

Supporting Information

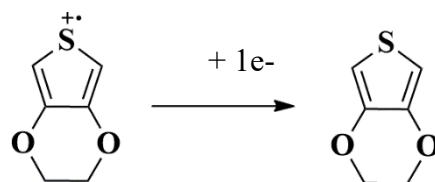
In-situ engineering of Au-Ag alloy embedded PEDOT nanohybrids at a solvent/non-solvent interface for the electrochemical enzyme-free detection of histamine

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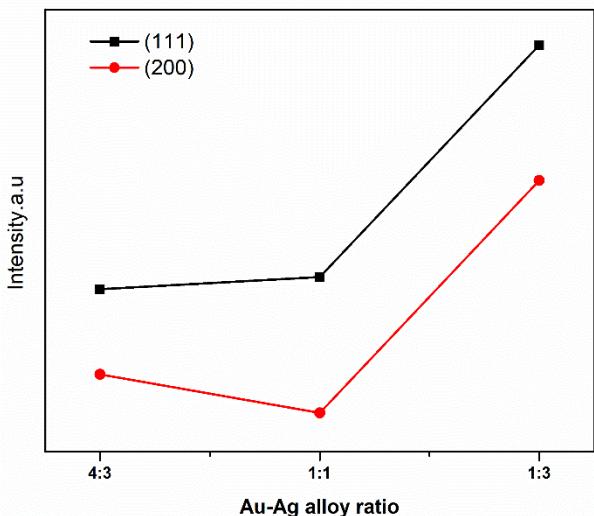
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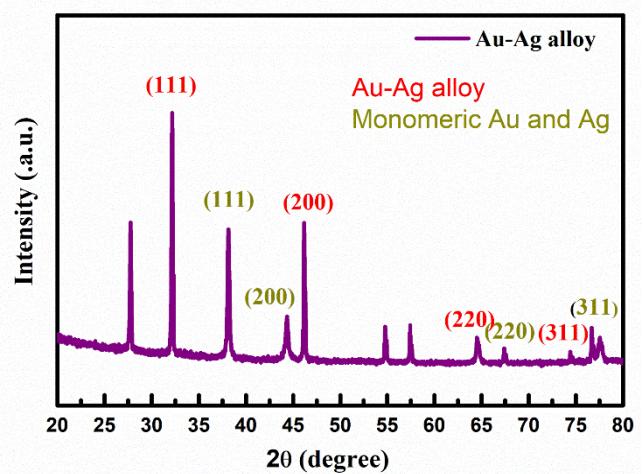
EDOT radical cation **EDOT** $E^o_{P\equiv} - 3.95 \text{ V}^\ddagger$

Sl. No.	Metal ion conversion	Standard Reduction Potential of Metal vs. NHE E°_M (V)	No of electrons involved in the reaction, (n)	E°_{cell} (V) = $E^\circ_M - E^\circ_P$	Standard Gibbs free energy, ΔG° (kcalmol ⁻¹)
1.	$\text{Ag}^+ + 1\text{e}^- \rightarrow \text{Ag}^0$	0.80 ²	1	4.75	-109.5
2.	$\text{Au}^{3+} + 3\text{e}^- \rightarrow \text{Au}^0$	0.99 ³	3	4.94	-341.6

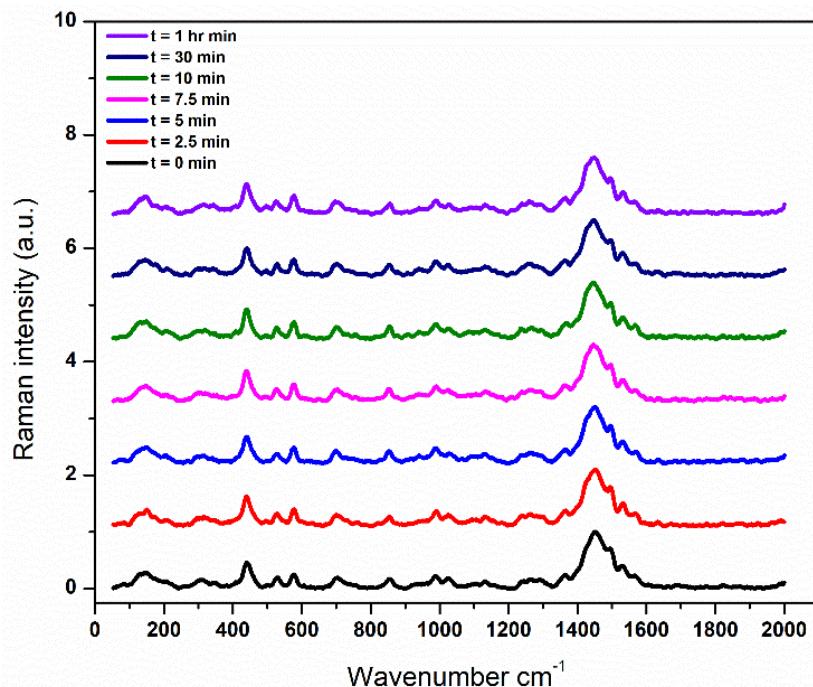
Supporting Table S1. Table showing the half-cell reaction of metal ions at the cathode, standard reduction potentials of monomer (E_P°) and metal ions (E_M°), number of electrons (n) involved in the redox reaction and the electrochemical cell potential (E_{cell}°). The thermodynamic free energy, $\Delta G^\circ = -nFE_{cell}^\circ$ was calculated to predict the thermodynamic feasibility of polymerization reactions.



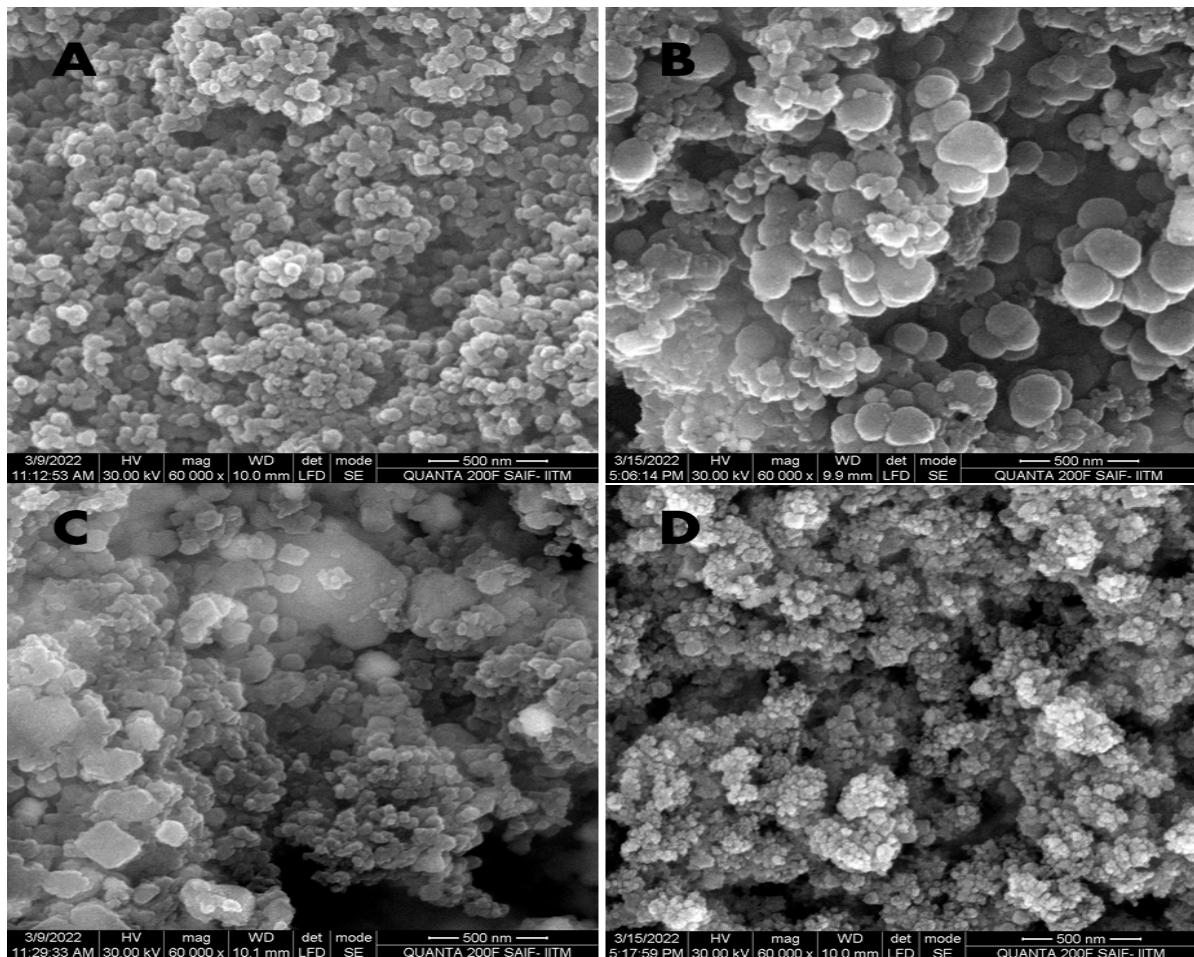
Supporting Fig. S1. Comparison of the variation in diffraction plane intensity of PEDOT/Au-Ag alloy with change in $\text{Au}^{3+}/\text{Ag}^+$ ratios.



Supporting Fig. S2. XRD spectra of Au-Ag alloy formed by the simple mixing of HAuCl_4 and AgNO_3 solutions in 1: 3 ratio.

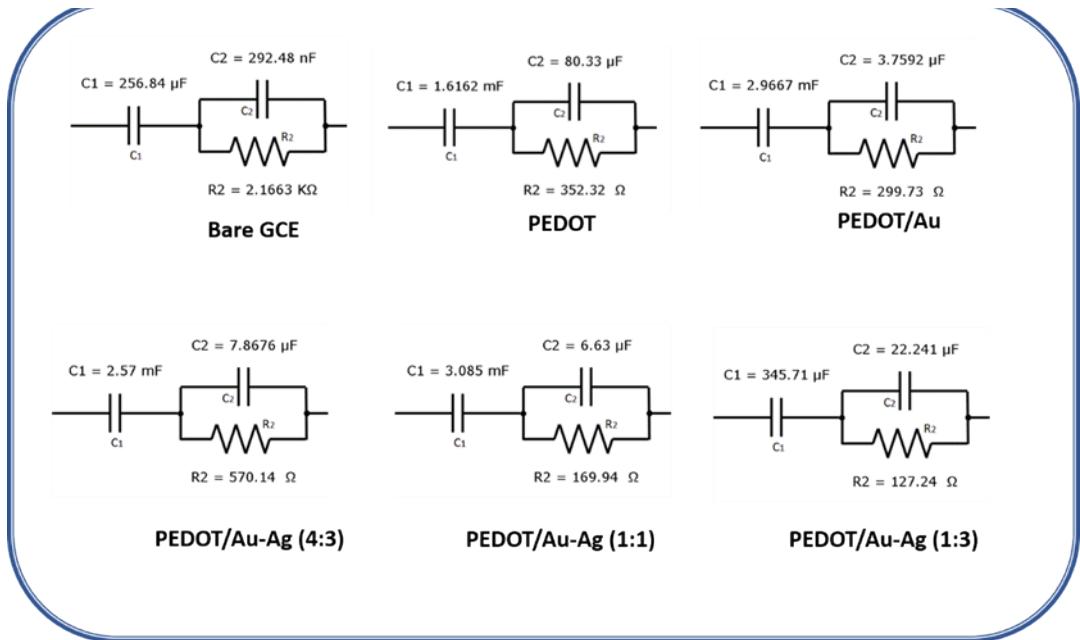


Supporting Figure S3. Raman spectra of PEDOT/Au-Ag alloy synthesized at various time intervals from $t = 0$ to $t = 1$ hr

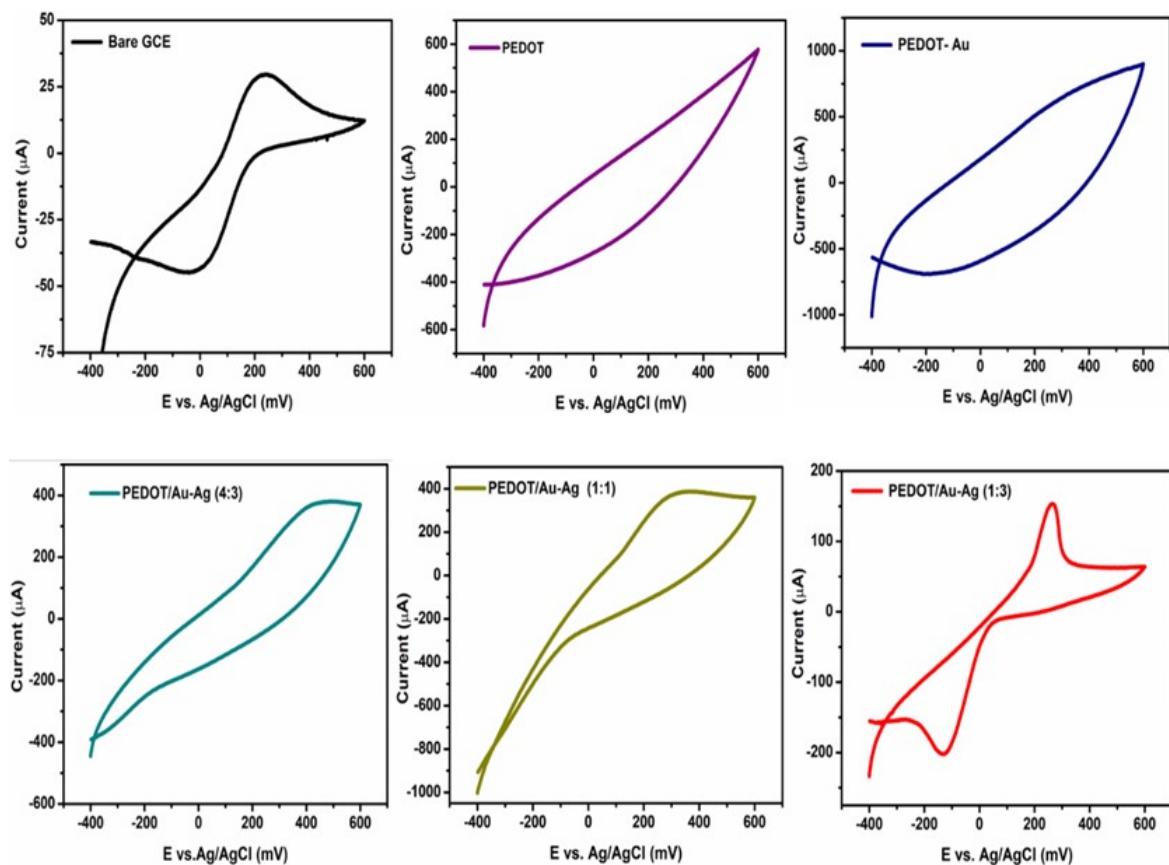


Supporting Fig. S4. SEM Images of PEDOT:Au-Ag alloy samples made with different Au/Ag mole ratios. (A) SEM image of PEDOT:Au without Ag, (B) SEM image of PEDOT alloy with Au/Ag mole ratio of 1:1, (C) Au/Ag mole ratio 4:3 and (D) Au/Ag mole ratio 1:3

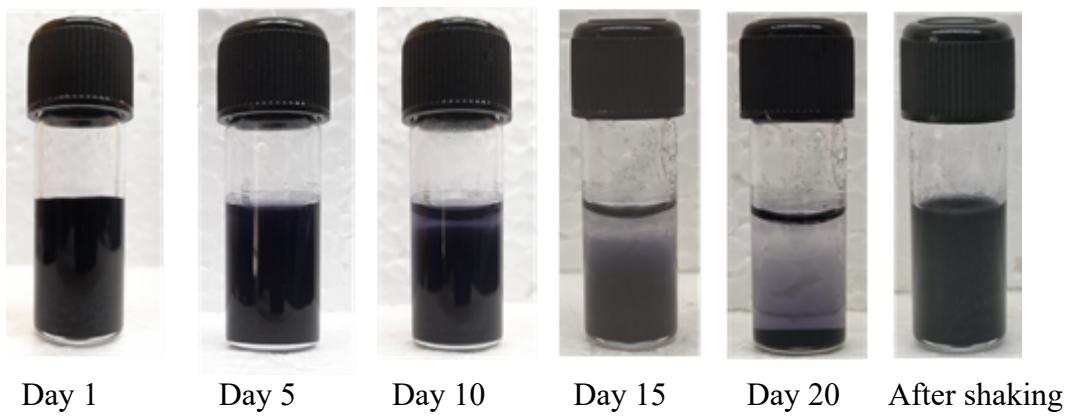
It can be noted that the optimized product PEDOT/Au- Ag (1:3) has smaller uniformly sized particles compared to the FE-SEM images of other compositions. It is reported that the $\text{Au}^{3+}/\text{Ag}^+$ molar ratio affects the average size of Au–Ag alloy NPs formed.⁴



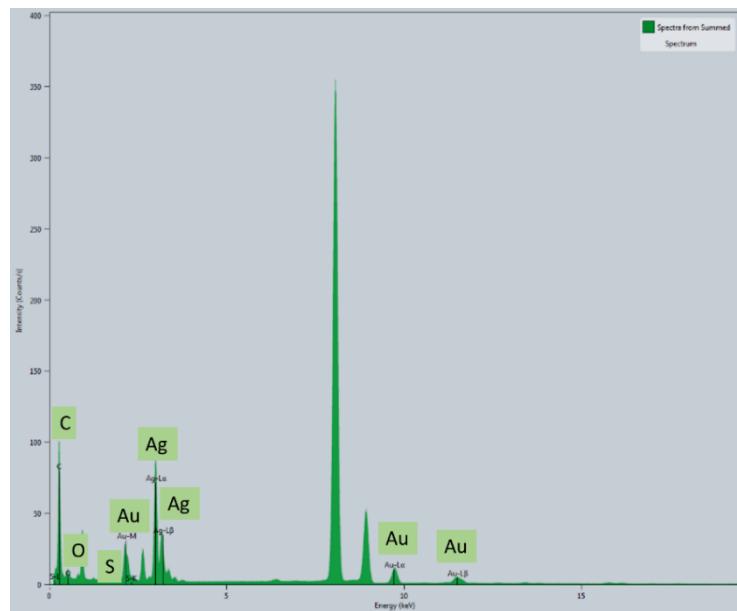
Supporting Fig. S5. The equivalent circuit diagrams corresponding to the Nyquist plots shown in Fig. 2d.



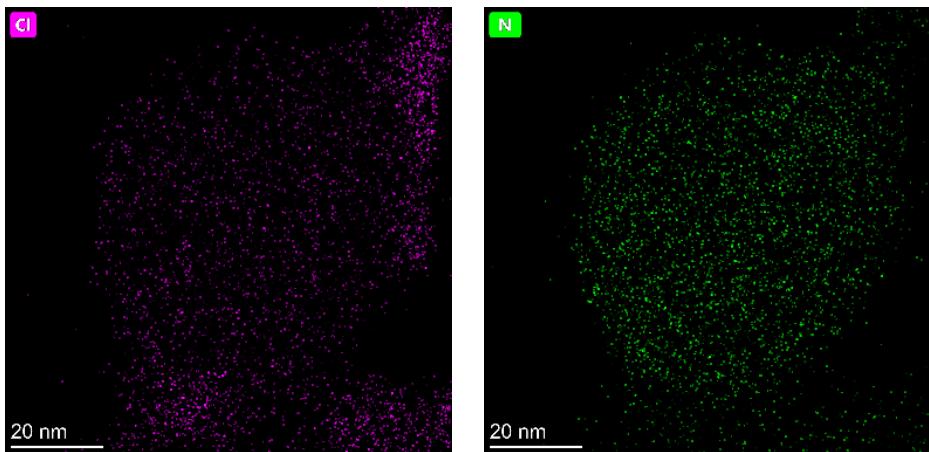
Supporting Fig. S6. Comparison of the electrochemical activity of Bare GCE, PEDOT, PEDOT- Au, and PEDOT/Au-Ag alloys with Au^{3+}/Ag^+ mole ratio (4:3), (1:1) and (1:3) in 5mM $K_4[Fe(CN)_6]$ in 0.1MKCl.



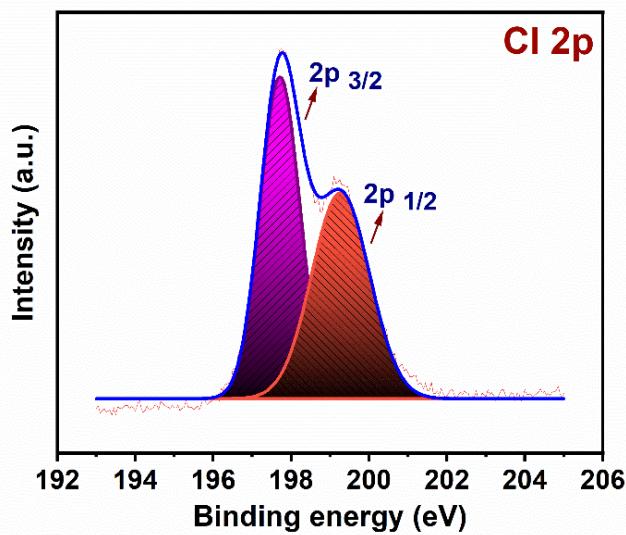
Supporting Fig. S7. Dispersion stability of PEDOT/Au-Ag (1:3) in water



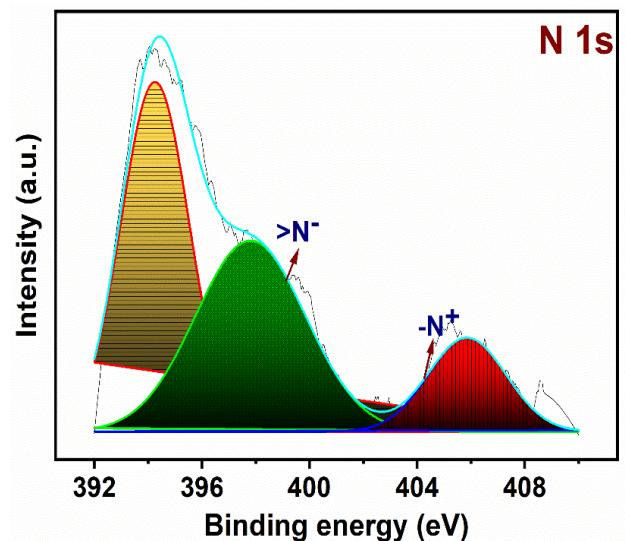
Supporting Fig. S8. HRTEM-EDS spectrum of PEDOT/Au-Ag alloy (1:3)



Supporting Fig. S9. Elemental mapping images of Chlorine and Nitrogen present in PEDOT/Au-Ag (1:3)



Supporting Fig. S10. XPS spectra of Chlorine



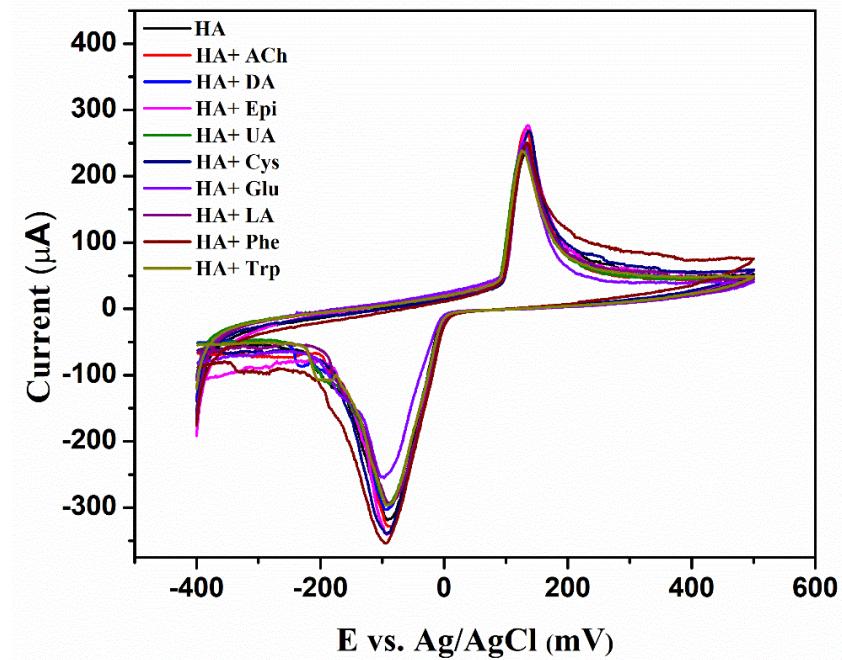
Supporting Fig. S11. XPS spectra of Nitrogen

Sl. No.	Type of material	Detection technique	Linear range	Oxidation peak potential (V)	LOD	Ref.
1	Au-NPs	Fluorescence	0.01–1.0 μM	Not applicable	2.04 nM	S5
2	Ag-Ag ₂ O/MWCNTs/GCE	DPV	5-200 mM	0.97	0.018 μM	S6
3	Au NPs	Colorimetry (LSPR)	0.4-2.63 mM	Not applicable	38 nM	S7
4	rGO-ppy/GCE	DPV	10-800 μM	0.67	3.01 μM	S8
5	TGA-CdTe QDs	Fluorescence	9.6-570 μM	Not applicable	9.6 μM	S9
6	Cu@Pd core shell NPs	Amperometry	0.1-10 μM	0.55	3.2 nM	S10
7	Nafion-MWCNTs/GCE	DPV	20-200 μM	1.12	0.39 μM	S11
8	MIP/L-cysteine/AuNPs /GCE	DPV	1-107 μM	1.2	0.6 μM	S12
9	MIP-apt/AuNPs /CNT/GCE	EIS	0.35-35 nM	0.3	0.11 nM	S13
10	MWCNTs/CPE	DPV	0.1-100 μM	1.12	1. 26 μM	S14
11	BDD electrode	CV	0.5–100 μM	1.4	0.5 μM	S15
12	Lingnin/GCE	CV	5-200 μM	1.35	0.38 μM	S16
13	SWCNT/CPE	DPV	4.5- 720 μM	1.25	1.26 μM	S17
14	FrGO/GCE	Amperometry	0.2-80 μM	0.74	0.007 μM	S18
15.	PEDOT/Au-Ag (1:3) alloy Present work	CV	100 nM - 1 μM	0.13	1.25 nM	

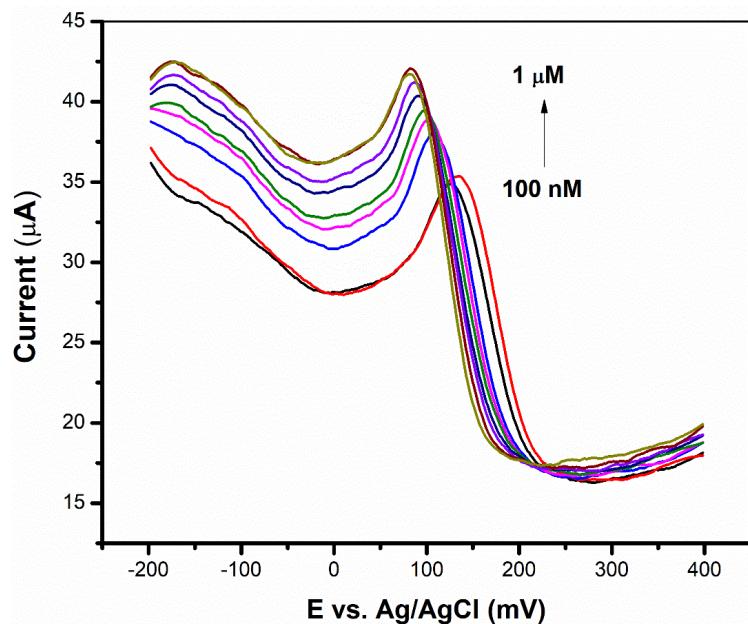
Supporting Table S2. Performance comparison of PEDOT/Au-Ag (1:3) sensor for HA detection with other previously reported sensors.

Sl. No.	Materials	Synthesis method	Analyte	Detection technique	LOD	Ref.
1.	PEDOT/NiNPs/GO	Electro-polymerization	Glucose	Amperometry	0.8 μM	S19
2.	PEDOT/PEDOT-SH/Au	Electro-polymerization	Nitrite	Amperometry	0.051 μM	S20
3.	PEDOT/PdNPs	Chemical Oxidation	Peroxide	Amperometry	2.84 μM	S21
4.	PtNPt/MWCNT-PEDOT:PSS	Electrochemical method	DA	CV	50 nM	S22
5.	PEDOT:PSS-RGO/AuNPs	Chemical oxidation	Peroxide	Amperometry	0.08 μM	S23
6.	PEDOT-PPY/Ag	Chemical oxidation	DNA	EIS	5.4 fM	S24
7.	PEDOT/GO/CuNPs	Electrochemical method	Glucose	Amperometry	47 nM	S25
8.	PEDOT/CNTs/ Cu-Co	Electrochemical method	Nitrite	Amperometry	60 nM	S26
9.	PEDOT/Ag NPs	Electrochemical method	Caffeic acid	CV	1.9 μM	S27
10.	PEDOT /AuNPs /CNT	Chemical oxidation	AA and UA	CV, DPV	0.283 μM	S28
11.	PEDOT/Au-Ag alloy Present work	Liquid-liquid interface-assisted polymerization	HA	CV	1.5 nM	

Supporting Table S3. The synthesis method and performance comparison of PEDOT-based composites for the non-enzymatic detection of various analytes.



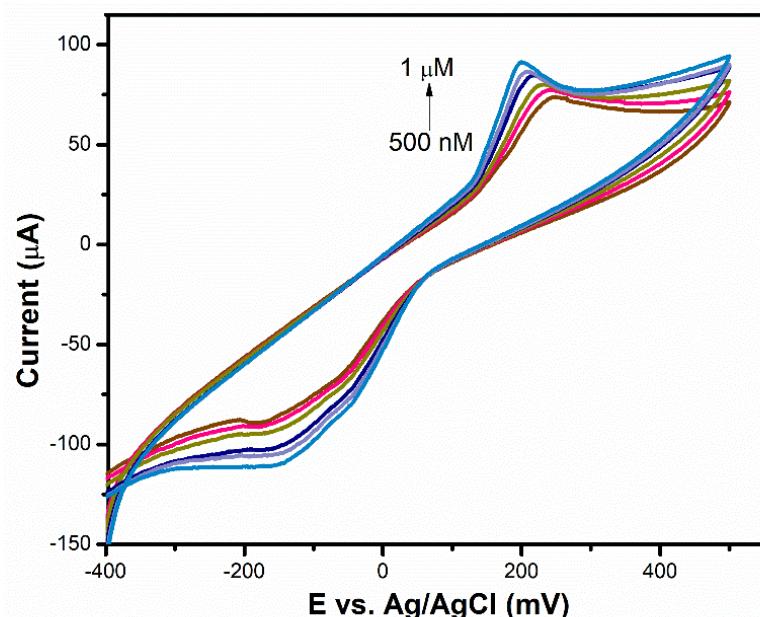
Supporting Fig. S12. Cyclic Voltammograms of PEDOT/Au-Ag (1:3) in $100\mu\text{M}$ HA in the presence of various interfering groups at a scan rate of 50mV/s



Supporting Fig. S13. DPV plots of PEDOT/Au-Ag (1:3) modified GCE towards varying concentrations of HA

Sl. No.	Added Concentration (nM)	Output current in std soln (μ A)	Output current in fish extract solution (μ A)	Recovery Concentration (nM)	% Recovery	RSD (%)
1	0	—	35	106	—	—
2	500	39.5	36	378	75.6	1.89
3	600	40.36	38.7	451	75.1	1.65
4	700	41.2	41.47	723	103.28	2.65
5	800	41.7	43.37	899	112.37	1.74
6	900	42	44.82	992	110.2	2.14
7	1000	43.5	45.97	1110	111.0	2.81

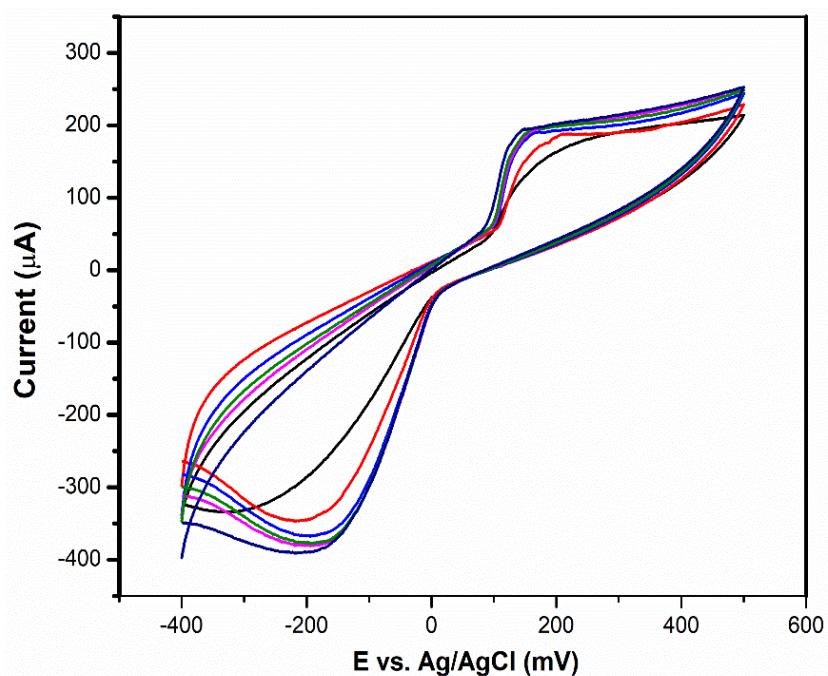
Supporting Table S4. Recovery results of HA in Canned Tuna Fish.



Supporting Fig. S14. CV plots of PEDOT/Au-Ag (1:3) modified GCE at different concentrations of HA in artificial sweat sample

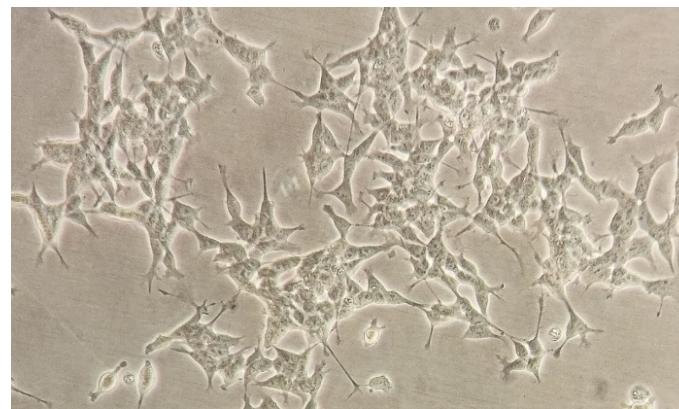
Sl No.	Added Concentration (nM/ μ M)	Output current in std soln (μ A)	Output current in artificial sweat solution (μ A)	Recovery Concentration (nM)	% Recovery	RSD (%)
1	50	175	115.3	55	110	2.9
1	500	111	401	80.2	2.89	
2	60	187	180.7	72	120	2.7
2	600	113	117.76	496	82.6	2.5
3	70	191	187	77	110	2.9
3	700	118	122.08	627	89.5	3.1
4	80	192	192	79	98.7	1.95
4	800	121	125.77	727	90.8	3.5
5	90	195	197.2	84	93.33	3.85
5	900	123	129.47	794	88.5	2.47
6	100	197	200.7	86	86	2.91
6	1000	129	131.52	1004	100.4	1.81

Supporting Table S5. Recovery results of HA in artificial sweat sample.



Supporting Fig. S15. CV plots of PEDOT/Au-Ag (1:3) modified GCE at different concentrations of HA in human serum.

Supporting Table S6. Recovery results of HA in human serum sample.



Supporting Fig. S16. Microscopic image of SH-SY5Y neural cells cultured in DMEM medium after two hrs of the addition of L-Histidine precursor.



References

- S1 S. Lee and H. Lee, *J. Electrochem. Sci. Technol.*, 2012, **3**, 85–89.
- S2 P. Arul, S. T. Huang, V. Mani and C. H. Huang, *ACS Appl. Nano Mater.*, 2022, **5**, 6340–6351.
- S3 N. S. K. Gowthaman, B. Sinduja and S. A. John, *RSC Adv.*, 2016, **6**, 63433–63444.
- S4 L. Sun, Y. Yin, P. Lv, W. Su and L. Zhang, *RSC Adv.*, 2018, **8**, 3964–3973.
- S5 J. Bi, C. Tian, G. L. Zhang, H. Hao and H. M. Hou, *Foods*.

Supporting Fig. S17. Image showing the electrochemical detection set up for the detection of HA released by human neural cells using DPV technique.

- S6 N. Butwong, J. Khajonklin, A. Thongbor and J. H. T. Luong, *Microchim. Acta*, ,
- S7 C. Huang, S. Wang, W. Zhao, C. Zong, A. Liang, Q. Zhang and X. Liu, *Microchim. Acta*, 2017, **184**, 2249–2254.
- S8 B. Hu, N. Zhang, H. Li and C. Sun, *Int. J. Electrochem. Sci.*, 2021, **16**, 1–13.
- S9 S. Khan, L. S. A. Carneiro, M. S. Vianna, E. C. Romani and R. Q. Aucelio, *J. Lumin.*, 2017, **182**, 71–78.
- S10 Z. Wu, E. Xu, A. Jiao, Z. Jin and J. Irudayaraj, *RSC Adv.*, 2017, **7**, 44933–44944.
- S11 R. K. R. Gajjala and S. K. Palathedath, *Biosens. Bioelectron.*, 2018, **102**, 242–246.
- S12 S. Li, T. Zhong, Q. Long, C. Huang, L. Chen, D. Lu, X. Li, Z. Zhang, G. Shen and X. Hou, *Microchem. J.*, 2021, **171**, 106844.
- S13 A. M. Mahmoud, S. A. Alkahtani, B. A. Alyami and M. M. El-Wekil, *Anal. Chim. Acta*, 2020, **1133**, 58–65.
- S14 A. Geto, M. Tessema and S. Admassie, *Synth. Met.*, 2014, **191**, 135–140.
- S15 B. V Sarada, T. N. Rao, D. A. Tryk and A. Fujishima, *Anal. Chem.*, 2000, **72**, 1632–1638.
- S16 H. Degefui, M. Amare, M. Tessema and S. Admassie, *Electrochim. Acta*, 2014, **121**, 307–314.
- S17 Z. S. Stojanović, E. Mehmeti, K. Kalcher, V. Guzsvány and D. M. Stanković, *Food Anal. Methods*, 2016, **9**, 2701–2710.
- S18 F. Shahzad, S. A. Zaidi and C. M. Koo, *ACS Appl. Mater. Interfaces*, 2017, **9**, 24179–24189.
- S19 N. Hui, S. Wang, H. Xie, S. Xu, S. Niu and X. Luo, *Sensors Actuators B. Chem.*, 2015, **221**, 606–613.
- S20 Y. Ge, R. Jamal, R. Zhang, W. Zhang, Z. Yu, Y. Yan and Y. Liu, 2020, 1–10.
- S21 F. Jiang, R. Yue, Y. Du, J. Xu and P. Yang, *Biosens. Bioelectron.*, 2013, **44**, 127–131.
- S22 Z. Lu, S. Xu, H. Wang, E. He, J. Liu, Y. Dai, J. Xie, Y. Song, Y. Wang, Y. Wang, L. Qu and X. Cai, *ACS Appl. Bio Mater.*, 2021, **4**, 4872–4884.

- S23 S. Correa, *Appl. Surf. Sci.*
- S24 S. Radhakrishnan, C. Sumathi, A. Umar, S. Jae, J. Wilson and V. Dharuman, *Biosens. Bioelectron.*, 2013, **47**, 133–140.
- S25 N. Hui, W. Wang and X. Luo, 2015, 556–561.
- S26 J. Wang, G. Xu, W. Wang, S. Xu and X. Luo, *Chem. - An Asian J.*, 2015, **10**, 1892–1897.
- S27 J. J. García-Guzmán, D. López-Iglesias, L. Cubillana-Aguilera, D. Bellido-Milla, J. M. Palacios-Santander, M. Marin, S. D. Grigorescu, C. Lete and S. Lupu, *Electrochim. Acta*
- S28 Chen, W. Chen, Y. Wang, X. Wang, Y. Ding, D. Zhao and J. Liu, *RSC Adv.*, 2022, **12**, 15038–15045.