## Table S1. Binding parameters for the formations of DHB<sub>DAXX</sub>/SPEP complexes determined

Peptide	Ele	Vdw	Polar	Nonpolar	ΔG*
PEPI	-836.9 (40)	-66.2 (1.4)	829.2 (33.1)	-12.7 (0.2)	-84.1 (7.1)
SPEP1	-929.4 (41)	-73.1 (1.4)	931.5 (42)	-13.5 (0.2)	-84.3 (2.3)
SPEP2	-850.5 (13)	-67.1 (0.6)	834.8 (8)	-12.5 (0.3)	-92.1 (1.8)
SPEP3	-853 (1.1)	-67.9 (2.3)	839.8 (7)	-12.7 (0.7)	-82.4 (3.3)
SPEP4	-564.4 ( 66)	-69.3 (1.6)	877 (31)	-11.9 (0.5)	-73.7 (3.3)
SPEP5	-892.6 (26)	-71.7 (7.2)	649 (67)	-11.3 (0.8)	-70.9 (5.5)
SPEP6	-641.1 (74)	-70.3 (3.6)	721 (25)	-12.3 (0.1)	-80.1 (4.0)
SPEP7	-726 (31)	-68.1 (1.1)	574 (64)	-11.2 (0.3)	-86.3 (1.4)

## using computational methods<sup>\$</sup>

<sup>\$</sup>All values are calculated in kcal mol<sup>-1</sup> at 300K. Errors of 1 SD are shown in parentheses. \*Entropy changes are not calculated, therefore the binding energy calculated here corresponds to only the enthalpy contribution.

Peptide	Ele	Vdw	Polar	Nonpolar	∆G*	Kd (nM)
STPEP1	-814.2 (52)	-63.7 (2.1)	799.5 (47)	-11.7 (0.5)	-90.1 (7)	n.d
STPEP2	-726.3 (15)	-62.1 (1.0)	712. 0 (14)	-11.8 (0.2)	-88.2 (1.5)	32.4 (2.6)
STPEP3	-630.7 (18)	-61.0 (3)	614.9 (17)	-11.5 (0.4)	-88.2 (4.3)	34.9 (2.4)
STPEP4	-447.7 (34)	-60.5 (1.8)	439.9 (31)	-11.2 (0.1)	-79.5 (1.0)	147.8 (12.4)
STPEP5	-772 (36)	-55.0 (3)	763 (32)	-10.7 (0.4)	-71.6 (7.5)	n.d
STPEP6	-694 (45)	-52.2 (6)	648.8 (44)	-10.4 (0.8)	-71.2 (9.1)	n.d
STPEP7	-662.7 (51)	-57.8 (0.7)	654.5 (45)	-10.8 (0.2)	-76.8 (5.5)	268.9 (17.3)

Table S2: Binding parameters for the formation of DHB<sub>DAXX</sub>/STPEP complexes determined using computational and experimental methods.

<sup>\$</sup>Binding energies are calculated using MMPBSA method. All values are calculated in kcal mol<sup>-1</sup> at 300K. Errors of 1 SD are shown in parentheses. \*Entropy changes are not calculated, therefore the binding energy calculated here corresponds to only the enthalpy contribution. Kd values were determined from fluorescence anisotropy measurments.

## **Supplementary Figures:**



Figure S1. ITC isotherms for binding of DHB<sub>DAXX</sub> to peptide fragments of ATRX.



**Figure S2. Stability of docked DHB**<sub>DAXX</sub>/PEP<sub>RassF1C</sub> and DHB<sub>DAXX</sub>/PEP<sub>ATRX</sub> complexes. A Cartoon representations showing 7 poses of the PEP<sub>ATRX</sub> helix (multiple colours) docked with DHB<sub>DAXX</sub> (grey). **B** RMSD of conformations of PEP<sub>Rassf1C</sub> sampled during MD simulations starting from the DHB<sub>DAXX</sub>/PEP<sub>Rassf1C</sub> solution structure. **C** RMSD of conformations of PEP<sub>ATRX</sub> peptides sampled during MD simulations starting from different docked poses of PEP<sub>ATRX</sub>; (left) RMSD of the peptides with stable binding; (right) RMSD of peptides with unstable binding during MD simulations.



Figure S3. Structure and stability analysis of PEP<sub>ATRX</sub> and DHB<sub>DAXX</sub> in apo and bound forms. **A** Time evolution of secondary structures in apo PEP<sub>ATRX</sub> sampled during BP-REMD simulations (blue:  $\alpha$  - helix, gray: 3<sub>10</sub>- helix, yellow: turn, green: bend, white: coil). **B** CD spectrum of PEP<sub>ATRX</sub> showing % helicity. **C** Probability distributions of RMSD of conformations sampled during simulations; (left) apo DHB<sub>DAXX</sub>; (middle) bound DHB<sub>DAXX</sub> from DHB<sub>DAXX</sub>/PEP<sub>ATRX</sub> complex, and (right) bound PEP<sub>ATRX</sub> from DHB<sub>DAXX</sub>/PEP<sub>ATRX</sub> complex. Black and red corresponds to RMSD of conformations with or without the flexible N- and/or C- terminal residues described in the main text, respectively.



Figure S4. Conformational analysis of stapled and stitched peptides. Time evolution of the secondary structure of SPEP 1-7 and STPEP 1-7 during BP-REMD simulations (blue:  $\alpha$  - helix, grey: 3<sub>10</sub>- helix, yellow: turn, green: bend, white: coil, red: beta strand). The overall peptide conformations sampled were used to calculate percentage helicity ('Calc'). For the stapled peptides, SPEP 1-7, percentage helicity obtained by fitting CD spectra ('Exp') is plotted alongside.



**Figure S5. Computational analysis of the stability of DHB**<sub>DAXX</sub>/**SPEP complexes.** RMSD of conformations sampled during MD simulations of DHB<sub>DAXX</sub>/SPEP complexes; (top) DHB<sub>DAXX</sub> and (bottom) stapled peptides (SPEP). The different colors, black, red, green, blue, yellow, brown and grey corresponds to different DHB<sub>DAXX</sub> /SPEP 1 to 7 complexes in numerical order.



**Figure S6: Per-residue contributions to binding of DHB**<sub>DAXX</sub> **by SPEP1-7 calculated from MD simulations.** Binding free energies of individual SPEP residues to DHB<sub>DAXX</sub> were calculated using the MMPBSA approach (see Methods). The staple linker positions are highlighted in blue.



Figure S7. ITC isotherms for binding of DHB<sub>DAXX</sub> to SPEP1 to 7.



**Figure S8. Competitive displacement of FAM-SPEP7 from DHB**<sub>DAXX</sub> **by non-fluorescent SPEP peptides 1-7.** Binding data with standard error bars derived from three independent measurements are shown for each peptide (closed symbols). Curves obtained from fitting the data to a competitive binding model (solid black line) are shown with simulated curves using Kd values obtained from ITC data (dashed red line) included for comparative purposes.



Figure S9. <sup>1</sup>H-<sup>15</sup>N HSQC spectra showing the formation of the <sup>15</sup>N NSIM-DHB<sub>DAXX</sub>/FAM-SPEP7 pre-complex and subsequent binding of SUMO-1 in sequential titrations. A. Overlaid <sup>1</sup>H-<sup>15</sup>N HSQC spectra of unbound <sup>15</sup>N-NSIM-DHB<sub>DAXX</sub> (blue) and a saturated complex formed with FAM-SPEP7 (red) are shown. The transition of a single resonance projected in 1D (inset) with an additional mid-titration point is included (black), indicating slow exchange. **B.** (Top) Three panels showing overlaid sections of the <sup>1</sup>H-<sup>15</sup>N HSQC spectra of <sup>15</sup>N-NSIM-DHB<sub>DAXX</sub>/FAM-SPEP7 in the absence (black) and presence (grey) of increasing quantities of SUMO-1. Multicoloured arrows indicate 9 individual resonances whose discrete chemical shift changes, in fast exchange, were measured during the titration. (Bottom) Titration curves of the 9 resonances, coloured as indicated above. The reported Kd value is the mean  $\pm$  SD of the 9 individually fitted curves. Combined amide chemical shift changes, Dd (ppm), were calculated as ( $(Dd_{1H})^2+(0.2Dd_{15N})^2$ )<sup>1/2</sup>.



**Figure S10. Cellular toxicity and localization of stapled peptides. A** HCT116 cells were titrated with peptides in the presence of 2 % serum and LDH release was assessed after 4 hrs incubation. **B** Cellular uptake was assessed by live cell imaging after treating HCT116 cells for 4 hrs with 25 uM of FAM-SPEP7 in the same conditions.



**Figure S11. Computational analysis of the stability of DHB**<sub>DAXX</sub>/**STPEP complexes.** RMSD of conformations sampled during MD simulations of DHB<sub>DAXX</sub>/STPEP complexes; (top) DHB<sub>DAXX</sub> and (bottom) stapled peptides (SPEP). The different colors; black, red, green, blue, yellow, brown and grey, correspond to different DHB<sub>DAXX</sub>/STPEP1 to 7 complexes in numerical order.