Supporting Information

Supramolecular Catalytic Nanomedicines Based on Coordination Self-Assembly of Amino Acid for Cascade-Activated and -Amplified Synergetic Cancer Therapy Enhui Song, Qiong Wu, Ren Gao, Xiaopeng Lan, Yanhui Zhang, Hao Geng, Chunlei Liu^{*}, Feijie Xu^{*}, Yongxin Li^{*} and Chunzhao Liu^{*}

[a] E. Song, R. Gao, X. Lan, Y. Zhang, H. Geng, C. Liu, Y. Li and C. Liu

State Key Laboratory of Bio-fibers and Eco-textiles, Institute of Biochemical Engineering, The Affiliated Qingdao Central Hospital of Qingdao University, College of Materials Science and Engineering, Qingdao University, Qingdao 266071, China.

E-mail: 18660229131@163.com, liyx@qdu.edu.cn, czliu@qdu.edu.cn

[b] Q. Wu

Department of Laboratory, Qingdao Hospital of Traditional Chinese Medicine (Qingdao Hiser hospital), Qingdao 266033, China.

[c] F. Xu

Department of Biomedical Sciences, City University of Hong Kong, Tat Chee Avenue, Kowloon, Hong Kong (China).

Email: feijiexu2-c@my.cityu.edu.hk



Figure S1. The SEM image of Fmoc-D/Cu.



Figure S2. The SEM image of Fmoc-D/Cu/G.



Figure S3. The UV-Vis absorption spectra of Fmoc-D/Cu and Fmoc-D/Cu/FITC-G.



Figure S4. The digital image of Fmoc-D/Cu nanoparticles before and after adding the GSH.



Figure S5. (a) Digital picture of the light-yellow product after lyophilization of the supernatant from the reaction between Fmoc-D/Cu nanoparticles and GSH. (b) FT-IR spectra of commercial GSSG and the light-yellow product. (c) HR-MS of the light-yellow product.



Figure S6. The GSH/GSSG ratio change with time after adding the Fmoc-D/Cu to GSH.



Figure S7. The degradation process of MB after addition of Fmoc-D/Cu nanoparticles at different time points.



Figure S8. Reaction of TPA with the generated ·OH-induced enhancement of fluorescence.



Figure S9. The H₂O₂ generation of Fmoc-D/Cu/G nanoparticles in glucose solution.



Figure S10. Reaction of TPA with the generated ·OH-induced enhancement of fluorescence.



Figure S11. ¹HNMR of CPT-TK in Chloroform-d.



Figure S12. HR-MS spectrum of CPT-TK-DOX.



Figure S13. HPLC analysis of CPT-TK-DOX, the purity is over 99%.



Figure S14. The SEM image of CPT-TK-DOX.



Figure S15. The fluorescence spectra of Fmoc-D/Cu/G@C and CPT-TK-DOX monomer.



Figure S16. The cleavage of CPT-TK-DOX in Fmoc-D/Cu/G@C nanomedicine after incubated with GSH and H_2O_2 (a), only GSH (b) and only H_2O_2 (c) for 4 h.



Figure S17. Size and polymer dispersity index (PDI) of (a) Fmoc-D/Cu nanoparticles, (b) Fmoc-D/Cu/G nanoparticles and (c) Fmoc-D/Cu/G@C nanoparticles in culture medium supplemented with 10% (v/v) FBS at 37 °C for 24 h.



Figure S18. (a) Quantification of fluorescence intensities of H_2O_2 . (b) Quantification of fluorescence intensities of generated $\cdot OH.s$



Figure S19. In vivo fluorescent imaging of the tumor-bearing mice after intravenous injection of FL-labeled Fmoc-D/Cu/G@C nanoparticles and free FL.



Figure S20. H&E-stained images of major tissues collected from tumor-bearing mice after different treatments. Scale bar, $100 \ \mu m$.