Mechanochemical Protocol Facilitates the Generation of Arylmanganese Nucleophiles from Unactivated Manganese Metal

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1. General and Materials

Materials were obtained from commercial suppliers and purified using standard procedures, unless otherwise noted. Solvents were purchased from commercial suppliers and further dried over molecular sieve (MS 4Å). Manganese powder (> 98%, product no. 130-06732) was purchased from Wako Pure Chemical Industries, Co., Ltd. All reactions were performed using grinding vessels in the Retsch MM 400 (Figure S1). Both jars and balls were made of stainless steel (SUS400B and SUS420J2, respectively, Figure S2), tungsten carbide, or zirconia. NMR spectra were recorded on JEOL JNM-ECZ400S and JNM-ECS400 spectrometers (1H: 392, 399, or 401 MHz, 13C: 99 or 100 MHz, 19F: 375 or 377 MHz). Tetramethylsilane (1H), CDCl₃ (13C), and (Trifluoromethyl)benzene (19F) were employed as internal standards, respectively. Multiplicity was recorded as follows: s = singlet, brs =broad singlet, d = doublet, t = triplet, q = quartet, and m = multiplet. Fluorobenzene, 1,2difluorobenzene, and hexafluorobenzene were used as an internal standard to determine the NMR yields. Thermographic images were obtained using the InfRec Thermo GEAR (NEC Avio Infrared Technologies Co., Ltd.). Recycling preparative gel permeation chromatography (GPC) was conducted with the JAI LC-9101 using CHCl₃ as the eluent with the JAIGEL-1H. X-Ray photoelectron spectroscopy (XPS) was performed using JEOL JPS-9200. High-resolution mass spectra were recorded at the Global Facility Center, Hokkaido University.



Figure S1. Retsch MM400 used in this study.



Figure S2. Stainless jars and balls used in this study.

2. List of Substrates Used in This Study

Aryl halides except 1k-1n were obtained from commercial suppliers and were used as received. 1k,^[1] 1l,^[2] 1m,^[3] and 1n^[4] were synthesized according to the reported procedures.

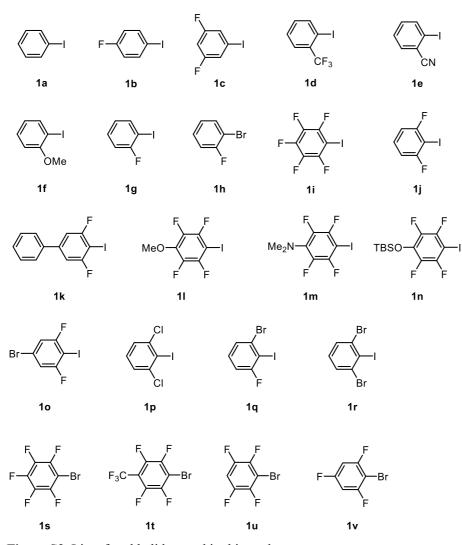


Figure S3. List of aryl halides used in this study.

All electrophiles were obtained from commercial suppliers. **2a** and **2b** were distilled before use. **2c**, **2d** and **5a**, **5b** were used as received.

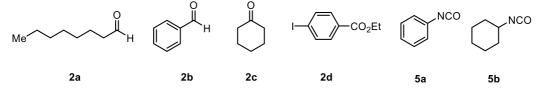
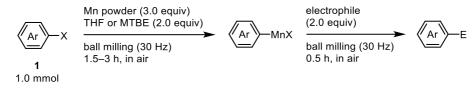


Figure S4. List of electrophiles used in this study.

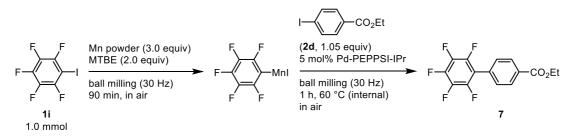
3. General Procedure for Synthesis of Organomanganese Reagents Using a Ball Mill

Procedure for the nucleophilic addition to various electrophiles (A)



All synthetic operations were carried out under atmospheric conditions. Manganese powder (3.0 mmol, 3.0 equiv) were placed in a milling jar (5 mL) with a ball (10 mm, diameter). An aryl halide (1, 1.0 mmol, 1.0 equiv) and liquid additive [2.0 mmol, 2.0 equiv, THF (162 μ L) or MTBE (236 μ L)] were added to the jar using a syringe. After the jar was closed without purging with inert gas, the jar was placed in the ball mill (Retsch MM 400, 1.5–3 h, 30 Hz). After ball milling, the jar was opened in air and charged with an electrophile (2.0 mmol, 2.0 equiv). In order to prevent undesired protonation of mechanochemically generated arylmanganese nucleophiles by atmospheric moisture, an electrophile should be added as rapidly as possible (< 1 min). The jar was then closed without purging with inert gas, and was placed in the ball mill (Retsch MM 400, 0.5 h, 30 Hz). After ball milling, the reaction mixture was quenched with 1 M HCl and extracted with ethyl acetate (30 mL×3). The solution was washed with brine and dried over MgSO₄. After the removal of the solvents under reduced pressure, the crude material was purified by flash column chromatography (SiO₂, hexane/ethyl acetate, 100:0 to 80:20) to give the corresponding product. In some cases, the product was further purified by recycling GPC.

Procedure for the palladium-catalyzed cross-coupling reactions (B)



The reaction was conducted based on a solution-state reaction reported by P. Knochel *et al.*^[5] All synthetic operations were carried out under atmospheric conditions. Manganese powder (3.0 mmol, 3.0 equiv) were placed in a milling jar (5 mL) with a ball (10 mm, diameter). 1,2,3,4,5-Pentafluoro-6-iodobenzene (**1i**, 1.0 mmol, 1.0 equiv) and MTBE (2.0 mmol, 2.0 equiv, 236 μ L) were added to the jar using a syringe. After the jar was closed without purging with inert gas, the jar was placed in the ball mill (Retsch MM 400, 90 min, 30 Hz). After ball milling, the jar was opened in air and charged with Pd-PEPPSI-IPr (0.05 mmol, 5 mol%) and ethyl 4-iodobenzoate (**2d**, 1.05 mmol, 1.05 equiv). The

jar was then closed without purging with inert gas, and was placed in the ball mill (Retsch MM 400, 1 h, 30 Hz) and a heat gun (Takagi HG-1450B). After ball milling while applying heated air to the outside of the milling jar (the preset temperature at 120 °C), the reaction mixture was quenched with 1 M HCl and extracted with ethyl acetate (30 mL×3). The solution was washed with brine and dried over MgSO₄. After the removal of the solvents under reduced pressure, the crude material was purified by flash column chromatography (SiO₂, hexane) and recycling GPC using CHCl₃ as the eluent.

Set-up procedure for high-temperature ball-milling

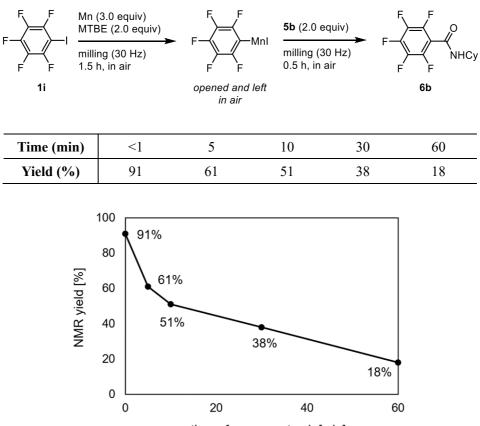
The heat gun was fixed with clamps and placed directly above the ball milling jar (distance between the heat gun and ball milling jar: ca. 1 cm) (Figure S5). The mechanochemical cross-coupling reactions were conducted while applying heated air to the outside of the milling jar (the preset temperature at 120 °C). The temperature inside the milling jar after the solid-state coupling reactions was confirmed by observation with a thermography camera immediately after opening the milling jar.



Figure S5. The set-up procedure for a heat gun on MM400.

4. Air Stability of Organomanganese Reagents Prepared by Ball Milling

We conducted a number of nucleophilic addition reactions with **5b** after exposing the mechanochemically generated arylmanganese nucleophile **1i** to air for different lengths of time (5–60 min; Figure S6). The results showed that the yield of **6b** decreased when the generated arylmanganese species were exposed to air for 5 min or more.



time of exposure to air [min]

Figure S6. Stability of the mechanochemically generated organomanganese reagent in air.

We also checked the air stability of the mechanochemically generated organomanganese reagent from **10** (Figure S7). We found that the yield of **6i** decreased when the generated arylmanganese species were exposed to air for 5 min or more, which was a result similar to that of **1i**.

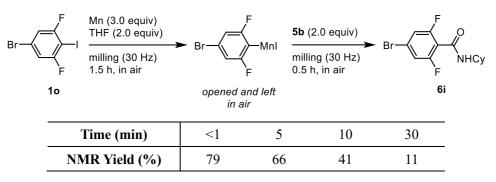
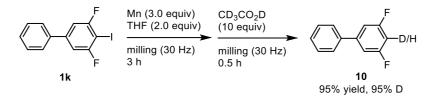


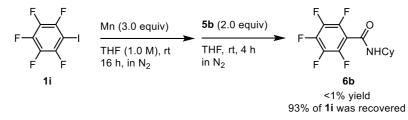
Figure S7. Stability of the mechanochemically generated organomanganese reagent from 10 in air.

5. Deuteration Experiments



All synthetic operations were carried out under atmospheric conditions. Manganese powder (3.0 mmol, 3.0 equiv) and 3,5-difluoro-4-iodo-1,1'-biphenyl (**1k**) were placed in a milling jar (5 mL) with a ball (10 mm, diameter). THF (2.0 mmol, 2.0 equiv, 162 μ L) were added to the jar using a syringe. After the jar was closed without purging with inert gas, the jar was placed in the ball mill (Retsch MM 400, 3 h, 30 Hz). After ball milling, the reaction was quenched by adding acetic acid-d₄ (10 mmol, 10 equiv) immediately after opening the jar in order to prevent undesired protonation by atmospheric moisture. The jar was then closed without purging with inert gas, and was placed in the ball mill (Retsch MM 400, 0.5 h, 30 Hz). After grinding, the reaction mixture was quenched with 1 M HCl, extracted with ethyl acetate (30 mL×3), washed with saturated aqueous Na₂CO₃, and dried over MgSO₄. After the removal of the solvents under reduced pressure, the resulting crude mixture was analyzed by ¹H NMR with dibromomethane as an internal standard to determine the NMR yield of 3,5-difluoro-1,1'-biphenyl (**10**, 95% yield, 95% D).

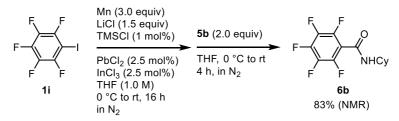
6. Details of Solution-based Reactions



Solution-based reaction without activation of manganese

An oven-dried reaction vial was charged with manganese powder (3.0 equiv) in air. After the vial was sealed with a screw cap containing a Teflon®-coated rubber septum, the vial was connected to a vacuum/nitrogen manifold through a needle. It was evacuated and then backfilled with nitrogen. This cycle was repeated three times. THF (1.0 mL) was added to the vial via syringe. **1i** was then added dropwise and the reaction mixture was stirred for 16 h. After 16 h, cyclohexyl isocyanate (**5b**, 249.8 mg, 2.0 mmol) was added to the reaction mixture. The reaction mixture was stirred at room temperature for 16 h followed by quenching with 1M HCl. The aqueous layer was extracted with ethyl acetate (30 mL×3). The combined organic phases were dried over MgSO₄, the solvent was removed in vacuo. The resulting crude mixture was analyzed by ¹⁹F NMR with fluorobenzene as internal standards to determine the NMR yield of **6b** and the conversion of **1i**.

Solution-based reaction under the conditions reported by P. Knochel et al.



The reaction was conducted according to the reported procedure.^[5] Mn powder (\geq 99.9%, product No. 463728, purchased from Aldrich), LiCl (99.9%, product No. 125-03322, purchased from Wako), PbCl₂ (-10 mesh, 99.999%, product No. 203440, purchased from Aldrich) and InCl₃ (99.999%, product No. 203440, purchased from Aldrich) were used for this reaction. An oven-dried reaction vial was charged with manganese powder (3.0 equiv), LiCl (1.5 equiv), InCl₃ (2.5 mol %) and PbCl₂ (2.5 mol %) in a glovebox under argon atmosphere. After the reaction vial was removed from the glove box, the mixture was dried for 5 min with a heat gun under high vacuum. The vial was then evacuated and backfilled with nitrogen three times and THF (4.0 mL) and TMSCl (1 mol%) were added to the mixture via syringe. **1i** (579.5 mg, 2.0 mmol) was then added dropwise at 0 °C and the reaction mixture was stirred for 16 h to give the solution of arylmanganese(II) iodide. This solution was added to a THF solution of cyclohexyl isocyanate (**5b**, 249.4 mg, 2.0 mmol, 1.3 M) at 0 °C.

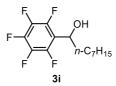
allowed to warm to room temperature and continuously stirred for 4 h, followed by quenching with brine. The aqueous layer was extracted with CH_2Cl_2 (30 mL×3). The combined organic phases were dried over MgSO₄, the solvent was removed in vacuo. The resulting crude mixture was analyzed by ¹⁹F NMR with fluorobenzene as an internal standard to determine the NMR yield of **6b**.

7. Sample Preparation for XPS Analysis

X-Ray photoelectron spectroscopy (XPS) was used to analyze the surface of manganese metals. The ball-milled sample was prepared via the following conditions: Manganese powder (160 mg) was placed in a milling jar (5 mL) with a ball (10 mm, diameter) under atmospheric conditions. After the jar was closed without purging with inert gas, the jar was placed in the ball mill (Retsch MM 400, 1.5 h, 30 Hz). After ball milling, the jar was opened in a glove box, and the ball-milled sample was transferred to the vessel for XPS analysis.

8. Characterization of Products

1-(Perfluorophenyl)octan-1-ol (3i).



The first metalation step was conducted with **1i** (293.7 mg, 1.0 mmol) and MTBE (236 μ L, 2.0 mmol) for 1.5 h and the second addition step was conducted with **2a** (246.5 mg, 1.9 mmol) for 0.5 h. The product **3i** was purified by flash column chromatography (SiO₂, hexane/ethyl acetate, 100:0 to 90:10) and GPC. The product **3i** was obtained in 54% yield (160.7 mg, 0.54 mmol) as a white solid.

¹H NMR (399 MHz, CDCl₃, δ): 0.88 (t, *J* = 6.6 Hz, 3H), 1.17–1.37 (m, 9H), 1.39–1.50 (m, 1H), 1.77–1.88 (m, 1H), 1.96–2.06 (m, 1H), 2.10 (d, *J* = 7.6 Hz, 1H), 5.04 (q, *J* = 7.6 Hz, 1H). ¹³C NMR (100 MHz, CDCl₃, δ): 14.1 (CH₃), 22.7 (CH₂), 26.0 (CH₂), 29.26 (CH₂), 29.31 (CH₂), 31.9 (CH₂), 36.9 (CH₂), 66.5 (CH), 117.3 (t, *J*_{C-F} = 15.3 Hz, *C*), 137.6 (dm, *J*_{C-F} = 252.0 Hz, *C*), 140.6 (dm, *J*_{C-F} = 254.0 Hz, *C*), 144.8 (dm, *J*_{C-F} = 247.3 Hz, *C*). ¹⁹F NMR (375 MHz, CDCl₃, δ): –162.8 (t, *J* = 23.1 Hz, 2F), –156.4 (d, *J* = 23.3 Hz, 1F), –145.0 (t, *J* = 22.9 Hz, 2F). HRMS-EI (m/z): [M]⁺ calcd for C₁₄H₁₇F₅O, 296.1200; found, 296.1198.

1-(2,6-Difluorophenyl)octan-1-ol (3j).



The first metalation step was conducted with 1j (240.5 mg, 1.0 mmol) and THF (165 μ L, 2.0 mmol) for 3 h and the second addition step was conducted with 2a (255.9 mg, 2.0 mmol) for 0.5 h. The product 3j was purified by flash column chromatography (SiO₂, hexane/ethyl acetate, 100:0 to 90:10) and GPC. The product 3j was obtained in 67% yield (96.6 mg, 0.68 mmol) as colorless oil.

¹H NMR (399 MHz, CDCl₃, δ): 0.87 (t, *J* = 6.8 Hz, 3H), 1.18–1.36 (m, 9H), 1.39–1.51 (m, 1H), 1.77–1.88 (m, 1H), 1.94–2.05 (m, 1H), 2.17 (dt, *J* = 2.4, 9.0 Hz, 1H), 5.03 (q, *J* = 7.8 Hz, 1H), 6.83–6.92 (m, 2H), 7.16–7.25 (m, 1H). ¹³C NMR (100 MHz, CDCl₃, δ): 14.2 (*C*H₃), 22.7 (*C*H₂), 26.0 (*C*H₂), 29.3 (*C*H₂), 29.4 (*C*H₂), 31.9 (*C*H₂), 37.4 (*C*H₂), 66.6 (*C*H), 111.4–112.0 (m, *C*H), 119.9 (t, *J*_{C–F} = 16.7 Hz, *C*), 128.9 (t, *J*_{C–F} = 11.0 Hz, *C*H), 161.1 (dd, *J*_{C–F} = 8.7, 248.0 Hz, *C*). ¹⁹F NMR (377 MHz, CDCl₃, δ): –116.5 (s, 2F). HRMS-EI (m/z): [M]⁺ calcd for C₁₄H₂₀F₂O, 242.1482; found, 242.1481.

1-(2-Fluorophenyl)octan-1-ol (3g).



The first metalation step was conducted with **1g** (222.2 mg, 1.0 mmol) and THF (162 μ L, 2.0 mmol) for 3 h and the second addition step was conducted with **2a** (257.0 mg, 2.0 mmol) for 0.5 h. The product **3g** was purified by flash column chromatography (SiO₂, hexane/ethyl acetate, 100:0 to 90:10) and GPC. The product **3g** was obtained in 25% yield (55.1 mg, 0.25 mmol) as colorless oil.

¹H NMR (399 MHz, CDCl₃, δ): 0.87 (t, J = 6.8 Hz, 3H), 1.18–1.37 (m, 9H), 1.39–1.50 (m, 1H), 1.69–1.82 (m, 2H), 1.85 (d, J = 4.8 Hz, 1H), 5.01 (q, J = 5.9 Hz, 1H), 7.02 (ddd, J = 1.4, 8.2, 10.4 Hz, 1H), 7.15 (td, J = 1.1, 7.5 Hz, 1H), 7.21–7.26 (m, 1H), 7.46 (td, J = 1.9, 7.6 Hz, 1H). ¹³C NMR (100 MHz, CDCl₃, δ): 14.2 (CH₃), 22.8 (CH₂), 25.8 (CH₂), 29.3 (CH₂), 29.5 (CH₂), 31.9 (CH₂), 38.2 (CH₂), 68.5 (CH), 115.3 (d, $J_{C-F} = 21.9$ Hz, CH), 124.3 (d, $J_{C-F} = 2.9$ Hz, CH), 127.4 (d, $J_{C-F} = 4.8$ Hz, CH), 128.8 (d, $J_{C-F} = 8.7$ Hz, CH), 132.0 (d, $J_{C-F} = 13.3$ Hz, C), 159.9 (d, $J_{C-F} = 245.2$ Hz, C). ¹⁹F NMR (377 MHz, CDCl₃, δ): –120.8 (s, 1F). HRMS-EI (m/z): [M]⁺ calcd for C₁₄H₂₁FO, 224.1576; found, 224.1568.

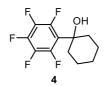
(Perfluorophenyl)(phenyl)methanol (3k).



The first metalation step was conducted with **1i** (293.8 mg, 1.0 mmol) and MTBE (236 μ L, 2.0 mmol) for 1.5 h and the second addition step was conducted with **2b** (215.4 mg, 2.0 mmol) for 0.5 h. The product **3k** was purified by flash column chromatography (SiO₂, hexane/ethyl acetate, 100:0 to 90:10) and obtained in 60% yield (165.0 mg, 0.60 mmol) as a white solid. ¹H, ¹³C, and ¹⁹F NMR of the product **3k** were in agreement with the literature.^[6]

¹H NMR (401 MHz, CDCl₃, δ): 2.64 (d, *J* = 8.0 Hz, 1H), 6.25 (d, *J* = 8.0 Hz, 1H), 7.30–7.42 (m, 5H). ¹³C NMR (100 MHz, CDCl₃, δ): 67.6 (*C*H), 117.0 (t, *J*_{C-F} = 13.8 Hz, *C*), 125.4 (*C*H), 128.3 (*C*H), 128.8 (*C*H), 137.7 (dm, *J*_{C-F} = 252.7 Hz, *C*), 140.6 (*C*), 140.9 (dm, *J*_{C-F} = 254.6 Hz, *C*), 144.7 (dm, *J*_{C-F} = 249.8 Hz, *C*). ¹⁹F NMR (377 MHz, CDCl₃, δ): –162.4 (t, *J* = 23.0 Hz, 2F), –155.5 (d, *J* = 34.7 Hz, 1F), –144.0 (t, *J* = 23.0 Hz, 2F). HRMS-EI (m/z): [M]⁺ calcd for C₁₃H₇F₅O, 274.0417; found, 274.0414.

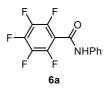
1-(Perfluorophenyl)cyclohexan-1-ol (4).



The first metalation step was conducted with **1i** (295.6 mg, 1.0 mmol) and MTBE (236 μ L, 2.0 mmol) for 1.5 h and the second addition step was conducted with **2c** (117.0 mg, 1.2 mmol) for 0.5 h. The product **4** was purified by flash column chromatography (SiO₂, hexane/ethyl acetate, 100:0 to 90:10) and GPC. The product **4** was obtained in 45% yield (120.1 mg, 0.45 mmol) as a white solid. ¹H, ¹³C, and ¹⁹F NMR of the product **4** were in agreement with the literature.^[6]

¹H NMR (392 MHz, CDCl₃, δ): 1.29 (qt, *J* = 3.8, 12.8 Hz, 1H), 1.56–1.64 (m, 2H), 1.67–1.89 (m, 3H), 1.97–2.09 (m, 4H), 2.42 (t, *J* = 3.4 Hz, 1H). ¹³C NMR (99 MHz, CDCl₃, δ): 21.4 (CH₂), 25.1 (CH₂), 37.2 (t, *J*_{C-F} = 4.3 Hz, CH₂), 75.3 (*C*), 121.3–121.7 (m, *C*), 137.5 (dm, *J*_{C-F} = 248.2 Hz, *C*), 140.0 (dm, *J*_{C-F} = 252.0 Hz, *C*), 145.1 (dm, *J*_{C-F} = 248.8 Hz, *C*). ¹⁹F NMR (377 MHz, CDCl₃, δ): –162.7 (t, *J* = 23.1 Hz, 2F), –157.1 (d, *J* = 23.1 Hz, 1F), –140.7 (t, *J* = 23.1 Hz, 2F). HRMS-EI (m/z): [M]⁺ calcd for C₁₂H₁₁F₅O, 266.0730; found, 266.0728.

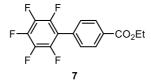
2,3,4,5,6-Pentafluoro-N-phenylbenzamide (6a).



The first metalation step was conducted with **1i** (293.8 mg, 1.0 mmol) and MTBE (236 μ L, 2.0 mmol) for 1.5 h and the second amidation step was conducted with **5a** (237.4 mg, 2.0 mmol) for 0.5 h. The product **6a** was purified by flash column chromatography (SiO₂, hexane/ethyl acetate, 100:0 to 85:15) and obtained in 69% yield (197.0 mg, 0.69 mmol) as a white solid. ¹H and ¹⁹F NMR of the product **6a** were in agreement with the literature.^[7]

¹H NMR (401 MHz, DMSO-d₆, δ): 7.23 (t, *J* = 7.2 Hz, 1H), 7.41 (t, *J* = 8.0 Hz, 2H), 7.52 (brs, 1H), 7.61 (d, *J* = 8.8 Hz, 2H). ¹³C NMR (100 MHz, CDCl₃, δ): 112.5 (t, *J*_{C-F} = 20.5 Hz, *C*), 119.6 (CH), 124.8 (CH), 129.1 (CH), 137.1 (dm, *J*_{C-F} = 253.7 Hz, *C*), 138.0 (*C*), 141.5 (dm, *J*_{C-F} = 250.9 Hz, *C*), 143.2 (dm, *J*_{C-F} = 251.3 Hz, *C*), 154.9 (*C*). ¹⁹F NMR (375 MHz, CDCl₃, δ): -160.4 (t, *J* = 23.1 Hz, 2F), -150.6 (t, *J* = 23.3 Hz, 1F), -140.9 (s, 2F). HRMS-EI (m/z): [M]⁺ calcd for C₁₃H₆F₅NO, 287.0370; found, 287.0372.

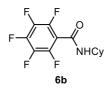
Ethyl 2',3',4',5',6'-pentafluoro-[1,1'-biphenyl]-4-carboxylate (7).



The reaction was conducted with **1i** (293.5 mg, 1.0 mmol) according to the general procedure B. The product **7** was purified by GPC. The product **7** was obtained in 71% yield (224.6 mg, 0.71 mmol) as a white solid. ¹H, ¹³C, and ¹⁹F NMR of the product **7** were in agreement with the literature.^[8]

¹H NMR (401 MHz, CDCl₃, δ): 1.42 (t, *J* = 7.1 Hz, 3H), 4.42 (q, *J* = 7.0 Hz, 2H), 7.51 (d, *J* = 8.2 Hz, 2H), 8.17 (d, *J* = 8.7 Hz, 2H). ¹³C NMR (100 MHz, CDCl₃, δ): 14.2 (CH₃), 61.3 (CH₂), 114.7–115.2 (m, *C*), 129.7 (CH), 130.2 (CH), 130.7 (C), 131.2 (C), 137.8 (dm, *J*_{C-F} = 252.2 Hz, *C*), 140.7 (dm, *J*_{C-F} = 254.1 Hz, *C*), 144.0 (dm, *J*_{C-F} = 249.3 Hz, *C*), 165.8 (*C*). ¹⁹F NMR (377 MHz, CDCl₃, δ): -162.2 (t, *J* = 23.1 Hz, 2F), -154.8 (d, *J* = 34.7 Hz, 1F), -143.4 (s, 2F). HRMS-EI (m/z): [M]⁺ calcd for C₁₅H₉F₅O₂, 316.0523; found, 316.0520.

N-Cyclohexyl-2,3,4,5,6-pentafluorobenzamide (6b).



For reaction with **1i**: the first metalation step was conducted with **1i** (293.3 mg, 1.0 mmol) and MTBE (236 μ L, 2.0 mmol) for 1.5 h and the second amidation step was conducted with **5b** (246.4 mg, 2.0 mmol) for 0.5 h. The product **6b** was purified by flash column chromatography (SiO₂, hexane/ethyl acetate, 100:0 to 80:20) and obtained in 72% yield (210.7 mg, 0.72 mmol) as a white solid.

For the reaction with **1s**: the first metalation step was conducted with **1s** (246.8 mg, 1.0 mmol) and MTBE (236 μ L, 2.0 mmol) for 1.5 h and the second arylation step was conducted with **5b** (248.2 mg, 2.0 mmol) for 0.5 h. The product **6b** was purified by flash column chromatography (SiO₂, hexane/ethyl acetate, 100:0 to 85:15) and obtained in 57% yield (168.2 mg, 0.57 mmol) as a white solid.

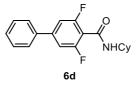
¹H NMR (392 MHz, CDCl₃, δ): 1.15–1.32 (m, 3H), 1.36–1.49 (m, 2H), 1.61–1.69 (m, 1H), 1.71– 1.80 (m, 2H), 1.99–2.09 (m, 2H), 3.94–4.06 (m, 1H), 5.76 (brs, 1H). ¹³C NMR (100 MHz, CDCl₃, δ): 24.8 (CH₂), 25.4 (CH₂), 32.7 (CH₂), 49.6 (CH), 112.2 (t, *J*_{C-F} = 18.6 Hz, *C*), 137.6 (dm, *J*_{C-F} = 255.1 Hz, *C*), 142.1 (dm, *J*_{C-F} = 256.1 Hz, *C*), 144.0 (dm, *J*_{C-F} = 251.7 Hz, *C*), 156.5 (*C*). ¹⁹F NMR (377 MHz, CDCl₃, δ): –161.1 (s, 2F), –152.1 (d, *J* = 23.0 Hz, 1F), –141.7 (d, *J* = 23.4 Hz, 2F). HRMS-EI (m/z): [M]⁺ calcd for C₁₃H₁₂F₅NO, 293.0839; found, 293.0833. N-Cyclohexyl-2,6-difluorobenzamide (6c).



The first metalation step was conducted with **1b** (242.1 mg, 1.0 mmol) and THF (162 μ L, 2.0 mmol) for 1.5 h and the second amidation step was conducted with **5b** (248.9 mg, 2.0 mmol) for 0.5 h. The product **6c** was purified by flash column chromatography (SiO₂, hexane/ethyl acetate, 100:0 to 80:20) and obtained in 79% yield (191.1 mg, 0.80 mmol) as a white solid. ¹H, ¹³C, and ¹⁹F NMR of the product **6c** were in agreement with the literature.^[9]

¹H NMR (392 MHz, CDCl₃, δ): 1.18–1.32 (m, 3H), 1.36–1.50 (m, 2H), 1.59–1.69 (m, 1H), 1.70– 1.80 (m, 2H), 2.01–2.10 (m, 2H), 3.97–4.07 (m, 1H), 5.77 (brs, 1H), 6.90–6.98 (m, 2H), 7.30–7.39 (m, 1H). ¹³C NMR (100 MHz, CDCl₃, δ): 24.8 (*C*H₂), 25.6 (*C*H₂), 33.0 (*C*H₂), 49.0 (*C*H), 111.9–112.1 (m, *C*H), 114.9 (t, $J_{C-F} = 20.5$ Hz, *C*), 131.5 (t, $J_{C-F} = 10.1$ Hz, *C*H), 159.5 (*C*), 160.0 (dd, $J_{C-F} = 6.7$, 251.7 Hz, *C*). ¹⁹F NMR (377 MHz, CDCl₃, δ): –113.6 (s, 2F). HRMS-EI (m/z): [M]⁺ calcd for C₁₃H₁₅F₂NO, 239.1122; found, 239.1120.

N-Cyclohexyl-3,5-difluoro-[1,1'-biphenyl]-4-carboxamide (6d).



The first metalation step was conducted with **1k** (316.3 mg, 1.0 mmol) and THF (162 μ L, 2.0 mmol) for 3 h and the second amidation step was conducted with **5b** (247.6 mg, 2.0 mmol) for 0.5 h. The product **6d** was purified by flash column chromatography (SiO₂, CH₂Cl₂) and the obtained solid was washed with hexane. The product **6d** was obtained in 71% yield (224.0 mg, 0.71 mmol) as a white solid.

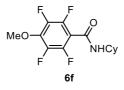
¹H NMR (401 MHz, CDCl₃, δ): 1.16–1.34 (m, 3H), 1.38–1.50 (m, 2H), 1.60–1.69 (m, 1H), 1.71– 1.81 (m, 2H), 2.02–2.13 (m, 2H), 3.98–4.10 (m, 1H), 5.84 (brs, 1H), 7.14–7.20 (m, 2H), 7.40–7.50 (m, 3H), 7.52–7.57 (m, 2H). ¹³C NMR (100 MHz, DMSO-*d*₆, δ): 24.5 (*C*H₂), 25.2 (*C*H₂), 32.2 (*C*H₂), 48.3 (*C*H), 109.8 (d, $J_{C-F} = 24.8$ Hz, *C*H), 114.4 (t, $J_{C-F} = 6.2$ Hz, *C*), 126.9 (*C*H), 128.9 (*C*H), 129.1 (*C*H), 137.2 (*C*H), 143.3 (t, $J_{C-F} = 9.6$ Hz, *C*), 158.5 (*C*), 159.2 (dd, $J_{C-F} = 9.0$, 247.5 Hz, *C*). ¹⁹F NMR (377 MHz, CDCl₃, δ): –113.1 (s, 2F). HRMS-EI (m/z): [M]⁺ calcd for C₁₉H₁₉F₂NO, 315.1435; found, 315.1430. N-Cyclohexyl-2-fluorobenzamide (6e).



The first metalation step was conducted with **1g** (221.3 mg, 1.0 mmol) and 1,2-dimethoxyethane (207 μ L, 2.0 mmol) for 3 h and the second amidation step was conducted with **5b** (253 μ L, 2.0 mmol) for 0.5 h. The product **6e** was purified by flash column chromatography (SiO₂, hexane/ethyl acetate, 100:0 to 80:20) and obtained in 38% yield (84.1 mg, 0.38 mmol) as a white solid. ¹H, ¹³C, and ¹⁹F NMR of the product **6e** were in agreement with the literature.^[10]

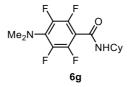
¹H NMR (401 MHz, CDCl₃, δ): 1.18–1.34 (m, 3H), 1.38–1.51 (m, 2H), 1.60–1.69 (m, 1H), 1.71– 1.80 (m, 2H), 1.98–2.09 (m, 2H), 3.96–4.09 (m, 1H), 6.62 (brs, 1H), 7.10 (dd, *J* = 8.6, 11.4 Hz, 1H), 7.22–7.29 (m, 1H), 7.40–7.50 (m, 1H), 8.09 (dt, *J* = 3.9, 11.3 Hz, 1H). ¹³C NMR (100 MHz, CDCl₃, δ): 24.8 (*C*H₂), 25.6 (*C*H₂), 33.0 (*C*H₂), 48.6 (*C*H), 115.9 (d, *J*_{C-F} = 24.8 Hz, *C*H), 121.6 (d, *J*_{C-F} = 12.3 Hz, *C*), 124.7 (d, *J*_{C-F} = 2.8 Hz, *C*H), 132.0 (*C*H), 133.0 (d, *J*_{C-F} = 8.7 Hz, *C*H), 160.5 (d, *J*_{C-F} = 247.0 Hz, *C*), 162.3 (d, *J*_{C-F} = 2.9 Hz, *C*). ¹⁹F NMR (377 MHz, CDCl₃, δ): –114.9 (s, 1F). HRMS-EI (m/z): [M]⁺ calcd for C₁₃H₁₆FNO, 221.1216; found, 221.1216.

N-Cyclohexyl-2,3,5,6-tetrafluoro-4-methoxybenzamide (6f).



The first metalation step was conducted with **11** (308.7 mg, 1.0 mmol) and THF (162 μ L, 2.0 mmol) for 1.5 h and the second amidation step was conducted with **5b** (249.4 mg, 2.0 mmol) for 0.5 h. The product **6f** was purified by flash column chromatography (SiO₂, hexane/ethyl acetate, 100:0 to 80:20) and obtained in 70% yield (215.2 mg, 0.70 mmol) as a white solid.

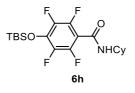
¹H NMR (401 MHz, CDCl₃, δ): 1.18–1.30 (m, 3H), 1.36–1.49 (m, 2H), 1.60–1.68 (m, 1H), 1.70– 1.79 (m, 2H), 1.98–2.09 (m, 2H), 3.94–4.04 (m, 1H), 4.10–4.12 (m, 3H), 5.77 (brs, 1H). ¹³C NMR (100 MHz, CDCl₃, δ): 24.7 (CH₂), 25.4 (CH₂), 32.5 (CH₂), 49.3 (CH), 62.0 (t, $J_{C-F} = 3.8$ Hz, CH₃), 110.0 (t, $J_{C-F} = 19.6$ Hz, C), 138.9–139.5 (m, C), 140.4 (dm, $J_{C-F} = 239.9$ Hz, C), 143.9 (dm, $J_{C-F} =$ 249.8 Hz, C), 157.3 (C). ¹⁹F NMR (377 MHz, CDCl₃, δ): –157.9 (s, 2F), –143.6 (d, J = 23.4 Hz, 2F). HRMS-EI (m/z): [M]⁺ calcd for C₁₄H₁₅F₄NO₂, 305.1039; found, 305.1039. N-Cyclohexyl-4-(dimethylamino)-2,3,5,6-tetrafluorobenzamide (6g).



The first metalation step was conducted with **1m** (318.1 mg, 1.0 mmol) and THF (162 μ L, 2.0 mmol) for 1.5 h and the second amidation step was conducted with **5b** (249.2 mg, 2.0 mmol) for 0.5 h. The product **6g** was purified by flash column chromatography (SiO₂, hexane/ethyl acetate, 100:0 to 80:20) and obtained in 69% yield (220.5 mg, 0.69 mmol) as a white solid.

¹H NMR (401 MHz, CDCl₃, δ): 1.16–1.30 (m, 3H), 1.35–1.48 (m, 2H), 1.59–1.68 (m, 1H), 1.69– 1.79 (m, 2H), 1.98–2.08 (m, 2H), 2.99 (t, J = 2.2 Hz, 6H), 3.93–4.05 (m, 1H), 5.78 (brs, 1H). ¹³C NMR (100 MHz, CDCl₃, δ): 24.8 (CH₂), 25.5 (CH₂), 32.8 (CH₂), 43.1 (t, $J_{C-F} = 4.3$ Hz, CH₃), 49.1 (CH), 107.1 (t, $J_{C-F} = 18.1$ Hz, C), 132.9 (t, $J_{C-F} = 8.6$ Hz, C), 141.2 (dm, $J_{C-F} = 244.1$ Hz, C), 144.8 (dm, $J_{C-F} = 245.1$ Hz, C), 158.0 (C). ¹⁹F NMR (377 MHz, CDCl₃, δ): –152.5 (d, J = 23.0 Hz, 2F), –144.6 (s, 2F). HRMS-EI (m/z): [M]⁺ calcd for C₁₅H₁₈F₄N₂O₁, 318.1355; found, 318.1346.

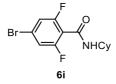
4-[(tert-Butyldimethylsilyl)oxy]-N-cyclohexyl-2,3,5,6-tetrafluorobenzamide (6h).



The first metalation step was conducted with **1n** (406.6 mg, 1.0 mmol) and THF (162 μ L, 2.0 mmol) for 90 min and the second amidation step was conducted with **5b** (248.7 mg, 2.0 mmol) for 30 min. The product **6h** was purified by flash column chromatography (SiO₂, hexane/ethyl acetate, 100:0 to 80:20) and obtained in 28% yield (113.4 mg, 0.28 mmol) as a white solid.

¹H NMR (401 MHz, CDCl₃, δ): 0.23 (s, 6H), 1.01 (s, 9H), 1.15–1.31 (m, 3H), 1.36–1.49 (m, 2H), 1.60–1.68 (m, 1H), 1.70–1.79 (m, 2H), 2.00–2.08 (m, 2H), 3.94–4.06 (m, 1H), 5.78 (brs, 1H). ¹³C NMR (100 MHz, CDCl₃, δ): –4.9 (*C*H₃), 18.5 (*C*), 24.8 (*C*H₂), 25.3 (*C*H₃), 25.5 (*C*H₂), 32.8 (*C*H₂), 49.3 (*C*H), 108.6 (t, $J_{C-F} = 18.6$ Hz, *C*), 136.1–136.5 (m, *C*), 140.7 (dm, $J_{C-F} = 248.4$ Hz, *C*), 144.3 (dm, $J_{C-F} = 251.7$ Hz, *C*), 157.7 (*C*). ¹⁹F NMR (377 MHz, CDCl₃, δ): –157.6 (d, J = 23.0 Hz, 2F), –144.4 (s, 2F). HRMS-EI (m/z): [M]⁺ calcd for C₁₉H₂₇F₄NO₂Si, 405.1747; found, 405.1740.

4-Bromo-N-cyclohexyl-2,6-difluorobenzamide (6i).



The first step was conducted with **10** (319.1 mg, 1.0 mmol) and THF (162 μ L, 2.0 mmol) for 1.5 h and the second amidation step was conducted with **5b** (247.6 mg, 2.0 mmol) for 0.5 h. The product **6i** was purified by flash column chromatography (SiO₂, hexane/ethyl acetate, 100:0 to 80:20) and obtained in 75% yield (237.9 mg, 0.75 mmol) as a white solid.

¹H NMR (401 MHz, CDCl₃, δ): 1.14–1.30 (m, 3H), 1.35–1.49 (m, 2H), 1.60–1.69 (m, 1H), 1.70– 1.79 (m, 2H), 1.98–2.10 (m, 2H), 3.93–4.06 (m, 1H), 5.73 (brs, 1H), 7.11–7.17 (m, 2H). ¹³C NMR (100 MHz, CDCl₃, δ): 24.8 (CH₂), 25.5 (CH₂), 32.9 (CH₂), 49.1 (CH), 114.2 (t, *J*_{C-F} = 21.0 Hz, *C*), 116.0 (d, *J*_{C-F} = 28.6 Hz, CH), 123.8 (t, *J*_{C-F} = 11.9 Hz, *C*), 158.6 (*C*), 159.8 (dd, *J*_{C-F} = 8.6, 255.6 Hz, *C*). ¹⁹F NMR (377 MHz, CDCl₃, δ): –111.7 (s, 2F). HRMS-ESI (m/z): [M+Na]⁺ calcd for C₁₃H₁₄BrF₂NONa, 340.0119; found, 340.0122.

2,6-Dichloro-N-cyclohexylbenzamide (6j).



The first metalation step was conducted with **1p** (273.3 mg, 1.0 mmol) and THF (162 μ L, 2.0 mmol) for 1.5 h and the second amidation step was conducted with **5b** (248.5 mg, 2.0 mmol) for 0.5 h. The product **6j** was purified by flash column chromatography (SiO₂, hexane/ethyl acetate, 100:0 to 80:20) and obtained in 55% yield (149.4 mg, 0.55 mmol) as a white solid.

¹H NMR (401 MHz, CDCl₃, δ): 1.13–1.33 (m, 3H), 1.37–1.51 (m, 2H), 1.61–1.69 (m, 1H), 1.71– 1.82 (m, 2H), 2.01–2.16 (m, 2H), 3.98–4.12 (m, 1H), 5.60 (brs, 1H), 7.21–7.26 (m, 1H), 7.29–7.33 (m, 2H). ¹³C NMR (100 MHz, CDCl₃, δ): 24.8 (*C*H₂), 25.4 (*C*H₂), 32.6 (*C*H₂), 48.7 (*C*H), 127.8 (*C*H), 130.3 (*C*H), 132.1 (*C*), 136.2 (*C*), 163.4 (*C*). HRMS-EI (m/z): [M]⁺ calcd for C₁₃H₁₅Cl₂NO, 271.0531; found, 271.0531.

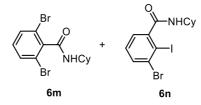
2-Bromo-N-cyclohexyl-6-fluorobenzamide (6k).



The first metalation step was conducted with **1q** (299.0 mg, 1.0 mmol) and THF (162 μ L, 2.0 mmol) for 1.5 h and the second amidation step was conducted with **5b** (249.7 mg, 2.0 mmol) for 0.5 h. The product **6k** was purified by flash column chromatography (SiO₂, hexane/ethyl acetate, 100:0 to 80:20) and obtained in 69% yield (208.4 mg, 0.69 mmol) as a white solid.

¹H NMR (401 MHz, CDCl₃, δ): 1.13–1.32 (m, 3H), 1.37–1.50 (m, 2H), 1.60–1.69 (m, 1H), 1.71– 1.81 (m, 2H), 2.04–2.15 (m, 2H), 3.97–4.10 (m, 1H), 5.64 (brs, 1H), 7.07 (td, J = 0.8, 8.6 Hz, 1H), 7.23 (td, J = 5.9, 8.2 Hz, 1H), 7.37 (d, J = 8.0 Hz, 1H). ¹³C NMR (100 MHz, CDCl₃, δ): 24.9 (CH₂), 25.5 (CH₂), 32.7 (CH₂), 48.9 (CH), 114.9 (d, $J_{C-F} = 22.0$ Hz, CH), 120.6 (d, $J_{C-F} = 4.8$ Hz, C), 127.8 (d, $J_{C-F} = 21.9$ Hz, C), 128.5 (d, $J_{C-F} = 3.8$ Hz, CH), 131.1 (d, $J_{C-F} = 8.6$ Hz, CH), 159.2 (d, $J_{C-F} = 251.7$ Hz, C), 162.4 (C). ¹⁹F NMR (377 MHz, CDCl₃, δ): –113.4 (s, 1F). HRMS-EI (m/z): [M]⁺ calcd for C₁₃H₁₅BrFNO, 299.0321; found, 299.0313.

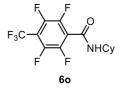
2,6-Dibromo-N-cyclohexylbenzamide (6m).



The first metalation step was conducted with 1r (361.7 mg, 1.0 mmol) and THF (162 µL, 2.0 mmol) for 1.5 h and the second amidation step was conducted with **5b** (249.7 mg, 2.0 mmol) for 0.5 h. The product **6m** and the byproduct **6n** were obtained in 54% NMR yield (**6m**:**6n** = 78:22).

¹H NMR for **6m** (401 MHz, CDCl₃, δ): 1.14–1.34 (m, 3H), 1.37–1.51 (m, 2H), 1.60–1.69 (m, 1H), 1.72–1.82 (m, 2H), 2.03–2.17 (m, 2H), 3.98–4.11 (m, 1H), 5.56 (brs, 1H), 7.09 (t, *J* = 8.0 Hz, 1H), 7.52 (d, *J* = 7.6 Hz, 2H). ¹³C NMR for **6m** (100 MHz, CDCl₃, δ): 24.9 (*C*H₂), 25.6 (*C*H₂), 32.8 (*C*H₂), 48.8 (*C*H), 120.5 (*C*), 131.2 (*C*H), 131.8 (*C*H), 140.0 (*C*), 165.3 (*C*). HRMS-EI (m/z): [M]⁺ calcd for C₁₃H₁₅Br₂NO, 358.9520; found, 358.9517.

N-Cyclohexyl-2,3,5,6-tetrafluoro-4-(trifluoromethyl)benzamide (60).



The first step was conducted with **1t** (296.2 mg, 1.0 mmol) and MTBE (236 μ L, 2.0 mmol) for 1.5 h and the second amidation step was conducted with **5b** (249.2 mg, 2.0 mmol) for 0.5 h. The product **6o** was purified by flash column chromatography (SiO₂, hexane/ethyl acetate, 100:0 to 80:20) and obtained in 69% yield (235.1 mg, 0.68 mmol) as a white solid.

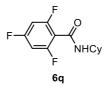
¹H NMR (401 MHz, CDCl₃, δ): 1.18–1.32 (m, 3H), 1.37–1.50 (m, 2H), 1.62–1.70 (m, 1H), 1.72–1.80 (m, 2H), 1.99–2.10 (m, 2H), 3.95–4.07 (m, 1H), 5.80 (brs, 1H). ¹³C NMR (100 MHz, DMSO-*d*₆, δ): 24.2 (*C*H₂), 25.1 (*C*H₂), 32.0 (*C*H₂), 48.7 (*C*H), 108.0–109.3 (m, *C*), 120.4 (d, *J*_{C-F} = 229.8 Hz, *C*), 121.9 (d, *J*_{C-F} = 21.9 Hz, *C*), 143.3 (dm, *J*_{C-F} = 262.3 Hz, *C*), 155.1 (*C*). ¹⁹F NMR (377 MHz, CDCl₃, δ): –139.1 (s, 2F), –138.5 (q, *J* = 23.1 Hz, 2F), –56.6 (t, *J* = 23.2 Hz, 3F). HRMS-EI (m/z): [M]⁺ calcd for C₁₄H₁₂F₇NO, 343.0807; found, 343.0799.

N-Cyclohexyl-2,3,5,6-tetrafluorobenzamide (6p).



The first metalation step was conducted with **1u** (229.1 mg, 1.0 mmol) and THF (162 μ L, 2.0 mmol) for 90 min and the second amidation step was conducted with **5b** (249.7 mg, 2.0 mmol) for 30 min. The product **6p** was purified by flash column chromatography (SiO₂, hexane/ethyl acetate, 100:0 to 80:20) and obtained in 45% yield (124.6 mg, 0.45 mmol) as a white solid.

¹H NMR (392 MHz, CDCl₃, δ): 1.14–1.32 (m, 3H), 1.36–1.49 (m, 2H), 1.61–1.70 (m, 1H), 1.71– 1.81 (m, 2H), 2.01–2.11 (m, 2H), 3.96–4.07 (m, 1H), 5.77 (brs, 1H), 7.07–7.17 (m, 1H). ¹³C NMR (100 MHz, CDCl₃, δ): 24.8 (CH₂), 25.5 (CH₂), 32.8 (CH₂), 49.4 (CH), 107.2 (t, $J_{C-F} = 22.4$ Hz, CH), 117.7 (t, $J_{C-F} = 19.1$ Hz, C), 144.6 (dm, $J_{C-F} = 495.4$ Hz, C), 144.5–145.0 (m, C), 157.3 (C). ¹⁹F NMR (377 MHz, CDCl₃, δ): –142.5 (s, 2F), –138.3 (s, 2F). HRMS-EI (m/z): [M]⁺ calcd for C₁₃H₁₃F₄NO, 275.0933; found, 275.0932. N-Cyclohexyl-2,4,6-trifluorobenzamide (6q).

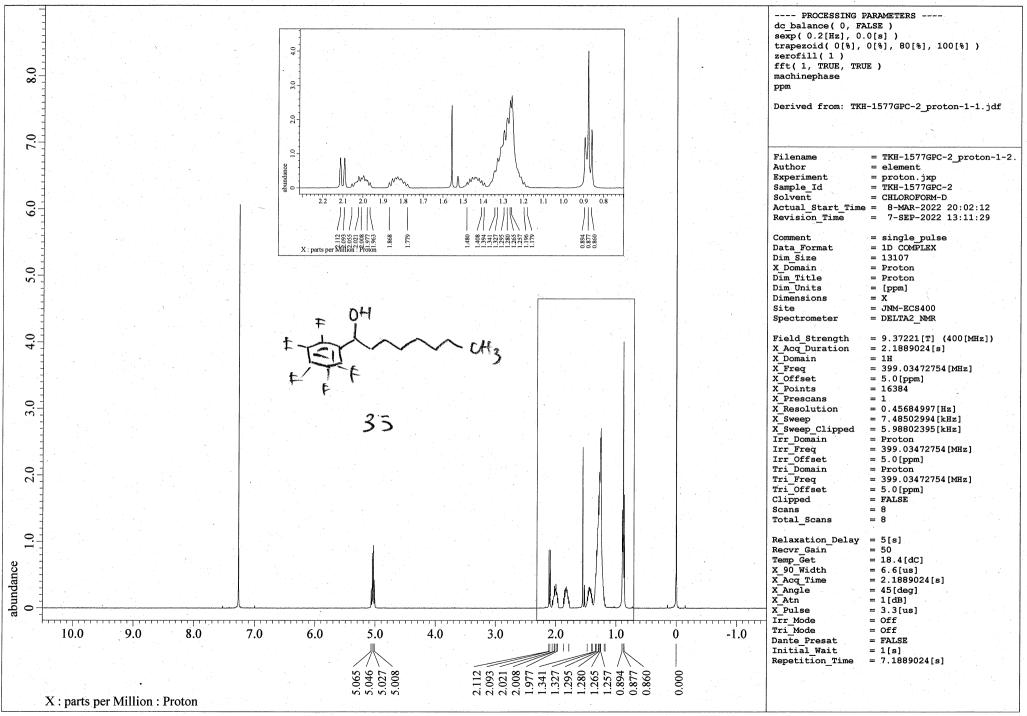


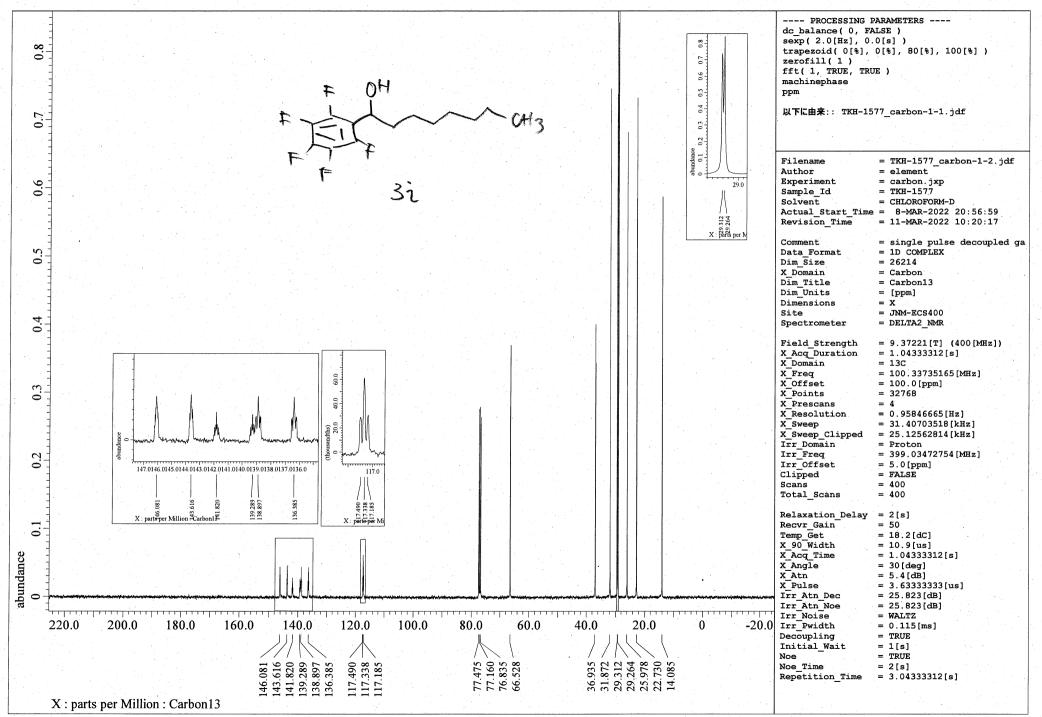
The first metalation step was conducted with 1v (211.4 mg, 1.0 mmol) and THF (162 μ L, 2.0 mmol) for 3 h and the second amidation step was conducted with 5b (250.4 mg, 2.0 mmol) for 0.5 h. The product 6q was purified by flash column chromatography (SiO₂, hexane/ethyl acetate, 100:0 to 85:15) and obtained in 39% yield (101.4 mg, 0.39 mmol) as a white solid.

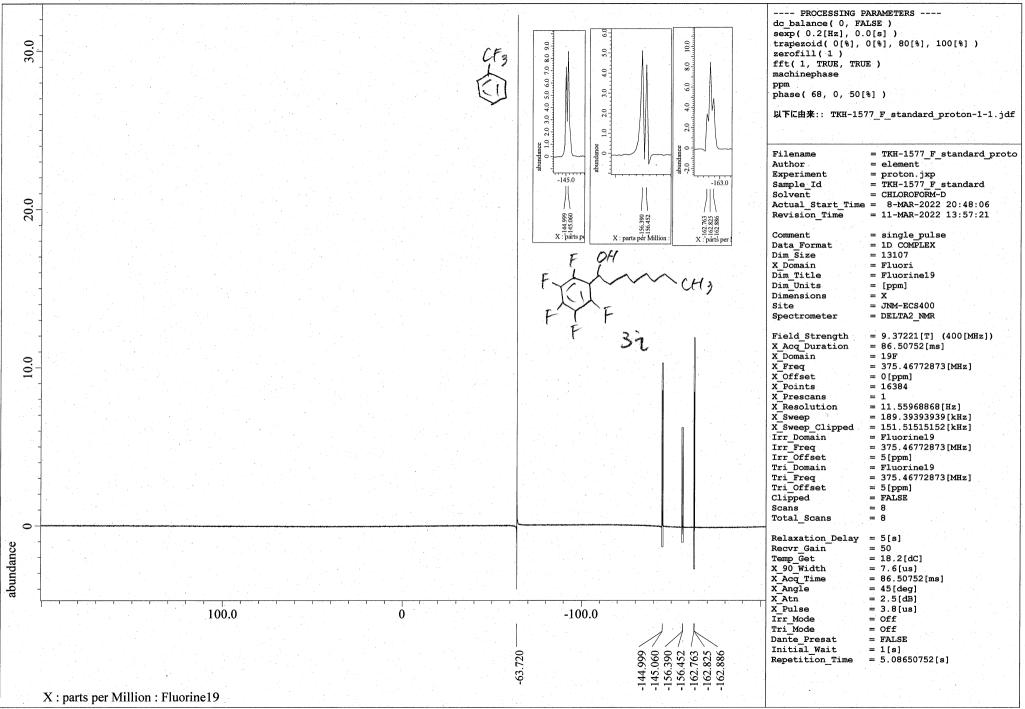
¹H NMR (401 MHz, CDCl₃, δ): 1.14–1.32 (m, 3H), 1.35–1.49 (m, 2H), 1.60–1.69 (m, 1H), 1.70–1.80 (m, 2H), 1.99–2.09 (m, 2H), 3.94–4.05 (m, 1H), 5.74 (brs, 1H), 6.66–6.75 (m, 2H). ¹³C NMR (100 MHz, CDCl₃, δ): 24.8 (*C*H₂), 25.5 (*C*H₂), 32.8 (*C*H₂), 49.1 (*C*H), 100.7 (td, $J_{C-F} = 3.5$, 26.2 Hz, *C*H), 111.7 (td, $J_{C-F} = 4.1$, 21.2 Hz, *C*), 158.7 (*C*), 160.4 (ddd, $J_{C-F} = 9.8$, 15.1, 253.0 Hz, *C*), 163.2 (dt, $J_{C-F} = 15.3$, 252.7 Hz, *C*). ¹⁹F NMR (377 MHz, CDCl₃, δ): –110.1 (s, 2F), –105.4 (s, 1F). HRMS-EI (m/z): [M]⁺ calcd for C₁₃H₁₄F₃NO, 257.1028; found, 257.1025.

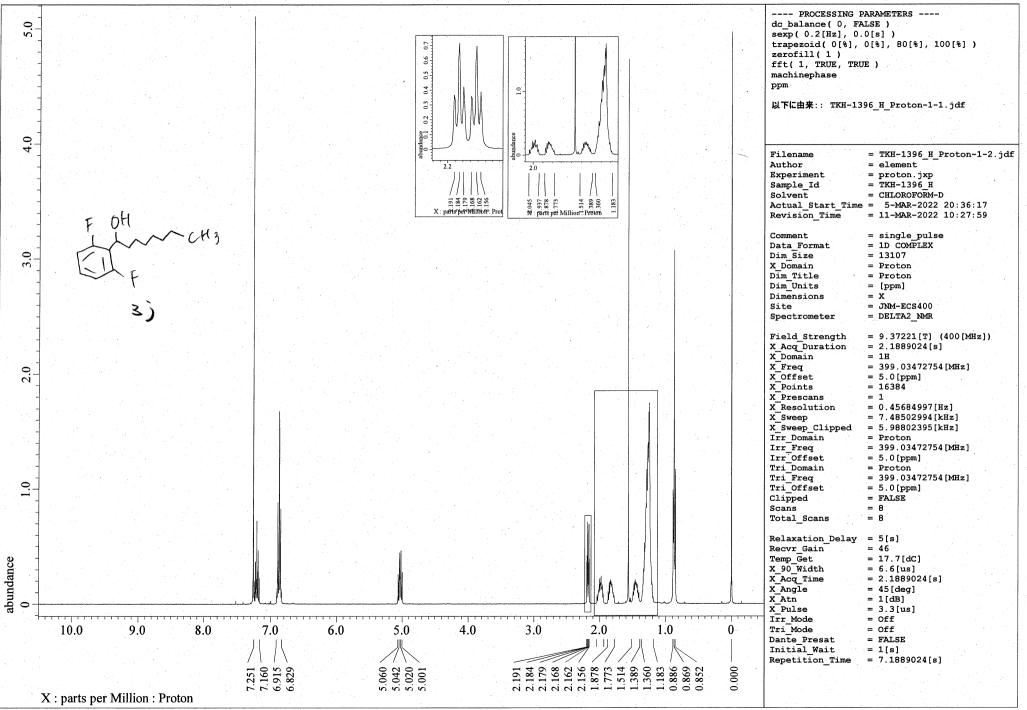
9. References

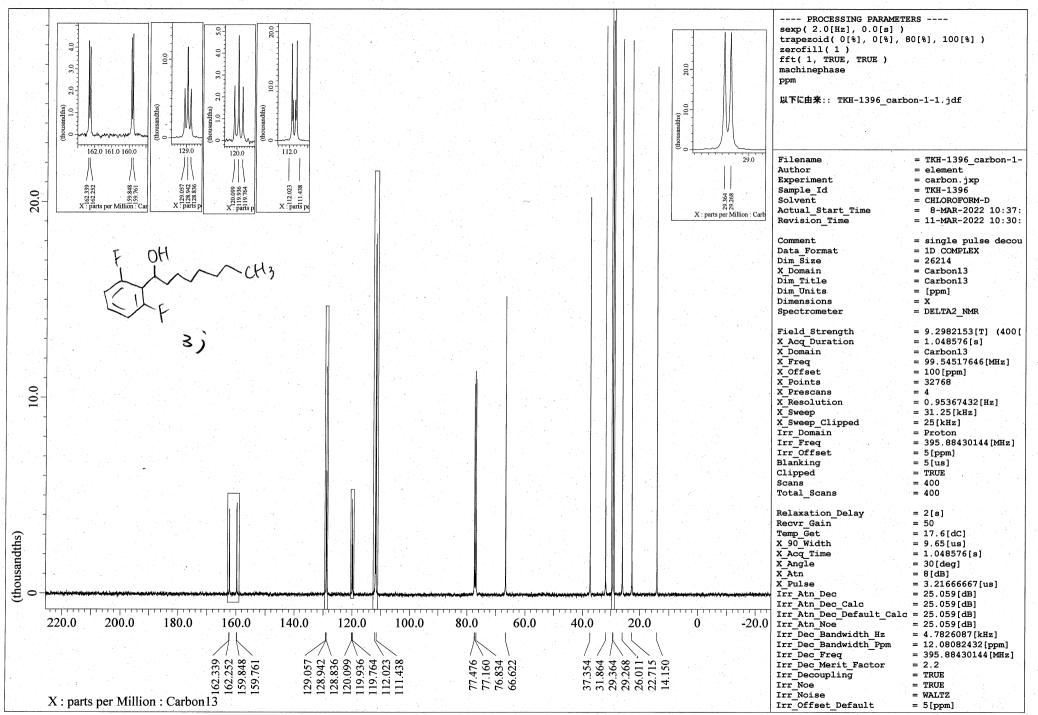
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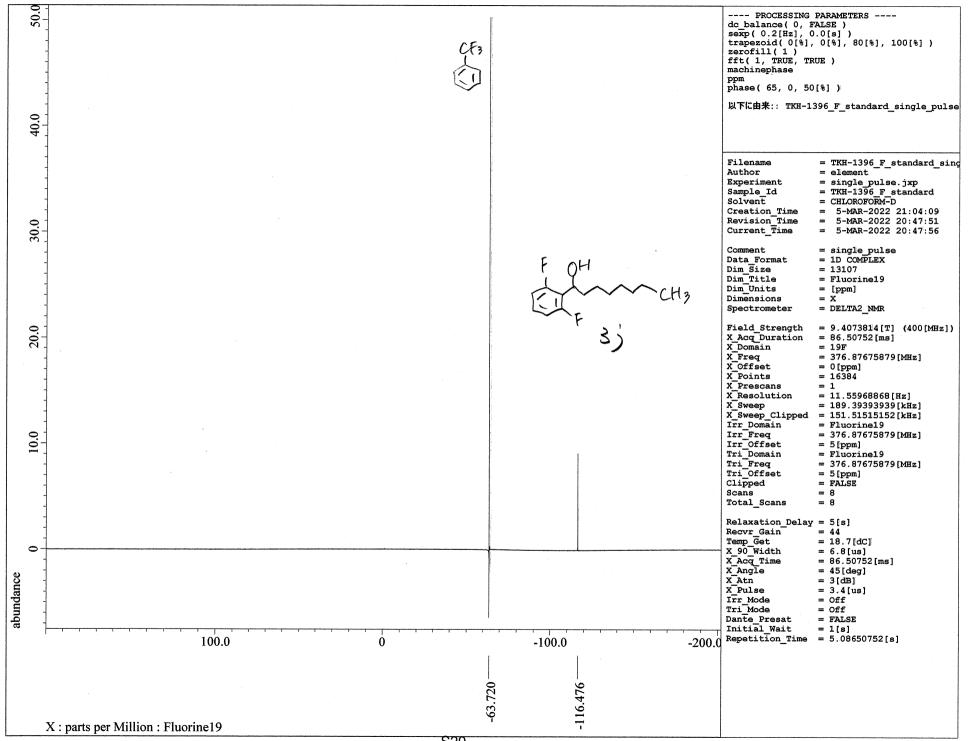


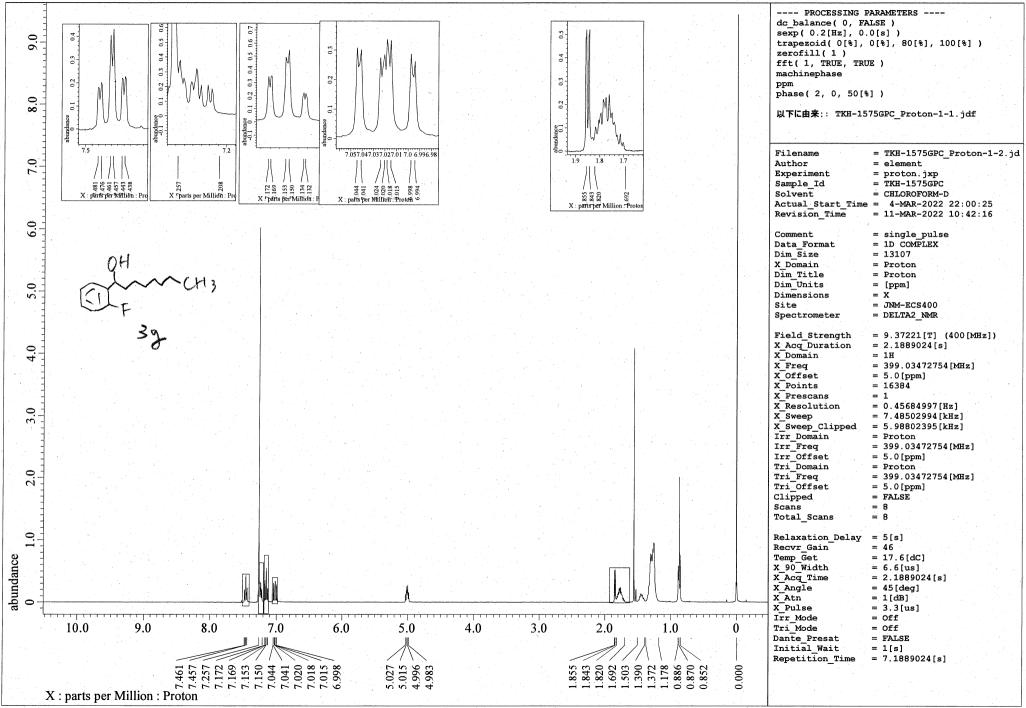


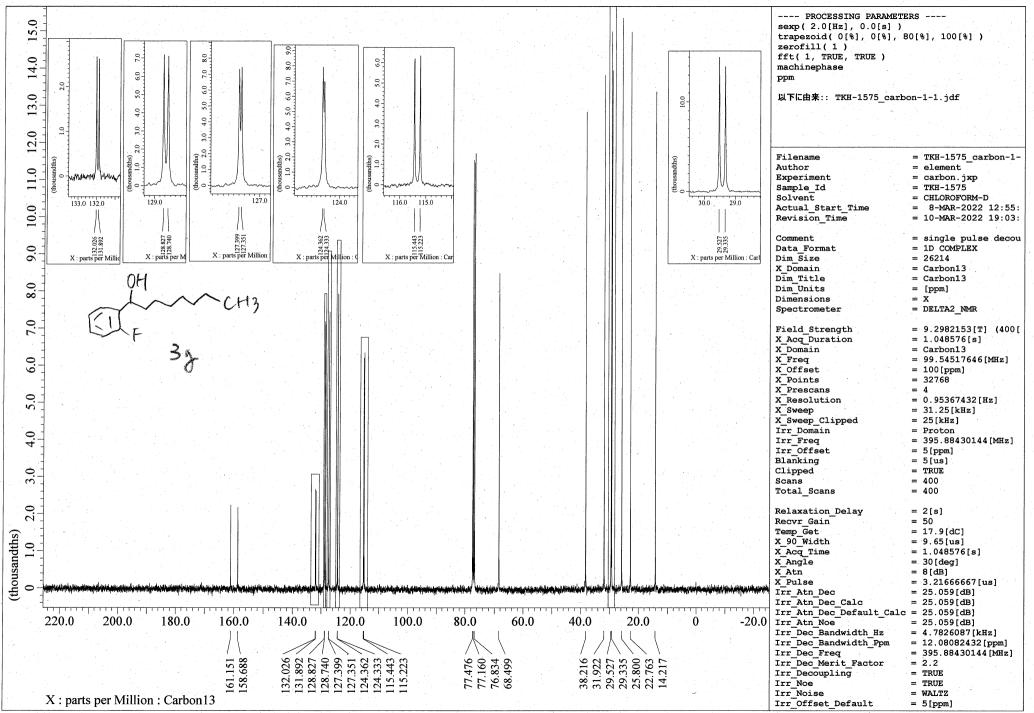


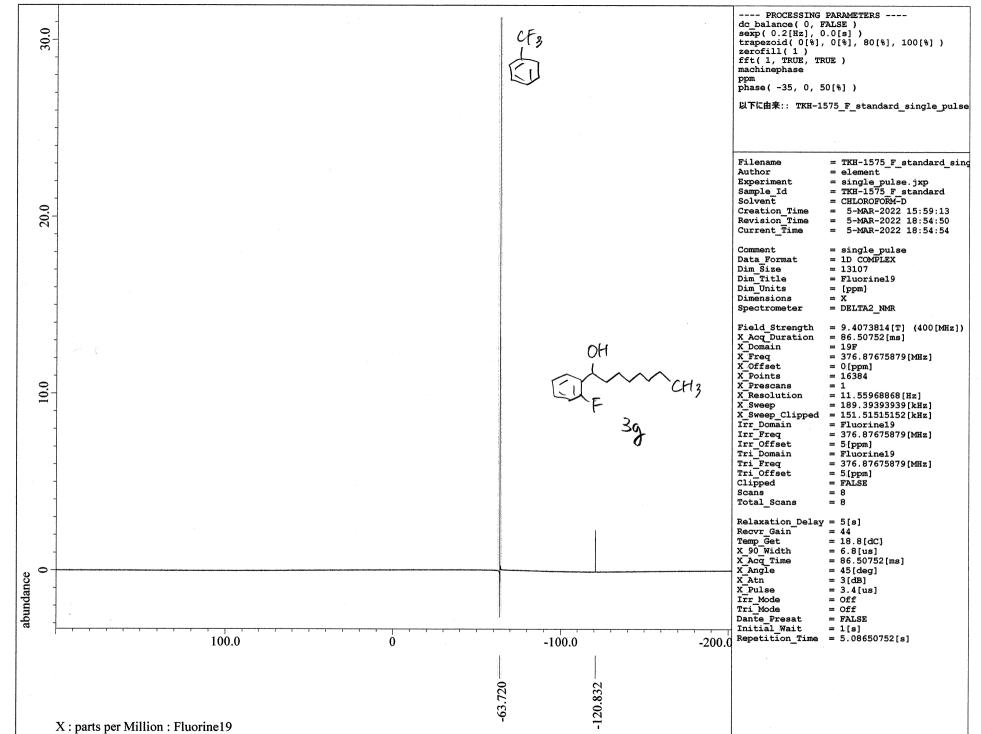


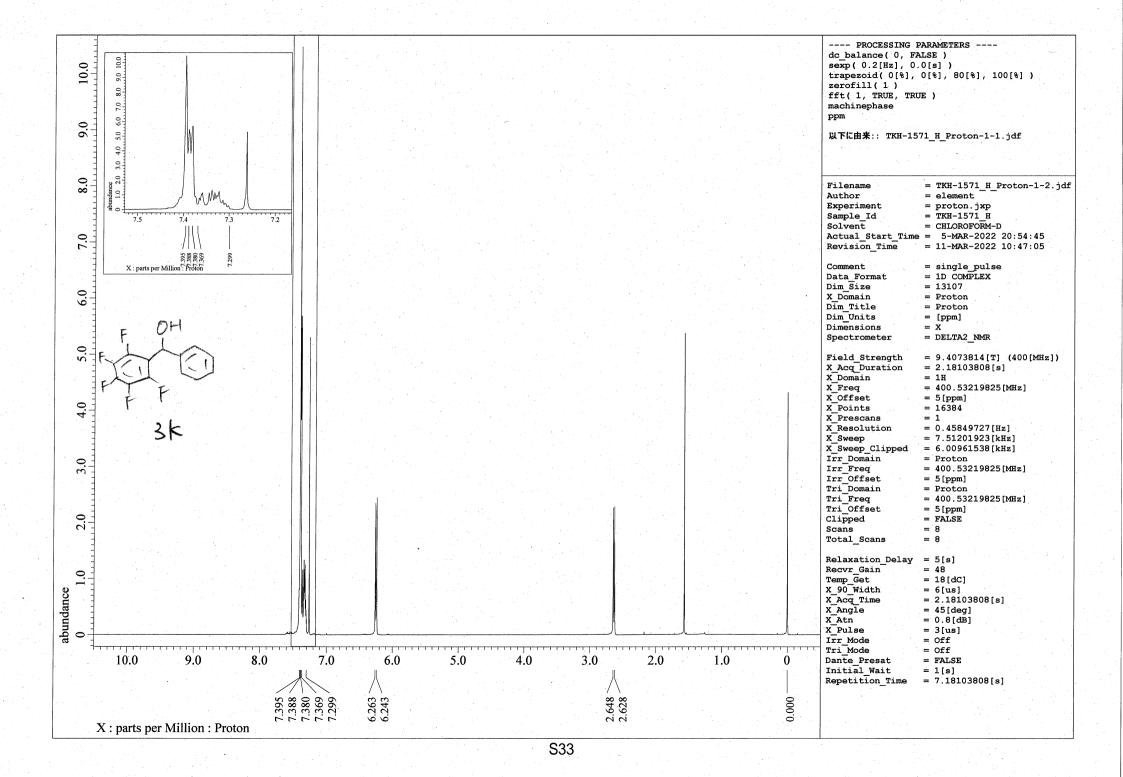


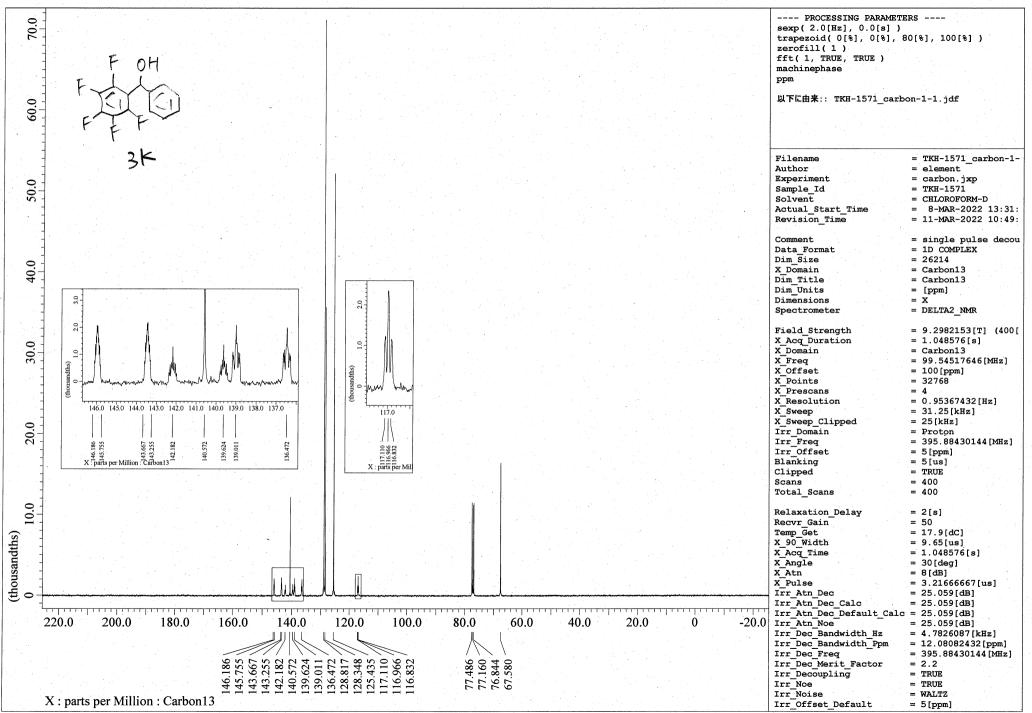


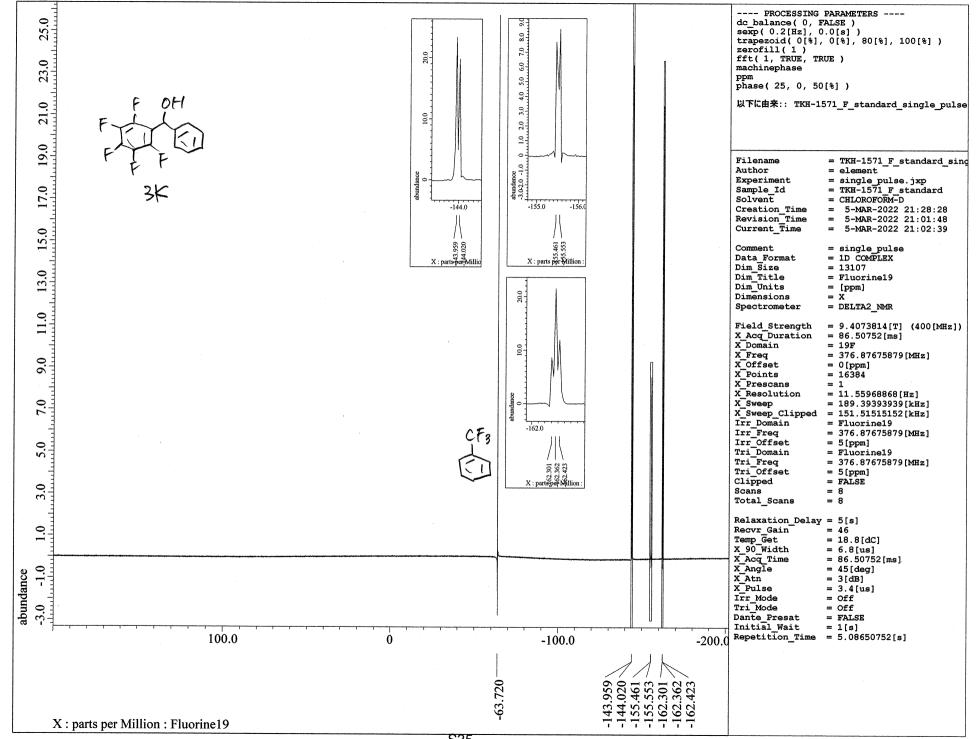


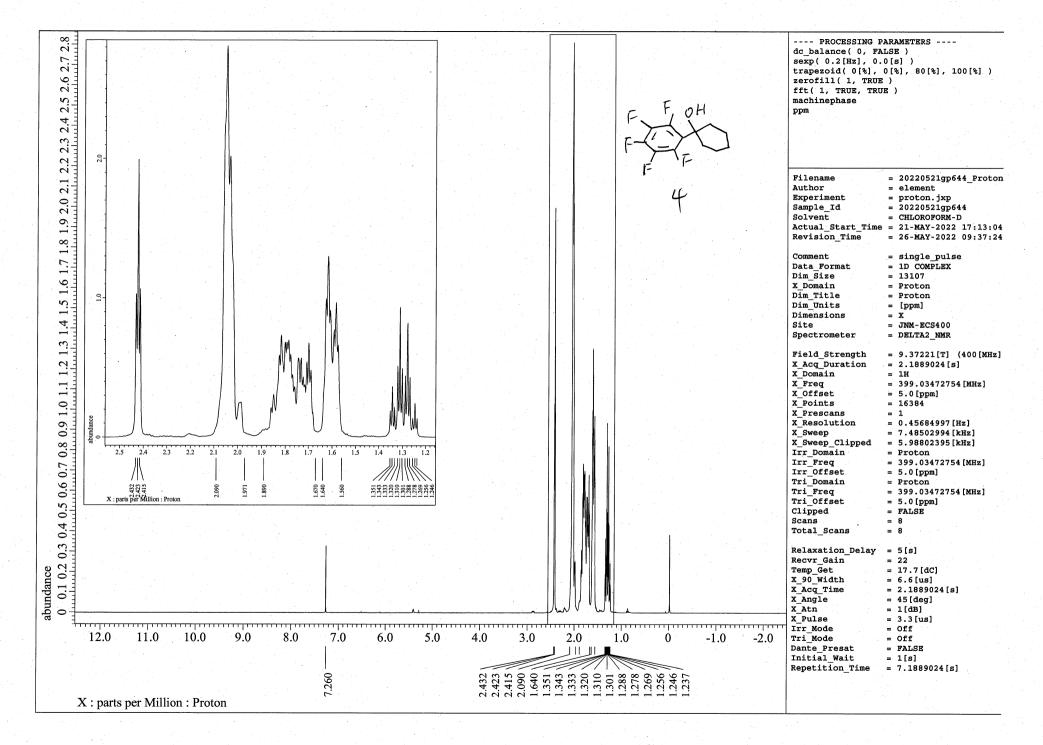


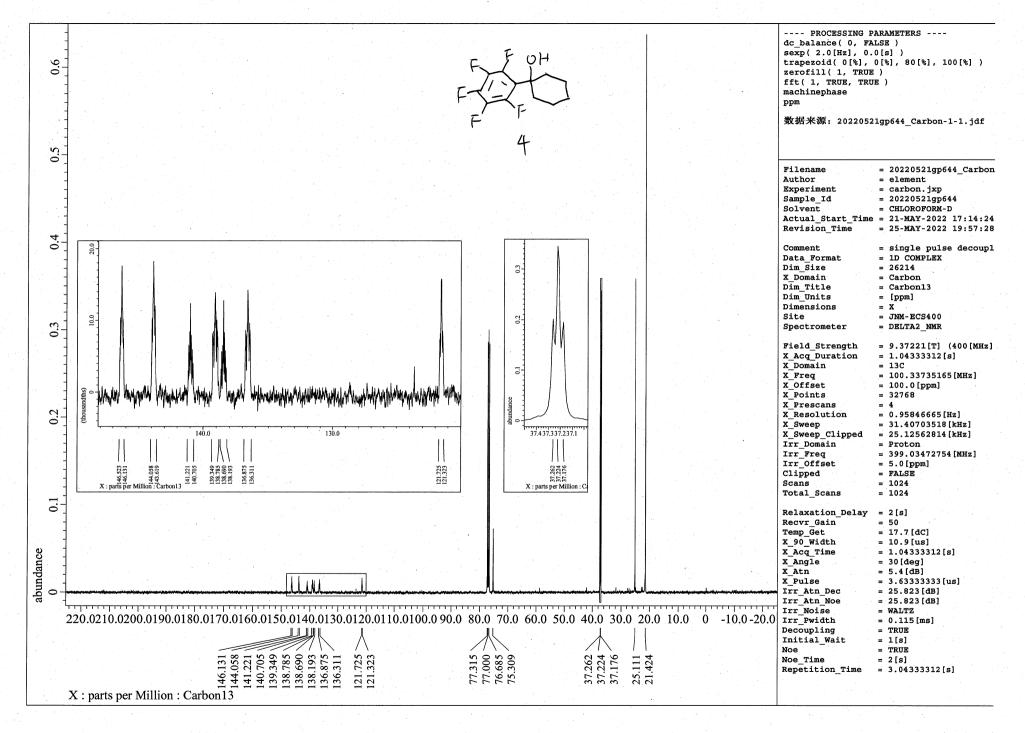


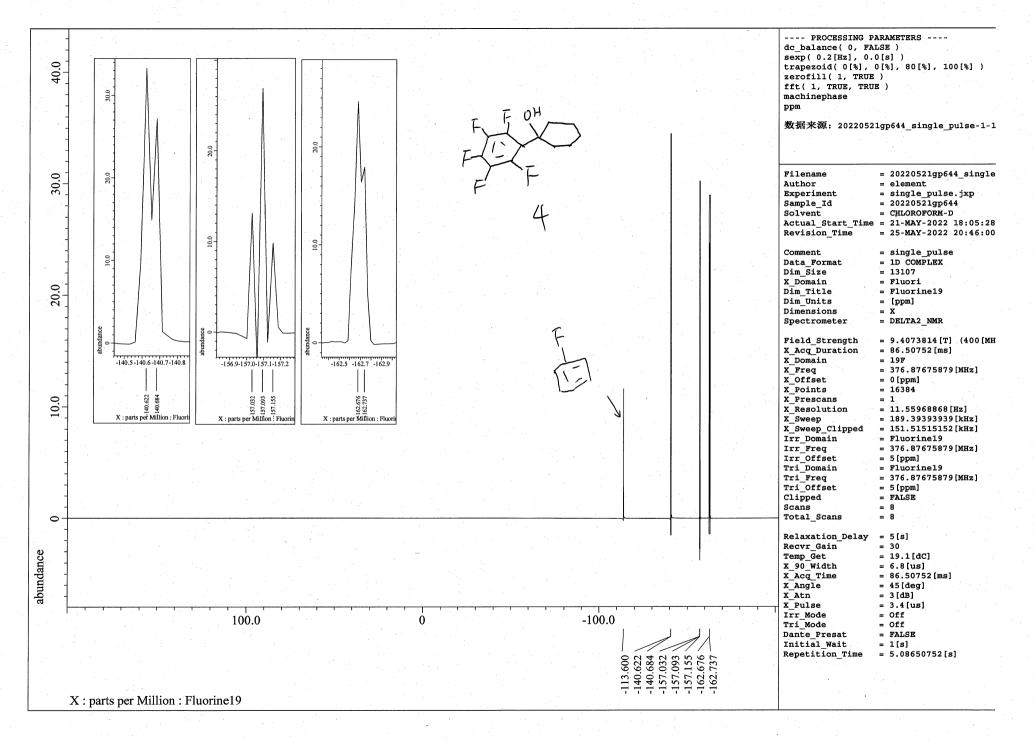


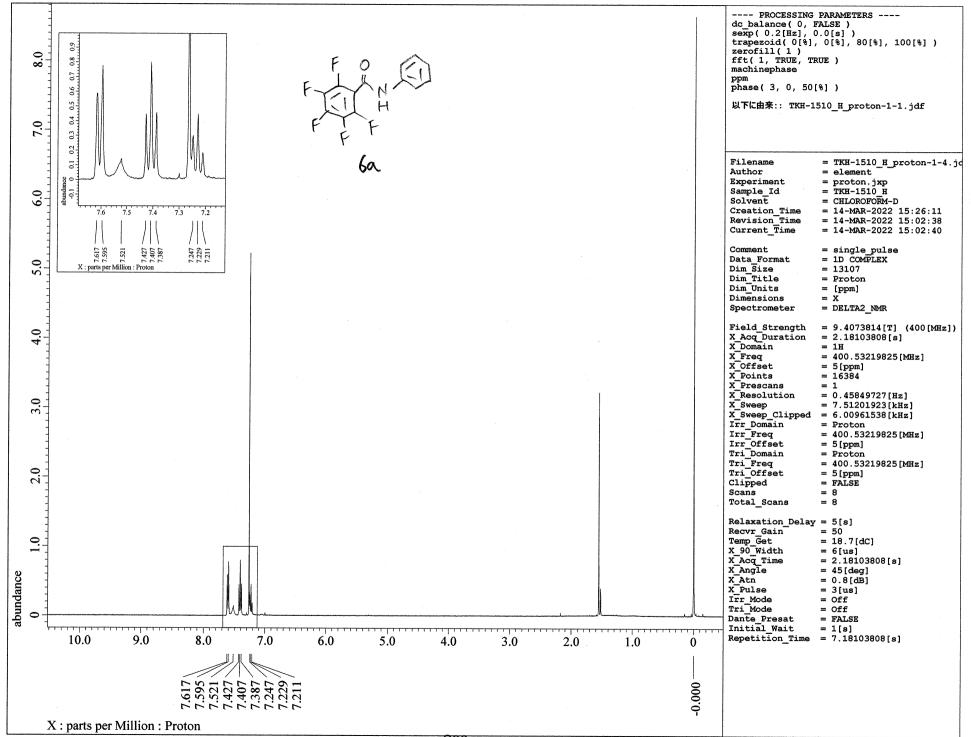




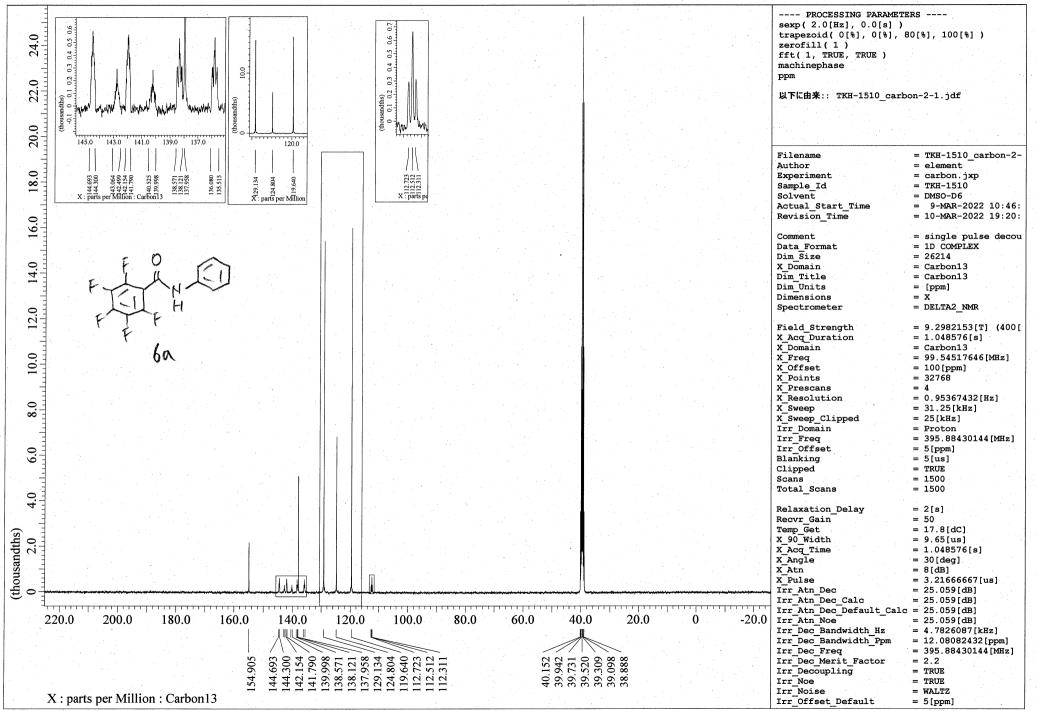


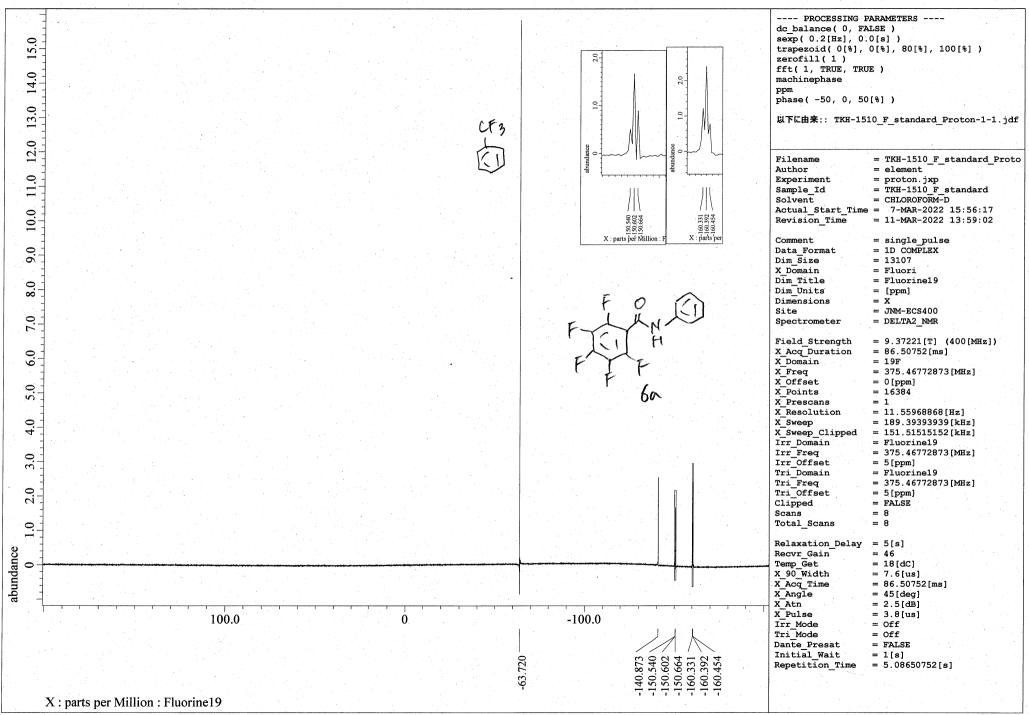


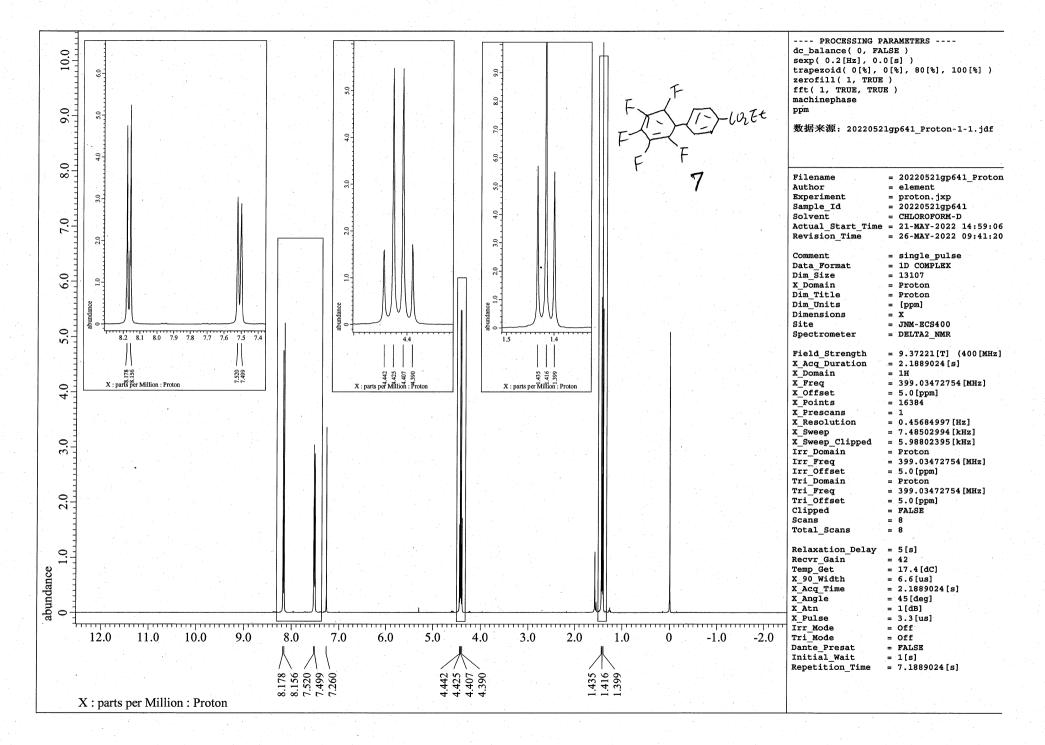


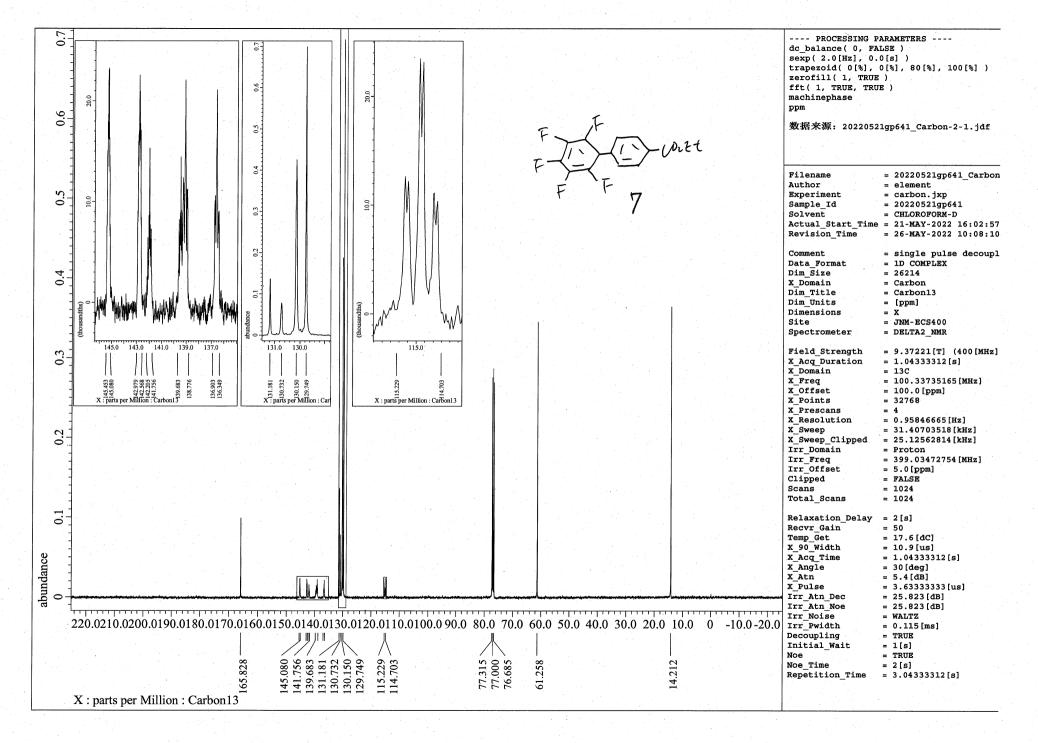


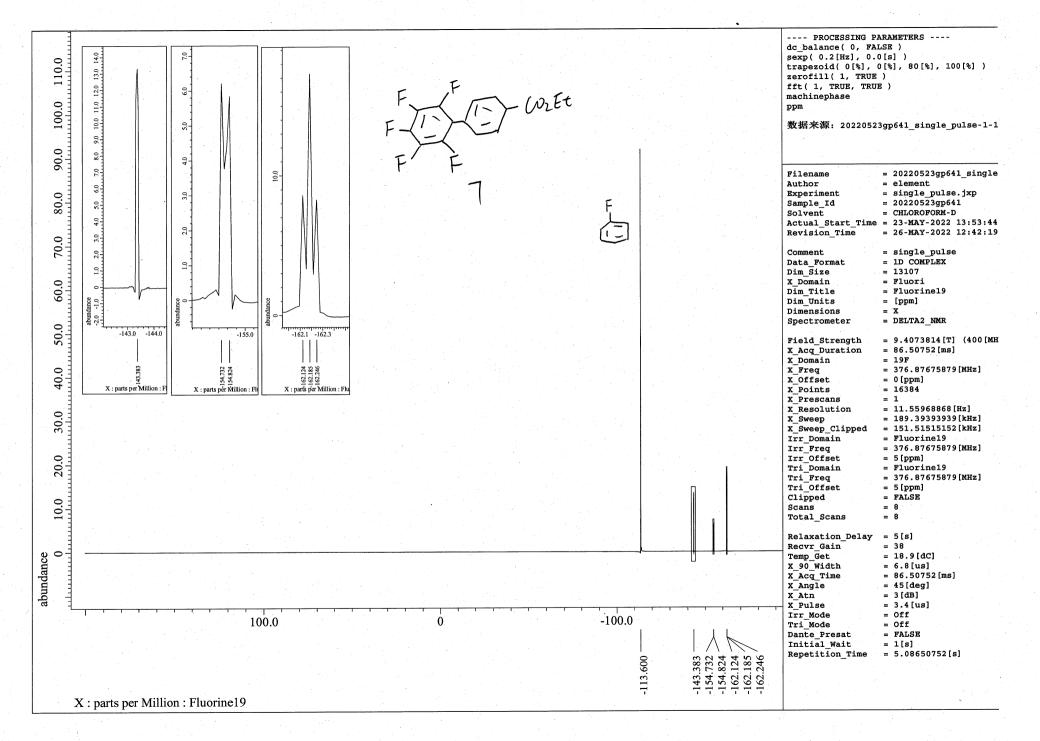
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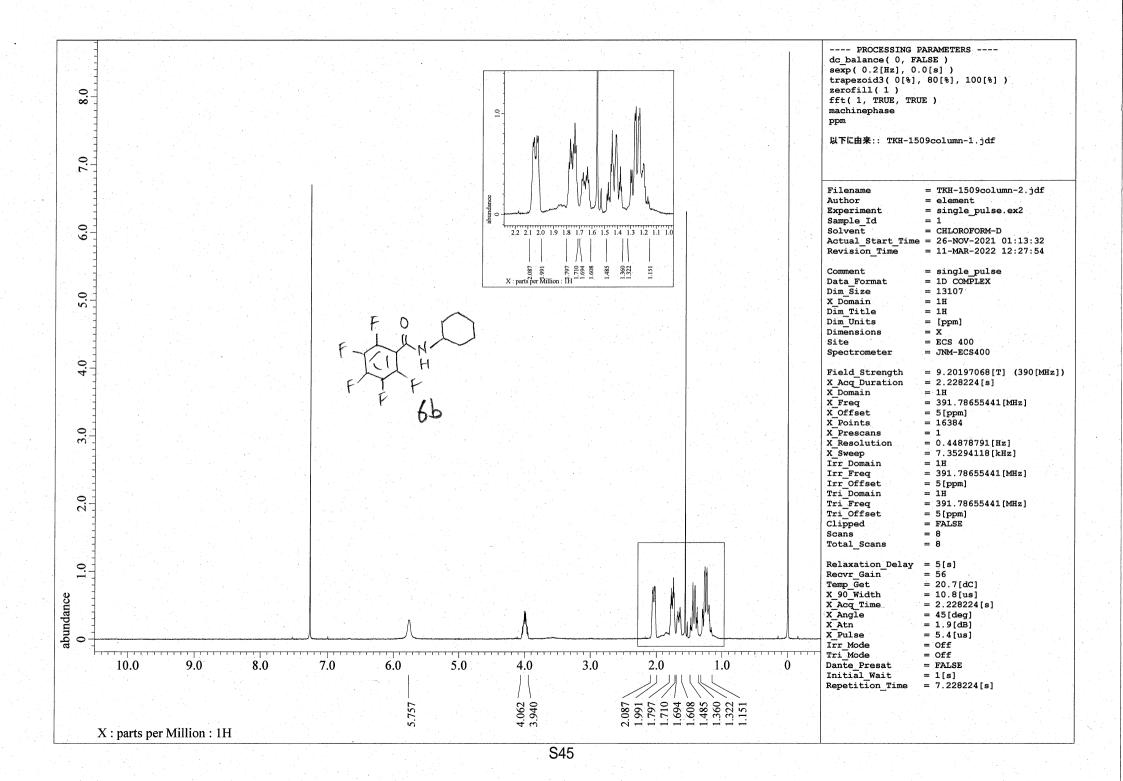


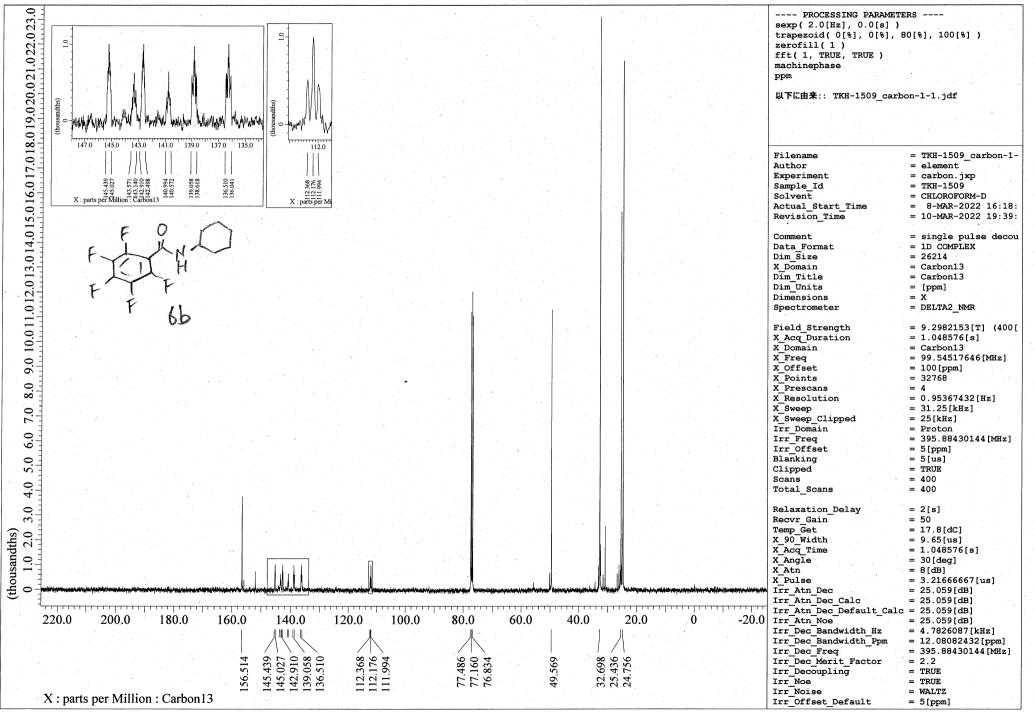


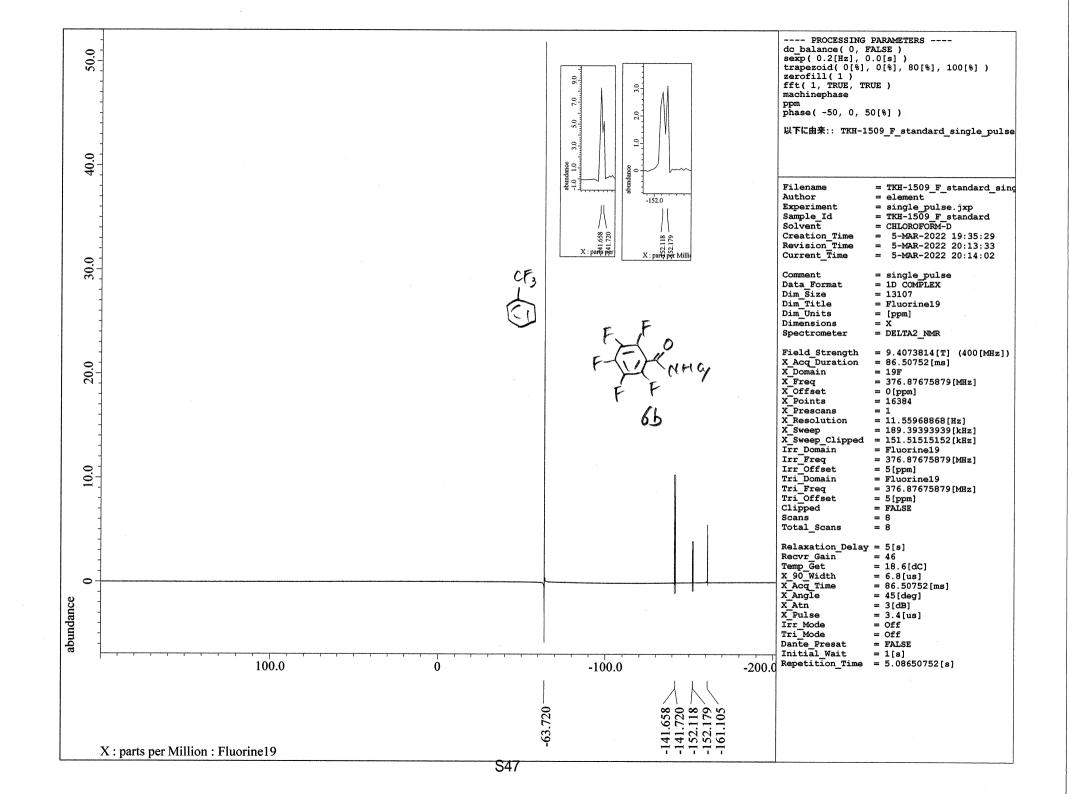


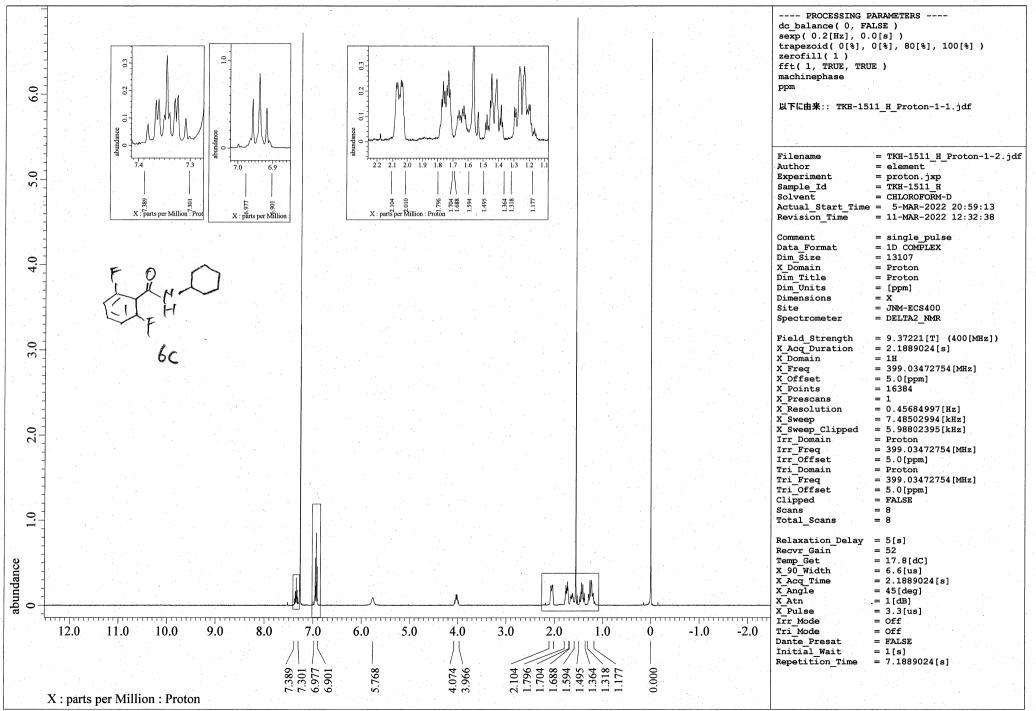


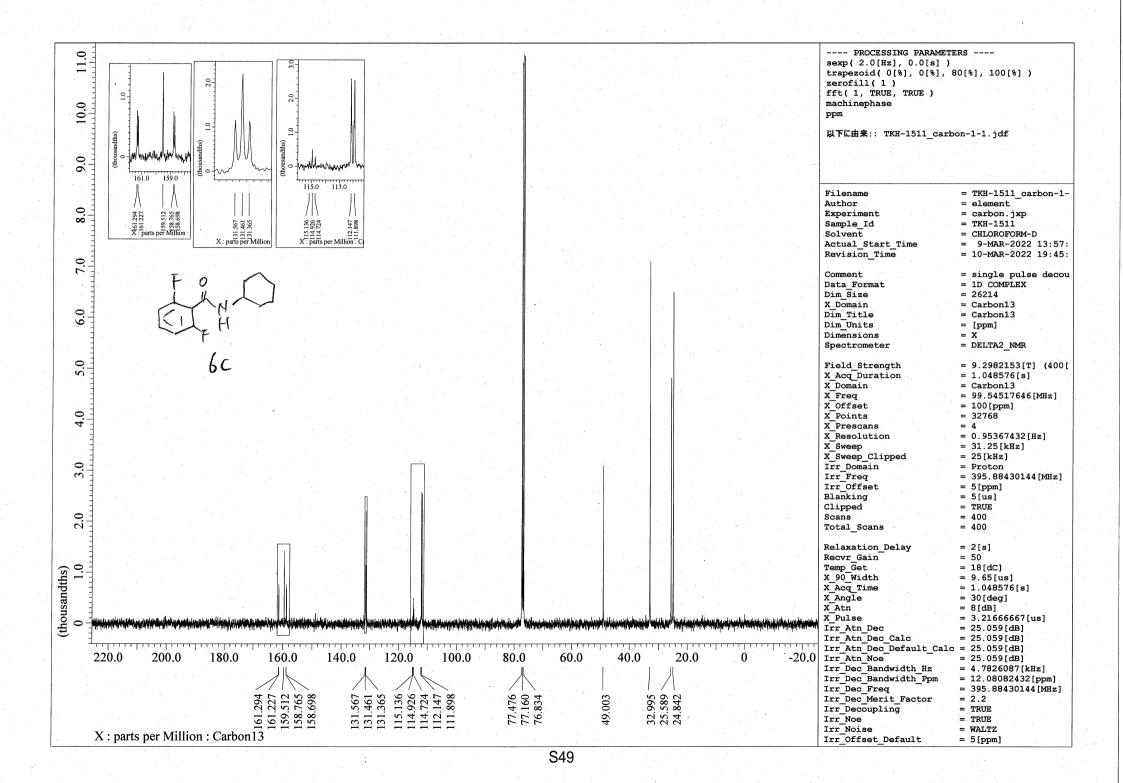


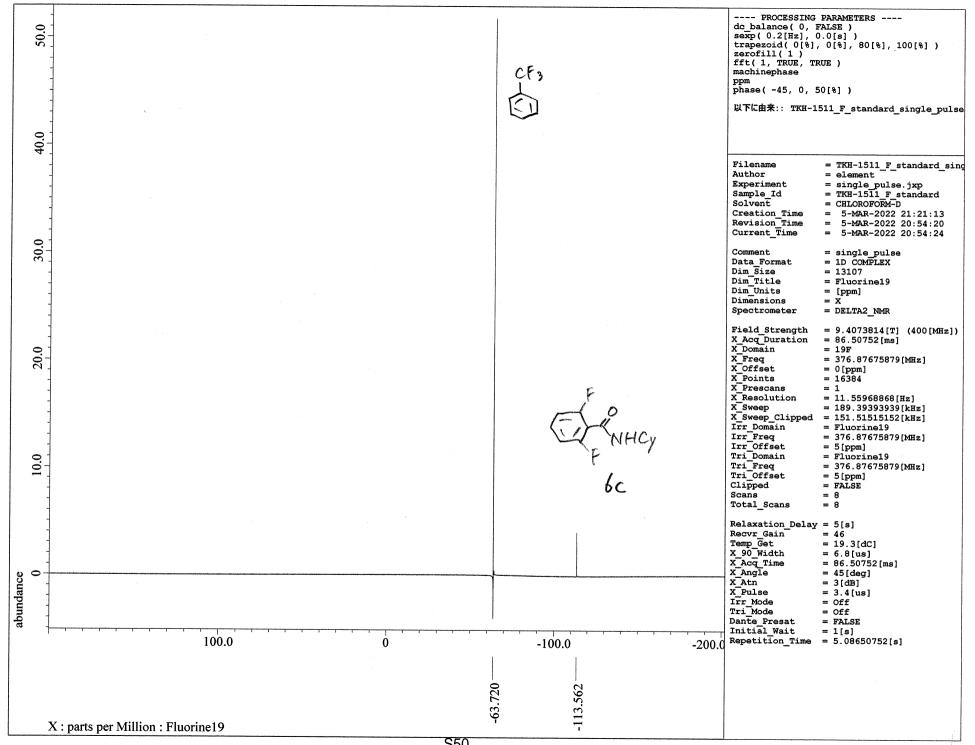


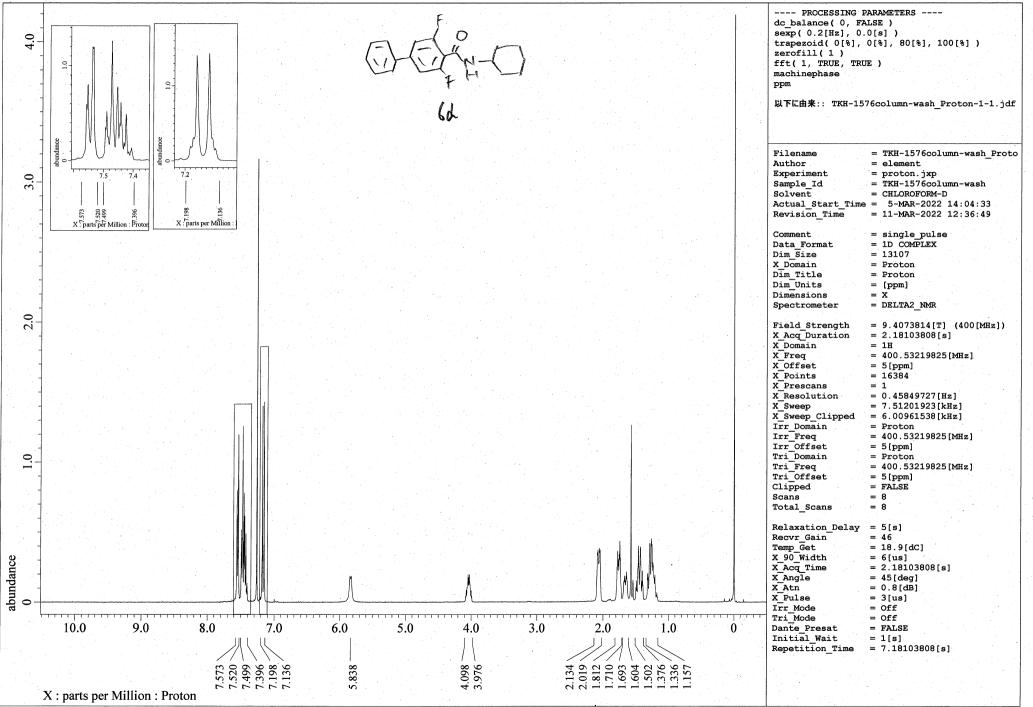


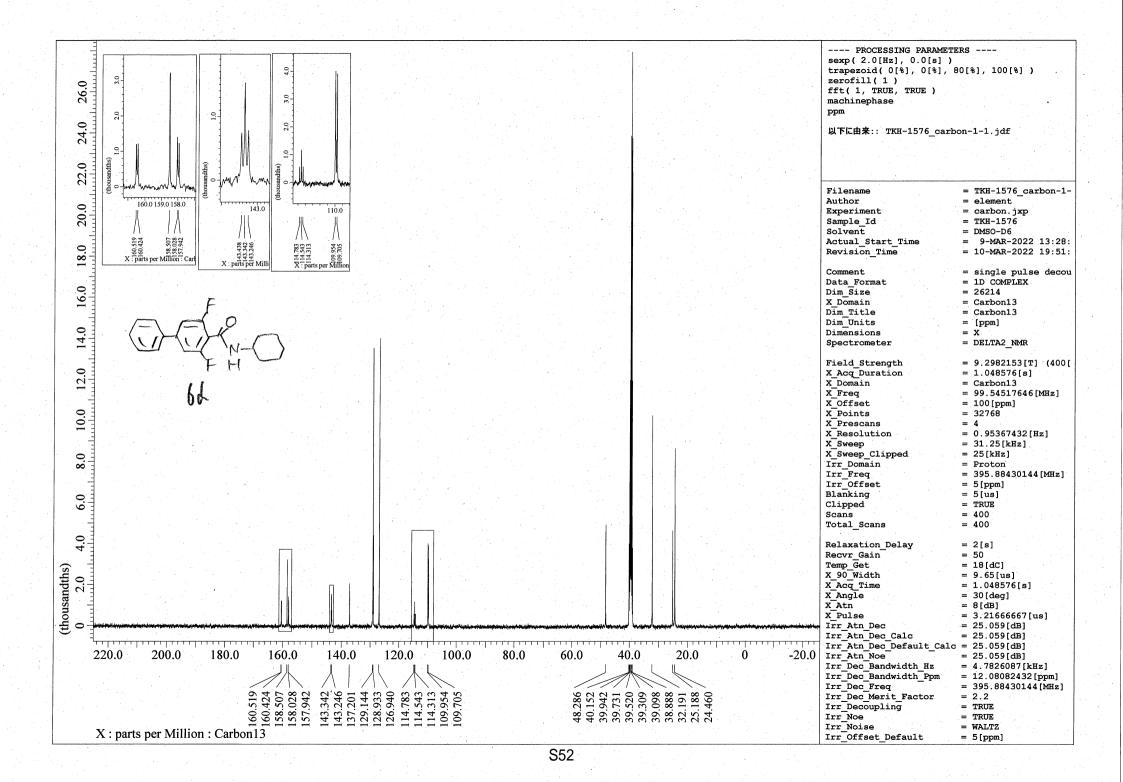


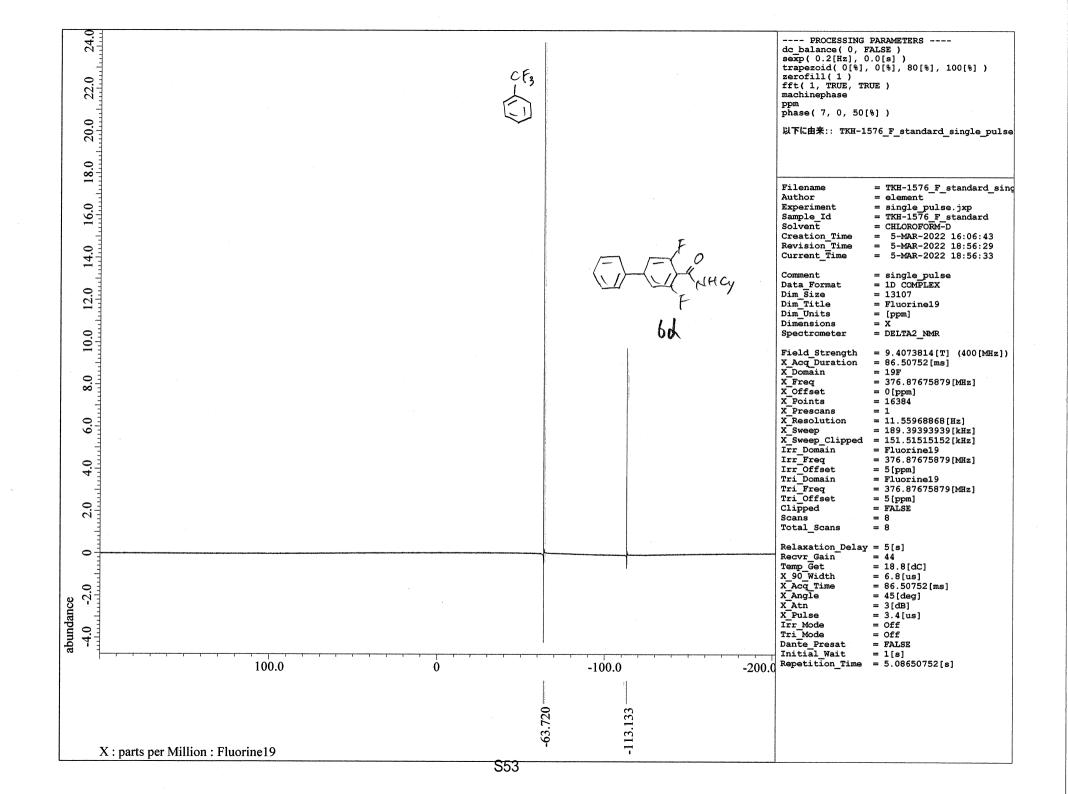


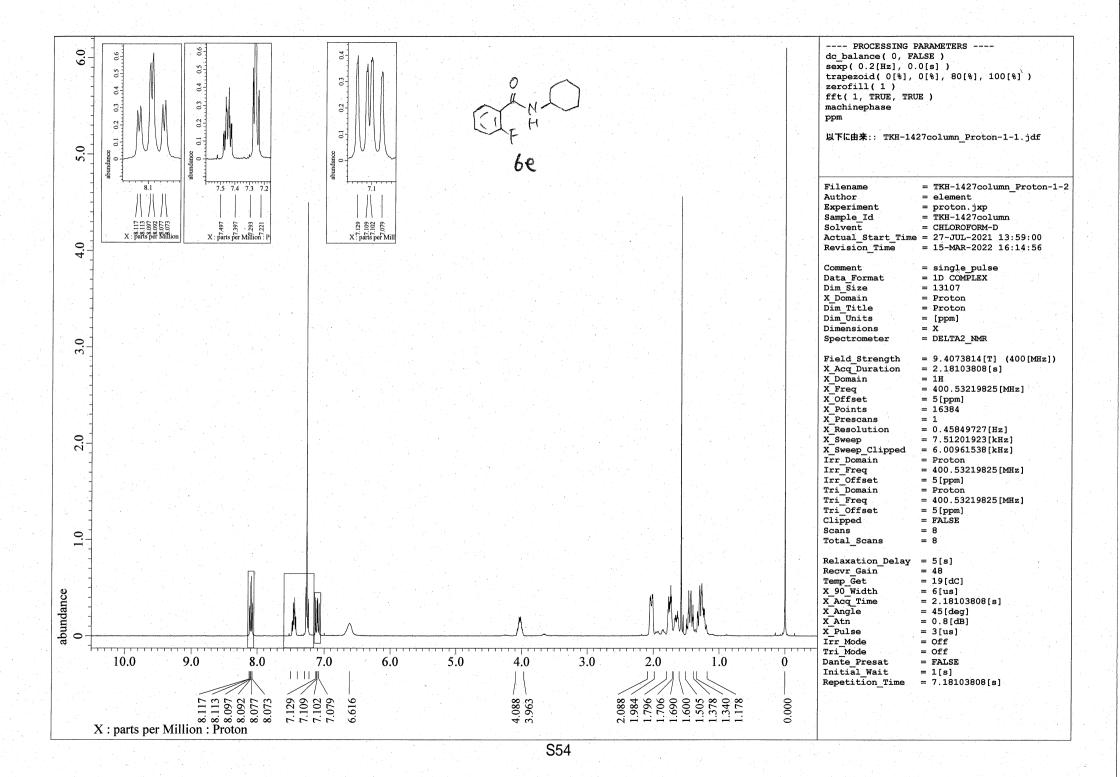


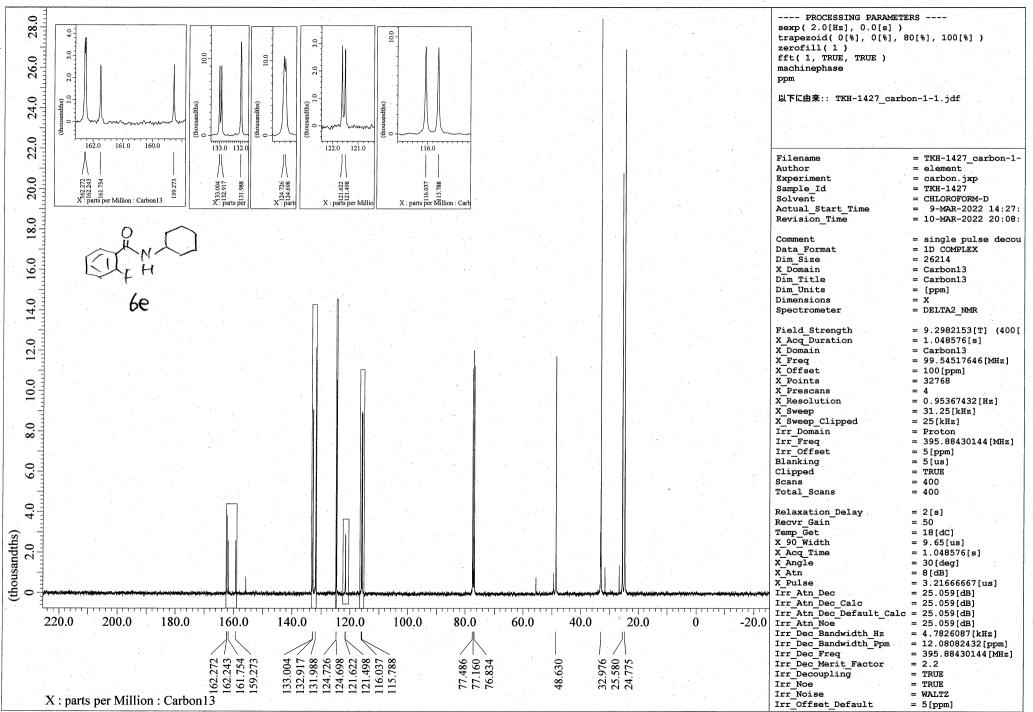




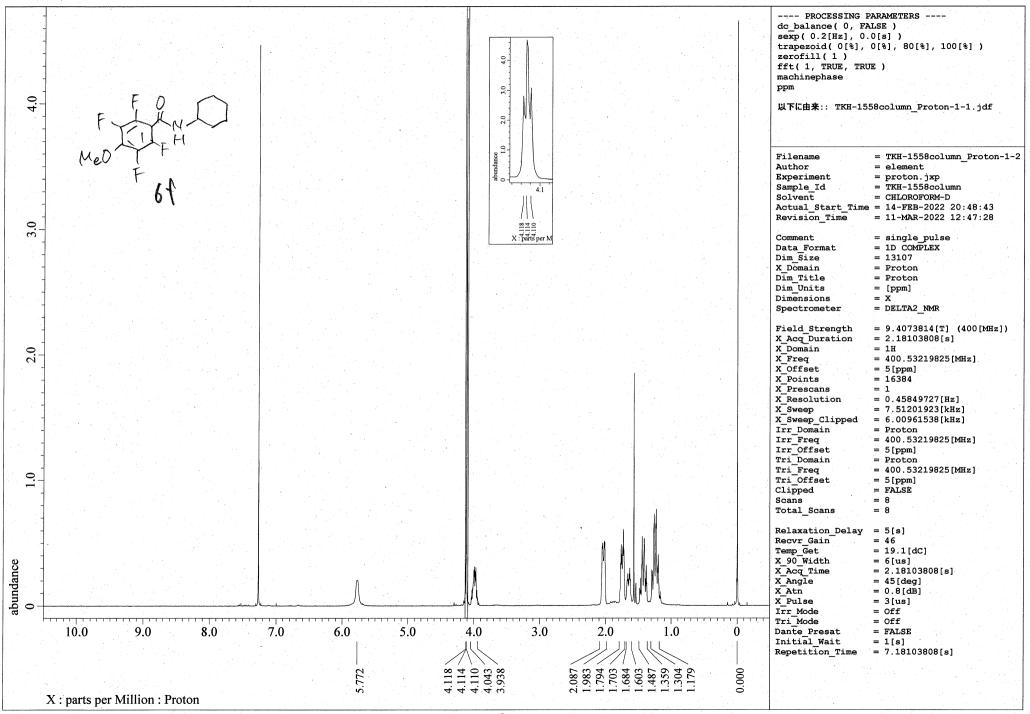


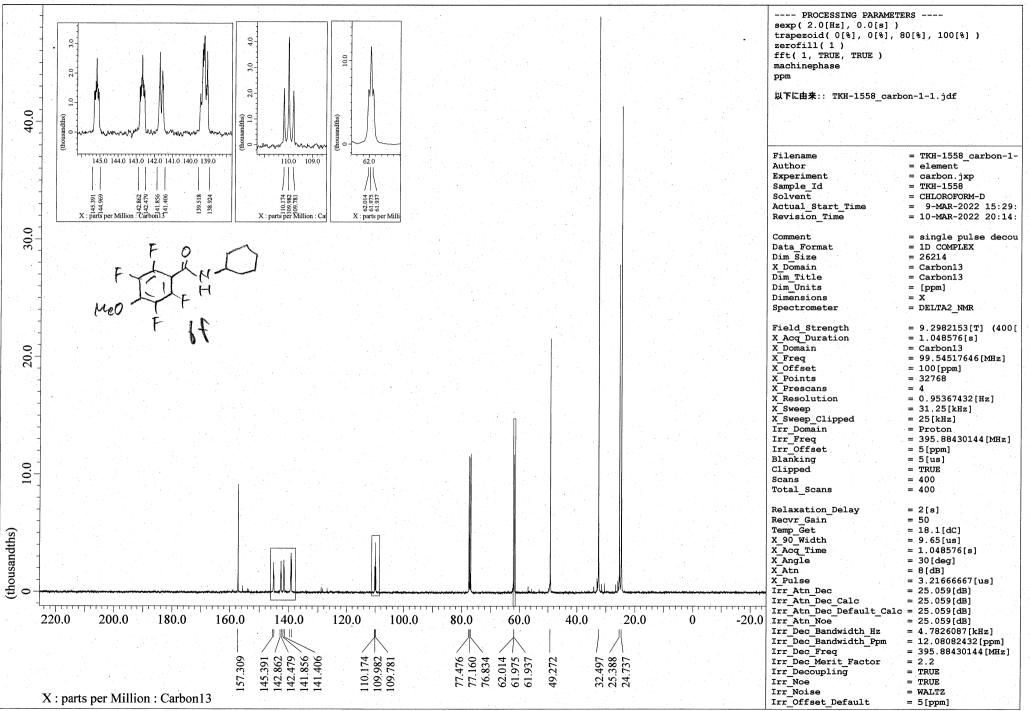


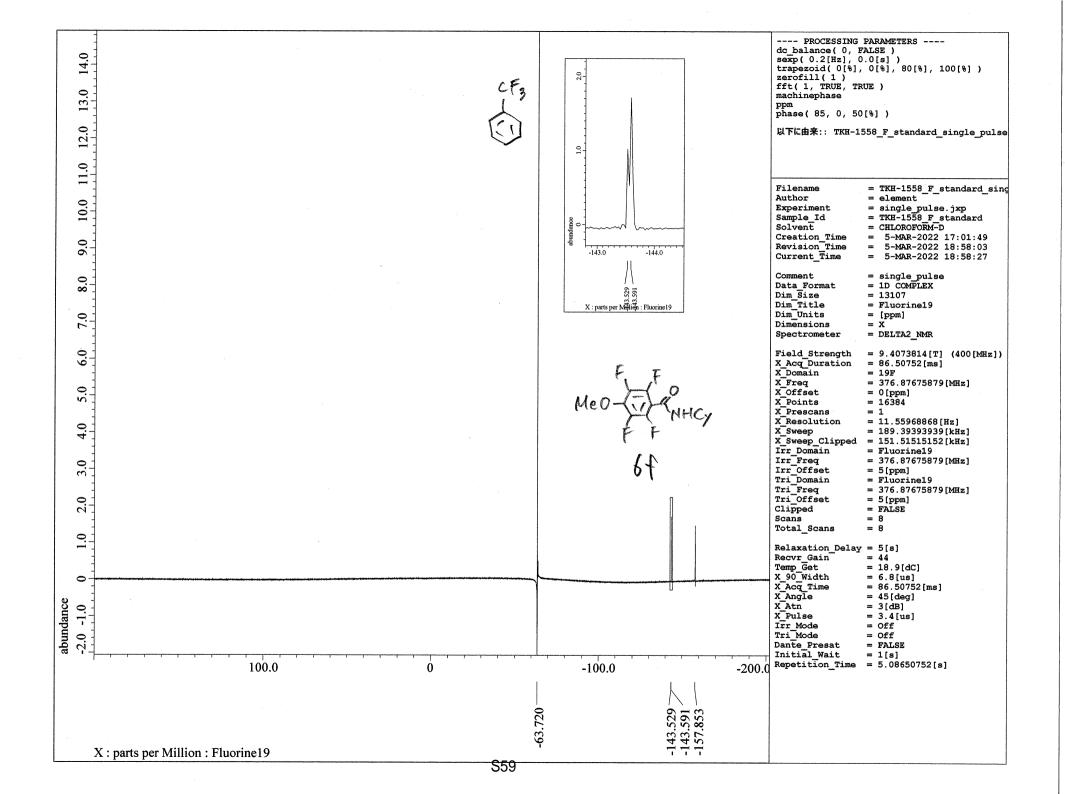


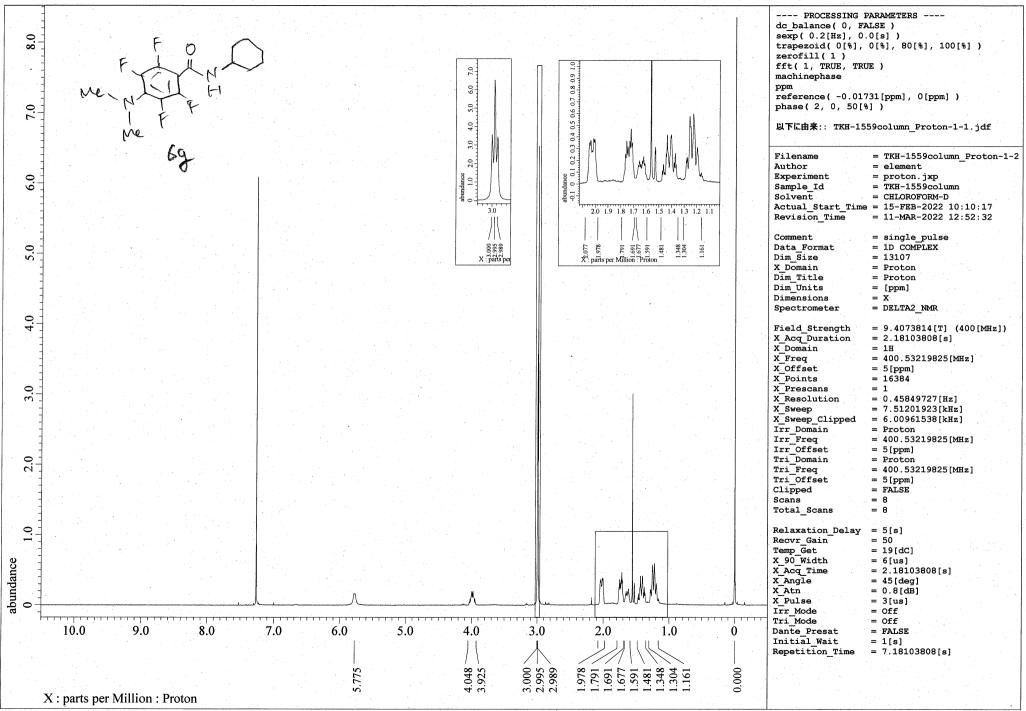


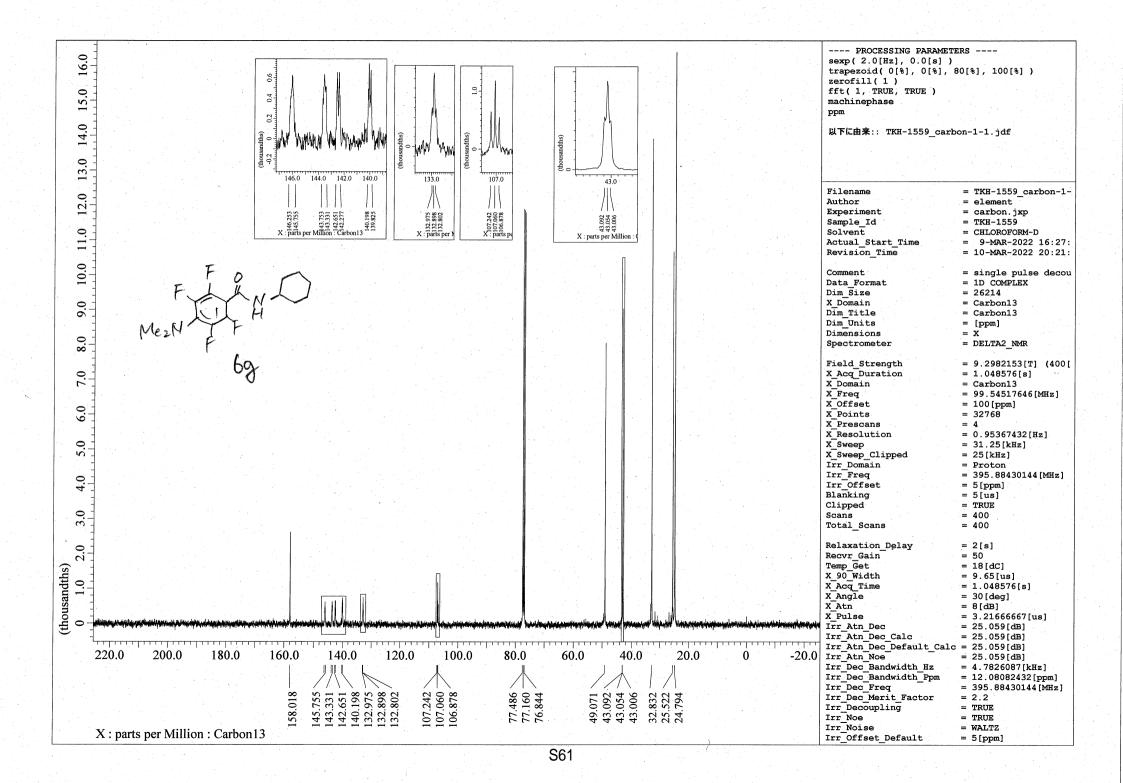
---- PROCESSING PARAMETERS ---dc balance(0, FALSE) 0 sexp(0.2[Hz], 0.0[s])
trapezoid(0[%],0[%],80[%],100[%]) 13. zerofill(1) CF3 blip(16, 64, 32) fft(1, TRUE, TRUE) 0 machinephase 2 ppm phase(-25, 0, 50[%]) 0 以下に由来:: TKH-1427_F_standard_single_pulse Ξ 0 10. Filename = TKH-1427 F standard sind Author = element = single_pulse.jxp = TKH-1427_F_standard Experiment 0 Sample Id **6** Solvent = CHLOROFORM-D Creation Time = 5-MAR-2022 18:54:28 Revision Time = 5-MAR-2022 18:27:49 0 Current Time = 5-MAR-2022 18:27:55 °, Comment = single pulse Data Format = 1D COMPLEX 0 Dim Size = 13107 ~ Dim_Title = Fluorine19 Dim Units = [ppm] Dimensions = X 0 Spectrometer = DELTA2 NMR 6 Field Strength = 9.4073814[T] (400[MHz])X Acq Duration = 86.50752 [ms]0 X Domain = 19FS X Freq = 376.87675879[MHz] K_{NHGy} F 6e X Offset = 0 [ppm]4.0 X Points = 16384 X Prescans = 1 X Resolution = 11.55968868[Hz] X Sweep = 189.39393939[kHz] 0 = 151.51515152[kHz] X Sweep Clipped 3 Irr Domain = Fluorine19 Irr Freq = 376.87675879[MHz] Irr Offset = 5[ppm] 0 Tri Domain = Fluorine19 d' Tri Freq = 376.87675879[MHz] Tri Offset = 5[ppm]Clipped = FALSE 0 Scans = 8 Total Scans = 8 Relaxation Delay = 5[s] Recvr Gain = 42 0 Temp Get = 18.3[dC]X 90 Width = 6.8[us]X Acq Time = 86.50752[ms] 0 X Angle = 45[deg] ÷ abundance X Atn = 3[dB]X Pulse = 3.4[us]0 Irr Mode = Off Tri Mode = Off Ģ Dante Presat = FALSE Initial Wait = 1[s]100.0 Repetition_Time = 5.08650752[s] 0 -100.0 -200.0 -63.720 -114.851 X : parts per Million : Fluorine19

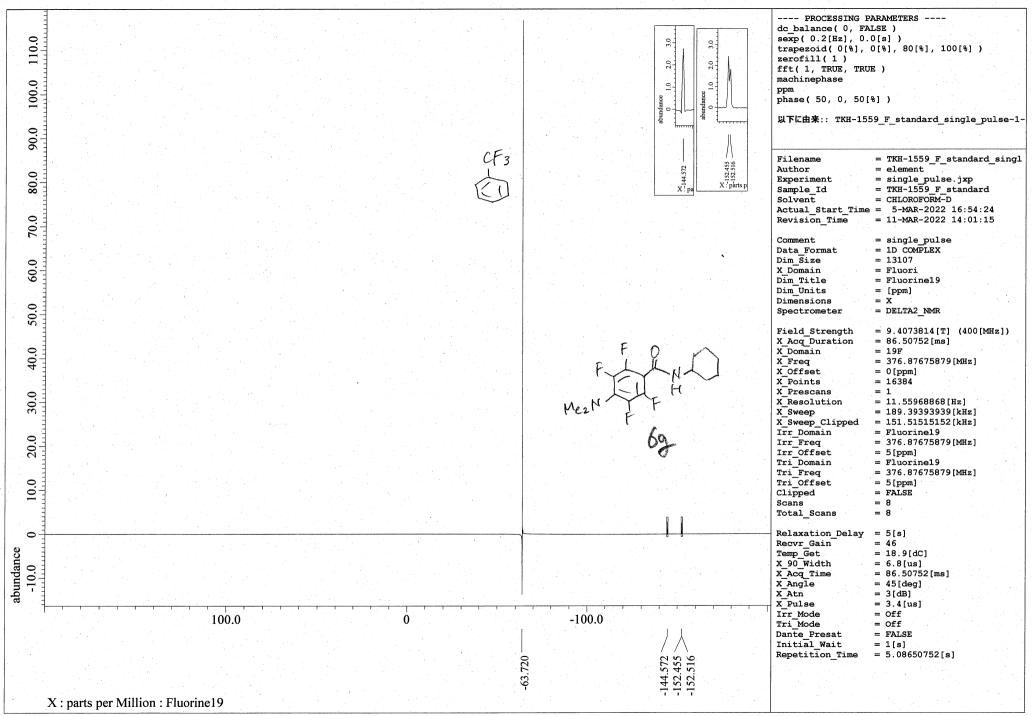


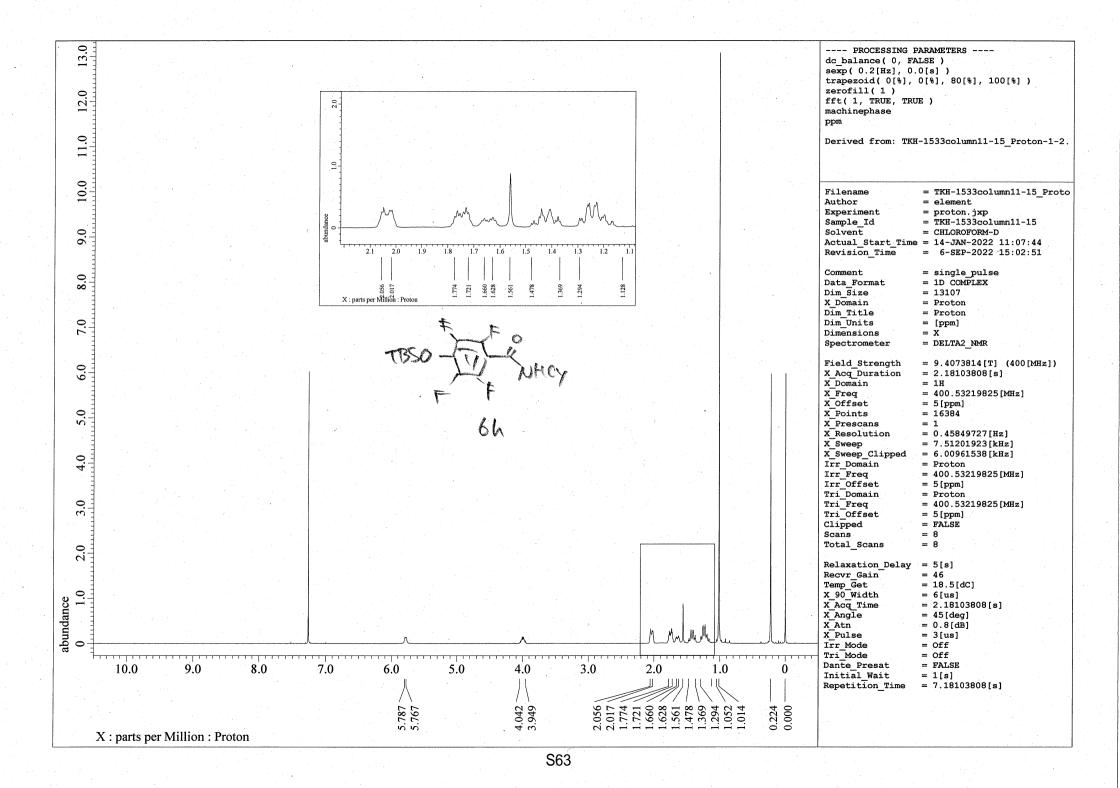


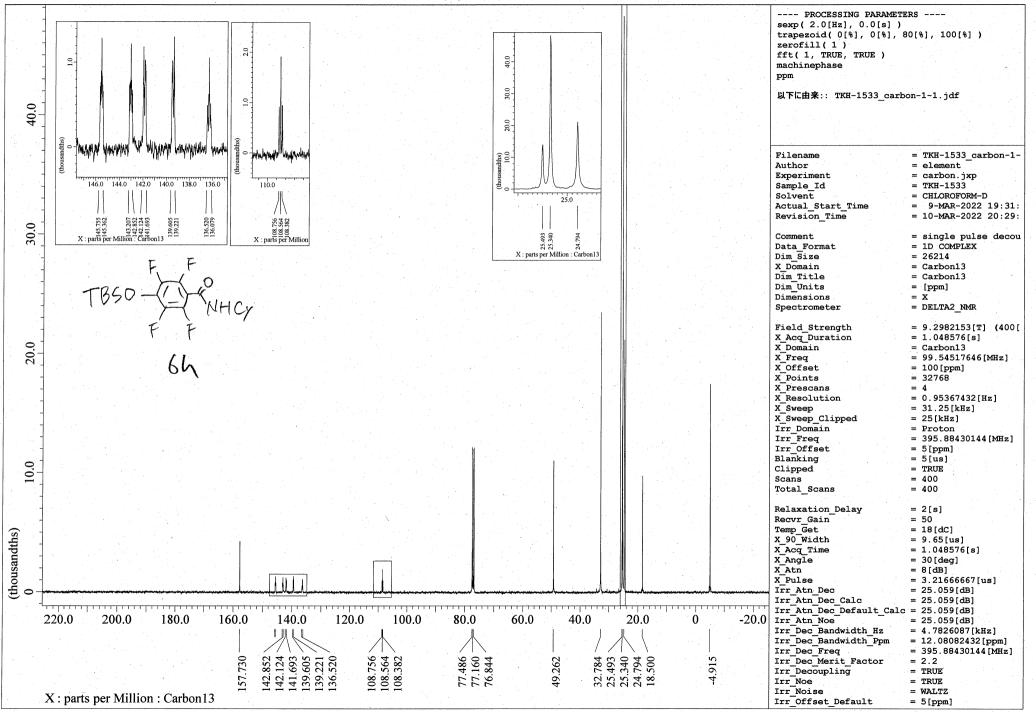


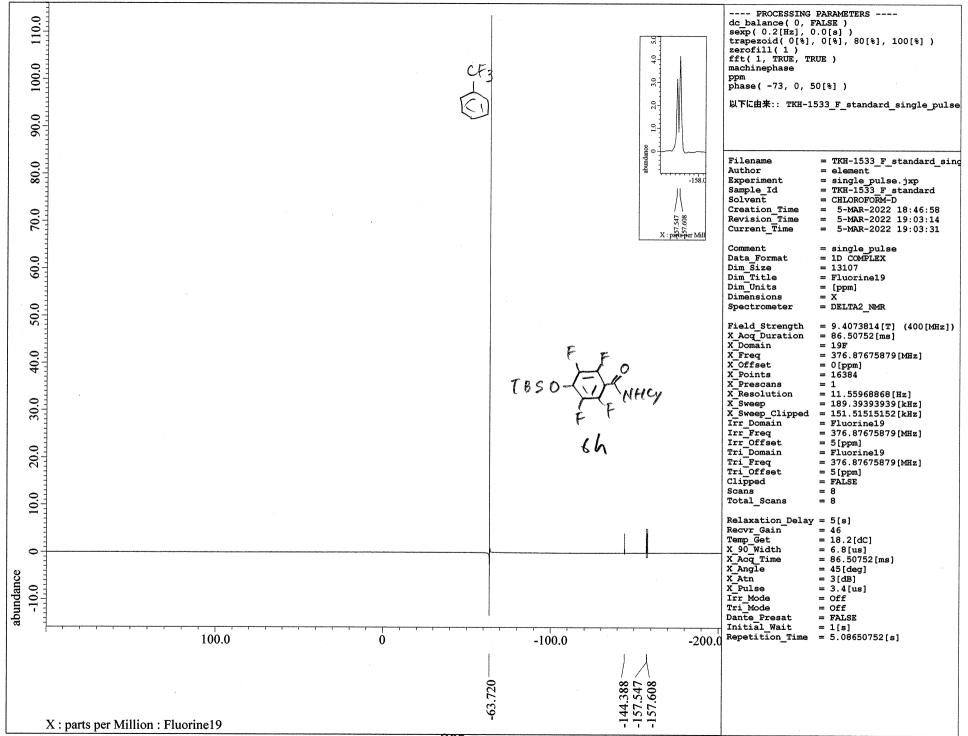


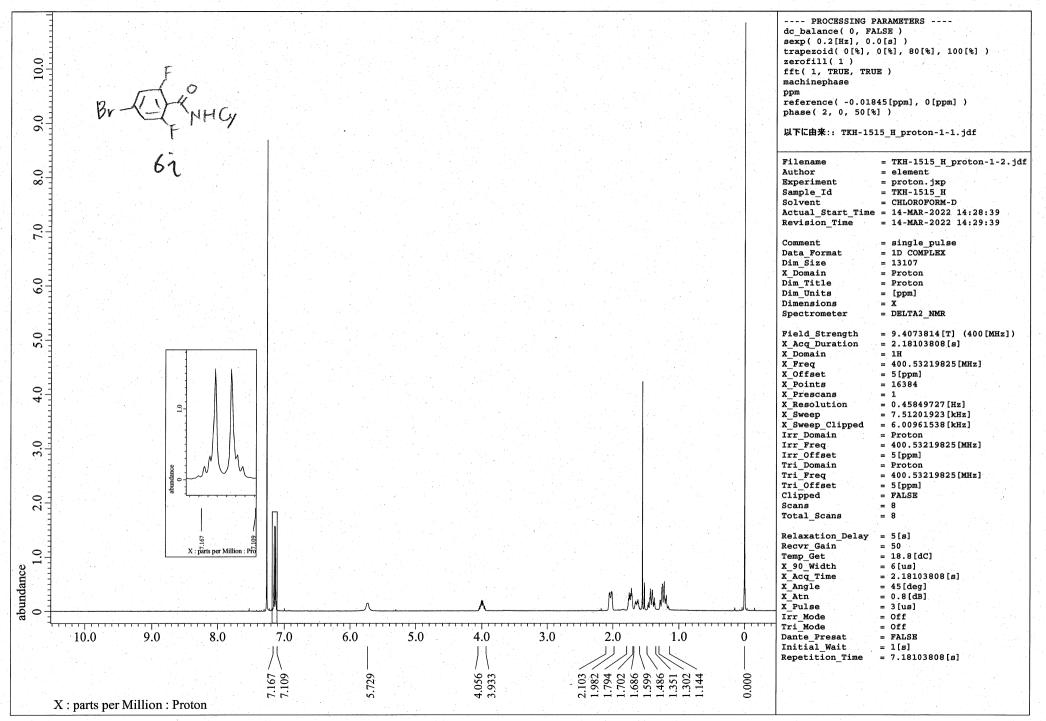


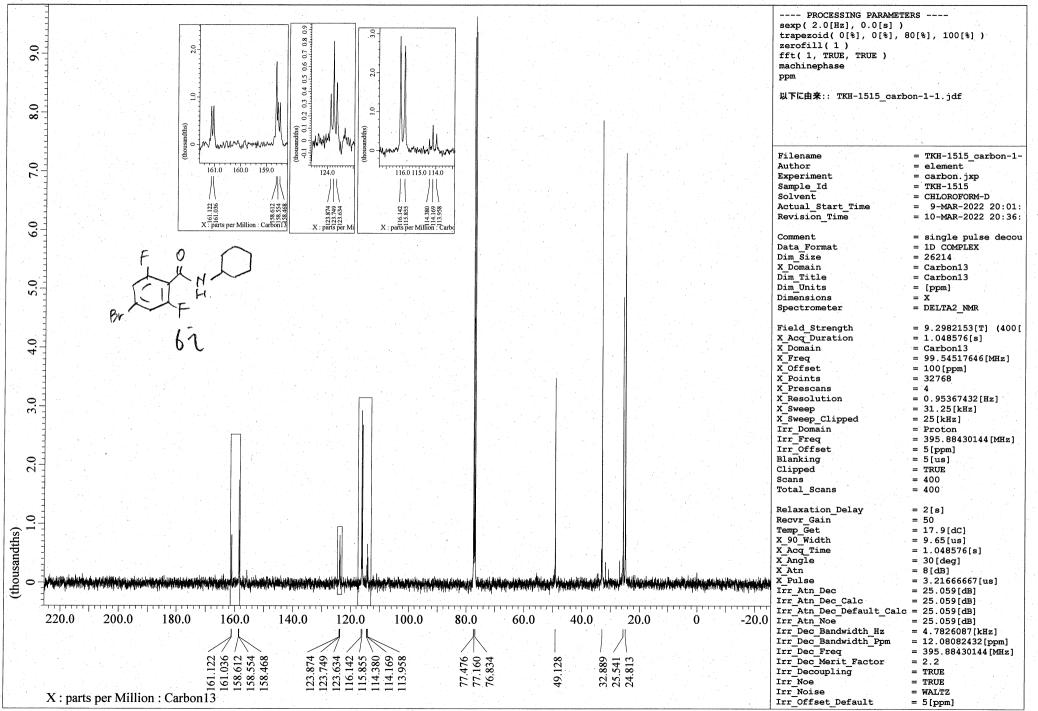


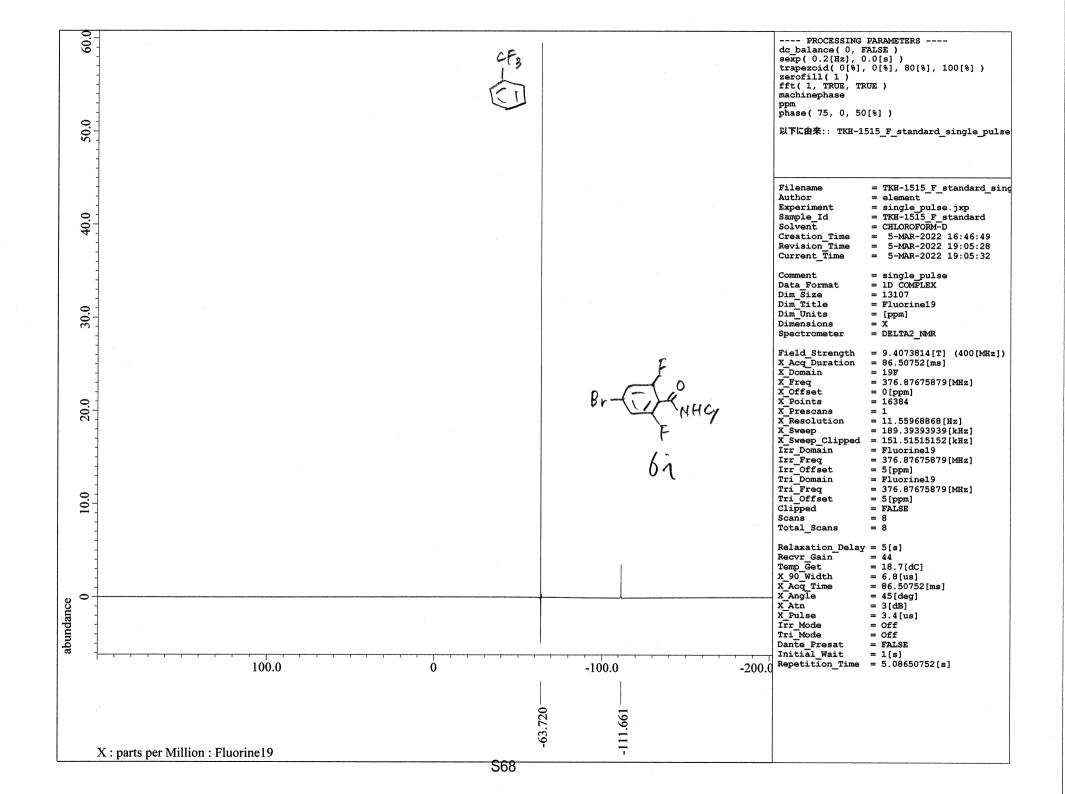


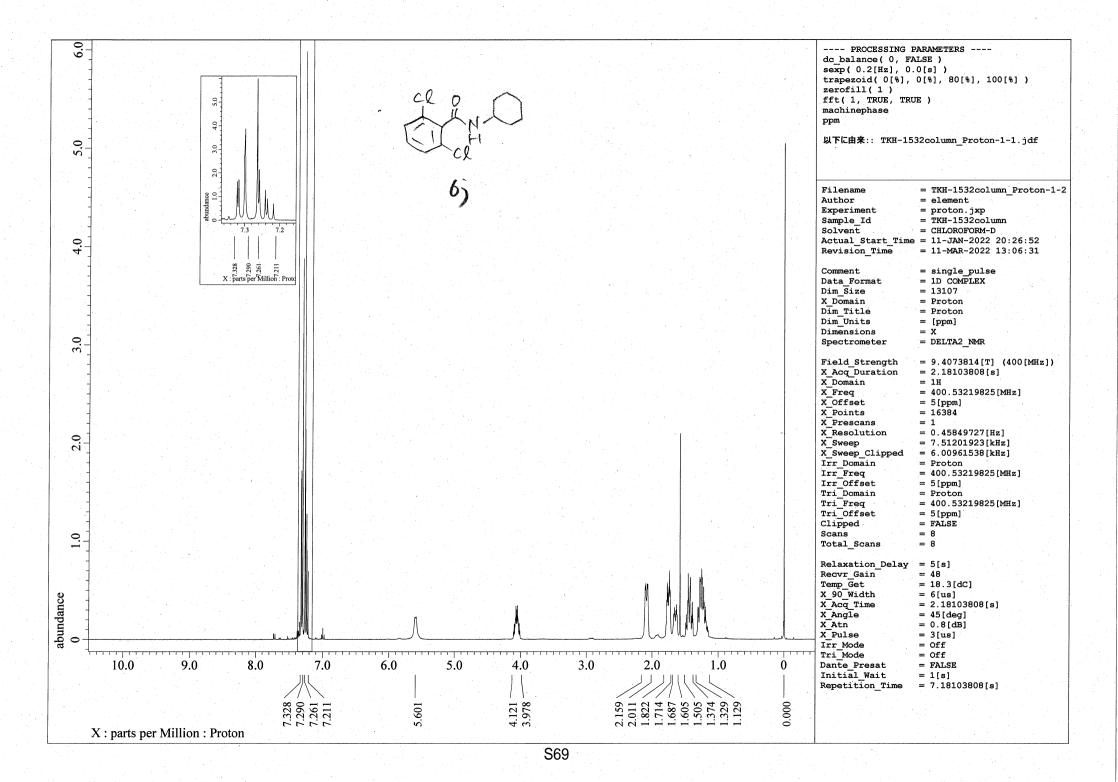


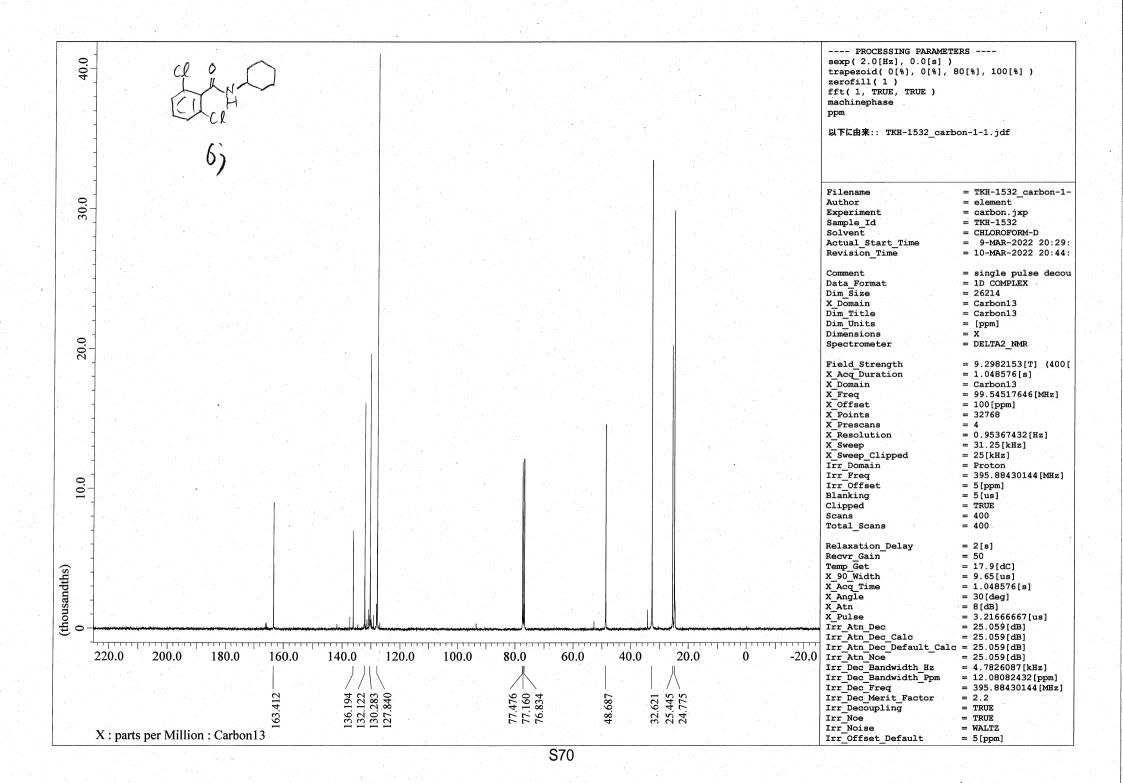


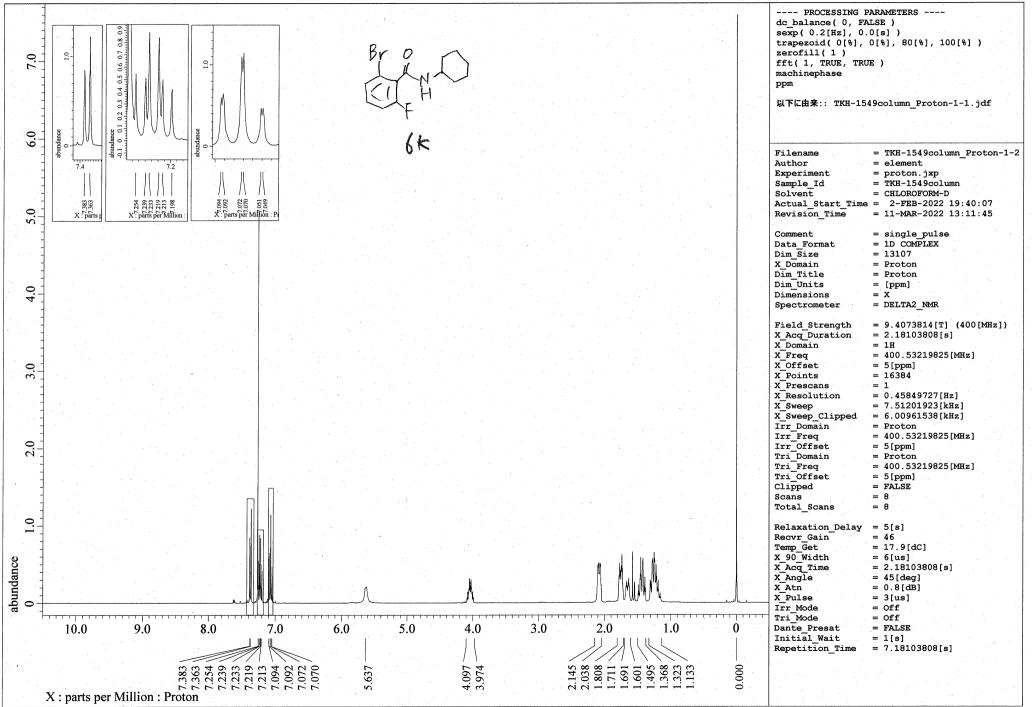


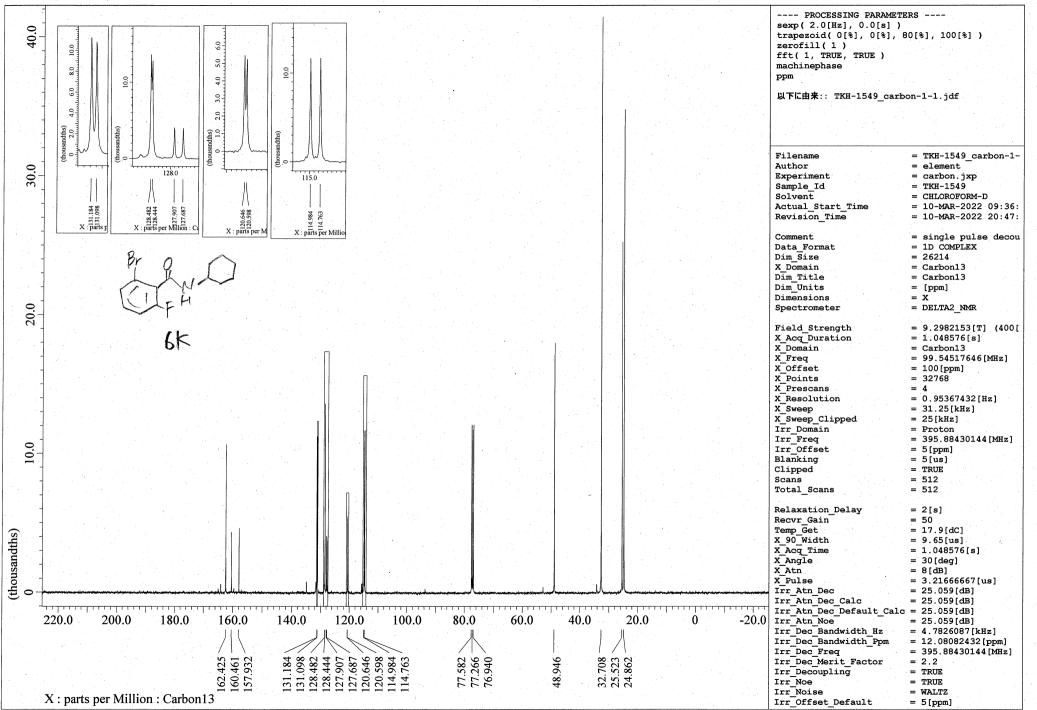












17.0 ---- PROCESSING PARAMETERS ----CF3 dc_balance(0, FALSE)
sexp(0.2[Hz], 0.0[s]) trapezoid(0[\$], 0[\$], 80[\$], 100[\$]) 16.0 zerofill(1) fft(1, TRUE, TRUE) machinephase 15.0 maa phase(61, 0, 50[%]) 以下に由来:: TKH-1549 F standard single pulse 14.0 13.0 Filename = TKH-1549 F standard sind Author = element 0 Experiment 12 = single_pulse.jxp Sample Id = TKH - 1549 F standard = CHLOROFORM-D Solvent 11.0 Creation Time = 5-MAR-2022 16:38:05 = 5-MAR-2022 19:06:15 Revision Time Current Time = 5-MAR-2022 19:06:19 0 10. Comment = single pulse = 1D COMPLEX Data Format Dim Size = 131070 Dim Title = Fluorine19 9 Dim Units = [ppm] Dimensions = X 0 Spectrometer = DELTA2 NMR ∞ Field Strength = 9.4073814[T] (400[MHz]) 0 X Acq Duration = 86.50752[ms] 1 X Domain = 19FX Freq = 376.87675879[MHz] 0 X Offset = 0[ppm] X Points = 16384 6 X Prescans = 1 X Resolution = 11.55968868[Hz] 0 X Sweep = 189.39393939[kHz] 5 X Sweep Clipped = 151.51515152[kHz] Irr Domain = Fluorine19 0 Irr Freq = 376.87675879[MHz] 4 Irr Offset = 5[ppm] Tri Domain = Fluorine19 0 Tri Freq = 376.87675879[MHz] ω Tri Offset = 5[ppm] Clipped = FALSE 0 Scans = 8 21 Total Scans = 8 Relaxation Delay = 5[s] 0 Recvr Gain = 46 Temp Get = 18.9[dC]X 90 Width = 6.8[us]X Acq Time = 86.50752[ms] 0 X Angle = 45[deg]abundance X Atn = 3[dB]-1.0 X Pulse = 3.4[us]Irr Mode = Off Tri Mode 0 = Off Dante Presat = FALSE Ģ Initial Wait = 1[s]Repetition Time = 5.08650752[s] 100.0 0 -100.0 -200.0 -63.720 -113.378 X : parts per Million : Fluorine19 S73

