

ICAM-1-decorated extracellular vesicles loaded with miR-146a and *Glut1* drive immunomodulation and hinder tumor progression in a murine model of breast cancer

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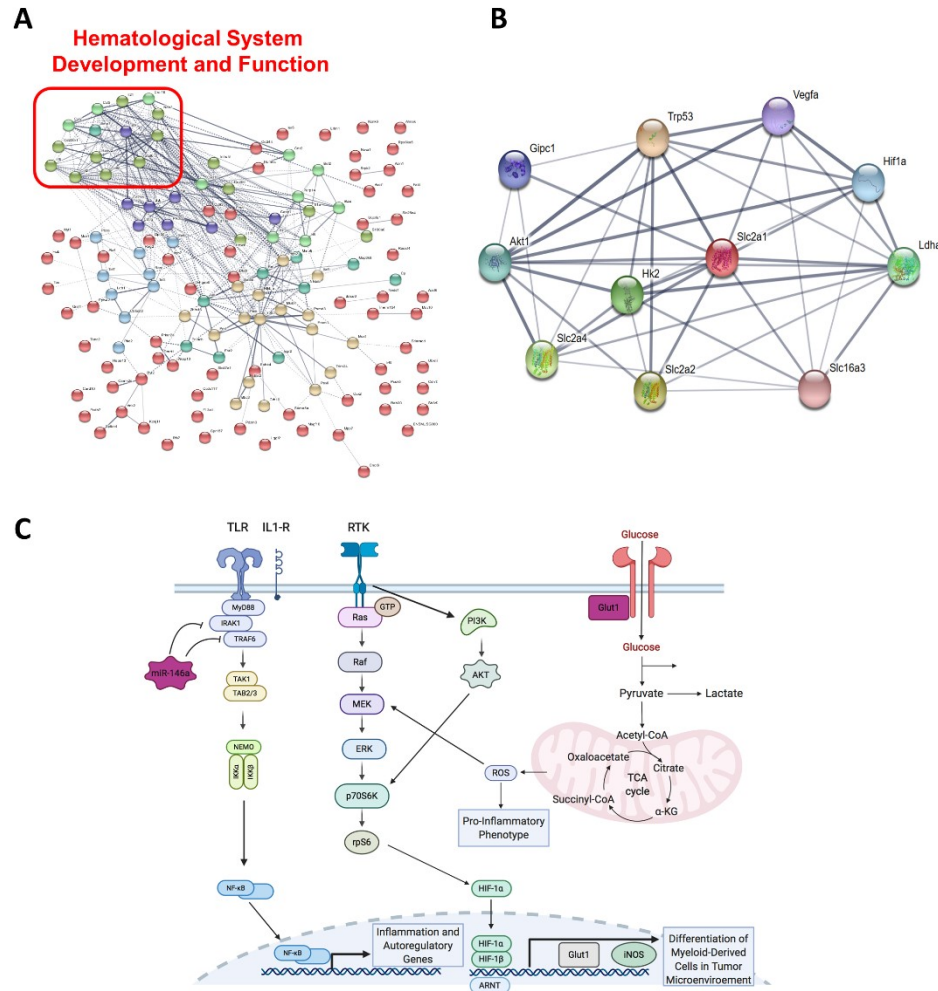
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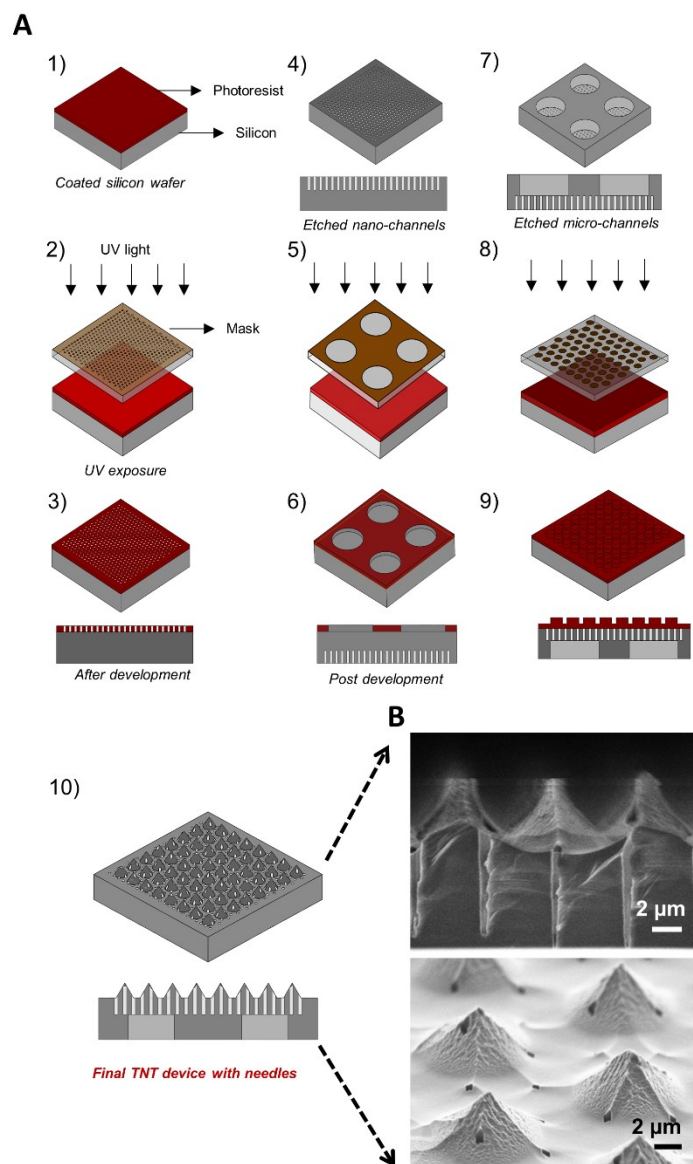
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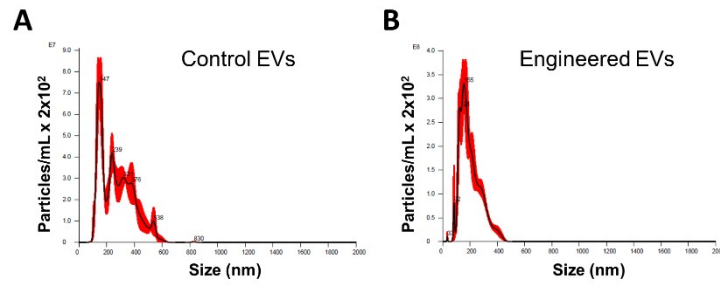
SUPPORTING INFORMATION



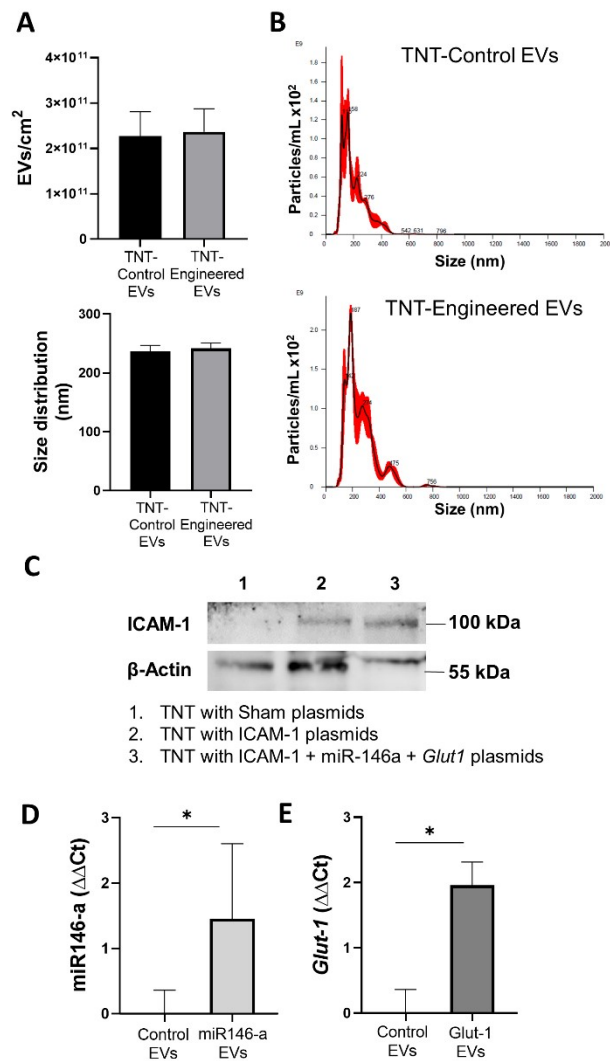
Supplementary Figure 1. *In silico* functional analysis identifying signaling pathways common to miR-146a and *Glut1*. (A) Functional interacting network of miR-146a-5p and miR-146a-3p targets leads to the identification of a subgroup of interacting genes involved in *Hematological System Development and Function*. (B) Functional interacting network of *Glut1* (Slca2a1). (C). Common factors between miR-146a and *Glut1* functional interacting networks suggest that the pro-inflammatory repolarization seen in MDSCs could potentially be partially mediated by the Hypoxia-Inducible Factor (HIF)-1 regulatory pathway.



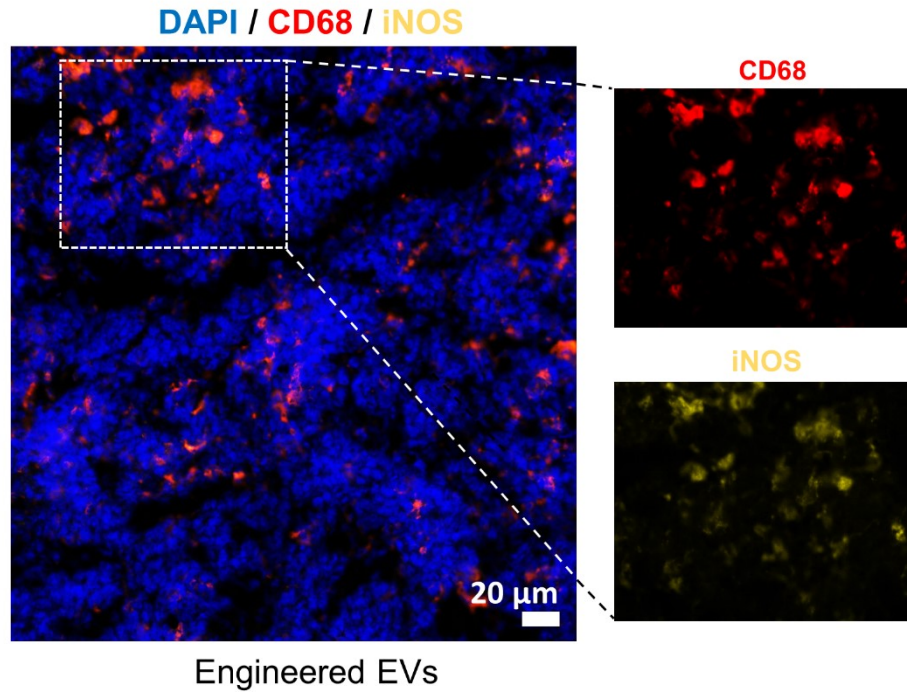
Supplementary Figure 2. TNT chip fabrication. (A) Schematic diagram showing the fabrication process for TNT devices. First, nanoscale openings are patterned and etched into the Si wafer surface using a combination of lithography and DRIE (1-4). Next, microscale reservoirs are drilled on the back side of the wafer using lithography and DRIE to gain fluidic access to the nanochannels (5-7). Lastly, microscale photoresist (PR) pillars were patterned on the nanochannel side and used as an etch mask to define the needles using SF₆, which undercuts the PR to form the needle (8-10). (B) SEM images of the cross-section and surface of the TNT device.



Supplemental Figure 3. Characterization of in vitro derived engineered EVs. NanoSight analysis showing the concentration (dilution factor 2×10^2) and size distribution of **(A)** control EVs and **(B)** engineered EVs produced and released by the ‘donor’ cells 24 hours after transfection.



Supplemental Figure 4. Characterization of skin derived EVs after TNT treatment. (A) NanoSight analysis of the amount and average size and (B) individual plots showing the particle distribution in terms of concentration (dilution factor 1X10²) and size of the EVs obtained from 1 cm² of TNT-treated skin. (C) Western blot analysis confirmed the presence of ICAM-1 protein in the ICAM-1 decorated EVs loaded with sham, and engineered EVs (ICAM-1 decorated EVs loaded with miR-146-a + *Glut1*) compared to control EVs (without ICAM-1), using β-actin as the internal control. RT-qPCR quantification of (D) miR-146a and (E) *Glut1* loading inside engineered skin EVs compared to skin-derived control skin EVs.



Supplemental Figure 5. Co-localization analysis of iNOS expression with macrophages. Representative fluorescence micrograph of tumor tissue showing the expression of the pro-inflammatory marker iNOS (yellow), the expression of the classically activated (M1-like) macrophage marker CD68 (red), and the nuclear stain DAPI (blue). Analyses of the double positive signal of iNOS and CD68 in tumor tissue revealed a co-localization of ~70%.