Supplementary Information

A highly sensitive fluorescence probe for on-site detection of nerve agent mimic diethylchlorophosphonate DCP

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1. Experimental details

1.1. Synthesis of compound 1a

4-Bromo-1,8-naphthalic anhydride (554 mg, 2 mmol) and 2-nitrobenzene-1,4diamine (306 mg, 2 mmol) were placed in a 50 ml round bottom flask. The reaction was heated to 80 °C under nitrogen atmosphere for 6 h. After completion of the reaction, the reaction mixture was cooled to room temperature and a yellow solid was gradually deposited within the solution, filtered in vacuum and washed 2-3 times with anhydrous ethanol. After drying, compound 1a was obtained (774 mg, 91% yield).

1.2. Synthesis of NZOH

4-bromo-1, 8-naphthalenedicarboxylic anhydride (1 g, 4 mmol) and 2-aminophenol (0.4 g, 4 mmol) were dissolved in 20 mL of ethanol under a nitrogen atmosphere. After heating at reflux for 8h at 80°C and cooling, a white solid precipitate was gradually formed in the reaction system. The precipitate was finally filtered out, washed three times with anhydrous ethanol and dried to obtain the compound 2 (1.1 g, 75%).

¹H NMR (500 MHz, DMSO-*d*₆) δ 9.68 (s, 1H), 8.59 (ddd, J = 18.9, 7.8, 1.1 Hz, 2H), 8.41 (d, J = 7.9 Hz, 1H), 8.07 – 7.98 (m, 2H), 7.34 – 7.28 (m, 1H), 6.88 (ddd, J = 8.3, 2.3, 1.1 Hz, 1H), 6.80 (dt, J = 6.3, 1.9 Hz, 2H).

The compound 2 (400 mg, 1.13 mmol) and n-butylamine (1 mL, 5 mmol) were added into 10 mL 2-methoxyethanol under a nitrogen atmosphere, and the mixture was magnetic stirred and refluxed at 130°C for 8 h. After the reaction, the resulting mixture was subjected to chloroform extraction, leading to gradual formation of an orange solid precipitate within the reaction system. Finally, the precipitate was filtered out, washed three times with anhydrous ethanol and dried to obtain the orange NZOH powder (268 mg, 70%).

¹H NMR (500 MHz, DMSO-*d*₆) δ 9.6 (s, 1H), 8.7 (dd, J = 8.6, 1.2 Hz, 1H), 8.4 (dd, J = 7.3, 1.0 Hz, 1H), 8.3 (d, J = 8.6 Hz, 1H), 7.8 (t, J = 5.5 Hz, 1H), 7.7 (dd, J = 8.4, 7.3 Hz, 1H), 7.3 (t, J = 8.0 Hz, 1H), 6.9 – 6.8 (m, 2H), 6.7 – 6.7 (m, 2H), 3.4 (d, J = 7.1 Hz, 2H), 1.8 – 1.7 (m, 2H), 1.5 (dt, J = 14.8, 7.4 Hz, 2H), 1.0 (t, J = 7.4 Hz, 3H). HR-MS (ESI, m/z): $[M+H]^+$ calcd for C₂₂H₂₀N₂O₃: 361.1507; found: 361.1542.

1.3. Synthesis of NZN

4-Bromo-1, 8-naphthalic anhydride (1 g, 4 mmol) and isophthalamide (0.4 g, 4 mmol) were dissolved in 20 mL ethanol under a nitrogen atmosphere. Subsequently, the mixture was magnetic stirred and refluxed at 80°C for 6 h. After cooling, a dark yellow solid precipitate appeared in the reaction system. The precipitate was obtained through filtration and dried by rotary evaporation to remove the residual solvent. The crude product was purified by column chromatography (eluent: dichloromethane/methanol = 10/1) to obtain compound 3 (548 mg, 61%).

¹H NMR (500 MHz, DMSO-*d*₆) δ 8.61 (dd, J = 8.4, 1.2 Hz, 1H), 8.57 (dd, J = 7.3, 1.2 Hz, 1H), 8.42 (d, J = 7.9 Hz, 1H), 8.08 – 7.99 (m, 2H), 7.14 (t, J = 7.9 Hz, 1H), 6.65 (ddd, J = 8.1, 2.3, 1.0 Hz, 1H), 6.54 – 6.47 (m, 2H), 5.22 (s, 2H).

The compound 3 (500 mg, 1.13 mmol) and n-butylamine (1 mL, 1 mmol) were added into 10 mL 2-methoxyethanol under a nitrogen atmosphere nitrogen. The mixture was magnetic stirred and refluxed at 130°C for 8 h. After the reaction, the resulting mixture was extracted with chloroform and then subjected to rotary evaporation to give the crude product of NZN. Finally, the crude product was purified by column chromatography (eluent: dichloromethane/methanol = 10/1) yielding pure dark orange NZN powder (300 mg, 70%).

¹H NMR (500 MHz, DMSO- d_6) δ 8.79 (dd, J = 8.4, 1.2 Hz, 1H), 8.56 (dd, J = 7.3, 1.2 Hz, 1H), 8.40 (d, J = 8.3 Hz, 1H), 7.91 (dd, J = 8.4, 7.3 Hz, 1H), 7.08 (ddd, J = 8.4, 7.3, 1.4 Hz, 1H), 7.03 (dd, J = 8.0, 1.4 Hz, 1H), 6.92 (dd, J = 8.1, 1.6 Hz, 1H), 6.81 (d, J = 8.3 Hz, 1H), 6.66 (td, J = 7.6, 1.6 Hz, 1H), 5.15 (s, 2H), 4.04 - 3.98 (m, 2H), 1.71 - 1.60 (m, 2H), 1.50 - 1.38 (m, 2H), 0.92 (t, J = 7.4 Hz, 3H).

HR-MS (ESI, m/z): $[M+H]^+$ calcd for $C_{22}H_{21}N_3O_2$: 360.1667; found: 361.1702.

1.4 Determination of the detection limit for NZNN test strip

The calibration curve was obtained by plotting the G values of the NZNN strips as a function of DCP level, and a regression curve equation was obtained.

Detection limit (LOD) = $3 \times \sigma/k$

where k is the slope of the curve equation, and σ represents the standard deviation for the G values of the NZNN strips in the absence of DCP.



Figure S1.(a) Synthetic route of NZNN. (b) Synthetic route of NZOH. (c) Synthetic route of NZN.



Figure S2. (a) UV spectral changes of NZOH (10 μ M) in chloroform. (b) UV spectral changes of NZN (10 μ M) in chloroform.



Figure S3. (a) UV-visible spectral response of probe NZNN (10 μ M) in chloroform with gradual addition of DCP in the concentration range of 0-18 μ M. (b) The UV absorption intensity of NZNN was linearly correlated with the concentration of DCP at an absorption wavelength of 435 nm.



Figure S4. Fluorescence spectra changes of NZNN (10 μ M) in the presence of Phosgene (1.2 mM) in chloroform. $\lambda_{ex} = 360$ nm, slits: 2/2 nm.



Figure S5. ¹H NMR spectrum of compound 1b in DMSO-*d*₆ (500 MHz).



Figure S6. ¹³C NMR spectrum of compound 1b in DMSO- d_6 (500 MHz).



Figure S7. ¹H NMR spectrum of NZNN in DMSO-*d*₆ (500 MHz).



Figure S8. ¹³C NMR spectrum of NZNN in DMSO- d_6 (500 MHz).





Figure S9. ¹H NMR spectrum of compound 2 in DMSO-*d*₆ (500 MHz).



Figure S10. ¹³C NMR spectrum of compound 2 in DMSO- d_6 (500 MHz).



Figure S11. ¹H NMR spectrum of NZOH in DMSO- d_6 (500 MHz).



Figure S12. ¹³C NMR spectrum of NZOH in DMSO- d_6 (500 MHz).





Figure S13. ¹H NMR spectrum of compound 3 in DMSO-*d*₆ (500 MHz).



Figure S14. ¹³C NMR spectrum of compound 3 in DMSO- d_6 (500 MHz).



Figure S15. ¹H NMR spectrum of NZN in DMSO-*d*₆ (500 MHz).



Figure S16. ¹³C NMR spectrum of NZN in DMSO- d_6 (500 MHz).



Figure S17. HR-MS (ESI) spectrum of NZNN.



Figure S18. HR-MS (ESI) spectrum of NZOH.



Figure S19. HR-MS (ESI) spectrum of NZN.



Fig S20. Some fluorescent probes used to detect DCP.

Probe	Emission	Fluorescence	Response time	Selectivity	LOD	Paper	Application	Devitrization	Reference
		response	(5)			strip			
1	414	Turn on	60	Good	13.6 µM	No	No	No	[1]
2	455	Turn on	60	Bad	200 µM	No	No	No	[2]
3	453	Turn on	6	Good	0.186 µM	Yes	No	No	[3]
4	515	Turn on	120	Good	10 µM	Yes	No	No	[4]
5	480	Turn on	No	Bad	8.7 nM	Yes	Yes	No	[5]
6	475	Turn on	No	Bad	0.1 μΜ	Yes	Yes	No	[6]
NZNN	500	Turn on	< 6	Good	30.1 nM	Yes	Yes	Yes	This work

Table S1. Comparison of data from this study with other studies

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