

Pd Catalyzed Synthesis of Hetero 1,2-Interlinked C-Disaccharides by Coupling of *iodo* Glycals with Glycals

Bisma Rasool^{a,b}, Irshad Ahmad Zargar^{a,b}, Nazar Hussain^{*c}, Debaraj Mukherjee^{*a,b}

^aNatural Products and Medicinal Chemistry Division, Indian Institute of Integrative Medicine (CSIR-IIIM), Jammu-180001, India.

^bAcademy of Scientific and Innovative Research (AcSIR), Ghaziabad-201002, India.

^cDepartment of Medicinal Chemistry Institute of Medical Sciences Banaras Hindu University/221005 Varanasi (India)

^d Department of Chemical Sciences, Bose Institute Kolkata, EN 80, Sector V, Bidhan Nagar, Kolkata-700091, WB, India

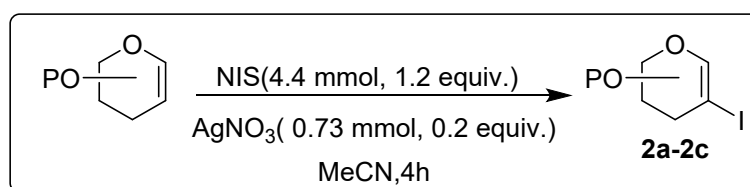
Table of Contents	
1. General Consideration	S3
2. General procedures	S3-S4
3. Characterization data: products	S4-S15
4. NMR spectra	S16-S56

1. General Consideration:

All reagents were purchased from commercial sources and used without treatment. Silica gel-coated aluminium plates were used for TLC. The products were purified by column chromatography on silica gel (60-120 mesh) using hexane-ethyl acetate as the eluent. ^1H NMR (400 MHz) spectra were recorded on a Bruker Avance 400 spectrometer in CDCl_3 and Acetone- D_6 , using CDCl_3 (for ^1H , $\delta = 7.26$) and acetone- d_6 (for ^1H , $\delta = 2.05$) as the internal standard. ^{13}C NMR (101 MHz) spectra were recorded on a Bruker Avance 400 spectrometer in CDCl_3 using CDCl_3 (for ^{13}C , $\delta = 77.0$) and acetone- d_6 (for ^{13}C , $\delta = 29.8$) as the internal standard. Chemical shifts are expressed in parts per million (δ ppm). The following abbreviations were used to explain the multiplicities: s = singlet, d = doublet, t = triplet, q = quartet, dd = doublet of the doublet, dt = doublet of triplet, m = multiplet, s br = singlet broad. The exact mass of all products was analysed by using HRMS having a QTOF analyser.

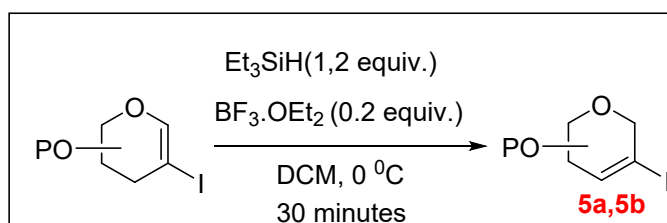
2. General procedures

2.1 General procedure for the Synthesis of 2-iodo glycals¹



By taking Tri-*O*-acetyl-D-glucal as an example, To a stirred solution of Tri-*O*-acetyl-D-glucal (1 g) in dry CH_3CN (10 mL) at 80 °C under N_2 atmosphere was added successively NIS (991 mg, 4.4 mmol) and AgNO_3 (124 mg, 0.73 mmol) and stirred for 4 h. On consumption of starting material (TLC monitoring), the reaction mixture was filtered through sintered funnel and the filtrate was evaporated to give a crude product which was purified by silica gel column chromatography (30% of EtOAc/hexane) to obtain the resulting 2-iodo glycal.

2.2 Procedure for the Synthesis of C2-C3 unsaturated 2-iodo glycals:



To a stirred solution of 2-iodo glycal (1 equiv.) in DCM at 0 °C, add Et_3SiH (1.2 equiv.) and $\text{BF}_3 \cdot \text{OEt}_2$ (0.2 equiv.) and stir for 30 minutes. On consumption of starting material (TLC monitoring), the reaction mixture was quenched with triethyl amine and then diluted with ethyl acetate and washed with brine (20 mL). The organic layer was dried over magnesium sulphate

and then evaporated to dryness. The residue was purified by column chromatography over silica gel (60-120 mesh) with hexane/ ethyl acetate as eluent to afford the desired compound.

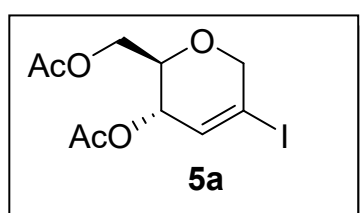
2.3 General experimental procedure for the synthesis of C2-C1 linked disaccharides (A):

To a solution of glycal (1 equiv.) in DMF (3 mL) taken in an oven dried sealed tube charged with magnetic bead, 2-iodo glycal (1.5 equiv.), Pd(OAc)₂ (0.10 equiv.) and PPh₃ (0.10 equiv.) were added. In the same solution, AgOAc (2 equiv.) was added and the reaction mixture was allowed to stir at 80 °C for 6 h. After the completion of the reaction, the reaction mixture was passed through celite, then filtered and extracted using ethyl acetate and washed with 20 mL of brine. The organic layer was dried over magnesium sulphate and evaporated to dryness. The residue was purified by column chromatography over silica gel (60-120 mesh) with hexane/ ethyl acetate as eluent to afford the desired compound.

2.4 General experimental procedure for the ring opening C2-C1 linked disaccharides (B):

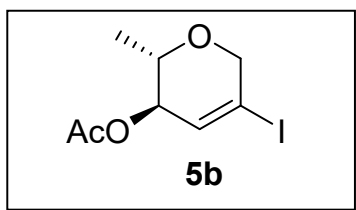
To a stirred solution of C2-C1 linked disaccharide (1 equiv.) in DCM at 0 °C, add BF₃.OEt₂ (0.2 equiv.) and stir for 30 minutes. On consumption of starting material (TLC monitoring), the reaction mixture was quenched with triethyl amine and then diluted with ethyl acetate and washed with brine (20 mL). The organic layer was dried over magnesium sulphate and then evaporated to dryness. The residue was purified by column chromatography over silica gel (60-120 mesh) with hexane/ ethyl acetate as eluent to afford the desired compound.

3. Characterization of the products:



Prepared by general procedure 3.2 using Tri-*O*-acetyl-2-iodo-D-glucal (200 mg, 0.5 mmol), Et₃SiH (1.2 equiv.) and BF₃.OEt₂ (0.2 equiv.) in solvent DCM. The reaction mixture was stirred at 0 °C for 30 minutes and then quenched with triethyl amine. The resulting mixture was extracted with EtOAc and the combined organic layer was washed with brine, dried over MgSO₄ and concentrated. The residue was purified by flash column chromatography (Hexane: EtOAc; 80:20) to afford **5a** as yellow oil in 71 % yield, 120 mg.

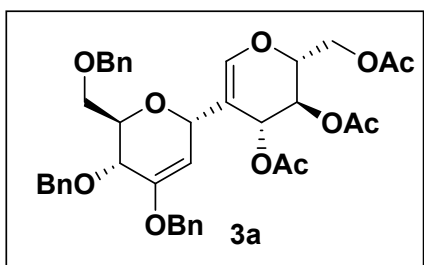
¹H NMR (400 MHz, CDCl₃) δ 6.39 (dd, *J* = 4.2, 2.2 Hz, 1H), 5.25 (dq, *J* = 8.1, 2.4 Hz, 1H), 4.24 (dd, *J* = 3.1, 2.2 Hz, 2H), 4.21 (d, *J* = 2.8 Hz, 1H), 4.19 – 4.15 (m, 1H), 3.80 (ddd, *J* = 8.4, 5.7, 2.9 Hz, 1H), 2.10 (s, 3H), 2.09 (s, 2H). **¹³C NMR (101 MHz, CDCl₃)** δ 170.80 (Cq), 170.04 (Cq), 133.46 (CH), 97.78 (Cq), 73.32 (CH), 72.98 (CH), 67.34 (CH), 62.87 (CH), 20.93 (CH₃), 20.81 (CH₃). **HRMS (ESI)** calcd for C₁₀H₁₇INO₅ [M+NH₄]⁺:358.0151, found 358.0153



Prepared by general procedure **3.2** using Tri-*O*-acetyl-2-*iodo*-D-glucal (200 mg, 0.5 mmol), Et₃SiH (1.2 equiv.) and BF₃·OEt₂ (0.2 equiv.) in solvent DCM. The reaction mixture was stirred at 0 °C for 30 minutes and then quenched with triethyl amine. The

resulting mixture was extracted with EtOAc and the combined organic layer was washed with brine, dried over MgSO₄ and concentrated. The residue was purified by flash column chromatography (Hexane: EtOAc; 85:15) to afford **5b** as yellow oil in 70 % yield, 98 mg.

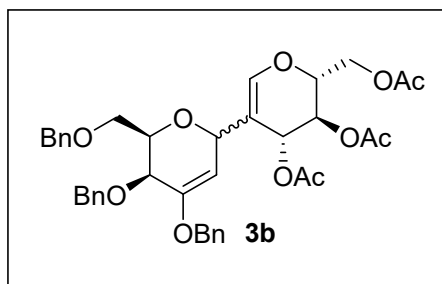
¹H NMR (400 MHz, CDCl₃) δ 6.36 (dd, *J* = 4.6, 2.1 Hz, 1H), 5.00 (ddd, *J* = 7.3, 5.0, 2.4 Hz, 1H), 4.18 (t, *J* = 2.2 Hz, 2H), 3.71 (dd, *J* = 7.2, 6.4 Hz, 1H), 2.09 (s, 3H), 1.24 (d, *J* = 6.4 Hz, 3H). **¹³C NMR (101 MHz, CDCl₃)** δ 170.30, 133.58, 98.62, 72.54, 72.35, 71.49, 21.03, 17.51. **HRMS (ESI)** calcd for C₈H₁₂IO₃ [M+H]⁺: 281.9753, found 281.9755.



Prepared by general procedure **A** using Tri-*O*-benzyl-D-glucal (50 mg, 0.12 mmol), Pd(OAc)₂ (0.006 mmol, 0.05 equiv.), PPh₃ (0.012 mmol, 0.10 equiv.), Tri-*O*-acetyl-2-*iodo*-D-glucal (0.179 mmol, 1.5 equiv.) and AgOAc (0.24 mmol, 2.0 equiv.). The reaction mixture was stirred at 80

°C for 6 h. The resulting mixture was extracted with EtOAc and the combined organic layer was washed with brine, dried over MgSO₄ and concentrated. The residue was purified by flash column chromatography (Hexane: EtOAc; 80:20) to afford **3a** as yellow oil in 75 % yield, 61.85 mg.

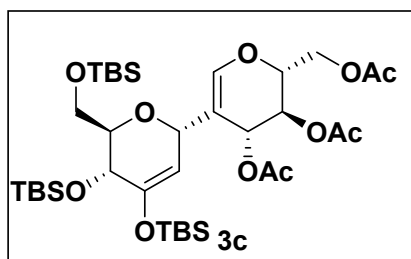
¹H NMR (400 MHz, CDCl₃) δ 7.39 – 7.37 (m, 4H), 7.35 – 7.31 (m, 5H), 7.27 – 7.24 (m, 4H), 7.19 (dt, *J* = 6.6, 2.4 Hz, 2H), 6.41 (s, 1H), 5.63 – 5.54 (m, 1H), 5.19 (dd, *J* = 6.5, 5.0 Hz, 1H), 4.87 (s, 3H), 4.74 (d, *J* = 4.1 Hz, 1H), 4.60 (d, *J* = 12.2 Hz, 1H), 4.48 (d, *J* = 4.2 Hz, 1H), 4.45 (d, *J* = 2.8 Hz, 1H), 4.38 (dd, *J* = 12.0, 6.3 Hz, 1H), 4.25 – 4.18 (m, 2H), 4.11 (dd, *J* = 12.0, 3.5 Hz, 1H), 3.75 (ddd, *J* = 8.6, 3.9, 2.6 Hz, 1H), 3.65 (dd, *J* = 10.5, 4.1 Hz, 1H), 3.60 (dd, *J* = 10.5, 2.5 Hz, 1H), 2.06 (s, *J* = 1.9 Hz, 3H), 2.04 (s, 3H), 1.96 (s, 3H). **¹³C NMR (101 MHz, CDCl₃)** δ 170.6 (Cq), 170.2 (Cq), 169.6 (Cq), 155.2 (Cq), 146.4 (CH), 138.2 (Cq), 138.2 (Cq), 136.7 (Cq), 128.6 (CH), 128.5 (CH), 128.4 (CH), 128.4 (CH), 128.0 (CH), 127.8 (CH), 127.7 (CH), 127.6 (CH), 127.2 (CH), 109.5 (Cq), 95.7 (CH), 74.8 (CH₂), 73.6 (CH), 73.4 (CH₂), 71.5 (CH), 70.7 (CH), 69.2 (CH₂), 69.0 (CH₂), 68.8 (CH), 67.4 (CH), 66.0 (CH), 61.3 (CH₂), 20.9 (CH₃), 20.8 (CH₃), 20.75 (CH₃). 6 CH resonances are missing in ¹³C NMR due to the overlap. **HRMS (ESI)** calcd for C₃₉H₄₃O₁₁ [M+H]⁺: 687.2805, found 687.2808.



Prepared by general procedure A using Tri-*O*-benzyl-D-galactal (50 mg, 0.12 mmol), Pd(OAc)₂ (0.006 mmol, 0.05 equiv.), PPh₃ (0.012 mmol, 0.10 equiv.), Tri-*O*-acetyl-2-iodo-D-glucal (0.179 mmol, 1.5 equiv.) and AgOAc (0.24 mmol, 2.0 equiv.). The reaction mixture was stirred at 80 °C for 6 h. The resulting mixture was

extracted with EtOAc and the combined organic layer was washed with brine, dried over MgSO₄ and concentrated. The residue was purified by flash column chromatography (Hexane: EtOAc; 80:20) to afford **3b** as yellow oil as (α/β;4/1) mixture in 70 % yield, 57 mg.

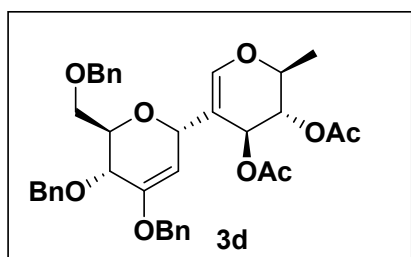
¹H NMR (400 MHz, CDCl₃) δ 7.30 – 7.27 (m, 4H), 7.25 – 7.23 (m, 5H), 7.21 – 7.18 (m, 6H), 6.19 (s, 1H), 5.56 (d, *J* = 5.2 Hz, 1H), 5.12 (dd, *J* = 6.8, 5.2 Hz, 1H), 4.89 (d, *J* = 3.6 Hz, 1H), 4.83 (d, *J* = 11.9 Hz, 1H), 4.77 (d, *J* = 11.7 Hz, 2H), 4.73 (d, *J* = 4.0 Hz, 1H), 4.58 (d, *J* = 11.7 Hz, 1H), 4.49 (d, *J* = 11.7 Hz, 1H), 4.41 (s, 1H), 4.31 (dd, *J* = 12.0, 6.1 Hz, 1H), 4.16 (dd, *J* = 6.5, 3.3 Hz, 1H), 4.05 (dd, *J* = 12.1, 3.3 Hz, 1H), 3.80 (dd, *J* = 6.2, 2.2 Hz, 1H), 3.75 (d, *J* = 2.2 Hz, 1H), 3.69 (dd, *J* = 9.7, 6.2 Hz, 1H), 3.57 (dd, *J* = 9.7, 6.2 Hz, 1H), 2.00 (s, 3H), 1.96 (s, 3H), 1.84 (s, 3H). **¹³C NMR (101 MHz, CDCl₃)** δ 170.6 (Cq), 170.1 (Cq), 169.8 (Cq), 154.1 (Cq), 145.4 (CH), 138.5 (Cq), 138.3 (Cq), 136.5 (Cq), 128.6 (CH), 128.6 (CH), 128.4 (CH), 128.2 (CH), 128.1 (CH), 128.0 (CH), 127.8 (CH), 127.6 (CH), 127.5 (CH), 127.4 (CH), 127.3 (CH), 109.4 (Cq), 97.0 (CH), 73.7 (CH), 73.7 (CH₂), 73.1 (CH₂), 71.6 (CH), 70.6 (CH), 69.5 (CH₂), 69.2 (CH₂), 68.4 (CH), 67.4 (CH), 66.1 (CH), 61.3 (CH₂), 20.8 (CH₃), 20.7 (CH₃), 20.6 (CH₃). 4 CH resonances are missing in ¹³C NMR due to the overlap. **HRMS(ESI)** calcd for C₃₉H₄₃O₁₁ [M+H]⁺: 687.2805, found 687.2813.



Prepared by general procedure A using Tri-*O*-tertbutyldimethylsilyl-D-glucal (50 mg, 0.10 mmol), Pd(OAc)₂ (0.005 mmol, 0.05 equiv.), PPh₃ (0.01 mmol, 0.10 equiv.), Tri-*O*-acetyl-2-iodo-D-glucal (0.15 mmol, 1.5 equiv.) and AgOAc (0.2 mmol, 2.0 equiv.). The reaction mixture was stirred at 80 °C for 6 h. The resulting mixture was extracted with EtOAc and the combined organic layer was washed with brine, dried over MgSO₄ and concentrated. The residue was purified by flash column chromatography (Hexane: EtOAc; 90:10) to afford **3c** as yellow oil in 65 % yield, 49 mg.

¹H NMR (400 MHz, CDCl₃) δ 6.45 (s, 1H), 5.52 (d, *J* = 5.5 Hz, 1H), 5.12 (dd, *J* = 7.4, 5.5 Hz, 1H), 4.70 (d, *J* = 3.8 Hz, 1H), 4.61 (d, *J* = 3.7 Hz, 1H), 4.31 (dd, *J* = 12.0, 5.9 Hz, 1H), 4.21 –

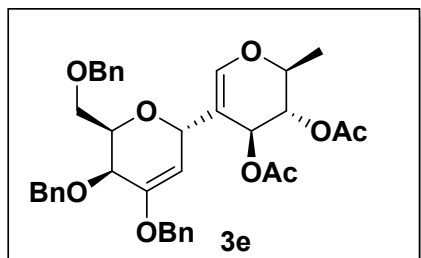
4.15 (m, 1H), 4.12 – 4.06 (m, 1H), 3.95 (dd, $J = 7.0, 1.3$ Hz, 1H), 3.64 (dd, $J = 5.4, 4.4$ Hz, 2H), 3.46 (dd, $J = 7.0, 4.7$ Hz, 1H), 2.00 (s, 3H), 1.95 (s, 2H), 1.95 (s, 3H), 0.86 (s, 9H), 0.80 (s, 9H), 0.79 (s, 9H), 0.12 (s, 3H), 0.10 (s, 3H), 0.01 (s, 3H), -0.00 (s, 3H), -0.04 (s, 3H), -0.05 (s, 3H). **^{13}C NMR (101 MHz, CDCl_3)** δ 170.6 (Cq), 170.1 (Cq), 169.8 (Cq), 151.7 (Cq), 145.9 (CH), 110.5 (Cq), 102.7 (CH), 75.4 (CH), 73.8 (CH), 68.4 (CH), 67.6 (CH), 66.7 (CH), 66.6 (CH), 62.8 (CH_2), 61.5 (CH_2), 26.3 (CH_3), 26.0 (CH_3), 26.0 (CH_3), 25.8 (CH_3), 25.8 (CH_3), 24.7 (CH_3), 22.7 (CH_3), 20.9 (CH_3), 20.8 (CH_3), 20.8 (CH_3), 18.9 (Cq), 18.4 (Cq), 18.3 (Cq), -3.4 (CH_3), -4.6 (CH_3), -5.4 (CH_3), -5.4 (CH_3). 1 CH_3 resonance is missing in ^{13}C NMR due to overlap. **HRMS** calcd for $\text{C}_{36}\text{H}_{67}\text{O}_{11}\text{Si}_3$ $[\text{M}+\text{H}]^+$: 759.3991, found 759.3994.



Prepared by general procedure A using Tri-*O*-benzyl-D-glucal (50 mg, 0.12 mmol), $\text{Pd}(\text{OAc})_2$ (0.002 mmol, 0.05 equiv.), PPh_3 (0.012 mmol, 0.10 equiv.), Di-*O*-acetyl-2-iodo-L-rhamnal (0.18 mmol, 1.5 equiv.) and AgOAc (0.24 mmol, 2.0 equiv.). The reaction mixture was stirred at 80 °C

for 6 h. The resulting mixture was extracted with EtOAc and the combined organic layer was washed with brine, dried over MgSO_4 and concentrated. The residue was purified by flash column chromatography (Hexane: EtOAc; 85:15) to afford **3d** as yellow oil in 68 % yield, 51 mg.

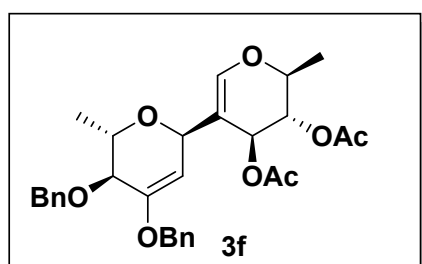
^1H NMR (400 MHz, CDCl_3) δ 7.38 (d, $J = 4.5$ Hz, 4H), 7.33 – 7.31 (m, 5H), 7.26 (d, $J = 1.8$ Hz, 4H), 7.22 (dd, $J = 7.3, 2.4$ Hz, 2H), 6.55 (s, 1H), 5.45 (d, $J = 3.9$ Hz, 1H), 4.94 (dd, $J = 4.8, 4.1$ Hz, 1H), 4.87 (d, $J = 5.4$ Hz, 2H), 4.86 – 4.83 (m, 2H), 4.74 (d, $J = 3.8$ Hz, 1H), 4.51 (d, $J = 11.1$ Hz, 1H), 4.47 (d, $J = 4.1$ Hz, 2H), 4.22 – 4.15 (m, 2H), 3.88 – 3.81 (m, 1H), 3.66 – 3.61 (m, 1H), 3.49 (dd, $J = 10.4, 2.9$ Hz, 1H), 2.07 (s, 3H), 1.90 (s, 3H), 1.29 (d, $J = 6.7$ Hz, 3H). **^{13}C NMR (101 MHz, CDCl_3)** δ 170.6 (Cq), 169.9 (Cq), 154.3 (Cq), 146.7 (CH), 138.3 (Cq), 138.0 (Cq), 136.8 (Cq), 128.5 (CH), 128.4 (CH), 128.4 (CH), 128.3 (CH), 128.0 (CH), 127.9 (CH), 127.8 (CH), 127.7 (CH), 127.7 (CH), 127.4 (CH), 127.3 (CH), 107.5 (Cq), 97.1 (CH), 74.3 (CH_2), 73.3 (CH_2), 72.2 (CH), 71.9 (CH), 71.5 (CH), 71.2 (CH), 70.8 (CH), 69.1 (CH_2), 68.7 (CH_2), 67.6 (CH), 21.01 (CH_3), 21.01 (CH_3), 16.0 (CH_3). 4 CH resonances are missing in ^{13}C NMR due to overlap. **HRMS (ESI)** calculated $\text{C}_{37}\text{H}_{41}\text{O}_9$ $[\text{M}+\text{H}]^+$: 629.2751, Found: 629.2755



Prepared by general procedure A using Tri-*O*-benzyl-D-galactal (50 mg, 0.12 mmol), $\text{Pd}(\text{OAc})_2$ (0.002 mmol, 0.05 equiv.), PPh_3 (0.012 mmol, 0.10 equiv.), Di-*O*-acetyl-2-

iodo-L- rhamnal (0.18 mmol, 1.5 equiv.) and AgOAc (0.24 mmol, 2.0 equiv.). The reaction mixture was stirred at 80 °C for 6 h. The resulting mixture was extracted with EtOAc and the combined organic layer was washed with brine, dried over MgSO₄ and concentrated. The residue was purified by flash column chromatography (Hexane: EtOAc; 85:15) to afford **3e** as yellow oil in 70% yield, 52 mg.

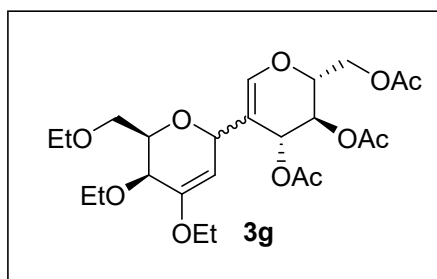
¹H NMR (400 MHz, CDCl₃) δ 7.38 (d, *J* = 4.5 Hz, 4H), 7.33 – 7.31 (m, 5H), 7.26 (d, *J* = 1.8 Hz, 4H), 7.24 – 7.21 (m, 2H), 6.55 (s, 1H), 5.45 (d, *J* = 3.9 Hz, 1H), 4.94 (dd, *J* = 4.8, 4.1 Hz, 1H), 4.87 (d, *J* = 5.4 Hz, 2H), 4.85 – 4.83 (m, 2H), 4.74 (d, *J* = 3.8 Hz, 1H), 4.52 (d, *J* = 6.5 Hz, 1H), 4.50 – 4.47 (m, 2H), 4.22 – 4.14 (m, 2H), 3.88 – 3.81 (m, 1H), 3.63 (dd, *J* = 10.4, 4.2 Hz, 1H), 3.49 (dd, *J* = 10.4, 2.9 Hz, 1H), 2.07 (s, 3H), 1.90 (s, 3H), 1.29 (d, *J* = 6.7 Hz, 3H). **¹³C NMR (101 MHz, CDCl₃)** δ 170.6 (Cq), 169.9 (Cq), 153.5 (Cq), 145.9 (CH), 138.6 (Cq), 138.1 (Cq), 136.6 (Cq), 128.6 (CH), 128.4 (CH), 128.2 (CH), 128.1 (CH), 128.0 (CH), 127.6 (CH), 127.5 (CH), 127.4 (CH), 107.2 (Cq), 97.9 (CH), 73.3 (CH₂), 73.1 (CH₂), 71.95 (CH), 72.0 (CH), 71.7 (CH), 71.5 (CH), 70.2 (CH), 69.2 (CH₂), 69.0 (CH₂), 67.8 (CH), 21.0 (CH₃), 21.0 (CH₃), 16.1 (CH₃). 7 CH resonances are missing in ¹³C NMR due to overlap. **HRMS (ESI)** calculated C₃₇H₄₁O₉ [M+H]⁺: 629.2772, Found; 629.2774.



Prepared by general procedure **A** using Di-*O*-benzyl-L-rhamnal (50 mg, 0.16 mmol), Pd(OAc)₂ (0.007 mmol, 0.05 equiv.), PPh₃ (0.016 mmol, 0.10 equiv.), Di-*O*-acetyl-2-*iodo*-L- rhamnal (0.24 mmol, 1.5 equiv.) and AgOAc (0.32 mmol, 2.0 equiv.). The reaction mixture was stirred at 80 °C for 6 h. The resulting mixture was extracted with EtOAc and the combined organic layer was washed with brine, dried over MgSO₄ and concentrated. The residue was purified by flash column chromatography (Hexane: EtOAc; 87:13) to afford **3f** as yellow oil in 72 % yield, 60 mg.

¹H NMR (400 MHz, CDCl₃) δ 7.38 (dd, *J* = 3.7, 1.8 Hz, 4H), 7.32 – 7.30 (m, 4H), 7.29 (dd, *J* = 2.7, 1.5 Hz, 1H), 7.26 (s, 1H), 6.36 (s, 1H), 5.56 (d, *J* = 5.5 Hz, 1H), 4.98 (dd, *J* = 7.3, 5.7 Hz, 1H), 4.91 (d, *J* = 11.0 Hz, 1H), 4.87 (d, *J* = 1.1 Hz, 2H), 4.76 (d, *J* = 3.9 Hz, 1H), 4.74 (d, *J* = 3.9 Hz, 1H), 4.61 (d, *J* = 11.1 Hz, 1H), 4.13 – 4.08 (m, 1H), 3.77 (dd, *J* = 7.6, 6.0 Hz, 1H), 3.75 – 3.72 (m, 1H), 2.06 (s, 2H), 2.03 (s, 2H), 1.28 (d, *J* = 6.6 Hz, 3H), 1.23 (d, *J* = 5.9 Hz, 3H). **¹³C NMR (101 MHz, CDCl₃)** δ 170.5 (Cq), 170.1 (Cq), 154.6 (Cq), 146.5 (CH), 138.3 (Cq), 136.8 (Cq), 128.6 (CH), 128.4 (CH), 128.4 (CH), 127.9 (CH), 127.8 (CH), 127.3 (CH),

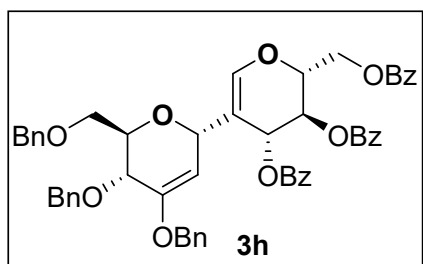
109.5 (Cq), 96.3 (CH), 77.3 (CH₂), 74.5 (CH₂), 72.3 (CH), 72.2 (CH), 69.1 (CH), 68.3 (CH), 67.2 (CH), 67.0 (CH), 20.9 (CH₃), 20.9 (CH₃), 18.3 (CH₃), 16.5 (CH₃). 4 CH resonances are missing in ¹³C NMR due to overlapping. **HRMS (ESI)** calculated C₃₀H₃₅O₈ [M+H]⁺: 523.2332, Found; 523.2335.



Prepared by general procedure A using Tri-*O*-ethyl-D-glucal (50 mg, 0.21 mmol), Pd(OAc)₂ (0.010 mmol, 0.05 equiv.), PPh₃ (0.021 mmol, 0.10 equiv.), Tri-*O*-acetyl-2-iodo-D-glucal (0.31 mmol, 1.5 equiv.) and AgOAc (0.42 mmol, 2.0 equiv.). The reaction mixture was stirred at 80 °C for 6 h. The resulting mixture was extracted with

EtOAc and the combined organic layer was washed with brine, dried over MgSO₄ and concentrated. The residue was purified by flash column chromatography (Hexane: EtOAc; 88:12) to afford **3g** as (α/β;4/1) as yellow oil in 75 % yield, 79 mg.

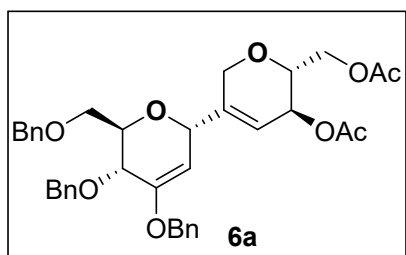
¹H NMR (400 MHz, CDCl₃) δ 6.51 (s, 1H), 5.49 (d, *J* = 3.5 Hz, 1H), 5.13 (ddd, *J* = 6.9, 5.1, 2.1 Hz, 1H), 4.77 (s, 1H), 4.55 – 4.51 (m, 1H), 4.37 – 4.30 (m, 2H), 4.19 (d, *J* = 2.3 Hz, 1H), 4.07 (dd, *J* = 12.1, 3.1 Hz, 1H), 3.90 (d, *J* = 7.7 Hz, 1H), 3.83 (ddd, *J* = 9.2, 4.8, 2.2 Hz, 1H), 3.70 – 3.66 (m, 2H), 3.53 (s, 3H), 3.48 – 3.42 (m, 2H), 2.01 (s, 3H), 1.98 (s, 3H), 1.97 (s, 3H), 1.18 (d, 3H), 1.14 (d, *J* = 1.9 Hz, 3H), 1.12 (d, *J* = 1.9 Hz, 3H). **¹³C NMR (101 MHz, CDCl₃)** δ 170.6 (Cq), 170.2 (Cq), 169.7 (Cq), 155.6 (Cq), 146.2 (CH), 109.9 (Cq), 94.0 (CH), 73.6 (CH), 71.5 (CH), 70.6 (CH), 69.4 (CH₂), 68.7 (CH), 67.9 (CH₂), 67.5 (CH), 67.0 (CH₂), 66.1 (CH), 62.7 (CH₂), 61.3 (CH₂), 20.9 (CH₂), 20.8 (CH₂), 20.7 (CH₂), 15.6 (CH₃), 15.1 (CH₃), 14.5 (CH₃). **HRMS (ESI)** calculated C₂₄H₃₇O₁₁ [M+H]⁺: 501.2336, Found: 501.2338.



Prepared by general procedure A using Tri-*O*-benzyl-D-glucal (50 mg, 0.12 mmol), Pd(OAc)₂ (0.006 mmol, 0.05 equiv.), PPh₃ (0.012 mmol, 0.10 equiv.), Tri-*O*-benzoyl-2-iodo-D-glucal (0.18 mmol, 1.5 equiv.) and AgOAc (0.24 mmol, 2.0 equiv.). The reaction mixture was stirred at 80

°C for 6 h. The resulting mixture was extracted with EtOAc and the combined organic layer was washed with brine, dried over MgSO₄ and concentrated. The residue was purified by flash column chromatography (Hexane: EtOAc; 88:12) to afford **3h** as yellow oil in 75 % yield, 92 mg.

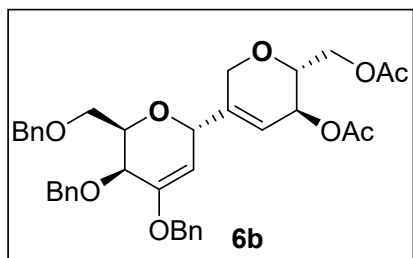
¹H NMR (400 MHz, Acetone d₆) δ 7.93 – 7.82 (m, 7H), 7.53 – 7.47 (m, 3H), 7.38 – 7.31 (m, 7H), 7.31 – 7.22 (m, 4H), 7.19 – 7.12 (m, H), 6.62 (s, 1H), 5.97 (d, *J* = 4.1 Hz, 1H), 5.68 (t, *J* = 4.3 Hz, 1H), 4.97 (d, *J* = 4.1 Hz, 1H), 4.90 – 4.86 (m, 2H), 4.83 (d, *J* = 12.0 Hz, 1H), 4.78 (d, *J* = 11.2 Hz, 1H), 4.71 (dt, *J* = 11.2, 5.3 Hz, 2H), 4.52 (dd, *J* = 11.1, 3.7 Hz, 1H), 4.33 (d, *J* = 12.3 Hz, 1H), 4.28 (d, *J* = 12.3 Hz, 1H), 3.99 (dd, *J* = 8.5, 1.0 Hz, 1H), 3.86 – 3.81 (m, 1H), 3.45 – 3.42 (m, 1H). **¹³C NMR (101 MHz, Acetone d₆)** δ 164.5 (Cq), 164.7 (Cq), 162.4 (Cq), 154.8 (Cq), 146.2 (CH), 138.9 (Cq), 138.8 (Cq), 137.4 (Cq), 138.9 (Cq), 138.8 (Cq), 137.4 (Cq), 133.4 (CH), 133.3 (CH), 133.2 (CH), 129.6 (CH), 129.5 (CH), 129.4 (CH), 128.6 (CH), 128.5 (CH), 128.4 (CH), 128.1 (CH), 128.0 (CH), 127.7 (CH), 127.6 (CH), 127.4 (CH), 127.3 (CH), 127.1 (CH), 109.6 (Cq), 96.5 (CH), 73.5 (CH₂), 73.2 (CH), 72.8 (CH₂), 71.7 (CH), 71.2 (CH), 69.6 (CH₂), 68.9 (CH), 68.8 (CH₂), 67.7 (CH), 65.2 (CH), 61.8 (CH₂). 14 CH resonances are missing in ¹³C NMR due to overlapping. **HRMS (ESI)** calculated C₅₄H₄₉O₁₁ [M+H]⁺: 872.3197, Found: 872.3198.



Prepared by general procedure A using Tri-O-benzyl-D-glucal (50 mg, 0.12 mmol), Pd(OAc)₂ (0.006 mmol, 0.05 equiv.), PPh₃ (0.012 mmol, 0.10 equiv.), Di-O-acetyl-2-iodo-D-glucal (0.18 mmol, 1.5 equiv.) and AgOAc (0.24 mmol, 2.0 equiv.). The reaction mixture was stirred at 80 °C

for 6 hrs. The resulting mixture was extracted with EtOAc and the combined organic layer was washed with brine, dried over MgSO₄ and concentrated. The residue was purified by flash column chromatography (Hexane: EtOAc; 85:15) to afford 6a as yellow oil in 75 % yield, 56 mg.

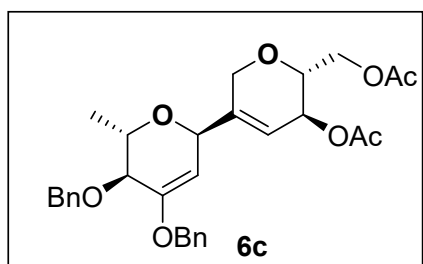
¹H NMR (400 MHz, CDCl₃) δ 7.39 – 7.36 (m, 5H), 7.33 – 7.31 (m, 5H), 7.25 (d, *J* = 1.1 Hz, 3H), 7.22 – 7.18 (m, 2H), 5.63 (s, 1H), 5.29 (d, *J* = 8.5 Hz, 1H), 4.89 – 4.84 (m, 2H), 4.73 (d, *J* = 3.8 Hz, 1H), 4.70 (d, *J* = 3.6 Hz, 1H), 4.56 (d, *J* = 12.2 Hz, 1H), 4.46 (dd, *J* = 16.7, 11.8 Hz, 3H), 4.22 – 4.18 (m, 2H), 4.17 – 4.13 (m, 2H), 3.83 – 3.78 (m, 1H), 3.64 – 3.58 (m, 2H), 3.55 (dd, *J* = 10.4, 3.1 Hz, 1H), 2.10 (s, 3H), 2.07 (s, *J* = 3.8 Hz, 3H). **¹³C NMR (101 MHz, CDCl₃)** δ 171.0 (Cq), 170.2 (Cq), 154.5 (Cq), 140.9 (Cq), 138.1 (Cq), 137.9 (Cq), 136.6 (Cq), 128.5 (CH), 128.4 (CH), 128.3 (CH), 128.3 (CH), 127.9 (CH), 127.9 (CH), 127.7 (CH), 127.3 (CH), 122.9 (CH), 96.0 (CH), 74.0 (CH₂), 73.9 (CH), 73.3 (CH₂), 72.3 (CH), 71.5 (CH), 71.1 (CH), 69.2 (CH₂), 68.5 (CH₂), 66.0 (CH₂), 65.3 (CH), 63.12 (CH₂), 21.1 (CH₃), 20.9 (CH₃). 7 CH resonances are missing in ¹³C NMR due to overlapping. **HRMS (ESI)** calculated C₃₇H₄₄NO₉ [M+NH₄]⁺: 646.3016, Found; 646.3017.



Prepared by general procedure A using Tri-*O*-benzyl-D-galactal (50 mg, 0.12 mmol), Pd(OAc)₂ (0.006 mmol, 0.05 equiv.), PPh₃ (0.012 mmol, 0.10 equiv.), Di-*O*-acetyl-2-*iodo*-D-glucal (0.18 mmol, 1.5 equiv.) and AgOAc (0.24 mmol, 2.0 equiv.). The reaction mixture was stirred at 80 °C

for 6 h. The resulting mixture was extracted with EtOAc and the combined organic layer was washed with brine, dried over MgSO₄ and concentrated. The residue was purified by flash column chromatography (Hexane: EtOAc; 85:15) to afford **6b** as yellow oil in 70 % yield, 53 mg.

¹H NMR (400 MHz, CDCl₃) δ 7.39 – 7.26 (m, 15H), 5.52 (s, J = 9.7 Hz, 1H), 5.28 (d, J = 8.0 Hz, 1H), 4.91 (d, J = 11.9 Hz, 1H), 4.83 (dd, J = 15.5, 3.6 Hz, 3H), 4.77 (s, 1H), 4.65 (d, J = 11.6 Hz, 1H), 4.54 – 4.45 (m, 3H), 4.23 (dd, J = 11.6, 2.0 Hz, 1H), 4.17 (dd, J = 12.3, 5.6 Hz, 2H), 3.86 (td, J = 6.2, 2.0 Hz, 1H), 3.83 (d, J = 2.1 Hz, 1H), 3.71 – 3.60 (m, 3H), 2.10 (s, 3H), 2.08 (s, J = 0.6 Hz, 3H). **¹³C NMR (101 MHz, CDCl₃)** δ 171.0 (Cq), 170.3 (Cq), 154.0 (Cq), 140.6 (Cq), 138.4 (Cq), 138.1 (Cq), 136.5 (Cq), 128.6 (CH), 128.4 (CH), 128.2 (CH), 128.1 (CH), 127.9 (CH), 127.7 (CH), 127.6 (CH), 127.6 (CH), 127.3 (CH), 122.6 (CH), 96.7 (CH), 73.9 (CH), 73.5 (CH₂), 73.2 (CH₂), 71.7 (CH), 71.4 (CH), 71.1 (CH), 69.2 (CH₂), 69.2 (CH₂), 66.3 (CH₂), 65.4 (CH), 63.1 (CH), 21.1 (CH₃), 20.9 (CH₃). 6 CH resonances are missing in ¹³C NMR due to overlapping. **HRMS (ESI)** calculated C₃₇H₄₁O₉ [M+H]⁺: 629.2751, Found; 629.2755.

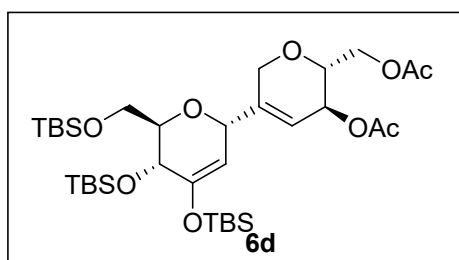


Prepared by general procedure A using Di-*O*-benzyl-L-rhamnal (50 mg, 0.16 mmol), Pd(OAc)₂ (0.008 mmol, 0.05 equiv.), PPh₃ (0.016 mmol, 0.10 equiv.), Di-*O*-acetyl-2-*iodo*-D-glucal (0.192 mmol, 1.5 equiv.) and AgOAc (0.32 mol, 2.0 equiv.). The reaction mixture was stirred at 80 °C

for 6 h. The resulting mixture was extracted with EtOAc and the combined organic layer was washed with brine, dried over MgSO₄ and concentrated. The residue was purified by flash column chromatography (Hexane: EtOAc; 85:15) to afford **6c** as yellow oil in 72% yield, 60 mg.

¹H NMR (400 MHz, CDCl₃) δ 7.37 (ddd, J = 5.3, 3.7, 2.1 Hz, 4H), 7.32 – 7.26 (m, 6H), 5.67 – 5.64 (m, 1H), 5.29 – 5.24 (m, 1H), 4.87 – 4.83 (m, 3H), 4.78 (d, J = 3.4 Hz, 1H), 4.69 – 4.66 (m, 1H), 4.63 (d, J = 11.3 Hz, 1H), 4.27 (d, J = 1.7 Hz, 2H), 4.22 (dd, J = 12.1, 2.8 Hz, 1H),

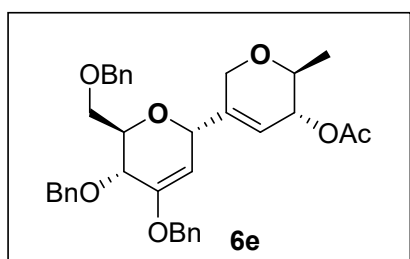
4.15 (dd, $J = 12.1, 6.0$ Hz, 1H), 3.91 – 3.86 (m, 1H), 3.72 (dd, $J = 5.9, 1.3$ Hz, 1H), 3.68 (ddd, $J = 8.6, 6.0, 2.8$ Hz, 1H), 2.10 (s, 3H), 2.08 (s, $J = 1.5$ Hz, 3H), 1.23 (d, $J = 6.4$ Hz, 3H). **^{13}C NMR (101 MHz, CDCl_3)** δ 171.0 (Cq), 170.5 (Cq), 153.6 (Cq), 141.3 (Cq), 138.2 (Cq), 136.6 (Cq), 128.5 (CH), 128.4 (CH), 128.2 (CH), 128.0 (CH), 127.8 (CH), 127.4 (CH), 122.2 (CH), 96.3 (CH), 76.5 (CH), 74.0 (CH), 73.6 (CH_2), 71.4 (CH), 69.6 (CH), 69.1 (CH_2), 66.3 (CH_2), 65.4 (CH), 63.2 (CH_2), 21.2 (CH_3), 20.9 (CH_3), 17.6 (CH_3). **HRMS (ESI)** calculated $\text{C}_{30}\text{H}_{38}\text{NO}_8$ $[\text{M}+\text{NH}_4]^+$: 540.2597, Found; 540.2597.



Prepared by general procedure A using Tri-*O*-tertbutyldimethylsilyl-D-glucal (50 mg, 0.10 mmol), $\text{Pd}(\text{OAc})_2$ (0.005 mmol, 0.05 equiv.), PPh_3 (0.01 mmol, 0.10 equiv.), Di-*O*-acetyl-2-iodo-D-glucal (0.15 mmol, 1.5 equiv.) and AgOAc (0.2 mmol, 2.0 equiv.). The reaction mixture was stirred at 80 °C for 6 h. The

resulting mixture was extracted with EtOAc and the combined organic layer was washed with brine, dried over MgSO_4 and concentrated. The residue was purified by flash column chromatography (Hexane: EtOAc; 90:10) to afford **6d** as yellow oil in 65 % yield, 45 mg.

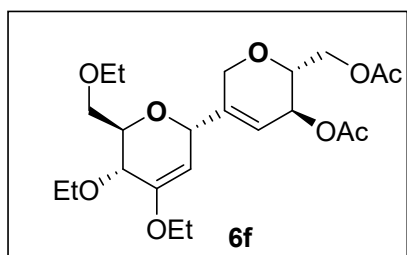
^1H NMR (400 MHz, CDCl_3) δ 5.62 (s, 1H), 5.20 – 5.14 (m, 2H), 4.56 (d, $J = 2.8$ Hz, 1H), 4.51 (s, 1H), 4.23 (d, $J = 5.1$ Hz, 1H), 4.20 – 4.19 (m, 1H), 4.15 (dd, $J = 5.2, 2.3$ Hz, 1H), 4.12 (d, $J = 2.8$ Hz, 1H), 4.09 (dd, $J = 5.7, 1.4$ Hz, 1H), 4.08 – 4.01 (m, 2H), 3.88 – 3.86 (m, 1H), 3.61 – 3.58 (m, 2H), 2.02 (s, 3H), 1.98 (s, 3H), 1.96 (s, 2H), 0.85 (s, 8H), 0.80 (s, 7H), 0.79 (s, 9H), 0.10 (d, $J = 5.2$ Hz, 6H), 0.00 (d, $J = 2.8$ Hz, 6H), -0.05 (d, $J = 1.6$ Hz, 6H). **^{13}C NMR (101 MHz, CDCl_3)** δ 171.0 (Cq), 170.5 (Cq), 150.1 (Cq), 142.1 (Cq), 121.1 (CH), 103.0 (CH), 78.8 (CH), 74.0 (CH), 72.5 (CH), 67.4 (CH), 66.8 (CH), 65.7 (CH), 65.1 (CH_2), 63.3 (CH_2), 62.0 (CH_2), 31.6 (CH_3), 26.0 (CH_3), 25.9 (CH_3), 25.7 (CH_3), 22.7 (CH_3), 21.1 (CH_3), 20.9 (CH_3), 18.4 (Cq), 18.2 (Cq), 18.1 (Cq), 14.1 (CH_3), -3.9 (CH_3), -4.1 (CH_3), -4.1 (CH_3), -4.7 (CH_3), -5.4 (CH_3), -5.4 (CH_3). 3 CH_3 resonances are missing in ^{13}C NMR due to overlap. **HRMS (ESI)** calculated $\text{C}_{34}\text{H}_{65}\text{O}_9\text{Si}_3$ $[\text{M}+\text{H}]^+$: 701.3936, Found; 701.3937.



Prepared by general procedure A using Tri-*O*-benzyl-D-glucal (50 mg, 0.12 mmol), $\text{Pd}(\text{OAc})_2$ (0.006 mmol, 0.05 equiv.), PPh_3 (0.012 mmol, 0.10 equiv.), 4-*O*-acetyl-2-iodo-L-rhamnal (0.18 mmol, 1.5 equiv.) and AgOAc (0.24 mmol, 2.0 equiv.). The reaction mixture was stirred at 80 °C for 6

h. The resulting mixture was extracted with EtOAc and the combined organic layer was washed with brine, dried over MgSO_4 and concentrated. The residue was purified by flash column chromatography (Hexane: EtOAc; 90:10) to afford **6e** as yellow oil in 68 % yield, 46 mg.

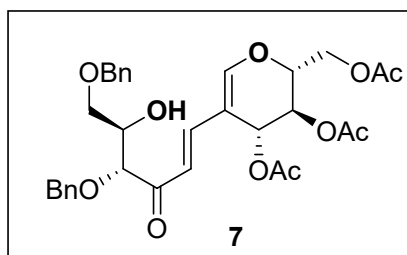
^1H NMR (400 MHz, CDCl_3) δ 7.40 – 7.37 (m, 4H), 7.33 (d, J = 4.0 Hz, 5H), 7.26 (dd, J = 4.8, 2.0 Hz, 4H), 7.19 (dd, J = 6.7, 2.9 Hz, 2H), 5.61 (s, 1H), 5.05 – 5.00 (m, 1H), 4.87 – 4.84 (m, 2H), 4.81 (d, J = 1.9 Hz, 1H), 4.74 (s, 1H), 4.62 – 4.57 (m, 1H), 4.49 (d, J = 4.0 Hz, 1H), 4.34 – 4.29 (m, 1H), 4.20 – 4.16 (m, 1H), 4.15 – 4.11 (m, 3H), 3.82 (ddd, J = 7.6, 4.5, 3.1 Hz, 1H), 3.62 (d, J = 4.6 Hz, 1H), 3.60 – 3.55 (m, 2H), 2.05 (s, 3H), 1.22 (d, J = 6.3 Hz, 3H). **^{13}C NMR (101 MHz, CDCl_3)** δ 170.8 (Cq), 154.3 (Cq), 140.9 (Cq), 138.0 (Cq), 138.0 (Cq), 136.6 (Cq), 128.5 (CH), 128.4 (CH), 128.4 (CH), 127.9 (CH), 127.8 (CH), 127.7 (CH), 127.4 (CH), 123.1 (CH), 96.0 (CH), 74.2 (CH_2), 73.4 (CH_2), 72.3 (CH), 72.1 (CH), 72.0 (CH), 71.4 (CH), 70.7 (CH), 69.2 (CH_2), 68.7 (CH_2), 65.9 (CH_2), 21.3 (CH_3), 17.7 (CH_3). 8 CH resonances are missing in ^{13}C NMR due to overlapping. **HRMS (ESI)** calculated $\text{C}_{35}\text{H}_{42}\text{NO}_7$ $[\text{M}+\text{NH}_4]^+$: 606.2961, Found; 606.2963.



Prepared by general procedure **A** using Tri-*O*-ethyl-D-glucal (50 mg, 0.12 mmol), $\text{Pd}(\text{OAc})_2$ (0.006 mmol, 0.05 equiv.), PPh_3 (0.012 mmol, 0.10 equiv.), Di-*O*-acetyl-2-iodo-D-glucal (0.18 mmol, 1.5 equiv.) and AgOAc (0.24 mmol, 2.0 equiv.). The reaction mixture was stirred at 80 °C for 6 h.

The resulting mixture was extracted with EtOAc and the combined organic layer was washed with brine, dried over MgSO_4 and concentrated. The residue was purified by flash column chromatography (Hexane: EtOAc; 85:15) to afford **6f** as yellow oil in 65 % yield, 62 mg.

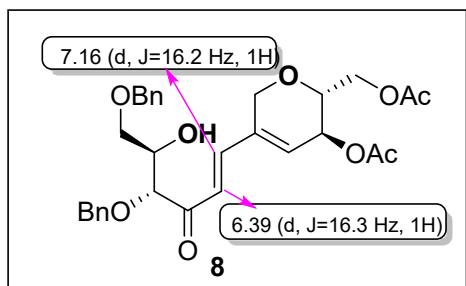
^1H NMR (400 MHz, CDCl_3) δ 5.76 (s, 1H), 5.30 (dd, J = 5.7, 2.4 Hz, 1H), 4.68 (d, J = 2.8 Hz, 1H), 4.60 (d, J = 3.8 Hz, 1H), 4.48 – 4.41 (m, 1H), 4.22 (dd, J = 5.3, 2.8 Hz, 1H), 4.19 (dd, J = 5.6, 4.5 Hz, 2H), 3.91 – 3.86 (m, 1H), 3.86 – 3.82 (m, 1H), 3.81 – 3.73 (m, 2H), 3.73 – 3.68 (m, 1H), 3.65 (ddd, J = 8.6, 5.6, 3.1 Hz, 1H), 3.60 – 3.56 (m, 3H), 3.55 – 3.45 (m, 2H), 2.11 (s, 3H), 2.08 (s, 3H), 1.34 (t, J = 7.0 Hz, 3H), 1.22 (t, J = 4.4 Hz, 3H), 1.19 (t, J = 5.9, 2.9 Hz, 3H). **^{13}C NMR (101 MHz, CDCl_3)** δ 170.9 (Cq), 170.3 (Cq), 155.0 (Cq), 141.3 (Cq), 122.5 (CH), 94.4 (CH), 73.9 (CH), 72.3 (CH), 71.6 (CH), 71.4 (CH), 69.3 (CH_2), 67.1 (CH_2), 66.9 (CH_2), 66.0 (CH_2), 65.5 (CH), 63.2 (CH_2), 62.7 (CH_2), 21.1 (CH_3), 20.9 (CH_3), 15.6 (CH_3), 15.1 (CH_3), 14.5 (CH_3). **HRMS (ESI)** calculated $\text{C}_{22}\text{H}_{35}\text{O}_9$ $[\text{M}+\text{H}]^+$: 444.2281, Found; 444.2283.



Prepared by general procedure **B** using **3a** (50 mg, 0.072 mmol), $\text{BF}_3 \cdot \text{OEt}_2$ (0.014 mmol, 0.20 equiv.). The reaction mixture was stirred at 0 °C for 30 minutes. The resulting mixture was quenched with Et_3N , and then extracted with EtOAc and the combined organic layer was washed with

brine, dried over MgSO_4 and concentrated. The residue was purified by flash column chromatography (Hexane: EtOAc; 50:50) to afford **7** as gummy liquid in 76 % yield, 33 mg.

^1H NMR (400 MHz, CDCl_3) δ 7.37 – 7.35 (m, 1H), 7.34 (d, J = 2.1 Hz, 1H), 7.33 – 7.32 (m, 2H), 7.30 (d, J = 2.8 Hz, 3H), 7.29 – 7.26 (m, 4H), 7.04 (s, 1H), 6.42 (d, J = 15.8 Hz, 1H), 5.65 – 5.60 (m, 1H), 5.19 – 5.14 (m, 1H), 4.70 – 4.61 (m, 1H), 4.54 – 4.51 (m, 1H), 4.51 – 4.48 (m, 2H), 4.46 (d, J = 3.4 Hz, 1H), 4.43 (dd, J = 7.6, 4.4 Hz, 1H), 4.19 (dd, J = 12.0, 4.2 Hz, 1H), 4.05 (t, J = 4.7 Hz, 1H), 4.01 (d, J = 6.1 Hz, 1H), 3.59 (d, J = 4.6 Hz, 2H), 2.10 (s, 3H), 2.09 (s, 3H), 2.00 (s, 3H). **^{13}C NMR (101 MHz, CDCl_3)** δ 199.0 (Cq), 170.4 (Cq), 170.0 (Cq), 169.4 (Cq), 154.0 (CH), 140.5 (CH), 137.8 (Cq), 137.2 (Cq), 129.1 (CH), 128.8 (CH), 128.6 (CH), 128.5 (CH), 128.4 (CH), 128.1 (CH), 128.0 (CH), 127.8 (CH), 127.8 (CH), 127.0 (CH), 118.1 (CH), 110.1 (Cq), 83.8 (CH), 74.6 (CH), 73.4 (CH_2), 72.9 (CH_2), 71.3 (CH), 70.2 (CH_2), 66.5 (CH), 62.7 (CH), 61.1 (CH_2), 20.8 (CH_3), 20.7 (CH_3), 20.6 (CH_3). **HRMS (ESI)** calculated $\text{C}_{32}\text{H}_{37}\text{O}_{11}$ $[\text{M}+\text{H}]^+$: 597.2336, Found; 597.2339.

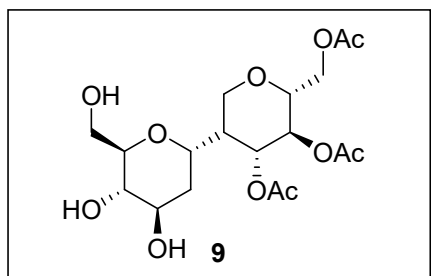


Prepared by general procedure **B** using **6b** (50 mg, 0.079 mmol), $\text{BF}_3 \cdot \text{OEt}_2$ (0.015 mmol, 0.20 equiv.). The reaction mixture was stirred at 0 °C for 30 minutes. The resulting mixture was quenched with Et_3N , and then extracted with EtOAc and the combined organic layer was washed with brine, dried over MgSO_4 and

concentrated. The residue was purified by flash column chromatography (Hexane: EtOAc; 50:50) to afford **8** as gummy liquid in 70% yield, 30 mg.

^1H NMR (400 MHz, CDCl_3) δ 7.36 – 7.32 (m, 6H), 7.29 (dd, J = 6.1, 3.7 Hz, 5H), 7.16 (d, J = 16.2 Hz, 1H), 6.39 (d, J = 16.3 Hz, 1H), 6.12 (s, 1H), 5.39 (d, J = 6.5 Hz, 1H), 4.64 (d, J = 11.5 Hz, 1H), 4.49 (d, J = 3.5 Hz, 2H), 4.47 – 4.43 (m, 1H), 4.32 – 4.17 (m, 4H), 4.14 – 4.09 (m, 1H), 3.70 (ddd, J = 8.4, 5.7, 2.7 Hz, 1H), 3.54 (dd, J = 10.5, 4.8 Hz, 2H), 2.12 (s, 3H), 2.11 (s, 3H). **^{13}C NMR (101 MHz, CDCl_3)** δ 199.6 (Cq), 170.9 (Cq), 170.2 (Cq), 140.0 (CH), 137.7 (Cq), 136.8 (Cq), 136.7 (Cq), 133.2 (CH), 128.6 (CH), 128.4 (CH), 128.4 (CH), 128.3 (CH),

127.9 (CH), 127.8 (CH), 121.8 (CH), 83.5 (CH), 73.9 (CH), 73.5 (CH₂), 73.3 (CH₂), 71.4 (CH), 70.3 (CH₂), 65.5 (CH), 64.8 (CH₂), 63.0 (CH₂), 21.0 (CH₃), 20.8 (CH₃). 4 CH resonances are missing in ¹³C NMR due to overlapping. **HRMS (ESI)** calculated C₃₀H₃₅O₉ [M+H]⁺: 540.2306, Found; 540.2308.



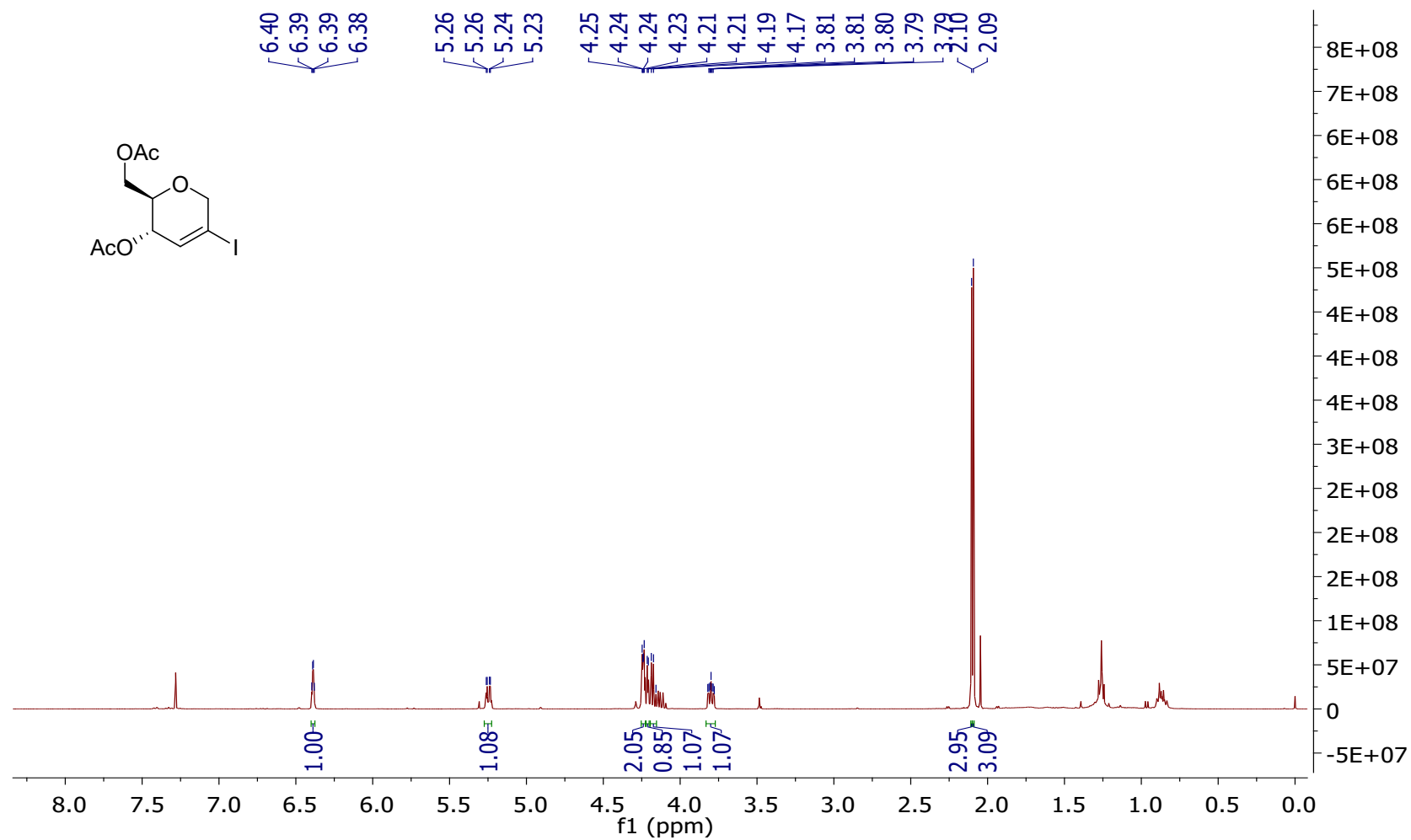
Compound 3a (50 mg, 0.072 mmol) was dissolved in MeOH (10 mL). To this solution 10 % Pd/C (60 mg) was added. The reaction was stirred under hydrogen atmosphere for 3 h. The reaction mixture was filtered through a Celite bed using methanol as eluent. Filtrate was concentrated under reduced pressure and dried under

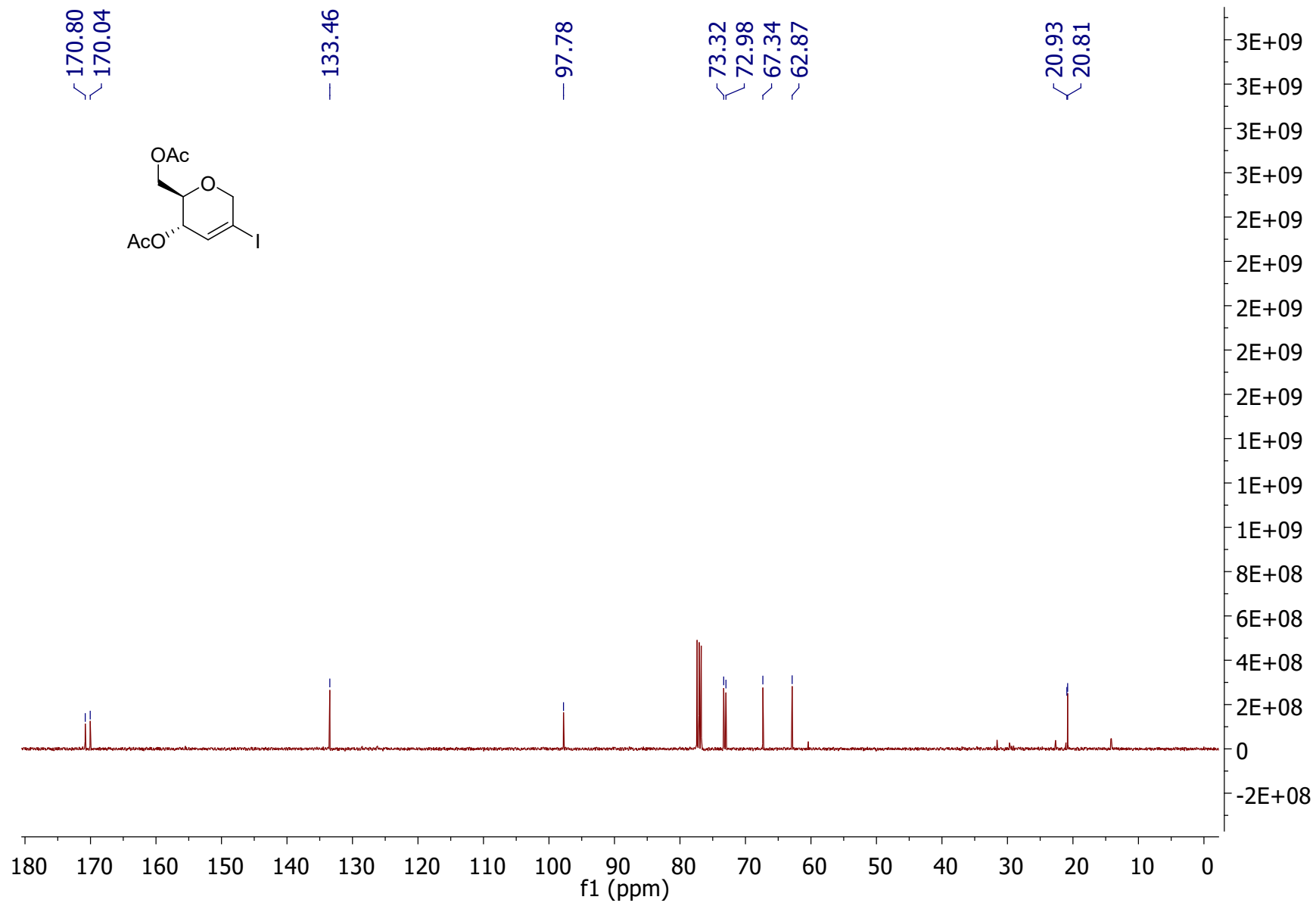
high vacuum to obtain **9** (27.5 mg, 90%) as white foam.

¹H NMR (400 MHz, MeOD) δ 5.06 – 4.95 (m, 2H), 4.47 (s, 1H), 4.11 (ddd, *J* = 12.1, 7.9, 3.4 Hz, 1H), 3.98 (tdd, *J* = 11.0, 6.7, 4.1 Hz, 1H), 3.86 (ddd, *J* = 9.7, 8.2, 1.9 Hz, 1H), 3.73 – 3.65 (m, 1H), 3.61 – 3.53 (m, 2H), 3.52 – 3.48 (m, 2H), 3.42 – 3.35 (m, 1H), 3.23 – 3.19 (m, 1H), 1.95 (s, 3H), 1.95 (s, *J* = 1.4 Hz, 3H), 1.93 (s, 3H), 1.76 – 1.63 (m, 1H), 1.60 – 1.46 (m, 2H). **¹³C NMR (101 MHz, MeOD)** δ 171.2 (Cq), 170.6 (Cq), 170.4 (Cq), 76.6 (CH), 74.4 (CH), 73.1 (CH), 72.1 (CH), 71.8 (CH), 66.8 (CH₂), 66.6 (CH), 63.3 (CH₂), 62.6 (CH₂), 48.5 (CH), 39.0 (CH), 29.4 (CH₂), 19.5 (CH₃), 19.4 (CH₃), 19.3 (CH₃).

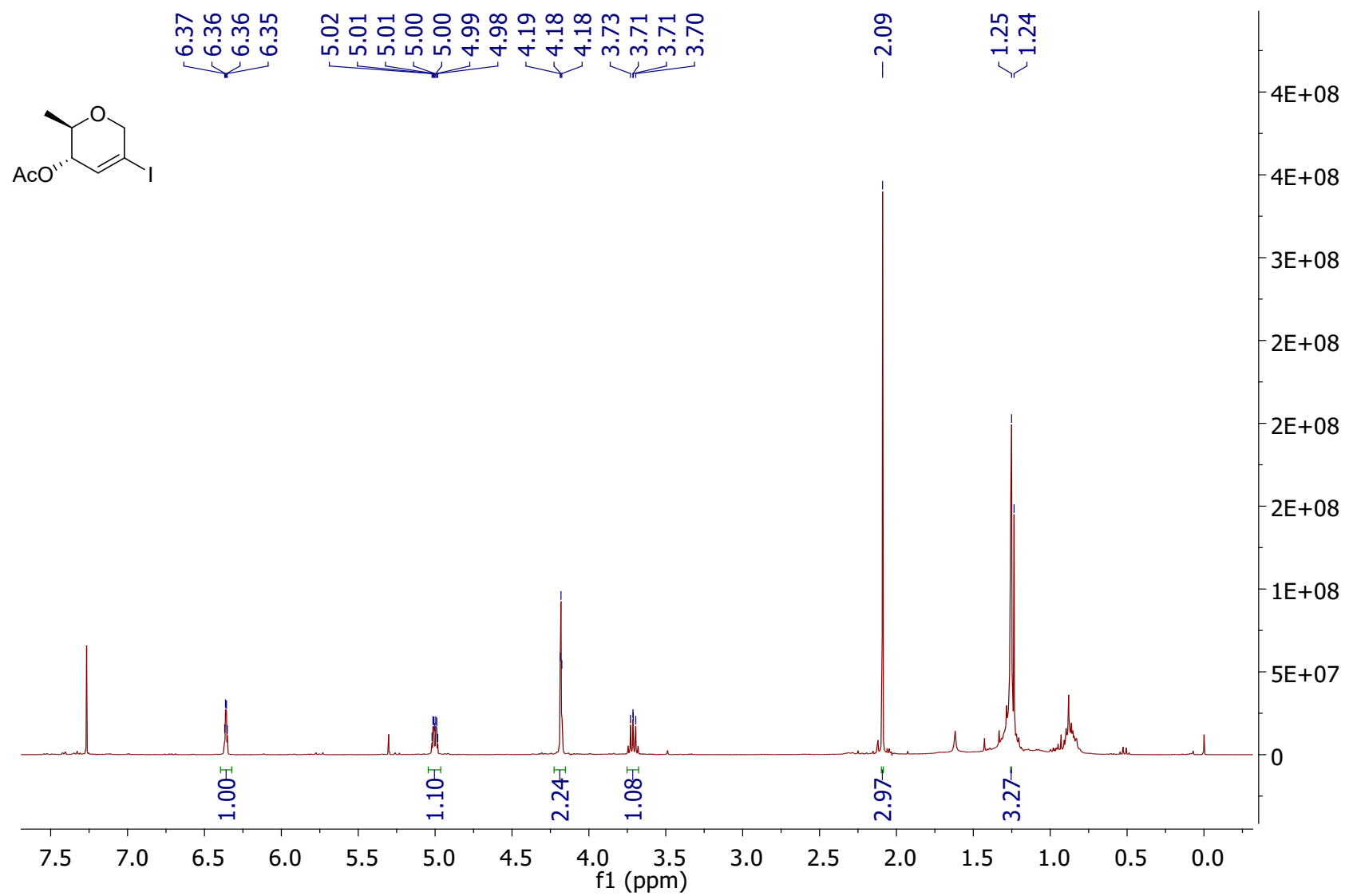
4. NMR Spectra:

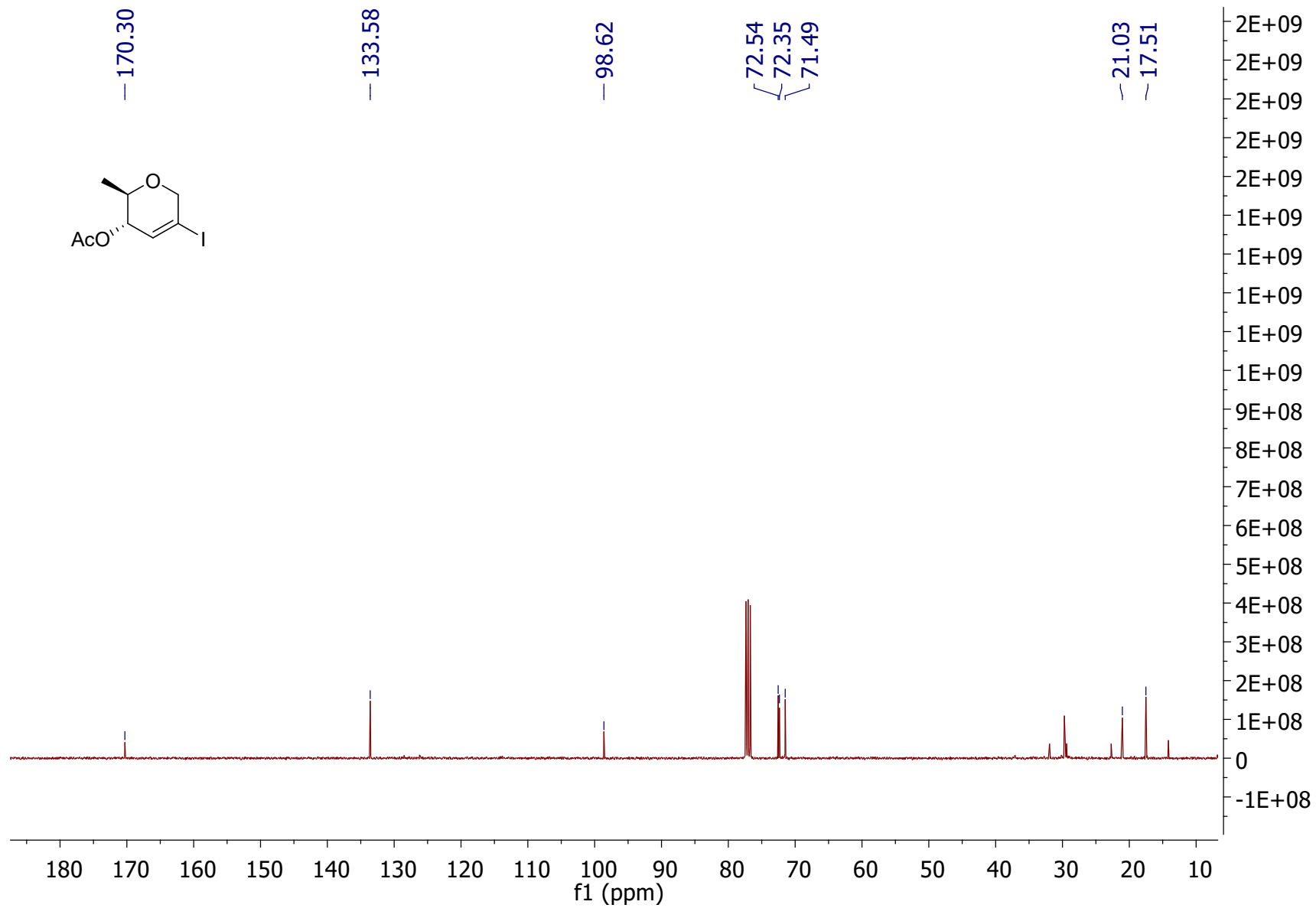
^1H NMR (400 MHz, CDCl_3) and ^{13}C $\{^1\text{H}\}$ NMR (101 MHz, CDCl_3) of compound 5a



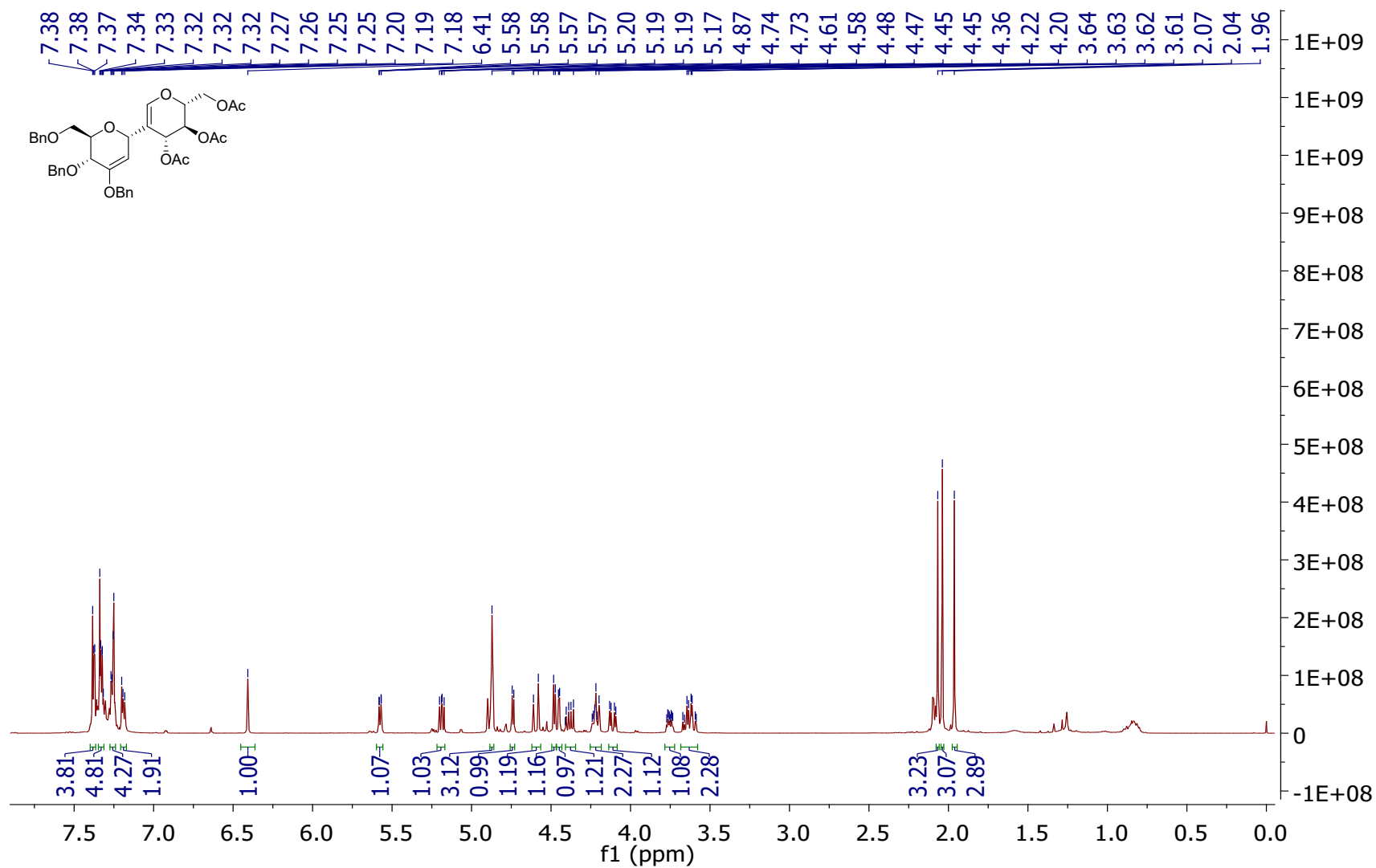


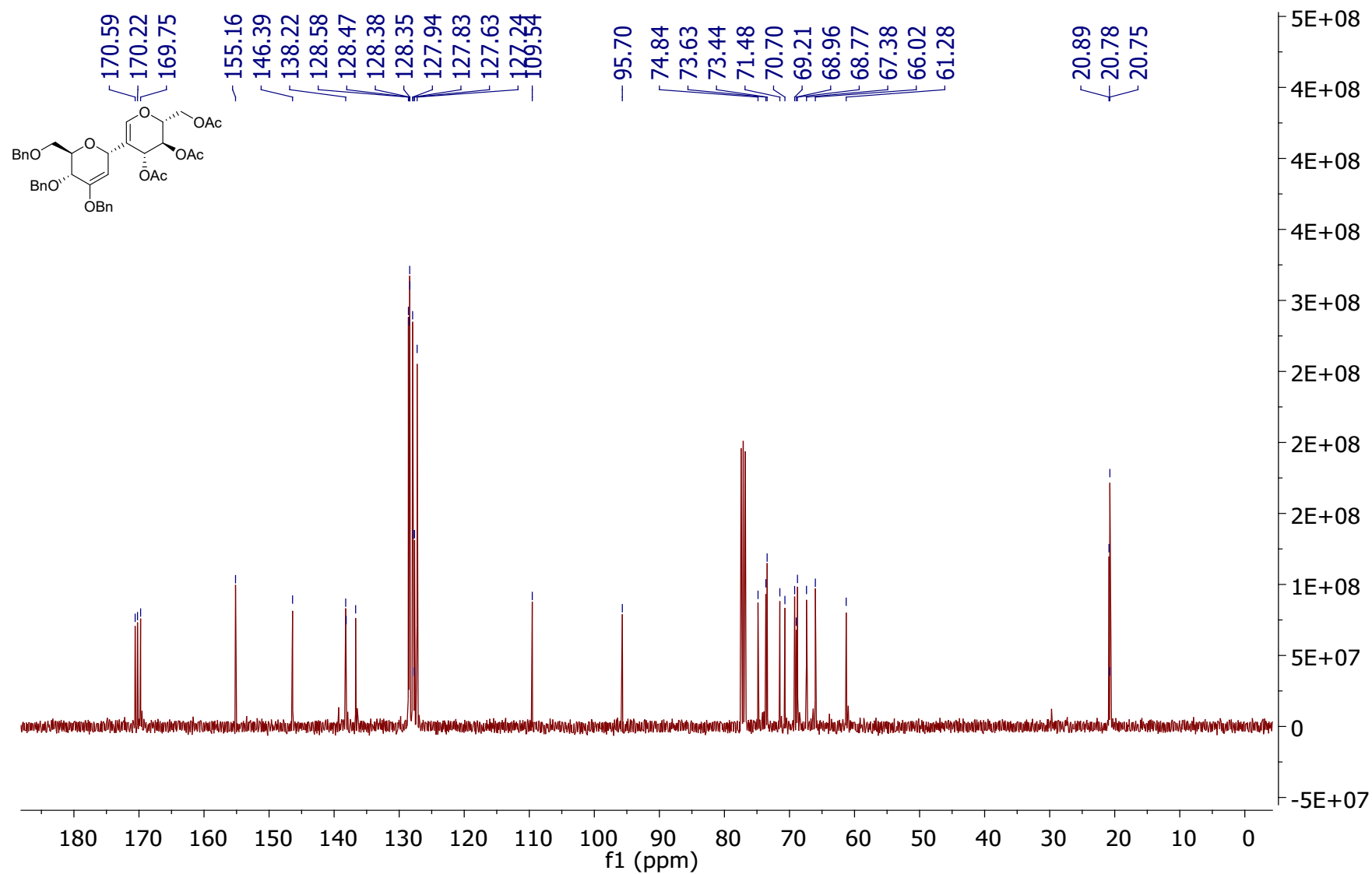
^1H NMR (400 MHz, CDCl_3) and ^{13}C $\{^1\text{H}\}$ NMR (101 MHz, CDCl_3) of compound 5b



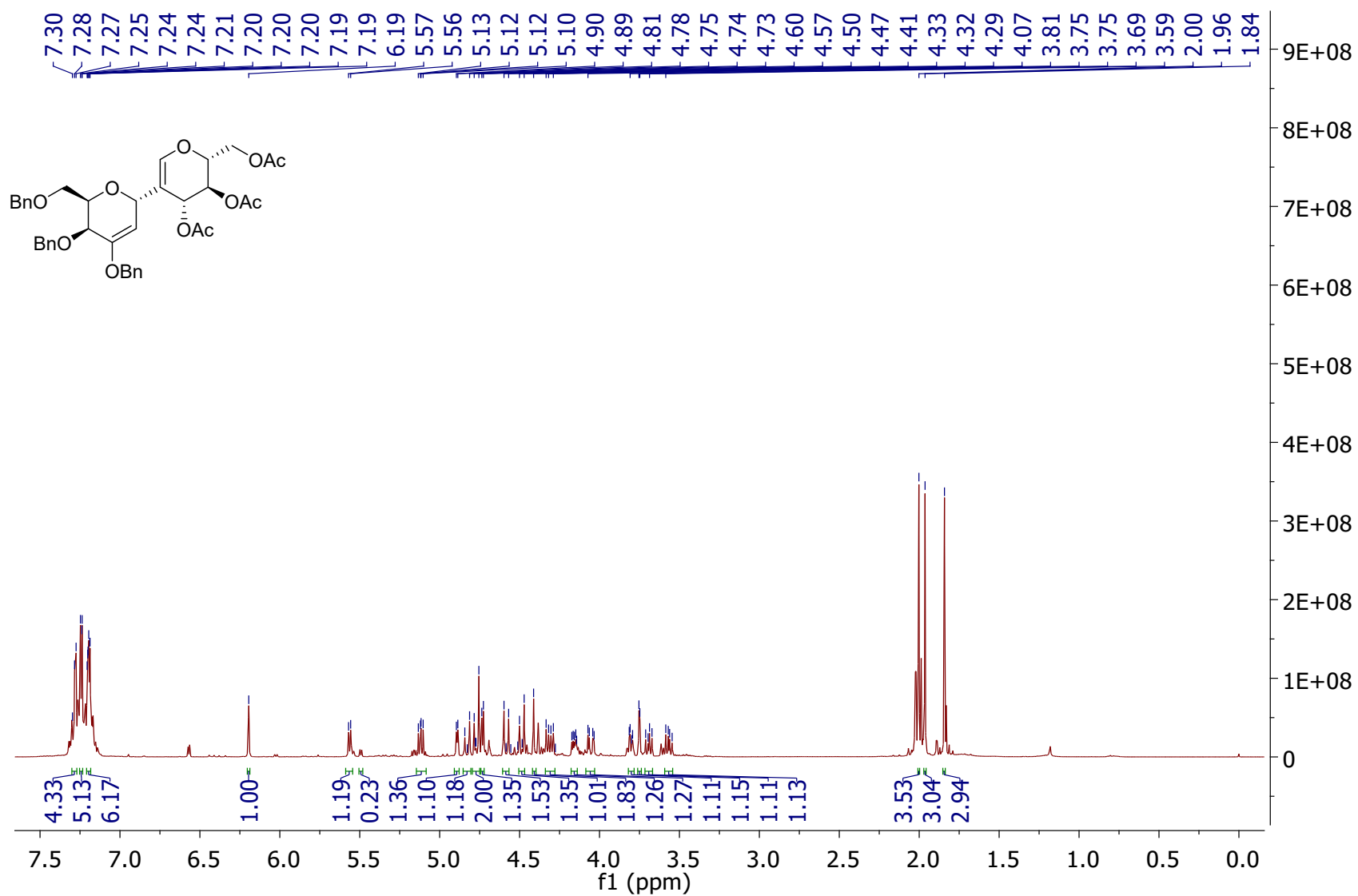


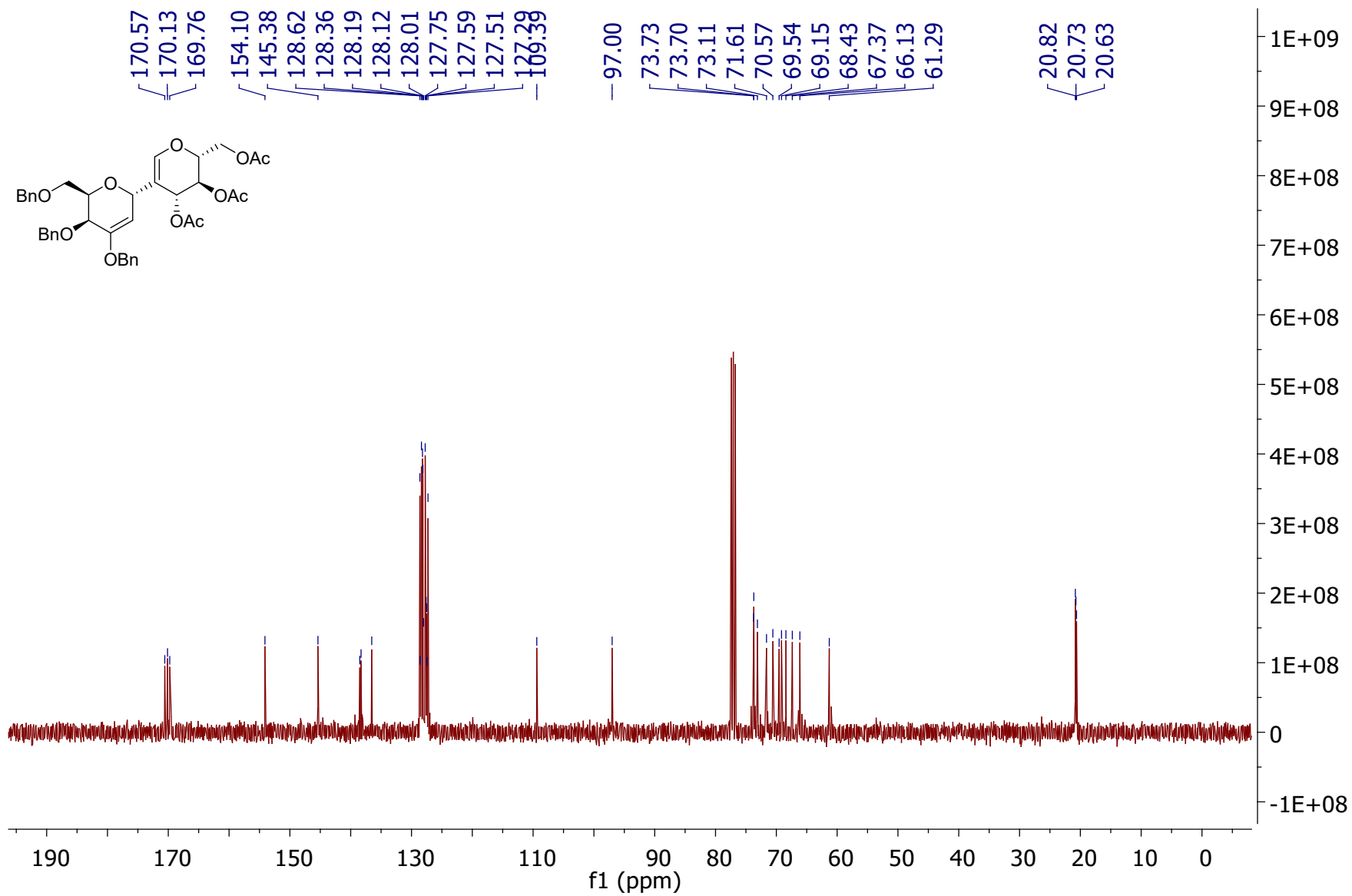
^1H NMR (400 MHz, CDCl_3) and ^{13}C $\{^1\text{H}\}$ NMR (101 MHz, CDCl_3) of compound 3a:



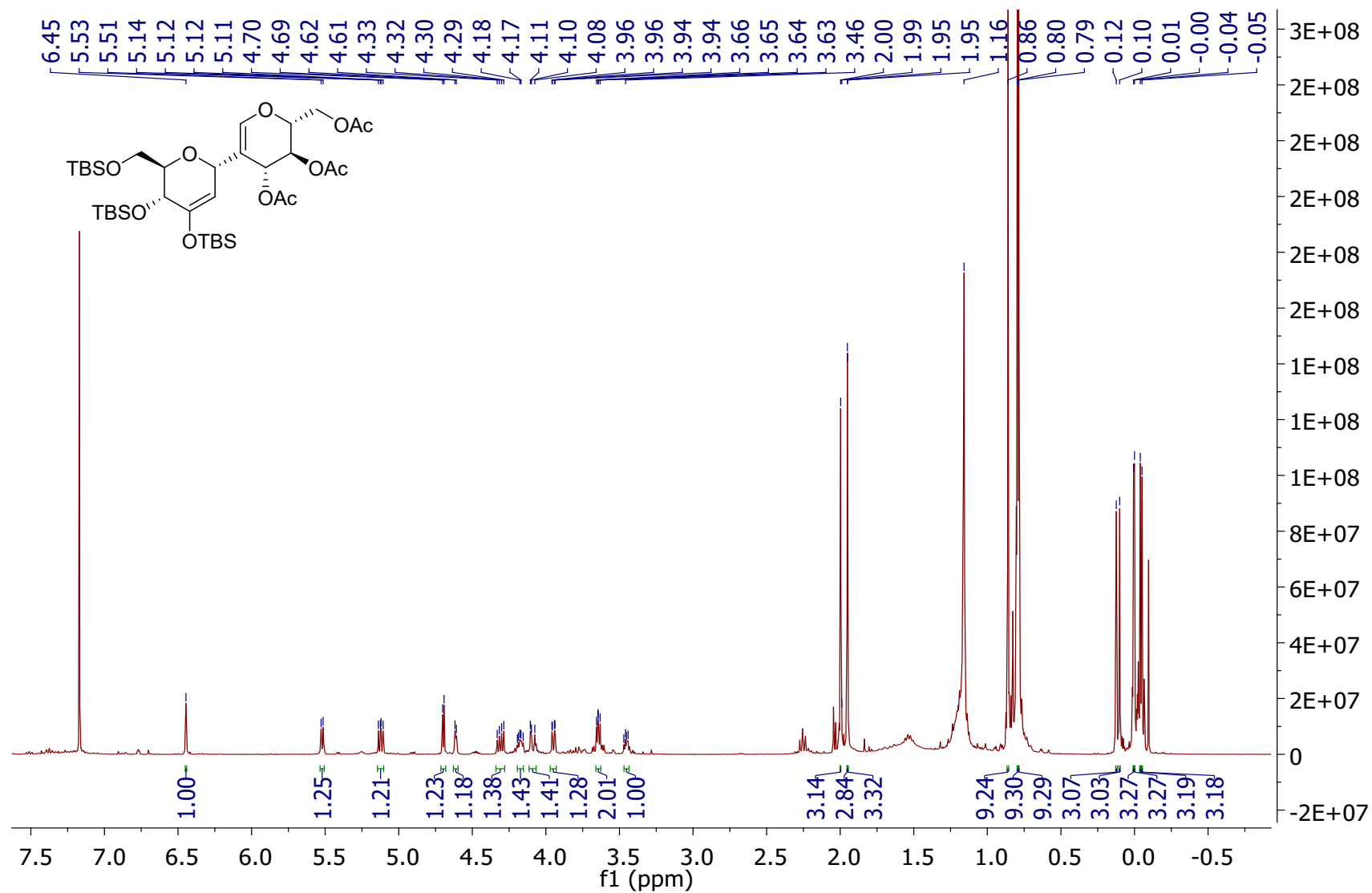


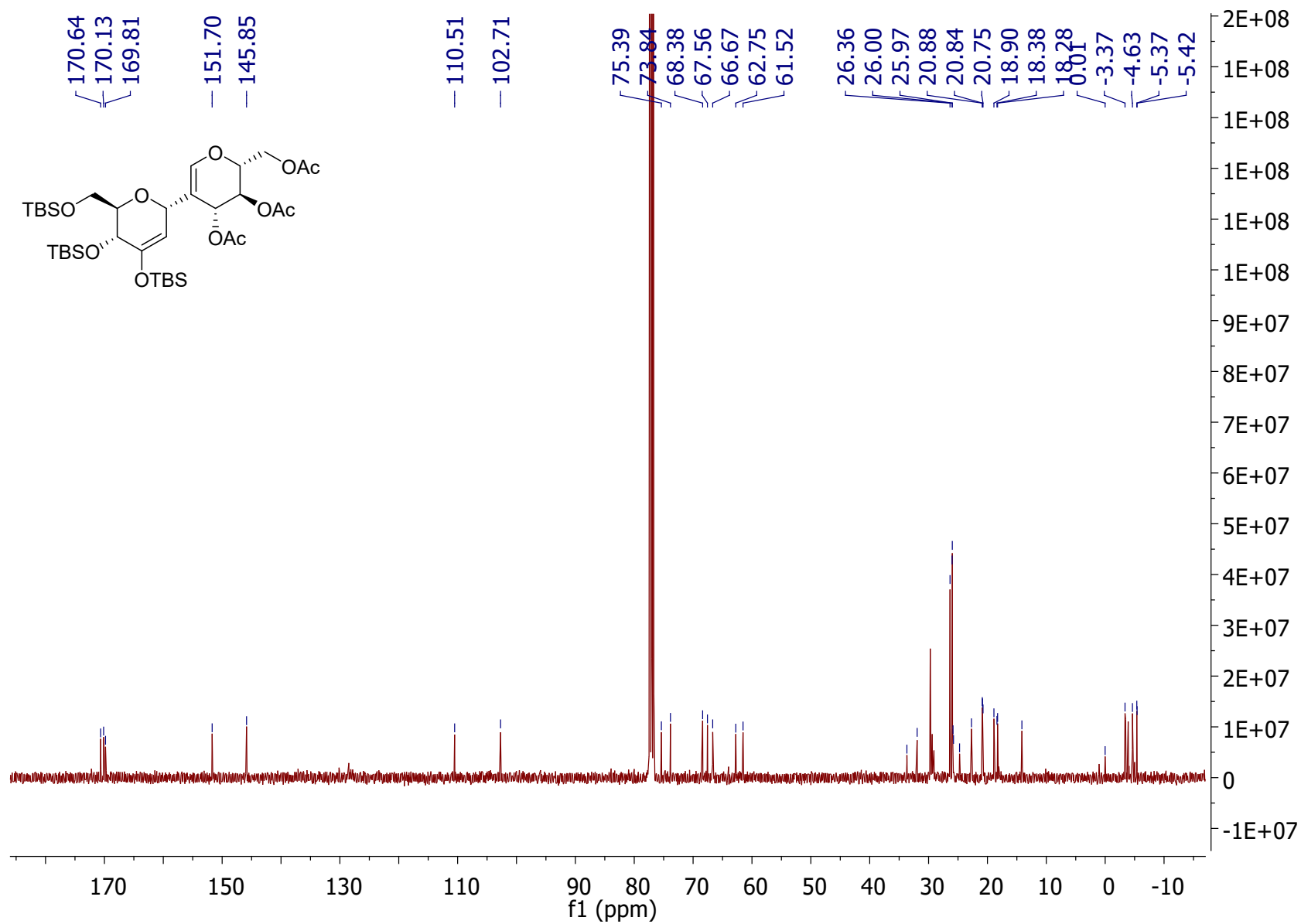
^1H NMR (400 MHz, CDCl_3) and ^{13}C $\{^1\text{H}\}$ NMR (101 MHz, CDCl_3) of compound 3b (α/β ; 4:1):



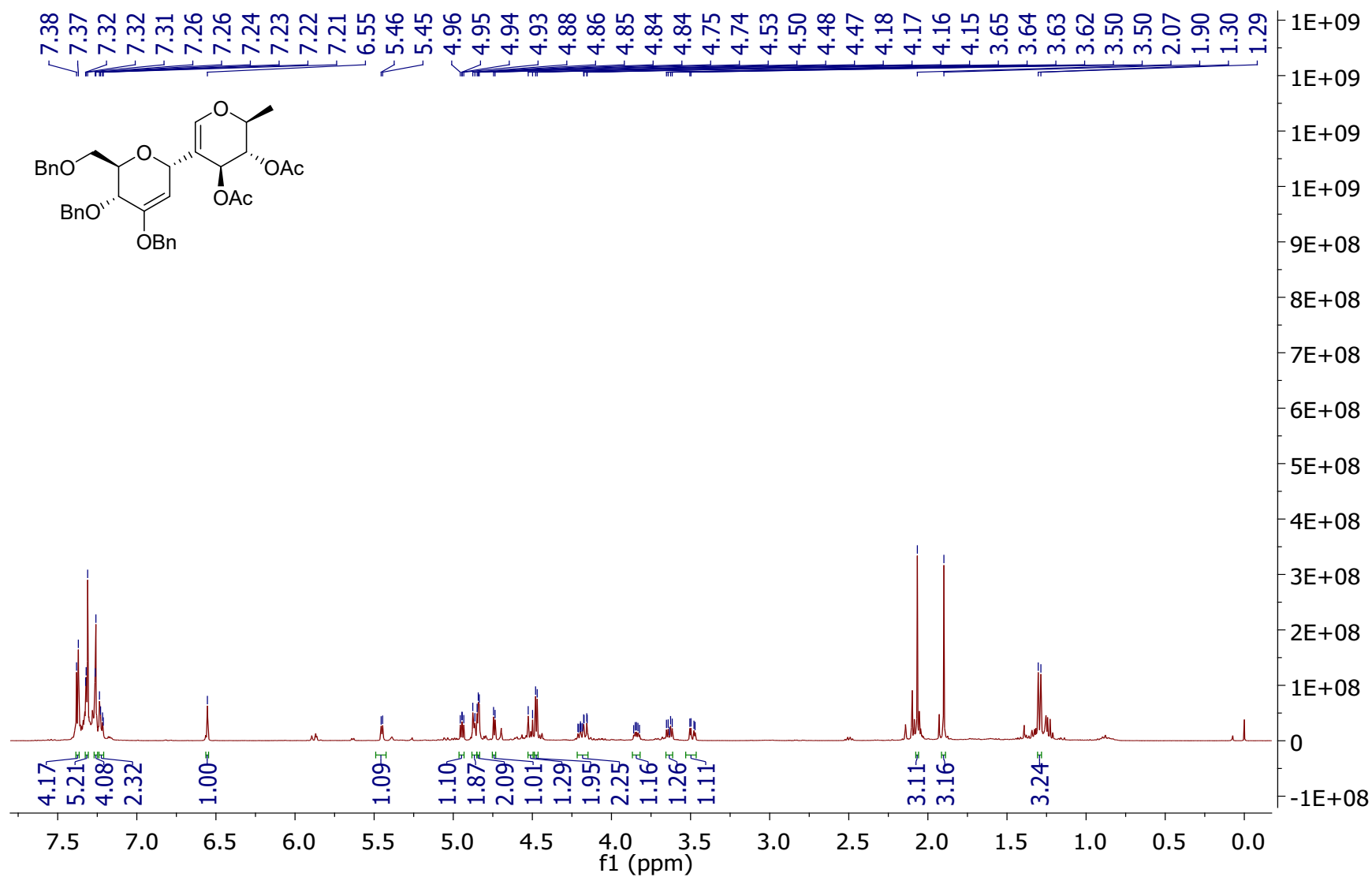


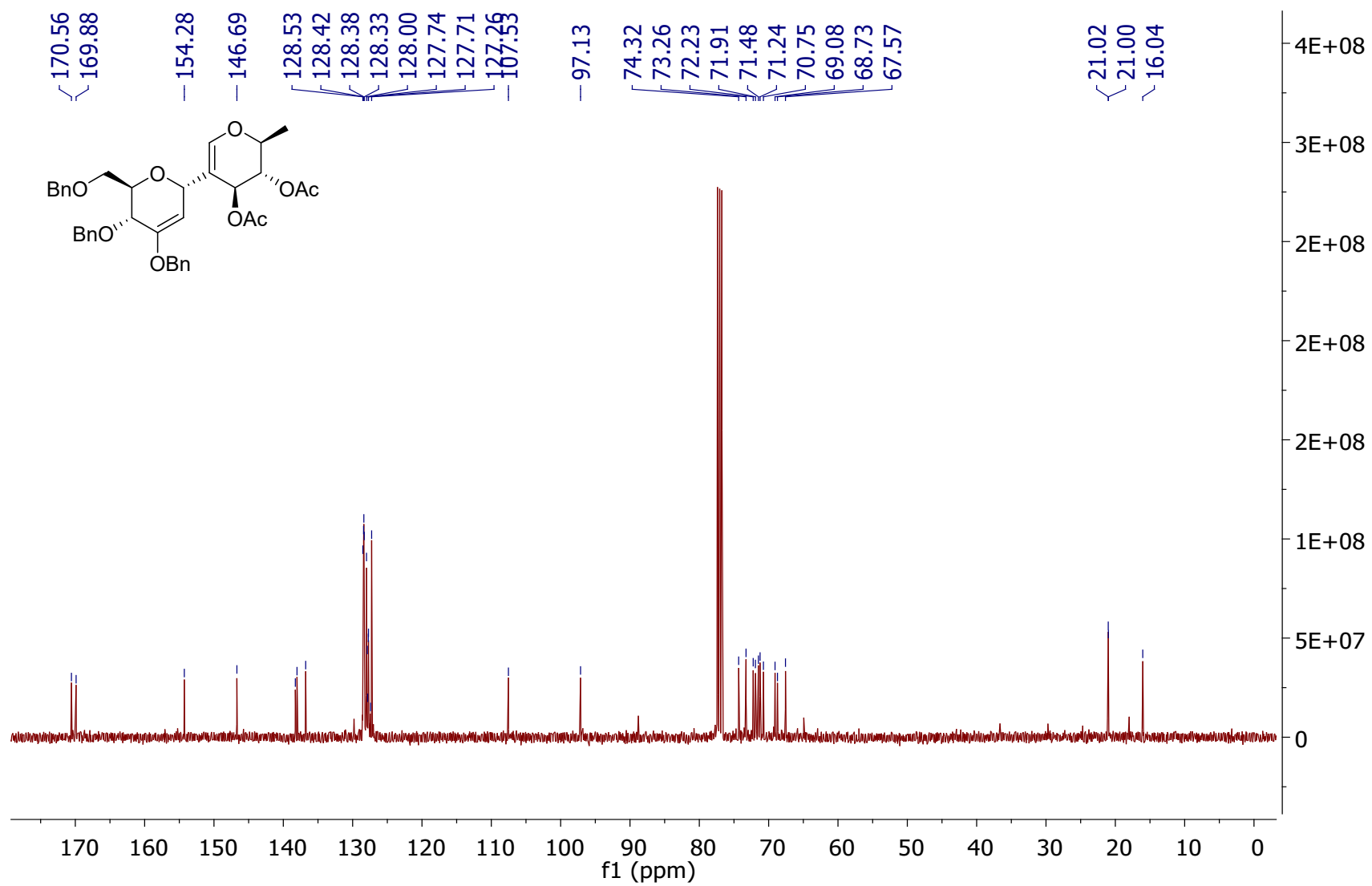
^1H NMR (400 MHz, CDCl_3) and ^{13}C $\{^1\text{H}\}$ NMR (101 MHz, CDCl_3) of compound 3c:



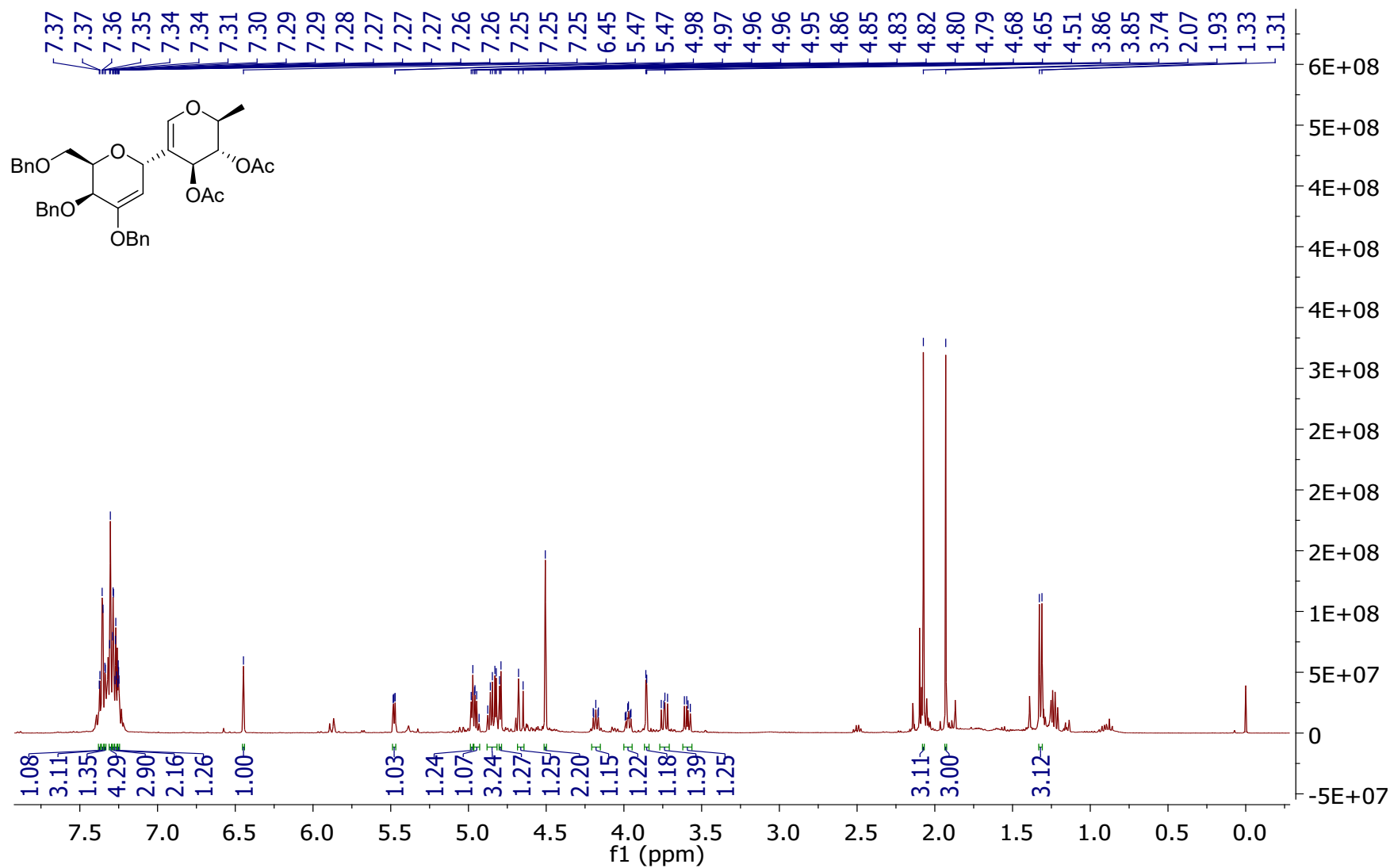


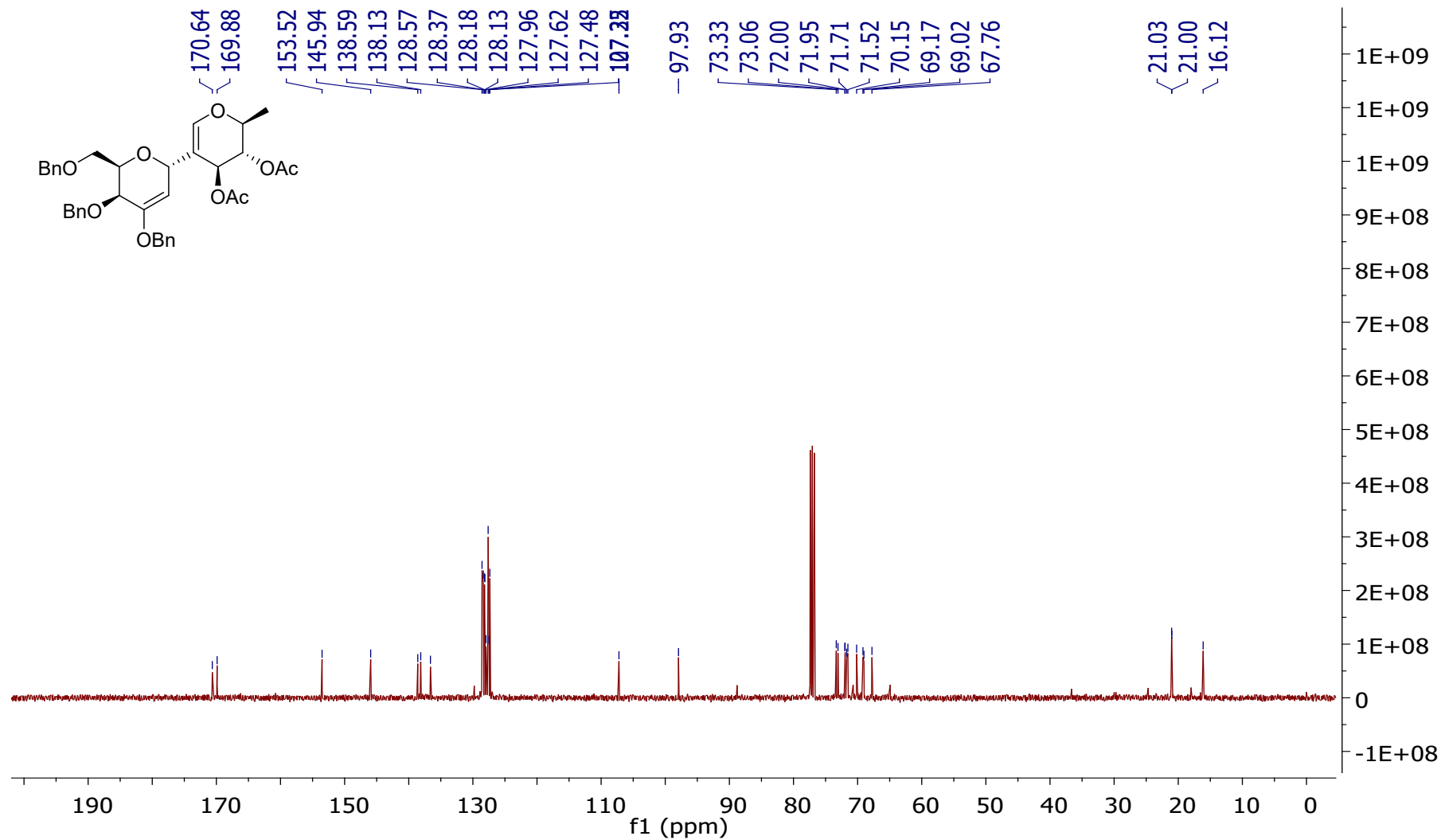
^1H NMR (400 MHz, CDCl_3) and ^{13}C { ^1H } NMR (101 MHz, CDCl_3) of compound 3d:



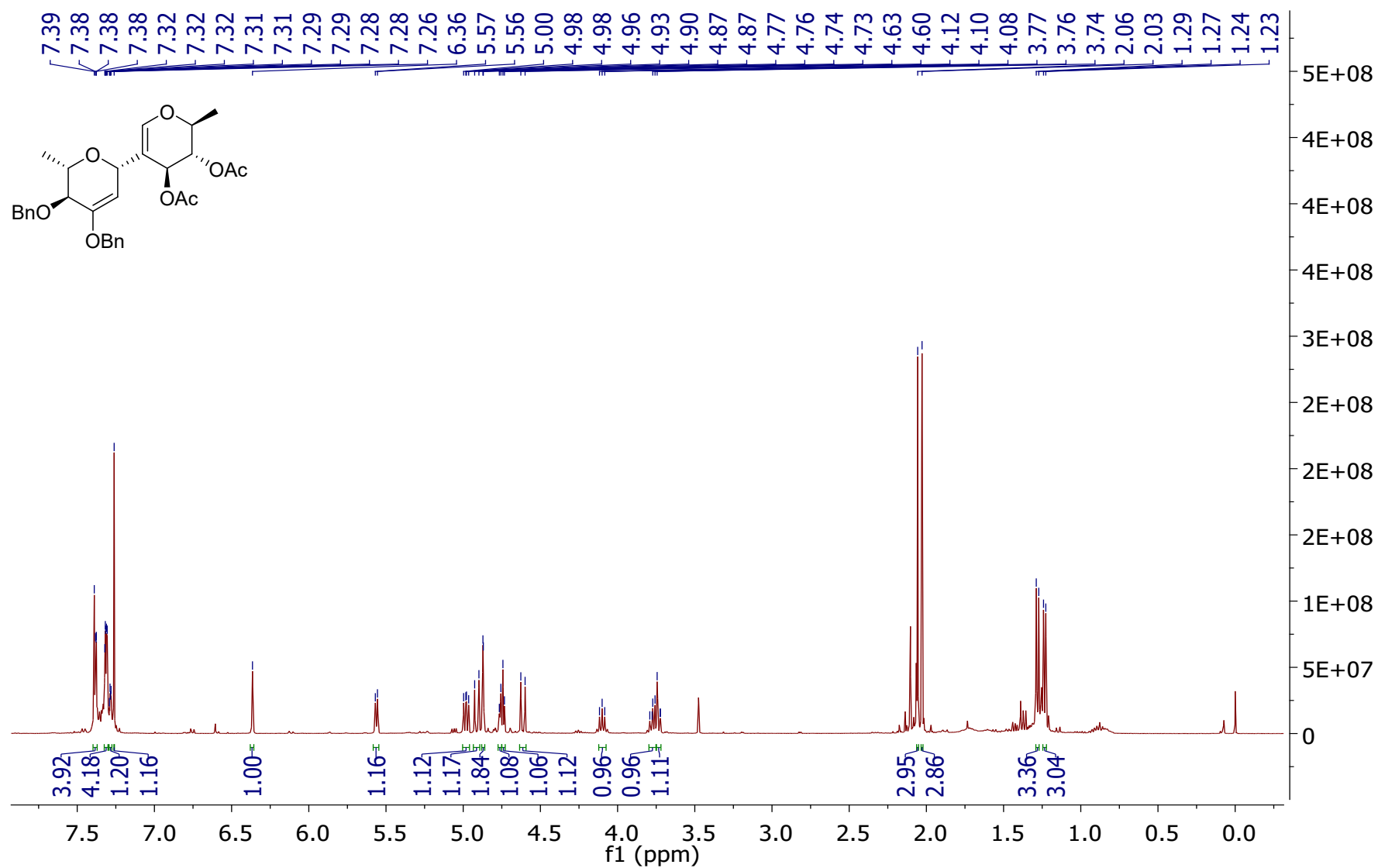


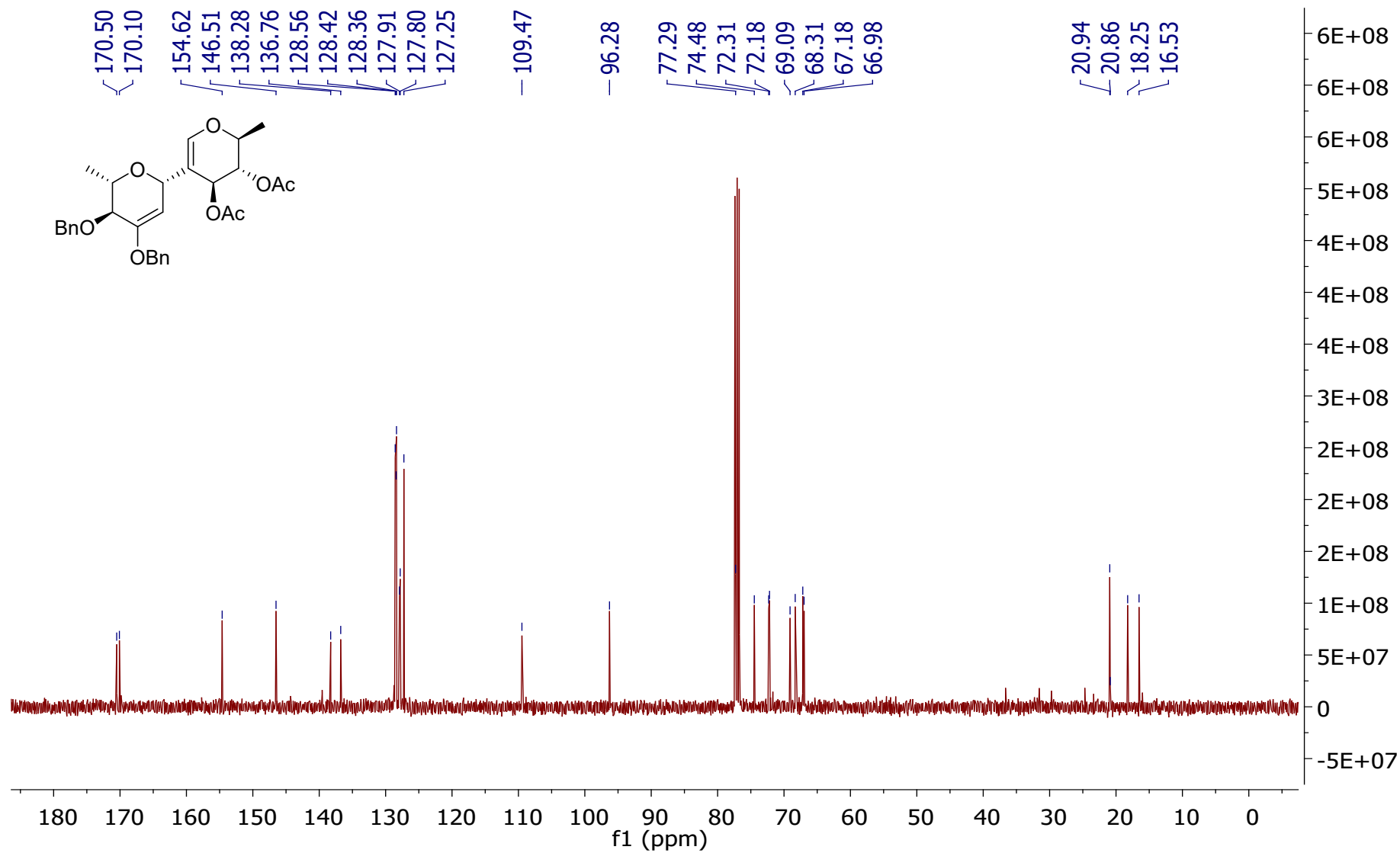
^1H NMR (400 MHz, CDCl_3) and ^{13}C { ^1H } NMR (101 MHz, CDCl_3) of compound 3e:



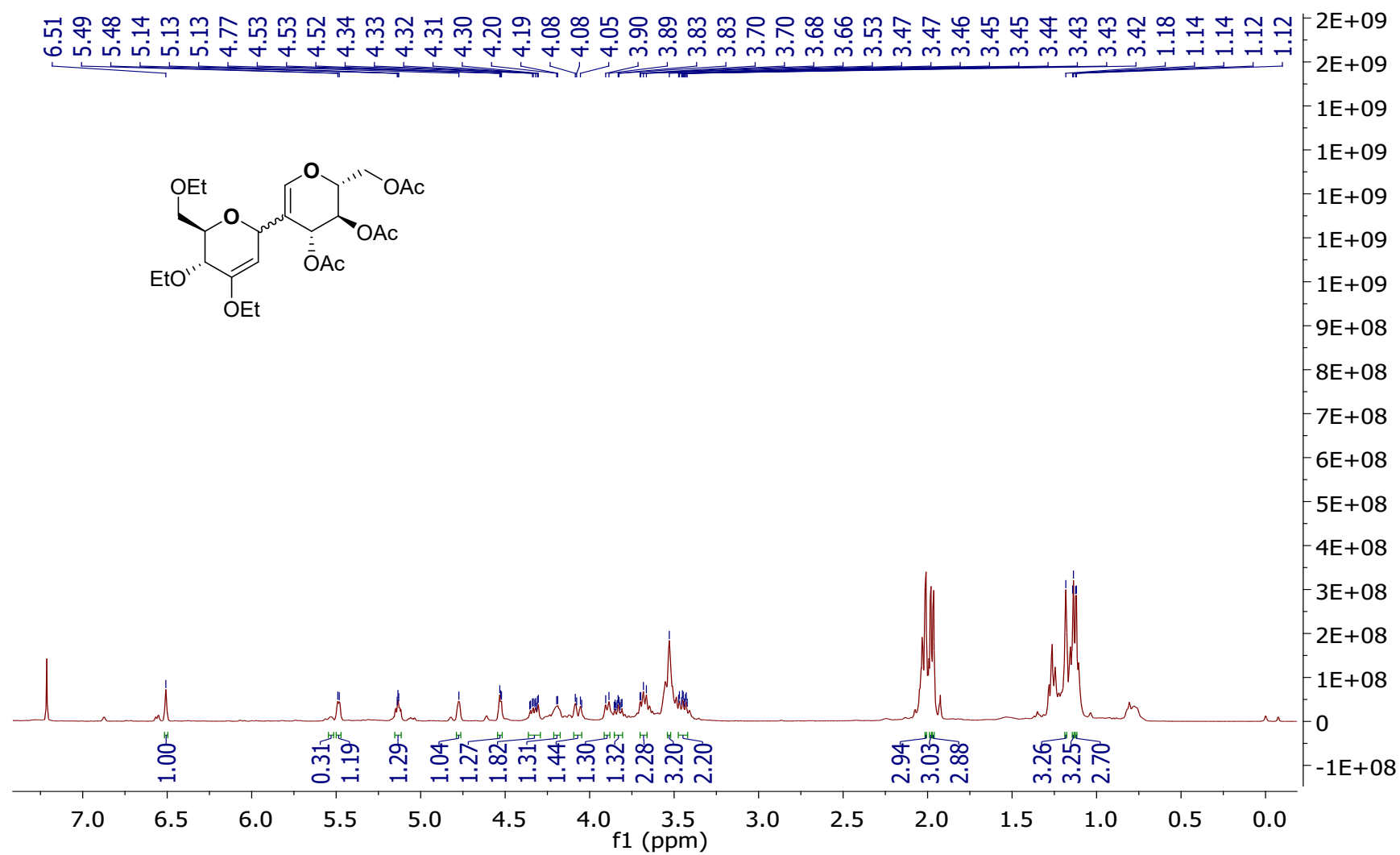


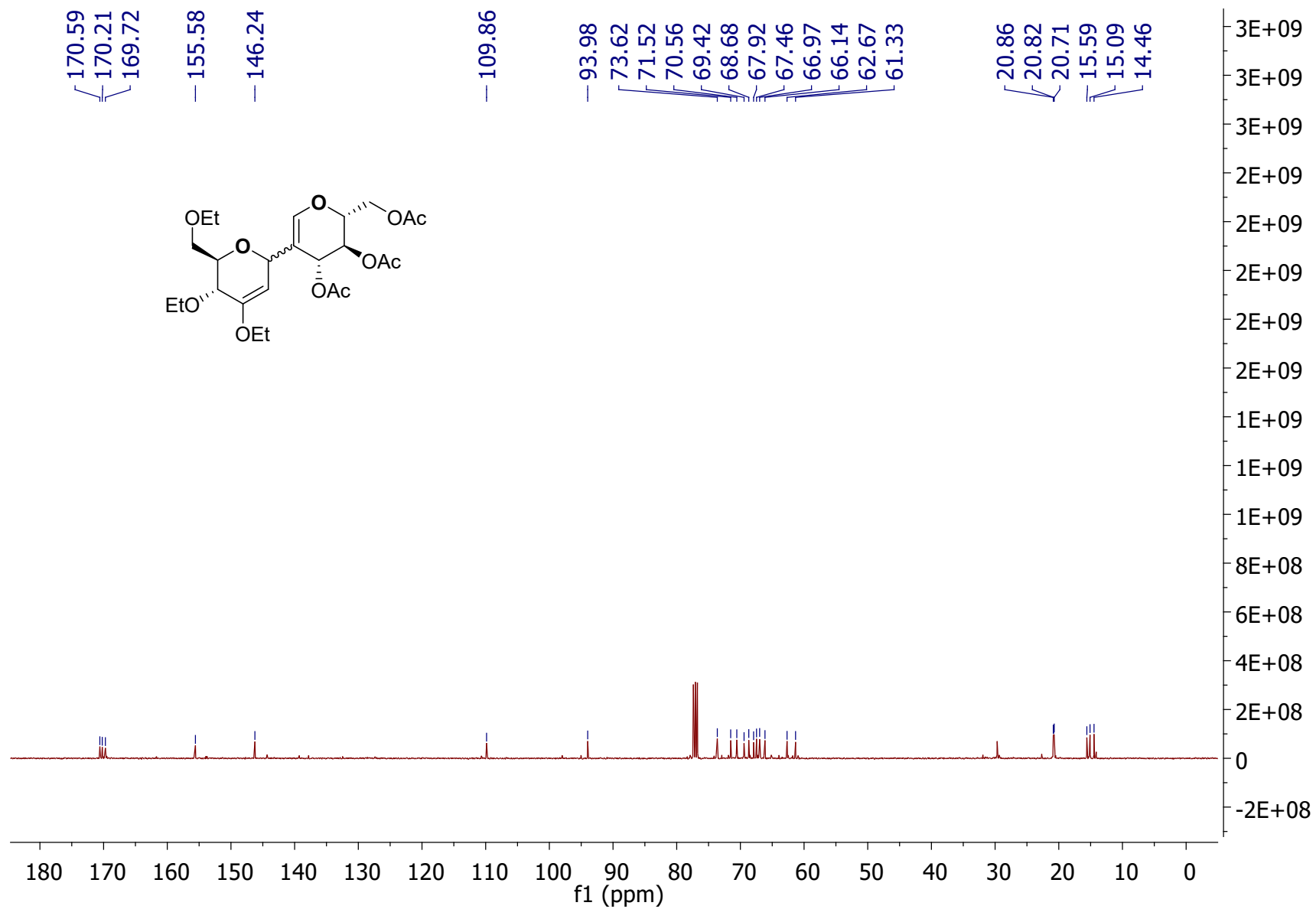
^1H NMR (400 MHz, CDCl_3) and ^{13}C NMR $\{^1\text{H}\}$ (101 MHz, CDCl_3) of compound 3f:



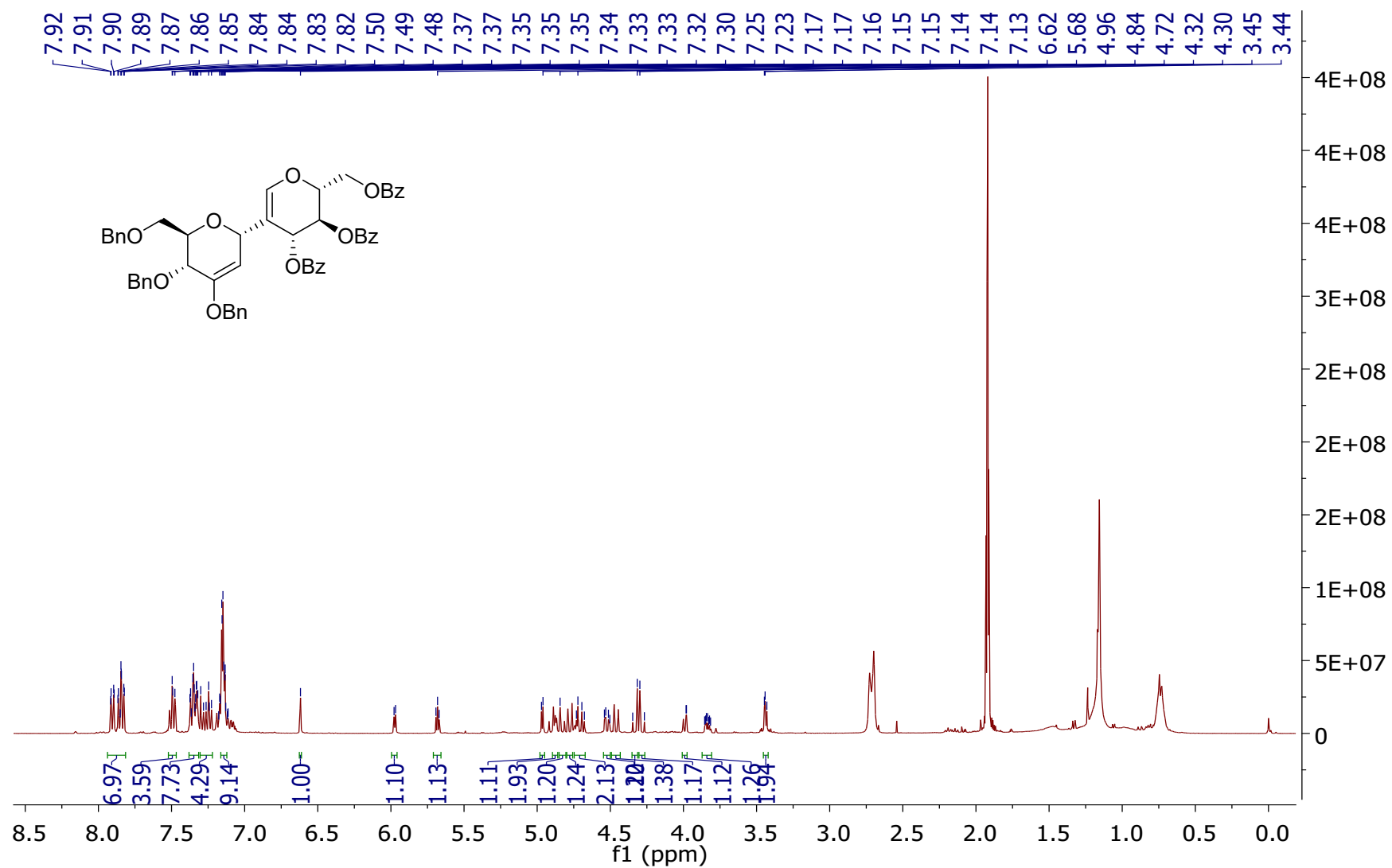


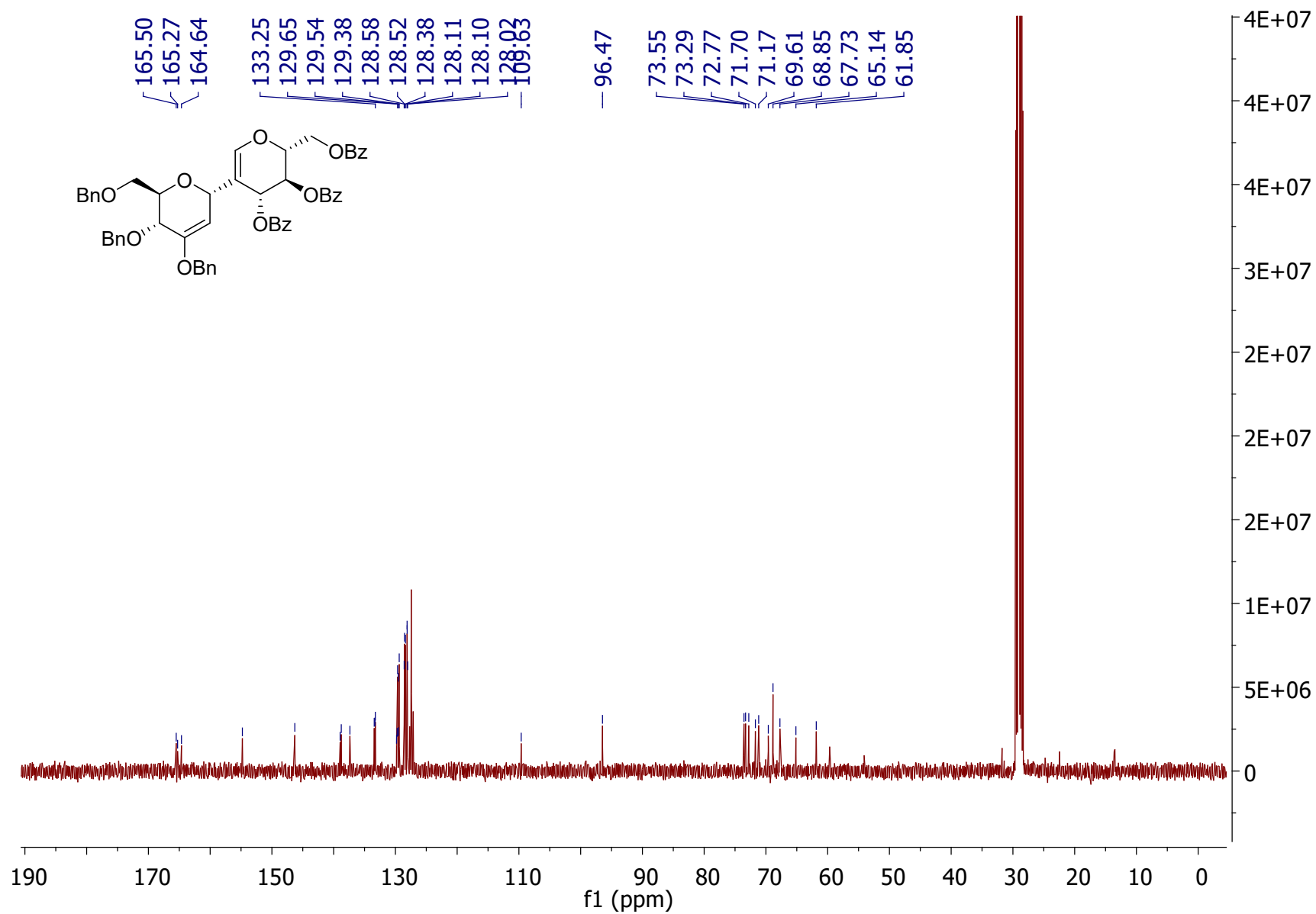
^1H NMR (400 MHz, CDCl_3) and ^{13}C $\{^1\text{H}\}$ NMR (101 MHz, CDCl_3) of compound 3g (α/β ; 4:1):



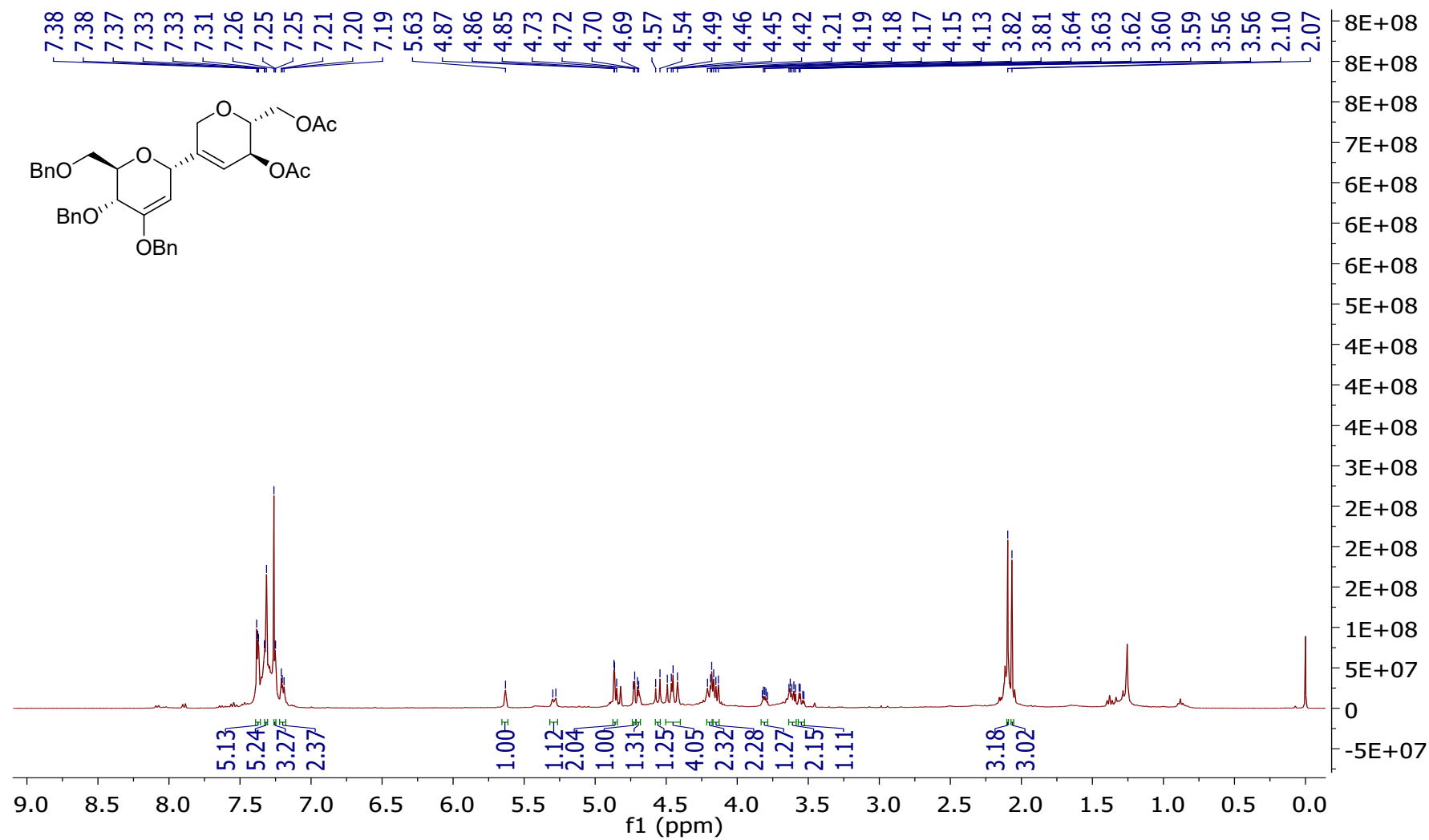


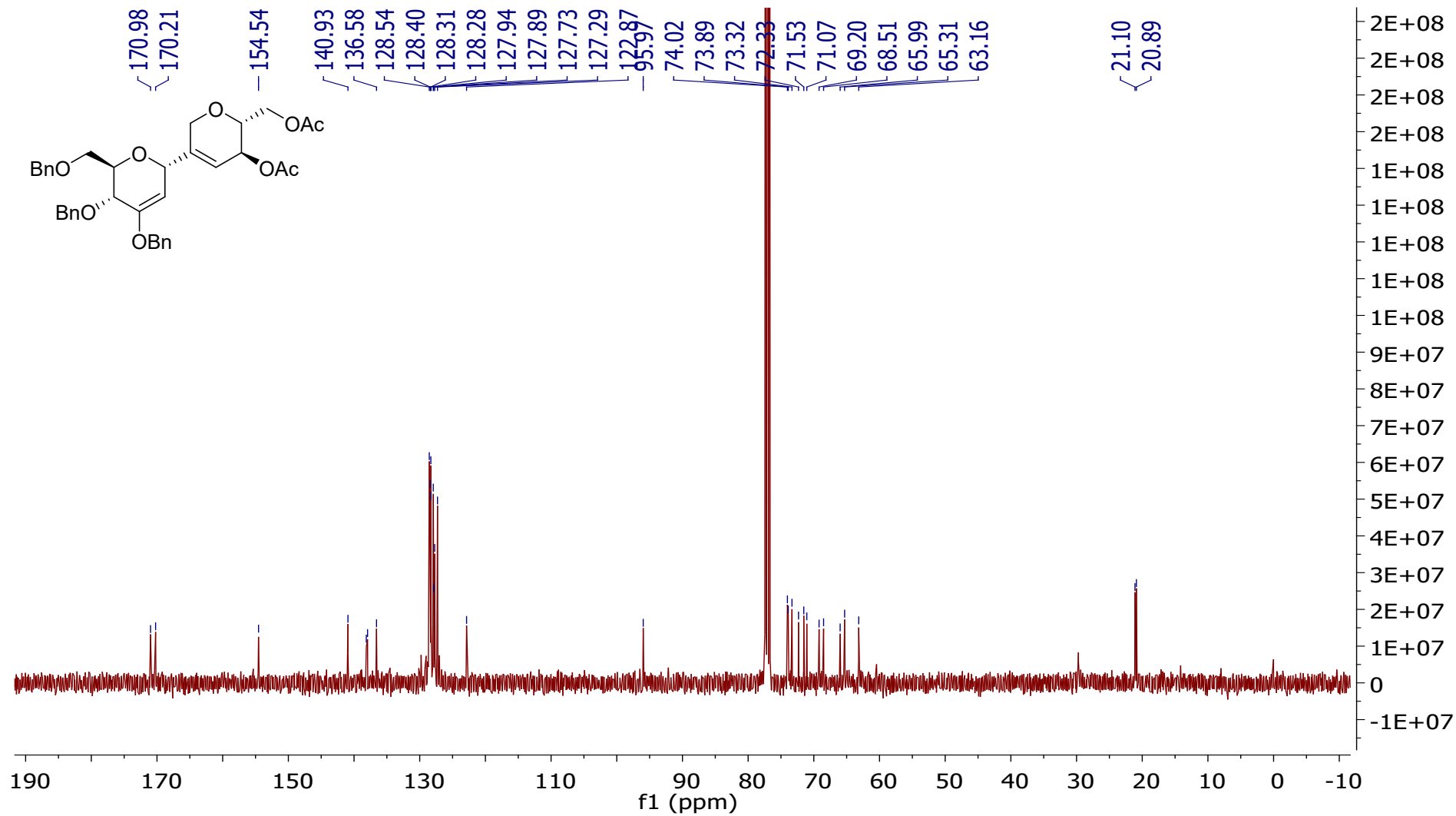
^1H NMR (400 MHz, acetone- d_6) and ^{13}C $\{^1\text{H}\}$ NMR (101 MHz, acetone- d_6) of compound 3h



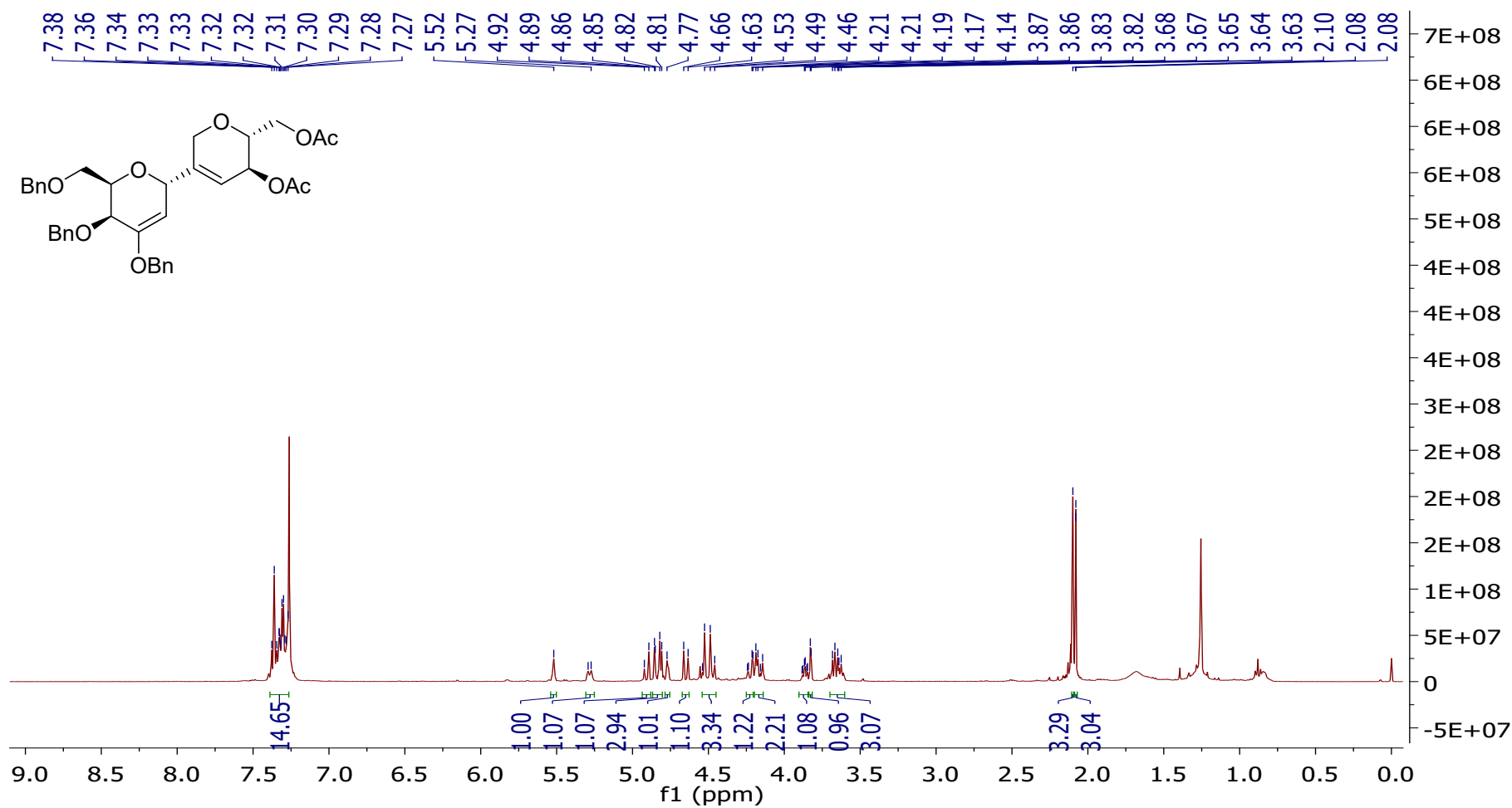


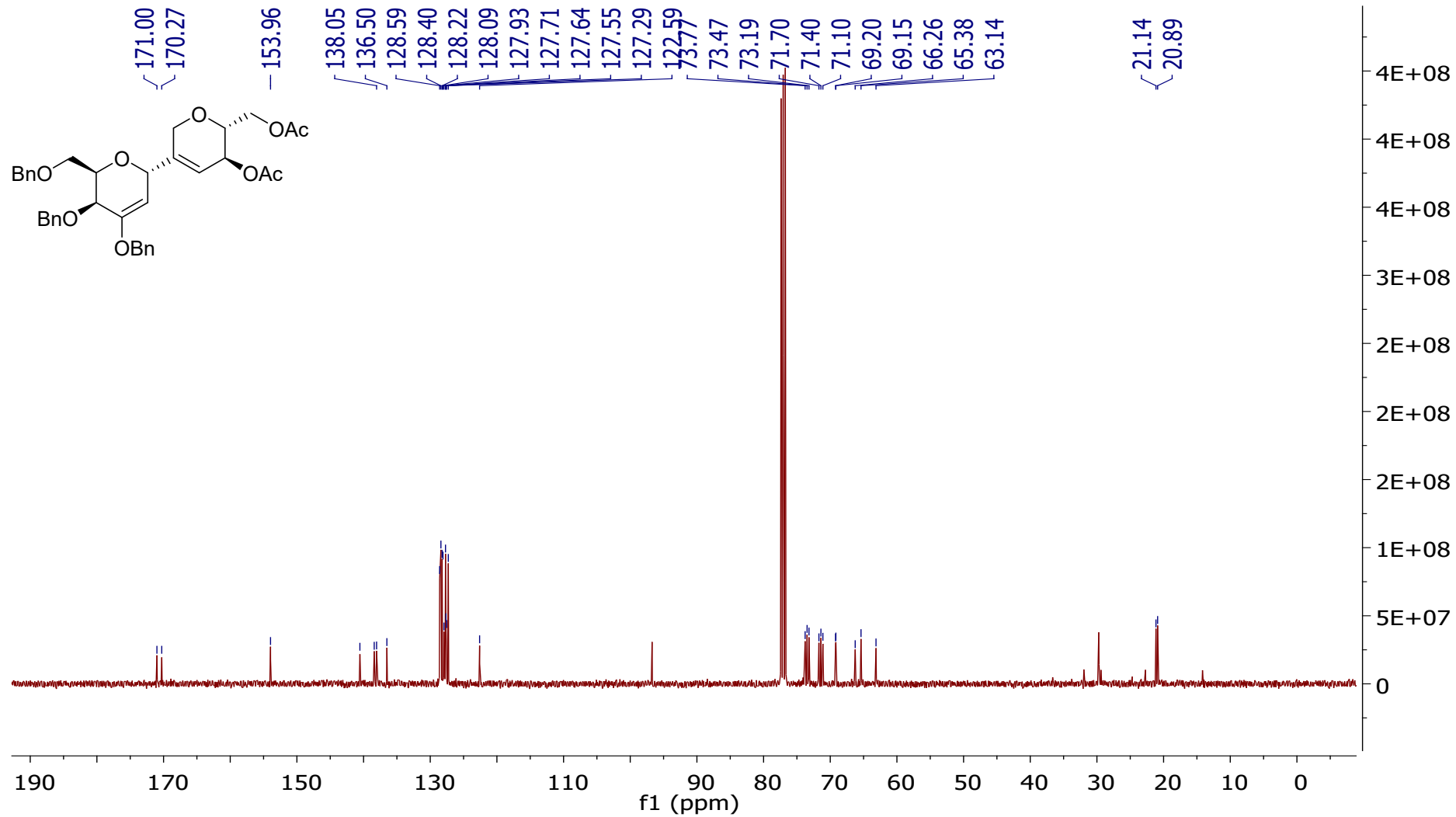
^1H NMR (400 MHz, CDCl_3) and ^{13}C $\{^1\text{H}\}$ NMR (101 MHz, CDCl_3) of compound 6a



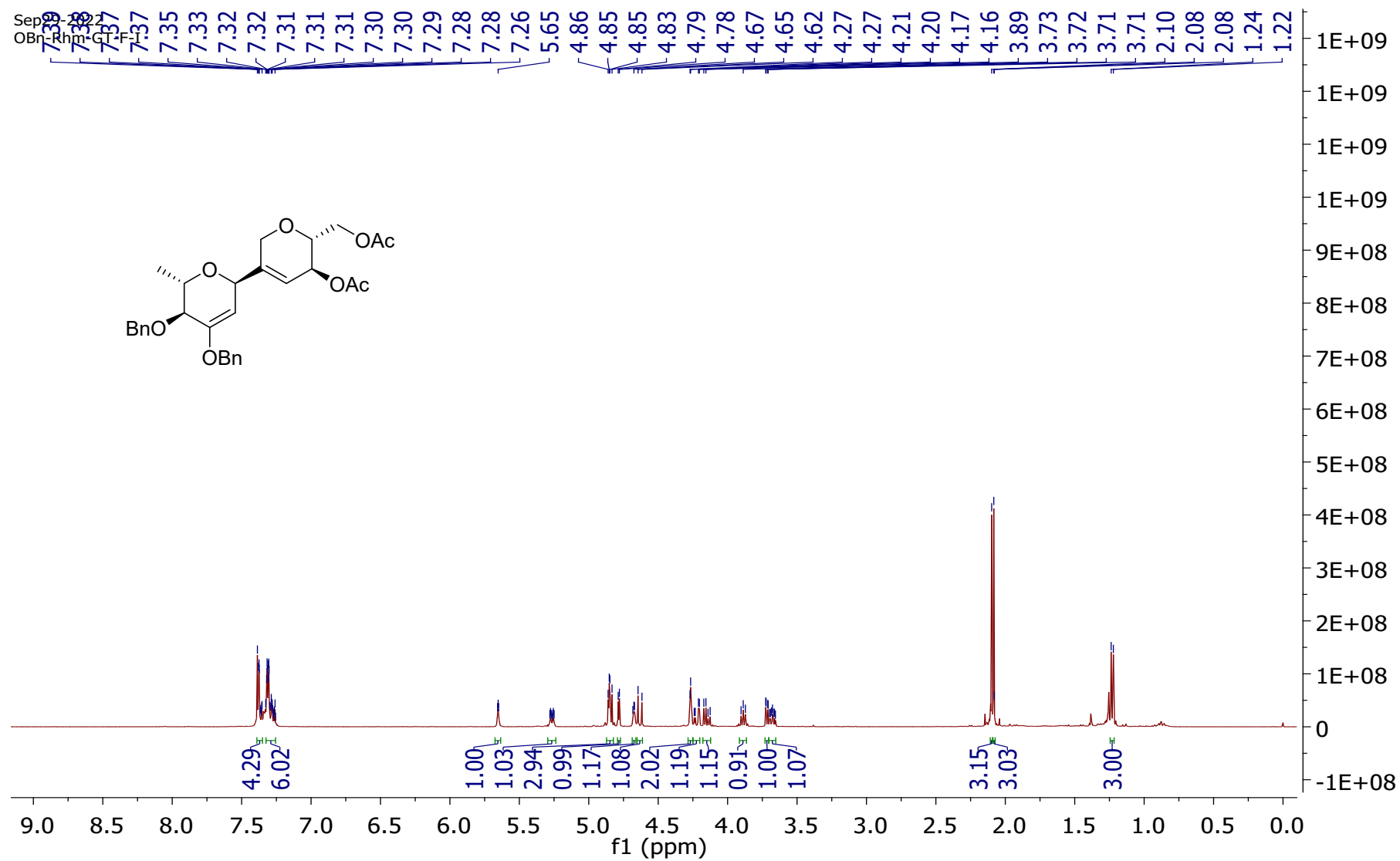


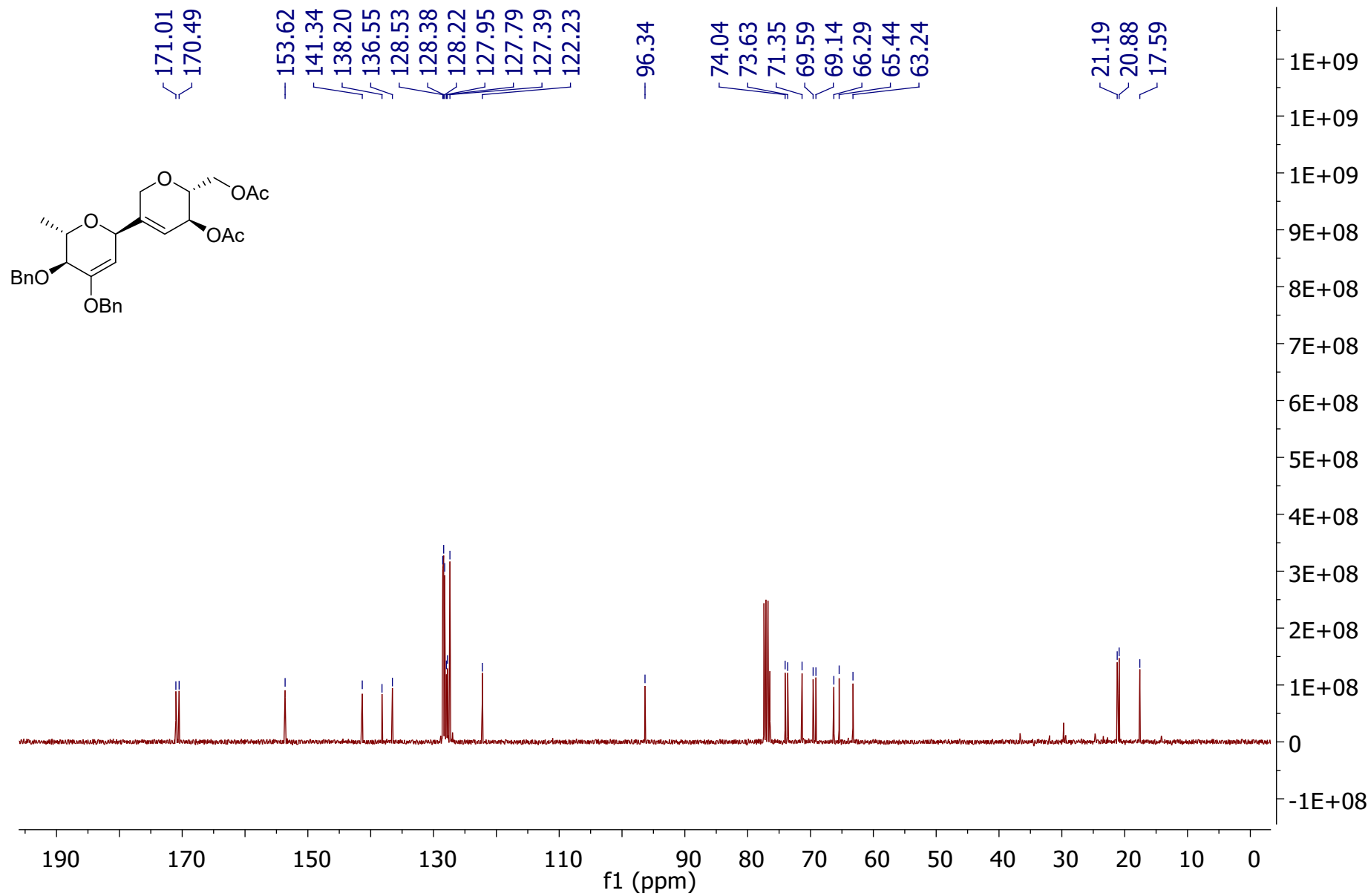
^1H NMR (400 MHz, CDCl_3) and ^{13}C $\{^1\text{H}\}$ NMR (101 MHz, CDCl_3) of compound 6b



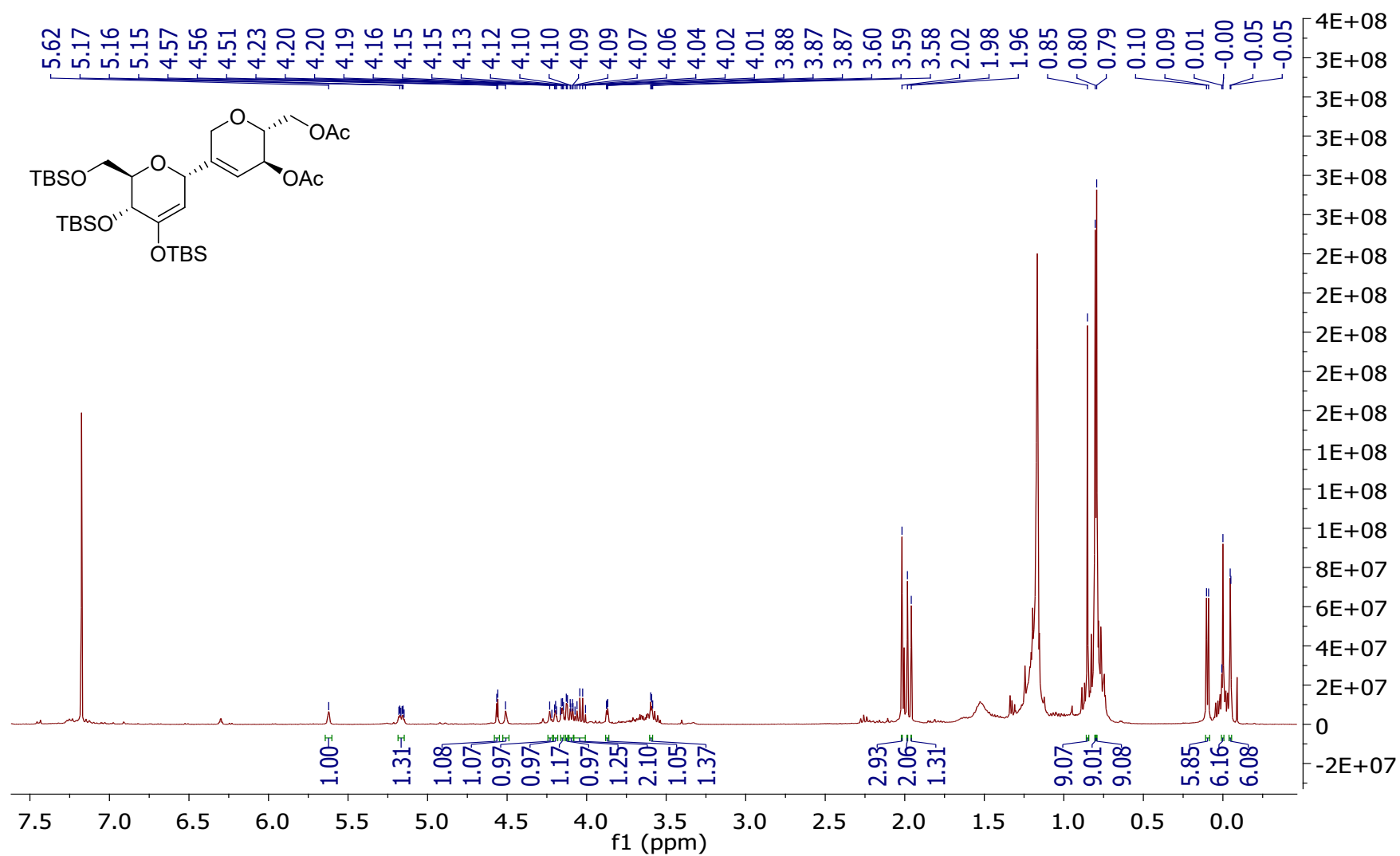


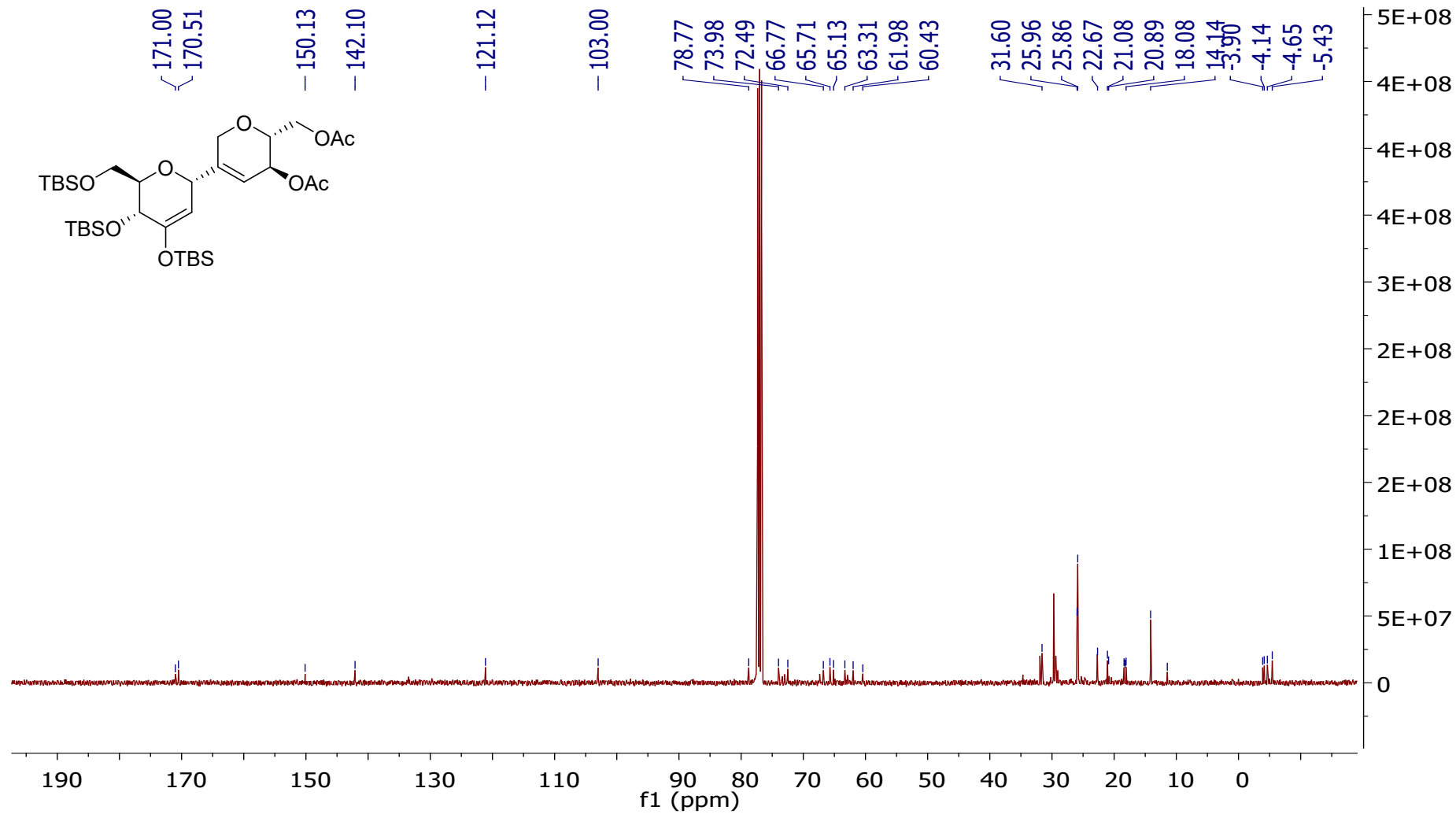
^1H NMR (400 MHz, CDCl_3) and ^{13}C $\{^1\text{H}\}$ NMR (101 MHz, CDCl_3) of compound 6c



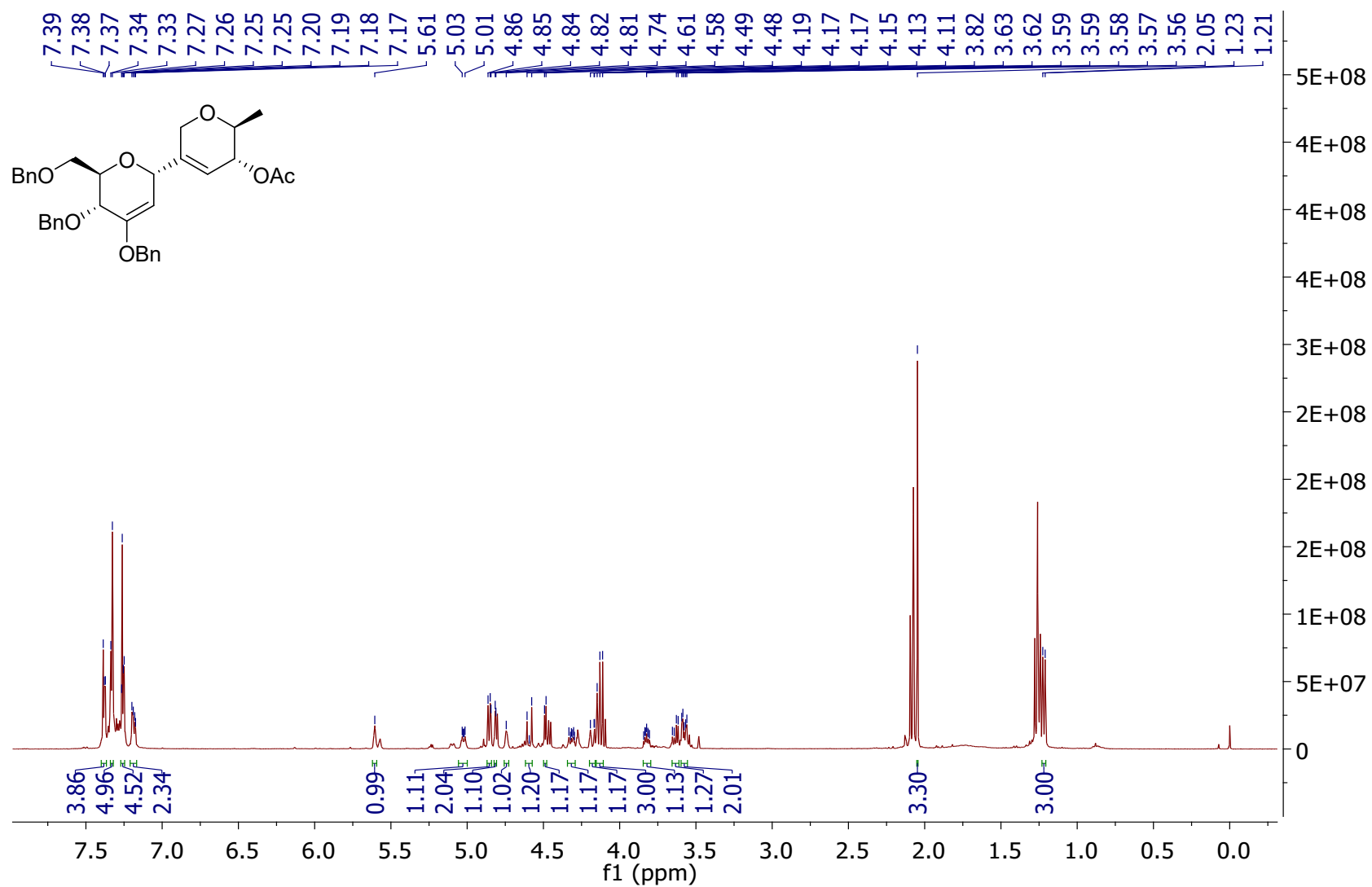


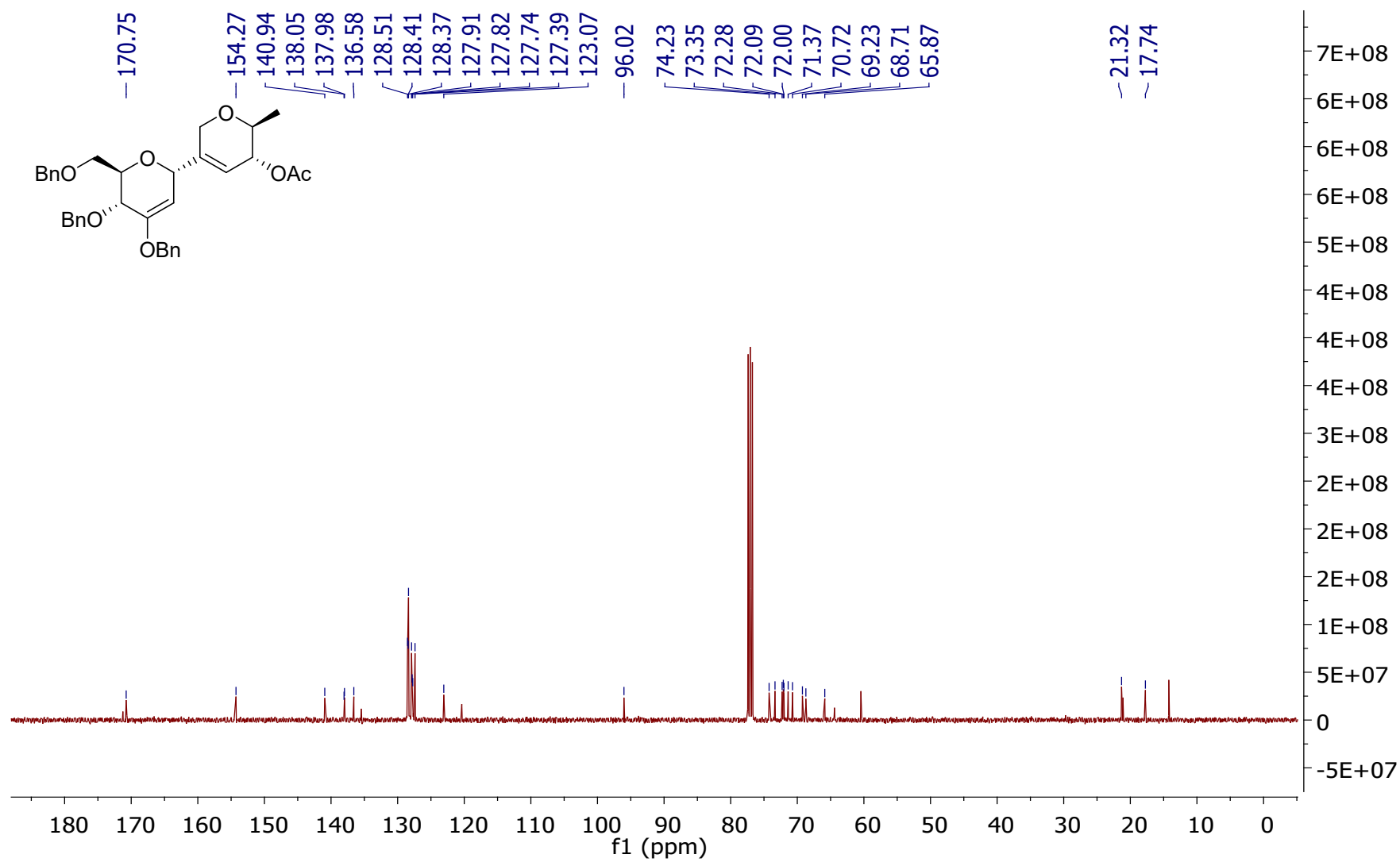
^1H NMR (400 MHz, CDCl_3) and ^{13}C $\{^1\text{H}\}$ NMR (101 MHz, CDCl_3) of compound 6d



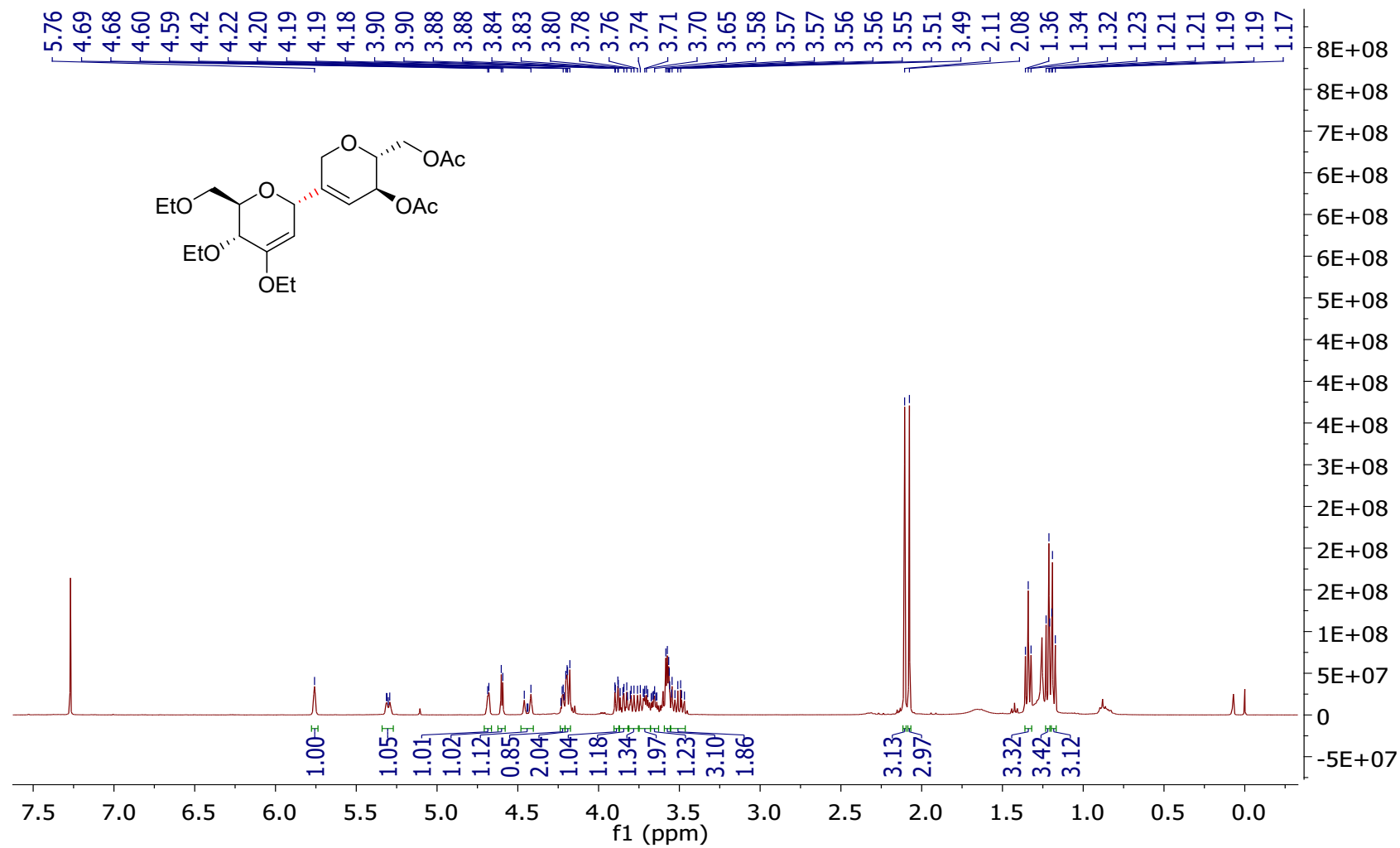


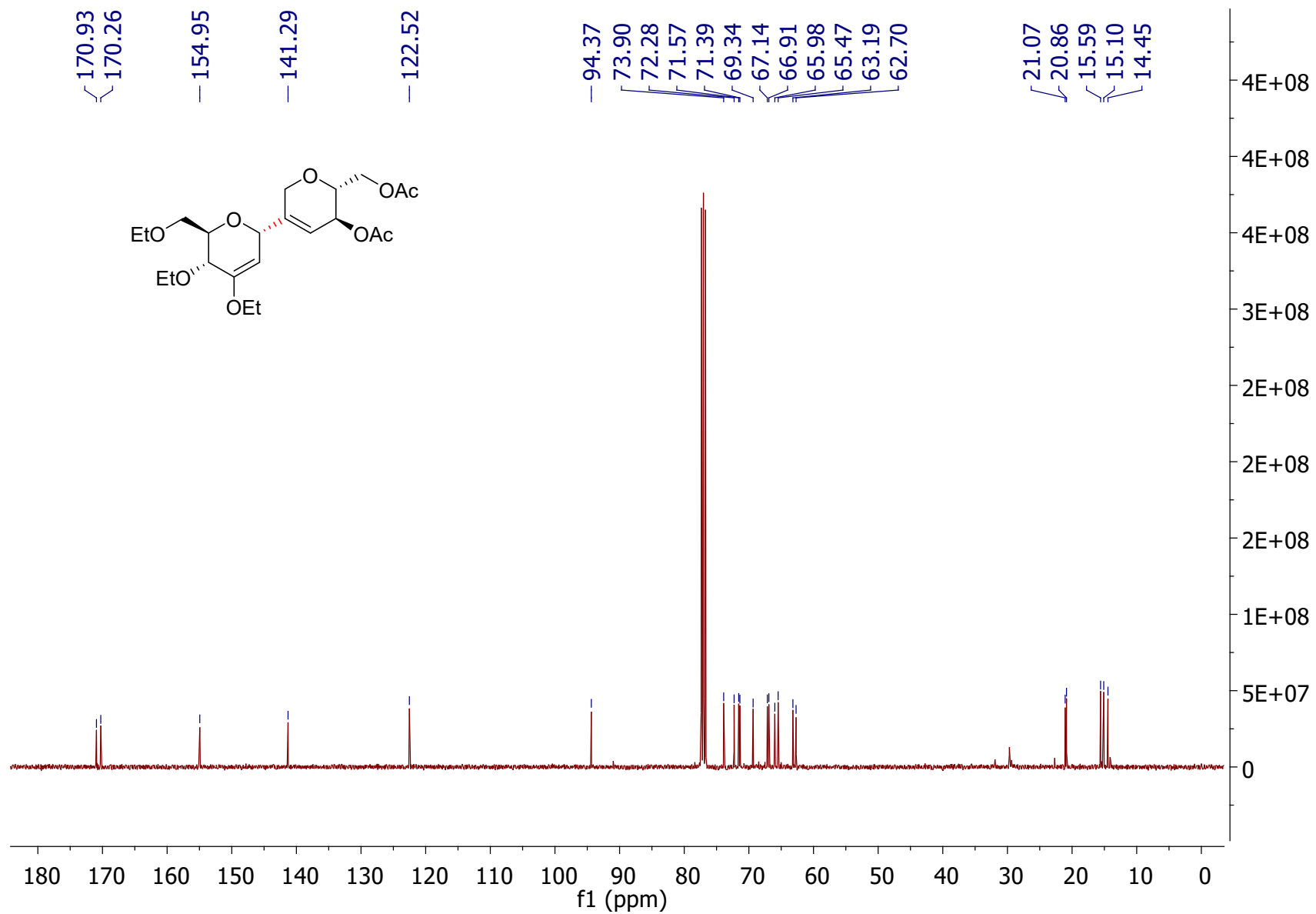
¹H NMR (400 MHz, CDCl₃) and ¹³C {¹H} NMR (101 MHz, CDCl₃) of compound 6e:





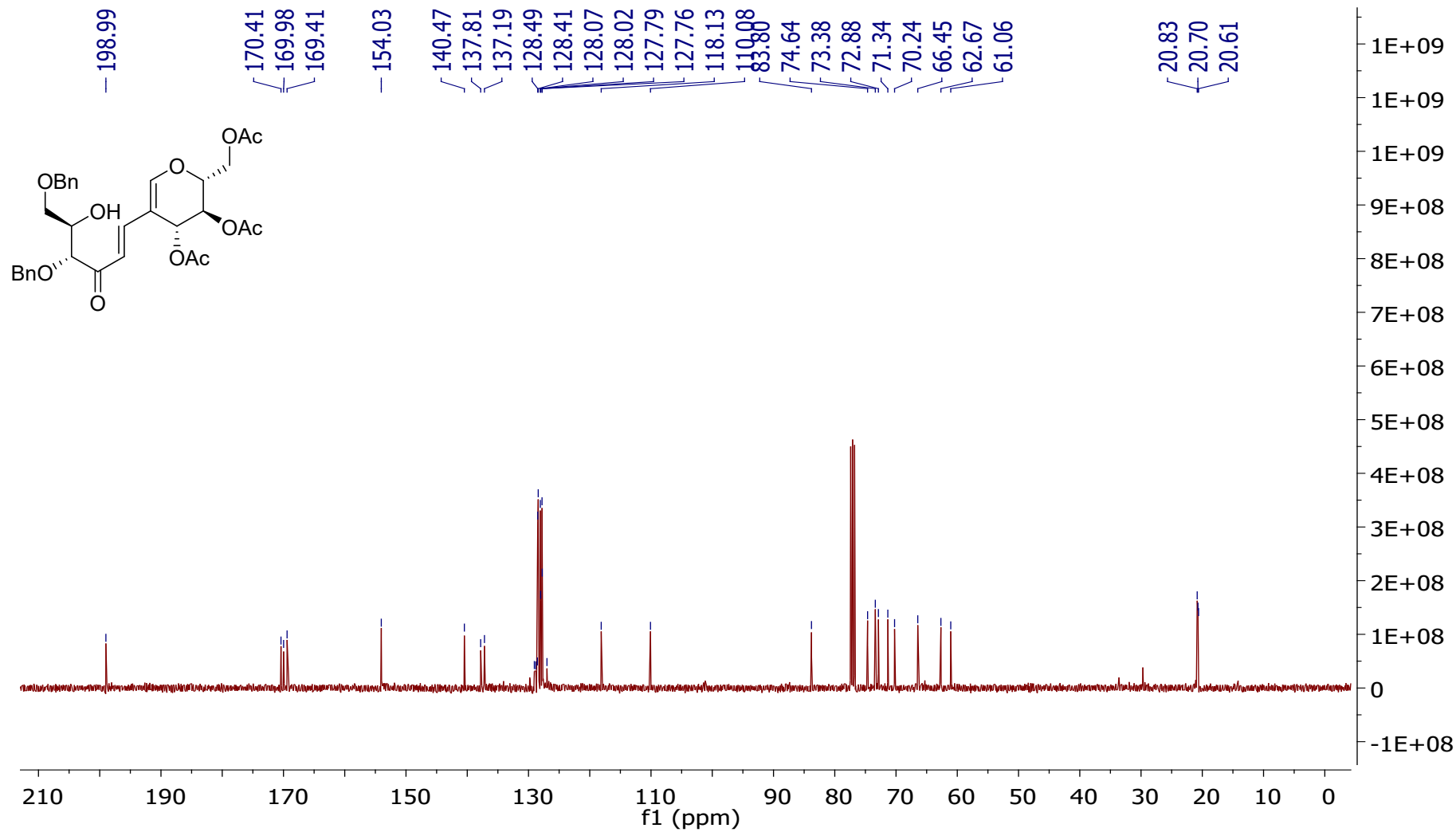
^1H NMR (400 MHz, CDCl_3) and ^{13}C $\{^1\text{H}\}$ NMR (101 MHz, CDCl_3) of compound 6f



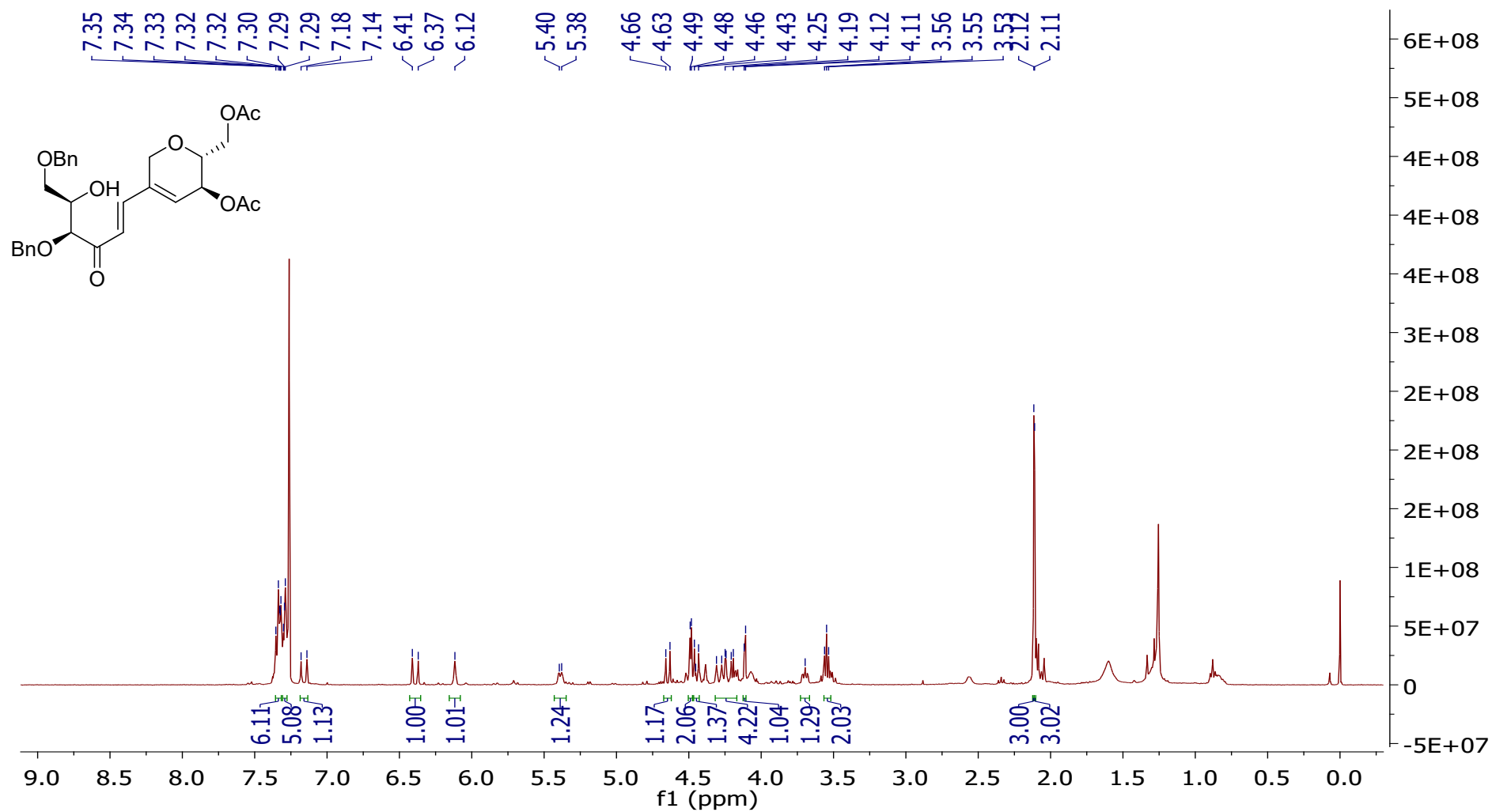


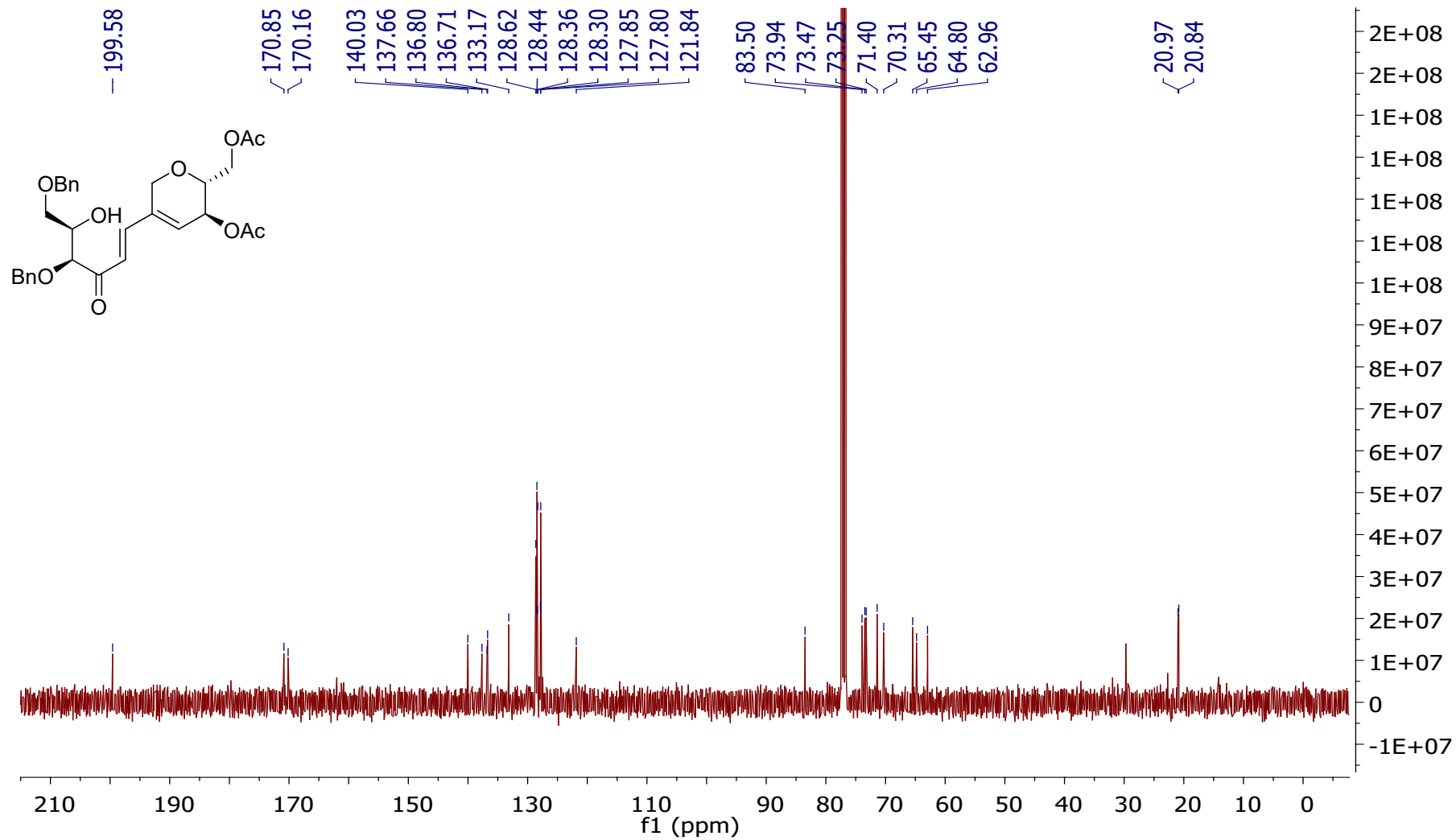
Chemical structure of compound 10 is shown in the top left. The ^1H NMR spectrum (CDCl₃) is displayed below, with chemical shift (f1, ppm) on the x-axis (0.0 to 9.0 ppm) and intensity on the y-axis (0 to 4E+08). The spectrum shows several peaks, with integration values provided below the baseline and peak labels on the right side.

Chemical Shift (ppm)	Integration	Label
7.36	1.38	
7.35	1.14	
7.34	2.15	
7.32	2.98	
7.30	2.13	
7.28	2.38	
7.27	1.03	
7.26	1.09	
7.04	1.06	
6.44	1.12	
6.40	1.42	
5.63	0.99	
5.62	2.11	
5.17	1.06	
5.17	1.31	
4.68	1.19	
4.65	1.19	
4.62	1.12	
4.53	2.05	
4.50		
4.49		
4.46		
4.45	3.12	
4.43	3.13	
4.42	3.00	
4.21		
4.05		
4.02		
4.00		
3.60		
3.58		
2.10		
2.09		
2.00		

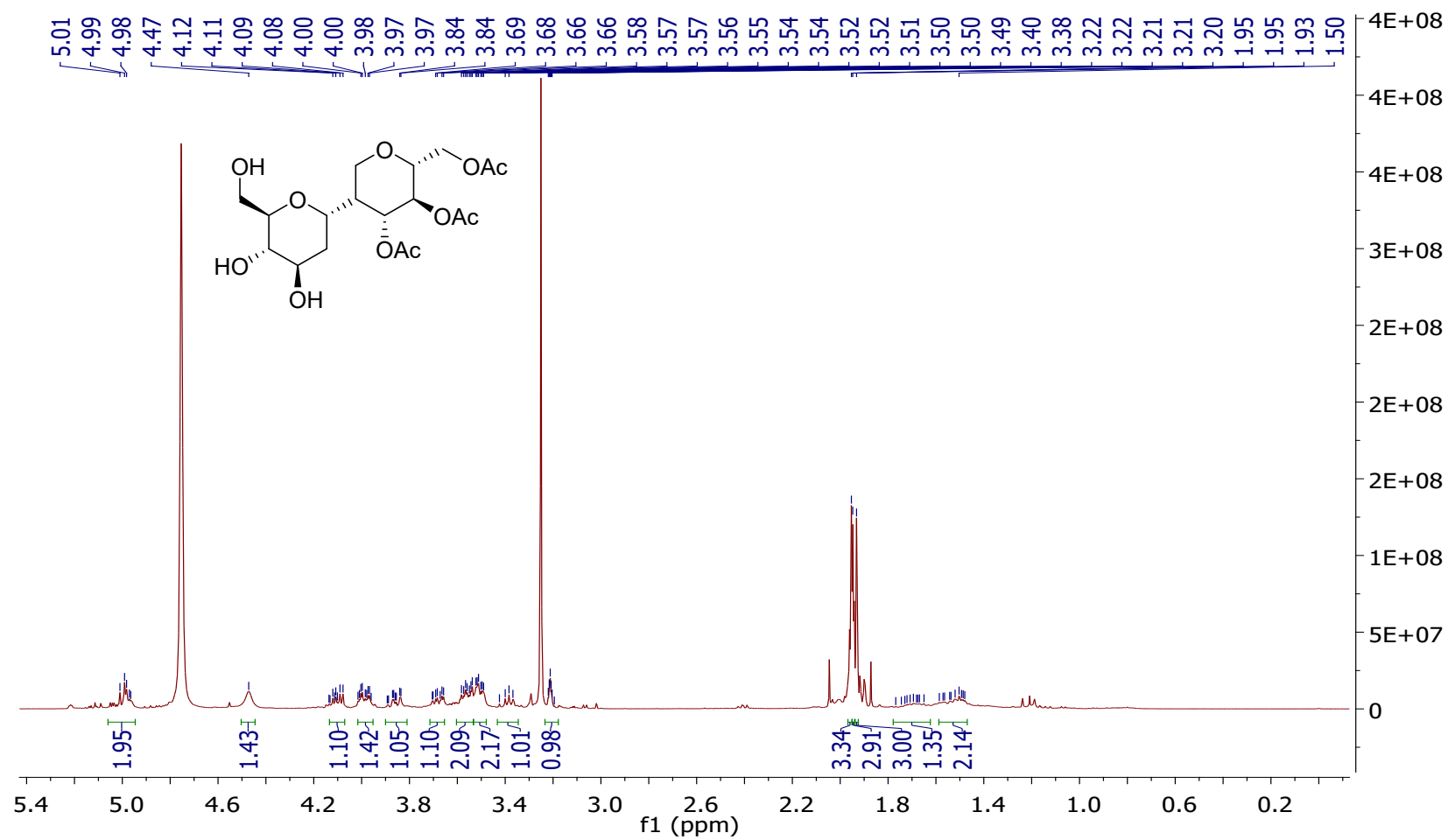


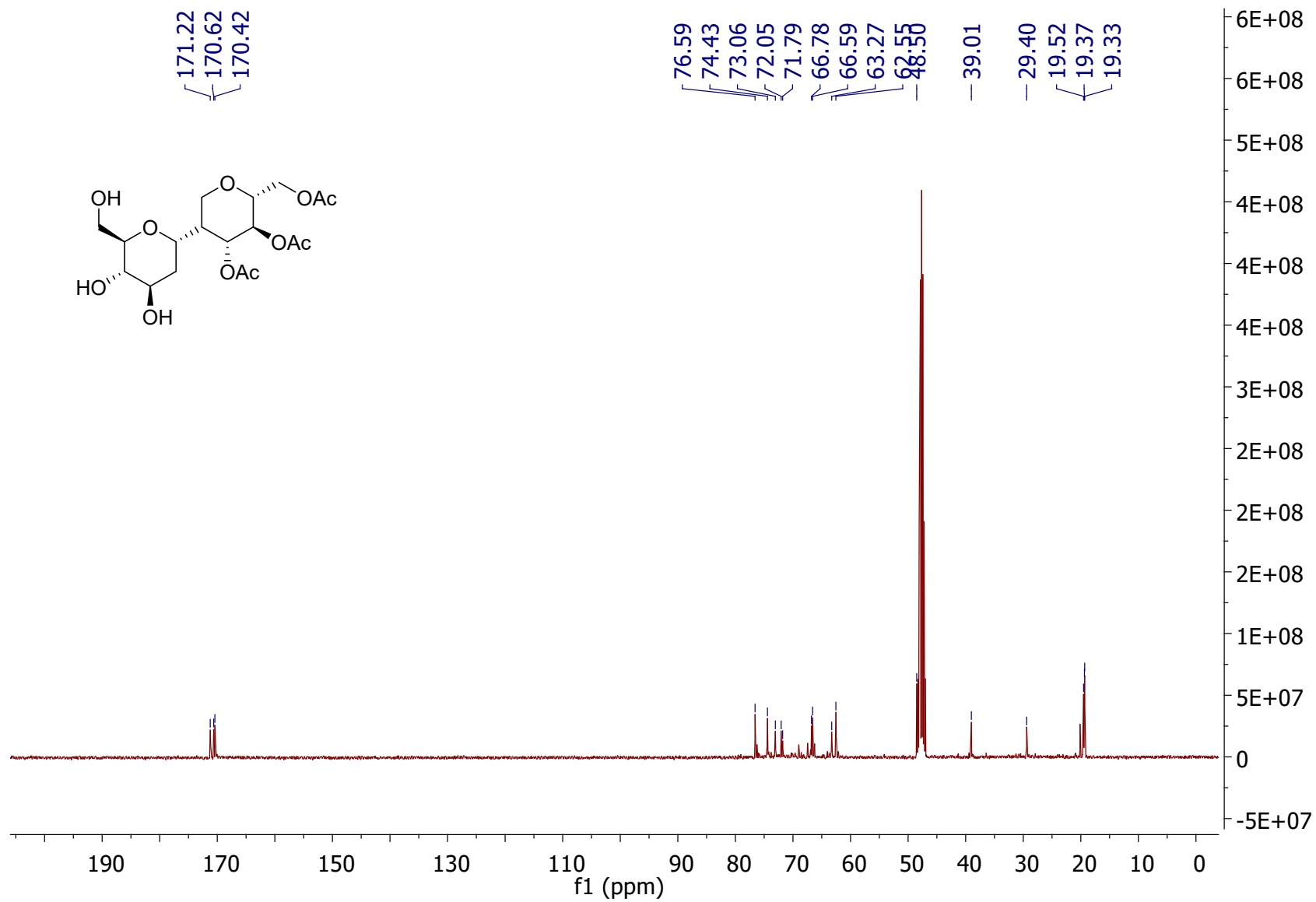
¹H (400 MHz, CDCl₃) & ¹³C {¹H} (101 MHz, CDCl₃) NMR of compound 8





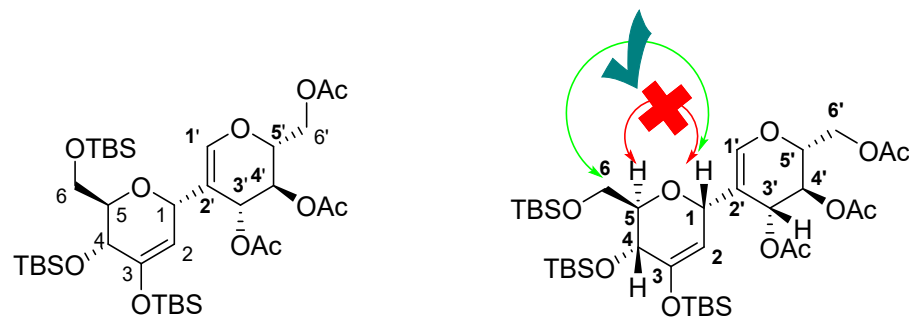
^1H (400 MHz, CDCl_3) & ^{13}C $\{^1\text{H}\}$ (101 MHz, CDCl_3) NMR of compound 9





4.1 2D Data of Compound 3d:

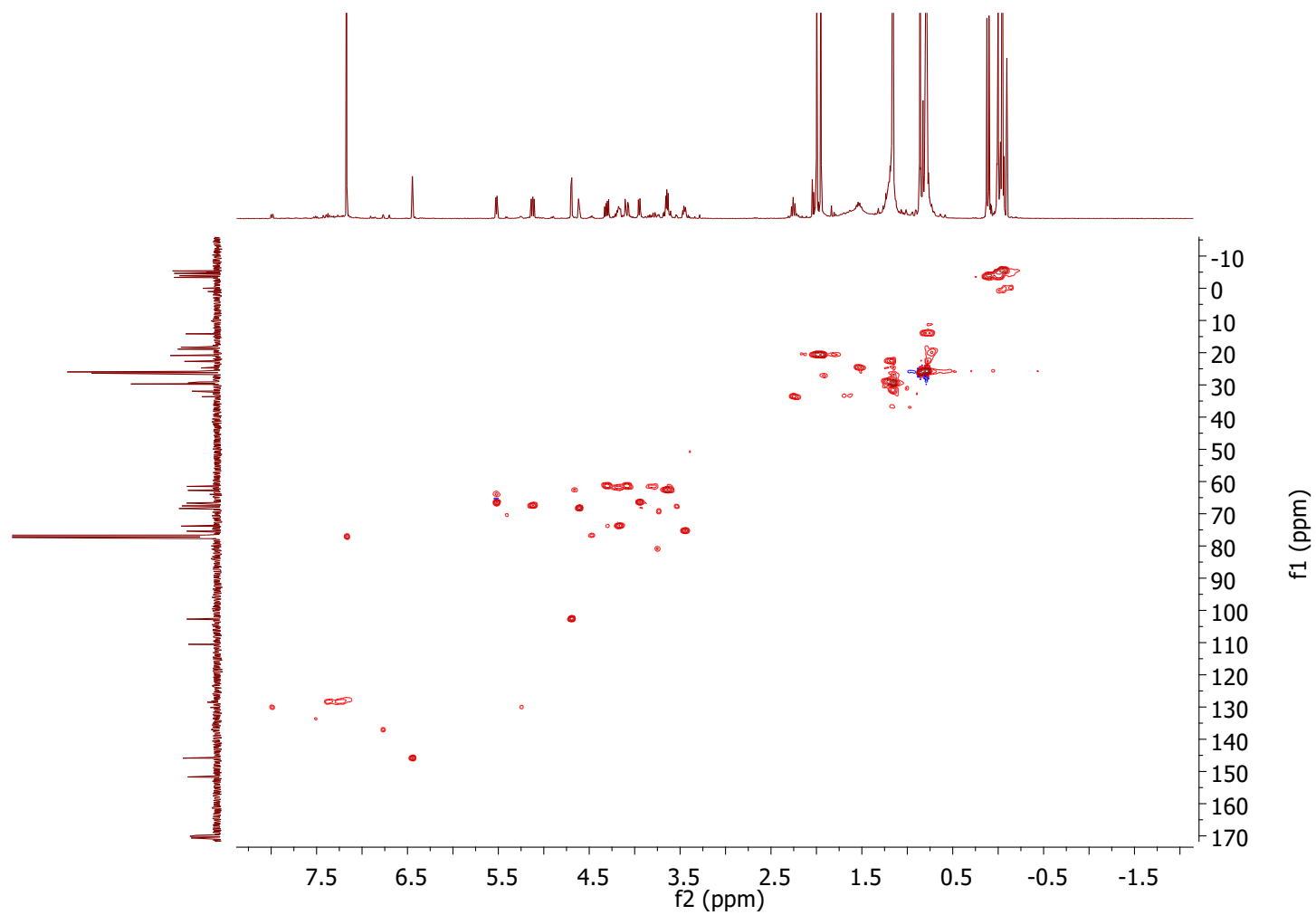
4.1.1 Structure determination of compound 3d:



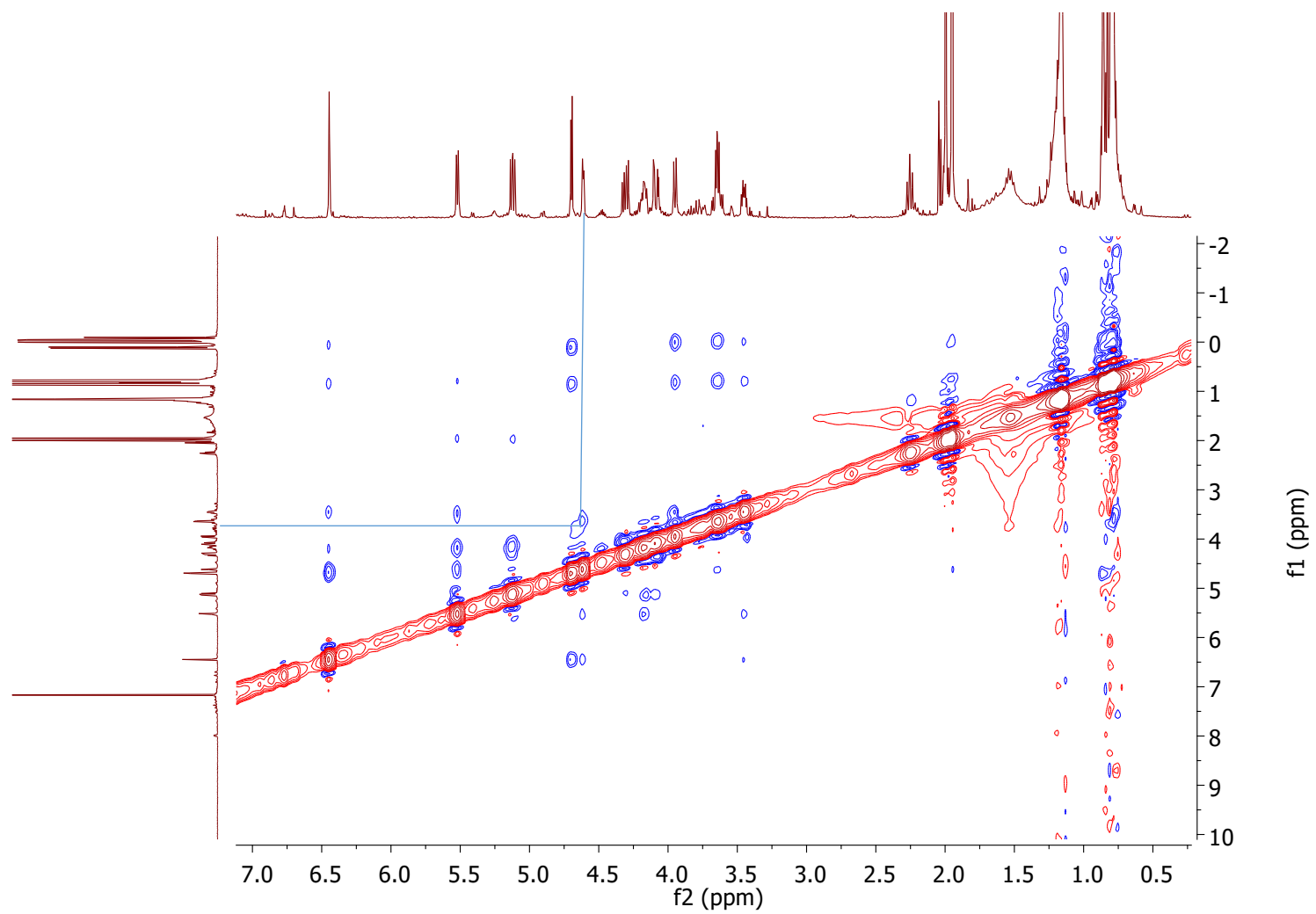
Atom Position	Type Of Atom	¹ H(ppm)	¹³ C(ppm)
1	CH	4.61	68.4
2	CH	4.70	102.7
3	C		151.7
4	CH	3.95	66.56
5	CH	3.45	75.39
6	CH ₂	3.64,3.65	62.8
1'	CH	6.45	145.9
2'	C		110.51
3'	CH	5.52	66.56
4'	CH	5.12	67.56
5'	CH	4.18	73.84
6'	CH ₂	4.09,4.31	61.52

The structure of product 3d was confirmed by 2D NMR analysis. In NOESY experiment we observed the strong correlation between H1 and H6 but no correlation was seen between H1 and H5 which confirmed the alpha stereochemistry at anomeric carbon.

4.1.2 HSQC of 3d



4.1.3 NOESY of 3d:



5. References:

- 1 S. Dharuman and Y. D. Vankar, 2014, 1172–1175.