

Supporting Information

Electroreductive formylation of activated alcohols via radical-polar crossover

Jungtaek Kang, Heyjin Cho and Hyunwoo Kim*

Department of Chemistry, Korea Advanced Institute of Science and Technology (KAIST),

Daejeon 34141, Republic of Korea

correspondence to: hwkim@kaist.ac.edu

Table of contents

1.	General information	S-2
2.	General procedure for electroreductive formylation	S-3
3.	Unsuccessful substrates	S-4
4.	Cyclic voltammetry experiments	S-5
5.	Gram-scale synthesis	S-6
6.	Synthesis of starting materials	S-7
7.	Structural determination of 27	S-10
8.	Reactivity towards other electrophiles	S-11
9.	Characterization of compounds	S-12
10.	NMR spectra	S-28
11.	References	S-92

1. General information

All reactions were conducted under an argon atmosphere unless otherwise noted. Commercial reagents were purchased from Sigma Aldrich, Alfa Aesar, Acros, TCI and Combi Blocks used as received with the following exceptions: *N,N*-dimethylformamide (DMF) was distilled over CaH_2 and stored in flame-dried Schlenk tube with 3 Å molecular sieves. The electrolyte lithium triflate (LiOTf) was purchased from Alfa Aesar (or Acros) and stored in the glove box. Analytical thin layer chromatography (TLC) was performed on silica gel 60 F254 aluminum plates (Merck). TLC plates were visualized by exposure to ultraviolet light (254 nm or 366 nm). Flash column chromatography was performed on Merck silica gel (40-63 mesh). ^1H NMR (400 or 500 MHz), $^{13}\text{C}\{^1\text{H}\}$ NMR (100 or 126 MHz), and $^{31}\text{P}\{^1\text{H}\}$ NMR (162 MHz or 202 MHz) spectra were recorded on a Bruker Ascend 400 or 500, or a Bruker Avance III HD spectrometer and were reported in ppm, relative to residual solvent peak. All coupling constants (J values) were reported in Hertz (Hz). Mass spectra were obtained using a Bruker Daltonik micro TOF-Q II high-resolution mass spectrometer (ESI) at the KAIST Analyst Center for Research Advancement.

Electrolysis experiments were performed using a Biologics VMP3 multichannel potentiostat or a DC power supply. Carbon felt was purchased from Fuel Cell Store. Custom-made 3-hole undivided cells were utilized for electrochemical reactions. The carbon felt was cut into $0.5 \times 0.5 \times 0.3 \text{ cm}^3$ pieces and connected to an electrical feed-through on the Teflon cap of the electrochemical cell via a piece of graphite (4H pencil lead, 2 mm in diameter). Magnesium electrode (Mg, 99.9 % purity) was purchased from Alfa Aesar. The Mg electrode was cut into $2.5 \times 0.5 \times 0.1 \text{ cm}^3$ and was connected to an electrical feed-through on the Teflon cap of the electrochemical cell via a piece of graphite (4H pencil lead, 2 mm in diameter).

2. General procedure for electroreductive formylation

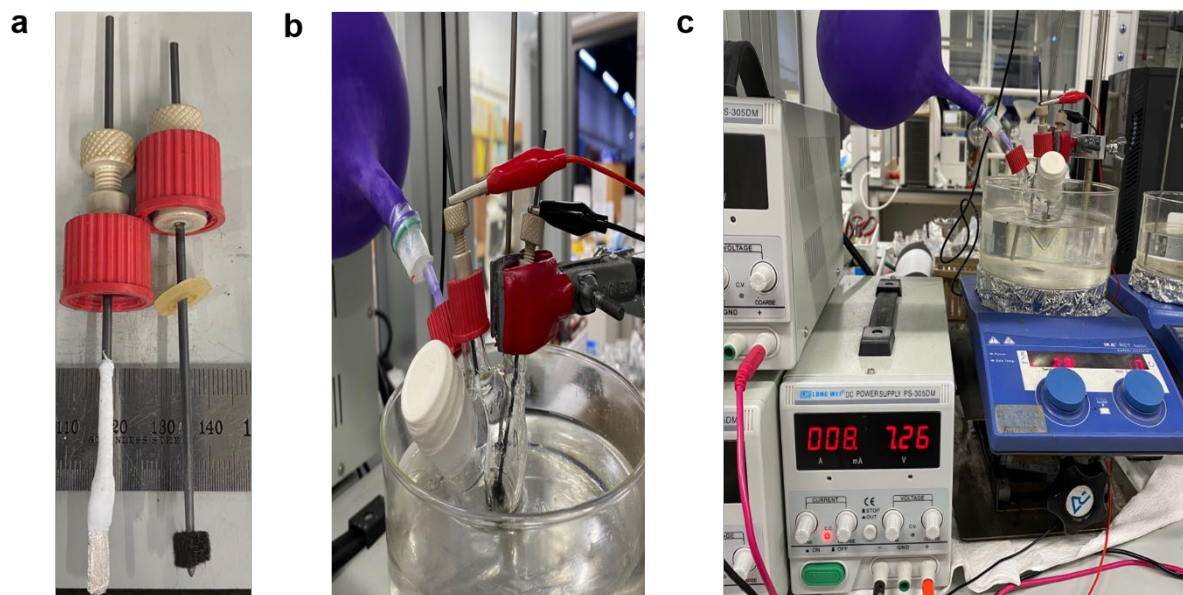
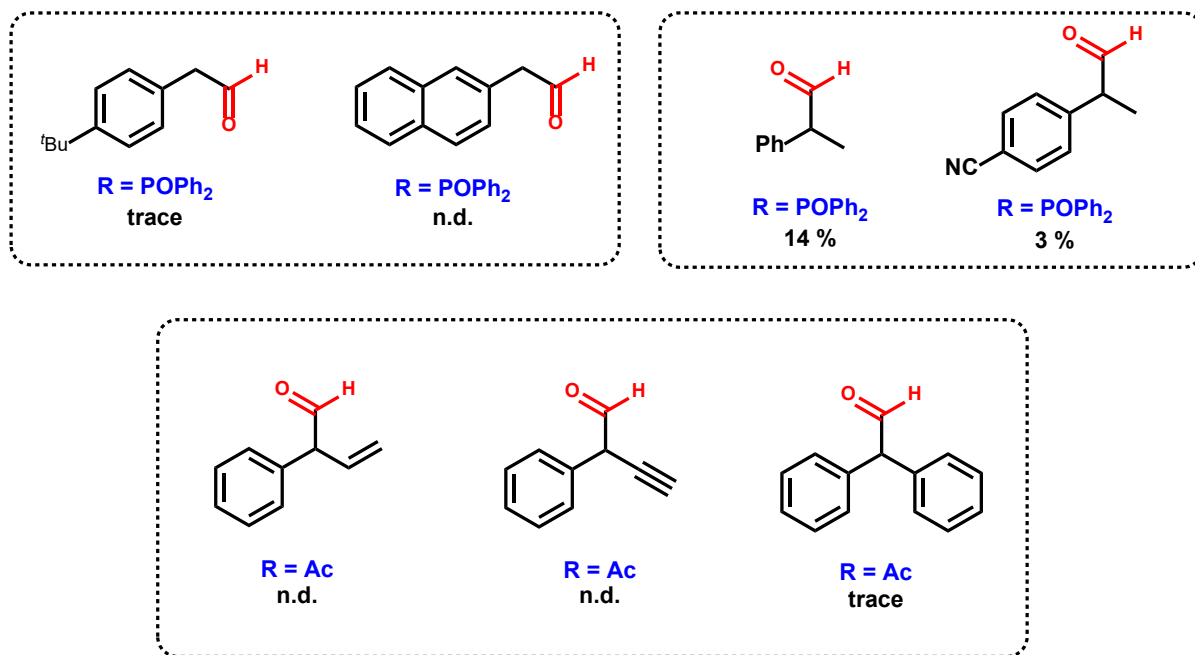


Figure S1. Set up for 1 mmol scale formylation of activated alcohols. a) Electrodes; b) Assembled reactor; c) Reaction in progress

An oven-dried, three-hole custom-designed glass electrochemical cell was equipped with a magnetic stir bar, a rubber septum, a magnesium anode ($2.5 \times 0.5 \times 0.1 \text{ cm}^3$) and a carbon felt cathode ($0.5 \times 0.5 \times 0.3 \text{ cm}^3$) connected to Teflon cap fitted with each separate electrical feed-throughs via 4H pencil leads (2 mm in diameter). This equipped reaction vessel was transferred into Ar-filled glove box. To this reaction vessel, substrate (1 mmol), LiOTf (172 mg, 1.1 mmol), and DMF (5 mL) were added and bring it out of glovebox. Pre-stirring the resulting mixture for 2~3 minutes, and then the reaction mixture was electrolyzed at a constant current of 8 mA electrolysis until passing 3.2 F/mol of a charge at 25 °C under Ar balloon. Then, the crude mixture was further diluted with Et₂O (40 mL). The resulting mixture was washed with saturated NH₄Cl (aq, 30 mL) and H₂O (30 mL x 5 times). The organic layer was dried over with anhydrous magnesium sulfate and concentrated under reduced pressure. The residue was subjected to flash column chromatography on silica gel (eluted with *n*-hexane/ethyl acetate) to yield the desired product.

3. Unsuccessful substrates



Detected byproducts

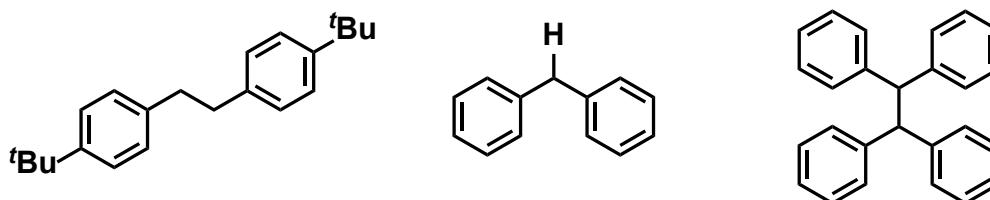


Figure S2. Suboptimal and unsuccessful substrates. Yields were determined by ^1H NMR analysis using 0.2 mmol mesitylene as the internal standard. Even though most reactions had a conversion rate of over 90%, the only detected byproducts were protonated or dimerized.

4. Cyclic voltammetry (CV) experiments

General information: CV was conducted in a 10 mL glass vial fitted with a glassy carbon working electrode (3 mm in diameter, BASi), a platinum wire and Ag/Ag⁺ as counter electrode and reference electrode. The 10 mM substrates in 0.1 M LiOTf in DMA (2 mL) or in 0.1 M LiOTf in DMF (2 mL) solutions were utilized. The solution of interest was sparged with Ar for 5 minutes before data collection. The redox potential of ferrocene/ferrocenium (Fc⁺/Fc) was measured and used to provide an internal reference. Scan rate is 100 mV/s unless specified.

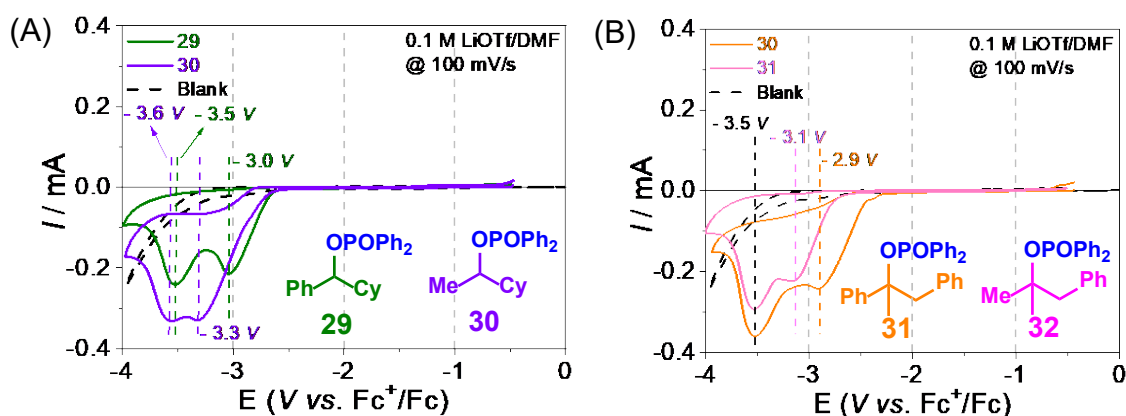


Figure S3. Cyclic voltammetry (CV) study of 2,3°-alcohol derivatives in 0.1 M LiOTf/DMF supporting electrolyte. (A) 2°-phosphinates. (B) 3°-phosphinates.

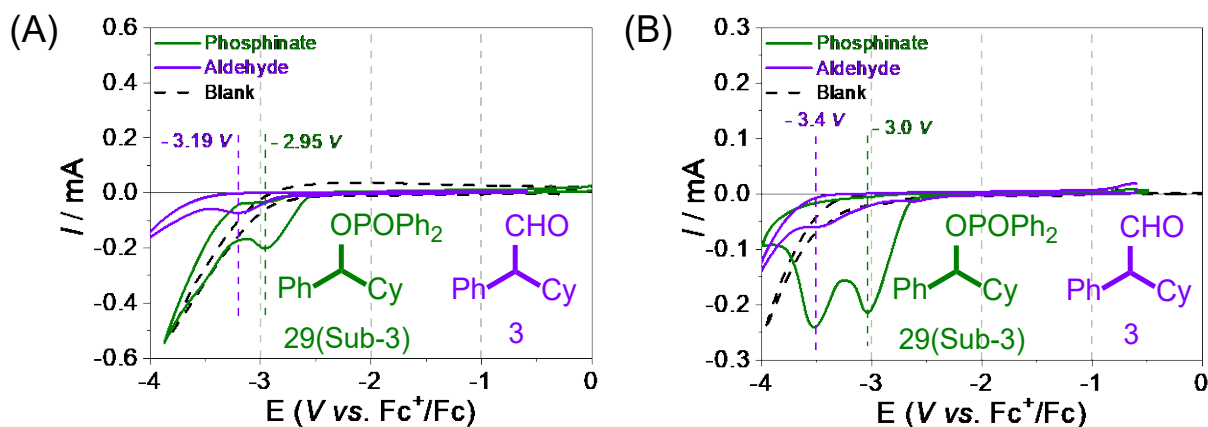


Figure S4. CV datas of aldehyde and phosphinate. (A) 0.1 M LiOTf/DMA as supporting electrolyte. (B) 0.1 M LiOTf/DMF as supporting electrolyte.

5. Gram-scale synthesis

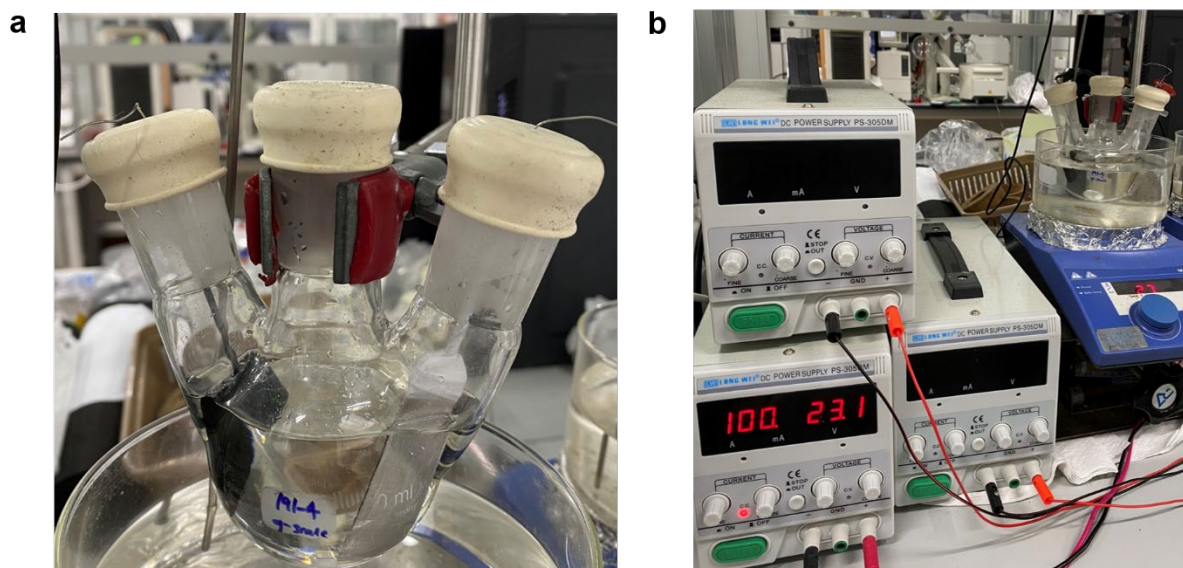
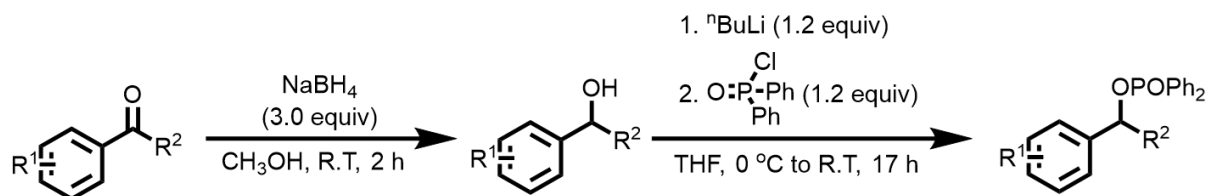


Figure S5. Set up for 20 mmol scale formylation of activated alcohols. a) Assembled reactor; b) Reaction in progress

An oven-dried, three-neck 250 mL Round bottom flask (RBF) was equipped with a magnetic stir bar, a rubber septum, a magnesium anode ($4 \times 1.5 \times 0.1 \text{ cm}^3$) and a carbon felt cathode ($0.3 \times 4 \times 4 \text{ cm}^3$) connected to septum lubber fitted with each separate electrical feed-throughs via 4H pencil leads (2 mm in diameter) and stainless wire (1 mm in diameter). This equipped reaction vessel was transferred into Ar-filled glove box. To this reaction vessel, substrate (20 mmol), LiOTf (3.43 g, 22 mmol), and DMF (100 mL) were added, and bring it was taken out of glovebox. Pre-stirring the resulting mixture for 2~3 minutes, and then the reaction mixture was electrolyzed at a constant current of 100 mA electrolysis was operated at 25 °C until passing 2.4 F/mol of a charge. Then, the crude mixture was further diluted with Et₂O (40 mL). The resulting mixture was washed with saturated NH₄Cl (aq. 30 mL) and H₂O (30 mL x 5 times). The organic layer was dried over with anhydrous magnesium sulfate and concentrated under reduced pressure. The residue was subjected to flash column chromatography on silica gel (eluted with *n*-hexane/ethyl acetate) to give **2** (1.07 g, 33% isolated yield) as a colorless oil.

6. Synthesis of starting materials

6.1. General Procedure A: Preparation of secondary benzyl alcohol derivatives.

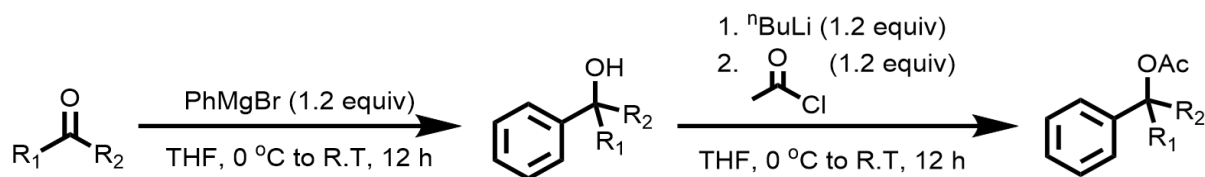


Scheme S1. Preparation of secondary benzyl alcohol derivatives.

General Procedure A: At $0\text{ }^\circ\text{C}$, to a solution of ketone (5.0 mmol, 1.0 equiv) in 20 mL MeOH was added NaBH_4 (567 mg, 3.0 equiv) in 2 portions. The reaction mixture was stirred at $0\text{ }^\circ\text{C}$ for 10 min at room temperature for 2 h. Solvent was removed under vacuum. The residue dissolved in EtOAc was washed with water. The organic layers were dried over MgSO_4 and evaporated. The crude mixture was used for the next step.

Phosphinates were prepared according to the literature procedure.¹ At $0\text{ }^\circ\text{C}$, $n\text{BuLi}$ (2.0 M in hexane, 1.2 equiv) was added dropwise to a solution of alcohol in 20 mL anhydrous THF under an Ar atmosphere. The resulting mixture was stirred at $0\text{ }^\circ\text{C}$ for 1 h, followed by the dropwise addition of diphenyl phosphinic chloride (1.2 equiv). Stirring continued for 1 h at $0\text{ }^\circ\text{C}$ and then the reaction was allowed to room temperature and stirred for additional 17 h. The mixture was quenched by sat. aq. NH_4Cl (10 mL). The solvent THF was removed under reduced pressure and the resulting mixture was dissolved in EtOAc. The organic layers were washed with water and dried over MgSO_4 . The solvent was evaporated under reduced pressure. The residue was purified by flash column chromatography on silica gel using a mixture of pentane and EtOAc as eluent to afford the desired product.

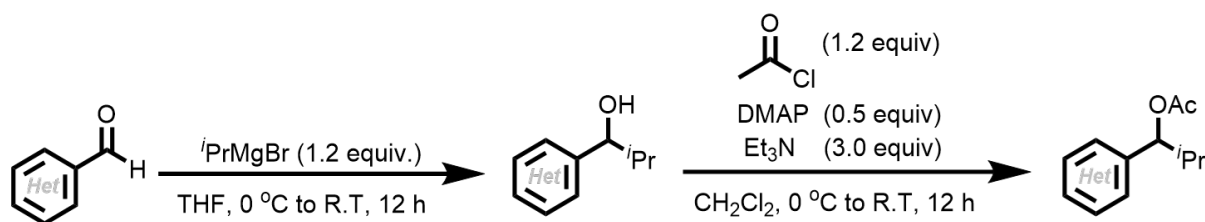
6.2. General Procedure **B**: Preparation of tertiary benzyl alcohol derivatives.



Scheme S2. Preparation of tertiary benzyl alcohol derivatives.

General Procedure **B**: At $0\text{ }^{\circ}\text{C}$, PhMgBr (1.0 M in THF, 1.2 equiv) was added dropwise to a solution of ketone (10.0 mmol, 1.0 equiv) in 20 mL anhydrous THF under an Ar atmosphere. The resulting mixture was stirred at $0\text{ }^{\circ}\text{C}$ for 1 h, and then allowed to at room temperature overnight. The mixture was quenched by sat. aq. NH_4Cl (10 mL). THF was removed under reduced pressure. The resulting mixture was dissolved in EtOAc and washed with water. The organic layers were dried over MgSO_4 and the solvent was evaporated under reduced pressure. The residue was purified by flash column chromatography on silica gel using a mixture of *n*-hexane and EtOAc as eluent. $n\text{BuLi}$ (2.0 M in hexane, 1.2 equiv) was added dropwise to a solution of isolated alcohol (5.0 mmol) in 20 mL anhydrous THF under an Ar atmosphere. The resulting mixture was stirred at $0\text{ }^{\circ}\text{C}$ for 1 h, followed by the dropwise addition of acetyl chloride (1.2 equiv). Stirring continued for 1 h at $0\text{ }^{\circ}\text{C}$ and then the reaction was allowed to room temperature and stirred for 12 h. The mixture was quenched by sat. aq. NH_4Cl (10 mL). THF was removed under reduced pressure. The resulting mixture was dissolved in EtOAc and washed with water. The organic layers were dried over MgSO_4 and the solvent was evaporated under reduced pressure. The residue was purified by flash column chromatography on silica gel using a mixture of *n*-hexane and EtOAc as eluent to afford pure product.

6.3. General Procedure C: Preparation of (Hetero)benzyl alcohol derivatives



Scheme S3. Preparation of Secondary-(Hetero)benzyl alcohol derivatives.

General Procedure C: At 0 °C, $i\text{PrMgBr}$ (2.0 M in THF, 6.0 mmol, 1.2 equiv) was added dropwise to a solution of aldehyde (5.0 mmol, 1.0 equiv) in 20 mL anhydrous THF under an Ar atmosphere. The resulting mixture was stirred at 0 °C for 1 h, and allowed to room temperature overnight. The mixture was quenched by sat. aq. NH_4Cl (10 mL). THF was removed under reduced pressure. The resulting mixture was dissolved in EtOAc and washed with water. The organic layers were dried over MgSO_4 and the solvent was evaporated under reduced pressure. The residue was purified by flash column chromatography on silica gel using a mixture of *n*-hexane and EtOAc as an eluent. At 0 °C, the isolated alcohol (4.0 mmol) was treated with 20 mL CH_2Cl_2 , Et_3N (12.0 mmol, 3.0 equiv), 4-dimethylaminopyridine (2.0 mmol, 0.5 equiv) and acetyl chloride (4.8 mmol, 1.2 equiv). The reaction mixture was stirred at room temperature overnight. Solvent was removed under vacuum and the residue was purified by flash column chromatography using a mixture of *n*-hexane and EtOAc to give the desired carboxylates.

7. Structural determination of **27**.

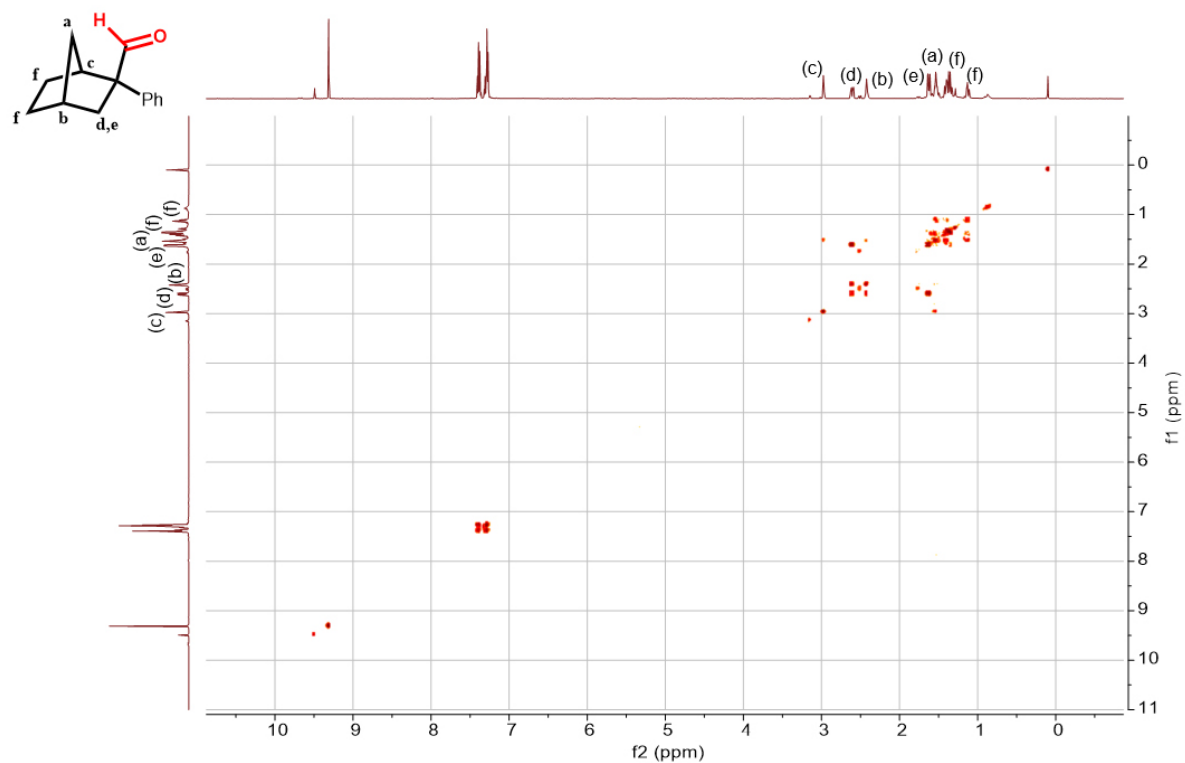


Figure S6. 2D-COSY NMR spectrum of **27** (500 MHz, CDCl_3).

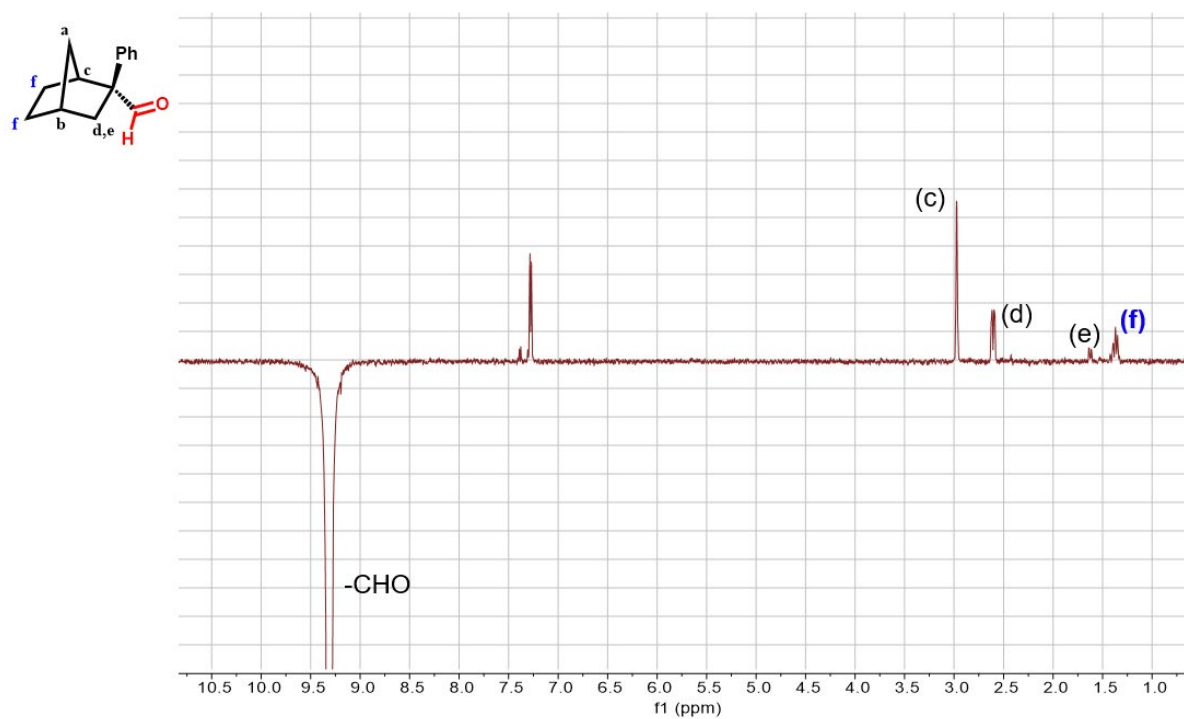
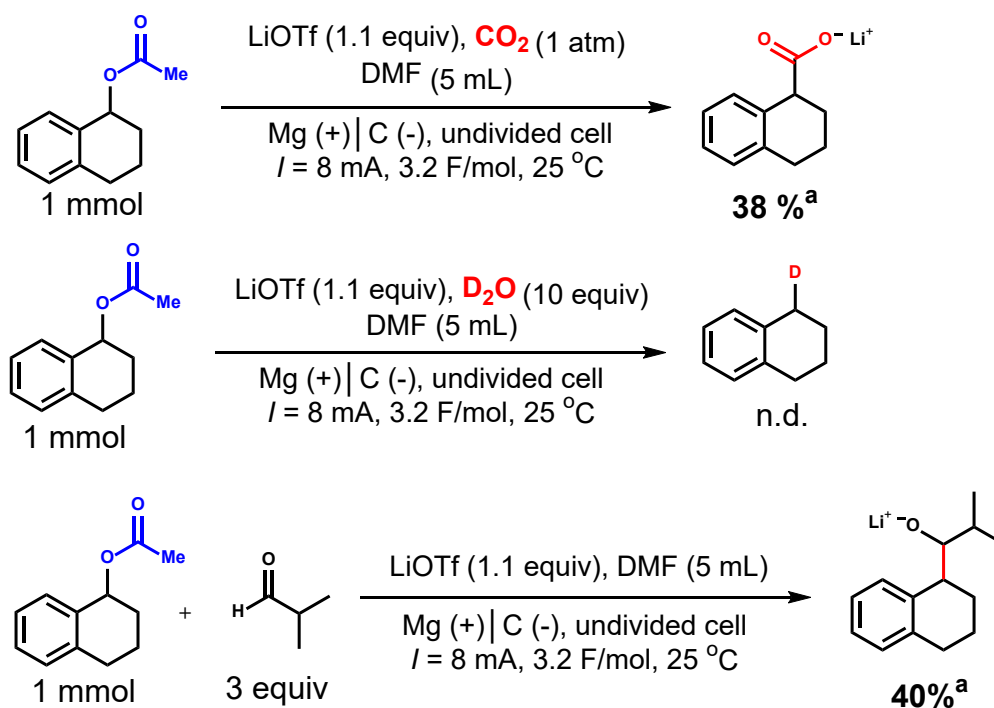


Figure S7. ^1H -1D NOESY NMR spectrum of **27** at 9.310 ppm (500 MHz, CDCl_3).

8. Reactivity towards other electrophiles

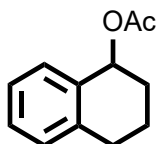


a) ^1H NMR yield using 0.2 mmol mesitylene as internal standard

Scheme S4. Reactivity towards other electrophiles

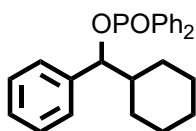
9. Characterization of compounds

9.1. Starting materials



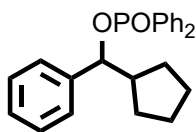
1 (2-sub)

Colorless liquid. $^1\text{H NMR}$ (500 MHz, CDCl_3) δ 7.31 – 7.17 (m, 3H), 7.14 (dd, J = 7.5, 1.4 Hz, 1H), 6.02 (t, J = 4.4 Hz, 1H), 2.88 (dt, J = 16.3, 4.9 Hz, 1H), 2.76 (ddd, J = 16.9, 8.8, 6.2 Hz, 1H), 2.09 (s, 3H), 2.05 – 1.94 (m, 3H), 1.89 – 1.79 (m, 1H). $^{13}\text{C}\{^1\text{H}\}$ NMR (126 MHz, CDCl_3) δ 170.8, 138.0, 134.6, 129.5, 129.1, 128.1, 126.1, 70.0, 29.1, 29.0, 21.5, 18.8. Data are consistent with the literature.²



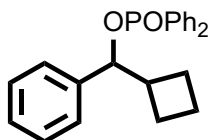
3-sub

White liquid. $^1\text{H NMR}$ (500 MHz, CDCl_3) δ 7.87 – 7.79 (m, 2H), 7.54 – 7.40 (m, 5H), 7.32 (td, J = 7.4, 1.5 Hz, 1H), 7.21 – 7.09 (m, 7H), 5.04 (dd, J = 9.9, 7.5 Hz, 1H), 2.06 (dt, J = 13.1, 3.2 Hz, 1H), 1.89 (tdt, J = 11.2, 7.0, 3.4 Hz, 1H), 1.80 – 1.71 (m, 1H), 1.62 (td, J = 14.1, 12.9, 7.2 Hz, 2H), 1.45 – 1.32 (m, 1H), 1.29 – 0.99 (m, 5H), 0.88 (qd, J = 12.3, 3.7 Hz, 1H). $^{13}\text{C}\{^1\text{H}\}$ NMR (126 MHz, CDCl_3) δ 139.6, 133.0, 132.0, 132.0, 131.9, 131.9, 131.6, 131.6, 131.5, 131.4, 130.9, 128.5, 128.3, 127.9, 127.9, 127.8, 127.7, 127.3, 82.6, 82.6, 44.4, 44.4, 29.4, 28.5, 26.3, 25.9. $^{31}\text{P}\{^1\text{H}\}$ NMR (162 MHz, CDCl_3) δ 30.7. HRMS (ESI) calculated for $\text{C}_{25}\text{H}_{27}\text{O}_2\text{P} [\text{M}+\text{Na}]^+$: 413.1641, Found: 413.1644.



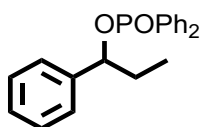
4-sub

Colorless oil. $^1\text{H NMR}$ (500 MHz, CDCl_3) δ 7.88 – 7.73 (m, 2H), 7.57 – 7.39 (m, 5H), 7.32 (td, J = 7.4, 1.4 Hz, 1H), 7.19 – 7.12 (m, 7H), 5.13 (t, J = 9.1 Hz, 1H), 2.57 – 2.38 (m, 2H), 2.00 – 1.88 (m, 1H), 1.73 – 1.61 (m, 2H), 1.60 – 1.51 (m, 2H), 1.50 – 1.43 (m, 1H), 1.42 – 1.30 (m, 1H), 1.19 – 1.06 (m, 1H). $^{13}\text{C}\{^1\text{H}\}$ NMR (101 MHz, CDCl_3) δ 140.6, 140.5, 132.0, 131.9, 131.9, 131.6, 131.6, 131.4, 131.3, 128.4, 128.3, 128.0, 127.9, 127.8, 127.8, 127.1, 82.4, 82.4, 47.2, 47.1, 30.2, 29.0, 25.3. $^{31}\text{P}\{^1\text{H}\}$ NMR (162 MHz, CDCl_3) δ 30.7. HRMS (ESI) calculated for $\text{C}_{24}\text{H}_{25}\text{O}_2\text{P} [\text{M}+\text{Na}]^+$: 399.1484, Found: 399.1488.



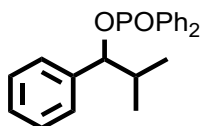
5-sub

Light yellow oil. $^1\text{H NMR}$ (500 MHz, CDCl_3) δ 7.82 (dddd, $J = 12.3, 8.6, 7.4, 1.4$ Hz, 3H), 7.57 – 7.48 (m, 3H), 7.44 (tdd, $J = 6.6, 3.5, 1.4$ Hz, 2H), 7.38 – 7.32 (m, 1H), 7.24 – 7.15 (m, 7H), 5.25 (dd, $J = 9.6, 8.1$ Hz, 1H), 2.91 – 2.78 (m, 1H), 2.18 – 2.07 (m, 1H), 2.06 – 1.98 (m, 1H), 1.83 – 1.68 (m, 4H). $^{13}\text{C}\{^1\text{H}\}$ NMR (126 MHz, CDCl_3) δ 139.5, 139.5, 132.6, 132.6, 132.1, 132.0, 132.0, 131.9, 131.7, 131.7, 131.7, 131.5, 131.4, 131.1, 128.7, 128.6, 128.4, 128.3, 128.1, 128.1, 128.0, 127.9, 126.8, 81.8, 81.8, 41.7, 41.6, 25.3, 24.1, 17.5. $^{31}\text{P}\{^1\text{H}\}$ NMR (202 MHz, CDCl_3) δ 31.0. HRMS (ESI) calculated for $\text{C}_{23}\text{H}_{23}\text{O}_2\text{P}$ $[\text{M}+\text{Na}]^+$: 385.1328, Found: 385.1330.



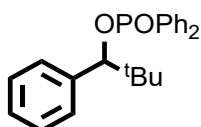
6-sub

Light yellow oil. $^1\text{H NMR}$ (500 MHz, CDCl_3) δ 7.83 (ddt, $J = 12.3, 6.9, 1.4$ Hz, 2H), 7.58 (ddt, $J = 12.4, 6.7, 1.4$ Hz, 2H), 7.53 – 7.48 (m, 1H), 7.47 – 7.41 (m, 2H), 7.41 – 7.36 (m, 1H), 7.29 – 7.17 (m, 7H), 5.27 (ddd, $J = 9.4, 7.0, 5.8$ Hz, 1H), 2.10 (tt, $J = 13.6, 7.4$ Hz, 1H), 1.95 (dt, $J = 13.7, 7.3$ Hz, 1H), 0.84 (t, $J = 7.4$ Hz, 3H). $^{13}\text{C}\{^1\text{H}\}$ NMR (126 MHz, CDCl_3) δ 140.5, 140.4, 133.0, 132.0, 132.0, 132.0, 131.9, 131.8, 131.8, 131.5, 131.4, 131.0, 128.6, 128.5, 128.4, 128.2, 128.2, 128.1, 127.9, 126.6, 79.6, 79.6, 31.3, 9.5. $^{31}\text{P}\{^1\text{H}\}$ NMR (202 MHz, CDCl_3) δ 31.0. Data are consistent with the literature.³



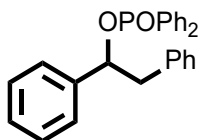
7-sub

Colorless oil. $^1\text{H NMR}$ (400 MHz, CDCl_3) δ 7.89 – 7.78 (m, 2H), 7.56 – 7.41 (m, 5H), 7.35 – 7.31 (m, 1H), 7.23 – 7.11 (m, 7H), 5.07 (dd, $J = 9.8, 7.1$ Hz, 1H), 2.22 (hept, $J = 6.8$ Hz, 1H), 1.04 (d, $J = 6.7$ Hz, 3H), 0.79 (d, $J = 6.8$ Hz, 3H). $^{13}\text{C}\{^1\text{H}\}$ NMR (126 MHz, CDCl_3) δ 139.4, 139.4, 133.0, 132.0, 132.0, 132.0, 131.9, 131.7, 131.6, 131.5, 131.4, 130.9, 128.5, 128.4, 128.0, 127.9, 127.7, 127.2, 83.2, 83.2, 35.1, 35.0, 19.0, 18.1. $^{31}\text{P}\{^1\text{H}\}$ NMR (202 MHz, CDCl_3) δ 30.9. HRMS (ESI) calculated for $\text{C}_{22}\text{H}_{23}\text{O}_2\text{P}$ $[\text{M}+\text{Na}]^+$: 373.1328, Found: 373.1329.



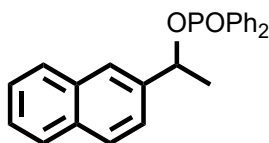
8-sub

White solid. (m.p.: 111-115°C) $^1\text{H NMR}$ (500 MHz, CDCl_3) δ 7.93 – 7.79 (m, 2H), 7.50 – 7.37 (m, 5H), 7.25 – 7.21 (m, 1H), 7.13 – 7.06 (m, 7H), 5.06 (d, $J = 10.4$ Hz, 1H), 0.97 (s, 9H). $^{13}\text{C}\{^1\text{H}\}$ NMR (126 MHz, CDCl_3) δ 138.4, 138.4, 132.8, 132.0, 132.0, 131.9, 131.9, 131.7, 131.5, 131.5, 131.4, 130.8, 128.5, 128.4, 128.1, 127.8, 127.7, 127.5, 127.3, 85.3, 85.2, 36.1, 36.0, 26.2. $^{31}\text{P}\{^1\text{H}\}$ NMR (202 MHz, CDCl_3) δ 30.6. HRMS (ESI) calculated for $\text{C}_{23}\text{H}_{25}\text{O}_2\text{P}$ $[\text{M}+\text{Na}]^+$: 387.1484, Found: 387.1485.



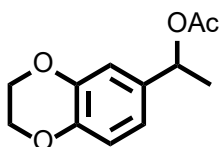
9-sub

White solid. (m.p.: 110-114°C) $^1\text{H NMR}$ (500 MHz, CDCl_3) δ 7.71 – 7.57 (m, 4H), 7.48 (t, J = 7.5 Hz, 1H), 7.37 (td, J = 7.7, 2.9 Hz, 3H), 7.27 (td, J = 7.6, 3.1 Hz, 2H), 7.23 – 7.19 (m, 3H), 7.17 (dd, J = 4.7, 2.2 Hz, 5H), 7.01 (dd, J = 6.6, 2.9 Hz, 2H), 5.54 (dt, J = 8.8, 6.8 Hz, 1H), 3.40 (dd, J = 13.5, 6.4 Hz, 1H), 3.22 (dd, J = 13.5, 7.1 Hz, 1H). $^{13}\text{C}\{^1\text{H}\}$ NMR (126 MHz, CDCl_3) δ 140.0, 140.0, 136.5, 132.6, 132.1, 132.0, 132.0, 131.9, 131.9, 131.8, 131.6, 131.5, 131.5, 131.1, 130.0, 128.5, 128.4, 128.3, 128.2, 128.0, 126.7, 126.6, 79.0, 79.0, 45.3, 45.3. $^{31}\text{P}\{^1\text{H}\}$ NMR (202 MHz, CDCl_3) δ 31.4. Data are consistent with the literature.⁴



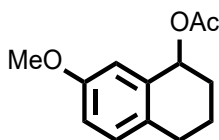
10-sub

White solid. (m.p.: 78-81°C) $^1\text{H NMR}$ (400 MHz, CDCl_3) δ 7.91 – 7.74 (m, 5H), 7.70 – 7.62 (m, 3H), 7.55 – 7.43 (m, 6H), 7.41 – 7.35 (m, 1H), 7.30 – 7.22 (m, 2H), 5.70 (dq, J = 9.2, 6.5 Hz, 1H), 1.76 (d, J = 6.5 Hz, 3H). $^{13}\text{C}\{^1\text{H}\}$ NMR (126 MHz, CDCl_3) δ 139.4, 133.1, 133.1, 132.1, 132.1, 132.0, 132.0, 131.9, 131.8, 131.6, 131.5, 128.5, 128.4, 128.4, 128.3, 128.2, 128.1, 127.7, 126.2, 126.1, 125.0, 123.9, 74.8, 74.8, 25.0, 24.9. $^{31}\text{P}\{^1\text{H}\}$ NMR (202 MHz, CDCl_3) δ 31.3. HRMS (ESI) calculated for $\text{C}_{24}\text{H}_{21}\text{O}_2\text{P}$ $[\text{M}+\text{Na}]^+$: 395.1171, Found: 395.1183.



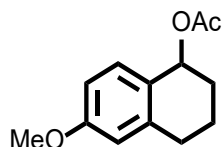
11-sub

Colorless oil. $^1\text{H NMR}$ (500 MHz, CDCl_3) δ 6.87 (d, J = 1.3 Hz, 1H), 6.82 (d, J = 1.4 Hz, 2H), 5.77 (q, J = 6.6 Hz, 1H), 4.22 (s, 4 Hz), 2.04 (s, 3H), 1.49 (d, J = 6.6 Hz, 3H). $^{13}\text{C}\{^1\text{H}\}$ NMR (126 MHz, CDCl_3) δ 170.3, 143.4, 143.2, 135.0, 119.3, 117.2, 115.2, 71.9, 64.3, 64.3, 22.0, 21.3. Data are consistent with the literature.⁵



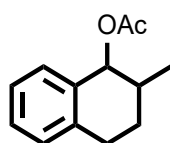
12-sub

Yellow oil. $^1\text{H NMR}$ (500 MHz, CDCl_3) δ 7.27 – 7.23 (m, 1H), 7.06 – 6.97 (m, 2H), 6.18 (t, J = 4.4 Hz, 1H), 3.99 (s, 3H), 3.01 (dt, J = 15.9, 4.8 Hz, 1H), 2.89 (ddd, J = 16.5, 8.8, 5.7 Hz, 1H), 2.31 (s, 3H), 2.23 – 2.11 (m, 3H), 2.03 (ttt, J = 15.1, 6.0, 5.2, 2.5 Hz, 1H). $^{13}\text{C}\{^1\text{H}\}$ NMR (126 MHz, CDCl_3) δ 170.9, 157.8, 135.4, 130.1, 130.0, 114.8, 113.7, 70.2, 55.3, 29.1, 28.2, 21.5, 19.0. Data are consistent with the literature.⁶



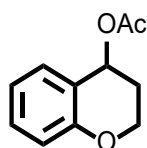
13-sub

Yellow oil. **¹H NMR** (400 MHz, CDCl₃) δ 7.21 (d, *J* = 8.5 Hz, 1H), 6.77 – 6.71 (m, 1H), 6.64 (d, *J* = 2.7 Hz, 1H), 5.96 (t, *J* = 3.8 Hz, 1H), 3.79 (s, 3H), 2.88 – 2.66 (m, 2H), 2.06 (s, 3H), 2.03 – 1.91 (m, 3H), 1.85 – 1.76 (m, 1H). **¹³C{¹H} NMR** (126 MHz, CDCl₃) δ 170.9, 159.3, 139.5, 131.0, 126.9, 113.4, 112.5, 69.8, 55.2, 29.4, 29.2, 21.6, 18.6. Data are consistent with the literature.⁷



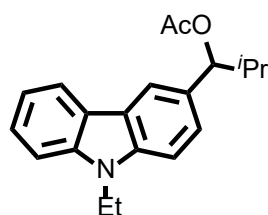
14-sub

Colorless solid. (m.p.: 55-57°C) **¹H NMR** (500 MHz, CDCl₃) δ 7.20 (qd, *J* = 6.7, 2.0 Hz, 3H), 7.13 (d, *J* = 7.3 Hz, 1H), 5.79 (d, *J* = 6.7 Hz, 1H), 2.90 – 2.78 (m, 2H), 2.15 (s, 3H), 2.06 (dddd, *J* = 22.8, 12.8, 6.5, 3.4 Hz, 2H), 1.64 (dtd, *J* = 13.9, 8.1, 5.9 Hz, 1H), 1.04 (d, *J* = 6.9 Hz, 3H). **¹³C{¹H} NMR** (126 MHz, CDCl₃) δ 171.2, 137.4, 134.5, 128.8, 128.8, 127.8, 126.1, 75.7, 33.8, 27.1, 26.9, 21.4, 17.3. Data are consistent with the literature.⁸



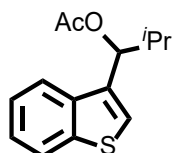
15-sub

Colorless oil. **¹H NMR** (500 MHz, CDCl₃) δ 7.29 (dd, *J* = 7.7, 1.7 Hz, 1H), 7.23 (ddd, *J* = 8.7, 7.3, 1.7 Hz, 1H), 6.91 (td, *J* = 7.5, 1.2 Hz, 1H), 6.86 (dd, *J* = 8.3, 1.2 Hz, 1H), 4.30 (dtd, *J* = 11.1, 4.1, 1.0 Hz, 1H), 4.24 (td, *J* = 11.3, 2.5 Hz, 1H), 2.22 (ddt, *J* = 14.7, 11.6, 4.2 Hz, 1H), 2.14 – 2.03 (m, 4H). **¹³C{¹H} NMR** (126 MHz, CDCl₃) δ 170.5, 155.2, 130.7, 130.2, 120.6, 120.2, 117.1, 65.2, 62.1, 28.4, 21.4. Data are consistent with the literature.⁹



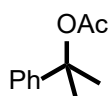
16-sub

Orange oil. **¹H NMR** (500 MHz, CDCl₃) δ 8.10 (dt, *J* = 7.8, 1.0 Hz, 1H), 8.04 (d, *J* = 1.7 Hz, 1H), 7.50 – 7.31 (m, 4H), 7.28 – 7.15 (m, 1H), 5.64 (d, *J* = 8.4 Hz, 1H), 4.36 (q, *J* = 7.3 Hz, 2H), 2.24 (dp, *J* = 8.4, 6.7 Hz, 1H), 2.09 (s, 3H), 1.43 (t, *J* = 7.2 Hz, 3H), 1.06 (d, *J* = 6.6 Hz, 3H), 0.81 (d, *J* = 6.8 Hz, 3H). **¹³C{¹H} NMR** (126 MHz, CDCl₃) δ 170.6, 140.3, 139.6, 130.4, 125.8, 125.1, 122.9, 122.7, 120.5, 119.4, 118.9, 108.5, 108.1, 82.0, 37.6, 33.9, 21.4, 19.1, 19.0, 13.9. **HRMS** (ESI) calculated for C₂₀H₂₃NO₂ [M+Na]⁺: 332.1621, Found: 332.1619.



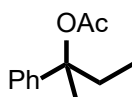
17-sub

Colorless oil. $^1\text{H NMR}$ (500 MHz, CDCl_3) δ 7.98 – 7.92 (m, 1H), 7.88 – 7.81 (m, 1H), 7.41 – 7.31 (m, 3H), 5.92 (d, J = 7.9 Hz, 1H), 2.38 (dp, J = 8.0, 6.6 Hz, 1H), 2.09 (s, 3H), 1.03 (d, J = 6.6 Hz, 3H), 0.87 (d, J = 6.8 Hz, 3H). $^{13}\text{C}\{^1\text{H}\}$ NMR (126 MHz, CDCl_3) δ 170.4, 140.6, 137.5, 135.0, 124.4, 124.1, 124.1, 122.9, 122.7, 76.5, 32.5, 21.1, 19.2, 18.6. HRMS (ESI) calculated for $\text{C}_{14}\text{H}_{16}\text{O}_2\text{S}$ $[\text{M}+\text{Na}]^+$: 271.0763, Found: 271.0769.



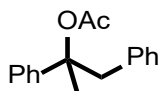
18-sub

Light yellow liquid. $^1\text{H NMR}$ (500 MHz, CDCl_3) δ 7.39 – 7.31 (m, 4H), 7.27 – 7.22 (m, 1H), 2.04 (s, 3H), 1.77 (s, 6H). $^{13}\text{C}\{^1\text{H}\}$ NMR (126 MHz, CDCl_3) δ 169.8, 145.9, 128.3, 127.0, 124.2, 81.5, 28.7, 22.3. Data are consistent with the literature.



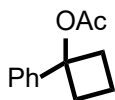
19-sub

Colorless liquid. $^1\text{H NMR}$ (400 MHz, CDCl_3) δ 7.36 – 7.28 (m, 4H), 7.27 – 7.20 (m, 1H), 2.07 (s, 3H), 2.03 (tt, J = 7.2, 3.8 Hz, 2H), 1.81 (s, 3H), 0.79 (t, J = 7.4 Hz, 3H). $^{13}\text{C}\{^1\text{H}\}$ NMR (126 MHz, CDCl_3) δ 169.7, 144.9, 128.1, 126.8, 124.7, 84.3, 35.1, 24.4, 22.2, 8.1. Data are consistent with the literature.¹⁰



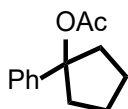
20-sub

Light yellow oil. $^1\text{H NMR}$ (500 MHz, CDCl_3) δ 7.35 – 7.31 (m, 2H), 7.30 – 7.19 (m, 6H), 6.98 – 6.90 (m, 2H), 3.31 (d, J = 13.5 Hz, 1H), 3.19 (d, J = 13.5 Hz, 1H), 2.08 (s, 3H), 1.86 (s, 3H). $^{13}\text{C}\{^1\text{H}\}$ NMR (126 MHz, CDCl_3) δ 169.7, 144.4, 136.3, 130.8, 128.1, 127.7, 127.0, 126.5, 124.9, 83.7, 49.2, 24.1, 22.3. HRMS (ESI) calculated for $\text{C}_{17}\text{H}_{18}\text{O}_2$ $[\text{M}+\text{Na}]^+$: 277.1199, Found: 277.1201.



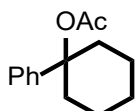
21-sub

Colorless liquid. $^1\text{H NMR}$ (400 MHz, CDCl_3) δ 7.53 – 7.49 (m, 2H), 7.41 – 7.35 (m, 2H), 7.32 – 7.26 (m, 1H), 2.74 – 2.56 (m, 4H), 2.01 (s, 4H), 1.77 (dp, J = 11.2, 8.8 Hz, 1H). $^{13}\text{C}\{^1\text{H}\}$ NMR (126 MHz, CDCl_3) δ 169.5, 142.7, 128.3, 127.3, 125.6, 82.2, 34.9, 21.7, 14.2. Datas are consistent with the literature.¹¹



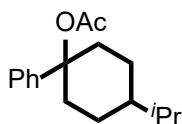
22-sub

Colorless oil. **¹H NMR** (400 MHz, CDCl₃) δ 7.40 – 7.36 (m, 2H), 7.32 (ddd, *J* = 7.8, 6.7, 1.2 Hz, 2H), 7.26 – 7.20 (m, 1H), 2.44 (dddd, *J* = 12.7, 7.2, 2.9, 1.3 Hz, 2H), 2.19 – 2.05 (m, 2H), 2.00 (s, 3H), 1.93 – 1.69 (m, 4H). **¹³C{¹H} NMR** (126 MHz, CDCl₃) δ 170.0, 143.6, 128.1, 127.0, 125.2, 91.5, 38.7, 23.4, 22.1. **HRMS** (ESI) calculated for C₁₃H₁₆O₂ [M+Na]⁺: 227.1043, Found: 227.1042.



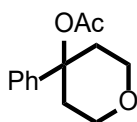
23-sub

Yellow liquid. **¹H NMR** (500 MHz, CDCl₃) δ 7.38 – 7.31 (m, 4H), 7.26 – 7.22 (m, 1H), 2.56 – 2.45 (m, 2H), 2.06 (s, 3H), 1.80 – 1.70 (m, 3H), 1.67 (ddt, *J* = 10.0, 6.5, 3.0 Hz, 4H), 1.31 (dtd, *J* = 12.4, 9.2, 4.7 Hz, 1H). **¹³C{¹H} NMR** (126 MHz, CDCl₃) δ 169.6, 145.7, 128.3, 127.0, 124.5, 82.5, 36.3, 25.4, 22.1, 22.1. Data are consistent with the literature.¹²



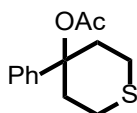
24-sub

Colorless oil. **¹H NMR** (500 MHz, CDCl₃) δ 7.52 – 7.47 (m, 2H), 7.37 – 7.30 (m, 2H), 7.27 – 7.21 (m, 1H), 2.69 – 2.62 (m, 2H), 1.99 – 1.93 (m, 2H), 1.92 (s, 3H), 1.75 (dd, *J* = 10.8, 5.4 Hz, 2H), 1.48 – 1.38 (m, 1H), 1.32 – 1.23 (m, 3H), 0.83 (d, *J* = 6.7 Hz, 6H). **¹³C{¹H} NMR** (126 MHz, CDCl₃) δ 169.6, 142.1, 128.0, 127.3, 126.7, 83.1, 42.7, 35.0, 30.7, 26.1, 22.4, 20.4. **HRMS** (ESI) calculated for C₁₇H₂₄O₂ [M+Na]⁺: 283.1669, Found: 283.1671.



25-sub

Light yellow oil. **¹H NMR** (500 MHz, CDCl₃) δ 7.39 – 7.34 (m, 4H), 7.28 (ddp, *J* = 8.1, 5.4, 2.6 Hz, 1H), 3.90 (ddd, *J* = 11.6, 5.0, 2.2 Hz, 2H), 3.81 (td, *J* = 11.7, 2.0 Hz, 2H), 2.44 (dq, *J* = 14.5, 2.6 Hz, 2H), 2.14 (ddd, *J* = 13.8, 11.7, 4.9 Hz, 2H), 2.07 (s, 3H). **¹³C{¹H} NMR** (126 MHz, CDCl₃) δ 169.5, 144.0, 128.5, 127.5, 124.5, 79.7, 63.7, 36.2, 22.1. **HRMS** (ESI) calculated for C₁₃H₁₆O₃ [M+Na]⁺: 243.0992, Found: 243.1003.



26-sub

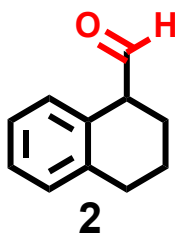
White solid. (m.p.: 83-87°C) **¹H NMR** (500 MHz, CDCl₃) δ 7.41 – 7.20 (m, 5H), 3.08 (ddd, *J* = 14.2, 12.5, 2.3 Hz, 2H), 2.88 – 2.72 (m, 2H), 2.63 – 2.43 (m, 2H), 2.11 (s, 5H). **¹³C{¹H} NMR** (126 MHz, CDCl₃) δ 169.3, 145.2, 128.6, 127.4, 124.1, 80.9, 37.4, 24.1, 22.0. **HRMS** (ESI) calculated for C₁₃H₁₆O₂S [M+Na]⁺: 259.0763, Found: 259.0765.



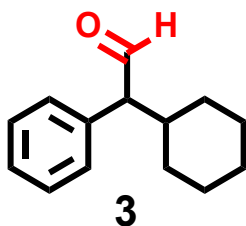
27-sub

Colorless oil. **¹H NMR** (500 MHz, CDCl₃) δ 7.44 (d, *J* = 7.7 Hz, 2H), 7.32 (t, *J* = 7.7 Hz, 2H), 7.22 (t, *J* = 7.5 Hz, 1H), 3.16 (d, *J* = 5.1 Hz, 1H), 2.50 – 2.25 (m, 2H), 1.98 (s, 3H), 1.89 – 1.76 (m, 2H), 1.57 (d, *J* = 9.6 Hz, 2H), 1.45 – 1.25 (m, 3H). **¹³C{¹H} NMR** (126 MHz, CDCl₃) δ 170.1, 145.8, 128.1, 126.7, 125.8, 88.4, 45.8, 45.5, 37.5, 36.6, 29.0, 23.0, 21.8. **HRMS** (ESI) calculated for C₁₅H₁₈O₂ [M+Na]⁺: 253.1199, Found: 253.1206.

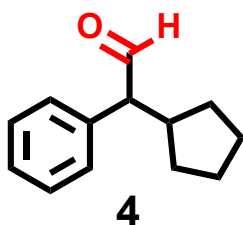
9.2. Products



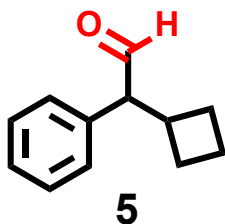
Light yellow oil. **¹H NMR** (500 MHz, CDCl₃) δ 9.73 (d, *J* = 2.2 Hz, 1H), 7.30 – 7.18 (m, 4H), 3.69 – 3.61 (m, 1H), 2.85 (t, *J* = 6.4 Hz, 2H), 2.34 – 2.24 (m, 1H), 1.99 (dddd, *J* = 13.3, 9.8, 6.4, 3.7 Hz, 1H), 1.86 (dq, *J* = 13.4, 6.4, 3.4 Hz, 2H). **¹³C{¹H} NMR** (126 MHz, CDCl₃) δ 202.1, 138.1, 130.9, 129.8, 129.7, 127.2, 126.2, 51.7, 29.2, 23.0, 20.5. **HRMS** (ESI) calculated for C₁₁H₁₂O [M+Na]⁺: 183.0780, Found: 183.0773.



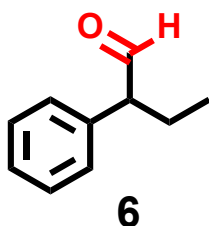
Colorless oil. **¹H NMR** (500 MHz, CDCl₃) δ 9.70 (d, *J* = 3.5 Hz, 1H), 7.38 – 7.33 (m, 2H), 7.31 – 7.26 (m, 1H), 7.20 – 7.16 (m, 2H), 3.25 (dd, *J* = 9.6, 3.5 Hz, 1H), 2.16 – 2.02 (m, 1H), 1.88 – 1.80 (m, 1H), 1.75 (dtd, *J* = 13.1, 3.5, 1.8 Hz, 1H), 1.68 – 1.61 (m, 2H), 1.41 (ddt, *J* = 13.2, 3.7, 2.0 Hz, 1H), 1.36 – 1.00 (m, 5H), 0.84 – 0.75 (m, 1H). **¹³C{¹H} NMR** (101 MHz, CDCl₃) δ 201.2, 135.3, 129.3, 128.9, 127.4, 65.9, 38.2, 31.8, 30.2, 26.3, 26.1, 26.1. **HRMS** (ESI) calculated for C₁₄H₁₈O [M+Na]⁺: 225.1248, Found: 225.1250.



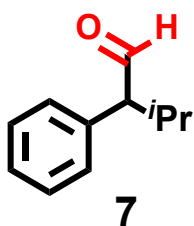
Colorless oil. **¹H NMR** (400 MHz, CDCl₃) δ 9.71 (d, *J* = 3.1 Hz, 1H), 7.41 – 7.35 (m, 2H), 7.34 – 7.29 (m, 1H), 7.25 – 7.21 (m, 2H), 3.32 (dd, *J* = 10.5, 3.1 Hz, 1H), 2.60 – 2.50 (m, 1H), 2.04 – 1.94 (m, 1H), 1.76 – 1.61 (m, 1H), 1.56 – 1.43 (m, 1H), 1.28 (dq, *J* = 12.7, 8.4 Hz, 1H), 1.15 – 1.03 (m, 1H). **¹³C{¹H} NMR** (101 MHz, CDCl₃) δ 200.8, 136.2, 129.1, 128.9, 127.4, 65.3, 40.3, 31.1, 30.8, 25.2, 24.5. **HRMS** (ESI) calculated for C₁₃H₁₆O [M+Na]⁺: 211.1093, Found: 211.1085.



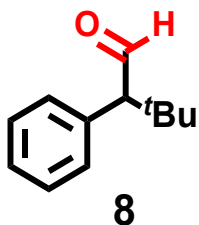
Light yellow oil. **¹H NMR** (500 MHz, CDCl₃) δ 9.63 (d, *J* = 2.4 Hz, 1H), 7.36 (t, *J* = 7.5 Hz, 2H), 7.31 – 7.27 (m, 1H), 7.20 – 7.14 (m, 2H), 3.53 (dd, *J* = 10.5, 2.4 Hz, 1H), 2.98 (dt, *J* = 11.1, 7.4 Hz, 1H), 2.30 – 2.16 (m, 1H), 1.96 – 1.78 (m, 4H), 1.72 – 1.61 (m, 1H). **¹³C{¹H} NMR** (126 MHz, CDCl₃) δ 200.5, 135.1, 129.0, 127.5, 65.7, 35.5, 27.5, 26.5, 18.6. **HRMS** (ESI) calculated for C₁₂H₁₄O [M+Na]⁺: 197.0937, Found: 197.0930.



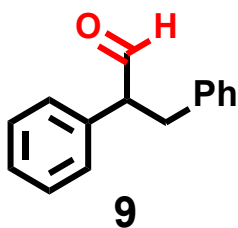
Colorless oil. **¹H NMR** (500 MHz, CDCl₃) δ 9.68 (d, *J* = 2.0 Hz, 1H), 7.38 (t, *J* = 7.5 Hz, 2H), 7.30 (t, *J* = 7.4 Hz, 1H), 7.19 (d, *J* = 7.5 Hz, 2H), 3.41 (t, *J* = 7.5 Hz, 1H), 2.12 (dt, *J* = 14.1, 7.1 Hz, 1H), 1.77 (dt, *J* = 14.5, 7.4 Hz, 1H), 0.91 (t, *J* = 7.4 Hz, 3H). **¹³C{¹H} NMR** (126 MHz, CDCl₃) δ 201.1, 136.3, 129.0, 128.8, 127.5, 60.9, 22.3, 11.7. **HRMS** (ESI) calculated for C₁₀H₁₂O [M+Na]⁺: 197.0937, Found: 197.0930.



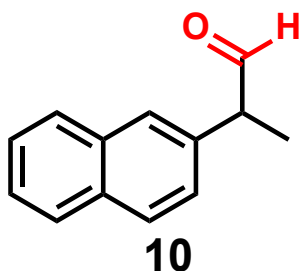
Colorless oil. **¹H NMR** (400 MHz, CDCl₃) δ 9.73 (d, *J* = 3.3 Hz, 1H), 7.42 – 7.35 (m, 2H), 7.34 – 7.27 (m, 1H), 7.24 – 7.19 (m, 2H), 3.21 (dd, *J* = 9.5, 3.3 Hz, 1H), 2.44 (dp, *J* = 9.5, 6.7 Hz, 1H), 1.07 (d, *J* = 6.6 Hz, 3H), 0.80 (d, *J* = 6.7 Hz, 3H). **¹³C{¹H} NMR** (101 MHz, CDCl₃) δ 201.2, 135.5, 129.3, 128.9, 127.5, 66.9, 28.8, 21.2, 20.1. **HRMS** (ESI) calculated for C₁₁H₁₄O [M+Na]⁺: 185.0937, Found: 185.0938.



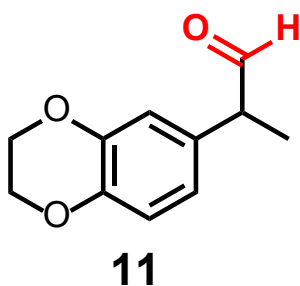
Colorless oil. **¹H NMR** (400 MHz, CDCl₃) δ 10.01 (d, *J* = 3.4 Hz, 1H), 7.38 – 7.29 (m, 3H), 7.25 – 7.20 (m, 2H), 3.29 (d, *J* = 3.4 Hz, 1H), 1.03 (s, 9H). **¹³C{¹H} NMR** (101 MHz, CDCl₃) δ 202.3, 135.3, 130.4, 128.3, 127.3, 68.4, 29.7, 28.2. **HRMS** (ESI) calculated for C₁₂H₁₈O [MH₂+Na]⁺: 201.1250, Found: 201.1240. HRMS data of the reduced alcohol was obtained.



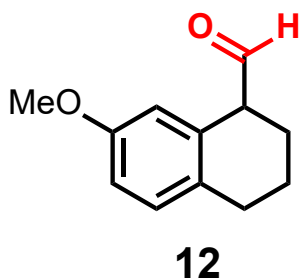
Yellow solid. (m.p.: 54-57 °C). **¹H NMR** (400 MHz, CDCl₃) δ 9.78 (d, *J* = 1.5 Hz, 1H), 7.40 – 7.29 (m, 3H), 7.26 – 7.14 (m, 5H), 7.10 – 7.06 (m, 2H), 3.86 (ddd, *J* = 8.1, 6.8, 1.5 Hz, 1H), 3.50 (dd, *J* = 14.0, 6.8 Hz, 1H), 3.00 (dd, *J* = 14.0, 7.9 Hz, 1H). **¹³C{¹H} NMR** (101 MHz, CDCl₃) δ 199.9, 138.8, 135.7, 129.1, 128.4, 127.7, 126.3, 61.0, 36.2. **HRMS** (ESI) calculated for C₁₂H₁₈O [MH₂+Na]⁺: 235.1093, Found: 235.1098. HRMS data of the reduced alcohol was obtained.



White solid. (m.p.: 53-58 °C). **¹H NMR** (400 MHz, CDCl₃) δ 9.77 (d, *J* = 1.4 Hz, 1H), 7.89 – 7.76 (m, 3H), 7.68 (d, *J* = 1.8 Hz, 1H), 7.54 – 7.45 (m, 2H), 7.32 (dd, *J* = 8.4, 1.8 Hz, 1H), 3.81 (dd, *J* = 7.0, 1.4 Hz, 1H), 1.55 (d, *J* = 8.8 Hz, 3H). **¹³C{¹H} NMR** (126 MHz, CDCl₃) δ 135.1, 133.6, 128.9, 127.7, 127.2, 126.5, 126.2, 126.1, 53.1, 14.7. Data are consistent with the literature.¹³

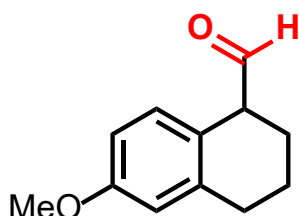


Colorless oil. **¹H NMR** (500 MHz, CDCl₃) δ 9.63 (d, *J* = 1.5 Hz, 1H), 6.86 (d, *J* = 8.2 Hz, 1H), 6.71 (s, 1H), 6.67 (dd, *J* = 8.2, 2.1 Hz, 1H), 4.25 (s, 4H), 3.51 (dd, *J* = 7.0, 1.5 Hz, 1H), 1.39 (d, *J* = 7.1 Hz, 3H). **¹³C{¹H} NMR** (126 MHz, CDCl₃) δ 201.0, 143.9, 143.0, 130.8, 121.3, 117.8, 117.0, 64.4, 52.2, 31.6, 22.7, 14.5. **HRMS** (ESI) calculated for C₁₁H₁₂O₃ [M+Na]⁺: 215.0684, Found: 205.0679.



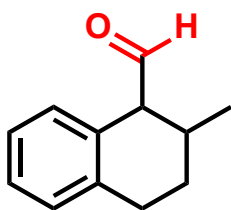
Light yellow oil. **¹H NMR** (400 MHz, CDCl₃) δ 9.69 (d, *J* = 2.3 Hz, 1H), 7.10 (d, *J* = 8.4 Hz, 1H), 6.81 (dd, *J* = 8.5, 2.7 Hz, 1H), 6.70 (d, *J* = 2.7 Hz, 1H), 3.81 (t, *J* = 0.7 Hz, 3H), 3.60 – 3.52 (m, 1H), 2.74 (t, *J* = 6.3 Hz, 2H), 2.22 (dq, *J* = 13.3, 5.6 Hz, 1H), 1.99

– 1.89 (m, 1H), 1.80 (dq, $J = 9.2, 5.9$ Hz, 2H). $^{13}\text{C}\{^1\text{H}\}$ NMR (101 MHz, CDCl_3) δ 202.0, 157.8, 131.8, 130.7, 130.1, 114.2, 113.5, 52.0, 28.3, 23.1, 20.7. HRMS (ESI) calculated for $\text{C}_{12}\text{H}_{14}\text{O}_2$ $[\text{M}+\text{Na}]^+$: 213.0886, Found: 213.0880.



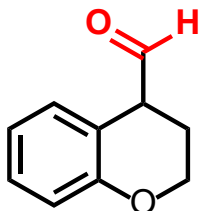
13

Light yellow oil. ^1H NMR (500 MHz, CDCl_3) δ 9.63 (d, $J = 2.3$ Hz, 1H), 7.05 (d, $J = 8.4$ Hz, 1H), 6.78 (dd, $J = 8.4, 2.7$ Hz, 1H), 6.69 (d, $J = 2.7$ Hz, 1H), 3.79 (s, 3H), 3.53 (td, $J = 5.7, 2.2$ Hz, 1H), 2.76 (t, $J = 6.4$ Hz, 2H), 2.23 – 2.15 (m, 1H), 1.92 (tdd, $J = 13.3, 6.4, 4.1$ Hz, 1H), 1.78 (pd, $J = 6.4, 3.1$ Hz, 2H). $^{13}\text{C}\{^1\text{H}\}$ NMR (126 MHz, CDCl_3) δ 202.3, 158.6, 139.4, 130.6, 122.9, 114.5, 112.5, 55.3, 51.0, 29.5, 23.2, 20.4. HRMS (ESI) calculated for $\text{C}_{12}\text{H}_{14}\text{O}_2$ $[\text{M}+\text{Na}]^+$: 213.0886, Found: 213.0889.



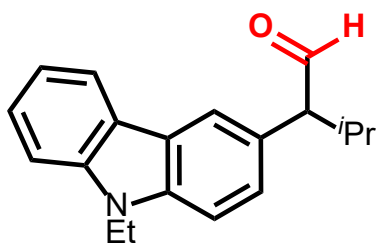
14

Light yellow oil. dr=1:1 in crude mixture. ^1H NMR (400 MHz, CDCl_3) δ 9.49 (d, $J = 3.9$ Hz, 1H), 7.24 – 7.17 (m, 3H), 7.09 – 7.03 (m, 1H), 3.28 (dd, $J = 8.2, 3.9$ Hz, 1H), 2.90 – 2.82 (m, 2H), 2.30 – 2.20 (m, 1H), 1.98 (dtd, $J = 13.5, 5.2, 3.3$ Hz, 1H), 1.59 – 1.51 (m, 1H), 1.12 (d, $J = 6.7$ Hz, 3H). $^{13}\text{C}\{^1\text{H}\}$ NMR (101 MHz, CDCl_3) δ 201.6, 130.2, 129.7, 129.5, 127.5, 127.2, 126.3, 59.8, 31.4, 28.9, 28.5, 28.2, 19.9. HRMS (ESI) calculated for $\text{C}_{12}\text{H}_{14}\text{O}$ $[\text{M}+\text{Na}]^+$: 197.0937, Found: 197.0941.



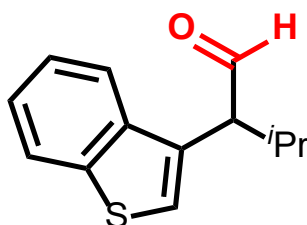
15

Light yellow oil. ^1H NMR (400 MHz, CDCl_3) 9.74 (t, $J = 1.2$ Hz, 1H), 7.24 – 7.17 (m, 2H), 6.97 (td, $J = 7.5, 1.3$ Hz, 1H), 6.89 (dd, $J = 8.1, 1.2$ Hz, 1H), 4.23 (dddd, $J = 11.2, 4.9, 3.6, 1.1$ Hz, 1H), 4.00 (ddd, $J = 11.2, 10.1, 2.6$ Hz, 1H), 3.61 (t, $J = 5.3$ Hz, 1H), 2.44 (dtd, $J = 14.0, 4.7, 2.7$ Hz, 1H), 2.20 – 1.97 (m, 1H). $^{13}\text{C}\{^1\text{H}\}$ NMR (101 MHz, CDCl_3) δ 200.4, 155.3, 129.9, 129.0, 121.0, 117.7, 116.4, 63.8, 47.1, 21.7. HRMS (ESI) calculated for $\text{C}_{10}\text{H}_{10}\text{O}_2$ $[\text{M}+\text{Na}]^+$: 185.0573, Found: 185.0569



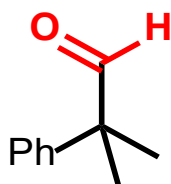
16

Yellow oil. **¹H NMR** (500 MHz, CDCl₃) δ 9.82 (d, *J* = 3.2 Hz, 1H), 8.12 (dt, *J* = 7.8, 1.0 Hz, 1H), 7.94 (d, *J* = 1.7 Hz, 1H), 7.54 – 7.40 (m, 3H), 7.34 – 7.23 (m, 2H), 4.39 (q, *J* = 7.2 Hz, 2H), 3.39 (dd, *J* = 9.7, 3.3 Hz, 1H), 2.56 (dhept, *J* = 9.5, 6.6 Hz, 1H), 1.47 (t, *J* = 7.2 Hz, 3H), 1.14 (d, *J* = 6.5 Hz, 3H), 0.84 (d, *J* = 6.7 Hz, 3H). **¹³C{¹H} NMR** (126 MHz, CDCl₃) δ 201.3, 140.3, 139.4, 126.9, 126.0, 125.5, 123.5, 122.6, 121.2, 120.5, 119.0, 108.9, 108.6, 67.0, 37.6, 28.9, 21.4, 20.2, 13.9. **HRMS** (ESI) calculated for C₁₉H₂₁NO [M+Na]⁺: 197.0337, Found: 197.0941.



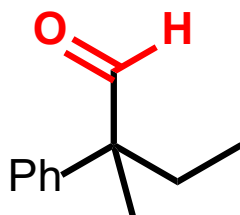
17

Yellow oil. **¹H NMR** (500 MHz, CDCl₃) δ 9.69 (d, *J* = 3.3 Hz, 1H), 7.93 – 7.89 (m, 1H), 7.84 – 7.80 (m, 1H), 7.42 (pd, *J* = 7.1, 1.4 Hz, 2H), 7.31 (s, 1H), 3.74 (dd, *J* = 9.3, 3.3 Hz, 1H), 2.60 (dhept, *J* = 9.2, 6.7 Hz, 1H), 1.13 (d, *J* = 6.6 Hz, 3H), 0.92 (d, *J* = 6.7 Hz, 3H). **¹³C{¹H} NMR** (126 MHz, CDCl₃) δ 199.3, 140.4, 138.8, 130.0, 124.7, 124.4, 124.3, 123.0, 121.8, 59.8, 28.9, 21.3, 20.3. **HRMS** (ESI) calculated for C₁₃H₁₄OS [M+Na]⁺: 214.0658, Found: 214.0660.



18

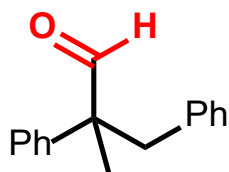
Light yellow oil. **¹H NMR** (500 MHz, CDCl₃) δ 9.53 (s, 1H), 7.36 (dt, *J* = 48.8, 7.8 Hz, 5H), 1.49 (s, 6H). **¹³C{¹H} NMR** (126 MHz, CDCl₃) δ 202.3, 141.2, 128.9, 127.3, 126.7, 50.5, 22.5. **HRMS** (ESI) calculated for C₁₀H₁₂O [M+H]⁺: 149.0961, Found: 149.0957.



19

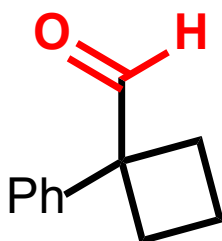
Light yellow oil. **¹H NMR** (500 MHz, CDCl₃) δ 9.54 (s, 1H), 7.40 (t, *J* = 7.6 Hz, 2H), 7.33 – 7.24 (m, 3H), 1.97 (ddt, *J* = 38.1, 14.2, 7.1 Hz, 2H), 1.46 (s, 3H), 0.82 (t, *J* =

7.7 Hz, 3H). $^{13}\text{C}\{^1\text{H}\}$ NMR (126 MHz, CDCl_3) δ 202.8, 128.8, 127.2, 127.2, 54.3, 28.6, 18.3, 8.4. HRMS (ESI) calculated for $\text{C}_{11}\text{H}_{14}\text{O}$ $[\text{M}+\text{H}]^+$: 167.1117, Found: 167.1110.



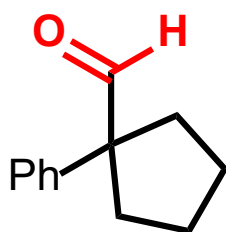
20

Yellow oil. ^1H NMR (500 MHz, CDCl_3) δ 9.63 (s, 1H), 7.37 (t, $J = 7.4$ Hz, 2H), 7.33 – 7.28 (m, 1H), 7.21 – 7.11 (m, 5H), 6.82 – 6.77 (m, 2H), 3.20 (q, $J = 13.7$ Hz, 2H), 1.38 (s, 3H). $^{13}\text{C}\{^1\text{H}\}$ NMR (126 MHz, CDCl_3) δ 202.0, 139.3, 136.8, 130.4, 128.8, 127.9, 127.6, 127.5, 126.4, 55.1, 42.7, 18.2. HRMS (ESI) calculated for $\text{C}_{16}\text{H}_{16}\text{O}$ $[\text{M}+\text{Na}]^+$: 247.1093, Found: 247.1105.



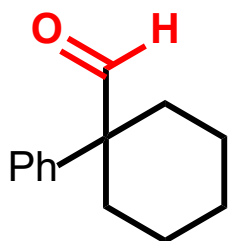
21

Colorless oil. ^1H NMR (500 MHz, CDCl_3) δ 9.56 (s, 1H), 7.43 – 7.38 (m, 2H), 7.32 – 7.28 (m, 1H), 7.20 – 7.16 (m, 2H), 2.80 – 2.73 (m, 2H), 2.44 (qd, $J = 9.3, 2.5$ Hz, 2H), 2.10 – 1.90 (m, 2H). $^{13}\text{C}\{^1\text{H}\}$ NMR (126 MHz, CDCl_3) δ 199.5, 141.0, 128.8, 127.0, 126.4, 57.6, 28.4, 15.9. HRMS (ESI) calculated for $\text{C}_{11}\text{H}_{12}\text{O}$ $[\text{M}+\text{Na}]^+$: 183.0780, Found: 183.0778.



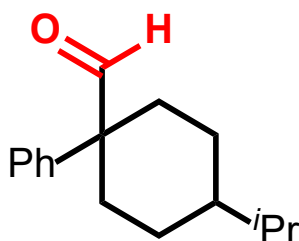
22

Colorless oil. ^1H NMR (500 MHz, CDCl_3) δ 9.43 (s, 1H), 7.38 (dd, $J = 8.7, 6.5$ Hz, 2H), 7.33 – 7.26 (m, 3H), 2.56 (ddt, $J = 15.0, 5.8, 2.2$ Hz, 2H), 1.96 – 1.87 (m, 2H), 1.78 (ttq, $J = 12.7, 8.6, 4.4$ Hz, 2H), 1.69 (pd, $J = 7.5, 3.3$ Hz, 2H). $^{13}\text{C}\{^1\text{H}\}$ NMR (126 MHz, CDCl_3) δ 200.8, 140.3, 128.8, 127.7, 127.2, 32.4, 24.2. HRMS (ESI) calculated for $\text{C}_{12}\text{H}_{14}\text{O}$ $[\text{M}+\text{H}]^+$: 175.1117, Found: 175.1123.



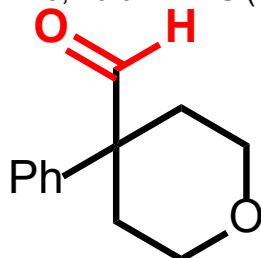
23

Colorless oil. **¹H NMR** (500 MHz, CDCl₃) δ 9.40 (s, 1H), 7.40 (dd, *J* = 8.4, 6.9 Hz, 2H), 7.37 – 7.33 (m, 2H), 7.31 – 7.27 (m, 1H), 2.33 (dt, *J* = 13.7, 4.5 Hz, 2H), 1.88 (ddd, *J* = 14.0, 10.8, 3.7 Hz, 2H), 1.76 – 1.58 (m, 3H), 1.51 (dtt, *J* = 13.6, 10.4, 3.4 Hz, 2H), 1.41 – 1.25 (m, 1H). **¹³C{¹H} NMR** (126 MHz, CDCl₃) δ 202.5, 139.8, 128.88, 127.21, 127.15, 31.29, 25.63, 22.84. **HRMS** (ESI) calculated for C₁₃H₁₆O [M+Na]⁺: 211.1093, Found: 211.1096.



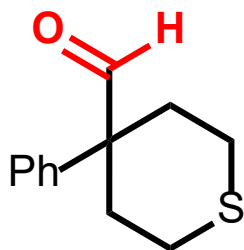
24

Colorless oil. **¹H NMR** (500 MHz, CDCl₃) δ 9.38 (s, 1H), 7.45 – 7.38 (m, 4H), 7.32 – 7.28 (m, 1H), 2.47 (ddd, *J* = 13.8, 4.2, 2.1 Hz, 2H), 1.82 – 1.68 (m, 4H), 1.35 (hept, *J* = 6.6 Hz, 1H), 1.17 (ddp, *J* = 11.8, 7.7, 4.4, 3.8 Hz, 3H), 0.82 (d, *J* = 6.7 Hz, 6H). **¹³C{¹H} NMR** (126 MHz, CDCl₃) δ 202.2, 137.3, 129.0, 128.2, 127.0, 54.6, 43.8, 32.1, 29.5, 24.9, 20.0. **HRMS** (ESI) calculated for C₁₆H₂₂ [M+Na]⁺: 256.1563, Found: 256.1569.



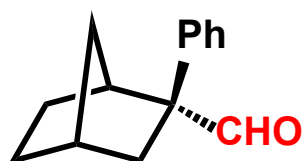
25

Colorless oil. **¹H NMR** (500 MHz, CDCl₃) δ 9.41 (s, 1H), 7.41 (ddd, *J* = 7.9, 6.4, 1.2 Hz, 2H), 7.33 – 7.27 (m, 3H), 3.91 (dt, *J* = 12.0, 4.0 Hz, 2H), 3.60 (ddd, *J* = 11.8, 10.9, 2.5 Hz, 2H), 2.40 (ddt, *J* = 13.7, 4.0, 2.1 Hz, 2H), 2.09 (ddd, *J* = 14.5, 10.8, 4.4 Hz, 2H). **¹³C{¹H} NMR** (126 MHz, CDCl₃) δ 200.9, 129.2, 129.0, 127.8, 126.8, 64.9, 31.1. **HRMS** (ESI) calculated for C₁₂H₁₄O₂ [M+Na]⁺: 213.0886, Found: 213.0891.



26

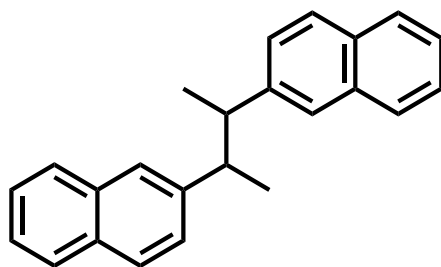
Yellow oil. $^1\text{H NMR}$ (400 MHz, CDCl_3) δ 9.35 (s, 1H), 7.43 – 7.37 (m, 2H), 7.33 – 7.26 (m, 3H), 2.84 – 2.75 (m, 2H), 2.71 – 2.63 (m, 2H), 2.58 (ddd, J = 14.0, 6.5, 2.7 Hz, 2H), 2.29 (ddd, J = 13.9, 10.5, 3.2 Hz, 2H). $^{13}\text{C}\{^1\text{H}\}$ NMR (101 MHz, CDCl_3) δ 201.4, 138.6, 129.2, 127.7, 127.0, 53.6, 32.5, 24.8. HRMS (ESI) calculated for $\text{C}_{12}\text{H}_{14}\text{OS}$ $[\text{M}+\text{H}]^+$: 207.0838, Found: 207.0842.



27

Colorless oil. dr = 11:1 $^1\text{H NMR}$ (500 MHz, CDCl_3) δ 9.31 (s, 1H), 7.39 (t, J = 7.6 Hz, 2H), 7.30 – 7.26 (m, 3H), 2.98 – 2.94 (m, 1H), 2.61 (ddd, J = 12.7, 4.9, 2.4 Hz, 1H), 2.42 (d, J = 4.5 Hz, 1H), 1.65 – 1.60 (m, 2H), 1.53 (dt, J = 9.5, 4.4 Hz, 2H), 1.43 – 1.32 (m, 4H), 1.15 – 1.09 (m, 1H). $^{13}\text{C}\{^1\text{H}\}$ NMR (126 MHz, CDCl_3) δ 199.8, 137.9, 129.3, 128.8, 127.1, 64.6, 41.2, 37.2, 36.7, 35.1, 29.8, 23.4. HRMS (ESI) calculated for $\text{C}_{14}\text{H}_{16}\text{O}$ $[\text{M}+\text{H}]^+$: 201.1274, Found: 201.1278.

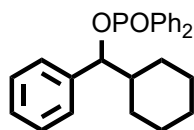
9.3. Mechanistic experiments



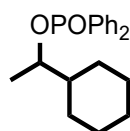
28

White solid. (m.p.: 85-88 °C). *dl* : *meso* = 1:1. $^1\text{H NMR}$ (400 MHz, CDCl_3) δ 7.93 – 7.84 (m, 3H), 7.80 – 7.66 (m, 4H), 7.59 – 7.38 (m, 6H), 7.29 (d, J = 8.5 Hz, 1H), 3.32 (h, J = 7.4 Hz, 1H) (for *dl*), 3.16 (dt, J = 7.8, 3.9 Hz, 1H) (for *meso*), 1.44 (d, J = 5.8 Hz, 3H), 1.19 (d, J = 5.2 Hz, 3H). $^{13}\text{C}\{^1\text{H}\}$ NMR (101 MHz, CDCl_3) δ 144.0, 143.4, 133.7, 133.4, 132.4, 132.1, 128.1, 127.7, 127.7, 127.5, 127.4, 126.8, 126.3, 126.2, 126.1, 126.0, 125.7, 125.3, 125.1, 47.4, 46.3, 21.3, 18.1. Data are consistent with the literatures.^{14, 15}

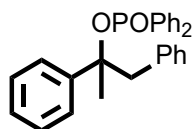
9.4. Substrates for cyclic voltammetry



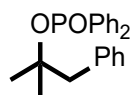
29(3-sub) Light yellow oil. $^1\text{H NMR}$ (500 MHz, CDCl_3) δ 7.87 – 7.79 (m, 2H), 7.54 – 7.40 (m, 5H), 7.32 (td, J = 7.4, 1.5 Hz, 1H), 7.21 – 7.09 (m, 7H), 5.04 (dd, J = 9.9, 7.5 Hz, 1H), 2.06 (dt, J = 13.1, 3.2 Hz, 1H), 1.89 (tdt, J = 11.2, 7.0, 3.4 Hz, 1H), 1.80 – 1.71 (m, 1H), 1.62 (td, J = 14.1, 12.9, 7.2 Hz, 2H), 1.45 – 1.32 (m, 1H), 1.29 – 0.99 (m, 5H), 0.88 (qd, J = 12.3, 3.7 Hz, 1H). $^{13}\text{C}\{^1\text{H}\}$ NMR (126 MHz, CDCl_3) δ 139.6, 133.0, 132.0, 132.0, 131.9, 131.9, 131.6, 131.6, 131.5, 131.4, 130.9, 128.5, 128.3, 127.9, 127.9, 127.8, 127.7, 127.3, 82.6, 82.6, 44.4, 44.4, 29.4, 28.5, 26.3, 25.9. $^{31}\text{P}\{^1\text{H}\}$ NMR (162 MHz, CDCl_3) δ 30.7. **HRMS** (ESI) calculated for $\text{C}_{25}\text{H}_{27}\text{O}_2\text{P}$ $[\text{M}+\text{Na}]^+$: 413.1641, Found: 413.1644.



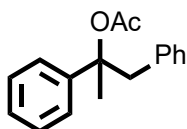
30 Colorless oil. $^1\text{H NMR}$ (500 MHz, CDCl_3) δ 7.80 (dddd, J = 13.7, 12.2, 8.3, 1.4 Hz, 4H), 7.53 – 7.46 (m, 2H), 7.42 (dddt, J = 8.4, 6.6, 3.2, 1.5 Hz, 4H), 4.43 – 4.32 (m, 1H), 1.82 – 1.68 (m, 4H), 1.64 (dt, J = 12.3, 3.2, 1.6 Hz, 1H), 1.54 (tdt, J = 11.7, 5.2, 3.2 Hz, 1H), 1.23 (d, J = 6.4 Hz, 3H), 1.21 – 0.87 (m, 5H). $^{13}\text{C}\{^1\text{H}\}$ NMR (126 MHz, CDCl_3) δ 133.4, 133.0, 132.3, 131.9, 131.9, 131.8, 131.8, 131.7, 131.7, 131.6, 128.4, 128.3, 77.4, 77.4, 44.1, 44.1, 28.5, 28.0, 26.5, 26.1, 26.1, 18.8, 18.8. $^{31}\text{P}\{^1\text{H}\}$ NMR (202 MHz, CDCl_3) δ 29.3.



31 White liquid. $^1\text{H NMR}$ (500 MHz, CDCl_3) δ 7.80 – 7.72 (m, 2H), 7.72 – 7.65 (m, 2H), 7.45 (q, J = 8.2 Hz, 2H), 7.36 (dd, J = 7.6, 3.5 Hz, 4H), 7.33 – 7.23 (m, 5H), 7.15 (t, J = 7.4 Hz, 1H), 7.08 (t, J = 7.5 Hz, 2H), 6.86 (d, J = 7.4 Hz, 2H), 3.52 – 3.40 (m, 2H), 1.89 (s, 3H). $^{13}\text{C}\{^1\text{H}\}$ NMR (126 MHz, CDCl_3) δ 144.4, 144.4, 136.2, 135.0, 134.8, 133.9, 133.7, 131.6, 131.6, 131.6, 131.5, 131.3, 131.2, 131.0, 128.4, 128.4, 128.3, 128.3, 128.0, 127.6, 127.4, 126.5, 125.6, 87.6, 87.5, 51.5, 51.5, 26.8, 26.8. $^{31}\text{P}\{^1\text{H}\}$ NMR (202 MHz, CDCl_3) δ 27.1. **HRMS** (ESI) calculated for $\text{C}_{27}\text{H}_{25}\text{O}_2\text{P}$ $[\text{M}+\text{Na}]^+$: 435.1484, Found: 435.1486.

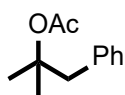


32 Light yellow oil. $^1\text{H NMR}$ (400 MHz, CDCl_3) δ 7.73 – 7.66 (m, 4H), 7.46 (td, J = 7.3, 1.5 Hz, 2H), 7.38 (ddd, J = 8.5, 6.8, 3.5 Hz, 4H), 7.33 – 7.25 (m, 5H), 3.08 (d, J = 1.0 Hz, 2H), 1.51 (s, 3H). $^{13}\text{C}\{^1\text{H}\}$ NMR (101 MHz, CDCl_3) δ 137.1, 135.1, 134.0, 131.5, 131.5, 131.3, 131.3, 131.0, 85.3, 85.2, 50.1, 50.1, 28.6, 28.5. $^{31}\text{P}\{^1\text{H}\}$ NMR (202 MHz, CDCl_3) δ 26.5. **HRMS** (ESI) calculated for $\text{C}_{25}\text{H}_{23}\text{O}_2\text{P}$ $[\text{M}+\text{Na}]^+$: 373.1328, Found: 373.1327.



33 (20-sub)

Light yellow oil. **¹H NMR** (500 MHz, CDCl₃) δ 7.35 – 7.31 (m, 2H), 7.30 – 7.19 (m, 6H), 6.98 – 6.90 (m, 2H), 3.31 (d, *J* = 13.5 Hz, 1H), 3.19 (d, *J* = 13.5 Hz, 1H), 2.08 (s, 3H), 1.86 (s, 3H). **¹³C{¹H} NMR** (126 MHz, CDCl₃) δ 169.7, 144.4, 136.3, 130.8, 128.1, 127.7, 127.0, 126.5, 124.9, 83.7, 49.2, 24.1, 22.3. **HRMS** (ESI) calculated for C₁₇H₁₈O₂ [M+Na]⁺: 277.1199, Found: 277.1201.



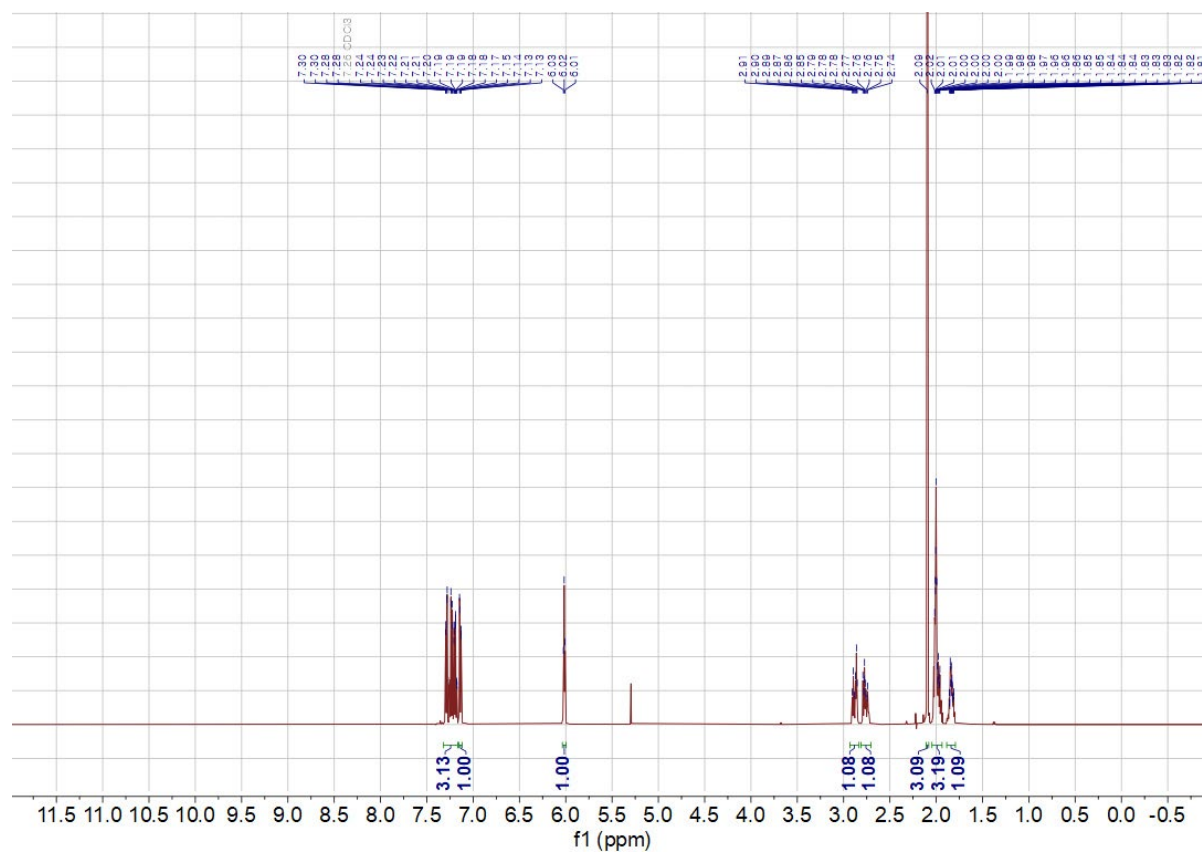
34

Colorless oil. **¹H NMR** (400 MHz, CDCl₃) δ 7.32– 7.22 (m, 3H), 7.20 – 7.16 (m, 2H), 3.06 (s, 2H), 1.98 (s, 3H), 1.45 (s, 6H). **¹³C{¹H} NMR** (101 MHz, CDCl₃) δ 170.7, 137.3, 130.6, 128.0, 126.4, 82.1, 46.4, 26.0, 22.6. Data are consistent with the literature.¹⁶

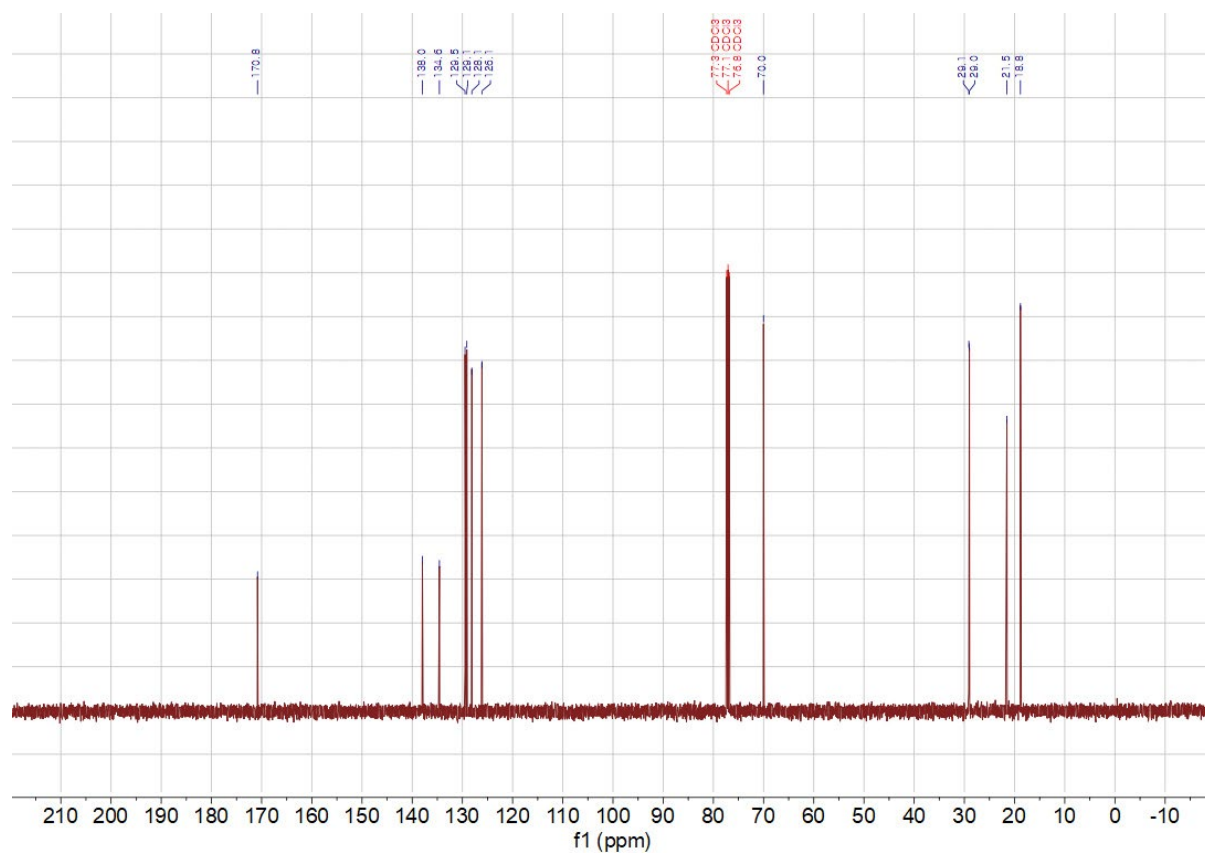
10. NMR Spectra

10.1. Starting materials

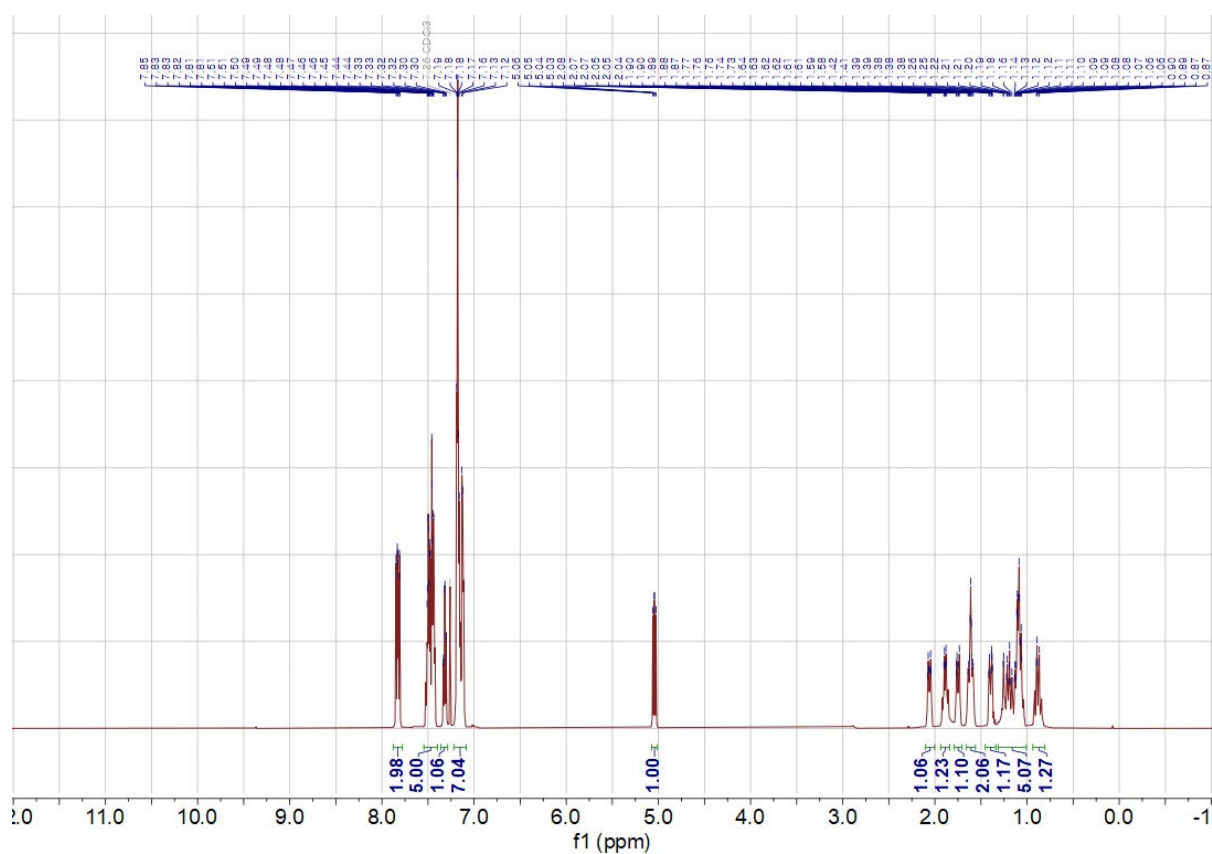
^1H NMR spectrum (500 MHz, CDCl_3) of **1(2-sub)**



$^{13}\text{C}\{^1\text{H}\}$ NMR spectrum (126 MHz, CDCl_3) of **1(2-sub)**

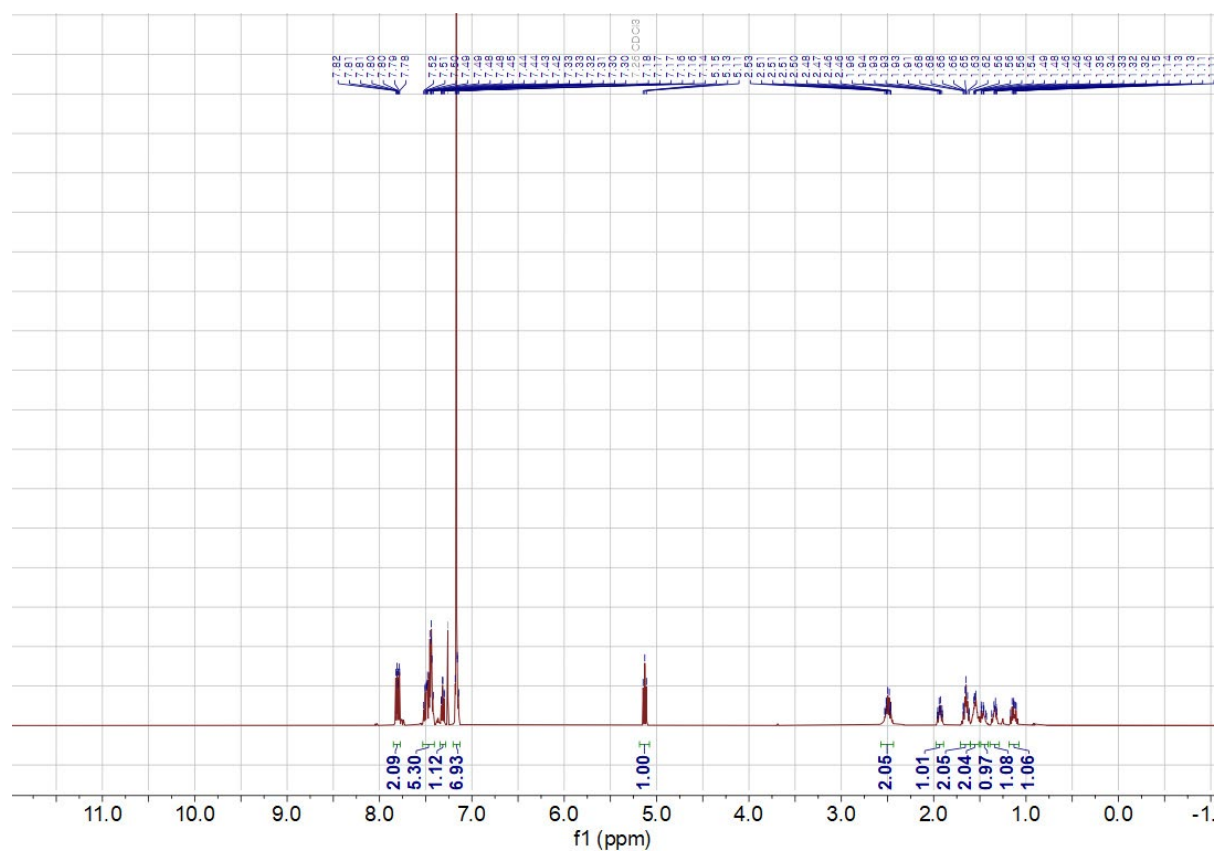


^1H NMR spectrum (500 MHz, CDCl_3) of **3-sub**

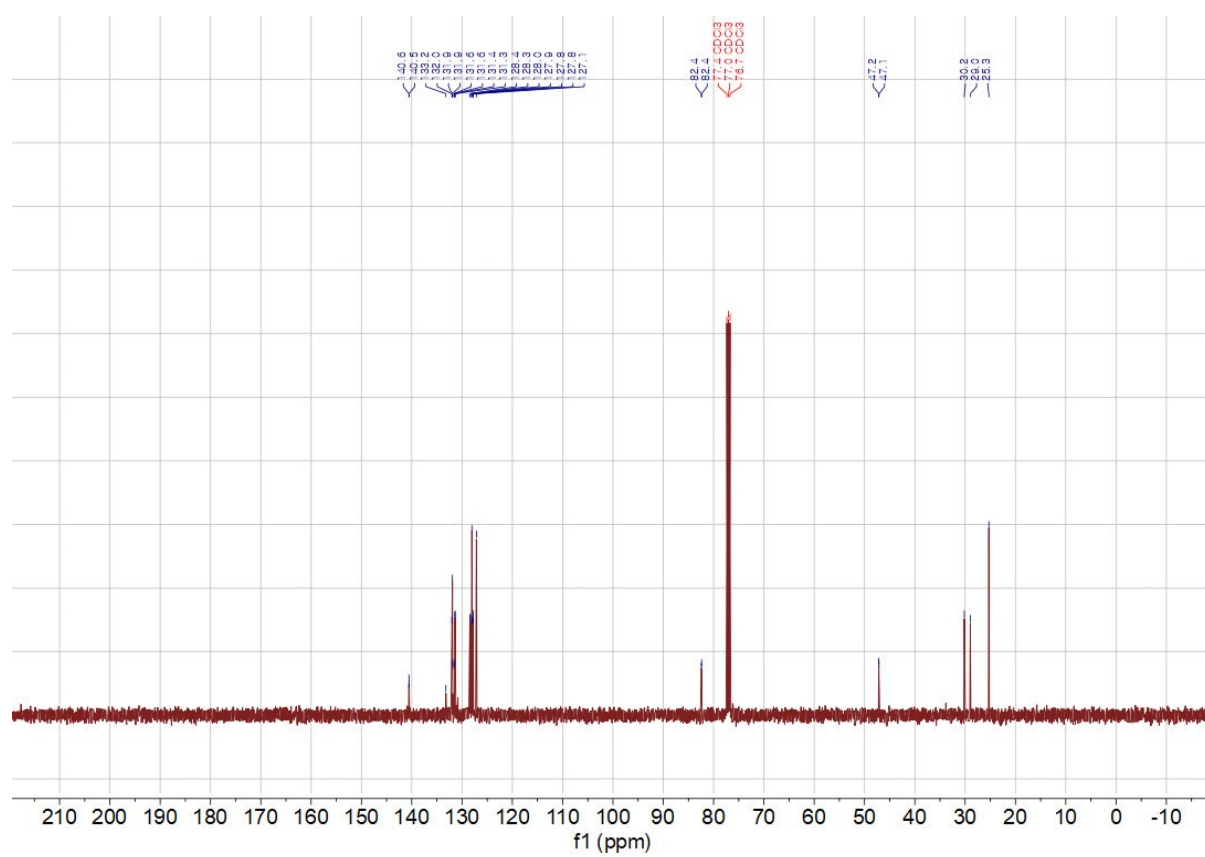


The image shows a 1H NMR spectrum with a single sharp peak at approximately 3.7 ppm. The x-axis is labeled 'f1 (ppm)' and ranges from 140 to -240. The peak is labeled '2.00'.

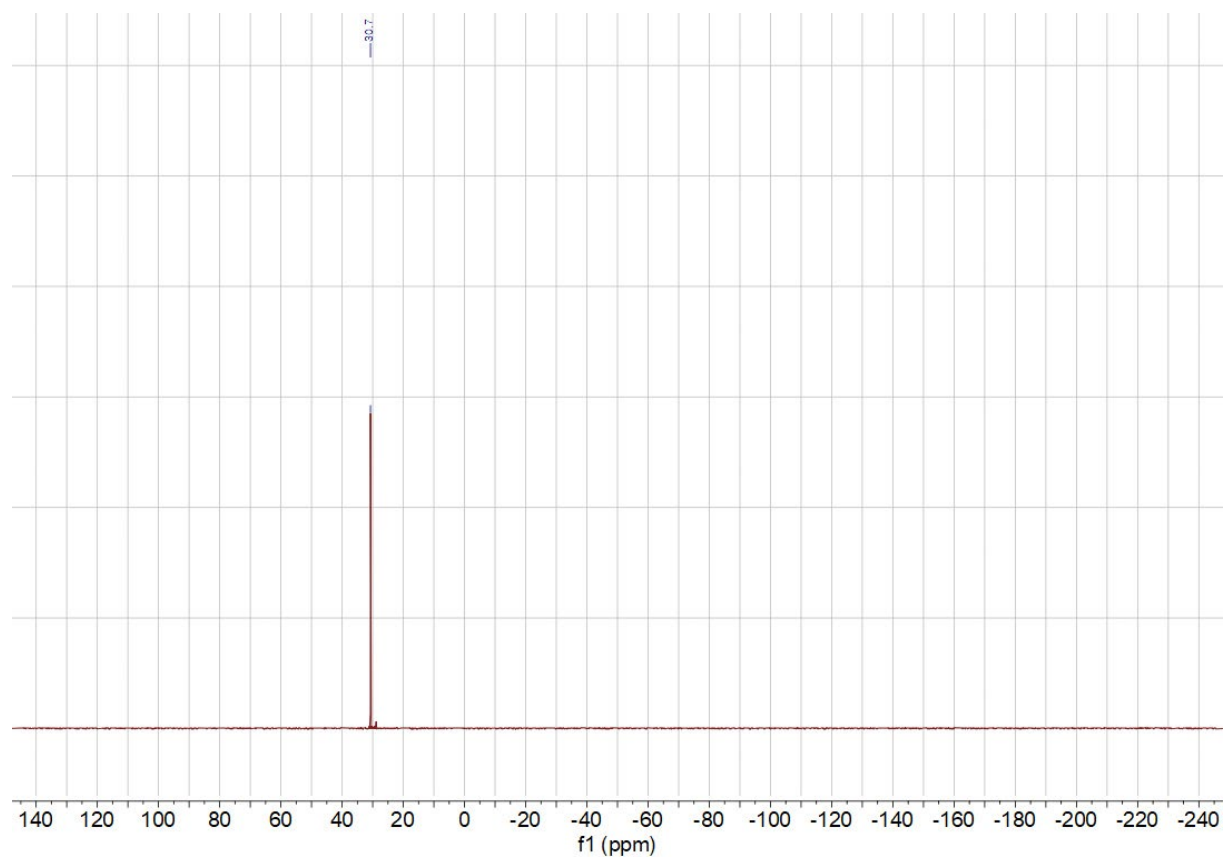
^1H NMR spectrum (500 MHz, CDCl_3) of **4-sub**



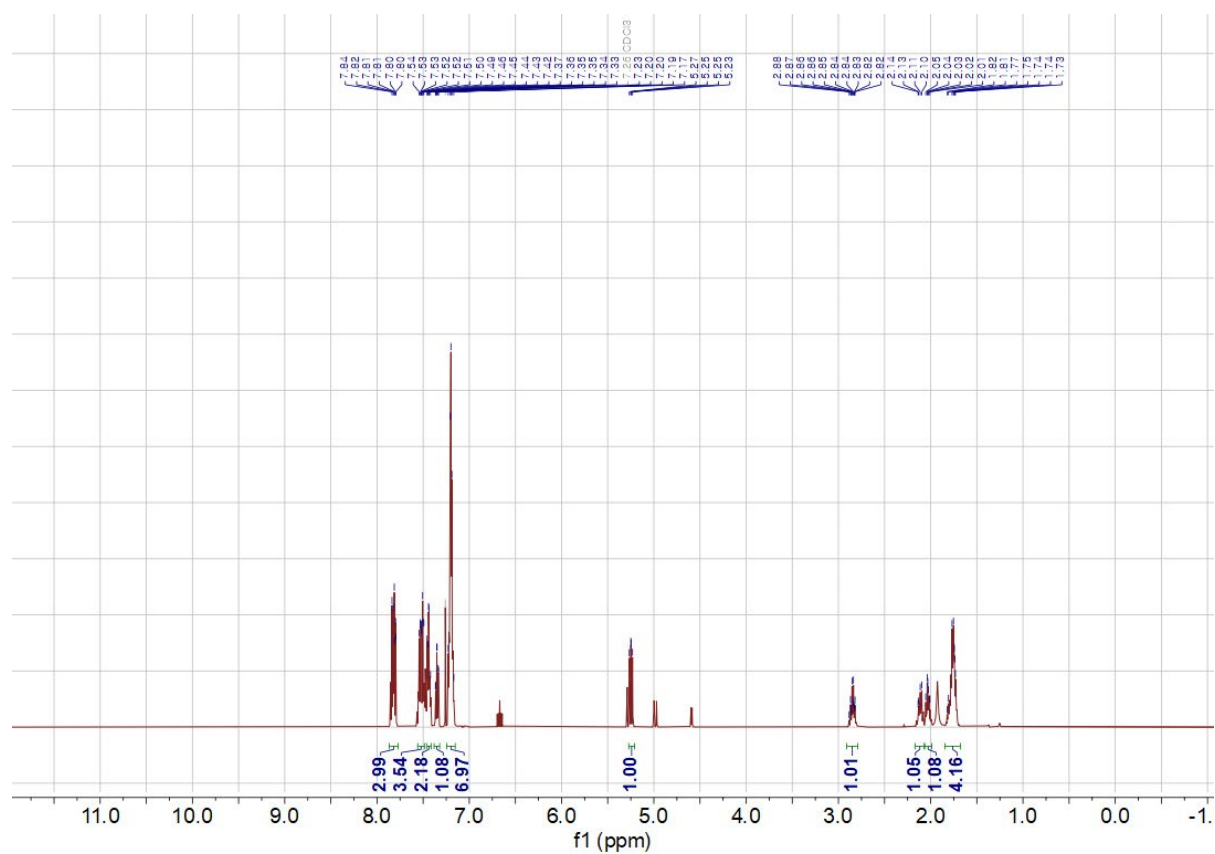
$^{13}\text{C}\{^1\text{H}\}$ NMR spectrum (126 MHz, CDCl_3) of **4-sub**



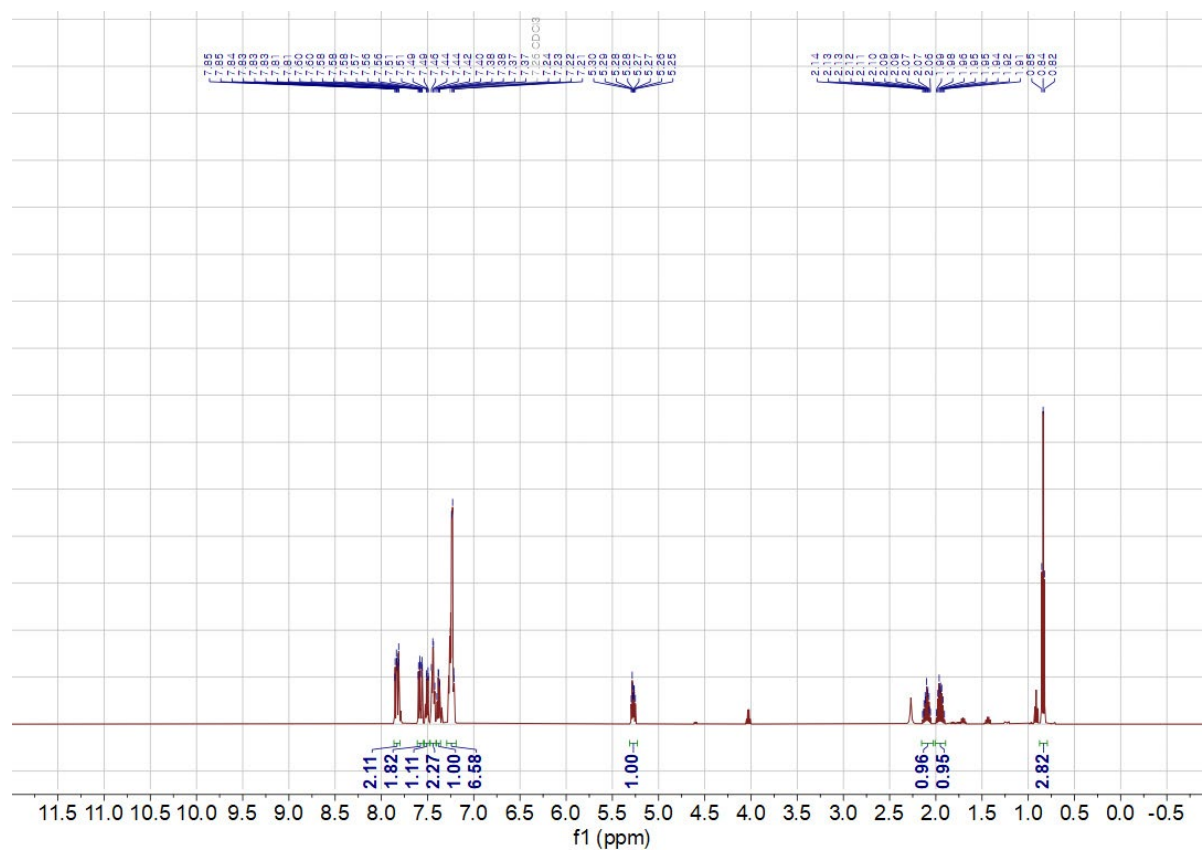
$^{31}\text{P}\{^1\text{H}\}$ NMR spectrum (162 MHz, CDCl_3) of **4-sub**



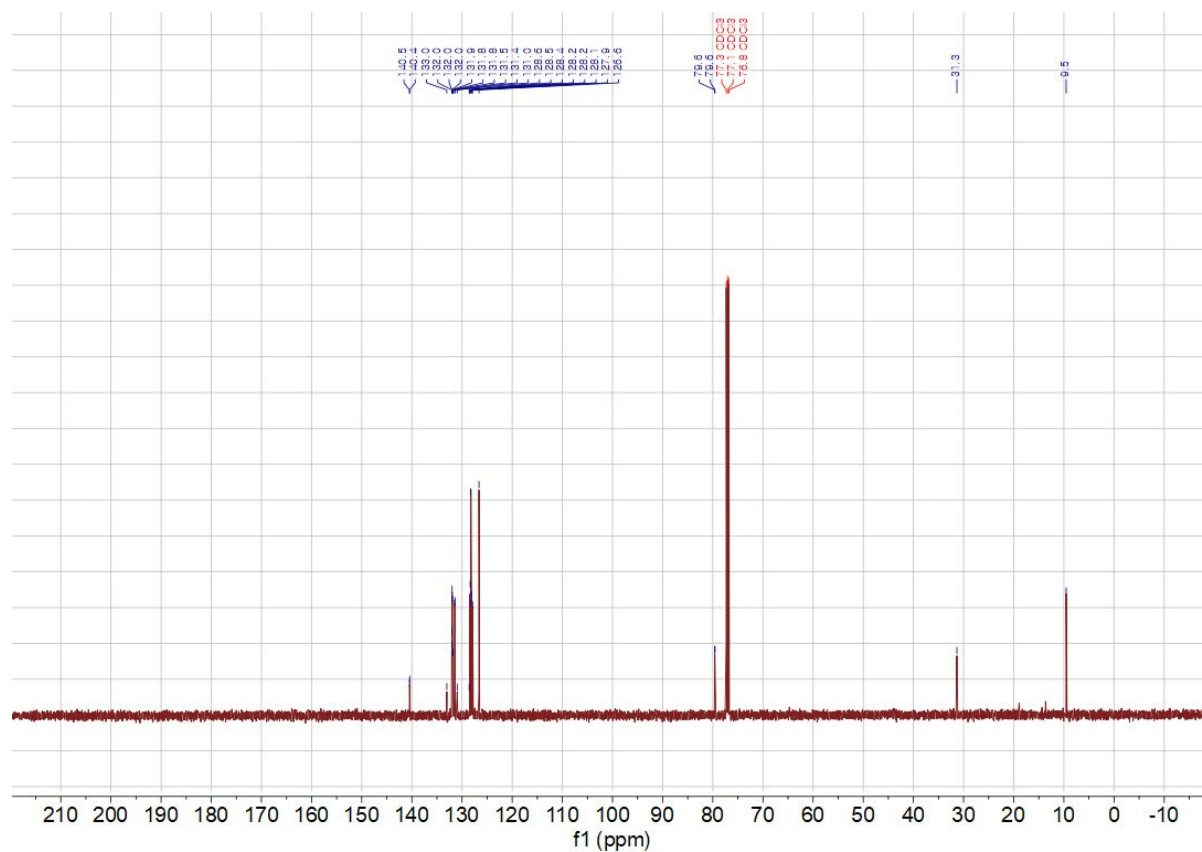
^1H NMR spectrum (500 MHz, CDCl_3) of **5-sub**



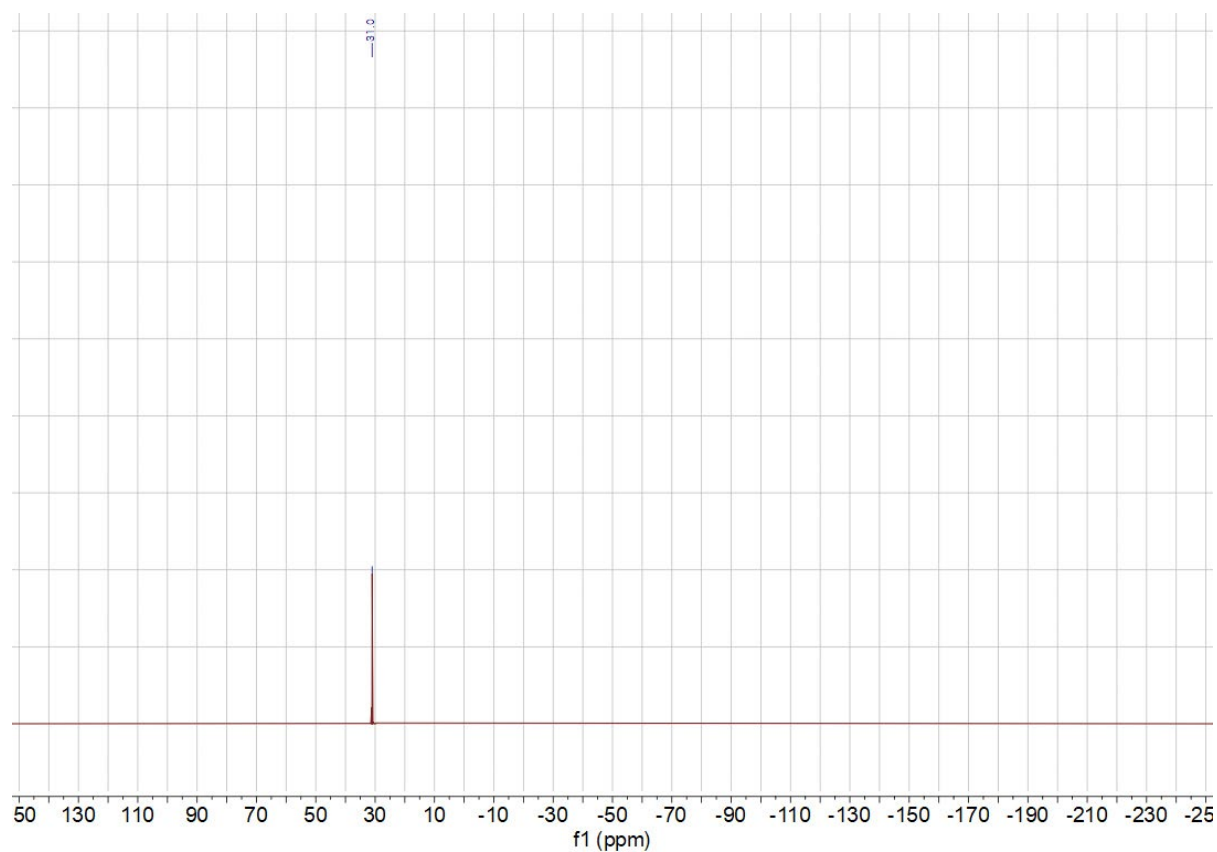
^1H NMR spectrum (500 MHz, CDCl_3) of **6-sub**



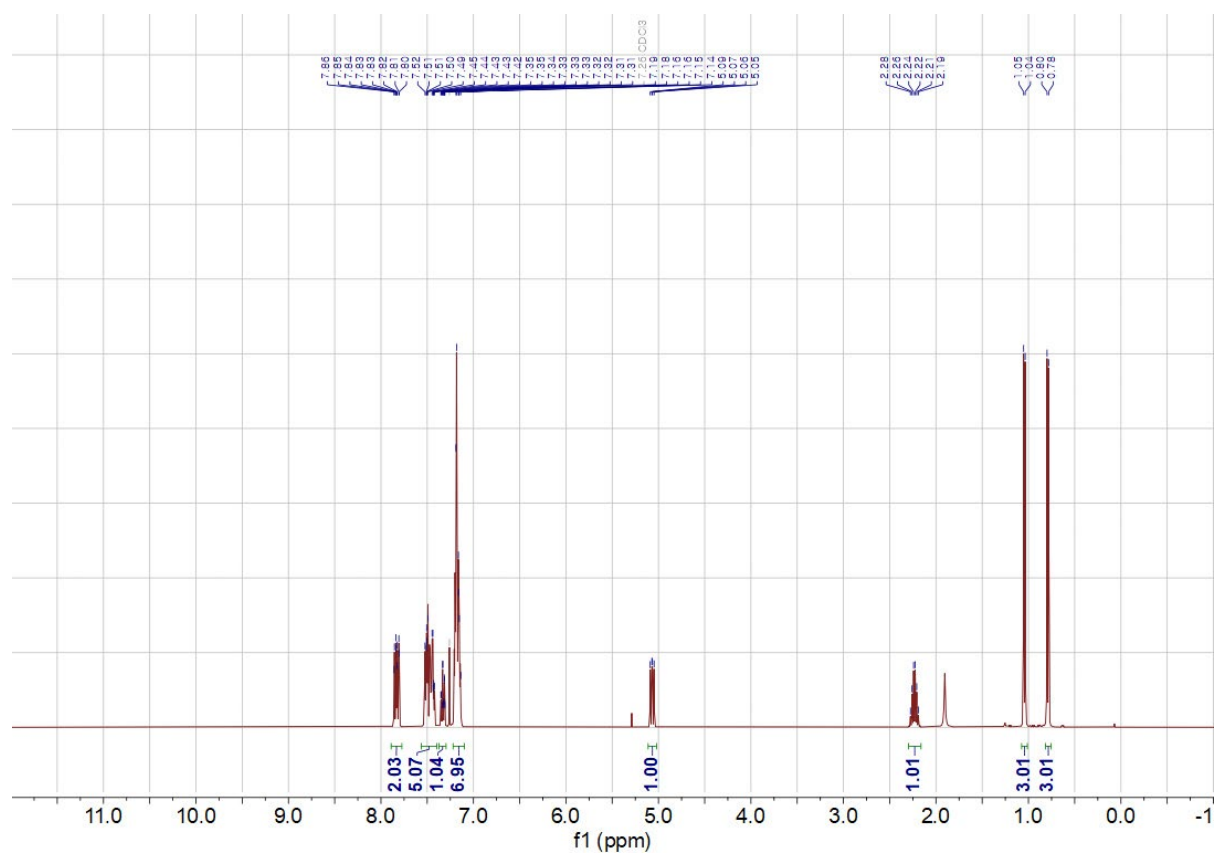
$^{13}\text{C}\{^1\text{H}\}$ NMR spectrum (126 MHz, CDCl_3) of **6-sub**



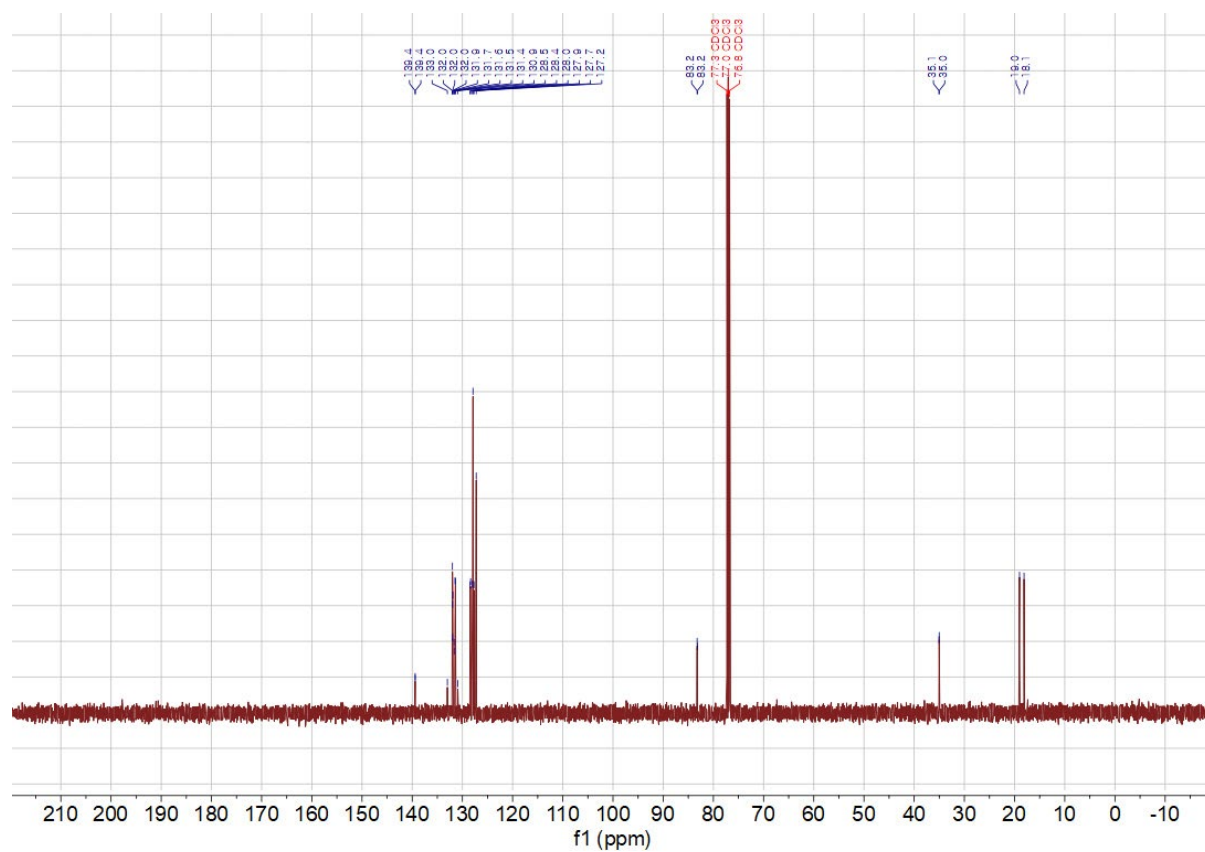
$^{31}\text{P}\{^1\text{H}\}$ NMR spectrum (202 MHz, CDCl_3) of **6-sub**



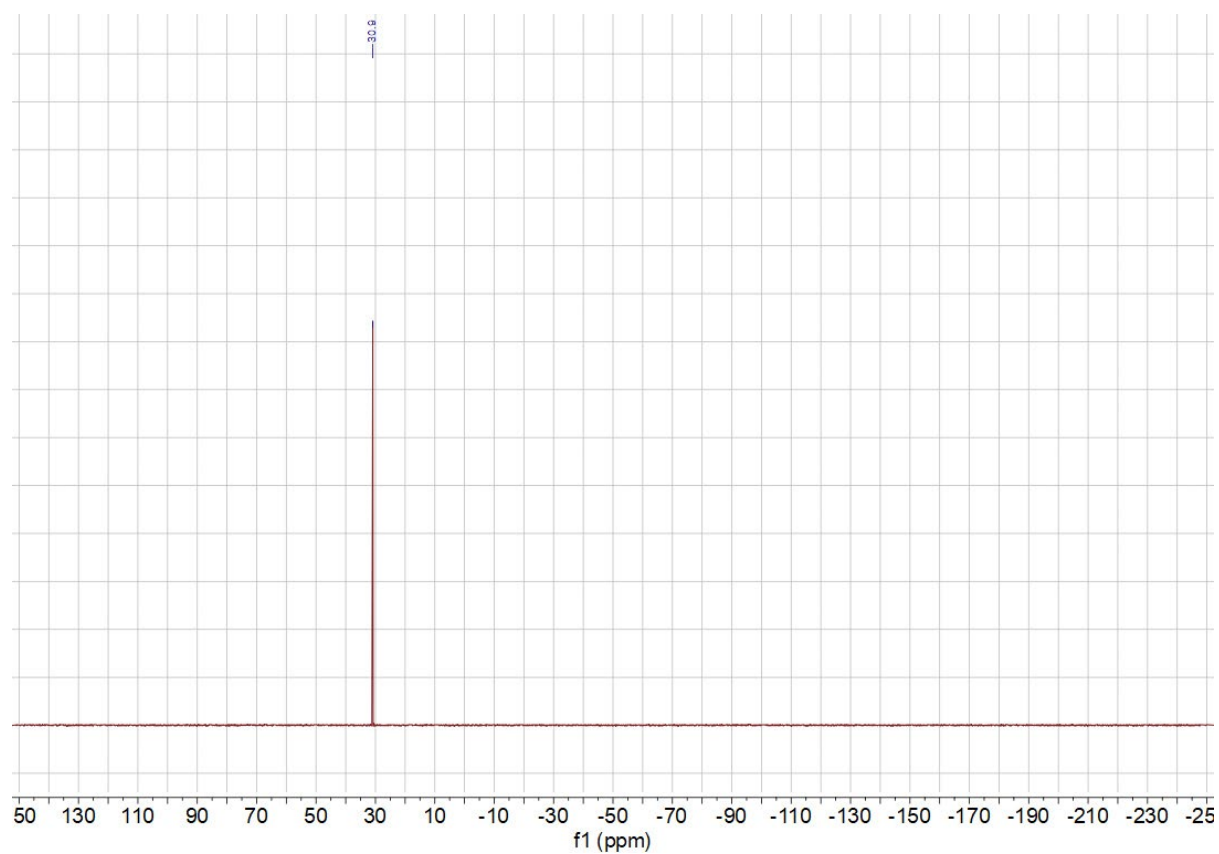
^1H NMR spectrum (400 MHz, CDCl_3) of **7-sub**



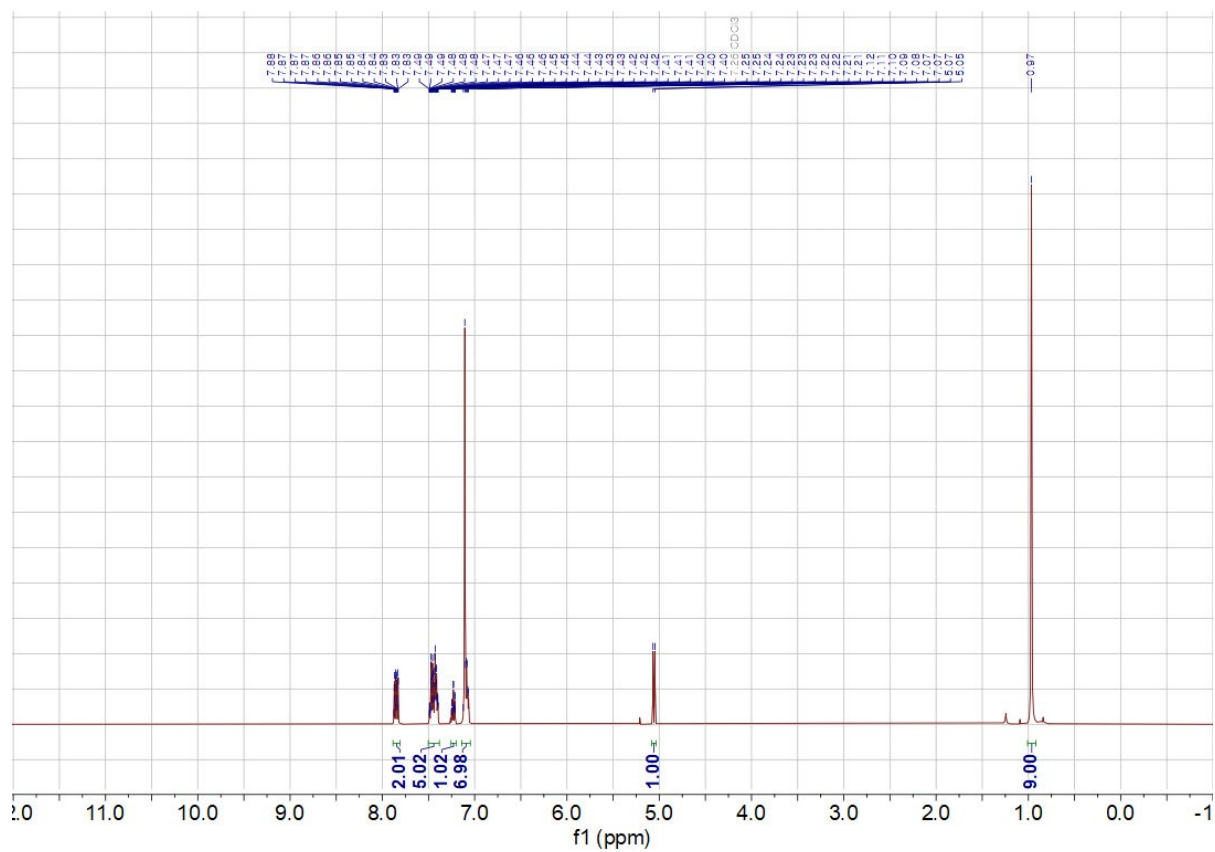
$^{13}\text{C}\{^1\text{H}\}$ NMR spectrum (126 MHz, CDCl_3) of **7-sub**



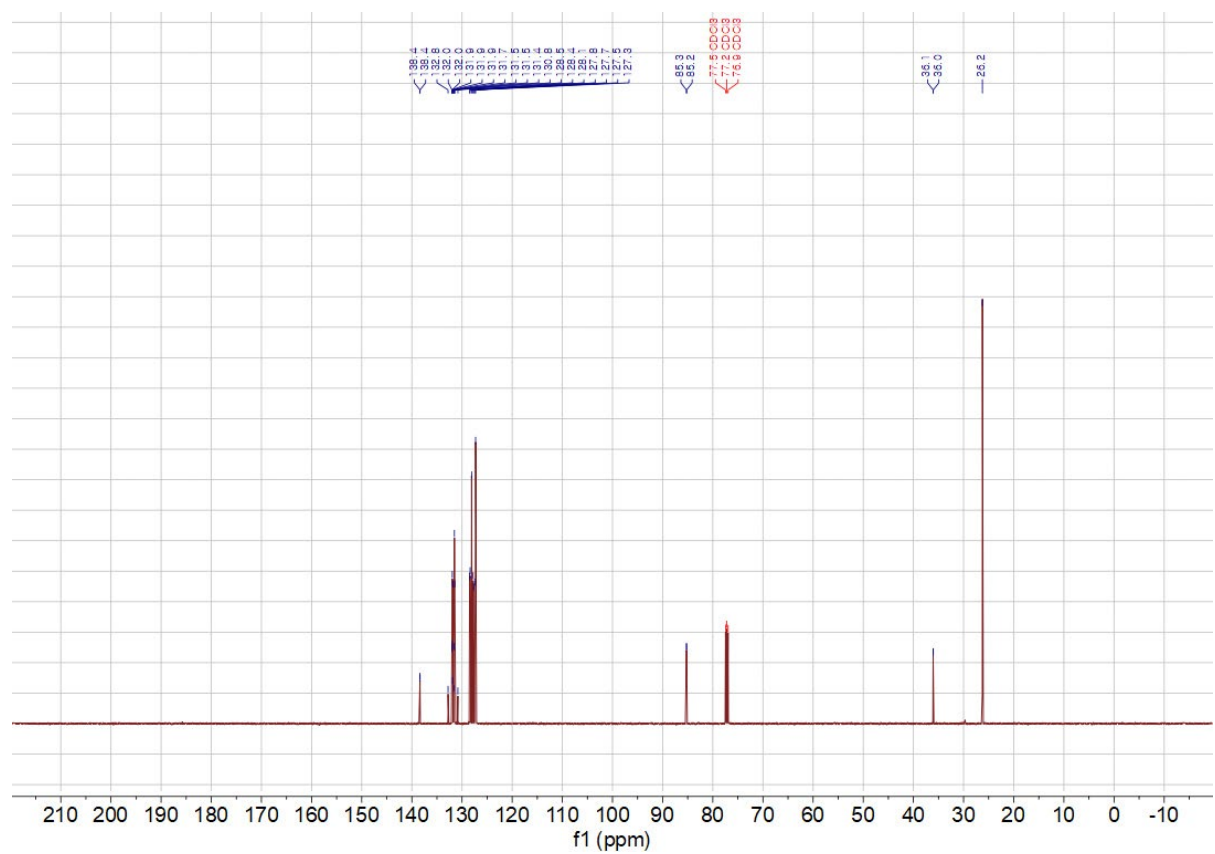
$^{31}\text{P}\{^1\text{H}\}$ NMR spectrum (202 MHz, CDCl_3) of **7-sub**



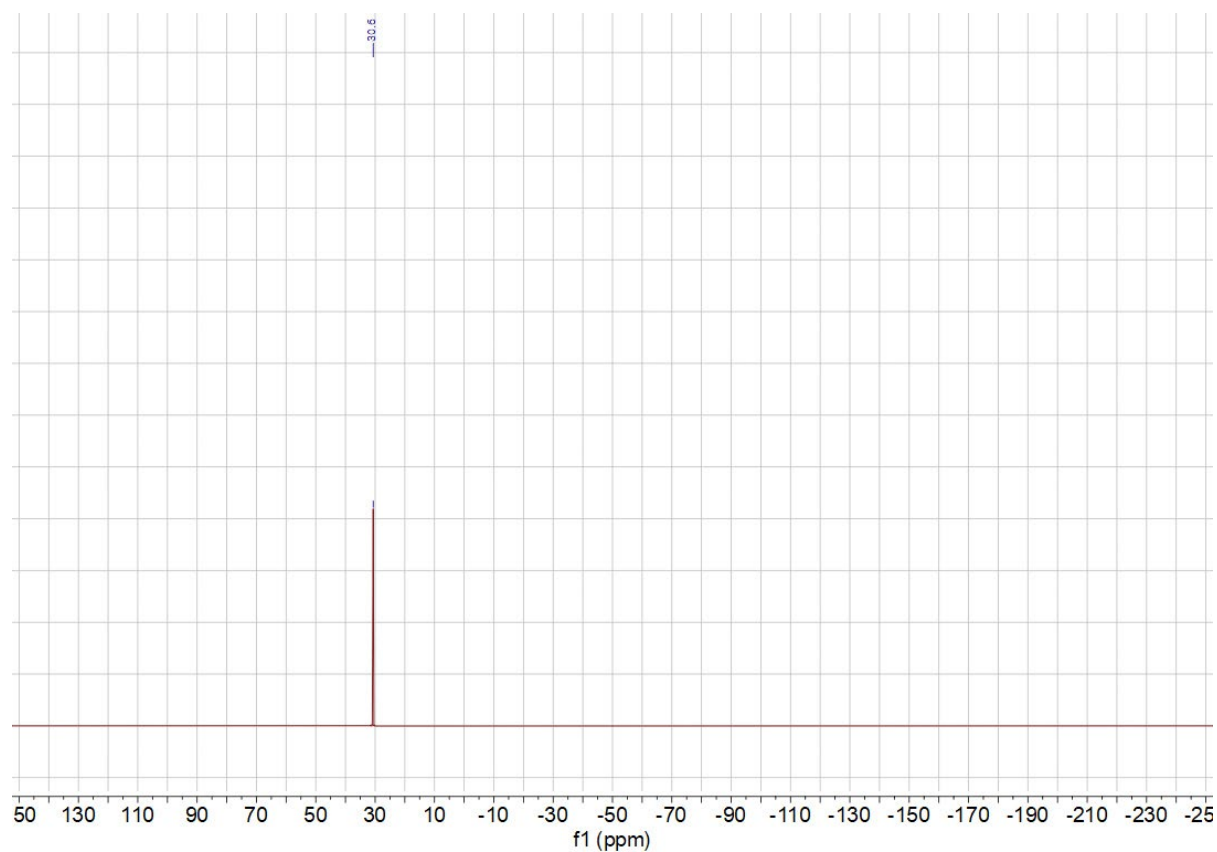
^1H NMR spectrum (500 MHz, CDCl_3) of **8-sub**



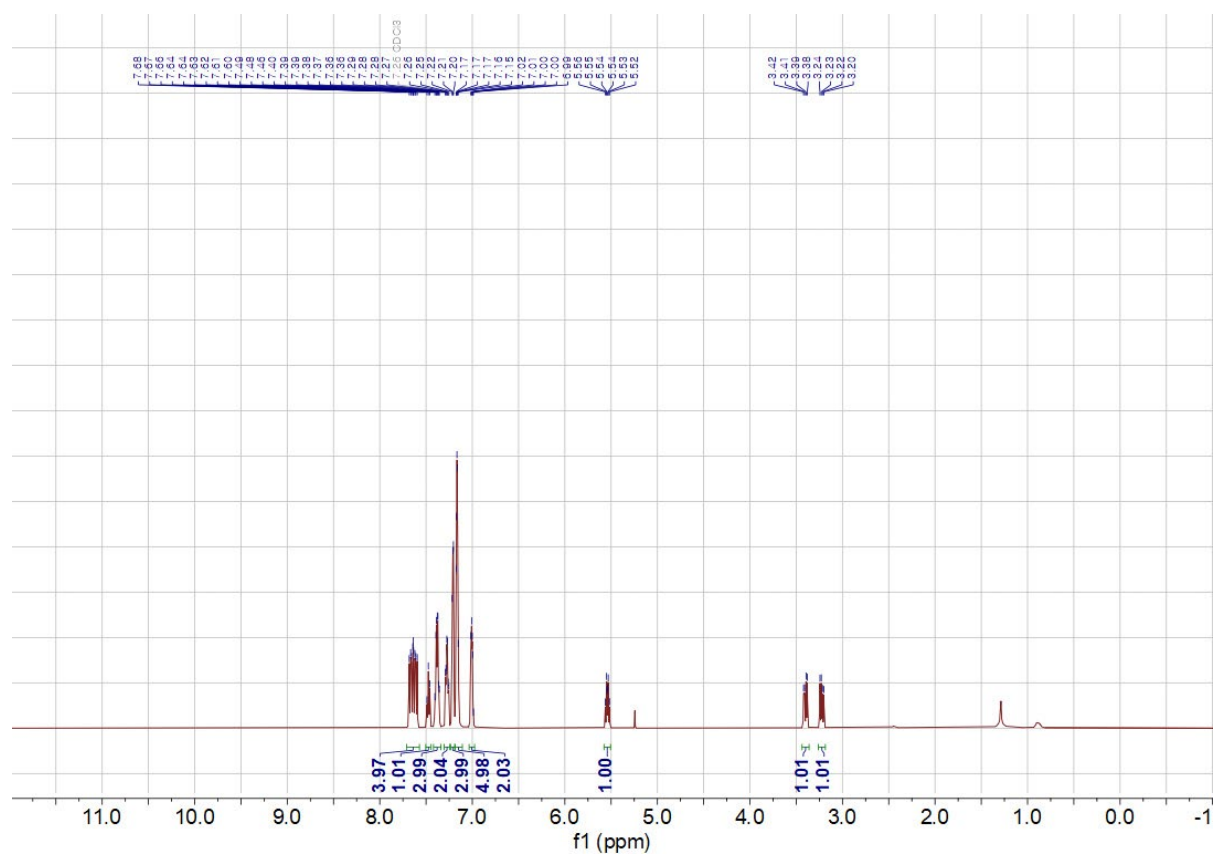
$^{13}\text{C}\{^1\text{H}\}$ NMR spectrum (126 MHz, CDCl_3) of **8-sub**



$^{31}\text{P}\{^1\text{H}\}$ NMR spectrum (202 MHz, CDCl_3) of **8-sub**

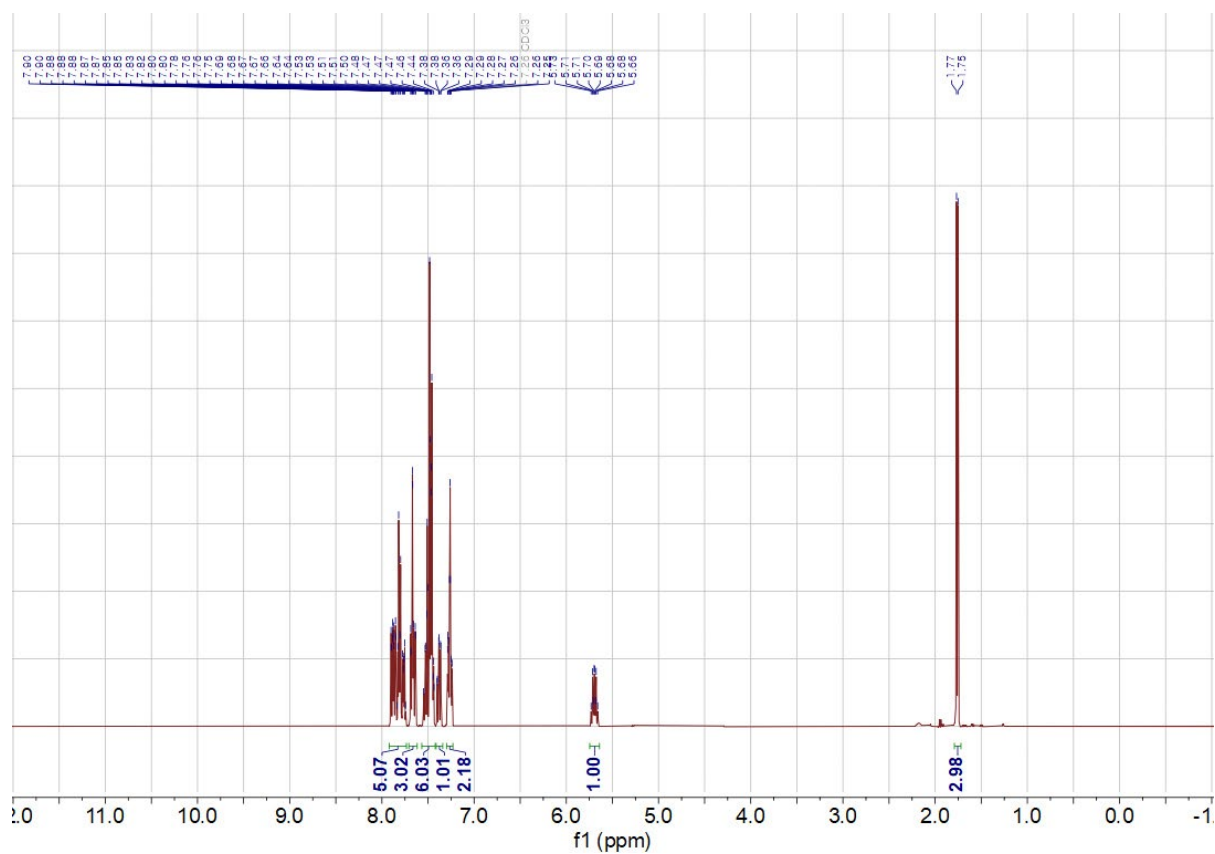


^1H NMR spectrum (500 MHz, CDCl_3) of **9-sub**

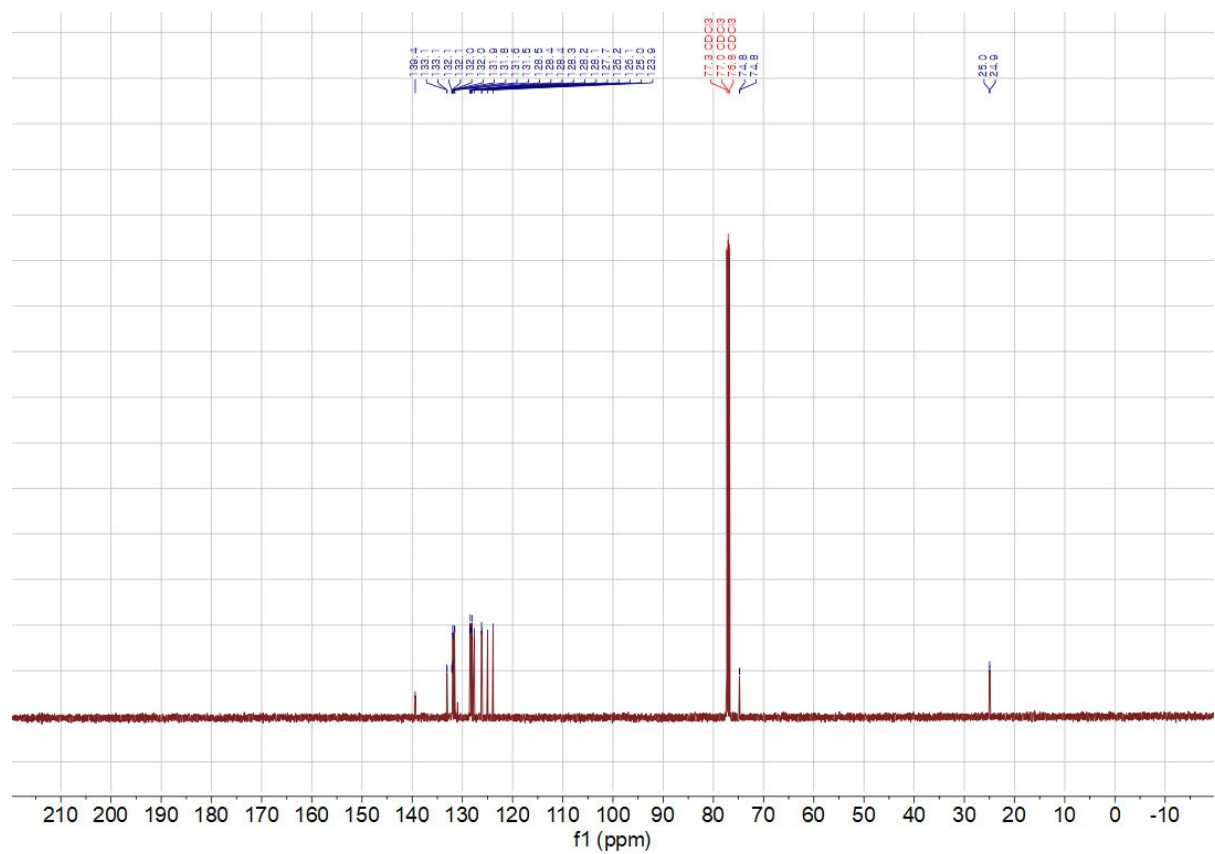


The figure shows a ^{13}C NMR spectrum with the x-axis labeled 'f1 (ppm)'. The axis scale is reversed, with values 50, 130, 110, 90, 70, 50, 30, 10, -10, -30, -50, -70, -90, -110, -130, -150, -170, -190, -210, -230, and -250. A single sharp peak is present at 30.4 ppm, indicated by a vertical line and the label '30.4'.

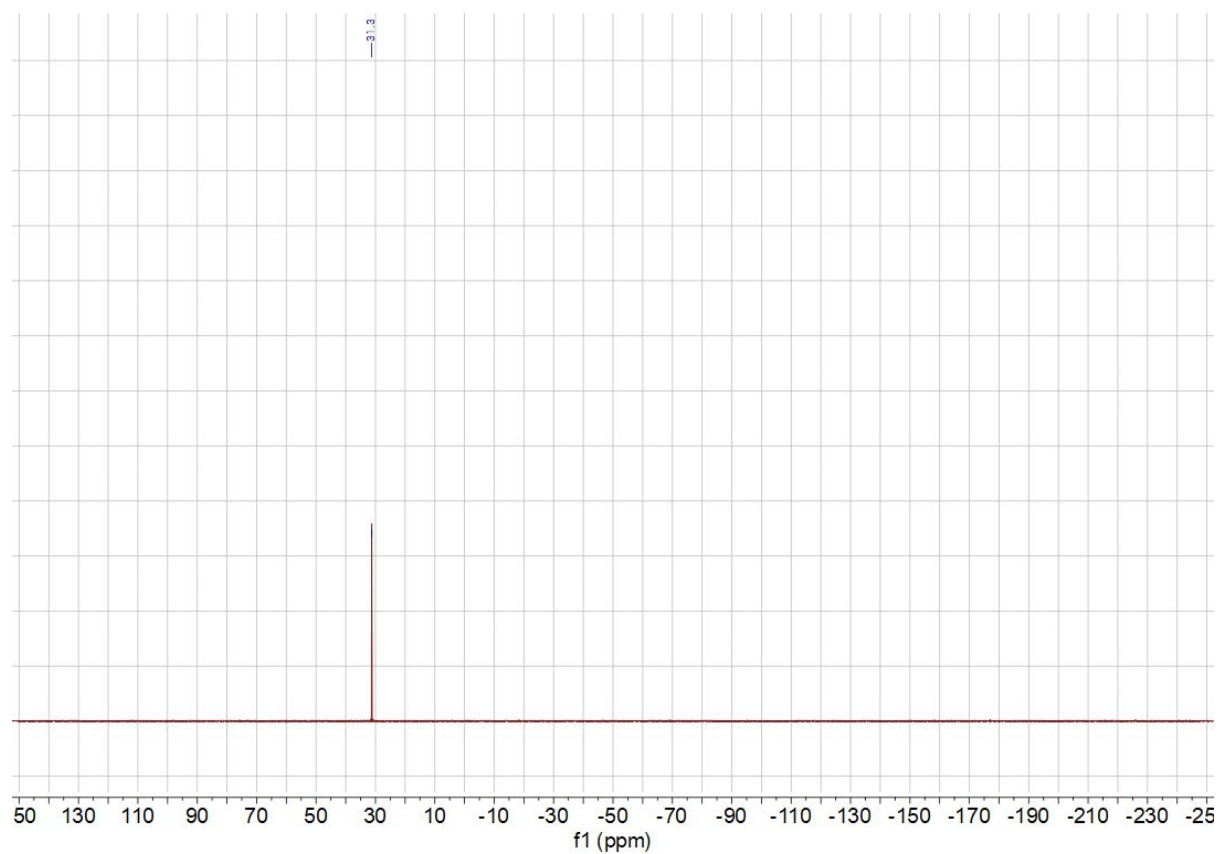
^1H NMR spectrum (400 MHz, CDCl_3) of **10-sub**



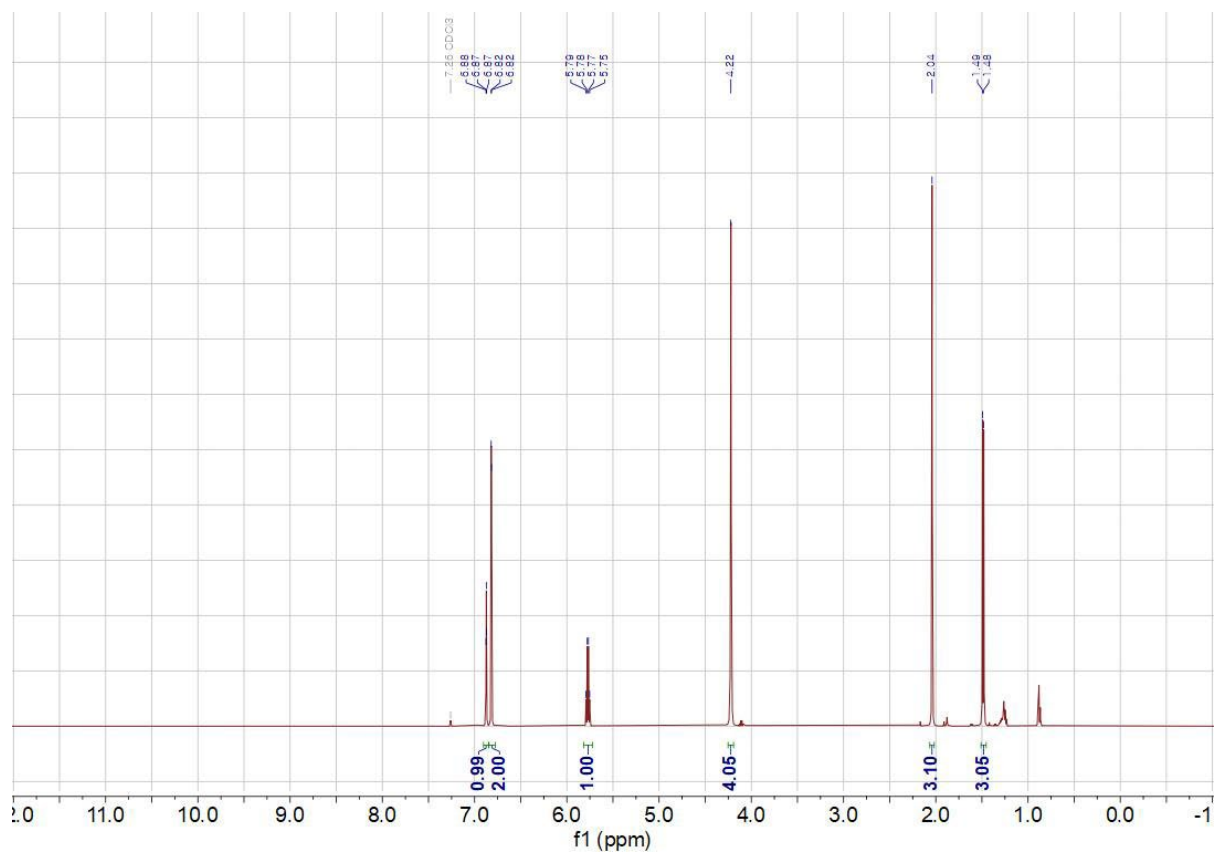
$^{13}\text{C}\{^1\text{H}\}$ NMR spectrum (126 MHz, CDCl_3) of **10-sub**



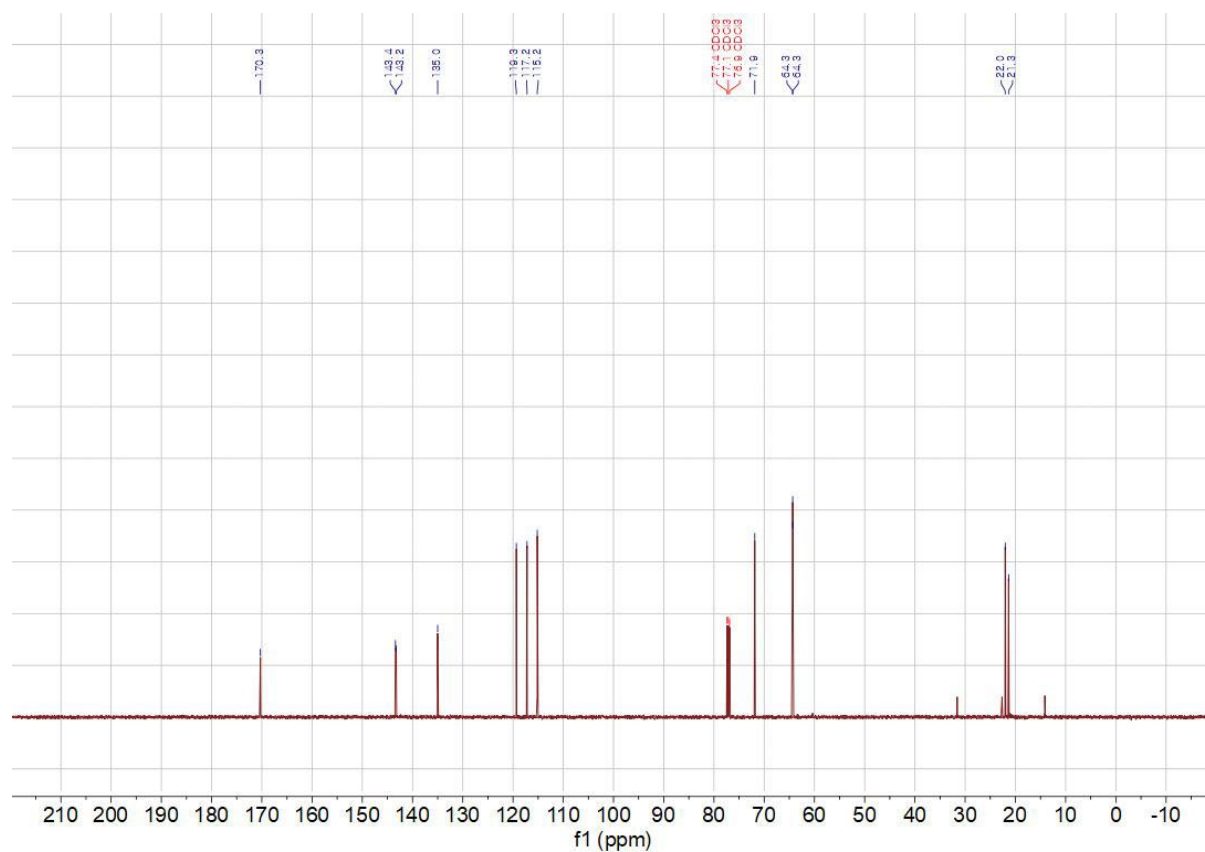
$^{31}\text{P}\{^1\text{H}\}$ NMR spectrum (202 MHz, CDCl_3) of **10-sub**



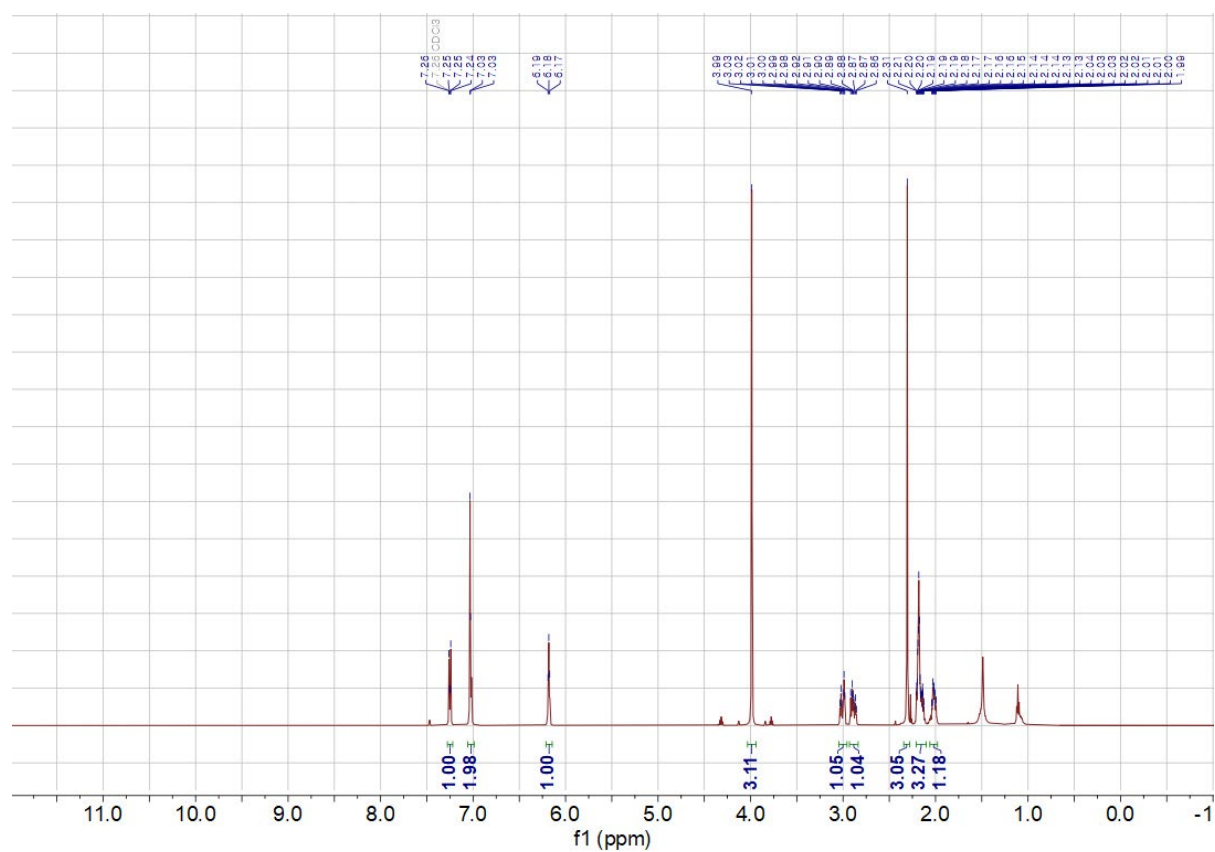
^1H NMR spectrum (500 MHz, CDCl_3) of **11-sub**



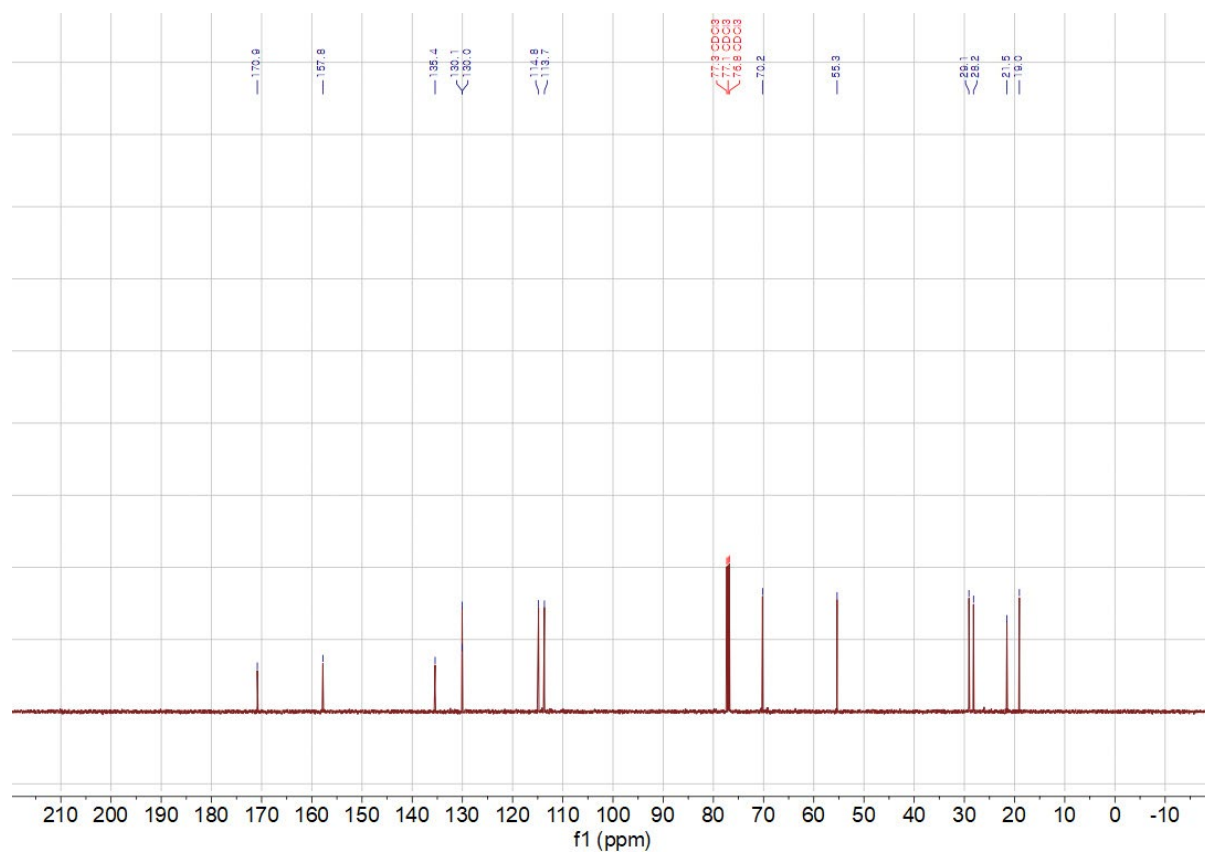
$^{13}\text{C}\{^1\text{H}\}$ NMR spectrum (126 MHz, CDCl_3) of **11-sub**



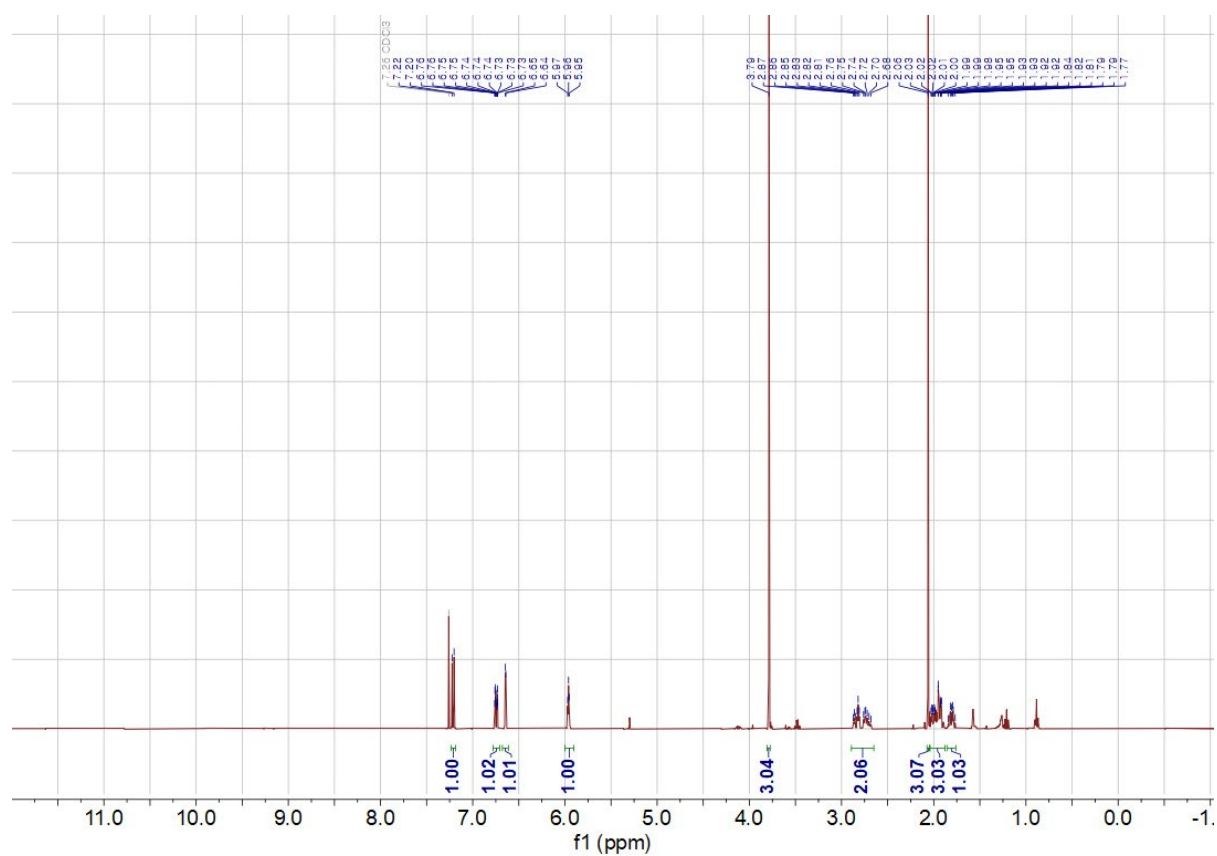
^1H NMR spectrum (500 MHz, CDCl_3) of **12-sub**



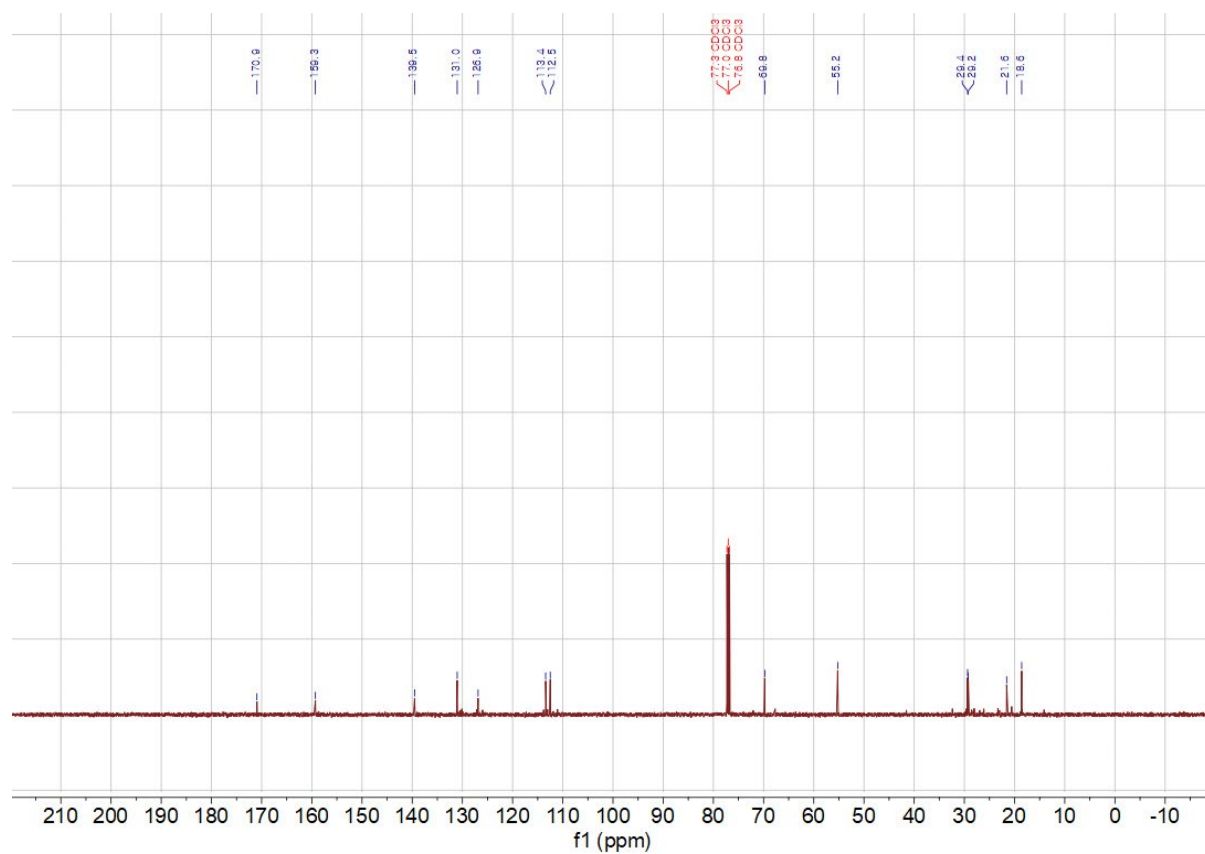
$^{13}\text{C}\{^1\text{H}\}$ NMR spectrum (126 MHz, CDCl_3) of **12-sub**



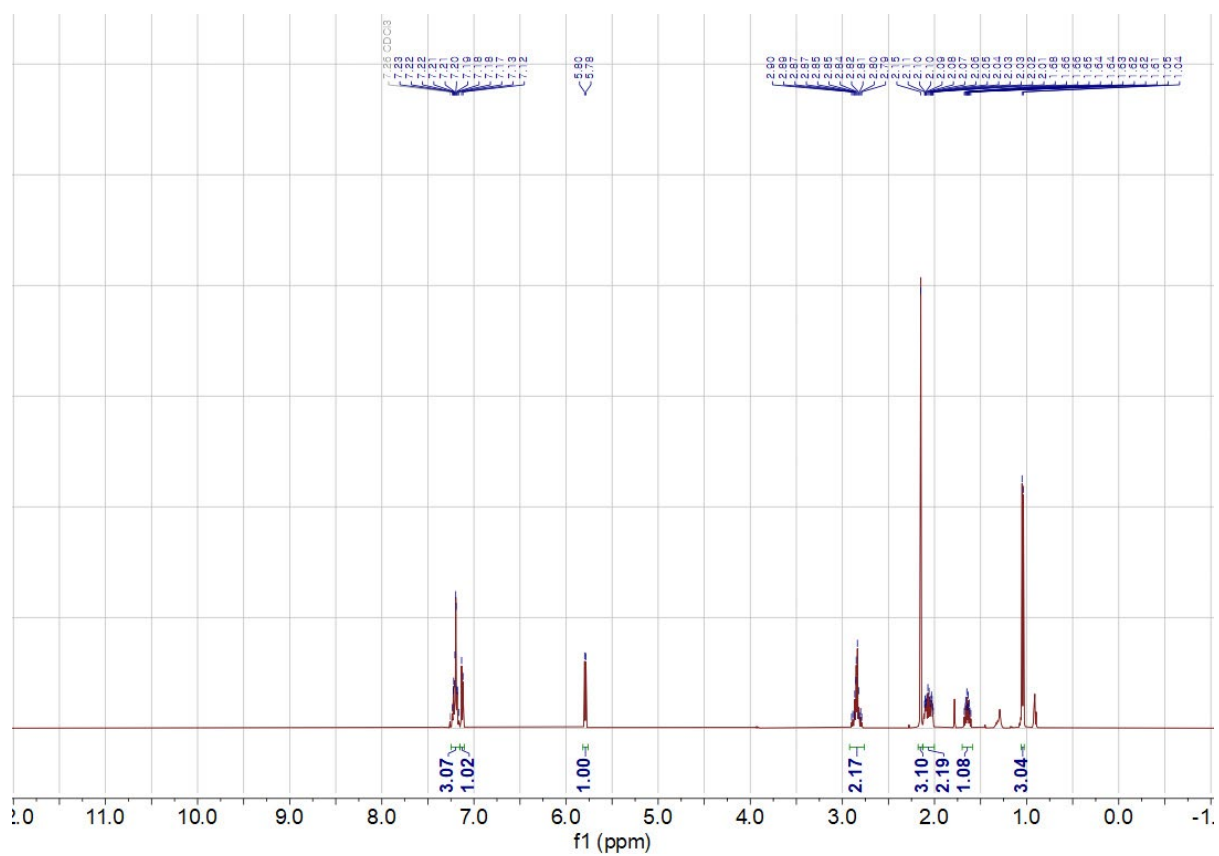
^1H NMR spectrum (400 MHz, CDCl_3) of **13-sub**



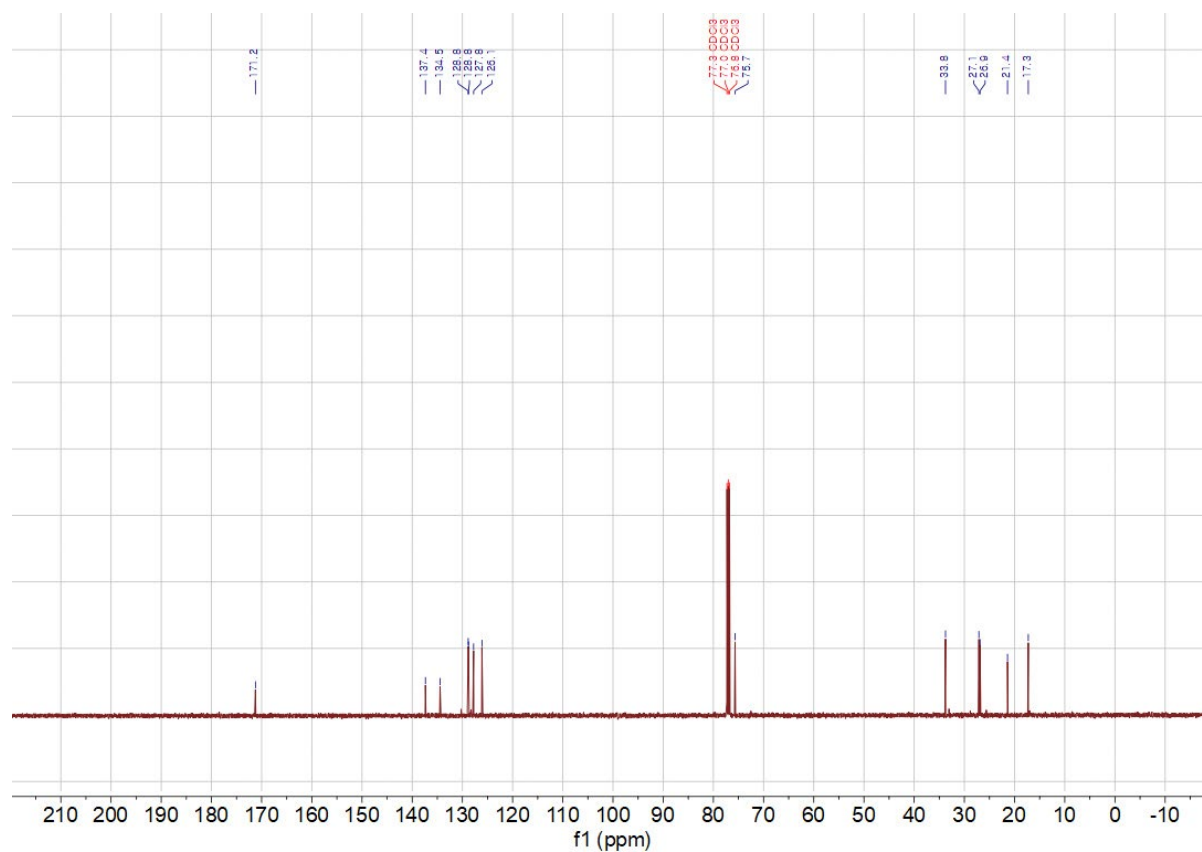
$^{13}\text{C}\{^1\text{H}\}$ NMR spectrum (126 MHz, CDCl_3) of **13-sub**



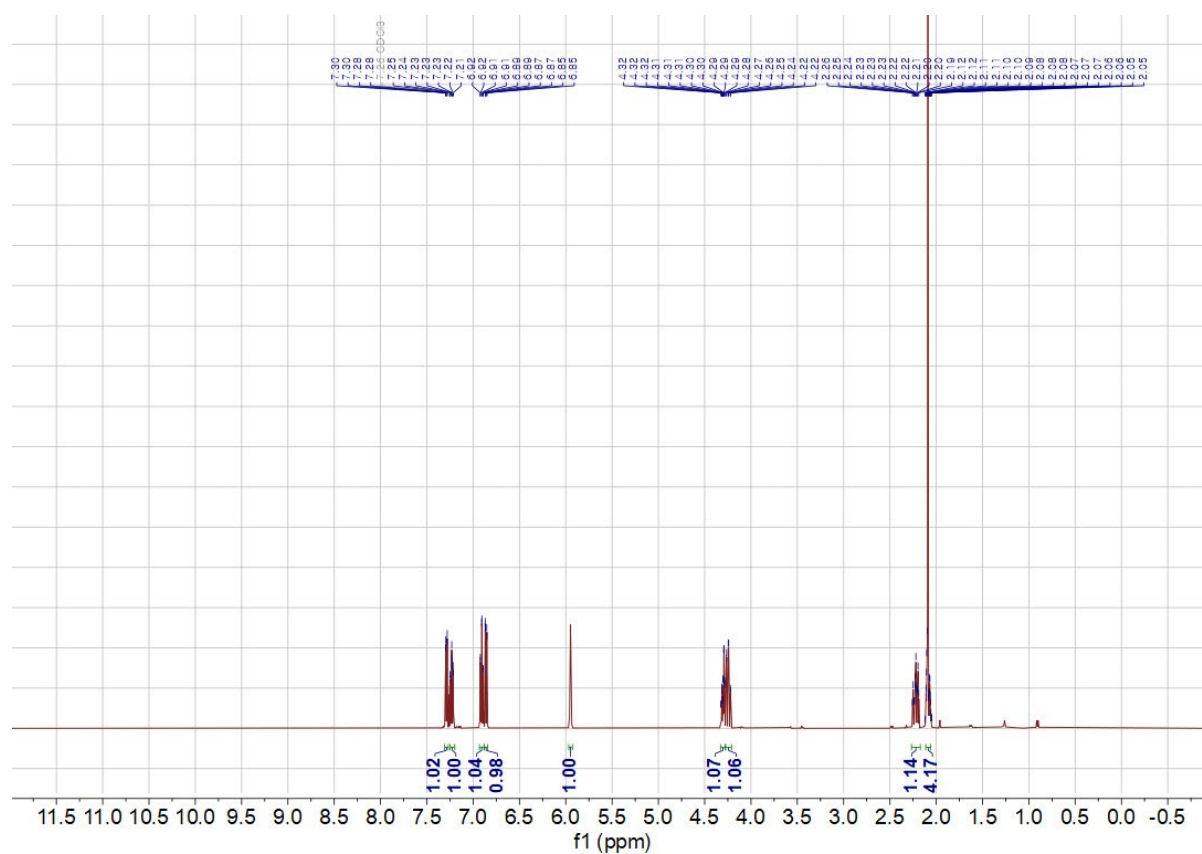
^1H NMR spectrum (500 MHz, CDCl_3) of **14-sub**



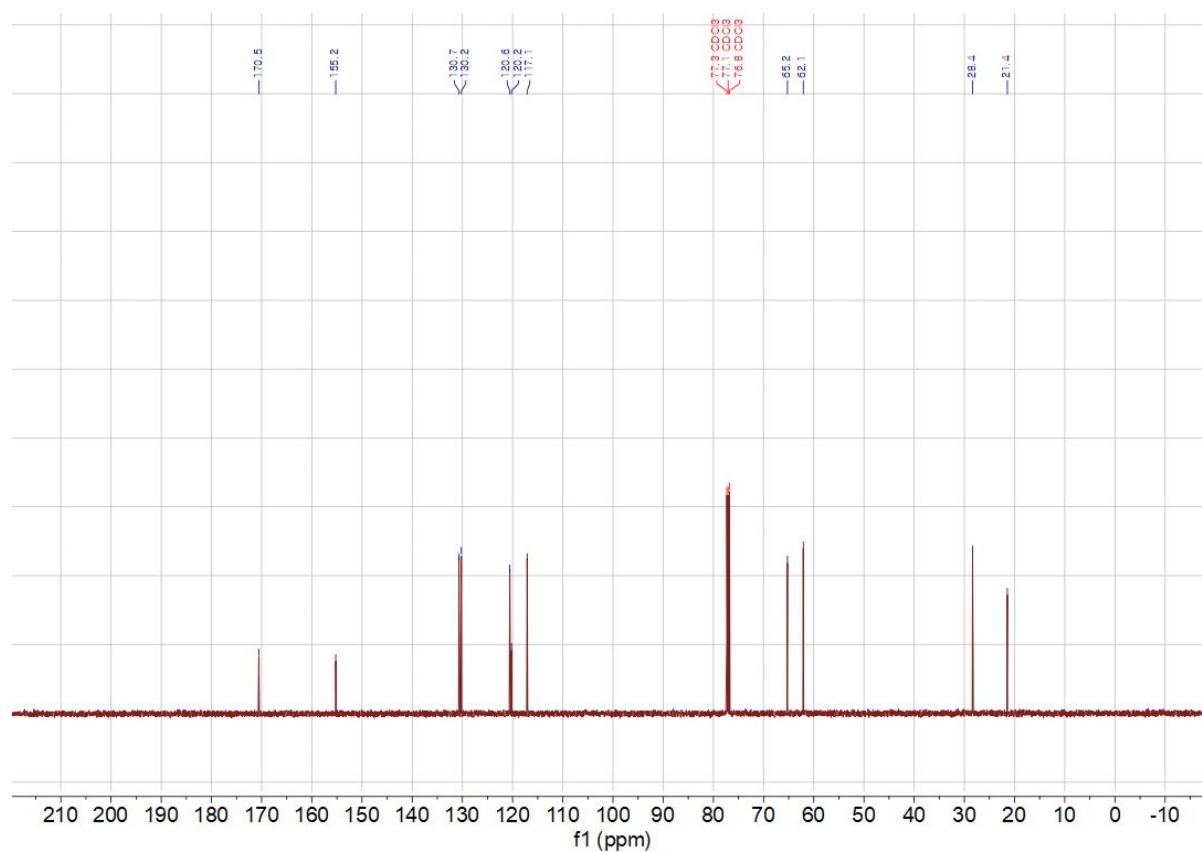
$^{13}\text{C}\{^1\text{H}\}$ NMR spectrum (126 MHz, CDCl_3) of **14-sub**



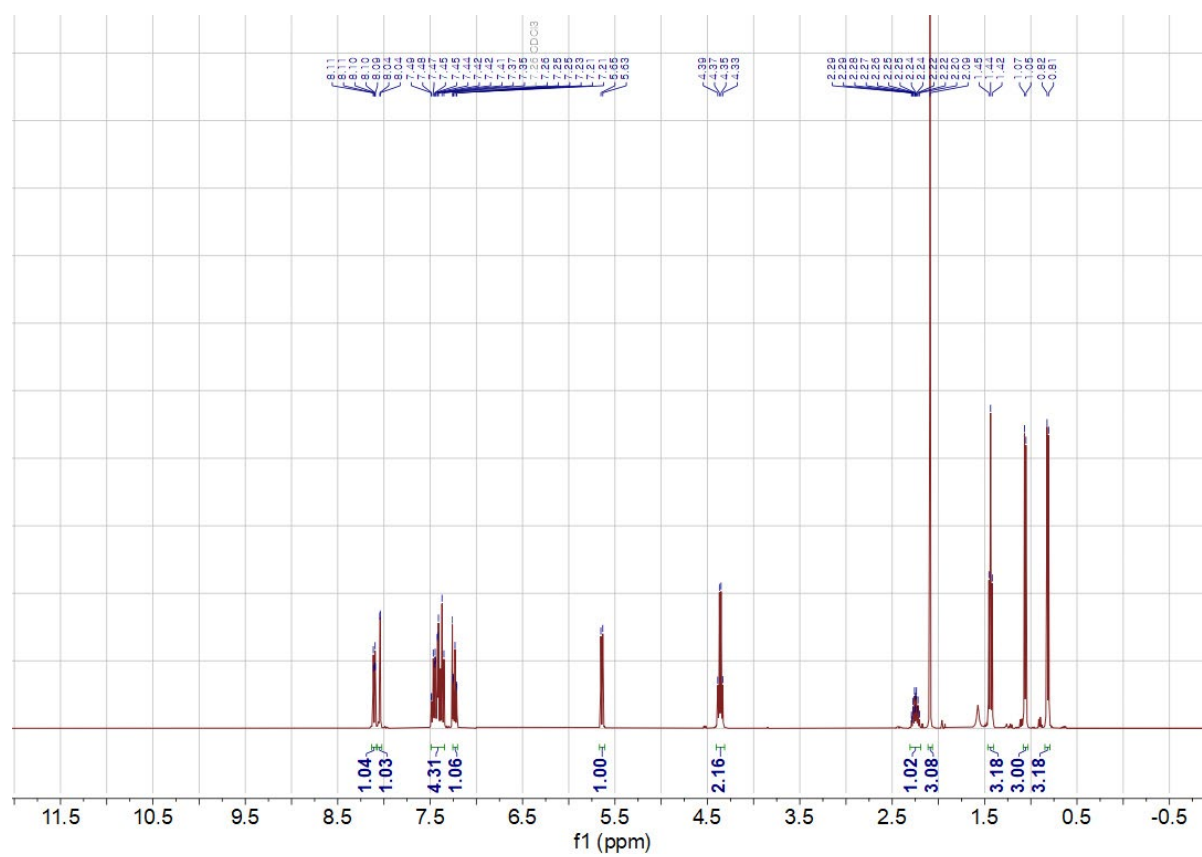
^1H NMR spectrum (500 MHz, CDCl_3) of **15-sub**



$^{13}\text{C}\{^1\text{H}\}$ NMR spectrum (126 MHz, CDCl_3) of **15-sub**

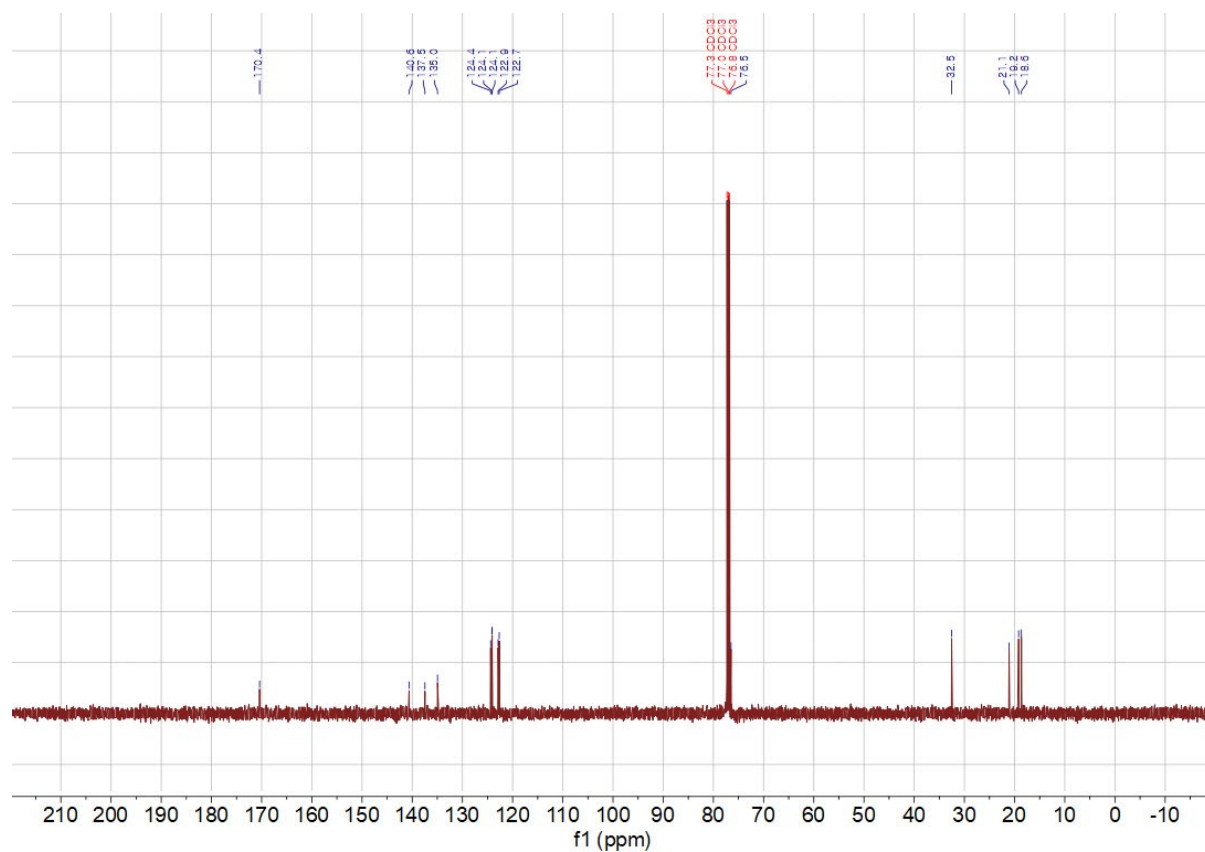


^1H NMR spectrum (500 MHz, CDCl_3) of **16-sub**

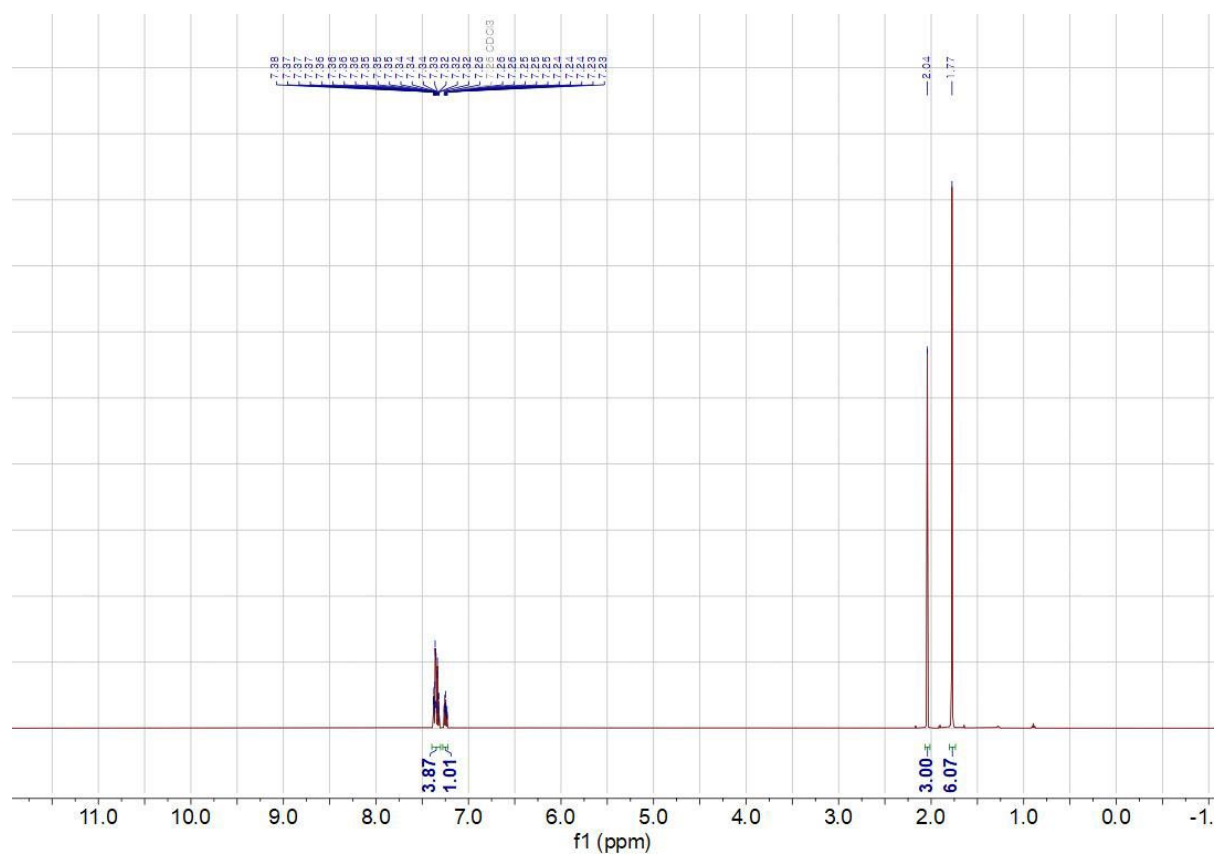


¹H NMR spectrum (CDCl₃) of 1,2-dichloroethane. The x-axis represents the chemical shift f1 in ppm, ranging from -1 to 11.0. The spectrum shows a triplet at approximately 1.0 ppm (integration 3.11, 3.23), a quartet at approximately 2.1 ppm (integration 1.08, 2.98), and a multiplet at approximately 7.2 ppm (integration 1.03, 1.01, 3.05). A solvent peak for CDCl₃ is visible at approximately 7.26 ppm, and a water peak is at approximately 3.3 ppm.

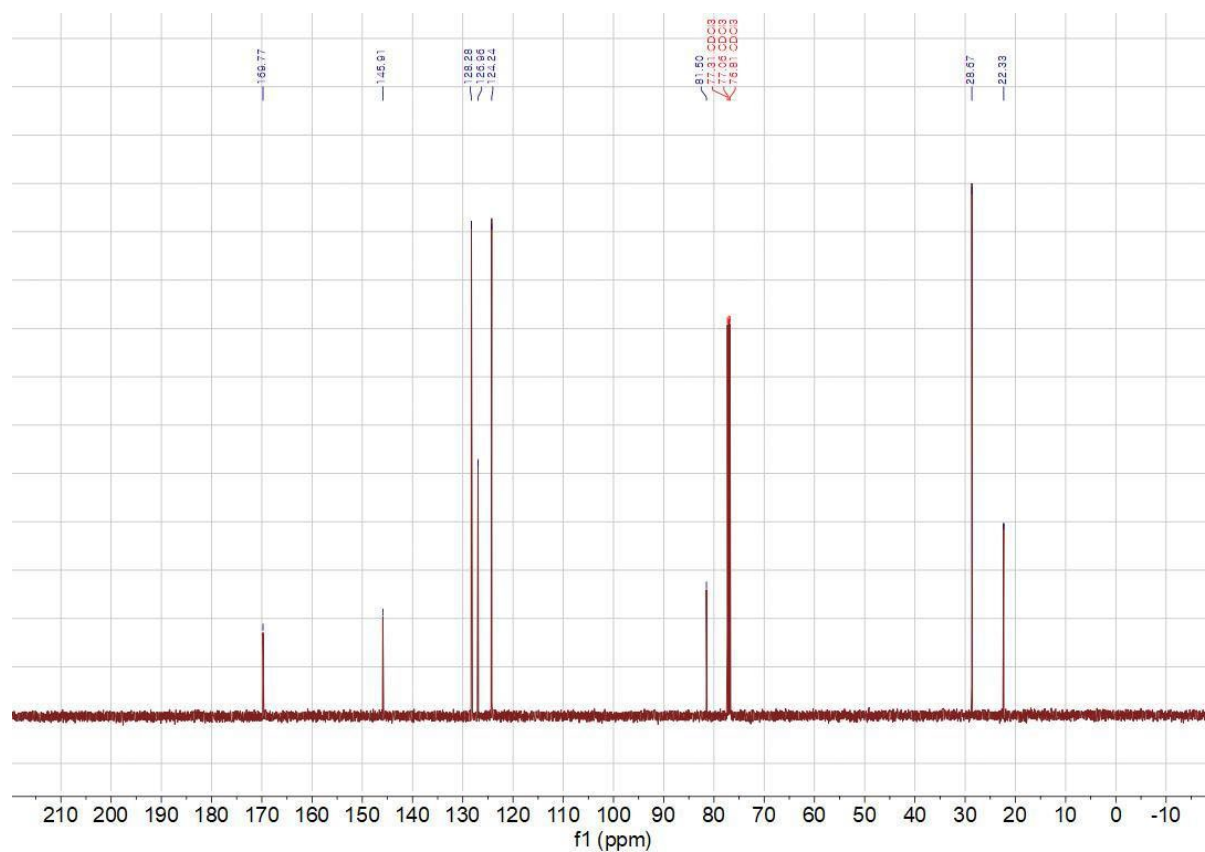
$^{13}\text{C}\{^1\text{H}\}$ NMR spectrum (126 MHz, CDCl_3) of **17-sub**



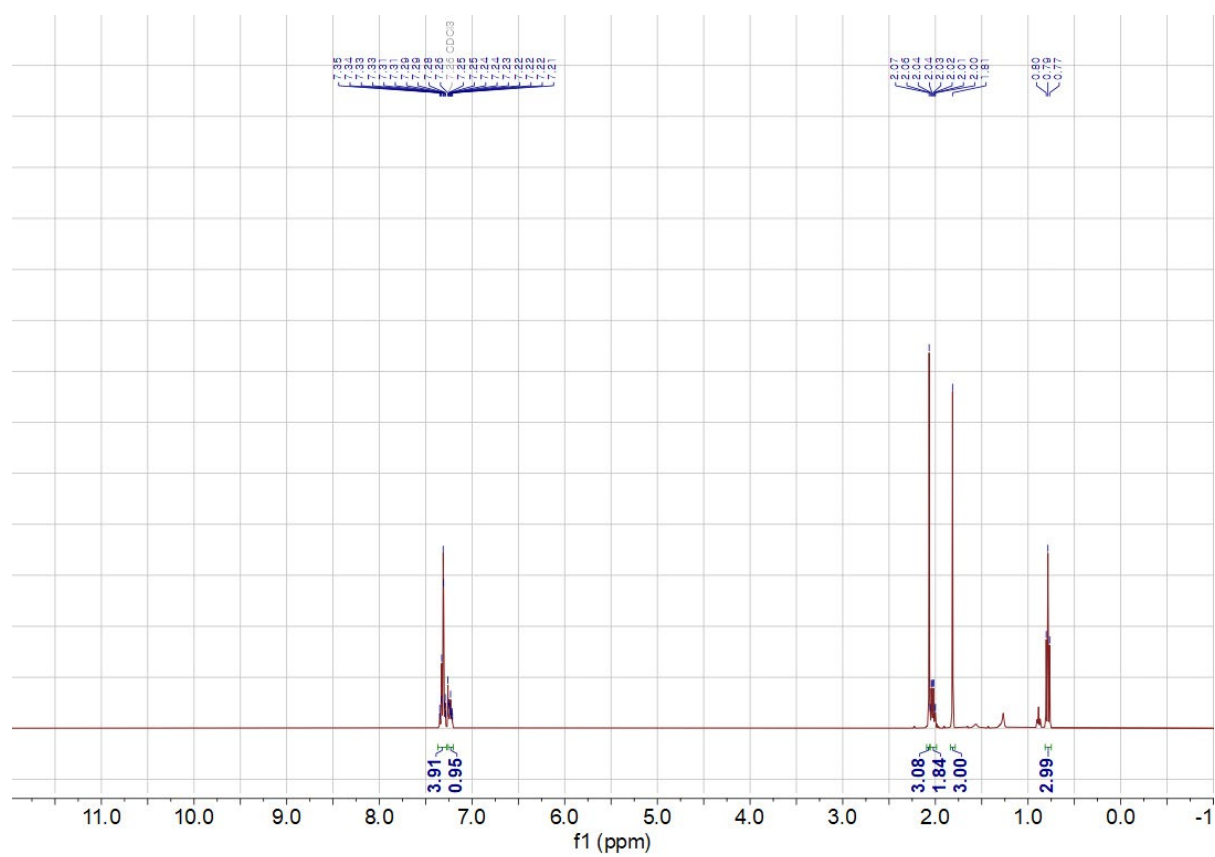
^1H NMR spectrum (500 MHz, CDCl_3) of **18-sub**



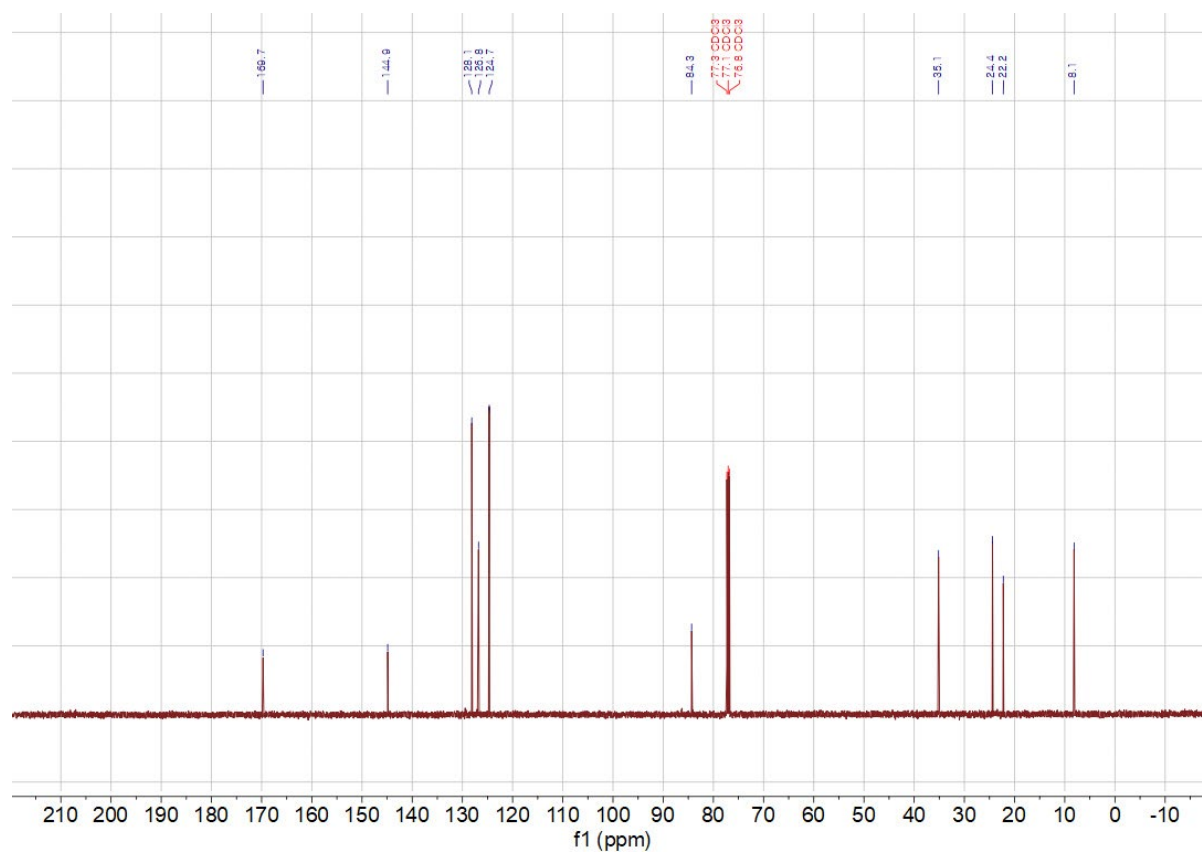
$^{13}\text{C}\{^1\text{H}\}$ NMR spectrum (126 MHz, CDCl_3) of **18-sub**



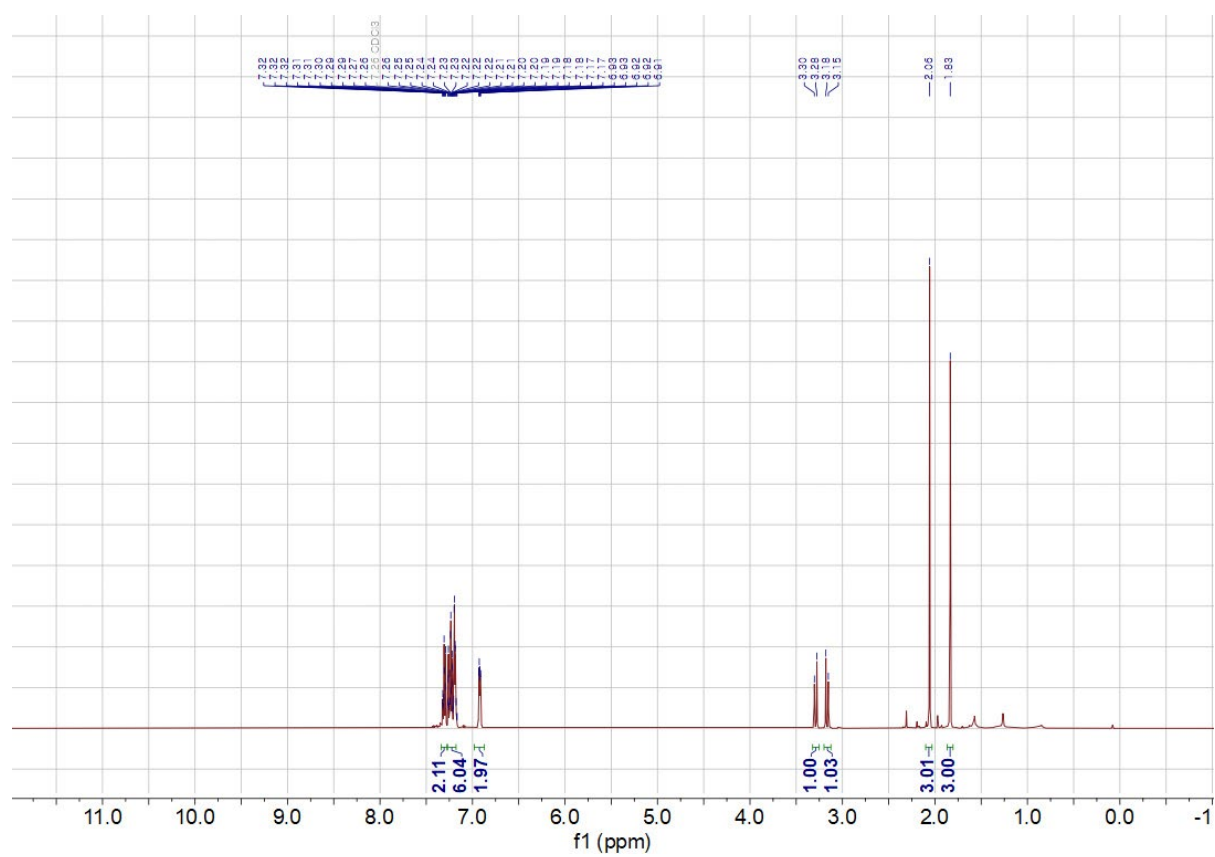
^1H NMR spectrum (400 MHz, CDCl_3) of **19-sub**



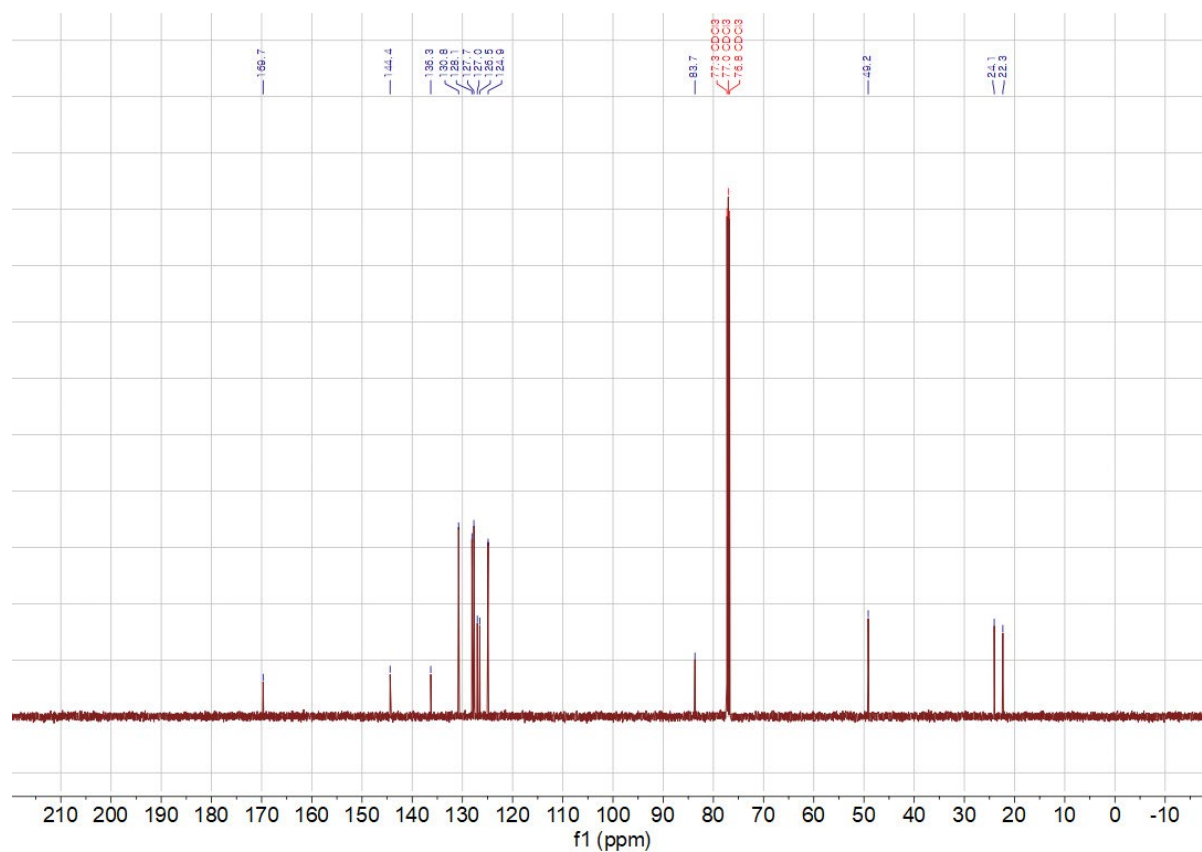
$^{13}\text{C}\{^1\text{H}\}$ NMR spectrum (126 MHz, CDCl_3) of **19-sub**



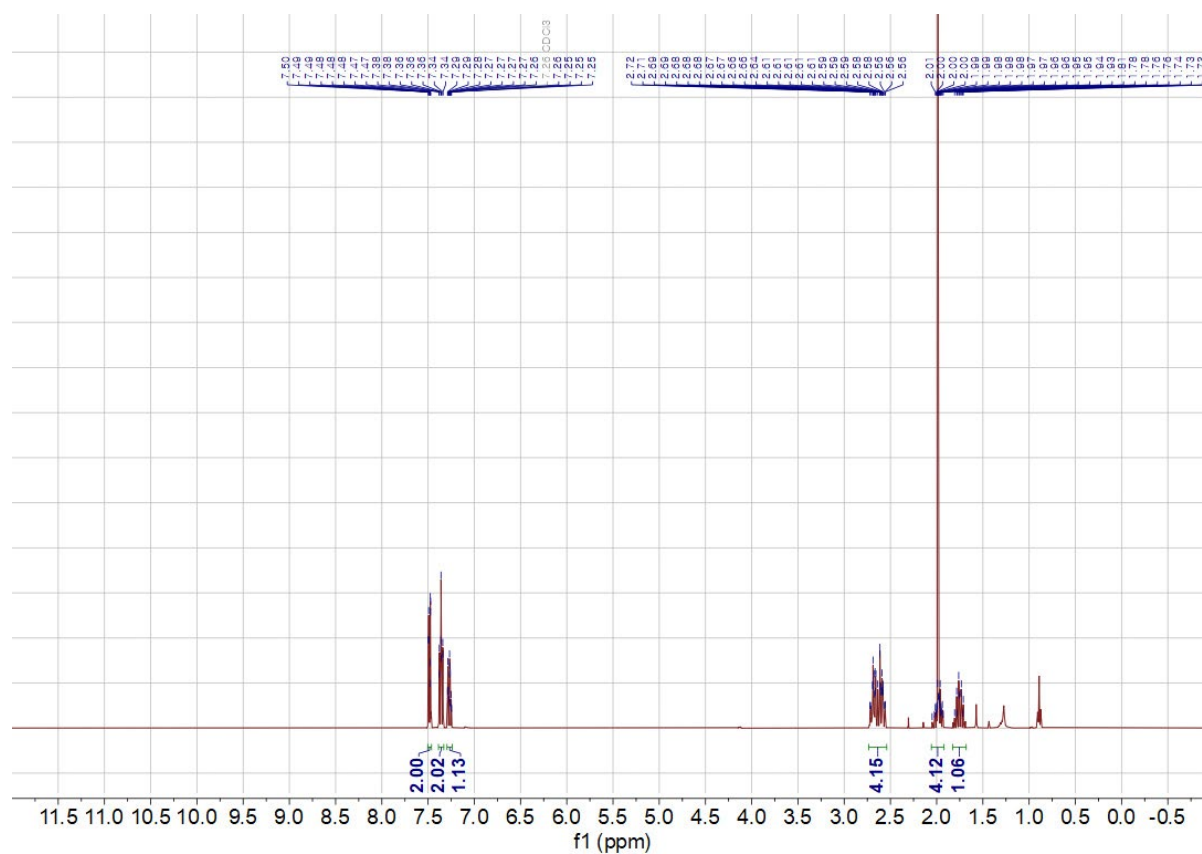
^1H NMR spectrum (500 MHz, CDCl_3) of **20-sub**



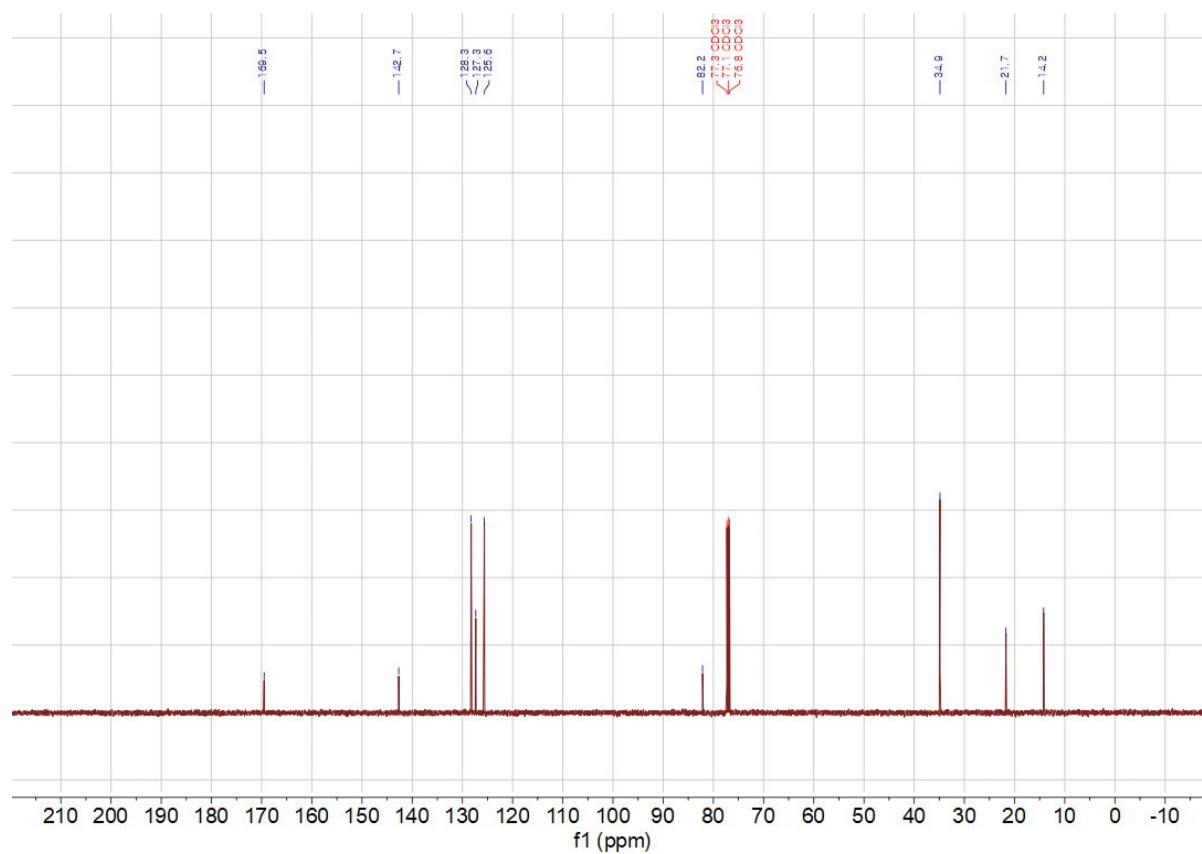
$^{13}\text{C}\{^1\text{H}\}$ NMR spectrum (126 MHz, CDCl_3) of **20-sub**



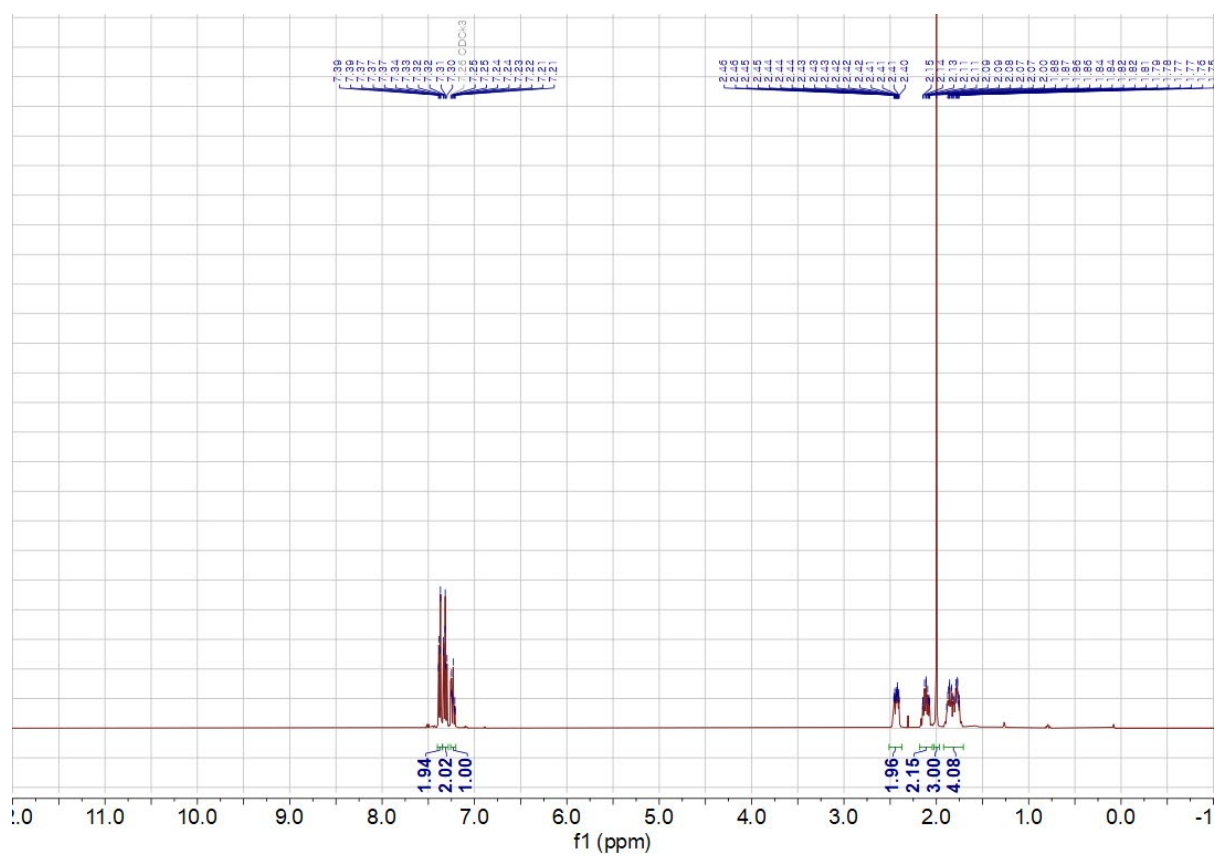
^1H NMR spectrum (400 MHz, CDCl_3) of **21-sub**



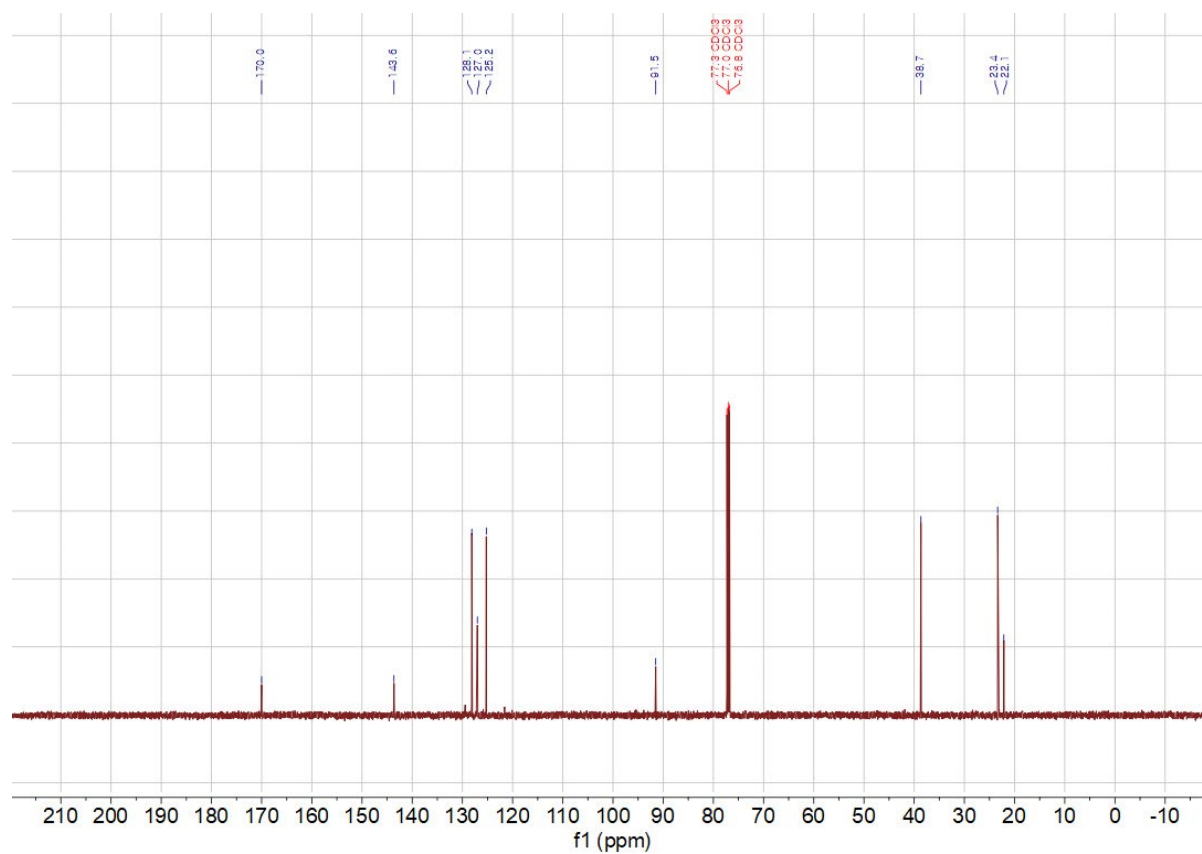
$^{13}\text{C}\{^1\text{H}\}$ NMR spectrum (126 MHz, CDCl_3) of **21-sub**



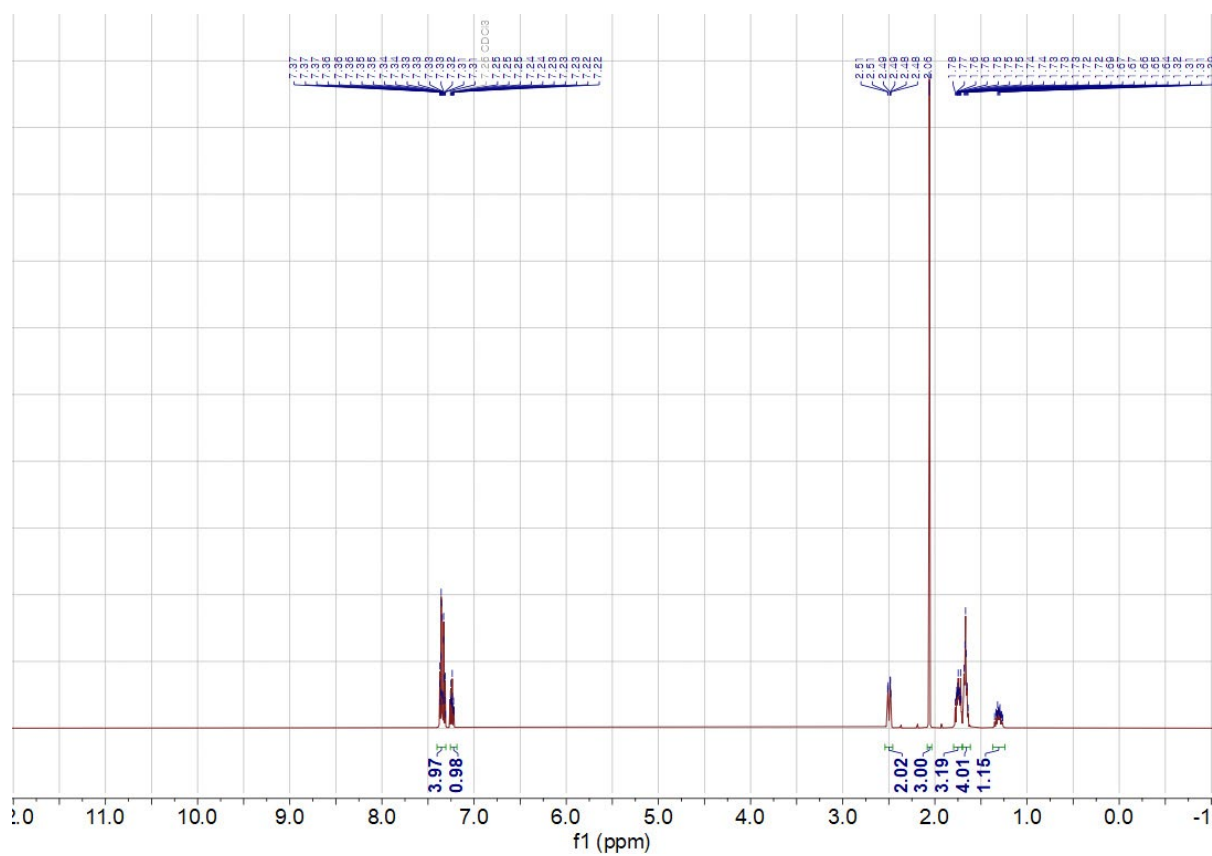
^1H NMR spectrum (400 MHz, CDCl_3) of **22-sub**



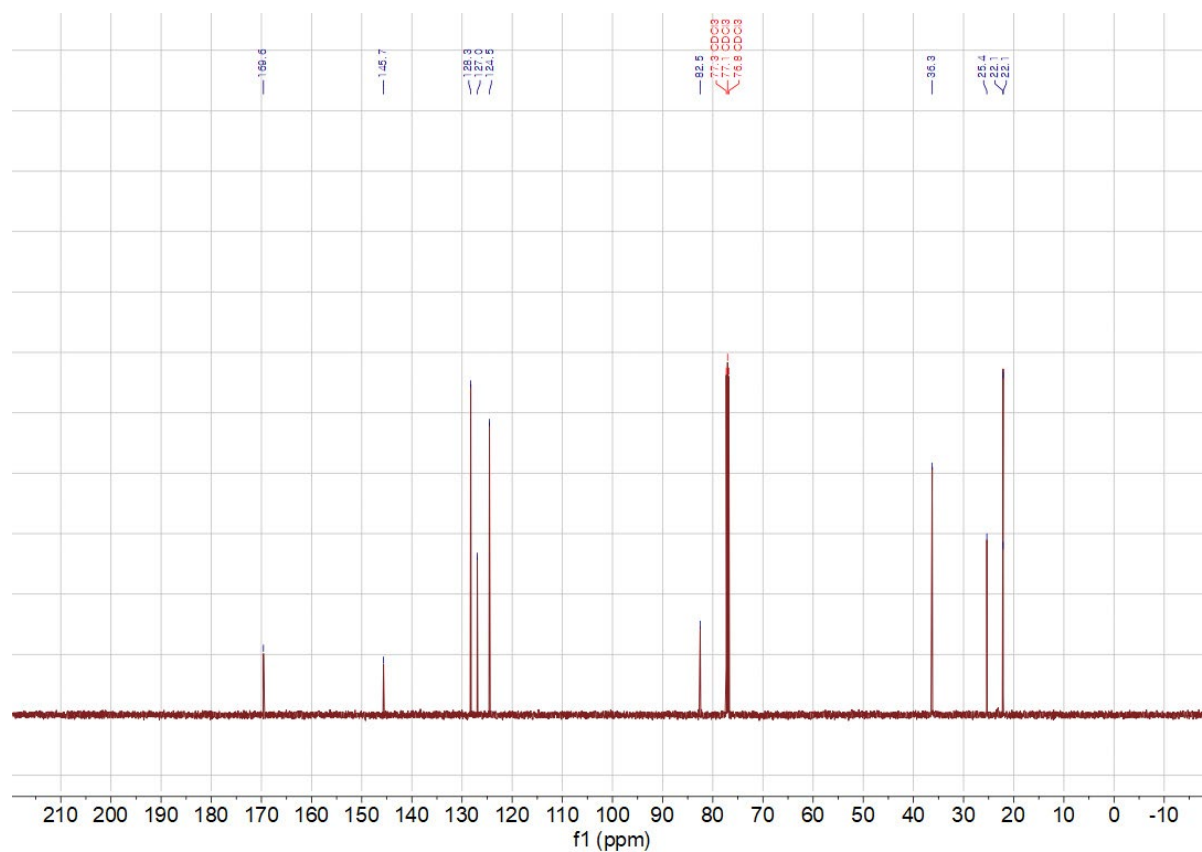
$^{13}\text{C}\{^1\text{H}\}$ NMR spectrum (126 MHz, CDCl_3) of **22-sub**



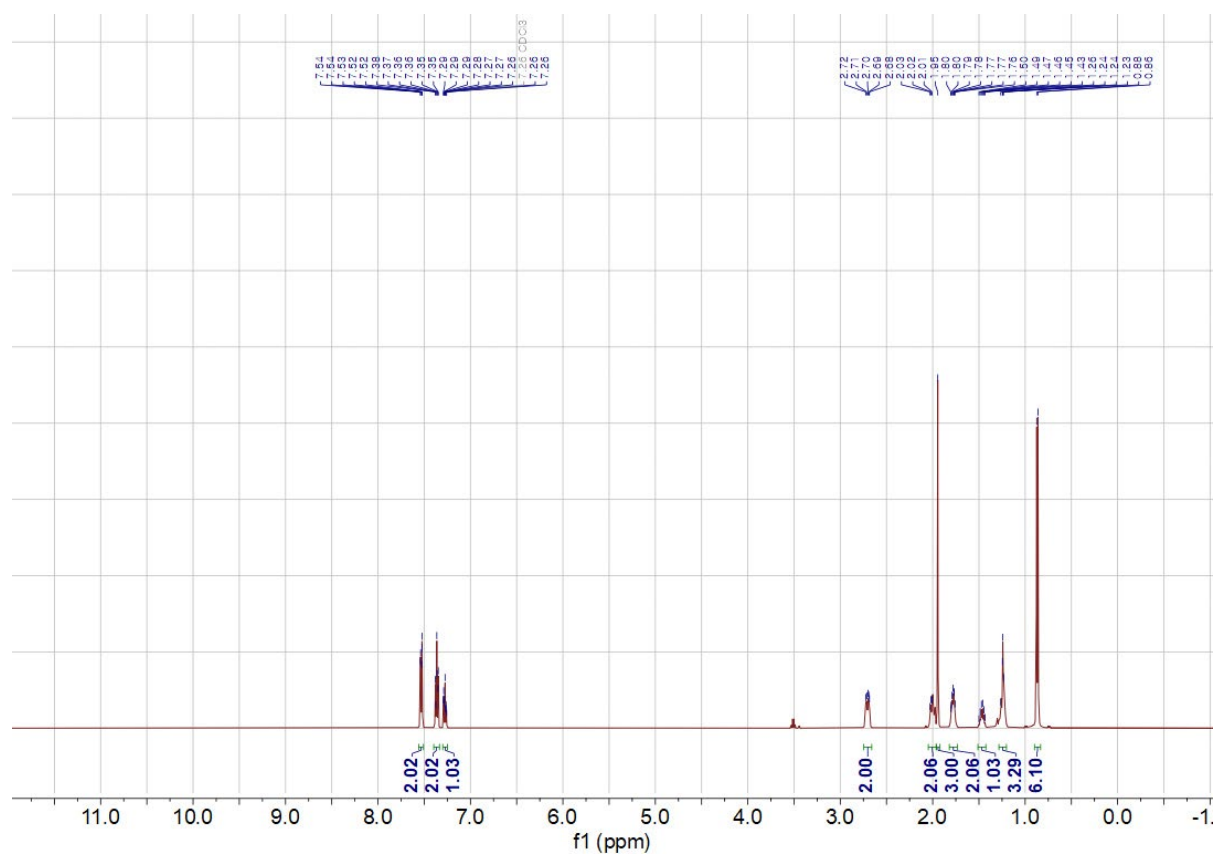
^1H NMR spectrum (500 MHz, CDCl_3) of **23-sub**



$^{13}\text{C}\{^1\text{H}\}$ NMR spectrum (126 MHz, CDCl_3) of **23-sub**



^1H NMR spectrum (500 MHz, CDCl_3) of **24-sub**

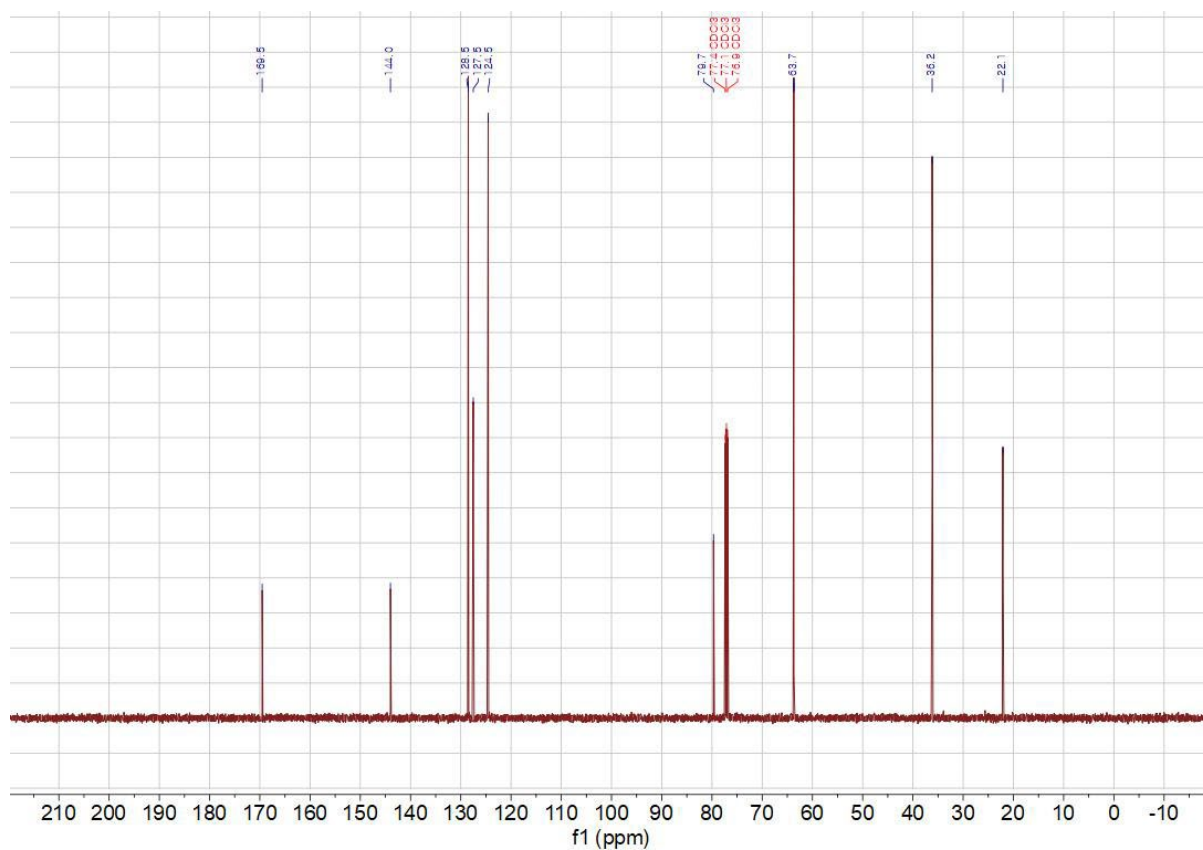


13C NMR spectrum of compound 10a in CDCl₃. The x-axis is labeled 'f1 (ppm)' and ranges from -10 to 210. The spectrum shows several peaks with the following chemical shifts and integration values:

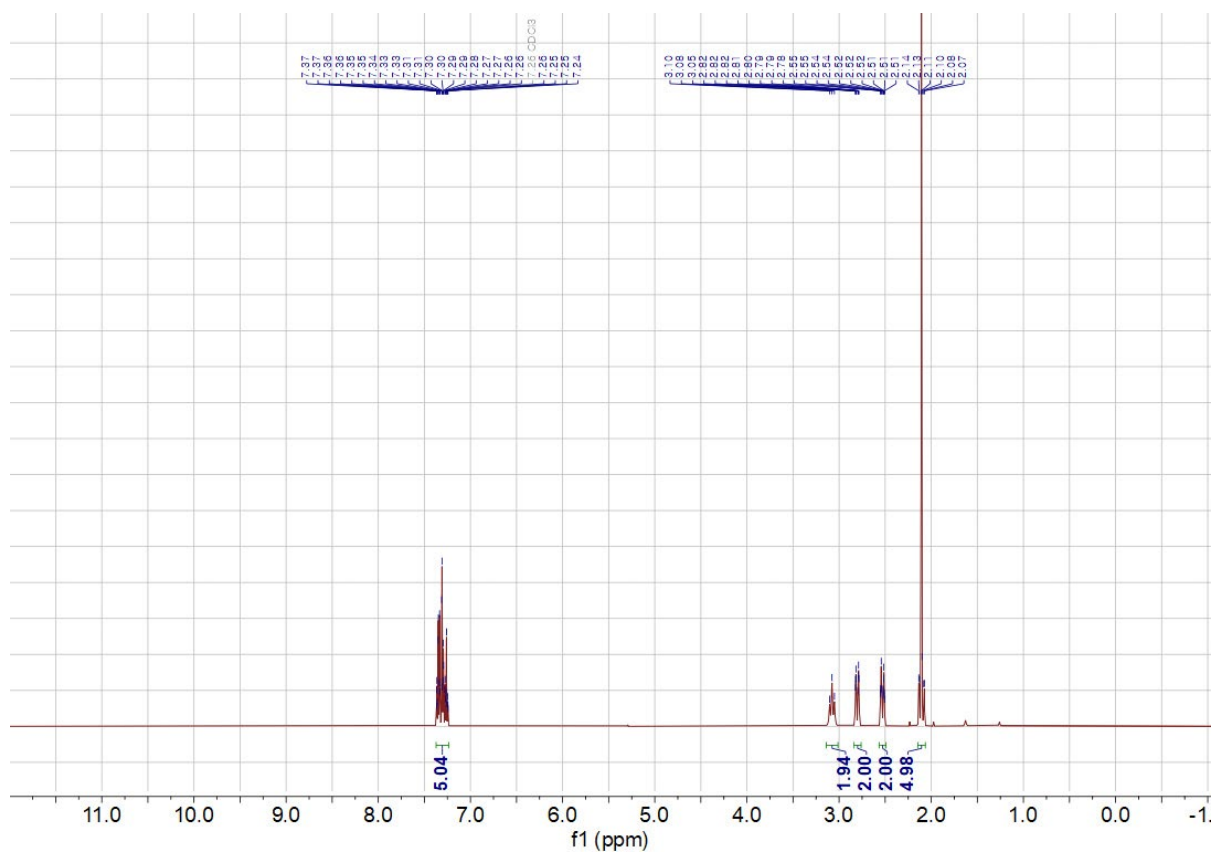
Chemical Shift (ppm)	Integration
170.6	1.00
142.1	1.00
128.1	0.99
127.1	0.99
126.1	0.99
125.1	0.99
82.1	0.99
77.0	0.99
77.0	0.99
77.0	0.99
40.1	0.99
35.1	0.99
30.1	0.99
25.1	0.99
20.1	0.99

1H NMR spectrum of 1,4-dichlorobenzene in CDCl₃. The spectrum shows a multiplet at 7.26 ppm (integration 3.37, 1.01), a multiplet at 3.74 ppm (integration 2.06, 2.05), and a multiplet at 2.36 ppm (integration 2.05, 2.09, 3.00). A large solvent peak is visible at 7.26 ppm.

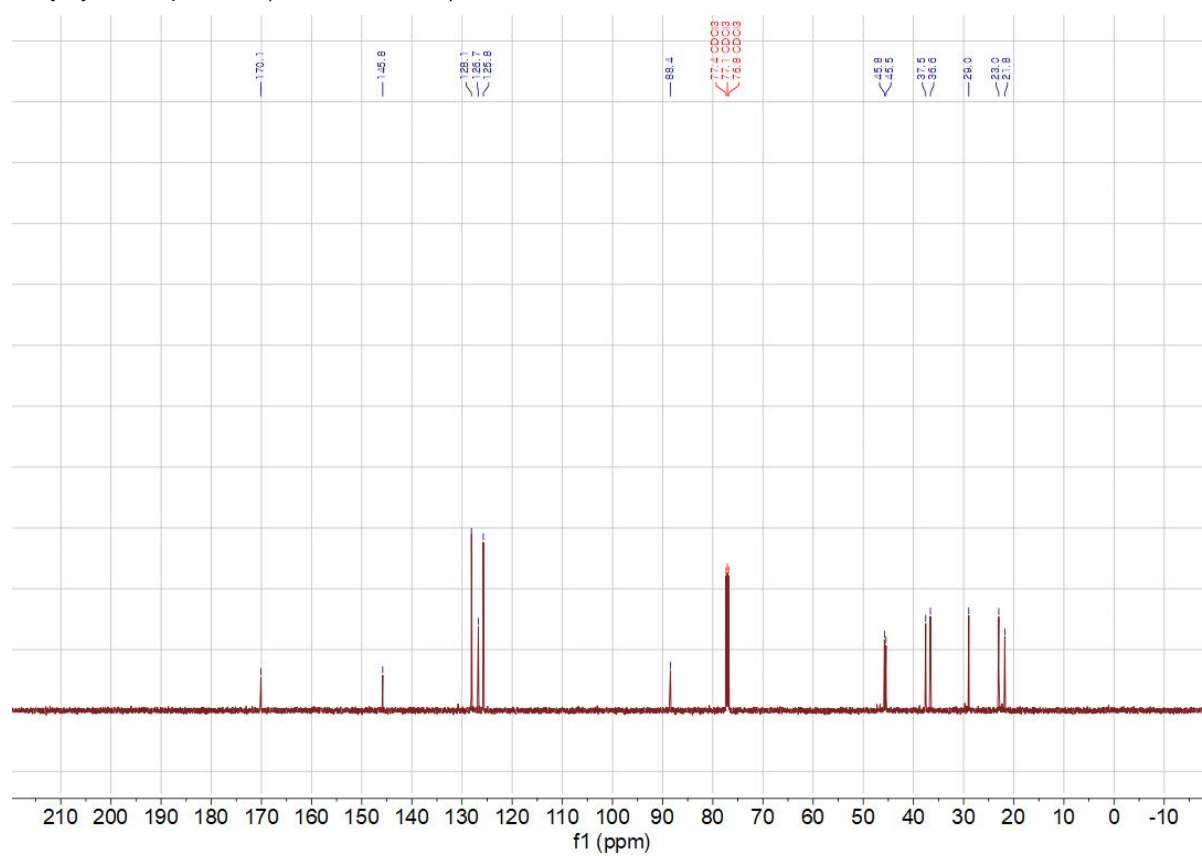
$^{13}\text{C}\{^1\text{H}\}$ NMR spectrum (126 MHz, CDCl_3) of **25-sub**



^1H NMR spectrum (500 MHz, CDCl_3) of **26-sub**

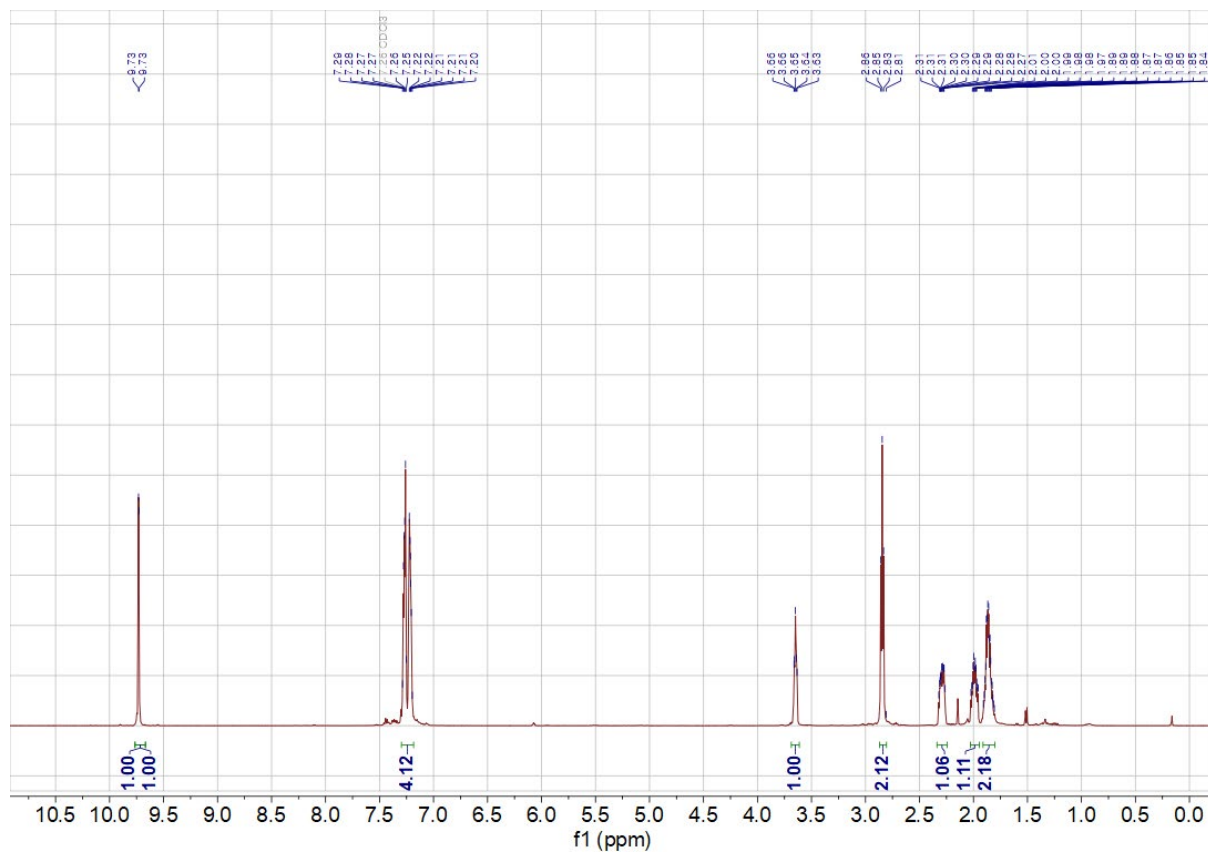


$^{13}\text{C}\{^1\text{H}\}$ NMR spectrum (126 MHz, CDCl_3) of **27-sub**

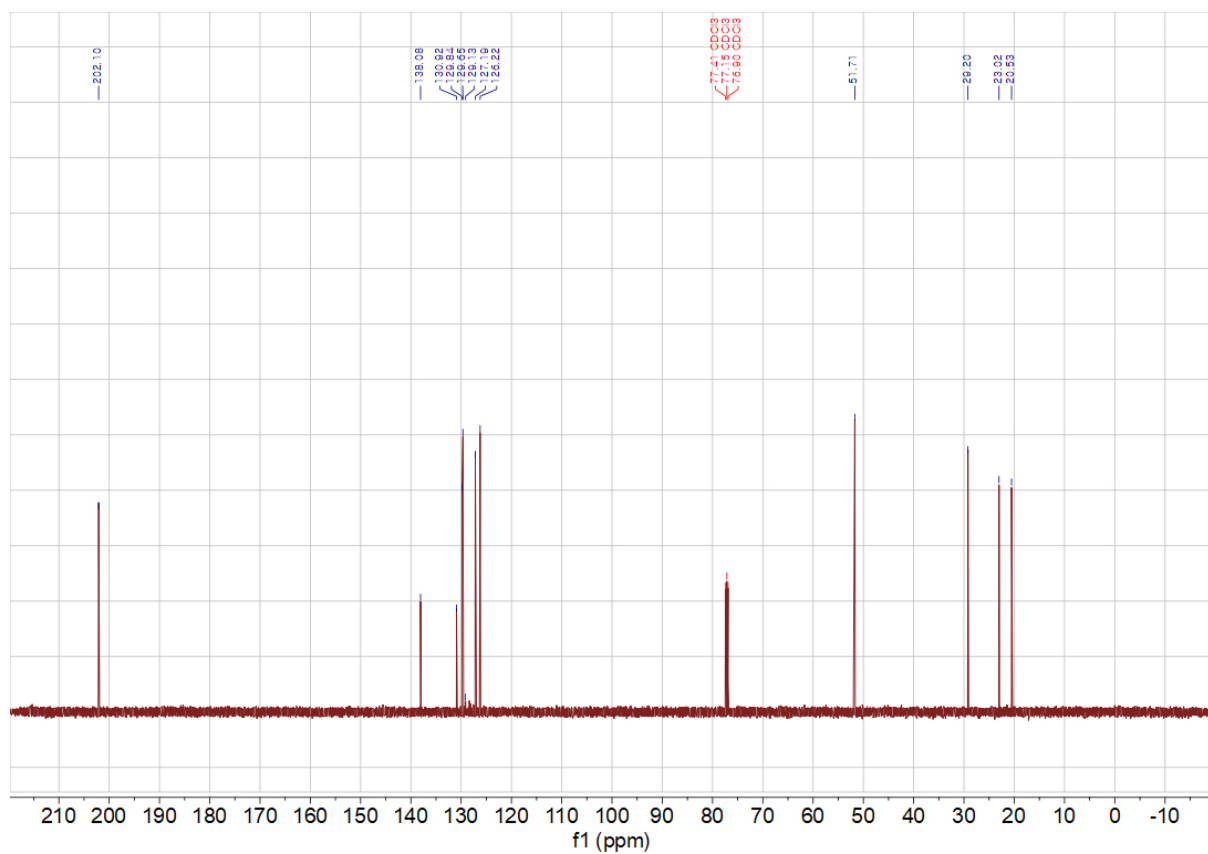


10.2. Products

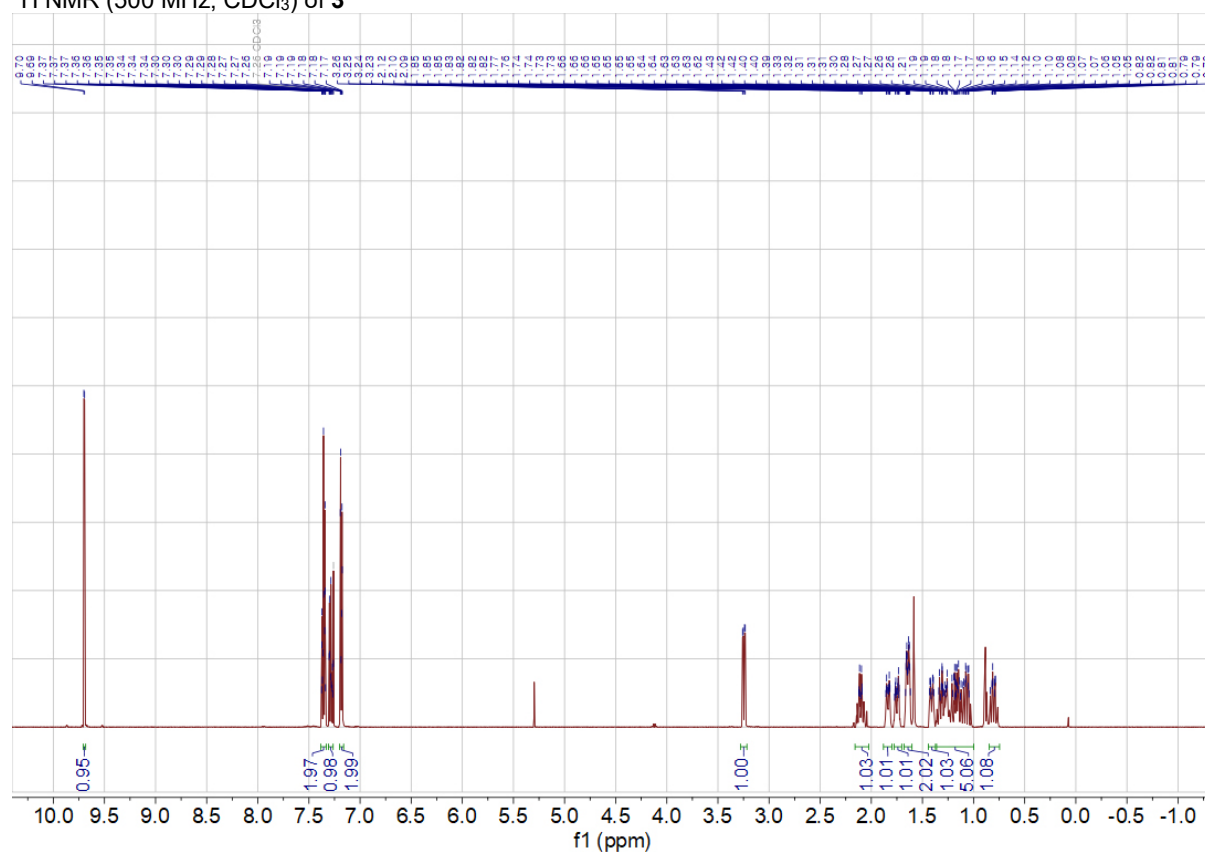
^1H NMR (500 MHz, CDCl_3) of **2**



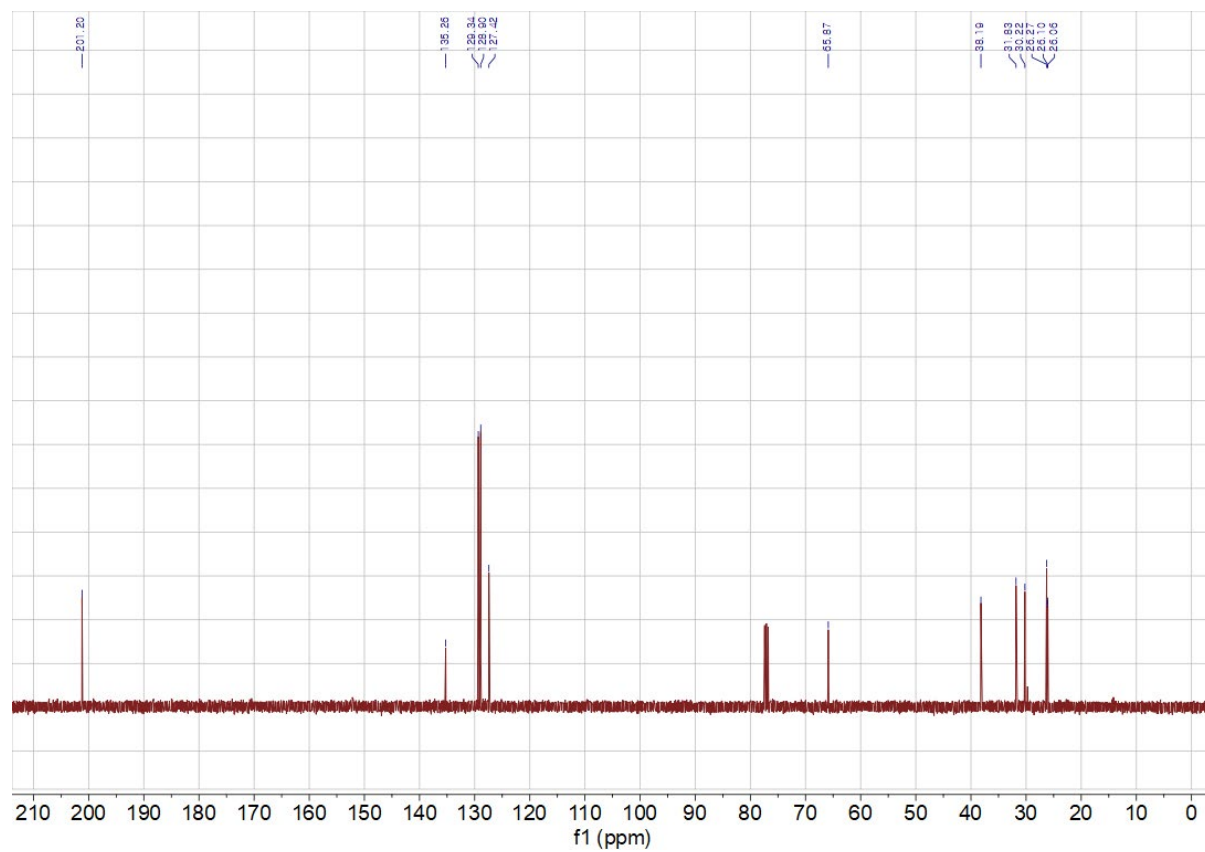
$^{13}\text{C}\{^1\text{H}\}$ NMR (126 MHz, CDCl_3) of **2**



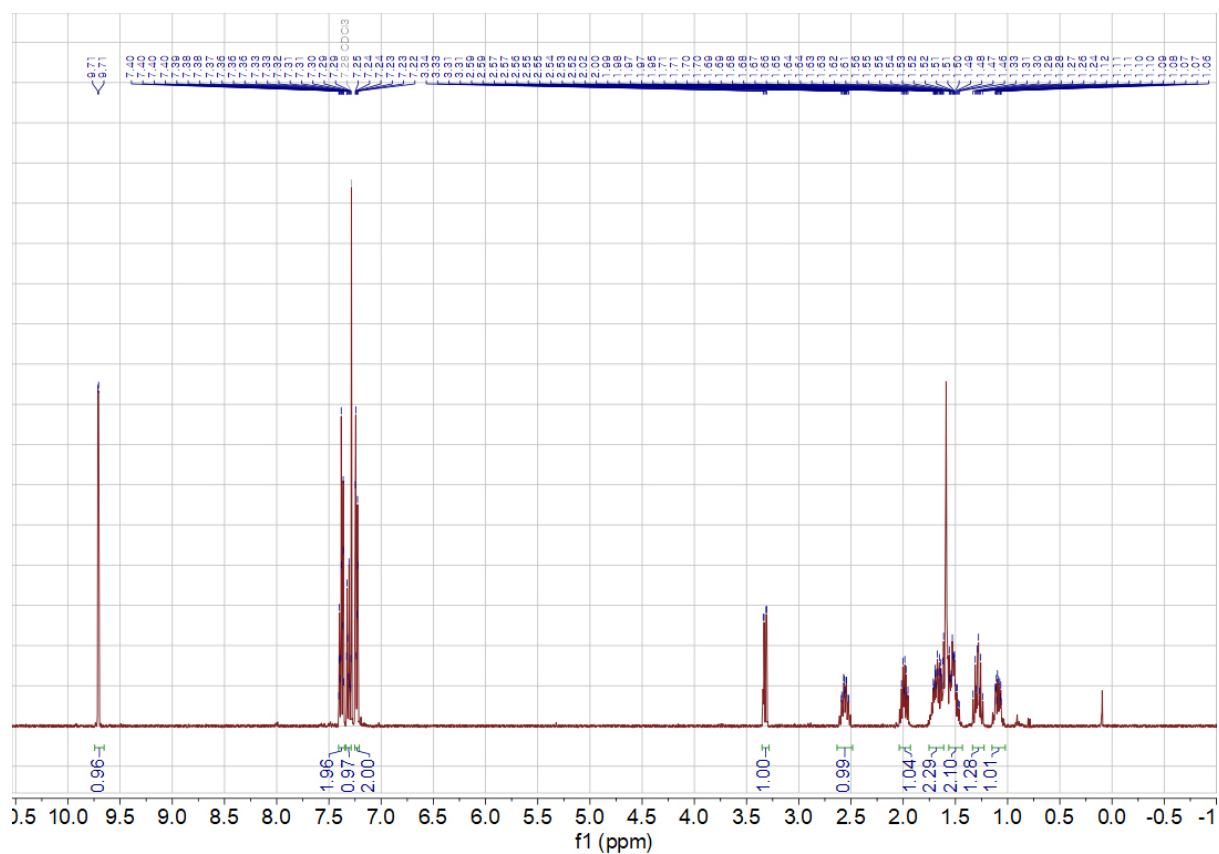
^1H NMR (500 MHz, CDCl_3) of **3**



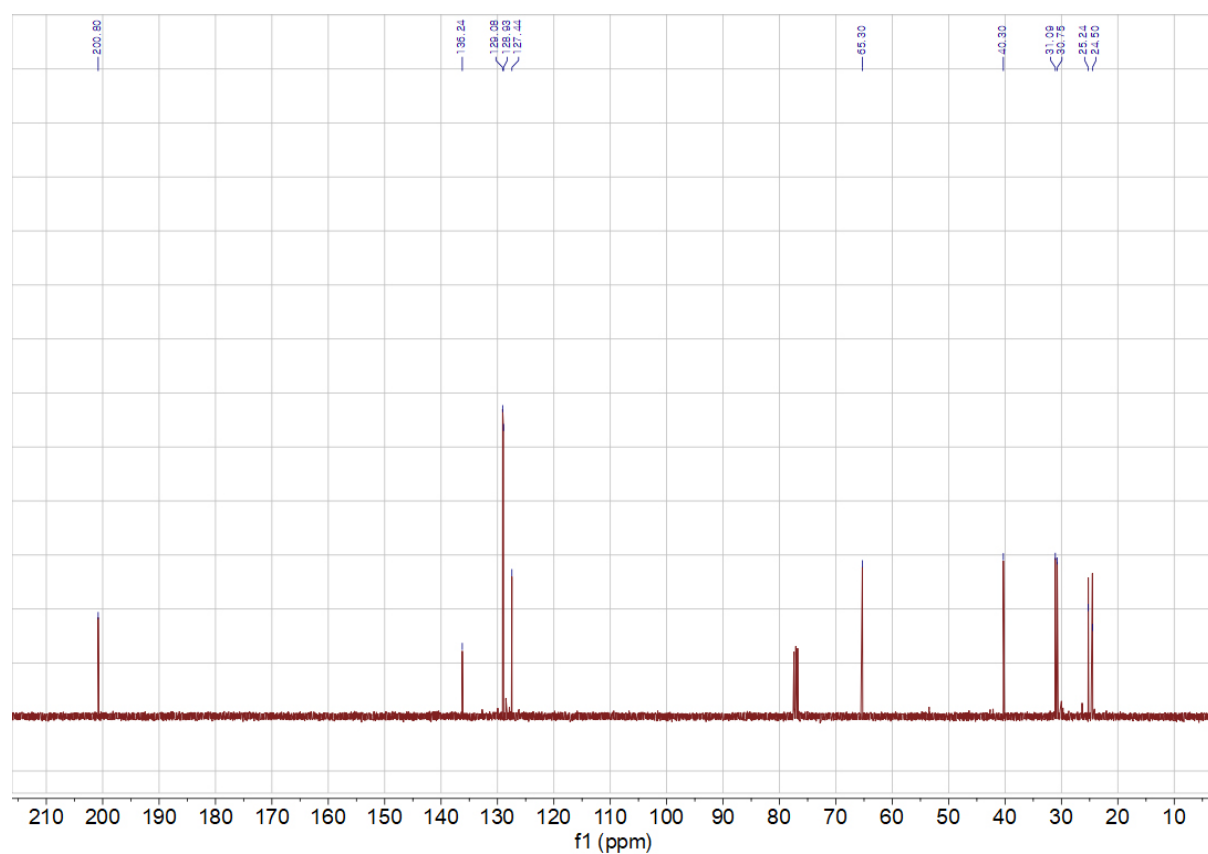
$^{13}\text{C}\{^1\text{H}\}$ NMR (101 MHz, CDCl_3) of **3**



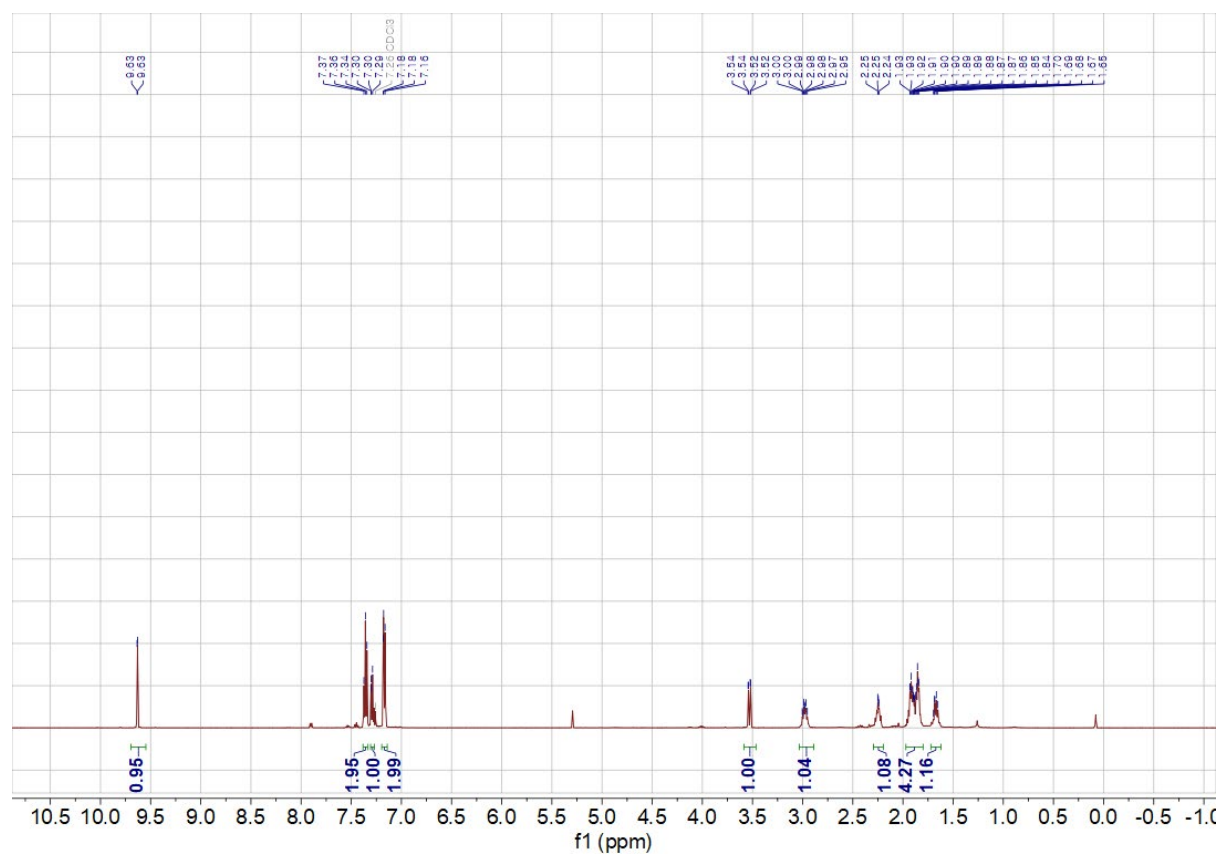
^1H NMR (400 MHz, CDCl_3) of **4**



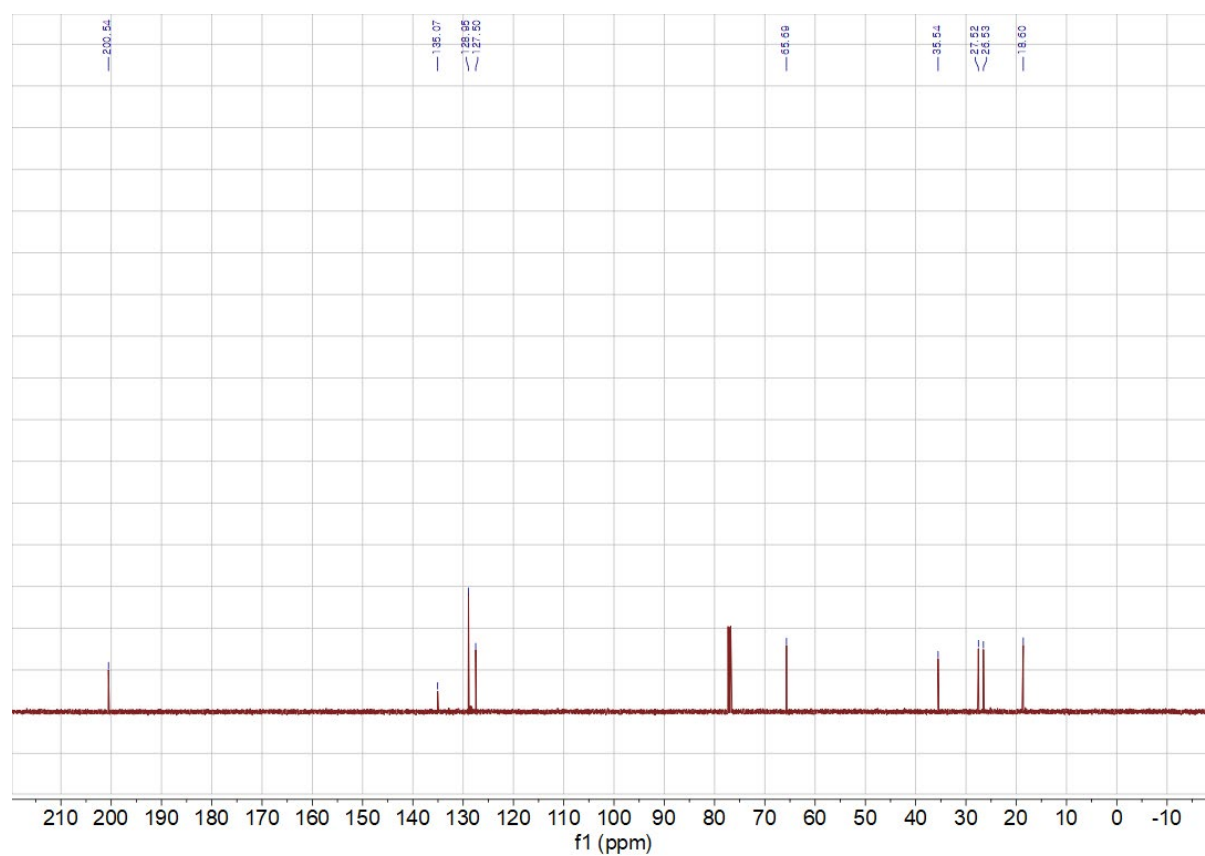
$^{13}\text{C}\{^1\text{H}\}$ NMR (101 MHz, CDCl_3) of **4**



^1H NMR (500 MHz, CDCl_3) of **5**

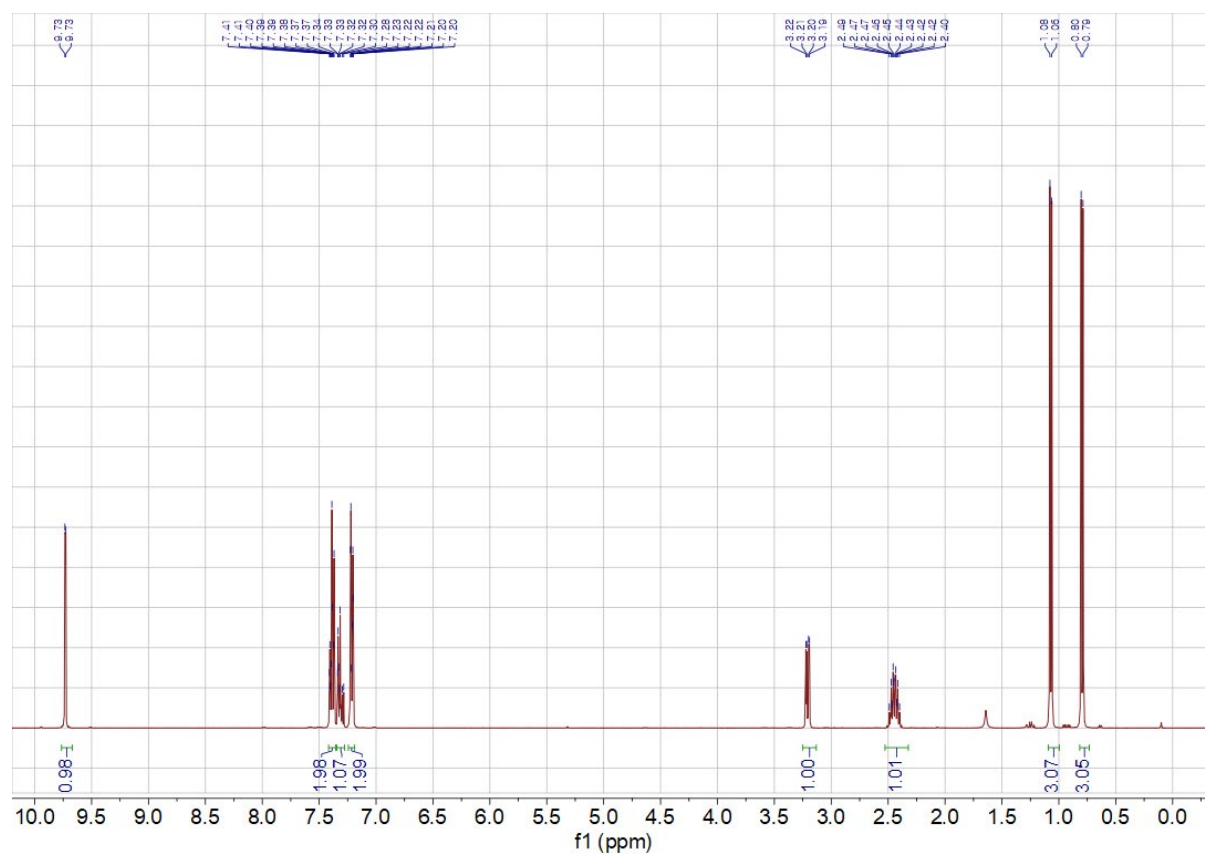


$^{13}\text{C}\{^1\text{H}\}$ NMR (126 MHz, CDCl_3) of **5**

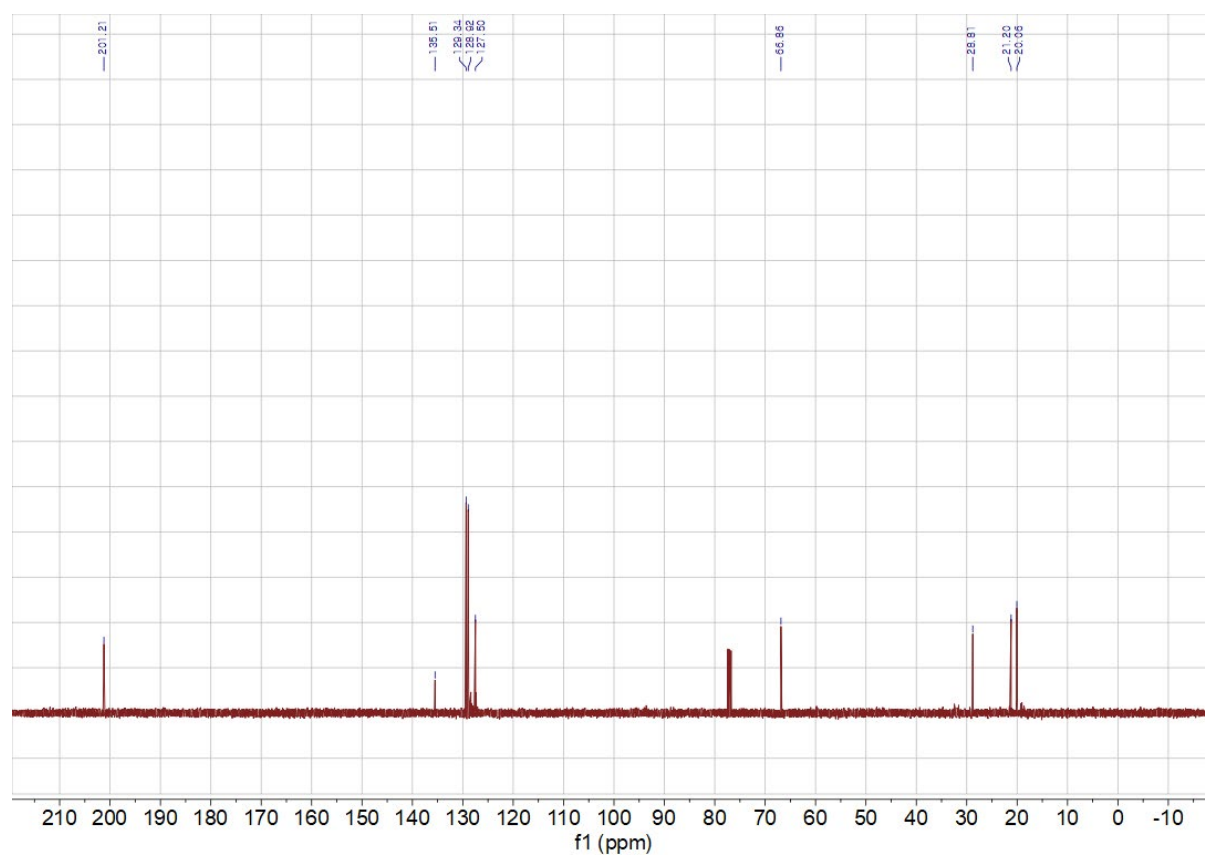


¹H NMR spectrum (400 MHz, CDCl₃) of 2,2,4,4-tetramethyl-5-oxotetrahydro-2H-pyran-3-ol. The spectrum shows peaks at 9.7 (s, 1H), 7.2-7.5 (m, 4H), 5.2 (s, 1H), 3.5 (s, 1H), 2.1 (s, 1H), 1.9 (s, 1H), 1.5 (s, 1H), and 1.0 (s, 3H). Integration values are 0.97, 2.02, 1.08, 2.04, 1.00, 1.01, 1.22, and 3.34. Chemical shift ranges are indicated by brackets at the top.

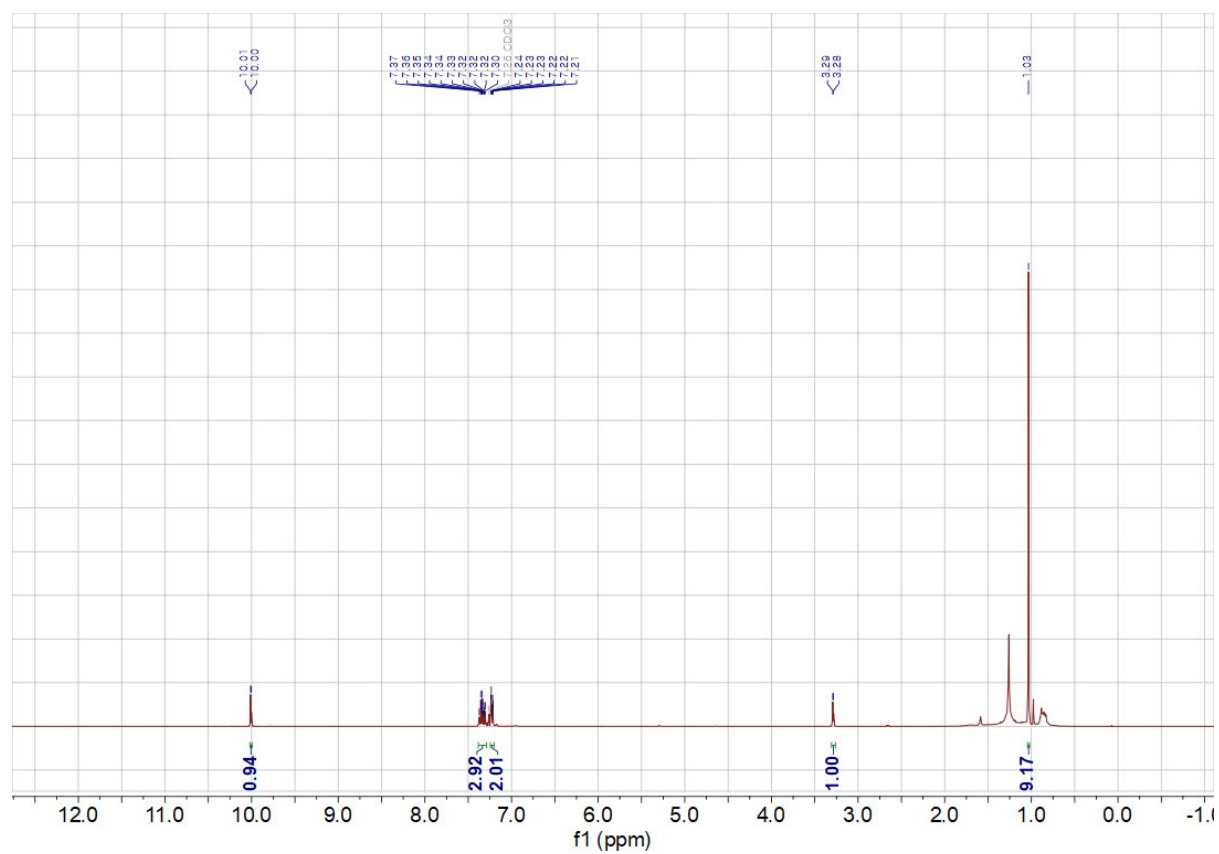
^1H NMR (400 MHz, CDCl_3) of **7**



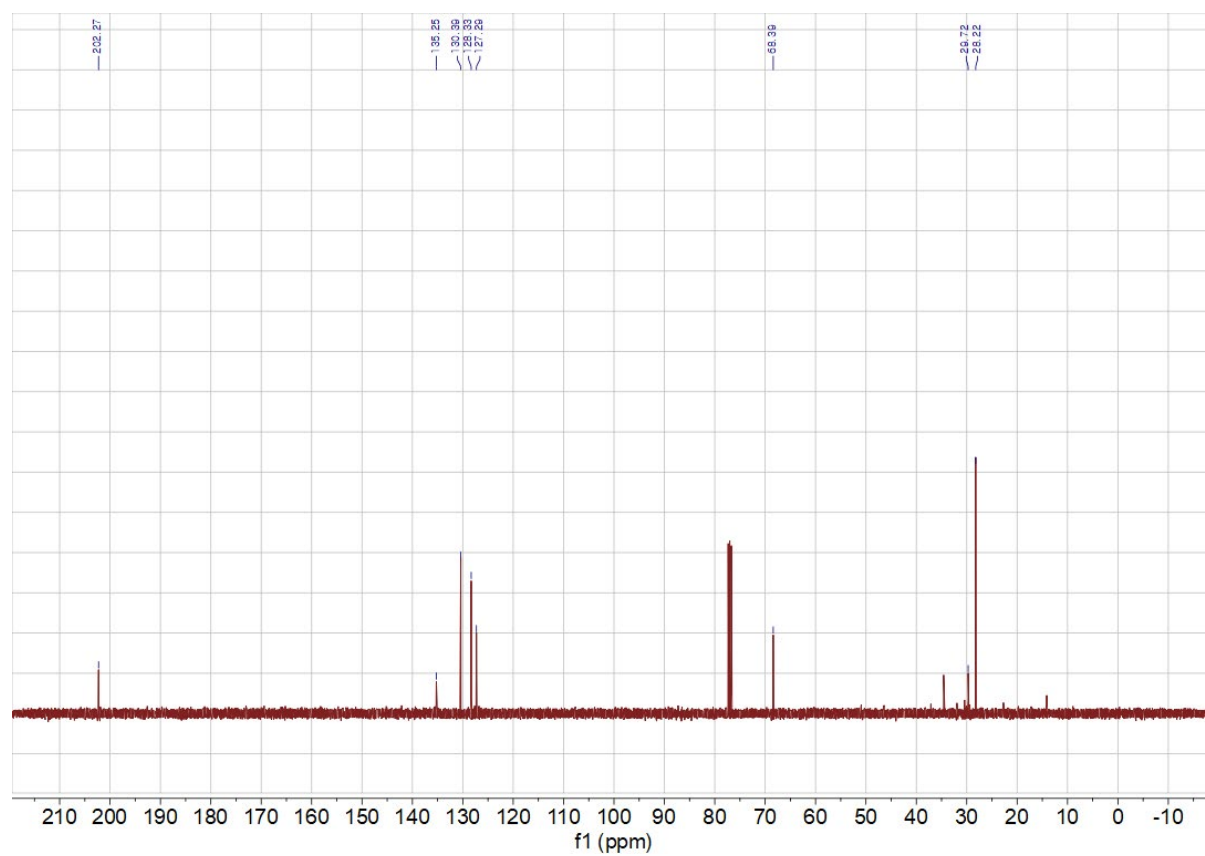
$^{13}\text{C}\{^1\text{H}\}$ NMR (101 MHz, CDCl_3) of **7**



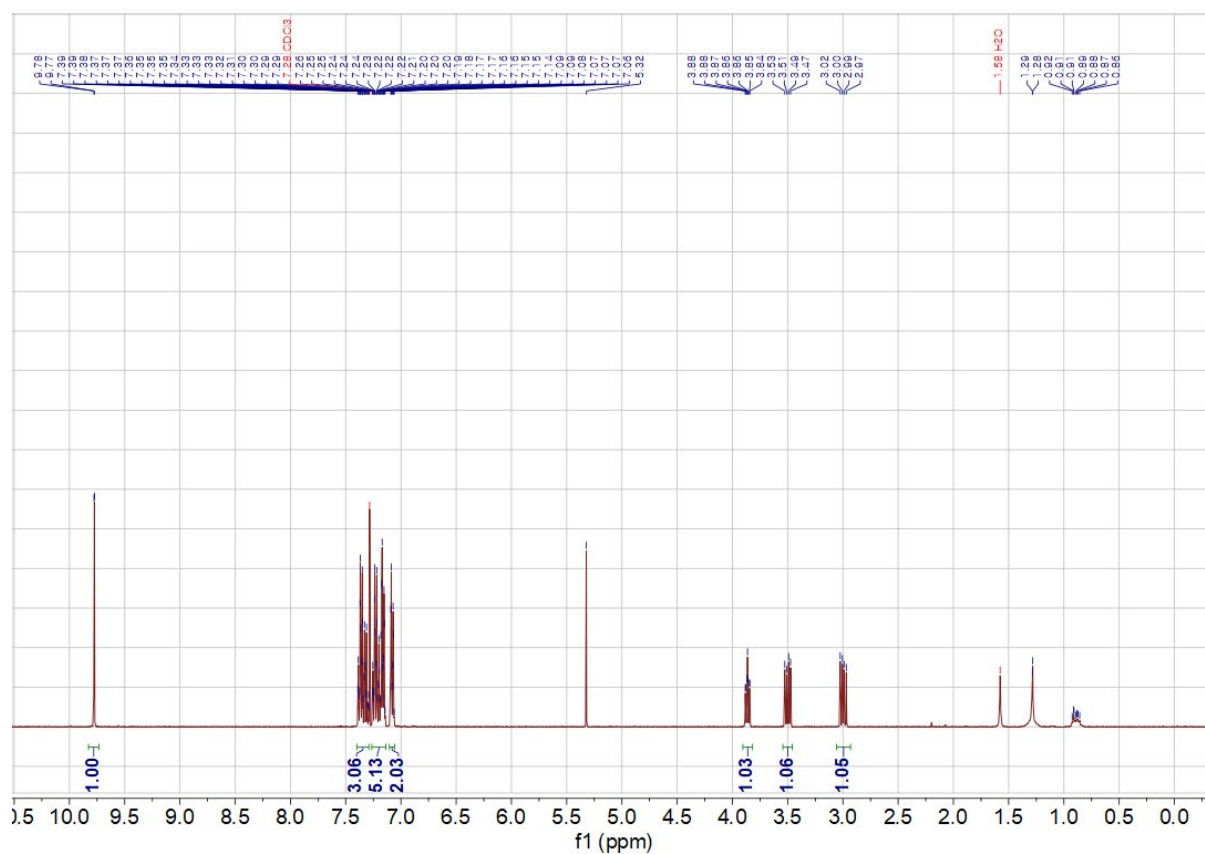
^1H NMR (400 MHz, CDCl_3) of **8**



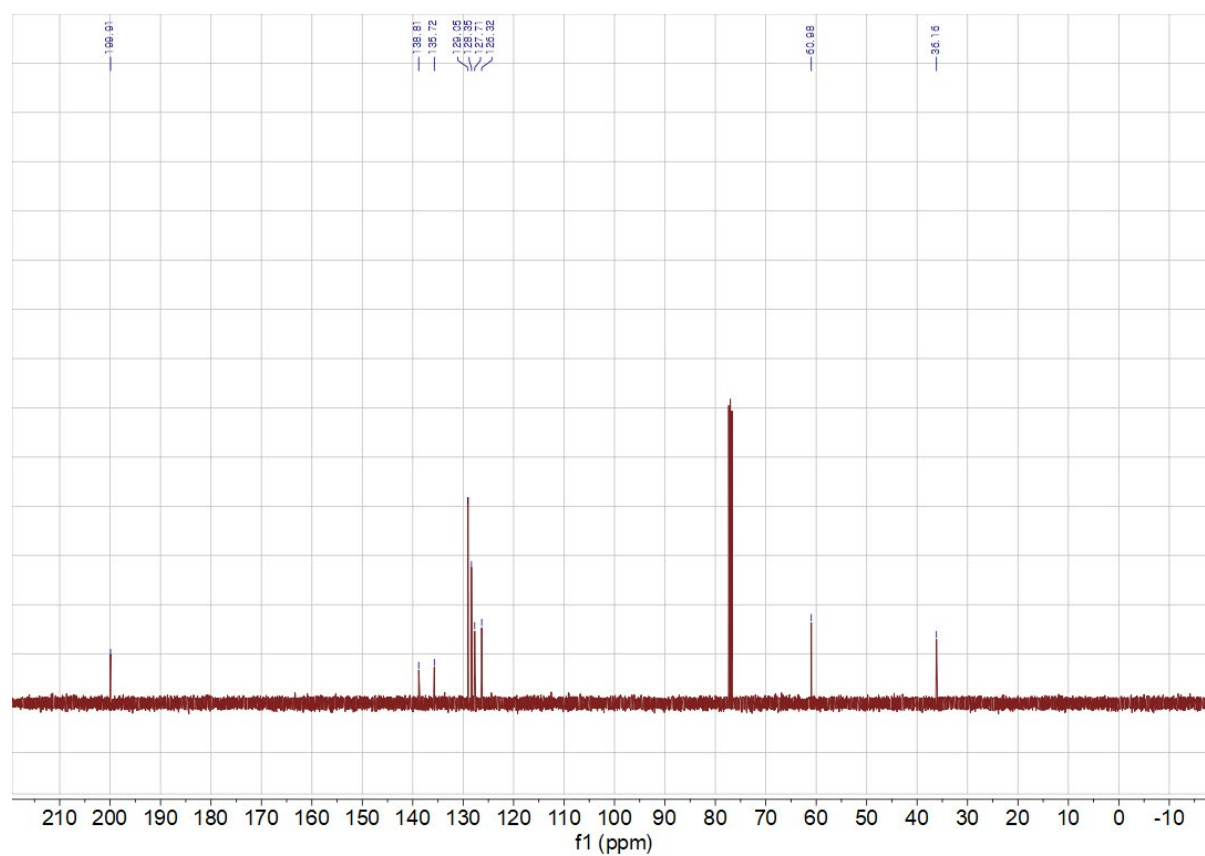
$^{13}\text{C}\{^1\text{H}\}$ NMR (101 MHz, CDCl_3) of **8**



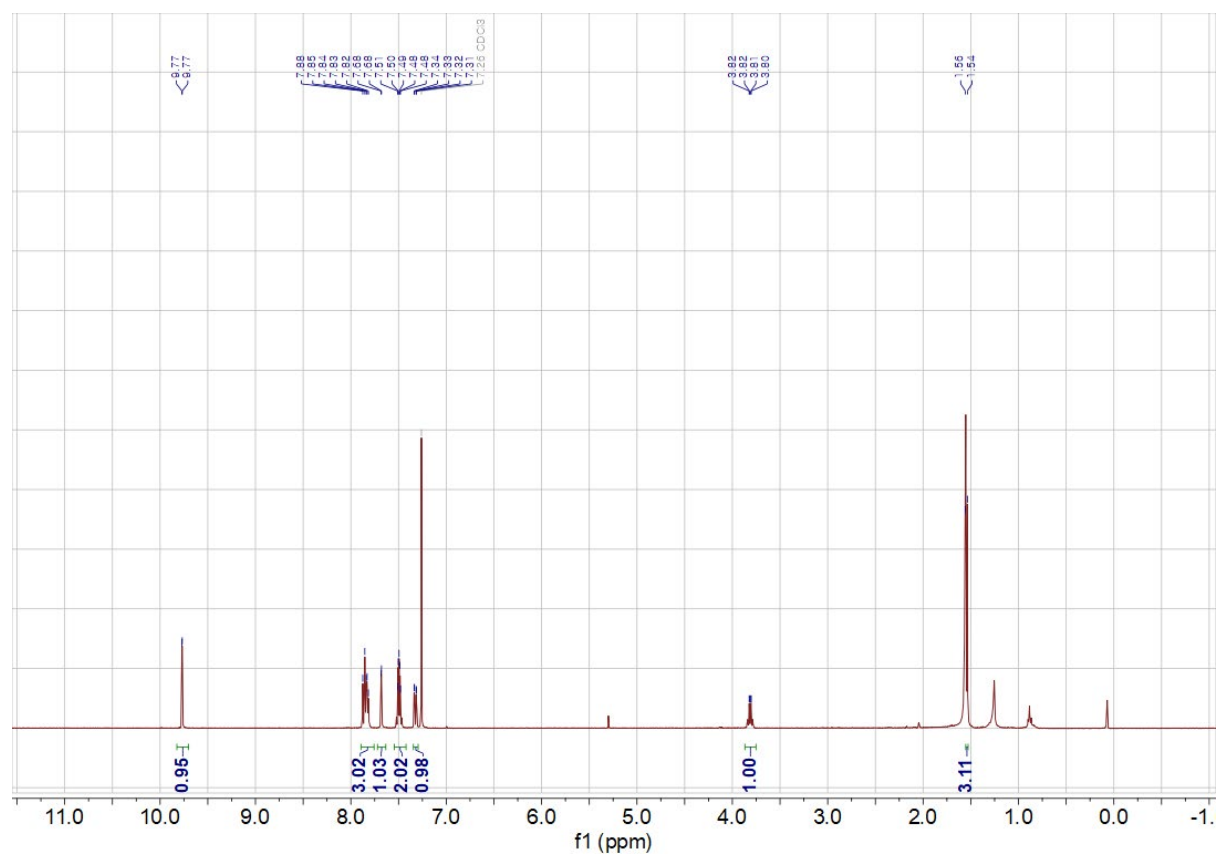
^1H NMR (400 MHz, CDCl_3) of **9**



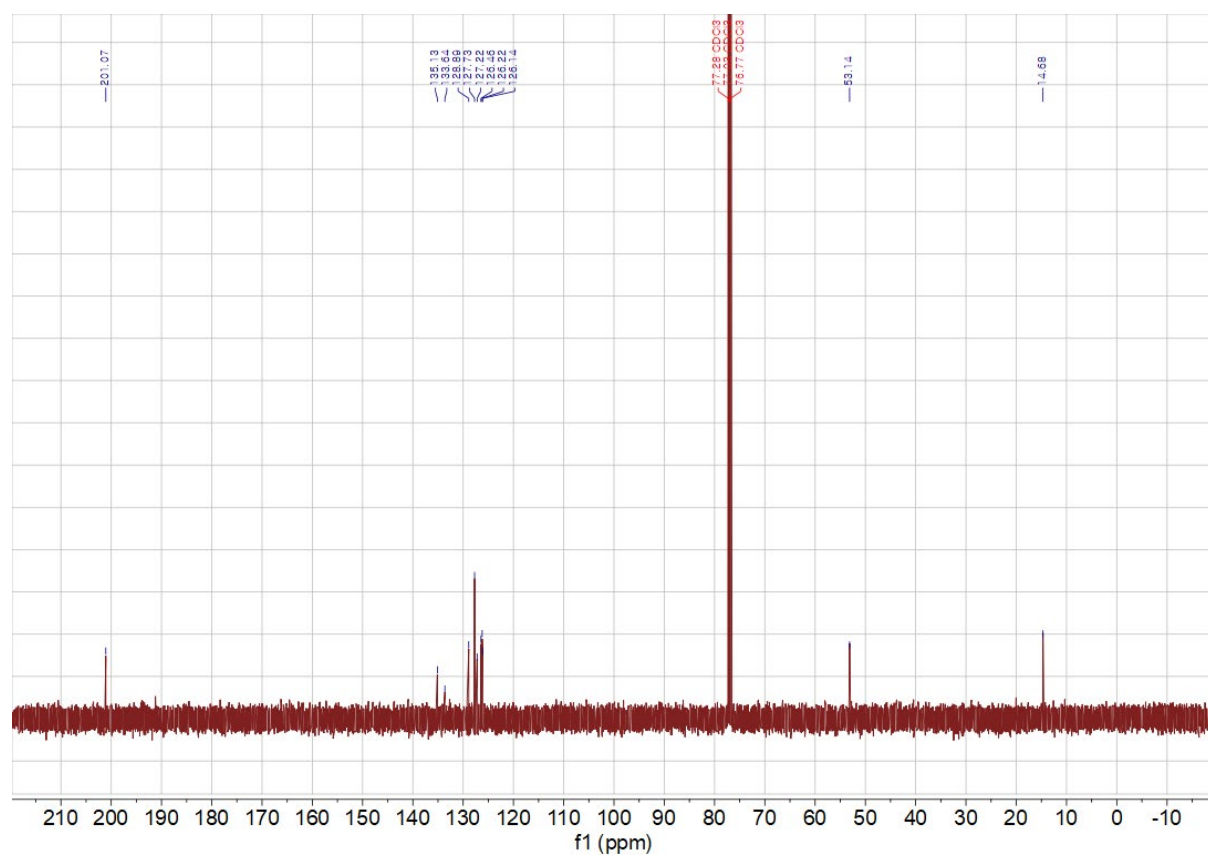
$^{13}\text{C}\{^1\text{H}\}$ NMR (101 MHz, CDCl_3) of **9**



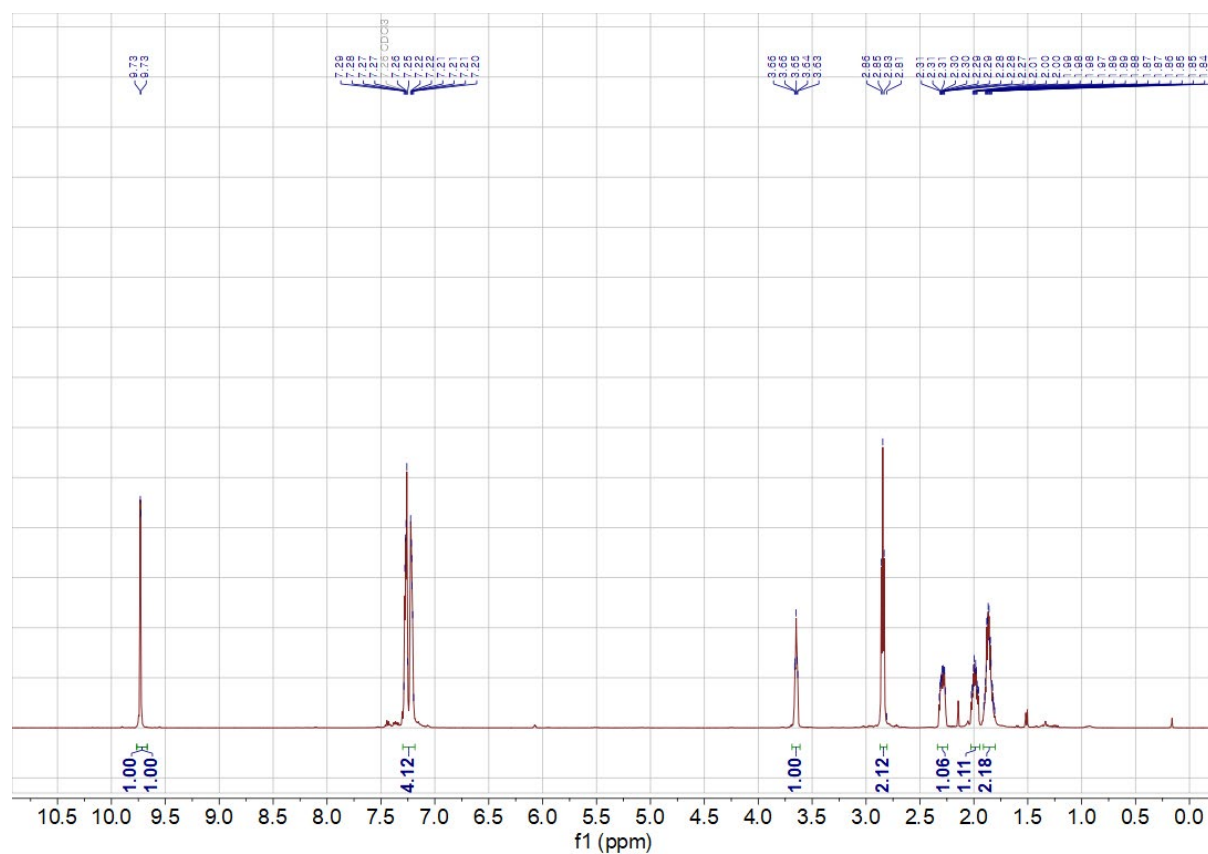
^1H NMR (400 MHz, CDCl_3) of **10**



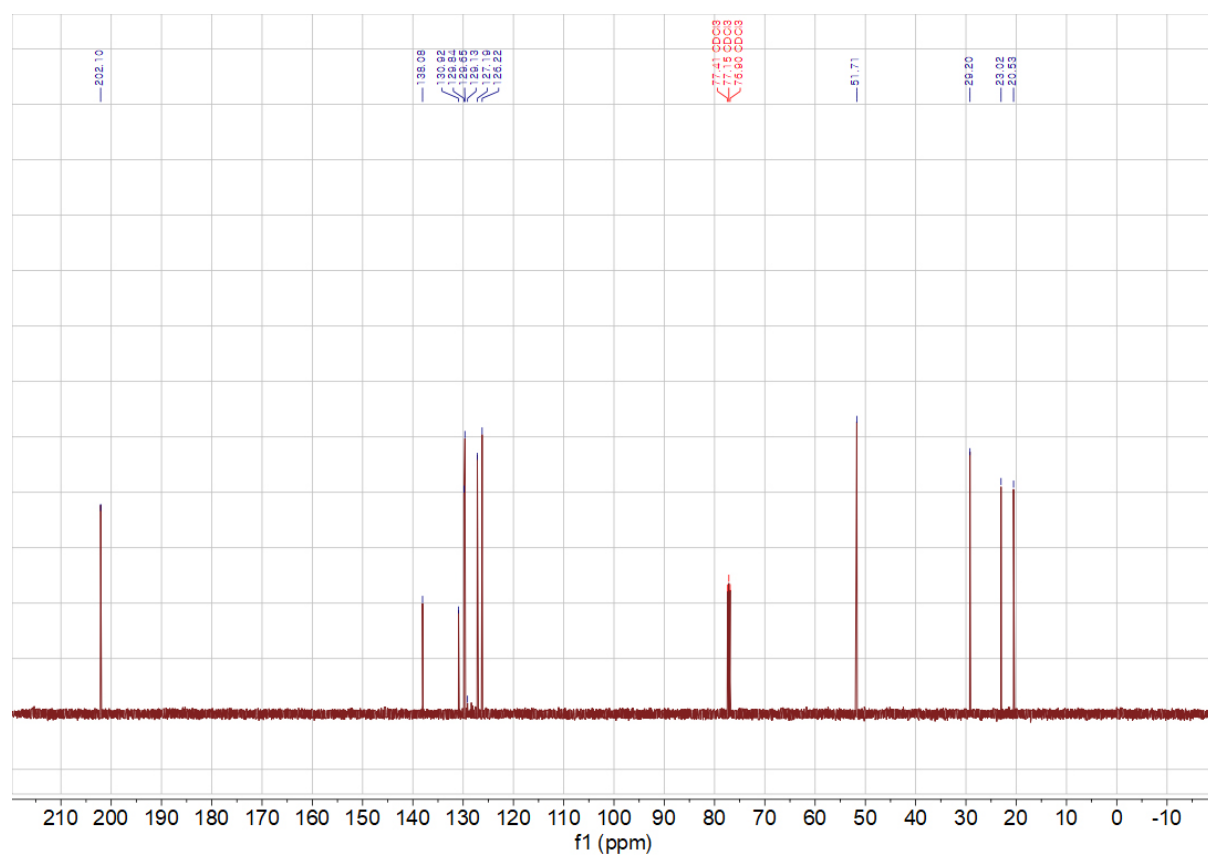
$^{13}\text{C}\{^1\text{H}\}$ NMR (126 MHz, CDCl_3) of **10**



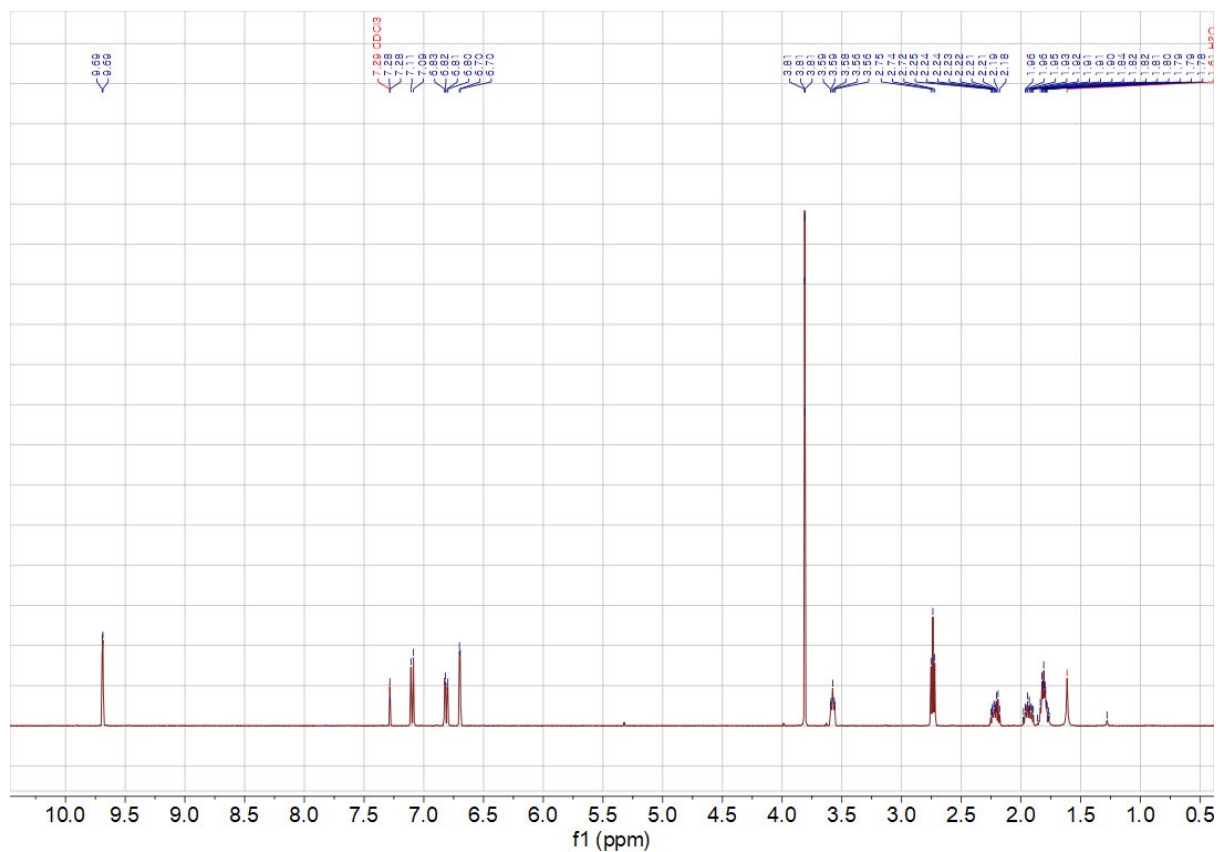
^1H NMR (500 MHz, CDCl_3) of **11**



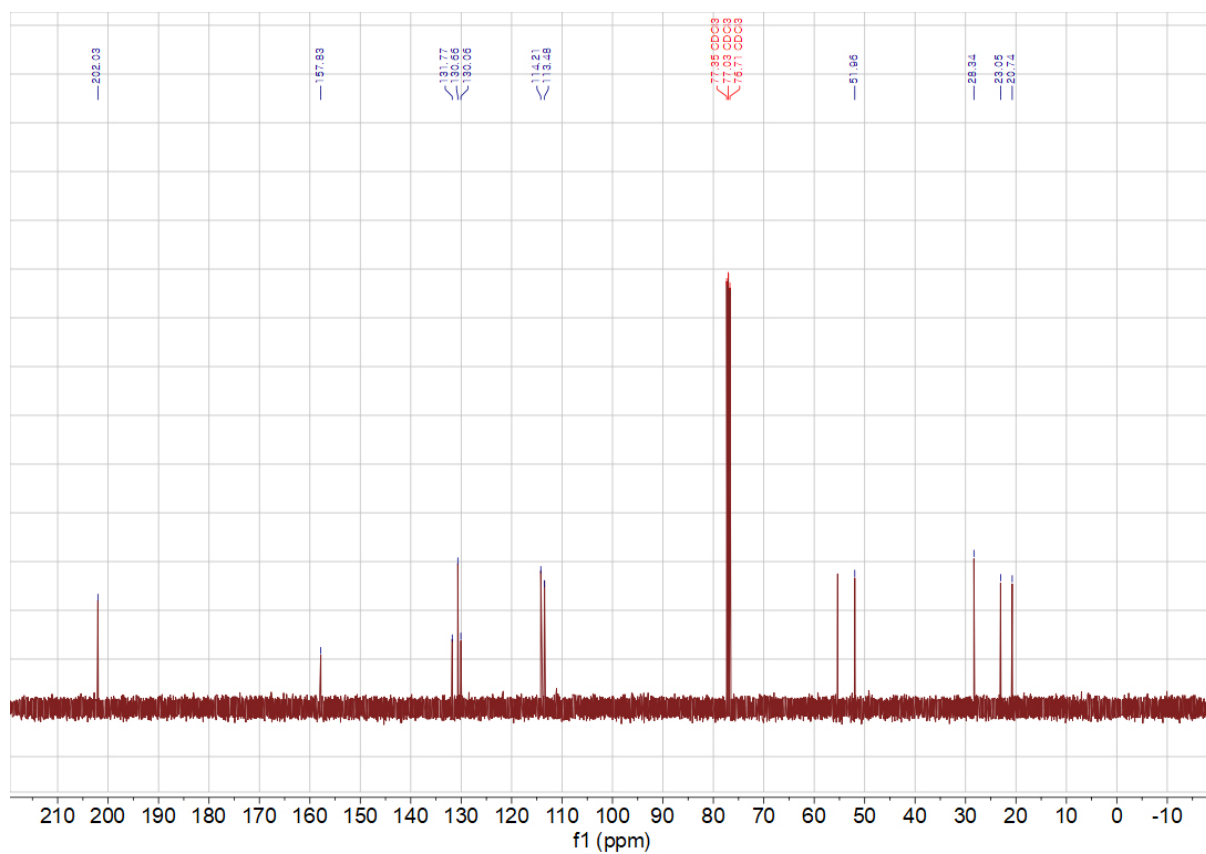
$^{13}\text{C}\{^1\text{H}\}$ NMR (126 MHz, CDCl_3) of **11**



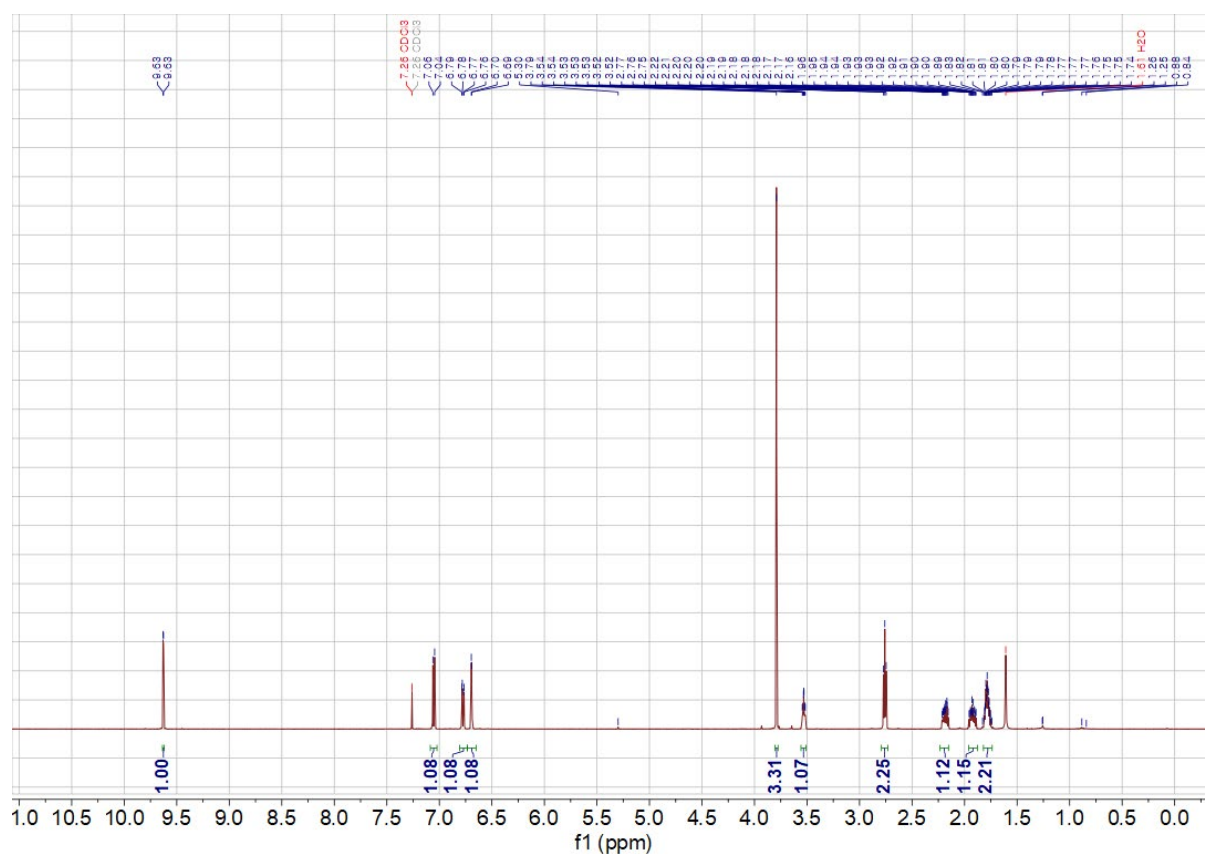
^1H NMR (400 MHz, CDCl_3) of **12**



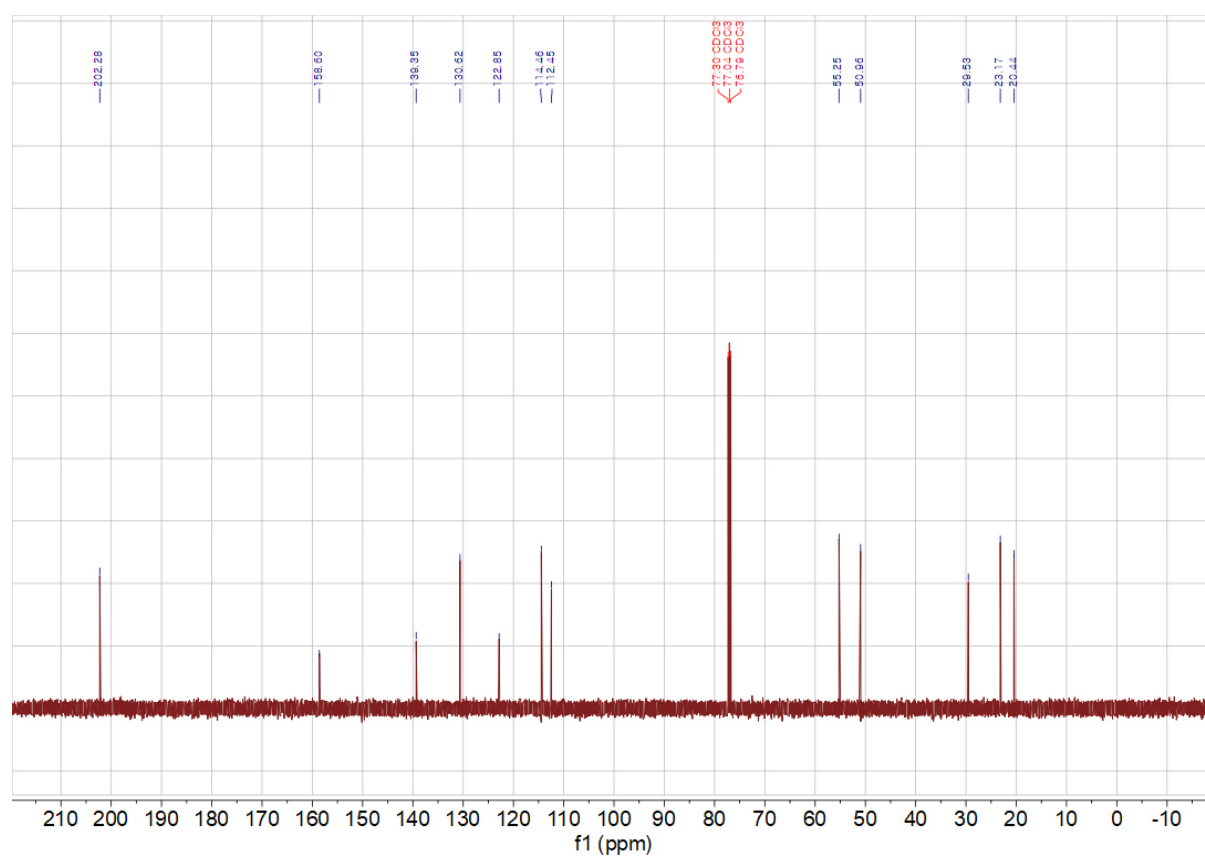
$^{13}\text{C}\{^1\text{H}\}$ NMR (101 MHz, CDCl_3) of **12**



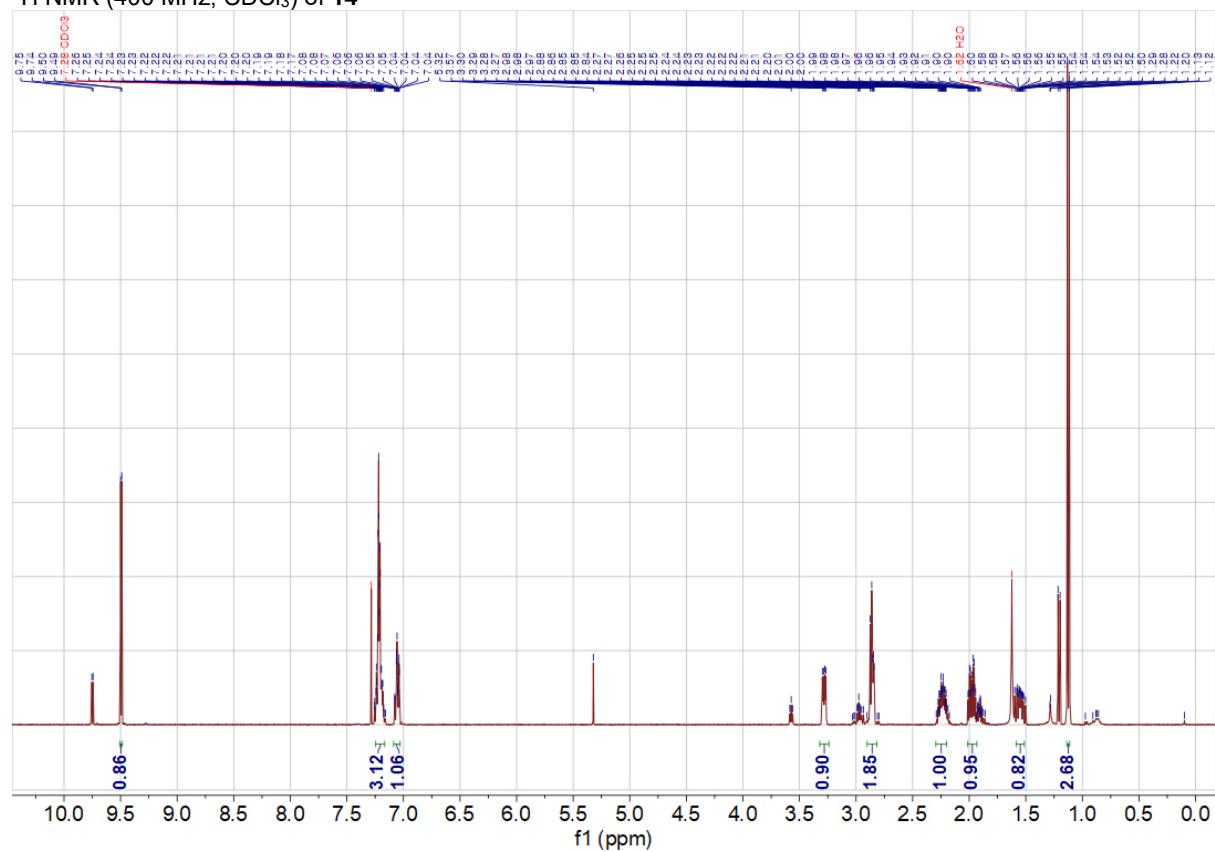
^1H NMR (500 MHz, CDCl_3) of **13**



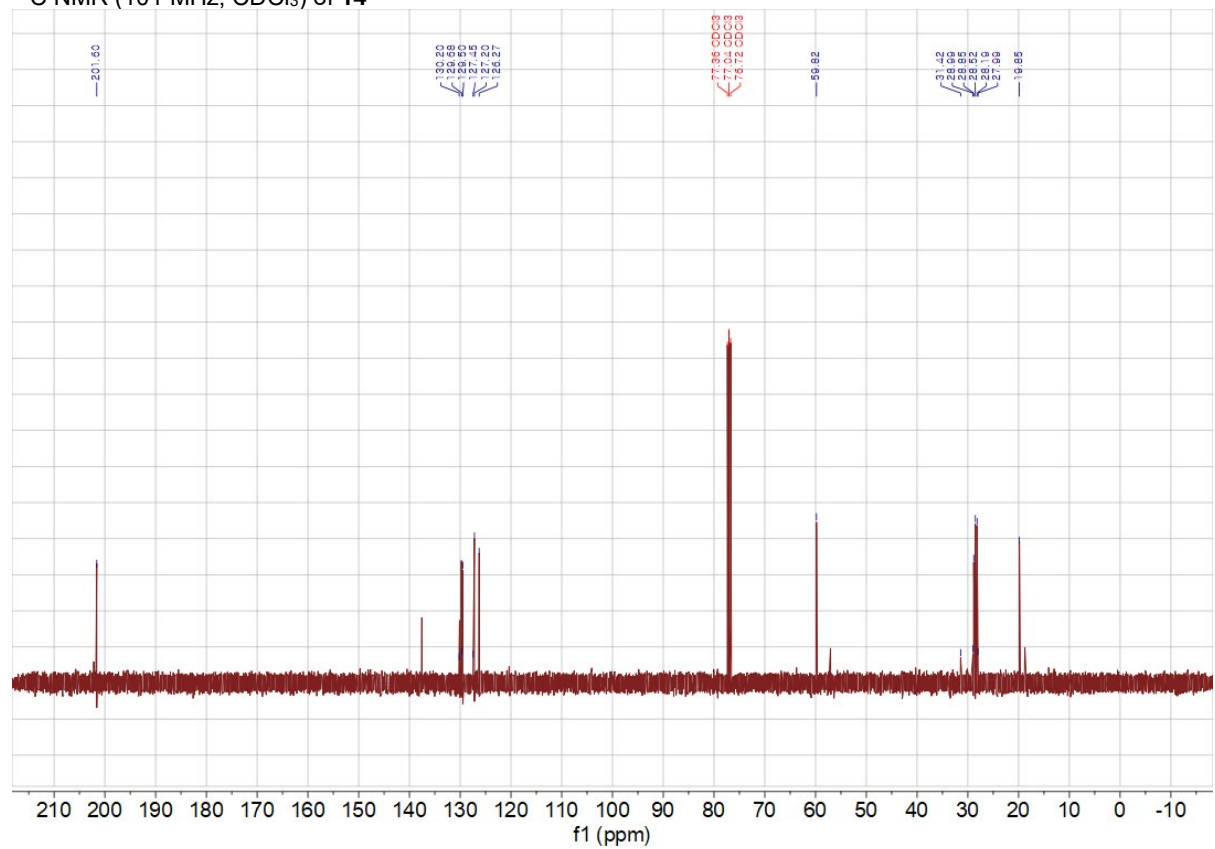
$^{13}\text{C}\{^1\text{H}\}$ NMR (126 MHz, CDCl_3) of **13**



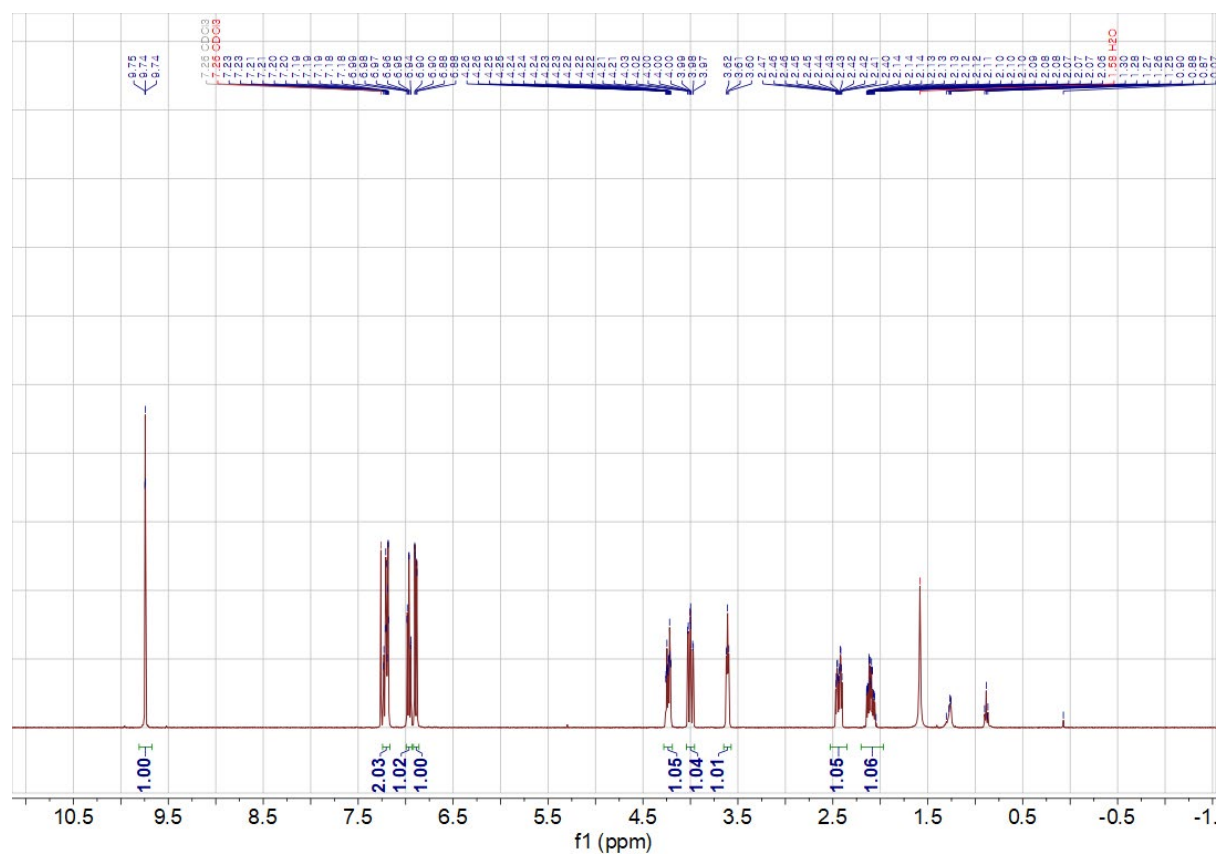
^1H NMR (400 MHz, CDCl_3) of **14**



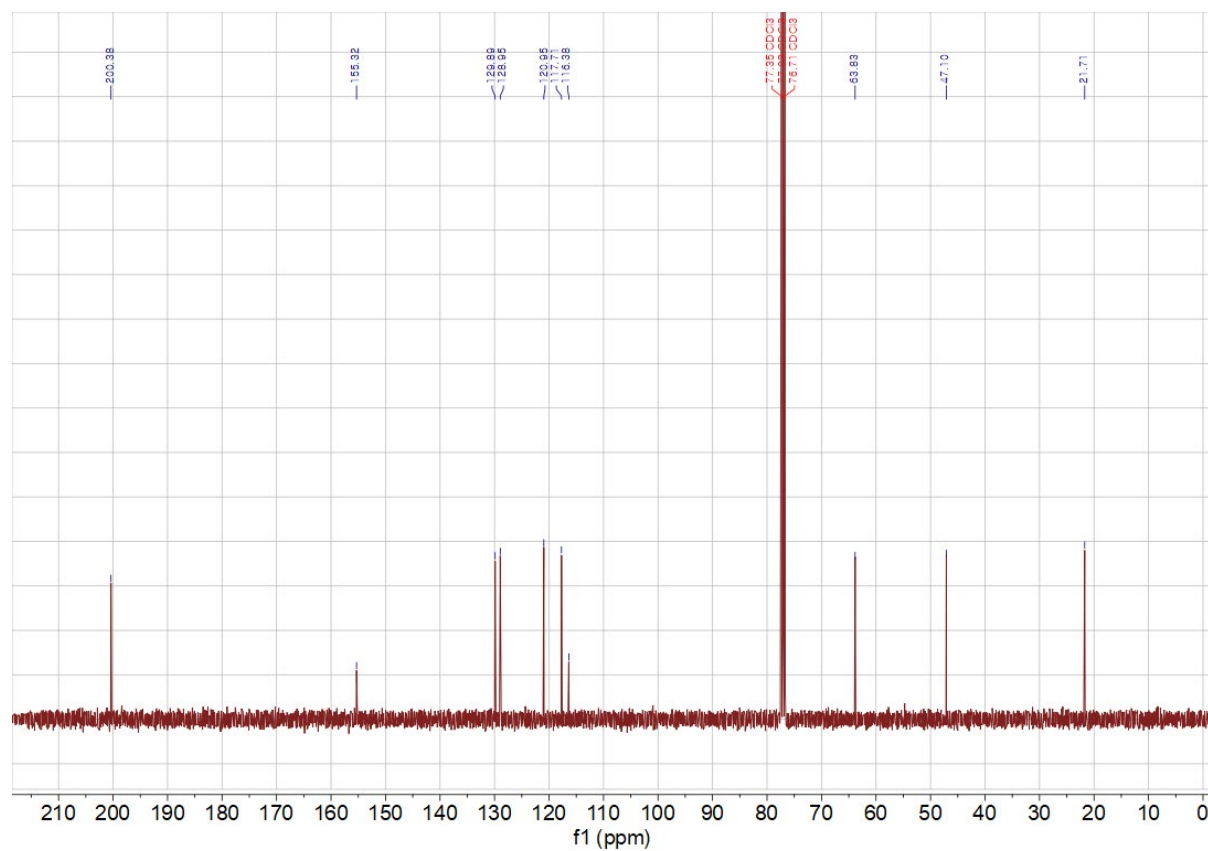
^{13}C NMR (101 MHz, CDCl_3) of **14**



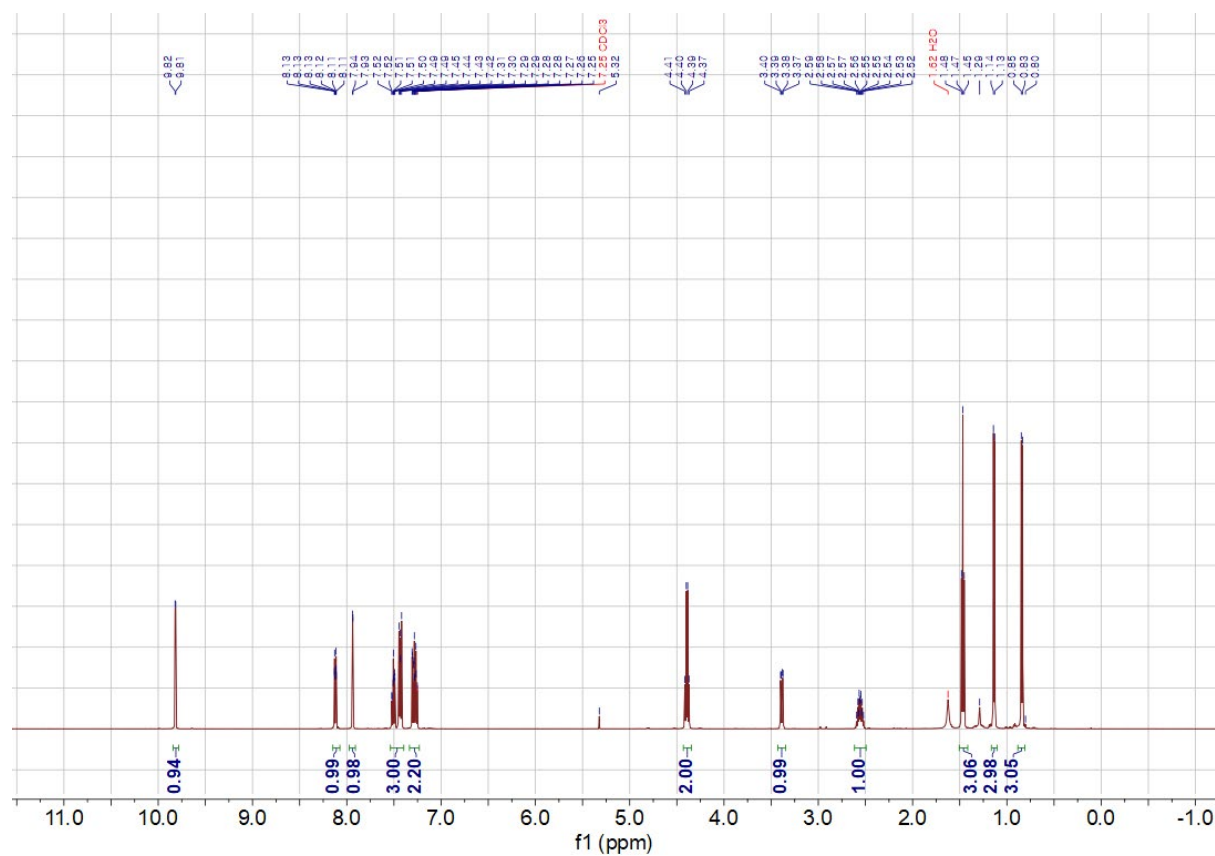
^1H NMR (400 MHz, CDCl_3) of **15**



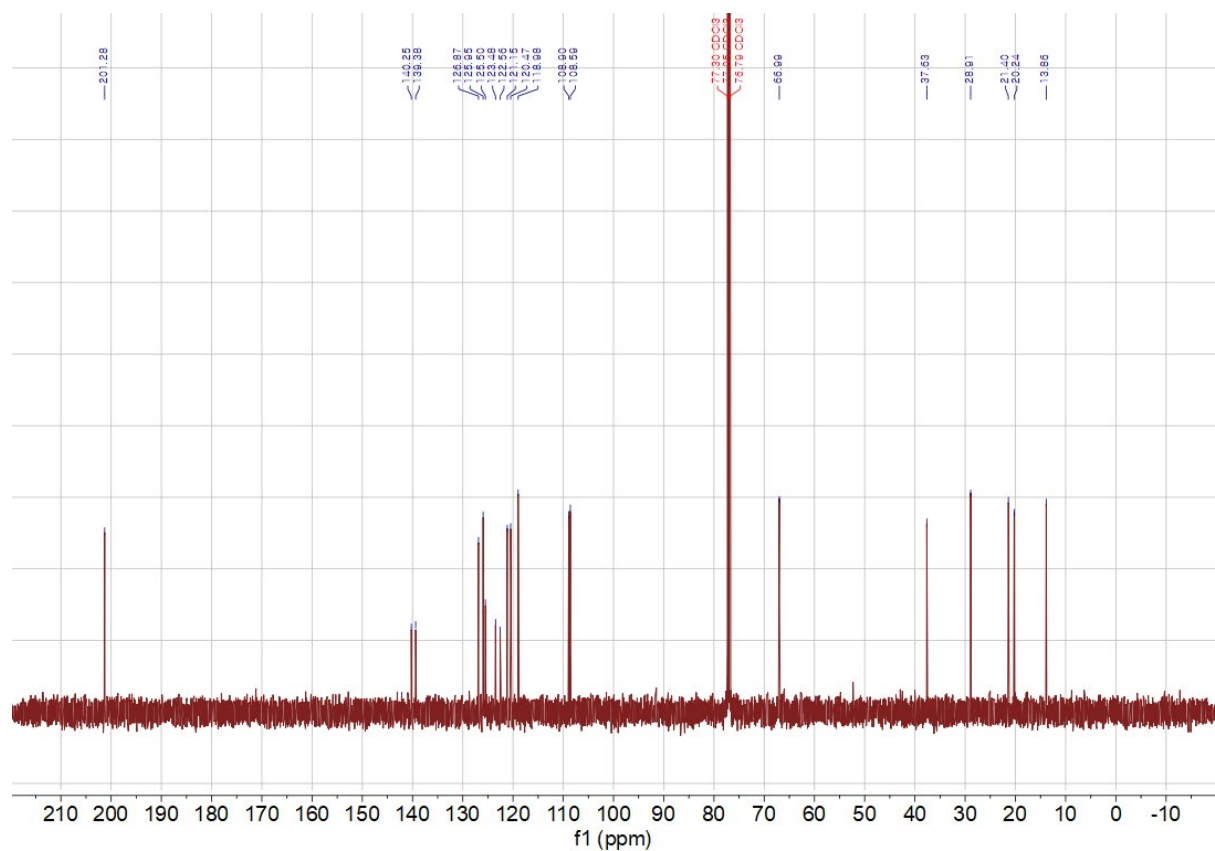
$^{13}\text{C}\{^1\text{H}\}$ NMR (101 MHz, CDCl_3) of **15**



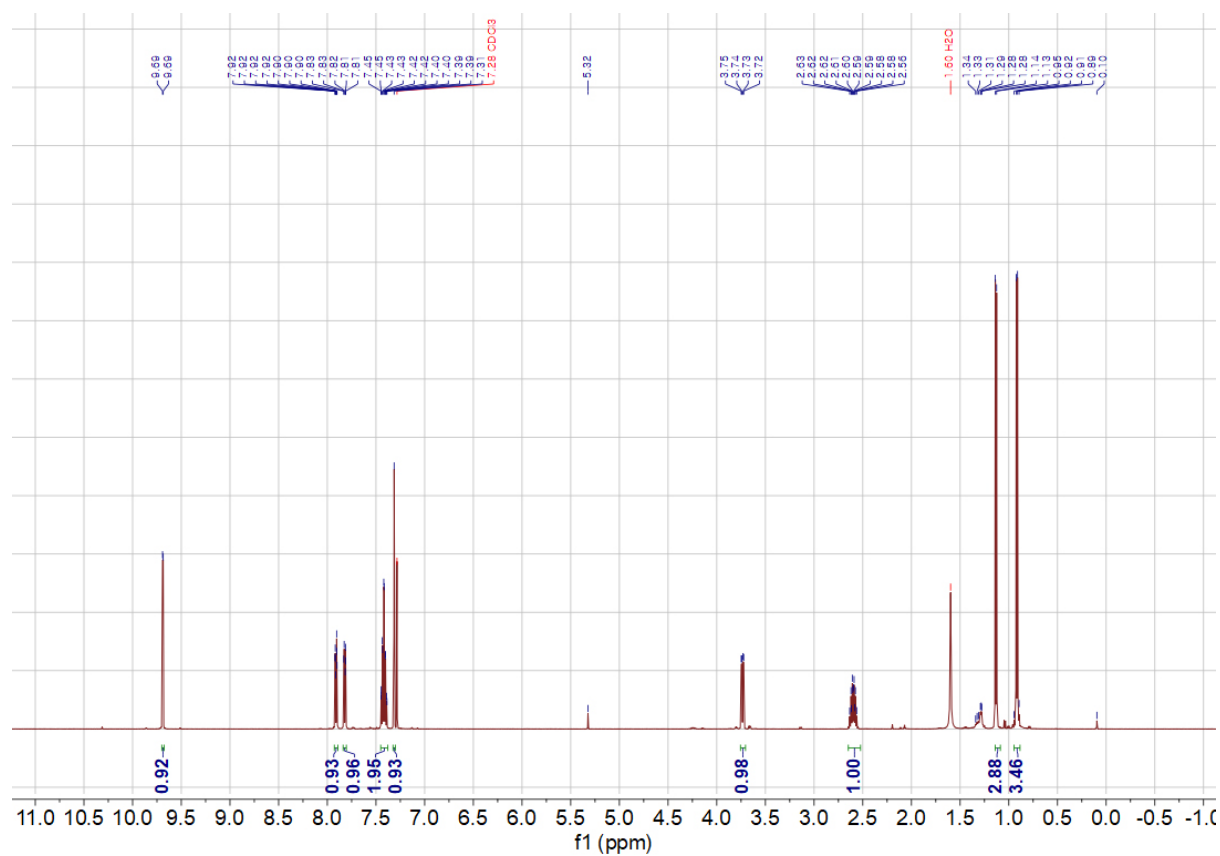
^1H NMR (500 MHz, CDCl_3) of **16**



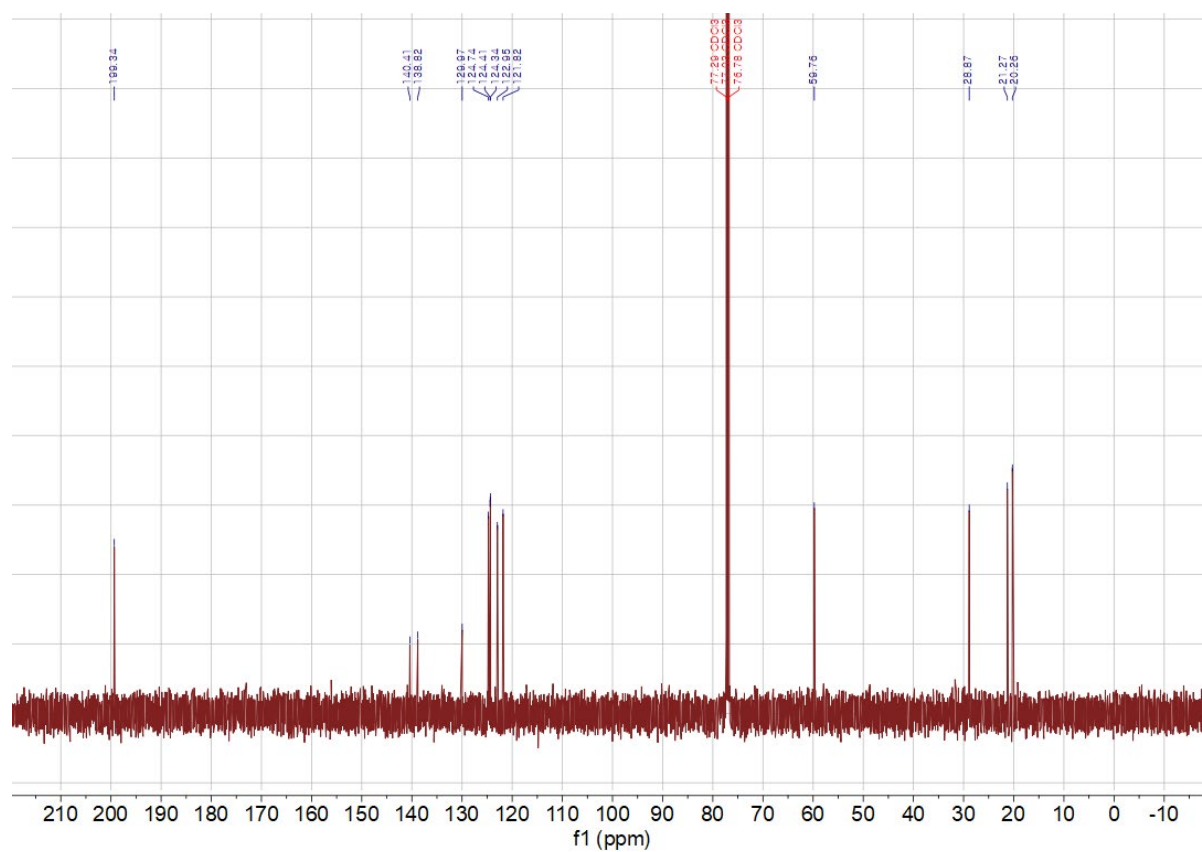
$^{13}\text{C}\{^1\text{H}\}$ NMR (126 MHz, CDCl_3) of **16**



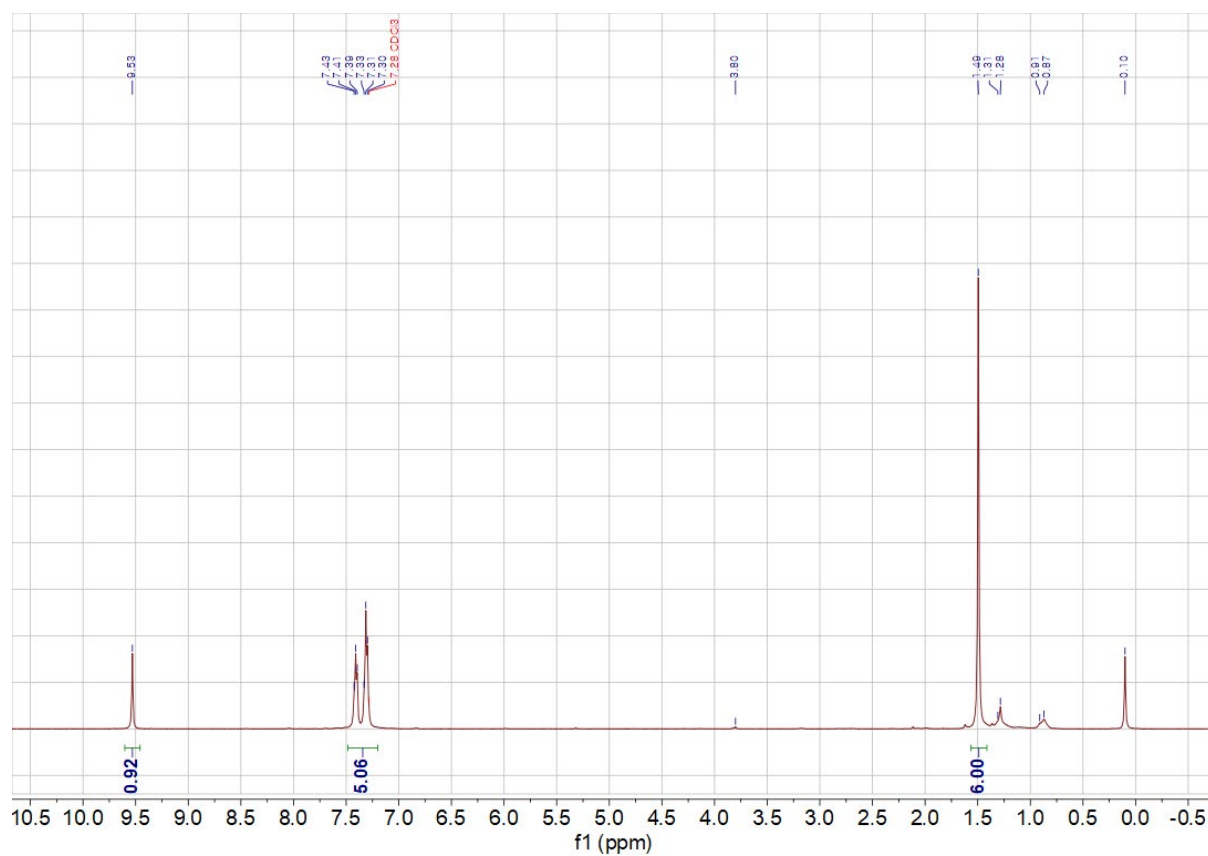
^1H NMR (500 MHz, CDCl_3) of **17**



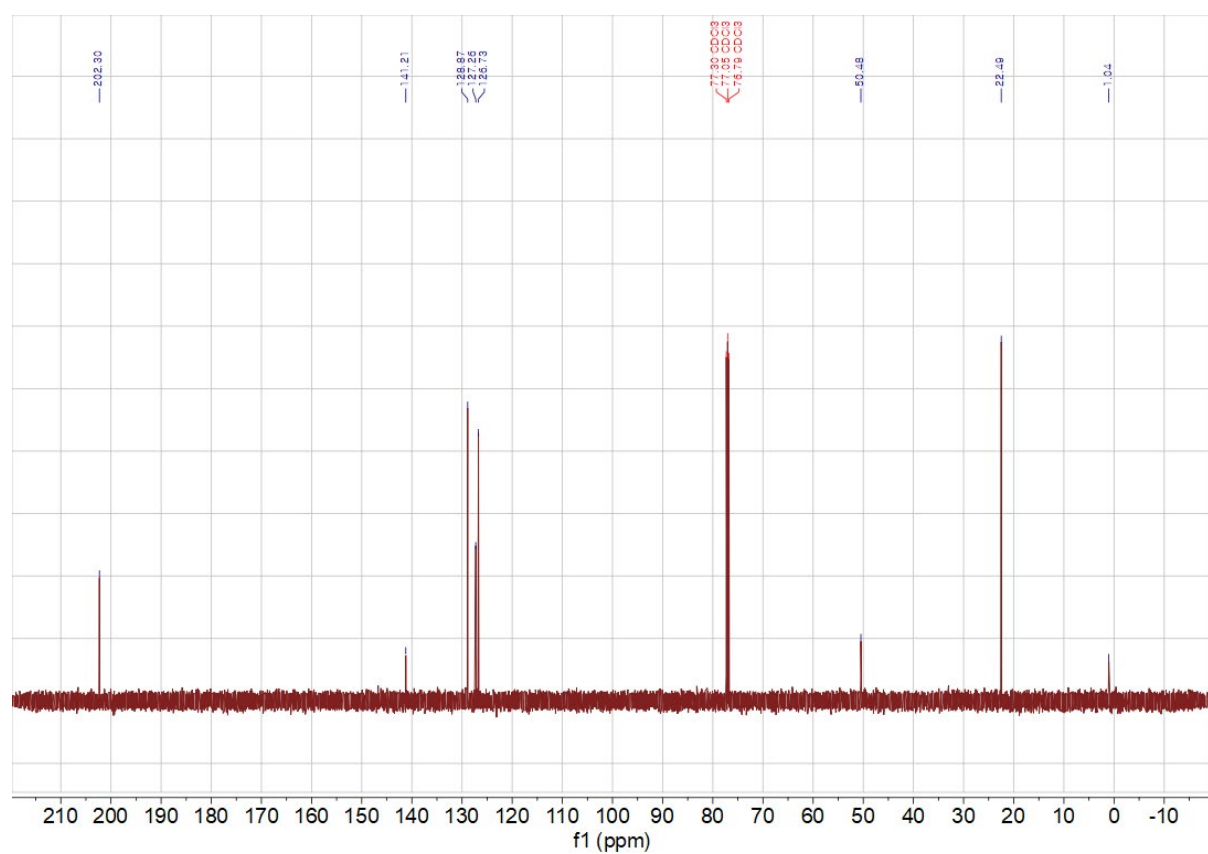
$^{13}\text{C}\{^1\text{H}\}$ NMR (126 MHz, CDCl_3) of **17**



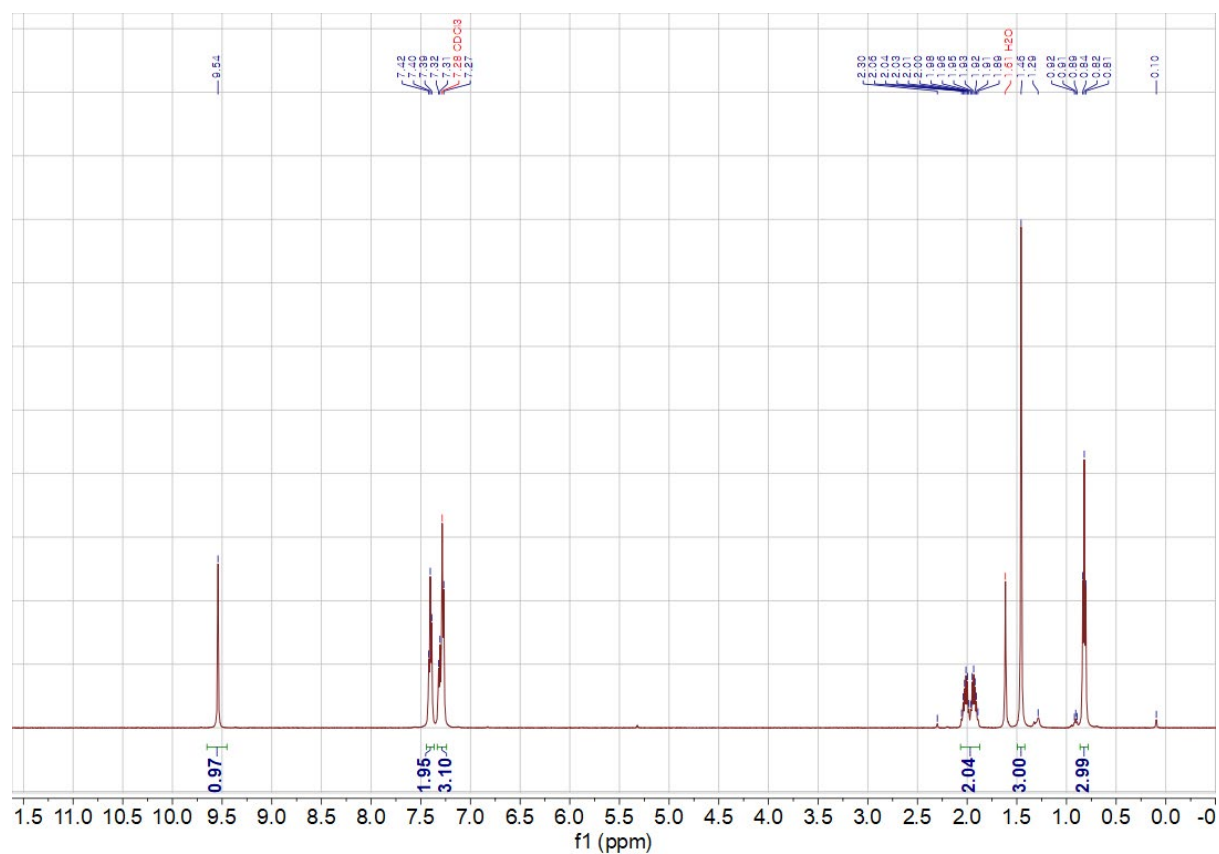
^1H NMR (500 MHz, CDCl_3) of **18**



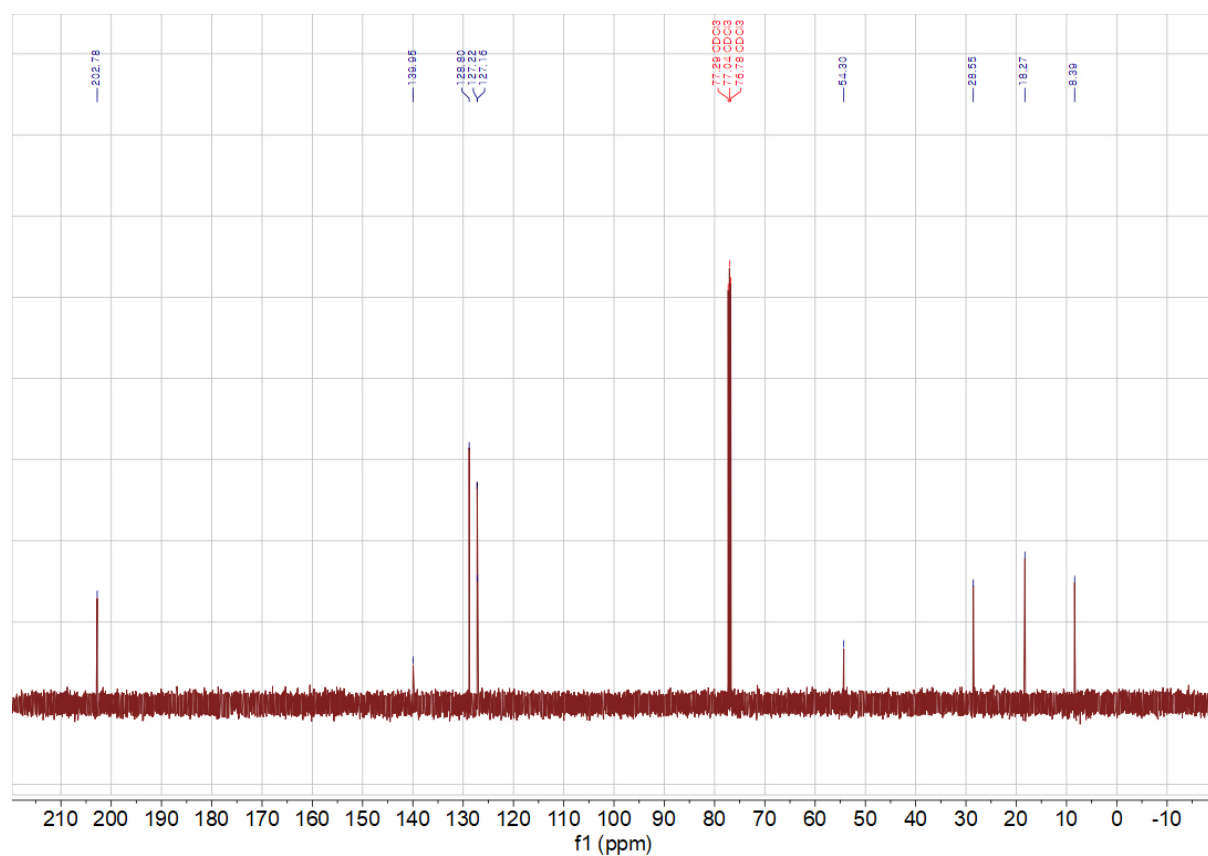
$^{13}\text{C}\{^1\text{H}\}$ NMR (126 MHz, CDCl_3) of **18**



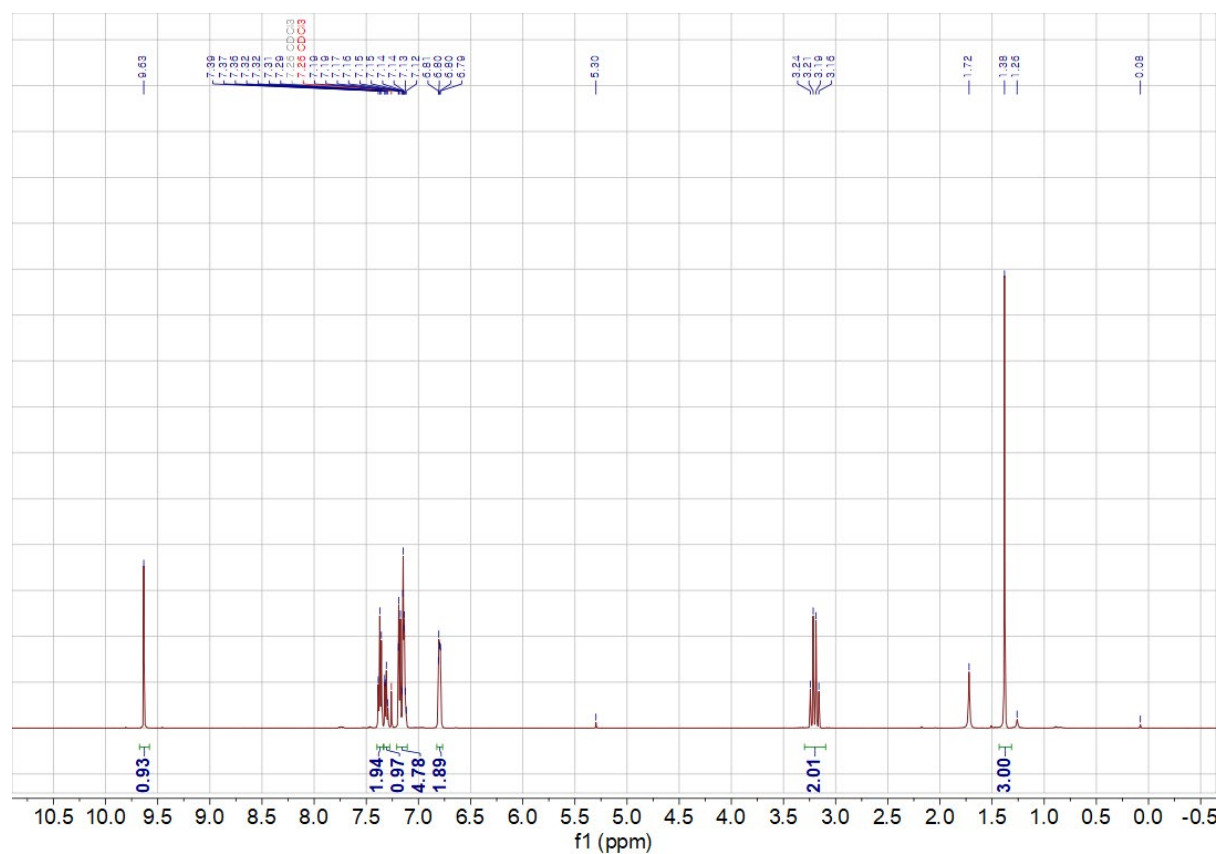
^1H NMR (500 MHz, CDCl_3) of **19**



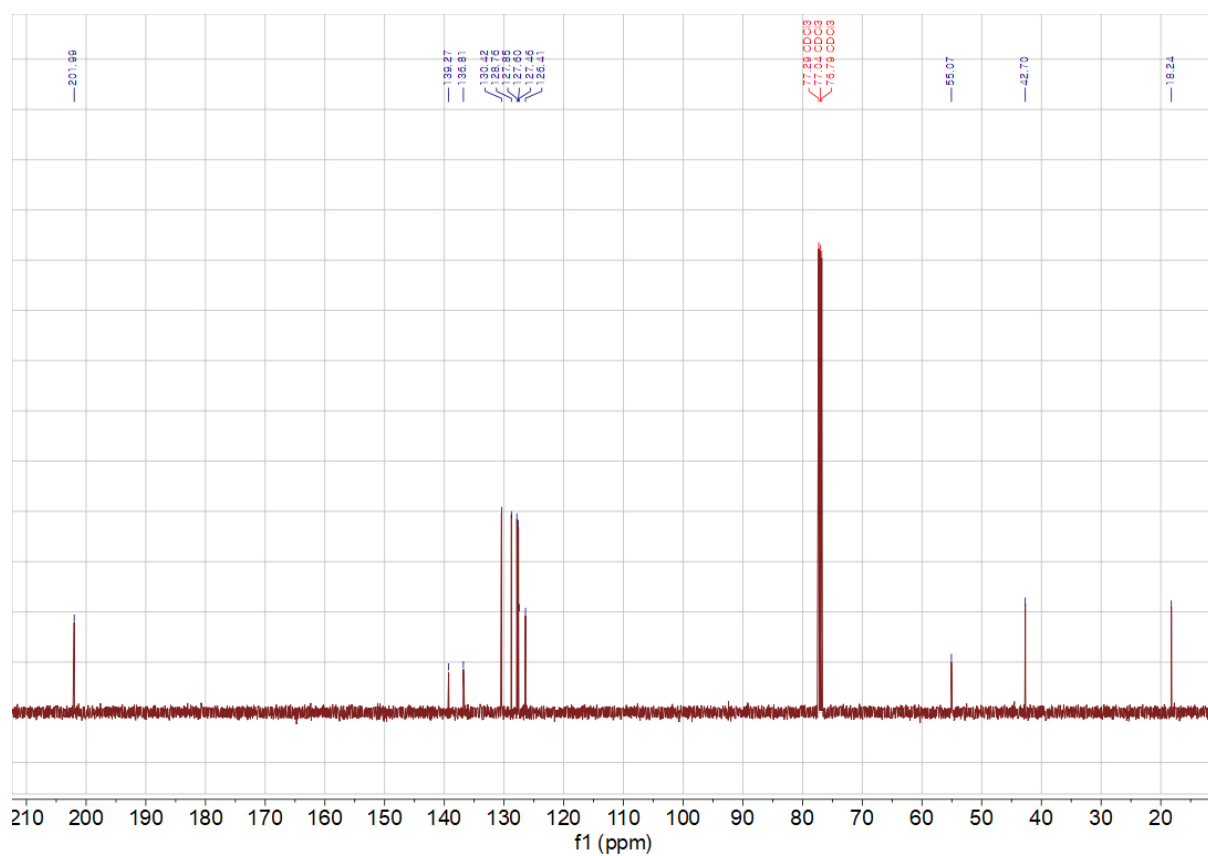
$^{13}\text{C}\{^1\text{H}\}$ NMR (126 MHz, CDCl_3) of **19**



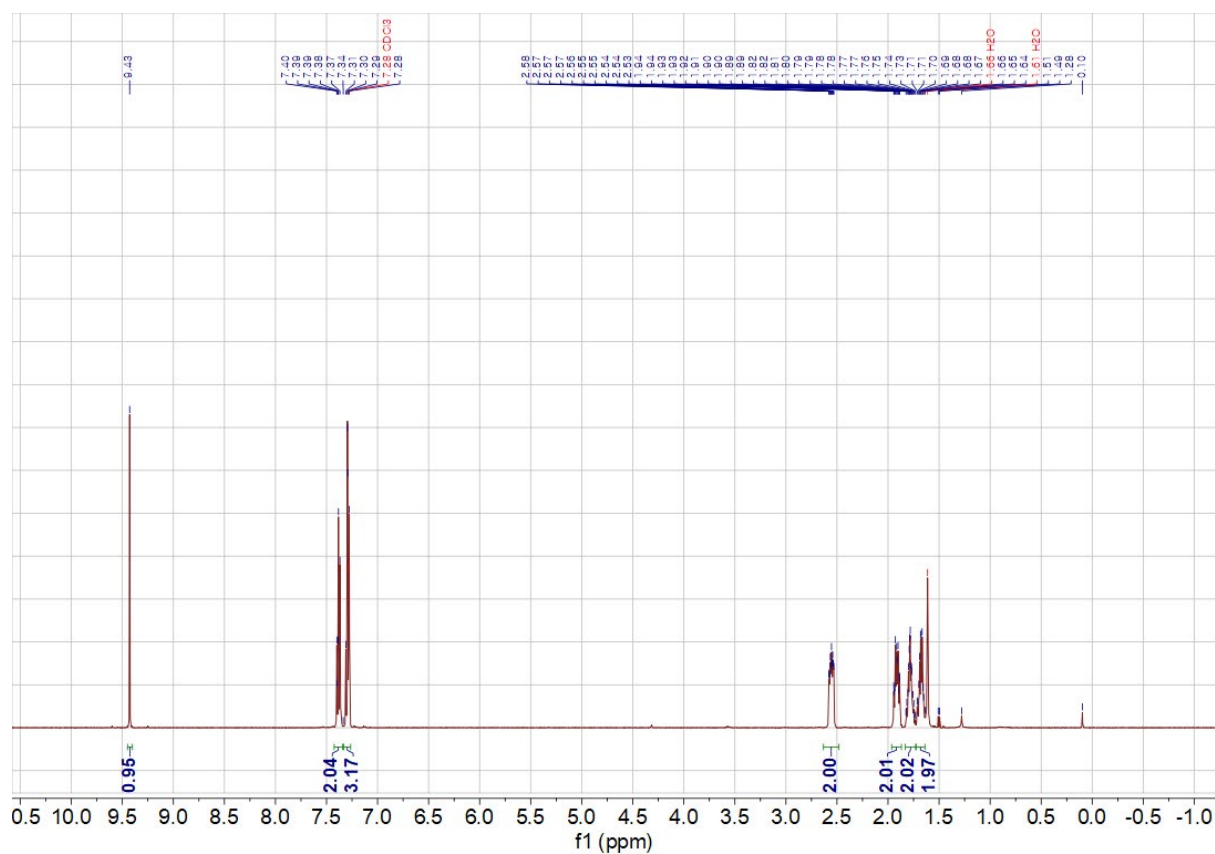
^1H NMR (500 MHz, CDCl_3) of **20**



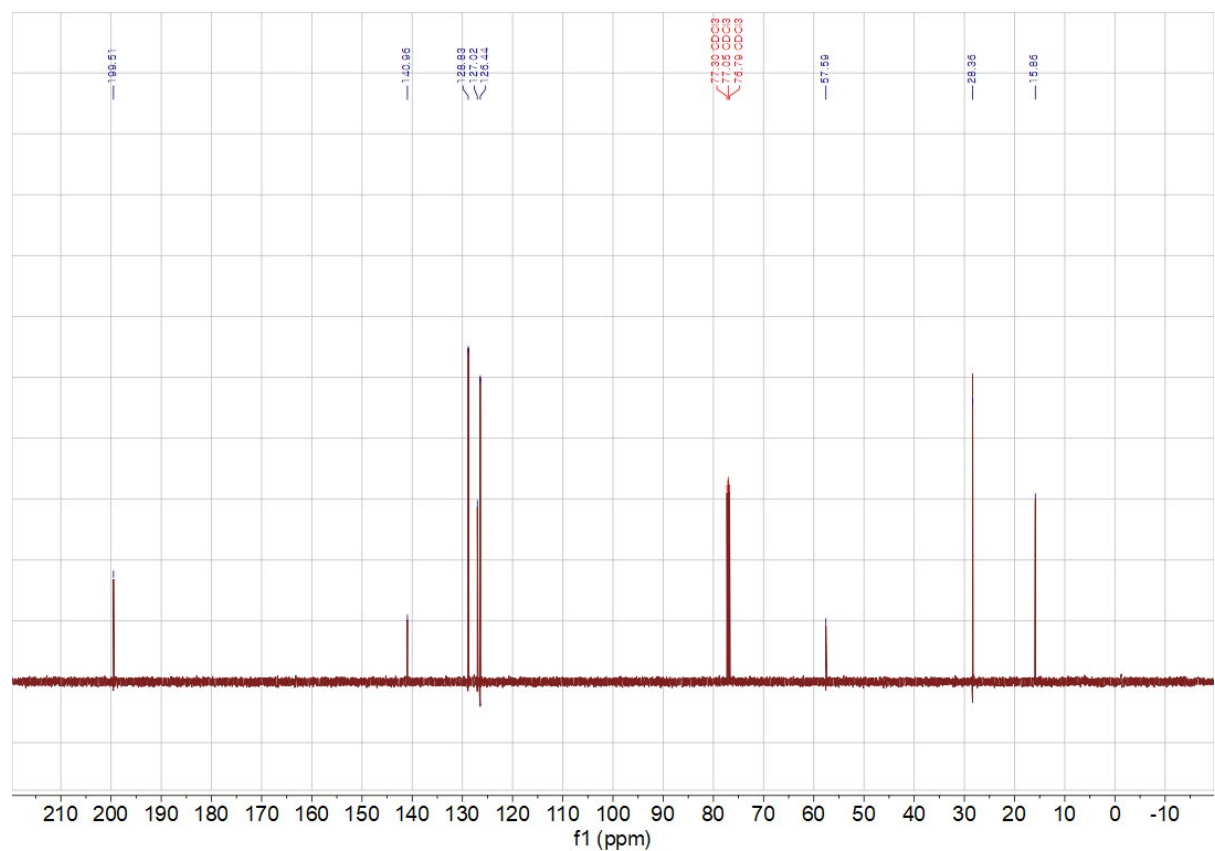
$^{13}\text{C}\{^1\text{H}\}$ NMR (126 MHz, CDCl_3) of **20**



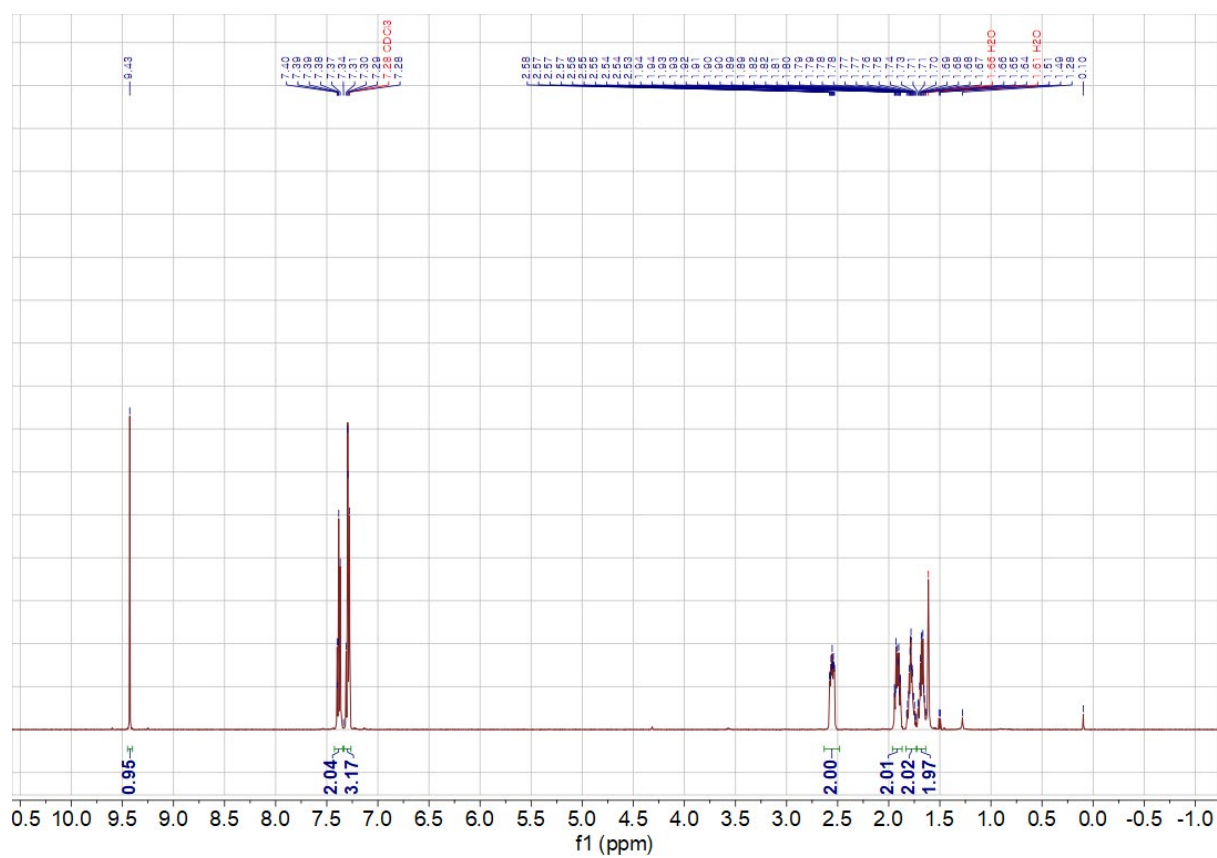
^1H NMR (500 MHz, CDCl_3) of **21**



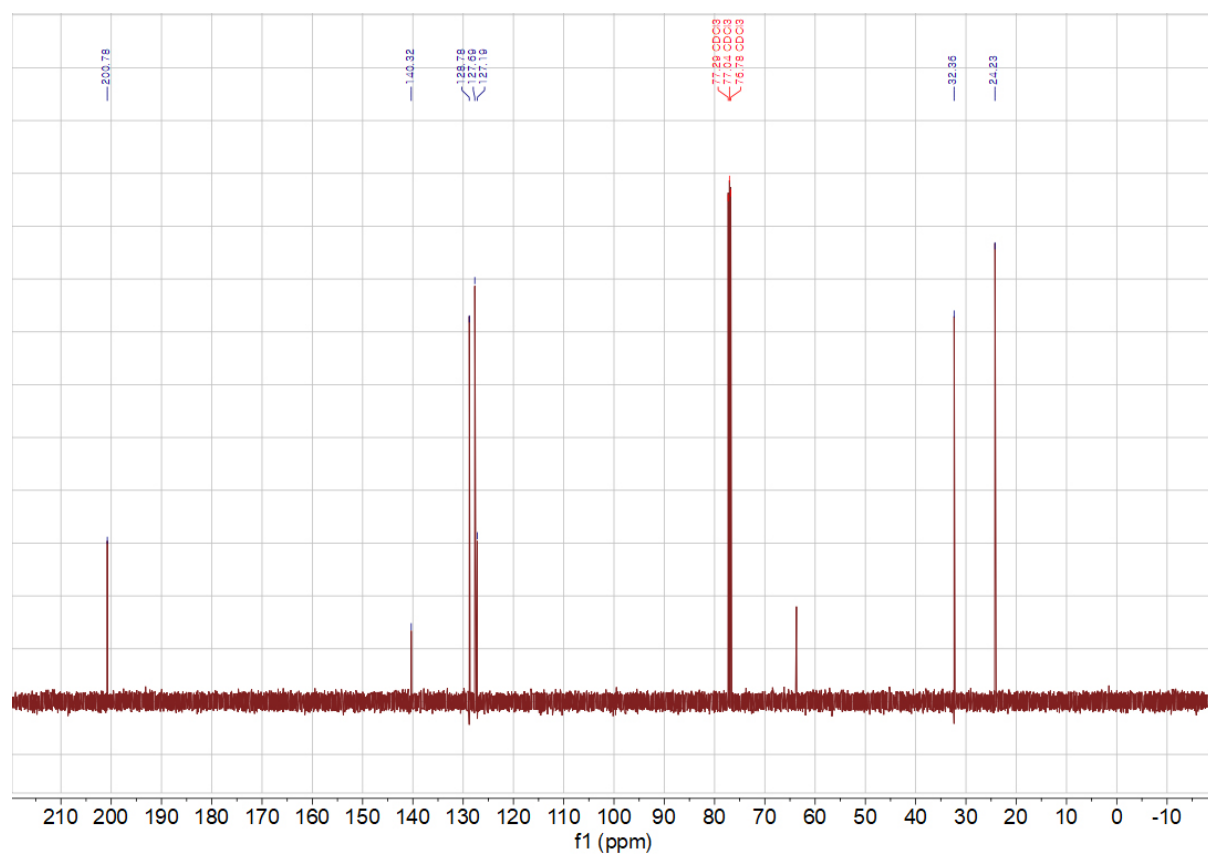
$^{13}\text{C}\{^1\text{H}\}$ NMR (126 MHz, CDCl_3) of **21**



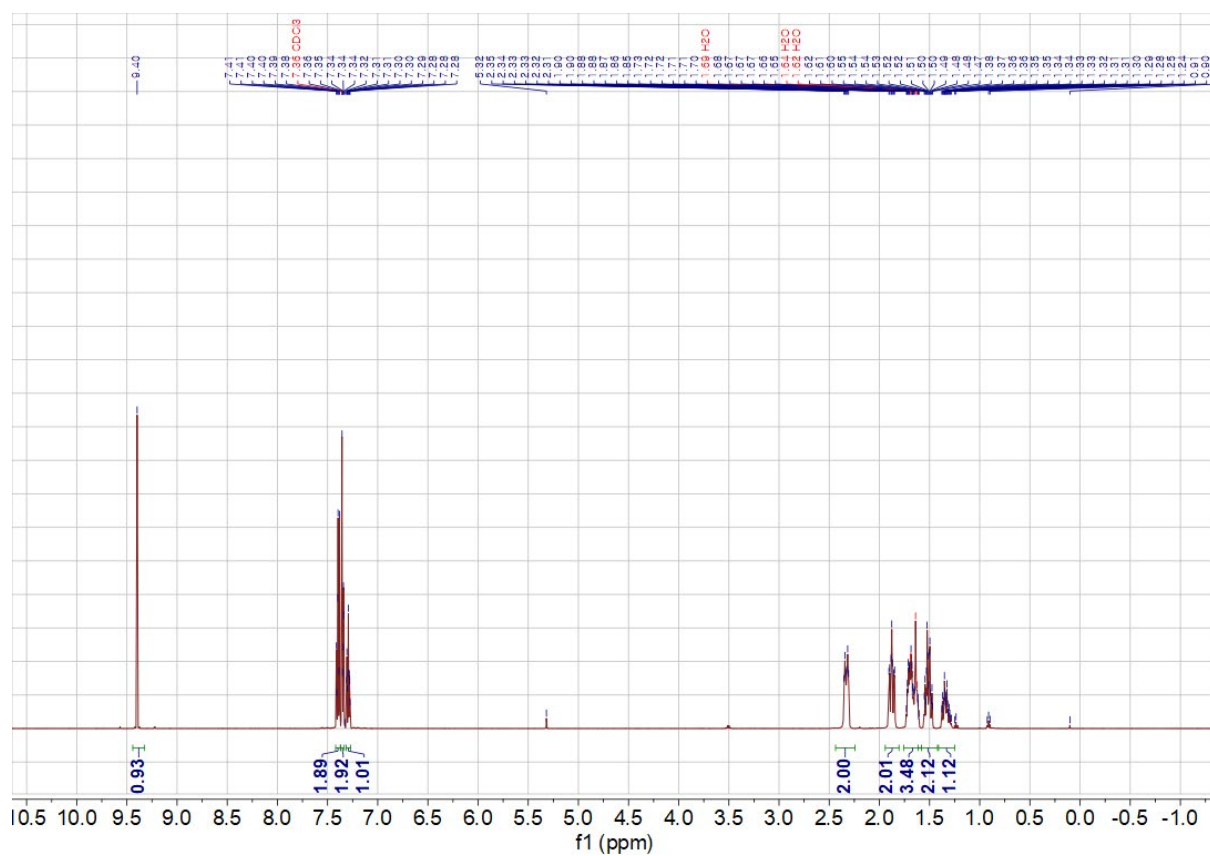
^1H NMR (500 MHz, CDCl_3) of **22**



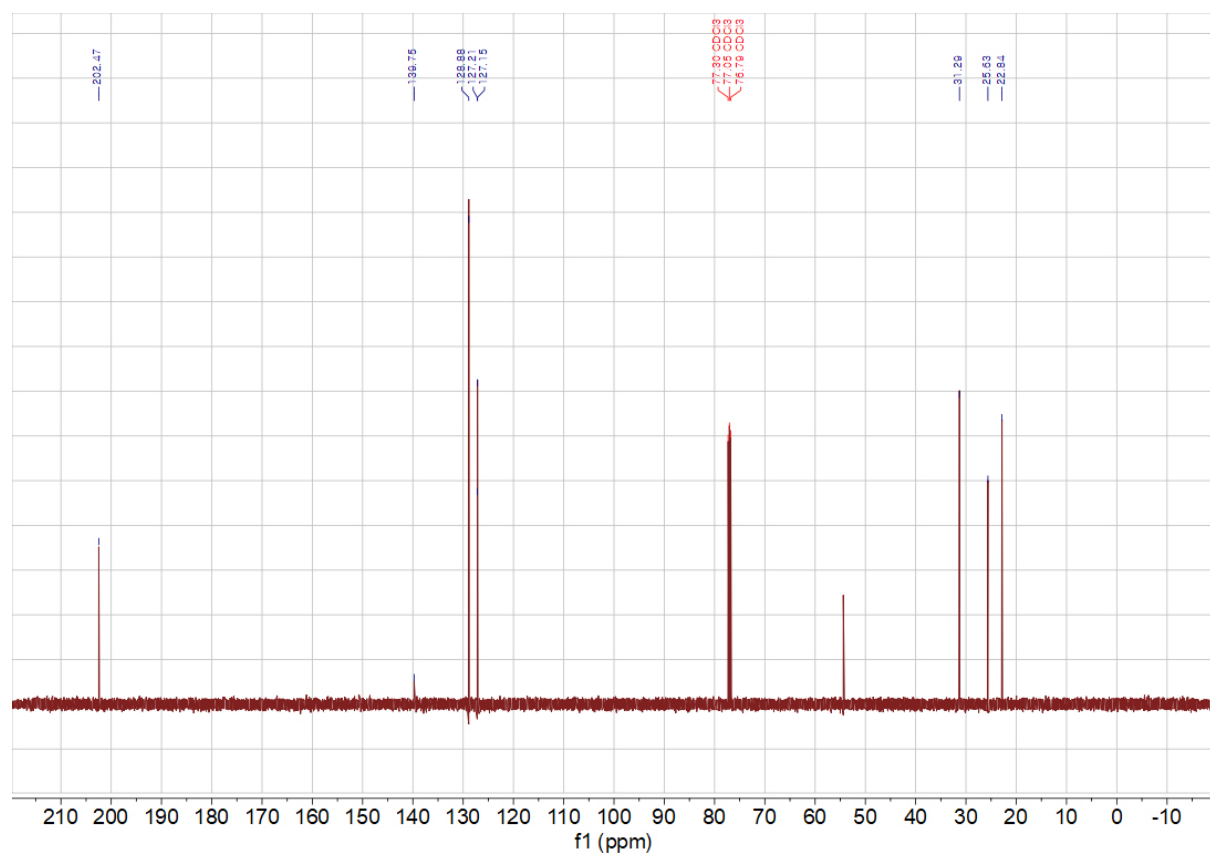
$^{13}\text{C}\{^1\text{H}\}$ NMR (126 MHz, CDCl_3) of **22**



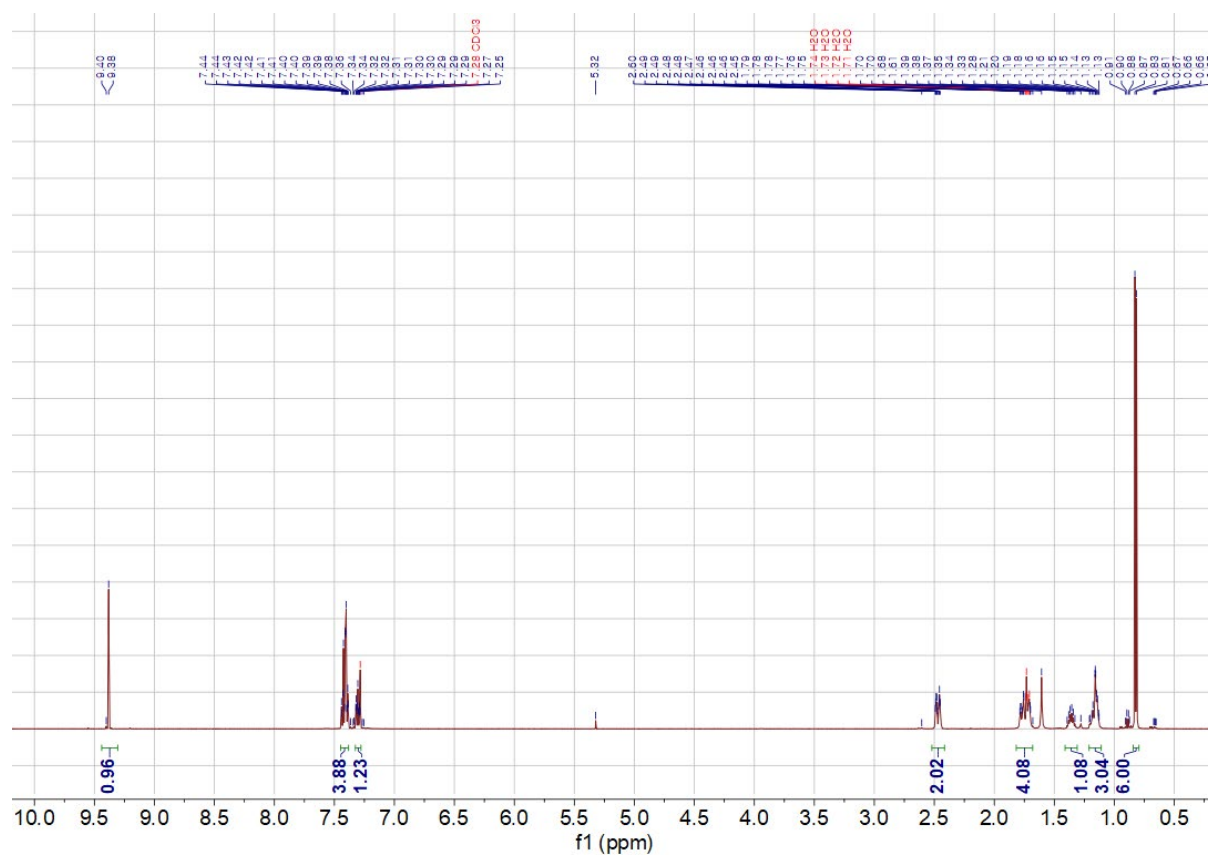
^1H NMR (500 MHz, CDCl_3) of **23**



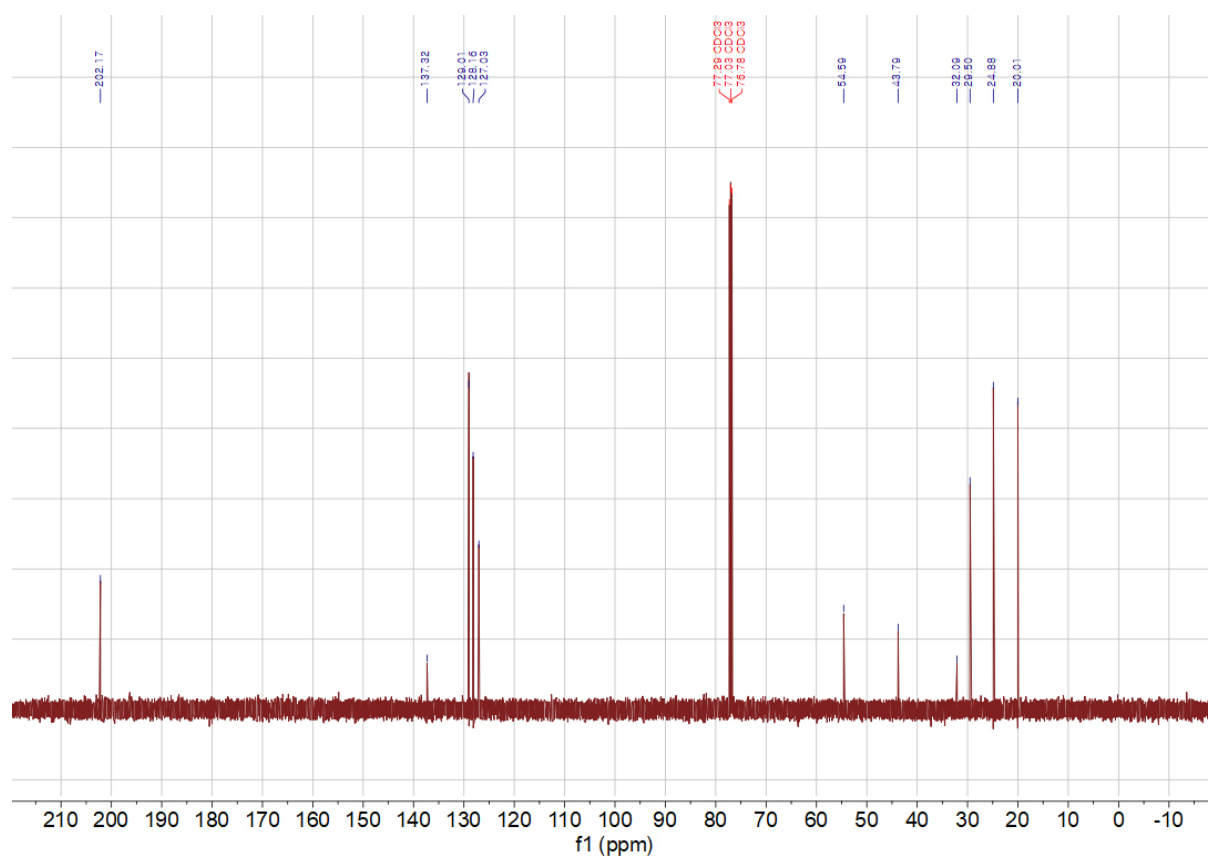
$^{13}\text{C}\{^1\text{H}\}$ NMR (126 MHz, CDCl_3) of **23**



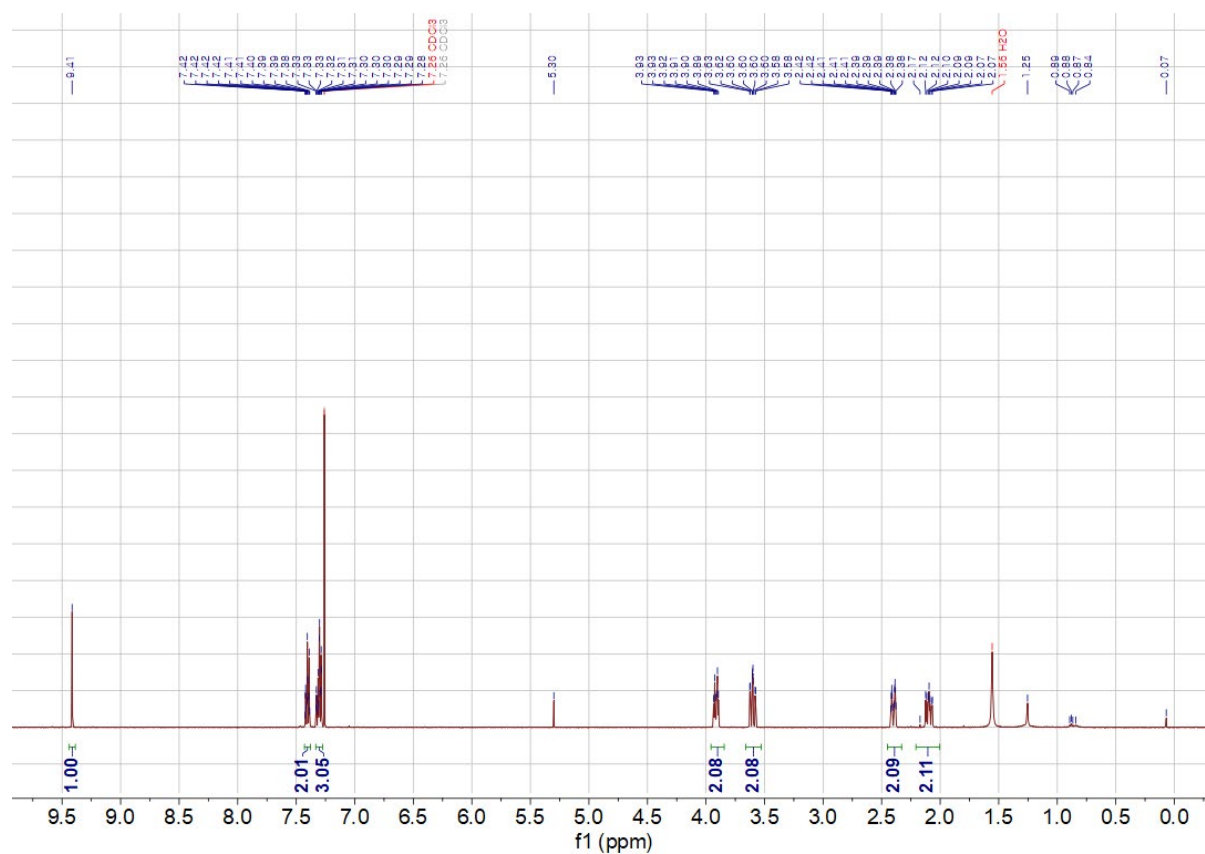
^1H NMR (500 MHz, CDCl_3) of **24**



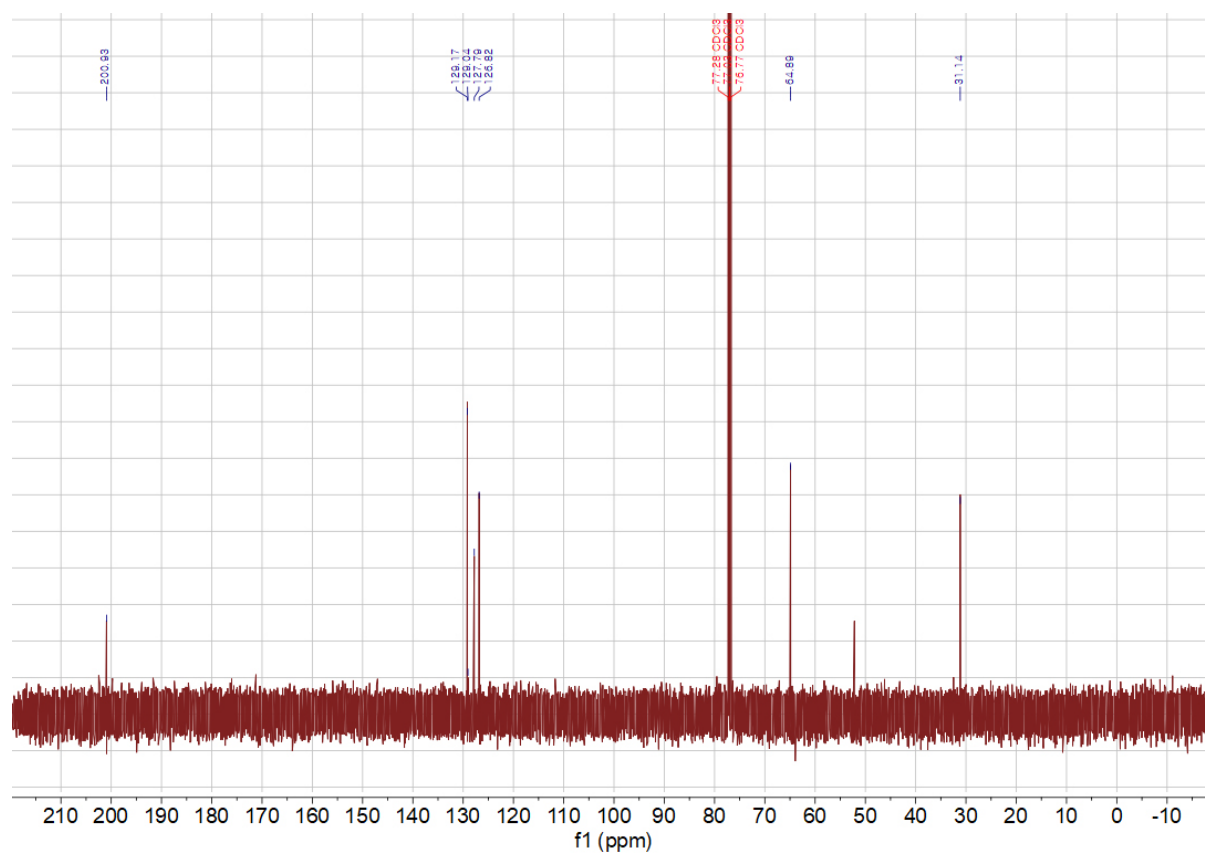
$^{13}\text{C}\{^1\text{H}\}$ NMR (126 MHz, CDCl_3) of **24**



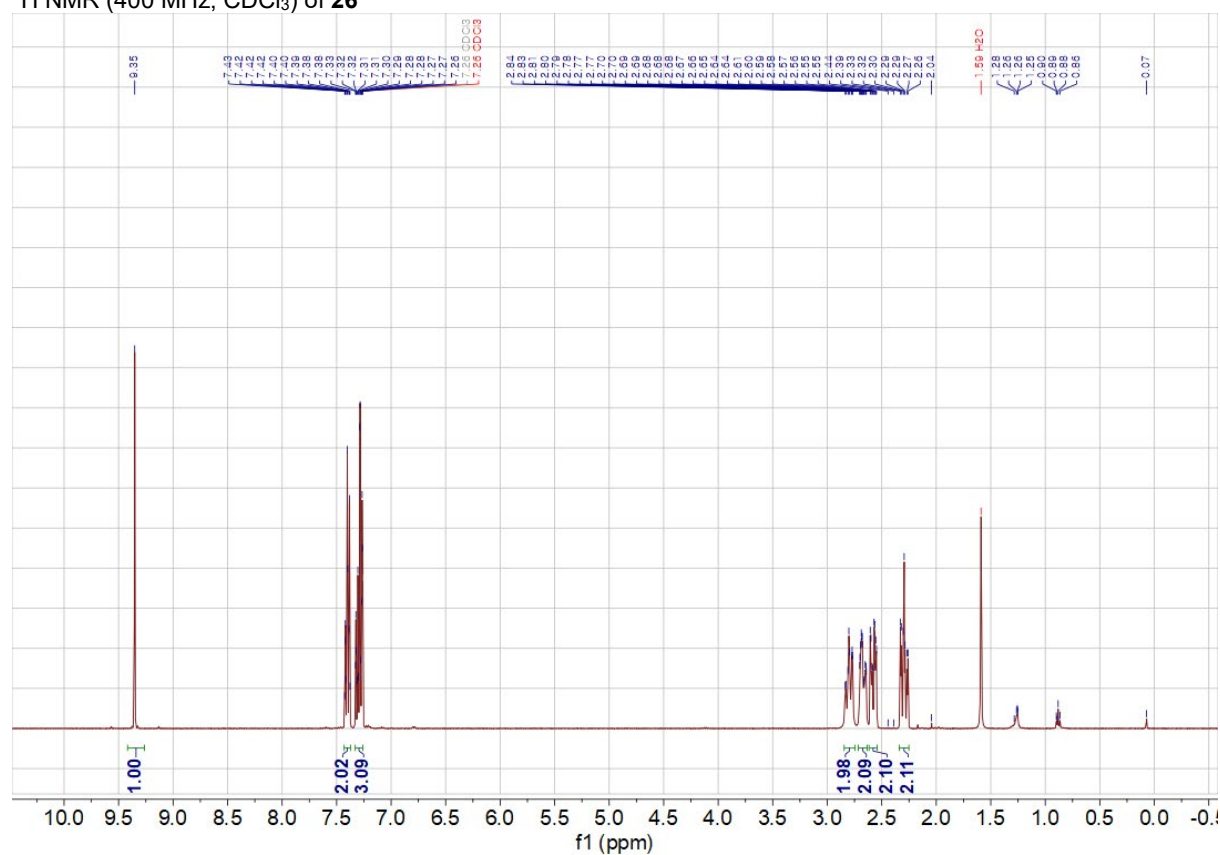
^1H NMR (500 MHz, CDCl_3) of **25**



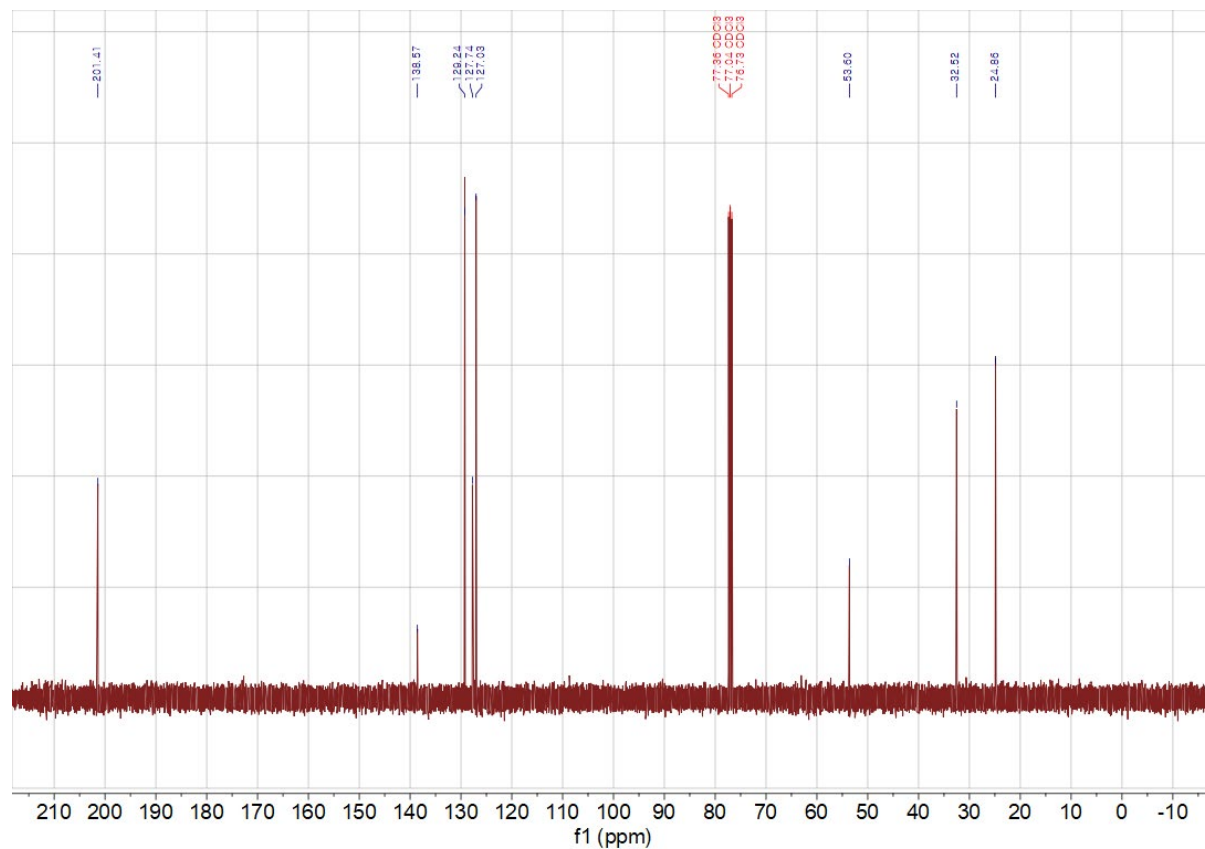
$^{13}\text{C}\{^1\text{H}\}$ NMR (126 MHz, CDCl_3) of **25**



^1H NMR (400 MHz, CDCl_3) of **26**



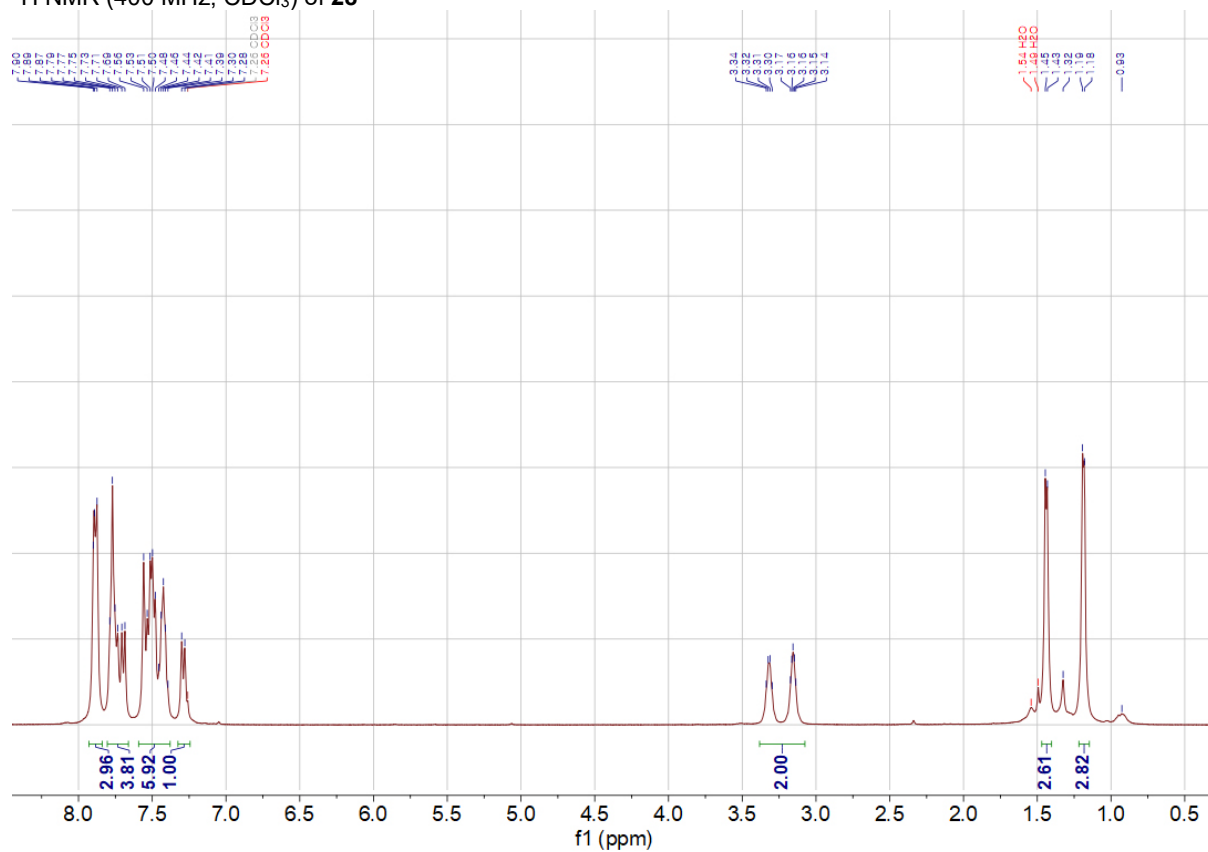
$^{13}\text{C}\{^1\text{H}\}$ NMR (101 MHz, CDCl_3) of **26**



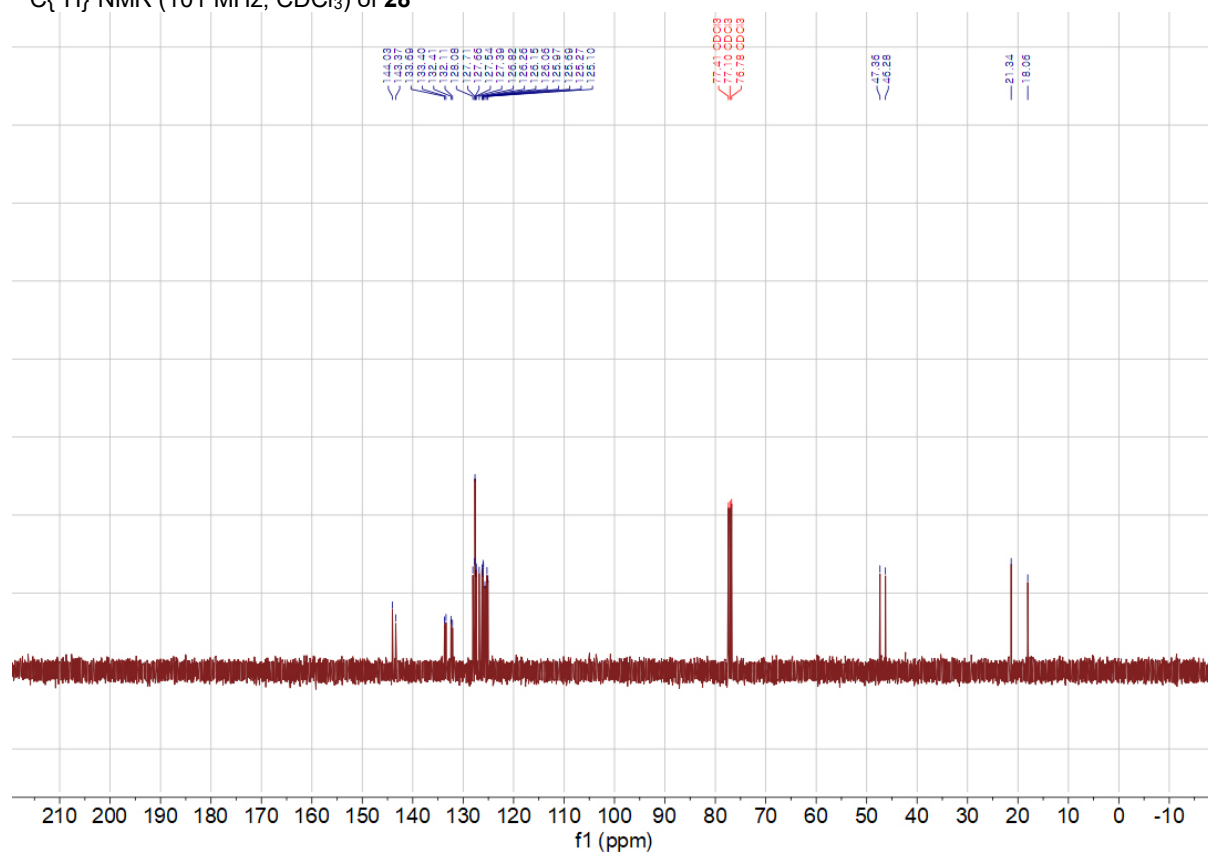
¹H NMR spectrum (CDCl₃) of compound 10b. The x-axis represents the chemical shift f1 (ppm) from 10.0 to -0.5. The spectrum shows several peaks with corresponding integration values:

- Peak at ~9.45 ppm (s, 1H) with integration 0.09.
- Peak at ~7.25 ppm (d, 2H) with integration 1.00.
- Peak at ~2.95 ppm (s, 3H) with integration 2.19.
- Peak at ~2.65 ppm (s, 3H) with integration 3.01.
- Peak at ~1.55 ppm (s, 3H) with integration 1.04.
- Peak at ~1.45 ppm (s, 3H) with integration 1.09.
- Peak at ~1.35 ppm (s, 3H) with integration 1.15.
- Peak at ~1.55 ppm (s, 3H) with integration 1.79.
- Peak at ~1.45 ppm (s, 3H) with integration 1.98.
- Peak at ~1.35 ppm (s, 3H) with integration 3.52.
- Peak at ~1.25 ppm (s, 3H) with integration 1.04.

^1H NMR (400 MHz, CDCl_3) of **28**

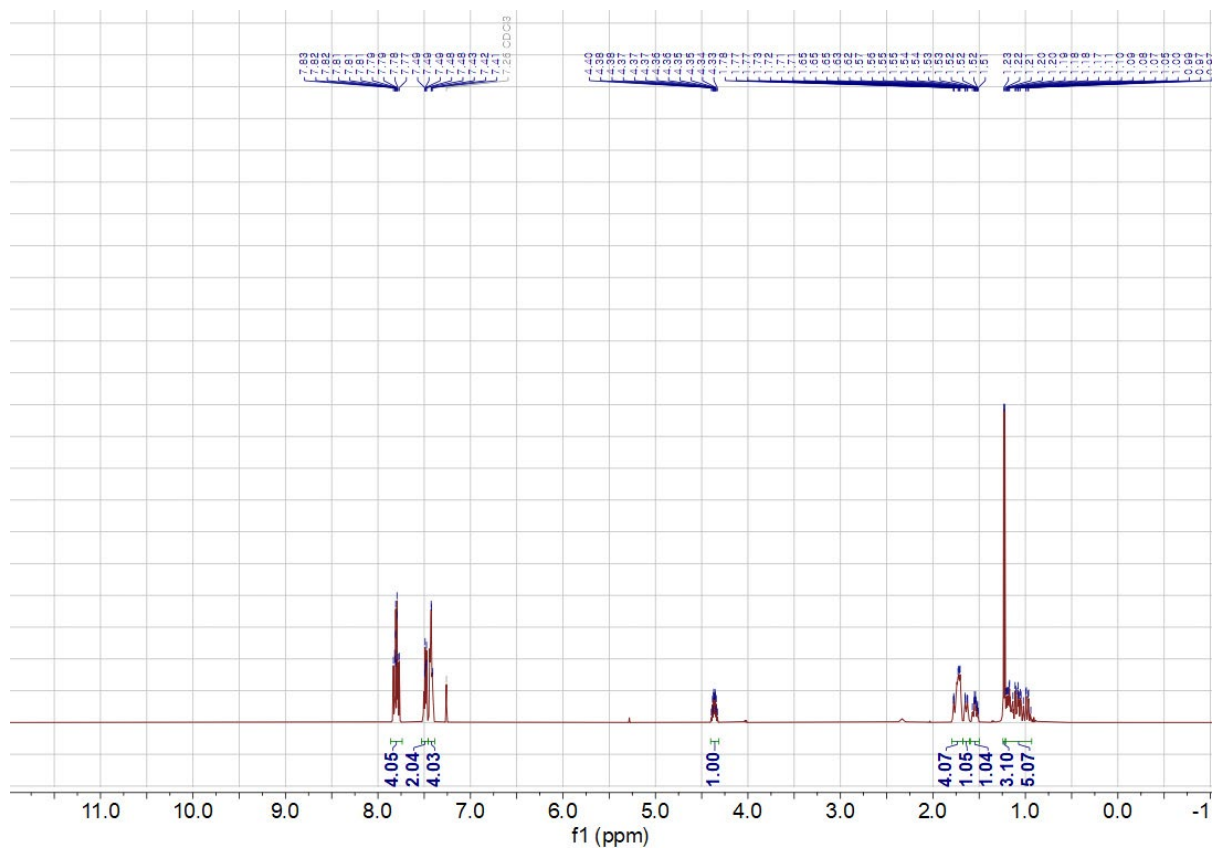


$^{13}\text{C}\{^1\text{H}\}$ NMR (101 MHz, CDCl_3) of **28**

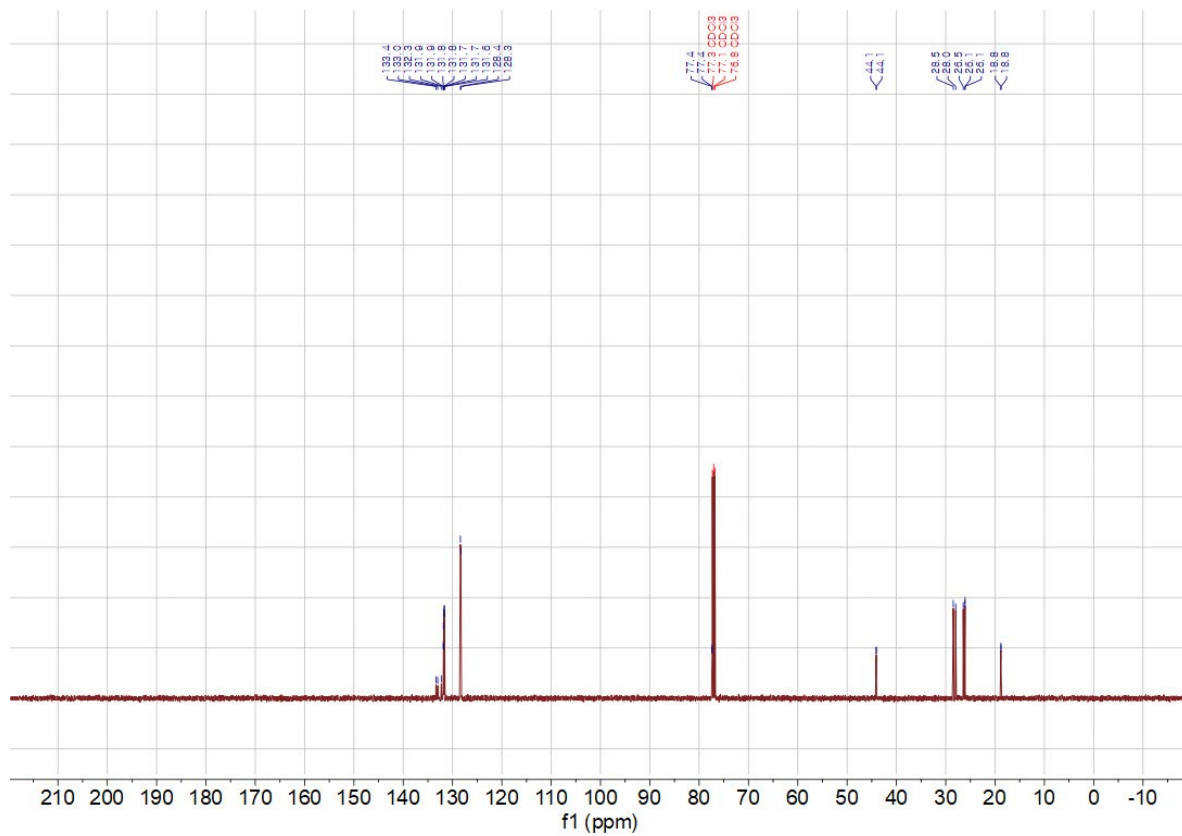


10.3. Substrates for cyclic voltammetry

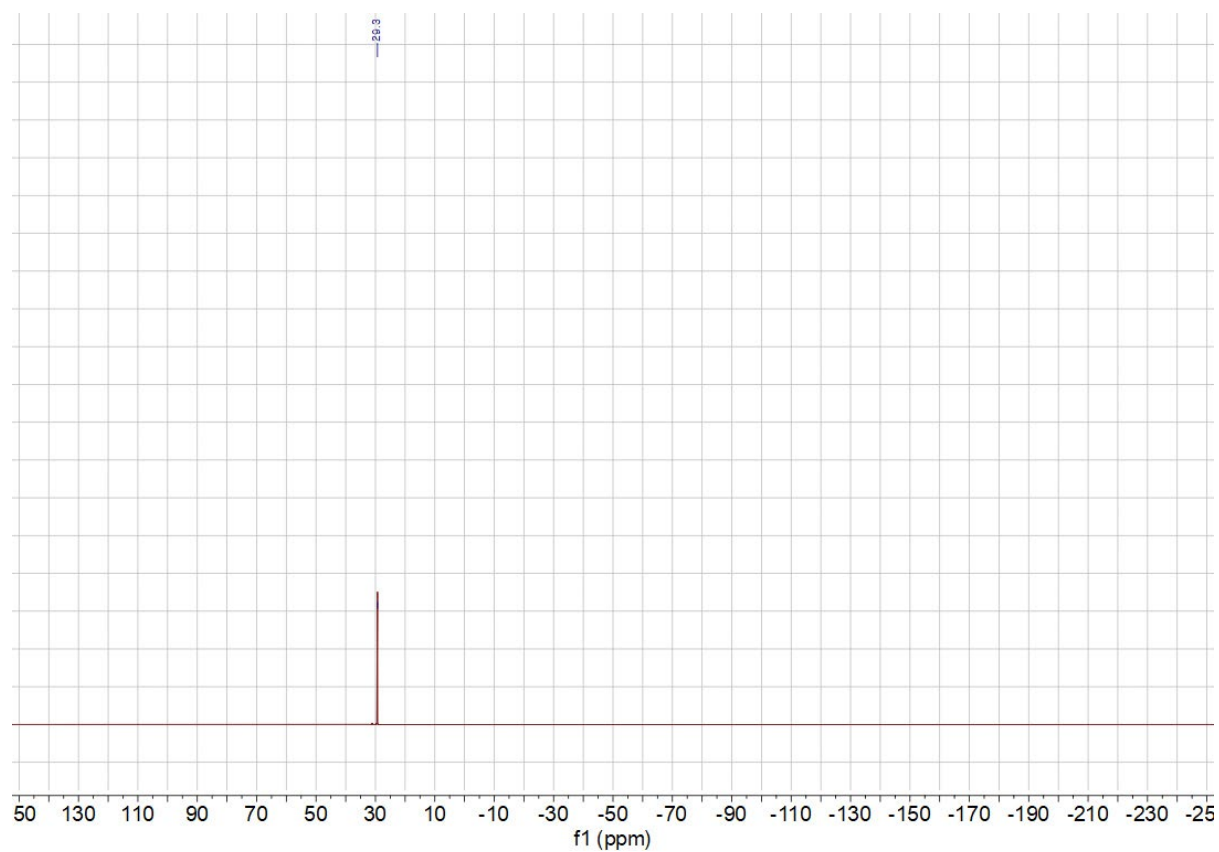
^1H NMR spectrum (500 MHz, CDCl_3) of **30**



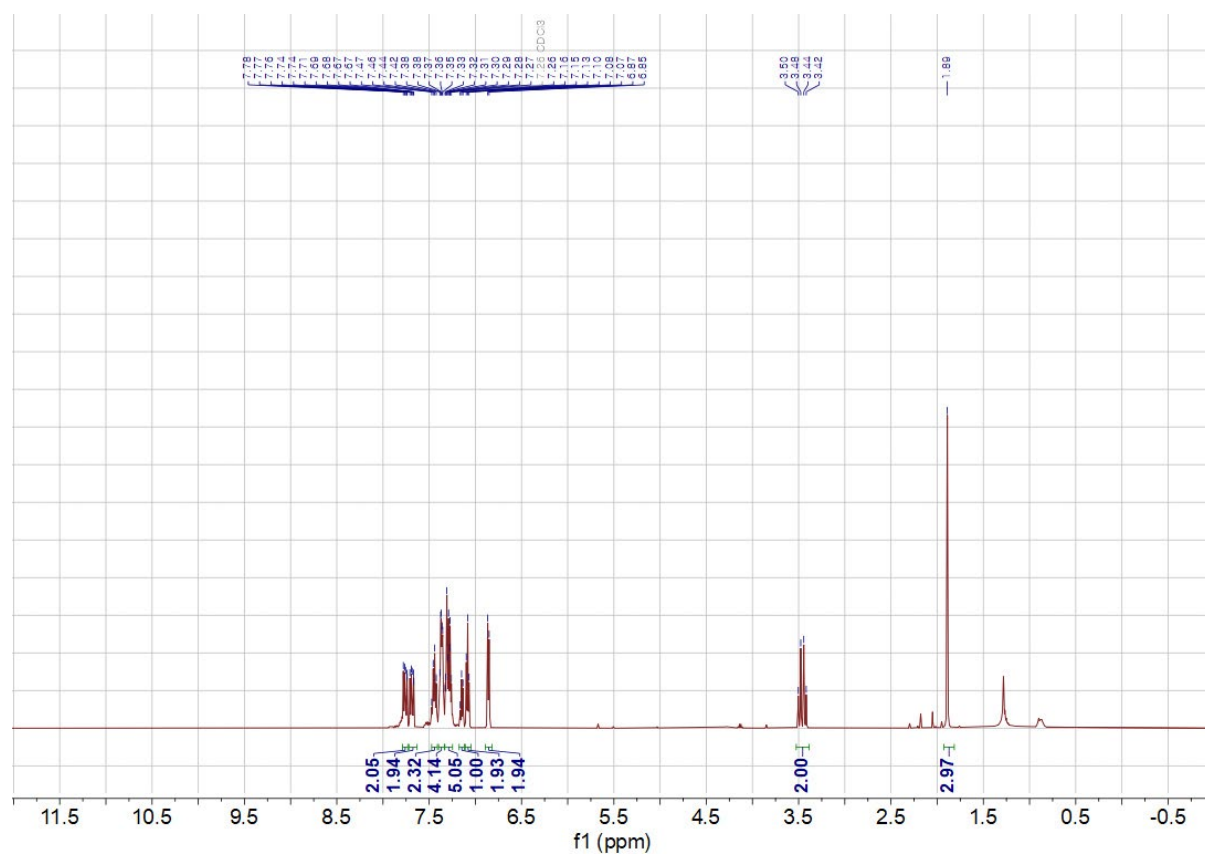
$^{13}\text{C}\{^1\text{H}\}$ NMR spectrum (126 MHz, CDCl_3) of **30**



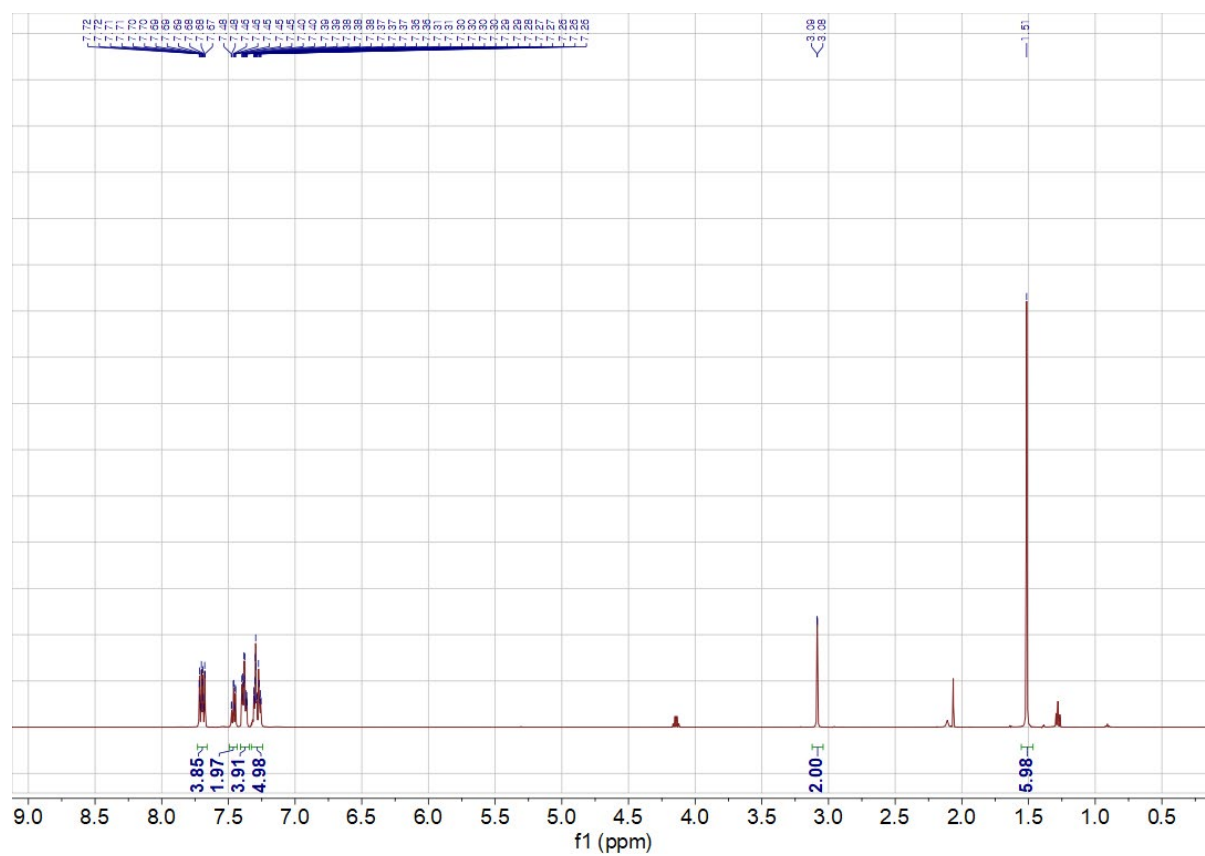
$^{31}\text{P}\{^1\text{H}\}$ NMR spectrum (202 MHz, CDCl_3) of **30**



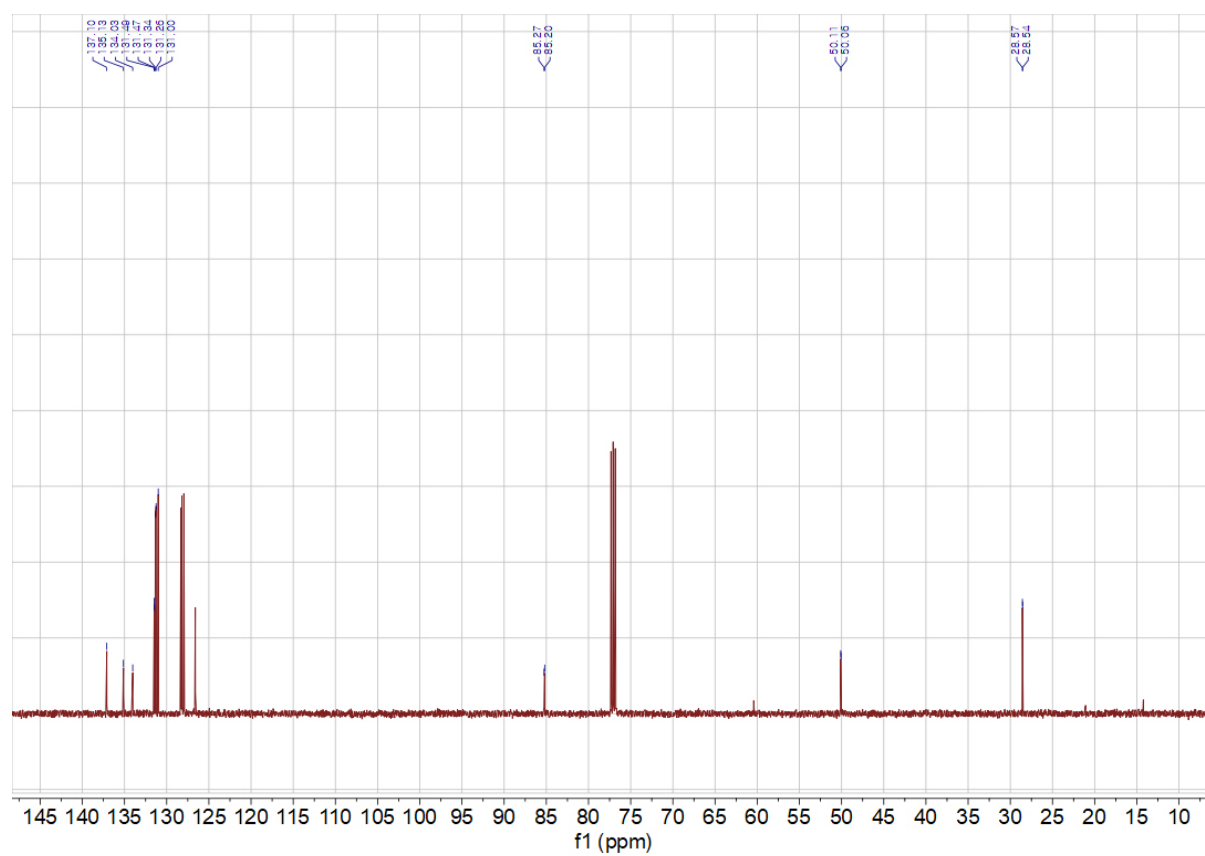
^1H NMR spectrum (500 MHz, CDCl_3) of **31**



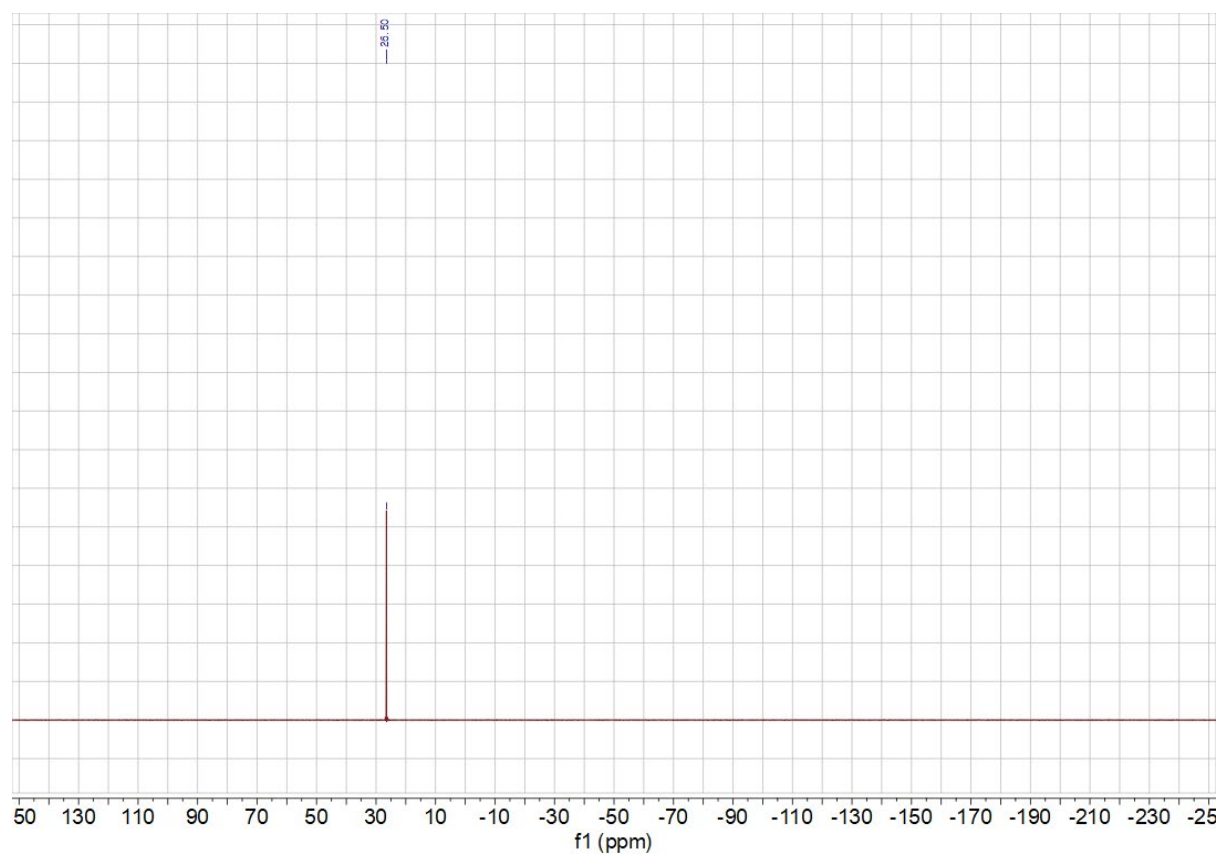
^1H NMR (400 MHz, CDCl_3) of **32**



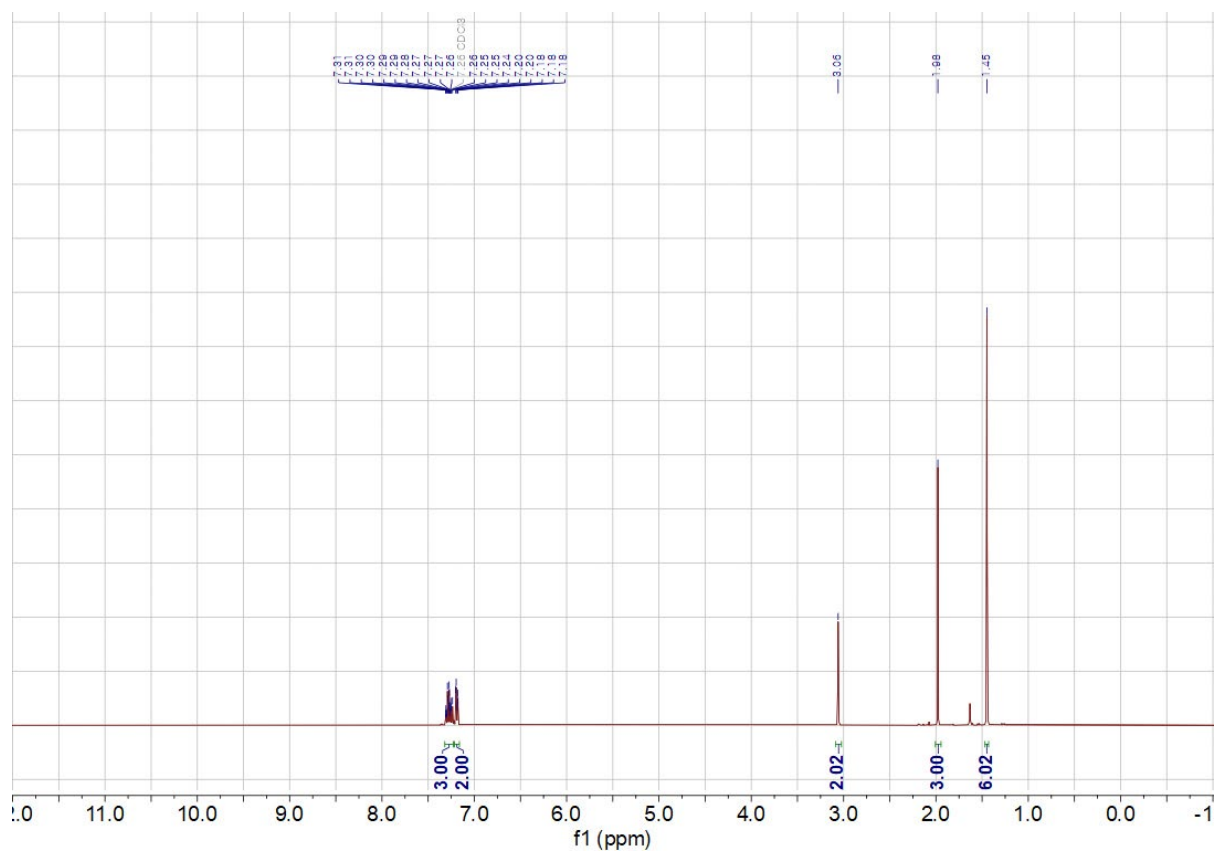
$^{13}\text{C}\{^1\text{H}\}$ NMR (101 MHz, CDCl_3) of **32**



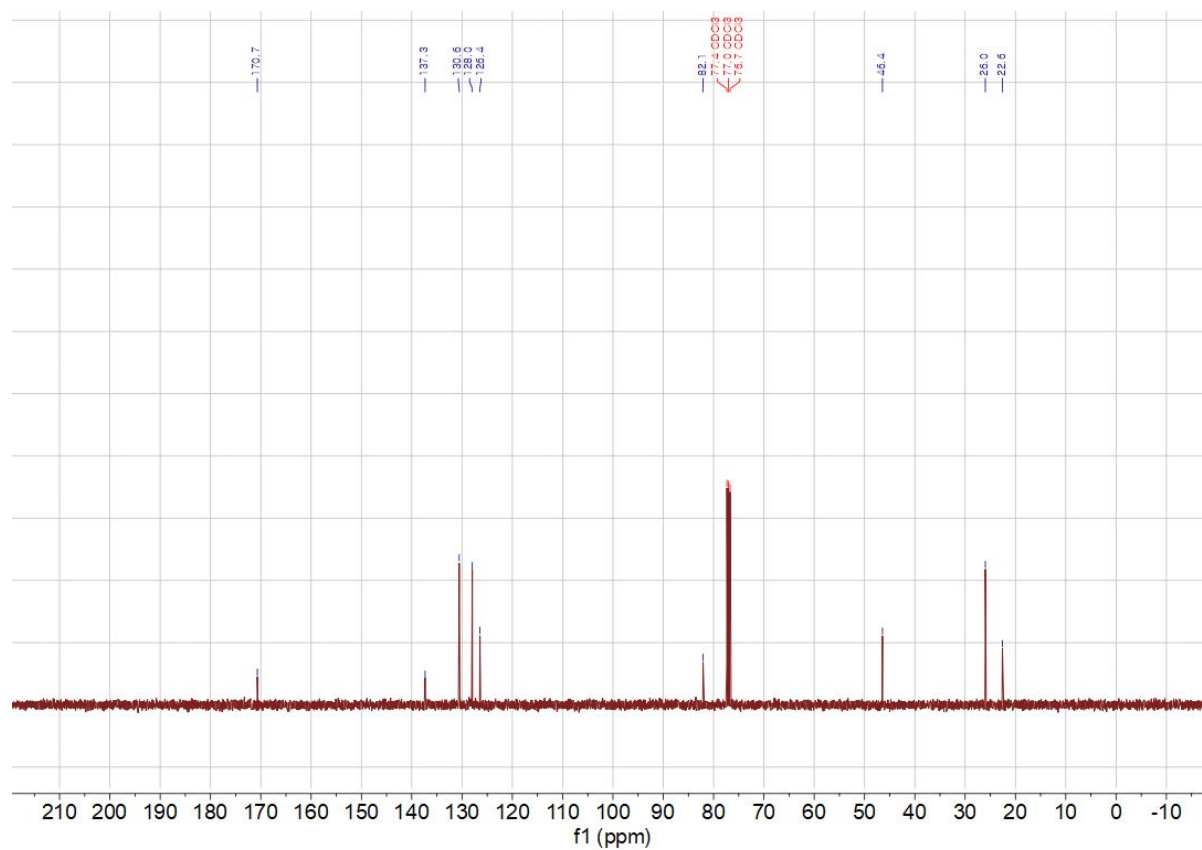
$^{31}\text{P}\{^1\text{H}\}$ NMR (202 MHz, CDCl_3) of **32**



^1H NMR spectrum (400 MHz, CDCl_3) of **34**



$^{13}\text{C}\{^1\text{H}\}$ NMR spectrum (101 MHz, CDCl_3) of **34**



11. References

- S1. K. Lam and I. E. Markó, *Org. Lett.*, 2011, **13**, 406-409.
- S2. A. P. Atkins, A. C. Rowett, D. M. Heard, J. A. Tate and A. J. Lennox, *Org. Lett.*, 2022, **24**, 5105-5108.
- S3. J. Margalef, M. Biosca, P. de la Cruz-Sánchez, X. Caldentey, C. Rodríguez-Escrich, O. Pàmies, M. A. Pericàs and M. Diéguez, *Adv. Synth. Catal.*, 2021, **363**, 4561-4574.
- S4. X. Tian, T. A. Karl, S. Reiter, S. Yakubov, R. de Vivie-Riedle, B. König and J. P. Barham, *Angew. Chem. Int. Ed.*, 2021, **60**, 20817-20825.
- S5. R. Gazzaeva, A. Fedotov, E. Trofimova, O. Popova, S. Mochalov and N. Zefirov, *Russ. J. Org. Chem.*, 2006, **42**, 87-99.
- S6. Y. Suzuki, K. Taniguchi, H. N. Hoang, M. Tamura and T. Matsuda, *Tetrahedron Lett.*, 2022, **99**, 153837.
- S7. R. Singha and J. K. Ray, *Tetrahedron Lett.*, 2016, **57**, 5395-5398.
- S8. M. Kasai, C. Froussios and H. Ziffer, *J. Org. Chem.*, 1983, **48**, 459-464.
- S9. H. Miura, Y. Hachiya, H. Nishio, Y. Fukuta, T. Toyomasu, K. Kobayashi, Y. Masaki and T. Shishido, *ACS Catal.*, 2021, **11**, 758-766.
- S10. R. V. Ottenbacher, A. A. Bryliakova, M. V. Shashkov, E. P. Talsi and K. P. Bryliakov, *ACS Catal.*, 2021, **11**, 5517-5524.
- S11. J. W. Wilt and D. D. Roberts, *J. Org. Chem.*, 1962, **27**, 3430-3434.
- S12. T. Barber, S. P. Argent and L. T. Ball, *ACS Catal.*, 2020, **10**, 5454-5461.
- S13. J. A. Friest, Y. Maezato, S. Broussy, P. Blum and D. B. Berkowitz, *J. Am. Chem. Soc.*, 2010, **132**, 5930-5931.
- S14. H. Lund, *J. Electroanal. Chem.*, 2005, **584**, 174-181.
- S15. S. Khan, A. Ghatak and S. Bhar, *Tetrahedron Lett.*, 2015, **56**, 2480-2487.
- S16. A. K. Chakraborti and Shivani, *J. Org. Chem.*, 2006, **71**, 5785-5788.