

Real-time detection of SARS-CoV-2 in clinical samples via ultrafast ligation-dependent RNA transcription amplification

Peng Zhang ‡,^a Yang Li ‡,^a Dongmei Zhang ^c, Xinghao Zhu ^a, Jinling Guo ^a, Cuiping Ma ^b, Chao Shi ^{*,a}

^a *Qingdao Nucleic Acid Rapid Testing International Science and Technology Cooperation Base, College of Life Sciences; Department of Pathogenic Biology, School of Basic Medicine; Department of Clinical Laboratory, the Affiliated Hospital of Qingdao University, Qingdao University, Qingdao, 266071, PR China.*

^b *Shandong Provincial Key Laboratory of Biochemical Engineering, Qingdao Nucleic Acid Rapid Detection Engineering Research Center, College of Marine Science and Biological Engineering, Qingdao University of Science and Technology, Qingdao, 266042, PR China.*

^c *Qingdao special servicemen recuperation center of PLA navy, Qingdao, 266071, PR China.*

* To whom correspondence should be addressed.

‡ These two authors contributed equally to this work.

Tel (Fax): +86-532-85955399

E-mail: sc169@163.com

Supplementary Information

Table S1. Sequences of nucleic acids used in this work

Name	Sequence (5'-3')
Promoter probe	^a AACACTATTAGCATAAGCAGTTGTGCCCTATAGTGAGTCGTATTAGG AAGG
Reporter probe	^b TCGACCTAGCATATCCATGATATCTGTTAGTTTTTTTCTGAAAGACC GCCGGACAAGTGACAGCTTGACAAATGTTAAA
Reporter probe (no terminator)	CCGCCGGACAAGTGACAGCTTGACAAATGTTAAA
T7 promoter	20-nt TAATACGACTCACTATAGGG
	22-nt CCTAATACGACTCACTATAGGG
	24-nt TTCCTAATACGACTCACTATAGGG
	26-nt CCTTCCTAATACGACTCACTATAGGG
Molecular beacon	MB1 FAM- CCGCCGGACAAGGAGGCTGGAGTCAACCGGCGG -DABCYL
	MB2 FAM-AGTATACCTGCAGTGACTCTTAAGATCCAGGTATACT-DABCYL
	MB3 FAM-ACCTTCTATAGCAAACACCATTGTCACACTCCACCTCAGCCTAT AGAAGGT-DABCYL
	MB4 FAM-ACTATAGGTATACAACCTACTACCTCACCTATAGT-DABCYL
qRT-PCR primer set (^c LR898678.2)	F GTGATGTAGAAAACCCCTCACC
	R AGCAAGAACAAGTGAGGC
qRT-LAMP primer set (^c LR898678.2)	F3 TGCTTCAGTCAGCTGATG
	B3 TTAAATTGTCATCTTCGTCCTT
	FIP TCAGTACTAGTGCCTGTGCCACAATCGTTTTTAAACGGGT
	BIP TCGTATACAGGGCTTTTGACATCTATCTTGGAAGCGACAACAA
	LF CTGCACTTACACCGCAA
	LB GTAGCTGGTTTTGCTAAATTCC
RNA fragment with SARS-CoV-2 <i>orf1ab</i> gene specific sequence (^c OQ117226.1)	AUGCCACAACUGCUUAUGCUAAUAGUGUUUUUAACAUUUGUCAAGC UGUCACGGCC
RNA fragment with SARS-CoV <i>orf1ab</i> gene specific sequence (^c MN312831.1)	^d AUGCUACAACUGCUUAUGCUAAUAGUGUCUUUAACAUUUGUCAAG CUGUUACAGCC

^a Sequences marked in red and blue were the sequence complementary to the part of the target RNA and T7 promoter binding site, respectively.

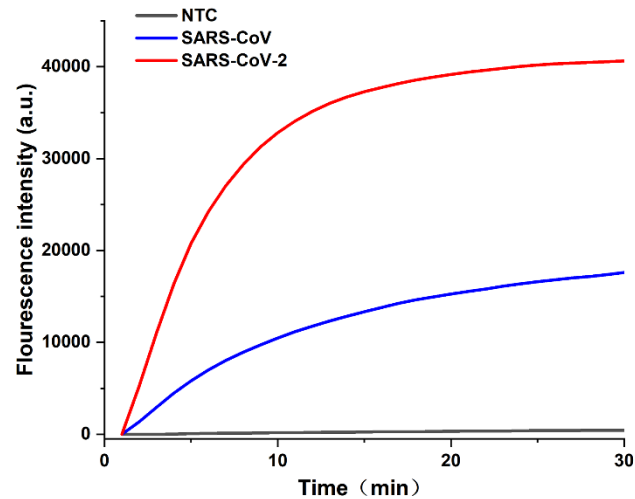
^b Sequences marked in orange, green, and red were the VSV terminator, the sequence consistent with the loop part of the MB, and the sequence complementary to the part of the target RNA, respectively.

^c GenBank accession number.

^d The portion in italic type represented the nucleotides variation compared to the RNA fragment with SARS-CoV-2 *orf1ab* gene specific sequence.

Procedure of qRT-PCR and qRT-LAMP

The reaction system of qRT-PCR with a total volume of 20 μL consists of 2 μL sample, 0.4 μL dNTPs (10 mM each), 0.8 μL forward and backward primer (10 μM each), 0.3 μL *Taq* DNA polymerase (5 U/ μL), 1.3 μL 10 \times *Taq* buffer, 0.5 μL EvaGreen Dye, 0.2 μL HiScript II Reverse Transcriptase (200 U/ μL), 1.3 μL 5 \times HiScript II Buffer, 0.2 μL Murine RNase inhibitor (40 U/ μL), 0.08 MgSO_4 solution (100 mM) and 12.12 μL DEPC-treated water. The reaction procedure included reverse transcription at 55 $^\circ\text{C}$ for 5 min, denaturation at 95 $^\circ\text{C}$ for 3 min, and 40 cycles consisting of 95 $^\circ\text{C}$ for 10 s, and 58 $^\circ\text{C}$ for 15 s for amplification. qRT-LAMP was performed by adding 2.5 μL sample, 4 μL dNTPs (10 mM each), 1 μL F3 and B3 (10 μM each), 0.8 μL FIP and BIP (100 μM each), 0.4 μL LF and LB (100 μM each), 0.5 μL *Bst* 2.0 WarmStart DNA polymerase (8000 U/mL), 2.5 μL 10 \times ThermoPol reaction buffer, 0.5 μL MgSO_4 solution (100 mM), 0.625 μL EvaGreen Dye, 0.3 μL HiScript II Reverse Transcriptase (200 U/ μL) and 11.875 μL DEPC-treated water into a total volume of 25 μL at 65 $^\circ\text{C}$ for 45 min. Both qRT-PCR and qRT-LAMP assays were carried out by a CFX ConnectTM Real-Time PCR System (Bio-Rad, CA, USA). The fluorescence signal of the qRT-PCR was read after each thermal cycling, while that of the qRT-LAMP was monitored at 1-min intervals.



SARS-CoV-2 AUGCCACAACUGCUUAUGC^{*UAAUAGUGUUUUUAACA*}UUUGUCAAGCUGUCACGGCC
SARS-CoV AUGCUACAACUGCUUAUGC^{*UAAUAGUGU*}CUUUUAACAUUUGUCAAGCUGUUACAGCC

Fig.S1. Capability of SCAN to distinguish RNA with similar sequence to the target. The concentration of the RNA fragments was 1 μ M. The portion in italic type represented the ligation site of the two probes (between the two nucleotides), while the variation nucleotides of the RNA fragment with SARS-CoV *orflab* gene specific sequence were marked in red. NTC represented no template control.