Electronic Supplementary Information

Functional Characterization, Structural Basis, and Regio-Selectivity Control of a Promiscuous Flavonoid 7,4'-di-*O*glycosyltransferase from *Ziziphus jujuba* var. *spinosa*

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1. Experimental procedures

1.1 General

Compounds 1-8, 10-12, 14, 18 and 20-34 were purchased from Shanghai Yuanye Bio-Technology Co., Ltd (Shanghai, China). Compounds 9, 13, 15-17 and 19 were previously purified and characterized in our lab. Methanol and acetonitrile (Fisher Scientific, USA) were of HPLC grade. All other chemicals and reagents were purchased from Sigma-Aldrich (St. Louis, MO, USA) and Beijing Chemical Corporation (Beijing, China) unless otherwise specified. Substrate specificity and conversion rates were analyzed by Vanquish UHPLC (Thermo Fisher Scientific, Massachusetts, USA). The glycosylated products were isolated and purified by semi-preparative HPLC on an Agilent 1200 instrument (Agilent Technologies, Waldbronn, Germany). LC/MS analysis was performed on an UHPLC-Q-Exactive HRMS (Thermo Fisher Scientific, Massachusetts, USA) equipped with an electrospray ionization (ESI) source. NMR spectra were recorded on a Bruker AVANCE III-400 instrument at 400 (¹H) and 100 (¹³C) MHz in DMSO-*d*₆. Chemical shifts (δ) are given in parts per million (ppm) and coupling constants (*J*) are given in Hertz (Hz).

1.2 Plant materials

Seeds of *Ziziphus jujuba* var. *spinosa* were obtained from Shandong Province (China). The seed kernels were collected and washed, then immediately frozen in liquid nitrogen for RNA extraction.

1.3 Transcriptome data assembly and candidate gene screening

Transcriptome data of *Ziziphus jujuba* var. *spinosa* (SRR9721936, SRR9721937, SRR9721938, SRR9721939, SRR9721941 and SRR9721944) were download from NCBI (https://www.ncbi.nlm.nih.gov/), and were assembled through the SOAPdenovo-Trans-src-v1.04 program. For BLAST analysis, 24 functionally characterized plant glycosyltransferase gene sequences were used as homology-based templates to analyze the transcriptome data (Table S1). By blast with these query sequences, 24 open reading frames (ORFs) were obtained as candidate genes (Figure S90). PCR primers were designed containing the homologous sequences of pET28a (+) as shown in Table S2.

1.4 Molecular cloning, heterologous expression, and protein purification

1.4.1 Molecular cloning

The total RNA of the seed kernels of *Ziziphus jujuba* var. *spinosa* was extracted using the TransZol Reagent (Transgen Biotech, China) following the manufacturer's instructions. RNA degradation and contamination was monitored on 1% agarose gel. Purity and concentration of RNA were measured using NanoDrop (Thermo Scientific). The first-stranded complementary DNA (cDNA) was synthesized using TransScript one-step genomic DNA (gDNA) removal and cDNA synthesis SuperMix (Transgen Biotech, China). The PCR was performed using 2 μ L of first strand cDNA as a template, ZjOGT3-F and ZjOGT3-R as primers, and *FastPfu* DNA Polymerase under the following conditions: 95°C for 3 min, 35 cycles of 95°C for 30 s, 55°C for 30 s, 72°C for 30 s, followed by 72°C for 10 min. The amplified fragments were cloned into pET28a (+) vector (Invitrogen, USA) by Quick-change method. The genes were sequenced by Tsingke biological technology (Beijing, China). The recombinant vector pET28a(+)-ZjOGT3 was transformed into *E. coli* BL21(DE3) (TransGen Biotech, China) for heterologous expression. Transformed cells were selected on agar plates containing 50 μ g/mL kanamycin.

1.4.2 Heterologous expression and protein purification

Single colonies harboring the desired expression construct were inoculated overnight at 37°C with shaking in LB culture medium containing 50 µg/mL kanamycin. Isopropyl thiogalactoside (0.1 mM) was added into the medium for the expression of recombinant protein at 16°C when optical density at 600 nm was about 0.6. After incubation with shaking for 20 h, *E. coli* was harvested by centrifugation. The recombinant proteins were purified using a Nickel-affinity column (Proteinlso Ni-NTA Resin, TransGen Biotech, Beijing, China), and concentrated using Amicon Ultra-15 Ultracel-30K (Merck Millipore). The purity of recombinant ZjOGT3 determined by SDS-PAGE is shown in Figure S1. The concentration of recombinant ZjOGT3 was measured using NanoDrop under parameter of E 1%=12 (Thermo Scientific).

1.5 Effects of reaction time, pH, temperature, and divalent metal ions

To investigate the enzymatic properties of ZjOGT3, the reaction time of glycosylation was firstly determined for 8 h (Figure S2). To optimize the reaction pH, the enzymatic reactions were carried out

in various reaction buffers ranged in pH value from 4.0-6.0 (Citric acid-sodium citrate buffer), 6.0-8.0 (Na₂HPO₄-NaH₂PO₄ buffer), 7.0-9.0 (Tirs-HCl buffer), and 9.0-10.0 (Na₂CO₃-NaHCO₃ buffer). To study the optimal reaction temperature, the enzymatic reactions were incubated at different temperatures (4-60°C). To test the dependence of divalent metal ions for ZjOGT3 activity, different divalent cations including Ca²⁺, Mn²⁺, Mg²⁺, Fe²⁺, Co²⁺, Ni²⁺, Ba²⁺, Zn²⁺, Cu²⁺ and EDTA in the final concentration of 5 mM were used individually. All enzymatic reactions (100 µL reaction mixtures including 0.5 mM UDP-Glc, 0.2 mM 1, and 25 µg of purified ZjOGT3) were conducted in three parallel experiments (*n*=3). All the reactions were terminated with pre-cooling methanol (MeOH) and centrifuged at 15,000 rpm for 20 min for further UHPLC analysis as described in general methods. The samples were separated on an ACQUITY UPLC® HSS T3 column (100 mm × 2.1 mm, 1.8 µm) at a flow rate of 0.3 mL min⁻¹. The mobile phase was a gradient elution of solvents A (MeOH) and B (0.1% formic acid aqueous solution), and the gradient programs were listed in Table S3. The conversion rates in percent were calculated from peak areas of glycosylated products and substrates as analyzed by UHPLC (Figure S3).

1.6 Kinetic studies

All assays were performed in a final volume of 50 μ L containing 50 mM NaH₂PO₄-Na₂HPO₄ buffer (pH 8.0) at 45°C for 10 min. For the first step (4'-*O*-glycosylation), 0.3365 μ g protein, 2 mM of saturated UDP-Glc, and different concentrations of 1 (1, 5, 10, 20, 50 μ M). For the second step (7-*O*-glycosylation), 3.0925 μ g protein, 2 mM of saturated UDP-Glc, and different concentrations of **1a** (10, 20, 50, 80, 100, 200, 500 μ M). The reactions were quenched with 200 μ L ice cold MeOH. Samples were centrifuged at 15,000 rpm for 20 min and analyzed by UHPLC as described above. All experiments were performed in triplicate. The Michaelis-Menten constants (*K*_m) values were calculated by using the Lineweaver-Burk plot method (Figure S4).

1.7 Substrate promiscuity and sugar donor selectivity of ZjOGT3

To investigate the substrate promiscuity and sugar donor selectivity of ZjOGT3, different acceptors including compounds **1-34** and different sugar donors including UDP-glucose (UDP-Glc), UDP-glactose (UDP-Gal), UDP-xylose (UDP-Xyl), UDP-arabinose (UDP-Ara), UDP-glucuronic acid (UDP-GlcA), UDP-*N*-acetylglucosamine (UDP-GlcNAc), and UDP-rhamnose (UDP-Rha) were used. The reaction mixtures were individually performed in a final volume of 100 µL containing 50 mM

NaH₂PO₄-Na₂HPO₄ buffer (pH 8.0), 25 μ g of purified ZjOGT3, 0.1 mM aglycone, and 0.5 mM sugar donor. All reactions were incubated at 45°C for 8 h and quenched with 200 μ L ice cold MeOH. The mixtures were then centrifuged at 15,000 rpm for 20 min and analyzed by UHPLC/MS. The samples were analyzed by an ACQUITY UPLC® HSS T3 column (100 mm × 2.1 mm, 1.8 μ m) at a flow rate of 0.3 mL min⁻¹. The mobile phase was a gradient elution of solvents A (MeOH) and B (0.1% formic acid aqueous solution), and the gradient programs were listed in Table S3. The conversion rates in percent were calculated from peak areas of glycosylated products and substrates as analyzed by UHPLC (Figure S5-S38). The flow rate was 0.3 mL/min. The column temperature was 55°C. The detection wavelength was 254 nm, 300 nm, and 340 nm. MS analysis was performed on a UHPLC-Q-Exactive HRMS instrument (Thermo Fisher Scientific, Massachusetts, USA). The MS parameters were as follows: sheath gas pressure 45 arb, aux gas pressure 10 arb, discharge voltage 4.5 kV, capillary temperature 350°C. MS¹ resolution was set as 70,000 FWHM, AGC target 1*E⁶, maximum injection time 50 ms, and scan range *m/z* 100-1000. MS² resolution was set as 17,500 FWHM, AGC target 1*E⁵, maximum injection time 100 ms, NCE 35.

1.8 Preparative-scale reactions and structural identification of glycosylated products

The substrates (50 mM) were dissolved in dimethyl sulfoxide (DMSO) as sugar acceptor, with 50 mM UDP-Glc as sugar donor. Preparative-scale reactions contain 2 ml buffer (50 mM NaH₂PO₄-Na₂HPO₄, pH 8.0), 40 µL sugar acceptor (50 mM), 90 µL sugar donor (50 mM), and ZjOGT3. The reactions were performed at the optimum condition for 12 h and terminated by adding MeOH. The reaction mixtures were then centrifuged at 15,000 rpm for 30 min, and the supernatants were concentrated and dissolved in 1.5 mL methanol. The glycosylated products were subsequently purified by reversed-phase semi-preparative HPLC and characterized by HR-ESI-MS and NMR analyses (Figures S39-S79).

1.9 Protein expression, purification, and crystallization

The full-length cDNA of ZjOGT3 was cloned into pET-28a (+) vector. The S-tag of pET-28a was removed. A TrxA-tag and 6 His-tag followed by thrombin site were added before N-terminus of the target protein to facilitate purification. The TrxA-His-thrombin-ZjOGT3 protein was expressed in *E. coli* (DE3) strain and purified by Ni-NTA affinity chromatography (GE healthcare). After purification, the recombinant protein was digested by thrombin to cut the TrxA-His tag. The sample was put on the Ni-NTA affinity beads for the second time to purify the protein. The flow-through was concentrated

and then applied to molecular sieve on a SuperdexTM 200 Increase 10/300 GL column (GE healthcare, Sweden) for further purification. Then the protein buffer was changed to elution buffer (20 mM Tris-HCl, pH 7.5, 50 mM NaCl). Fractions containing ZjOGT3 were collected and concentrated to 20 mg/mL. The purified ZjOGT3 protein (20 mg/mL) was incubated with UDP-Glc (10 mM) for 2 hours. The crystals were prepared by hanging drop vapor diffusion. Crystals of ZjOGT3 were obtained after 2 days at 16°C in hanging drops containing 1.0 μ L of protein solution and 1.5 μ L of reservoir solution 0.2 M sodium acetate trihydrate, 26% *w/v* PEG 4000, 10 mM Tris-HCl, pH 8.0 (Figures S81).

1.10 Crystal structure determination and refinement

Diffraction data of the ZjOGT3 crystals were collected at beamlines BL18U1, Shanghai Synchrotron Radiation Facility (SSRF) and processed with HKL-2000. The structure was solved by molecular replacement with Phaser using the previously reported UGT74AC1 structure (PDB ID: 6L8X) as the search model. Crystallographic refinement was performed repeatedly using Phenix and COOT. The refined structure was validated by Phenix and the PDB validation server (https://validate-rcsb-1.wwpdb.org/). The final refined structure was deposited in Protein Data Bank with the access ID 8INH. The diffraction data and structure refinement statistics are summarized in Table S4.

1.11 Site-directed mutagenesis of ZjOGT3 and enzyme activity assay

The mutants of ZjOGT3 were constructed using a Fast Mutagenesis System kit (TransGen Biotech, China) according to the manufacturer's instructions. The corresponding degenerate primers designed to construct the site-directed mutants are listed in Table S2. After verification of the mutant sequences, the recombinant plasmids were transformed into *E. coli* BL21(DE3) for heterologous expression. The protein expression, purification, and enzyme activity assays of mutants were performed under the same conditions as described above (Figure S85).

1.12 DFT calculations

DFT calculations were performed with the Gaussian 16 package¹. The geometry optimizations of minima and transition states involved were carried out at the B3LYP-D3/6-31+G(d,p) level. The vibrational frequency calculations were calculated at the same level to ensure that all of the stationary points were transition states (one imaginary frequency) or minima (no imaginary frequency) and to evaluate zero-point vibrational energies (ZPVE) and thermal corrections at 298K. Single-point energy calculations were performed at the B3LYP-D3 level with the 6-311++G(2d,p) basis set. Solvation by

water was taken into account by using the CPCM model²⁻⁴ for all above calculations. Gibbs free energy is the sum of the electronic energy and ZPVE and thermal corrections.

1.13 Initial structural preparation

The initial structure of wild type ZjOGT3 in complex with UDP-Glc was based on the crystal structure (PDB ID: 7EFK). The geometry of UDP-Glc was fully optimized at the B3LYP-D3/6-31+G(d) level of Gaussian 16 using the CPCM model in water, and the partial charges were fitted with HF/6-31G(d) calculations and the restrained electrostatic potential (RESP)⁵⁻⁶ protocol implemented by the Antechamber module. The protonation states of charged residues were determined at constant pH 8.0 based on pKa calculations via the H++ program⁷ and the consideration of the local hydrogen bonding network. His19, 44, 155, 158, 191, 323, 351, and 360 were assigned as HIE, and the rest His were HID. All Asp and Glu residues were deprotonated, while Lys and Arg were protonated. The system was neutralized by adding Na⁺ ions and solvated into a rectangular TIP3P⁸ water box with a 10-Å buffer distance on each side. A representative MD pre-equilibrated binary complex structure was used as the target receptor for molecular docking.

1.14 Molecular docking

20000 snapshots uniformly distributed at equal intervals from the last 40 ns MD simulation (with time intervals of 2ps) were picked up and divided into ten groups using hierarchical agglomerative (bottom-up) approach⁹. The anionic intermediates of substrates **2**, **10** and **12** were fully optimized at the B3LYP-D3/6-31+G(d,p) level of Gaussian 16 using the CPCM model in water, and then docked into the active site of one representative group snapshot to gain the ternary complex. Molecular docking was performed using the Lamarckian genetic algorithm local search method in the AutoDock 4.2 and AutoDockTools-1.5.6¹⁰. The docking approach was employed on rigid-receptor conformation, while all the rotatable torsional bonds of all substrates were set free. A grid box was centered on the carbon atom being attacked of UDP-Glc and its size was set to 40 Å × 40 Å × 40 Å points with a 0.375 Å spacing. A total of 500 independent docking runs were undertaken with a maximum energy evaluation of 2.5×10^7 . The obtained 500 docked conformations were clustered with 2.0 Å RMSD and ranked depending on an energy-based scoring function. The possible catalytically active binding modes were selected as initial configurations to perform MD simulations of the ternary complexes according to scoring function and reasonable conformation.

1.15 Molecular dynamics simulations

All molecular dynamics (MD) simulations were performed by Amber 16 package¹¹. The preequilibrated ZjOGT3 in complex with UDP-Glc binary complex structures and possible catalytically reactive binding modes of substrates were used as the starting conformations for MD simulations on the ternary complexes. Acceptor substrates were fully optimized at the B3LYP-D3/6-31+G(d,p) level of Gaussian 16 using the CPCM model in water, and the partial charges were fitted with HF/6-31G(d) calculations and RESP protocol implemented by the Antechamber module. The force field parameters for substrates were adapted from the standard general amber force field 2.0 (gaff2)¹² parameters, while the standard Amber14SB force field was applied to describe the protein.

Each system was neutralized by adding Na⁺ ions and solvated into a rectangular TIP3P water box with a 10-Å buffer distance on each side. After equilibrated with a series of minimizations interspersed by short MD simulations during which restraints on the protein backbone heavy atoms were gradually released [with force constant of 10, 2, 0.1 and 0 kcal/(mol·Å²)], each system was heated from 0 to 318 K for 50 ps. Finally, 200 ns MD simulations with periodic boundary condition at constant temperature and pressure were carried out. The pressure was maintained at 1 atm and coupled with isotropic position scaling. The temperature was controlled at 318 K with Berendsen thermostat method. Longrange electrostatic interactions were treated with particle mesh Ewald (PME)¹³ method and 12-Å cutoff was applied to both PME and van der Waals (vdW) interactions. Time step of 2 fs was employed along with SHAKE algorithm for hydrogen atoms, and periodic boundary condition was used. Each system was checked for stability (structure, energy, and temperature fluctuations) and convergence (root mean square deviations-RMSD of structures).

1.16 MM/GBSA calculations

The binding free energies of ZjOGT3 in complex with UDP-Glc and substrates were calculated using the Molecular Mechanics Generalized Born Surface Area (MM/GBSA) method in the MMPBSA.py program¹⁴ in AMBER 16 package. Before calculation, the water and ions were stripped from the last 1 ns of MD trajectory of the complexes. ΔG_{bind} is based on the following equation: $\Delta G_{bind} = \Delta E_{vdw} + \Delta E_{ele} + \Delta G_{pol} + \Delta G_{nonpol} - T\Delta S$, where ΔE_{vdw} and ΔE_{ele} are van der Waals and electrostatic components in the gas phase, and ΔG_{pol} and ΔG_{nonpol} indicate the polar and nonpolar solvent interaction energies, respectively. ΔE_{vdw} and ΔE_{ele} were calculated using the Amber14SB force field (ff14sb). ΔG_{pol} was calculated by solving the Generalized Born (GB) equations. $\Delta G_{nonpol} = 0.0050 \times SASA + 0.000$, where SASA referred to the solvent accessible surface buried upon binding. Since the normal mode calculation is extremely time-consuming for large systems, we estimated the ΔG_{bind} ignoring entropy. Here, we used GB model with igb=1 and the mbondi2 radii set.

2. MS, ¹H and ¹³C NMR spectral data for glycosylated products



Luteolin 4'-*O*-*β*-**D**-glucoside (3a): ESI-MS *m*/*z* 447.0922 [M–H]⁻. ¹H NMR (DMSO-*d*₆, 400 MHz): δ 12.91 (1H, s, 5-OH), 7.51 (1H, dd, *J*=8.5 2.1 Hz, H-6'), 7.49 (1H, d, *J*=2.1 Hz, H-2'), 7.24 (1H, d, *J*=8.5 Hz, H-5'), 6.83 (1H, s, H-3), 6.50 (1H, d, *J*=2.0 Hz, H-8), 6.21 (1H, d, *J*=2.0 Hz, H-6), 4.89 (1H, d, *J*=7.2 Hz, H-1").¹⁵

¹³C NMR (DMSO-*d*₆, 100 MHz): δ 181.8 (C-4), 164.3 (C-7), 163.2 (C-2), 161.5 (C-5), 157.4 (C-9), 148.6 (C-4'), 147.0 (C-3'), 124.7 (C-1'), 118.6 (C-6'), 116.1 (C-5'), 113.6 (C-2'), 104.0 (C-10), 103.8 (C-3), 101.2 (C-1''), 99.0 (C-6), 94.1 (C-8), 77.3 (C-5''), 75.9 (C-3''), 73.3 (C-4''), 69.8 (C-2''), 60.7 (C-6'').¹⁵



Luteolin 7,4'-di-*O***-glucoside** (**3b**): ESI-MS *m*/*z* 609.1448 [M–H][–]. ¹H NMR (DMSO-*d*₆, 400 MHz): δ 12.91 (1H, s, 5-OH), 7.55 (1H, dd, *J*=8.4 2.1 Hz, H-6'), 7.54 (1H, d, *J*=2.1 Hz, H-2'), 7.26 (1H, d, *J*=8.4 Hz, H-5'), 6.92 (1H, s, H-3), 6.86 (1H, d, *J*=2.0 Hz, H-8), 6.45 (1H, d, *J*=2.0 Hz, H-6), 5.09 (1H, d, *J*=7.2 Hz, H-1″'), 4.89 (1H, d, *J*=7.2 Hz, H-1″).¹⁵

¹³C NMR (DMSO-*d*₆, 100 MHz): δ 182.0 (C-4), 163.7(C-2), 163.0 (C-7), 161.1 (C-5), 157.0 (C-9), 148.7 (C-4'), 146.9 (C-3'), 124.5 (C-1'), 118.7 (C-6'), 116.0 (C-2'), 113.7 (C-5'), 105.4 (C-3), 104.2 (C-10), 101.3 (C-1''), 99.8 (C-1'''), 99.6 (C-6), 94.8 (C-8), 77.3 (C-3''), 77.1 (C-5''), 76.4 (C-5'''), 75.8 (C-3'''), 73.2 (C-2'''), 73.1 (C-2''), 69.8 (C-4''), 69.5 (C-4'''), 60.7 (C-6''), 60.6 (C-6''').¹⁵



Quercetin 4'-*O*-*β*-**D**-glucoside (7a): ESI-MS *m*/*z* 463.0872 [M–H]⁻. ¹H NMR (DMSO-*d*₆, 400 MHz): δ 12.41 (1H, s, 5-OH), 7.70 (1H, d, *J*=2.1 Hz, H-2'), 7.62 (1H, dd, *J*=8.8 2.1 Hz, H-6'), 7.24 (1H, d, *J*=8.8 Hz, H-5'), 6.45 (1H, d, *J*=2.0 Hz, H-8), 6.20 (1H, d, *J*=2.0 Hz, H-6), 4.84 (1H, d, *J*=7.2 Hz, H-1'').¹⁶

¹³C NMR (DMSO-*d*₆, 100 MHz): δ 176.0 (C-4), 164.1 (C-7), 160.7 (C-5), 156.2 (C-9), 146.8 (C-3'), 146.4 (C-2), 145.9 (C-4'), 136.4 (C-3), 125.1 (C-1'), 118.7 (C-6'), 115.8 (C-2'), 115.1 (C-5'), 103.1 (C-10), 101.4 (C-1''), 98.3 (C-6), 93.5 (C-8), 77.3 (C-3''), 76.0 (C-5''), 73.3 (C-2''), 69.8 (C-4''), 60.7 (C-6'').¹⁶



Quercetin 7,4'-di-*O*-glucoside (7b): ESI-MS *m*/*z* 625.1355 [M–H]⁻. ¹H NMR (DMSO-*d*₆, 400 MHz): δ 12.42 (1H, s, 5-OH), 7.75 (1H, d, *J*=2.1 Hz, H-2'), 7.65 (1H, dd, *J*=8.8, 2.1 Hz, H-6'), 7.26 (1H, d, *J*=8.8 Hz, H-5'), 6.82 (1H, d, *J*=2.0 Hz, H-8), 6.42 (1H, d, *J*=2.0 Hz, H-6), 5.09 (1H, d, *J*=7.2 Hz, H-1''), 4.85 (1H, d, *J*=6.8 Hz, H-1'').¹⁶

¹³C NMR (DMSO-*d*₆, 100 MHz): δ 176.3 (C-4), 162.8 (C-7), 160.3 (C-5), 155.8 (C-9), 147.0 (C-3'), 146.6 (C-2), 146.4 (C-4'), 136.8 (C-3), 125.0 (C-1'), 119.6 (C-2'), 115.8 (C-5'), 105.7 (C-10), 101.4 (C-1''), 99.8 (C-1'''), 98.8 (C-6), 94.3 (C-8), 77.3 (C-3''), 77.1 (C-5'''), 76.4 (C-3'''), 75.9 (C-5''), 73.3 (C-2''), 73.1 (C-2'''), 69.8 (C-4''), 69.5 (C-4'''), 60.7 (C-6''), 60.6 (C-6''').¹⁶



Naringenin 7-*O***-β-D-glucoside** (**10a**): ESI-MS *m/z* 433.1100 [M–H]⁻. ¹H NMR (DMSO-*d*₆, 400 MHz): δ 12.06 (1H, s, 5-OH), 7.33 (2H, d, *J*=8.4 Hz, H-2', H-6'), 6.80 (2H, d, *J*=8.4 Hz, H-3', 5'), 6.15 (1H, d, *J*=2.2 Hz, H-8), 6.14 (1H, d, *J*=2.2 Hz, H-6), 5.50 (1H, dt, *J*=12.8 2.8 Hz, H-2), 4.98 (1H, d, *J*=7.6 Hz, H-1'').¹⁷

¹³C NMR (DMSO-*d*₆, 100 MHz): δ 197.2 (C-4), 165.3 (C-7), 162.9 (C-5), 162.8 (C-9), 157.8 (C-4'), 128.6 (C-2', C-6'), 128.4 (C-1'), 115.2 (C-3', C-5'), 103.2 (C-10), 99.6 (C-1''), 96.5 (C-6), 95.4 (C-8), 78.7 (C-2), 77.1 (C-5''), 76.3 (C-3''), 73.0 (C-2''), 69.5 (C-4''), 60.6 (C-6''), 42.1 (C-3).¹⁸



Eriodictyol 7-*O***-β-D-glucoside** (**11a**): ESI-MS *m/z* 449.1050 [M–H]⁻. ¹H NMR (DMSO-*d*₆, 400 MHz): δ 12.06 (1H, s, 5-OH), 6.88 (1H, s, H-2'), 6.75 (2H, s, H-5', 6'), 6.14 (2H, m, H-6, 8), 5.44 (1H, dd, *J*=12.0 3.6 Hz, H-2), 4.97(1H, m, H-1"), 3.14 (1H, dd, *J*=18.4, 9.2 Hz, H_{ax}-3), 2.73 (1H, dd, *J*=18.4, 4.4 Hz, H_{eq}-3). ¹⁹

¹³C NMR (DMSO-*d*₆, 100 MHz): δ 197.2 (C-4), 165.3 (C-7), 162.9 (C-5), 162.7 (C-9), 145.8 (C-3'),
145.2 (C-4'), 129.2 (C-1'), 118.1 (C-6'), 115.3(C-2'), 114.4 (C-5'), 103.2 (C-10), 99.6 (C-1''), 96.4 (C-6), 95.4 (C-8), 78.7 (C-2), 77.0 (C-5''), 76.3 (C-3''), 73.0 (C-2''), 69.5 (C-4''), 60.6 (C-6''), 42.2 (C-3).



Astragalin 7-*O*-β-D-glucopyranoside (12a): ESI-MS *m*/*z* 609.1443 [M–H]⁻. ¹H NMR (DMSO-*d*₆,

400 MHz): δ 12.6 (1H, s, 5-OH), 8.06 (2H, d, *J*=8.8 Hz, H-2', 6'), 6.90 (2H, d, *J*=8.8 Hz, H-3', 5'), 6.79 (1H, d, *J*=2.0 Hz, H-8), 6.44 (1H, d, *J*=2.0 Hz, H-6), 5.48 (1H, d, *J*=7.2 Hz, H-1"), 5.08 (1H, d, *J*=7.2 Hz, H-1"").²⁰

¹³C NMR (DMSO-*d*₆, 100 MHz): δ 177.6 (C-4), 162.8 (C-7), 160.8 (C-5), 160.1 (C-4'), 156.8 (C-9), 131.0 (C-6'), 120.8 (C-1'), 115.1 (C-3', C-5'), 105.6 (C-10), 100.7 (C-1''), 99.7 (C-1'''), 99.3 (C-6), 94.5 (C-8), 77.5 (C-5''), 77.2 (C-5'''), 76.4 (C-3'', C-3'''), 74.2 (C-2''), 73.1 (C-2'''), 69.9 (C-4''), 69.6 (C-4'''), 60.8 (C-6''), 60.6 (C-6'''). ²⁰



Spinosin 4'-*O*-*β***-D**-glucoside (20a): ESI-MS *m/z* 769.2177 [M–H]⁻. ¹H NMR (DMSO-*d*₆, 400 MHz): δ 8.10 (2H, d, *J*=9.0 Hz, H-2', 6'), 7.22 (2H, d, *J*=9.0 Hz, H-3', 5'), 5.04 (1H, d, *J*=7.2 Hz, H-1'''), 4.69 (1H, d, *J*=6.9 Hz, H-1''), 4.17 (1H, d, *J*=8.0 Hz, H-1''').

¹³C NMR (DMSO-*d*₆, 100 MHz): δ 182.1 (C-4), 165.1 (C-7), 163.2 (C-2), 160.8 (C-4'), 160.5 (C-5), 157.3 (C-9), 128.2 (C-2', 6'), 123.9 (C-1'), 116.6 (C-3', 5'), 108.7 (C-6), 105.4 (C-1'''), 105.2 (C-10), 104.3 (C-3), 100.0 (C-1'''), 90.4 (C-8), 81.9 (C-5''), 81.9 (C-2''), 78.7 (C-3''), 77.2 (C-5'''), 77.2 (C-5'''), 76.7 (C-3'''), 76.6 (C-3'''), 74.6 (C-2'''), 73.2 (C-2'''), 71.1 (C-1''), 70.4 (C-4''), 69.7 (C-4'''), 69.5 (C-4''''), 61.5 (C-6''), 60.6 (C-6'''), 56.6 (7-OCH₃).



Swertisin 4'-*O*-β-D-glucoside (21a): ESI-MS *m*/*z* 667.1614 [M–H]⁻. ¹H NMR (DMSO-*d*₆, 400 S12

MHz):δ 13.40 (1H, s, 5-OH), 8.09 (2H, d, *J*=9.0 Hz, H-2', 6'), 6.97 (2H, d, *J*=9.0 Hz, H-3', H-5'), 5.04 (1H, d, *J*=7.2 Hz, H-1'''), 4.59(1H, m, H-1'').²¹

¹³C NMR (DMSO-*d*₆, 100 MHz): δ 182.4 (C-4), 165.1 (C-7), 163.8 (C-2), 160.4 (C-5), 159.5 (C-4'), 156.9 (C-9), 128.2 (C-2', 6'), 123.7 (C-1'), 116.6 (C-3', 5'), 109.8 (C-6), 104.7 (C-3), 104.0 (C-10), 99.8 (C-1''), 90.3 (C-8), 81.9 (C-5''), 79.1 (C-3''), 77.2 (C-5'''), 76.6 (C-3'''), 73.2 (C-2'''), 72.8 (C-1''), 70.8 (C-4''), 69.7 (C-2''), 69.6 (C-4'''), 61.7 (C-6''), 60.7 (C-6'''), 56.5 (7-OCH₃). ²¹



Isovitexin 4'-*O*-*β***-D**-glucoside (22a): ESI-MS *m*/*z* 593.1497 [M–H]⁻. ¹H NMR (DMSO-*d*₆, 400 MHz): δ 13.49 (1H, s, 5-OH), 8.04 (2H, d, *J*=8.9 Hz, H-2', 6'), 7.19 (2H, d, *J*=8.9 Hz, H-3', 5'), 6.89 (1H, s, H-3), 6.55 (1H, s, H-8), 5.03 (1H, d, *J*=7.3 Hz, H-1''), 4.60 (1H, d, *J*=9.8 Hz, H-1''), 4.04 (1H, dd, *J*=9.8, 7.7 Hz, H-2''), 3.71 (1H, overlapped, H-6'''), 3.69 (1H, overlapped, H-6'''), 3.48 (1H, m, H-6''').

¹³C NMR (DMSO-*d*₆, 100 MHz): δ 182.0 (C-4), 163.4 (C-7), 162.9 (C-2), 160.6 (C-5), 160.3 (C-4'), 156.3 (C-9), 128.2 (C-2', 6'), 123.9 (C-1'), 116.6 (C-3', 5'), 109.0 (C-6), 103.8 (C-3), 103.5 (C-10), 99.9 (C-1''), 93.7 (C-8), 81.6 (C-5''), 78.9 (C-3''), 77.2 (C-5'''), 76.5 (C-3'''), 73.2 (C-2'''), 73.0 (C-1''), 70.6 (C-4''), 70.2 (C-2''), 69.6 (C-4'''), 61.5 (C-6''), 60.6 (C-6'''). ²²



Acacetin 7-*O*-β-D-glucoside (25a): ESI-MS *m*/*z* 491.1181 [M–H+HCOOH]⁻. ¹H NMR (DMSO-*d*₆, 400 MHz): δ 12.91 (1H, s, 5-OH), 8.07 (2H, d, *J* = 9.0 Hz, H-2', 6'), 7.13 (2H, d, *J* = 9.0 Hz, H-3', 5'), 6.95 (1H, s, H-3), 6.46 (1H, d, *J*=2.2 Hz, H-8), 6.86 (1H, d, *J*=2.2 Hz, H-6), 5.07 (1H, d, *J*=7.7 Hz, H-1''), 3.86 (3H, s, 4'-OCH₃).²³

¹³C NMR (DMSO-*d*₆,100 MHz): δ 182.1 (C-4), 163.1 (C-2), 162.9 (C-7), 162.1 (C-4'), 161.1 (C-5),

157.0 (C-9), 128.5 (C-2', 6'), 122.7 (C-1'), 114.7 (C-3', 5'), 105.5 (C-10), 103.4 (C-3), 100.0 (C-1"), 99.6 (C-6), 95.0 (C-8), 77.2 (C-3"), 76.5 (C-5"), 73.2 (C-2"), 69.5 (C-4"), 60.6 (C-6"), 55.6 (4'-OCH₃).²³

3. Supplementary Tables

Name	Organism	Genbank accession number	
ZjOGT3	Ziziphus jujuba var. spinosa	UGT84A68	
NmF4'GT (UGT88P1)	Nemophila menziesii	LC328827	
UGT88A1	Arabidopsis thaliana	Q9LK73	
UGT73A10	Lycium barbarum	AB360612	
MdPh-4'-OGT	Malus domestica	AY786997	
OsUGT-3	Oryza sativa	CT830931	
UGT73A10	Lycium barbarum	AB360612	
UGT74W1	Bacopa monniera	FJ586244	
Sb3GT1 (UGT78B4)	Scutellaria baicalensis	MK577650	
TcCGT1	Trollius chinensis	MK644229	
UGT708D1	Glycine max	BAR73279	
UGT708A6	Zea mays	NP_001132650.1	
SbCGTa	Scutellaria baicalensis	MK894443	
SbCGTb	Scutellaria baicalensis	MK894444	
GgCGT	Glycyrrhiza glabra	MH998596	
OsCGT	Oryza sativa	CAQ77160	
MiCGT	Mangifera indica	ALD83754	
GuCGTa	Glycyrrhiza uralensis	MK894447	
GuCGTb	Glycyrrhiza uralensis	MK894448	
GtUF6CGT1	Gentiana triflora	BAQ19550	
PlUGT43	Pueraria lobata	A0A172J2G3.2	

Table S1. Accession numbers of plant GTs used in this study.

Name		Sequence (5'-3')
ZjOGT3	F	TGGACAGCAAATGGGTCGCCGGATGGGGCTACAACCTCCAATCC
	R	TGTCGACGGAGCTCGAATTCGGAGTGGCAGCTGGTTTTGAATTG
ZjOGT 2	F	TGGACAGCAAATGGGTCGCCGGATGGAATCTGAACCTCTTGCCC
	R	TGTCGACGGAGCTCGAATTCGGAATTACTGAATCGACAATCCCA
ZjOGT 6	F	TGGACAGCAAATGGGTCGCCGGATGAAGAAATCATTAGCAGAAC
	R	TGTCGACGGAGCTCGAATTCGGGGGAAATATTCTCTATTACATCA
ZjOGT 10	F	TGGACAGCAAATGGGTCGCCGGATGAAGAAAATAAACCTTGTAG
	R	TGTCGACGGAGCTCGAATTCGGAGAGATGTTTGCTAGAATAGCC
ZjOGT 13	F	TGGACAGCAAATGGGTCGCCGGATGCAGAGCTCTAAACCACACG
	R	TGTCGACGGAGCTCGAATTCGGGGGCATTCTGTTGAACCTCACAA
ZjOGT 15	F	TGGACAGCAAATGGGTCGCCGGATGAAGAAAGCAGAGTTAGTT
	R	TGTCGACGGAGCTCGAATTCGGGGGACACATTCTTCAAGATATCA
F14A	F	gcCCCAGCTCAAGGCCATATCAATCCCATG
	R	GCCTTGAGCTGGGgcGCAAACTAGAAAGAG
H19A	F	gcTATCAATCCCATGCTCAGATTAGCAAAAC
	R	CATGGGATTGATAgcGCCTTGAGCTGGGAAG
H44A	F	gcCGCCGGCAAAGACATACGACAAGCCAAC
	R	GTCTTTGCCGGCGgcCTCGGTTGTAGAGAAG
R85A	F	gcAGACCTCGACTTCTACGTTCCTCAGCTTG
	R	GAAGTCGAGGTCTgcTCGTTTCGGATCGTC
L87A	F	gcCGACTTCTACGTTCCTCAGCTTGAACTC
	R	GAACGTAGAAGTCGgcGTCTCGTCGTTTCGG
F124A	F	gcaATCCCTTGGGTCTGCGATGTCGCAGAAG
	R	GACCCAAGGGATtgcAGGGTTGTTCACGAAG
Q145A	F	gcATCCTGTGCTGTTTTCAGCTGTTACTATC
	R	AAACAGCACAGGATgcAATCCAAAGAGTTGC
L199A	F	gcAGGAACCGCCATTCTAGGCCAGTTCAAG
	R	GAATGGCGGTTCCTgcGATTTTATAAGGACTAG
I285A	F	gcAGCTTACATCAAGCAAGAGCAAGTGGAAG
	R	CTTGATGTAAGCTgcGGTTCCGAATGAAAT
W382A	F	gcGGGGGATCAGGTAACGAACGCCAAGTTC
	R	TACCTGATCCCCCgcTTGAGGATAAGCCAG

Table S2. PCR primers used in this study.

Note: The mutation sites are showed in lower case.

Method	Solvent A	Solvent B	Elution gradient	Substrates
A	Water containing 0.1% formic acid	МеОН	10-10% B, 3 min; 10-50% B, 7 min; 50-50% B, 3 min; 50-100% B, 1 min.	1, 3, 5, 6, 12, 14, 16, 20-22, 30
В	Water containing 0.1% formic acid	МеОН	20-45% B, 3 min; 45-45% B, 1 min; 45-100% B, 4.5 min; 100-100% B, 3.5 min.	2, 4, 7-11, 13, 15, 17-19, 23- 29, 31-34

Table S3. UHPLC methods used in this study.

	ZjOGT3/UDP
Data collection [*]	
Space group	<i>P</i> 2 ₁
Cell dimensions	
<i>a</i> , <i>b</i> , <i>c</i> (Å)	61.73, 84.81, 103.63
α, β, γ (°)	90, 93.33, 90
Resolution (Å)	84.81-2.50 (2.60-2.50) ^a
$R_{\rm pim}^{[b]}(\%)$	9.6 (61.2)
$CC_{1/2}$ (%)	98.4 (74.7)
Ι/σΙ	5.4 (1.1)
Completeness (%)	97.5 (98.3)
Redundancy	4.1 (4.1)
Refinement	
No. reflections	35988
No. reflections (test sets)	1737
$R_{ m work}/R_{ m free}$ (%)	23.98/27.61
No. atoms	7169
Proteins	7160
UDP/H ₂ O	50/114
B-factors (Å ²)	
Proteins	48.28
UDP/H ₂ O	41.05/41.53
R.m.s.deviations	
Bond lengths (Å)	0.002
Bond angles (°)	0.481
Ramachandran Plot (%)	
Favored region	96.12%
Allowed region	3.66%
Outliers	0.22%

Table S4. Crystallographic data collection and refinement statistics^[a].

^[a] Numbers in brackets indicate values for the outermost resolution shell. One crystal was used to obtain this dataset.

^[b] R_{pim} is a redundancy-independent R factor used to evaluate the diffraction data quality as proposed by Evans.

Table S5. The overall glycosylation free energy barriers (in kcal/mol) for substrates **2**, **10** and **12** from DFT calculations and the binding free energies (in kcal/mol) with standard errors for the anionic intermediates of substrate **2**, **10** and **12** calculated by MM/GBSA.

substrates	$\Delta G_{act,4'}$	$\Delta G_{act,7}$	$\Delta G_{act,4}$ '- $\Delta G_{act,7}$	$\Delta G_{\text{bind},4'}$	$\Delta G_{bind,7}$	$\Delta G_{bind,4}$ - $\Delta G_{bind,7}$
2	26.8	25.9	0.9	-21.4 ± 0.9	-15.0 ± 0.4	-6.4
10	30.6	27.3	3.3	-22.4 ± 0.7	-22.1 ± 0.9	-0.3
12	25.0	25.4	-0.4	-32.2 ± 0.8	-36.8 ± 1.1	4.6

4. Supplementary Figures



Figure S1. SDS-PAGE of the His-tagged ZjOGT3 purified by affinity chromatography. Lane M: Protein marker.



Figure S2. A time-course study of the enzymatic reaction of ZjOGT3. Kumatakenin B (1) was used as the acceptor and UDP-Glc was used as the sugar donor.



Figure S3. Effects of reaction time (A), reaction buffer (B), temperature (C), and divalent metal ions (D) on glycosylation activity of ZjOGT3. Kumatakenin B (1) was used as the acceptor and UDP-Glc was used as the sugar donor. An optimized reaction time of 480 min was used. ZjOGT3 exhibited its maximum activity at pH 8.0 (50 mM NaH₂PO₄-Na₂HPO₄ buffer) and 45°C, and this enzyme was independent of divalent cations.



Figure S4. Determination of kinetic parameters for recombinant ZjOGT3. **1** and **1a** were used as sugar acceptor, respectively.



Figure S5. UHPLC/MS analysis of the enzyme catalytic reaction products of **1** by recombinant ZjOGT3. (A) HPLC chromatograms of the reaction products and reference standard **1**. (B) The reaction pathway. (C) (-)-ESI-MS and MS/MS spectra of **1a** and **1b**. The analytical conditions are given in Table S3.



Figure S6. UHPLC/MS analysis of the enzyme catalytic reaction products of **2** by recombinant ZjOGT3. (A) HPLC chromatograms of the reaction products and reference standard **2**. (B) The reaction pathway. (C) (-)-ESI-MS and MS/MS spectra of **2a** and **2b**. The analytical conditions are given in Table S3.



Figure S7. UHPLC/MS analysis of the enzyme catalytic reaction products of **3** by recombinant ZjOGT3. (A) HPLC chromatograms of the reaction products and reference standard **3**. (B) The reaction pathway. (C) (-)-ESI-MS and MS/MS spectra of **3a** and **3b**. The analytical conditions are given in Table S3.



Figure S8. UHPLC/MS analysis of the enzyme catalytic reaction products of **4** by recombinant ZjOGT3. (A) HPLC chromatograms of the reaction products and reference standard **4**. (B) The reaction pathway. (C) (-)-ESI-MS and MS/MS spectra of **4a** and **4b**. The analytical conditions are given in Table S3.



Figure S9. UHPLC/MS analysis of the enzyme catalytic reaction products of 5 by recombinant ZjOGT3. (A) HPLC chromatograms of the reaction products and reference standard 5. (B) The reaction pathway. (C) (-)-ESI-MS and MS/MS spectra of 5a and 5b. The analytical conditions are given in Table S3.



Figure S10. UHPLC/MS analysis of the enzyme catalytic reaction products of **6** by recombinant ZjOGT3. (A) HPLC chromatograms of the reaction products and reference standard **6**. (B) The reaction pathway. (C) (-)-ESI-MS and MS/MS spectra of **6a** and **6b**. The analytical conditions are given in Table S3.

10 -

0 | 200

250 300 350

400 450

m/z

В

10 =

0 ^{⊒∥}∺ 300

400

500

m/z

600

700



Figure S11. UHPLC/MS analysis of the enzyme catalytic reaction products of 7 by recombinant ZjOGT3. (A) HPLC chromatograms of the reaction products and reference standard 7. (B) The reaction pathway. (C) (-)-ESI-MS and MS/MS spectra of 7a and 7b. The analytical conditions are given in Table S3.

400

500

m/z

10

700

600

0 300

400

500

600

m/z

800

700



Figure S12. UHPLC/MS analysis of the enzyme catalytic reaction products of **8** by recombinant ZjOGT3. (A) HPLC chromatograms of the reaction products and reference standard **8**. (B) The reaction pathway. (C) (-)-ESI-MS and MS/MS spectra of **8a** and **8b**. The analytical conditions are given in Table S3.



Figure S13. UHPLC/MS analysis of the enzyme catalytic reaction products of **9** by recombinant ZjOGT3. (A) HPLC chromatograms of the reaction products and reference standard **9**. (B) The reaction pathway. (C) (-)-ESI-MS and MS/MS spectra of **9a** and **9b**. The analytical conditions are given in Table S3.



Figure S14. UHPLC/MS analysis of the enzyme catalytic reaction products of **10** by recombinant ZjOGT3. (A) HPLC chromatograms of the reaction products and reference standard **10**. (B) The reaction pathway. (C) (-)-ESI-MS and MS/MS spectra of **10a**. The analytical conditions are given in Table S3.



Figure S15. UHPLC/MS analysis of the enzyme catalytic reaction products of **11** by recombinant ZjOGT3. (A) HPLC chromatograms of the reaction products and reference standard **11**. (B) The reaction pathway. (C) (-)-ESI-MS and MS/MS spectra of **11a**. The analytical conditions are given in Table S3.



Figure S16. UHPLC/MS analysis of the enzyme catalytic reaction products of **12** by recombinant ZjOGT3. (A) HPLC chromatograms of the reaction products and reference standard **12**. (B) The reaction pathway. (C) (-)-ESI-MS and MS/MS spectra of **12a**. The analytical conditions are given in Table S3.



Figure S17. UHPLC/MS analysis of the enzyme catalytic reaction products of **13** by recombinant ZjOGT3. (A) HPLC chromatograms of the reaction products and reference standard **13**. (B) The reaction pathway. (C) (-)-ESI-MS and MS/MS spectra of **13a**. The analytical conditions are given in Table S3.


Figure S18. UHPLC/MS analysis of the enzyme catalytic reaction products of **14** by recombinant ZjOGT3. (A) HPLC chromatograms of the reaction products and reference standard **14**. (B) The reaction pathway. (C) (-)-ESI-MS and MS/MS spectra of **14a**. The analytical conditions are given in Table S3.



Figure S19. UHPLC/MS analysis of the enzyme catalytic reaction products of **15** by recombinant ZjOGT3. (A) HPLC chromatograms of the reaction products and reference standard **15**. (B) The reaction pathway. (C) (-)-ESI-MS and MS/MS spectra of **15a**. The analytical conditions are given in Table S3.



Figure S20. UHPLC/MS analysis of the enzyme catalytic reaction products of **16** by recombinant ZjOGT3. (A) HPLC chromatograms of the reaction products and reference standard **16**. (B) The reaction pathway. (C) (-)-ESI-MS and MS/MS spectra of **16a**. The analytical conditions are given in Table S3.



Figure S21. UHPLC/MS analysis of the enzyme catalytic reaction products of 17 by recombinant ZjOGT3. (A) HPLC chromatograms of the reaction products and reference standard 17. (B) The reaction pathway. (C) (-)-ESI-MS and MS/MS spectra of 17a. The analytical conditions are given in Table S3.



Figure S22. UHPLC/MS analysis of the enzyme catalytic reaction products of **18** by recombinant ZjOGT3. (A) HPLC chromatograms of the reaction products and reference standard **18**. (B) The reaction pathway. (C) (-)-ESI-MS and MS/MS spectra of **18a**. The analytical conditions are given in Table S3.



Figure S23. UHPLC/MS analysis of the enzyme catalytic reaction products of 19 by recombinant ZjOGT3. (A) HPLC chromatograms of the reaction products and reference standard 19. (B) The reaction pathway. (C) (-)-ESI-MS and MS/MS spectra of 19a. The analytical conditions are given in Table S3.



Figure S24. UHPLC/MS analysis of the enzyme catalytic reaction products of 20 by recombinant ZjOGT3. (A) HPLC chromatograms of the reaction products and reference standard 20. (B) The reaction pathway. (C) (-)-ESI-MS and MS/MS spectra of 20a. The analytical conditions are given in Table S3.



Figure S25. UHPLC/MS analysis of the enzyme catalytic reaction products of 21 by recombinant ZjOGT3. (A) HPLC chromatograms of the reaction products and reference standard 21. (B) The reaction pathway. (C) (-)-ESI-MS and MS/MS spectra of 21a. The analytical conditions are given in Table S3.



Figure S26. UHPLC/MS analysis of the enzyme catalytic reaction products of **22** by recombinant ZjOGT3. (A) HPLC chromatograms of the reaction products and reference standard **22**. (B) The reaction pathway. (C) (-)-ESI-MS and MS/MS spectra of **22a**. The analytical conditions are given in Table S3.



Figure S27. UHPLC/MS analysis of the enzyme catalytic reaction products of 23 by recombinant ZjOGT3. (A) HPLC chromatograms of the reaction products and reference standard 23. (B) The reaction pathway. (C) (-)-ESI-MS and MS/MS spectra of 23a. The analytical conditions are given in Table S3.



Figure S28. UHPLC/MS analysis of the enzyme catalytic reaction products of **24** by recombinant ZjOGT3. (A) HPLC chromatograms of the reaction products and reference standard **24**. (B) The reaction pathway. (C) (-)-ESI-MS and MS/MS spectra of **24a**. The analytical conditions are given in Table S3.



Figure S29. UHPLC/MS analysis of the enzyme catalytic reaction products of **25** by recombinant ZjOGT3. (A) HPLC chromatograms of the reaction products and reference standard **25**. (B) The reaction pathway. (C) (-)-ESI-MS and MS/MS spectra of **25a**. The analytical conditions are given in Table S3.



Figure S30. UHPLC/MS analysis of the enzyme catalytic reaction products of 26 by recombinant ZjOGT3. (A) HPLC chromatograms of the reaction products and reference standard 26. (B) The reaction pathway. (C) (-)-ESI-MS and MS/MS spectra of 26a. The analytical conditions are given in Table S3.



Figure S31. UHPLC/MS analysis of the enzyme catalytic reaction products of 27 by recombinant ZjOGT3. (A) HPLC chromatograms of the reaction products and reference standard 27. (B) The reaction pathway. (C) (-)-ESI-MS and MS/MS spectra of 27a. The analytical conditions are given in Table S3.



Figure S32. UHPLC/MS analysis of the enzyme catalytic reaction products of 28 by recombinant ZjOGT3. (A) HPLC chromatograms of the reaction products and reference standard 28. (B) The reaction pathway. (C) (-)-ESI-MS and MS/MS spectra of 28a. The analytical conditions are given in Table S3.



В



Figure S33. UHPLC/MS analysis of the enzyme catalytic reaction products of 29 by recombinant ZjOGT3. (A) HPLC chromatograms of the reaction products and reference standard 29. (B) The reaction pathway. (C) (-)-ESI-MS and MS/MS spectra of 29a and 29b. The analytical conditions are given in Table S3. 29a and 29b were confirmed by comparing with reference standards.



В



Figure S34. UHPLC/MS analysis of the enzyme catalytic reaction products of **30** by recombinant ZjOGT3. (A) HPLC chromatograms of the reaction products and reference standard **30**. (B) The reaction pathway. (C) (-)-ESI-MS and MS/MS spectra of **30a** and **30b**. The analytical conditions are given in Table S3.



Figure S35. UHPLC/MS analysis of the enzyme catalytic reaction products of **31** by recombinant ZjOGT3. (A) HPLC chromatograms of the reaction products and reference standard **31**. (B) The reaction pathway. (C) (-)-ESI-MS and MS/MS spectra of **31a**. The analytical conditions are given in Table S3.



Figure S36. UHPLC/MS analysis of the enzyme catalytic reaction products of 32 by recombinant ZjOGT3. (A) HPLC chromatograms of the reaction products and reference standard 32. (B) The reaction pathway. (C) (-)-ESI-MS and MS/MS spectra of 32a. The analytical conditions are given in Table S3.



Figure S37. UHPLC/MS analysis of the enzyme catalytic reaction products of 33 by recombinant ZjOGT3. (A) HPLC chromatograms of the reaction products and reference standard 33. (B) The reaction pathway. (C) (-)-ESI-MS and MS/MS spectra of 33a. The analytical conditions are given in Table S3. 33a were confirmed by comparing with reference standards. 29b and 33a are the same compound.

Α

В



Figure S38. UHPLC/MS analysis of the enzyme catalytic reaction products of 34 by recombinant ZjOGT3. (A) HPLC chromatograms of the reaction products and reference standard 34. (B) The reaction pathway. (C) (-)-ESI-MS and MS/MS spectra of 34a and 34b. The analytical conditions are given in Table S3.

m/z

m/z

m/z

m/z



Figure S40. ¹³C NMR spectrum of **3a** in DMSO- d_6 (100 MHz).



Figure S41. HMBC spectrum of 3a in DMSO-*d*₆ (400 MHz).



Figure S42. HSQC spectrum of 3a in DMSO-*d*₆ (400 MHz).



Figure S44. ¹³C NMR spectrum of **3b** in DMSO- d_6 (100 MHz).



Figure S45. HMBC spectrum of 3b in DMSO-*d*₆ (400 MHz).



Figure S46. ¹H NMR spectrum of 7a in DMSO- d_6 (400 MHz).



Figure S47. ¹³C NMR spectrum of 7a in DMSO- d_6 (100 MHz).



Figure S48. HMBC spectrum of 7a in DMSO-*d*₆ (400 MHz).



Figure S49. HSQC spectrum of 7a in DMSO-d₆ (400 MHz).



Figure S50. ¹H NMR spectrum of 7b in DMSO- d_6 (400 MHz).



Figure S51. ¹³C NMR spectrum of 7b in DMSO- d_6 (100 MHz).



Figure S52. HMBC spectrum of 7b in DMSO-d₆ (400 MHz).



Figure S53. HSQC spectrum of 7b in DMSO-d₆ (400 MHz).



Figure S54. ¹H NMR spectrum of 10a in DMSO- d_6 (400 MHz).





Figure S56. HMBC spectrum of 10a in DMSO-*d*₆ (400 MHz).



Figure S57. HSQC spectrum of 10a in DMSO- d_6 (400 MHz).



Figure S58. ¹H NMR spectrum of 11a in DMSO- d_6 (400 MHz).



Figure S59. ¹³C NMR spectrum of 11a in DMSO- d_6 (100 MHz).



Figure S60. HMBC spectrum of 11a in DMSO- d_6 (400 MHz).



Figure S62. ¹³C NMR spectrum of 12a in DMSO- d_6 (100 MHz).



Figure S63. HMBC spectrum of 12a in DMSO- d_6 (400 MHz).



Figure S64. HSQC spectrum of 12a in DMSO- d_6 (400 MHz).



Figure S65. ¹H NMR spectrum of 20a in DMSO- d_6 (400 MHz).



Figure S66. ¹³C NMR spectrum of **20a** in DMSO- d_6 (100 MHz).



Figure S67. HMBC spectrum of 20a in DMSO- d_6 (400 MHz).



Figure S68. HSQC spectrum of 20a in DMSO- d_6 (400 MHz).



Figure S69. ¹H-¹H COSY spectrum of 20a in DMSO- d_6 (400 MHz).



Figure S70. ¹H NMR spectrum of 21a in DMSO- d_6 (400 MHz).



Figure S71. ¹³C NMR spectrum of 21a in DMSO- d_6 (100 MHz).



Figure S72. HMBC spectrum of 21a in DMSO-*d*₆ (400 MHz).



Figure S73. HSQC spectrum of 21a in DMSO-d₆ (400 MHz).



Figure S74. ¹H NMR spectrum of 22a in DMSO- d_6 (400 MHz).



Figure S75. ¹³C NMR spectrum of 22a in DMSO- d_6 (100 MHz).



Figure S76. HMBC spectrum of 22a in DMSO-*d*₆ (400 MHz).



Figure S77. HSQC spectrum of 22a in DMSO- d_6 (400 MHz).



Figure S78. ¹H NMR spectrum of 25a in DMSO- d_6 (400 MHz).



Figure S79. ¹³C NMR spectrum of 25a in DMSO- d_6 (100 MHz).



Figure S80. Sugar donor promiscuity of ZjOGT3. (A) Chemical structures of sugar donors used in this study. (B) Conversion rates of compound **1** using different sugar donors. N.D., products were not detected.



Figure S81. Photo of ZjOGT3 crystals. Purified ZjOGT3 (20 mg/mL) was incubated with 5 mM UDP-Glc at 4°C for 1 h before setting up the crystallization trays. The crystals were prepared by hanging drop vapor diffusion. The crystals were obtained in 0.2 M sodium acetate trihydrate, 0.1 M Tris-HCl 8.00, 26% *w/v* PEG 4000 after 2 days.


Figure S82. Root-mean-square deviation (RMSD) of backbone heavy atoms relative to the first snapshot during 200 ns MD simulation of these two anionic intermediates for the ternary complex (A) ZjOGT3:UDP-Glc:2, (B) ZjOGT3:UDP-Glc:10, (C) ZjOGT3:UDP-Glc:12.



Figure S83. Distances between the imidazole ring hydrogen atom (HE2) of H16 and the phenol anion oxygen atom (O) of the two anionic intermediates during 200 ns MD simulation for the ternary complex ZjOGT3:UDP-Glc:**2**.



Figure S84. Per-residue contribution of binding energies and key residues of ZjOGT3 (| contribution $| \ge 1.0$ kcal/mol) with 4' and 7 anion orientation of acceptor substrates (A) 2, (B) 10 and (C) 12.



Figure S85. UHPLC analysis of reaction mixtures catalyzed by site-directed mutagenesis based on the results of molecule docking of ZjOGT3. The substrate was apigenin (2). The reaction mixtures were individually performed in a final volume of 100 μ L containing 50 mM NaH₂PO₄-Na₂HPO₄ buffer (pH 8.0), 25 µg of purified ZjOGT3, 0.1 mM aglycone, 0.5 mM sugar-donor, and 45°C for 2 h.



Figure S86. The molecular details for 7-*O*-glycosylation of **2** catalyzed by ZjOGT3-W382A mutant. (A) Close-up view of representative ternary complex MD snapshots of ZjOGT3-W382A active site in complex with UDP-Glc and the anionic intermediates of acceptor **2**. (B) MD plots for the distances to form the O-C bond d(O-C1) and the angles between the forming O-C bond and the breaking C-O bond a(O-C1-O1) for the two anionic intermediates in the ternary complex. (C) Root-mean-square deviation (RMSD) of backbone heavy atoms relative to the first snapshot during 200-ns MD simulation of these two anionic intermediates for the ternary complex ZjOGT3-W382A/UDP-Glc/**2**. (D) Distances between the imidazole ring hydrogen atom (HE2) of H16 and the phenolic anion oxygen atom (O) of these two anionic intermediates during 200-ns MD simulation for the ternary complex ZjOGT3-W382A/UDP-Glc/**2**.



Figure S87. The molecular details for 7-*O*-glycosylation of ZjOGT3-L199A mutant. (A) Close-up view of representative ternary complex MD snapshots of ZjOGT3-L199A active site in complex with UDP-Glc and the anionic intermediates of acceptor **2**. (B) MD plots for the distances to form the O-C bond d(O-C1) and the angles between the forming O-C bond and the breaking C-O bond a(O-C1-O1) for the two anionic intermediates in the ternary complex. (C) Root-mean-square deviation (RMSD) of backbone heavy atoms relative to the first snapshot during 200-ns MD simulation of these two anionic intermediates for the ternary complex ZjOGT3-L199A/UDP-Glc/**2**. (D) Distances between the imidazole ring hydrogen atom (HE2) of H16 and the phenolic anion oxygen atom (O) of these two anionic intermediates during 200-ns MD simulation for the ternary complex ZjOGT3-L199A/UDP-Glc/**2**.



Figure S88. (A) Gibbs free energy profiles and DFT-optimized transition state structures for 7-*O*-glycosylation of **2a**. Computed at the CPCM(water)-B3LYP-D3/6-311++G(2d,p)//CPCM(water)-B3LYP-D3/6-31+G(d,p) level of theory (Carbon, gray; Hydrogen, white; Oxygen, red; Nitrogen, blue; Phosphorus, orange; distances are shown in Å). (B) Close-up view of representative ternary complex MD snapshots of ZjOGT3 active site in complex with UDP-Glc donor and the anionic intermediate of acceptor **2a** (2a-Int). (C) MD plots for the distances to form the O-C bond d(O-C1) and the angles between the forming O-C bond and the breaking C-O bond a(O-C1-O1) for the anionic intermediate in the ternary complex. (D) Root-mean-square deviation (RMSD) of backbone heavy atoms relative to the first snapshot during 200-ns MD simulation of the anionic intermediate for the ternary complex ZjOGT3/UDP-Glc/**2a**. (E) Distance between the imidazole ring hydrogen atom (HE2) of H16 and the phenolic anion oxygen atom (O) of the anionic intermediate during 200-ns MD simulation for the ternary complex ZjOGT3/UDP-Glc/**2a**.



Figure S89. Close-up view of docking models of ZjOGT3 active site in complex with donor UDP-Glc and the anionic intermediate of acceptors (A) **10a**, (B) **12a** and (C) **22a**.



Figure S90. Functional characterization of ZjOGT 2/3/6/10/13/15. Apigenin and kaempferol were used as sugar acceptor, respectively. UDP-Glc was used as sugar donor.

Through blast with reported UGTs, 24 open reading frames (ORFs) were obtained as candidate genes from the transcriptome of *Z. jujuba* var. *spinosa*. After cloning and functional characterization, eight UGTs (ZjOGT 1/2/3/4/6/10/13/15) exhibited glycosylation activities. Among them, ZjOGT1 and ZjOGT4 were identified as 2-hydroxy flavanone CGTs, which were respectively named as ZjCGT1 and ZjCGT2 in our previous report (Zhang YQ, *et al. Chem. Commun.* 2022, *58*, 2472-2475.). ZjOGT 2/3/6/10/13/15 exhibited *O*-glycosylation activities. Apigenin and kaempferol were used as sugar acceptors to investigate their catalytic features.

ZjOGT2 and ZjOGT3 showed similar activities to produce 4'-O- and 7,4'-di-O-glycosides. ZjOGT6 and ZjOGT10 exhibited moderate 7-O-glycosylation activities towards apigenin, but potent 3-O-glycosylation activities towards kaempferol. ZjOGT13 and ZjOGT15 mainly showed moderate 7-O-glycosylation activities.

5. Calculated coordinates

UDP-Glc

G(Water) = -2656.886855 Hartree

Ν	-4.143224	0.432193	0.537576
С	-5.388458	0.978026	0.849138
Ν	-5.547703	2.296203	0.467188
С	-4.628995	3.118981	-0.200051
С	-3.374095	2.464210	-0.492144
С	-3.184873	1.177688	-0.117024
0	-6.283144	0.352294	1.414914
0	-4.935027	4.282315	-0.478838
С	-3.879037	-0.968454	0.928263
С	-3.791101	-1.938227	-0.261177
0	-4.375184	-3.171081	0.145791
С	-2.275639	-2.106528	-0.480323
С	-1.711894	-1.925331	0.947968
0	-2.639333	-1.037411	1.615775
0	-2.035191	-3.376513	-1.045520
С	-0.350993	-1.244593	1.085443
0	0.736769	-2.025191	0.573051
Р	1.211249	-2.101199	-0.999632
0	0.511860	-3.271217	-1.672930
0	2.719752	-2.050594	-1.016776
0	0.515174	-0.762507	-1.656498
Р	0.847793	0.857448	-1.693217
0	-0.482407	1.563422	-1.681857
0	1.857524	1.134912	-2.791730
0	1.563348	0.958489	-0.221160
С	2.426562	1.934363	0.371696
С	3.778284	2.072071	-0.357840
С	4.411175	0.686436	-0.444606
С	4.465865	-0.001460	0.930696
С	3.183454	0.155475	1.770607
С	3.413368	-0.099376	3.252975
0	3.658982	2.725851	-1.609009
0	5.737150	0.810267	-0.977605
0	4.763381	-1.390185	0.761688
0	2.644847	1.499127	1.689788
0	4.334283	0.826108	3.838585
Н	-6.449629	2.703647	0.690541
Н	-2.589286	2.998384	-1.008518
Н	-2.242752	0.690003	-0.326118
Н	-4.696182	-1.254241	1.588845
Н	-3.867232	-3.872634	-0.296926

Н	-1.075555	-3.417304	-1.322523
Н	-4.303269	-1.541792	-1.145642
Н	-1.903278	-1.311611	-1.138047
Н	-1.706309	-2.901188	1.449372
Н	-0.377004	-0.262254	0.612496
Н	-0.135620	-1.099852	2.147172
Н	1.918143	2.901567	0.432028
Н	4.399107	2.719287	0.272371
Н	3.802486	0.070815	-1.114215
Н	5.305195	0.433100	1.483850
Н	2.434198	-0.553635	1.408111
Н	2.447079	-0.063471	3.774049
Н	3.842768	-1.095914	3.381248
Н	6.116439	-0.080770	-0.994146
Н	4.051145	-1.758954	0.189201
Н	3.073876	2.172053	-2.183728
Н	4.002525	1.717879	3.656892

HPO4²⁻

G(Water) = -643.4070798 Hartree

781
621
256
990
54

H₂PO₄⁻

G(Water) = -643.8805478 Hartree

Р	0.000843	0.095080	0.127473
0	-1.298632	-0.346939	-0.807045
0	0.017942	1.602809	0.135435
0	-0.011526	-0.736627	1.393960
0	1.294370	-0.375447	-0.801316
Н	1.508652	-1.306107	-0.639683
Н	-1.538532	-1.270462	-0.640677

2

G(Water) = -953.9117166 Hartree

С	-4.188341	-0.556969	0.024937
С	-3.478586	0.639240	-0.025248
С	-2.056390	0.634461	-0.018159

С	-1.403234	-0.610385	0.039485			
С	-2.087527	-1.817560	0.089310			
С	-3.486491	-1.772094	0.080584			
Н	-5.273098	-0.535398	0.019135			
С	-1.279432	1.858199	-0.076839			
Н	-1.557679	-2.760756	0.133613			
С	0.723430	0.453904	-0.007701			
С	0.149883	1.692173	-0.080500			
Н	0.766189	2.577440	-0.161883			
С	2.160785	0.161623	0.002392			
С	3.104052	1.163423	0.309214			
С	2.631592	-1.131095	-0.296146			
С	4.464938	0.888693	0.307674			
Н	2.777595	2.163438	0.572459			
С	3.994036	-1.414452	-0.301552			
Н	1.927412	-1.919329	-0.535428			
С	4.916372	-0.403187	0.000020			
Н	5.188327	1.658994	0.552722			
Н	4.341355	-2.415243	-0.541972			
0	-4.146085	1.809759	-0.082145			
0	-4.130929	-2.969548	0.129060			
Н	-5.091358	-2.845899	0.115526			
0	6.263082	-0.611779	0.016379			
Н	6.471251	-1.533190	-0.197215			
0	-0.035506	-0.675338	0.054467			
0	-1.836992	2.992915	-0.133794			
Н	-3.449084	2.526022	-0.113528			
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С	-4.222927	-0.613105	0.027418			
С	-3.527277	0.575858	-0.028021			
С	-2.094856	0.598689	-0.019290			
С	-1.438772	-0.657616	0.048452			
С	-2.106149	-1.860675	0.104105			
С	-3.548518	-1.890499	0.095095			
Н	-5.308856	-0.600761	0.018782			
С	-1.338264	1.811942	-0.085068			
Н	-1.552696	-2.792270	0.156432			
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С

H C

С

С

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0.707673

2.131161

3.061026

2.620597

1.663260

2.555301

0.160508

1.163218

-1.115953

С	4.426730	0.906639	0.339939
Н	2.718779	2.150389	0.636499
С	3.987862	-1.382465	-0.334574
Н	1.926049	-1.905420	-0.589415
С	4.896155	-0.369177	-0.002561
Н	5.139000	1.679272	0.609806
Н	4.348366	-2.372195	-0.601010
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0	-0.057351	-0.703217	0.066965
0	-1.890904	2.960389	-0.150820
Н	-3.491259	2.469624	-0.128747

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G(Water) = -953.4589379 Hartree

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С	-2.009209	0.636949	-0.001198
С	-1.362690	-0.609662	0.001407
С	-2.057379	-1.814583	0.004946
С	-3.455620	-1.760497	0.006116
Н	-5.234411	-0.510707	0.004548
С	-1.218509	1.859672	-0.006460
Н	-1.533746	-2.762509	0.007021
С	0.785902	0.440708	-0.002896
С	0.194086	1.691768	-0.008224
Н	0.809562	2.581064	-0.015520
С	2.196071	0.143576	-0.001300
С	3.174655	1.175955	0.021581
С	2.674026	-1.196169	-0.021821
С	4.524495	0.897723	0.022615
Н	2.864404	2.216536	0.040968
С	4.022726	-1.487295	-0.021292
Н	1.958196	-2.011643	-0.039564
С	5.033463	-0.455630	0.001101
Н	5.248560	1.708547	0.041803
Н	4.358750	-2.521405	-0.038351
0	-4.083771	1.833088	-0.002767
0	-4.109270	-2.957989	0.009776
Н	-5.068277	-2.824973	0.010368
0	6.284355	-0.716817	0.002899
0	0.002377	-0.684941	0.001141
0	-1.791248	2.999486	-0.010477

-0.086919

-0.176084

0.005941

0.345283

-0.326459

Н	-3.364924	2.537328	-0.005805	Н	-2.127229	-4.586523	0.347808
				Н	1.303981	-4.012858	1.904947
2-TSa				Н	-0.749229	-2.889568	1.321315
G(Water	r) = -3610.304	581 Hartree		Н	-0.471082	-3.711065	-1.596662
				Н	-1.193499	-1.029923	-0.265018
Ν	2.252945	-1.755290	0.987418	Н	-0.918249	-1.263327	-2.003061
С	3.564224	-1.768758	1.457971	Н	-1.545551	2.018285	0.551895
Ν	3.920697	-0.661790	2.202908	Н	-3.953265	3.289508	-0.026139
С	3.111322	0.419953	2.577519	Н	-4.655471	0.339593	0.299506
С	1.764420	0.333987	2.064015	Н	-5.057583	2.118907	-2.125812
С	1.393218	-0.728336	1.309148	Н	-3.114086	-0.109860	-2.393697
0	4.349538	-2.689066	1.237490	Н	-1.809389	1.554490	-3.642443
0	3.573927	1.317708	3.287867	Н	-3.497490	1.569676	-4.191038
С	1.838607	-2.880019	0.106833	Н	-6.753569	1.206425	-0.239239
С	1.061561	-3.983122	0.836379	Н	-5.341413	-0.669970	-1.629032
0	1.381999	-5.223597	0.214520	Н	-3.857374	1.464399	2.171285
С	-0.408889	-3.611107	0.570596	Н	-2.376420	3.461746	-2.331146
С	-0.340864	-2.943255	-0.823812	С	0.213775	4.886062	0.253272
0	1.000585	-2.394390	-0.919583	С	1.544660	4.807578	0.626469
0	-1.184568	-4.792667	0.616300	С	2.371752	3.736897	0.169406
С	-1.257228	-1.748036	-1.083380	С	1.777661	2.787152	-0.688878
0	-2.633343	-2.079281	-1.313804	С	0.459903	2.858383	-1.099245
Р	-3.696585	-2.603312	-0.162534	С	-0.371308	3.905880	-0.605184
0	-3.639935	-4.131428	-0.149234	Н	-0.408198	5.688856	0.635306
0	-5.021248	-1.946691	-0.517386	С	3.747398	3.589352	0.570978
0	-3.098552	-2.140506	1.247734	Н	0.051019	2.094212	-1.747505
Р	-2.672435	-0.825833	2.287286	С	3.803453	1.534286	-0.759023
0	-2.016798	-1.568669	3.448481	С	4.427526	2.418545	0.071579
0	-4.000757	-0.104136	2.611826	Н	5.440882	2.238305	0.404245
0	-1.726368	0.045310	1.439893	С	4.377387	0.299675	-1.309987
С	-2.407680	1.869975	-0.075853	С	5.764742	0.054280	-1.269030
С	-3.778927	2.291865	0.398697	С	3.544228	-0.679805	-1.880421
С	-4.815556	1.317945	-0.162458	С	6.298157	-1.126149	-1.769436
С	-4.692315	1.187503	-1.681869	Н	6.441519	0.793961	-0.855735
С	-3.248281	0.947979	-2.168301	С	4.070171	-1.866303	-2.382561
С	-2.836789	1.817138	-3.353373	Н	2.472040	-0.533481	-1.906282
0	-3.804605	2.400920	1.798521	С	5.450162	-2.094951	-2.325510
0	-6.114651	1.803877	0.177779	Н	7.366755	-1.310683	-1.738693
0	-5.545196	0.150268	-2.153056	Н	3.406150	-2.616431	-2.802830
0	-2.205267	1.197086	-1.141418	0	2.076732	5.743952	1.453487
0	-2.969280	3.201942	-3.088068	0	-1.650090	3.940056	-0.876975
Н	4.875762	-0.660410	2.544601	0	6.030427	-3.238952	-2.790862
Н	1.055540	1.116533	2.295049	н	5.360789	-3.840524	-3.148109
Н	0.379916	-0.797972	0.937743	0	2.515067	1.723556	-1.153647
Н	2.760430	-3.276265	-0.315838	0	4.319471	4.429391	1.333463
Н	0.550392	-5.729510	0.190683	- Н	3.028089	5.475176	1.596459
					2.220002		

2-TSb				Н	0.257549	2.839379	0.763948
<i>G</i> (Water) = -3610.303106 Hartree			Н	0.038256	2.700776	-2.270417	
				Н	1.232452	0.730210	-0.224428
Ν	-2.365017	1.112288	0.808576	Н	1.110054	0.463712	-1.971066
С	-3.625303	1.205852	1.391859	Н	2.333236	-1.971098	0.849125
Ν	-3.807586	0.406644	2.502678	Н	4.955009	-2.852068	0.533369
С	-2.859029	-0.410580	3.133061	Н	5.102716	0.194547	0.573533
С	-1.557633	-0.368401	2.511354	Н	6.002951	-1.667642	-1.642079
С	-1.362934	0.370552	1.392715	Н	3.679138	0.065808	-2.263722
0	-4.511075	1.937447	0.953465	Н	2.836601	-1.946653	-3.369949
0	-3.180701	-1.071449	4.126753	Н	4.545737	-1.719805	-3.781965
С	-2.141735	1.931087	-0.418310	Н	7.352518	-0.305724	0.259072
С	-1.672965	3.352981	-0.090008	Н	5.626208	1.142425	-1.435557
0	-2.165319	4.222588	-1.104133	Н	4.312127	-0.846015	2.476145
С	-0.141473	3.232905	-0.179226	Н	3.536024	-3.576220	-1.816077
С	0.037033	2.185901	-1.301615	С	-6.672862	0.463054	-1.575180
0	-1.141838	1.338400	-1.214176	С	-6.458906	-0.404552	-0.507534
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С	1.219474	1.226764	-1.195809	С	-4.262976	-0.921226	-1.386691
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0	3.002371	4.204086	-0.569578	Н	-7.609520	1.006492	-1.644872
0	4.797466	2.323726	-0.508834	С	-4.969196	-2.027468	0.709728
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0	4.059091	0.732740	2.665006	Н	-3.449891	-3.349617	1.502360
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Н	-4.737928	0.428421	2.906488	0	2.602091	-4.020295	-0.507030
Н	-0.745723	-0.943087	2.933183	0	-3.061932	-1.570284	-1.319983
Н	-0.392939	0.394612	0.916246	0	-5.834569	-2.223048	1.618518
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С	5.739655	0.192761	0.834017	Н	-6.977931	-2.164580
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0	6.892285	0.154560	1.674767			
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Н	3.299499	0.762039	-2.017568	С	5.228743	1.635430
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Н	4.979424	-0.377574	1.375060	С	6.105168	-2.009797
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Н	6.300912	-2.330736	-1.515064	О	5.058900	2.352139
Н	7.035639	-2.233668	0.100864	О	6.864228	0.187005
Н	5.859387	2.133994	2.406631	0	5.015372	-0.124554
Н	7.645529	0.562544	1.219904	0	4.993062	-2.633221
Н	4.316040	3.074447	-1.369563	Н	3.282784	0.697321
Н	4.227564	-2.244361	-0.235158	Н	3.101342	1.501165
С	-7.234103	0.985548	0.138698	Н	5.969547	2.146069
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С	-5.313922	-0.524347	0.152393	Н	6.946512	-0.109102
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С	-4.735352	-1.851759	0.234174	Н	4.294227	3.026877
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С	1.099605	-1.769284	0.098497	Н	-8.342181	1.194323
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С	0.931689	0.485203	-0.792293	Н	-4.305564	2.772332
Н	-1.049654	1.275402	-0.948631	С	-2.585994	-0.757723
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Н	1.377903	1.393900	-1.177479	С	-1.119484	-0.713132
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Н	-1.068027	1.270403	-0.991299	О	-7.253163	-2.151182	-0.347345
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Н	1.669802	-2.596948	0.456687	0	-3.678875	-0.225679	-1.291257
Н	1.363327	1.367965	-1.216400	0	-5.285073	0.780437	-3.489961
0	-7.601070	-1.307247	0.350091	Н	2.352098	3.015774	2.960121
0	-6.887315	3.377981	-0.058969	Н	-1.663164	1.882217	2.310227
0	3.049225	-0.688383	-0.505160	Н	-0.805424	-0.153378	1.275972
0	-3.145632	0.482014	-0.071509	Н	2.743981	-0.555984	0.467512
0	-5.507322	-2.848880	0.326421	Н	2.572698	-3.836485	0.983975
Н	-7.014044	-2.115700	0.372516	Н	-0.239458	-4.676437	0.820696
				Н	1.815993	-2.019063	2.601003
2a-TS				Н	-0.382402	-2.484123	1.718813
G(Water)	= -4221.130	185 Hartree		Н	0.740951	-2.981639	-1.065869
				Н	-1.667069	-1.329225	-0.087559
Ν	1.230388	0.329858	1.534176	Н	-1.088271	-1.407558	-1.762774
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Ν	1.665100	2.361154	2.601603	Н	-6.473101	0.353902	-0.560440
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Н	3.306551	4.787122	-0.392143	Н	1.635450	-2.797237	0.065277
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0	8.110304	0.530317	-2.205765	Н	3.432730	2.531569	-0.094770
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10				С	1.326931	1.773536	-0.156346
G(Wa	(ter) = -955.09781	79 Hartree		Н	1.632251	-2.830429	0.050526
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С	3.472988	0.641982	-0.005239	Н	-0.458933	1.596072	-1.311516
С	2.046517	0.610661	-0.015325	С	-2.130048	0.085703	0.265895
С	1.404136	-0.658322	0.014146	С	-3.020782	0.209174	1.339619
С	2.135715	-1.837158	0.034188	С	-2.642433	-0.262683	-0.991810
С	3.536938	-1.761491	0.033311	С	-4.390504	0.001253	1.168897
Н	5.299112	-0.487224	0.025350	Н	-2.644390	0.472532	2.324566
С	1.272085	1.820870	-0.151365	С	-4.007130	-0.484352	-1.176638

	Н	-1.968147	-0.372979	-1.835750		
	С	-4.884484	-0.347436	-0.092173		
	Н	-5.079226	0.098958	2.001904		
	Н	-4.389810	-0.759775	-2.155891		
	0	4.163525	1.781939	-0.023407		
	0	4.269243	-2.960250	0.076499		
	0	-6.236704	-0.546035	-0.206040		
	Н	-6.468710	-0.780650	-1.115940		
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	0	1.849686	2.926663	-0.218504		
	Н	3.451719	2.477870	-0.095711		
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1	10-IMb					
<i>G</i> (Water) = -954.6330653 Hartree						
	С	4.170384	-0.570059	0.021838		
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C	+.17050+	-0.570057	0.021050
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Н	3.421416	2.505950	0.116939

Н	-0.526889	0.436671	1.495468		
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10-TSa

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N	2.151527	-2.565361	0.664900
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С	1.429464	-1.565007	1.276653
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0	3.333751	-1.194232	4.293268
С	1.782363	-3.115691	-0.678322
С	0.935851	-4.386590	-0.551818
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С	-0.499676	-3.836581	-0.609881
С	-0.349831	-2.644267	-1.580825
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0	-1.355640	-4.862182	-1.063647
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0	-4.989913	-1.571278	-0.785282
0	-3.057215	-2.351045	0.805335
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С	-2.353578	1.867400	0.530285
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С	-4.793065	1.457928	0.371417
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С	-3.267946	1.473609	-1.698159
С	-2.838056	2.533787	-2.707593
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0	-2.958243	3.855656	-2.210957
Н	4.385502	-3.010144	2.964021
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Н	0.564559	-1.199299	0.742159
Н	2.717969	-3.309008	-1.201961

Н	0.360683	-5.630098	-1.915453	Н	-0.246994	6.306674
Н	-2.301437	-4.523868	-1.072610	Н	5.705288	3.376815
Н	1.145233	-4.928379	0.377101	Н	3.859744	2.667027
Н	-0.800739	-3.473437	0.381197			
Н	-0.459183	-3.007318	-2.609882	10-TSb		
Н	-1.143156	-1.063776	-0.315671	G(Water) =	-3611.482	523 Hartree
Н	-0.900046	-0.625235	-2.014223			
Н	-1.466898	1.895741	1.141568	Ν	-2.153979	1.644656
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Н	-4.993276	2.716324	-1.342374	С	-3.008354	0.076080
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Н	-1.811942	2.313684	-3.033226	С	-1.264545	0.848411
Н	-3.497958	2.444628	-3.576940	0	-4.281489	2.525554
Н	-6.735226	1.479627	0.313768	0	-3.480556	-0.599123
Н	-5.406846	-0.089176	-1.542269	С	-1.746194	2.421252
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Н	-2.396257	3.963011	-1.400488	0	-1.436486	4.688062
С	0.349788	5.438794	0.366918	С	0.403778	3.452037
С	1.729182	5.535026	0.331524	С	0.491750	2.357540
С	2.552260	4.402728	0.012580	0	-0.795193	1.682279
С	1.881186	3.181360	-0.286596	0	1.103109	4.641245
С	0.507245	3.079827	-0.283729	С	1.513611	1.236816
С	-0.306553	4.204518	0.068658	0	2.876741	1.623466
С	3.978685	4.459414	0.073450	Р	3.794935	2.525804
Н	0.057070	2.139941	-0.572263	0	3.643928	3.991777
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С	4.483849	0.809615	-1.195356	Р	2.714887	1.335359
С	4.404517	0.148725	-2.429275	0	1.885330	2.254036
С	4.938125	0.089602	-0.082281	0	4.069773	0.872200
С	4.726339	-1.203947	-2.548896	0	1.941856	0.175897
Н	4.052763	0.688659	-3.304448	С	2.865934	-1.865960
С	5.282666	-1.256128	-0.191169	С	4.212370	-2.019747
Н	5.009417	0.564488	0.890788	С	5.226407	-1.128956
С	5.139176	-1.917599	-1.418860	С	5.239426	-1.400434
Н	4.626949	-1.721590	-3.497545	С	3.835953	-1.469434
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0	2.583295	2.043874	-0.561264	0	6.036954	-0.429843
0	4.615561	5.511592	0.348376	0	2.714124	-1.546819
Н	3.304827	6.559566	0.583677	0	3.911070	-3.905524
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Н	5.171527	-3.701568	-0.706239	Н	-0.906633	-0.530915
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Н	-2.653228	2.556986	-1.348822	Н	-2.042156	-0.5401
Н	-0.642338	5.234594	-1.590291	Н	-8.144635	0.59223
Н	2.084354	4.442563	-0.822710	Н	-2.218009	-1.2583
Н	-1.439117	4.145610	0.554292	Н	-3.527002	-3.1300
Н	0.716803	3.045417	0.523119			
Н	0.614873	2.838767	-2.514411	12		
Н	1.424319	0.791603	-0.357119	G(Water) = -1639.9942	289 Har
Н	1.303521	0.466057	-2.095485			
Н	1.959955	-1.994502	1.189069	С	-5.522887	-1.1053
Н	4.491047	-3.073529	1.142724	С	-4.310819	-1.6109
Н	4.954069	-0.085767	0.749869	С	-3.180836	-0.7535
Н	5.729803	-2.366577	-1.088677	С	-3.337983	0.6020
Н	3.629301	-0.539927	-2.104622	С	-4.536630	1.12900
Н	2.628361	-2.653387	-2.897006	С	-5.624536	0.25683
Н	4.354395	-2.581353	-3.312506	Н	-6.374799	-1.7698
Н	7.149064	-0.851030	0.615743	С	-1.897736	-1.2326
Н	5.713810	0.468714	-1.325079	Н	-4.626626	2.17708
Н	4.073680	-0.776164	2.785809	С	-1.058126	1.06702
Н	3.241069	-4.058214	-1.085462	С	-0.836689	-0.2515
С	-7.247796	-0.017471	-0.102061	С	-0.102768	2.17840
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С	-4.914996	-1.592348	-0.034657	С	1.863308	3.27953
С	-5.894972	-1.785840	0.928922	Н	1.146520	1.3760
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С	-3.981530	-0.331779	-1.962233	Н	-1.114645	3.27704
Н	-5.769289	-2.536167	1.699383	С	1.675858	4.35618
С	-2.971387	-2.497334	-1.120867	Н	2.696105	3.30202
С	-2.717838	-1.129707	-1.767295	Н	0.452335	5.1777
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С	-0.825479	-3.677643	-1.699891	0	-6.786345	0.79638
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С	0.984917	-3.995700	-0.073121	Н	-3.276318	-3.0802
Н	1.122061	-4.461496	-2.182542	С	1.302811	-1.1660
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0	2.255826	-4.213677	0.206294	C	4.035462	-1.9079
0	-3,802494	-2,358106	0.061025	C	3.405927	-0.5651
0	-4.048480	0.561307	-2.838017	C	4.391261	0.5945
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Н	-8,740415	-0.643226	1.747961	0	3.571740	-4.1798
н	0.417721	-3.533442	1.961144	0	5.012847	-2.2366
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Н	-2.042156	-0.540129	-1.137756
Н	-8.144635	0.592254	-0.125682
Н	-2.218009	-1.258360	-2.730437
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С	-5.522887	-1.105334	0.413103
С	-4.310819	-1.610919	-0.042365
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С	-4.536630	1.129068	0.626472
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Н	-6.374799	-1.769893	0.511099
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Н	-4.626626	2.177081	0.882536
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С	-0.836689	-0.251542	-0.692238
С	-0.102768	2.178408	-0.406357
С	0.984704	2.202894	-1.301849
С	-0.284082	3.273380	0.461749
С	1.863308	3.279530	-1.322618
Н	1.146520	1.376074	-1.979020
С	0.597923	4.348481	0.454713
Н	-1.114645	3.277046	1.158281
С	1.675858	4.356180	-0.442213
Н	2.696105	3.302025	-2.017491
Н	0.452335	5.177757	1.140569
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0	2.575378	5.375549	-0.507263
Н	2.362121	6.066317	0.137510
0	-2.288370	1.468703	0.047771
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С	1.302811	-1.166036	-0.152544
С	1.808245	-2.546699	-0.580687
С	2.948589	-2.977575	0.341719
С	4.035462	-1.907902	0.420681
С	3.405927	-0.565149	0.809198
С	4.391261	0.594583	0.762092
0	0.794754	-3.534661	-0.498513
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Н	2.539108	-3.131419	1.351566
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Н	2.979535	-0.638683	1.821340
Н	5.208987	0.401258	1.459069
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Н	3.067969	2.013051	0.530849
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С	-5.554786	-1.069032	0.442550
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Н	-6.408512	-1.731066	0.550722
С	-1.948127	-1.204779	-0.611617
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С	-0.109058	2.191599	-0.403653
С	0.970465	2.214908	-1.307371
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Н	1.114346	1.394736	-1.996930
С	0.633959	4.339467	0.481531
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С	1.704769	4.343992	-0.422336
Н	2.695460	3.296700	-2.020325
Н	0.505452	5.161345	1.179976
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0	-6.830943	0.782685	1.214913
0	2.621421	5.352593	-0.479260
Н	2.422238	6.034908	0.178547
0	-2.307149	1.505781	0.048081
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Н	-3.309043	-3.040186	-0.611486

С	1.254820	-1.165445	-0.163232
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С	2.870270	-3.002376	0.339061
С	3.976970	-1.953182	0.410643
С	3.371120	-0.597890	0.793504
С	4.375685	0.545112	0.742028
0	0.706558	-3.526118	-0.501276
0	3.469576	-4.220564	-0.107593
0	4.951093	-2.295046	1.394957
0	2.341163	-0.257445	-0.145804
0	3.785471	1.784940	1.128820
Н	2.119099	-2.471077	-1.613365
Н	2.459229	-3.141942	1.350445
Н	4.448146	-1.866981	-0.579983
Н	2.944241	-0.660608	1.806441
Н	5.188931	0.343759	1.442019
Н	4.793497	0.609510	-0.272847
Н	2.789294	-4.909389	-0.098341
Н	5.227813	-3.208218	1.227205
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Н	3.070024	1.979044	0.502648
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Н	0.761231	-1.199980	0.820089

12-IMb

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С	-4.514829	1.088002	0.677452
С	-5.592518	0.202316	0.781468
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С	2.939337	-2.937241	0.379460	Р	4.367272	0.186681	2.264126
С	4.041285	-1.881209	0.438502	0	3.977422	0.902622	3.555067
С	3.424538	-0.520079	0.781230	0	5.592292	-0.753273	2.365251
С	4.419601	0.630906	0.713763	0	3.209386	-0.446309	1.473129
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0	3.548708	-4.162268	-0.034455	С	4.893821	-2.893343	0.026831
0	5.005647	-2.197148	1.441084	С	6.054812	-2.060516	-0.522406
0	2.405439	-0.213155	-0.178391	С	5.879849	-1.758874	-2.012716
0	3.820155	1.872253	1.076866	С	4.467659	-1.262890	-2.385483
Н	2.198758	-2.457350	-1.590971	С	3.845909	-1.981676	-3.579873
Н	2.520663	-3.056276	1.390396	0	4.974383	-3.126514	1.409147
Н	4.523181	-1.816365	-0.548603	0	7.264121	-2.790884	-0.314898
Н	2.986280	-0.560712	1.790631	0	6.858450	-0.815530	-2.433958
Н	5.232241	0.446344	1.419553	0	3.464521	-1.402605	-1.297925
Н	4.841235	0.678995	-0.300588	0	3.759997	-3.381793	-3.387216
Н	2.871664	-4.854077	-0.016627	Н	-2.766551	1.437855	3.552337
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