

Supplementary Information

Ten-step Asymmetric Total Syntheses of Potent Antibiotic Anthracimycin and Anthracimycin B

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Table of content

General Information.....	3
Comparison of intramolecular Diels-Alder reaction.....	4
Ten-step synthetic route for anthracimycin.....	4
Preparation of compound 2	5
Preparation of compound 3	6
Preparation of compound S-1	7
Preparation of compound 5a and 7	8
Preparation of compound 9	9
Preparation of compound 10	10
Preparation of compound 11	11
Preparation of compound 12a	12
Preparation of compound S-4	13
Preparation of compound 13a	14
Preparation of compound S-6	15
Preparation of compound 14a	16
Preparation of compound S-7 and 16a	17
Preparation of compound S-8	19
Preparation of compound 13b	20
Preparation of compound S10 and 14b	21
Preparation of compound 14c	22

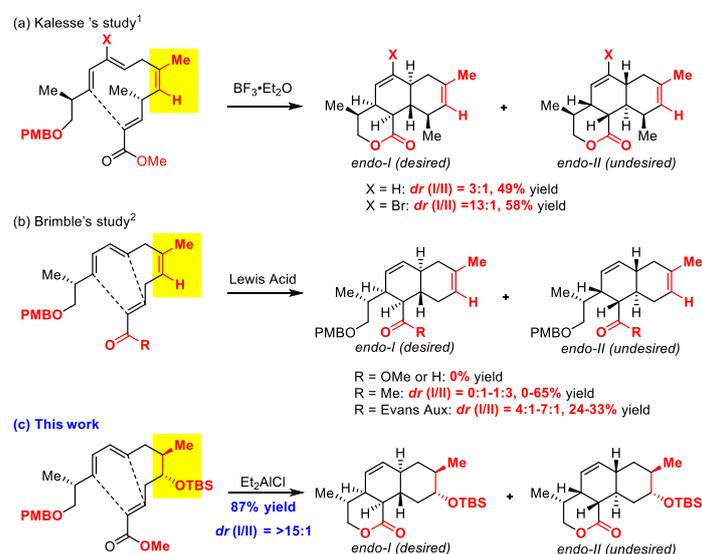
Preparation of compound 16b	23
Preparation of compound 16c and S-5	24
Preparation of compound 5b	25
Preparation of compound 8a and 8e	26
Preparation of compound 13b	27
Preparation of anthracimycin.....	28
Preparation of anthracimycin B.....	29
NMR comparison of natural and synthetic anthracimycin and anthracimycin B.....	31
X-Ray Crystal Structure of compound 10	35
X-Ray Crystal Structure of anthracimycin.....	36
Antibacterial assays.....	37
References.....	40
Copies of ¹ H NMR and ¹³ C NMR spectra.....	41

General Information

Reactions were carried out in oven-dried glassware under an argon atmosphere, unless otherwise noted. Tetrahydrofuran (THF) was freshly distilled before use from sodium using benzophenone as indicator. Dichloromethane was freshly distilled before use from calcium hydride (CaH₂). Solvents used in workup, extraction and column chromatography were used as received from commercial suppliers without further purification. Reactions were magnetically stirred and monitored by thin layer chromatography (TLC, 0.25 mm) on Merck pre-coated silica gel plates. Flash chromatography was performed with silica gel 60 (particle size 0.040 – 0.062 mm) supplied by Grace. ¹H and ¹³C NMR spectra were recorded on a Bruker AV-400 spectrometer (400 MHz for ¹H, 101 MHz for ¹³C). Chemical shifts are reported in parts per million (ppm) as values relative to residual chloroform peaks (7.26 ppm for ¹H and 77.0 ppm for ¹³C). Abbreviations for signal coupling are as follows: s, singlet; d, doublet; t, triplet; q, quartet; m, multiplet. Optical rotations were measured on a JASCO Perkin-Elmer model P-2000 polarimeter and RUDOLPH API, A28576-T-LED. High resolution mass spectra were measured at the Hong Kong University of Science and Technology Mass Spectrometry Service Center on either an Agilent GC/MS 5975C system or an API QSTAR XL System.

Comparison of intramolecular Diels–Alder reaction

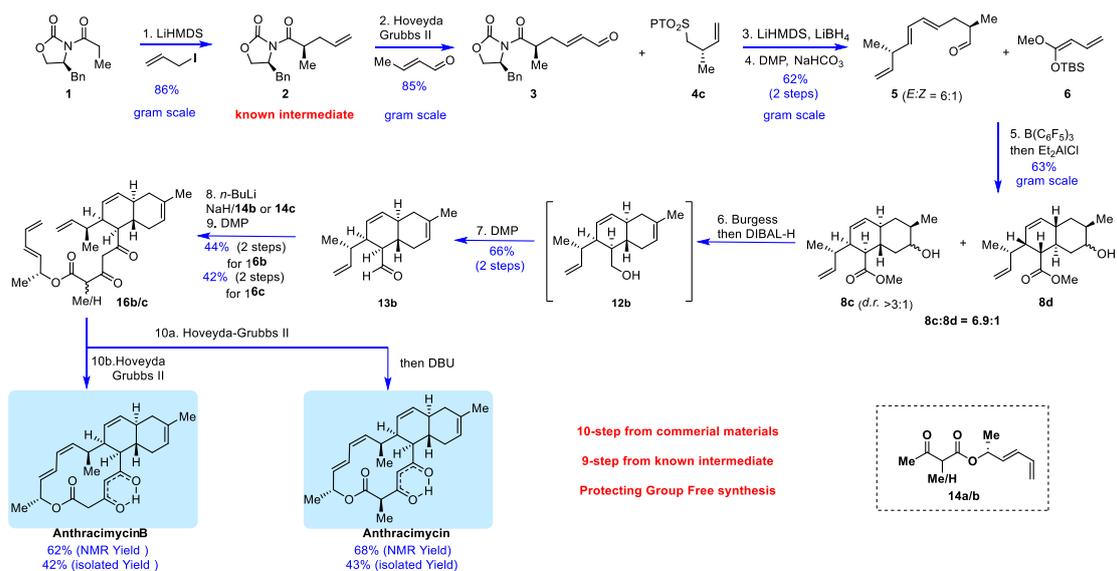
To address the challenge of the bioinspired intramolecular Diels–Alder reaction of tetraene substrates employed by Kalesse¹ and Brimble² and highlight our design of the intramolecular Diels–Alder reaction substrate, we summarized these results in Scheme S1. Kalesse *et al.* found the bromodiene enhanced yield and selectivity, while Brimble *et al.* identified Evans chiral auxiliary to be advantageous for the bioinspired Diels–Alder reaction. Our strategically designed substrate delivered the desired decalin with excellent yield (87%) and high selectivity (*d.r.*>15:1).



Scheme S1. Construction of the *trans*-decalin core through Diels–Alder reaction

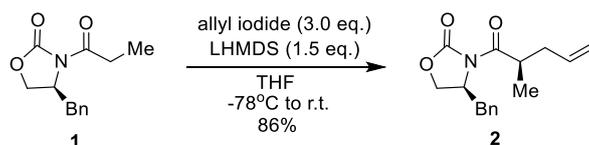
Ten-step synthetic route for anthracimycin and anthracimycin B

With our optimized synthesis of decalin intermediate **13b**, the total number of steps for anthracimycin and anthracimycin B was reduced to 10 without using any protecting groups and with substantially improved overall yield (>3%) (Scheme S2).



Scheme S2. Ten-step synthetic route for anthracimycin and anthracimycin B

Preparation of Compound 2



Compound **2** was prepared according to a slightly modified procedure described by Liu and coworkers³. To a solution of compound **1** (25.00 g, 107.2 mmol, 1.0 equiv.) in 250 mL THF at -78°C was added LHMDS (1.0 M in THF, 160.7 mL, 160.7 mmol, 1.5 equiv. , dropwise). After the reaction mixture was stirred at the same temperature for 2 hours, allyl iodide (29.4 mL, 321.5 mmol, 3 equiv.) was added and the resulting solution was stirred for 10 hours. The reaction was allowed to warm up to room temperature and stirred for another 30 minutes before being quenched with sat. aq. NH₄Cl (100 mL). The two phases were separated and the aqueous phase was extracted with EtOAc (3 × 150 mL). The combined organic phases were washed with brine, dried over Na₂SO₄ and concentrated *in vacuo*. The crude residue was purified by flash chromatography on silica gel (eluent: EtOAc/hexane, 1/15 to 1/10) to give compound **2** (25.2 g, 86%) as a colorless oil.

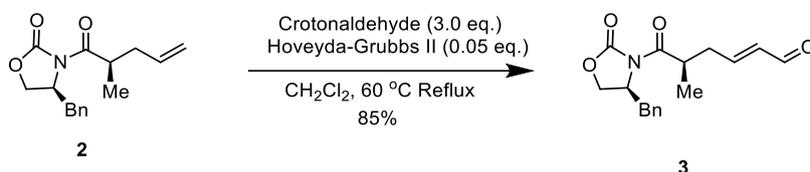
$[\alpha]_D^{25} = +31.60$ (*c* 1, CHCl₃);

¹H NMR (400 MHz, CDCl₃) δ 7.39 – 7.14 (m, 5H), 5.94 – 5.71 (m, 1H), 5.27 – 4.97 (m, 2H), 4.80 – 4.59 (m, 1H), 4.29 – 4.08 (m, 2H), 3.96 – 3.74 (m, 1H), 3.29 (dd, *J* = 13.3, 3.4 Hz, 1H), 2.71 (dd, *J* = 13.3, 9.8 Hz, 1H), 2.63 – 2.37 (m, 1H), 2.30 – 2.15 (m, 1H), 1.19 (d, *J* = 6.8 Hz, 3H).

¹³C NMR (101 MHz, CDCl₃) δ 176.4, 153.0, 135.3, 135.2, 129.3, 128.8, 127.2, 117.1, 65.9, 55.3, 38.0, 37.9, 37.1, 16.3.

HRMS (TOF, ES⁺) *m/z* calculated for [C₁₇H₁₉NO₄+Na]⁺ 296.1257, found 296.1259.

Preparation of Compound 3



A solution of compound **2** (25.2 g, 92.2 mmol, 1.0 equiv.) in 460 mL CH₂Cl₂, was degassed with argon sparging for 5 minutes. Then crotonaldehyde (22.8 ml, 276.9 mmol, 3.0 equiv.) and Hoveyda Grubbs II catalyst (2.89 g, 4.6 mmol, 0.05 equiv.) were added sequentially to the solution. The reaction mixture was heated to reflux for 3 days, and then cooled to room temperature. The CH₂Cl₂ solvent was removed under reduced pressure and the resulting residue was purified by flash chromatography on silica gel (eluent: EtOAc/hexane, 1/10 to 1/3) to give compound **3** (27.8 g, 85%) as a brown oil.

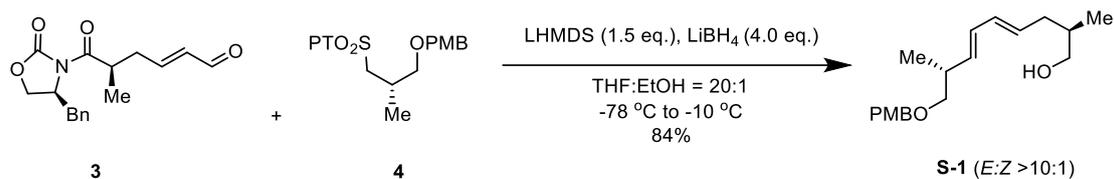
[α]_D²⁵ = +32.10 (*c* 1, CHCl₃);

¹H NMR (400 MHz, CDCl₃): δ 9.51 (d, *J* = 7.8 Hz, 1H), 7.36 – 7.27 (m, 3H), 7.21 – 7.14 (m, 2H), 6.85 (dt, *J* = 15.7, 7.1 Hz, 1H), 6.24 – 6.10 (m, 1H), 4.72 – 4.62 (m, 1H), 4.26 – 4.14 (m, 2H), 4.00 – 3.90 (m, 1H), 3.24 (dd, *J* = 13.3, 3.4 Hz, 1H), 2.86 – 2.68 (m, 2H), 2.48 (dtd, *J* = 14.5, 7.1, 1.4 Hz, 1H), 1.23 (d, *J* = 6.8 Hz, 3H).

¹³C NMR (101 MHz, CDCl₃): δ 193.6, 175.3, 154.6, 153.1, 135.0, 134.7, 129.3, 128.9, 127.4, 66.2, 55.4, 37.9, 36.7, 36.3, 16.8.

HRMS (TOF, ES⁺) *m/z* calculated for [C₁₇H₁₉NO₄+Na]⁺ 324.1206, found 324.1211.

Preparation of Compound S-1



To a solution of aldehyde **3** (4.91 g, 16.3 mmol, 1.0 equiv.) and sulfone **4**⁴ (7.87 g, 19.6 mmol, 1.2 equiv.) in 160 mL THF at -78 °C was added dropwise LHMDS (1.0 M in THF, 21.2 mL, 21.2 mmol, 1.3 equiv.). The reaction mixture was stirred at -78 °C for 2 hours before adding 8.2 mL ethanol. Then reaction mixture was allowed to warm up to -10 °C and then LiBH₄ (2.0 M in THF, 32.6 mL, 65.2 mmol, 4.0 equiv.) was added dropwise to the reaction mixture. After stirring for another 2 hours or until completion of the reaction as indicated by TLC, the reaction was allowed to warm up to 0 °C and quenched with 1 N NaOH (70 mL). The organic phase was collected and the aqueous phase was extracted with EtOAc (3 × 100 mL). The combined organic phases were washed with brine, dried over Na₂SO₄ and concentrated *in vacuo*. The crude residue was purified by flash chromatography on silica gel (eluent: EtOAc/hexane, 1/5 to 1/2) to give compound **S-1** (4.15 g, 84%) as a light yellow oil.

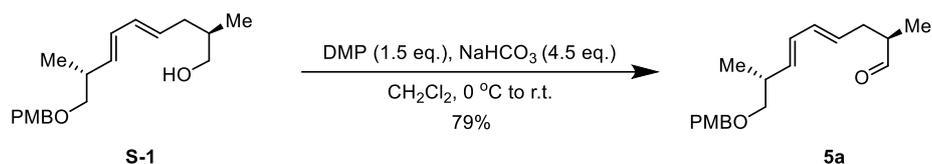
$[\alpha]_D^{25} = -0.15$ (*c* 2, CHCl₃);

¹H NMR (400 MHz, CDCl₃): δ 7.32 – 7.21 (m, 2H), 6.94 – 6.85 (m, 2H), 6.14 – 5.98 (m, 2H), 5.68 – 5.49 (m, 2H), 4.47 (s, 2H), 3.83 (s, 3H), 3.59 – 3.43 (m, 2H), 3.32 (ddd, *J* = 32.6, 9.2, 6.7 Hz, 2H), 2.60 – 2.46 (m, 1H), 2.26 – 2.15 (m, 1H), 2.04 – 1.92 (m, 1H), 1.82 – 1.65 (m, 1H), 1.54 (s, 1H), 1.05 (d, *J* = 6.8 Hz, 3H), 0.94 (d, *J* = 6.8 Hz, 3H).

¹³C NMR (101 MHz, CDCl₃): δ 159.1, 134.9, 132.0, 130.6, 130.1, 129.7, 129.2, 113.7, 74.9, 72.6, 67.9, 55.2, 36.8, 36.5, 36.1, 17.0, 16.5.

HRMS (TOF, ES⁺) *m/z* calculated for [C₁₉H₂₈O₃+Na]⁺ 327.1931, found 327.1932.

Preparation of Compound 5a



To a solution of alcohol **S-1** (4.15 g, 13.6 mmol, 1.0 equiv.) in 136 mL CH₂Cl₂ at 0 °C were added NaHCO₃ (5.15 g, 61.3 mmol, 4.5 equiv.) and Dess-Martin periodinane (8.67 g, 20.4 mmol, 1.5 equiv.). The reaction mixture was stirred at 0 °C for 2 hours, and then the reaction was quenched by addition of sat. aq. NaHCO₃ (30 mL) and sat. aq. Na₂S₂O₃ (60 mL). The reaction mixture was stirred for another 30 minutes. The organic phase was collected and the aqueous phase was extracted with CH₂Cl₂ (3 × 60 mL). The combined organic phases were washed with brine, dried over Na₂SO₄, concentrated under reduced pressure. The resulting residue was purified by flash chromatography on silica gel (eluent: EtOAc/hexane, 1/5) to afford aldehyde **5a** (3.26 g, 79%) as a pale-yellow oil.

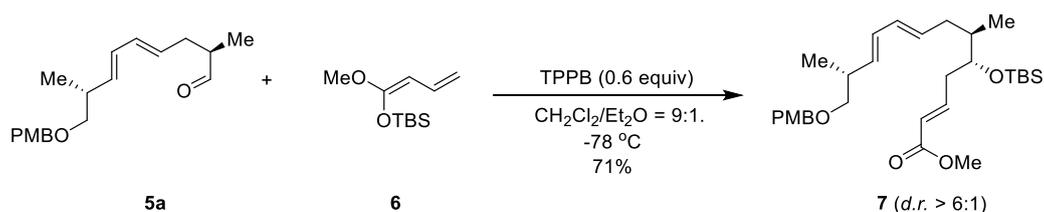
$[\alpha]_D^{25} = -0.40$ (*c* 2, CHCl₃);

¹H NMR (400 MHz, CDCl₃): δ 9.65 (d, *J* = 1.4 Hz, 1H), 7.25 (d, *J* = 8.8 Hz, 2H), 6.87 (d, *J* = 8.6 Hz, 2H), 6.12 – 5.96 (m, 2H), 5.65 – 5.45 (m, 2H), 4.44 (s, 2H), 3.80 (s, 3H), 3.33 (dd, *J* = 9.1, 6.6 Hz, 1H), 3.26 (dd, *J* = 9.1, 6.8 Hz, 1H), 2.58 – 2.36 (m, 3H), 2.22 – 2.10 (m, 1H), 1.10 (d, *J* = 7.0 Hz, 3H), 1.03 (d, *J* = 6.8 Hz, 3H).

¹³C NMR (101 MHz, CDCl₃): δ 204.6, 159.0, 135.8, 1323.0, 130.6, 129.3, 129.1, 128.3, 113.7, 74.7, 72.5, 55.2, 46.2, 36.8, 33.6, 17.0, 13.1.

HRMS (TOF, ES⁺) *m/z* calculated for [C₁₉H₂₆O₃+Na]⁺ 325.1774, found 325.1776.

Preparation of Compound 7



for 18 hours before addition of 1.0 mL H₂O and 15.0 mL concentrated HCl at 0 °C. The reaction mixture was stirred for another 10 hours before addition of 2 N NaOH (100 mL). The organic phase was collected, and the aqueous phase was extracted by CH₂Cl₂ (3 × 50 mL). The combined organic phases were washed with brine, dried over Na₂SO₄, concentrated under reduced pressure. The resulting residue was purified by flash chromatography on silica gel (eluent: EtOAc/hexane, 1/2) to afford lactone **9** (1.58 g, 87%) as a white solid.

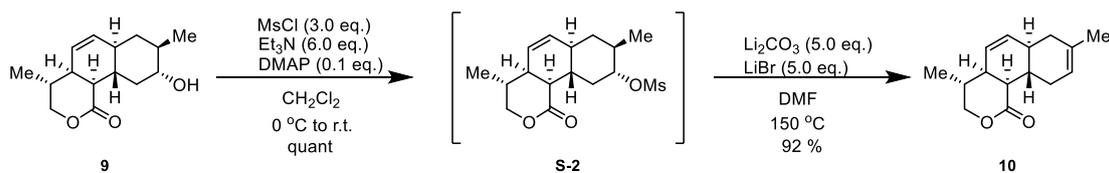
$[\alpha]_D^{25} = -26.67$ (*c* 0.3, CHCl₃);

¹H NMR (400 MHz, CDCl₃): δ 5.74 – 5.65 (m, 1H), 5.61 (d, *J* = 10.0 Hz, 1H), 4.28 (dd, *J* = 11.3, 4.7 Hz, 1H), 3.87 (t, *J* = 11.0 Hz, 1H), 3.25 – 3.12 (m, 1H), 2.61 (dd, *J* = 11.2, 6.0 Hz, 1H), 2.30 – 2.18 (m, 1H), 2.09 – 1.91 (m, 2H), 1.91 – 1.74 (m, 2H), 1.52 – 1.42 (m, 2H), 1.39 – 1.24 (m, 1H), 1.03 (dd, *J* = 15.6, 6.5 Hz, 6H), 0.88 (q, *J* = 12.5 Hz, 1H).

¹³C NMR (101 MHz, CDCl₃): δ 171.9, 132.8, 126.0, 76.0, 74.2, 45.4, 41.7, 39.8, 39.6, 39.2, 37.0, 32.1, 18.2, 14.3.

HRMS (TOF, ES⁺) *m/z* calculated for [C₁₅H₂₂O₃+Na]⁺ 273.1461, found 273.1461.

Preparation of Compound 10



To a solution of lactone **9** (1.58 g, 6.31 mmol, 1.0 equiv.), DMAP (77 mg, 0.63 mmol, 0.1 eq) and Et₃N (8.8 mL, 63.3 mmol, 10.0 eq.) in 63 mL CH₂Cl₂ was added dropwise MsCl (2.45 mL, 31.6 mmol, 5.0 equiv.) at 0 °C. The reaction mixture was stirred for 18 hours at room temperature before quenching by addition of sat. aq. NHCl₄ (30 mL). The organic phase was collected and the aqueous phase was extracted with CH₂Cl₂ (3 × 30 mL). The combined organic phases were washed with brine and dried over Na₂SO₄, concentrated under reduced pressure. The residue was dissolved in 32.0 mL DMF, and Li₂CO₃ (2.34 g, 31.7 mmol, 5.0 equiv.) and LiBr

(2.75 g, 31.7 mmol, 5.0 equiv.) were added. The reaction mixture was heated to 150 °C for 1 hour and then cooled to room temperature. 250 mL Et₂O was added to the reaction mixture. The organic phase was washed with H₂O (3 × 30 mL), dried over Na₂SO₄, concentrated under reduced pressure. The residue was purified by flash chromatography on silica gel (eluent: EtOAc/hexane, 1/5) to give lactone **10** (1.35 g, 92%) as a white solid.

See **X-Ray** crystal structure analysis in page-35 (CCDC 2175241)

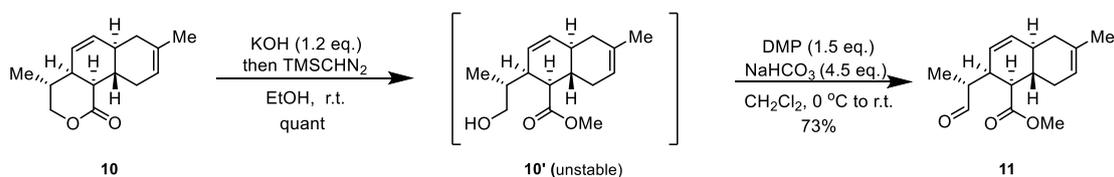
$[\alpha]_D^{25} = +94.10$ (*c* 1, CHCl₃);

¹H NMR (400 MHz, CDCl₃): δ 5.81 – 5.71 (m, 1H), 5.71 – 5.61 (m, 1H), 5.45 – 5.38 (m, 1H), 4.29 (dd, *J* = 11.3, 4.9 Hz, 1H), 3.85 (t, *J* = 11.0 Hz, 1H), 2.56 (dd, *J* = 11.6, 5.5 Hz, 1H), 2.27 – 2.16 (m, 1H), 2.17 – 1.93 (m, 5H), 1.85 – 1.70 (m, 1H), 1.72 – 1.62 (m, 3H), 1.01 (d, *J* = 6.6 Hz, 3H).

¹³C NMR (101 MHz, CDCl₃): δ 172.2, 133.1, 132.8, 125.7, 121.0, 74.6, 45.2, 40.0, 38.4, 37.3, 34.6, 32.1, 30.5, 23.2, 14.3.

HRMS (TOF, ES⁺) *m/z* calculated for [C₁₅H₂₀O₂+Na]⁺ 255.1356, found 255.1361.

Preparation of Compound 11



To a stirred solution of lactone **10** (112 mg, 0.48 mmol, 1.0 equiv.) in 2.5 mL EtOH was added 4 *N* KOH (0.15mL, 0.58 mmol, 1.2 equiv.). The reaction mixture was stirred for 14 hours at room temperature, then EtOH was removed under reduced pressure. The residue was suspended in H₂O (4 mL) and treated with 1 *N* HCl (2.3 ml) until pH of 4-5. The aqueous phase was extracted with EtOAc (3 × 10 mL) and the combined organic phases were washed with H₂O (3 × 30 mL), dried over Na₂SO₄, and concentrated under reduced pressure. The crude product was dissolved in 24.0 mL MeOH/EtOAc (2:1) and a solution of TMSCHN₂ (2.0 M hexane, 1.0 mL, 2 mmol, 4.2 equiv.) was added dropwise into the reaction mixture until the solution became

yellow and persisted for 1 minute. After the reaction solvent and excess of TMSCHN₂ were removed under reduced pressure, 5 mL CH₂Cl₂ was added to the residue. The resulting solution was cooled down to 0 °C, and then NaHCO₃ (182 mg, 2.17 mmol, 4.5 equiv.) and Dess-Martin periodinane (307 mg, 0.72 mmol, 1.5 equiv.) were added. The reaction mixture was stirred for another 1.5 hours at room temperature before addition of sat. aq. NaHCO₃ (2 mL) and sat. aq. Na₂S₂O₃ (5 mL) with stirring for another 30 minutes. The organic phase was collected and the aqueous phase was extracted with CH₂Cl₂ (3 × 10 mL). The combined organic phases were washed with brine, dried over Na₂SO₄, concentrated under reduced pressure. The residue was purified through flash chromatography (eluent: EtOAc/hexane, 1/50) to afford aldehyde **11** (92 mg, 73%) as a colourless oil.

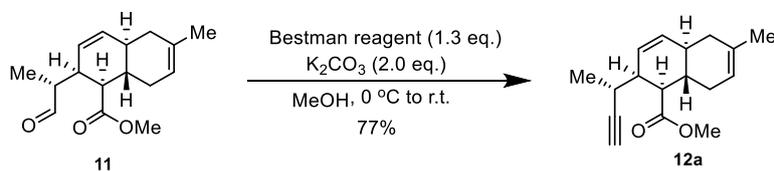
$[\alpha]_D^{25} = +249.9$ (*c* 1, CHCl₃);

¹H NMR (400 MHz, CDCl₃): δ 9.55 (s, 1H), 5.65 (d, *J* = 10.0 Hz, 1H), 5.43 – 5.34 (m, 1H), 5.31 (d, *J* = 5.4 Hz, 1H), 3.60 (s, 3H), 3.19 – 3.12 (m, 1H), 2.64 (dd, *J* = 11.6, 6.3 Hz, 1H), 2.56 – 2.42 (m, 1H), 2.22 – 2.12 (m, 1H), 2.03 – 1.91 (m, 2H), 1.84 – 1.48 (m, 6H), 1.06 (d, *J* = 7.3 Hz, 3H).

¹³C NMR (101 MHz, CDCl₃): δ 203.3, 174.2, 133.6, 133.1, 124.9, 120.5, 51.3, 48.7, 47.9, 37.3, 36.8, 35.1, 33.9, 30.4, 23.2, 11.2.

HRMS (TOF, CI⁺) *m/z* calculated for [C₁₆H₂₂O₃+Na]⁺ 285.1461, found 285.1467.

Preparation of Compound 12a



To a stirred solution of aldehyde **11** (380 mg, 1.45 mmol, 1.0 equiv.) in 20 mL MeOH at 0 °C were added K₂CO₃ (401 mg, 2.90 mmol, 2.0 equiv.) and dimethyl (1-diazo-2-oxopropyl)phosphonate (0.29 mL, 1.89 mmol, 1.3 equiv.). After the reaction mixture was allowed to warm to room temperature and stirred for 2 hours. The solvent was removed under reduced pressure. The residue was dissolved in

EtOAc (30 mL) and washed with sat. aq. NaHCO₃ (10 mL). The organic phase was concentrated under reduced pressure and purified via flash chromatography (eluent: EtOAc/hexane, 1/80) to afford alkyne **12a** (288 mg, 77%) as a colourless oil.

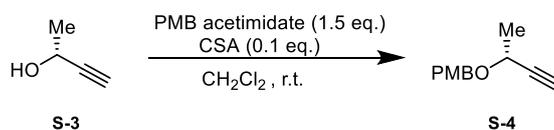
$[\alpha]_D^{25} = +230.33$ (*c* 0.3, CHCl₃);

¹H NMR (400 MHz, CDCl₃): δ 5.86 – 5.77 (m, 1H), 5.77 – 5.68 (m, 1H), 5.38 – 5.27 (m, 1H), 3.69 (d, *J* = 4.6 Hz, 3H), 2.86 – 2.76 (m, 1H), 2.64 (dd, *J* = 11.9, 6.8 Hz, 1H), 2.57 – 2.43 (m, 1H), 2.30 (dd, *J* = 16.9, 6.0 Hz, 1H), 2.06 (d, *J* = 2.4 Hz, 1H), 2.05 – 1.92 (m, 2H), 1.83 – 1.69 (m, 2H), 1.66 – 1.62 (m, 3H), 1.61 – 1.48 (m, 1H), 1.16 (d, *J* = 7.0 Hz, 3H).

¹³C NMR (101 MHz, CDCl₃): δ 173.7, 133.7, 133.1, 124.8, 120.7, 88.7, 68.5, 51.3, 49.2, 40.8, 37.3, 37.1, 33.8, 30.8, 27.8, 23.3, 17.8.

HRMS (TOF, CI⁺) *m/z* calculated for [C₁₇H₂₂O₂+H]⁺ 259.1693, found 259.1693.

Preparation of Compound S-4



To a stirred solution of alkyne **S-3** (1.50 g, 21.4 mmol, 1.0 equiv.) and *para*-methoxy benzyl acetimidate (9.07 g, 32.1 mmol, 1.5 equiv.) in 107 mL CH₂Cl₂ was added camphorsulfonic acid (497 mg, 2.14 mmol, 0.1 equiv.). The reaction mixture was stirred at room temperature for 15 hours before quenched with sat. aq. NaHCO₃ (50 mL). The organic phase was collected and the aqueous phase was extracted with CH₂Cl₂ (3 × 50 mL). The combined organic phases were washed with brine, dried over Na₂SO₄, concentrated under reduced pressure. The residue was purified through flash chromatography (eluent: EtOAc/hexane, 1/50) to afford alkyne **S-4** (3.63 g, 89%) as a yellow oil.

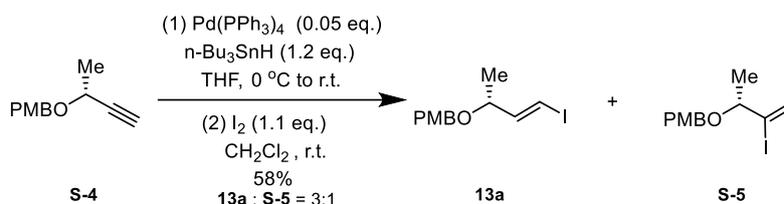
$[\alpha]_D^{25} = +133.60$ (*c* 1, CHCl₃);

¹H NMR (400 MHz, CDCl₃): δ 7.39 – 7.30 (m, 2H), 6.99 – 6.82 (m, 2H), 4.78 (d, *J* = 11.3 Hz, 1H), 4.49 (d, *J* = 11.3 Hz, 1H), 4.28 – 4.18 (m, 1H), 3.83 (s, 3H), 2.53 (d, *J* = 2.0 Hz, 1H), 1.51 (d, *J* = 6.6 Hz, 3H).

^{13}C NMR (101 MHz, CDCl_3): δ 159.2, 129.6, 113.7, 83.7, 73.0, 70.0, 63.7, 55.1, 21.9.

HRMS (TOF, CI^+) m/z calculated for $[\text{C}_{12}\text{H}_{14}\text{O}_2]^+$ 190.0988, found 190.0996.

Preparation of Compound 13a



To a stirred solution of alkyne **S-4** (3.63 g, 19.1 mmol, 1.0 equiv.) and $\text{Pd}(\text{PPh}_3)_4$ in 95.0 mL THF at 0 °C was added $n\text{-Bu}_3\text{SnH}$ (6.16 mL, 22.9 mmol, 1.2 equiv.). After the reaction mixture was allowed to warm to room temperature and stirred for 10 hours. The solvent was removed under reduced pressure. The residue was dissolved in 100 mL CH_2Cl_2 and titrated with a solution of I_2 (2.67 g, 21.0 mmol, 1.1 equiv.) in THF (30 mL) until the reddish color persisted. The reaction was quenched with sat. aq. $\text{Na}_2\text{S}_2\text{O}_3$ (50 mL). The organic phase was collected and the aqueous phase was extracted with CH_2Cl_2 (3×50 mL). The combined organic phases were washed with brine, dried over Na_2SO_4 , concentrated under reduced pressure. The residue was purified through flash chromatography (eluent: EtOAc/hexane, 1/100) to afford vinyl iodide **13a** (3.52 g, 58%) as a yellow oil.

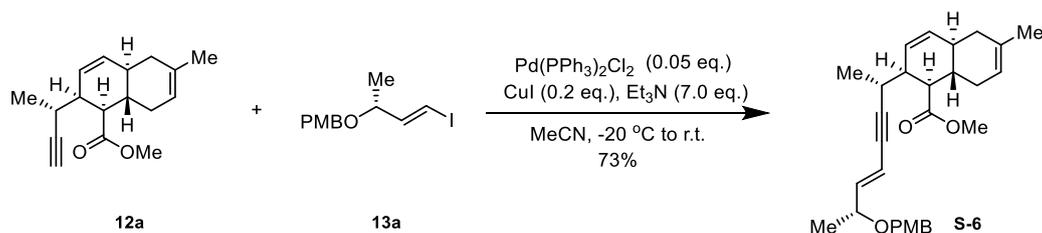
$[\alpha]_D^{25} = +71.20$ (c 1, CHCl_3);

^1H NMR (400 MHz, CDCl_3): δ 7.30 (d, $J = 8.6$ Hz, 2H), 6.98 – 6.91 (m, 2H), 6.57 (dd, $J = 14.5, 7.3$ Hz, 1H), 6.36 (dd, $J = 14.5, 0.8$ Hz, 1H), 4.55 (d, $J = 11.5$ Hz, 1H), 4.37 (d, $J = 11.5$ Hz, 1H), 3.99 – 3.87 (m, 1H), 3.83 (s, 3H), 1.32 (d, $J = 6.5$ Hz, 3H).

^{13}C NMR (101 MHz, CDCl_3): δ 158.9, 147.8, 129.0, 113.6, 79.4, 76.6, 69.7, 55.0, 20.6.

HRMS (TOF, CI^+) m/z calculated for $[\text{C}_{12}\text{H}_{15}\text{IO}_2]^+$ 318.0111, found 318.0113.

Preparation of Compound S-6



To a solution of alkyne **12a** (130 mg, 0.50 mmol, 1.0 equiv.) and vinyl iodide **13a** (304mg, 0.96 mmol, 1.9 equiv.) in 15 mL anhydrous acetonitrile at -78 °C were added Pd(PPh₃)Cl₂ (18mg, 0.03mmol, 0.05 equiv.) and CuI (19mg, 0.1mmol, 0.2 equiv.). The reaction solution was degassed with argon. The reaction mixture under argon atmosphere was allowed to warm up to -20 °C before triethylamine (0.49ml, 3.52 mmol, 7.0 equiv.) was added. The reaction mixture was stirred for 30 minutes and then warmed to room temperature and stirred for another 30 minutes before the reaction was quenched with phosphate buffer (pH 7, 10 mL). EtOAc (15 mL) was added, and the reaction mixture was stirred at room temperature for 20 minutes. The organic phase was collected and the aqueous phase was extracted with EtOAc (3 × 15 mL). The combined organic phases were washed with brine, dried over Na₂SO₄, concentrated under reduced pressure. The residue was purified through flash chromatography (eluent: EtOAc/hexane, 1/50) to afford compound **S-6** (165 mg, 73%) as a colourless oil.

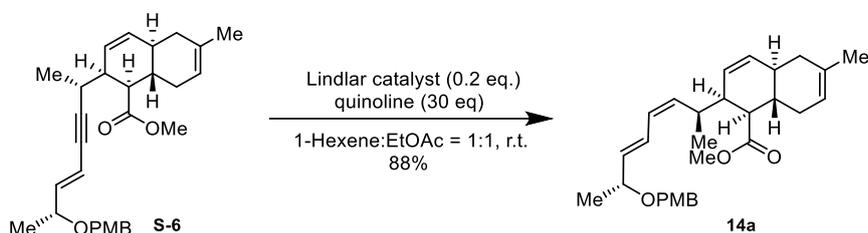
$[\alpha]_D^{25} = +172.00$ (*c* 0.2, CHCl₃);

¹H NMR (400 MHz, CDCl₃): δ 7.26 – 7.20 (m, 2H), 6.91 – 6.83 (m, 2H), 5.98 (dd, *J* = 15.9, 7.4 Hz, 1H), 5.89 – 5.76 (m, 1H), 5.73 (d, *J* = 10.1 Hz, 1H), 5.65 (dd, *J* = 15.9, 1.7 Hz, 1H), 5.42 – 5.33 (m, 1H), 4.49 (d, *J* = 11.6 Hz, 1H), 4.29 (d, *J* = 11.5 Hz, 1H), 3.97 – 3.86 (m, 1H), 3.80 (s, 3H), 3.70 (d, *J* = 1.7 Hz, 3H), 2.88 – 2.79 (m, 1H), 2.70 – 2.56 (m, 2H), 2.33 (d, *J* = 16.7 Hz, 1H), 2.09 – 1.93 (m, 2H), 1.84 – 1.77 (m, 1H), 1.77 – 1.68 (m, 1H), 1.64 (d, *J* = 14.7 Hz, 3H), 1.61 – 1.53 (m, 1H), 1.26 (d, *J* = 6.3 Hz, 3H), 1.17 (d, *J* = 6.9 Hz, 3H).

¹³C NMR (101 MHz, CDCl₃): δ 173.8, 143.7, 133.7, 133.0, 130.6, 129.2, 125.1, 120.8, 113.8, 111.4, 74.8, 69.8, 55.2, 51.3, 49.3, 41.0, 37.4, 37.1, 33.8, 30.9, 28.7, 23.3, 21.3, 17.9.

HRMS (TOF, ES⁺) m/z calculated for [C₂₉H₃₆O₄+Na]⁺ 471.2506, found 471.2510.

Preparation of Compound 14a



To a stirred solution of **S-6** (80 mg, 0.18 mmol, 1.0 equiv.) in 14 mL EtOAc/1-Hexene (1:1) were added quinoline (0.63 mL, 5.3 mmol, 30.0 equiv.) and Lindlar catalyst (Aldrich, 5 % Pd on CaCO₃ poisoned with Pb, 80 mg). The reaction mixture was stirred under H₂ (1 atm) atmosphere for 2.5 hours. The reaction mixture was passed through a plug of SiO₂ (eluent: EtOAc). The filtrates were concentrated under reduced pressure. The residue was purified by flash chromatography on silica gel (eluent: EtOAc/hexane, 1/50) to afford compound **14a** (70 mg, 88%) as a colourless oil.

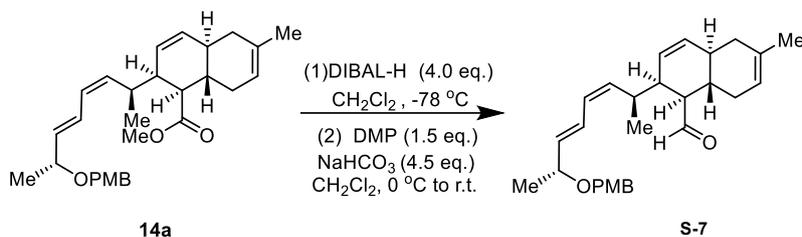
$[\alpha]_D^{25} = +101.25$ (*c* 0.8, CHCl₃);

¹H NMR (400 MHz, CDCl₃): δ 7.33 – 7.21 (m, 2H), 6.87 (dd, *J* = 9.0, 2.5 Hz, 2H), 6.40 – 6.28 (m, 1H), 5.97 – 5.85 (m, 1H), 5.77 – 5.54 (m, 3H), 5.45 – 5.32 (m, 2H), 4.50 (dd, *J* = 11.5, 9.0 Hz, 1H), 4.41 – 4.25 (m, 1H), 4.00 – 3.87 (m, 1H), 3.80 (s, 3H), 3.72 – 3.59 (m, 3H), 2.62 (dd, *J* = 9.7, 4.9 Hz, 2H), 2.52 – 2.41 (m, 1H), 2.11 – 1.95 (m, 2H), 1.83 (t, *J* = 4.4 Hz, 2H), 1.70 – 1.66 (m, 3H), 1.61 – 1.49 (m, 1H), 1.35 – 1.23 (m, 3H), 1.23 – 1.11 (m, 1H), 1.00 – 0.83 (m, 3H).

¹³C NMR (101 MHz, CDCl₃): δ 173.8, 159.0, 137.8, 135.6, 133.7, 132.5, 130.8, 129.4, 126.9, 126.4, 125.4, 121.0, 113.7, 74.9, 69.3, 55.2, 51.0, 49.5, 42.6, 37.5, 37.2, 34.1, 33.7, 31.1, 23.3, 21.6, 17.9.

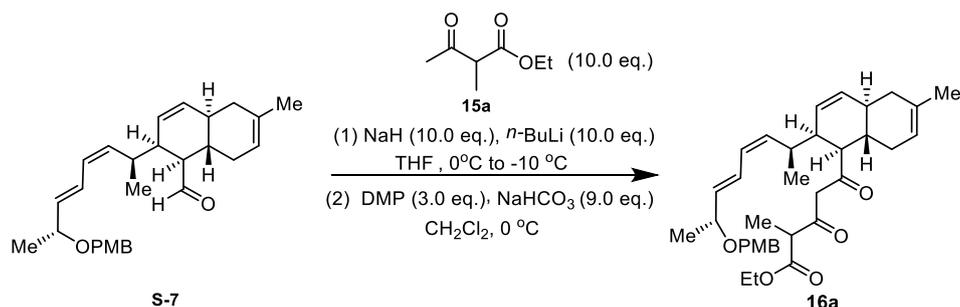
HRMS (TOF, ES⁺) m/z calculated for [C₂₉H₃₈O₄+Na]⁺ 473.2662, found 473.2665.

Preparation of Compound S-7



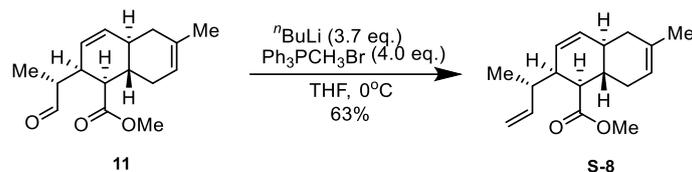
To a stirred solution of **14a** (70 mg, 0.16 mmol, 1.0 equiv.) in 3.0 mL CH₂Cl₂ at -78 °C was added dropwise DIBAL-H (1.0 M in hexane, 0.6 mL, 0.6 mmol, 4.0 equiv.). The reaction mixture was stirred for 2 hours and then quenched by 2 N HCl (5 mL). The resulting solution was stirred at room temperature for another 30 minutes. The organic phase was collected and the aqueous phase was extracted with CH₂Cl₂ (3 × 10 mL). The combined organic phases were washed with brine, dried over Na₂SO₄, concentrated under reduced pressure. The residue was dissolved in 3.0 mL CH₂Cl₂ was cooled down to 0 °C, and NaHCO₃ (59 mg, 0.70 mmol, 4.5 equiv.) and Dess-Martin periodinane (99 mg, 0.23 mmol, 1.5 equiv.) were added. The reaction mixture was stirred for another 1.5 hours at room temperature before sat. aq. NaHCO₃ (3 mL) and sat. aq. Na₂S₂O₃ (3 mL) were added. The organic phase was collected and the aqueous phase was extracted with CH₂Cl₂ (3 × 10 mL). The combined organic phases were washed with brine, dried over Na₂SO₄ and concentrated under reduced pressure. The residue was dissolved in 0.5 mL EtOAc and the suspension was filtered through a plug of SiO₂ (eluent: EtOAc/hexane, 1/5). The filtrates were concentrated under reduced pressure. The crude product **S-7** (56 mg, check by ¹H NMR) was used directly in the next step without other purifications.

Preparation of Compound 16a



To a stirred suspension of sodium hydride (60% dispersion in mineral oil, 53 mg, 1.33 mmol, 10.0 equiv.) in 4.5 mL THF at 0 °C was added a solution of **15a** (192 mg, 1.33 mmol, 10.0 equiv. , dropwise) in 4.5 mL THF. The reaction mixture was stirred for 30 minutes and then cooled to -10 °C before *n*BuLi (2.5 M in hexane, 0.53 mL, 1.33 mmol, 10.0 equiv. , dropwise) was added. After stirring for another 30 minutes at -10 °C, a solution of the crude product **S-7** (56 mg, 0.13 mmol, 1 equiv. , dropwise) in 2.0 mL THF was then added. The reaction mixture was stirred for 2 hours at the same temperature before water (2 mL) was added to quench the reaction. The reaction mixture was diluted with Et₂O (10 mL), and acidified with 1 N HCl until pH of 4-5. The organic layer was collected and the aqueous layer was extracted with Et₂O (3 × 15 mL). The combined organic phases were washed with brine, dried over Na₂SO₄, concentrated under reduced pressure. The crude product dissolved in CH₂Cl₂ (5 mL) was cooled to 0° C and then NaHCO₃ (98 mg, 1.17 mmol, 9.0 equiv.) and Dess-Martin periodinane (166 mg, 0.40 mmol, 3.0 equiv.) were added. The reaction mixture was stirred for another 1.5 hours at 0 °C (the product decomposed if the reaction mixture was allowed to warm to room temperature) before sat. aq. NaHCO₃ (2 mL) and sat. aq. Na₂S₂O₃ (5 mL) were added to quench the reaction. The resulting reaction mixture was allowed to warm to room temperature and stirred for another 30 minutes. The organic phase was collected and the aqueous phase was extracted with CH₂Cl₂ (3 × 10 mL). The combined organic phases were washed with brine, dried over Na₂SO₄ and concentrated under reduced pressure. The residue dissolved in 0.5 mL EtOAc was filtered through a plug of SiO₂ (eluent: EtOAc/hexane, 1/5). The filtrates were concentrated under reduced pressure. The crude product **16a** (6 mg, check by ¹H NMR) was used directly in the next step without other purifications.

Preparation of Compound S-8



To a stirred suspension of $\text{Ph}_3\text{PCH}_2\text{Br}$ (381 mg, 1.08 mmol, 4.0 equiv.) in 11 mL THF at 0 °C was added ${}^n\text{BuLi}$ (2.5 M in hexane, 0.4 mL, 1.0 mmol, 3.7 equiv. , dropwise). The reaction mixture was allowed to warm to room temperature. After stirring for 1 hour at room temperature, the solution was cooled to -78 °C and a solution of aldehyde **11** (56 mg, 0.13 mmol, 1.0 equiv. , dropwise) in 2.7 mL THF was added. The reaction mixture was allowed to warm to 0 °C and stirred for another 1 hour before quenching with sat. aq. NH_4Cl (10 mL). The organic phase was collected and the aqueous phase was extracted with Et_2O (3×10 mL). The combined organic phases were washed with brine, dried over Na_2SO_4 , concentrated under reduced pressure. The residue was purified through flash chromatography (eluent: $\text{EtOAc}/\text{hexane}$, 1/50) to afford compound **S-8** (35 mg, 63%) as a colourless oil.

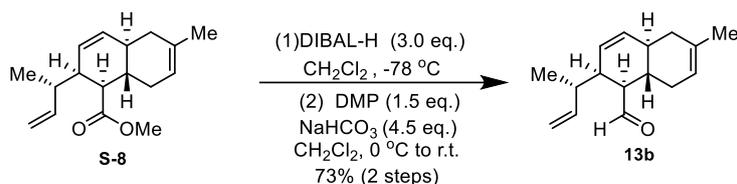
$[\alpha]_D^{25} = +271.6$ (c 1, CHCl_3);

${}^1\text{H NMR}$ (400 MHz, CDCl_3): δ 5.89 – 5.77 (m, 1H), 5.66 (d, $J = 10.2$ Hz, 1H), 5.62 – 5.51 (m, 1H), 5.39 – 5.32 (m, 1H), 5.03 – 4.88 (m, 2H), 3.68 (s, 3H), 2.70 – 2.60 (m, 2H), 2.43 – 2.33 (m, 1H), 2.20 – 2.12 (m, 1H), 2.07 – 1.90 (m, 2H), 1.79 (tdd, $J = 14.8, 8.2, 4.9$ Hz, 2H), 1.69 – 1.64 (m, 3H), 1.60 – 1.47 (m, 1H), 0.98 (d, $J = 6.8$ Hz, 3H).

${}^{13}\text{C NMR}$ (101 MHz, CDCl_3): δ 174.1, 143.6, 133.7, 132.5, 125.0, 120.9, 113.2, 51.1, 49.7, 41.2, 38.9, 37.4, 37.2, 33.8, 31.1, 23.3, 15.6.

HRMS (TOF, CI^+) m/z calculated for $[\text{C}_{17}\text{H}_{24}\text{O}_2+\text{H}]^+$ 261.1849, found 261.1855.

Preparation of Compound 13b



To a stirred solution of compound **S-8** (528 mg, 2.03 mmol, 1.0 equiv.) in 10 mL CH₂Cl₂ at -78 °C was added dropwise DIBAL-H (1.0 M in hexane, 6.1 mL, 6.10 mmol, 3.0 equiv.). The reaction mixture was stirred for 2 hours at -78 °C and then quenched with 2 N HCl (10 mL). The resulting mixture was stirred at room temperature for another 30 minutes. The organic phase was collected and the aqueous phase was extracted with CH₂Cl₂ (3 × 15 mL). The combined organic phases were washed with brine, dried over Na₂SO₄, concentrated under reduced pressure. The residue was dissolved in 10 mL CH₂Cl₂ cooled down to 0 °C and then NaHCO₃ (767 mg, 9.13 mmol, 4.5 equiv.) and Dess-Martin periodinane (1.29 g, 3.04 mmol, 1.5 equiv.) were added. The reaction mixture was stirred for another 1.5 hours at room temperature before quenching with sat. aq. NaHCO₃ (3 mL) and sat. aq. Na₂S₂O₃ (3 mL). The organic phase was collected and the aqueous phase was extracted with CH₂Cl₂ (3 × 15 mL). The combined organic phases were washed with brine, dried over Na₂SO₄, concentrated under reduced pressure. The residue was purified by flash column chromatography on silica gel (eluent: EtOAc/hexane, 1/60) to afford compound **13b** (341 mg, 73%) as a colourless oil.

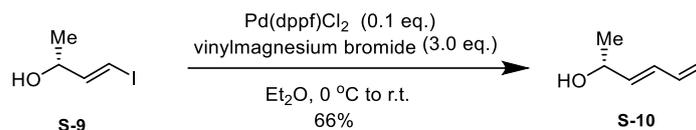
$[\alpha]_D^{25} = +41.80$ (*c* 1, CHCl₃);

¹H NMR (400 MHz, CDCl₃): δ 9.86 (d, *J* = 3.9 Hz, 1H), 5.86 – 5.71 (m, 1H), 5.70 – 5.59 (m, 2H), 5.37 (dd, *J* = 5.1, 2.6 Hz, 1H), 5.06 – 4.92 (m, 2H), 2.75 – 2.66 (m, 1H), 2.51 – 2.34 (m, 2H), 2.34 – 2.23 (m, 1H), 2.10 – 1.87 (m, 3H), 1.83 (d, *J* = 13.9 Hz, 1H), 1.67 (d, *J* = 2.4 Hz, 3H), 1.64 – 1.57 (m, 1H), 1.02 (d, *J* = 6.9 Hz, 3H).

¹³C NMR (101 MHz, CDCl₃): δ 205.6, 143.8, 134.2, 132.5, 126.0, 120.5, 114.2, 55.8, 41.9, 39.3, 37.5, 36.9, 32.8, 30.8, 23.4, 17.2.

HRMS (TOF, ES⁻) *m/z* calculated for [C₁₇H₂₄O₂-H]⁺ 229.1587, found 229.1601.

Preparation of Compound S-10



To a stirred solution of known vinyl iodide **S-9**⁵ (2.56 g, 12.9 mmol, 1.0 equiv.) in 65 mL Et₂O at 0 °C were added dropwise Pd(dppf)Cl₂ (946 mg, 0.13 mmol, 0.1 equiv.) and vinylmagnesium bromide (1.0 M in THF, 38.8 mL, 38.8 mmol, 3 equiv.). The resulting mixture was allowed to warm to room temperature over 15 hours before the reaction was diluted with Et₂O (50 mL) and quenched with H₂O (50 mL) at 0 °C. The organic phase was collected and the aqueous phase was extracted with Et₂O (3 × 50 mL). The combined organic phases were washed with brine, dried over Na₂SO₄, concentrated under reduced pressure. The residue was purified by flash column chromatography on silica gel (eluent: Et₂O/Hexane, 1/3) to afford compound **S-10** (836 mg, 66%) as a colourless oil.

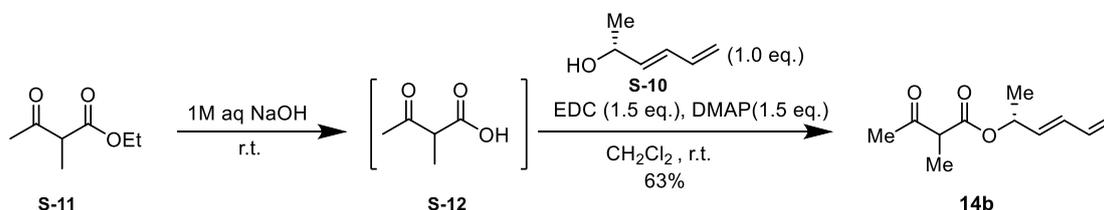
$[\alpha]_D^{25} = -6.00$ (*c* 0.35, CHCl₃);

¹H NMR (400 MHz, CDCl₃): δ 6.34 – 6.18 (m, 1H), 6.12 (dd, *J* = 15.2, 10.4 Hz, 1H), 5.73 – 5.62 (m, 1H), 5.29 – 4.98 (m, 2H), 4.31 – 4.20 (m, 1H), 1.20 (dd, *J* = 6.5, 2.0 Hz, 3H).

¹³C NMR (101 MHz, CDCl₃): δ 137.7, 136.3, 129.6, 129.6, 117.0, 67.8, 67.8, 22.9.

HRMS (TOF, CI⁺) *m/z* calculated for [C₆H₁₀O]⁺ 98.0732, found 98.0734.

Preparation of Compound 14b



The solution of ethyl-2-methylacetoacetate **S-11** (3.65 g, 25.3 mmol, 1 equiv.) in 35 mL 1 *N* NaOH was stirred vigorously at room temperature for 16 h. The reaction mixture was cooled to 0 °C and acidified with 1 *N* HCl until pH of 3-4. The reaction mixture was extracted with CH₂Cl₂ (3 × 50 mL). The combined organic phases were

washed with brine, dried over Na₂SO₄ and concentrated under reduced pressure at 10 °C. To the solution of the crude product **S-12** (2.36 g, 20.3 mmol, 1.5 equiv.) and alcohol **S-10** (1.33 g, 13.6 mmol, 1 equiv.) in 136 mL CH₂Cl₂ were added 1-ethyl-3-(3-dimethylaminopropyl)carbodiimide hydrochloride salt (3.89 g, 20.3 mmol, 1.5 equiv.) and 4-dimethylaminopyridine (2.48 g, 20.3 mmol, 1.5 equiv.). The reaction mixture was stirred for 18 hours before it was diluted with addition of CH₂Cl₂ (150 mL). The reaction solution was washed by sat. aq. NH₄Cl (3 × 50 mL). The organic phase was collected, washed with brine, dried over Na₂SO₄, concentrated under reduced pressure. The resulting residue was purified by flash column chromatography on silica gel (eluent: Et₂O/Hexane, 1/5) to afford compound **14b** (1.68 g, 63%, 1:1 mixture of diastereomers) as a colourless oil.

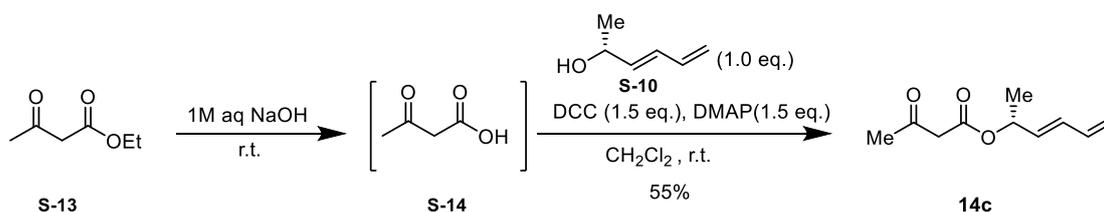
$[\alpha]_D^{25} = +20.70$ (*c* 1, CHCl₃);

¹H NMR (400 MHz, CDCl₃): δ 6.30 – 6.00 (m, 2H), 5.62 – 5.48 (m, 1H), 5.39 – 5.24 (m, 1H), 5.17 – 5.07 (m, 1H), 5.07 – 4.96 (m, 1H), 3.45 – 3.33 (m, 1H), 2.10 (dd, *J* = 5.8, 1.3 Hz, 3H), 1.35 – 1.11 (m, 6H).

¹³C NMR (101 MHz, CDCl₃): δ 203.0, 203.0, 169.4, 169.3, 135.6, 132.4, 132.3, 131.8, 131.7, 118.4, 71.2, 53.4, 53.4, 28.1, 28.0, 19.7, 19.6, 12.3, 12.3.

HRMS (TOF, ES⁺) *m/z* calculated for [C₁₁H₁₆O₃+Na]⁺ 219.0992, found 219.0992.

Preparation of Compound 14c



Following the same procedure for preparation of **14a**, compound **14c** (1.03 g, 55%) was obtained as a colourless oil from ethyl acetoacetate **S-13** (2.53 g, 19.4 mmol, 1 equiv.) and alcohol **S-10** (1.01 g, 10.3 mmol, 1 equiv.).

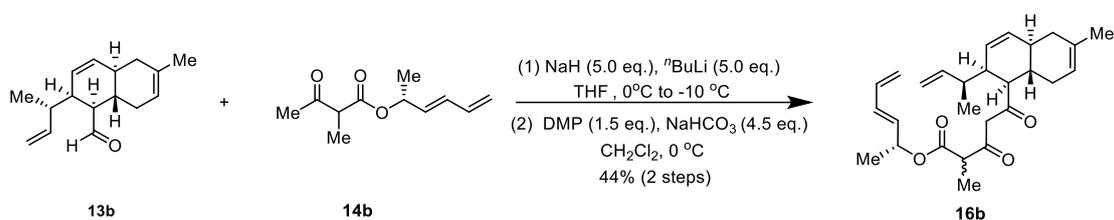
$[\alpha]_D^{25} = +44.90$ (*c* 1, CHCl₃);

¹H NMR (400 MHz, CDCl₃): δ 6.31 – 6.16 (m, 2H), 5.70 – 5.57 (m, 1H), 5.47 – 5.34 (m, 1H), 5.25 – 5.15 (m, 1H), 5.14 – 5.04 (m, 1H), 3.39 (s, 2H), 2.21 (s, 3H), 1.30 (dd, *J* = 6.5, 2.9 Hz, 3H).

¹³C NMR (101 MHz, CDCl₃): δ 200.4, 166.2, 135.7, 132.5, 131.9, 118.6, 71.6, 50.2, 30.0, 19.9.

HRMS (TOF, ES⁺) *m/z* calculated for [C₁₁H₁₆O₃+Na]⁺ 205.0835, found 205.0830.

Preparation of Compound 16b



Following the same procedure for preparation of compound **16a**, compound **16b** (45 mg, 44%) was obtained as a colourless oil from **14b** (234 mg, 1.19 mmol, 5.0 equiv.) and aldehyde **13b** (55 mg, 0.24 mmol, 1 equiv.).

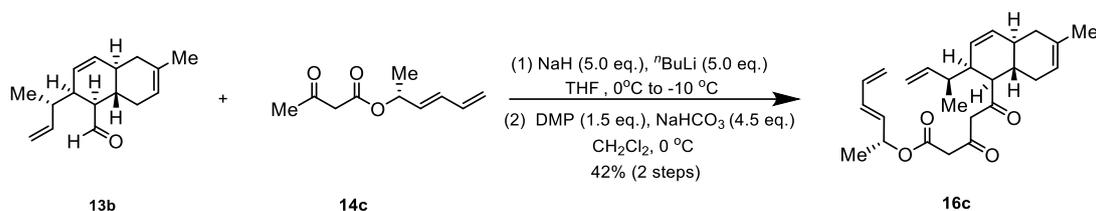
[α]_D²⁵ = +82.4 (*c* 1, CHCl₃);

¹H NMR (400 MHz, CDCl₃): δ 15.34 (brs, 1H), 6.36 – 6.17 (m, 2H), 5.87 – 5.73 (m, 1H), 5.70 – 5.60 (m, 3H), 5.59 – 5.52 (m, 1H), 5.50 – 5.37 (m, 1H), 5.37 – 5.31 (m, 1H), 5.30 – 5.21 (m, 1H), 5.21 – 5.06 (m, 1H), 5.00 – 4.87 (m, 2H), 3.43 – 3.32 (m, 1H), 2.55 – 2.45 (m, 2H), 2.31 (d, *J* = 17.8 Hz, 2H), 2.25 – 2.08 (m, 2H), 2.02 (d, *J* = 15.0 Hz, 1H), 2.00 – 1.93 (m, 1H), 1.83 (d, *J* = 4.2 Hz, 1H), 1.76 – 1.69 (m, 1H), 1.66 (s, 3H), 1.45 – 1.37 (m, 3H), 1.32 (d, *J* = 6.8 Hz, 3H), 1.00 – 0.94 (m, 3H).

¹³C NMR (101 MHz, CDCl₃): δ 193.6, 193.5, 192.3, 192.0, 170.1, 170.0, 143.9, 135.9, 133.8, 133.8, 132.6, 132.5, 132.5, 132.5, 132.1, 132.1, 125.2, 125.1, 120.9, 120.9, 118.7, 118.7, 113.1, 100.0, 99.9, 77.3, 77.0, 76.7, 71.6, 71.5, 52.1, 49.6, 49.5, 43.2, 37.9, 37.9, 37.7, 37.3, 33.6, 33.5, 31.1, 23.3, 20.1, 19.9, 15.5, 15.4, 13.9, 13.8.

HRMS (TOF, ES⁺) *m/z* calculated for [C₂₇H₃₆O₄+Na]⁺ 447.2506, found 447.2509.

Preparation of Compound 16c



Following the same procedure for preparation of compound **16a**, compound **16c** (32 mg, 42%) was obtained as a colourless oil from **14c** (172 mg, 0.94 mmol, 5.0 equiv.) and aldehyde **13b** (43 mg, 0.19 mmol, 1 equiv.).

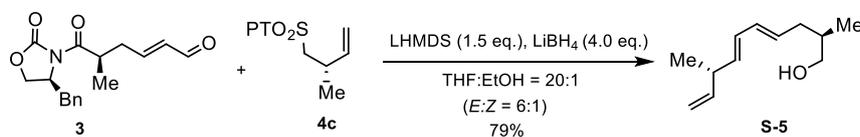
$[\alpha]_D^{25} = +48.30$ (*c* 1, CHCl₃);

¹H NMR (400 MHz, CDCl₃): δ 15.35 (brs, 1H), 6.37 – 6.20 (m, 2H), 5.82 (ddd, *J* = 16.8, 10.6, 6.1 Hz, 1H), 5.71 – 5.64 (m, 3H), 5.60 – 5.53 (m, 1H), 5.45 (p, *J* = 6.7 Hz, 1H), 5.35 (s, 1H), 5.32 – 5.21 (m, 1H), 5.15 (ddd, *J* = 8.8, 4.9, 1.7 Hz, 1H), 4.98 – 4.89 (m, 2H), 3.35 (d, *J* = 5.1 Hz, 2H), 2.57 – 2.47 (m, 2H), 2.35 (s, 1H), 2.30 (s, 1H), 2.07 – 1.90 (m, 3H), 1.90 – 1.77 (m, 2H), 1.35 (t, *J* = 6.7 Hz, 3H), 1.07 – 0.94 (m, 4H).

¹³C NMR (101 MHz, CDCl₃): δ 194.5, 186.9, 166.8, 143.9, 135.9, 133.8, 132.7, 132.5, 132.0, 125.1, 120.9, 118.8, 113.1, 101.6, 71.8, 52.4, 45.4, 43.1, 37.9, 37.7, 37.3, 33.6, 31.1, 23.3, 20.1, 15.4.

HRMS (TOF, ES⁺) *m/z* calculated for [C₂₆H₃₄O₄+Na]⁺ 433.2349, found 433.2353.

Preparation of Compound S-5



To a solution of aldehyde **3** (2.58 g, 8.6 mmol, 1.0 equiv.) and sulfone **4c**⁶ (2.86 g, 10.3 mmol, 1.2 equiv.) in 43 mL THF at -78 °C was added dropwise LHMDS (1.0 M in THF, 11.2 mL, 11.2 mmol, 1.3 equiv.). The reaction mixture was stirred at -78 °C for 2 hours and then 2.2 mL ethanol was added. The reaction mixture was allowed to warm up to -10 °C and LiBH₄ (2.0 M in THF, 34.3 mL, 34.3 mmol, 4.0 equiv.) was added dropwise into the reaction mixture. After the reduction was completed as

indicated by TLC (~2 hours), the reaction mixture was allowed to warm up to 0 °C and then quenched with 1 N NaOH (30 mL). The organic phase was collected and the aqueous phase was extracted with EtOAc (3 × 50 mL). The combined organic phases were washed with brine, dried over Na₂SO₄ and concentrated *in vacuo*. The crude residue was purified by flash column chromatography on silica gel (eluent: EtOAc/hexane, 1/20) to give compound **S-5** (1.22 g, 79%) as a light yellow oil.

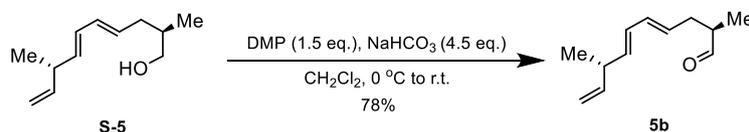
$[\alpha]_D^{25} = +8.20$ (*c* 2, CHCl₃);

¹H NMR (400 MHz, CDCl₃): δ 6.09 – 5.89 (m, 2H), 5.85 – 5.70 (m, 1H), 5.67 – 5.47 (m, 2H), 4.98 (t, *J* = 15.1 Hz, 2H), 3.52 – 3.37 (m, 2H), 2.95 – 2.82 (m, 1H), 2.39 – 2.08 (m, 2H), 1.92 (dtt, *J* = 14.3, 7.4, 3.0 Hz, 1H), 1.77 – 1.65 (m, 1H), 1.14 – 1.05 (m, 3H), 0.94 – 0.86 (m, 3H).

¹³C NMR (101 MHz, CDCl₃): δ 142.4, 135.8, 131.8, 130.7, 128.9, 112.9, 67.6, 40.1, 36.4, 35.9, 19.6, 16.4.

HRMS (TOF, CI⁺) *m/z* calculated for [C₁₂H₂₀O]⁺ 180.1514, found 180.1517.

Preparation of Compound **5b**



To a solution of alcohol **S-5** (1.22 g, 6.8 mmol, 1.0 equiv.) in 34 mL CH₂Cl₂ at 0 °C were added NaHCO₃ (2.56 g, 30.5 mmol, 4.5 equiv.) and Dess-Martin periodinane (4.31 g, 10.2 mmol, 1.5 equiv.). The reaction mixture was stirred at 0 °C for 2 hours and then quenched by addition of sat. aq. NaHCO₃ (20 mL) and sat. aq. Na₂S₂O₃ (20 mL). The organic phase was collected and the aqueous phase was extracted with CH₂Cl₂ (3 × 30 mL). The combined organic phases were washed with brine, dried over Na₂SO₄, concentrated under reduced pressure. The residue was purified by flash column chromatography on silica gel (eluent: EtOAc/hexane, 1/50) to afford aldehyde **5b** (941 mg, 79%) as a pale-yellow oil.

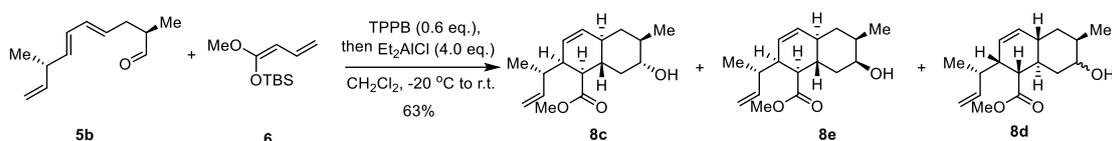
$[\alpha]_D^{25} = +0.20$ (*c* 1, CHCl₃);

¹H NMR (400 MHz, CDCl₃): δ 9.61 (d, *J* = 1.3 Hz, 1H), 6.09 – 5.85 (m, 2H), 5.81 – 5.67 (m, 1H), 5.66 – 5.44 (m, 2H), 5.01 – 4.87 (m, 2H), 2.91 – 2.78 (m, 1H), 2.54 – 2.33 (m, 2H), 2.23 – 2.06 (m, 1H), 1.07 (dd, *J* = 6.9, 2.6 Hz, 6H).

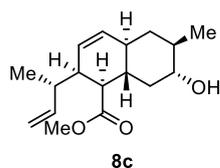
¹³C NMR (101 MHz, CDCl₃): δ 204.4, 142.2, 136.7, 132.8, 128.4, 128.3, 113.0, 46.1, 40.1, 33.5, 19.5, 13.0.

HRMS (TOF, CI⁺) *m/z* calculated for [C₁₂H₁₈O]⁺ 178.1358, found 178.1356.

Preparation of Compound **8c** and **8e**



To a solution of aldehyde **5b** (1.16 g, 6.51 mmol, 1.0 equiv.) and compound **6** (2.79 g, 13.0 mmol, 2.0 equiv.) in 65 mL CH₂Cl₂ at -20 °C was added tris(pentafluorophenyl)-borane (2.00 g, 3.91 mmol, 0.6 equiv.). The reaction mixture was stirred at -20 °C for 30 minutes before Et₂AlCl (2.0 M in hexane, 13.0 mL, 26.0 mmol, 4.0 equiv.) was added dropwise. The reaction mixture was allowed to warm to room temperature and stirred for 13 hours before 1.0 mL H₂O and 8.0 mL concentrated HCl was added at 0 °C. The reaction mixture was stirred for another 10 hours before quenching by addition of 2 *N* NaOH (50 mL). The organic phase was collected and the aqueous phase was extracted by CH₂Cl₂ (3 × 50 mL). The combined organic phases were washed with brine, dried over Na₂SO₄, concentrated under reduced pressure. The residue was purified by flash column chromatography on silica gel (eluent: EtOAc/hexane, 1/20) to afford **8c** (998 mg, 55%) as a colourless oil (mixed with **8d** inseparable, **8c**:**8d** > 3:1) and **8e** (145 mg, 8%) as a colourless oil.



[α]_D²⁵ = +26.0 (*c* 0.1, CHCl₃);

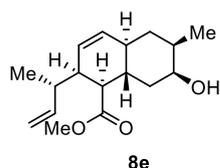
¹H NMR (400 MHz, CDCl₃): δ 5.89 – 5.77 (m, 1H), 5.63 – 5.56 (m, 1H), 5.56 – 5.47 (m, 1H), 5.04 – 4.89 (m, 2H), 3.69 (d, *J* = 3.0

Hz, 3H), 3.34 – 3.22 (m, 1H), 2.74 – 2.64 (m, 1H), 2.64 – 2.50 (m, 1H), 2.29 – 2.19 (m, 1H), 2.19 – 2.05 (m, 1H), 1.95 – 1.80 (m, 1H), 1.80 – 1.65 (m, 2H), 1.65 – 1.51

(m, 1H), 1.51 – 1.37 (m, 1H), 1.05 (dd, $J = 6.8, 3.2$ Hz, 3H), 0.97 (dd, $J = 7.1, 1.1$ Hz, 3H).

^{13}C NMR (101 MHz, CDCl_3): δ 173.8, 143.5, 132.2, 125.3, 113.4, 76.1, 51.2, 49.4, 40.8, 40.7, 40.1, 39.4, 39.2, 38.8, 36.4, 18.3, 15.3.

HRMS (TOF, ES^+) m/z calculated for $[\text{C}_{17}\text{H}_{26}\text{O}_3+\text{Na}]^+$ 301.1774, found 301.1772.



$[\alpha]_{\text{D}}^{25} = -28.00$ (c 0.25, CHCl_3);

^1H NMR (400 MHz, CDCl_3): δ 5.86 – 5.67 (m, 1H), 5.61 – 5.53 (m, 1H), 5.51 – 5.41 (m, 1H), 5.00 – 4.87 (m, 2H), 3.89 – 3.79 (m,

1H), 3.66 (d, $J = 3.7$ Hz, 3H), 2.66 – 2.51 (m, 2H), 2.18 – 2.06 (m, 2H), 1.95 – 1.86 (m, 1H), 1.73 – 1.64 (m, 1H), 1.64 – 1.54 (m, 1H), 1.51 – 1.41 (m, 1H), 1.30 – 1.21 (m, 1H), 1.13 – 1.06 (m, 1H), 1.05 – 1.01 (m, 1H), 0.99 – 0.94 (m, 6H).

^{13}C NMR (101 MHz, CDCl_3): δ 174.0, 143.6, 133.0, 125.0, 113.2, 70.5, 51.1, 49.5, 41.3, 41.1, 38.8, 38.0, 36.9, 34.6, 31.1, 18.0, 15.5.

HRMS (TOF, ES^+) m/z calculated for $[\text{C}_{17}\text{H}_{26}\text{O}_3+\text{Na}]^+$ 301.1774, found 301.1775.

Preparation of Compound 13b



To a solution of **8c/8e** (52 mg, 0.19 mmol, 1.0 equiv.) in 9.4 mL toluene was added Burgess reagent (178 mg, 0.75 mmol, 4.0 equiv.). The reaction mixture was heated to 85 °C for 2 hours. It was cooled to -78 °C and DIBAL-H (1.0 M in hexane, 1.5 mL, 1.5 mmol, 8.0 equiv.) was added dropwise. After stirring for 2 hours, the reaction was quenched by addition of 1 *N* HCl (10 mL). The organic phase was collected and the aqueous phase was extracted by EtOAc (3 × 15 mL). The combined organic phases were washed with brine, dried over Na_2SO_4 , concentrated under reduced pressure. The residue in 1.9 mL CH_2Cl_2 was cooled to 0 °C and NaHCO_3 (189 mg, 2.25 mmol, 12.0 equiv.) and Dess-Martin periodinane (359 mg, 0.85 mmol, 4.5

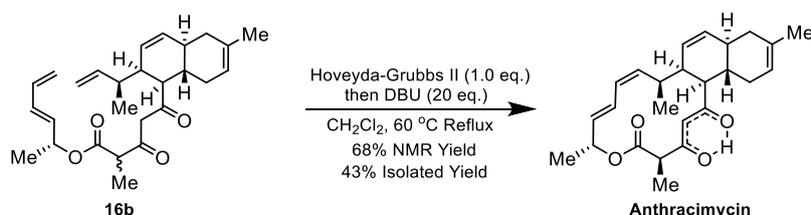
equiv.) were added. The reaction mixture was allowed to warm to room temperature and stirred for 1.5 hours before quenching with sat. aq. NaHCO₃ (1 mL) and sat. aq. Na₂S₂O₃ (2 mL). The organic phase was collected and the aqueous phase was extracted with CH₂Cl₂ (3 × 3 mL). The combined organic phases were washed with brine, dried over Na₂SO₄, concentrated under reduced pressure. The residue was purified by flash column chromatography on silica gel (eluent: EtOAc/hexane, 1/50) to afford aldehyde **13b** (31 mg, 65%) as a colourless oil.

¹H NMR (400 MHz, CDCl₃): δ 9.94 – 9.81 (m, 1H), 5.83 – 5.70 (m, 1H), 5.66 – 5.63 (m, 1H), 5.40 – 5.31 (m, 1H), 5.06 – 4.91 (m, 2H), 2.74 – 2.62 (m, 1H), 2.51 – 2.37 (m, 2H), 2.37 – 2.23 (m, 1H), 2.08 – 1.88 (m, 3H), 1.88 – 1.77 (m, 1H), 1.67 (dd, *J* = 2.6, 1.4 Hz, 3H), 1.64 – 1.55 (m, 1H), 1.07 (dd, *J* = 6.9, 0.8 Hz, 1H), 1.01 (dd, *J* = 7.0, 2.9 Hz, 3H).

¹³C NMR (101 MHz, CDCl₃): δ 205.5, 143.7, 134.1, 132.4, 126.0, 120.4, 114.2, 55.8, 41.8, 39.2, 37.5, 36.9, 32.7, 30.7, 23.4, 17.1.

HRMS (TOF, ES⁻) *m/z* calculated for [C₁₇H₂₄O₂-H]⁺ 229.1587, found 229.1601.

Preparation of Anthracimycin



The solution of compound **16b** (45 mg, 0.11 mmol, 1.0 equiv.) in 21 mL CH₂Cl₂ was degassed with argon sparging for 5 minutes and the Hoveyda Grubbs II catalyst (67 mg, 0.11 mmol, 1.0 equiv.) was added. The reaction mixture was heated to reflux for 3 hours and then cooled to room temperature. 1,8-Diazabicyclo[5.4.0]undec-7-ene (DBU) (0.32 mL, 2.14 mmol, 20.0 equiv.) was added to the reaction mixture, which was stirred at room temperature for another 30 minutes. The solvent was removed under reduced pressure to provide the crude anthracimycin with 68% yield based on ¹H NMR analysis using 1,3,5-trimethoxybenzene (6 mg, 0.036 mmol, 0.33 equiv.) as the internal standard. The crude anthracimycin was purified by column

chromatography on reversed-phase silica gel (water/acetonitrile, 10:1 to 0:1) to afford anthracimycin (18 mg, 43%) as a light brown solid. For biological testing, the sample was further purified by the reverse-phase HPLC (Phenomenex Luna ® C18 column, 100 Å, 250 x 4.6 mm, 5µm) using a gradient of H₂O: MeCN = 10:90 over 1 hour, the flow rate was 1 mL/min, with UV detection under 280nm wavelength (Waters 2998 Photodiode Array Detector; Milford, USA). Pure anthracimycin (11 mg) was collected at 44.0 min as a white solid. Total 25 mg of anthracimycin was obtained from two batches. A single crystal suitable for X-Ray diffraction analysis was obtained (see page 36) with CCDC deposition number 2194803.

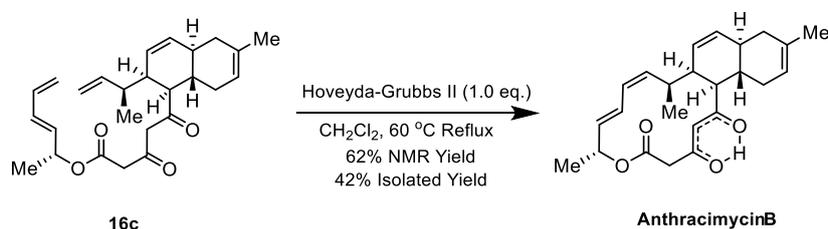
$[\alpha]_D^{25} = -144.00$ (*c* 1, CHCl₃);

¹H NMR (400 MHz, CDCl₃): δ 15.45 (s, 1H), 6.53 – 6.38 (m, 1H), 5.96 (s, 1H), 5.87 (t, *J* = 11.1 Hz, 1H), 5.72 (d, *J* = 10.2 Hz, 1H), 5.60 – 5.51 (m, 3H), 5.46 – 5.32 (m, 2H), 3.53 (q, *J* = 6.9 Hz, 1H), 2.75 – 2.49 (m, 3H), 2.39 (d, *J* = 17.4 Hz, 1H), 2.11 – 1.88 (m, 3H), 1.81 (d, *J* = 7.1 Hz, 1H), 1.67 (s, 3H), 1.57 – 1.49 (m, 1H), 1.45 – 1.37 (m, 3H), 1.34 (d, *J* = 6.7 Hz, 3H), 0.95 (d, *J* = 6.6 Hz, 3H).

¹³C NMR (101 MHz, CDCl₃): δ 194.2, 190.8, 168.9, 139.1, 134.0, 133.0, 131.7, 126.1, 124.9, 123.8, 121.0, 103.0, 70.0, 52.7, 49.2, 46.0, 37.5, 37.4, 33.0, 32.8, 31.4, 23.5, 21.0, 16.4, 11.8.

HRMS (TOF, ES⁺) *m/z* calculated for [C₂₅H₃₂O₄+Na]⁺ 419.2193, found 419.2197.

Preparation of Anthracimycin B



Following the same procedure for preparation of anthracimycin, anthracimycin B (7 mg, 42% yield; 62% NMR yield) was obtained from compound **16c** (18 mg, 0.04 mmol, 1.0 equiv.) and Hoveyda Grubbs II catalyst (28 mg, 0.04 mmol, 1.0 equiv.) as a light green solid. Anthracimycin B was further purified by reverse-phase HPLC (Phenomenex Luna ® C18 column, 100 Å, 250 x 4.6 mm, 5µm) using a gradient of

H₂O: MeCN =10:90 over 1 hour, the flow rate was 1 mL/min, with UV detection under 280nm wavelength (Waters 2998 Photodiode Array Detector; Milford, USA). Anthracimycin B (4.5 mg) as a white solid was collected at 45.7 min. Total 14 mg of anthracimycin B was obtained from two batches.

$[\alpha]_D^{25} = -169.00$ (*c* 0.45, CHCl₃);

¹H NMR (400 MHz, CDCl₃): δ 15.37 (s, 1H), 6.50 – 6.47 (m, 1H), 5.96 (s, 1H), 5.90 (t, *J* = 10.7 Hz, 1H), 5.73 (d, *J* = 10.0 Hz, 1H), 5.57 – 5.54 (m, 3H), 5.42 – 5.37 (m, 2H), 3.50 (d, *J* = 11.5 Hz, 1H), 3.22 (d, *J* = 11.4 Hz 1H), 2.63 – 2.61 (m, 3H), 2.43 (d, *J* = 17.4 Hz, 1H), 2.05 – 2.02 (m, 2H), 1.99 – 1.90 (m, 1H), 1.87 – 1.74 (m, 1H), 1.67 (s, 3H), 1.51 (d, *J* = 24.0 Hz, 1H), 1.35 (d, *J* = 6.6 Hz, 3H), 0.95 (d, *J* = 6.6 Hz, 3H).

¹³C NMR (101 MHz, CDCl₃): δ 196.5, 184.8, 165.4, 138.9, 133.8, 133.0, 131.3, 125.9, 124.6, 123.5, 120.9, 102.8, 70.0, 53.1, 46.5, 45.5, 37.3, 37.3, 32.8, 32.7, 31.2, , 23.4, 20.7, 16.2.

HRMS (TOF, ES⁺) *m/z* calculated for [C₂₄H₃₀O₄+Na]⁺ 405.2036, found 405.2040.

NMR comparison of natural and synthetic anthracimycin and anthracimycin B

Table S1 ¹H NMR comparison of natural and synthetic Anthracimycin

Natural sample ⁷ δ _H (500 MHz, CDCl ₃)	Brimble's synthetic sample ² δ _H (500 MHz, CDCl ₃)	Our synthetic sample δ _H (400 MHz, CDCl ₃)
Not observed	15.45 (brs)	15.45 (brs)
6.45 (dd, 15.2, 11.0)	6.46 (ddd, 15.4, 11.6, 1.8)	6.46 (m)
5.96 (s)	5.96 (s)	5.96 (s)
5.87 (dd, 11.0, 10.5)	5.87 (app. t, 10.9)	5.87 (t, 11.1)
5.71 (d, 10.0)	5.72 (d, 10.4)	5.72 (d, 10.2)
5.57 (dq, 6.5, 2.4)	5.58-5.56 (m) ^b	5.60 – 5.51 (m) ^b
5.56 (dd, 15.2, 2.4)	5.58-5.56 (m) ^b	5.60 – 5.51 (m) ^b
5.53 (dd, 10.0, 5.0)	5.54-5.52 (m)	5.60 – 5.51 (m) ^b
5.40 (dd, 10.5, 9.8)	5.40 (app. t, 9.4)	5.48 – 5.32 (m)
5.36 (brd, 4.5)	5.36 (brd, 4.6)	5.48 – 5.32 (m)
3.53 (q, 7.0)	3.53 (q, 7.0)	3.53 (q, 6.9)
2.64 ^a	2.66-2.57 (m) ^b	2.75 – 2.49 (m) ^b
2.60 ^a	2.66-2.57 (m) ^b	2.75 – 2.49 (m) ^b
2.58 (11.8, 6.7)	2.66-2.57 (m) ^b	2.75 – 2.49 (m) ^b
2.39 (ddd, 16.0, 4.5, 4.5)	2.39 (brd, 18.6)	2.39 (d, 17.4)
2.02 (dd, 16.5, 4.0)	2.03 (brd, 5.3)	2.11 – 1.88 (m) ^b
1.98 (ddd, 14.5, 12.5, 11.8)	2.00-1.97 (m)	2.11 – 1.88 (m) ^b
1.93 (m)	1.93 (dd, 10.6, 4.2)	2.11 – 1.88 (m) ^b
1.82 (dd, 16.5, 10.3)	1.85-1.81 (m)	1.81 (d, 7.1)
1.67 (s)	1.67 (s)	1.67 (s)
1.52(ddd, 16.0, 10.5, 4.5)	Not observed ^c	1.57 – 1.49 (m)
1.39 (d, 7.0)	1.40 (d, 6.8)	1.45 – 1.37 (m)
1.33 (d, 6.5)	1.34 (d, 6.8)	1.34 (d, 6.7)
0.94 (d, 7.0)	0.95 (d, 6.6)	0.95 (d, 6.6)

^aThe coupling constant was not measured because of overlapping signals

^bUnresolved signals

^c Not observed due to overlapping water signal.

Table S2 ^{13}C NMR comparison of natural and synthetic Anthracimycin

Natural sample ⁷ δ_c (125 MHz, CDCl_3)	Brimble's synthetic sample ² δ_c (125 MHz, CDCl_3)	Our synthetic sample δ_c (101 MHz, CDCl_3^a)
194.1	194.2	194.2
190.9	190.8	190.8
168.9	168.9	168.9
139.1	139.1	139.1
134.0	134.0	134.0
133.0	133.0	133.0
131.7	131.7	131.7
126.1	126.1	126.1
124.9	124.9	124.9
123.7	123.8	123.8
121.0	121.0	121.0
103.0	103.0	103.0
70.0	70.0	70.0
52.6	52.7	52.7
49.2	49.2	49.2
46.0	46.0	46.0
37.5	37.5	37.5
37.4	37.4	37.4
33.0	33.0	33.0
32.8	32.8	32.8
31.4	31.4	31.4
23.5	23.5	23.5
21.0	21.0	21.0
16.4	16.4	16.4

^a Chemical shifts are reported in parts per million (ppm) as values relative to the internal chloroform 77.16 ppm for ^{13}C

Table S3 ¹H NMR comparison of natural and synthetic Anthracimycin B

Natural sample ⁸ δ_{H} (500 MHz, CDCl ₃)	Our synthetic sample δ_{H} (400 MHz, CDCl ₃)
15.35 (brs)	15.37 (brs)
6.48 (dd, 13.0, 12.7)	6.50 – 6.47 (m)
5.96 (brs)	5.96 (s)
5.88 (dd, 10.8, 10.8)	5.90H(t, 10.7)
5.73 (d, 10.0)	5.73 (d, 10.0)
5.57 (m)	5.57 – 5.54 (m) ^a
5.55 (m)	5.57 – 5.54 (m) ^a
5.40 (m)	5.42 – 5.37 (m) ^a
5.37 (m)	5.42 – 5.37 (m) ^a
3.50 (d, 11.3)	3.50 (d, 11.5)
3.22 (d, 11.3)	3.22 (d, 11.4)
2.65 (m)	2.63-2.61 (m) ^a
2.64 (m)	2.63-2.61 (m) ^a
2.63 (m)	2.63-2.61 (m) ^a
2.42 (brd, 15.4)	2.43 (d, 17.4) ^a
2.05 (m)	2.05-2.02 (m) ^a
1.99 (m)	1.99-1.90 (m) ^a
1.96 (m)	1.96-1.93 (m) ^a
1.82 (brd, 16.9)	1.87-1.74 (m) ^a
1.68 (s)	1.67 (s)
1.51 (m)	1.51 (m)
1.35 (d, 6.7)	1.35 (d, 6.6)
0.95 (d, 6.0)	0.95 (d, 6.6)

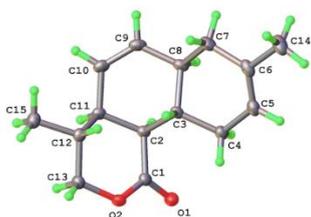
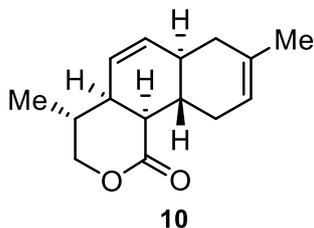
^aUnresolved signals

Table S4 ^{13}C NMR comparison of natural and synthetic Anthracimycin

Natural sample ⁸ δ_{C} (125 MHz, CDCl_3)	Our synthetic sample δ_{C} (101 MHz, CDCl_3 ^a)
196.5	196.5
184.7	184.8
165.5	165.4
139.0	138.9
133.8	133.8
133.0	133.0
131.3	131.3
125.9	125.9
124.6	124.6
123.6	123.5
120.9	120.9
102.8	102.8
70.0	70.0
53.1	53.1
46.5	46.5
45.5	45.5
37.3	37.3
37.3	32.3
32.9	32.8
32.7	32.7
31.2	31.2
23.4	23.4
20.7	20.7
16.3	16.2

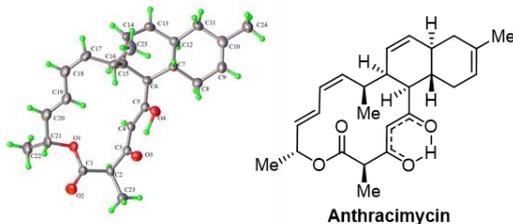
^a Chemical shifts are reported in parts per million (ppm) as values relative to the internal chloroform 77.00 ppm for ^{13}C

X-Ray Crystal Structure of compound 10



(Probability ellipsoid level: 40%;
CCDC Deposition Number 2175241)

Compound	ptian3CuLT
Formula	C ₁₅ H ₂₀ O ₂
<i>D</i> _{calc.} / g cm ⁻³	1.204
<i>m</i> /mm ⁻¹	0.615
Formula Weight	232.31
Colour	colourless
Shape	needle
Size/mm ³	0.40×0.08×0.07
<i>T</i> /K	99.97(10)
Crystal System	orthorhombic
Flack Parameter	0.12(9)
Hoofit Parameter	0.12(9)
Space Group	<i>P</i> 2 ₁ 2 ₁ 2 ₁
<i>a</i> /Å	5.60939(10)
<i>b</i> /Å	10.26643(19)
<i>c</i> /Å	22.2551(4)
<i>a</i> ^o	90
<i>b</i> ^o	90
<i>g</i> ^o	90
<i>V</i> /Å ³	1281.64(4)
<i>Z</i>	4
<i>Z</i> '	1
Wavelength/Å	1.54184
Radiation type	CuK _α
<i>Q</i> _{min} ^o	3.973
<i>Q</i> _{max} ^o	76.640
Measured Refl.	7631
Independent Refl.	2639
Reflections with <i>I</i> > 2(<i>I</i>)	2612
<i>R</i> _{int}	0.0230
Parameters	156
Restraints	0
Largest Peak	0.426
Deepest Hole	-0.170
GooF	1.057
<i>wR</i> ₂ (all data)	0.0912
<i>wR</i> ₂	0.0910
<i>R</i> ₁ (all data)	0.0351
<i>R</i> ₁	0.0348



(Probability ellipsoid level: 50%; CCDC
Deposition Number 2194803)

Crystal data and structure refinement	
Identification code	pre_Dr. WANG_s sample_auto
Empirical formula	C ₂₅ H ₃₂ O ₄
Formula weight	396.50
Temperature/K	100.0
Crystal system	orthorhombic
Space group	P2 ₁ 2 ₁ 2 ₁
a/Å	5.5419(2)
b/Å	15.3789(5)
c/Å	25.7913(8)
α/°	90
β/°	90
γ/°	90
Volume/Å ³	2198.15(13)
Z	4
ρ _{calc} /cm ³	1.198
μ/mm ⁻¹	0.634
F(000)	856.0
Crystal size/mm ³	0.12 × 0.1 × 0.05
Radiation	Cu Kα (λ = 1.54184)
2θ range for data collection/°	8.95 to 155.176
Index ranges	-6 ≤ h ≤ 6, -18 ≤ k ≤ 9, -31 ≤ l ≤ 31
Reflections collected	7767
Independent reflections	3739 [R _{int} = 0.0906, R _{sigma} = 0.0855]
Data/restraints/parameters	3739/0/270
Goodness-of-fit on F ²	1.063
Final R indexes [I ≥ 2σ (I)]	R ₁ = 0.0709, wR ₂ = 0.1844
Final R indexes [all data]	R ₁ = 0.0797, wR ₂ = 0.1917
Largest diff. peak/hole / e Å ⁻³	0.42/-0.38
Flack parameter	-0.1(3)

An issue was encountered during data collection for this crystal because the crystal specimen was decaying fast.

Antibacterial assays

The antibacterial activity of anthracimycin and anthracimycin B was examined with several pathogenic strains including gram-negative strains--*A. baumannii* B-65371, *E. cloacae* NRRL-B-425, *E. coli* k12, *K. pneumoniae* NRRL-B-408, and gram-positive strains-- MRSA ATCC 43300, MRSA ATCC 29213, MRSA Sa115, *S. aureus* ATCC 25923, *S. aureus* R22952, *B. subtilis* zk31 and *Micrococcus luteus* ATCC 10040. The minimal inhibition concentration (MIC) was determined using the broth microdilution method according to CLSI guidelines. The tested bacteria were overcultured and inoculated into 1×10^5 CFU/mL in MHB media; samples were injected into 96-well plates with different concentrations. Plates were incubated at 37 °C for 24 hours.

The minimum biofilm inhibitory concentration (MBIC) was measured by MTT (3-(4,5-Dimethylthiazol-2-yl)-2,5-diphenyltetrazolium bromide) staining assays. Similar to cytotoxicity tests, MTT could be reduced from bright yellow to blueviolet formazan. The color intensity was correlated to the number of live cells in the remaining biofilm, which dissolves in DMSO and can be quantified with 550 nm wavelength. The overnight cultured MRSA ATCC 29213 was diluted into 1×10^7 CFU/mL in LB broth with 0.5% glucose. Compounds with different concentration gradient were injected in to the diluted cultures in 96-well plates and incubated for 24 hours at 37 °C. Then each well was rinsed with 1X PBS in order to remove the planktonic and non-adhering cells. Inject 20 uL of MTT(5mg/mL) and 80 uL of 1X PBS to each well and keep incubation for another 2 hours, 100 uL DMSO was then added to dissolve the formazan. MBIC was determined as the minimum concentration with no violet color development.

Vancomycin

Anthracimycin

Anthracimycin B.

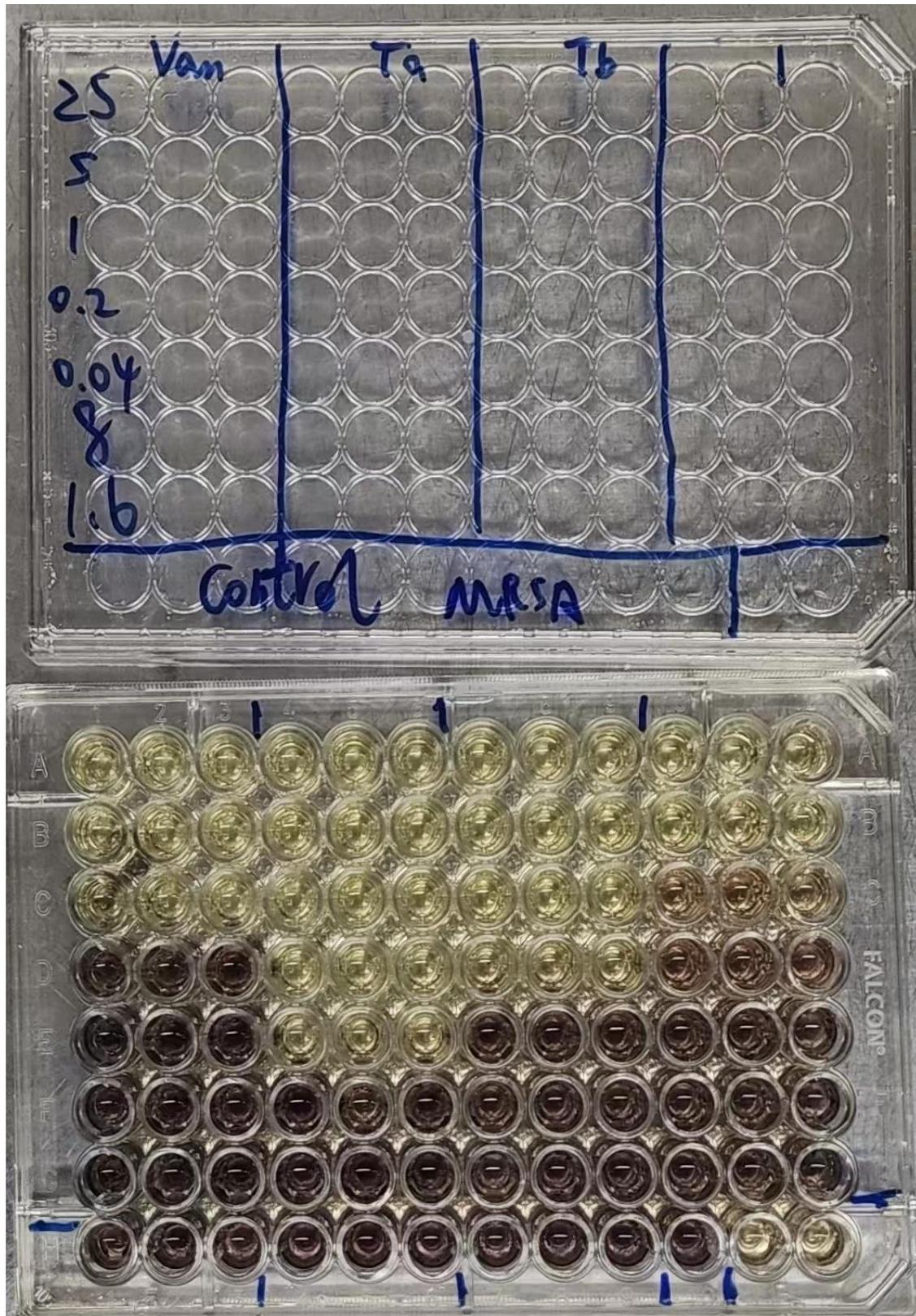


Figure S1. Inhibition effect of anthracimycins on biofilm formation

Cytotoxicity assays

Human epidermal keratinocyte cell line HaCaT cells were used in the MTT assays to evaluate samples' cytotoxicities. Cells were grown in DMEM with 10% fetal bovine serum and 1% penicillin and streptomycin at 37 °C with 5% CO₂. Each well of the 96-well plates was seeded with 5x10³ cells and incubated for 1 day. Then injection of compounds into each well achieved different concentrations and cultured for another 24 hours. After injecting 20uL of 5mg/mL MTT into each well for 3 hours at 37 °C, the culture liquid was removed and 100 uL DMSO was added to dissolve formazan MTT. The absorbance was measured at 570 nm wavelength using the MultiskanTM FC microplate photometer, and the IC₅₀ data was analyzed using GraphPad Prism software.

Table S5 Antimicrobial and cytotoxic activities of compounds

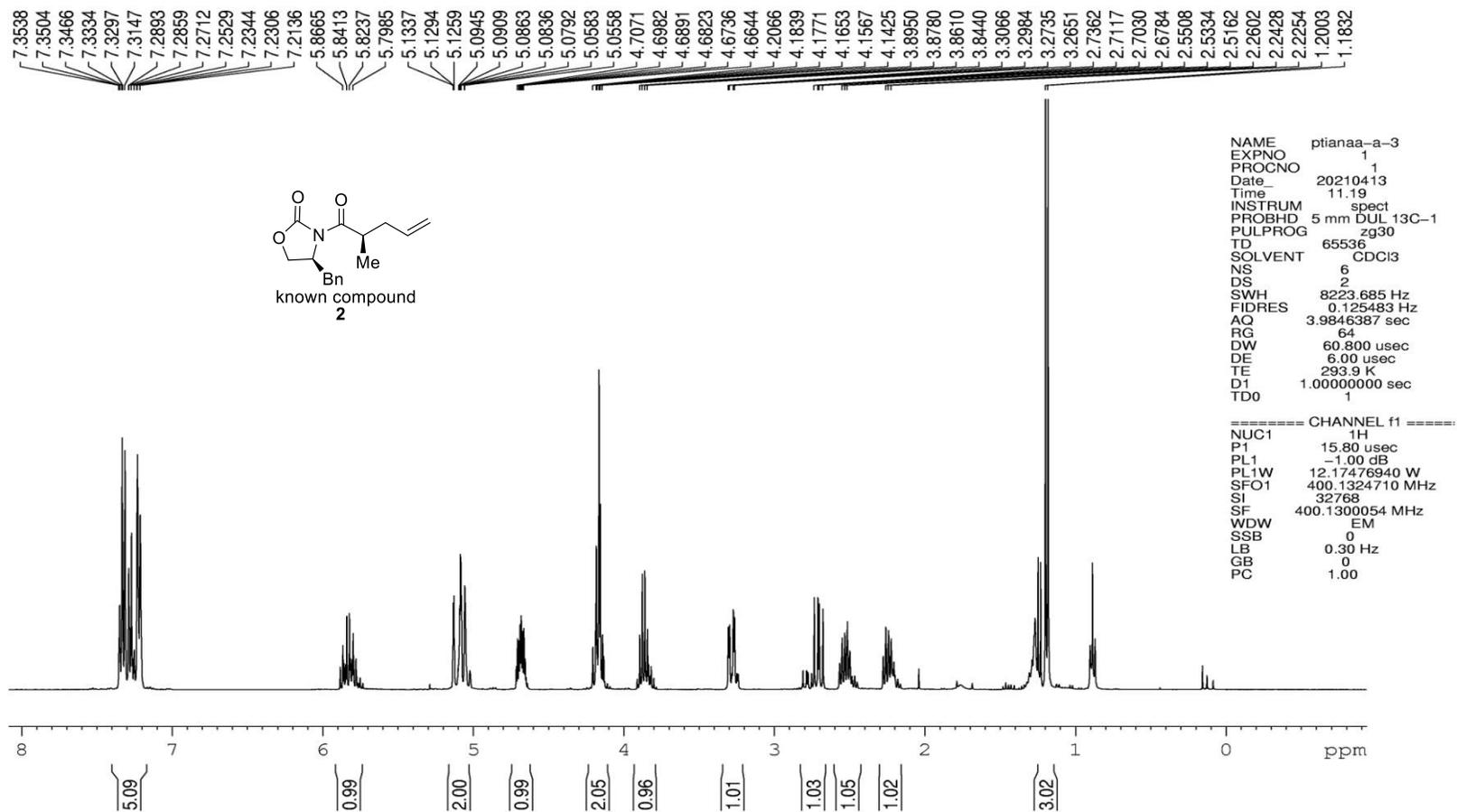
Strains/Cells	Anthracimycin (µg/mL)	Anthracimycin B (µg/mL)	vancomycin (µg/mL)
<i>A. baumannii</i> B65371 (-)	>40	>40	>40
<i>E.cloacae</i> NRRL-B-425 (-)	>40	>40	>40
<i>E. coli</i> K12 (-)	>40	>40	>40
<i>K. pneumoniae</i> NRRL-B-408 (-)	>40	>40	>40
MRSA ATCC43300 (+)	0.03	0.7	0.8
MRSA ATCC29213 (+)	0.04	1.0	0.4
MRSA ATCC29213 Biofilm	0.02	0.6	1.2
MRSA Sa115 (+)	0.04	1.0	0.4
<i>S. aureus</i> ATCC 25923 (+)	0.04	0.3	0.8
<i>S. aureus</i> R2952 (+)	0.04	1.0	0.4
<i>B. subtilis</i> zk31 (+)	0.04	0.5	0.07
<i>M. luteus</i> ATCC 10040 (+)	0.02	0.8	1.6
HaCaT cells	14.74	14.90	

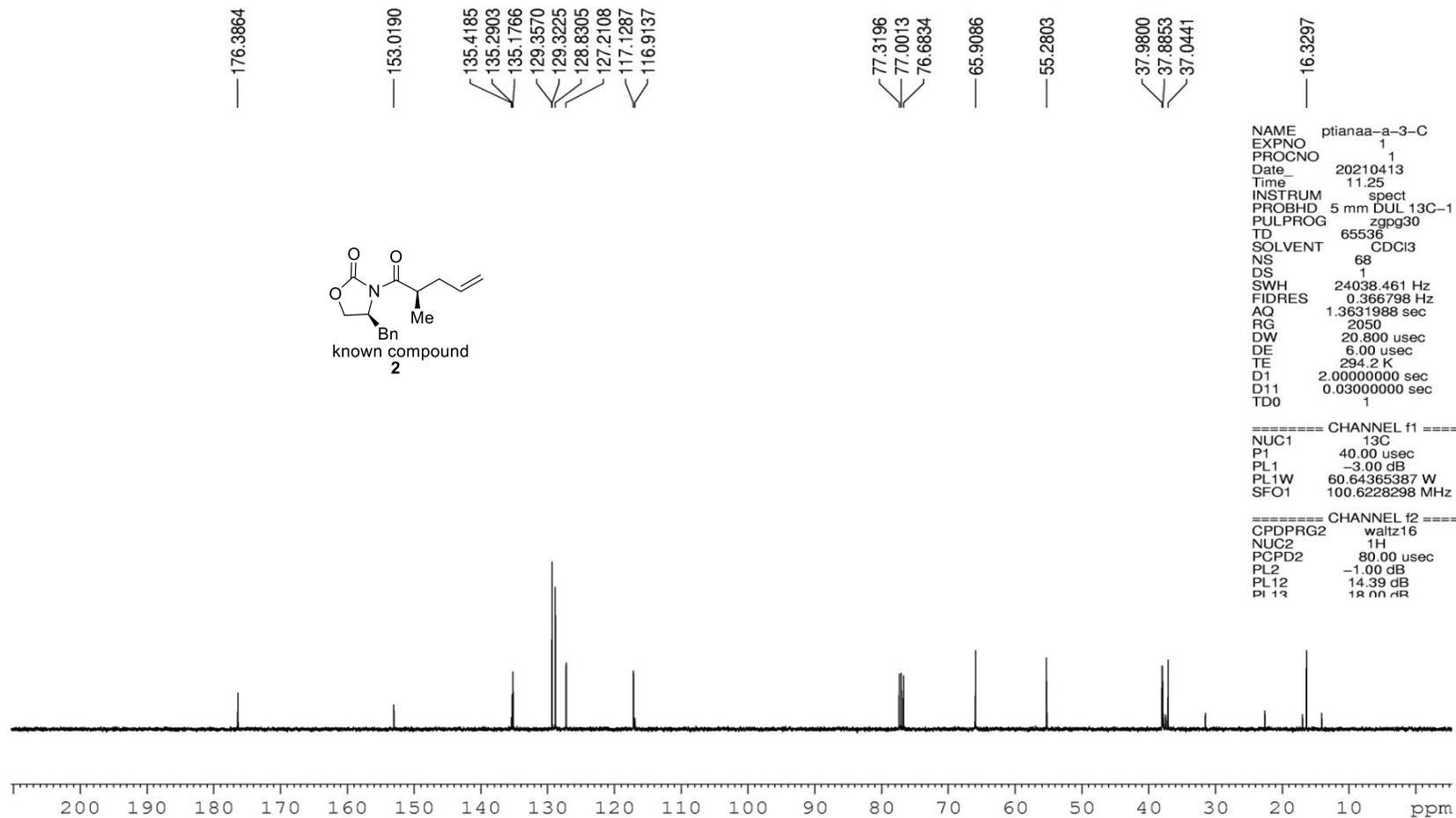
Note: (-) represents Gram-negative strains, and (+) for positive. The experimental results are expressed as MIC, MBIC and IC₅₀ (ug/mL). All the bioactivity assays of compounds were performed three times. In antimicrobial experiments, kanamycin and vancomycin were used as positive controls.

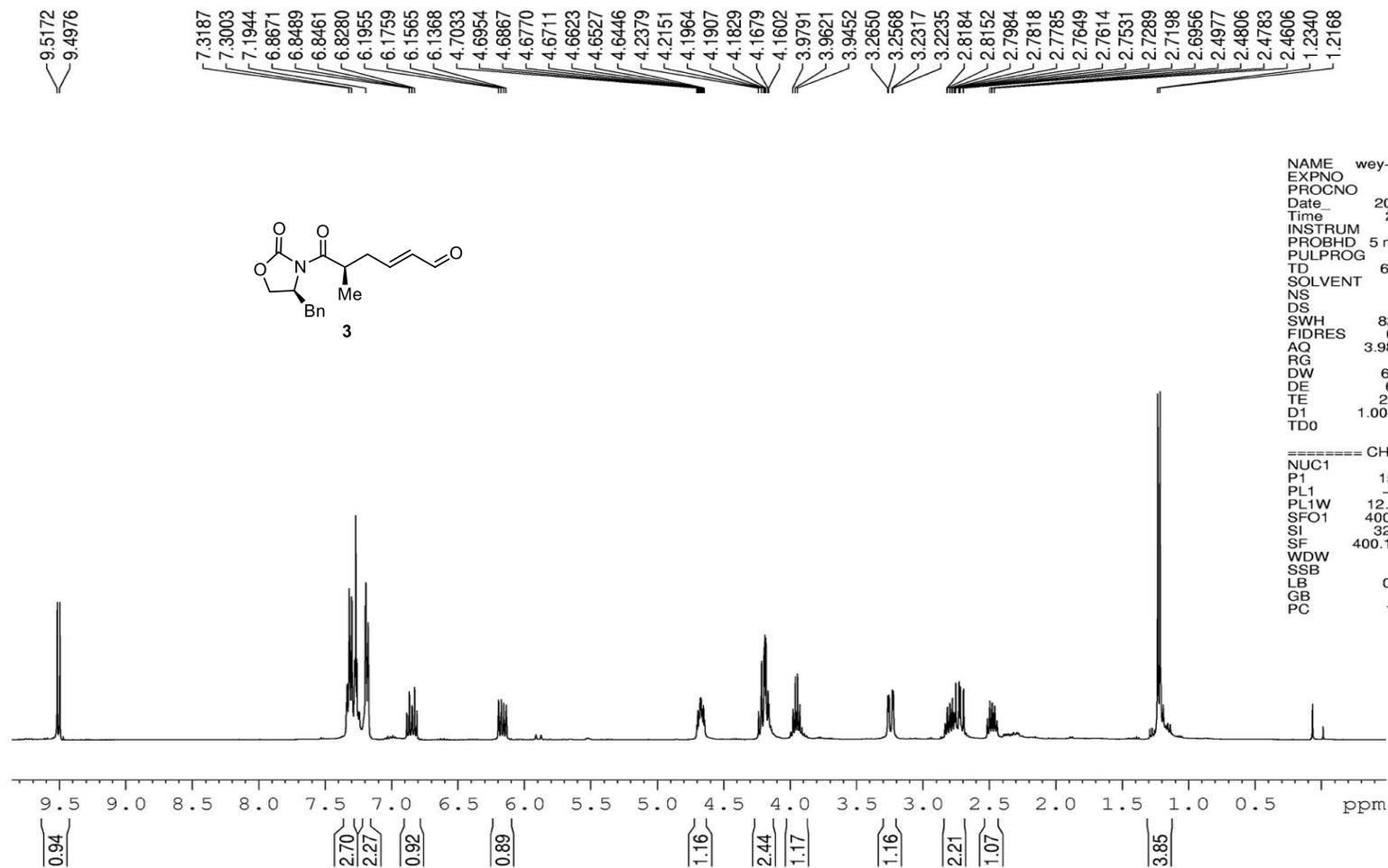
References:

1. Rahn N, Kalesse M. *Angew. Chem. Int. Ed.* **2008**, *47*, 597-599.
2. Davison, E. K., Freeman, J. L., Zhang, W., Wuest, W. M., Furkert, D. P., Brimble, M. A. *Org. Lett.* **2020**, *22*, 5550–5554.
3. Xiong X, Wu Y, Liu B. *Eur. J. Org. Chem.* **2020**, *8*, 948–960.
4. Freeman J L, Brimble M A, Furkert D . *Org. Chem. Front.*, **2019**, *6*, 2954-2963.
5. Weber F, Brückner R. *Org. Lett.* **2014**, *16*, 6428–6431.
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7. Jang K H, Nam S J, Locke J B, et al. a, Beatty, DS, Paul, L. a, Fenical, W. *Angew. Chem. Int. Ed.* **2013**, *52*, 7822 –7824.
8. Rodríguez, V., Martín, J., Sarmiento-Vizcaíno, A., De la Cruz, M., García, L. A., Blanco, G., Reyes, F. *Mar. Drugs*, **2018**, *16*, 406-413

¹H and ¹³C NMR Spectra

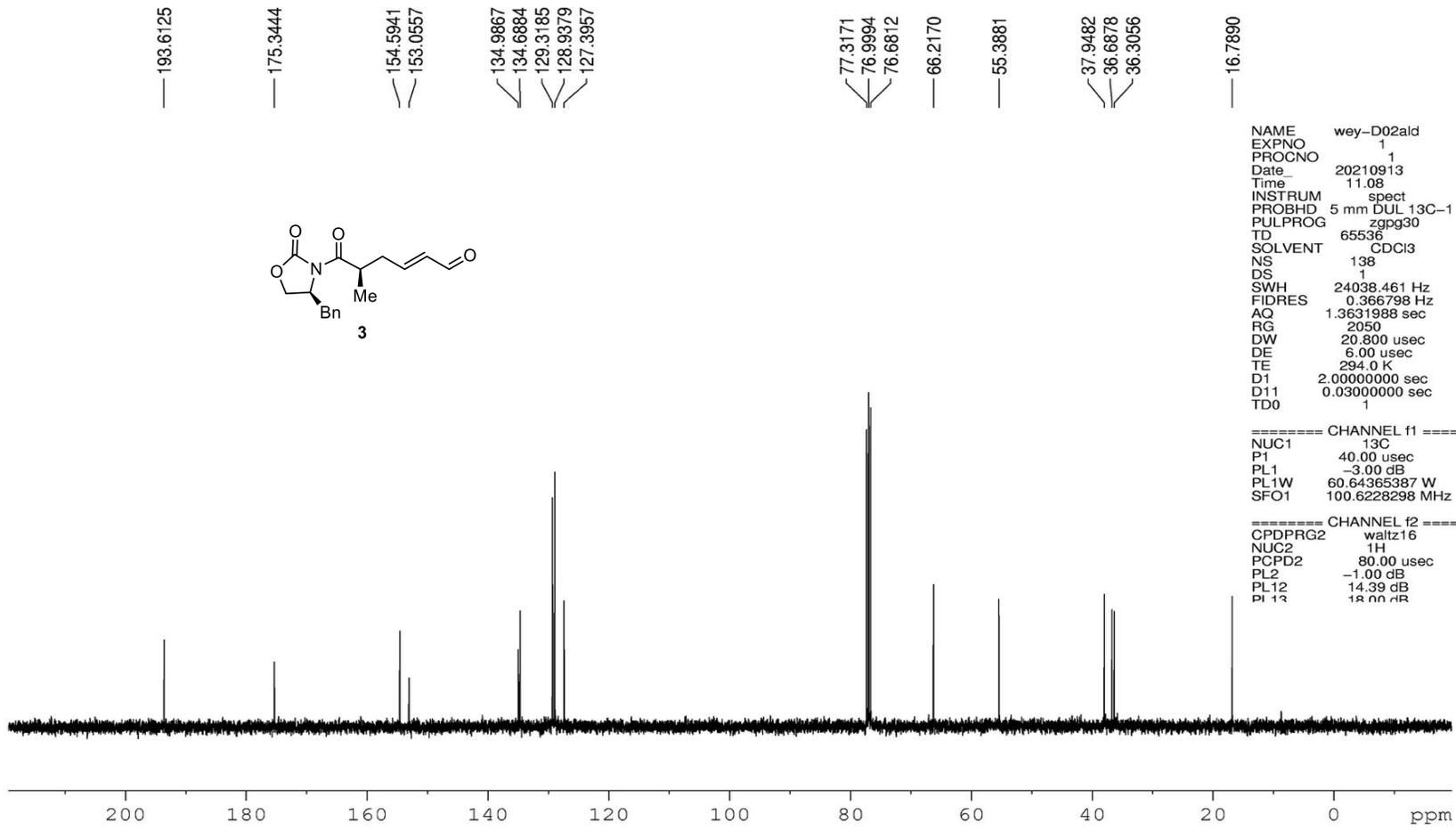






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 TD 65536
 SOLVENT CDCl3
 NS 16
 DS 2
 SWH 8223.685 Hz
 FIDRES 0.125483 Hz
 AQ 3.9846387 sec
 RG 64
 DW 60.800 usec
 DE 6.00 usec
 TE 292.5 K
 D1 1.00000000 sec
 TD0 1

===== CHANNEL f1 =====
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 PL1 -1.00 dB
 PL1W 12.17476940 W
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 SI 32768
 SF 400.1300054 MHz
 WDW EM
 SSB 0
 LB 0.30 Hz
 GB 0
 PC 1.00



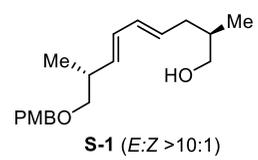
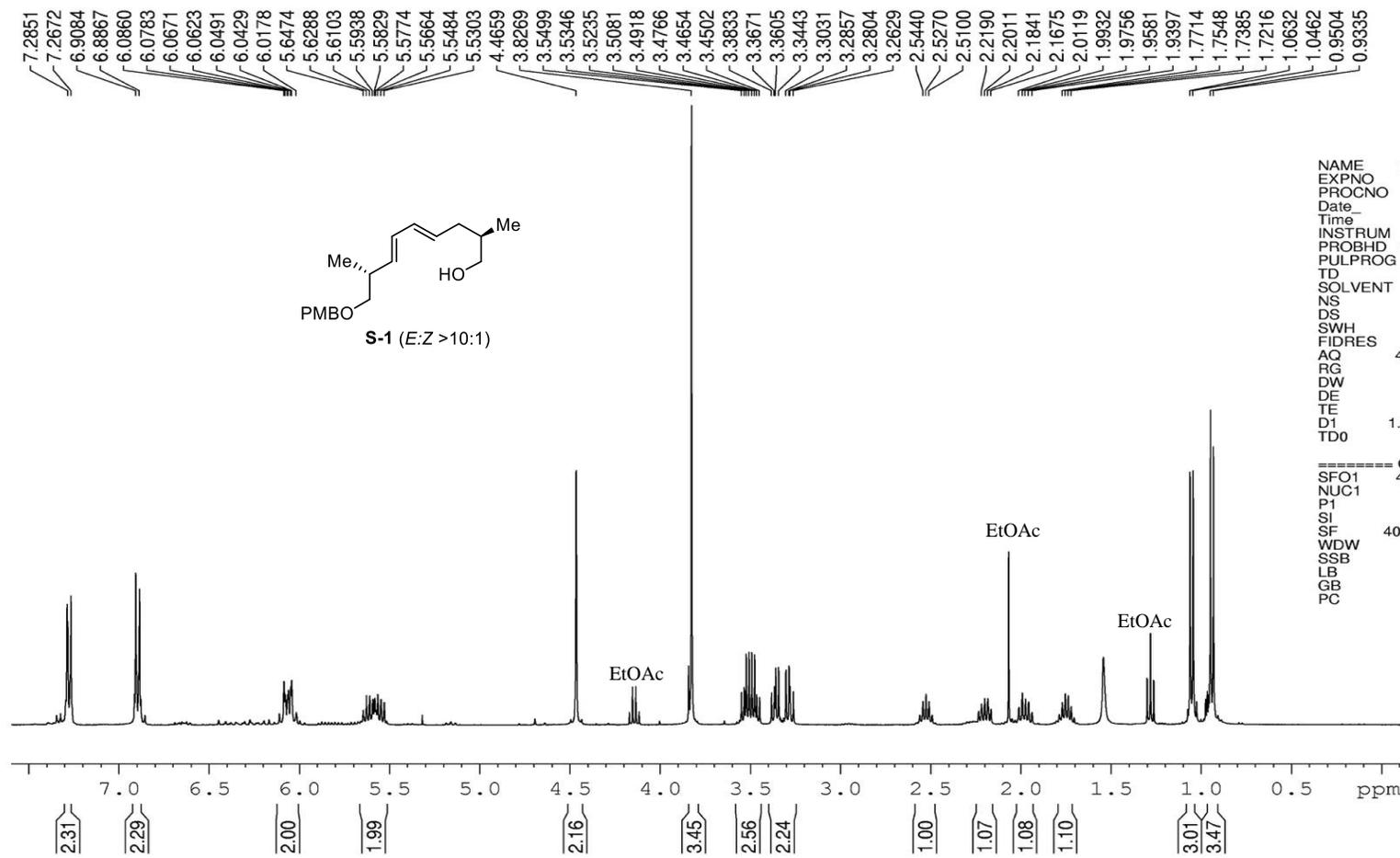
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TD         65536
SOLVENT   CDCl3
NS         138
DS         1
SWH        24038.461 Hz
FIDRES     0.366798 Hz
AQ         1.3631988 sec
RG         2050
DW         20.800 usec
DE         6.00 usec
TE         294.0 K
D1         2.00000000 sec
D11        0.03000000 sec
TD0        1

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NUC1       13C
P1         40.00 usec
PL1        -3.00 dB
PL1W       60.64365387 W
SFO1       100.6228298 MHz

===== CHANNEL f2 =====
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PCPD2      80.00 usec
PL2        -1.00 dB
PL12       14.39 dB
PL13       18.00 dB

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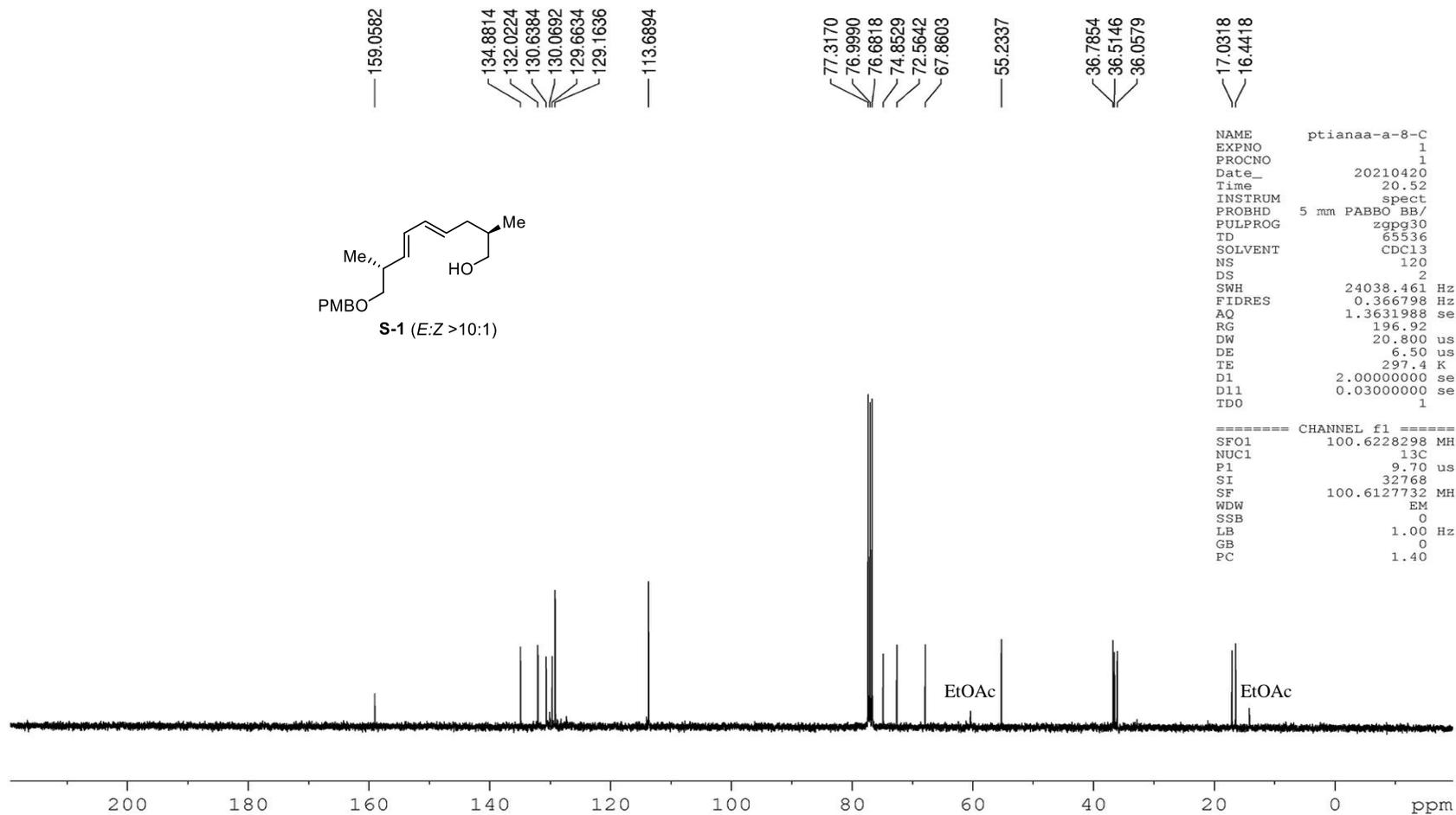
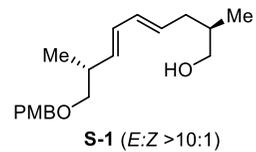


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PULPROG zg30
TD 65536
SOLVENT CDCl3
NS 7
DS 2
SWH 8012.820 Hz
FIDRES 0.122266 Hz
AQ 4.0894966 sec
RG 62.93
DW 62.400 usec
DE 6.50 usec
TE 296.7 K
D1 1.00000000 sec
TD0 1

===== CHANNEL f1 =====
SFO1 400.1324710 MHz
NUC1 1H
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SI 65536
SF 400.1300000 MHz
WDW EM
SSB 0
LB 0.30 Hz
GB 0
PC 1.00

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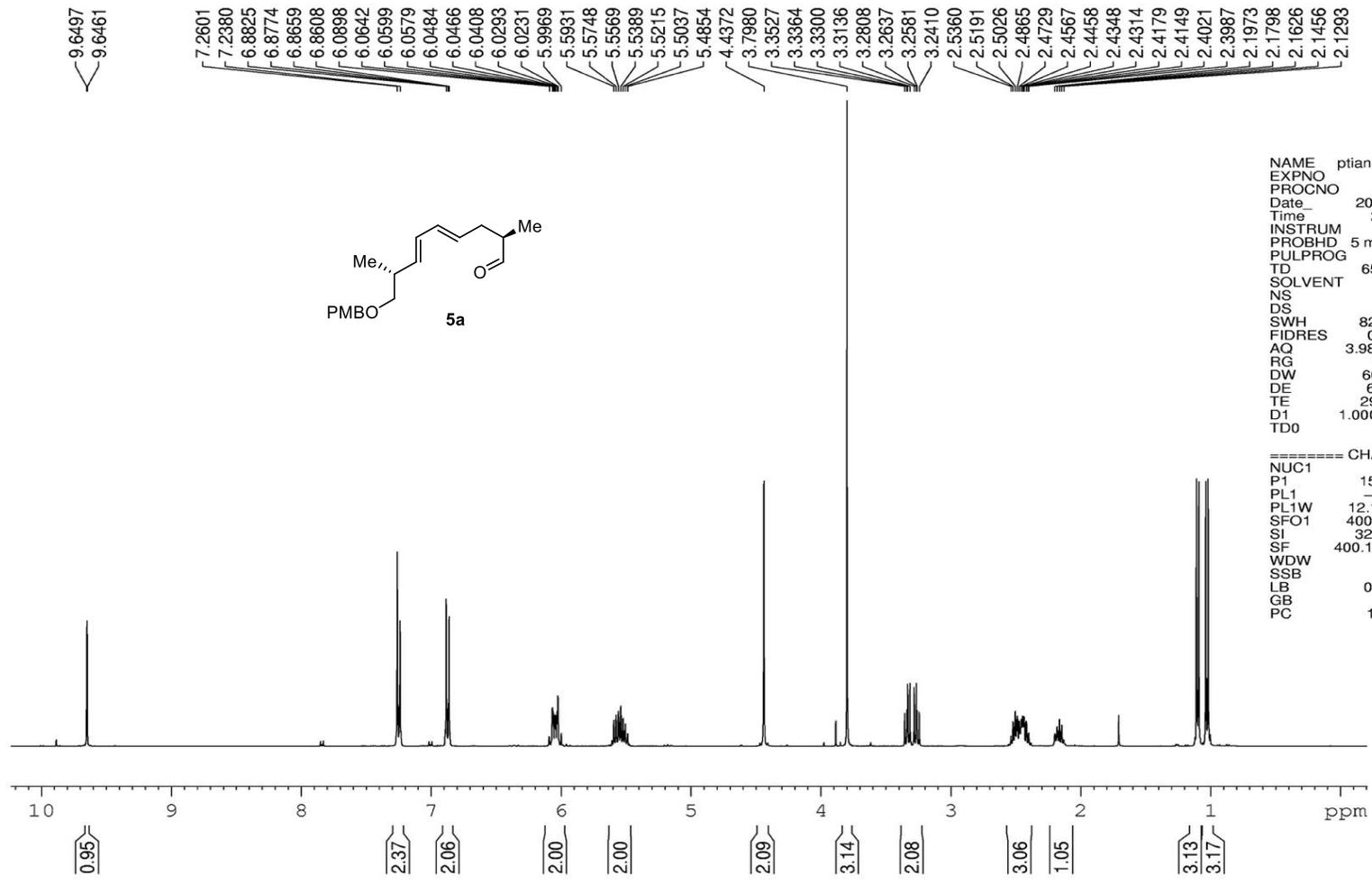


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PROCNO    1
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PULPROG   zgpg30
TD         65536
SOLVENT   CDCl3
NS         120
DS         2
SWH        24038.461 Hz
FIDRES     0.366798 Hz
AQ         1.3631988 se
RG         196.92
DW         20.800 us
DE         6.50 us
TE         297.4 K
DL         2.00000000 se
D11        0.03000000 se
TD0        1

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NUC1       13C
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SF         100.6127732 MH
WDW        EM
SSB        0
LB         1.00 Hz
GB         0
PC         1.40

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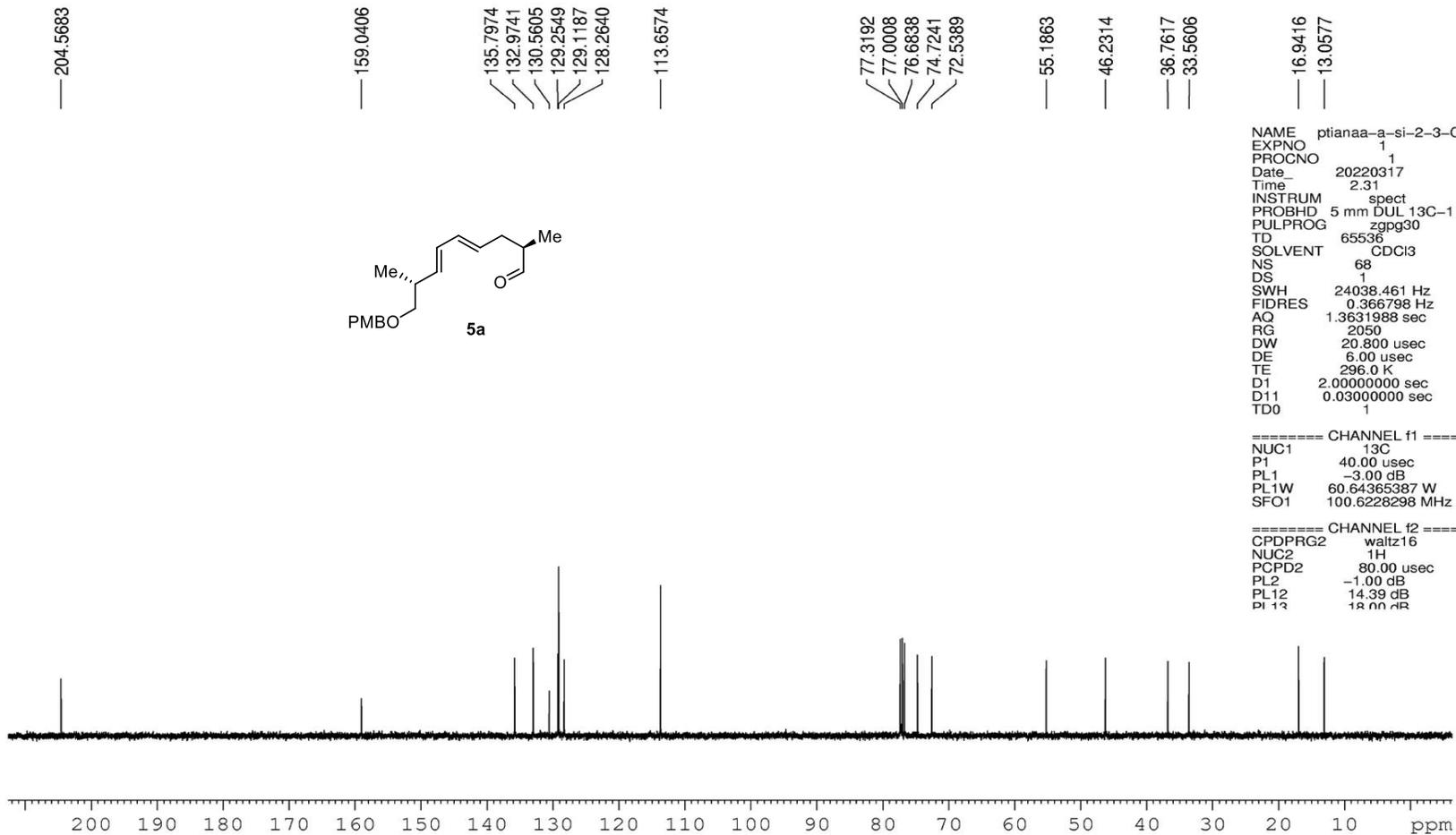


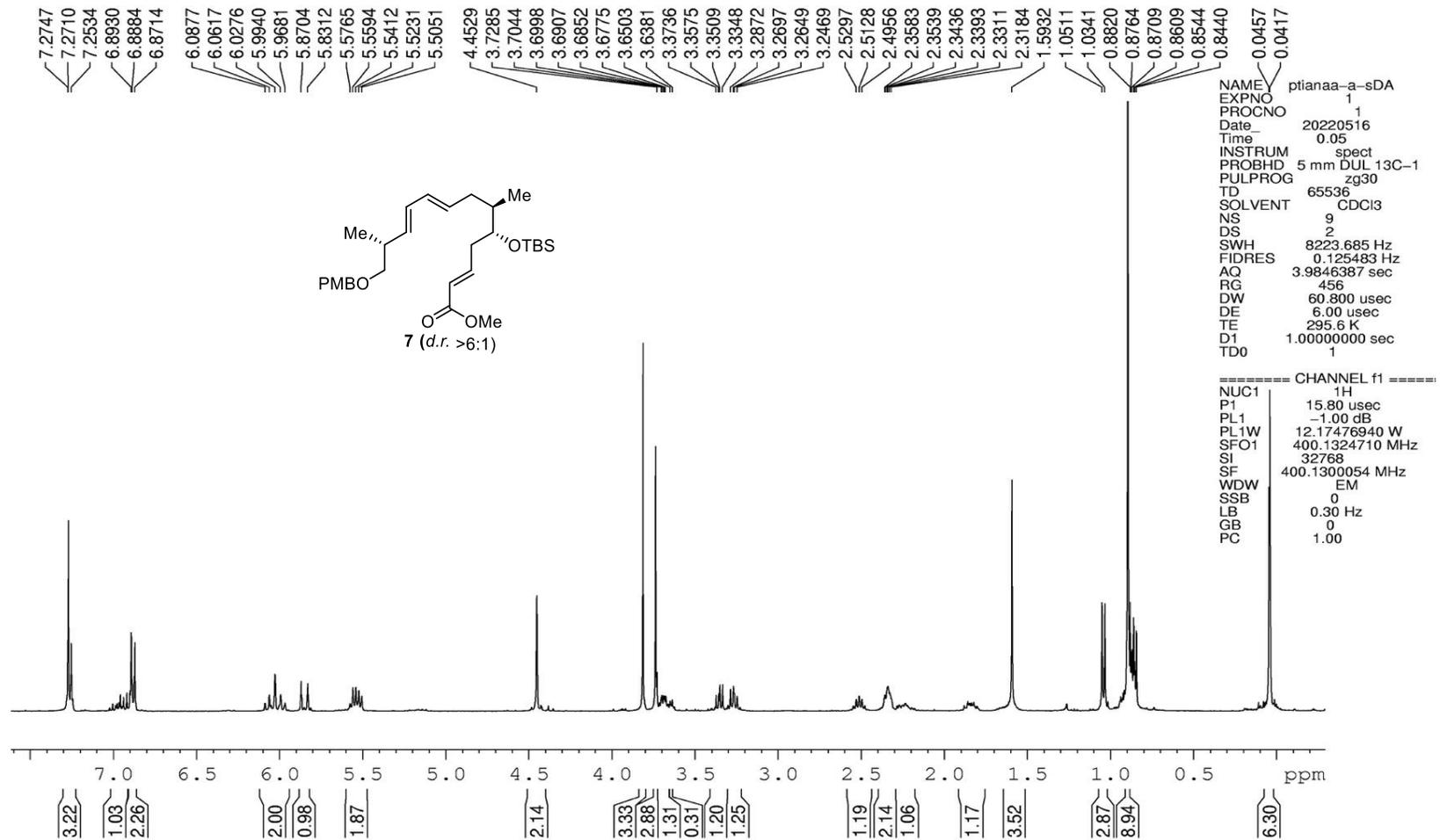
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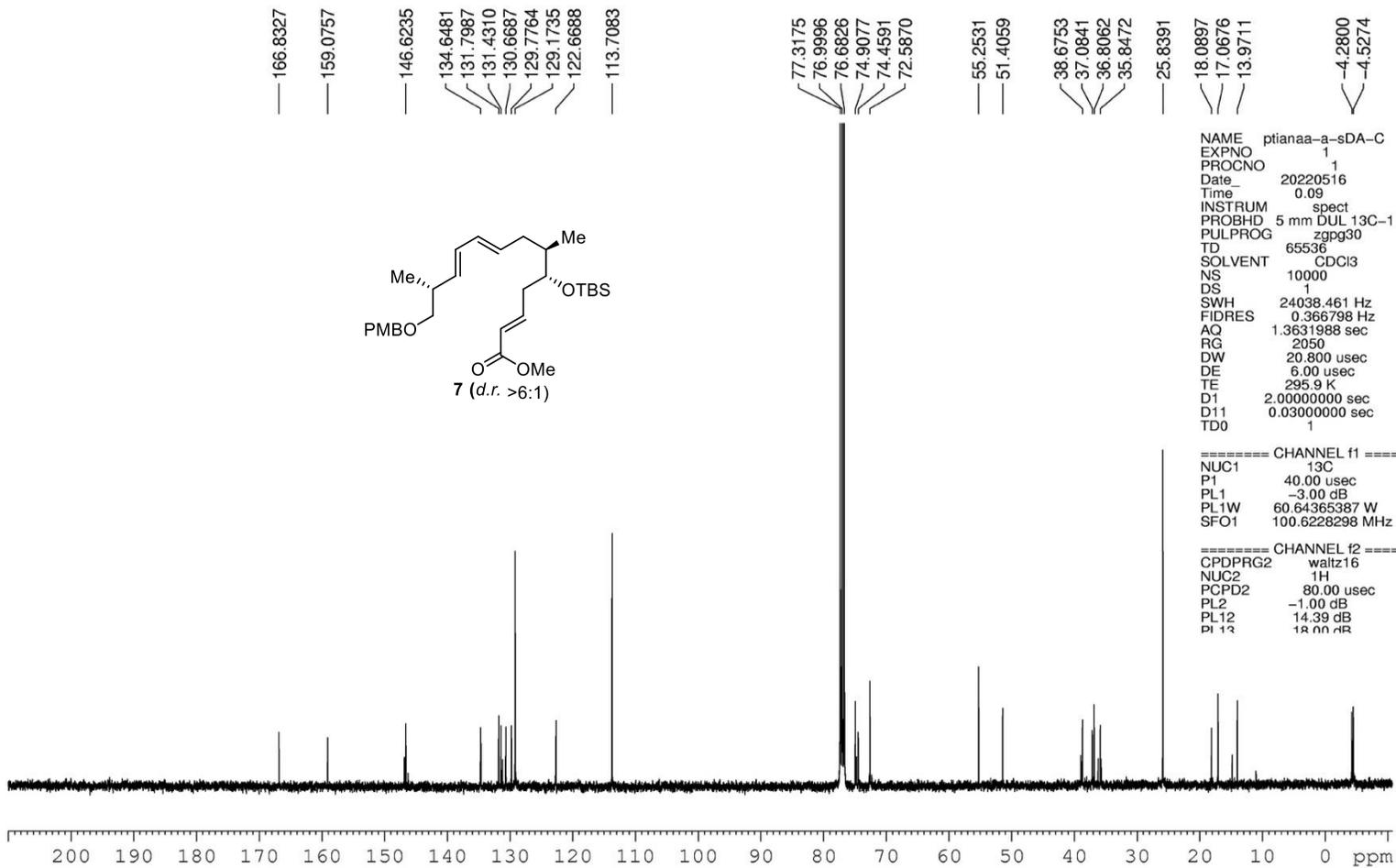
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PROCNO 1
Date_ 20220317
Time 2.27
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PULPROG zg30
TD 65536
SOLVENT CDCl3
NS 5
DS 2
SWH 8223.685 Hz
FIDRES 0.125483 Hz
AQ 3.9846387 sec
RG 144
DW 60.800 usec
DE 6.00 usec
TE 295.7 K
D1 1.00000000 sec
TD0 1

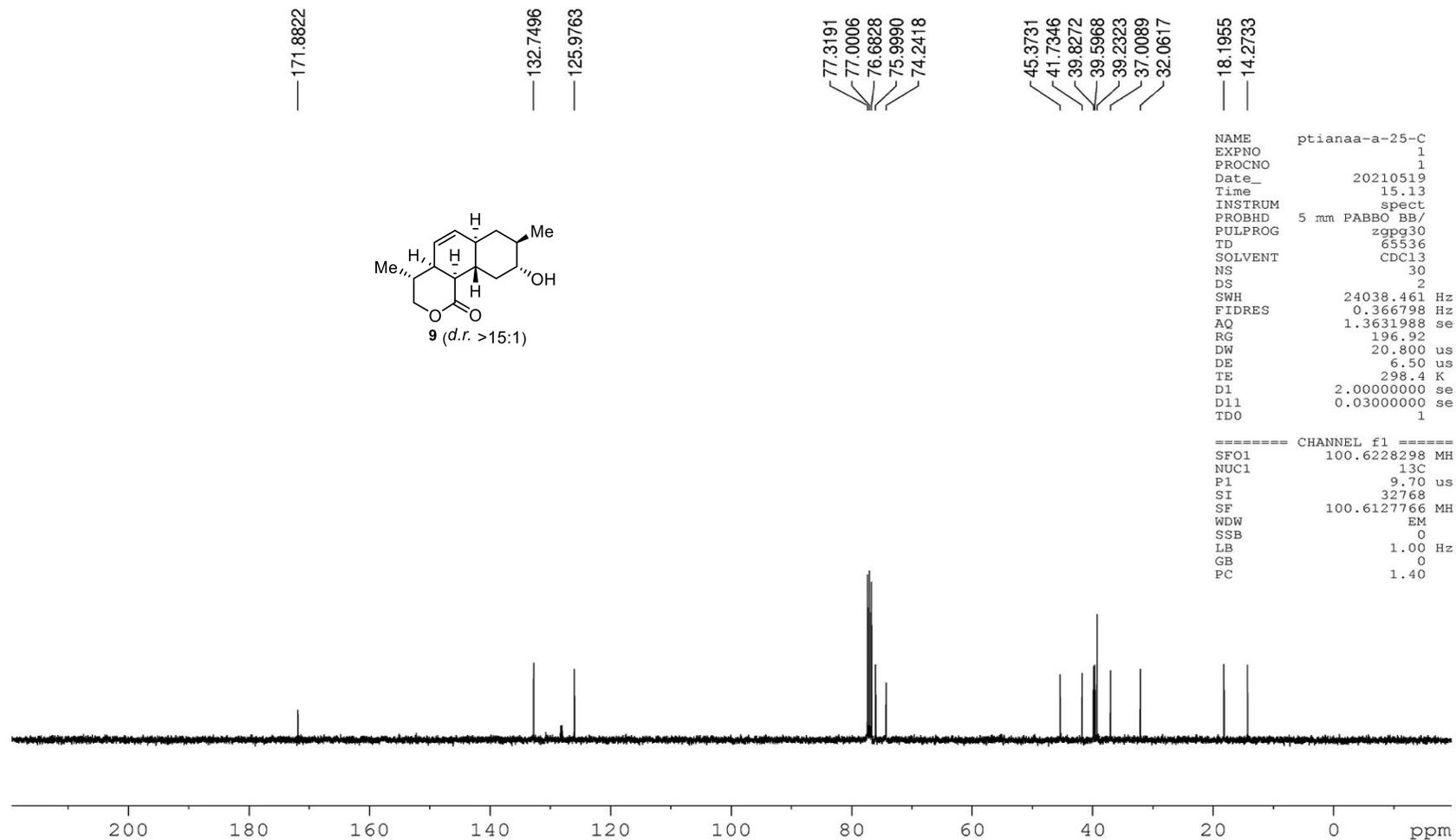
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NUC1 1H
P1 15.80 usec
PL1 -1.00 dB
PL1W 12.17476940 W
SFO1 400.1324710 MHz
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SF 400.1300096 MHz
WDW EM
SSB 0
LB 0.30 Hz
GB 0
PC 1.00

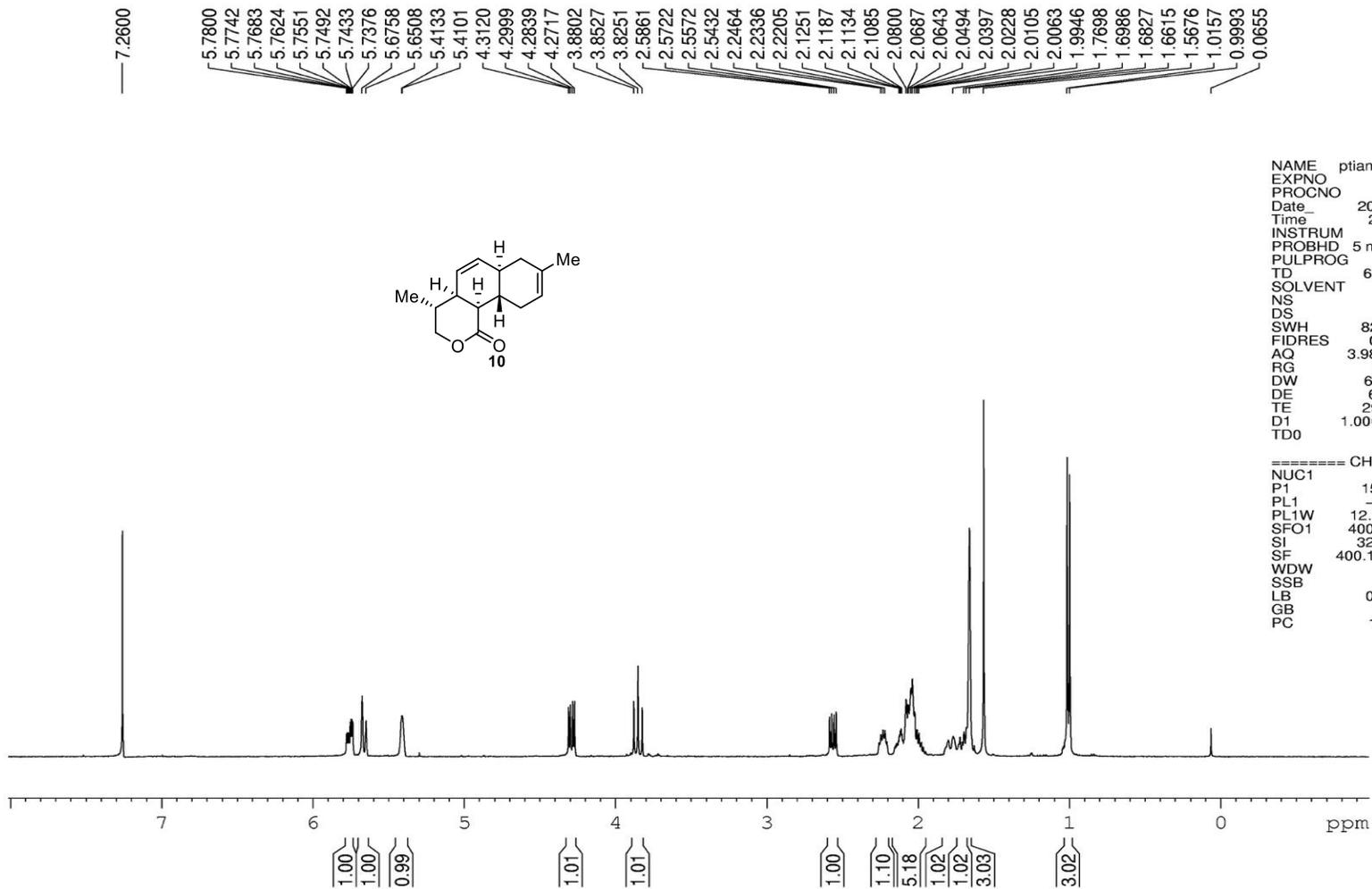
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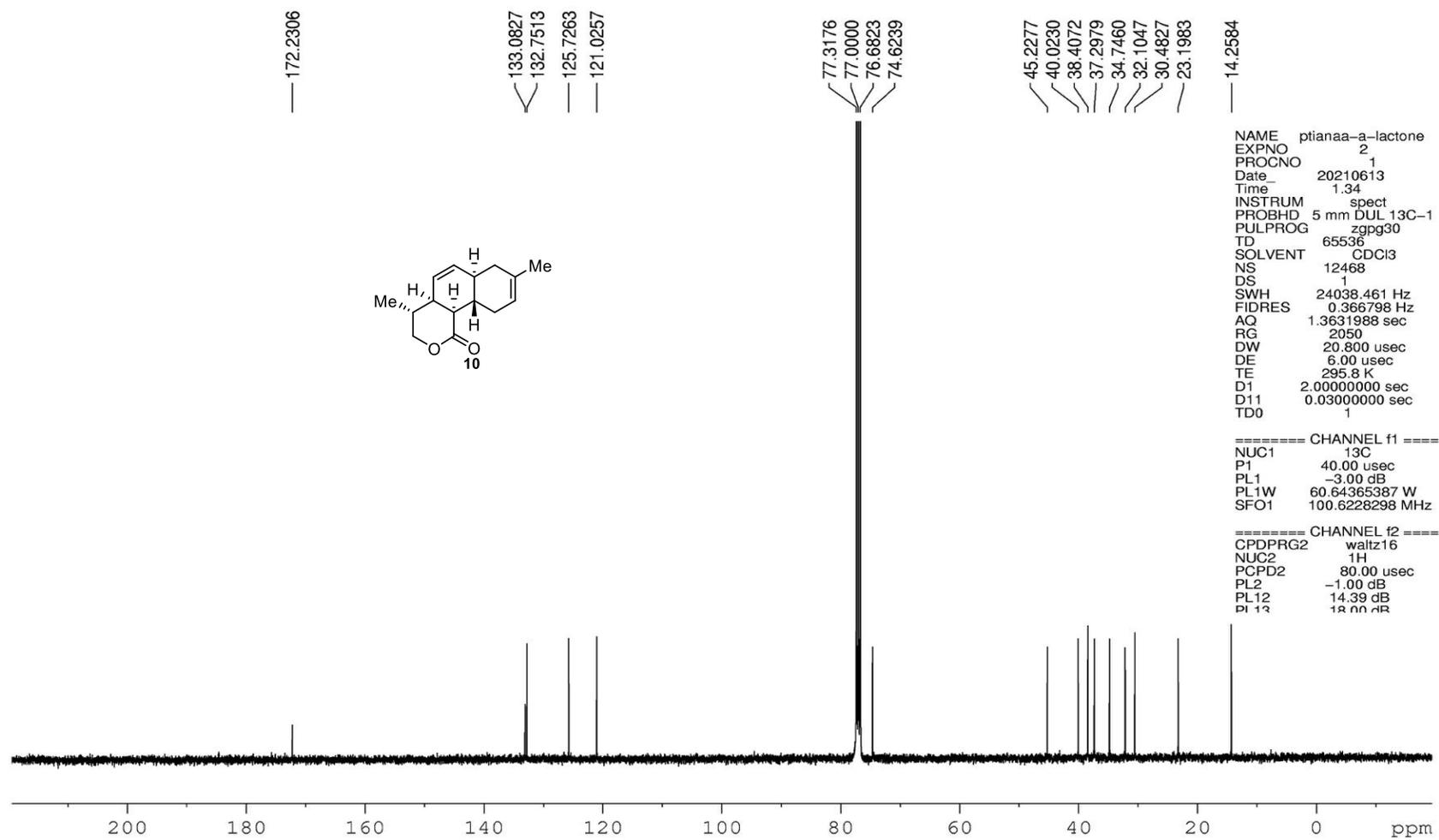


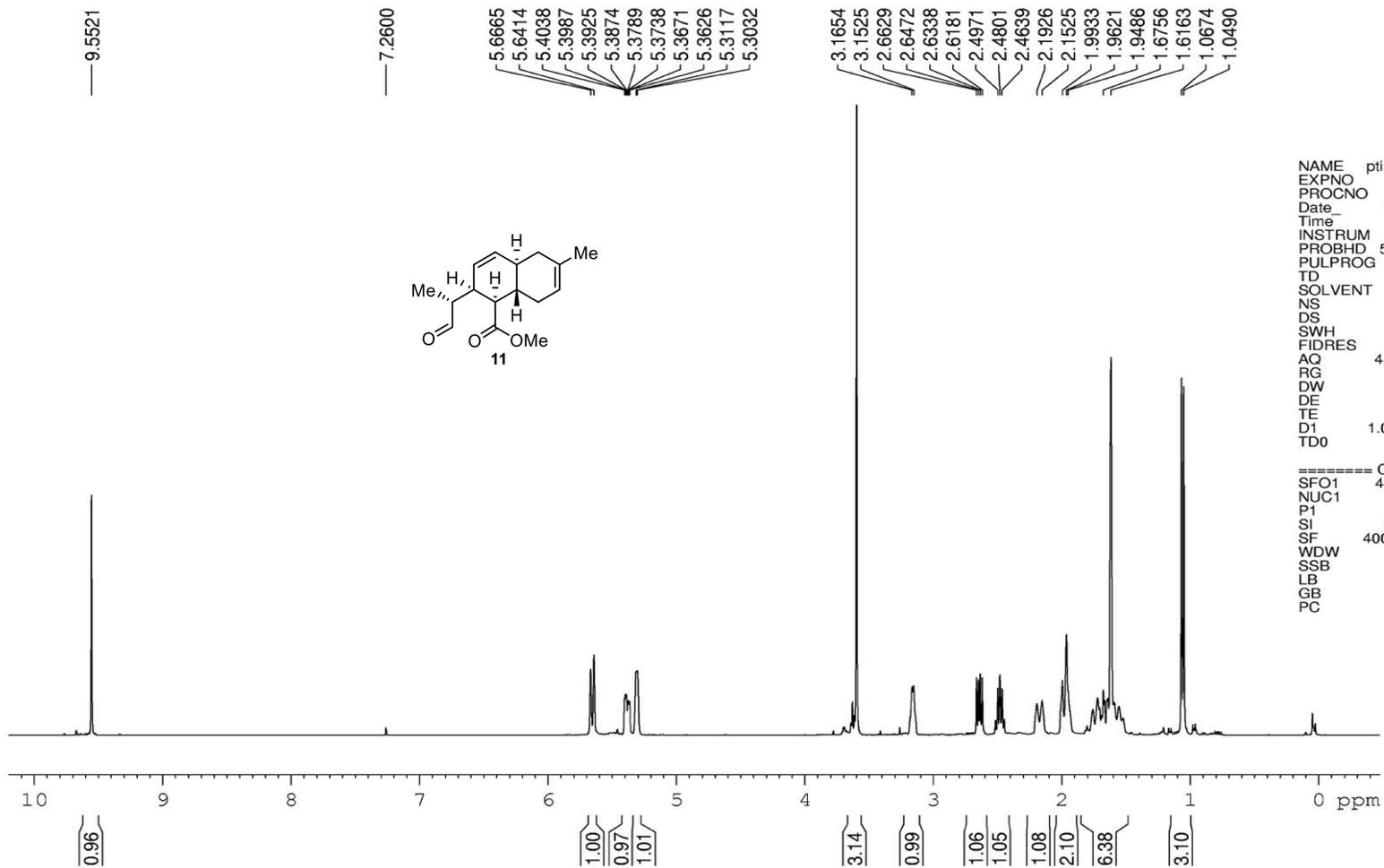




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 PULPROG zg30
 TD 65536
 SOLVENT CDCl3
 NS 12
 DS 2
 SWH 8223.685 Hz
 FIDRES 0.125483 Hz
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 RG 645
 DW 60.800 usec
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 D1 1.00000000 sec
 TD0 1

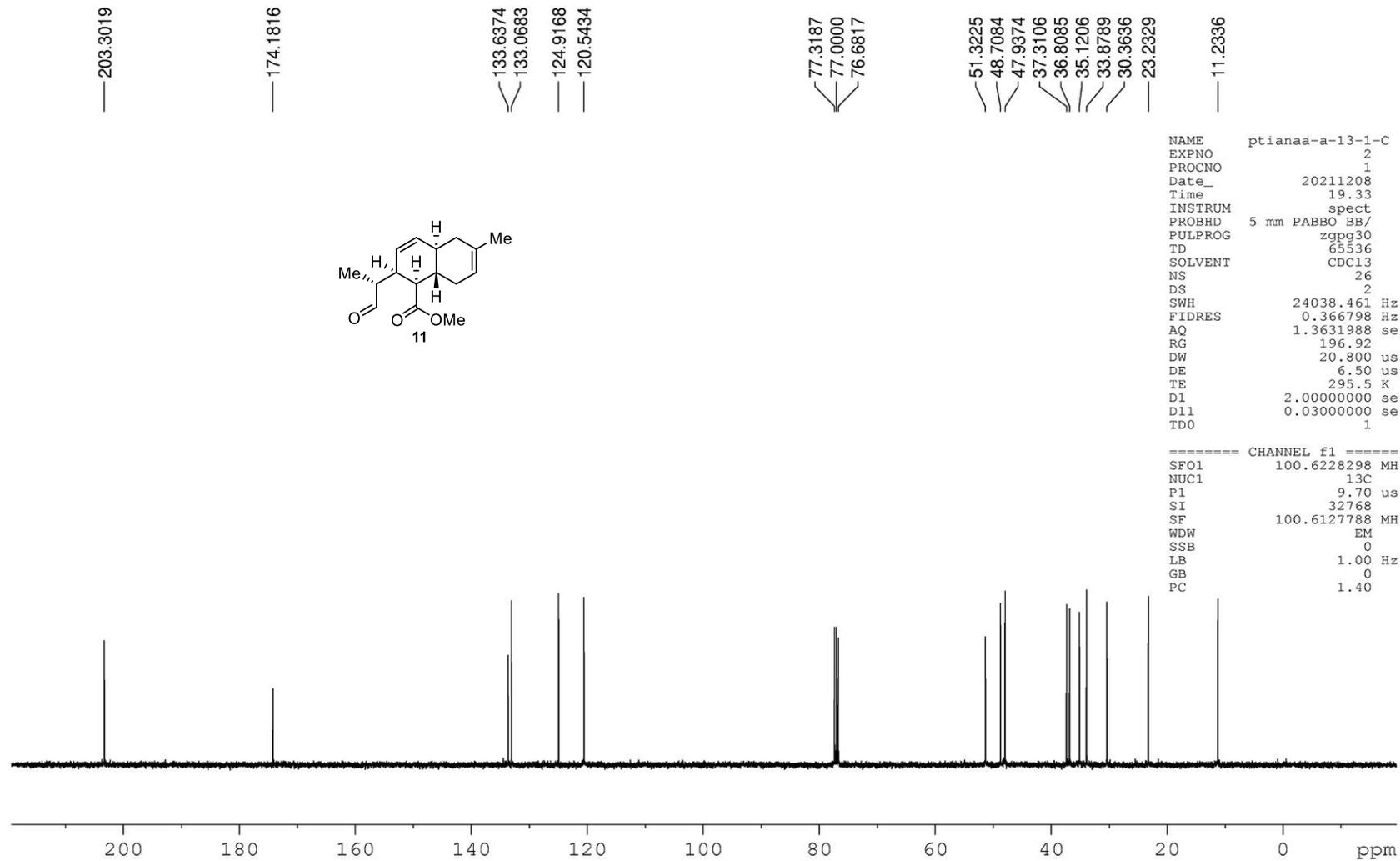
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 WDW EM
 SSB 0
 LB 0.30 Hz
 GB 0
 PC 1.00





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 PROCNO 1
 Date_ 20211208
 Time 19.31
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 PULPROG zg30
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 DS 2
 SWH 8012.820 Hz
 FIDRES 0.122266 Hz
 AQ 4.0894966 sec
 RG 15.71
 DW 62.400 usec
 DE 6.50 usec
 TE 294.9 K
 D1 1.00000000 sec
 TD0 1

===== CHANNEL f1 =====
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 SI 65536
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 SSB 0
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 GB 0
 PC 1.00



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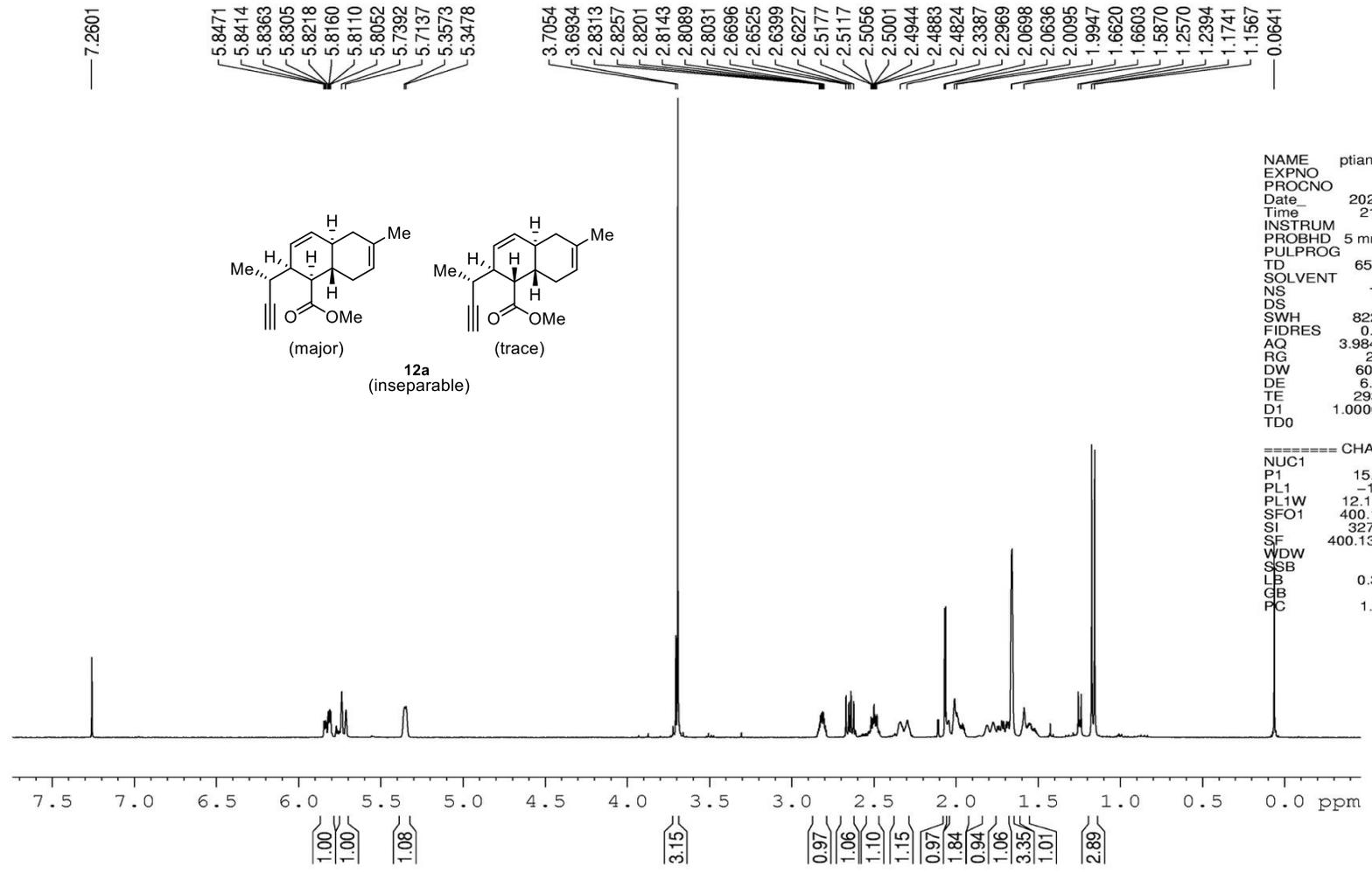
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PULPROG   zgpg30
TD         65536
SOLVENT   CDC13
NS         26
DS         2
SWH        24038.461 Hz
FIDRES     0.366798 Hz
AQ         1.3631988 se
RG         196.92
DW         20.800 us
DE         6.50 us
TE         295.5 K
D1         2.00000000 se
D11        0.03000000 se
TD0        1

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===== CHANNEL f1 =====
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NUC1       13C
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PC         1.40

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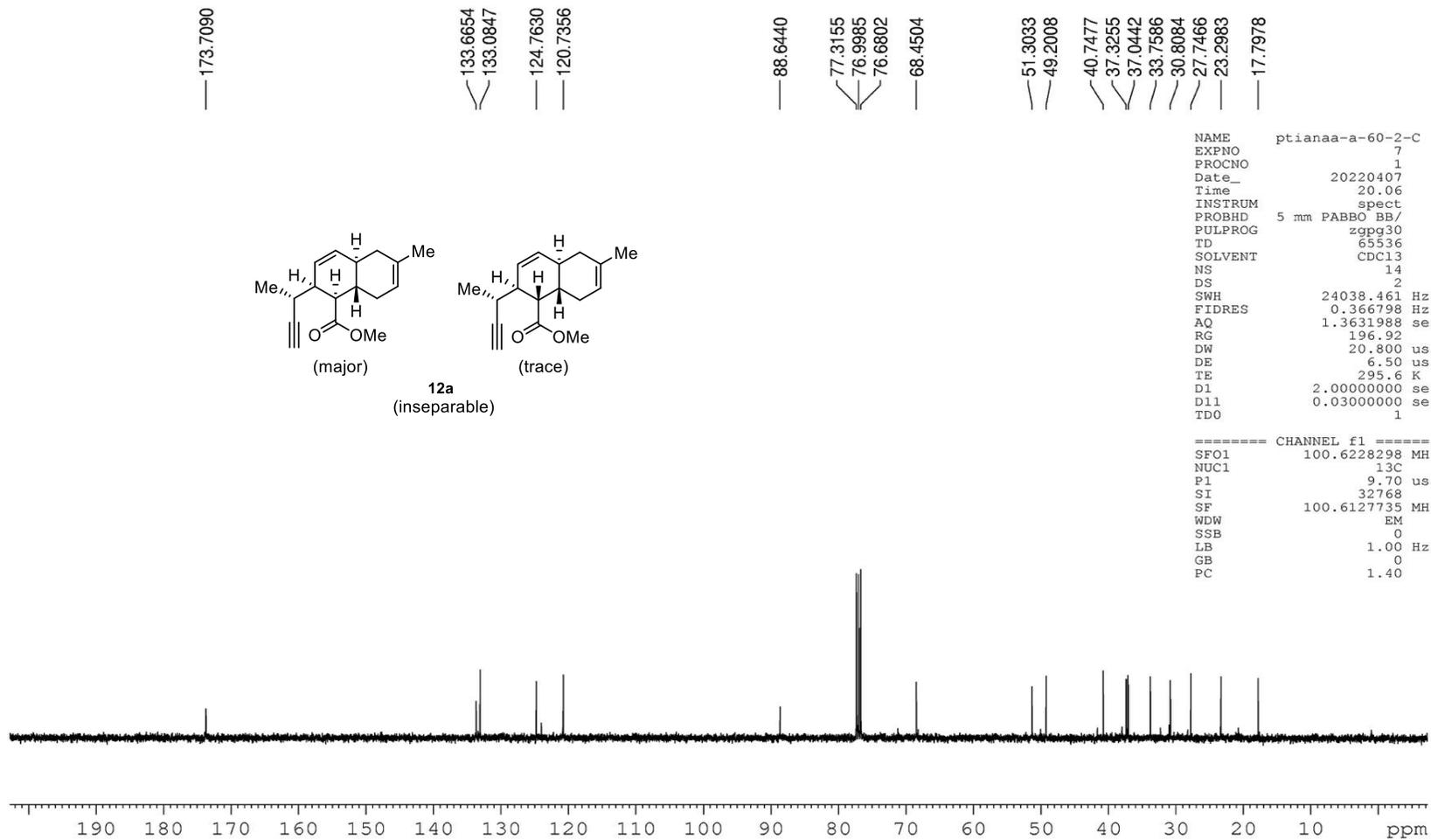
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SOLVENT   CDCl3
NS         11
DS         2
SWH        8223.685 Hz
FIDRES     0.125483 Hz
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RG         287
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DE         6.00 usec
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D1         1.0000000 sec
TD0        1

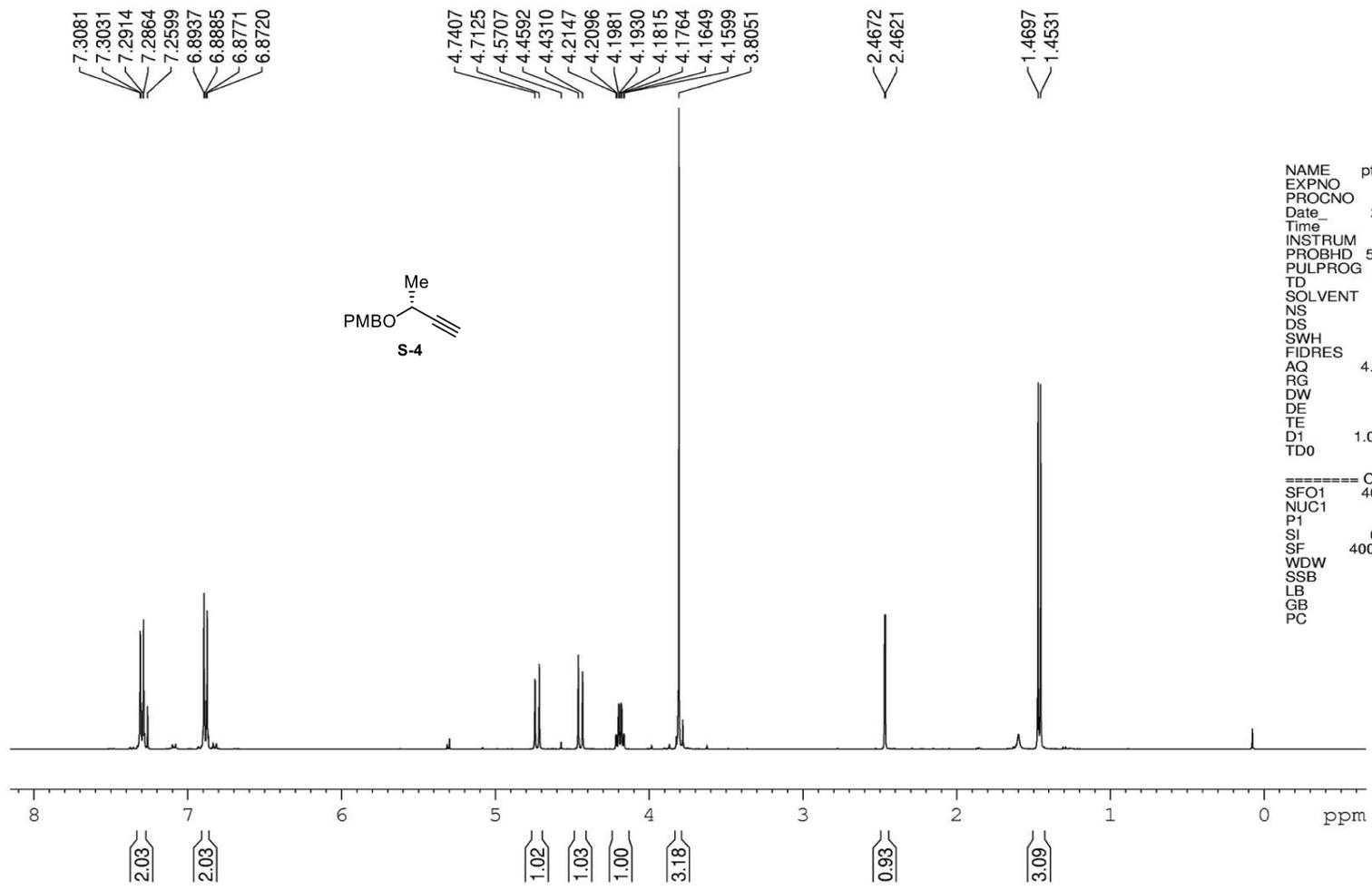
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===== CHANNEL f1 =====
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PL1       -1.00 dB
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SF        400.1300098 MHz
WDW       EM
SSB       0
LB        0.30 Hz
GB        0
PC        1.00

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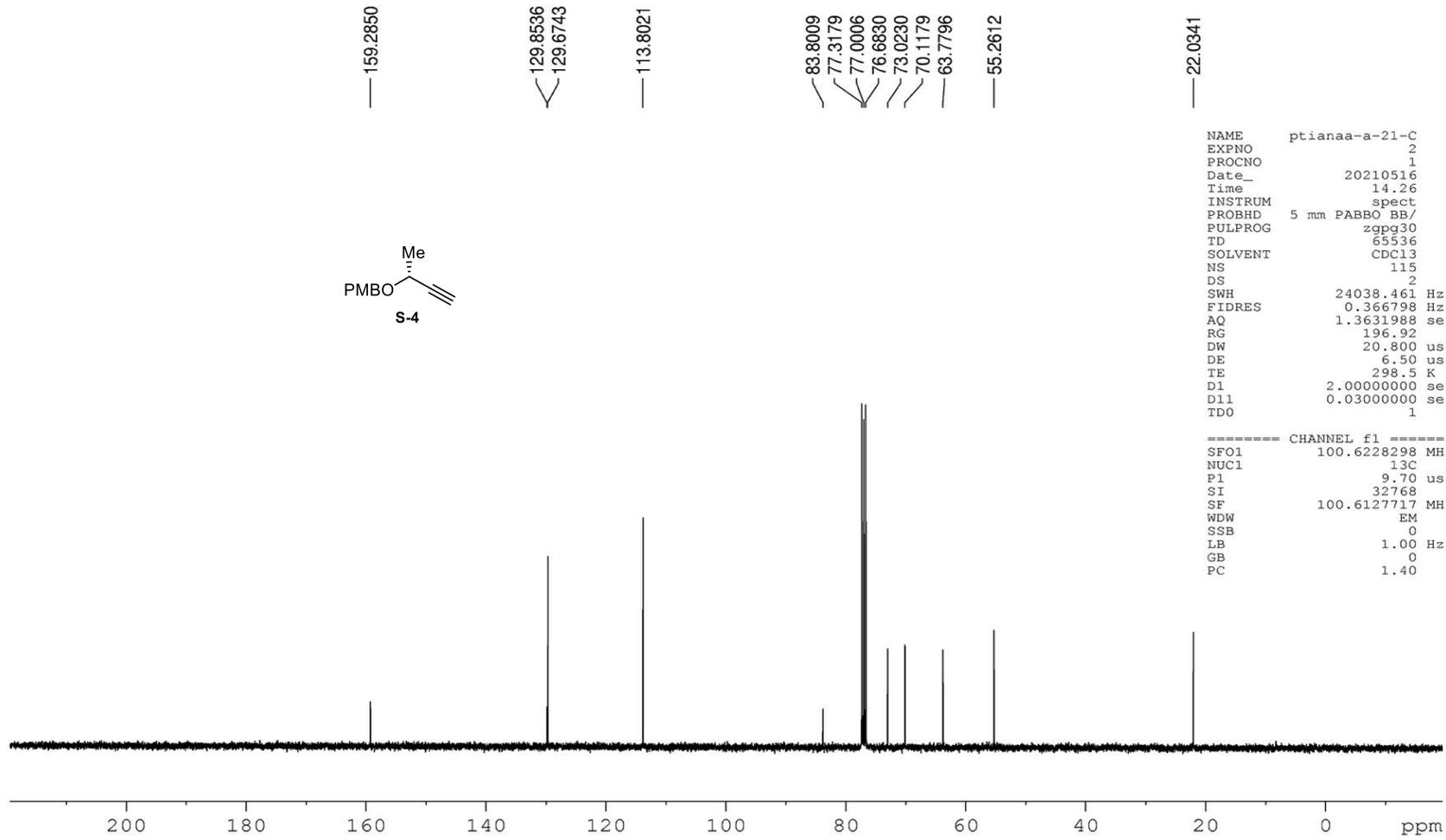
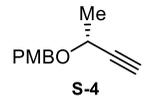


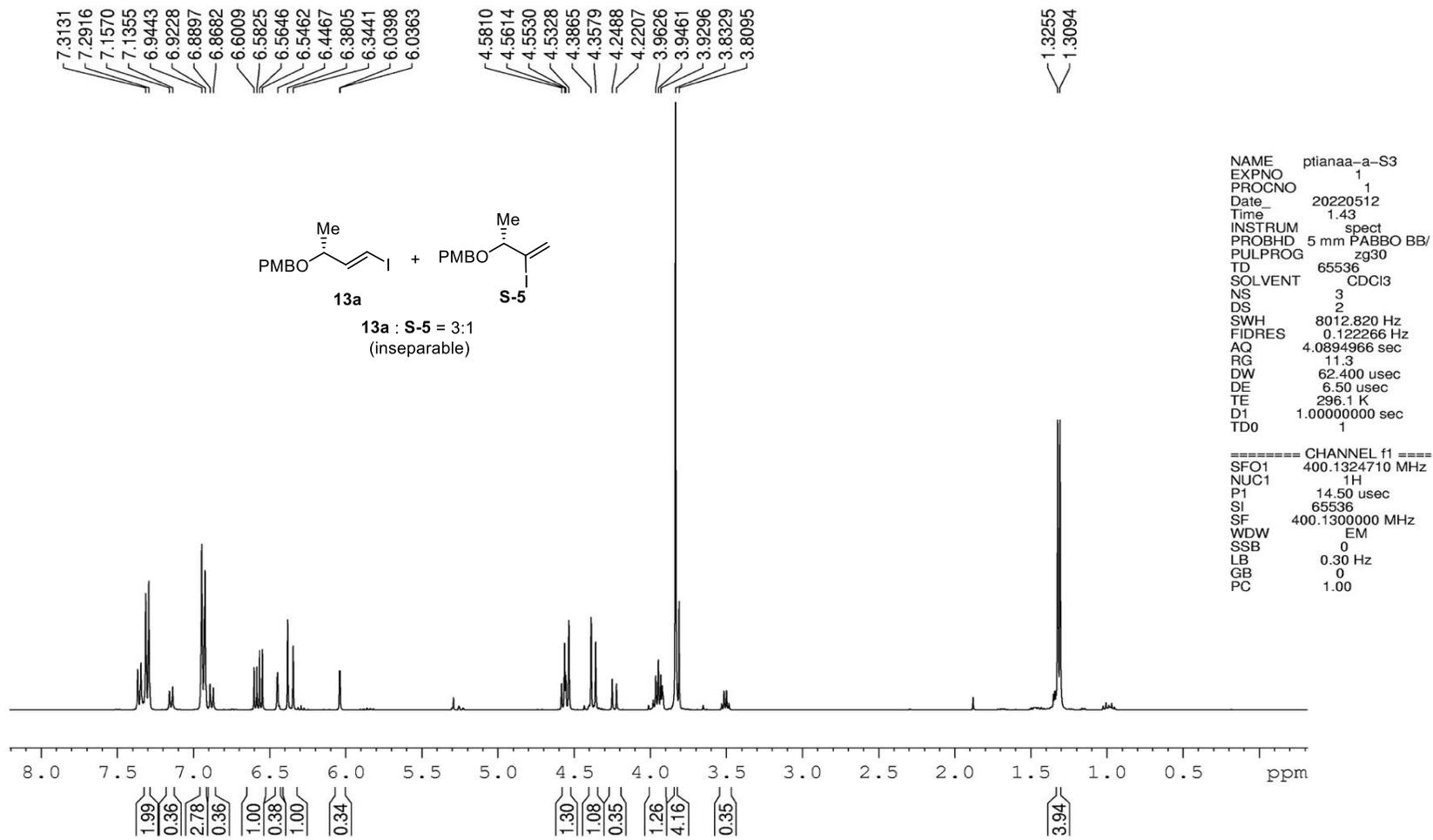
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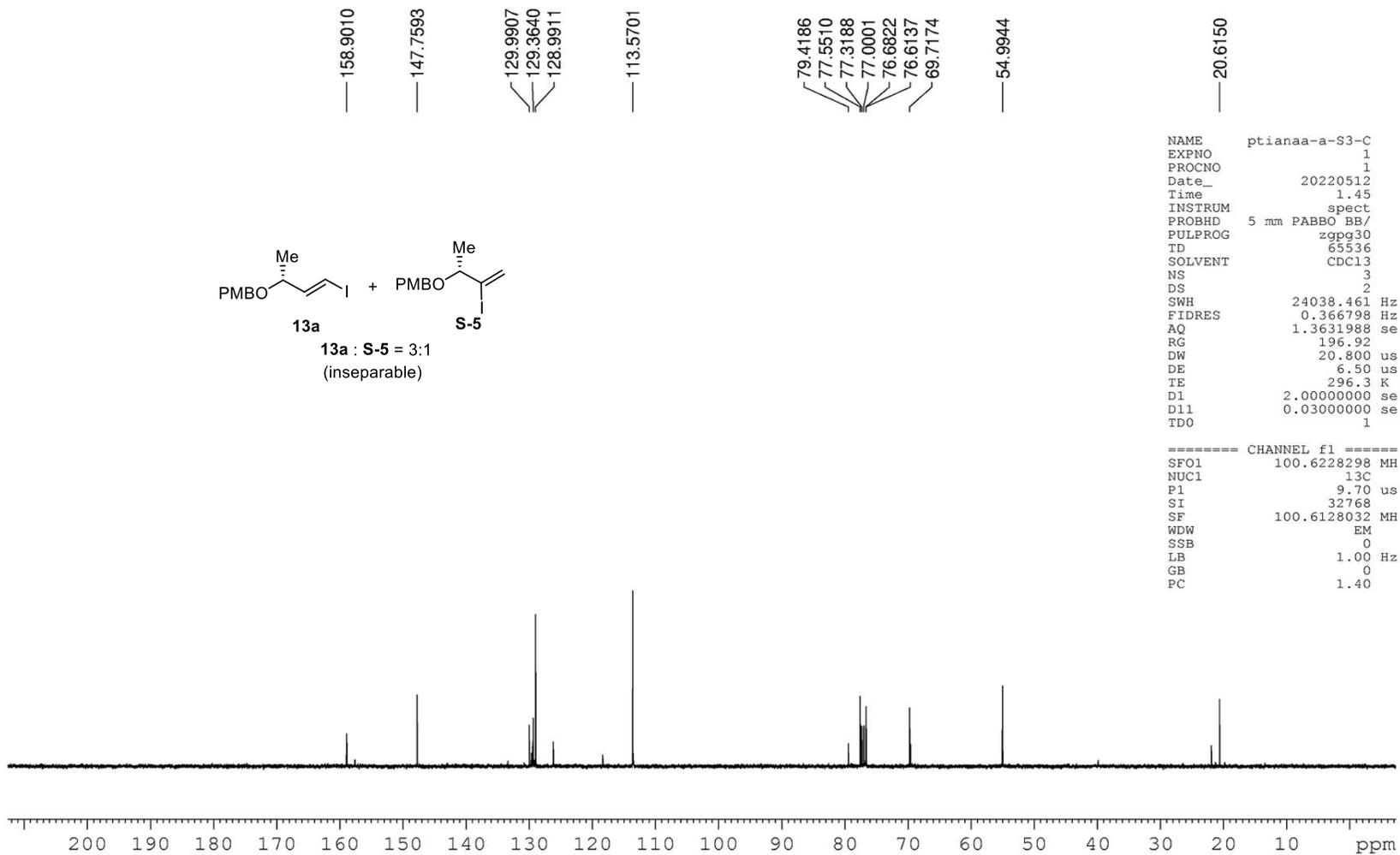
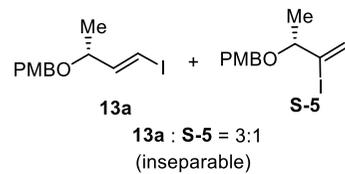
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EXPNO     2
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Time      14.24
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PULPROG   zg30
TD         65536
SOLVENT   CDCl3
NS         4
DS         2
SWH       8012.820 Hz
FIDRES    0.122266 Hz
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RG        88.84
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TE        297.9 K
D1        1.00000000 sec
TD0       1

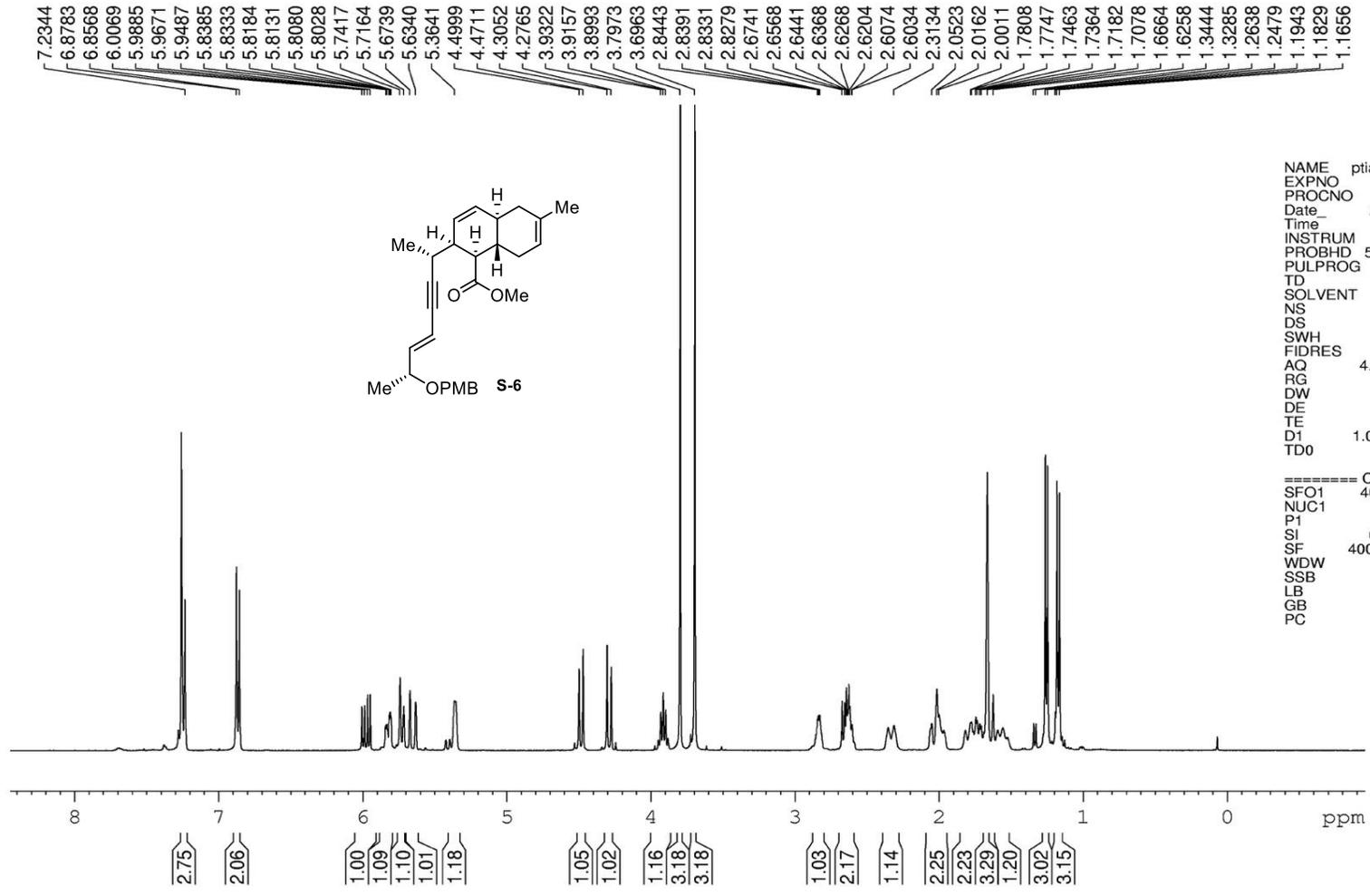
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GB        0
PC        1.00

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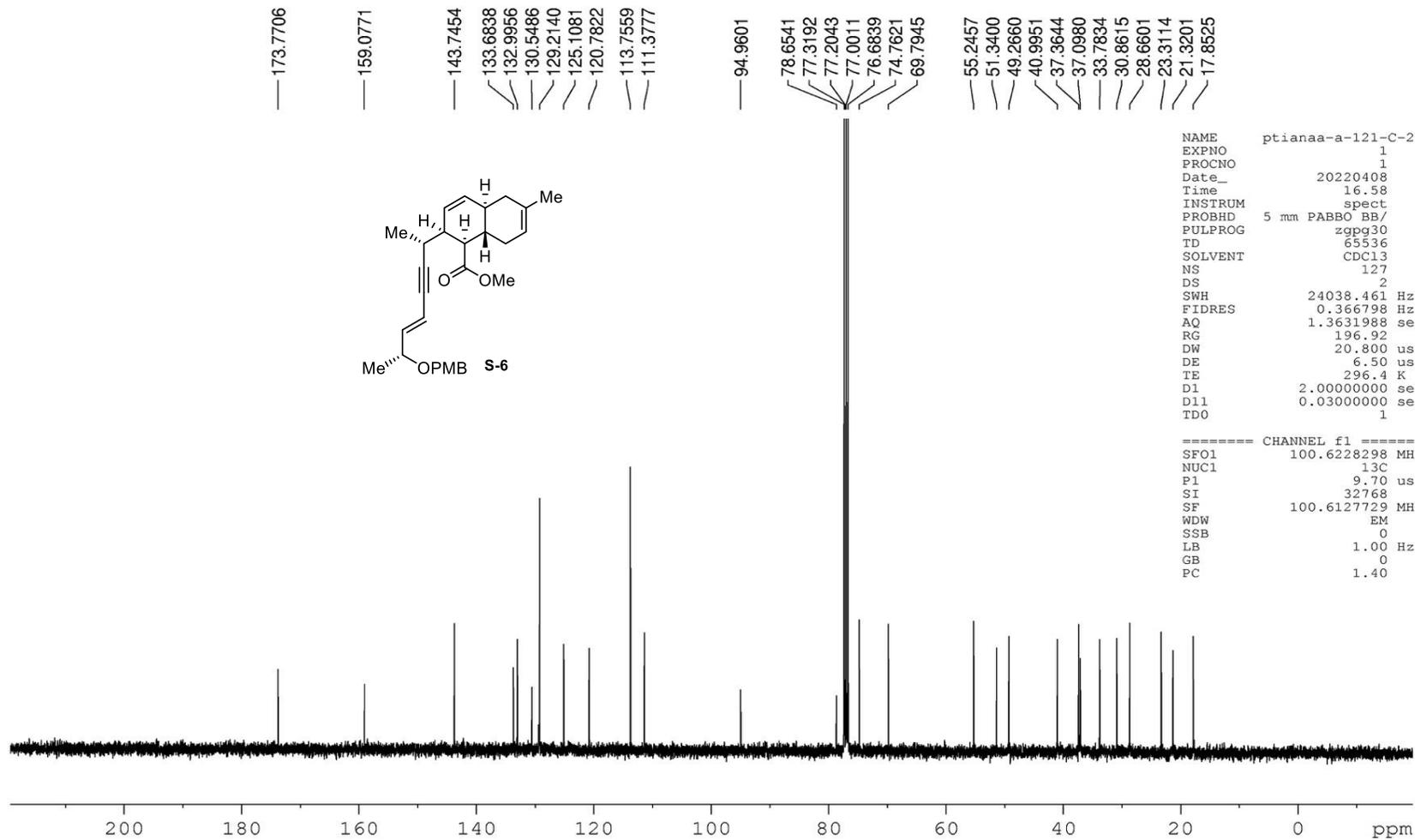


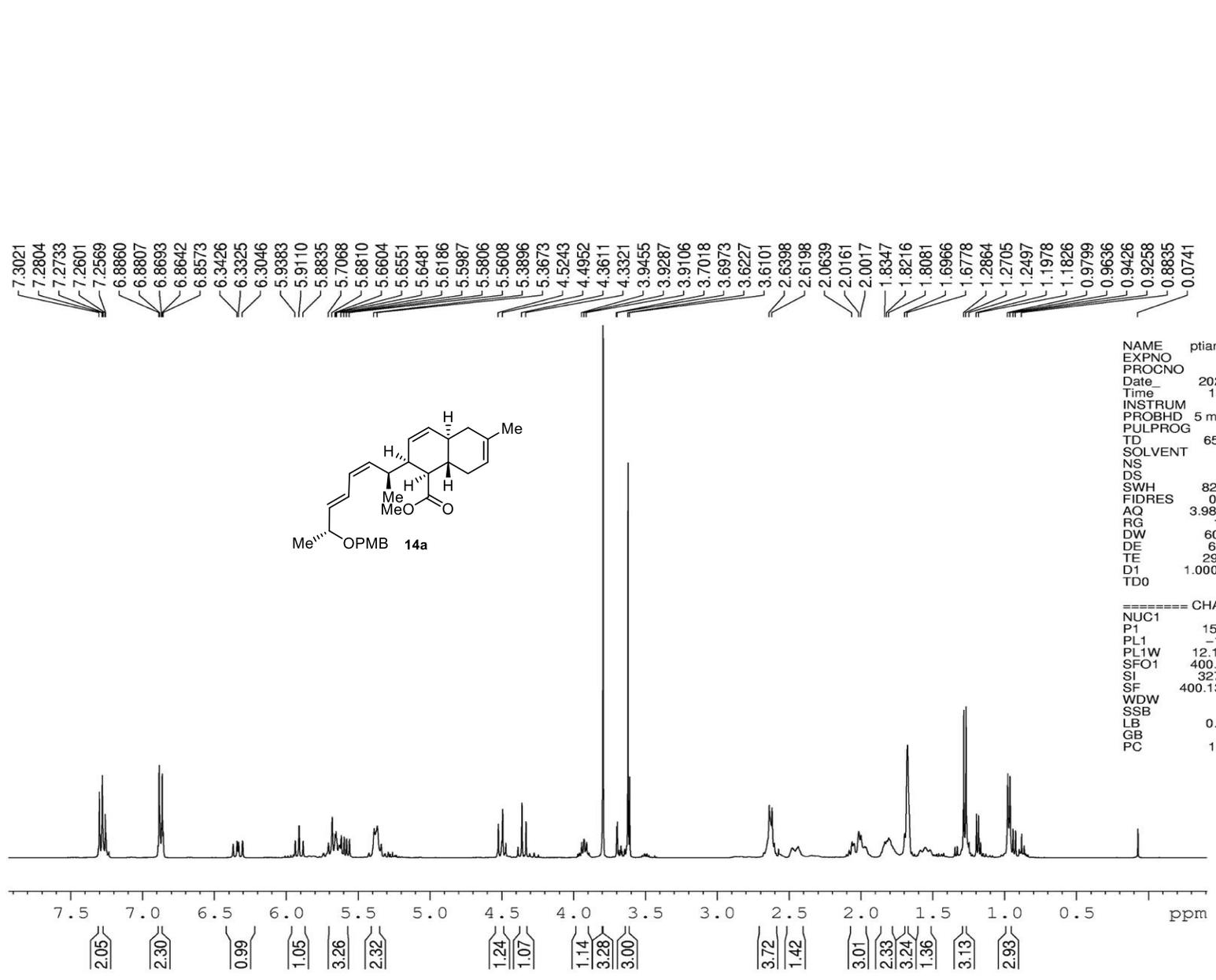
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PROCNO 1
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TD 65536
SOLVENT CDCl3
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DS 2
SWH 8012.820 Hz
FIDRES 0.122266 Hz
AQ 4.0894966 sec
RG 62.93
DW 62.400 usec
DE 6.50 usec
TE 295.5 K
D1 1.00000000 sec
TD0 1

===== CHANNEL f1 =====
SFO1 400.1324710 MHz
NUC1 1H
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GB 0
PC 1.00

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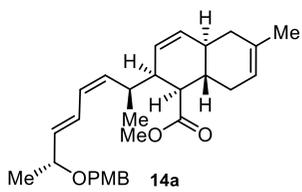
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PULPROG  zg30
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SOLVENT  CDCl3
NS        16
DS        2
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FIDRES   0.125483 Hz
AQ        3.9846387 sec
RG        114
DW        60.800 usec
DE        6.00 usec
TE        293.3 K
D1        1.0000000 sec
TD0       1

===== CHANNEL f1 =====
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PL1       -1.00 dB
PL1W      12.17476940 W
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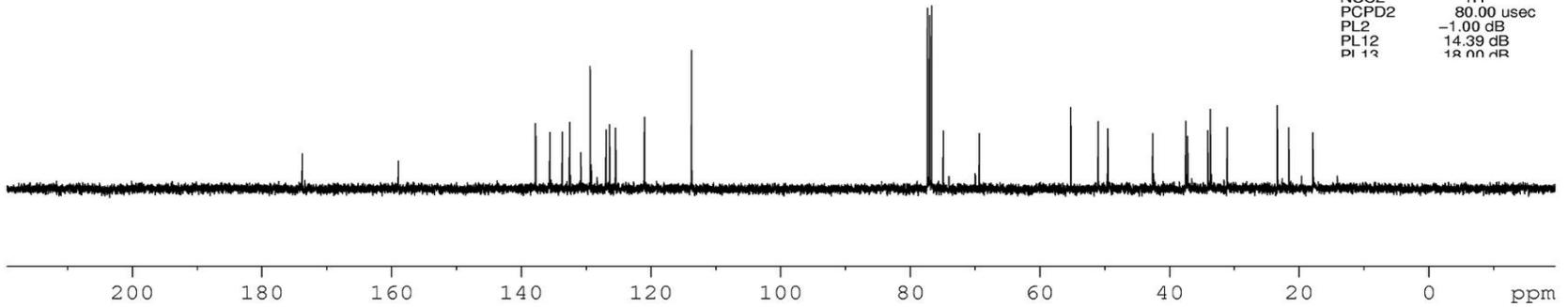
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 — 158.9887
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 126.3663
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 120.9994
 113.7064

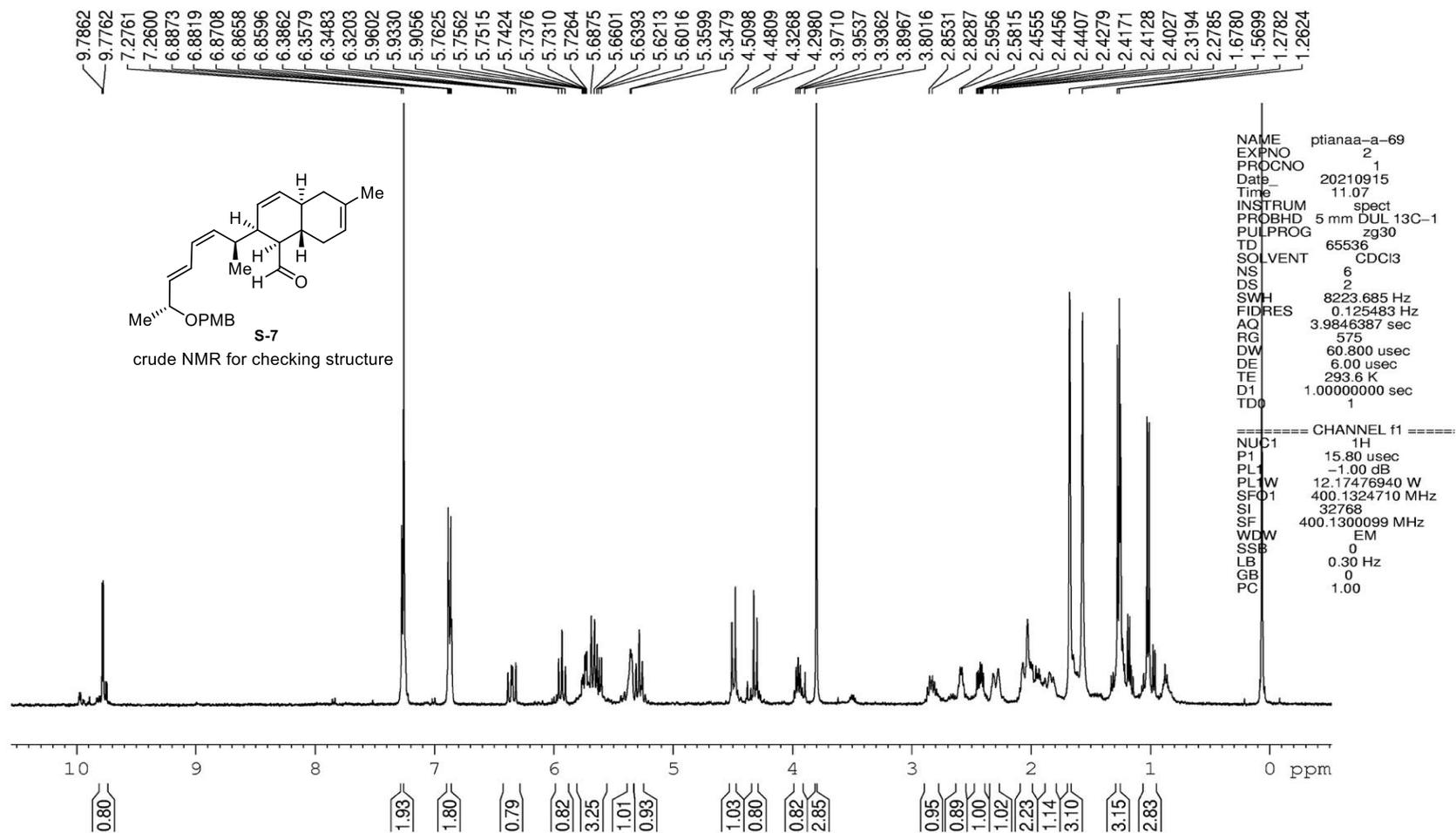
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 34.0629
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 21.5542
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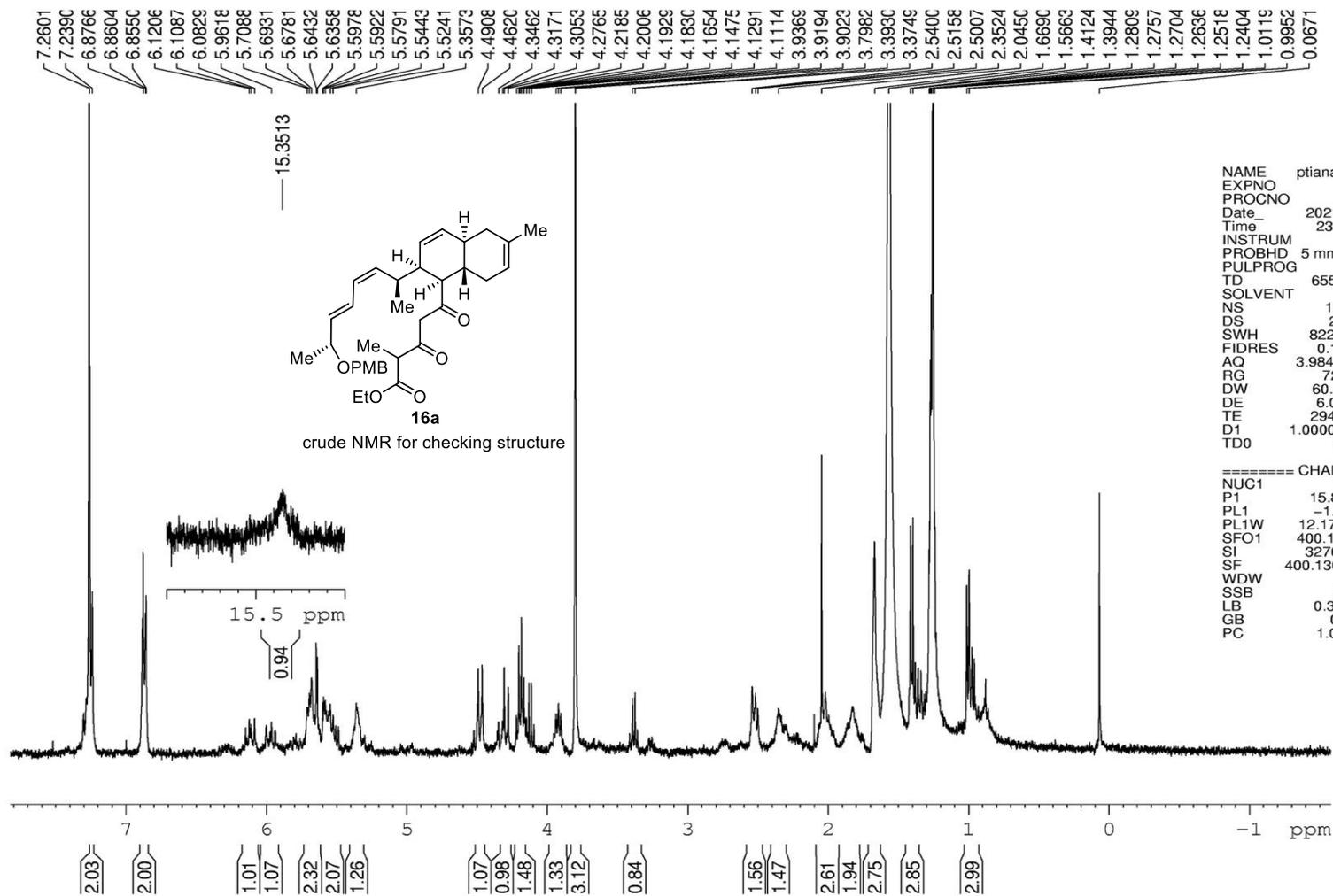


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 TD 65536
 SOLVENT CDCl3
 NS 117
 DS 1
 SWH 24038.461 Hz
 FIDRES 0.366798 Hz
 AQ 1.3631988 sec
 RG 2050
 DW 20.800 usec
 DE 6.00 usec
 TE 293.4 K
 D1 2.00000000 sec
 D11 0.03000000 sec
 TD0 1

===== CHANNEL f1 =====
 NUC1 13C
 P1 40.00 usec
 PL1 -3.00 dB
 PL1W 60.64365387 W
 SFO1 100.6228298 MHz
 ===== CHANNEL f2 =====
 CPDPRG2 waltz16
 NUC2 1H
 PCPD2 80.00 usec
 PL2 -1.00 dB
 PL12 14.39 dB
 PL13 18.00 dB





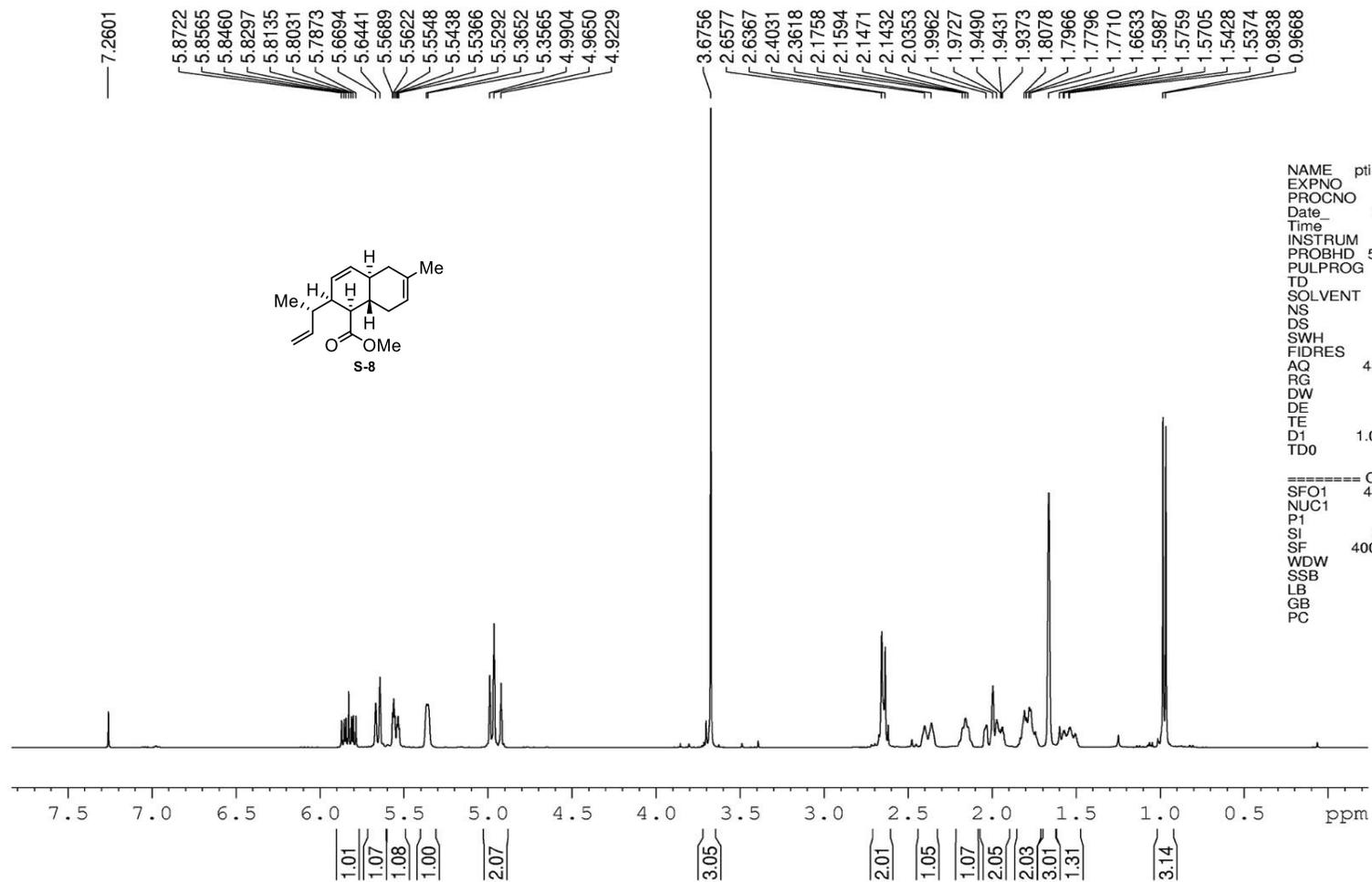


```

NAME ptianaa-a-55
EXPNO 1
PROCNO 1
Date_ 20210726
Time_ 23.50
INSTRUM spect
PROBHD 5 mm DUL 13C-1
PULPROG zg30
TD 65536
SOLVENT CDCl3
NS 16
DS 2
SWH 8223.685 Hz
FIDRES 0.125483 Hz
AQ 3.9846387 sec
RG 724
DW 60.800 usec
DE 6.00 usec
TE 294.7 K
D1 1.0000000 sec
TD0 1

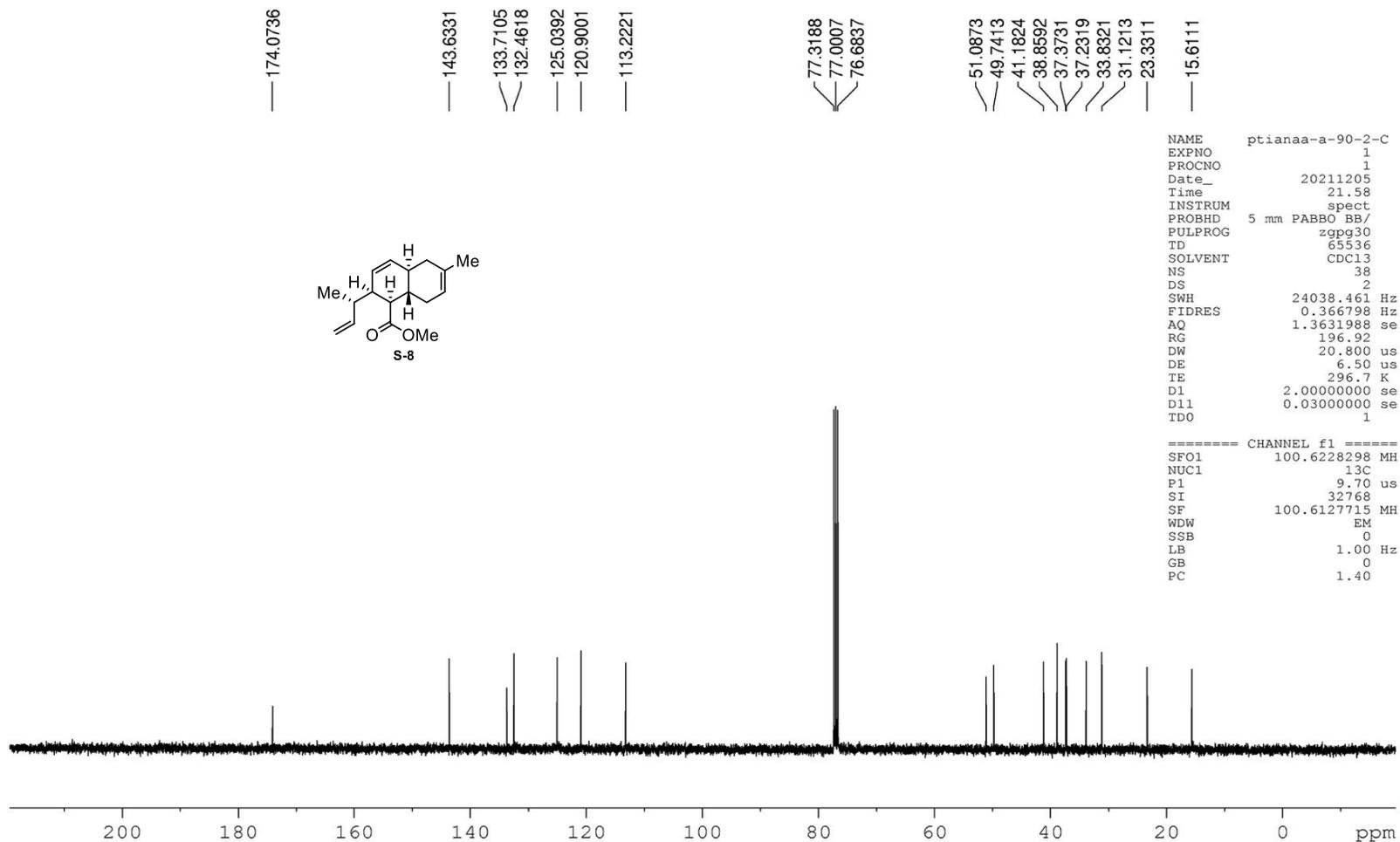
===== CHANNEL f1 =====
NUC1 1H
P1 15.80 usec
PL1 -1.00 dB
PL1W 12.17476940 W
SFO1 400.1324710 MHz
SI 32768
SF 400.1300099 MHz
WDW EM
SSB 0
LB 0.30 Hz
GB 0
PC 1.00

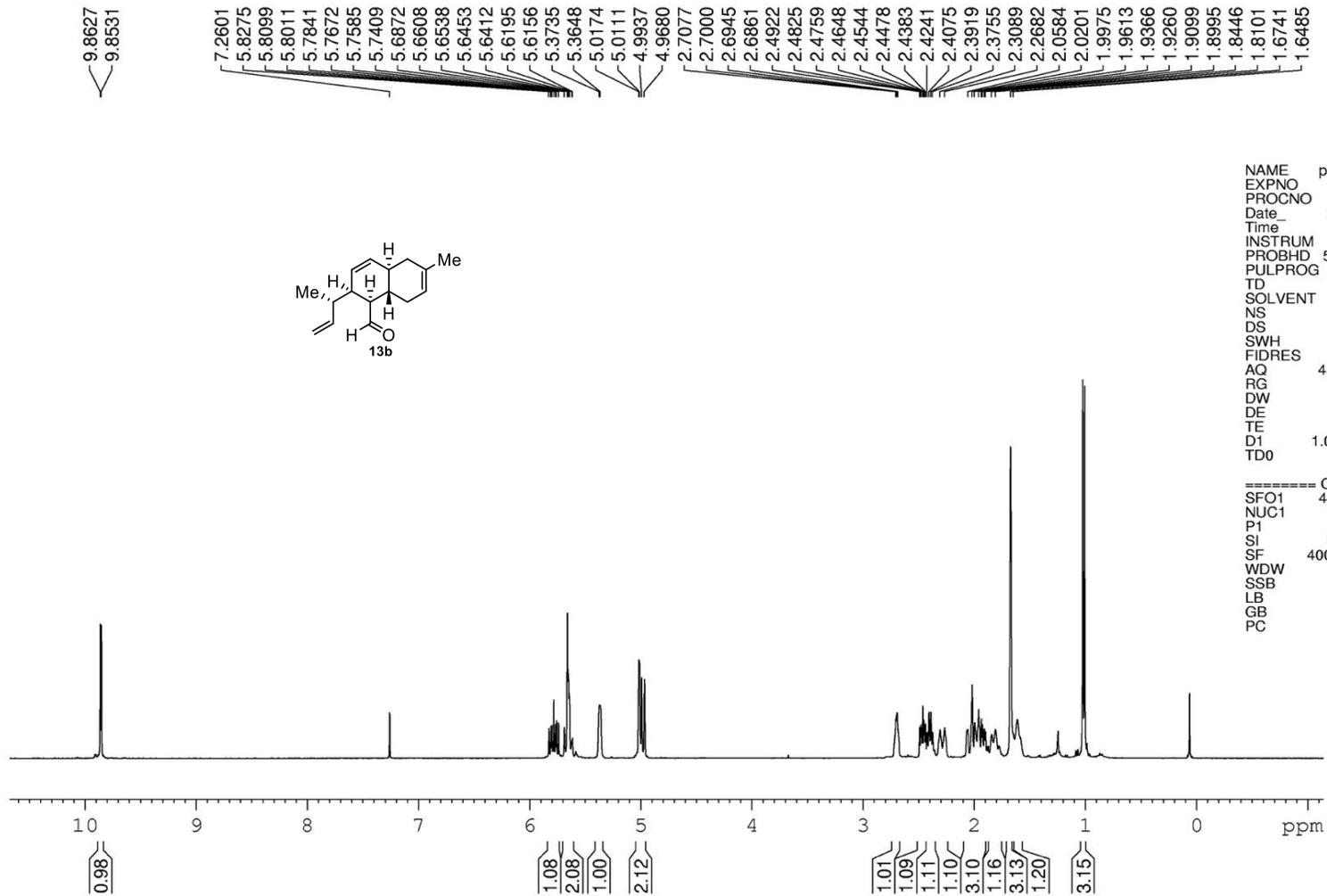
```



NAME ptianaa-a-90-2
 EXPNO 1
 PROCNO 1
 Date_ 20211205
 Time 21.57
 INSTRUM spect
 PROBHD 5 mm PABBO BB/
 PULPROG zg30
 TD 65536
 SOLVENT CDCl3
 NS 10
 DS 2
 SWH 8012.820 Hz
 FIDRES 0.122266 Hz
 AQ 4.0894966 sec
 RG 62.93
 DW 62.400 usec
 DE 6.50 usec
 TE 296.3 K
 D1 1.00000000 sec
 TD0 1

===== CHANNEL f1 =====
 SFO1 400.1324710 MHz
 NUC1 1H
 P1 14.50 usec
 SI 65536
 SF 400.1300100 MHz
 WDW EM
 SSB 0
 LB 0.30 Hz
 GB 0
 PC 1.00



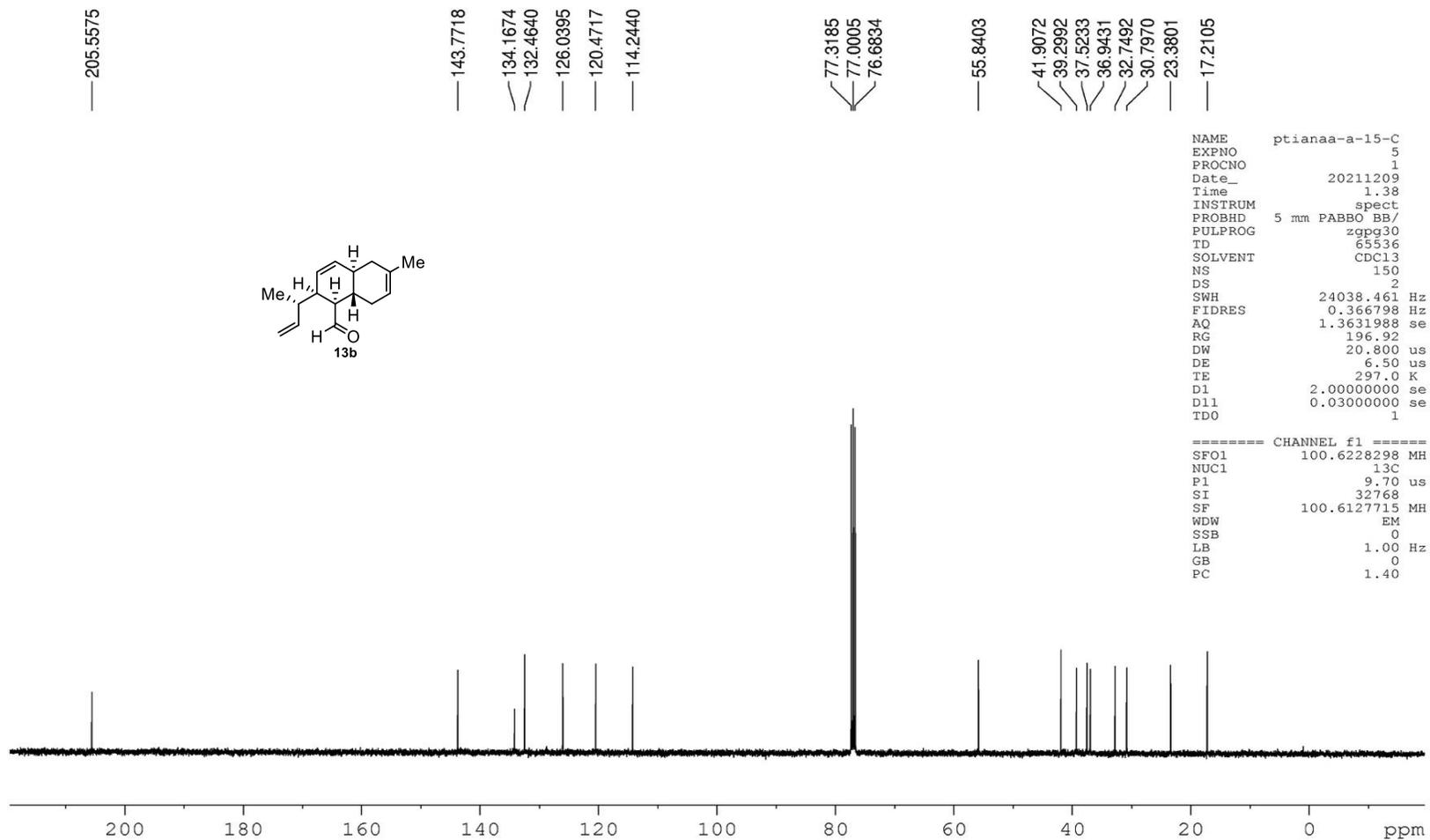


```

NAME ptianaa-a-15
EXPNO 5
PROCNO 1
Date_ 20211209
Time 1.35
INSTRUM spect
PROBHD 5 mm PABBO BB/
PULPROG zg30
TD 65536
SOLVENT CDCl3
NS 11
DS 2
SWH 8012.820 Hz
FIDRES 0.122266 Hz
AQ 4.0894966 sec
RG 62.93
DW 62.400 usec
DE 6.50 usec
TE 296.3 K
D1 1.00000000 sec
TD0 1

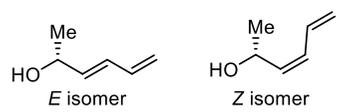
===== CHANNEL f1 =====
SFO1 400.1324710 MHz
NUC1 1H
P1 14.50 usec
SI 65536
SF 400.1300100 MHz
WDW EM
SSB 0
LB 0.30 Hz
GB 0
PC 1.00

```



6.3312
6.3219
6.3058
6.2804
6.2456
6.2431
6.2193
6.2168
6.2074
6.2061
6.1799
6.1790
5.7836
5.7678
5.7457
5.7449
5.7300
5.7291
5.2393
5.2353
5.1945
5.1117
5.1080
5.0863
5.0837
4.3744
4.3587
4.3438

1.3019
1.2858



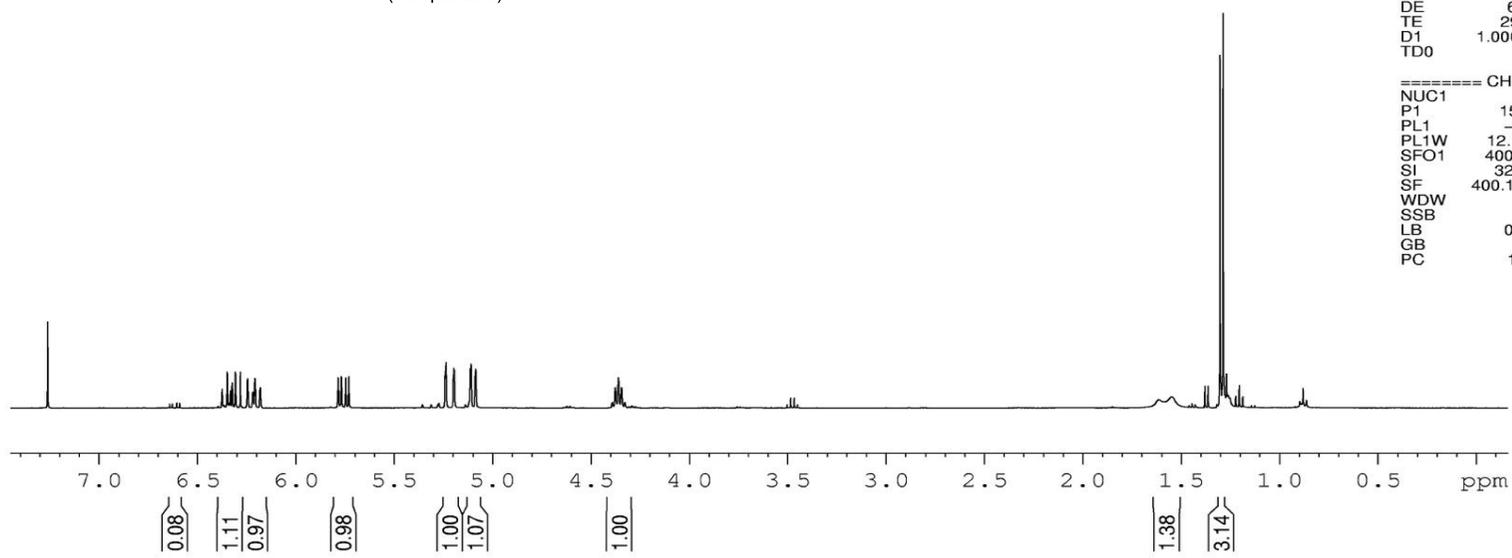
S-10
E/Z >13:1
 (inseparable)

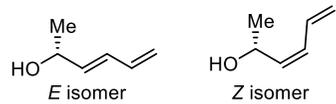
```

NAME ptianaa-a-s13
EXPNO 1
PROCNO 1
Date_ 20220515
Time 0.16
INSTRUM spect
PROBHD 5 mm DUL 13C-1
PULPROG zg30
TD 65536
SOLVENT CDCl3
NS 10
DS 2
SWH 8223.685 Hz
FIDRES 0.125483 Hz
AQ 3.9846387 sec
RG 512
DW 60.800 usec
DE 6.00 usec
TE 295.6 K
D1 1.0000000 sec
TD0 1
  
```

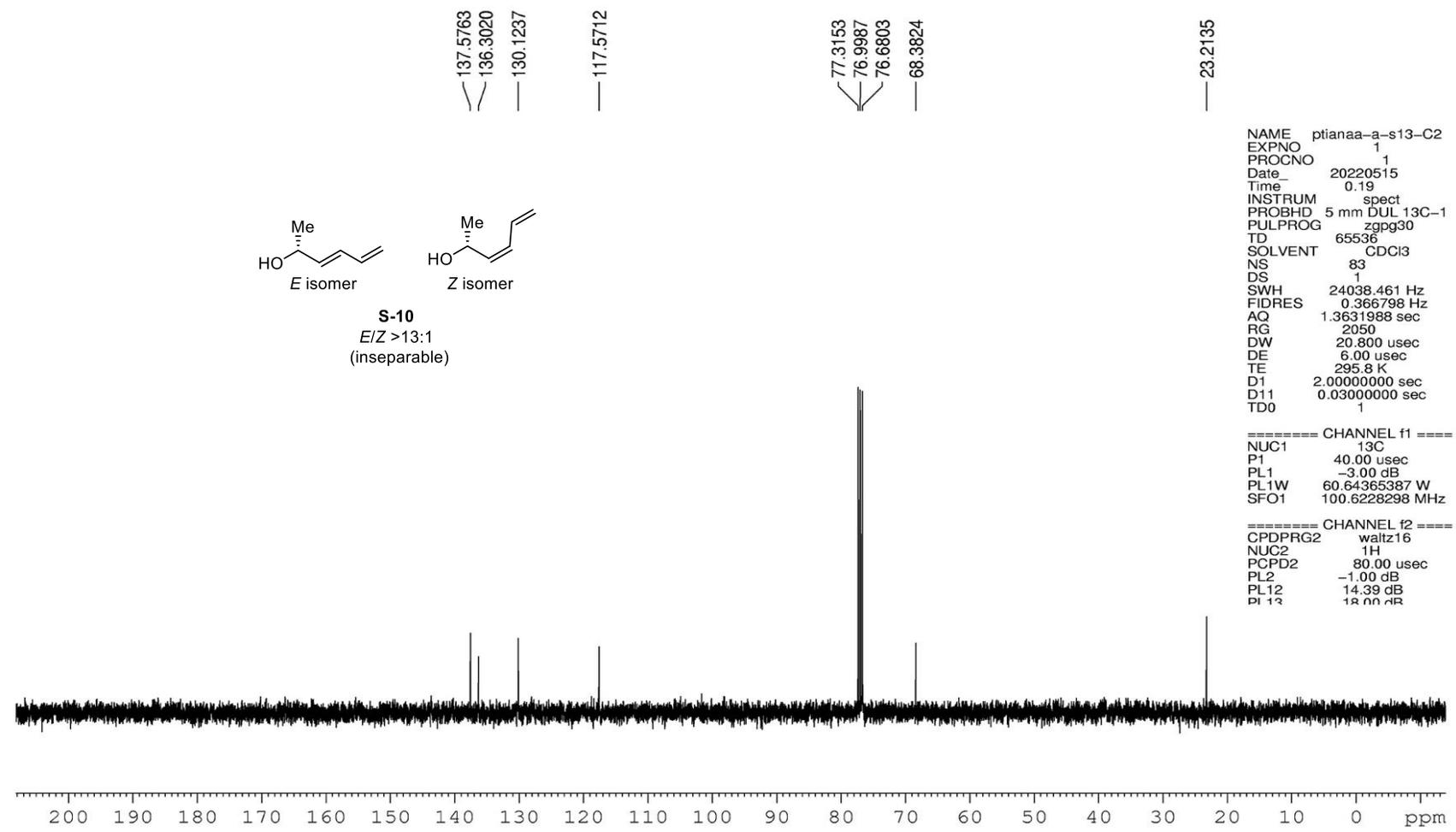
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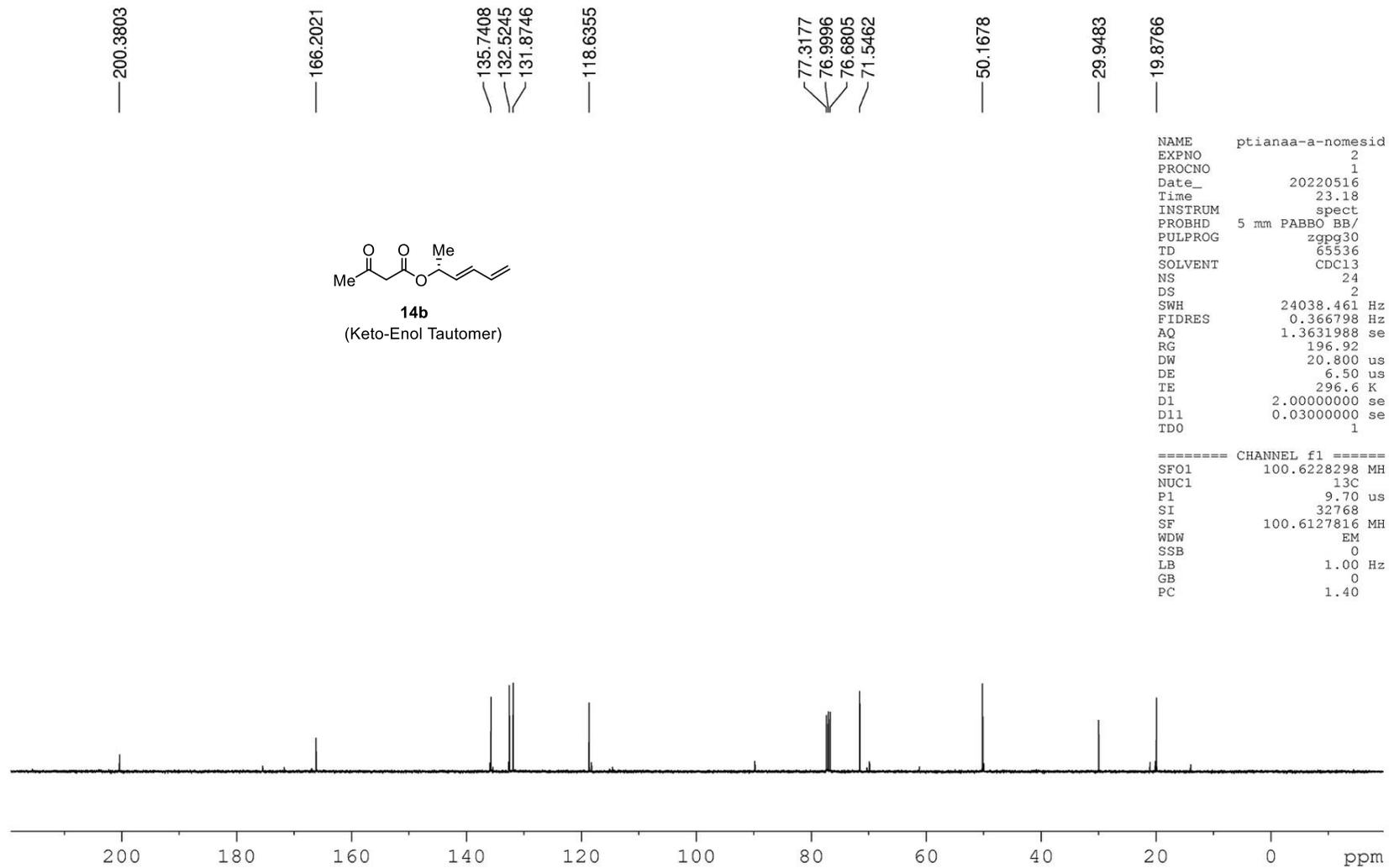
===== CHANNEL f1 =====
NUC1 1H
P1 15.80 usec
PL1 -1.00 dB
PL1W 12.17476940 W
SFO1 400.1324710 MHz
SI 32768
SF 400.1300098 MHz
WDW EM
SSB 0
LB 0.30 Hz
GB 0
PC 1.00
  
```





S-10
E/Z >13:1
 (inseparable)





```

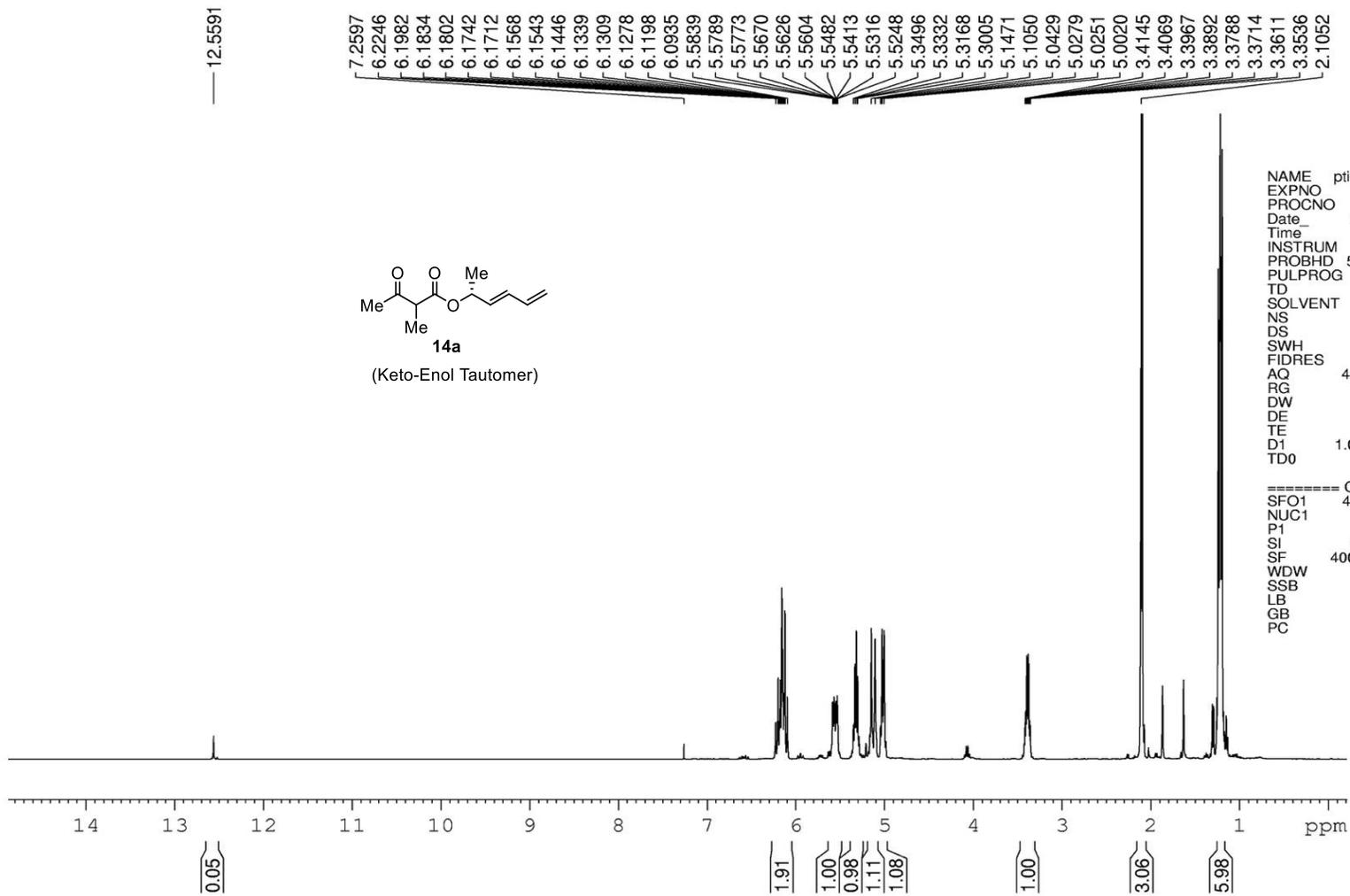
NAME      ptianaa-a-nomesid
EXPNO     2
PROCNO    1
Date_     20220516
Time      23.18
INSTRUM   spect
PROBHD    5 mm PABBO BB/
PULPROG   zgpg30
TD         65536
SOLVENT   CDC13
NS         24
DS         2
SWH        24038.461 Hz
FIDRES     0.366798 Hz
AQ         1.3631988 se
RG         196.92
DW         20.800 us
DE         6.50 us
TE         296.6 K
D1         2.00000000 se
D11        0.03000000 se
TD0        1

```

```

===== CHANNEL f1 =====
SFO1      100.6228298 MH
NUC1      13C
P1         9.70 us
SI        32768
SF         100.6127816 MH
WDW        EM
SSB        0
LB         1.00 Hz
GB         0
PC         1.40

```

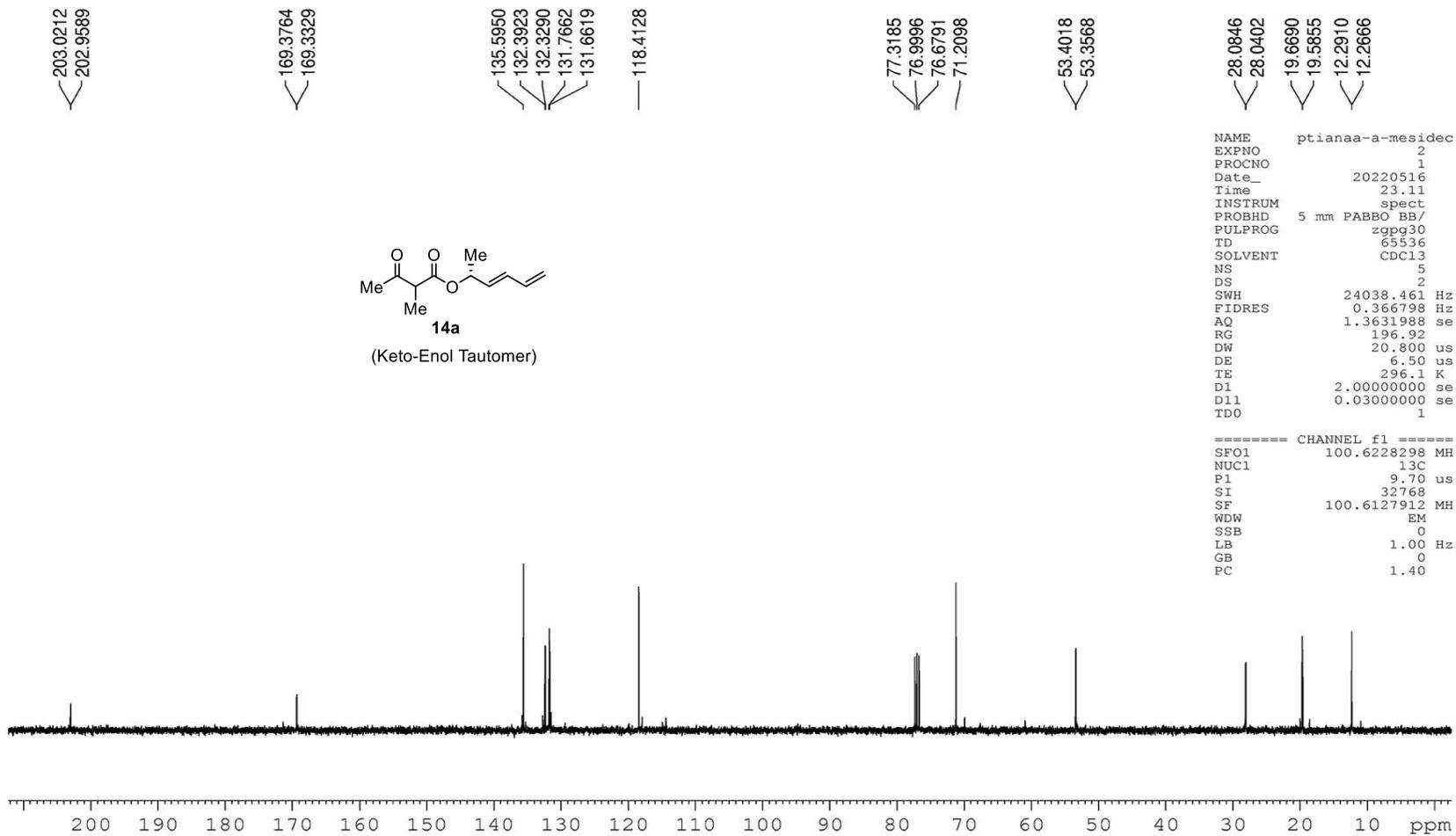


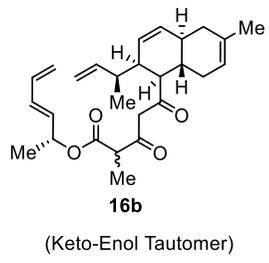
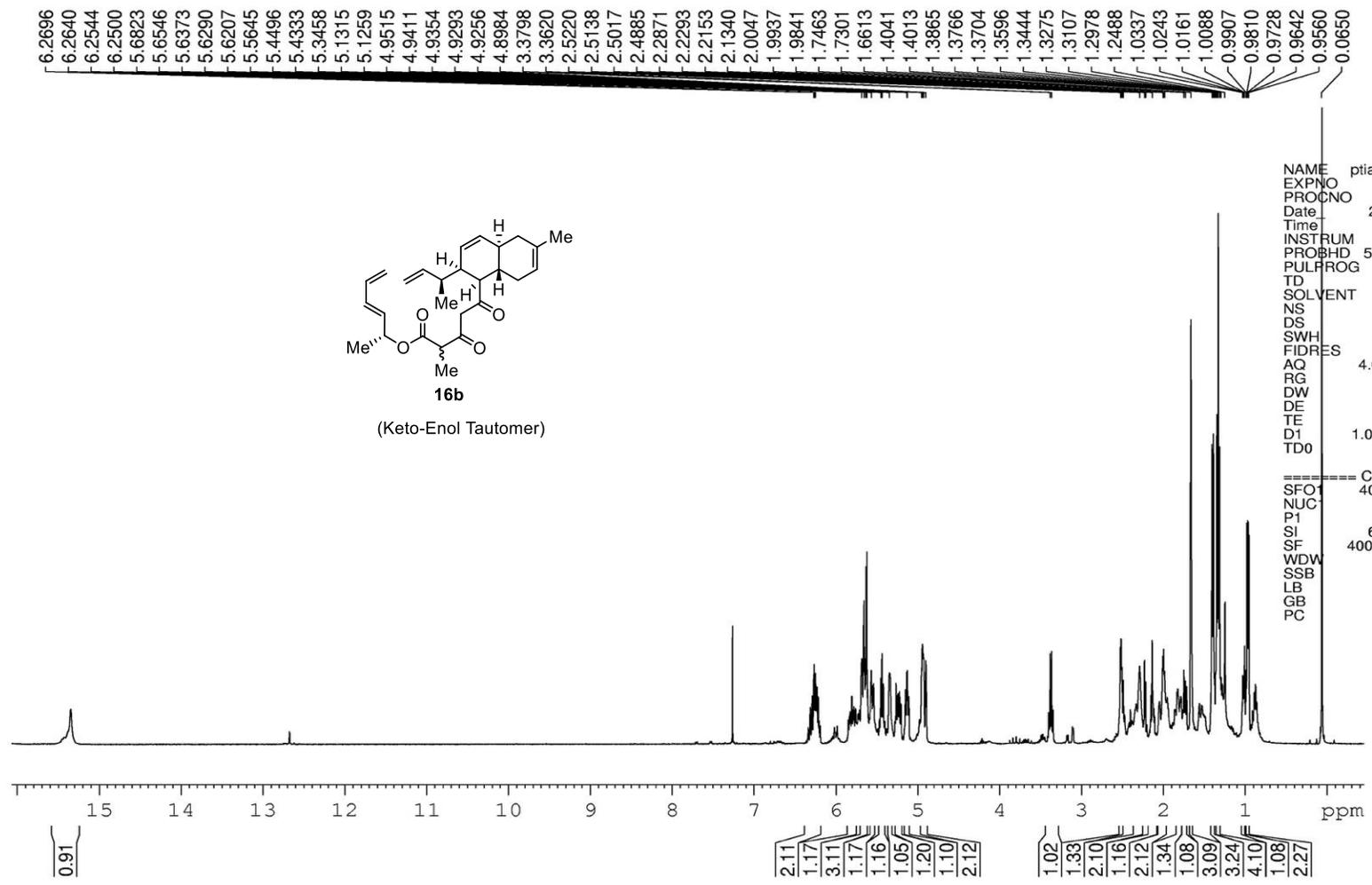
```

NAME ptianaa-a-mesidec
EXPNO 2
PROCNO 1
Date_ 20220516
Time 23.10
INSTRUM spect
PROBHD 5 mm PABBO BB/
PULPROG zg30
TD 65536
SOLVENT CDCl3
NS 2
DS 2
SWH 8012.820 Hz
FIDRES 0.122266 Hz
AQ 4.0894966 sec
RG 9.04
DW 62.400 usec
DE 6.50 usec
TE 296.0 K
D1 1.00000000 sec
TD0 1

===== CHANNEL f1 =====
SFO1 400.1324710 MHz
NUC1 1H
P1 14.50 usec
SI 65536
SF 400.1300095 MHz
WDW EM
SSB 0
LB 0.30 Hz
GB 0
PC 1.00

```



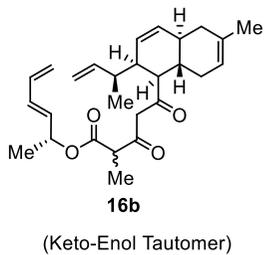
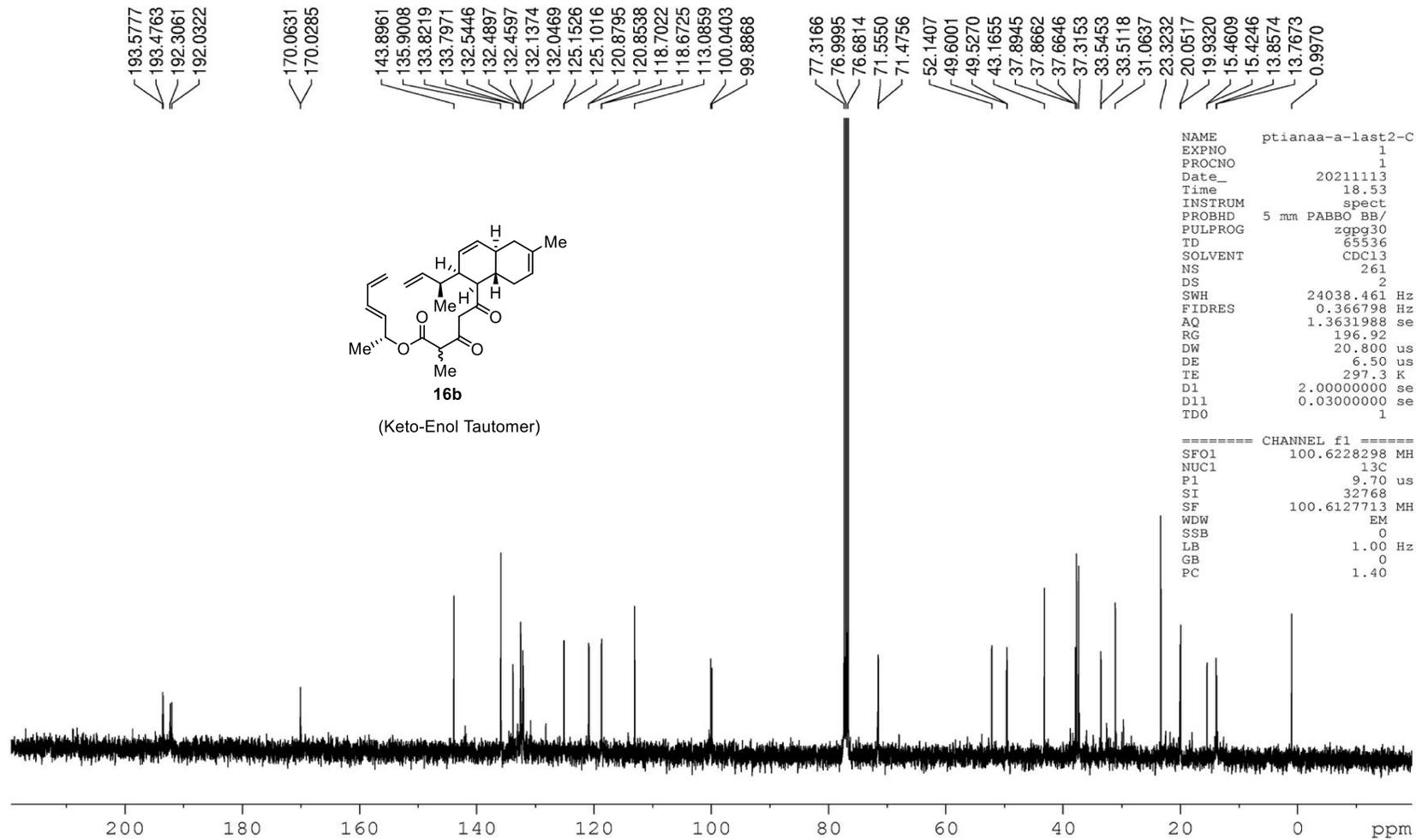


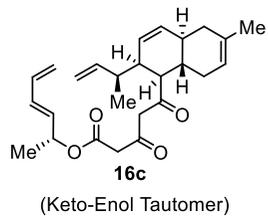
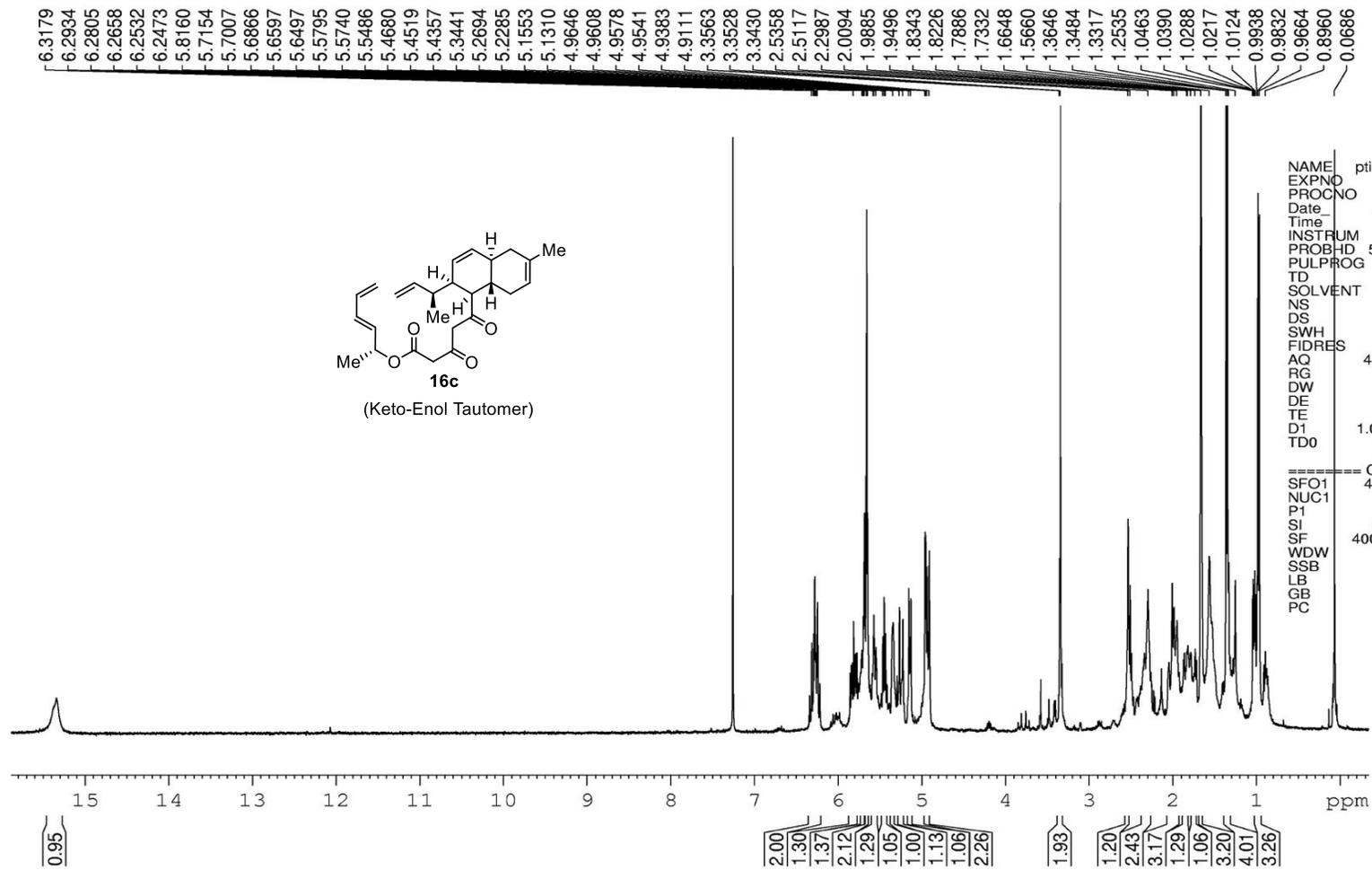
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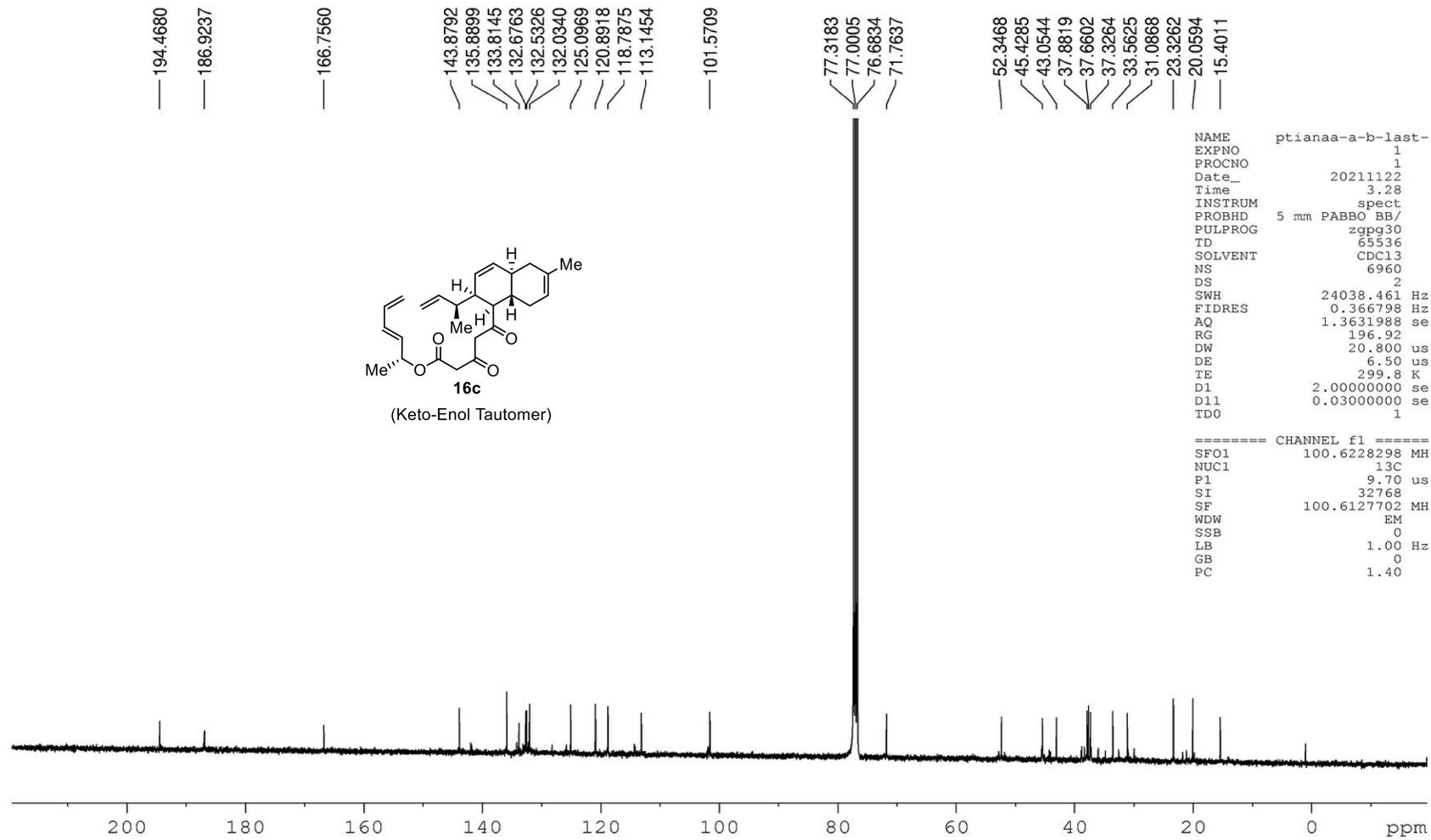
NAME      ptianaa-a-last2
EXPNO     1
PROCNO    1
Date_     20211113
Time      18.51
INSTRUM   spect
PROBHD    5 mm PABBO BB/
PULPROG   zg30
TD         65536
SOLVENT   CDCl3
NS         4
DS         2
SWH        8012.820 Hz
FIDRES     0.122266 Hz
AQ         4.0894966 sec
RG         31.55
DW         62.400 usec
DE         6.50 usec
TE         296.6 K
D1         1.00000000 sec
TD0        1

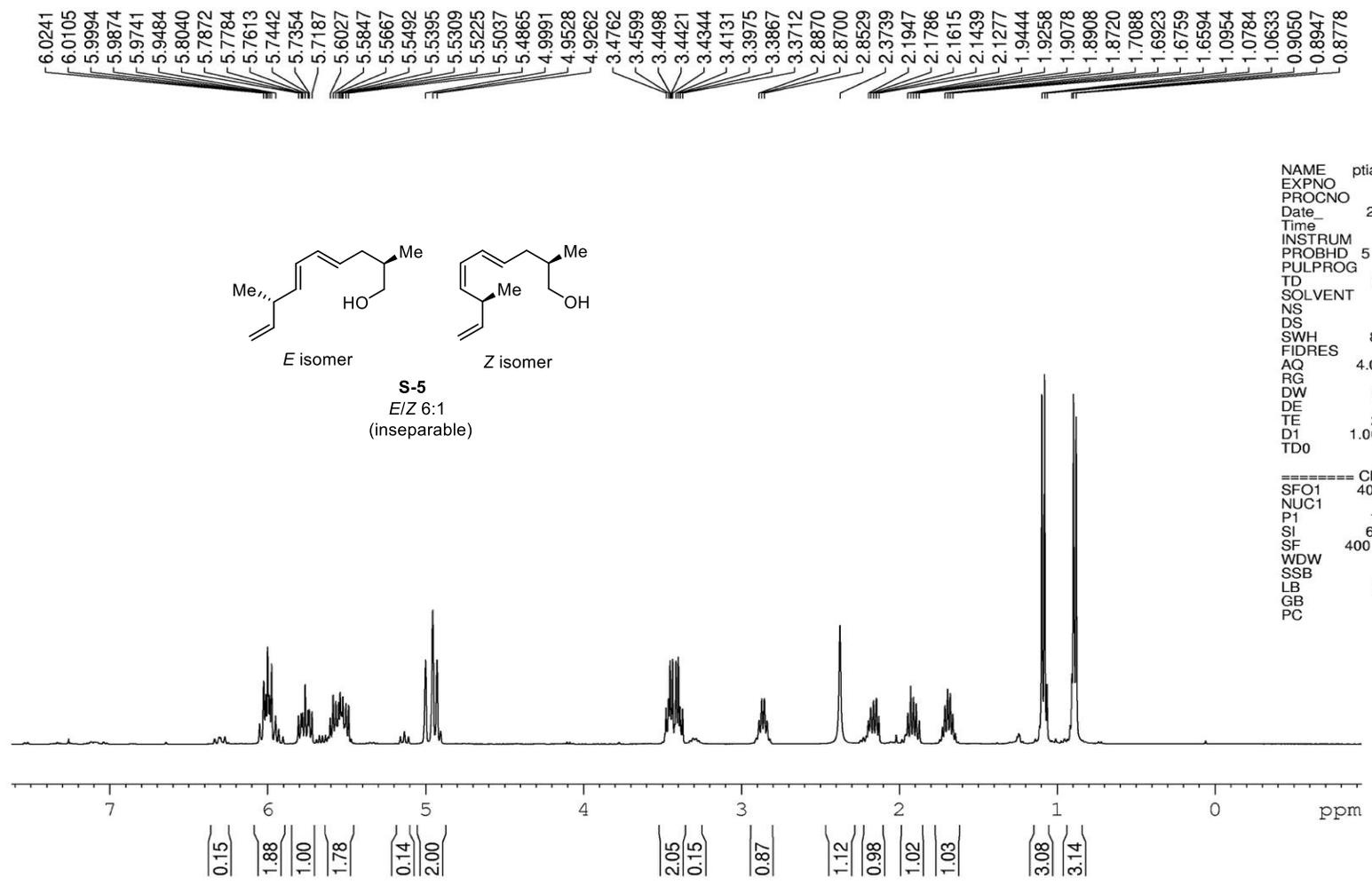
===== CHANNEL f1 =====
SFO1      400.1324710 MHz
NUC1       1H
P1         14.50 usec
SI         65536
SF         400.1300101 MHz
WDW        EM
SSB        0
LB         0.30 Hz
GB         0
PC         1.00

```







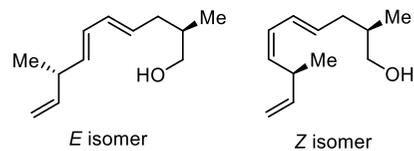


6.0241
 6.0105
 5.9994
 5.9874
 5.9741
 5.9484
 5.8040
 5.7872
 5.7784
 5.7613
 5.7442
 5.7354
 5.7187
 5.6027
 5.5847
 5.5667
 5.5492
 5.5395
 5.5309
 5.5225
 5.5037
 5.4865
 4.9991
 4.9528
 4.9262
 3.4762
 3.4599
 3.4498
 3.4421
 3.4344
 3.4131
 3.3975
 3.3867
 3.3712
 2.8870
 2.8700
 2.8529
 2.3739
 2.1947
 2.1786
 2.1615
 2.1439
 2.1277
 1.9444
 1.9258
 1.9078
 1.8908
 1.8720
 1.7088
 1.6923
 1.6759
 1.6594
 1.0954
 1.0784
 1.0633
 0.9050
 0.8947
 0.8778

```

NAME ptiaaa-a-3-2
EXPNO 2
PROCNO 1
Date_ 20220112
Time 22.23
INSTRUM spect
PROBHD 5 mm PABBO BB/
PULPROG zg30
TD 65536
SOLVENT CDCl3
NS 16
DS 2
SWH 8012.820 Hz
FIDRES 0.122266 Hz
AQ 4.0894966 sec
RG 11.3
DW 62.400 usec
DE 6.50 usec
TE 294.6 K
D1 1.00000000 sec
TD0 1

===== CHANNEL f1 =====
SFO1 400.1324710 MHz
NUC1 1H
P1 14.50 usec
SI 65536
SF 400.1300100 MHz
WDW EM
SSB 0
LB 0.30 Hz
GB 0
PC 1.00
  
```



S-5
E/Z 6:1
(inseparable)

142.4133
135.7916
131.7675
130.6601
128.8447

112.8824

77.3154
76.9967
76.6787

67.5314

40.1363
36.3835
35.9190

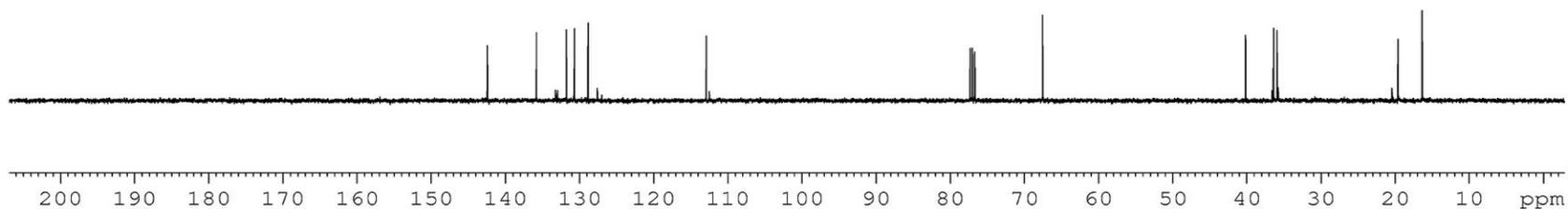
19.6095
16.3463

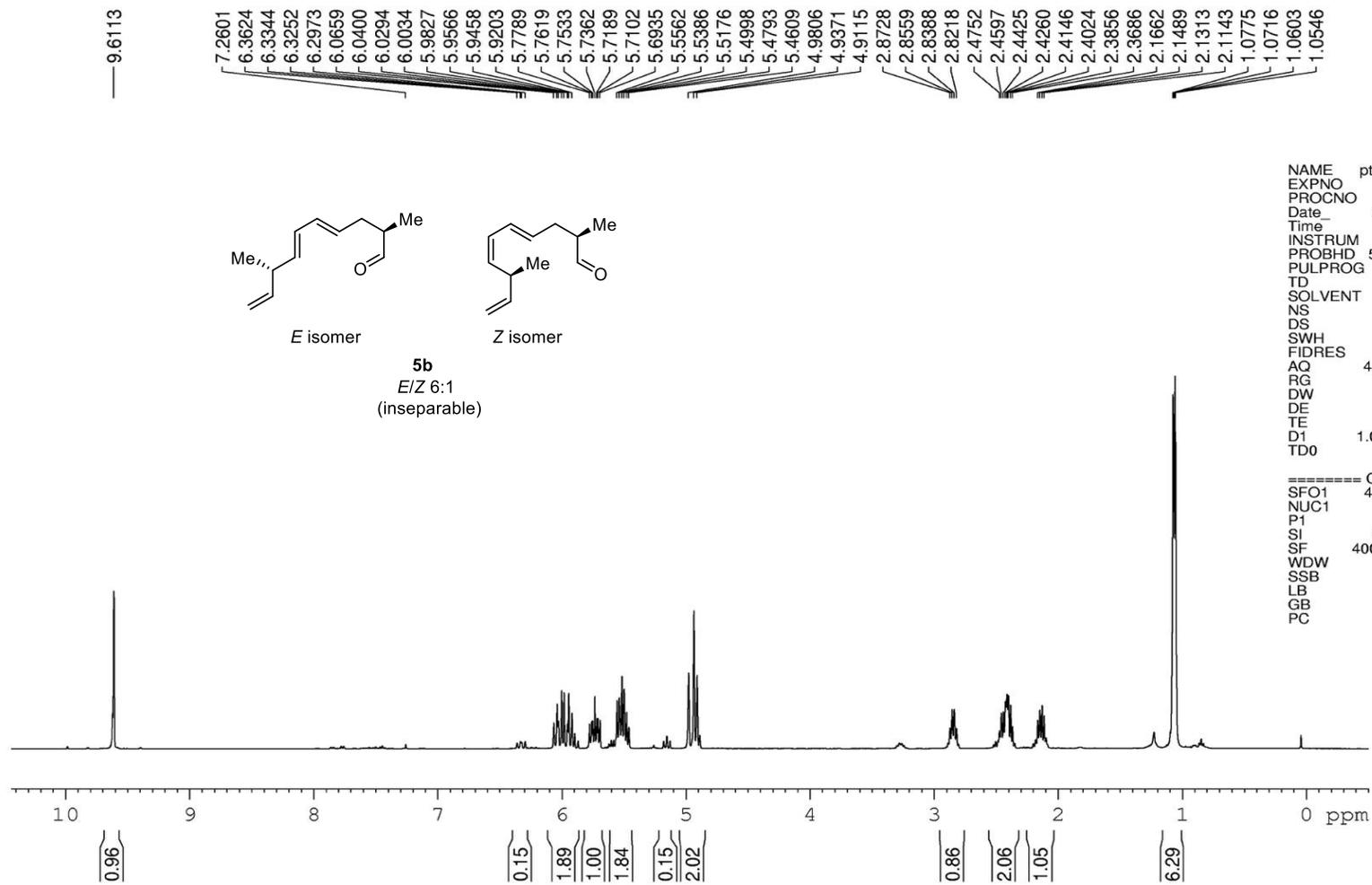
```

NAME      ptianaa-a-3-2-C
EXPNO     2
PROCNO    1
Date_     20220112
Time      22.25
INSTRUM   spect
PROBHD    5 mm PABBO BB/
PULPROG   zgpg30
TD        65536
SOLVENT   CDC13
NS         5
DS         2
SWH       24038.461 Hz
FIDRES    0.366798 Hz
AQ        1.3631988 se
RG        196.92
DW        20.800 us
DE        6.50 us
TE        294.8 K
D1        2.0000000 se
D11       0.0300000 se
TD0       1
  
```

```

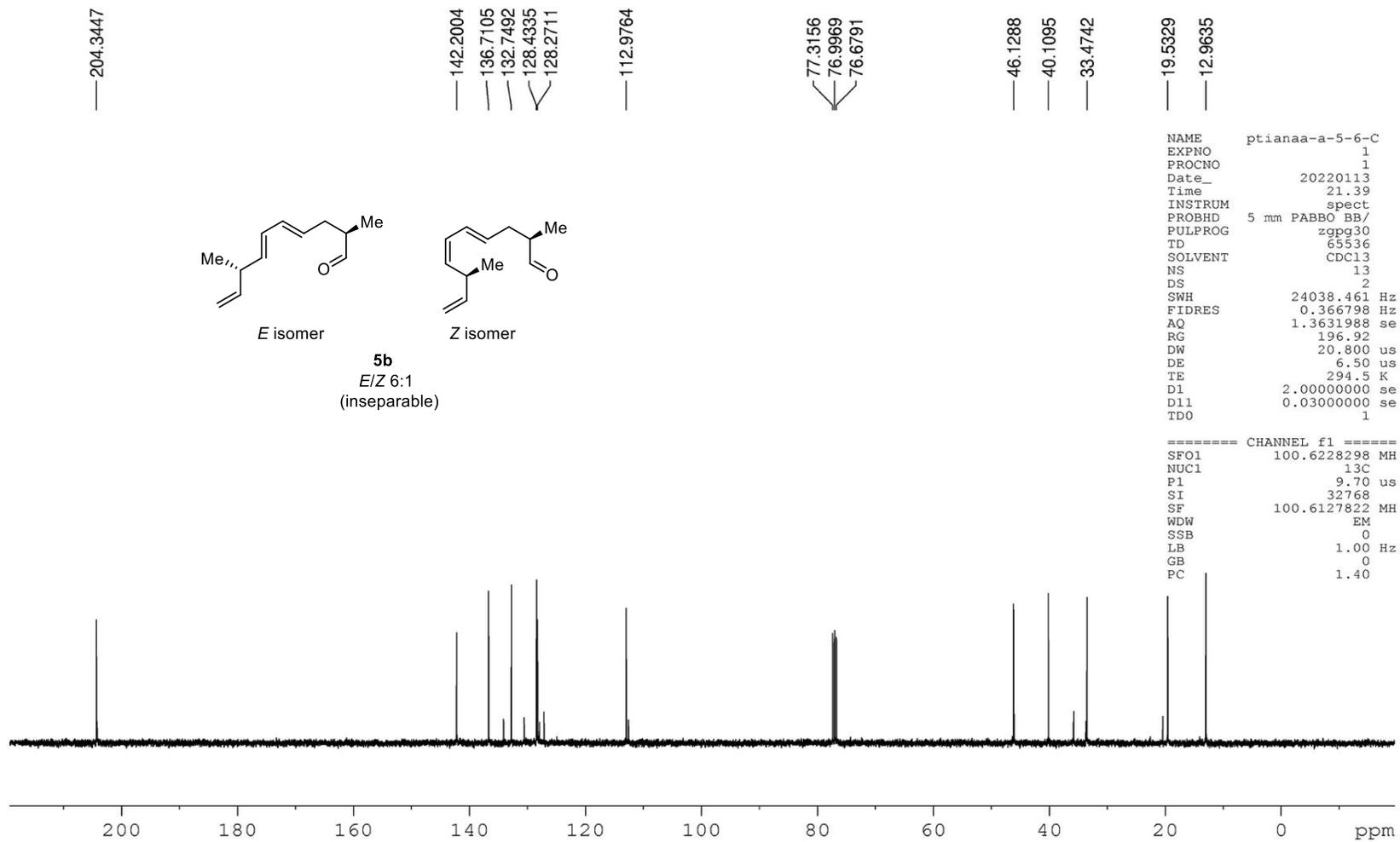
===== CHANNEL f1 =====
SFO1     100.6228298 MH
NUC1     13C
P1       9.70 us
SI       32768
SF       100.6127821 MH
WDW      EM
SSB      0
LB       1.00 Hz
GB       0
PC       1.40
  
```

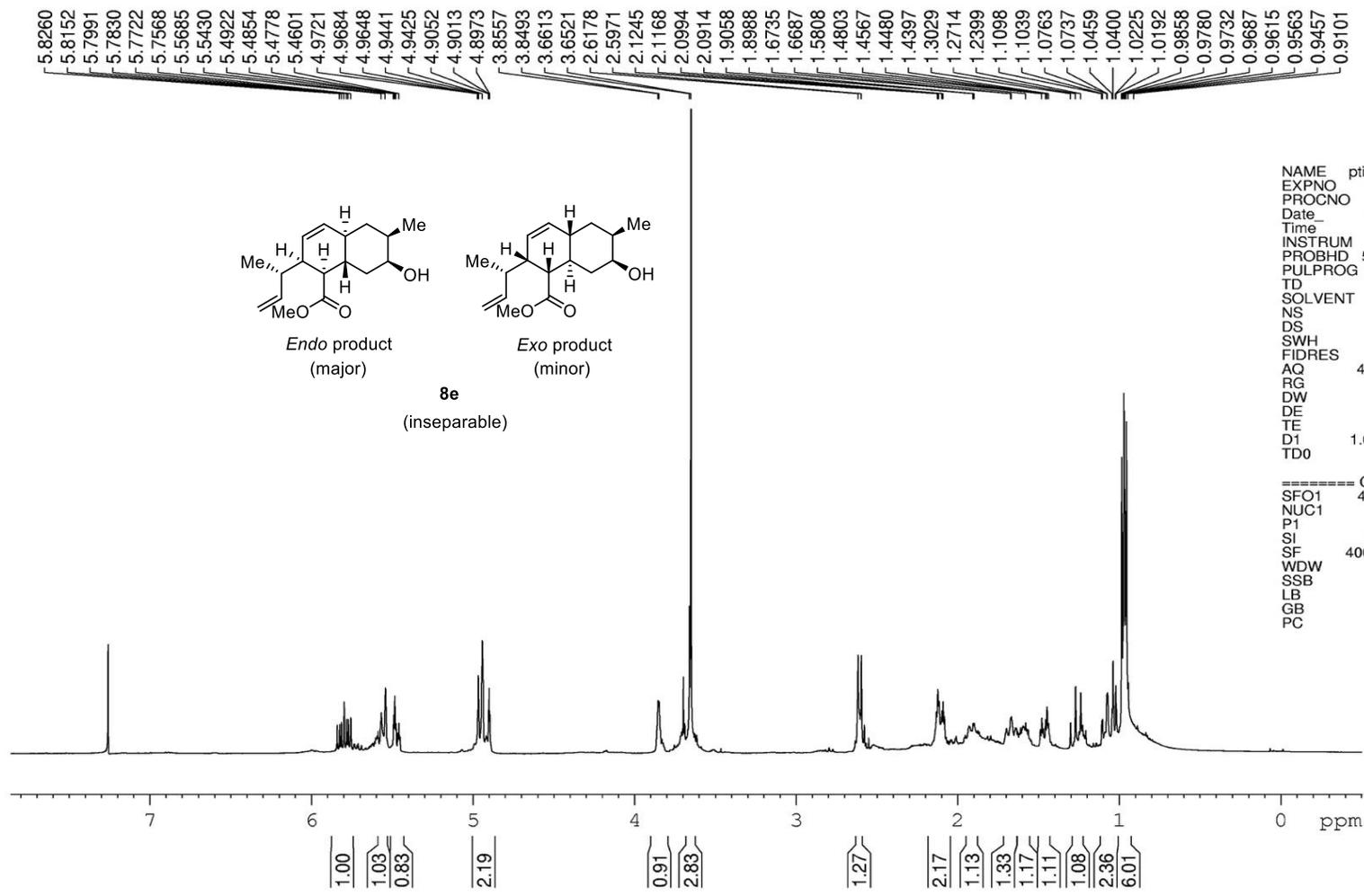




NAME ptianaa-a-5-6
 EXPNO 1
 PROCNO 1
 Date_ 20220113
 Time 21.37
 INSTRUM spect
 PROBHD 5 mm PABBO BB/
 PULPROG zg30
 TD 65536
 SOLVENT CDCl3
 NS 5
 DS 2
 SWH 8012.820 Hz
 FIDRES 0.122266 Hz
 AQ 4.0894966 sec
 RG 12.56
 DW 62.400 usec
 DE 6.50 usec
 TE 294.1 K
 D1 1.00000000 sec
 TD0 1

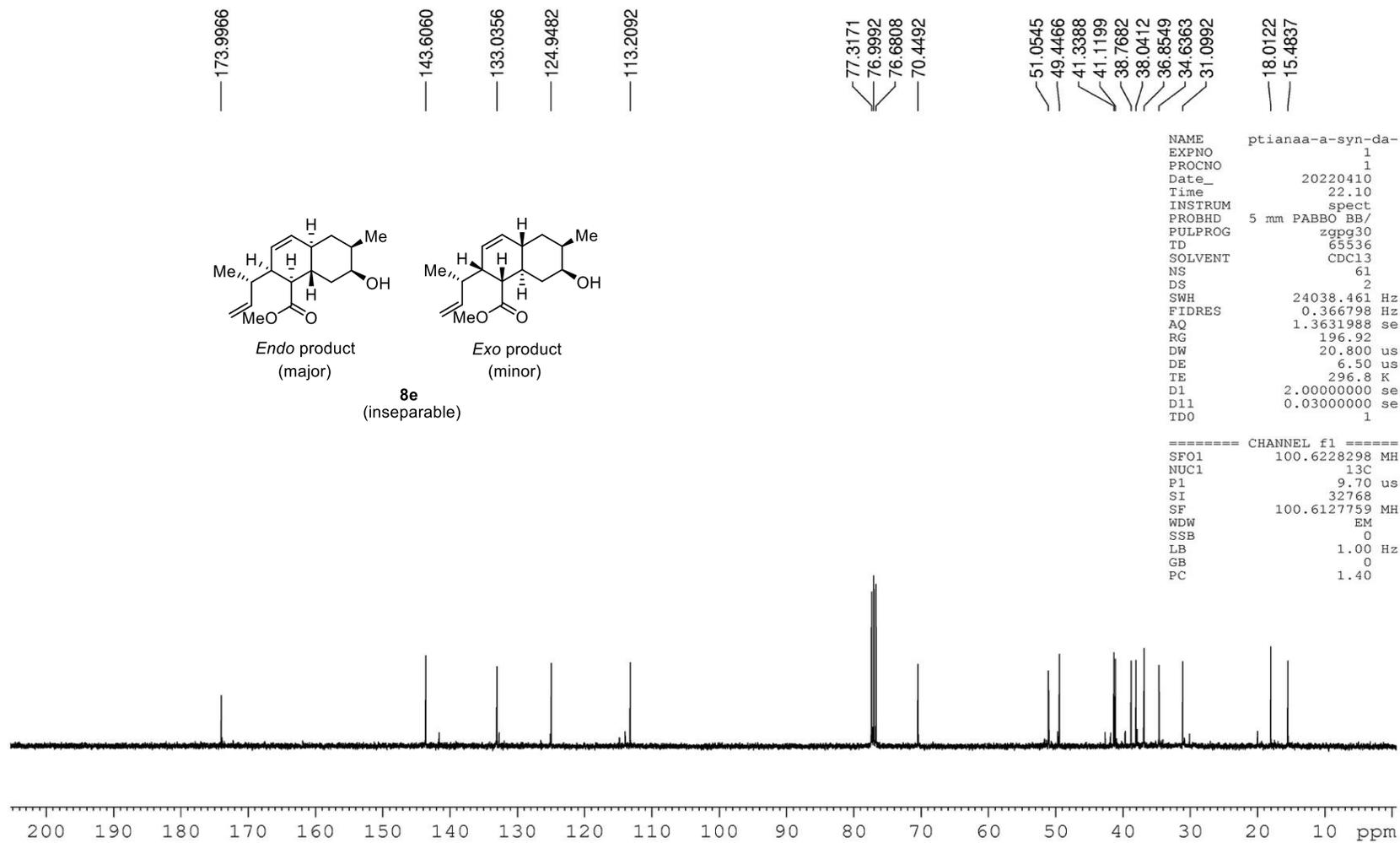
===== CHANNEL f1 =====
 SFO1 400.1324710 MHz
 NUC1 1H
 P1 14.50 usec
 SI 65536
 SF 400.1300098 MHz
 WDW EM
 SSB 0
 LB 0.30 Hz
 GB 0
 PC 1.00

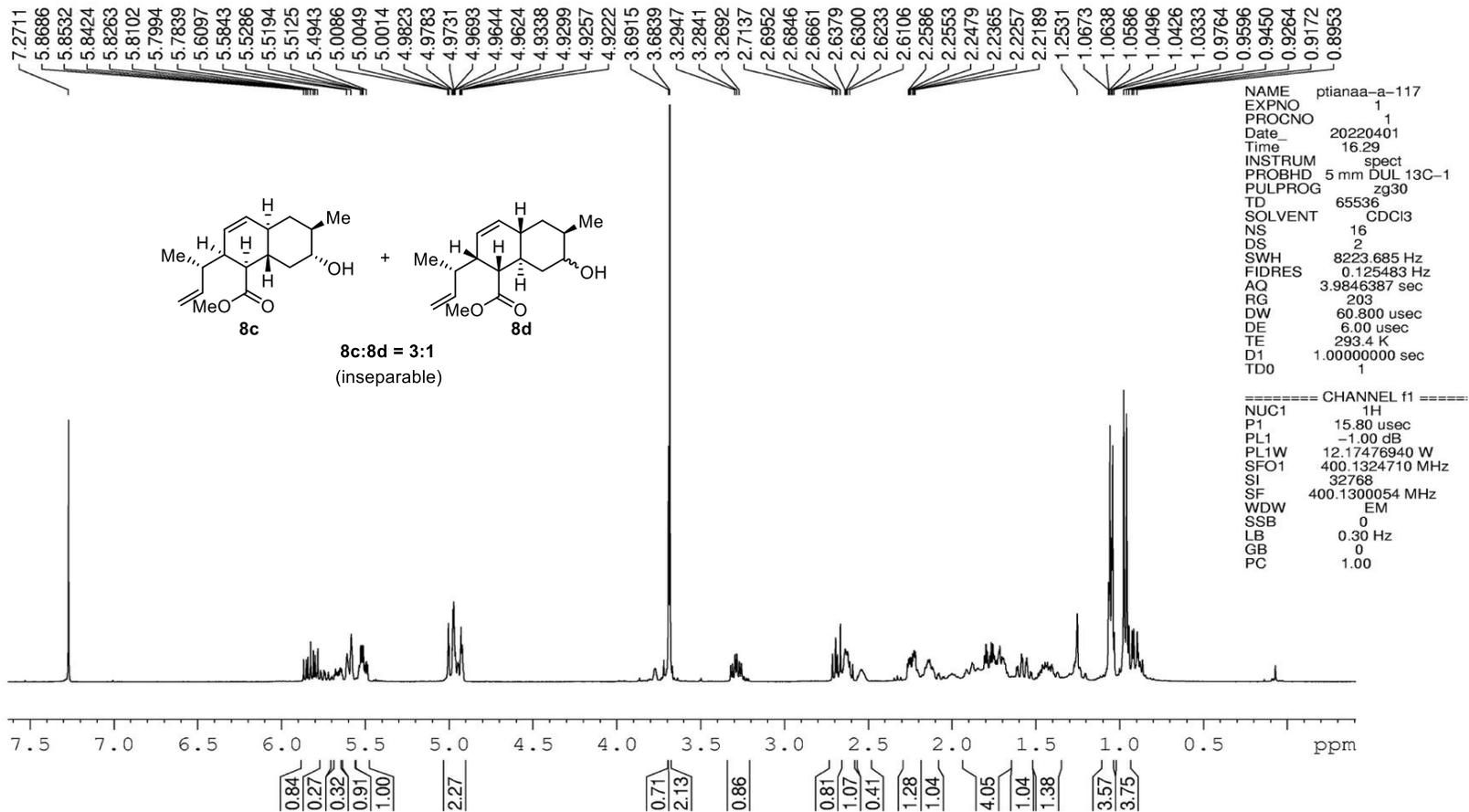




NAME ptianaa-a-syn-da
 EXPNO 1
 PROCNO 1
 Date 20220410
 Time 22.05
 INSTRUM spect
 PROBHD 5 mm PABBO BB/
 PULPROG zg30
 TD 65536
 SOLVENT GDCI3
 NS 16
 DS 2
 SWH 8012.820 Hz
 FIDRES 0.122266 Hz
 AQ 4.0894966 sec
 RG 27.78
 DW 62.400 usec
 DE 6.50 usec
 TE 296.3 K
 D1 1.00000000 sec
 TD0 1

===== CHANNEL f1 =====
 SFO1 400.1324710 MHz
 NUC1 1H
 P1 14.50 usec
 SI 65536
 SF 400.1300098 MHz
 WDW EM
 SSB 0
 LB 0.30 Hz
 GB 0
 PC 1.00



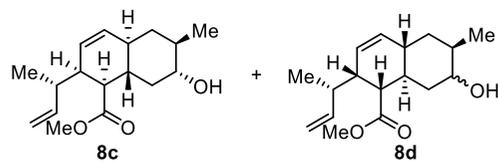


173.8276
143.4699
132.2092
125.3126
113.3738

77.3170
76.9991
76.6811
76.0602

49.4217
40.8118
40.7319
40.1207
39.3998
39.2321
38.7768
36.3707

18.3192
15.3490

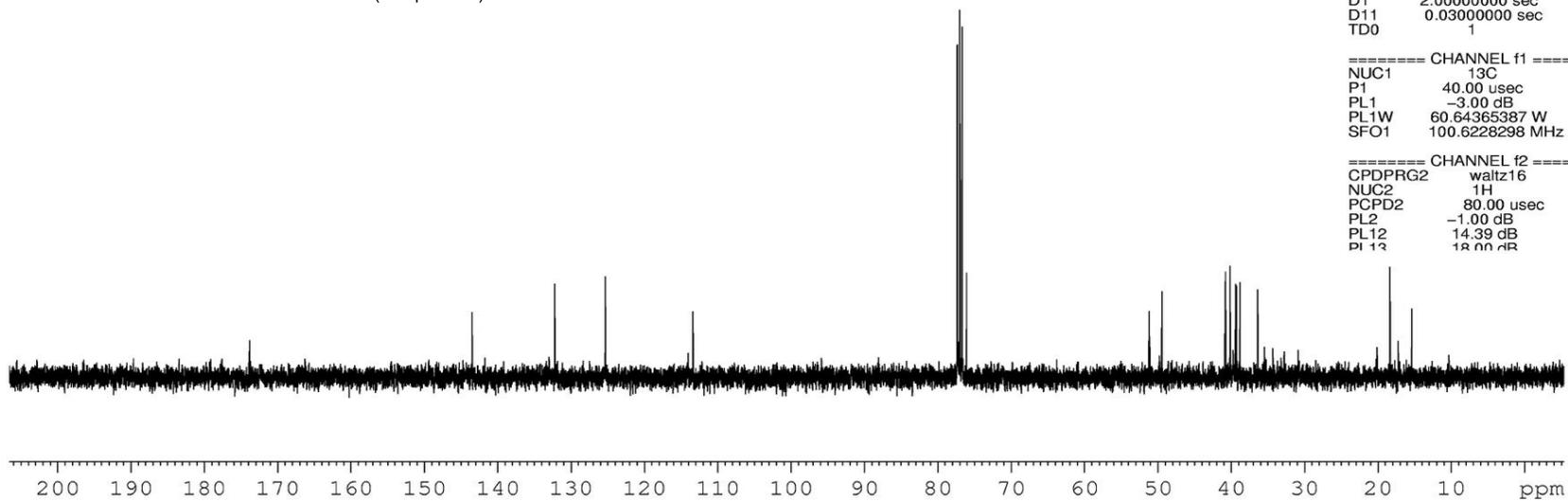


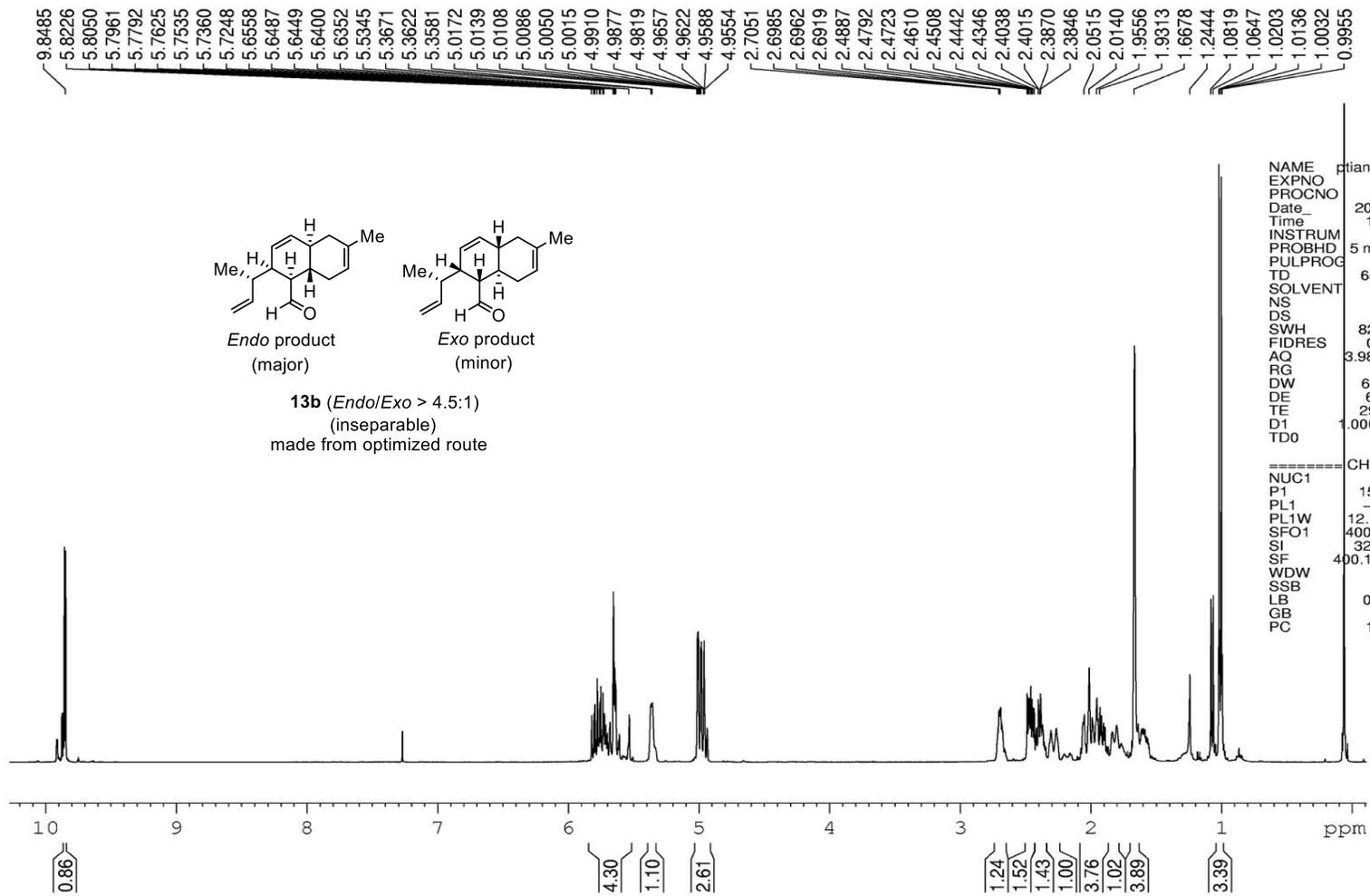
8c:8d = 3:1
(inseparable)

NAME ptianaa-a-117-C
EXPNO 1
PROCNO 1
Date_ 20220401
Time 16.34
INSTRUM spect
PROBHD 5 mm DUL 13C-1
PULPROG zgpg30
TD 65536
SOLVENT CDCl3
NS 66
DS 1
SWH 24038.461 Hz
FIDRES 0.366798 Hz
AQ 1.3631988 sec
RG 2050
DW 20.800 usec
DE 6.00 usec
TE 293.7 K
D1 2.0000000 sec
D11 0.03000000 sec
TD0 1

===== CHANNEL f1 =====
NUC1 13C
P1 40.00 usec
PL1 -3.00 dB
PL1W 60.64365387 W
SFO1 100.6228298 MHz

===== CHANNEL f2 =====
CPDPRG2 waltz16
NUC2 1H
PCPD2 80.00 usec
PL2 -1.00 dB
PL12 14.39 dB
PT 12 18.00 dB

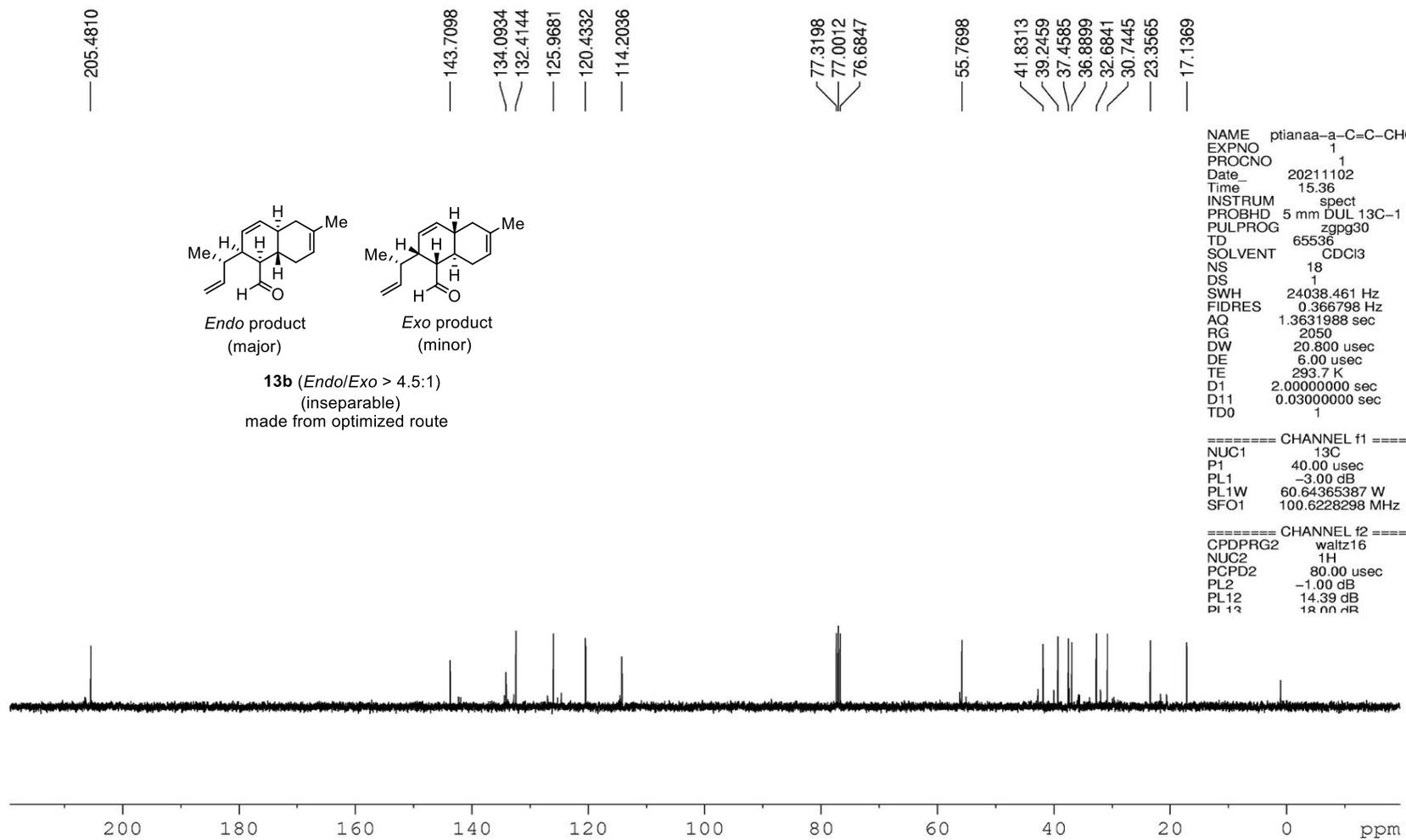


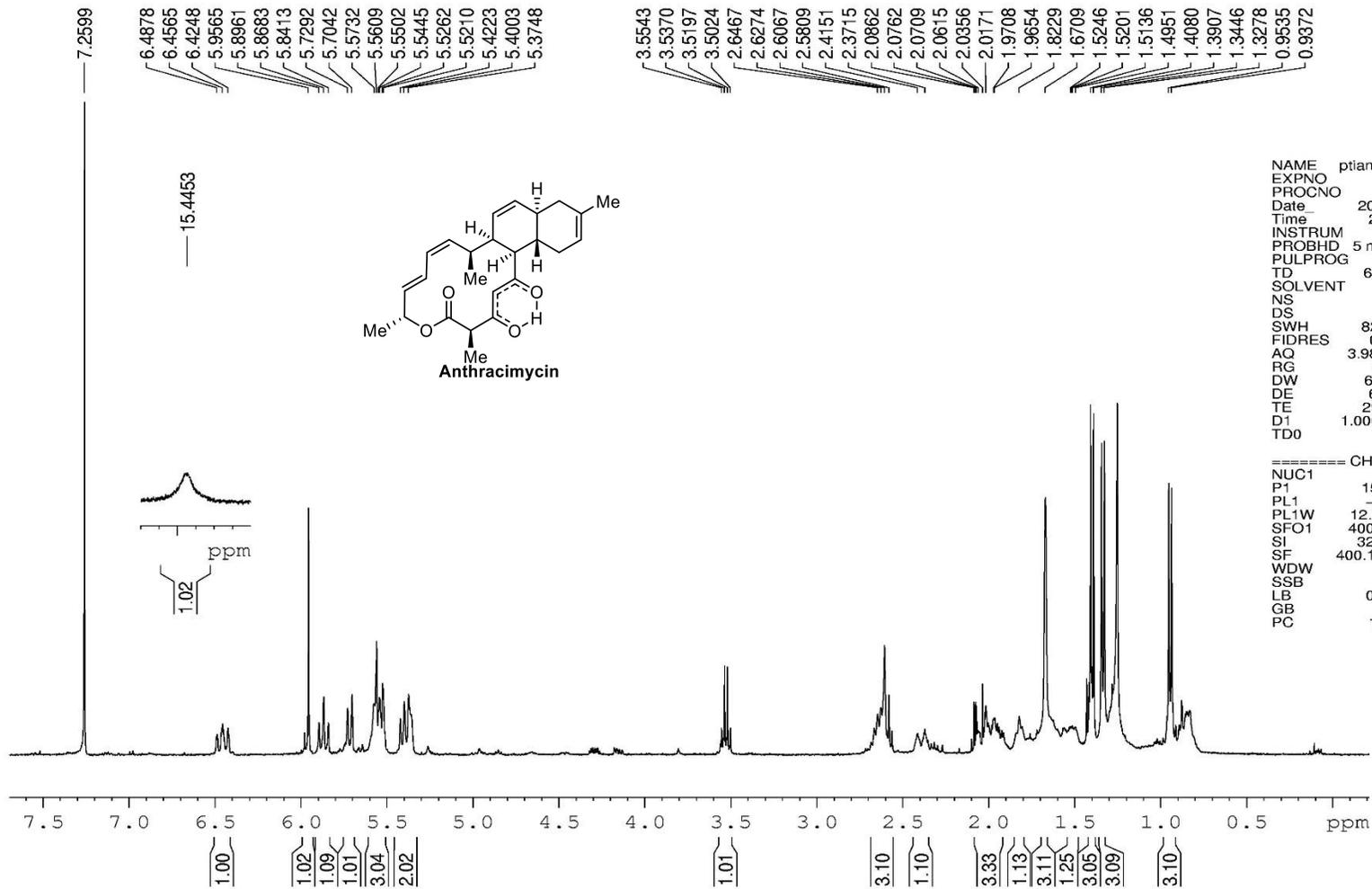


```

NAME ptianaa-a-C=C-CHO
EXPNO 1
PROCNO 1
Date_ 20211102
Time 15.35
INSTRUM spect
PROBHD 5 mm DUL 13C-1
PULPROG zg30
TD 65536
SOLVENT CDCl3
NS 4
DS 2
SWH 8223.685 Hz
FIDRES 0.125483 Hz
AQ 3.9846387 sec
RG 64
DW 60.800 usec
DE 6.00 usec
TE 293.4 K
D1 1.0000000 sec
TD0 1

===== CHANNEL f1 =====
NUC1 1H
P1 15.80 usec
PL1 -1.00 dB
PL1W 12.17476940 W
SFO1 400.1324710 MHz
SI 32768
SF 400.1300054 MHz
WDW EM
SSB 0
LB 0.30 Hz
GB 0
PC 1.00
  
```



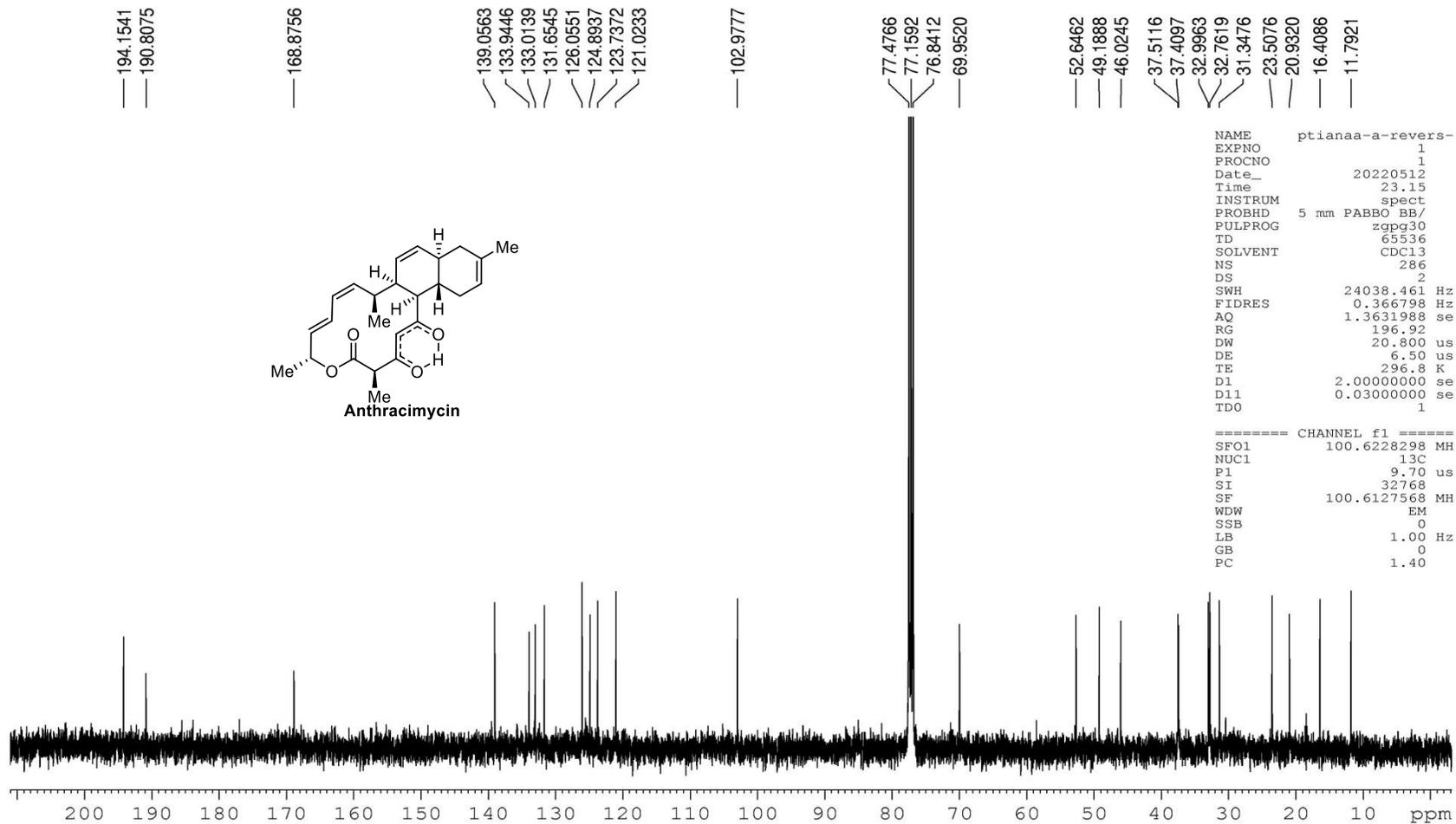


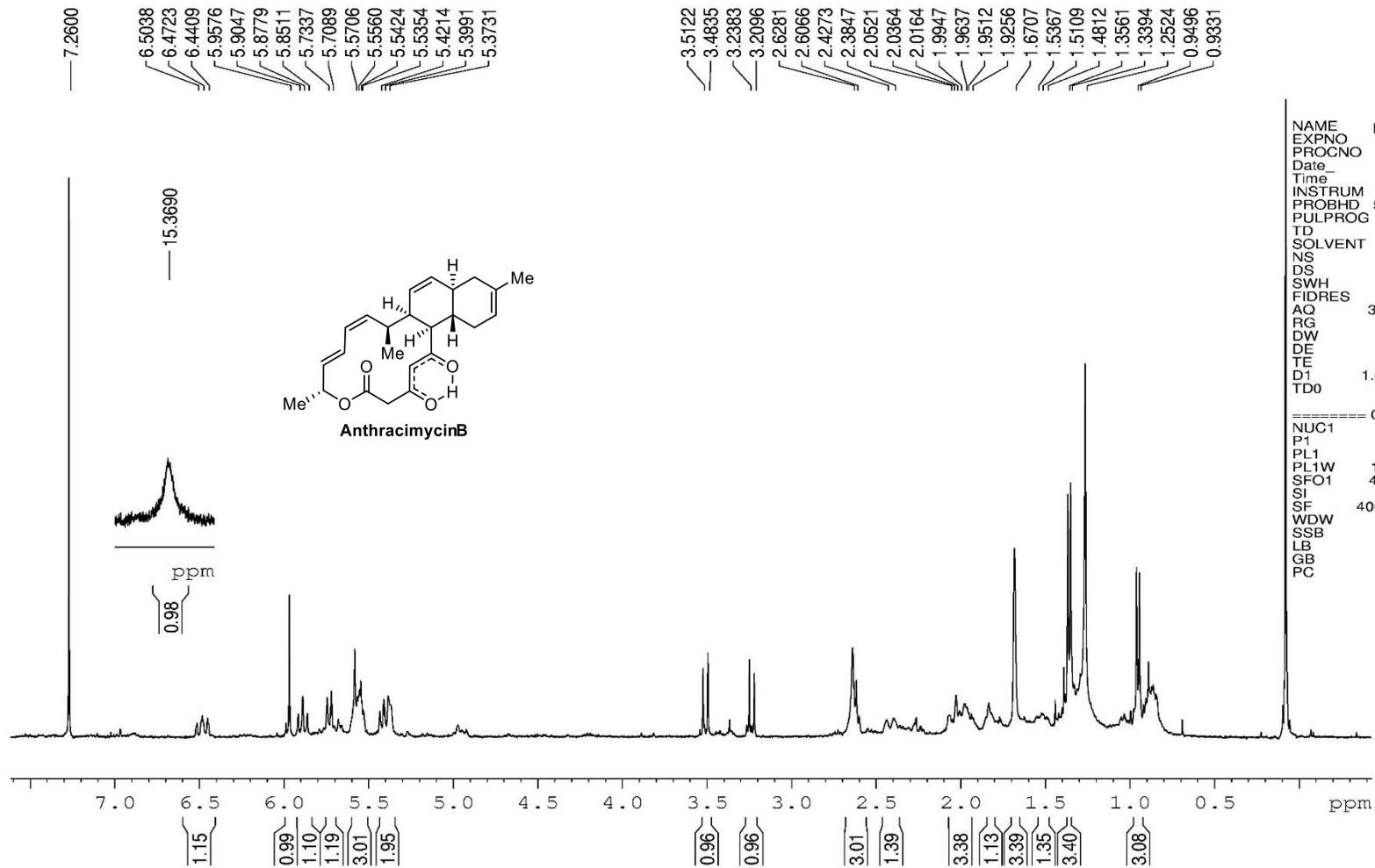
```

NAME ptianaa-anthracimycin
EXPNO 1
PROCNO 1
Date 20211117
Time 23.28
INSTRUM spect
PROBHD 5 mm DUL 13C-1
PULPROG zg30
TD 65536
SOLVENT CDCl3
NS 16
DS 2
SWH 8223.685 Hz
FIDRES 0.125483 Hz
AQ 3.9846387 sec
RG 645
DW 60.800 usec
DE 6.00 usec
TE 296.1 K
D1 1.00000000 sec
TD0 1

===== CHANNEL f1 =====
NUC1 1H
P1 15.80 usec
PL1 -1.00 dB
PL1W 12.17476940 W
SFO1 400.1324710 MHz
SI 32768
SF 400.1300099 MHz
WDW EM
SSB 0
LB 0.30 Hz
GB 0
PC 1.00

```





```

NAME      ptianaa-a-b
EXPNO     1
PROCNO    1
Date_     20211123
Time      22.58
INSTRUM   spect
PROBHD    5 mm DUL 13C-1
PULPROG   zg30
TD         65536
SOLVENT   CDCl3
NS         9
DS         2
SWH        8223.685 Hz
FIDRES     0.125483 Hz
AQ         3.9846387 sec
RG         575
DW         60.800 usec
DE         6.00 usec
TE         294.7 K
D1         1.00000000 sec
TD0        1

===== CHANNEL f1 =====
NUC1      1H
P1        15.80 usec
PL1       -1.00 dB
PL1W      12.17476940 W
SFO1      400.1324710 MHz
SI         32768
SF         400.1300054 MHz
WDW        EM
SSB        0
LB         0.30 Hz
GB         0
PC         1.00

```

