# Cage-like structures based on constrained cyclic arylopeptoids

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# I-Chemicals and general experimental information

Chemicals: Rink acid resin (0.62 mmol/g loading) and 2-chlorotrityl resin (1.06 mmol/g loading) was obtained from Novabiochem, dichloromethane from Carlo Erba; 3-chloromethylbenzoylchloride, 2-chloromethylbenzoylchloride, TFA, DIPEA, 4-DMAP isopropyl and propargyl amines from TCI; DMSO from Acros. 1,4-bis(azidomethyl)benzene was synthetized according to literature procedures. 10 mL jacketed reactors were purchased from Kamush and thermo-regulated using a Lauda thermostat. Purification was performed on a Buchi Pure Chromatography system.

NMR was recorded on a Bruker AC-400 spectrometer, operating at 400 MHz for <sup>1</sup>H and 100 MHz for <sup>13</sup>C J-modulation or a Bruker AC-500 spectrometer operating at 500 MHz for <sup>1</sup>H and 125 MHz for <sup>13</sup>C.

High-resolution mass spectra (HRMS) are recorded using electrospray ionization in positive mode (ESI+) on or a Q Exactive Quadrupole-Orbitrap Mass Spectrometer or on a Qtof-micro WATERS (3000V) spectrometer. Liquid chromatography mass spectroscopy (LC-MS) were recorded on a Q Exactive Quadrupole-Orbitrap Mass Spectrometer coupled to a UPLC Ultimate 3000 (Kinetex EVO C18; 1,7 $\mu$ m; 100mm x 2,1mm column with a flow rate of 0.45 ml.min-1 with the following gradient: a linear gradient of solvent B from 5% to 95% over 7.5 min (solvent A = H2O + 0.1% formic acid, solvent B = acetonitrile + 0.1% formic acid) equipped with a DAD UV/VIS 3000 RS detector) or on a Qtof-micro WATERS (3000V) with electrospray ionization coupled to a HPLC ALLIANCE WATERS system with a diode array detector (DAD) using a reverse-phase C18 Nucleosil column (100 mm x 2.1 mm, 5  $\mu$ m pore size) with an H2O/acetonitrile gradient and a flow rate of 0.2 mL/min.

Analytical HPLC was recorded on a Hitachi liquid chromatograph (Oven 5310, 30°C; Pump 5160; DAD detector 5430) equipped with a C18 Acclaim column (4.6 mm×250 mm, 5µm, 120Å). Detection wavelength was 240nm or 280nm and flow rate 0.5mL/min. Gradient elution used (A) water/0.1% TFA; (B) methanol according Method A: (Solvents A/B: 0 to 5 minutes isocratic at 95/5; 5 to 25 minutes gradient to 5/95; 25 to 35 minutes isocratic at 5/95; 35 to 45 minutes gradient to 95/5; 45 to 50 minutes 95/5) or Method B (Solvents A/B: 0 to 5 minutes isocratic at 95/5; 5 to 10 minutes gradient to 75/25; 10 to 50 minutes gradient to 40/60; 50 to 65 minutes gradient to 5/95; 65 to 70 minutes isocratic at 5/95; 70 to 80 minutes gradient to 95/5).

- II. General Procedures and compounds characterization
- II.1. Linear arylopeptoids
- II.1.a. Linear ortho-hexamer **Ia**



a) Swelling, CH<sub>2</sub>Cl<sub>2</sub>; b) 2-chloromethyl benzoylchloride, DIPEA, DMAP; CH<sub>2</sub>Cl<sub>2</sub>; c) Isopropyl amine or propargyl amine, dmso; d) 2-chloromethyl benzoylchloride, DIPEA, CH<sub>2</sub>Cl<sub>2</sub>; e) Cleavage: TFA/CH<sub>2</sub>Cl<sub>2</sub> (20:80).

Scheme S1. Solid-phase submonomer synthesis of Ia on Rink acid resin

1-For 100 mg of resin, swelling: 2 ml of  $CH_2Cl_2$  at RT for 10 min

2- 2-chloromethylbenzoylchloride (3 equiv. per mmol loading), 4-DMAP (3 equiv. per mmol loading) DIPEA (3 equiv. per mmol loading) dissolved in 1 mL  $CH_2Cl_2$  were added at RT, shaken 10 minutes. This step is repeated. The liquid was drained off, then the resin was washed with  $CH_2Cl_2$  (5x2 ml), then with DMSO (5x2 ml).

3- Isopropyl amine (20 equiv per mmol loading) dissolved in 0.5 mL of DMSO were added. The reaction was shaken for 1h at RT. The liquid was drained off, then the resin was washed with DMSO (5x2 ml), then with  $CH_2CL_2$  (5x2 ml).

4- 2-chloromethylbenzoylchloride (3 equiv. per mmol loading), DIPEA (6 equiv. per mmol loading) dissolved in 1 mL  $CH_2Cl_2$  were added at RT, shaken 10 minutes. The liquid was drained off, then the resin was washed with  $CH_2Cl_2$  (5x2 ml), then with DMSO (5x2 ml).

5- Isopropyl amine or propargyl amine (20 equiv per mmol loading, 1M) dissolved in 0.5 mL of DMSO were added. The reaction was shaken for 1h at RT. The liquid was drained off, then the resin was washed with DMSO (5x2 ml), then with  $CH_2CL_2$  (5x2 ml).

Steps 4 and 5 were repeated to grow the targeted arylopeptoid oligomer until the expected sequence length.

6-The arylopeptoid was cleaved from the resin by the addition of 1 ml of  $CH_2Cl_2/TFA$  (8:2) for 30 min at RT. The solution was collected and then the resin was washed with  $CH_2Cl_2$  (5x2 ml). The solution was evaporated under reduced pressure. The product was purified on Buchi LC auctomatic C18 column (Water + 0.1% TFA/MeOH) affording 90% of product **Ia** (HPLC purity 96%).

HRMS (TOF MS ES+): *m/z* calcd for C<sub>66</sub>H<sub>73</sub>N<sub>6</sub>O<sub>7</sub> [M+H]<sup>+</sup>: 1061.55352; found: 1061.5520 (-1.43 ppm).



Figure S1. HPLC chromatogram of the pure Ia.



Figure S2. <sup>1</sup>H-NMR in CDCl<sub>3.</sub>of the pure Ia.



Figure S3. LCMS spectrum of the crude Ia.





II.1.b. Linear meta-hexamer.**Ib** 



a) Swelling, CH<sub>2</sub>Cl<sub>2</sub>; b) 3-chloromethyl benzoic acid, DIPEA, CH<sub>2</sub>Cl<sub>2</sub>; c) Isopropyl amine or propargyl amine, dmso, 50°C; d) 3-chloromethyl benzoiylchloride, DIPEA, CH<sub>2</sub>Cl<sub>2</sub>;e) Cleavage: TFA/CH<sub>2</sub>Cl<sub>2</sub> (2:8).

Scheme S2. Synthesis of linear meta-arylopeptoid Ib

1-For 100 mg of resin, swelling: 2 ml of  $CH_2Cl_2$  at RT for 10 min

2- 3-chloromethylbenzoic acid (1.2 equiv. per mmol loading), DIPEA (5 equiv. per mmol loading) dissolved in 1 mL  $CH_2Cl_2$  were added at RT and shaken 40 minutes. This step is repeated. The liquid was drained off, then the resin was washed with  $CH_2Cl_2$  (5x2 ml), then with DMSO (5x2 ml).

3- Isopropyl amine or propargyl amine (20 equiv per mmol loading, 1M) dissolved in 0.5 mL of DMSO were added. The reaction was shaken for 1h at 50°C. The liquid was drained off, then the resin was washed with DMSO (5x2 ml), then with  $CH_2CL_2$  (5x2 ml).

4- 3-chloromethylbenzoylchloride (3 equiv. per mmol loading), DIPEA (6 equiv. per mmol loading) dissolved in 1 mL  $CH_2Cl_2$  were added at RT, shaken 10 minutes. The liquid was drawn down, then the resin was washed with  $CH_2Cl_2$  (5x2 ml), then with DMSO (5x2 ml).

5- Isopropyl amine or propargyl amine (20 equiv per mmol loading) dissolved in 0.5 mL of DMSO were added. The reaction was shaken for 1h at 50°C. The liquid was drained off, then the resin was washed with DMSO (5x2 ml), then with CH<sub>2</sub>CL<sub>2</sub> (5x2 ml).

Steps 4 and 5 were repeated to grow the targeted arylopeptoid oligomer until the expected sequence length.

6-The arylopeptoid was cleaved from the resin by the addition of 1 ml of  $CH_2Cl_2/TFA$  (8:2) for 30 min at RT. The solution was collected and then the resin was washed with  $CH_2Cl_2$  (5x2 ml). The solution was evaporated under reduced pressure. The product was purified on LC Buchi C18 column (water +0.1%TFA/MeOH) affording 93% of product **Ib** (HPLC purity 96%).



HRMS (TOF MS ES+): *m*/*z* calcd for C<sub>66</sub>H<sub>73</sub>N<sub>6</sub>O<sub>7</sub> [M+H]<sup>+</sup>: 1061.55352; found: 1061.5536 (0.06 ppm).

Figure S5. HPLC chromatogram of pure Ib.



Figure S6. <sup>1</sup>H-NMR spectrum in CDCl<sub>3</sub> of pure Ib.



Figure S7. LCMS spectrum of the crude Ib.





II.2. Cyclisation

II.2.a. ortho-cyclohexamer IIa.



Chemical Formula: C<sub>66</sub>H<sub>70</sub>N<sub>6</sub>O<sub>6</sub> Exact Mass: 1042,5357 Molecular Weight: 1043,3220

The ortho-hexamer **Ia** was dissolved in  $CH_2CI_2$  (5 mmol/L), then 5 equiv. of DIPEA was added followed by 1.2 equiv. of HATU. The reaction was stirred overnight at RT. The solvent was evaporated under reduced pressure and then the residue was evaporated with  $CH_2CI_2$  (2x20 ml). The residue is then

dissolved in EtOAc (20 ml), extracted with NaHCO<sub>3</sub> (2x10ml) then brine (1x10 ml). The organic layer was dried over MgSO<sub>4</sub>, filtered then evaporated under reduced pressure. The product was purified on C18 column (Water + 0.1% TFA/ MeOH) affording macrocycle **IIa** in 70% yield (HPLC purity 95%).





Figure S9. HPLC chromatogram of the pure *ortho*-macrocycle (IIa).



Figure S10. <sup>1</sup>H-NMR spectrum in CDCl<sub>3</sub> of pure *ortho*-macrocycle (IIa).



Figure S11. COSY spectrum in CDCl<sub>3</sub> of pure ortho-macrocycle (IIa).







Figure S13. IR spectra of ortho-macrocycle (IIa).

II.2.b. meta-cyclohexamer IIb.



Chemical Formula: C<sub>66</sub>H<sub>70</sub>N<sub>6</sub>O<sub>6</sub> Exact Mass: 1042,5357 Molecular Weight: 1043,3220

The *meta*-hexamer **Ib** was dissolved in  $CH_2CI_2$  (5 mmol/L), then 5 equiv. of DIPEA was added followed by 1.2 equiv. of HATU. The reaction was stirred overnight at RT. The solvent was evaporated under reduced pressure and then the residue was evaporated with  $CH_2CI_2$  (2x20 ml). The residue is then

dissolved in EtOAc (20 ml), extracted with NaHCO<sub>3</sub> (2x10 ml) then brine (1x10 ml). The organic layer was dried over MgSO<sub>4</sub>, filtered then evaporated under reduced pressure. The product was purified on C18 column (Water + 0.1% TFA/ MeOH) affording macrocycle **IIb** in 81% yield (HPLC purity 95%).

HRMS (TOF MS ES+): *m/z* calcd for C<sub>66</sub>H<sub>71</sub>N<sub>6</sub>O<sub>6</sub> [M+H]<sup>+</sup>: 1043.54296; found: 1043.5427 (-0.23 ppm).





Figure S14. HPLC of pure *meta*-macrocycle (IIb).



Figure S15. <sup>1</sup>H-NMR in CDCl<sub>3</sub> of pure *meta*-macrocycle (IIb).



Figure S16. COSY spectra in CDCl<sub>3</sub> of pure *meta*-macrocycle (IIb).



Figure S17. LCMS spectrum of the crude meta-macrocycle (IIb).



Figure S18. IR spectra of meta-macrocycle (IIb).

# II.3. CuAAC reactions

<u>High dilution procedure</u>: Macrocycle **IIa** or **IIb** (1.0 equiv.) was dissolved in methanol (C = 1.0 mM). 1,4bis(azidomethyl) benzene (1.1 equiv.) and catalyst **III** (10 mol-% per alkyne) were added. The reaction was stirred for 2 days at room temperature and the solvent was evaporated under reduced pressure.

<u>High concentration procedure for ortho series</u>: Macrocycle **IIa** (1.0 equiv.) was dissolved in methanol/ $CH_2Cl_2$  (1/1) at concentration of 0.1 M. 1,4-bis(azidomethyl) benzene (1.1 equiv.) and catalyst **III** (10 mol-% per alkyne) were added. The reaction was stirred for 2 days at room temperature and the solvent was evaporated under reduced pressure.

<u>High concentration procedure for meta series</u>: Macrocycle **IIb** was dissolved in methanol (C = 0.1 M). 1,4-bis(azidomethyl) benzene (1.1 equiv.) and catalyst **III** (10 mol-% per alkyne) were added. The reaction was stirred for 2 days at room temperature and the solvent was evaporated under reduced pressure.

#### II.3.a. Compound IVa.



High dilution procedure performed on 95 mg (0.09 mmol) of macrocycle IIa. HRMS of the crude indicates the presence of crown-like IVa as major compound and tube-like Va as traces. The product IVa was purified by column chromatography (EtOAc/MeOH 90:10 then  $CH_2Cl_2/MeOH$  95:5 to 70:30) followed by C18 purification (MeOH/H<sub>2</sub>O + 0.1%TFA) furnishing IVa (86 mg, 78%, 98% HPLC purity).

High concentration procedure performed on 50 mg (0.045 mmol) of **Ia**. HRMS of the crude also indicates the presence of crown-like **IVa** as major compound and tube-like **Va** as traces. The product **IVa** was purified by column chromatography (EtOAc/MeOH 90:10 then  $CH_2Cl_2/MeOH$  95:5 to 70:30) followed by C18 purification (MeOH/H<sub>2</sub>O + 0.1%TFA) furnishing **IVa** (40 mg, 76 %, 98% HPLC purity).



HRMS (TOF MS ES+): *m*/z calcd for C<sub>74</sub>H<sub>79</sub>N<sub>12</sub>O<sub>6</sub> [M+H]<sup>+</sup>: 1231.624; found: 1231.6248 (0.61 ppm)

Figure S19. HPLC of the pure ortho compound IVa.



Figure S20. LC-MS spectra of the pure ortho compound IVa.



**Figure S21.** <sup>1</sup>H-NMR in CDCl<sub>3</sub> of the pure *ortho* compound **IVa**.



**Figure S22.** COSY spectra in  $CDCI_3$  of the pure *ortho* compound **IVa**.



Figure S23. IR spectra of *ortho* compound IVa.

# II.3.b. Compounds IVb and Vb.



High dilution procedure performed on 200 mg (0.19 mmol) of macrocycle **IIb** furnishing 103 mg of cupola-like compound **IVb** (44 % yield, 98% HPLC purity) and 88 mg of tube-like compound **Vb** (37% yield, 98.5% HPLC purity).

High concentration procedure performed on 100 mg (0.1 mmol) of **IIb** furnishing 48 mg of cupola-like compound **IVb** (39 % yield, 98% purity) and 50 mg of tube-like compound **Vb** (41 % yield, 98% purity).

Purification procedure: The products were purified by column chromatography (EtOAc/MeOH 90:10 then  $CH_2Cl_2/MeOH$  95:5 to 70:30) affording mixture of cupola-like **IVb** and tube-like **Vb**. Further C18 purification (MeOH/H<sub>2</sub>O + 0.1%TFA) was performed for separation of **IVb** and **Vb**.

#### Compound IVb:



HRMS (TOF MS ES+): m/z calcd for  $C_{74}H_{79}N_{12}O_6$  [M+H]<sup>+</sup>: 1231.624; found: 1231.6257 (1.4 ppm).

Figure S24. HPLC chromatogram of the pure *meta* compound IVb.



Figure S25. LCMS spectrum of crude *meta* compound IVb.



**Figure S26.** <sup>1</sup>H-NMR spectra in CDCl<sub>3</sub> of the pure *meta* compound **IVb**.



**Figure S27.** COSY spectra in  $CDCI_3$  of the pure *meta* compound **IVb**.



#### Figure S28. Infra-red of *meta* compound IVb.

### Compound Vb:

# HRMS (TOF MS ES+): m/z calcd for $C_{148}H_{158}N_{24}O_{12}$ [M+2H]<sup>2+</sup>: 1231.624; found: 1231.6252 (1.01 ppm).



Figure S29. LCMS spectra of the pure *meta* compound Vb.



**Figure S30.** <sup>1</sup>H-NMR spectra in  $CDCl_3$  of the pure *meta* compound **Vb**.



Figure S31. COSY in  $CDCl_3$  of the pure *meta* compound Vb.



Figure S32. IR spectra of pure *meta* compound Vb.

### II.4. Triazolium formation

The methylation of cupola-like compounds IVa and IVb was performed in ace pressure tube in pure MeI at 70°C for 24 hours and produced the double methylated products VIa and VIb in quantitative yields.

II.4.a. ortho cupola-like bis-triazolium VIa.



HRMS (TOF MS ES+): *m/z* calcd for C<sub>76</sub>H<sub>84</sub>N<sub>12</sub>O<sub>6</sub> [M]<sup>2+</sup>: 630.33129; found: 630.3325 (1.95 ppm).



Figure S33. HPLC spectra of the crude ortho bis-triazolium compound VIa.



Figure S34. <sup>1</sup>H-NMR spectra in CDCl<sub>3</sub> of the crude *ortho* bis-triazolium compound VIa.







Figure S36. LCMS of the crude ortho bis-triazolium compound VIa.

II.4.b. Meta cupola-like triazolium VIb.



 $m_{crude}$  = 40 mg (HPLC purity 96%), isolated yield 98%

HRMS (TOF MS ES+): m/z calcd for  $C_{82}H_{90}N_{12}O_6$  [M]<sup>2+</sup>: 669.35477; found: 669.3538 (-1.41 ppm).



Figure S37. HPLC chromatogram of *meta* bis-triazolium compound VIb.



Figure S38. LCMS spectrum of meta bis-triazolium compound VIb.



Figure S39. <sup>1</sup>H-NMR in CDCl<sub>3</sub> of the pure *meta* bis-triazolium compound VIb.



Figure S40. IR spectra of *meta* bis-triazolium compound VIb.

# III-Conformational studies III.1. NMR study



Figure S41. Comparison of <sup>1</sup>H NMR spectra of IIa (red curve) and IIb (blue curve) in CDCl<sub>3</sub> at 298 K.



**Figure S42.** Variable temperature study of macrocycle **IIa** in DMSO (2.7 mg in 0.6 mL): 298 K (blue curve), 338 K (red curve), 358 K (green curve), 388 K (purple curve), 398 K (yellow curve), back to 298 K (orange curve).



**Figure S43.** VT study of macrocycle **IIa** in CDCl<sub>3</sub> (5 mg in 0.6 mL): 298 K (blue curve), 288K (red curve), 278 K (green curve), 268 K (purple curve), 265 K (yellow curve), back to 298 K (orange curve).



Figure S44. COSY experiment of *ortho* bicyclic compound IVa in CDCl<sub>3</sub> (8-10 mM) at 278 K.



**Figure S45.** Comparison of <sup>1</sup>H NMR spectra of *ortho* bicyclic compound **IVa** in  $CDCl_3$  at 298 K (green curve) and at 268 K (purple curve) and *meta* bicyclic compound **IVb** in  $CDCl_3$  at 298 K (blue curve) and at 268 K (red curve).



**Figure S46.** Comparison of <sup>1</sup>H NMR spectra of the *ortho* bis-triazolium bicyclic compound **VIa** at 298 K in various solvants: CDCl<sub>3</sub> (blue curve), CD<sub>3</sub>CN (red curve), CD<sub>3</sub>OD (green curve) and DMSO-d<sub>6</sub> (purple curve).



**Figure S47.** COSY NMR spectra of the *ortho* bis-triazolium bicyclic compound **VIa** (2 mM) in  $CD_3CN$  at 288K.



**Figure S48.** TOCSY NMR spectra of the *ortho* bis-triazolium bicyclic compound **VIa** (20 mM) in  $CD_3CN$  at 298K.



**Figure S49.** NOESY NMR spectra of the *ortho* bis-triazolium bicyclic compound **VIa** (2 mM) in CD<sub>3</sub>CN at 288K (d8=500).



**Figure S50.** NOESY NMR spectra (3-9.5 ppm region) of the *ortho* bis-triazolium bicyclic compound **VIa** (2 mM) in CD<sub>3</sub>CN at 288K (d8=500).



**Figure S51.** HSQC NMR spectra of the *ortho* bis-triazolium bicyclic compound **VIa** (20 mM) in  $CD_3CN$  at 298K.



**Figure S52.** HMBC NMR spectra of the *ortho* bis-triazolium bicyclic compound **VIa** (20 mM) in  $CD_3CN$  at 298K.



Protons number	На (СН)	Hb (CH₃)	b′ (CH₃)	c,c' ABsystem (CH <sub>2</sub> )
Residue 1	3.87 (hept <i>, J</i> = 7.5 Hz	-0.14 (d <i>, J</i> = 7.5 Hz)	1.03 (d <i>, J</i> = 7.5Hz)	4.12 (d, J = 15.9 Hz)
				4.53 (d <i>, J</i> = 15.9 Hz)
Residue 2	3.72 (hept <i>, J</i> = 6.5 Hz)	0.15 (d <i>, J</i> = 6.5 Hz)	0.53 (d <i>, J</i> = 6.5 Hz)	4.00 (d, J = 17.7 Hz)
				4.59 (d <i>, J</i> = 17.7 Hz)
Residue 3	3.66 (hept <i>, J</i> = 6.5 Hz)	0.87 (d <i>, J</i> = 6.5 Hz)	1.23 (d <i>, J</i> = 6.5 Hz)	4.16 (d, J = 16,0 Hz)
				5.34(d, <i>J</i> = 16.0 Hz)
Residue 4	4.06 (m)	1.38 (d <i>, J</i> = 6.8 Hz)	1.52 (d <i>, J</i> = 6.4 Hz)	4.23 (d, J = 14.9 Hz)
				5.07(d, J = 14.9 Hz)
	d,d' ABsystem (CH <sub>2</sub> )	e,e' ABsystem (CH <sub>2</sub> )	f,f' ABsystem (CH <sub>2</sub> )	g,g' ABsystem (CH <sub>2</sub> ) (CH <sub>2</sub> )
Residue 5	4.07 (d <i>, J</i> = 14 Hz)	4.48 (d <i>, J</i> = 18.6 Hz)		
	6.03 (d <i>, J</i> = 14 Hz)	4.97 (d, J = 18.6 Hz)		
Residue 6			4.16 (d <i>, J</i> = 18.5 Hz)	4.33 (d, J = 17.5 Hz)
			4.82 (d <i>, J</i> = 18.5 Hz)	5.77 (d <i>, J</i> = 17.5 Hz)
Triazolium	Hh (CH)	Hi (CH₃)	Нј (СН)	Hk(CH₃)
Triazolium 1	8.38 (s)	3.47 (s)		
Triazolium 2			9.21 (s)	4.14 (s)
Linker	I,I' ABsystem (CH <sub>2</sub> )	m,m' ABsystem (CH <sub>2</sub> )		
	5.35 (d <i>, J</i> = 14 Hz)	5.58 (d, J = 14.5 Hz)		
	5.69 (d, J = 14 Hz)	5.92 (d, J = 14.5 Hz)		

Table S1. H<sup>1</sup> NMR signals attribution of bis-triazolium bicyclic compound VIa in CD<sub>3</sub>CN at 298K.



**Figure S53.** Variable temperature NMR spectra of the *ortho* bis-triazolium bicyclic compound **VIa** (2 mM) in CD3CN: 298K (blue curve), 288K (red curve) and 273K (green curve).



**Figure S54.** Variable temperature NMR of the *ortho* bis-triazolium bicyclic compound **VIa** (3 mM) in DMSO-d<sub>6</sub>: 298K (blue curve), 338K (red curve), 358K (green curve), 378K (purple curve), 388K (Yellow curve) and back to 298K (orange curve).



**Figure S55.** Variable temperature NMR of the *meta* bis-triazolium bicyclic compound **VIb** (3 mM) in DMSO-d<sub>6</sub>: 298 K (blue curve), 358 K (red curve), 388 K (green curve) and back to 298 K (purple curve).

# III.2. Computational study



**Figure S56.** Model structure of *ortho*-arylopeptoid macrocycle **IIa** with all amides in the trans conformation and the two propargyl groups on the same side of the ring.

<u>Computational study of bicyclic bis-triazolium</u> **VIa** using Conformer-Rotamer Ensemble Sampling Tool (CREST)<sup>1</sup>

We first built a possible structure, without a priori, of the bicyclic system with Chimera software. This structure has been geometrically optimized with Gaussian 16<sup>2</sup> at DFT level of theory, using the B3LYPas a hybrid functional and 6-31g as a basis for the description of the electrons and acetonitrile as solvent in SCRF model (Table S2).<sup>3,4</sup> It was not necessary to do an optimization with a more precise electronic basis because only a geometrically valid starting system was needed. the rest of the conformational research being done by semi-empirical methods. CREST program is used using the conformer search module of ChimeraX software.<sup>5</sup> The charge is set to +2 to respect the two triazoliums and the six amide bonds are constrained in the *trans* conformation. The APLB implicit solvent model was used with the dielectric constant of acetonitrile (37,5). Energy threshold for energy and RMSD was set by default to 6 kcal and 0.125. There is no protonation screening. We used the method GFN2-xTB<sup>6</sup> with no empirical dispersion and with the default integration grid. 198 solutions proposed by CREST have energy in a 6 kcal window (Fig. S57). Structures of the 10 lowest energy solutions are represented in Figure 58.



Atom	X	Y	Z
Ν	5.31900	-1.61200	0.04500
С	4.43800	-2.80500	0.03800
Н	3.39500	-2.46700	0.04600
Н	4.60100	-3.34300	0.98100
С	6.20500	-1.37600	1.22800
Η	6.66700	-0.39400	1.06100
С	7.32600	-2.42000	1.34800

Н	6.93800	-3.41100	1.60800
Н	8.01700	-2.11500	2.14100
Н	7.90000	-2.50400	0.42000
С	5.37600	-1.29500	2.52200
Н	4.87000	-2.24100	2.74500
Н	4.61400	-0.51200	2.45100
Н	6.03200	-1.06300	3.36800
С	4.66100	-3.77800	-1.11500
C	3.59000	-4.54000	-1.63300
C	3.83500	-5.53600	-2.59600
Н	3.00700	-6.13100	-2.97400
С	5.12500	-5.76100	-3.08000
Н	5.30000	-6.53300	-3.82300
С	6.18300	-4.97900	-2.60300
Н	7.19000	-5.13200	-2.98000
С	5.94700	-4.00400	-1.62900
Н	6.77700	-3.40900	-1.26300
C	2.19400	-4.37400	-1.10300
0	1.96000	-4.59700	0.12000
N	1.17800	-4.06000	-1.97000
C	-0.18300	-3.94100	-1.42000
Н	-0.10200	-3.50100	-0.41400
Н	-0.74000	-3.23000	-2.03900
C	1.37100	-3.66100	-3.40200
Н	2.43400	-3.82200	-3.62000
С	0.53800	-4.55000	-4.33400
Н	-0.53800	-4.42300	-4.16300
Н	0.78400	-5.60900	-4.20500
Н	0.74300	-4.27600	-5.37500
С	1.07300	-2.16500	-3.60100
Н	0.03000	-1.91600	-3.37700
Н	1.25300	-1.89900	-4.64900
Н	1.73300	-1.55500	-2.97800
С	-0.98300	-5.23700	-1.34300
С	-2.36200	-5.17400	-1.03200
С	-3.12500	-6.35200	-0.96300
Н	-4.19100	-6.29300	-0.75700
С	-2.52800	-7.59700	-1.17800
Н	-3.12600	-8.50100	-1.12800
С	-1.15900	-7.66600	-1.45900
Н	-0.68300	-8.62900	-1.61900
С	-0.39800	-6.49400	-1.53800
Н	0.66100	-6.56100	-1.76400
С	-3.03700	-3.83900	-0.89000
0	-3.10300	-3.05600	-1.86900
Ν	-3.57900	-3.46500	0.32600
С	-4.23400	-2.13800	0.40200
Н	-3.63100	-1.44000	-0.19100
Н	-4.19100	-1.80300	1.44700
С	-3.51700	-4.28400	1.55200
Н	-3.28000	-5.31400	1.27500
Н	-4.50900	-4.30800	2.02500

С	-5.67500	-2.10000	-0.09100
С	-6.21700	-0.88500	-0.56800
Н	-7.95900	0.09800	-1.37900
С	-7.55500	-0.83400	-0.99300
С	-8.36500	-1.97100	-0.93700
Н	-9.39700	-1.91900	-1.27000
С	-7.83400	-3.17400	-0.46000
Н	-8.45200	-4.06600	-0.41600
С	-6.50100	-3.23200	-0.04000
Н	-6.10100	-4.17700	0.31200
С	-5.34800	0.32900	-0.78200
0	-4.60300	0.38100	-1.79900
N	-5.37500	1.36300	0.12000
С	-4.45600	2.48800	-0.13600
Н	-4.23900	2.98900	0.81600
Н	-3.50800	2.05800	-0.49700
C	-6.15400	1.33600	1.39500
Н	-6.71300	0.39100	1.38000
C	-5.21900	1.31300	2.61800
H	-4.70800	2.27100	2.76500
Н	-4.46000	0.52900	2.52200
Н	-5.80600	1.11300	3.52100
C	-7.16100	2,49400	1.47100
Н	-7.88200	2.45100	0.64900
Н	-6.66000	3.46800	1.44200
Н	-7.71600	2.43100	2.41400
C	-4.93100	3.52600	-1.14600
C	-4.05200	4.57600	-1.49500
C	-4.47000	5.58500	-2.37600
Н	-3,79100	6.39800	-2.62200
C	-5.74500	5.54000	-2.94900
Н	-6.06000	6.32300	-3.63300
С	-6.60100	4.47800	-2.64400
Н	-7.58600	4.42200	-3.09800
С	-6.19300	3.48100	-1.74900
Н	-6.86600	2.66500	-1.51200
С	-2.68800	4.65200	-0.85900
0	-2.56800	5.12100	0.30800
Ν	-1.59500	4.20000	-1.55500
С	-0.29800	4.17400	-0.84500
Н	0.21200	3.24600	-1.11800
Н	-0.52700	4.13300	0.22900
С	-1.65100	3.65400	-2.95100
Н	-2.68200	3.80600	-3.28900
С	-1.36100	2.14400	-2.97600
Н	-0.34500	1.91800	-2.63400
Н	-1.44600	1.78000	-4.00600
Н	-2.08100	1.58800	-2.36500
С	-0.71800	4.42800	-3.89900
Н	-0.90900	4.10900	-4.93000
Н	0.33600	4.23000	-3.67400
Н	-0.89400	5.50600	-3.83600

С	0.59600	5.37500	-1.11800
С	1.98100	5.21900	-1.34100
С	2.78500	6.34500	-1.59800
Н	3.84100	6.21700	-1.82000
С	2.23000	7.62800	-1.59800
Н	2.85800	8.48900	-1.80400
С	0.86400	7.79100	-1.33800
Н	0.42600	8.78500	-1.32500
С	0.05900	6.67300	-1.10300
Н	-1.00000	6.80200	-0.89900
С	2.57400	3.84300	-1.43900
0	2.27400	3.09200	-2.39900
N	3.42100	3.38700	-0.44500
С	3.92200	2.00100	-0.55900
H	3.75100	1.49700	0.40200
Н	3.29200	1.49200	-1.29900
C	3.87300	4,19500	0.70600
H	4.94400	3,99600	0.86000
H	3.79200	5.25900	0.46600
C	5.38300	1.85400	-0.97000
C	5.94500	0.55600	-1.03900
Н	7.69900	-0.60800	-1.51000
C	7.28200	0.39200	-1.43500
C	8.07200	1.50000	-1.75700
Н	9.10400	1.35900	-2.06300
C	7.51700	2.78100	-1.70100
Н	8.11500	3.64900	-1.96200
С	6.18200	2.95200	-1.31300
Н	5 76200	3 95200	-1 29800
С	5 07100	-0.66400	-0.91200
0	4.13100	-0.81700	-1.74000
C	1 98100	-0.09800	5 25600
C	1.50900	1.22200	5.21500
C	0.12600	1.46300	5.21500
C	-0.77300	0.39600	5.28700
C	-0.30000	-0.92400	5.35800
C	1.08200	-1.16600	5.32700
C	2.48800	2.37400	5.27100
C	-1.27000	-2.06700	5.56000
N	-1.56000	-2.84300	4.32300
N	2.59400	3.13500	3.99600
C	-2.71300	-2.85300	3.58700
C	-2.53100	-3.79700	2.58300
N	-1.24400	-4.27200	2.77400
N	-0.64900	-3.70200	3.83600
N	2.26500	4.74100	2.63900
N	1.90100	4.27400	3.84600
С	3.38200	2.87000	2.91000
С	3.17700	3.91100	2.01100
H	3.05000	-0.29500	5.26000
Н	-0.25100	2.48200	5.18800
Н	-1.84000	0.59400	5.31500
	1		

Н	1.45900	-2.18300	5.38500
Н	2.19100	3.10700	6.03200
Н	3.50000	2.02700	5.51000
Н	-2.24100	-1.70600	5.91900
Н	-0.88200	-2.78600	6.29200
Н	-3.57300	-2.25700	3.84600
Н	4.04200	2.01700	2.86000
С	-0.49100	-5.29500	2.02400
Н	-1.18300	-5.86500	1.40500
Н	0.26600	-4.81900	1.38600
Н	-0.01000	-5.96500	2.74100
С	1.69500	6.02800	2.19900
Н	2.24300	6.39000	1.32800
Н	0.64200	5.89100	1.93900
Н	1.78800	6.75000	3.01500

**Table S2.** Starting point for CREST: Representation (H atom are removed for clarity) and xyz file of the optimized structure of bicyclic bis-triazolium **VIa**.



Figure S57. Energies of the 198 conformers proposed by CREST



Figure S58. Structures of the 10 lowest energy solutions obtained by CREST

IV-Binding NMR study



**Figure S59.** <sup>1</sup>H NMR of the *ortho* bis-triazole bicyclic compound **IVa** (3 mM) in CD<sub>3</sub>CN at 298 K (blue curve), upon addition of TBAHSO<sub>4</sub>:  $\approx$  0.25 equiv. (red curve),  $\approx$  0.70 equiv. (green curve),  $\approx$  1.05 equiv. (purple curve) and  $\approx$  6.0 equiv. (yellow curve).



**Figure S60.** <sup>1</sup>H NMR of the *ortho* bis-triazole bicyclic compound **IVa** (3 mM) in CDCl<sub>3</sub> at 298 K (blue curve), upon addition of TBAHSO<sub>4</sub>:  $\approx$  0.5 equiv. (red curve),  $\approx$  2.5 equiv. (green curve) and  $\approx$  9 equiv. (purple curve).



**Figure S61.** <sup>1</sup>H NMR of the *ortho* bis-triazolium bicyclic compound **VIa** in CD<sub>3</sub>CN at 298 K (blue curve), upon addition of TBAHSO<sub>4</sub>:  $\approx$  1.5 equiv. (red curve),  $\approx$  3 equiv. (green curve),  $\approx$  5.3 equiv. (purple curve),  $\approx$  6 equiv. (yellow curve) and  $\approx$  20 equiv. (orange curve).



**Figure S62.** <sup>1</sup>H NMR of the *ortho* bis-triazolium bicyclic compound **VIa** (3 mM) in  $CDCI_3$  at 298 K (blue curve), upon addition of  $TBAHSO_4$ :  $\approx 1.9$  equiv. (red curve),  $\approx 2.4$  equiv. (green curve),  $\approx 5$  equiv. (purple curve),  $\approx 6.5$  equiv. (yellow curve) and  $\approx 8$  equiv. (orange curve).



**Figure S63.** COSY NMR spectra of the *ortho* bis-triazolium bicyclic compound **VIa** + 2.25 equiv. TBAHSO<sub>4</sub> in CDCl<sub>3</sub> (3 mM) at 298K.



**Figure S64.** HSQC NMR spectra of the *ortho* bis-triazolium bicyclic compound **VIa** + 2.25 equiv. TBAHSO<sub>4</sub> in CDCl<sub>3</sub> (3 mM) at 298K.



**Figure S65.** HMBC NMR spectra of the *ortho* bis-triazolium bicyclic compound **VIa** + 2.25 equiv. TBAHSO<sub>4</sub> in  $CDCI_3$  (3 mM) at 298K.



Protons number	Ha (CH)	Hb (CH₃)	b' (CH <sub>3</sub> )	c,c' ABsystem (CH <sub>2</sub> )
Residue 1,3	3.55 (hept, <i>J</i> = 6.6 Hz	0.67 (d <i>, J</i> = 6.6 Hz)	1.23 (d <i>, J</i> = 6.6Hz)	4.63 (d, J = 17.3 Hz)
				5.52 (d <i>, J</i> = 17.3 Hz)
Residue 2,4	4.08 (hept, <i>J</i> = 6.6 Hz)	0.88 (d <i>, J</i> = 6.6 Hz)	1.25 (d <i>, J</i> = 6.6 Hz)	3.88 (d, J = 17.1 Hz)
				5.45 (d <i>, J</i> = 17.1 Hz)
	d,d' ABsystem (CH <sub>2</sub> )	e,e' ABsystem (CH <sub>2</sub> )		
Residue 5,6	4.85 (d, J = 14.9 Hz)	4.39 (d <i>, J</i> = 15.5 Hz)		
	5.12 (d <i>, J</i> = 14.9 Hz)	5.17 (d <i>, J</i> = 15.5 Hz)		
Triazolium	Hh (CH)	Hi (CH₃)	Нј (СН)	Hk(CH₃)
Triazolium 1,2	9.21 (s)	4.46 (s)		
Linker	I,I' ABsystem (CH <sub>2</sub> )			
	5.25 (d, J = 15.2 Hz)			
	5.92 (d, J = 15.2 Hz)			

**Table S3.** H<sup>1</sup> NMR signals attribution of bicyclic bis-triazolium compound **VIa** + TBAHSO<sub>4</sub> in CD<sub>3</sub>CN at 298K.

# V. References:

- 1- P. Pracht, F. Bohle and S. Grimme, Physical Chemistry Chemical Physics, 2020, 22, 7169.
- 2- Gaussian 16, Revision C.01, M. J. Frisch, G. W. Trucks, H. B. Schlegel, G. E. Scuseria, M. A. Robb, J. R. Cheeseman, G. Scalmani, V. Barone, G. A. Petersson, H. Nakatsuji, X. Li, M. Caricato, A. V. Marenich, J. Bloino, B. G. Janesko, R. Gomperts, B. Mennucci, H. P. Hratchian, J. V. Ortiz, A. F. Izmaylov, J. L. Sonnenberg, D. Williams-Young, F. Ding, F. Lipparini, F. Egidi, J. Goings, B. Peng, A. Petrone, T. Henderson, D. Ranasinghe, V. G. Zakrzewski, J. Gao, N. Rega, G. Zheng, W. Liang, M. Hada, M. Ehara, K. Toyota, R. Fukuda, J. Hasegawa, M. Ishida, T. Nakajima, Y. Honda, O. Kitao, H. Nakai, T. Vreven, K. Throssell, J. A. Montgomery, Jr., J. E. Peralta, F. Ogliaro, M. J. Bearpark, J. J. Heyd, E. N. Brothers, K. N. Kudin, V. N. Staroverov, T. A. Keith, R. Kobayashi, J. Normand, K. Raghavachari, A. P. Rendell, J. C. Burant, S. S. Iyengar, J. Tomasi, M. Cossi, J. M. Millam, M. Klene, C. Adamo, R. Cammi, J. W. Ochterski, R. L. Martin, K. Morokuma, O. Farkas, J. B. Foresman and D. J. Fox, Gaussian, Inc., Wallingford CT, 2016.
- 3- A. D. Becke, "Density-functional thermochemistry. III. The role of exact exchange," J. Chem. *Phys.*, **98** (1993) 5648-52.
- 4- V. A. Rassolov, M. A. Ratner, J. A. Pople, P. C. Redfern and L. A. Curtiss, "6-31G\* Basis Set for Third-Row Atoms," *J. Comp. Chem.*, **22** (2001) 976-84.
- Pettersen, E. F.; Goddard, T. D.; Huang, C. C.; Meng, E. C.; Couch, G. S.; Croll, T. I.; Morris, J. H.; Ferrin, T. E. UCSF ChimeraX: Structure visualization for researchers, educators, and developers. *Protein Science* 2021, **30**, 70–82.
- 6- Grimme, S.; Bannwarth, C.; Shushkov, P. A robust and accurate tight-binding quantum chemical method for structures, vibrational frequencies, and noncovalent interactions of large molecular systems parametrized for all spd-block elements (Z= 1–86). *Journal of chemical theory and computation* 2017, **13**, 1989–2009.