

## Supporting Information for

# All-in-one detection of breast cancer-derived exosomal miRNA on pen-based paper chip

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## Preparation of lateral flow strip

### i. Pretreatment of Sample Pads

Immerse a 20 cm × 30 cm glass fiber membrane (GL-b04) in 50 mL of sample pad treatment solution. Air-dry at room temperature overnight and then dry in a 37 °C oven for 4 hours. After drying, cut into 2 cm × 30 cm strips, seal in aluminum foil bags, and store in a constant temperature and humidity chamber for later use.

### ii. Preparation of Detection Probes (AuNPs-FAM-Antibody)

Take 1.5 mL of 13 nm colloidal gold solution, add 0.2 M K<sub>2</sub>CO<sub>3</sub> to adjust the pH to 9.0. The mixture was thoroughly shaken and mixed well. Add 20 µg of streptavidin and react at room temperature for 2 hours. Add 10 µL of 10% BSA, incubate at room temperature for 1 hour, then centrifuge (10 min, 4 °C, 10000 rpm). The supernatant liquid was discarded while the labeled complex precipitate obtained after resuspending with a volume of 100 µL served as the detection probe (AuNPs-FAM-Antibody), which were stored away from light at 4 °C until further use.

### iii. Preparation of Conjugate Pads

Cut a 22 cm × 30 cm glass fiber membrane (8964) into 1 cm × 30 cm strips and set aside in a constant temperature and humidity chamber. Spray the detection probe onto the 1 cm × 30 cm glass fiber membrane at a rate of 6 µL/cm. Air-dry overnight in a 37 °C oven, seal in aluminum foil bags, and store in a constant temperature and humidity chamber for later use.

### iv. Preparation of Reaction Membrane

In the direction from conjugate pad to absorbent pad, coat 1 mg/mL anti-FAM antibody and 1 mg/mL biotin using a spray-coating instrument at a rate of 1 µL/cm onto nitrocellulose membrane (NC membrane), designated as the test line and control line, respectively. After drying overnight in a 37 °C convection oven, seal in aluminum foil bags and store in a constant temperature and humidity chamber for later use.

### v. Preparation of Absorbent Pads

Cut a 20 cm × 30 cm absorbent paper (H-2) into 2.5 cm × 30 cm strips and set aside in a constant temperature and humidity chamber.

### vi. Assembly of Test Strips

Assemble all components on a PVC baseplate in the following order: sample pad, conjugate pad, reaction membrane, and absorbent pad. Each section should overlap by 1-2 mm. Use a programmable KM-3100 strip cutter to cut the assembled strip into a width of 4 mm. Place the cut strips into aluminum foil bags, seal with desiccant, and store in a constant temperature and humidity chamber for later use.

**Table S1** Existing methods for microRNAs detection

Reference	Platform	Target	Detection Method	Mechanism	Limits of detection
Li et al. [1]	Electrochemical biosensor	Lytic Exosome/ miRNA-21	Electrochemical biosensor	Primer exchange reaction with MOF@Pt@MOF nanozyme	0.29 fM
Li et al. [2]	Carbon Nanotube Field-Effect Transistor Biosensor	Purified exosomal miRNA/ miRNA-21	Carbon Nanotube Field- Effect Transistor Biosensor	Hybridization between the immobilized DNA probe and target miRNA	0.87 aM
Yang et al. [3]	Micromixer Biochip	Intact Exosome/ miRNA-21 TTF-1 mRNA	Fluorescence- based	Cationic Lipoplex Nanoparticles	2.06 × 10 <sup>9</sup> and 3.71 × 10 <sup>9</sup> exosomes /mL
Zhao et al. [4]	Thermophoresis	Intact Exosome/ miRNA-375	Fluorescence- based	Hybridization of DNA probes by nanoflares	0.36 fM
Cho et al. [5]	96-well plates	Intact Exosome/ miRNA-21 miRNA -574-3p CD63 EpCAM	Fluorescence- based	Molecular beacon	-
Deng et al. [6]	Lateral flow assay	Lytic tumor cells/ miRNA-21	Quantum dots- labeled strip biosensor	Enzyme-free catalyzed hairpin assembly	200 aM
Qian et al. [7]	Agarose-based microfluidic chip	Intact Exosome/ miRNA-21	Fluorescence- based	Enzyme-free catalyzed hairpin assembly	1000 exosomes
This work	Pen-based paper chip	Lytic Exosome/ miRNA-21	Lateral flow assay	Enzyme-free catalyzed hairpin assembly	25 fM

**Table S2** Sequences of DNA probes and microRNA

<b>Name</b>	<b>Sequences (5'-3')</b>					
HP1	1	2	3	4*	3*	2*
	TCAACATC-AGTCTGA-TAAGCTA-CCATGTGTAGA-TAGCTTA-TCAGACT-FAM					
HP2	3	4	3*	2*	4*	
	TAAGCTA-TCTACACATGG-TAGCTTA-TCAGACT-CCATGTGTAGA-Biotin					
miR-21	UAGCUUAUCAGACUGAUGUUGA					
miR-210	CUGUGCGUGUGACAGCGGCUGA					
miR-214	ACAGCAGGCACAGACAGGCAGU					

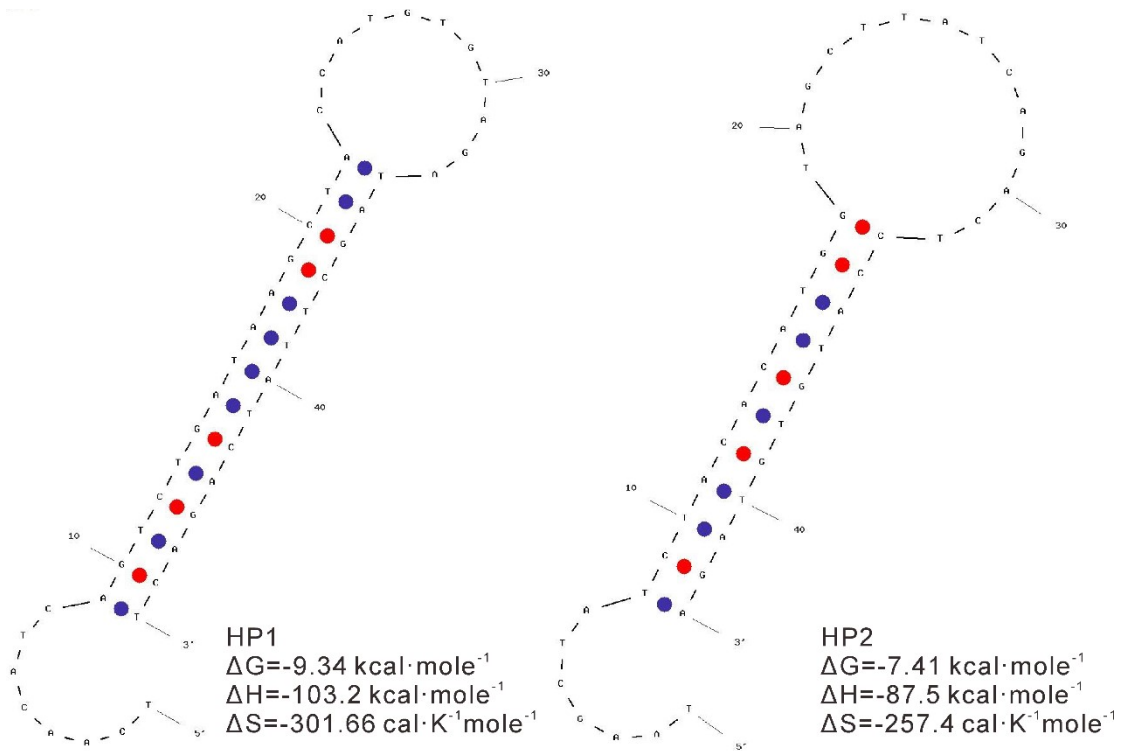


Figure S1. Structure image and thermal kinetic parameters of HP1 probe and HP2 probe.

## References

- [1] X. Li, X. Li, D. Li, M. Zhao, H. Wu, B. Shen, P. Liu, S. Ding, Electrochemical biosensor for ultrasensitive exosomal miRNA analysis by cascade primer exchange reaction and MOF@Pt@MOF nanozyme, *Biosensors & bioelectronics* 168 (2020) 112554, <https://doi.org/10.1016/j.bios.2020.112554>.
- [2] T.X. Li, Y.Q. Liang, J.H. Li, Y. Yu, M.M. Xiao, W. Ni, Z.Y. Zhang, G.J. Zhang, Carbon Nanotube Field-Effect Transistor Biosensor for Ultrasensitive and Label-Free Detection of Breast Cancer Exosomal miRNA21, *Analytical chemistry* 93(46) (2021) 15501-15507, <https://doi.org/10.1021/acs.analchem.1c03573>.
- [3] Y.C. Yang, E. Kannisto, S.K. Patnaik, M.E. Reid, L. Li, Y. Wu, Ultrafast Detection of Exosomal RNAs via Cationic Lipoplex Nanoparticles in a Micromixer Biochip for Cancer Diagnosis, *Acs Appl Nano Mater* 4(3) (2021) 2806-2819, <https://doi.org/10.1021/acsanm.0c03426>.
- [4] J.X. Zhao, C. Liu, Y.K. Li, Y. Ma, J.Q. Deng, L.L. Li, J. Sun, Thermophoretic Detection of Exosomal microRNAs by Nanoflakes, *J Am Chem Soc* 142(11) (2020) 4996-5001, <https://doi.org/10.1021/jacs.9b13960>.
- [5] J.H. Lee, J.A. Kim, S. Jeong, W.J. Rhee, Simultaneous and multiplexed detection of exosome microRNAs using molecular beacons, *Biosensors & bioelectronics* 86 (2016) 202-210, <https://doi.org/10.1016/j.bios.2016.06.058>.
- [6] H.P. Deng, Q.W. Liu, X. Wang, R. Huang, H.X. Liu, Q.M. Lin, X.M. Zhou, D. Xing, Quantum dots-labeled strip biosensor for rapid and sensitive detection of microRNA based on target-recycled nonenzymatic amplification strategy, *Biosensors & bioelectronics* 87 (2017) 931-940, <https://doi.org/10.1016/j.bios.2016.09.043>.
- [7] C.G. Qian, Y.J. Xiao, J. Wang, Y.W. Li, S.J. Li, B. Wei, W. Du, X.J. Feng, P. Chen, B.F. Liu, Rapid exosomes concentration and in situ detection of exosomal microRNA on agarose-based microfluidic chip, *Sensor Actuat B-Chem* 333(15) (2021) 129559, <https://doi.org/10.1016/j.snb.2021.129559>.