

Supporting Information

Title: Amplification of oxidative stress with hyperthermia-enhanced chemodynamic process and MTH1 inhibition for tumor sequential nanocatalytic therapy

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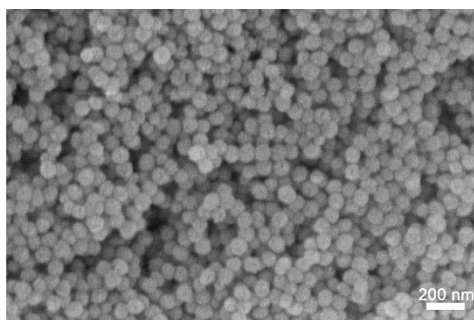


Fig. S1 SEM image of DMSN NPs.

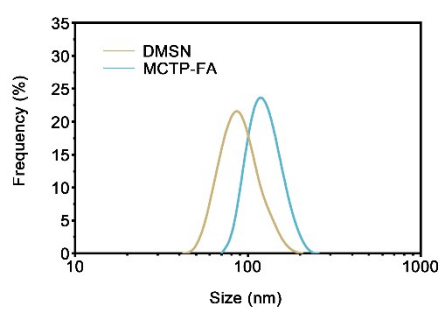


Fig. S2 DLS measurements of DMSN and MCTP-FA NPs.

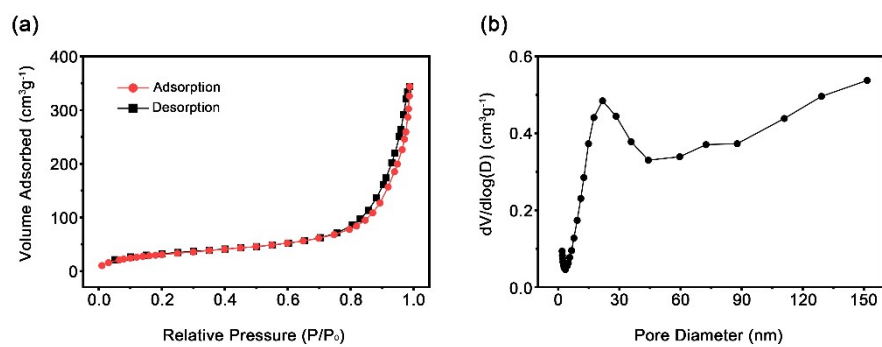


Fig. S3 (a) N₂ adsorption-desorption isotherms and (b) pore size distribution of the DMSN NPs.

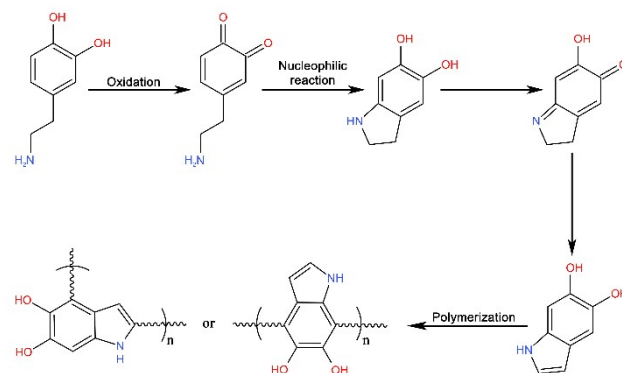


Fig. S4 Schematic illustration of the synthesis of PDA shell through oxidative polymerization.

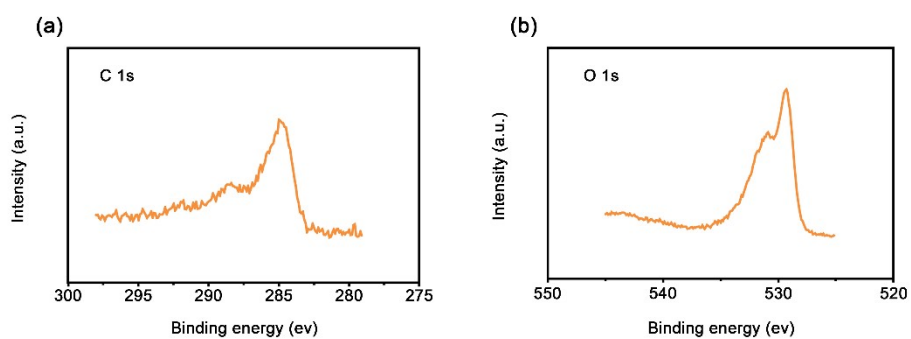


Fig S5 XPS high resolution C 1s (a) spectrum and O 1s (b) spectrum of CeO₂ NPs.

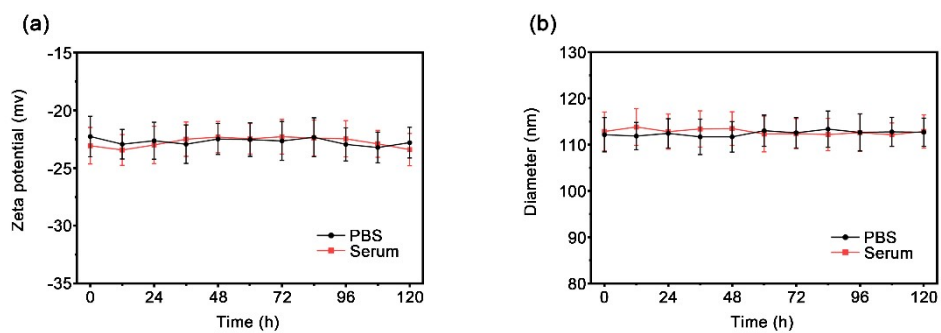


Fig. S6 Zeta potential (a) and DLS (b) changes of MCTP-FA NPs with time in PBS and serum every 12 h (mean $\pm$ SD, $n = 3$).

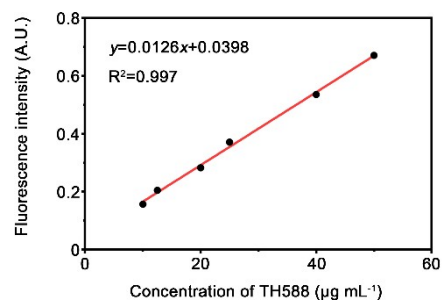


Fig. S7 The standard curve of absorption intensity of TH588 as a function of concentration.

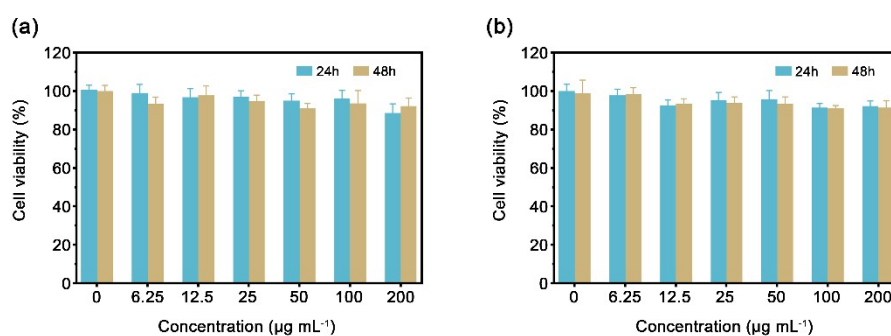


Fig. S8 Cell viabilities of L929 fibroblast cells (a) and HUVEC cells (b) incubated with MCTP-FA NPs at a series of concentrations for 24 h and 48 h.

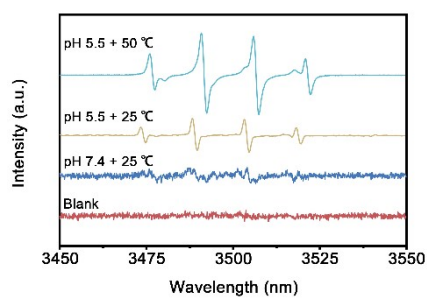


Fig. S9 ESR spectra of $\cdot\text{OH}$ generation in different reaction systems (Blank: ESR spectrum of H_2O).

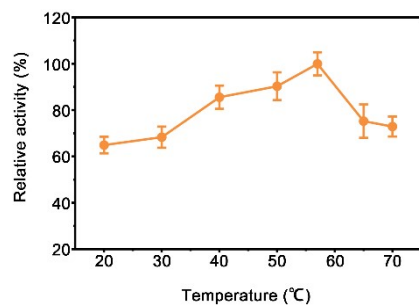


Fig. S10 Effect of temperature on POD-mimic catalytic activity of the MC NPs.

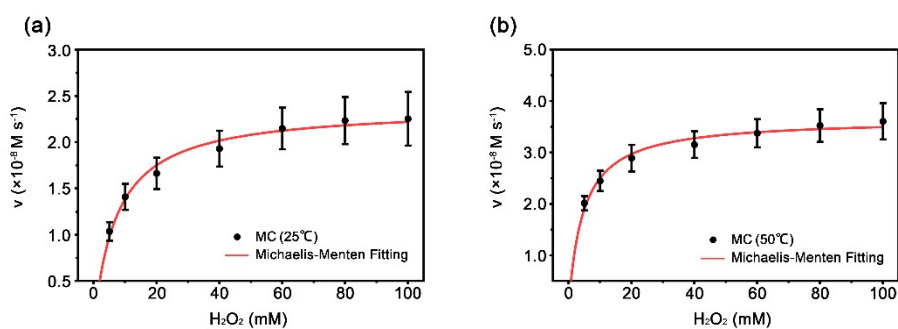


Fig. S11 Michaelis–Menten kinetics for MC NPs with H₂O₂ as the substrate at 25 °C (a) and 50 °C (b).

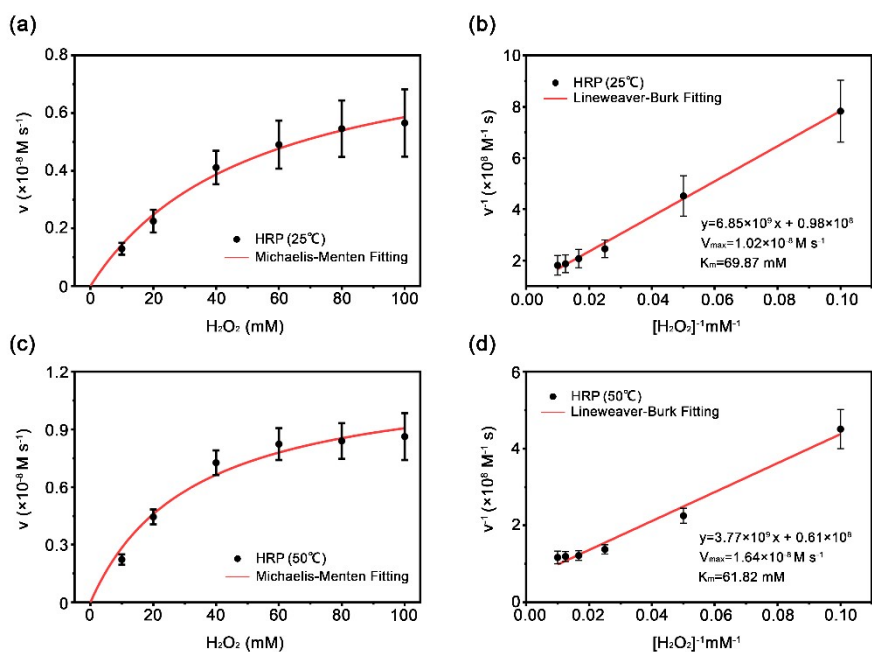


Fig. S12 Michaelis–Menten kinetics and Lineweaver–Burk plotting for HRP with H₂O₂ as the substrate at 25 °C (a, b) and 50 °C (c, d).

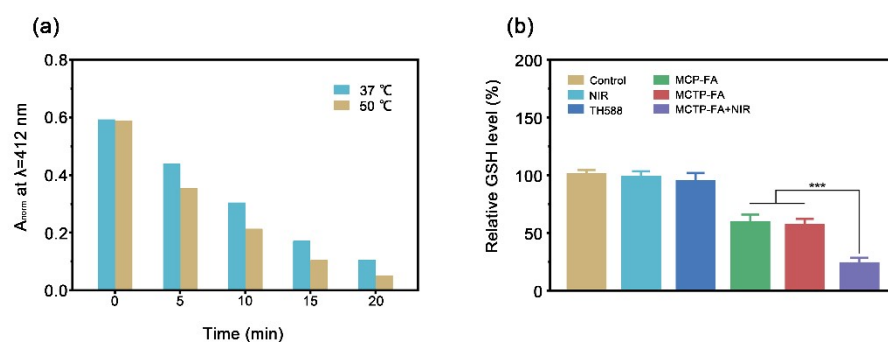


Fig. S13 (a) Effect of temperature on the velocity of GSH consumption. (b) The relative GSH level in MMNG/HOS cells of different treatment groups.

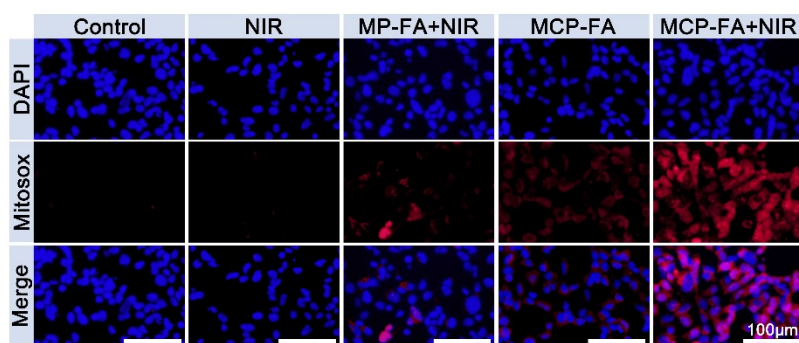


Fig. S14 Fluorescence images of MMNG/HOS cells stained by Mitosox Red after treatment with different formulations.

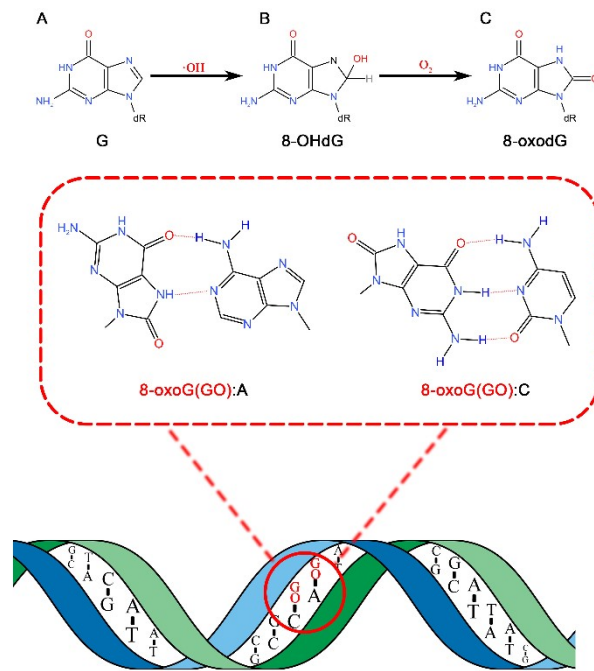


Fig. S15 Formation mechanism of 8-oxoG and changes in base pairing properties.

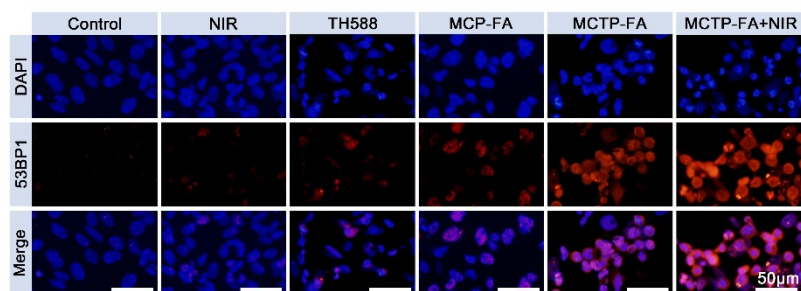


Fig. S16 Immunofluorescence images of 53BP1 after different treatments.

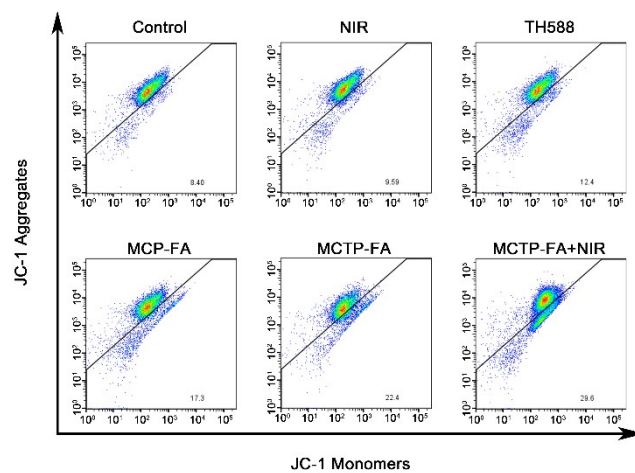


Fig. S17 The decrease of mitochondrial membrane potential after different treatments confirmed by JC-1 analysis.

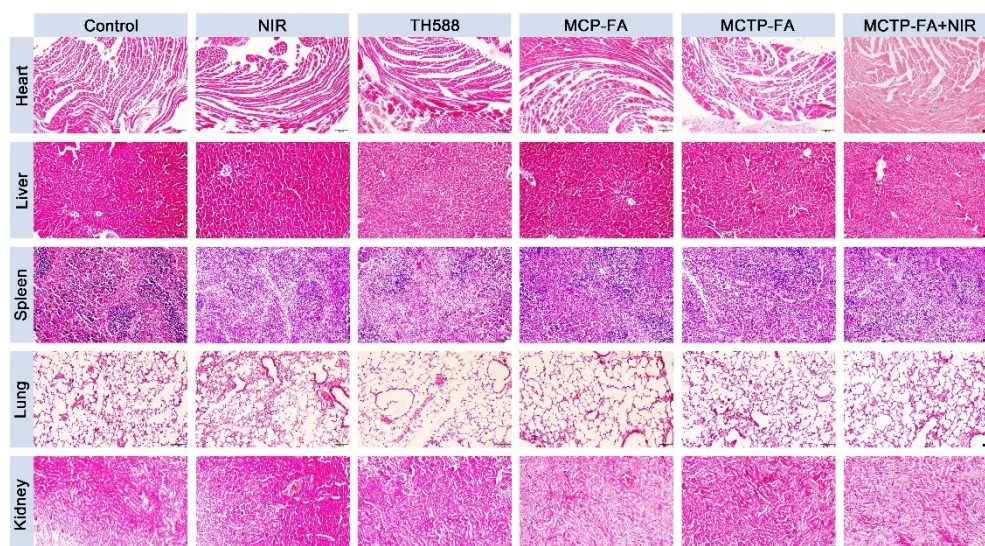


Fig. S18 H&E-stained images of major organs in different groups.

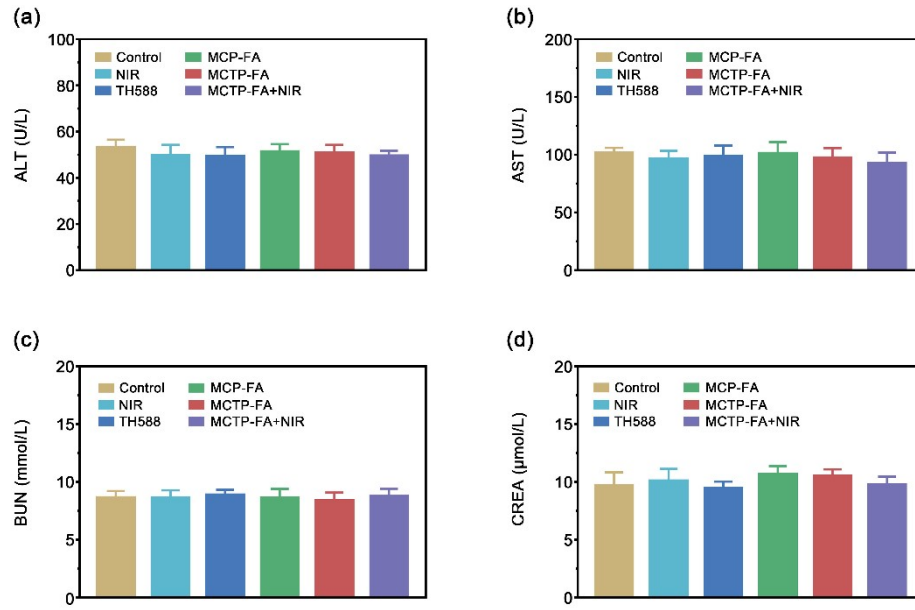


Fig. S19 Biosafety evaluation by blood biochemistry test of mice after intravenous injection with MCTP-FA NPs. (a, b) Serum levels of ALT and AST (liver function index). (c, d) Serum levels of BUN and CREA (kidney function index).

Table S1: Kinetic parameters (K_m , V_{max}) of various nanozymes with H_2O_2 as the substrate for POD-mimic catalysis.

Nanozyme	V_{max} (M s ⁻¹)	K_m (mM)	Reference
Fe ₂ O ₃	3.05×10^{-8}	86.43	1
Fe ₃ O ₄	1.13×10^{-8}	4.94	2
Zn-CuO	3.0×10^{-9}	71	3
CuO	1.61×10^{-7}	400	3
Ala-Fe ₃ O ₄	4.45×10^{-9}	226.6	4
PtFe	8.182×10^{-8}	217.6	5
Cu _{2-x} Te	7.3×10^{-7}	189	6
CeO ₂	2.63×10^{-8}	32.11	7

Reference

1. S. Tanaka, Y. Kaneti, R. Bhattacharjee, M. Islam, R. Nakahata, N. Abdullah, S. Yusa, N. Nguyen, M. Shiddiky, Y. Yamauchi and M. Hossain, *ACS Appl Mater Interfaces*, 2018, **10**, 1039-1049.
2. W. Feng, X. Han, R. Wang, X. Gao, P. Hu, W. Yue, Y. Chen and J. Shi, *Adv Mater*, 2019, **31**, e1805919.
3. A. Nagvenkar and A. Gedanken, *ACS Appl Mater Interfaces*, 2016, **8**, 22301-22308.
4. K. Fan, H. Wang, J. Xi, Q. Liu, X. Meng, D. Duan, L. Gao and X. Yan, *Chem Commun*, 2016, **53**, 424-427.
5. S. Li, L. Shang, B. Xu, S. Wang, K. Gu, Q. Wu, Y. Sun, Q. Zhang, H. Yang, F. Zhang, L. Gu, T. Zhang and H. Liu, *Angew Chem Int Ed Engl*, 2019, **58**, 12624-12631.
6. M. Wen, J. Ouyang, C. Wei, H. Li, W. Chen and Y. Liu, *Angew Chem Int Ed Engl*, 2019, **58**, 17425-17432.
7. S. Dong, Y. Dong, T. Jia, S. Liu, J. Liu, D. Yang, F. He, S. Gai, P. Yang and J. Lin, *Adv Mater*, 2020, **32**, e2002439.