

Supplementary Data

For

A novel electrochemical detection method for butylated hydroxyanisole (BHA) as antioxidant: BHA imprinted polymer based on nickel ferrite@graphene nanocomposite and its application

Bahar Bankoğlu Yola^a, Sena Bekerecioğlu^b, İlknur Polat^b, Necip Atar^c, Mehmet Lütfi Yola^{b*}

^a*Gaziantep Islam Science and Technology University, Faculty of Engineering and Natural Sciences, Department of Engineering Basic Sciences, Gaziantep, Turkey*

^b*Hasan Kalyoncu University, Faculty of Health Sciences, Department of Nutrition and Dietetics, Gaziantep, Turkey*

^c*Pamukkale University, Faculty of Engineering, Department of Chemical Engineering, Denizli, Turkey*

* To whom correspondence should be addressed:

E-mail: mlutfi.yola@hku.edu.tr

Tel.: +903422118080 Fax: +903422118081

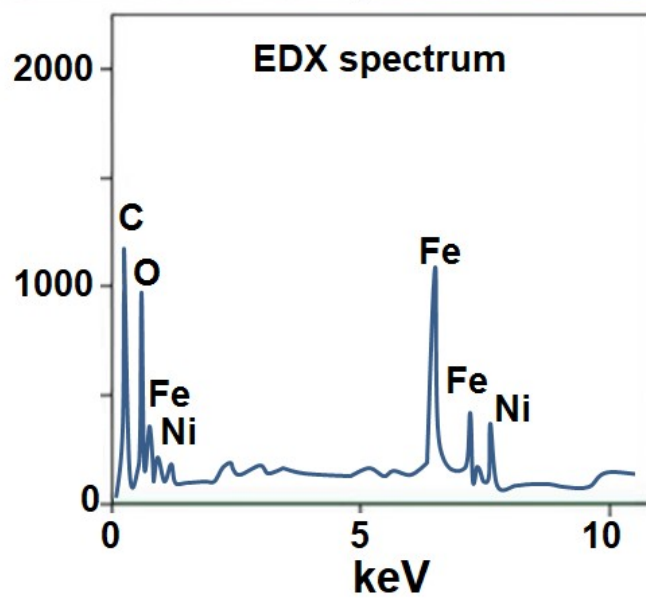
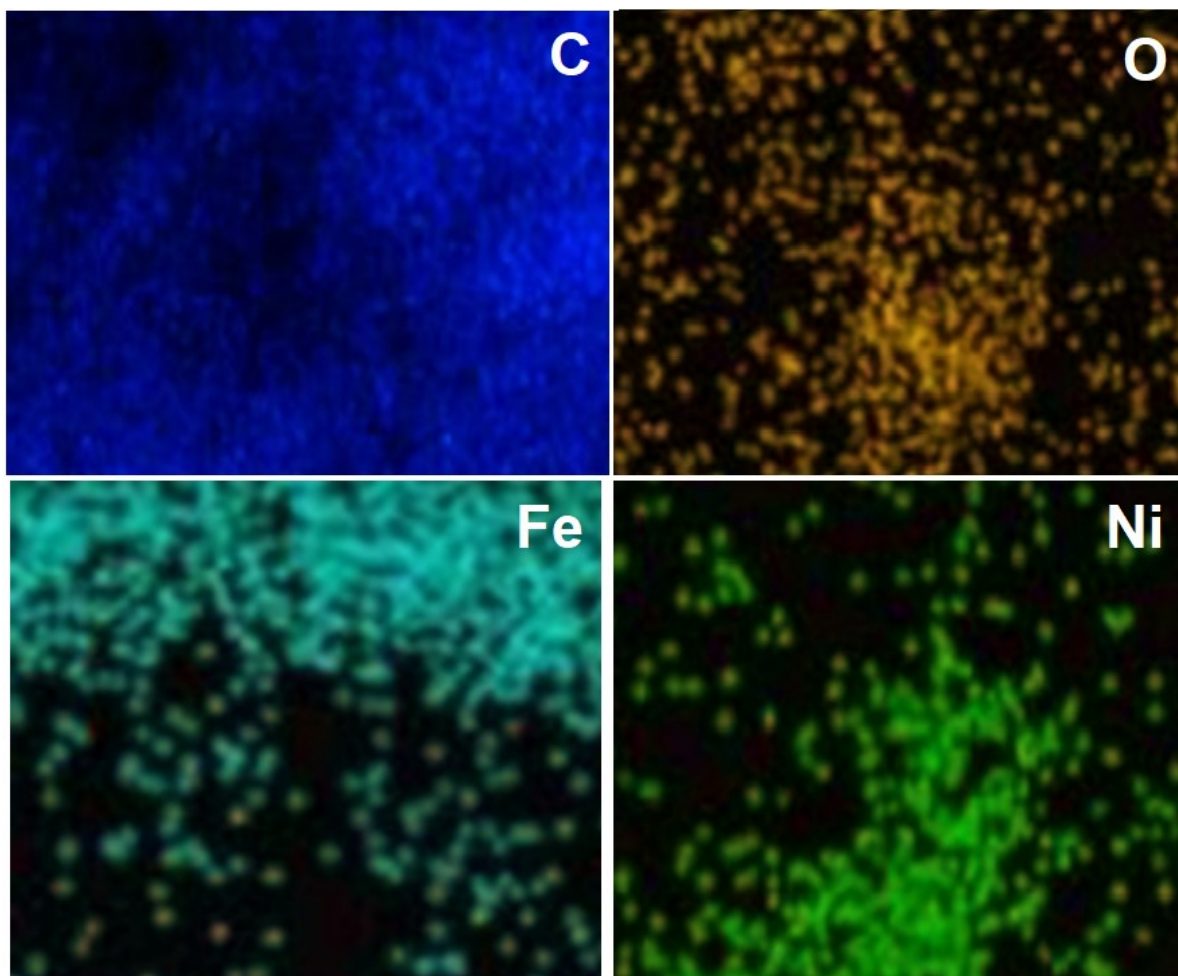


Fig. S1. EDS elemental distribution and EDX spectrum of $\text{NiFe}_2\text{O}_4@\text{Gr}(0.6)$

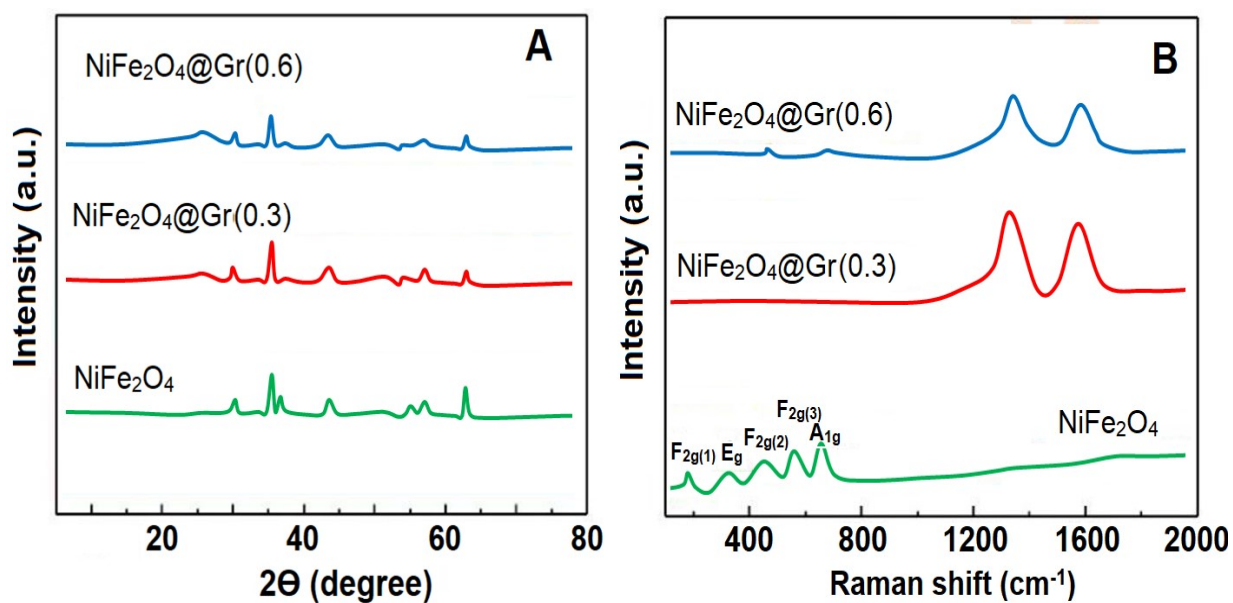


Fig. S2. (A) XRD patterns and (B) Raman spectra of NiFe₂O₄, NiFe₂O₄@Gr(0.3) and NiFe₂O₄@Gr(0.6)

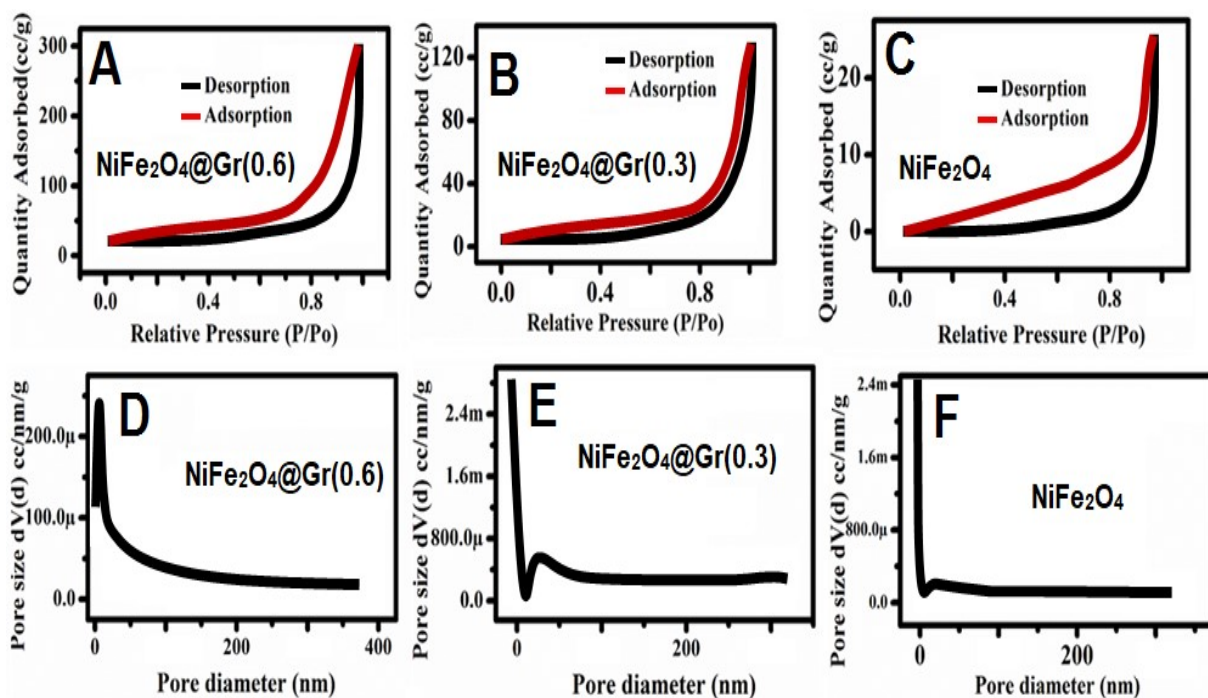


Fig. S3. (A), (B), (C) N₂ adsorption-desorption isotherms of NiFe₂O₄, NiFe₂O₄@Gr(0.3) and NiFe₂O₄@Gr(0.6), and (D), (E), (F) pore size distribution isotherms

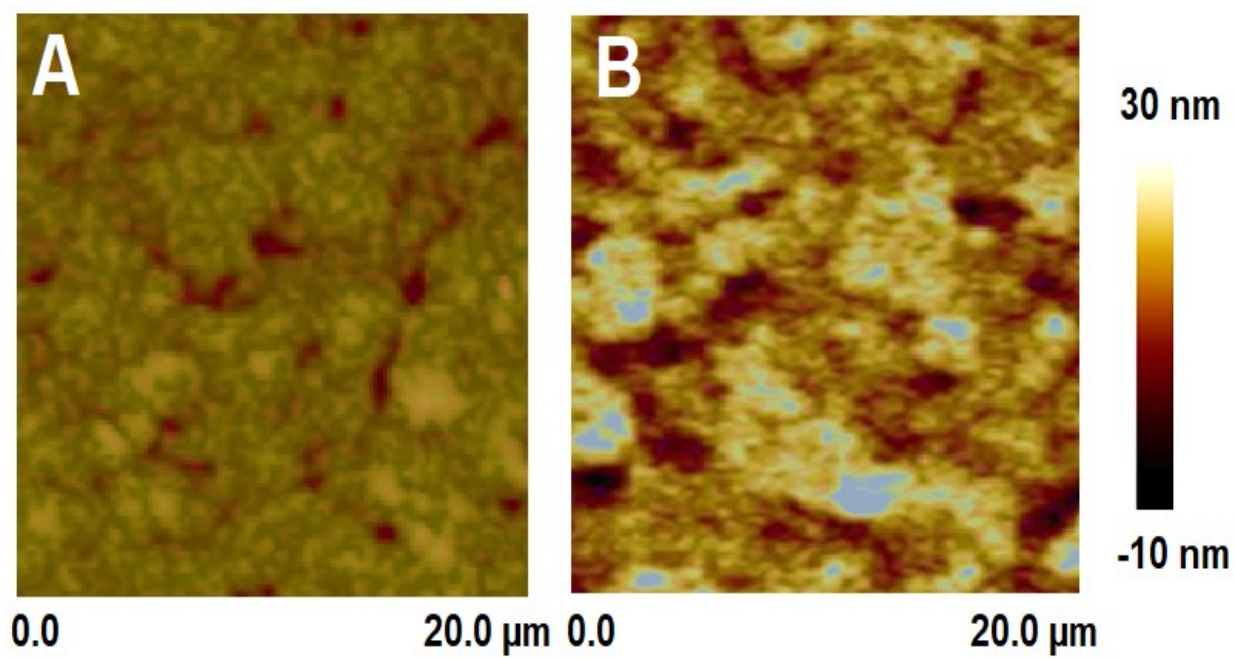


Fig. S4. AFM images of (A) bare GCE and (B) $\text{NiFe}_2\text{O}_4@\text{Gr}(0.6)/\text{GCE}$

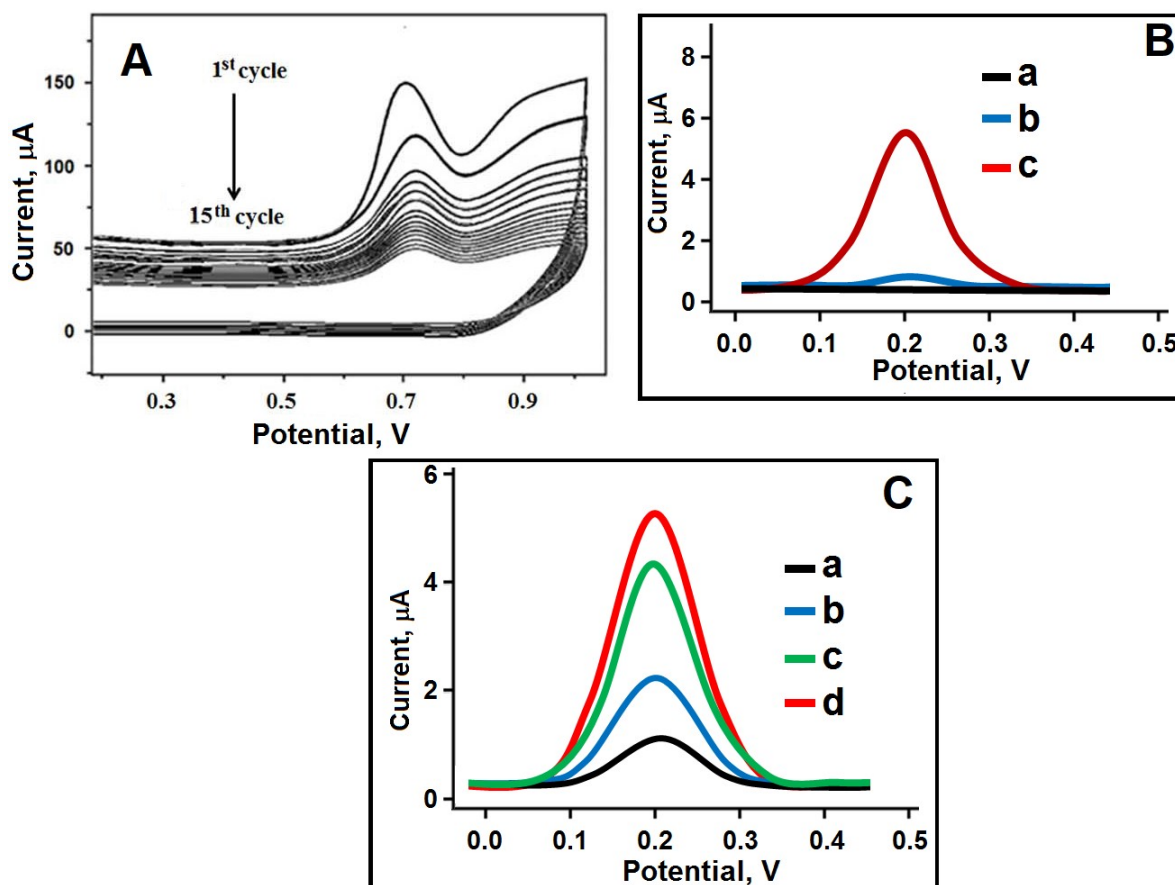


Fig. S5. (A) 100.0 mM Py polymerization containing 25.0 mM BHA on NiFe₂O₄@Gr(0.6)/GCE (Scan rate: 100 mV s⁻¹), (B) DPVs of the prepared electrodes in this study: (a) MIP/NiFe₂O₄@Gr(0.6) in blank buffer solution (pH 7.0), (b) NIP/NiFe₂O₄@Gr(0.6) after rebinding of 0.5 nM BHA in 0.1 M PBS (pH 7.0), (c) MIP/NiFe₂O₄@Gr(0.6) after rebinding of 0.5 nM BHA in 0.1 M PBS (pH 7.0), (C) DPVs of different molecularly imprinting electrodes after rebinding of 0.5 nM BHA in 0.1 M PBS (a) MIP/bare GCE, (b) MIP/NiFe₂O₄/GCE, (c) MIP/NiFe₂O₄@Gr(0.3)/GCE and (d) MIP/NiFe₂O₄@Gr(0.6)/GCE

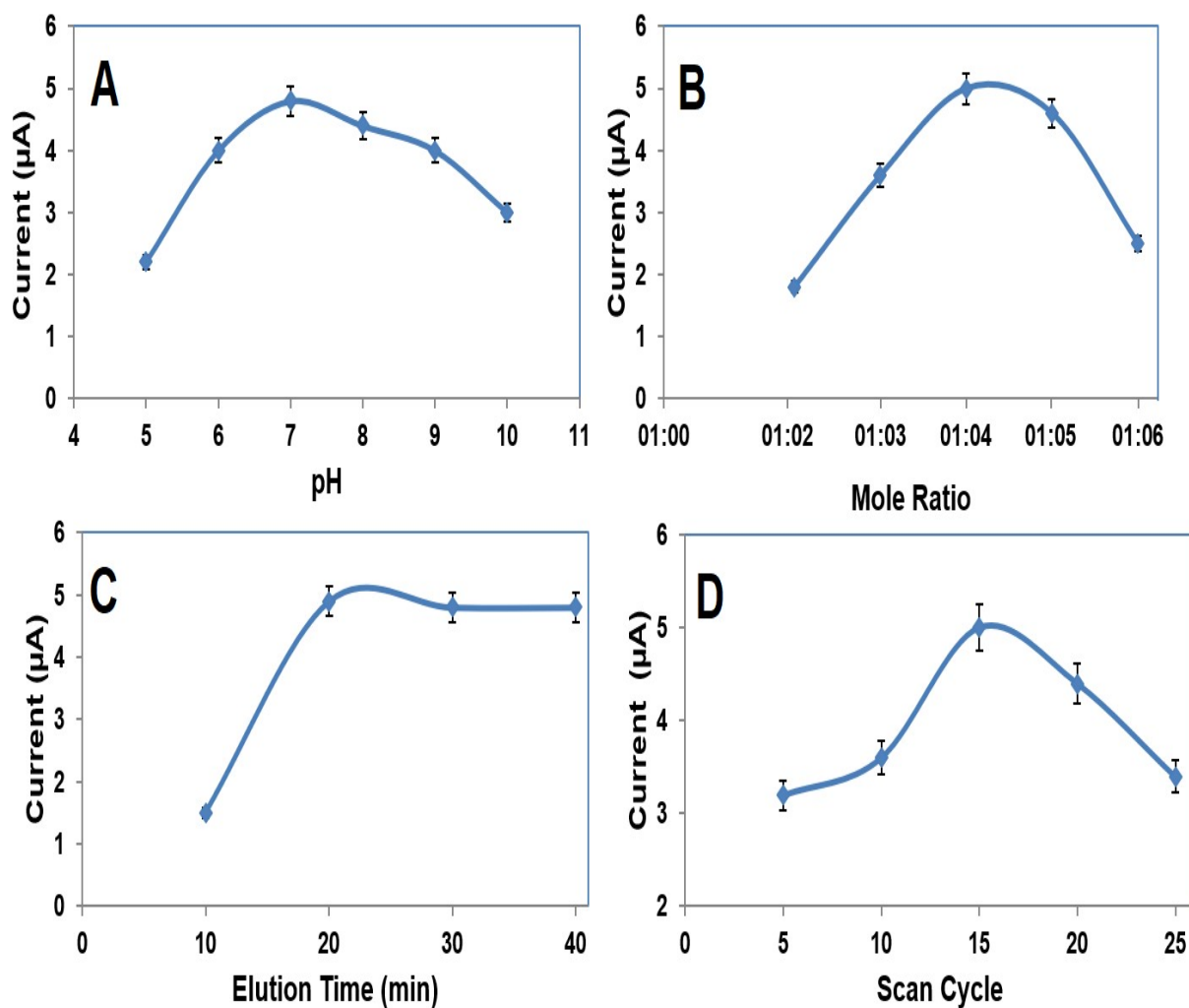


Fig. S6. Effect of (A) pH, (B) mole ratio, (C) desorption time, (D) scan cycle on signals of DPVs (in presence of 0.5 nM BHA) (n = 6)

Table S1. Recovery results of BHA in flour (n=6)

Sample	Added BHA (nM)	Found BHA (nM)	*Recovery (%)
Flour	-	0.273 ± 0.002	-
	0.100	0.374 ± 0.002	100.27 ± 0.01
	0.300	0.572 ± 0.005	99.83 ± 0.06
	0.500	0.772 ± 0.003	99.87 ± 0.04

*Recovery = Found BHA, nM / Real BHA, nM

Table S2. k and k' values of BHA imprinted electrodes (MIP/NiFe₂O₄@Gr(0.6)/GCE and NIP/NiFe₂O₄@Gr(0.6)/GCE)

	MIP		NIP		k'
	ΔI (μA)	k	ΔI (μA)	k	
BHA	5.20	-	0.50	-	-
BHT	0.50	10.40	0.30	1.67	6.23
PHE	0.40	13.00	0.20	2.50	5.20
AA	0.30	17.33	0.10	5.00	3.47
CA	0.20	26.00	0.05	10.00	2.60

Analyte concentrations: 0.5 nM BHA, 100.0 nM BHT, 100.0 nM PHE, 100.0 nM AA, 100.0 nM CA

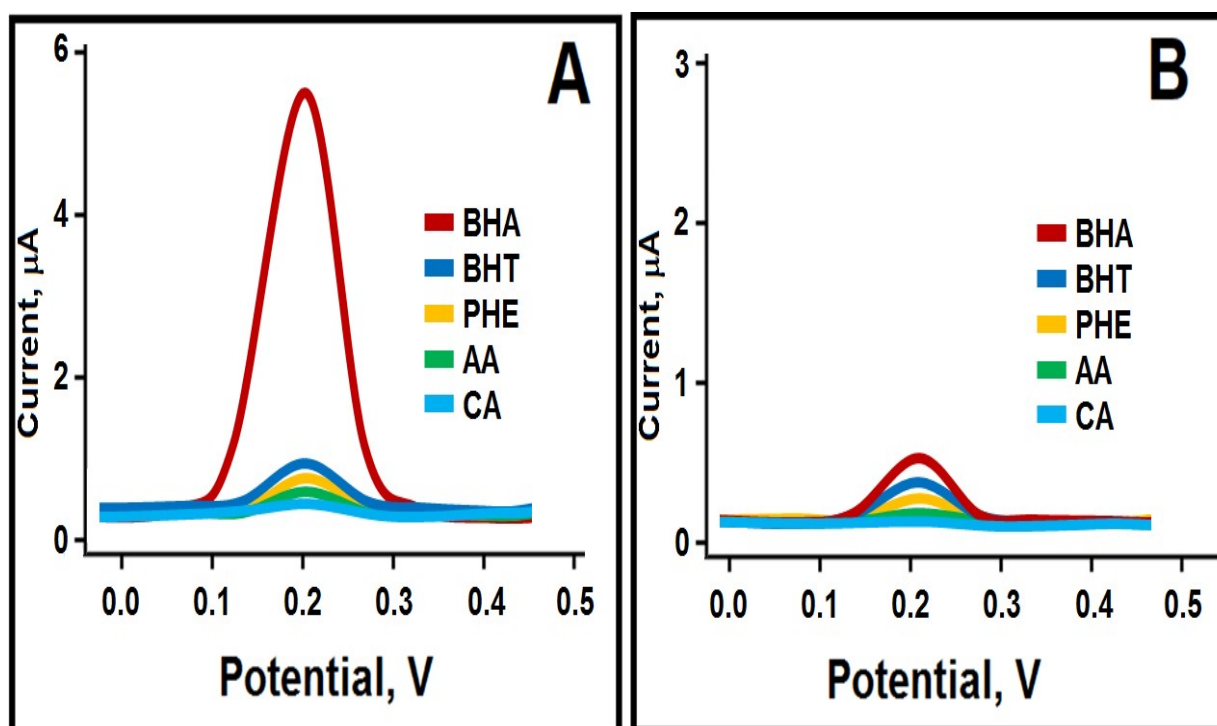


Fig. S7. DPVs of (A) MIP/NiFe₂O₄@Gr(0.6)/GCE and (B) NIP/NiFe₂O₄@Gr(0.6)/GCE in 0.5 nM BHA, 100.0 nM BHT, 100.0 nM PHE, 100.0 nM AA, 100.0 nM CA

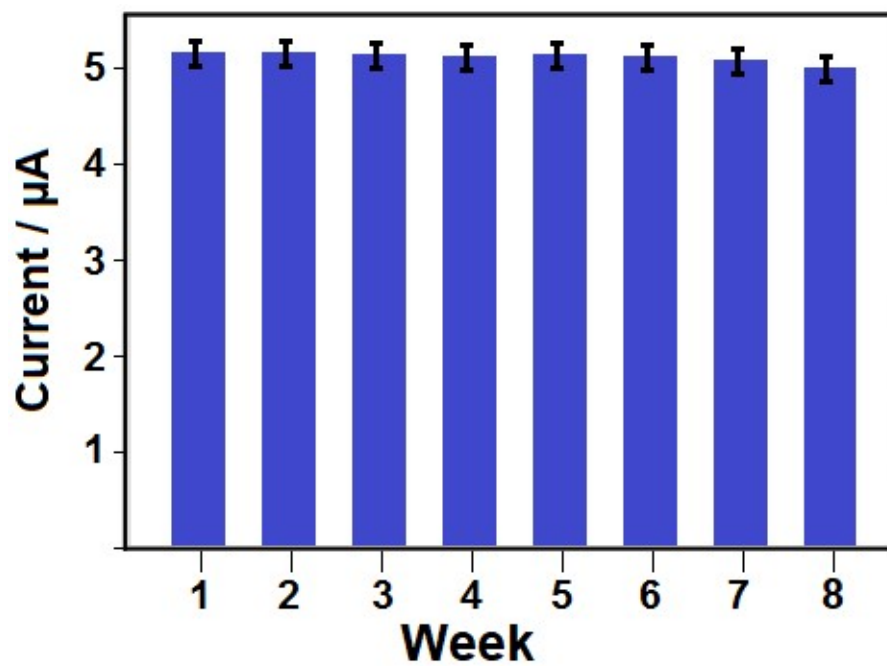


Fig. S8. Stability test of MIP/NiFe₂O₄@Gr(0.6)/GCE including 0.5 nM BHA (n = 6)