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Preparation of new chiral diene ligands and Their Use in the Rhodium Catalyzed Asymmetric Annulation of 2-Formylarylboronic acid with unactivated Alkynes

Yu Sun, Yuanmeng Li, Xiuqi Wang, Xu Cao, Hongping Deng, Xiaoli Bu, Mengtao Ma and Fei Xue

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1. General information

All anaerobic manipulations were carried out with standard Schlenk techniques under nitrogen gas. Analytical thin-layer chromatography (TLC) was performed on silica gel, irradiation with UV light. For column chromatography, 200-300 mesh silica gel was used. NMR spectra were recorded on a BRUKER 600 MHz (or 400 Hz) spectrometer and ¹³C NMR spectra were recorded in CDCl3 at 151 MHz (or 101 Hz). ¹H and ¹³C NMR were referenced to CDCl3 at δ 7.26 and 77.00 respectively. The following abbreviations were used to explain the multiplicities; s (singlet), bs (broad singlet), d (doublet), t (triplet), m (multiplet); coupling constants (J) are in Hertz (Hz). Optical rotations were measured on an Anton Paar MCP 100 automatic polarimeter. HRMS spectra were obtained by BRUKER microTOF-Q III instrument with ESI source. Enantiomeric excesses (ee) were determined by HPLC analysis on SHIMADZU HPLC system with Daicel chiral columns. GPC (gel permeation chromatography) was performed using LaboACE LC-5060 recycling preparative HPLC.

2. Materials

Unless otherwise noted, all the solvents and commercially available reagents were used as received for the reactions without further purification. The solvent and chemicals were purchased from Energy Chemical or J&K Scientific. Preparation of all these starting materials, $2c-2e^{-[1-3]}$ were synthesized according the reported references and they were assigned by comparison of their NMR spectra with the reported data. Chiral diene ligands, L5*-L13*^[4-8] were prepared according to the reported procedures.



3. Preparation of new chiral diene ligands

Scheme S1

Ligand (R,R)-IIa: To a solution of (1R,4R)-5-phenylbicyclo[2.2.2]octa-2,5-dien-2-yl trifluoromethanesulfonate⁹ (Ia, 1.32 g, 4 mmol), and Pd(OAc)₂ (22.4 mg, 0.10 mmol), and PPh3 (26.2 mg, 0.10 mmol) in DMF (10 mL) were added MeOH (0.50 mL, 12 mmol) and N-ethyldiisopropylamine (0.86 mL, 4.8 mmol). Further, the system was degassed with CO gas for 10 minnuts, and the mixture was stirred at room temperature for 5 h under CO atmosphere (1 atm). Then the reaction was quenched with saturated ammonium chloride aqueous solution and extracted with ether for three times, the combined organic layer was washed with 1 brine, dried over Na₂SO₄, filtered, and concentrated on a rotary evaporator. The crude product was purified by silica gel column chromatography using hexane/ethyl acetate (4/1) to afford the corresponding product (*R*,*R*)-IIa (1.10 g, 87% yield).



Ligand (*R*,*R*)-**IIa**, $[\alpha]^{25}{}_{D}$ = -38.5 (c 2.22, CHCl₃), ¹H NMR (CDCl₃, 400 MHz) δ 1.49-1.57 (m, 4H), 3.75 (s, 3H), 4.25-4.31 (m, 2H), 6.58 (dd, J = 1.9, 6.4Hz, 1H), 7.23-7.26 (m, 1H), 7.30-7.34 (m, 2H), 7.37-7.41 (m, 3H). ¹³C NMR (CDCl₃, 151 MHz) δ 24.6, 25.2, 37.2, 40.5, 51.6, 124.9, 127.1, 128.6, 129.4, 137.7, 139.4, 145.2, 145.6,

165.3.HRMS (ESI) Calcd for $C_{16}H_{17}O_2$ [M+H]⁺241.1223, found 241.1228.

Ligand I: To a solution of LDA (22.0 mmol) in THF (25 ml), freshly prepared from diisopropylamine (2.89 mL, 22.0 mmol) and n-butyllithium (2.5 M in hexane, 8.8 mL, 22.0 mmol) at 0 °C under N₂ for 10 minutes. The resulting mixture was cooled to -78 °C. A solution of 2-cyclohexenone (1.92 g, 20.0 mmol) in THF (25 mL) was added dropwise, and stirred for another 30 minutes at -78 °C. Then A solution of Tf₂NPh (7.15 g, 20.0 mmol) dissolved in THF (30 mL) was slowly added for 10 minutes. After that, the reaction mixture was warmed to 0 °C and stirred at 0 °C until the reaction was judged complete according to TLC. The solvent was evaporated, quenched by saturated ammonium chloride aqueous solution, and extracted with ether for three times, the combined organic layer was dried over MgSO4, and concentrated under vacuum to give a colorless oil, which directly used in copper-catalyzed cross-coupling reaction.

CuI (20 mg, 0.1 mmol) was added to a solution of the above dienyl triflate in THF (10 mL) at 0 °C under nitrogen. Grignard reagent (30 mmol) was then added slowly by syringe for 20 minutes. The mixture was stirred for a further 30 minutes. Then the reaction was quenched with saturated ammonium chloride aqueous solution and extracted with ether for three times, the combined organic layer was dried over MgSO4, filtered, dried over Na₂SO₄, and concentrated under vacuum. Considering the stability of the substituted cyclohexadiene products, which was directly used for synthesis of **Ib-Ie** without further purification.

To a solution of the above crude substituted cyclohexadiene, 2-naphthyl propiolate (3.5 g, 17.87 mmol) in CH₂Cl₂ (60 mL) and BHT (10.0 mg) was slowly added Me₂AlCl (1.0 M in heptane, 20.0 mL, 20.0 mmol) at -78 °C, and the mixture was warmed to room temperature and stirred for 16 h. Then the solution was carefully poured into a vigorously stirred, ice-cooled aqueous solution of 1N HCl (70 mL). Then the resulting solution was extracted with CH₂Cl₂ (50 mL) for three times, the combined organic layer was dried over MgSO₄, filtered, and concentrated under vacuum. Purification of the residue by column chromatography on silica gel with hexane/EtOAc (20/1~5/1) gave corresponding ligands *dl*-I.



Ligand *dl*-**Ib**, (4.2 g, 65% yield), ¹H NMR (CDCl₃, 400 MHz) δ 1.09-1.25 (m, 3H), 1.26-1.39 (m, 2H), 1.39-1.48 (m, 4H), 1.71-1.84 (m, 5H), 2.03-2.09 (m, 1H), 3.75-3.77 (m, 1H), 4.27 (dd, J = 2.1, 6.3 Hz, 1H), 5.97 (d, J = 6.3 Hz, 1H), 7.29 (dd, J = 2.3, 8.6 Hz, 1H), 7.45-7.51 (m, 1H), 7.51 (m, 1

2H), 7.59-7.62 (m, 2H), 7.80-7.82 (m, 1H), 7.84-7.88 (m, 2H). ¹³C NMR (CDCl₃, 101 MHz) δ 24.7, 25.6, 26.4, 31.4, 31.5, 36.7, 41.3, 42.0, 118.7, 121.5, 124.6, 125.5, 126.4, 127.6, 127.8, 129.3, 131.4, 133.9, 139.4, 148.2, 148.8, 151.6, 163.5. HRMS (ESI) Calcd for C₂₅H₂₇O₂ [M+H]⁺ 59.2006, found 359.2006.



Ligand *dl*-Ic, (5.1 g, 77% yield). ¹H NMR (CDCl₃, 400 MHz) δ 1.53-1.74 (m, 4H), 2.36 (s, 3H), 4.04-4.06 (m, 1H), 4.49-4.51 (m, 1H), 6.40 (dd, J = 1.8, 6.3 Hz, 1H), 7.16-7.18 (m, 1H), 7.21-7.26 (m, 3H), 7.36 (dd, J = 2.3, 8.8 Hz, 1H), 7.49-7.56 (m, 2H), 7.69 (d, J = 2.2 Hz, 1H),

7.81 (dd, J = 1.9, 6.4 Hz, 1H), 7.85-7.87 (m, 1H), 7.88-7.93 (m, 2H). ¹³C NMR (CDCl₃, 101 MHz) δ 20.8, 24.5, 25.4, 37.1, 43.9, 118.7, 121.5, 125.7, 125.8, 126.6, 127.3, 127.7, 127.8, 128.4, 129.4, 130.5, 131.5, 131.5, 13.3.9, 135.6, 139.1, 139.3, 146.3, 147.9, 148.7, 163.5. HRMS (ESI) Calcd for C₂₆H₂₃O₂ [M+H]⁺ 367.1703, found 367.1693. Resolution of *dl*-I**c** was carried out by use of a chiral stationary phase column [Chiral IG (2.0 cm I.D. × 25 cm), hexane/CH₂Cl₂/ IPA= 89:10/1 to give both enantiomers (*S*,*S*)-I**c** and (*R*,*R*)-I**c**, quantitatively. *t*₁ = 86 min for (*S*,*S*)-I**c**, *t*₂ = 97 min for (*R*,*R*)-I**c**. [α]²⁵_D = +78.1 (c 0.93, CHCl₃) for (*R*,*R*)-I**c**.



Ligand *dl*-**Id** (5.0 g, 71% yield). ¹H NMR (CDCl₃, 400 MHz) δ 1.1 (d, J = 6.8 Hz, 3H), 1.31 (d, J = 6.8 Hz, 3H), 1.51-1.72 (m, 4H), 3.01-3.07 (m, 1H), 3.95 (dd, J = 1.8, 6.3 Hz, 1H), 4.46 (dd, J = 1.9, 6.1 Hz, 1H), 6.31 (dd, J = 1.5, 6.3 Hz, 1H), 7.07 (d, J = 7.6 Hz, 1H), 7.16-7.20 (m, 1H), 7.29-7.36 (m, 3H), 7.47-7.53 (m, 2H),

7.67 (d, J = 1.6 Hz, 1H), 7.76-7.78 (m, 1H), 7.83-7.85 (m, 1H), 7.87-7.91 (m, 2H). ¹³C NMR (CDCl₃, 101 MHz) δ 24.1, 24.4, 24.5, 25.3, 29.9, 37.1, 44.8, 118.7, 121.5, 125.5, 125.6, 125.6, 126.5, 127.7, 127.8, 127.8, 128.7, 129.3, 130.8, 131.4, 133.9, 138.6, 138.9, 146.2, 146.7, 147.7,148.7, 163.5. HRMS (ESI) Calcd for C₂₈H₂₇O₂ [M+H]⁺ 395.2006, found 395.2015. Resolution of *dl*- Id was carried out by use of a chiral stationary phase column [Chiral IG (2.0 cm I.D. × 25 cm), hexane/CH₂Cl₂/ IPA= 89:10/1 to give both enantiomers (*S*,*S*)-Id and (*R*,*R*)-Id, quantitatively. t_1 = 41 min for (*S*,*S*)-Id, t_2 = 49 min for (*R*,*R*)-Id. [α]²⁵_D = +93.5 (c 0.92, CHCl₃) for (*R*,*R*)-Id.



Ligand *dl*-**Ie** (6.2 g, 87% yield). ¹H NMR (CDCl₃, 400 MHz) δ 1.57-1.82 (m, 4H), 4.16-4.19 (m, 1H), 4.57-4.59 (m, 1H), 6.60 (dd, J = 1.8, 6.3 Hz, 1H), 7.34 (dd, J = 1.1, 7.0 Hz, 1H), 7.39 (dd, J = 2.3, 8.8 Hz, 2H), 7.46-7.55 (m, 5H), 7.71 (d, J = 2.2 Hz, 1H),

7.82-7.95 (m, 7H). ¹³C NMR (CDCl₃, 101 MHz) δ 24.8, 25.5, 37.4, 44.8, 118.8, 121.5, 125.2, 125.4, 125.7, 125.7, 125.9, 126.1, 126.6, 127.7, 127.8, 128.5, 129.4, 131.5, 131.5, 132.6, 133.9, 137.6, 139.3, 145.4, 147.8, 148.7, 163.5. HRMS (ESI) Calcd for C₂₉H₂₃O₂ [M+H]⁺ 403.1693, found 403.1695. Resolution of *dl*-**Ie** was carried out by use of a chiral stationary phase column [Chiral IG (2.0 cm I.D.×25 cm), hexane/CH₂Cl₂/ IPA= 89:10/1 to give both enantiomers (*S,S*)-

Ie and (*R*,*R*)-Ie, quantitatively. $t_1 = 41 \text{ min for } (S,S)$ -Ie, $t_2 = 53 \text{ min for } (R,R)$ -Ie. ([α]²⁵ _D+125.7 (c 0.69, CHCl₃) for (*R*,*R*)-Ie.

Ligand II: A solution of the corresponding (R,R)-I or dl-Ic (2.0 mmol) and NaOMe (216 mg, 4.0 mmol) in MeOH (8 mL) was stirred at room temperature for 24 h. The reaction mixture was concentrated under vacuum. Purification of the residue by column chromatography on silica gel with hexane/EtOAc (10/1) gave corresponding ligands II.



Compound *dl*-**IIb** (467.4 mg, 95% yield), ¹H NMR (CDCl₃, 400 MHz) δ 1.06-1.15 (m, 3H), 1.20-1.34 (m, 6H), 1.64-1.75 (m, 5H), 1.94-2.00 (m, 1H), 3.62-3.64 (m, 1H), 3.71 (s, 3H), 4.08 (dd, J = 1.8, 6.3Hz, 1H), 5.83-5.86 (m, 1H), 7.26 (dd, J = 2.1, 6.5Hz, 1H). ¹³C NMR (CDCl₃, 101 MHz) δ 24.6, 25.4, 26.3, 31.3, 31.4, 36.5,

41.0, 41.9, 51.4, 124.4, 139.8, 145.9, 151.7, 165.5. HRMS (ESI) Calcd for $C_{16}H_{23}O_2$ [M+H]⁺ 247.1693, found 247.1701.



Ligand (*R*,*R*)-**IIc** (467.3 mg, 92% yield), $[\alpha]^{25}_{D} = +61.0$ (c 1.10, CHCl₃). ¹H NMR (CDCl₃, 400 MHz) δ 1.39-1.63 (m, 4H), 2.26 (s, 3H), 3.79 (s, 3H), 3.89-3.92 (m, 1H), 4.30-4.33 (m, 1H), 6.26-6.28 (m, 1H), 7.06-7.11 (m, 1H), 7.13-7.19 (m, 3H), 7.46 (dd, J = 1.87, 6.39 Hz, 1H). ¹³C NMR (CDCl₃, 101 MHz) δ 20.6, 24.4, 25.2, 36.9,

43.6, 51.5, 125.7, 127.1, 128.2, 130.3, 131.3, 135.5, 139.4, 139.4, 145.6, 146.4, 165.4. HRMS (ESI) Calcd for $C_{17}H_{19}O_2$ [M+H]⁺255.1380, found 255.1391.



Ligand (*R*,*R*)-**IId** (530.2 mg, 94% yield), $[\alpha]^{25}{}_{D}$ = +68.1 (c 1.62, CHCl₃), ¹H NMR (CDCl₃, 400 MHz) δ 1.12 (d, J = 6.8 Hz 3H), 1.25 (d, J = 6.9 Hz, 3H), 1.38-1.49 (m, 2H), 1.51-1.66 (m, 2H), 2.92-2.99 (m, 1H), 3.79 (s, 1H), 4.29-4.31 (m, 1H), 6.20-6.22 (m, 1H), 6.99-7.01 (m, 1H), 7.11-7.15 (m, 1H), 7.24-7.31 (m, 2H), 7.45-7.47 (m, 1H). ¹³C NMR (CDCl₃, 101

MHz) δ 23.9, 24.3, 24.4, 25.1, 29.7, 36.9, 44.4, 51.5, 125.3, 125.4, 127.5, 128.6, 130.7, 138.7, 139.2, 145.4, 146.3, 146.6, 165.4. HRMS (ESI) Calcd for C₂₈H₂₇O₂ [M+H]⁺ 283.1693, found 283.1694.



Ligand (*R*,*R*)-**IIe** (551.0 mg, 95% yield), $[\alpha]^{25}{}_{D} = +78.8$ (c 0.67, CHCl₃), ¹H NMR (CDCl3, 400 MHz) δ 1.45-1.55 (m, 2H), 1.62-1.71 (m, 2H), 3.82 (S, 3H), 4.02-4.06 (m, 1H), 4.39-4.41 (m, 1H), 6.47-6.49 (m, 1H), 7.25-7.27 (m, 1H), 7.41-7.51 (m, 3H), 7.58-7.60 (m, 1H), 7.77 (d, J = 8.2Hz, 1H), 7.82-7.87 (m, 2H). ¹³C NMR (CDCl3, 101 MHz) δ 24.7, 25.3,

37.2, 44.4, 51.6, 125.1, 125.3, 125.7, 125.9, 127.6, 128.4, 131.5, 132.5, 133.8, 137.8, 139.7, 145.5, 145.5, 165.4. HRMS (ESI) Calcd for $C_{20}H_{19}O_2$ [M+H]⁺291.1380, found 291.1379.

Ligand III: A solution of the corresponding (R,R)-II or dl-IIc (1.0 mmol) in MeOH (8 mL) and NaOH (160 mg, 4.0 mmol) was stirred at 60 °C for 8 h. Then 1N HCl (10 mL) was added in to the solution. The resulting solution was extracted with CH₂Cl₂ (10 mL) for three times, the combined organic layer was dried over MgSO₄, filtered, and concentrated under vacuum, which

directly used in next step (quantitative yields for all cases).



Ligand (*R*,*R*)-**IIIa**, $[\alpha]^{25}_{D}$ = -35.6 (c 0.62, CHCl₃), ¹H NMR (CDCl₃, 400 MHz) δ 1.47-1.63 (m, 4H), 4.31-4.33 (m, 2H), 6.62 (dd, J = 1.9, 6.4Hz, 1H), 7.26-7.28 (m, 1H), 7.34-7.37 (m, 2H), 7.41-7.43 (m, 2H), 7.59 (dd, J = 1.9, 6.5Hz, 1H). ¹³C NMR (CDCl₃, 101 MHz) δ 24.5, 25.2, 36.9, 40.7, 124.9, 127.2, 128.6, 129.3, 138.9, 145.0, 148.4,

169.7. HRMS (ESI) Calcd for C₁₅H₁₅O₂ [M+H]⁺ 227.1067, found 227.1069.



Compound *dl*-**IIIb**, ¹H NMR (CDCl₃, 400 MHz) δ 0.88-1.19 (m, 3H), 1.22-1.37 (m, 6H), 1.65-1.74 (m, 5H), 1.96-2.01 (m, 1H), 3.67 (d, J = 6.4Hz, 1H), 4.07 (d, J = 6.2Hz, 1H), 5.87 (d, J = 6.3Hz, 1H), 7.42 (dd, J = 1.9, 6.5Hz, 1H). ¹³C NMR (CDCl₃, 101 MHz) δ 24.5, 25.4, 26.3, 31.3, 31.5, 36.2, 41.2, 41.9, 124.5, 139.4, 148.8, 151.4,

170.2. HRMS (ESI) Calcd for $C_{15}H_{21}O_2$ [M+H]⁺233.1536, found 233.1544.



Ligand (*R*,*R*)-**IIIc**, $[\alpha]^{25}{}_{D}$ = +41.2 (c 1.65, CHCl₃), ¹H NMR (CDCl₃, 400 MHz) δ 1.41-1.66 (m, 4H), 2.26 (s, 3H), 3.94 (d, J = 6.1Hz, 3H), 4.31 (d, J = 6.1Hz, 1H), 6.29 (d, J = 6.3Hz, 1H), 7.08-7.09 (m, 1H), 7.14-7.20 (m, 3H), 7.64 (d, J = 6.4 Hz, 1H). ¹³C NMR (CDCl₃, 101 MHz) δ 20.6, 24.3, 25.2, 36.6, 43.8, 125.7, 127.1, 128.2, 130.4, 131.3,

135.5, 139.0, 139.3, 146.2, 148.6, 170.2. HRMS (ESI) Calcd for $C_{16}H_{17}O_2$ [M+H]⁺ 241.1223, found 241.1223.



Ligand (*R*,*R*)-**IIId** $[\alpha]^{25}_{D}$ = +68.7 (c 1.54, CHCl₃), ¹H NMR (CDCl₃, 400 MHz) δ 1.12 (d, J = 6.8 Hz 3H), 1.26 (d, J = 6.8 Hz, 3H), 1.40-1.50 (m, 2H), 1.53-1.65 (m, 2H), 2.91-2.98 (m, 1H), 3.86-3.88 (m, 1H), 4.29-4.31 (m, 1H), 6.22-6.24 (m, 1H), 7.01-7.02 (m, 1H), 7.12-7.16 (m, 1H), 7.25-7.32 (m, 2H), 7.62-7.65 (m, 1H). ¹³C NMR (CDCl₃, 101 MHz) δ 24.0, 24.3, 24.4, 25.1, 29.8,

36.7, 44.7, 125.4, 125.5, 127.6, 128.6, 130.7, 138.6, 138.9, 146.2, 146.7, 148.5, 170.3. HRMS (ESI) Calcd for $C_{18}H_{21}O_2$ [M+H]⁺ 269.1536, found 269.1545.



Ligand (*R*,*R*)-**IIIe**, $[\alpha]^{25}_{D} = +83.1$ (c 0.68, CHCl₃), ¹H NMR (CDCl3, 400 MHz) δ 1.48-1.61 (m, 2H), 1.64-1.74 (m, 2H), 4.09 (dd, J = 1.9,6.3Hz, 1H), 4.41 (dd, J = 1.9,6.2Hz, 1H), 6.54 (dd, J = 1.7, 6.3Hz, 1H), 7.26-7.28 (m, 1H), 7.42-7.51 (m, 3H), 7.77-7.79 (m, 2H), 7.82-7.87 (m, 2H). ¹³C NMR (CDCl3, 101 MHz) δ 24.6, 25.3, 36.9, 44.6, 125.1, 125.3, 125.6, 125.8, 126.0,

127.6, 128.4, 131.4, 132.4, 133.8, 137.6, 139.2, 145.2, 148.4, 170.1. HRMS (ESI) Calcd for $C_{19}H_{17}O_2$ [M+H]⁺ 277.1223, found 277.1234.

Ligand IV: To a solution of the carboxylic acid III (0.5 mmol) and DMF (7.5 μ L, 0.1 mmol) in CH₂Cl₂ (0.2 mL) was added dropwise oxalyl chloride (85 μ L, 1.0 mmol) at 0 °C, and the mixture was warmed to room temperature and stirred for 3 h. The solvent was removed under vacuum, and the residue was dissolved in THF (1.0 mL). To a solution of the corresponding amine (1.2 mmol) in THF (0.8 mL) was added the solution of the acid chloride dropwise at

0 °C. The mixture was stirred at room temperature for 20 h. saturated aqueous Na_2CO_3 solution was added, and the mixture was extracted with diethyl ether for three times. The organic layer was washed with brine, dried over MgSO₄, filtered, and concentrated under vacuum. Purification of the residue by column chromatography on silica gel with hexane/EtOAc (10/1) gave corresponding ligands **IV**.



Ligand (*R*,*R*)-**IVa** (133.5 mg, 96% yield), $[\alpha]^{25}{}_{D} = -97.6$ (c 0.05, CHCl₂), ¹H NMR (CDCl₃, 400 MHz) δ 1.41 (s, 9H), 1.50-1.50 (m, 4H), 4.21-4.26 (m, 2H), 5.57 (s, 1H), 6.61 (dd, J = 2.0, 6.4Hz, 1H), 6.84 (dd, J = 1.8, 6.4Hz, 1H), 7.23-7.27 (m, 1H), 7.32-7.36 (m, 2H), 7.40-7.42 (m, 2H). ¹³C NMR (CDCl₃, 101 MHz) δ 24.9,

25.4, 28.9, 37.5, 40.0, 51.1, 124.8, 127.0, 128.5, 129.5, 136.3, 137.7, 144.7, 145.6, 165.3. HRMS (ESI) Calcd for $C_{19}H_{21}NO [M+H]^+$ 282.1852, found 282.1855.



Compound *dl*-**IVb** (133.5 mg, 93% yield), ¹H NMR (CDCl₃, 400 MHz) δ 0.87-1.17 (m, 3H), 1.20-1.32 (m, 6H), 1.35 (s, 9H), 1.64-1.77 (m, 5H), 1.93-1.99 (m, 1H), 3.55 (d, J = 4.6Hz, 1H), 3.96 (d, J = 4.0Hz, 1H), 5.51 (s, 1H), 5.83 (d, J = 6.2Hz, 1H), 6.71 (t, 1H). ¹³C NMR (CDCl₃, 101 MHz) δ 24.9, 25.6, 26.3, 28.9, 31.2,

31.4, 36.8, 40.5, 41.9, 50.9, 124.3, 136.9, 144.8, 152.1, 165.3.HRMS (ESI) Calcd for C₁₉H₃₀NO $[M+H]^+$ 288.2322, found 288.2333. Resolution of *dl*- **IVb** was carried out by use of a chiral stationary phase column [Chiral IG (2.0 cm I.D. × 25 cm), hexane/IPA= 98:2 to give both enantiomers (*S*,*S*)-**IVb** and (*R*,*R*)-**IVb**, quantitatively. *t*₁ = 34 min for (*R*,*R*)-**IVb**, *t*₂ = 42 min for (*S*,*S*) -**IVb**. [α]²⁵_D = +8.5 (c 0.85, CHCl₃) for (*R*,*R*)-**IVb**.



Ligand (*R*,*R*)-**IVc** (143.1 mg, 97% yield), $[\alpha]^{25}{}_{D}$ = +59.5 (c 0.77, CHCl₃). ¹H NMR (CDCl₃, 400 MHz) δ 1.41(s, 4H), 1.46-1.58 (m, 4H), 2.26 (s, 3H), 3.82-3.84 (m, 1H), 4.18-4.20 (m, 1H), 5.59 (s, 1H), 6.25 (dd, J = 1.7, 6.3Hz, 1H), 6.89 (dd, J = 1.9, 6.3Hz, 1H), 7.05-7.07 (m, 1H), 7.12-7.17 (m, 3H). ¹³C NMR (CDCl₃,

101 MHz) δ 20.7, 24.6, 25.4, 28.9, 37.3, 43.1, 51.2, 125.6, 127.0, 128.2, 130.3, 131.5, 135.4, 136.6, 139.5, 144.4, 146.7, 165.2. HRMS (ESI) Calcd for C₂₀H₂₆NO [M+H]⁺ 296.2009, found 296.2022.



Ligand (*R*,*R*)-**IVd** (151.2 mg, 94% yield), $[\alpha]^{25}_{D}$ +60.9 (c 0.97, CHCl₃). ¹H NMR (CDCl₃, 400 MHz) δ 1.12 (d, J = 6.8 Hz 3H), 1.24 (d, J = 6.9 Hz, 3H), 1.42 (s, 1H), 1.46-1.61 (m, 3H), , 2.93-3.03 (m, 1H), 3.75-3.77 (m, 1H), 4.16-4.18 (m, 1H), 5.61 (s, 1H), 6.18-6.20 (m, 1H), 6.89-6.91 (m, 1H), 6.99-7.01 (m, 1H), 7.09-7.13 (m, 1H), 7.22-7.30 (m, 2H), ¹³C NMR (CDCl₃)

101 MHz) δ 24.1, 24.5, 24.6, 25.3, 28.2, 29.0, 29.7, 37.3, 44.1, 51.2, 125.4, 125.5, 127.4, 128.6, 130.8, 136.6, 138.8, 144.2, 146.6, 165.1. HRMS (ESI) Calcd for C₂₂H₃₀NO [M+H]⁺ 324.2322, found 324.2336.



Ligand (*R*,*R*)-**IVe** (158.9 mg, 96% yield), $[\alpha]^{25}{}_{D} = +79.1$ (c 0.69, CHCl₃), ¹H NMR (CDCl3, 400 MHz) δ 1.44 (s, 9H), 1.55-1.65 (m, 4H), 3.98-4.00 (m, 1H), 4.27-4.29 (m, 1H), 5.67 (s, 1H), 6.47 (dd, J = 1.8, 6.3Hz, 1H), 7.02 (dd, J = 1.9, 6.3Hz, 1H), 7.25-7.27 (m, 1H), 7.40-7.48 (m, 3H), 7.76 (d,

 $J = 8.2Hz, 1H), 7.84-7.88 (m, 2H). {}^{13}C NMR (CDCl3, 101 MHz) \delta 25.0, 25.5, 29.0, 37.6, 44.0, 51.2, 125.0, 125.3, 125.7, 125.8, 125.9, 127.4, 128.4, 131.4, 132.6, 133.9, 136.5, 137.9, 144.7, 145.8, 165.1. HRMS (ESI) Calcd for C₂₃H₂₆NO [M+H]⁺ 332.2009, found 332.2010.$



Ligand (*R*,*R*)-**IVf** (156.4 mg, 78% yield), $[\alpha]^{25}{}_{D}$ = +48.6 (c 0.84, CHCl₃), ¹H NMR (CDCl3, 400 MHz) δ 0.81-0.88 (m, 6H), 0.88-0.94 (m, 3H), 1.19-1.26 (m, 3H), 1.28-1.41 (m, 4H), 1.49-1.53 (m, 3H), 1.68-1.80 (m, 4H), 4.00 (dd, J = 1.9, 6.2Hz, 1H), 4.26 (dd, J = 2.0, 6.3Hz,

1H), 5.26 (s, 1H), 6.47 (dd, J = 1.8, 6.2Hz, 1H), 7.03 (dd, J = 1.8, 6.3Hz, 1H), 7.26-7.27 (m, 1H), 7.40-7.50 (m, 3H), 7.76 (d, J = 8.2Hz, 1H), 7.84-7.89 (m, 2H). ¹³C NMR (CDCl3, 101 MHz) δ 7.7, 14.2, 23.1, 25.0, 25.4, 25.6, 27.2, 29.7, 34.0, 37.7, 44.0, 59.4, 125.0, 125.3, 125.7, 125.8, 125.9, 127.4, 128.4, 131.4, 132.5, 133.9, 136.3, 137.9, 144.8, 145.8, 164.9. HRMS (ESI) Calcd for C₂₈H₃₆NO [M+H]⁺ 402.2791, found 402.2804.



Ligand (*R*,*R*)-**IVg** (173.7 mg, 76% yield), $[\alpha]^{25}{}_{D} = +59.3$ (c 1.00, CHCl₃), ¹H NMR (CDCl₃, 400 MHz) δ 0.93 (t, 9H), 1.17-1.25 (m, 6H), 1.30-1.37 (m, 6H), 1.50-1.53 (m, 2H), 1.63-1.75 (m, 8H), 3.99 (d, J = 4.6Hz, 1H), 4.25 (d, J = 4.6Hz, 1H), 5.23 (s, 1H), 6.47 (d, J = 5.0Hz, 1H), 7.01-7.03 (m, 1H), 7.27 (d, J = 6.8Hz, 1H), 7.40-7.48 (m, 3H), 7.76

(d, J = 8.2Hz, 1H), 7.84-7.89 (m, 2H). ¹³C NMR (CDCl₃, 101 MHz) δ 14.2, 23.1, 25.0, 25.5, 25.6, 35.1, 37.7, 44.0, 58.9, 125.0, 125.3, 125.7, 125.8, 127.4, 128.4, 131.4, 132.6, 133.9, 136.2, 144.9, 145.8, 164.8. HRMS (ESI) Calcd for C₃₂H₄₄NO [M+H]⁺ 458.3417, found 458.3426.

4. A typical procedure for rhodium-catalyzed domino reaction of 2-formyl phenylboric acid (1a) with 4-octyne (2a)





[RhCl(coe)₂]₂ (2.7 mg, 0.0038 mmol, 5 mol% of Rh) and (R,R)-IVg * (3.8 mg, 0.0083 mmol) were placed in an oven dried 10 mL schlenk tube. The tube was evacuated and re-filled with nitrogen gas for 3 cycles. Anhydrous 1,4-dioxane (0.20 mL) was added, and the resulting mixture was stirred at room temperature for 10 min. To the mixture, 4-octyne (**2a**, 16.5 mg, 0.15 mmol, 1.0 equiv.), 2-formyl phenylboric acid (**1a**, 34.8 mg, 0.225 mmol, 1.5 equiv.), the freshly degassed H₂O (0.025 mL), 40 mol% KOH (0.060 mmol, 40.0 mol%) and 1,4-dioxane (0.30 mL) were added. The resulting mixture was stirred at rt for 16 h. The mixture was passed

through a short pad of silica gel with ethyl acetate as eluent. The organic solution was concentrated under reduced pressure, and the residue was purified by flash chromatography on silica gel using 10/1 PE/EA as eluent to give the target compound **3aa** (29.6 mg, 91% yield). Subsequently, the ee value of the target product **3aa** was determined by HPLC.

5. Characterization of products



Compound **3aa** (29.6 mg, 91% yield). The ee was measured by HPLC (Daicel Chiralpak OD-H column), hexane/2-propanol = 90/10, flow 0.7 mL/min, 256 nm, $t_{major} = 7.7$ min, $t_{minor} = 8.4$ min. $[\alpha]^{25}_{D}$ +9.5 (c 0.80, CHCl₃) for **3aa** of 92% ee (S). The ee was measured by HPLC (Daicel Chiralpak OD-H column), hexane/2-propanol = 95/5, flow 0.7 mL/min,

256 nm, $t_{major} = 10.2 \text{ min}$, $t_{minor} = 7.4 \text{ min}$. $[\alpha]^{25}{}_{D}$ -5.8 (c 0.54, CHCl₃) for **3aa** of 90% ee (R). ¹H NMR (CDCl₃, 600 MHz) δ 0.98 (td, J = 2.76, 7.32 Hz, 6H), 1.50-1.65 (m, 4H), 2.38-2.44 (m, 4H), 4.97 (d, J = 9.47 Hz, 1H), 7.13-7.17 (m, 2H), 7.25-7.27 (m, 1H), 7.47 (d, J = 7.17 Hz, 1H); ¹³C NMR (CDCl₃, 151 MHz) δ 14.4, 21.8, 22.9, 27.3, 27.8, 118.7, 123.1, 124.9, 128.3, 137.9, 144.3, 145.0, 145.2. HRMS (ESI) Calcd for C₁₅H₂₀O [M+H]⁺ 217.1587, found 217.1584.



Compound **3ba** (28.0 mg, 76% yield). The ee was measured by HPLC (Daicel Chiralpak OD-H column), hexane/2-propanol = 95/5, flow 1 mL/min, 256 nm, $t_{major} = 9$ min, $t_{minor} = 8$ min. $[\alpha]^{25} _{D}$ +20.2 (c 0.47, CHCl₃) for **3ba** of 89% ee (S). ¹H NMR (CDCl₃, 600 MHz) δ 0.97 (td, J = 2.19, 7.35 Hz, 6H), 1.49-164 (m, 4H),

2.34-2.41 (m, 4H), 3.82 (s, 3H), 4.94 (s, 1H), 6.78 (dd, J = 2.40, 8.14 Hz, 1H), 7.04 (d, J = 8.14 Hz 1H), 7.10 (d, J = 2.29 Hz, 1H); ¹³C NMR (CDCl₃, 101 MHz) δ 14.3, 14.3, 21.8, 22.9, 27.4, 27.7, 55.6, 110.4, 112.9, 119.1, 137.0, 137.6, 143.0, 146.9, 158.1. HRMS (ESI) Calcd for C₁₆H₂₂O₂ [M+H]⁺ 247.1693, found 247.1692.



Compound **3ca** (28.0 mg, 81% yield). The ee was measured by HPLC (Daicel Chiralpak OD-H column), hexane/2-propanol = 98/2, flow 1 mL/min, 256 nm, $t_{major} = 5 \text{ min}$, $t_{minor} = 8 \text{ min}$. [α]²⁵ D +14.7 (c 0.47, CHCl₃) for **3ca** of 95% ee (S). ¹H NMR (CDCl₃, 600 MHz) δ 0.95-1.00 (m, 6H), 1.55-1,59 (m, 4H), 2.37-2.41 (m, 7H), 4.94 (s,

1H), 6.96 (d, J = 9.45 Hz, 2H), 7.35 (d, J = 7.26 Hz 1H); ¹³C NMR (CDCl₃, 151 MHz) δ 14.3, 14.4, 21.7, 21.8, 22.9, 27.3, 27.8, 119.6, 122.8, 125.4, 137.8, 138.1, 142.2, 144.5, 145.5. HRMS (ESI) Calcd for C₁₆H₂₂O [M+H]⁺ 231.1743, found 231.1742.



Compound **3da** (31.1 mg, 80% yield). The ee was measured by HPLC (Daicel Chiralpak OD-H column), hexane/2-propanol = 95/5, flow 1 mL/min, 256 nm, $t_{major} = 11$ min, $t_{minor} = 10$ min. $[\alpha]^{25}$ $_{D}$ +45.7 (c 0.47, CHCl₃) for **3da** of 84% ee (S). ¹H NMR (CDCl₃, 600 MHz) δ 0.97 (q, J = 6.57 Hz, 6H), 1.54-1.65 (m, 4H), 2.33-2.40 (m, 4H), 2.94 (s, 6H), 4.93 (s, 1H), 6.62 (dd, J = 2.23, 8.16)

Hz, 1H), 7.01-7.03 (m, 2H); ¹³C NMR (CDCl₃, 151 MHz) δ 14.3, 14.4, 21.9, 23.0, 27.4, 27.7, 41.3, 109.7, 111.9, 119.0, 133.6, 137.8, 141.3, 146.7, 149.3. HRMS (ESI) Calcd for C₁₇H₂₅NO [M+H]⁺ 260.2009, found 260.2007.



Compound **3ea** (34.1 mg, 80% yield). The ee was measured by HPLC (Daicel Chiralpak OD-H column), hexane/2-propanol = 95/5, flow 1 mL/min, 230 nm, $t_{major} = 5 \text{ min}$, $t_{minor} = 7 \text{ min}$. [α]²⁵ _D+1.8 (c 0.51, CHCl₃) for **3ea** of 96% ee (S). ¹H NMR (CDCl₃, 400 MHz) δ 0.98 (t, J = 7.31 Hz, 6H), 1.52-1.67 (m, 4H), 2.42-2.44 (m, 4H), 5.01 (s, 1H), 7.22 (d, J = 7.80 Hz, 2H), 7.53 (d, J = 7.77 Hz, 1H),

7.69 (s, 1H); ¹³C NMR (CDCl₃, 151 MHz) δ 14.3, 14.3, 21.7, 22.8, 27.2, 27.9, 118.6, 119.9, 125.8, 127.2, 137.5, 145.4, 147.7, 148.4. ¹⁹F NMR (CDCl₃, 565MHz) δ -113.2. HRMS (ESI) Calcd for C₁₆H₁₉F₃O [M+H]⁺285.1461, found 285.1460.



Compound **3fa** (25.6 mg, 73% yield). The ee was measured by HPLC (Daicel Chiralpak OD-H column), hexane/2-propanol = 90/10, flow 1 mL/min, 230 nm, $t_{major} = 7$ min, $t_{minor} = 7$ min. [α]²⁵ _D +17.7 (c 0.47, CHCl₃) for **3fa** of 88% ee (S). ¹H NMR (CDCl₃, 400 MHz) δ 0.97 (t, Hz, 6H), 1.51-1.64 (m, 4H), 2.35-2.42 (m, 4H), 4.92 (s, 1H), 6.91-6.96 (m, 1H), 7.03-7.06 (m, 1H), 7.17 (d, J = 8.16 Hz, 1H); ¹³C NMR (CDCl₃, 101 MHz) δ 14.3, 21.7, 22.8, 27.3, 27.8, 111.4 (d, J = 23.4

Hz), 114.4 (d, J = 22.5 Hz), 119.2 (d, J = 8.1 Hz), 137.2, 139.9, 144.8 (d, J = 4.0 Hz), 147.2 (d, J = 7.7 Hz), 161.56 (d, J = 243.9). ¹⁹F NMR (CDCl₃, 565MHz) δ -121.9 (m, 1F) .HRMS (ESI) Calcd for C₁₅H₁₉FO [M+H]⁺ 235.1493, found 235.1490.



Compound **3ga** (28.6 mg, 76% yield). The ee was measured by HPLC (Daicel Chiralpak OD-H column), hexane/2-propanol = 98/2, flow 1 mL/min, 256 nm, $t_{major} = 7$ min, $t_{minor} = 5$ min. [α]²⁵ _D +10.3 (c 0.37, CHCl₃) for **3ga** of 99% ee (S). ¹H NMR (CDCl₃, 600 MHz) δ 0.97 (td, J = 2.45, 7.35 Hz, 6H), 1.49-1.57 (m, 4H), 2.37-2.40 (m,

4H), 4.95 (d, J = 9.62 Hz, 1H), 7.05 (d, J = 7.95 Hz, 1H), 7.23 (dd, J = 1.86, 7.94 Hz, 1H), 7.43 (d, J = 1.58 Hz, 1H); ¹³C NMR (CDCl₃, 101 MHz) δ 14.3, 14.3, 21.7, 22.8, 27.2, 27.8, 119.5, 123.8, 128.2, 130.9, 137.4, 142.7, 145.6, 146.8. HRMS (ESI) Calcd for C₁₅H₁₉FO [M+H]⁺ 251.1197, found 251.1202



Compound **3ha** (25.0 mg, 71% yield). The ee was measured by HPLC (Daicel Chiralpak OD-H column), hexane/2-propanol = 95/5, flow 1 mL/min, 230 nm, $t_{major} = 10$ min, $t_{minor} = 8$ min. [α]²⁵ _D +4.0 (c 0.05, CHCl₂) for **3ha** of 99% ee (S). ¹H NMR (CDCl₃, 600 MHz) δ 1 (t, 6H), 1.50-1.69 (m, 4H), 2.39 (t, 2H), 2.51-2.52 (m, 2H), 5.00 (s, 1H), 6.94-6.97 (m, 1H), 7.10-7.14 (m, 1H), 7.27-7.28 (m, 1H); ¹³C NMR (CDCl₃, 151 MHz) δ

14.3, 14.3, 21.8, 22.7, 27.4, 27.7, 74.8, 112.6 (d, J = 2.1 Hz), 114.9 (d, J = 323.3 Hz), 129.3, 130.6 (d, J = 7.1 Hz), 137.8, 146.7 (d, J = 323.3 Hz), 159.0 (d, J = 248.0 Hz).s ¹⁹F NMR (CDCl₃, 565MHz) δ -121.9 (t, 1F). HRMS (ESI) Calcd for C₁₅H₁₉FO [M+H]⁺235.1493, found 235.1494.



Compound **3ia** (24.5 mg, 70% yield). The ee was measured by HPLC (Daicel Chiralpak OD-H column), hexane/2-propanol = 95/5, flow 1 mL/min, 230 nm, $t_{major} = 12$ min, $t_{minor} = 13$ min. [α]²⁵ _D +9.7 (c 0.05, CHCl₂) for **3ia** of 86% ee (S). ¹H NMR (CDCl₃, 600 MHz) δ 0.94-0.97 (m, 6H), 1.40-1.63 (m, 4H), 2.32-2.38 (m, 4H), 4.90 (s, 1H), 6.70-6.72 (m, 1H), 6.96-6.97 (m, 1H), 7.01 (d, J=2.1Hz, 1H);

¹³C NMR (CDCl₃, 151 MHz) δ 14.3, 14.3, 21.8, 22.9, 24.8, 27.4, 27.7, 111.8, 114.4, 119.3,



Compound **3ab** (31.1 mg, 85% yield). The ee was measured by HPLC (Daicel Chiralpak OD-H column), hexane/2-propanol = 95/5, flow 1 mL/min, 230 nm, $t_{major} = 6$ min, $t_{minor} = 7$ min. [α]²⁵ _D +5.07 (c 0.48, CHCl₂) for **3ab** of 86% ee (S). ¹H NMR (CDCl₃, 600 MHz) δ 0.94-0.96 (m, 6H), 1.37-1.44 (m, 4H), 1.45-1.62 (m, 4H), 2.40-2.46 (m, 4H), 4.97 (s, 1H), 7.13-7.16 (m, 2H), 7.25-7.28 (m, 1H), 7.47 (d, J = 7.1 Hz, 1H);

¹³C NMR (CDCl₃, 151 MHz) δ 14.0, 22.9, 25.0, 25.4, 30.8, 31.8, 118.7, 123.0, 124.9, 128.3, 137.9, 144.3, 145.0, 145.1. HRMS (ESI) Calcd for $C_{17}H_{24}O [M+H]^+$ 245.1900, found 245.1903



Compound **3ac** (36.3 mg, 85% yield). The ee was measured by HPLC (Daicel Chiralpak OD-H column), hexane/2-propanol = 95/5, flow 1 mL/min, 230 nm, $t_{major} = 11$ min, $t_{minor} = 12$ min. [α]²⁵ _D -0.4 (c 0.47, CHCl₃) for **3ac** of 77% ee (S). ¹H NMR (CDCl₃, 600 MHz) δ 1.98-2.14 (m, 4H), 2.58-2.67 (m, 4H), 2.57-3.62 (m, 7H), 4.97 (s, 1H), 7.17-7.20 (m, 2H), 7.28-7.31 (m, 1H), 7.48 (d, J = 7.1 Hz, 1H); ¹³C NMR

 $(CDCl_3, 101 \text{ MHz}) \, \delta \, 22.3, 23.0, 31.2, 32.1, 44.7, 44.7, 118.8, 123.3, 125.6, 128.6, 137.5, 143.3, 144.3, 144.8. \text{ HRMS} (ESI) Calcd for C_{15}H_{18}Cl_2O [M+H]^+ 285.0807, found 285.0801 \\$



COO^tBu HPLC (Daicel Chiralpak OD-H column), hexane/2-propanol = 95/5, flow 1 mL/min, 230 nm, $t_{major} = 7$ min, $t_{minor} = 6$ min. $[\alpha]^{25} D_{12}$ +2.9 (c 0.14, CHCl₃) for **3ad** of 71% ee (S). ¹H NMR (CDCl₃, 600 MHz) δ 1.43 (s, 9H), 1.44 (s, 9H), 2.41-2.44 (m, 2H), 2.52-2.55 (m, 2H), 2.72-2.76 (m, 4H), 4.95 (s, 1H), 7.16-7.18 (m, 2H),

7.26-7.28 (m, 1H), 7.48 (d, J = 7.5 Hz, 1H); ¹³C NMR (CDCl₃, 151 MHz) δ 20.7, 21.0, 24.4, 24.9, 28.1, 28.1, 34.1, 34.8, 80.6, 80.9, 118.7, 123.2, 125.5, 128.3, 137.2, 143.0, 144.2, 144.8, 172.3, 173.2. HRMS (ESI) Calcd for C₂₃H₃₂O₅ [M+H]⁺389.2323, found 389.2323.



Compound **3ae** (47.9 mg, 63% yield). The ee was measured by HPLC (Daicel Chiralpak OD-H column), hexane/2-propanol = 95/5, flow 1 mL/min, 230 nm, $t_{major} = 19$ min, $t_{minor} = 20$ min. $[\alpha]^{25}_{D} +20.4$ (c 0.05, CHCl₃) for **3ae** of 71% ee (R). ¹H NMR (CDCl₃, 600 MHz) δ 1.92-2.05 (m, 4H), 2.49-2.55 (m, 4H), 3.70-3.73 (m, 4H), 4.99 (d, J= 8.1Hz, 1H), 7.05-7.08 (m, 2H), 7.18 (t, 1H), 7.40 (d, J = 7.2 Hz, 1H), 7.66-7.70 (m, 4H), 7.76-7.79 (m, 2H), 7.80-7.82 (m, 2H); ¹³C NMR (CDCl₃, 151 MHz) δ 22.8, 23.3, 27.0, 28.0, 37.9, 37.9, 118.7, 123.2, 123.2, 125.3, 128.3, 132.2, 133.8, 133.9, 137.3, 143.2, 144.3, 144.9, 168.3,

168.4. HRMS (ESI) Calcd for $C_{31}H_{26}N_2O_5$ [M+H]⁺ 507.1914, found 507.1915.



Compound **3af** (20.7 mg, 64% yield). The ee was measured by HPLC (Daicel Chiralpak IC column), hexane/2-propanol = 95/5, flow 0.7 mL/min, 245 nm, $t_{major} = 10$ min, $t_{minor} = 13$ min. $[\alpha]^{25}$ D -4.0 (c 0.08, CHCl₂) for **3af** of 80% ee (S). ¹H NMR (CDCl₃, 600 MHz) δ 0.9 (t, 3H), 1.34-1.35 (m, 4H), 1.48-1.54 (m, 1H), 1.58-

1.64 (m, 1H), 1.99 (s, 3H), 2.42 (t, 2H), 4.97 (s, 1H), 7.13-7.17 (m, 2H), 7.27-7.30 (m, 1H), 7.46 (d, J=7.2Hz, 1H); 13 C NMR (CDCl₃, 151 MHz) δ 10.2, 14.1, 22.6, 25.5, 29.1, 31.9, 118.3, 122.9, 125.1, 128.4, 133.4, 144.7, 145.0, 145.1. HRMS (ESI) Calcd for C₁₅H₂₀O [M+H]⁺ 217.1587, found 217.1592.



Compound **3af**'(7.1 mg, 22% yield). The ee was measured by HPLC (Daicel Chiralpak IC column), hexane/2-propanol = 95/5, flow 0.7 mL/min, 245 nm, $t_{major} = 7$ min, $t_{minor} = 8$ min. [α]²⁵ _D+13.3 (c 0.09, CHCl₂) for **3af**' of 85% ee (S). ¹H NMR (CDCl₃, 600 MHz) δ 0.89 (t, 3H), 1.26-1.35 (m, 6H), 1.99 (s, 3H), 2.43 (t, 2H), 4.83 (d, J=6.5Hz, 1H), 7.13-7.15 (m, 2H), 7.26 (s, 1H), 7.46 (d, J=7.4Hz, 1H); ¹³C NMR (CDCl₃, 151 MHz) δ 11.1, 14.1, 22.6, 25.1, 27.9, 31.9, 78.9, 118.5, 123.0, 124.9, 128.3.

HRMS (ESI) Calcd for C₁₅H₂₀O [M+H]⁺ 217.1587, found 217.1590.



Compound **3ag** (30.8 mg, 71% yield). The ee was measured by HPLC (Daicel Chiralpak OD-H column), hexane/2-propanol = 90/10, flow 0.7 mL/min, 256 nm, $t_{major} = 14$ min, $t_{minor} = 17$ min. [α]²⁵ D -110.6 (c 0.62, CHCl₃) for **3ag** of 77% ee (R). ¹H NMR (CDCl₃, 600 MHz) δ 0.94 (t, 3H), 1.42-1.46 (m, 2H), 1.60-1.69 (m, 2H), 2.64-2.68 (m, 2H), 5.52 (s, 1H), 7.29-7.32 (m, 1H), 7.34-7.40

(m, 2H), 7.58-7.60 (m, 3H), 7.72 (d, J = 8.4Hz, 2H). ¹³C NMR (CDCl₃, 101 MHz) δ 13.8, 23.0, 25.9, 30.7, 110.5, 120.3, 123.5, 126.9, 128.9, 129.3, 132.3, 140.1, 141.3, 143.1, 143.3, 144.6. HRMS (ESI) Calcd for C₂₀H₂₀NO [M+H]⁺ 290.1539, found 290.1540.

6. Transformations





To a Schlenk tube (10 mL) under an atmosphere of nitrogen was added (S)-**3aa** (21.6 mg, 0.1 mmol) in NEt₃ (30.4 mg, 0.3 mmol). The mixture was stirring 16 h at 50 °C. The mixture was washed with NaHCO₃ the organic layer was extracted with EA for 3 times, then dried over Na₂SO₄ and concentrated under vacuum to give the compound **4a**^[10] in 95% yield. The ee was measured by HPLC (Daicel Chiralpak ODH column), hexane/2-propanol = 95/5, flow 0.7

mL/min, 245 nm, $t_{major} = 7 \text{ min}$, $t_{minor} = 8 \text{ min}$. $[\alpha]^{25} {}_{D} + 2.4$ (c 0.05, CHCl₂) for (S,S)-**4a** of 89% ee (R). ¹H NMR (CDCl₃, 600 MHz) δ 0.92-0.97 (m, 6H), 1.40-1.46 (m, 2H), 1.55-1.59 (m, 2H), 1.73-1.79 (m, 2H), 2.35-2.37 (m, 1H), 3.04-3.07 (m, 1H), 7.34-7.36 (m, 1H), 7.46-7.47 (m, 1H), 7.57-7.59 (m, 1H), 7.71 (d, J=7.7Hz, 1H). HRMS (ESI) Calcd for C₁₅H₂₁O [M+H]⁺ 217.1592, found 290.1587.

To a Schlenk tube (10 mL) under an atmosphere of nitrogen was added **3ab** (24.4 mg, 0.1 mmol) in NEt₃ (40.5 mg, 0.4mmol). The mixture was stirring 16 h at 50 °C. The mixture was washed with NaHCO₃ the organic layer was extracted with EA for 3 times, then dried over Na₂SO₄ and concentrated under vacuum to give the compound **5a**^[11] in 62% yield. The ee was measured by HPLC (Daicel Chiralpak IC column), hexane/2-propanol = 99.8/0.2, flow 0.5 mL/min, 245 nm, $t_{major} = 33 \text{ min}, t_{minor} = 50 \text{ min}. [\alpha]^{25} \text{ }_{D} + 5.2$ (c 0.5, CHCl₂) for (S,S)-**5a** of 84% ee. ¹H NMR (CDCl₃, 600 MHz) δ 0.90 (q, 6H), 1.28-1.39 (m, 10H), 1.75-1.85 (m, 2H), 2.34-2.37 (m, 1H), 3.05-3.07 (m, 1H), 7.36 (t, 1H), 7.48 (d, J=7.6Hz, 1H). 7.58-7.60 (m, 1H), 7.72 (d, J=7.6Hz, 1H). HRMS (ESI) Calcd for C₁₇H₂₅O [M+H]⁺ 245.1905, found 290.1901.

7. Reference

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8. Data for X-ray crystal structure of (S)-3aa

Crystals of (S)-**3aa** (92% ee) suitable for X-ray crystallographic analysis were obtained by recrystallization from hexane and ether. A suitable crystal of 3aa (recrystallized from n-Hexane at room temperature) was selected and mounted. The diffraction data were collected on a Bruker D8 venture microsource diffractometer using monochromatic Cu-K α radiation (1.54178 Å) at 150 K. Date reduction, scaling and absorption corrections were performed using SAINT (Bruker, V8.40A, 2016). The structures were solved with the ShelXT (Sheldrick, 2015) structure solution program using the Intrinsic Phasing solution method and by using Olex2 (Dolmanov et al., 2009) as the graphical interface. The model was refined with version 2016/6 of ShelXT (Sheldrick, 2008) using Least Squares minimization. The crystal structure has been deposited at the Cambridge Crystallographic Data Centre (CCDC 2262466).

Bond precision:	C-C = 0.0019 A	Wavelength=1.54178	
Cell:	a=4.7593(1)	b=16.2350(3)	c=16.5153(4)
	alpha=90	beta=90	gamma=90
Temperature: 150 K			
	Calculated	Reported	
Volume	1276.09(5)	127	(6.09(5)
Space group	P 21 21 21	P 21 21 21	
Hall group	P 2ac 2ab	P 2ac 2ab	
Moiety formula	C15 H20 O	C15 H20 O	
Sum formula	C15 H20 O	C1:	5 H20 O
Mr	216.31	216	.31
Dx,g cm-3	1.126	1.12	26
Z	4	4	
Mu (mm-1)	0.522	0.52	22
F000	472.0	816	5.0
F000'	473.23		
h,k,lmax	5,20,20	5,20	0,20
Nref	2602[1548]	259	1
Tmin,Tmax	0.963, 0.974	0.6	76, 0.754
Tmin'	0.934		

Table 1. Crystal data and structure refinement for (S)-3aa

Correction method= # Reported T Limits: Tmin=0.676 Tmax=0.754

AbsCorr =REFDELF

Data completeness= 1.67/1.00		Theta(max)= 74.575
R(reflections)= 0.0271(2543)		wR2(reflections)
= 0.0694(2591)		
S = 1.038	Npar= 251	

The ORTEP drawing of (S)-**3aa** with ellipsoid contour at 30% probability level is shown as follows:



9. NMR spectra













































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¹H NMR (600 MHz, CDCl₃) spectrum of **3aa**



¹³C NMR (151 MHz, CDCl₃) spectrum of **3aa**


¹H NMR (600 MHz, CDCl₃) spectrum of **3ba**



¹³C NMR (151 MHz, CDCl₃) spectrum of **3ba**



¹H NMR (600 MHz, CDCl₃) spectrum of **3ca**



¹³C NMR (151 MHz, CDCl₃) spectrum of **3ca**



¹H NMR (600 MHz, CDCl₃) spectrum of **3da**



¹³C NMR (151 MHz, CDCl₃) spectrum of **3da**



¹H NMR (600 MHz, CDCl₃) spectrum of **3ea**



¹³C NMR (151 MHz, CDCl₃) spectrum of **3ea**



¹⁹F NMR (565 MHz, CDCl₃,) spectrum of **3ea**



¹H NMR (600 MHz, CDCl₃) spectrum of **3fa**



¹³C NMR (151 MHz, CDCl₃) spectrum of **3fa**



¹⁹F NMR (565 MHz, CDCl₃,) spectrum of **3fa**



¹H NMR (600 MHz, CDCl₃) spectrum of **3ga**



¹³C NMR (151 MHz, CDCl₃) spectrum of **3ga**



¹H NMR (600 MHz, CDCl₃) spectrum of **3ha**



¹³C NMR (151 MHz, CDCl₃) spectrum of **3ha**



¹⁹F NMR (565 MHz, CDCl₃,) spectrum of **3ha**



¹H NMR (600 MHz, CDCl₃) spectrum of **3ia**



¹³C NMR (151 MHz, CDCl₃) spectrum of **3ia**



¹H NMR (600 MHz, CDCl₃) spectrum of **3ab**



¹³C NMR (151 MHz, CDCl₃) spectrum of **3ab**



¹H NMR (600 MHz, CDCl₃) spectrum of **3ac**



¹³C NMR (151 MHz, CDCl₃) spectrum of **3ac**



 ^1H NMR (600 MHz, CDCl₃) spectrum of 3ad



¹³C NMR (151 MHz, CDCl₃) spectrum of **3ad**



 1 H NMR (600 MHz, CDCl₃) spectrum of **3ae**



200 180 160 140 120 100 80 60 40 20 0 ppm

¹³C NMR (151 MHz, CDCl₃) spectrum of **3ae**



¹H NMR (600 MHz, CDCl₃) spectrum of **3af**



 ^{13}C NMR (151 MHz, CDCl₃) spectrum of 3af



¹H NMR (600 MHz, CDCl₃) spectrum of **3ag**



¹³C NMR (151 MHz, CDCl₃) spectrum of **3ag**



 ^1H NMR (600 MHz, CDCl₃) spectrum of 3ah



¹³C NMR (151 MHz, CDCl₃) spectrum of **3ah**



¹H NMR (600 MHz, CDCl₃) spectrum of **4a**



¹H NMR (600 MHz, CDCl₃) spectrum of **5a**

NOE NMR for 3af and 3af'







10. HPLC Data

(S)**-3aa**



reak%	Ket. lime	Area	neight	Area%
1	7.827	3520058	413283	95.897
2	8.597	150602	17204	4.103
Total		3670660	430487	





Peak#	Ret.Time	Area	Height	Area%
1	7.834	424435	52461	5.092
2	10.317	7910936	511553	94.908
总计		8335371	564014	





Peak#	Ret. Time	Area	Height	Area%
1	7.742	719410	59065	50.075
2	9.146	717263	47210	49.925
Total		1436673	106275	100.000



Peak#	Ret.Time	Area	Height	Area%
1	7.657	113672	9836	5.439
2	8.972	1976411	126091	94.561
Total		2090083	135927	



Peak#	Ret.Time	Area	Height	Area%
1	5.793	1845291	273526	50.027
2	8.168	1843315	176797	49.973
Total		3688606	450323	



Peak#	Ret.Time	Area	Height	Area%
1	5.533	8357479	1152818	97.632
2	7.914	202673	20113	2.368
Total		8560151	1172930	





Peak#	Ret.Time	Area	Height	Area%
1	10.176	11980919	1088933	49.866
2	11.040	12045273	1014226	50.134
Total		24026191	2103159	100.000





Peak#	Ret.Time	Area	Height	Area%
1	10.152	2066931	197771	8.139
2	11.004	23329300	1958181	91.861
Total		25396231	2155952	100.000





Peak#	Ret.Time	Area	Height	Area%
1	5.618	15595186	953384	50.053
2	6.919	15562122	501351	49.947
Total		31157308	1454735	100.000



Peak#	Ret.Time	Area	Height	Area%
1	5.662	1540188	201056	98.178
2	6.573	28582	2919	1.822
Total		1568770	203975	100.000



Prak#	ket.lime	Area	Height	Area%
1	6.833	188781	25730	5.836
2	7.195	3046160	328464	94.164
Total		3234942	354194	



Peak#	Ret.Time	Area	Height	Area%
1	5.178	1700872	245324	50.203
2	6.383	1687106	195864	49.797
Total		3387978	441188	100.000



Peak#	Ret.Time	Area	Height	Area%
1	5.077	1560	316	0.282
2	6.711	551193	48149	99.718
Total		552753	48465	100.000

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Peak#	Ret.Time	Area	Height	Area%
1	7.924	12330995	1084282	47.875
2	9.863	13425555	961391	52.125
Total		25756550	2045673	100.000



Peak#	Ret.Time	Area	Height	Area%
1	7.889	6511364	429330	99.816
2	9.832	11998	1204	0.184
Total		6523362	430534	100.000



Peak#	Ret.Time	Area	Height	Area%
1	12.043	2674350	190824	50.169
2	12.727	2656339	179925	49.831
Total		5330689	370750	100.000

mV



Peak#	Ret. Time	Area	Height	Area%
1	11.984	1049308	76485	92.625
2	12.685	83547	6027	7.375
Total		1132855	82512	

3ia:













Peak#	Ret.Time	Area	Height	Area%
1	11.356	430341	33228	88.668
2	12.231	54999	4439	11.332
Total		485340	37667	100.000





Peak#	Ret.Time	Area	Height	Area%
1	6.624	2910557	240288	49.979
2	7.558	2913032	203906	50.021
Total		5823589	444194	100.000



Peak# Ret. Time	Area	Height	Area%
1 6.576	709175	62977	14.277
2 7.507	4258183	290529	85.723
Total	4967358	353507	



Peak# Ret.Time	Area	Height	Area%
1 18.773	14707960	794762	50.921
2 20.036	14176023	707717	49.079
Total	28883982	1502479	



Peak#	Ret.Time	Area	Height	Area
1	19.036	9875934	526835	85.449
2	20.332	1681746	84129	14.551
Total		11557681	610964	



	1 Cava	Rec. IIme	ni ea	nergne	ALCO/0
	1	10.529	1508389	126409	90.125
	2	13.716	165270	11339	9.875
I	Total		1673659	137748	
I					



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				1111 0 0010
1	13.733	846777	40824	50.301
2	17.339	836642	30988	49.699
Tatol		1683419	71811	



Peak#	Ret. Time	Area	Height	Area%
1	13.668	2180809	105090	88.647
2	17.285	279299	12001	11.353
Tatol		2460108	117091	



Peak#	Ret.Time	Area	Height	Area%
1	7.994	23805128	1869790	49.166
2	8.702	24612374	1790622	50.834
Total		48417503	3660412	



Peak#	Ret.Time	Area	Height	Area%
1	7.780	119044	15993	5.491
2	8.511	2048901	187050	94.509
Total		2167945	203043	100.000

4a:



Peak#	Ret.Time	Area	Height	Area%
1	33.046	6271782	179767	92.291
2	50.108	523849	10570	7.709
Total		6795631	190337	