

Bio-based vitrimers from divanillic acid and epoxidized soybean oil

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S1 ^1H and ^{13}C NMR spectra of DVA and BuDVA

^1H NMR of DVA and BuDVA:

DVA: ^1H NMR (500 MHz, $\text{DMSO-}d_6$) δ 12.54 (s, 2H), 9.33 (s, 2H), 7.46 (d, $J = 2.0$ Hz, 2H), 7.42 (d, $J = 2.0$ Hz, 2H), 3.89 (s, 6H).

BuDVA: ^1H NMR (500 MHz, $\text{DMSO-}d_6$) δ 12.88 (s, 2H), 7.56 (d, $J = 2.0$ Hz, 2H), 7.42 (d, $J = 2.0$ Hz, 2H), 3.88 (s, 6H), 3.81 (t, $J = 6.3$ Hz, 4H), 1.34 (p, 4H), 1.05 (h, 4H), 0.66 (t, $J = 7.4$ Hz, 6H).

^{13}C NMR spectrum of DVA:

DVA: ^{13}C NMR (126 MHz, $\text{DMSO-}d_6$) δ 167.11 (C1), 148.28 (C2), 147.17 (C3), 125.40 (C4), 124.26 (C5), 120.52 (C6), 111.01 (C7), 55.83 (C8).

(The ^{13}C NMR spectrum of BuDVA was reported in our previous study.¹⁾)

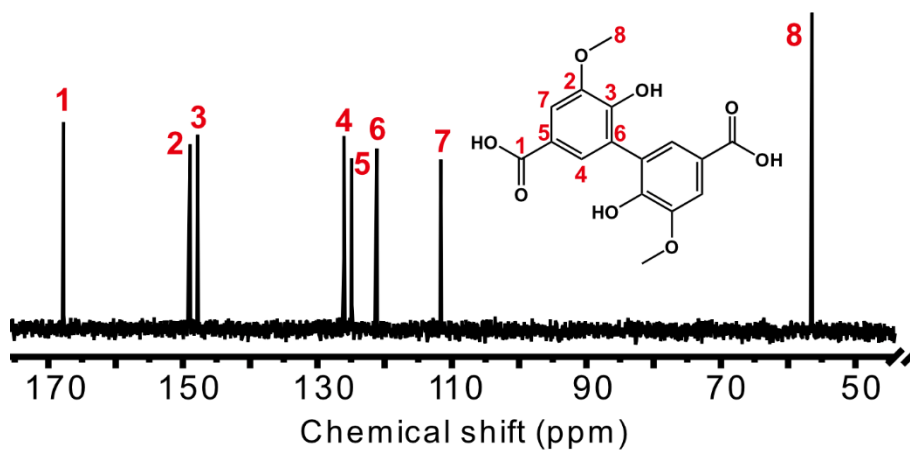


Fig. S1 ^{13}C NMR spectra of DVA in $\text{DMSO-}d_6$.

S2 Swelling experiment

The swell ratios and gel contents of OHESO-*x* and BuESO-*x* were measured to evaluate the crosslinking densities of cured vitrimers. A solvent extraction method according to ASTM D2765-16 with THF as solvent was used. Cured vitrimer films (10 mm × 5 mm × 0.3 mm) were weighted as W_s and enveloped using metal mesh (weighted W_{s+m}). The specimens were then immersed in THF, and the solvent was stirred and refluxed at 90 °C for 24 hours. After cooling to room temperature, the swollen specimens were removed from the solvent, wiped the THF on the surface, and then weighed (W_g). The specimens were dried at 100 °C under vacuum until the weight reached a constant (W_d). The swelling ratio and gel content can be given by:

$$\text{Swell ratio} = \left(1 + \frac{W_g - W_d}{W_d}\right) \times 100\%$$

$$\text{Gel content} = \left(1 - \frac{W_{s+m} - W_d}{W_s}\right) \times 100\%$$

Table S1 Swell ratios and gel contents of cured vitrimer films

| Code | Swelling ratio (%) | Gel content (%) |
|------------------|--------------------|-----------------|
| OHESO-0.7 | 159.5 ± 17.9 | 81.4 ± 3.8 |
| OHESO-1.0 | 154.9 ± 17.1 | 84.8 ± 2.3 |
| OHESO-1.5 | 149.4 ± 17.2 | 80.4 ± 1.7 |
| BuESO-0.7 | 165.8 ± 23.6 | 89.2 ± 1.9 |
| BuESO-1.0 | 175.5 ± 18.5 | 94.2 ± 7.0 |
| BuESO-1.5 | 146.5 ± 10.5 | 89.6 ± 1.1 |

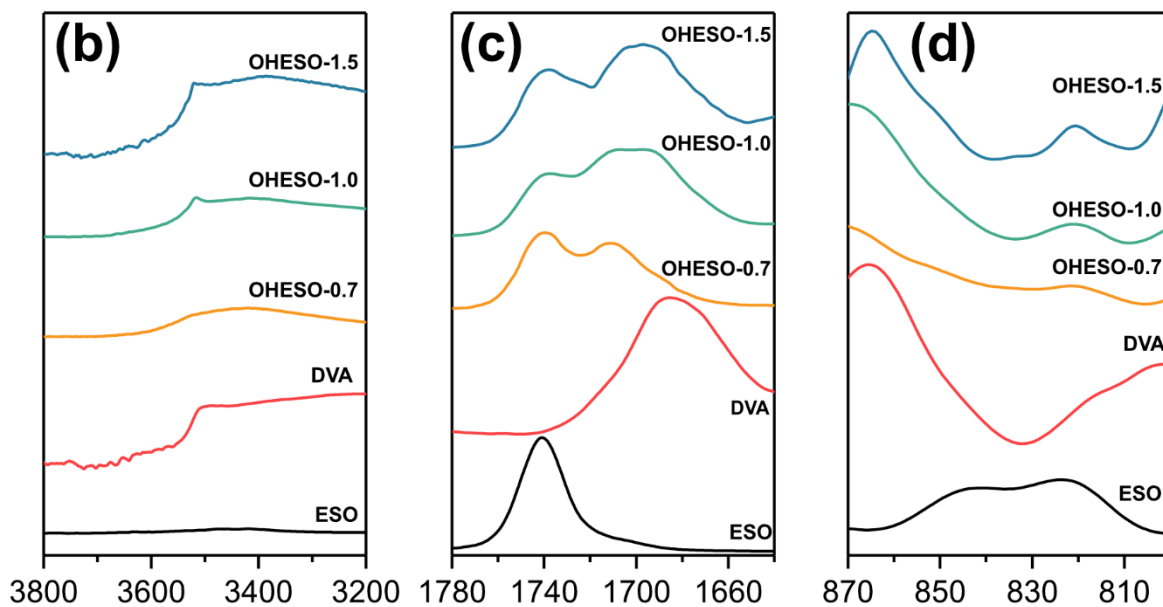
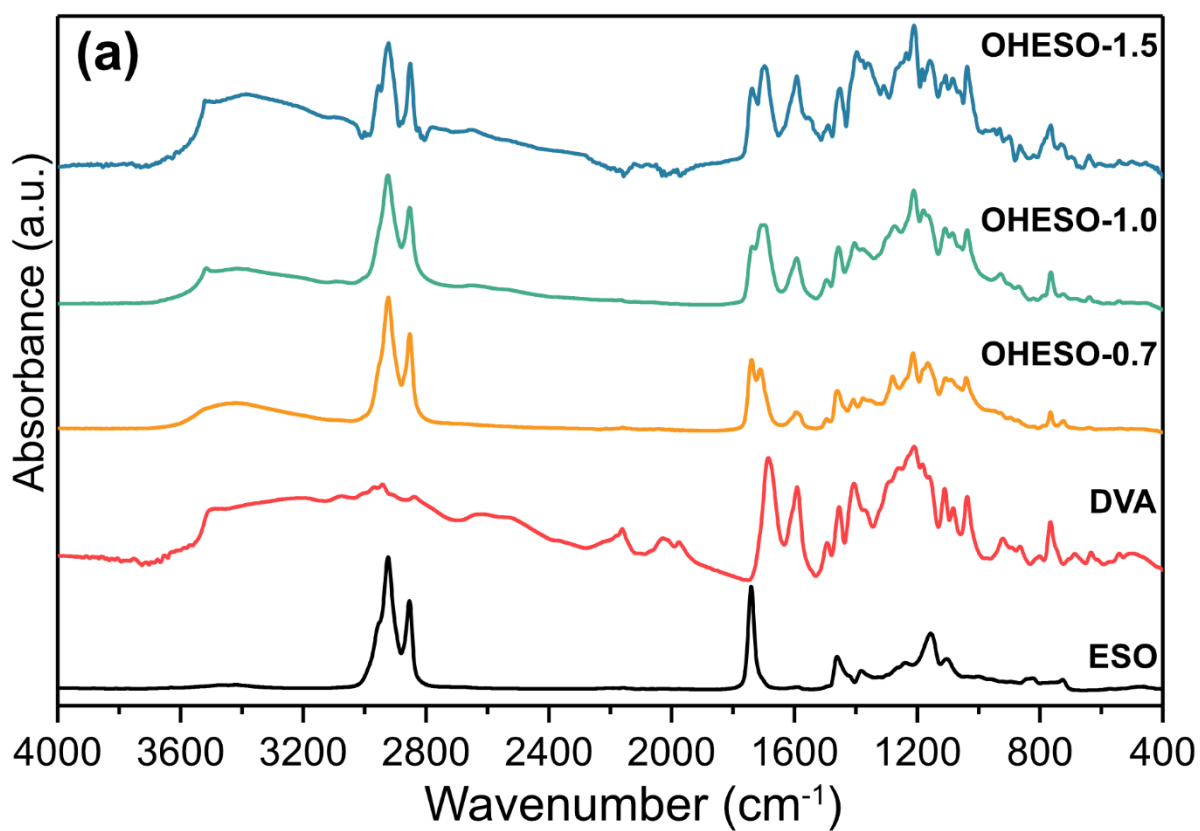


Fig. S2 FT-IR spectra of ESO, DVA and OHESO-*x* vitrimers.

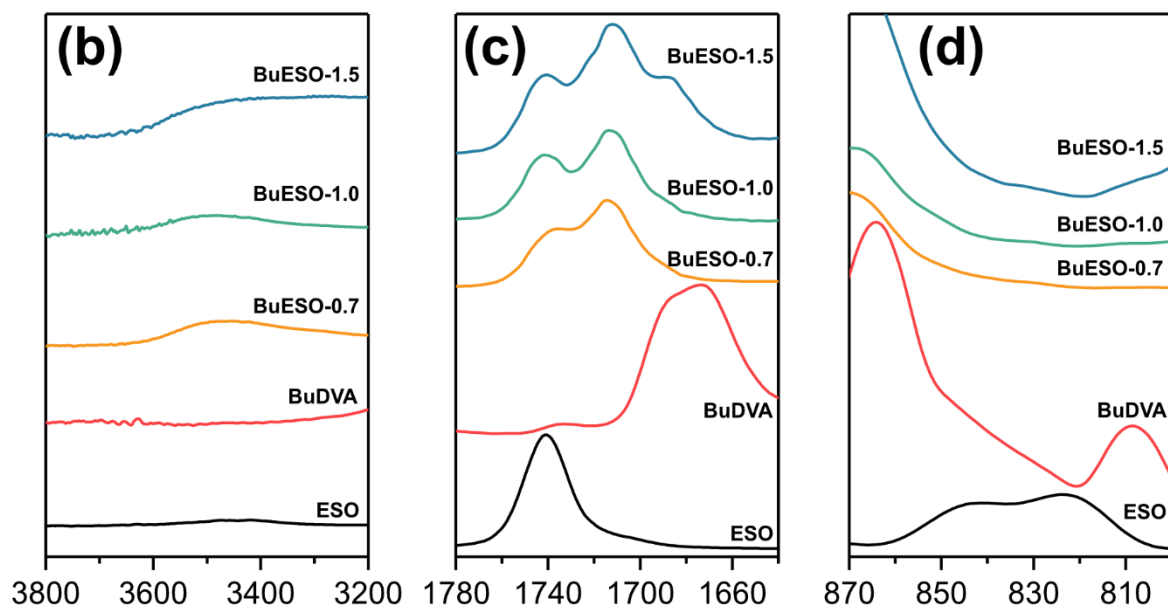
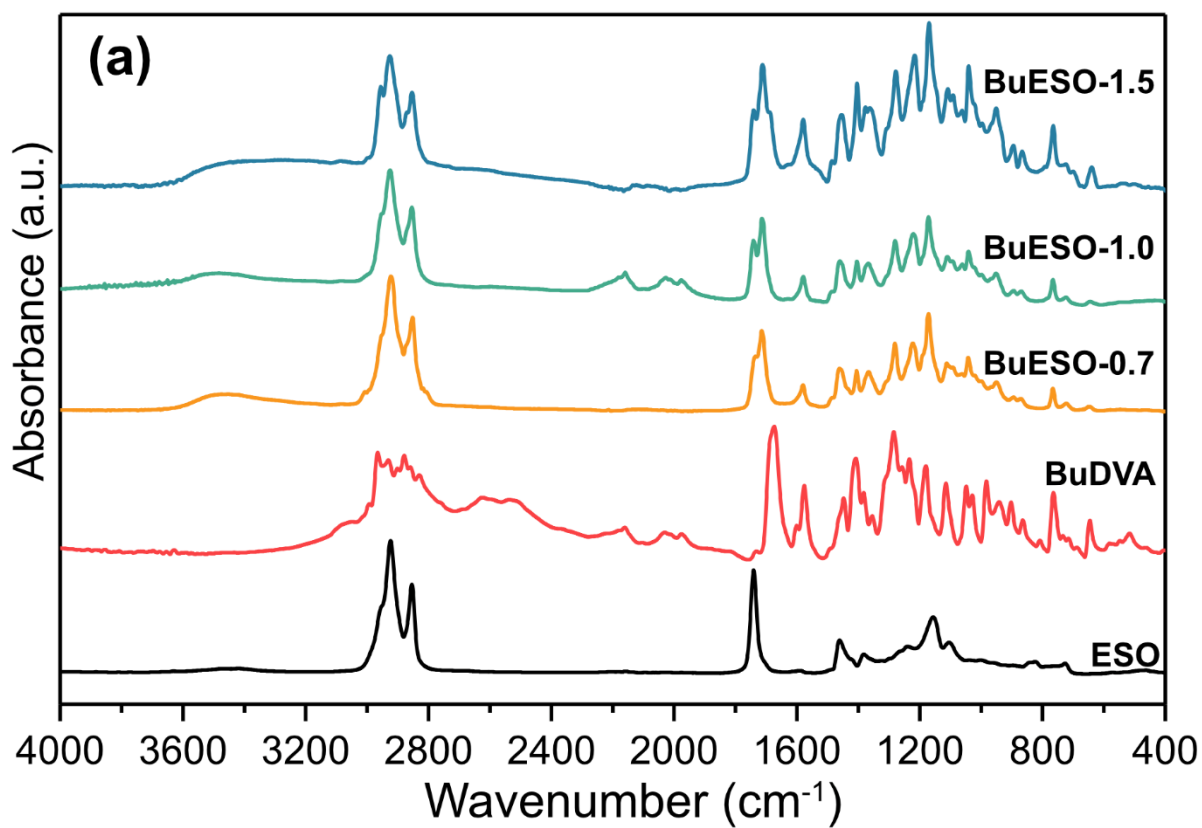


Fig. S3 FT-IR spectra of ESO, BuDVA and BuESO-*x* vitrimers.

S4 As-measured DMA curves

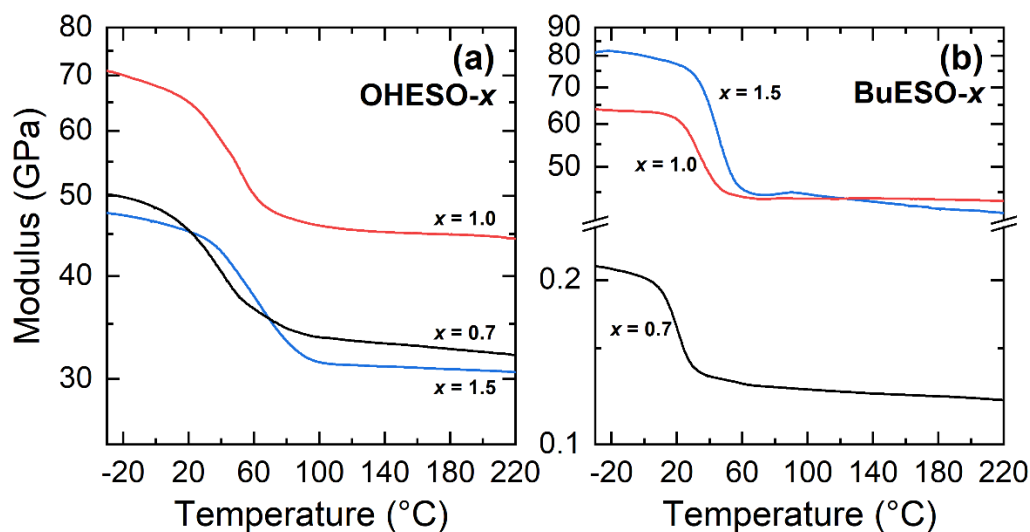


Fig. S4 DMA curves of as-measured storage moduli versus temperatures at a heating rate of 3 °C/min.

S5 Fitting function of vitrimer films and evaluation of activation energy

According to several previous works²⁻⁴, the stress relaxation curves of OHESO-x and BuESO-x at each temperature (except for OHESO-1.0 at $T = 190$ °C) were fitted by a stretched exponential function expressed as below,

$$G_t/G_0 = e^{\left(-\frac{t}{\tau_{\text{fit}}}\right)^\beta} + c$$

Table S2 Values of τ_{fit} and parameters of relaxation function

| Codes | T (°C) | τ_{fit} (10^3 s) | Parameters | |
|------------------|----------|---------------------------------|------------|-------|
| | | | β | c |
| OHESO-1.0 | 170 | 36.3 | 0.81 | -0.09 |
| | 180 | 25.3 | 0.91 | -0.11 |
| | 190 | Not well fitted ^a | | |
| | 200 | 5.6 | 0.80 | -0.32 |
| OHESO-0.7 | 200 | 21.9 | 0.26 | 0.18 |
| OHESO-1.5 | 200 | 40.1 | 0.53 | -0.14 |
| BuESO-1.0 | 170 | 221.9 | 0.75 | -0.04 |
| | 180 | 180.4 | 0.76 | -0.04 |
| | 190 | 76.4 | 0.73 | -0.04 |
| | 200 | 30.2 | 0.62 | -0.04 |
| BuESO-0.7 | 200 | 6.4 | 0.56 | 0.08 |
| BuESO-1.5 | 200 | 82.0 | 0.63 | -0.06 |

^a The relaxation curve cannot be well fitted by the relaxation function.

The fitting curves are shown as follow.

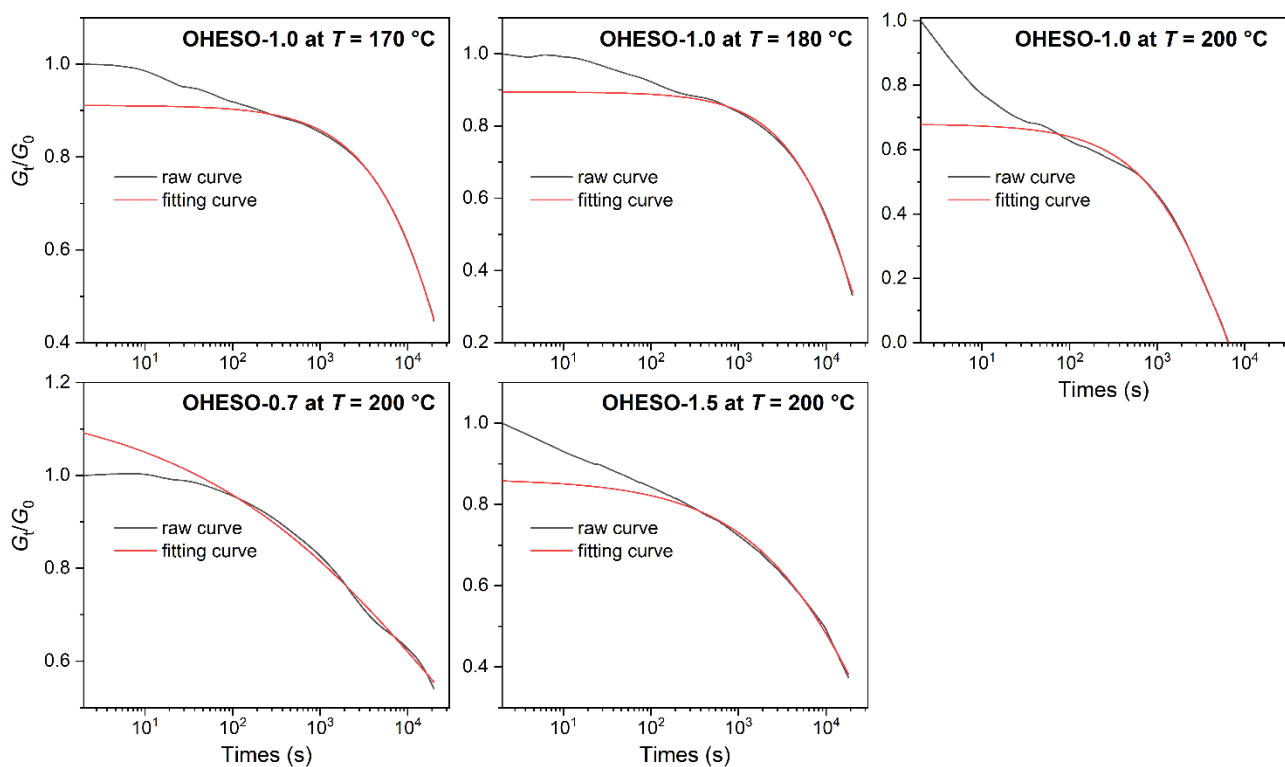


Fig. S5 Normalized stress relaxation curves of OHESO-x and their fitting curves.

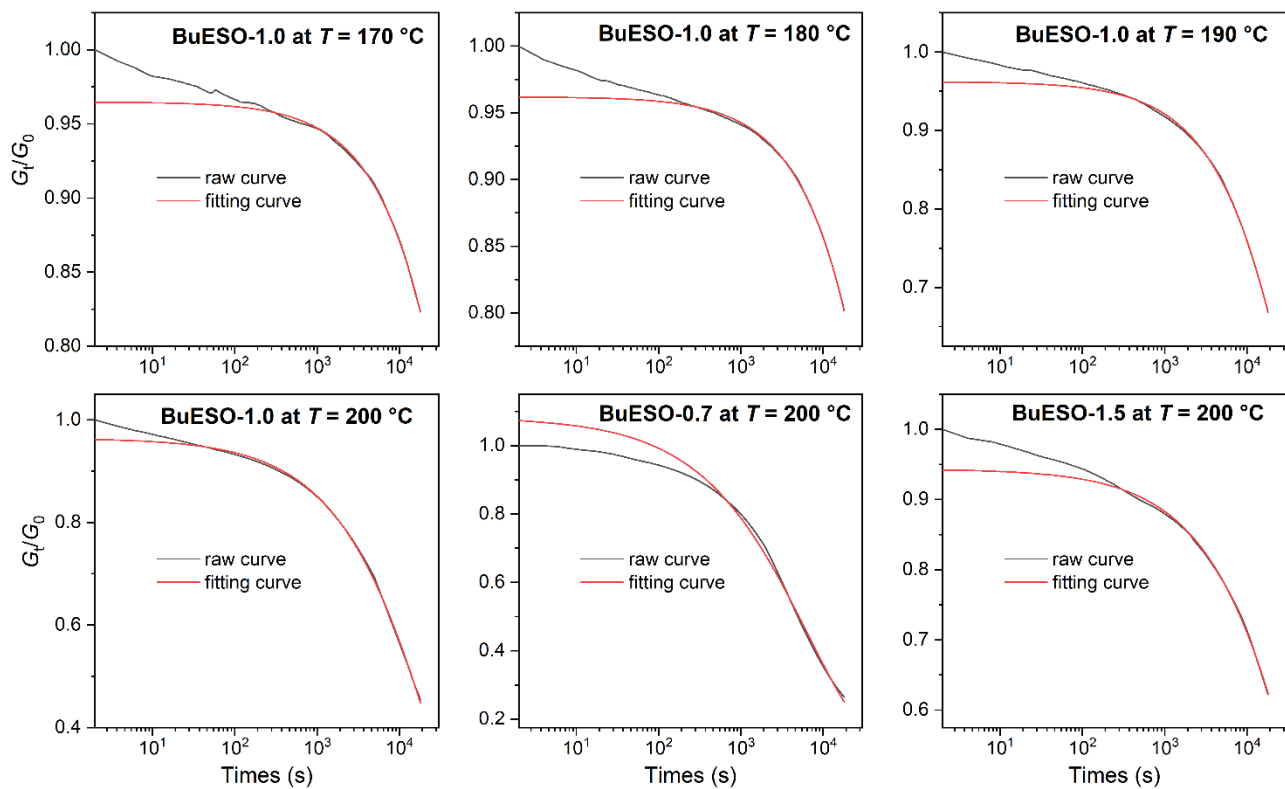


Fig. S6 Normalized stress relaxation curves of BuESO-x and their fitting curves.

For OHESO-1.0 and BuESO-1.0, the values of τ_{fit} at $T = 170\text{--}200\text{ }^{\circ}\text{C}$ are substituted into the following Arrhenius equation (The R represents the ideal gas constant) to obtain the E_a value from the slope of the fitting line.

$$\ln(\tau_{\text{fit}}) = A + \frac{E_a}{RT}$$

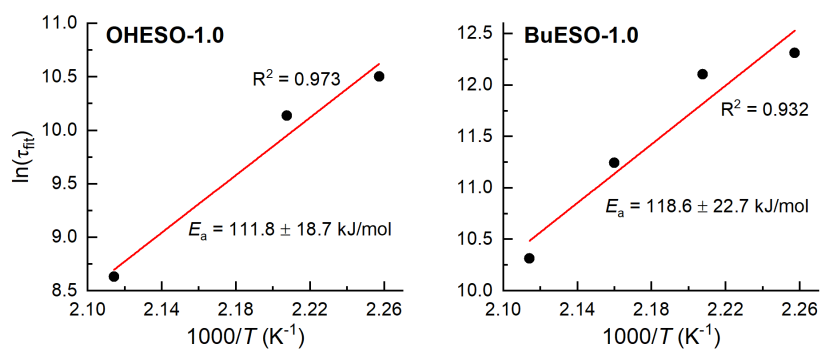


Fig. S7 Arrhenius plot of the relaxation time for OHESO-1.0 and BuESO-1.0.

S6 Reprocessed vitrimer films

Table S3 Mechanical properties of reprocessed vitrimer films

| Sample | Times of reprocessing | σ_{\max} (MPa) | ϵ_{\max} (%) | E (MPa) |
|------------------|-----------------------|-----------------------|-----------------------|--------------|
| OHESO-1.0 | 0 (pristine) | 13.6 ± 1.1 | 13.6 ± 2.0 | 286 ± 49 |
| | 1 | 10.1 ± 1.4 | 6.8 ± 3.1 | 228 ± 30 |
| | 2 | 7.7 ± 2.3 | 12.7 ± 10.7 | 168 ± 28 |
| | 3 | 8.8 ± 0.9 | 30.0 ± 17.1 | 199 ± 24 |
| | 4 | 6.9 ± 1.0 | 19.2 ± 7.9 | 163 ± 18 |
| | 5 | 8.6 ± 0.5 | 31.5 ± 19.5 | 174 ± 23 |
| BuESO-1.0 | 0 (pristine) | 4.8 ± 0.6 | 69 ± 12 | 57 ± 11 |
| | 1 | 2.0 ± 0.2 | 63 ± 10 | 22 ± 8 |
| | 2 | 2.5 ± 0.2 | 71 ± 17 | 29 ± 14 |
| | 3 | 2.1 ± 0.7 | 54 ± 16 | 38 ± 11 |
| | 4 | 2.6 ± 0.6 | 77 ± 19 | 30 ± 8 |
| | 5 | 2.7 ± 0.5 | 50 ± 17 | 32 ± 11 |

IR spectra of reprocessed vitrimer films

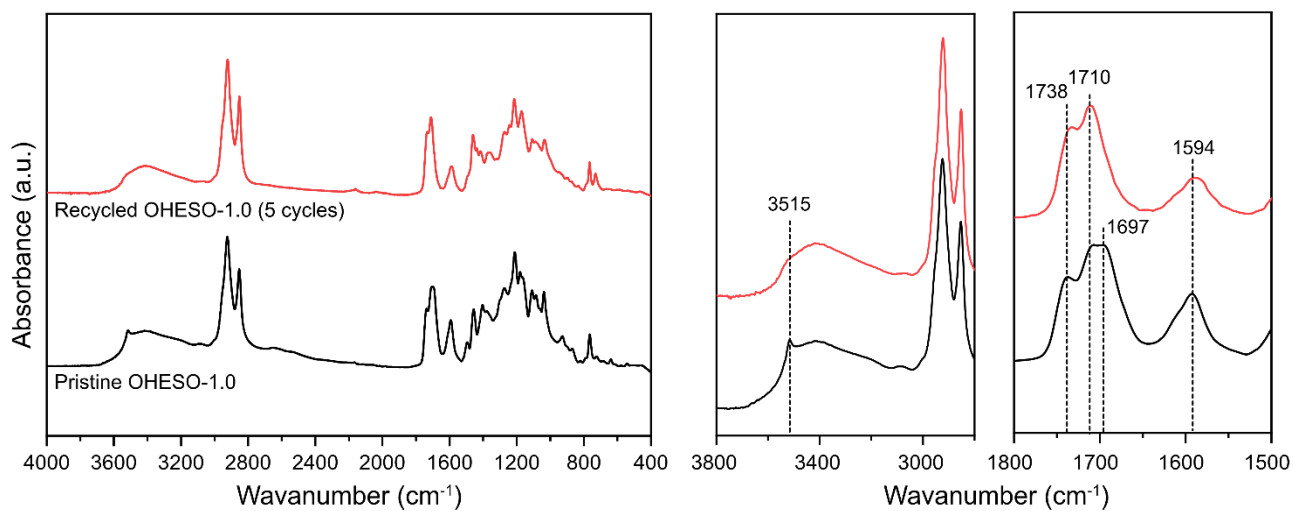


Fig. S8 FT-IR spectra of pristine and recycled (5 cycles) OHESO-1.0.

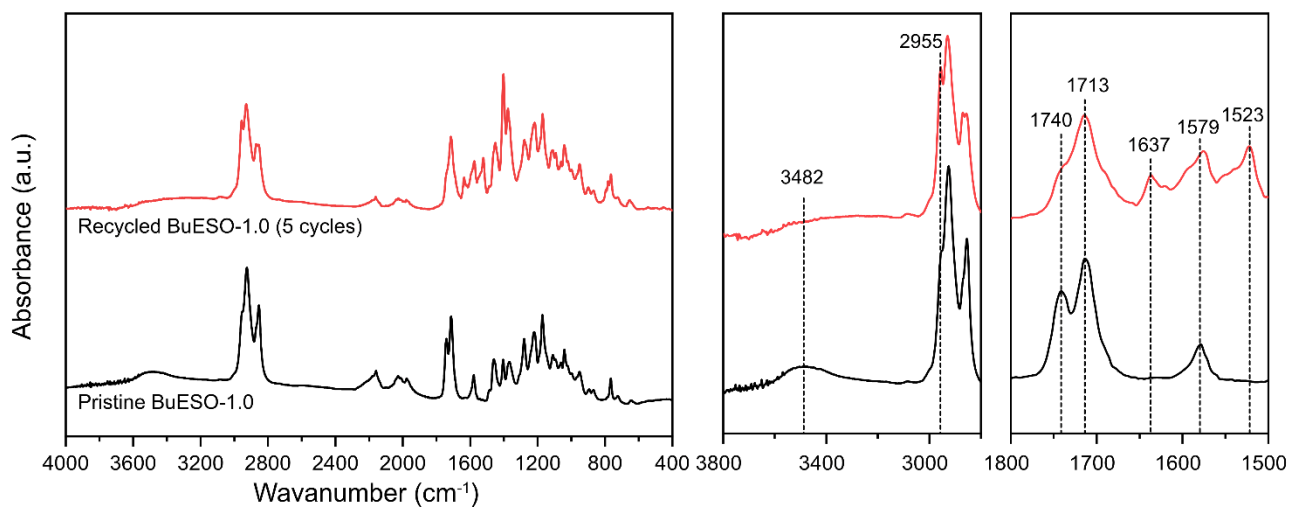


Fig. S9 FT-IR spectra of pristine and recycled (5 cycles) BuESO-1.0.

S7 Shape memory property of OHESO-1.0

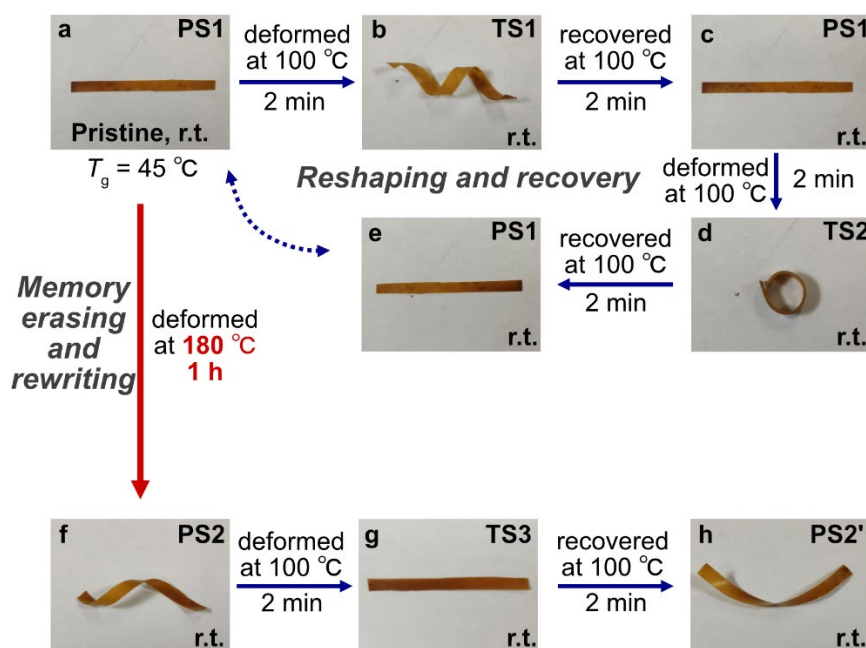


Fig. S10 Shape memory photographs of OHESO-1.0. PS represents the permanent shapes (i.e., shapes in a stable state), and TS represents the temporary shapes (i.e., shapes in a metastable state). All images were taken at room temperature.

The shape memory behavior of OHESO-1.0 was studied as an example in a series of heating-and-reheating processes shown in Fig. S10. A strip of OHESO-1.0 [PS1, Fig. S10 (a)] was firstly cut from a vitrimer film. At this initial state, the molecular chains were in the conformation with high entropy (i.e., thermally stable).⁵ Then, the specimen was heated at $T = 100\text{ °C}$ (higher than its T_g), and a force was applied to fix it in a helix shape

for 2 min. After cooling the reshaped specimens to room temperature and removing the force, a helical specimen was obtained (PS1, Fig. S10 (b)). At this state, the vitrimer was in a metastable stage because the molecular chains were frozen at $T < T_g$ with relatively low entropy. After reheating the helical specimen to $T = 100\text{ }^{\circ}\text{C}$, the specimen recovered almost to its original shape (PS1) in 2 min. During the reheating (TS1 to PS1), the frozen molecular chains became movable, and the entropy increased. The following PS1→TS2→PS1 [Fig. S10 (c)–(e)] was a similar reshaping and recovery process to PS1→TS1→PS1. Furthermore, we re-edited the permanent shape of the specimen by heating the specimen at $180\text{ }^{\circ}\text{C}$ for one hour [PS1→PS2, Fig. S10 (a) to (f)]. At $T = 180\text{ }^{\circ}\text{C}$, the transesterification was activated, and the networks of vitrimer were rearranged to form a new thermally stable permanent topology (PS2). As shown in [Fig. S10 (f) to (g)], although a complete shape recovery was not achieved, the specimen with re-edited permanent shape (PS2) also exhibited satisfactory shape memory behavior (PS2→TS3→PS2').

Reference

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