

Supplemental Information

Efficient clearance of periodontitis pathogens by *S. gordonii* membrane-coated H₂O₂ self-supplied nanocomposites in a “Jenga” style

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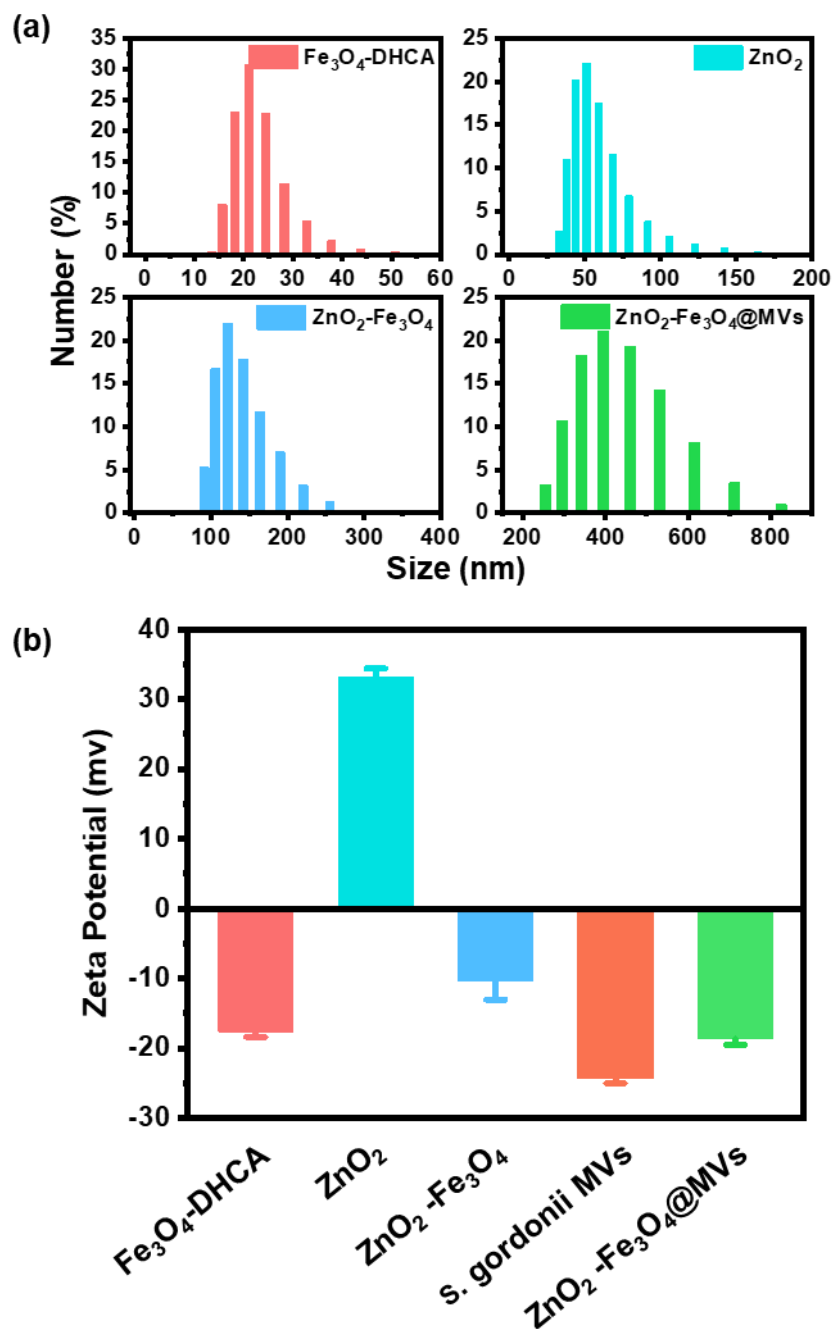


Fig. S1. DLS and zeta potential characterization of each part of the materials. (a) DLS characterization of Fe₃O₄-DHCA NPs, ZnO₂ NPs, ZnO₂-Fe₃O₄ NPs and ZnO₂-Fe₃O₄@MV NPs dispersed in water. (b) Surface zeta potentials of Fe₃O₄-DHCA NPs, ZnO₂ NPs, ZnO₂-Fe₃O₄ NPs, *S. gordonii* MVs and ZnO₂-Fe₃O₄@MV NPs. Error bar: standard deviation (n = 3).

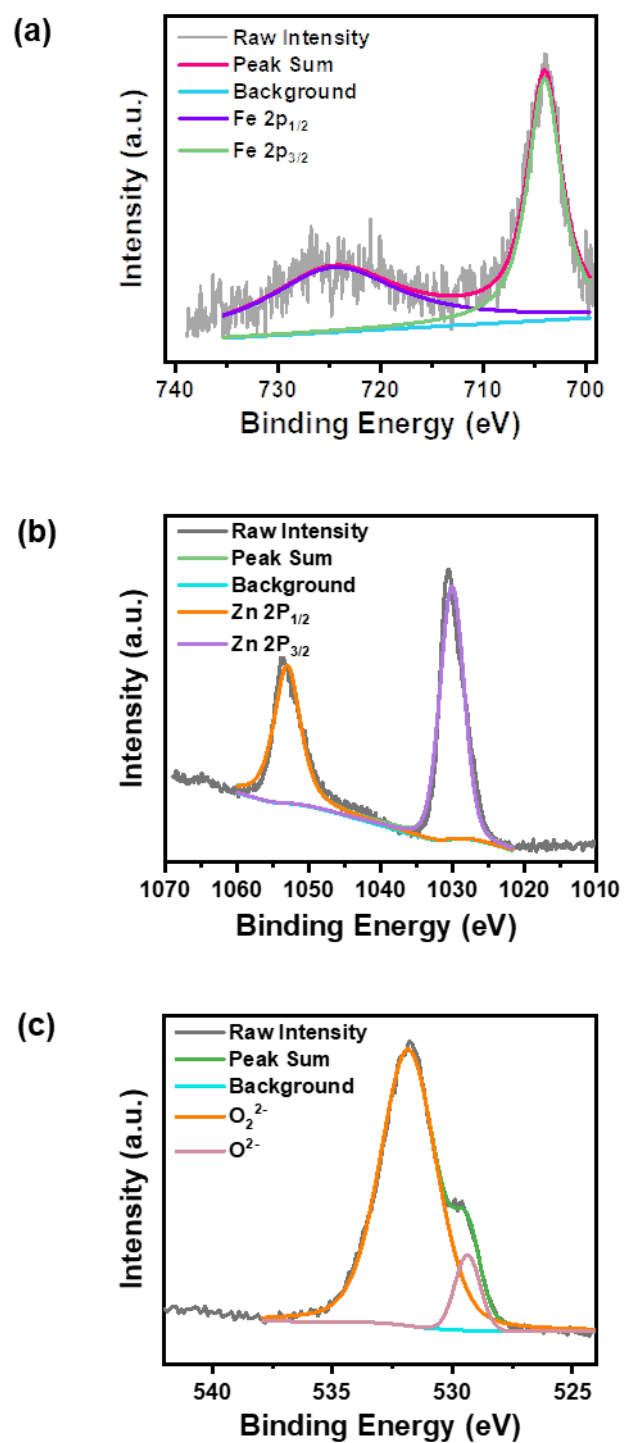


Fig. S2. X-ray photoelectron spectroscopy characterization of ZnO₂-Fe₃O₄@MV NPs. XPS spectra of Fe 2p (a), Zn 2p (b) and O 1s (c) for ZnO₂-Fe₃O₄@MV NPs.

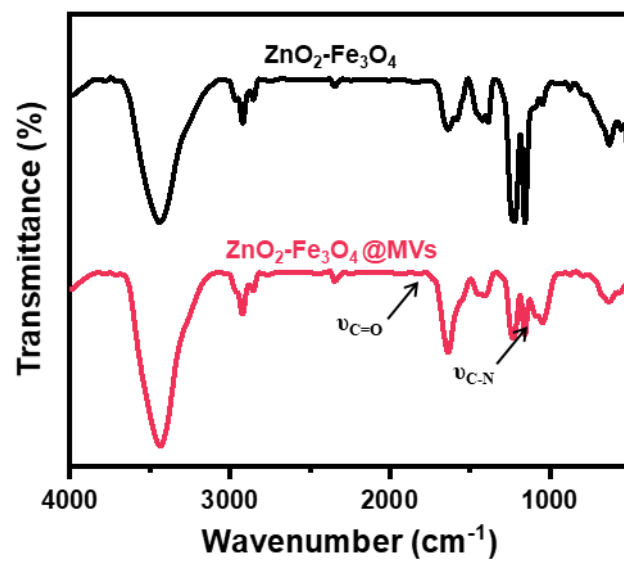


Fig. S3. Fourier-transform infrared spectroscopy characterization. FT-IR spectra of $\text{ZnO}_2\text{-Fe}_3\text{O}_4$ NPs and $\text{ZnO}_2\text{-Fe}_3\text{O}_4\text{@MV}$ NPs, respectively.

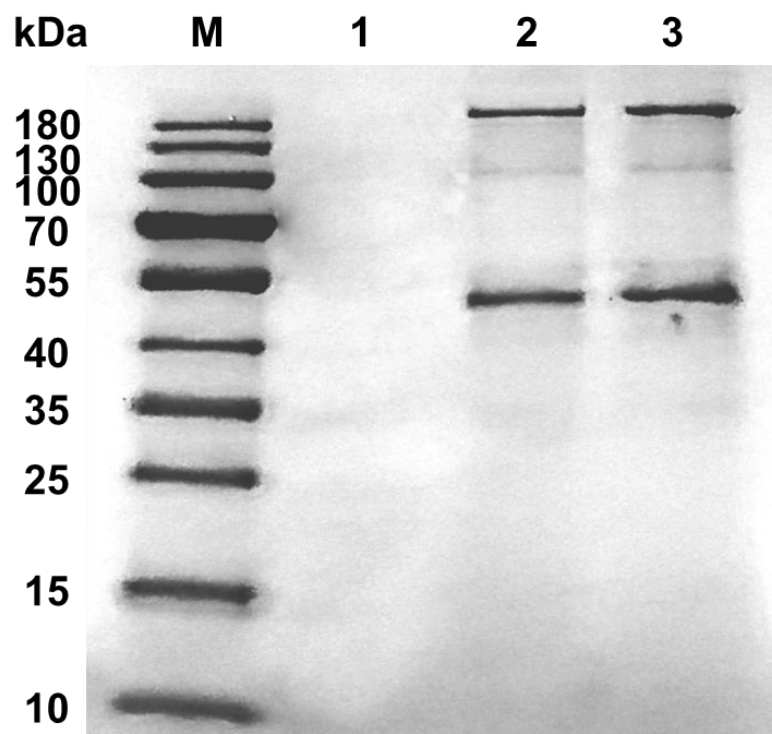


Fig. S4. Protein bands of *S. gordonii* MVs by Coomassie brilliant blue staining.

Lane M: Prestained marker

Lane 1: ZnO₂-Fe₃O₄ NPs

Lane 2: *S. gordonii* MVs

Lane 3: ZnO₂-Fe₃O₄@MV NPs

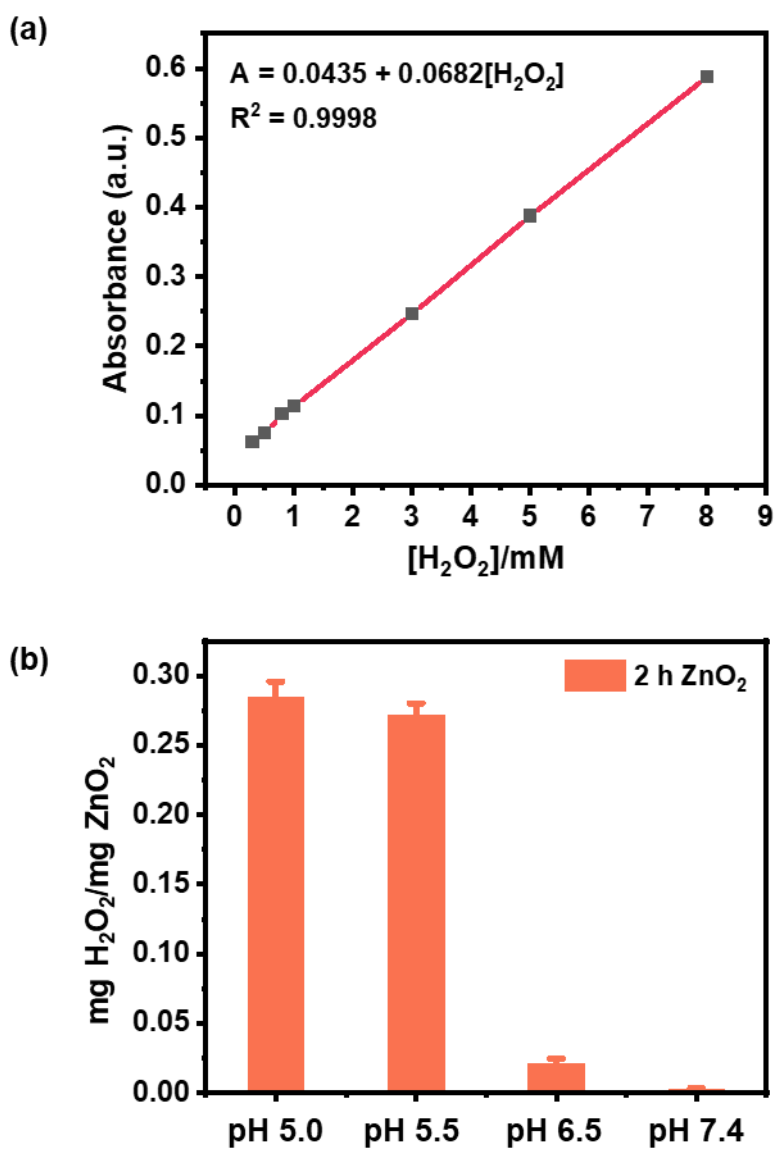


Fig. S5. Measurements of H₂O₂ production by ZnO₂ with Hydrogen Peroxide Content Assay Kit (a) Quantification of H₂O₂ in ZnO₂ NPs. (b) The ratios of the amount of H₂O₂ produced to the mass of ZnO₂ at different pH values. Error bar: standard deviation (n = 3).

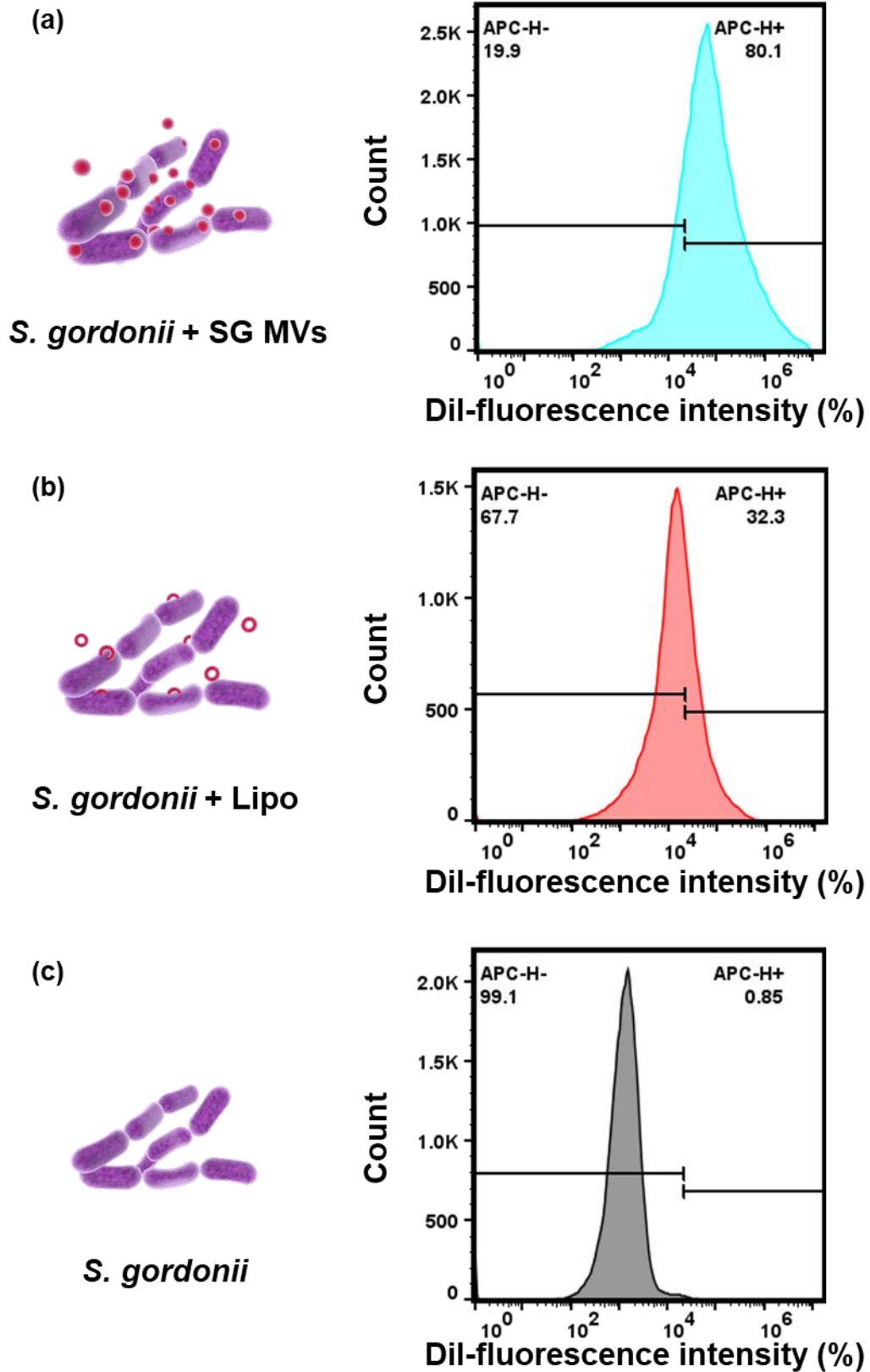


Fig. S6. Flow cytometry analysis of the uptake by individual bacteria of Dil-loaded SG MVs from parental *S. gordonii*. Lipo@Dil and PBS were used as control. Flow cytometry counts as a function of the red-fluorescence intensity due to SG MVs and Lipo@Dil uptake by *S. gordonii*.

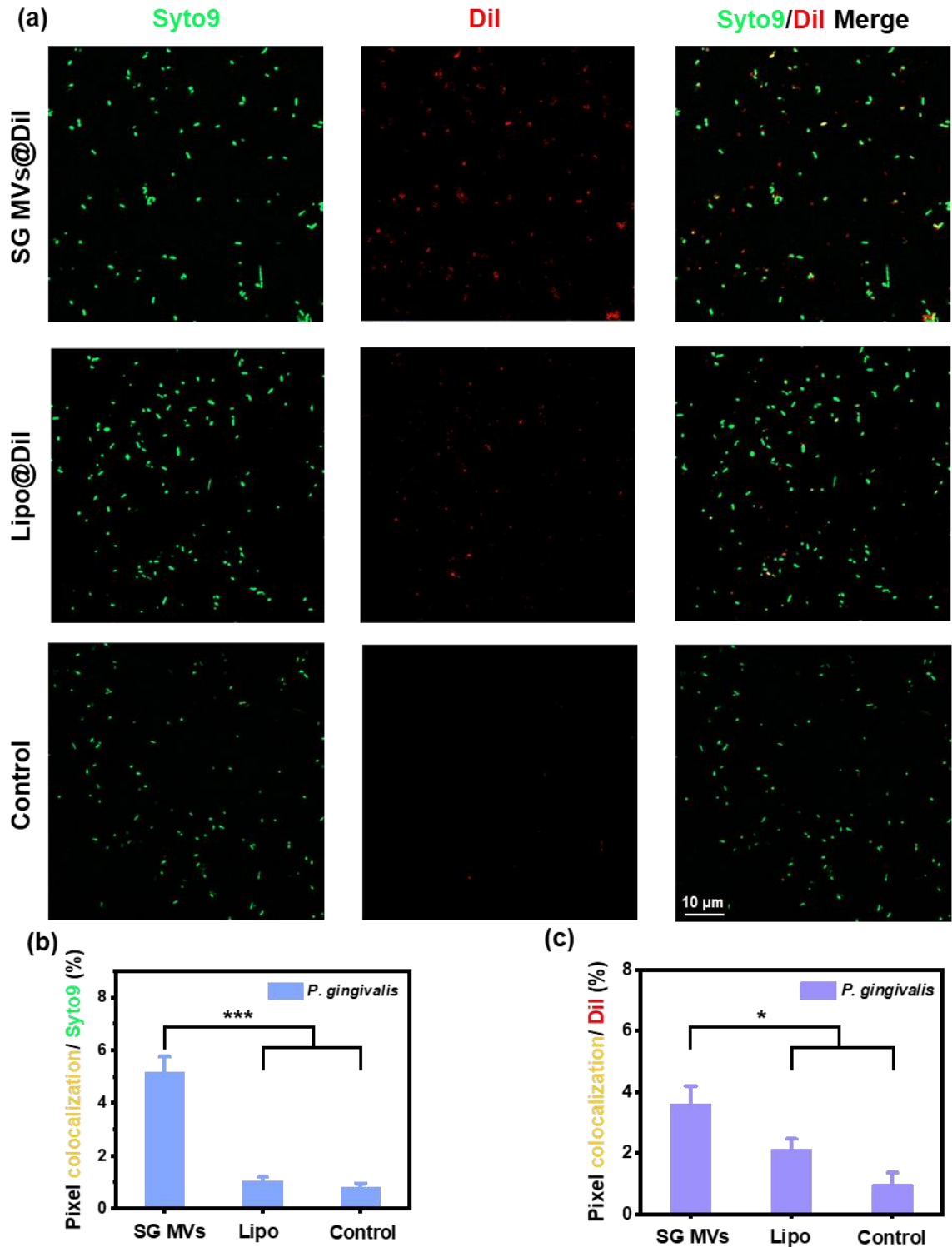


Fig. S7. Confocal fluorescence images of *P. gingivalis* incubated with *S. gordonii* MVs and image analysis. (a) Confocal fluorescence images of *P. gingivalis* incubated with SG MVs@Dil, Lipo@Dil, and PBS control for 2 h. Green fluorescence: *P. gingivalis* stained by Syto9, E_m : 488 nm; Red fluorescence: *S. gordonii* MVs and liposomes stained by Dil, E_m : 565 nm. Yellow: merge of the green and the red. Scale bar: 10 μm . (b) The area ratios of colocalized pixels of both the green and the red normalized by the pixels of the green (Syto9). (c) The area ratios of colocalized pixels of both the green and the red normalized by the pixels of the red (Dil). Error bar: standard deviation (n = 3). Statistical significance by one-way analysis of variance (*p < 0.05, ***p < 0.001).

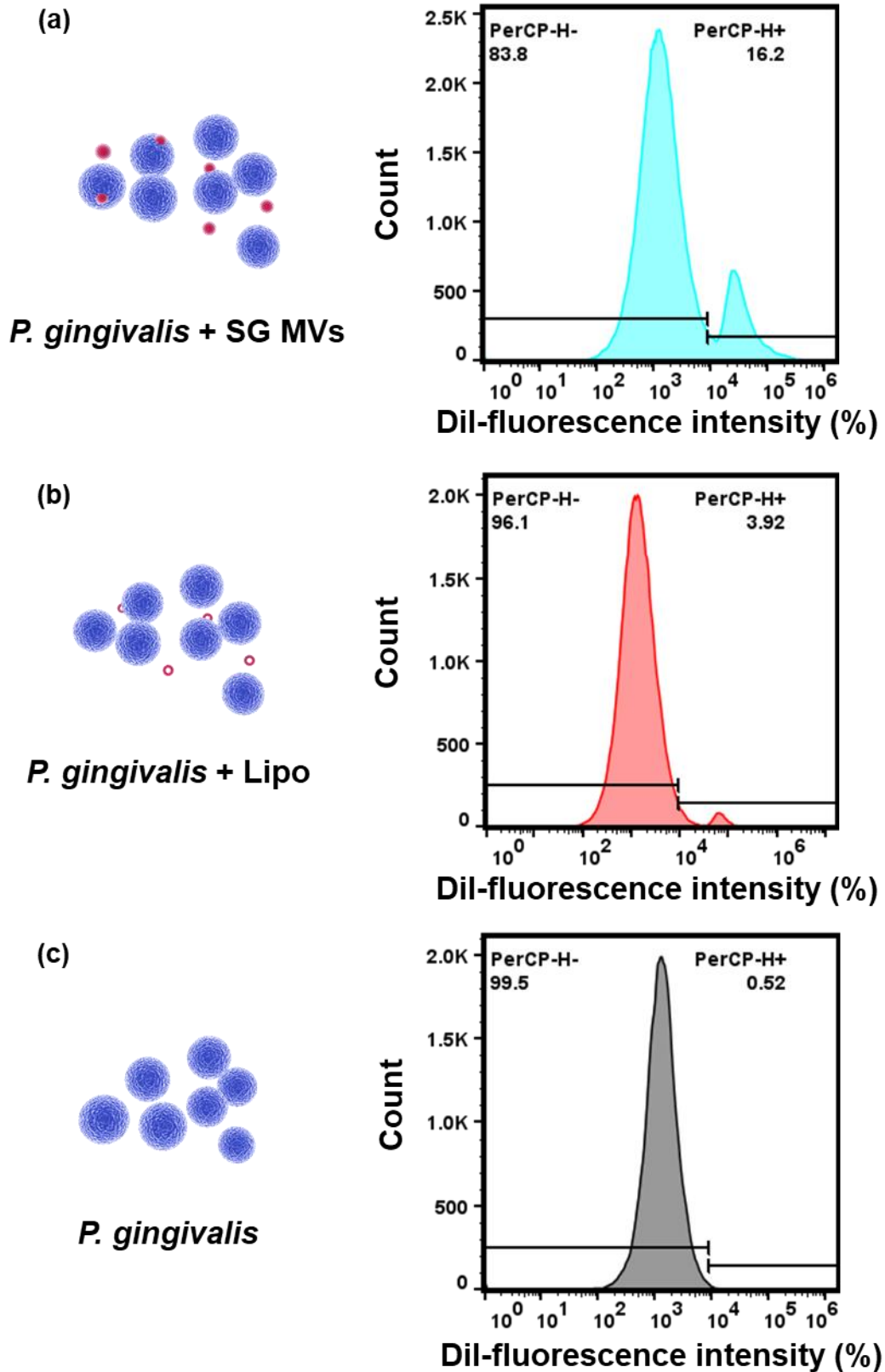


Fig. S8. Flow cytometry analysis of the uptake by individual bacteria of Dil-loaded SG MVs from *P. gingivalis*. Lipo@Dil and PBS were used as control. Flow cytometry counts as a function of the red-fluorescence intensity due to SG MVs and Lipo@Dil uptake by *P. gingivalis*.

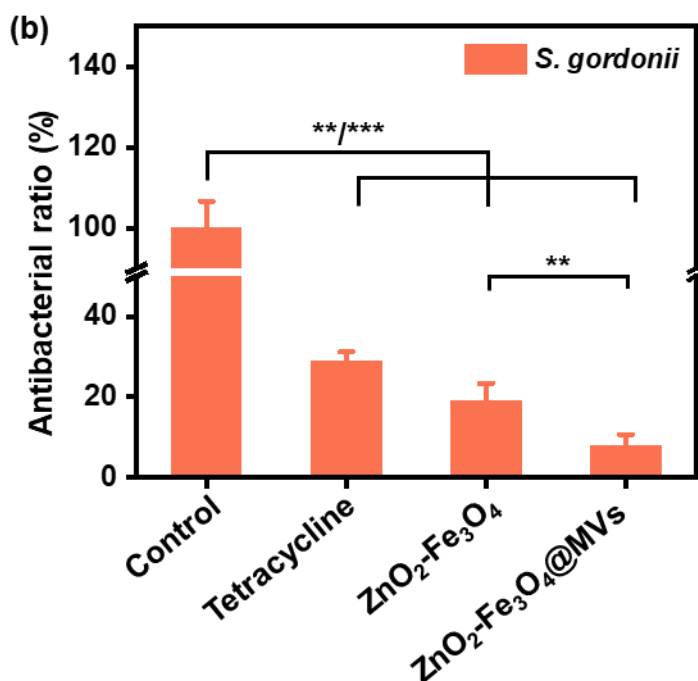
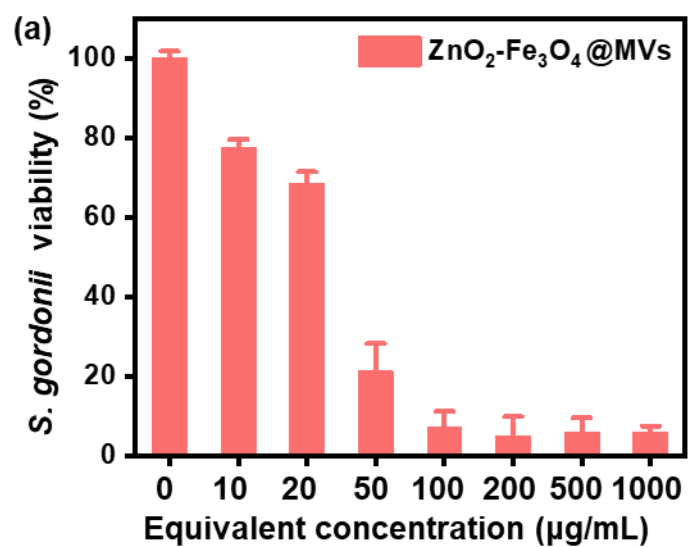


Fig. S9. Bacterial viability evaluation by colony counting after (a) the treatment of ZnO₂-Fe₃O₄@MV NPs at different concentrations; and (b) different treatments including PBS, tetracycline, ZnO₂-Fe₃O₄ NPs and ZnO₂-Fe₃O₄@MV NPs. Error bar: standard deviation (n = 3). Statistical significance by one-way analysis of variance (**p < 0.01, ***p < 0.001).

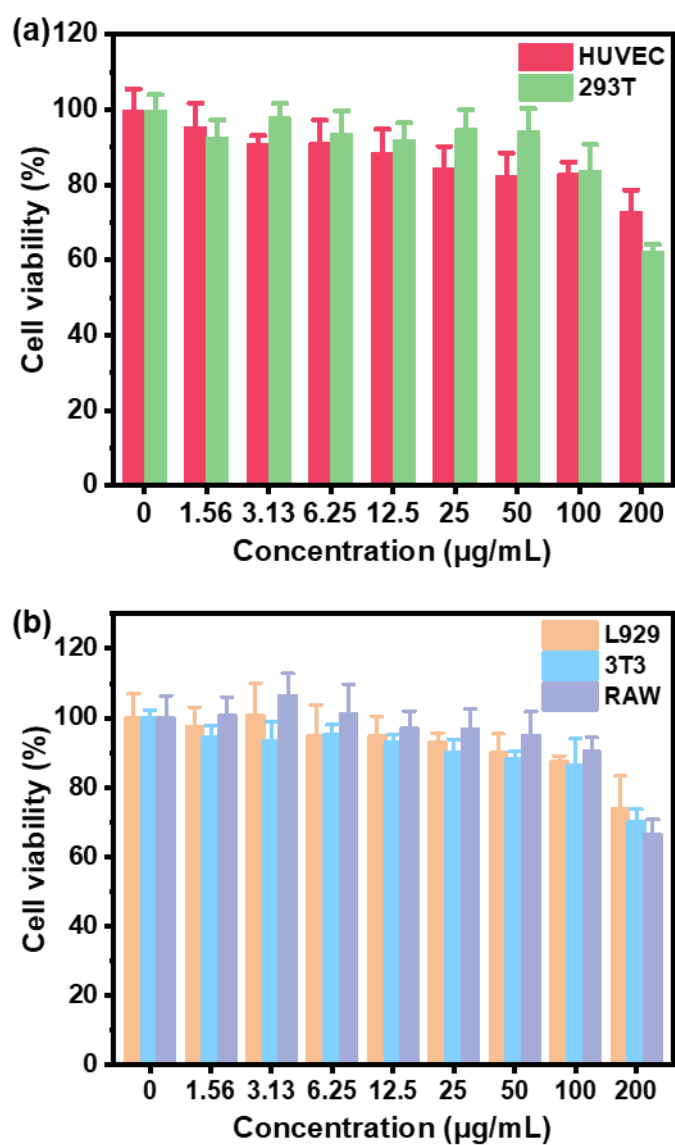


Fig. S10. Biocompatibility assessment of ZnO₂-Fe₃O₄@MV NPs. (a) Human derived cell viability evaluation by ZnO₂-Fe₃O₄@MV NPs, including 293T cells and HUVECs cells. (b) The evaluation of the biocompatibility of normal mouse derived cells by ZnO₂-Fe₃O₄@MV NPs, including L929 cells, 3T3 cells and RAW 264.7 cells. Incubation: 24 h. Error bar: standard deviation (n = 3).

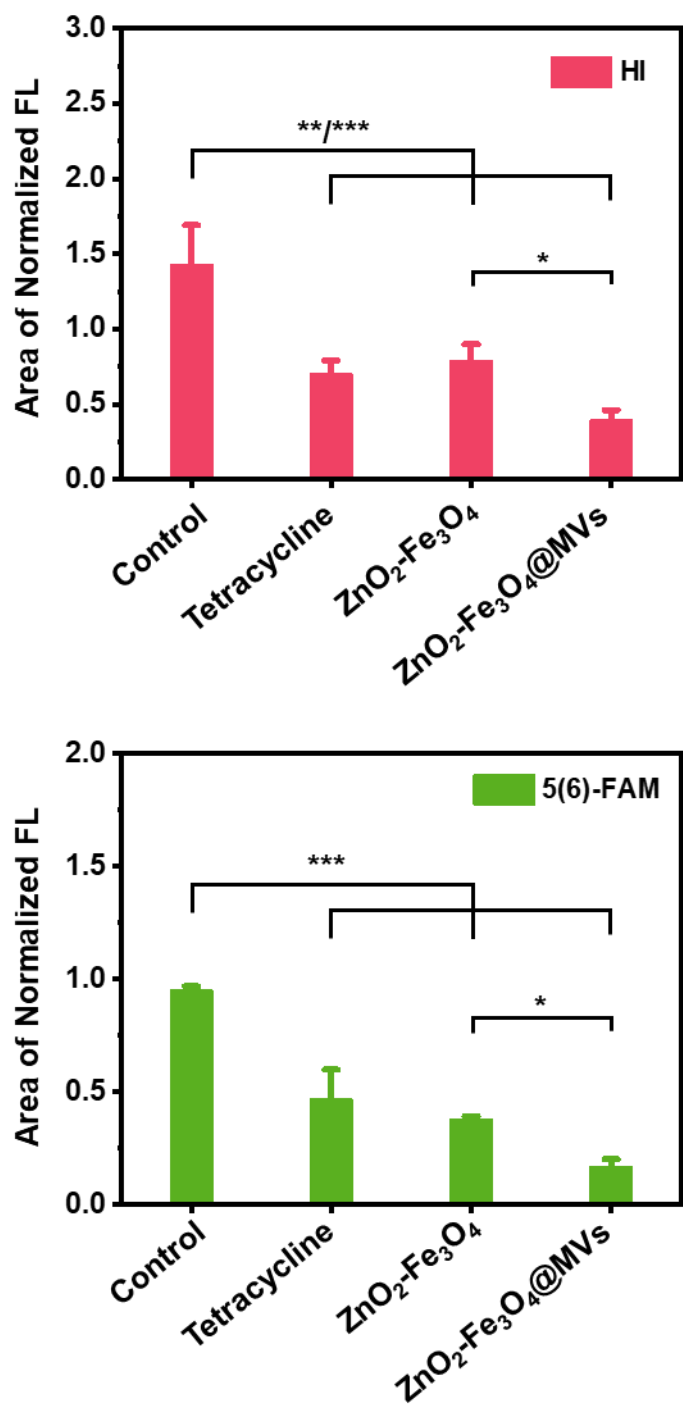
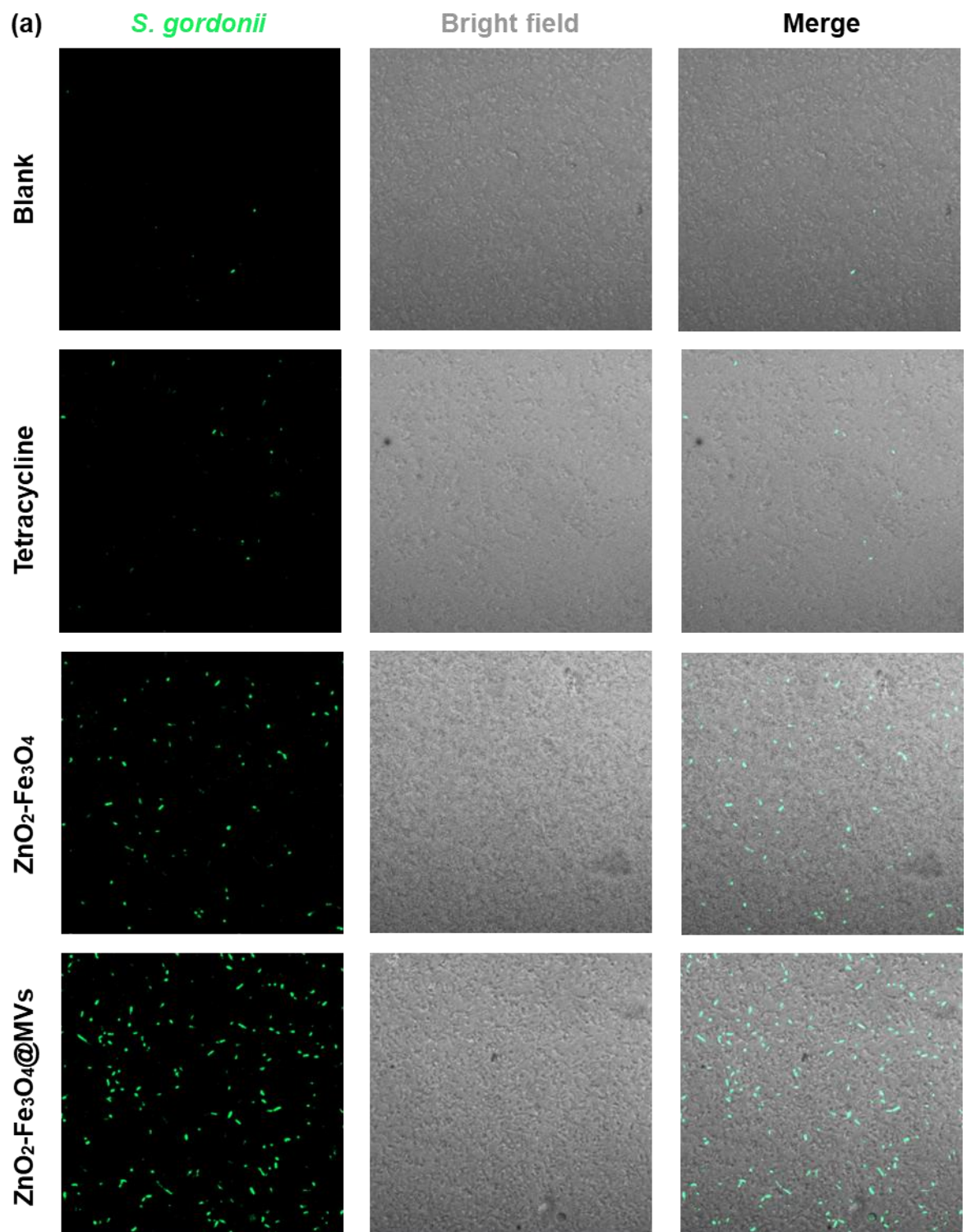


Fig. S11. Comparison of the accumulative peak areas from the curves in Fig. 4b-c. Error bar: standard deviation (n = 3). Statistical significance by one-way analysis of variance (*p < 0.05, **p < 0.01, ***p < 0.001).



(b)

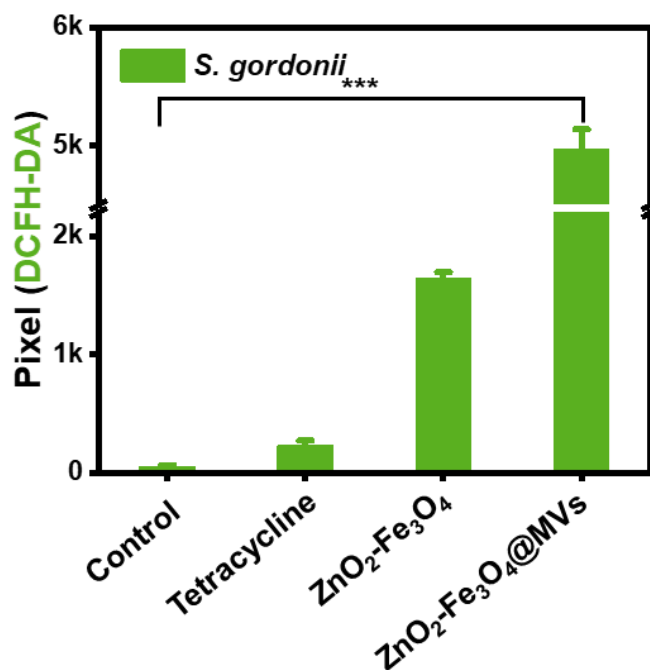


Fig. S12. Fluorescence images and analysis of ROS in planktonic bacteria (*S. gordonii*) by different treatments including PBS, tetracycline, ZnO₂-Fe₃O₄ NPs and ZnO₂-Fe₃O₄@MV NPs. (a) Fluorescence images of planktonic bacteria (*S. gordonii*). ROS probe: DCFH-DA. (b) Accumulative pixel areas of the fluorescence images at the channel of DCFH-DA. Error bar: standard deviation (n = 3).

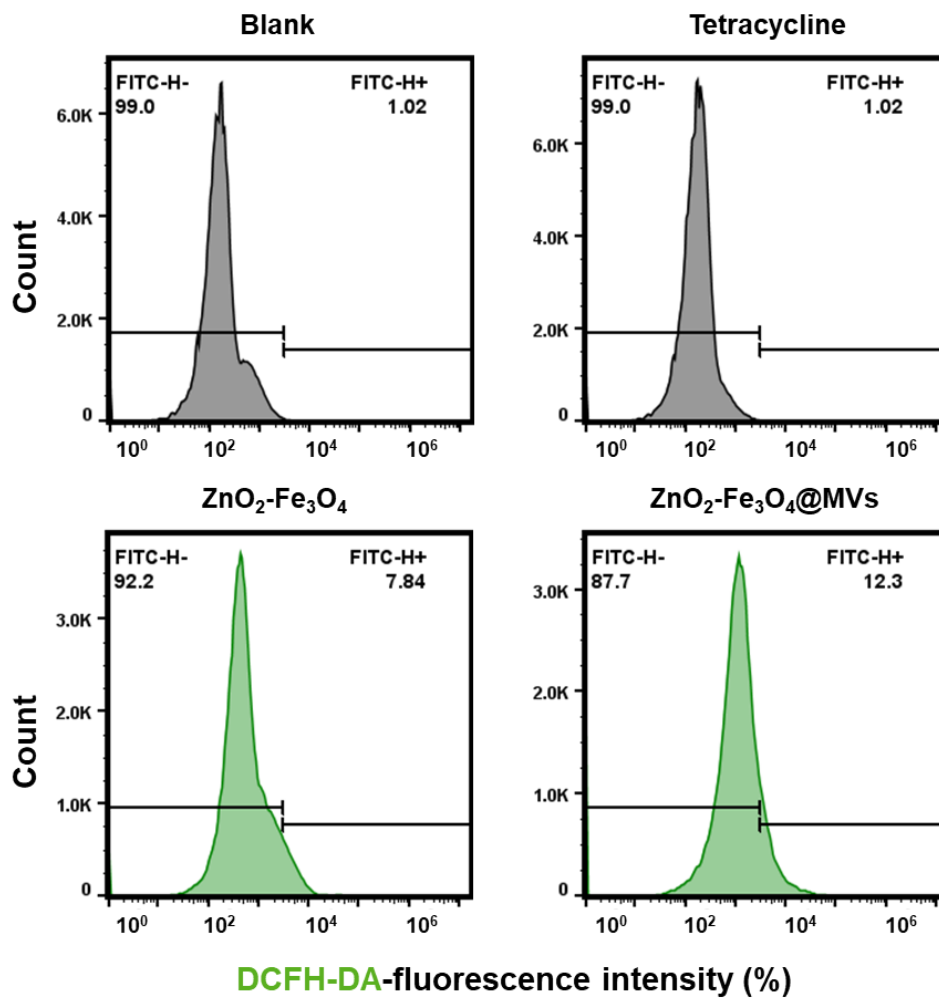


Fig. S13. Flow cytometry analysis of the uptake by ROS in planktonic bacteria (*S. gordonii*) by different treatments including PBS, tetracycline, ZnO₂-Fe₃O₄ NPs and ZnO₂-Fe₃O₄@MV NPs. ROS probe: DCFH-DA.

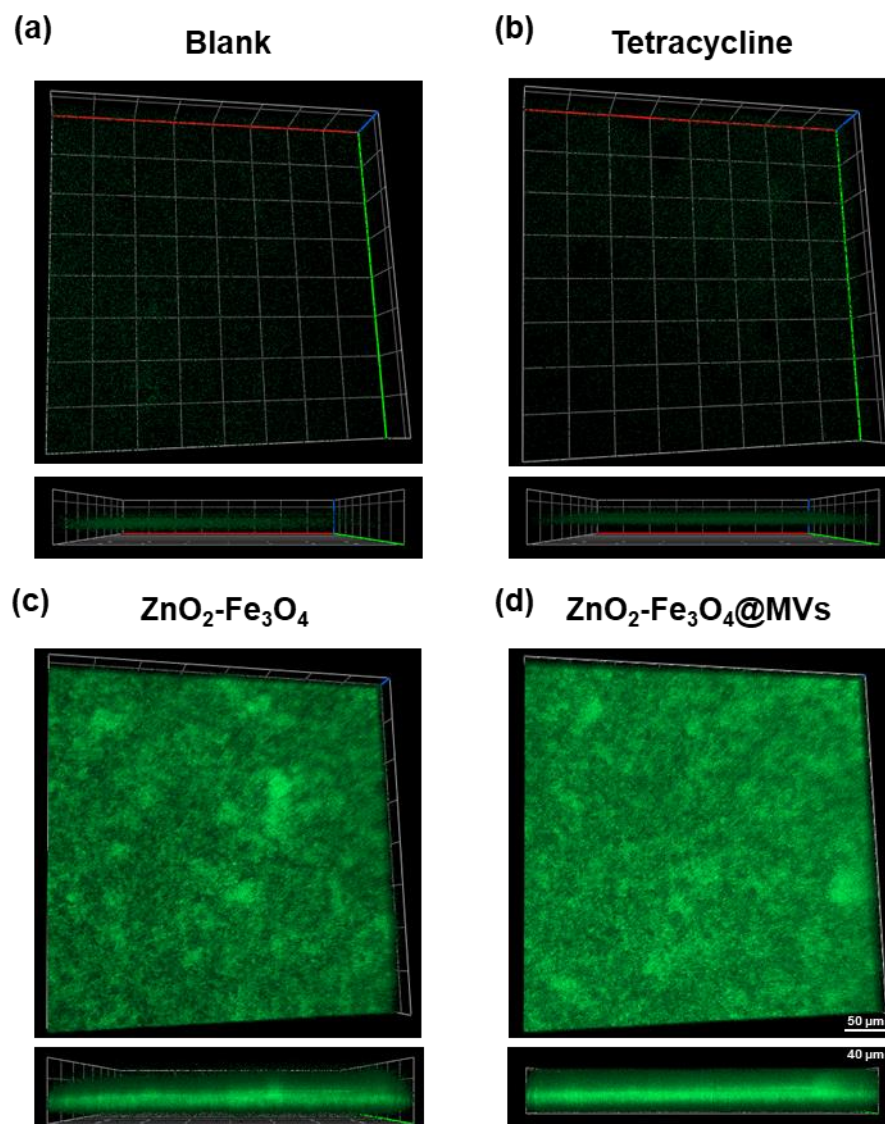


Fig. S14. Fluorescence images and analysis of ROS in symbiotic biofilms by different treatments including (a) PBS, (b) tetracycline, (c) $\text{ZnO}_2\text{-Fe}_3\text{O}_4$ NPs and (d) $\text{ZnO}_2\text{-Fe}_3\text{O}_4\text{@MV}$ NPs. ROS probe: DCFH-DA.

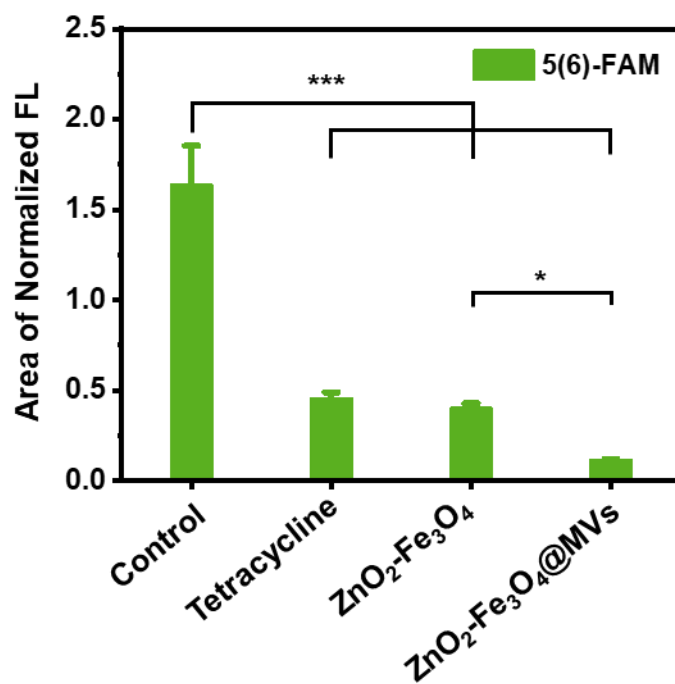
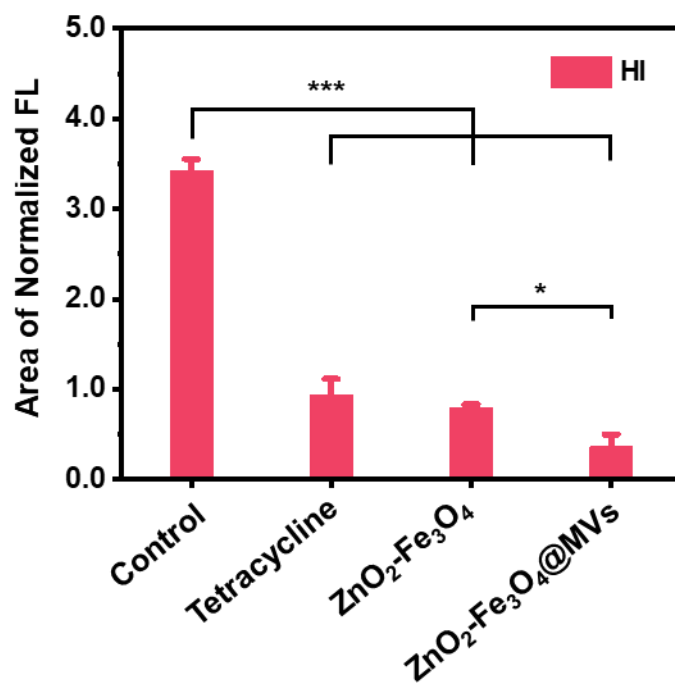


Fig. S15. Comparison of the accumulative peak areas from the curves in Fig. 5b-c. Error bar: standard deviation (n = 3). Statistical significance by one-way analysis of variance (*p < 0.05, ***p < 0.001).