

Small-Molecule Fluorogenic Probe for the Detection of Hypochlorite and Its Application in the Bio-imaging of Human Breast Cancer Cells

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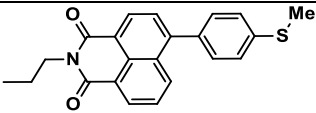
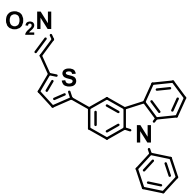
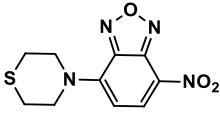
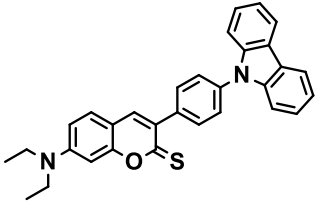
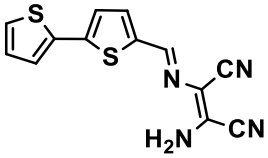
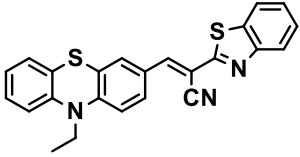
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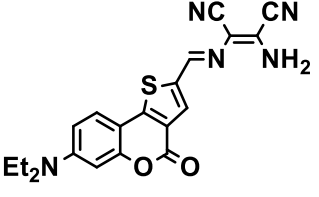
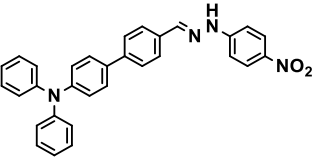
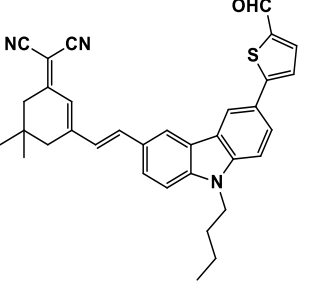
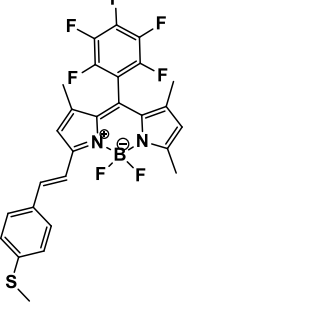
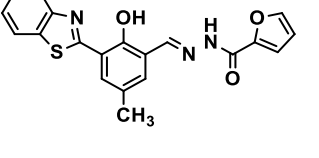
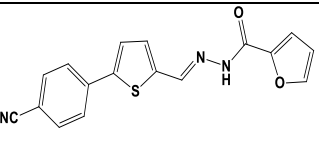
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1. Table S1 Comparison between previously reported ClO⁻ sensors with the current work

| Sl. No. | Probe structure | Solvent | Sensor type | LOD | Application | Reference |
|---------|---|------------------------|-------------|---------|--------------|---|
| 1. |  | DMF | Ratiometric | 0.02 μM | MCF-7 cells | Zhou et al., <i>Chem. Commun.</i> , 2021, 57 , 11366 |
| 2. |  | DMSO (1:1) | Ratiometric | 71.4 nM | HeLa cells | Li et al., <i>Spectrochimica Acta Part A: Molecular and Biomolecular Spectroscopy</i> , 2020, 241 , 118672 |
| 3. |  | Acetonitrile | Turn off | 72 nM | - | Podsiadły et al., <i>Dyes and Pigments</i> , 2021, 193 , 109563 |
| 4. |  | Acetonitrile | Turn on | 0.17 μM | HeLa cells | Yoon et al., <i>Sensors & Actuators: B. Chemical</i> , 2020, 317 , 128213 |
| 5. |  | ~100% aqueous solution | Turn on | 8.3 nM | HeLa cells | Niu et al., <i>Journal of Molecular Liquids</i> , 2020, 320 , 114396 |
| 6. |  | DMSO | Turn off | 0.64 μM | HepG-2 cells | Qu et al., <i>Ind. Eng. Chem. Res.</i> 2018, 57 , 7735–7741 |

| | | | | | | |
|-----|---|---------------|-------------|-------------------------------|--|--|
| 7. |  | DMF | Ratiometric | 94 nM | RAW264.7 cells | Shao et al., <i>New J. Chem.</i> , 2020, 44 , 6232 |
| 8. |  | DMSO | Turn on | 0.36 μM | Zebrafish | Liu et al., <i>Spectrochimica Acta Part A: Molecular and Biomolecular Spectroscopy</i> , 2021, 258 , 119827 |
| 9 |  | DMF | Ratiometric | $1.4 \times 10^{-7} \text{M}$ | HepG2 | Li et al., <i>Dyes and pigments</i> , 2022, 208 , 110879 |
| 10. |  | DMF | Ratiometric | 59nM | Zebrafish and mice (in vivo) | Kim et al., <i>Anal Chem</i> , 2019, 91 , 4172-4178 |
| 11. |  | DMSO (1:9) | Turn on | 1.44 nM | HepG2 cells | Yan et al., <i>J Fluoresc.</i> , 2021, 31 , 569–576 |
| 12. |  | THF/H2O (2:8) | Turn On | 53.8nM | Human breast cancer cells (MDA-MB 231) | This work |

2. Solid state ClO⁻ sensing:

Small pieces of TLC sticks were dipped into 1×10^{-5} M probe solution followed by drying in open air. The sticks were further dipped into different concentrated solutions of NaOCl and dried for 5 minutes. After that photographs were taken to examine the sensing behaviour of the probe TPHZ.

3. Computational method:

Theoretical calculations:

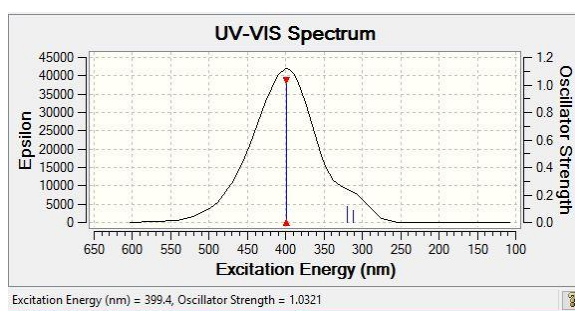


Figure S1. Absorption spectra of the Probe (TPHZ)

Table S2 The vertical main orbital transition of the TPHZ calculated by TDDFT method

| Energy (eV) | Wavelength (nm) | Osc. Strength (f) | Transition |
|-------------|-----------------|-------------------|----------------|
| 3.1042 | 399.40 | 1.0321 | HOMO → LUMO |
| 3.8713 | 320.27 | 0.1164 | HOMO-1 → LUMO |
| 3.9800 | 311.52 | 0.0907 | HOMO → LUMO +1 |

4. Live cell imaging study:

Cytotoxicity assay:

In the present study Human breast cancer cell line MDA-MB 231 and human normal lung fibroblast cell line WI-38 have been used and MTT cell proliferation assay¹ was performed to assess the cytotoxic effect of the ligand TPHZ. In brief, cells growing in a log phase were first seeded in 96-well plates at a concentration of 1×10^4 cells per well and were incubated overnight at 37 °C under 5% CO₂. The cells were then exposed to the different working concentration of ligand TPHZ (0 μM, 10 μM, 20 μM, 40 μM, 80 μM, 100 μM) for 24 hrs.

Following incubation, 0.5 mg/ml of MTT solution were added to each well and incubated for 4h, and the cells were then rinsed with 1X PBS. The formazan crystals that formed were then dissolved in DMSO, and the absorbance was measured at 570 nm using a microplate reader. Cell viability was calculated as a percentage of the experimental design used as the control.

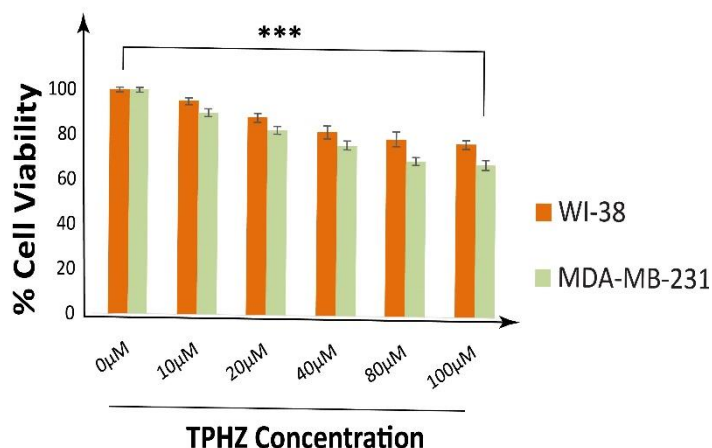


Figure S2. Cell survivability of MDA-MB 231 and WI-38 cells exposed to different ligand **TPHZ** concentration. Data are representative of at least three independent experiments and bar graph shows mean \pm SEM, * $p < 0.0001$, ** $p < 0.001$, *** $p < 0.01$ were interpreted as statistically significant, as compared with the control.

5. Calculation of Limit of detection:

From the plot of fluorescence intensity I_{399} vs concentration of ClO^- limit of detection was calculated by using the formula $\text{LOD} = k \times \delta / m$ where $k = 3$, δ is the standard deviation of the blank solution (4484.72) and m is the slope of the calibration curve.

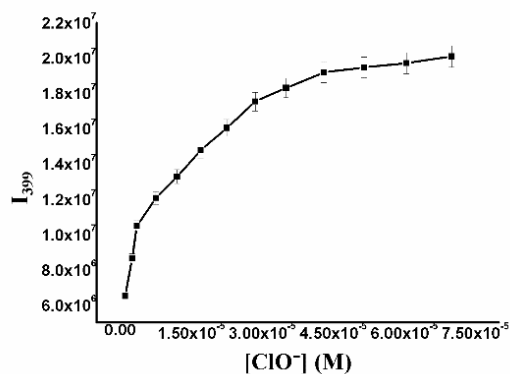


Figure S3. Plot of fluorescence intensity vs concentration of ClO^-

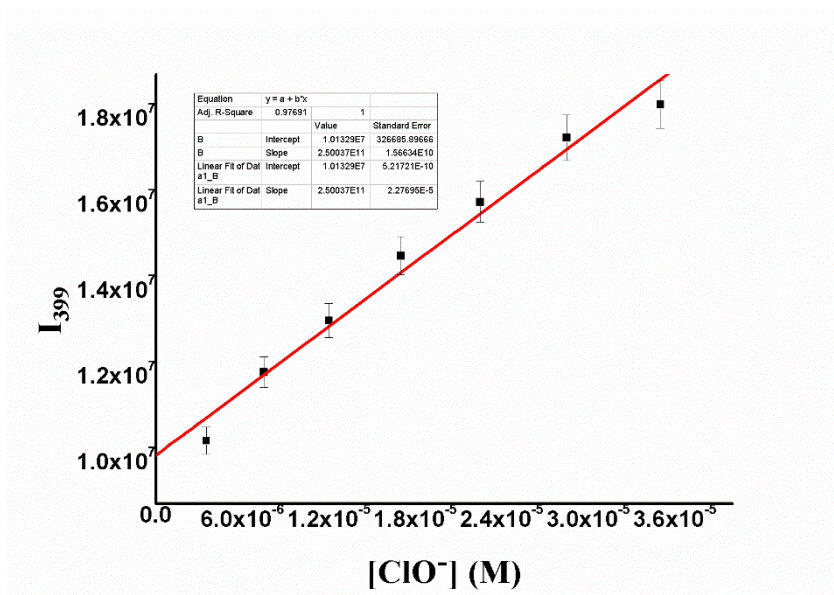


Figure S4. Calibration of the probe at an intensity I_{399} depending on ClO^- concentration.

LOD= 53.8 nM ($R^2=0.977$)

6.pH effect:

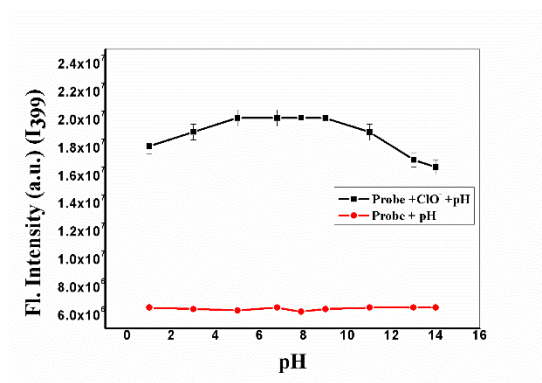


Figure S5. Effect of pH on fluorescence of TPHZ and TPHZ+ ClO^- in THF/ H_2O ($\lambda_{\text{ex}}=339\text{nm}$)

7. Application of the probe in commercial bleach and water samples:

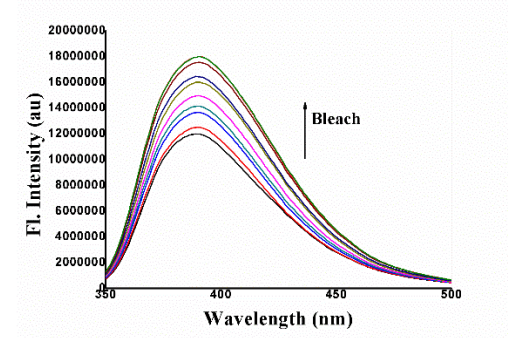


Figure S6: Fluorescence intensity changes of TPHZ upon gradual addition of bleach in THF-water (2:8 v/v) ($\lambda_{em}=399nm$)

Table S3: Water sample study for TPHZ

| Water sample | Spiked (μM) ClO^- | Found (μM) | % Recovery | RSD (%) |
|--------------|----------------------------|-----------------------|------------|---------|
| Tap water | 15 | 14.79 (± 0.33) | 98.6 | 2.2 |
| | 30 | 29.82 (± 0.51) | 99.4 | 1.7 |
| River water | 15 | 15.045 (± 0.36) | 100.3 | 2.4 |
| | 30 | 29.25 (± 0.58) | 97.5 | 2.0 |

8. Calculation of first order rate constant (k'):

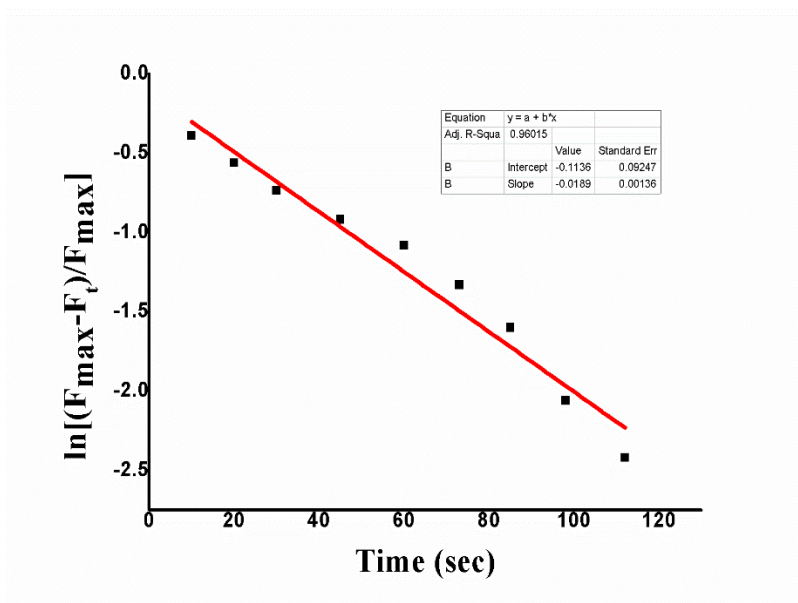


Figure S7. First order kinetic plot of probe ($1 \times 10^{-5} M$) in the presence of $1 \times 10^{-4} M$ ClO^- solution ($\lambda_{em}=399nm$)

First order rate constant $k' = 0.0189 s^{-1}$

9. Emission spectra of probe:

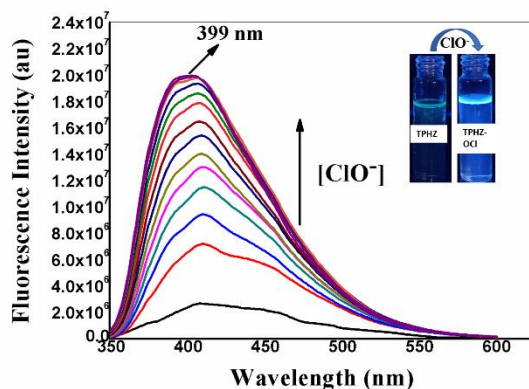


Figure S8. Fluorescence intensity changes of TPHZ (1×10^{-5} M) upon gradual addition of NaOCl in **THF-water** (2:8 v/v) ($\lambda_{em}=399$ nm).

10. Job's plot of the probe TPHZ for ClO⁻

Job's plots were drawn by plotting $\Delta F \cdot X(\text{host})$ vs $X(\text{host})$ (ΔF = change of intensity of the emission spectrum [I_{399}] for TPHZ during titration and $X(\text{host})$ is the mole fraction of the host in each case respectively).

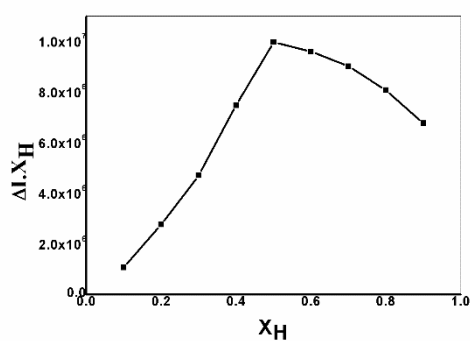
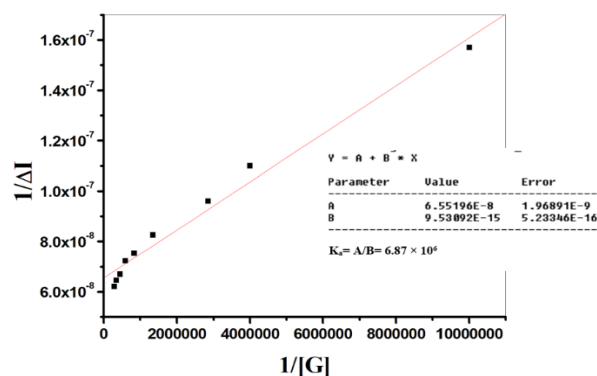


Figure S9. Job's plot of TPHZ with ClO⁻ using fluorescence data

11. Determination of binding constant value (K_a) using linear method for TPHZ

Binding constant value (K_a) was calculated by plotting $1/\Delta I$ vs $1/[G]$ [(ΔI = change of intensity of the emission spectrum at 399nm for TPHZ during titration and $[G]$ is the concentration of ClO^- in each case respectively).



Binding constant $K_a(A/B) = 6.87 \times 10^6$

Figure S10. Binding constant value of TPHZ with ClO^- using fluorescence data

13. NMR spectra: ^1H -NMR, ^{13}C -NMR

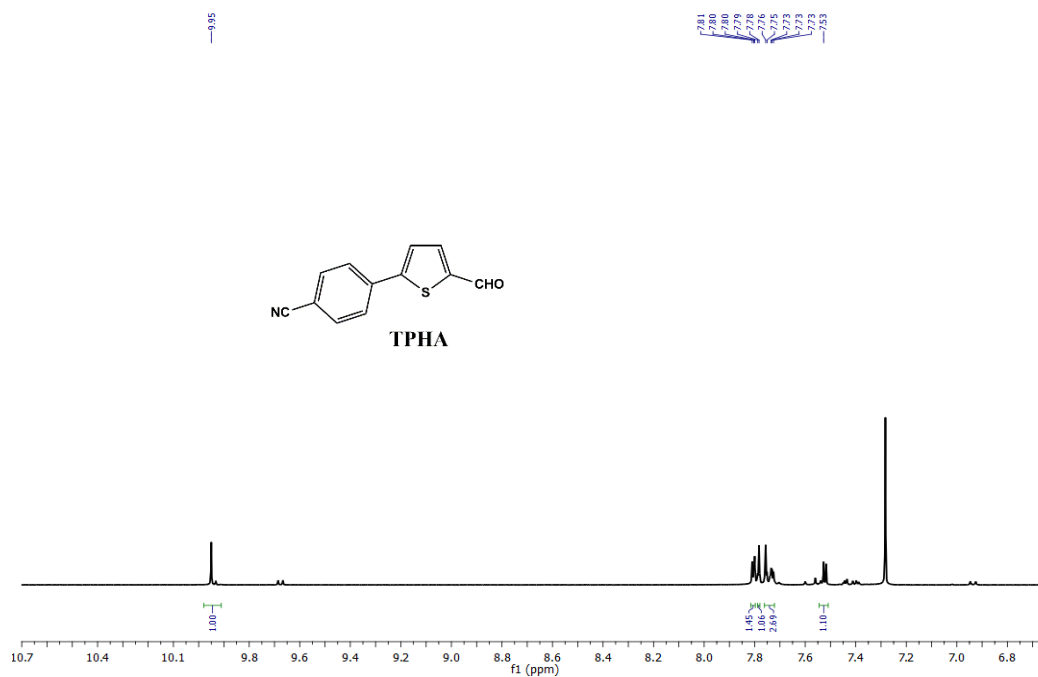


Figure S11: ^1H -NMR spectra of TPHZ in CDCl_3

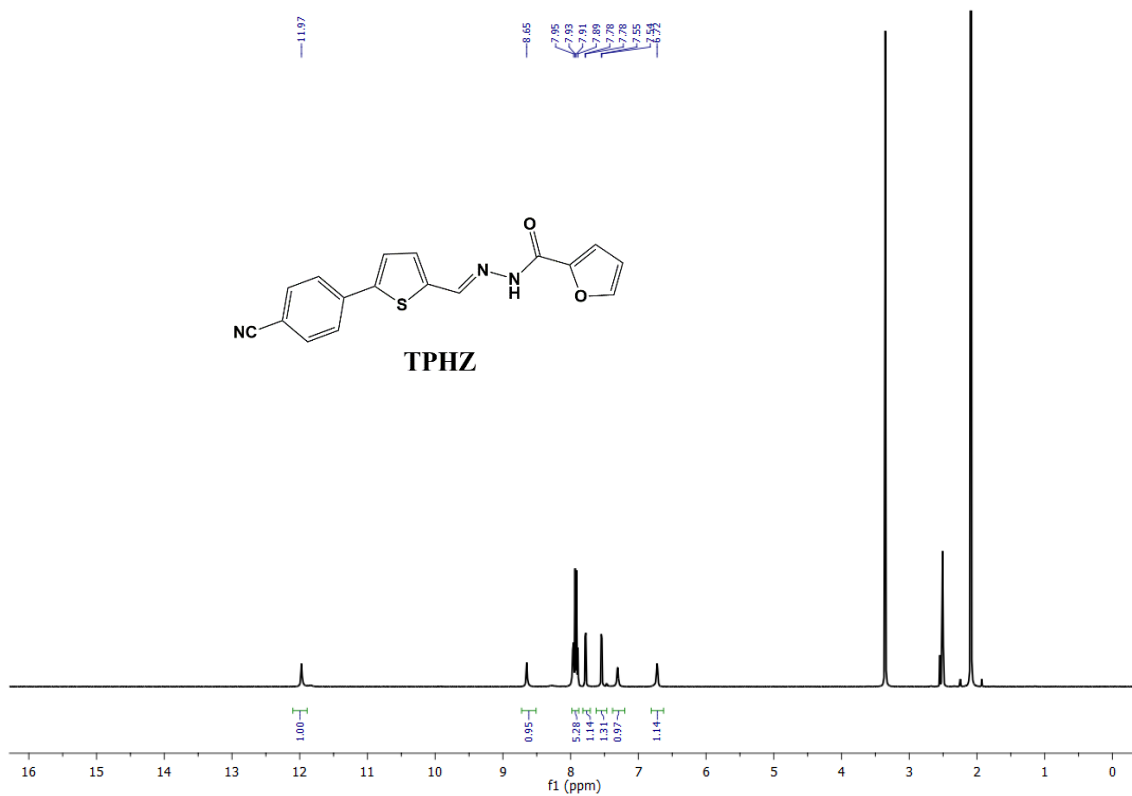


Figure S12: $^1\text{H-NMR}$ spectra of TPHZ in DMSO-d_6

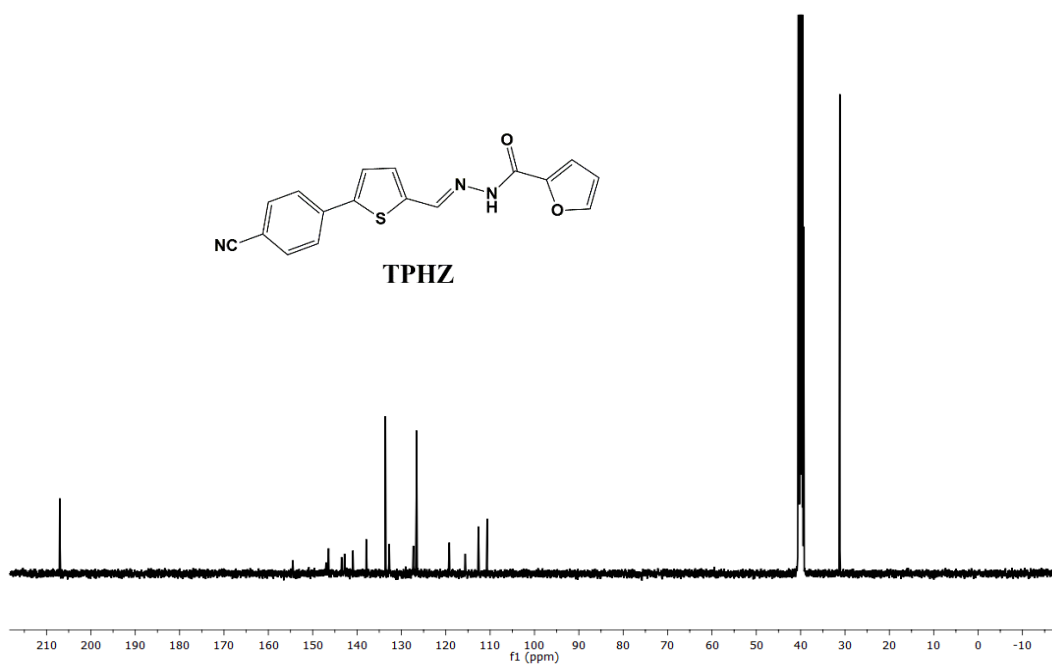


Figure S13 :¹³C-NMR spectra of TPHZ in DMSO-d⁶

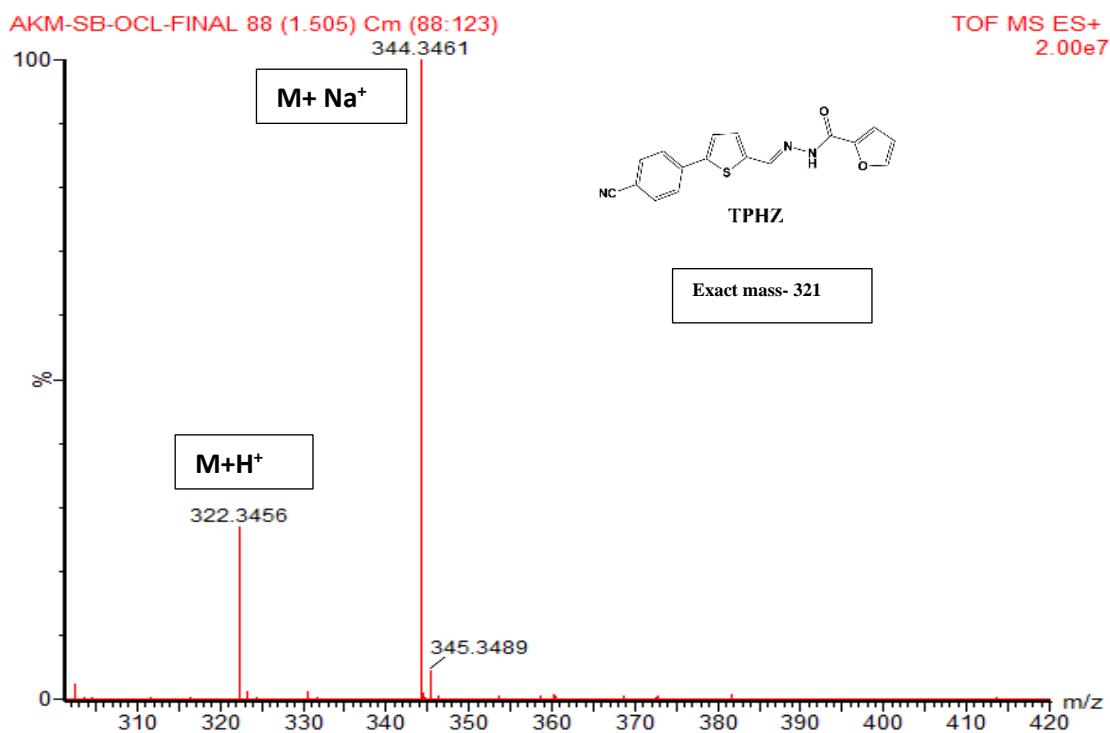


Figure S14 : ESI-MS of probe TPHZ

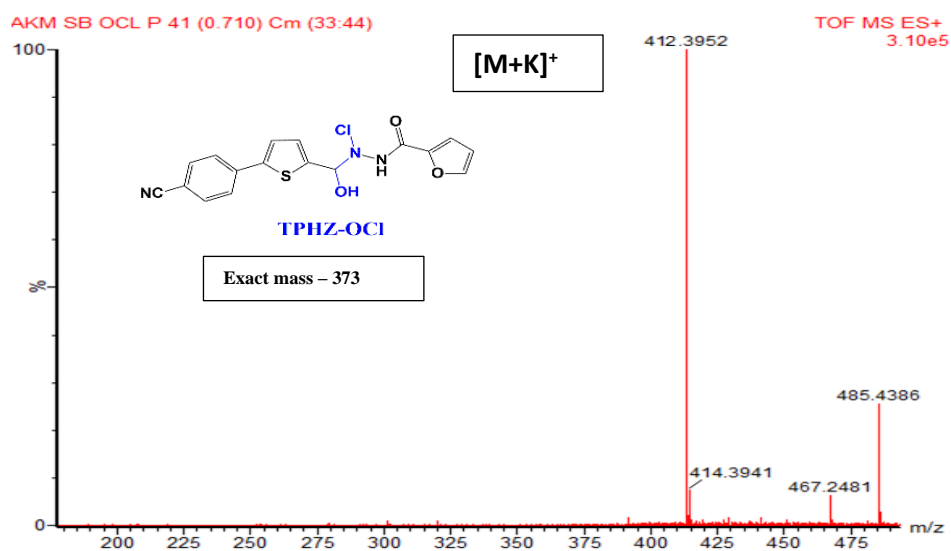


Figure S15: ESI-MS of product TPHZ-OCI

Reference:

- 1) P. R. Twentyman and M. Luscombe, *Br. J. Cancer*, 1987, **56**, 279–285.