

Supporting information

Solvent Directed Morphogenesis of Peptidic-Benzimidazolium Dipodal Receptor: Ratiometric Detection and Catalytic Degradation of Ochratoxin A

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Chemical structure of Dipodal receptor (SS4)

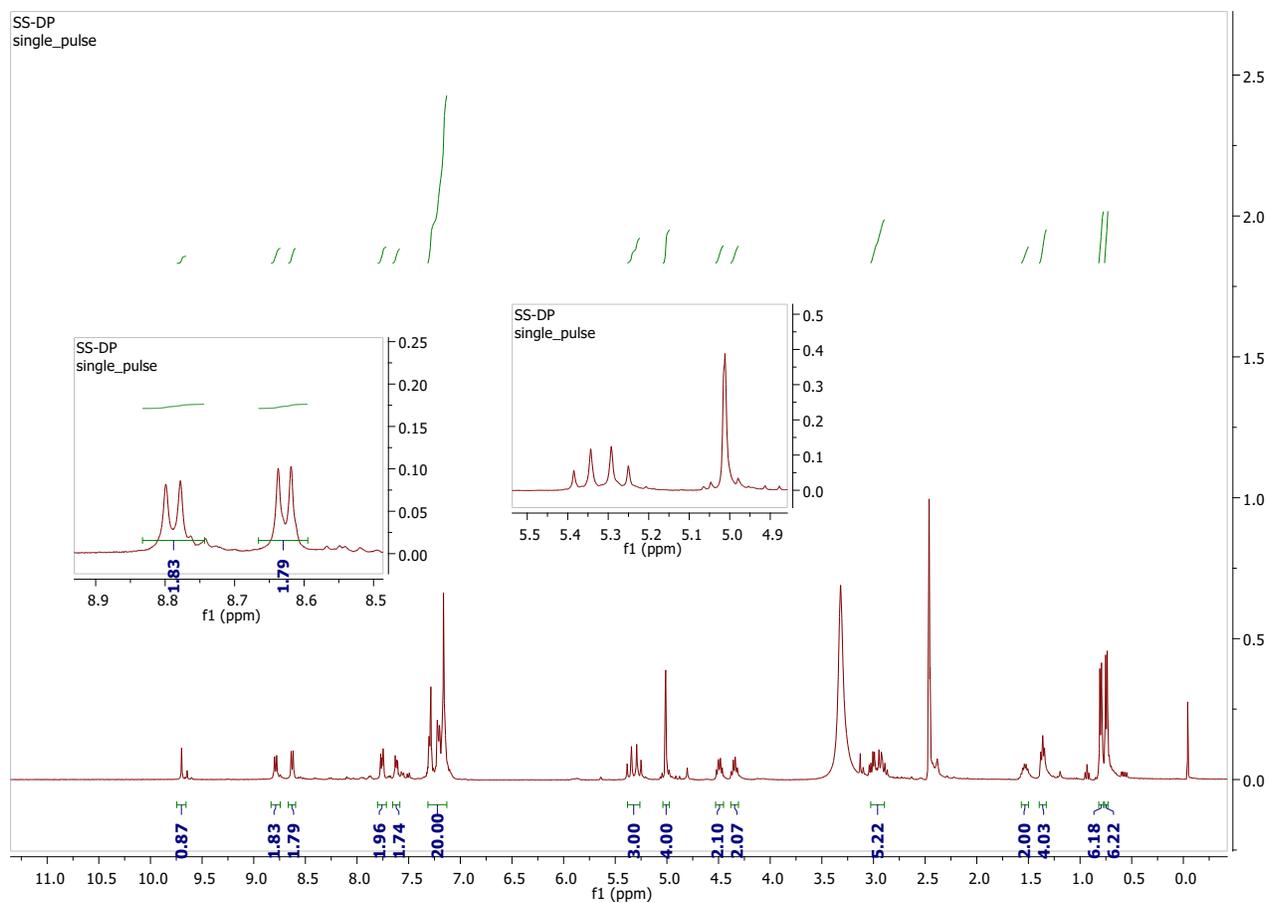
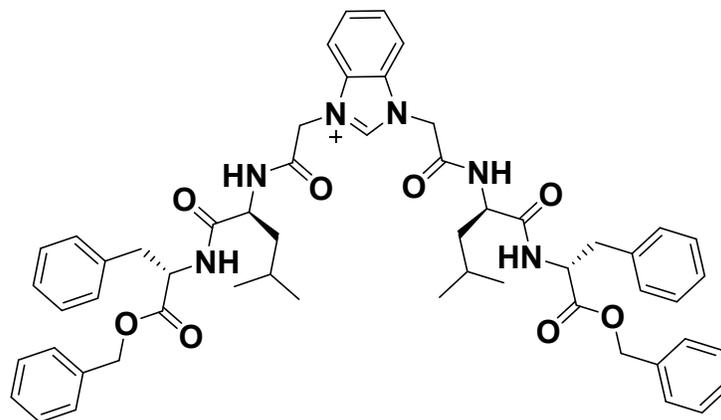


Figure SF1: ¹H NMR spectrum of peptidic-benzimidazole dipodal receptor (SS4).

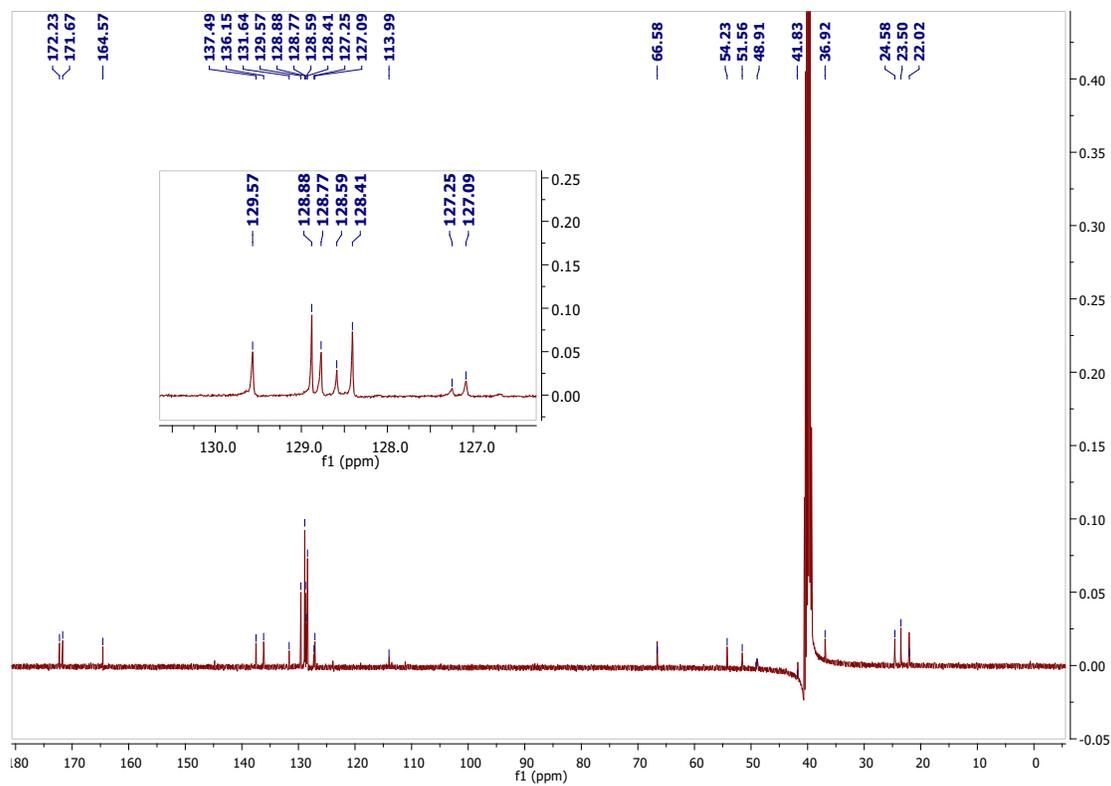


Figure SF2: ^{13}C NMR spectrum of peptidic-benzimidazole dipodal receptor (SS4).

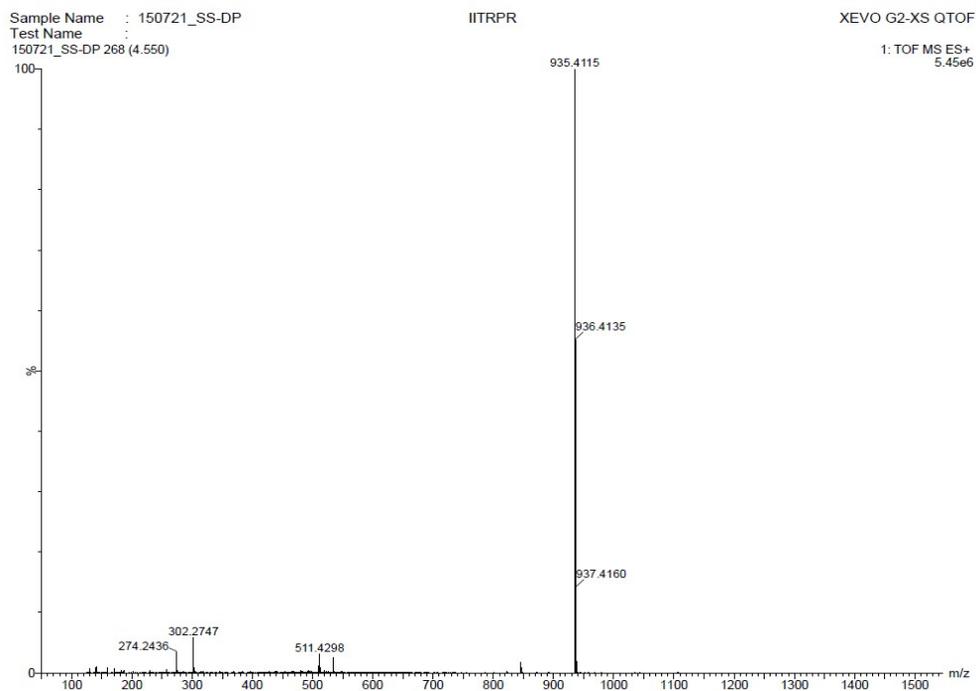


Figure SF3: Mass data of peptidic-benzimidazole dipodal receptor (SS4).

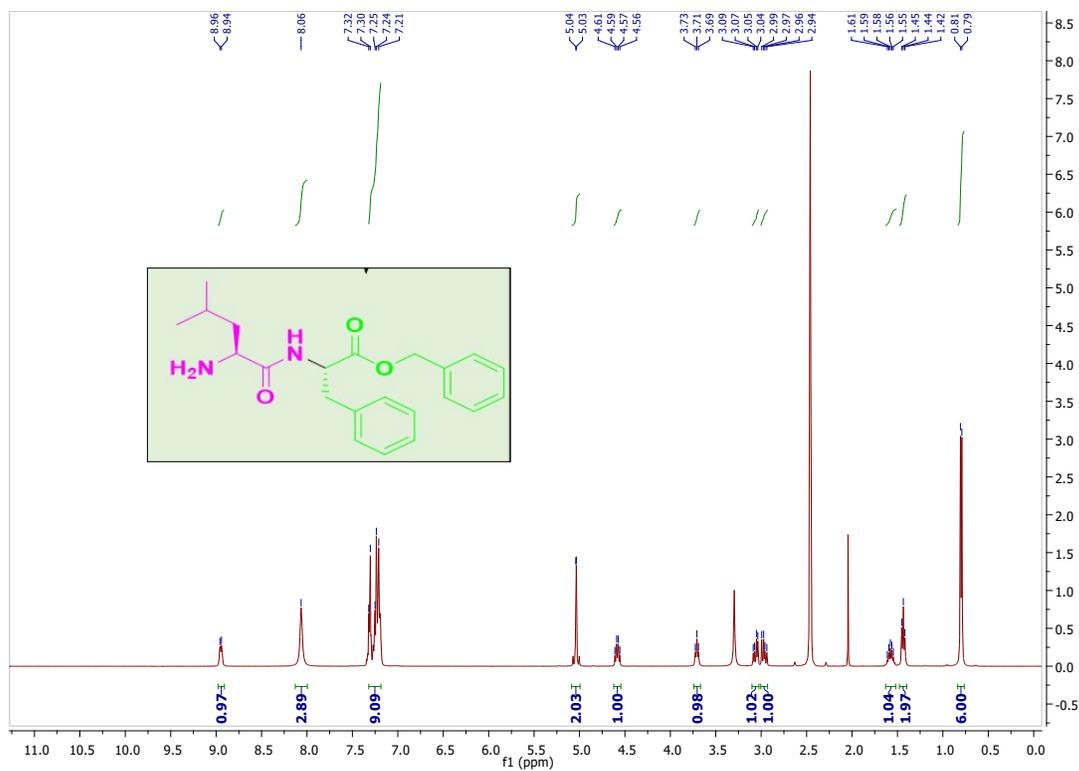


Figure SF4: ^1H NMR data of compound (2).

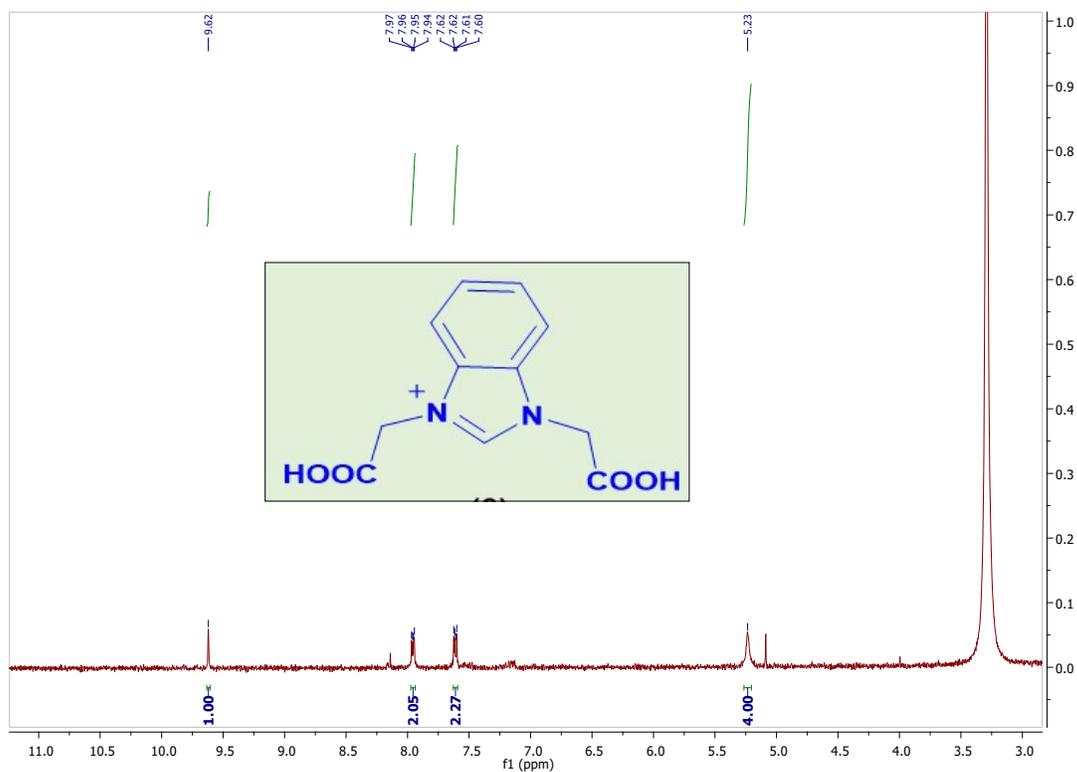
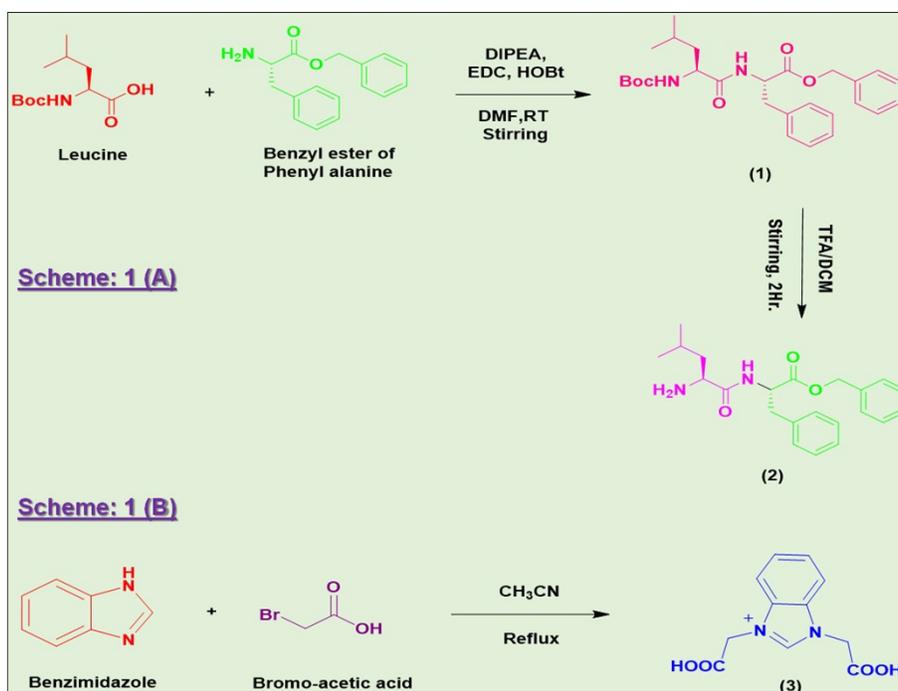


Figure SF5: ^1H NMR data of compound (3).



Scheme S1 (A) and (B): Reaction scheme for the preparation of dipeptide (2) and di-acid (3), respectively.

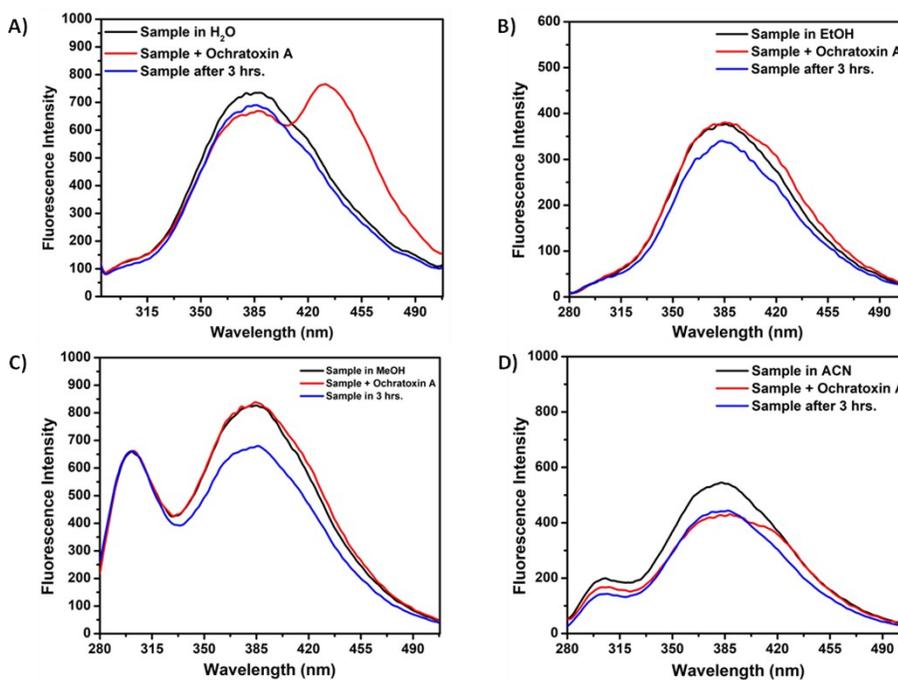


Figure SF6: A) Fluorescence emission profile of sample 4 on addition of Ochratoxin A. **B)** Fluorescence emission profile of sample 2 on addition of Ochratoxin A. **C)** Fluorescence emission profile of sample 1 on addition of Ochratoxin A. **D)** Fluorescence emission profile of sample 3 on addition of Ochratoxin A.

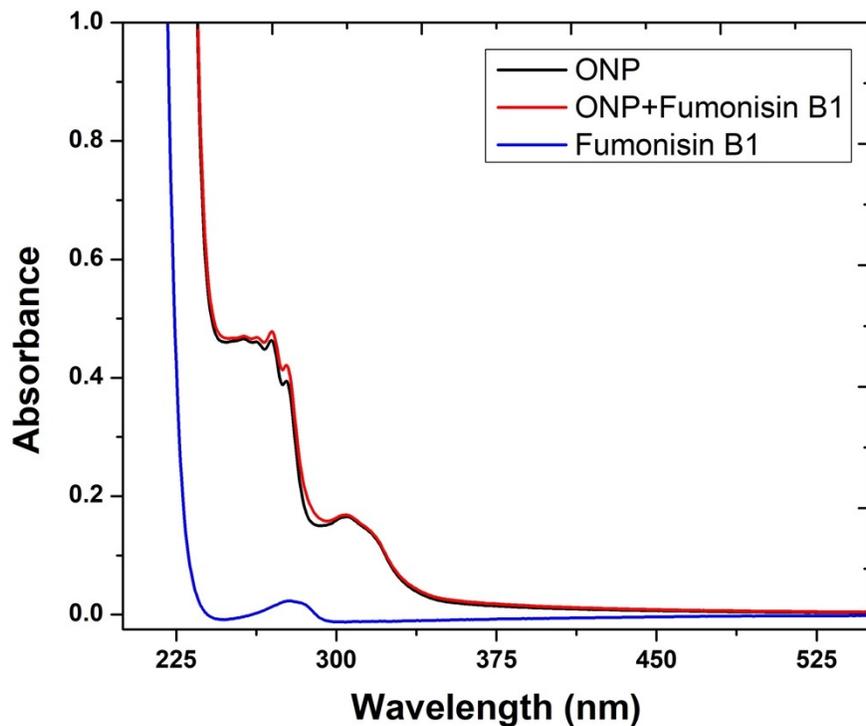


Figure SF7: UV-Visible absorption study of SS4 (ONPs) with Fumonisin B1.

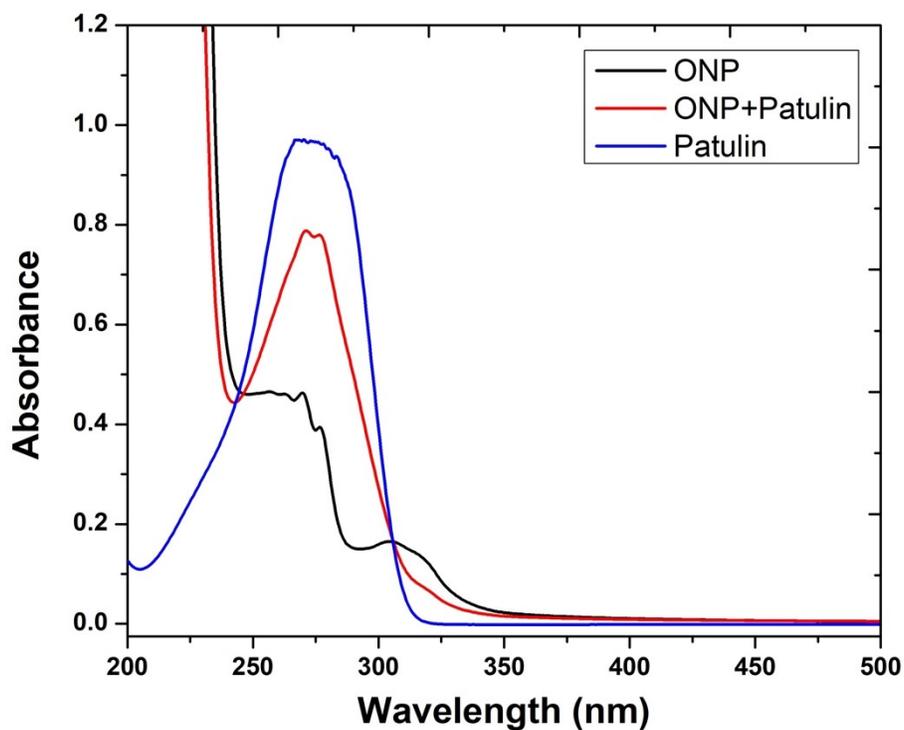


Figure SF8: UV-Visible absorption study of SS4 (ONPs) with Patulin.

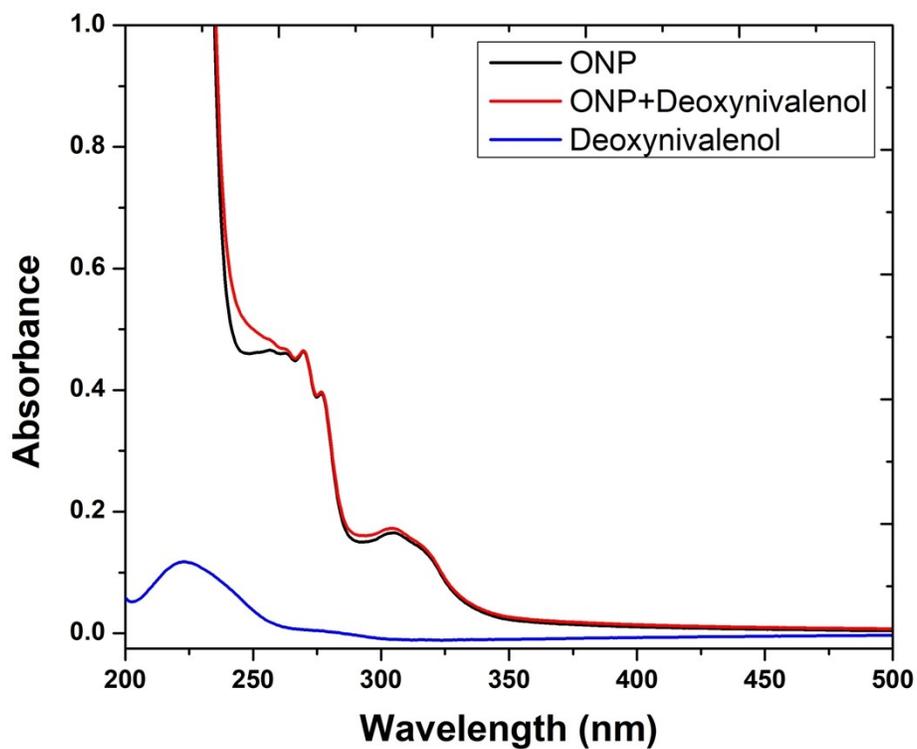


Figure SF9: UV-Visible absorption study of SS4 (ONPs) with Deoxynivalenol.

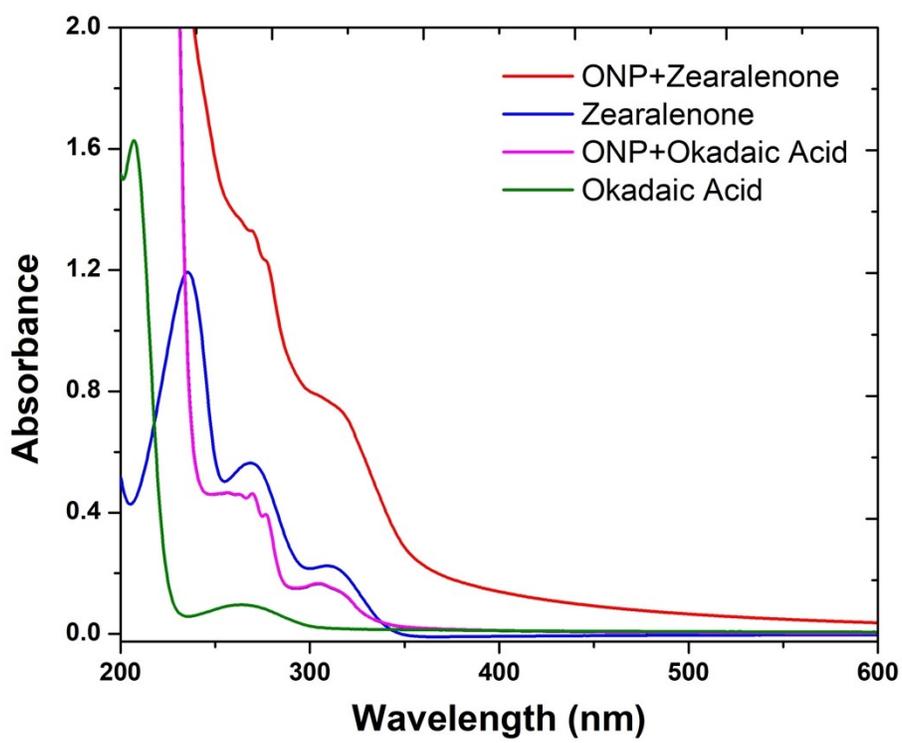


Figure SF10: UV-Visible absorption study of SS4 (ONPs) with Zearalenone and Okadaic acid.

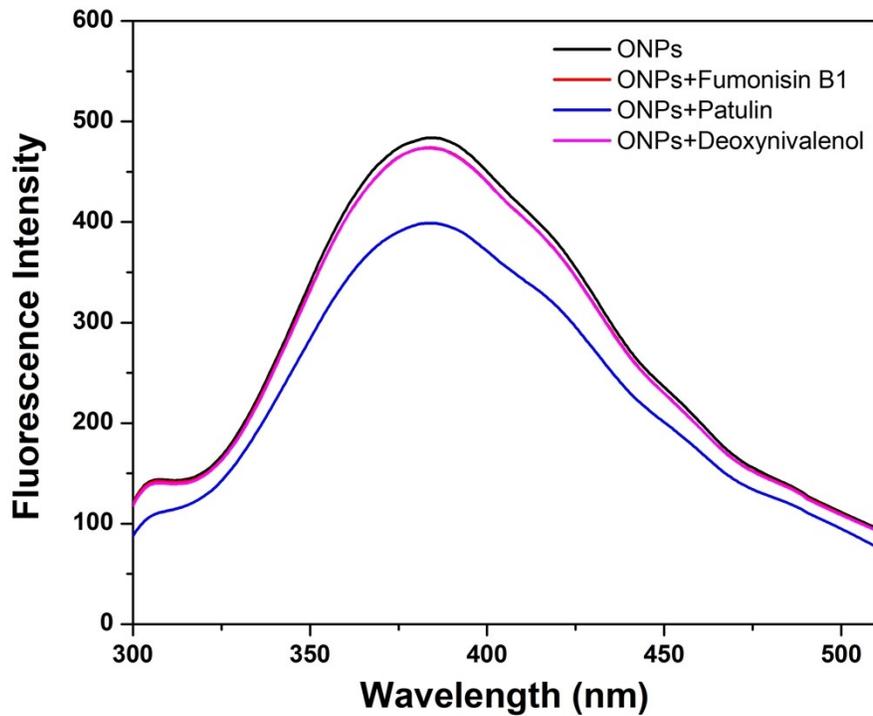


Figure SF11: Fluorescence emission profile of SS4 (ONPs) on addition of various toxins.

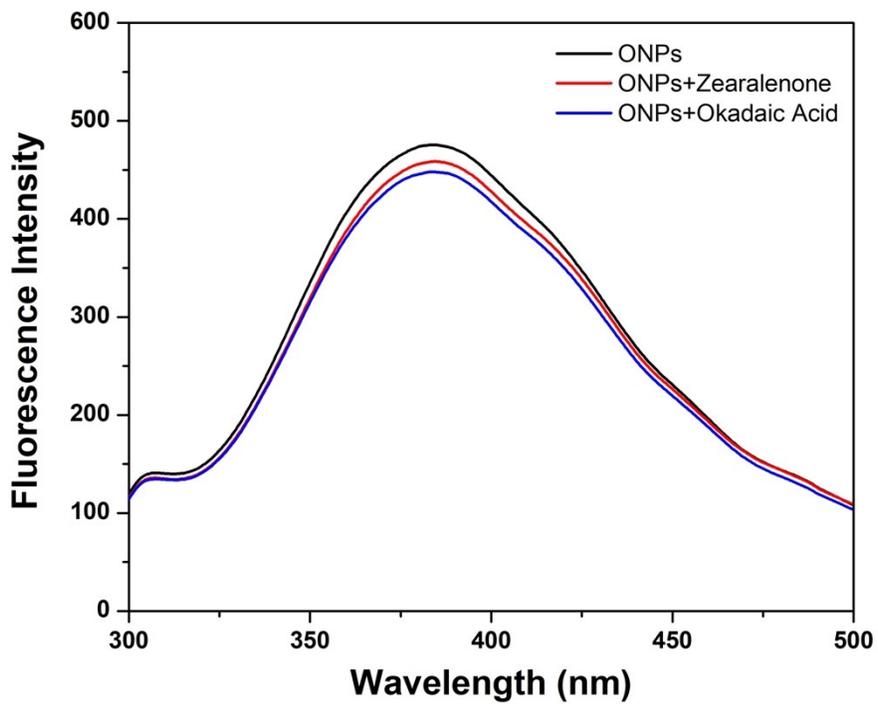


Figure SF12: Fluorescence emission profile of SS4 (ONPs) on addition of Zearalenone and Okadaic acid.

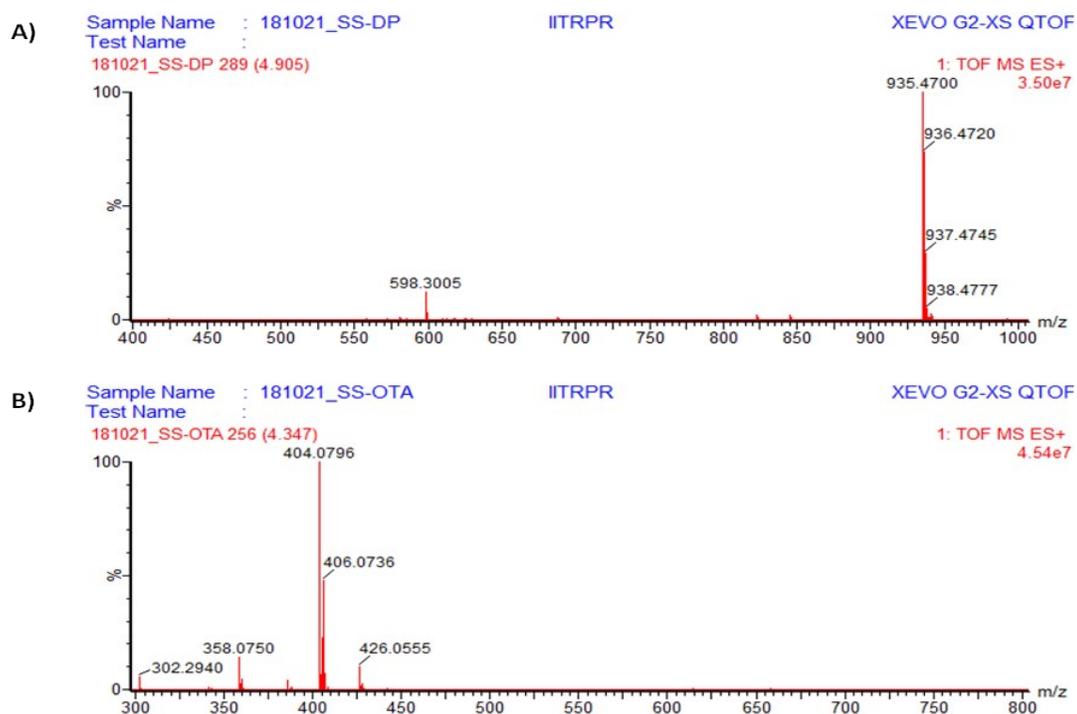


Figure SF13: LCMS data showing m/z value of dipodal receptor (SS4) and ochratoxin A.

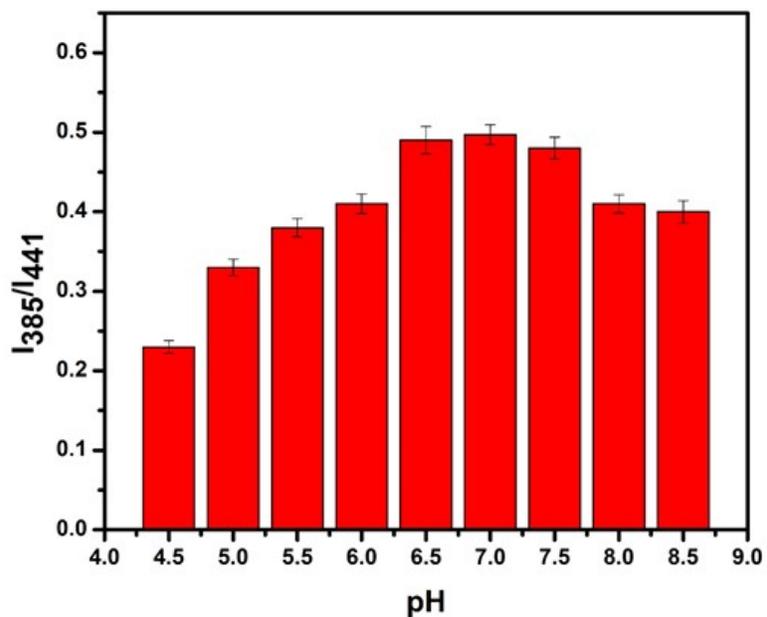


Figure SF14: Relative fluorescence intensity of sensing probe at different pH

The binding constant was calculated using Benesi-Hildebrand Equation stated below.

$$1/(A-A_0) = 1/ \{K (A_{\max} - A_0) [OTA]\} + 1/ [A_{\max} - A_0]$$

Here A_0 is the absorbance of receptor (SS4) in the absence of OTA, A is the absorbance recorded in the presence of OTA, A_{\max} is absorbance in presence of added $[OTA]_{\max}$ and K_a is the association constant. The association constant (K_a) could be determined from the slope of the straight line of the plot of $1/(A-A_0)$ against $1/[OTA]$ and is found to be $8.59 \times 10^4 \text{ M}^{-1}$

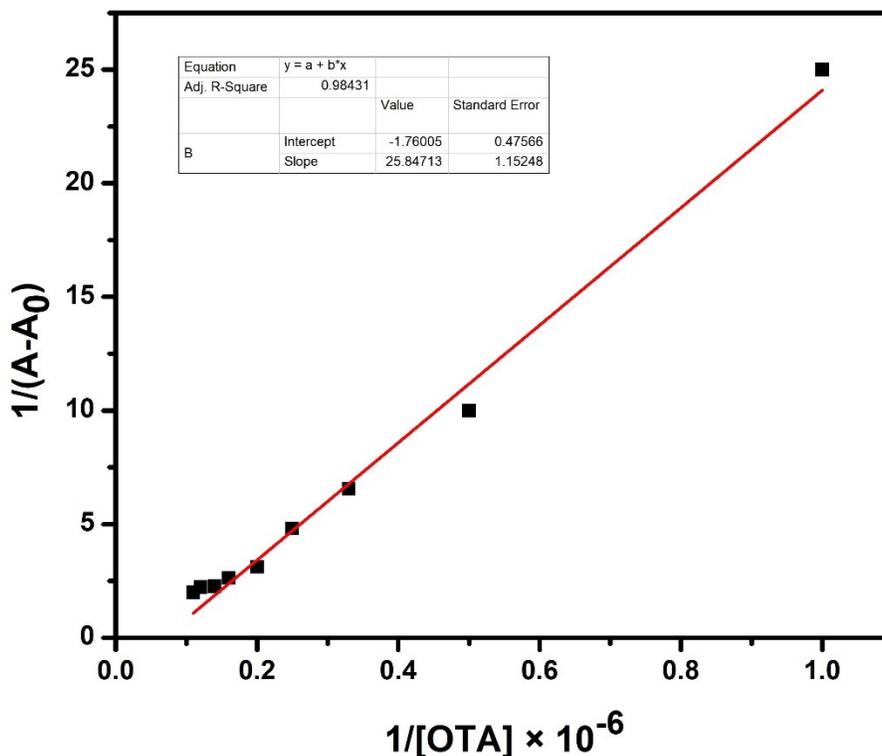


Figure SF15: Benesi–Hildebrand plot from UV/vis titration data of SS4 (50 μM) with OTA.

The following equation was used to calculate the quantum yields by comparison with fluorescein ($\Phi_s = 0.97$ in basic ethanol) as a reference.

$$\Phi_X = \Phi_S \times \left(\frac{I_X}{I_S}\right) \times \left(\frac{A_S}{A_X}\right) \times \left(\frac{n_X}{n_S}\right)^2$$

where A is the absorbance, n is the solvent's refractive index, Φ is the quantum yield, I is the integrated area under the fluorescence emission curve, and x and s , respectively, denote the unknown and standard solution.

Using the aforementioned equation, we were able to compute the quantum yields of SS4 and SS4-OTA, which come out to be 0.51 and 0.53 respectively.

Table S1: Ochratoxin A recovery experiment in spiked samples.

| S. No. | Sample | Spiked concentration | Found concentration | Recovery (%) | RSD |
|--------|-------------------------------------------------------------------------------------|----------------------|---------------------|--------------|------|
| 1. |  | 1.5 μM | 1.54 μM | 102.6 | 3.21 |
| | | 2.0 μM | 1.96 μM | 98 | 2.75 |
| | | 2.5 μM | 2.47 μM | 98.8 | 1.86 |
| 2. |  | 1.5 μM | 1.48 μM | 98.6 | 2.38 |
| | | 2.0 μM | 1.97 μM | 98.5 | 3.54 |
| | | 2.5 μM | 2.54 μM | 101.6 | 1.73 |

Table S2: Literature comparison with present work.

| S. No. | Type of Method | | Medium | LOD | References |
|--------|------------------------------------------|-------------------------------------------------------------------|------------------------------------------------------------------|------------------------|---------------------------------------------------|
| | Detection | Degradation | | | |
| 1. | Fluorescence based | × | Binding buffer (pH – 8.5) | 17.2 nM | Food Control 60 (2016) 296-301. |
| 2. | Colorimetric method | × | Buffer solution | 5.0 nM | Journal of Hazardous Materials 388 (2020) 1217583 |
| 3. | Colorimetric ELISA | × | Bicarbonate buffer pH (8.6) & PBS buffer (7.4) | 40 pg/mL | Sensors and Actuators B 262 (2018) 102–109 |
| 4. | Fluorescence based ratiometric detection | Catalytic degradation by peptidic-benzimidazole dipodal receptor | Aqueous medium | 2.1×10^{-7} M | Present work |
| 5. | HPLC | By hydrolytic action of <i>Alcaligenes faecalis</i> | LB medium | NA | Journal of Applied Microbiology 123, 661 – 668 |
| 6. | LCMS | By lactic acid bacteria | MRS medium | NA | Food and Chemical Toxicology 112 (2018) 60 - 66 |
| 7. | HPLC | commercial peroxidase (POD) enzyme (<i>Armoracia rusticana</i>) | Potassium phosphate buffer, water, H ₂ O ₂ | NA | Food research international 131 (2020) 109039 |