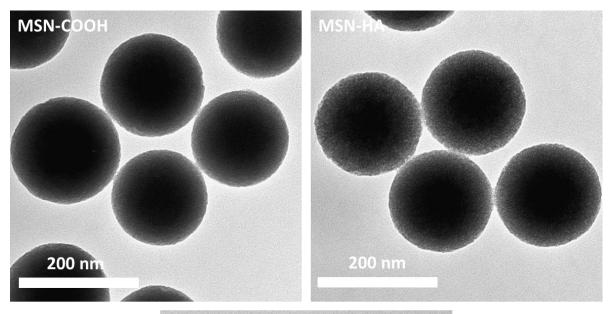
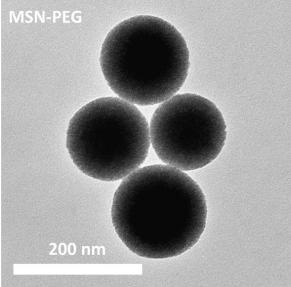
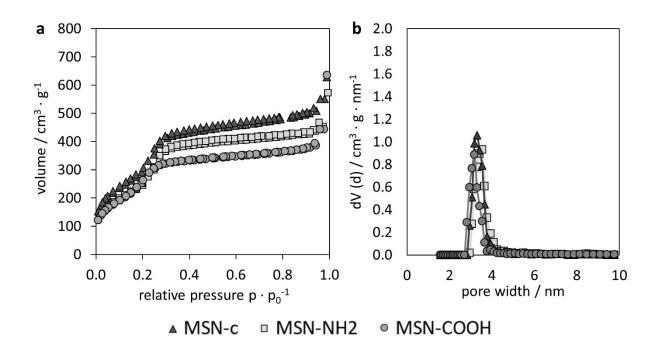
Electronic Supplementary Material (ESI) for Biomaterials Science. This journal is © The Royal Society of Chemistry 2023

## **Supporting Information**





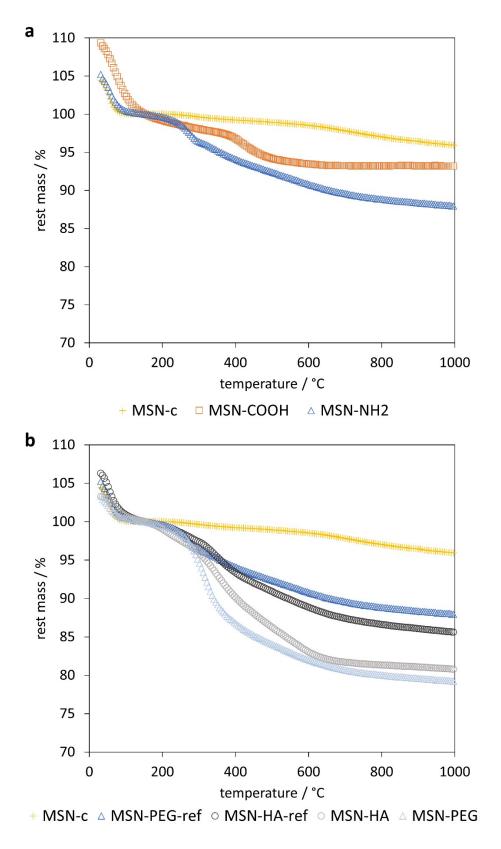
**Figure S1.** Exemplary transmission electron microscopy image of MSN-COOH, MSN-PEG and MSN-HA particles.



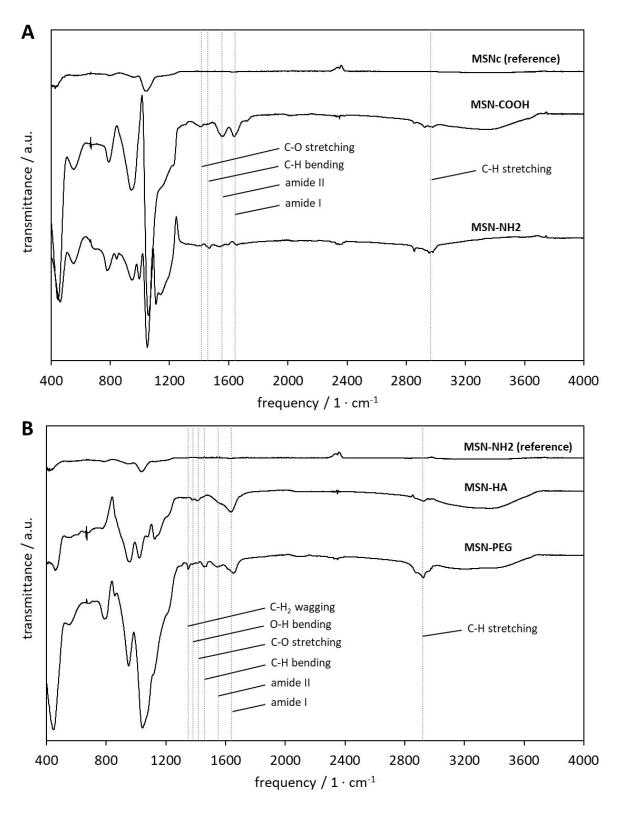
**Figure S2.** a) Nitrogen sorption isotherms of the basic particles MSN-NH<sub>2</sub>, MSN-COOH and calcined MSN-c. b) Corresponding pore size distributions calculated using the equilibrium NLDFT kernel developed for silica.

**Table S1.** Data from the nitrogen sorption measurements of MSN-COOH,  $MSN-NH_2$  and the basic calcined MSNc

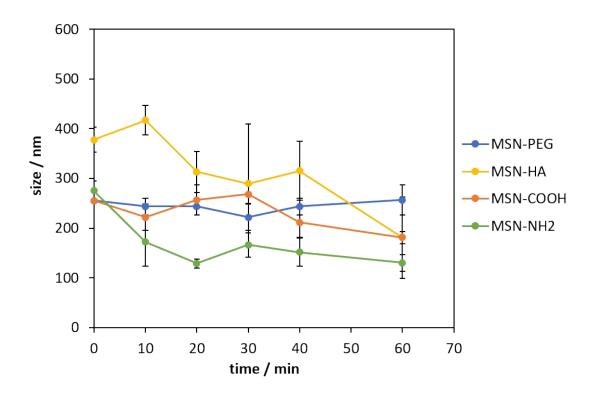
	specific surface area / m² · g <sup>-1</sup>	pore volume / cm³ • g-¹	pore diameter/ nm
MSN-c	1048	0.74	3.3
MSN-NH <sub>2</sub>	869	0.64	3.5
MSN-COOH	842	0.55	3.2



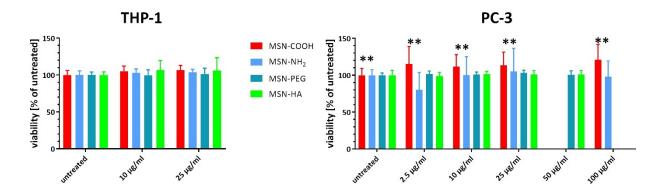
**Figure S3.** Exemplary thermogravimetric measurements of calcined MSN-c, MSN-COOH, MSN-NH2, MSN-HA and MSN-PEG. For the quantification of grafted polymer, the native particles were incubated in the respective solvent without HA or PEG to account for hydrolysis-related mass losses ('ref').



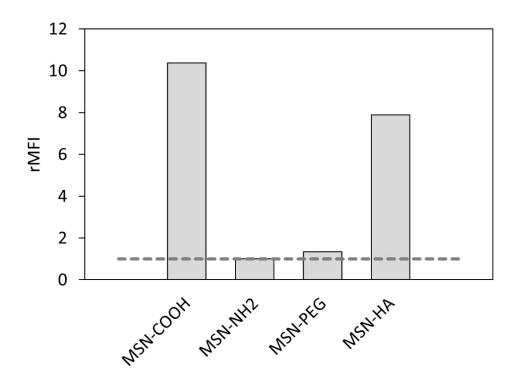
**Figure S4.** Exemplary ATR-IR spectroscopic analysis of different MSN types used in this study. A) IR spectra of MSN-COOH and MSN-NH2; IR spectrum of non-functionalized, calcined MSN-c particle was subtracted as background spectrum. B) IR spectra of MSN-HA and MSN-PEG; IR spectrum of the basic amino-functionalized MSN-NH2 particle was subtracted as background spectrum. Bands in the spectra of MSNs assignable to the various functional groups are highlighted in the figure (dashed lines). (doi.org/10.1002/bip.10245, doi.org/10.1016/0008-6215(94)00147-2, doi.org/10.3390/ijms13066639)



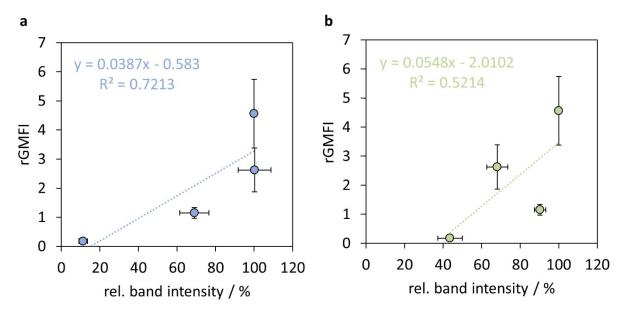
**Figure S5.** Mean hydrodynamic diameter measured by light-scattering as a function of incubation time in 10% FCS at a particle concentration of 10  $\mu$ g/mL.



**Figure S6.** Cell populations were incubated with nanoparticles at indicated amounts for 24 h prior to cell viability testing using the Cell Titer Blue assay. The graph shows mean viability  $\pm$  standard deviation (SD) as percentages of the untreated control, respectively. Three to four independent experiments were performed in triplicates (N = 9-12). Asterisks denote experiments which were carried out with parental, not genetically engineered PC-3 cells (please see methods section for details).



**Figure S7.** Mean fluorescence intensities (MFIs) of ATTO647N-labelled MSNs were measured in aqueous sucrose solution (10 %) at a particle concentration of 100  $\mu$ g ml<sup>-1</sup>. The respective MFIs were normalized on the MFI of MSN-NH<sub>2</sub> resulting in rated MFIs (rMFIs) to compensate for differences in particle fluorescence intensities. The dashed line indicates a rMFI of 1.



**Figure S8.** rGMFIs of THP-1 cells measured via flow cytometry after treatment with ATTO647-labelled MSNs at 10  $\mu$ g mL<sup>-1</sup> for 24 h and 37°C as a function of relative band intensity for the a) high molecular weight proteins and b) low molecular weight proteins (n = 3, error bars are SD).