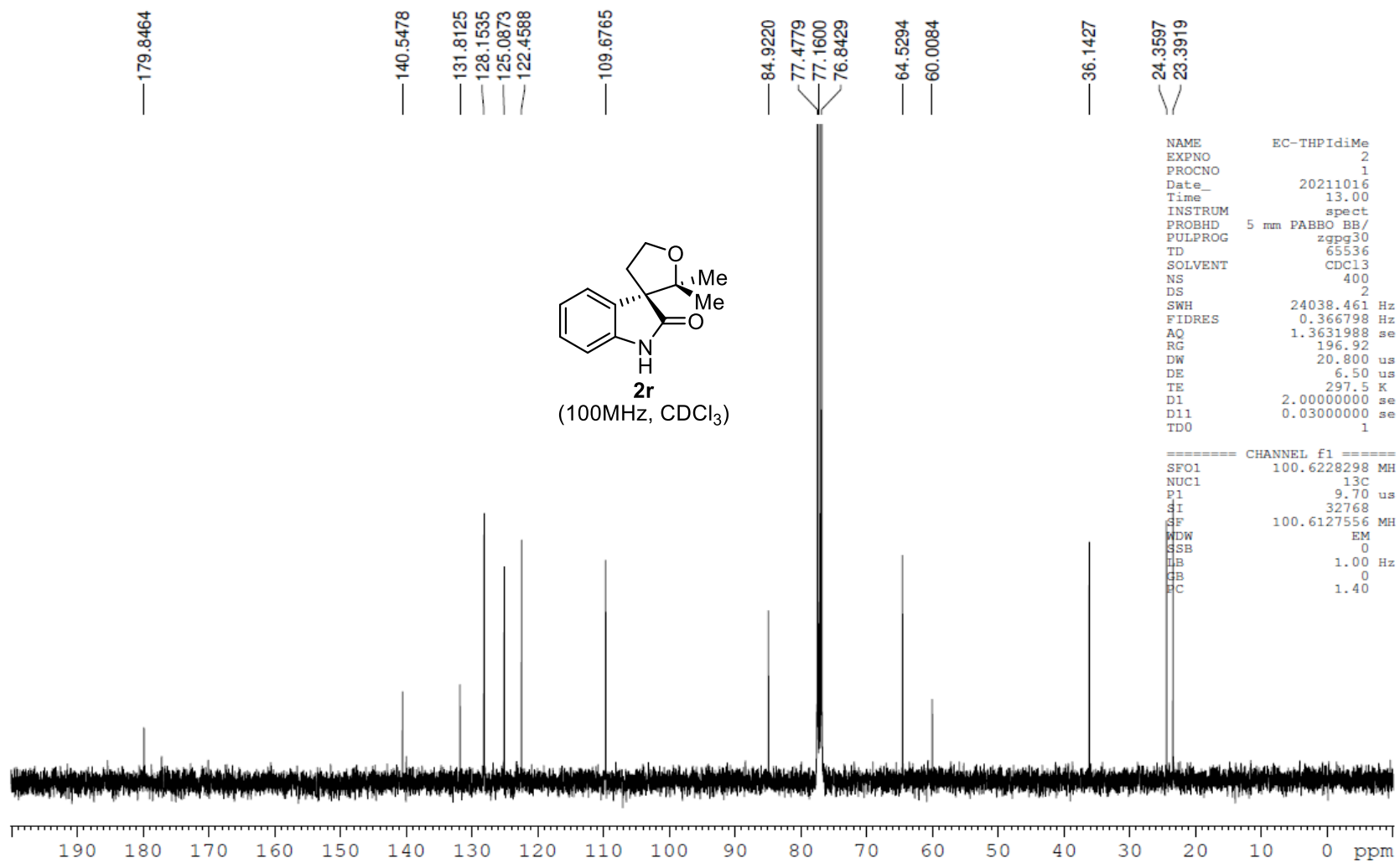
NAME      EC-THPI-Me
EXPNO    1
PROCNO   1
Date_    20211028
Time     12.40
INSTRUM  spect
PROBHD   5 mm PABBO BB/
PULPROG  zg30
TD        65536
SOLVENT  CDCl3
NS        16
DS        2
SWH       8012.820 Hz
FIDRES    0.122266 Hz
AQ        4.0894966 sec
RG        103.52
DW        62.400 usec
DE        6.50 usec
TE        297.3 K
D1        1.00000000 sec
TD0       1

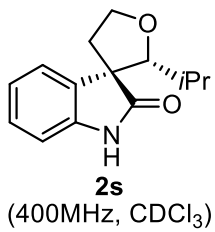
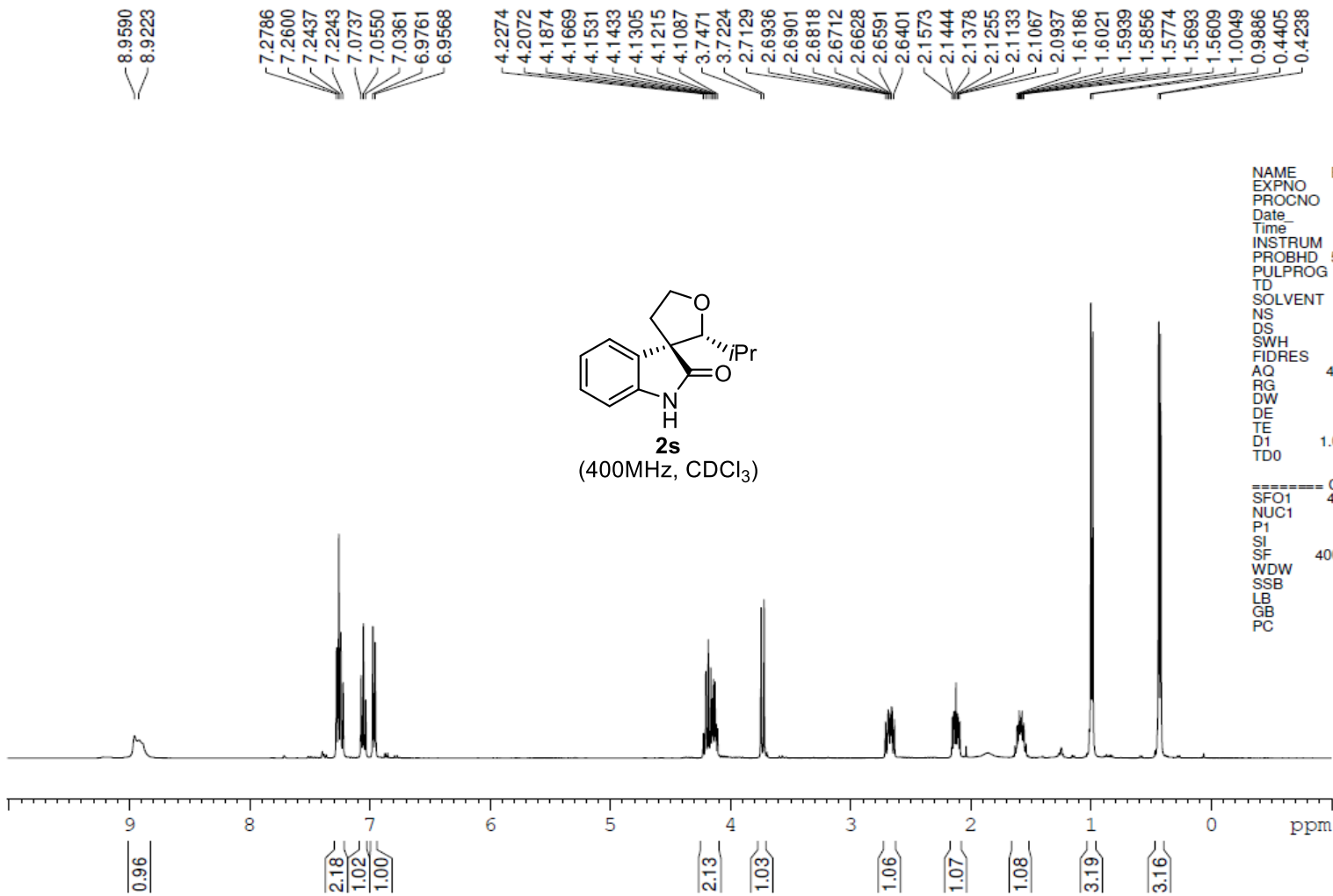
===== CHANNEL f1 =====
SFO1     400.1324710 MHz
NUC1     1H
P1       14.50 usec
SI       65536
SF       400.1300100 MHz
WDW      EM
SSB      0
LB       0.30 Hz
GB       0
PC       1.00

```





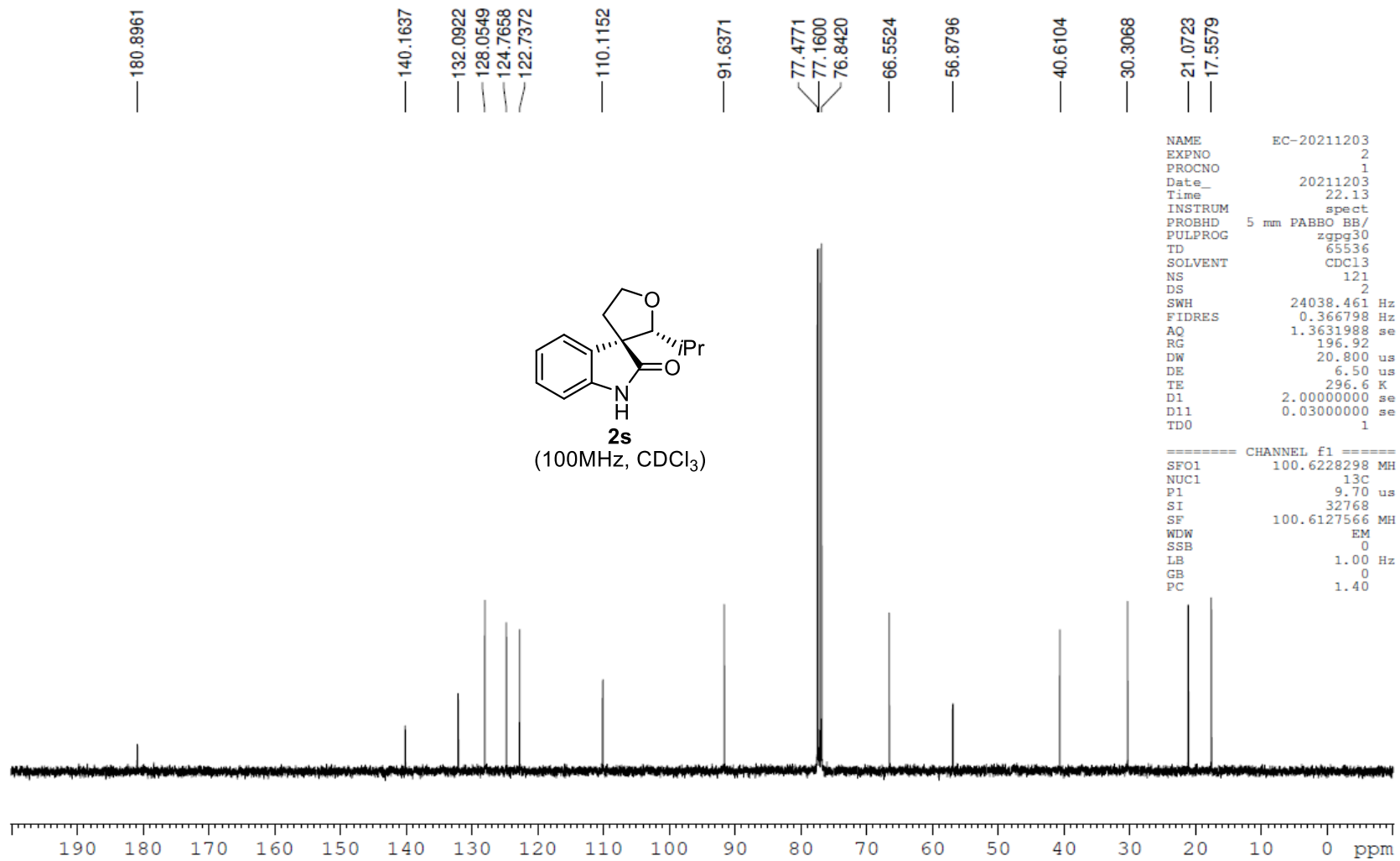


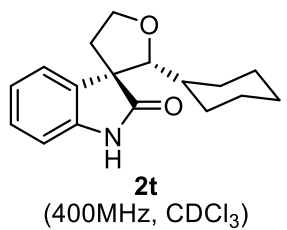
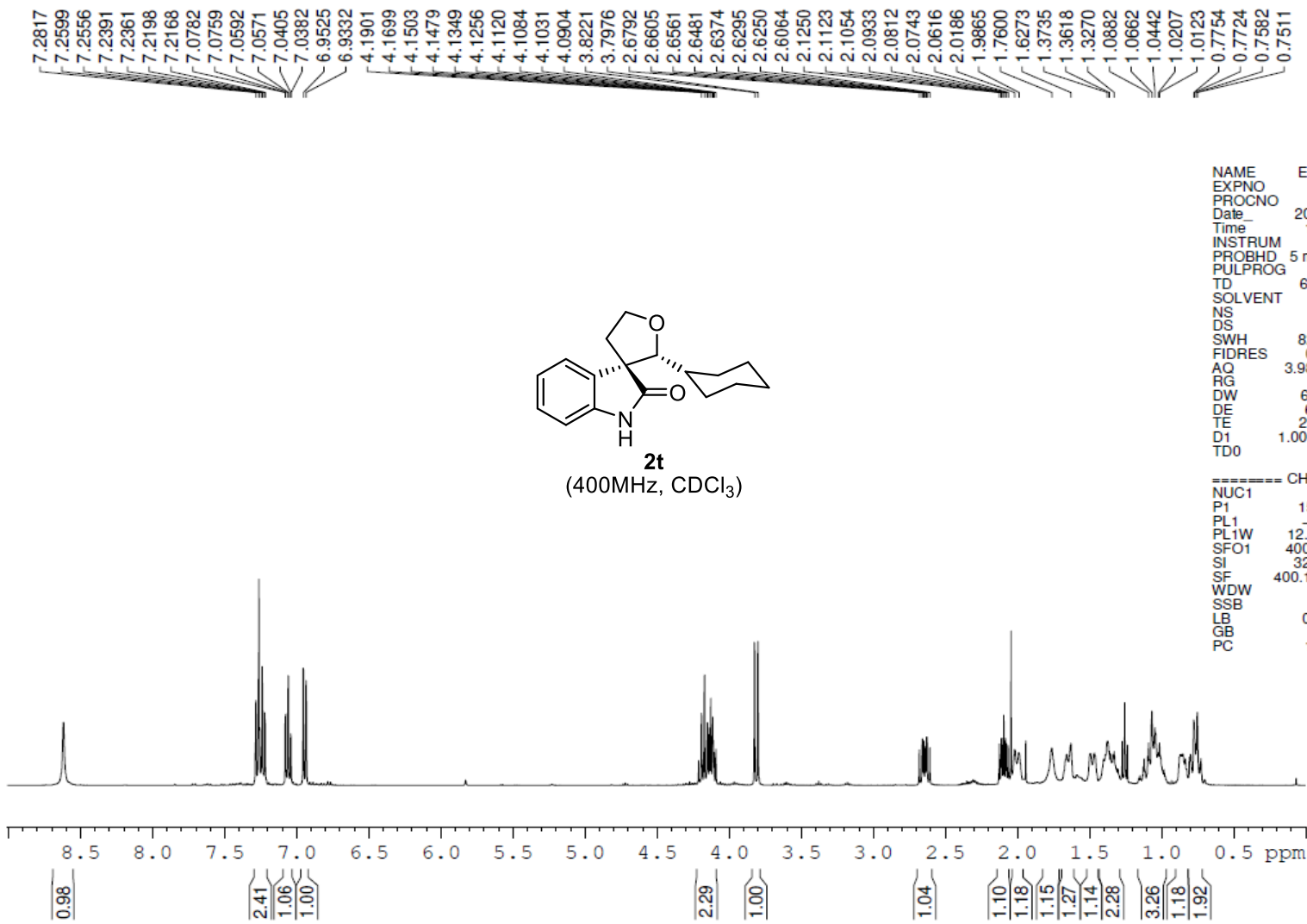


```

NAME      EC-20211203
EXPNO     1
PROCNO    1
Date_     20211203
Time      22.08
INSTRUM   spect
PROBHD    5 mm PABBO BB/
PULPROG   zg30
TD         65536
SOLVENT   CDCl3
NS         16
DS         2
SWH        8012.820 Hz
FIDRES     0.122266 Hz
AQ         4.0894966 sec
RG         70.97
DW         62.400 usec
DE         6.50 usec
TE         295.8 K
D1         1.00000000 sec
TD0        1

===== CHANNEL f1 =====
SFO1      400.1324710 MHz
NUC1       1H
P1         14.50 usec
SI         65536
SF         400.1300102 MHz
WDW        EM
SSB        0
LB         0.30 Hz
GB         0
PC         1.00
  
```

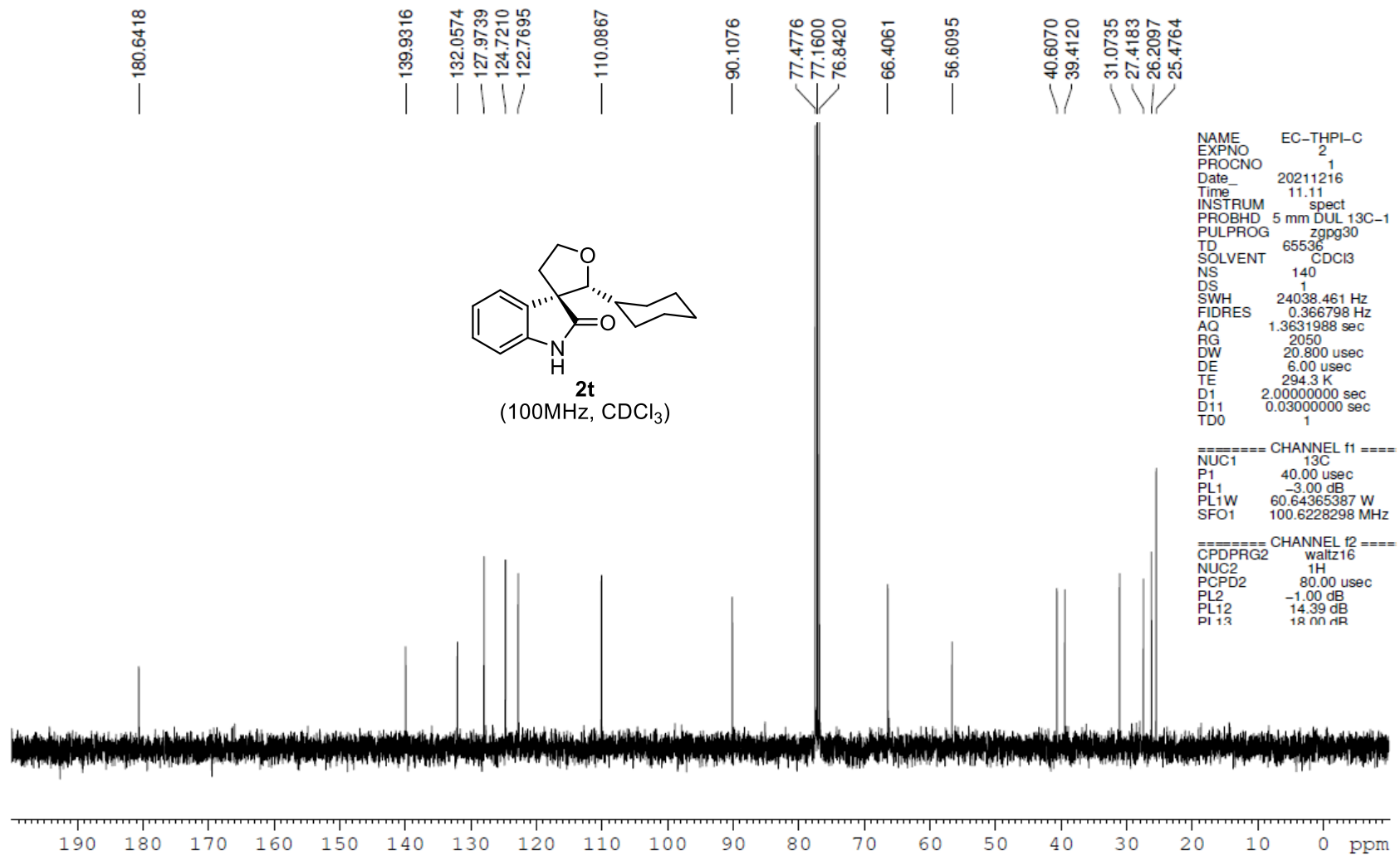


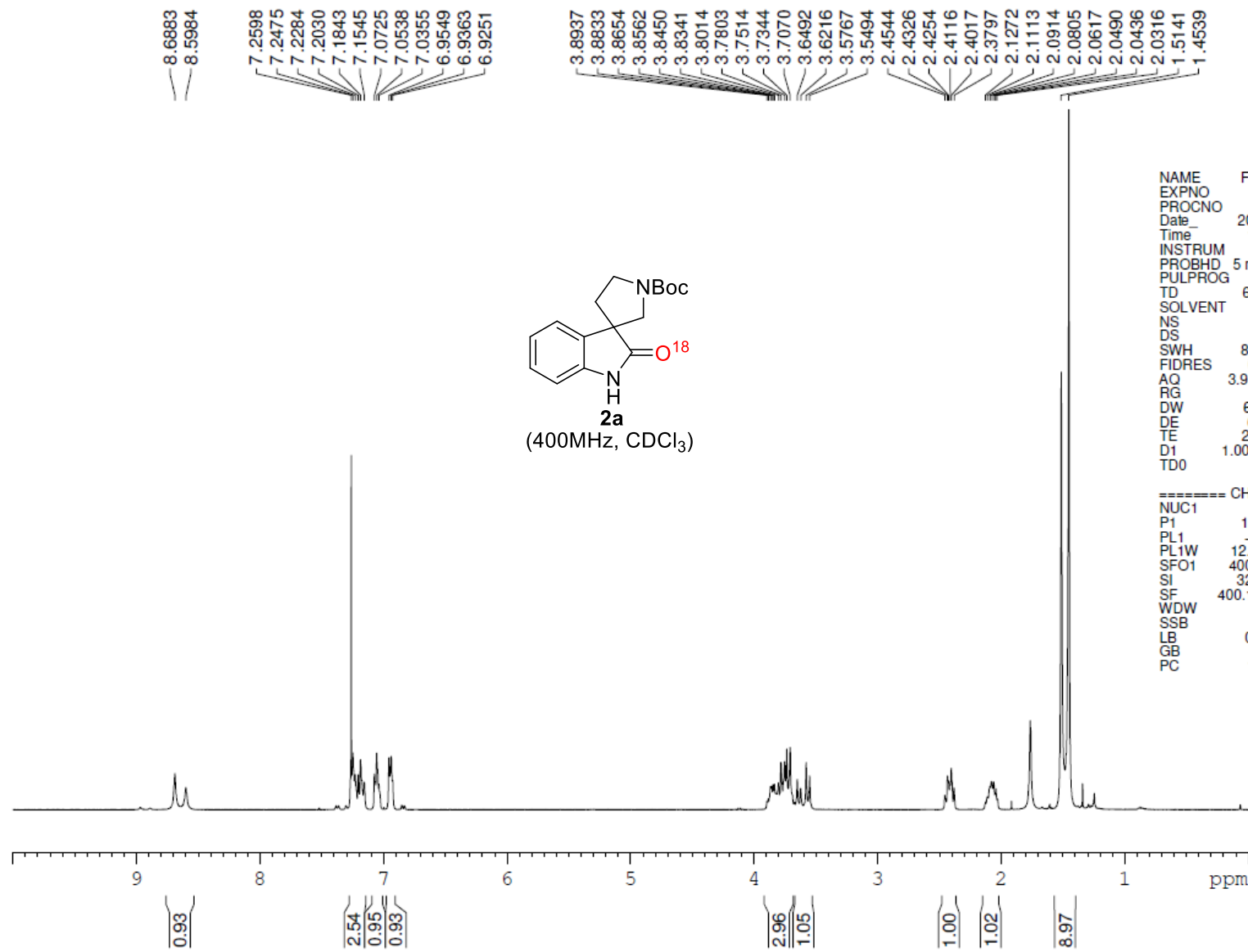


```

NAME      EC-THPI-C
EXPNO     1
PROCNO    1
Date_     20211216
Time      11.07
INSTRUM   spect
PROBHD    5 mm DUL 13C-1
PULPROG   zg30
TD         65536
SOLVENT   CDCl3
NS         16
DS         2
SWH        8223.685 Hz
FIDRES     0.125483 Hz
AQ         3.9846387 sec
RG         287
DW         60.800 usec
DE         6.00 usec
TE         294.0 K
D1         1.00000000 sec
TD0        1

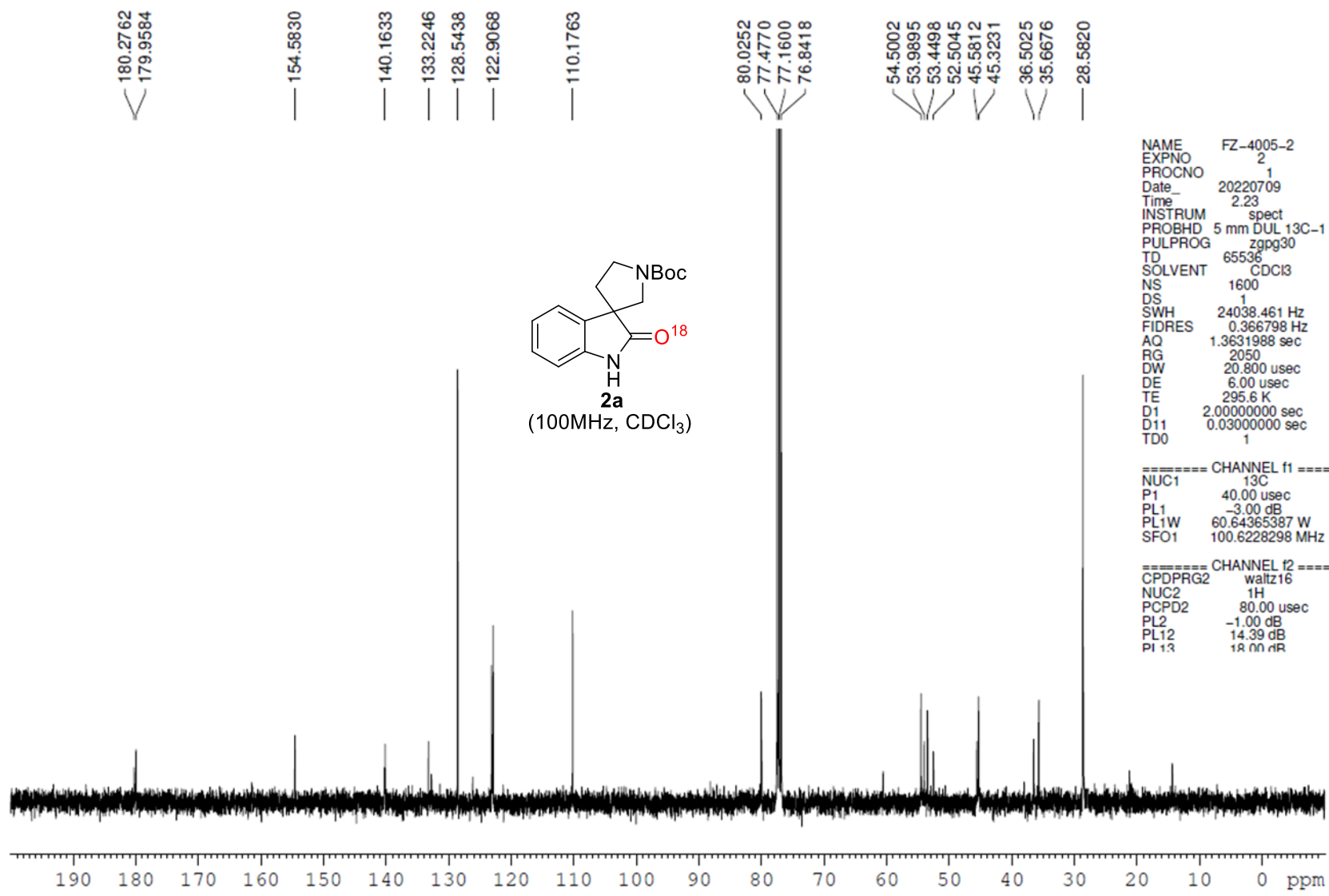
===== CHANNEL f1 =====
NUC1      1H
P1        15.80 usec
PL1       -1.00 dB
PL1W      12.17476940 W
SFO1      400.1324710 MHz
SI         32768
SF         400.1300099 MHz
WDW        EM
SSB        0
LB         0.30 Hz
GB         0
PC         1.00
  
```



NAME FZ-4005-2
 EXPNO 3
 PROCNO 1
 Date 20220709
 Time 16.39
 INSTRUM spect
 PROBHD 5 mm DUL 13C-1
 PULPROG zg30
 TD 65536
 SOLVENT CDCl3
 NS 16
 DS 2
 SWH 8223.685 Hz
 FIDRES 0.125483 Hz
 AQ 3.9846387 sec
 RG 406
 DW 60.800 usec
 DE 6.00 usec
 TE 292.8 K
 D1 1.00000000 sec
 TD0 1

===== CHANNEL f1 =====
 NUC1 1H
 P1 15.80 usec
 PL1 -1.00 dB
 PL1W 12.17476940 W
 SFO1 400.1324710 MHz
 SI 32768
 SF 400.1300099 MHz
 WDW EM
 SSB 0
 LB 0.30 Hz
 GB 0
 PC 1.00



```

NAME      FZ-4005-2
EXPNO     2
PROCNO    1
Date_     20220709
Time      2.23
INSTRUM   spect
PROBHD    5 mm DUL 13C-1
PULPROG   zgpg30
TD         65536
SOLVENT   CDCl3
NS         1600
DS         1
SWH       24038.461 Hz
FIDRES    0.366798 Hz
AQ         1.3631988 sec
RG         2050
DW         20.800 usec
DE         6.00 usec
TE         295.6 K
D1         2.00000000 sec
D11        0.03000000 sec
TD0        1

```

```

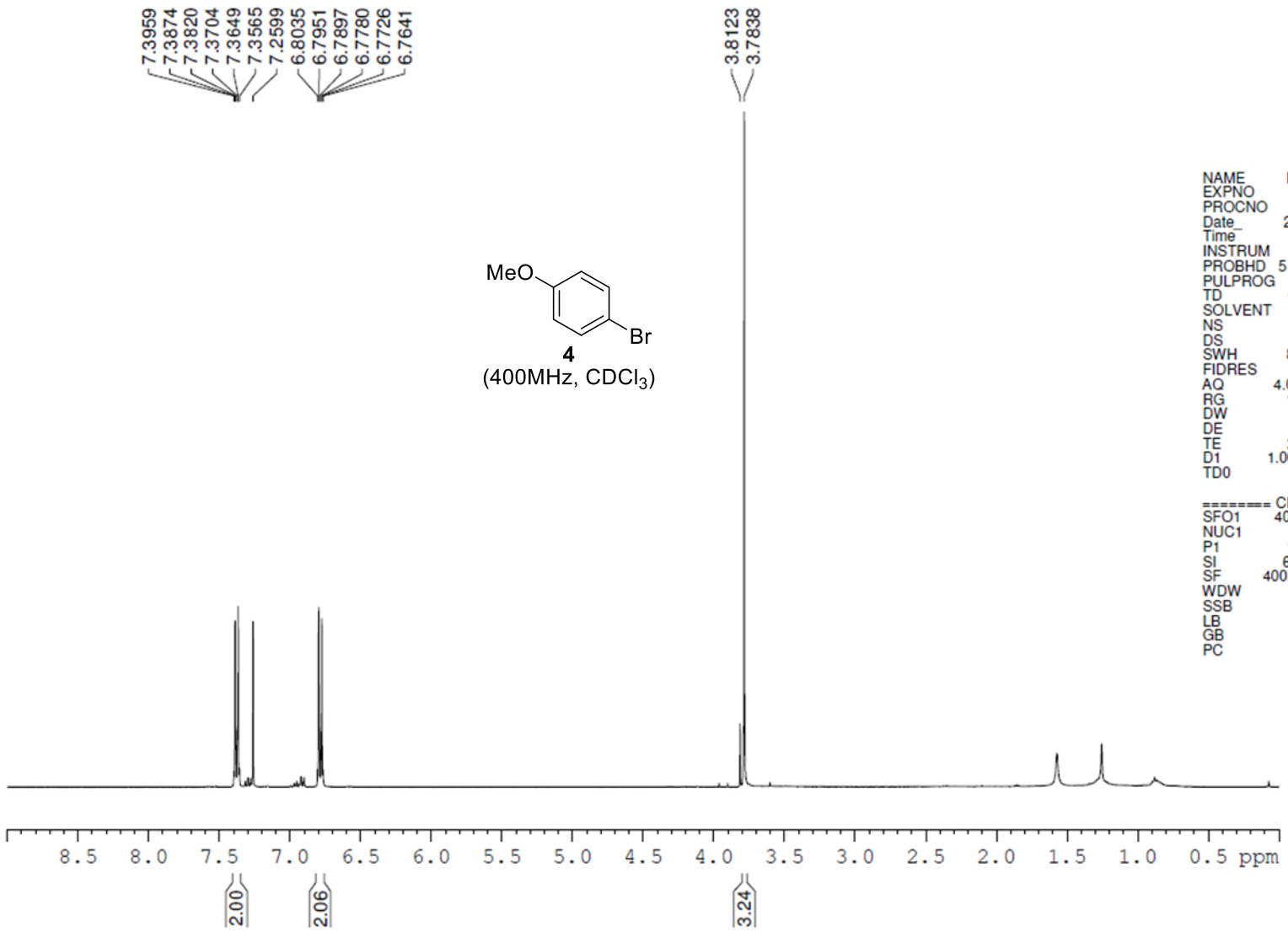
===== CHANNEL f1 =====
NUC1      13C
P1         40.00 usec
PL1       -3.00 dB
PL1W      60.64365387 W
SFO1      100.6228298 MHz

```

```

===== CHANNEL f2 =====
CPDPRG2   waltz16
NUC2       1H
PCPD2     80.00 usec
PL2       -1.00 dB
PL12      14.39 dB
PI 13     18.00 dB

```



```

NAME      FZ-4008-c
EXPNO     2
PROCNO    1
Date_     20220714
Time      20.21
INSTRUM   spect
PROBHD    5 mm PABBO BB/
PULPROG   zg30
TD         65536
SOLVENT   CDCl3
NS         16
DS         2
SWH        8012.820 Hz
FIDRES     0.122266 Hz
AQ         4.0894966 sec
RG         164.33
DW         62.400 usec
DE         6.50 usec
TE         295.5 K
D1         1.00000000 sec
TD0        1

===== CHANNEL f1 =====
SFO1      400.1324710 MHz
NUC1       1H
P1         14.50 usec
SI         65536
SF         400.1300103 MHz
WDW        EM
SSB        0
LB         0.30 Hz
GB         0
PC         1.00
  
```

