Supporting Information for

Bifunctional Thiourea-Based Organocatalysts Promoted Kinetic Resolution Polymerization of Racemic Lactide to Isotactic Polylactide

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TABLE OF CONTENTS	<u>Pages</u>
Reagents, Instruments, and Methods	S7-8
Synthesis of catalysts	S9
Scheme S1. Synthesis of catalysts	S9
1. Synthesis of (R)-BINAM-Ac-NCS	S9
2. Synthesis of (S)-BINAM-Ac-NCS	S10
3. Synthesis of (R)-BINAM-TFA-NCS	S11
4. Synthesis of (R)-H ₈ -BINAM-Ac-NCS	S12
5. Synthesis of (<i>R</i>)-DACH-dimethyl	S13
6. Synthesis of (S)-DACH-dimethyl	S14
7. Synthesis of (S)-DACH-pyrrolidine	S15
8. Synthesis of (S)-DACH- ⁱ Pr	S16
9. Synthesis of (S)-DACH-ethanol-methyl	S17
10. Synthesis of (S)-DACH-diethyl	S18
11. Synthesis of catalyst (<i>R</i> , <i>S</i>)-1	S18
12. Synthesis of catalyst (<i>R</i> , <i>R</i>)-1	S19
13. Synthesis of catalyst (<i>S</i> , <i>S</i>)-1	S20
14. Synthesis of catalyst (<i>S</i> , <i>R</i>)-1	S20
15. Synthesis of catalyst (<i>R</i> , <i>S</i>)-2	S21
16. Synthesis of catalyst (<i>R</i> , <i>S</i>)-3	S21
17. Synthesis of catalyst (<i>R</i> , <i>S</i>)-4	S22
18. Synthesis of catalyst (<i>R</i> , <i>S</i>)-5	S23
19. Synthesis of catalyst (<i>R</i> , <i>S</i>)- 6	S23
20. Synthesis of catalyst (<i>R</i> , <i>S</i>)-7	S24
NMR spectra of catalysts	S25

Figure S1. ¹ H NMR spectrum (400 MHz, CDCl ₃) of (<i>R</i> , <i>S</i>)-1	S25
Figure S2. ¹³ C NMR spectrum (100 MHz, CDCl ₃) of (<i>R</i> , <i>S</i>)-1	S25
Figure S3. 1 H NMR spectrum (400 MHz, CDCl ₃) of (R , R)-1	S26
Figure S4. ¹³ C NMR spectrum (100 MHz, CDCl ₃) of (<i>R</i> , <i>R</i>)-1	S26
Figure S5. ¹ H NMR spectrum (400 MHz, CDCl ₃) of (S,S)-1	S27
Figure S6. ¹³ C NMR spectrum (100 MHz, CDCl ₃) of (S,S)-1	S27
Figure S7. ¹ H NMR spectrum (400 MHz, CDCl ₃) of (<i>S,R</i>)-1	S28
Figure S8. ¹³ C NMR spectrum (100 MHz, CDCl ₃) of (S,R)-1	S28
Figure S9. ¹ H NMR spectrum (400 MHz, CDCl ₃) of (<i>R</i> , <i>S</i>)-2	S29
Figure S10. 13 C NMR spectrum (100 MHz, CDCl ₃) of (R , S)-2	S29
Figure S11. ¹ H NMR spectrum (400 MHz, CDCl ₃) of (<i>R</i> , <i>S</i>)-3	S30
Figure S12. 13 C NMR spectrum (100 MHz, CDCl ₃) of (R , S)-3	S30
Figure S13. ¹ H NMR spectrum (400 MHz, CDCl ₃) of (<i>R</i> , <i>S</i>)-4	S31
Figure S14. ¹³ C NMR spectrum (100 MHz, CDCl ₃) of (<i>R</i> , <i>S</i>)-4	S31
Figure S15. ¹ H NMR spectrum (400 MHz, CDCl ₃) of (<i>R</i> , <i>S</i>)-5	S32
Figure S16. 13 C NMR spectrum (100 MHz, CDCl ₃) of (R , S)-5	S32
Figure S17. ¹ H NMR spectrum (400 MHz, CDCl ₃) of (<i>R</i> , <i>S</i>)-6	S33
Figure S18. ¹³ C NMR spectrum (100 MHz, CDCl ₃) of (<i>R</i> , <i>S</i>)-6	S33
Figure S19. ¹ H NMR spectrum (400 MHz, CDCl ₃) of (<i>R</i> , <i>S</i>)-7	S34
Figure S20. 13 C NMR spectrum (100 MHz, CDCl ₃) of (R , S)-7	S34
X-Ray crystallographic analysis	S35
Figure S21. X-ray structure of (R,S) -1	S35
Table S1 . Crystallographic table for (<i>R</i> , <i>S</i>)-1	S36
Procedure for polymerization	S37
Table S2. Ring-opening polymerization results of LA by different catalysts	S37
Table S3 . Ring-opening polymerization results of <i>rac</i> -LA by (<i>R</i> , <i>S</i>)-1 and Base	S38
Procedure for HPLC chromatograms of the unreacted monomer	S39
Figure S22. HPLC chromatograms of $_{D}$ -LA/ $_{L}$ -LA = 4/11	S39
Figure S23. HPLC chromatograms of unreacted monomer obtained by (R,S) -1 (Table	S41
S2 Entry 1)	
Kinetic study of the ROP of rac-LA, D-LA, and L-LA	S42
Table S4 . The raw date of kinetic for ROP of <i>rac</i> -LA, D-LA, and L-LA catalyzed by	S42
(R,S)-1	
Figure S24. Plots of $ln([LA]_0/[LA]_t)$ versus time for polymerization catalyzed by	S42
(R,S)-1	

Table S5 . The raw date of kinetic for ROP of <i>rac-</i> LA, D-LA, and L-LA catalyzed by	S43
(S,R)-1	
Figure S25. Plots of $ln([LA]_0/[LA]_t)$ versus time for polymerization catalyzed by	S43
(S,R)-1	
Table S6 . The raw date of kinetic for ROP of <i>rac-</i> LA, _D -LA, and _L -LA catalyzed by	S44
(R,S)-1	
Figure S26. Plots of $ln([LA]_0/[LA]_t)$ versus time for polymerization catalyzed by	S44
(R,S)-1	
Table S7 . The raw date of kinetic for ROP of <i>rac-</i> LA, _D -LA, and _L -LA catalyzed by	S45
(R,S)-2	
Figure S27. Plots of $ln([LA]_0/[LA]_t)$ versus time for polymerization catalyzed by	S45
(R,S)-2	
Thermal properties of polymers	S46
Figure S28. DSC curves of PLA obtained by $[rac-LA]/[(R,S)-1]/[I] = 50/2/1$, solvent	S46
= DCM	
Figure S29. DSC curve of PLA obtained by $[rac\text{-LA}]/[(R,S)\text{-}1]/[I] = 50/2/1$, solvent	S46
= TOL	
Figure S30. DSC curve of PLA obtained by $[rac\text{-LA}]/[(R,S)\text{-}1]/[I] = 50/2/1$, solvent	S47
$= CHCl_3$	
Figure S31. DSC curve for PLA obtained by $[rac-LA]/[(S,R)-1]/[I] = 50/2/1$, solvent	S47
= DCM	
Figure S32. DSC curve for PLA obtained by $[rac\text{-LA}]/[(R,R)\text{-}1]/[I] = 50/2/1$, solvent	S48
= DCM	
Figure S33. DSC curve for PLA obtained by $[rac-LA]/[(S,S)-1]/[I] = 50/2/1$, solvent	S48
= DCM	
Figure S34. DSC curve for PLA obtained by $[rac-LA]/[(R,S)-2]/[I] = 50/2/1$, solvent	S49
= DCM	
Figure S35. DSC curve for PLA obtained by $[rac-LA]/[(R,S)-3]/[I] = 50/2/1$, solvent	S49
= DCM	
Figure S36. DSC curve for PLA obtained by $[rac-LA]/[(R,S)-6]/[I] = 50/2/1$, solvent	S50
= DCM	
Figure S37. DSC curve for PLA obtained by $[rac-LA]/[(R,S)-7]/[I] = 50/2/1$, solvent	S50
= DCM	
Figure S38. DSC curve for PLA obtained by $[rac-LA]/[(R,S)-1]/[I] = 200/20/1$,	S51
solvent = DCM	

Figure S39. DSC curve for PLA obtained by [rac-LA]/[(S,R)-1]/[I] = 200/20/1, S51 solvent = DCM**Figure S40.** DSC curves of PLA obtained by [rac-LA]/[(R,S)-1]/[I] = 50/2/1, solvent S52 = DCMFigure S41. DSC curves of PLA obtained by [D-LA]/[(R,S)-1]/[I] = 50/2/1, solvent = S52 CHC₁₃ **Figure S42.** DSC curves of PLA obtained by [rac-LA]/[(S,R)-1]/[I] = 50/2/1, solvent S53 $= CHCl_3$ Figure S43. DSC curves of PLA obtained by [L-LA]/[(S,R)-1]/[I] = 50/2/1, solvent = S53 CHCl₃ Figure S44. DSC curves of PLA obtained by [D-LA]/[(R,S)-1]/[I] = 50/2/1, solvent = DCM. Figure S45. DSC curves of PLA obtained by [rac-LA]/[(R,S)-2]/[I] = 50/2/1, solvent S54 = DCMFigure S46. DSC curves of PLA obtained by [D-LA]/[(R,S)-2]/[I] = 50/2/1, solvent = S55 **DCM SEC trace of PLAs** S56 Figure S47. SEC trace of PLA obtained by [rac-LA]/[(R,S)-1]/[I] = 50/2/1, solvent = S56 DCM ($M_n = 6.4 \text{ kg/mol}, D = 1.08$) Figure S48. SEC trace of PLA obtained by [rac-LA]/[(R,S)-1]/[I] = 50/2/1, solvent = TOL $(M_n = 5.6 \text{ kg/mol}, D = 1.08)$ Figure S49. SEC trace of PLA obtained by [rac-LA]/[(R,S)-1]/[I] = 50/2/1, solvent = S57 CHCl₃ ($M_n = 6.2 \text{ kg/mol}, D = 1.09$) **Figure S50.** SEC trace of PLA obtained by [rac-LA]/[(R,R)-1]/[I] = 50/2/1, solvent = S57 DCM ($M_n = 6.0 \text{ kg/mol}, D = 1.06$) Figure S51. SEC trace of PLA obtained by [rac-LA]/[(S,S)-1]/[1] = 50/2/1, solvent = S58 DCM ($M_n = 6.2 \text{ kg/mol}, D = 1.06$) **Figure S52.** SEC trace of PLA obtained by [rac-LA]/[(R,S)-2]/[I] = 50/2/1, solvent = S58 DCM ($M_n = 5.6 \text{ kg/mol}, D = 1.07$) Figure S53. SEC trace of PLA obtained by [rac-LA]/[(R,S)-3]/[I] = 50/2/1, solvent = DCM ($M_n = 5.3 \text{ kg/mol}, D = 1.07$) Figure S54. SEC trace of PLA obtained by [rac-LA]/[(R,S)-6]/[I] = 50/2/1, solvent = S59 DCM ($M_n = 6.0 \text{ kg/mol}, D = 1.07$) Figure S55. SEC trace of PLA obtained by [rac-LA]/[(R,S)-7]/[I] = 50/2/1, solvent = DCM ($M_n = 5.3 \text{ kg/mol}, D = 1.07$)

Figure S56. SEC trace of PLA obtained by $[rac\text{-LA}]/[(R,S)\text{-}1]/[I] = 200/20/1$, solvent	S60
= DCM (M_n = 19.7 kg/mol, D = 1.09)	
Figure S57. SEC trace of PLA obtained by $[rac\text{-LA}]/[(S,R)\text{-}1]/[I] = 200/20/1$, solvent	S61
= DCM (M_n = 19.1 kg/mol, D = 1.10)	
Figure S58. SEC trace of PLA obtained by $[rac-LA]/[(R,S)-1]/[I] = 50/2/1$, solvent =	S61
DCM ($M_n = 5.5 \text{ kg/mol}, D = 1.09$)	
Figure S59. SEC trace of PLA obtained by $[D-LA]/[(R,S)-1]/[I] = 50/2/1$, solvent =	S62
CHCl ₃ ($M_n = 12.4 \text{ kg/mol}, D = 1.08$)	
Figure S60. SEC trace of PLA obtained by $[rac\text{-LA}]/[(S,R)\text{-}1]/[I] = 50/2/1$, solvent =	S62
CHCl ₃ ($M_n = 6.7 \text{ kg/mol}, D = 1.08$)	
Figure S61. SEC trace of PLA obtained by $[L-LA]/[(S,R)-1]/[I] = 50/2/1$, solvent =	S63
CHCl ₃ ($M_n = 11.8 \text{ kg/mol}, D = 1.07$)	
Figure S62. SEC trace of PLA obtained by $[D-LA]/[(R,S)-1]/[I] = 50/2/1$, solvent =	S63
DCM ($M_n = 13.5 \text{ kg/mol}$, $D = 1.06$)	
Figure S63. SEC trace of PLA obtained by $[rac\text{-LA}]/[(R,S)\text{-2}]/[I] = 50/2/1$, solvent =	S64
DCM ($M_n = 7.0 \text{ kg/mol}, D = 1.07$)	
Figure S64. SEC trace of PLA obtained by $[D-LA]/[(R,S)-2]/[I] = 50/2/1$, solvent =	S64
DCM (14 1001 / 1 D 100)	
DCM $(M_n = 12.8 \text{ kg/mol}, D = 1.06)$	
Homonuclear decoupled ¹ H NMR spectra of PLAs	S65
	S65 S65
Homonuclear decoupled ¹ H NMR spectra of PLAs	
Homonuclear decoupled ¹ H NMR spectra of PLAs Table S8. Tetrad probabilities of ESC mechanisms based on non-Bernouillanin	S65
Homonuclear decoupled ¹ H NMR spectra of PLAs Table S8. Tetrad probabilities of ESC mechanisms based on non-Bernouillanin Figure S65. Homonuclear decoupled ¹ H NMR spectrum (400 MHz, CDCl ₃) of PLA	S65 S65
Homonuclear decoupled ${}^{1}H$ NMR spectra of PLAs Table S8. Tetrad probabilities of ESC mechanisms based on non-Bernouillanin Figure S65. Homonuclear decoupled ${}^{1}H$ NMR spectrum (400 MHz, CDCl ₃) of PLA obtained by $[rac\text{-LA}]/[(R,S)\text{-1}]/[I] = 50/2/1$, solvent = DCM ($P_{\rm m} = 0.96$)	S65 S65
Homonuclear decoupled ¹ H NMR spectra of PLAs Table S8. Tetrad probabilities of ESC mechanisms based on non-Bernouillanin Figure S65. Homonuclear decoupled ¹ H NMR spectrum (400 MHz, CDCl ₃) of PLA obtained by [rac-LA]/[(R,S)-1]/[I] = 50/2/1, solvent = DCM (P _m = 0.96) Figure S66. Homonuclear decoupled ¹ H NMR spectrum (400 MHz, CDCl ₃) of PLA	S65 S65 S66
Homonuclear decoupled ¹ H NMR spectra of PLAs Table S8. Tetrad probabilities of ESC mechanisms based on non-Bernouillanin Figure S65. Homonuclear decoupled ¹ H NMR spectrum (400 MHz, CDCl ₃) of PLA obtained by $[rac\text{-LA}]/[(R,S)\text{-1}]/[I] = 50/2/1$, solvent = DCM ($P_m = 0.96$) Figure S66. Homonuclear decoupled ¹ H NMR spectrum (400 MHz, CDCl ₃) of PLA obtained by $[rac\text{-LA}]/[(R,S)\text{-1}]/[I] = 50/2/1$, solvent = TOL ($P_m = 0.93$)	S65 S65 S66
Homonuclear decoupled ¹ H NMR spectra of PLAs Table S8. Tetrad probabilities of ESC mechanisms based on non-Bernouillanin Figure S65. Homonuclear decoupled ¹ H NMR spectrum (400 MHz, CDCl ₃) of PLA obtained by $[rac\text{-LA}]/[(R,S)\text{-1}]/[I] = 50/2/1$, solvent = DCM ($P_m = 0.96$) Figure S66. Homonuclear decoupled ¹ H NMR spectrum (400 MHz, CDCl ₃) of PLA obtained by $[rac\text{-LA}]/[(R,S)\text{-1}]/[I] = 50/2/1$, solvent = TOL ($P_m = 0.93$) Figure S67. Homonuclear decoupled ¹ H NMR spectrum (400 MHz, CDCl ₃) of PLA obtained by $[rac\text{-LA}]/[(R,S)\text{-1}]/[I] = 50/2/1$, solvent = CHCl ₃ ($P_m = 0.94$)	S65 S65 S66
Homonuclear decoupled ¹ H NMR spectra of PLAs Table S8. Tetrad probabilities of ESC mechanisms based on non-Bernouillanin Figure S65. Homonuclear decoupled ¹ H NMR spectrum (400 MHz, CDCl ₃) of PLA obtained by $[rac\text{-LA}]/[(R,S)\text{-1}]/[I] = 50/2/1$, solvent = DCM ($P_m = 0.96$) Figure S66. Homonuclear decoupled ¹ H NMR spectrum (400 MHz, CDCl ₃) of PLA obtained by $[rac\text{-LA}]/[(R,S)\text{-1}]/[I] = 50/2/1$, solvent = TOL ($P_m = 0.93$) Figure S67. Homonuclear decoupled ¹ H NMR spectrum (400 MHz, CDCl ₃) of PLA obtained by $[rac\text{-LA}]/[(R,S)\text{-1}]/[I] = 50/2/1$, solvent = CHCl ₃ ($P_m = 0.94$)	\$65 \$65 \$66 \$67
Homonuclear decoupled ¹ H NMR spectra of PLAs Table S8. Tetrad probabilities of ESC mechanisms based on non-Bernouillanin Figure S65. Homonuclear decoupled ¹ H NMR spectrum (400 MHz, CDCl ₃) of PLA obtained by [rac-LA]/[(R,S)-1]/[I] = 50/2/1, solvent = DCM (P _m = 0.96) Figure S66. Homonuclear decoupled ¹ H NMR spectrum (400 MHz, CDCl ₃) of PLA obtained by [rac-LA]/[(R,S)-1]/[I] = 50/2/1, solvent = TOL (P _m = 0.93) Figure S67. Homonuclear decoupled ¹ H NMR spectrum (400 MHz, CDCl ₃) of PLA obtained by [rac-LA]/[(R,S)-1]/[I] = 50/2/1, solvent = CHCl ₃ (P _m = 0.94) Figure S68. Homonuclear decoupled ¹ H NMR spectrum (400 MHz, CDCl ₃) of PLA	\$65 \$65 \$66 \$67 \$68
Homonuclear decoupled ¹ H NMR spectra of PLAs Table S8. Tetrad probabilities of ESC mechanisms based on non-Bernouillanin Figure S65. Homonuclear decoupled ¹ H NMR spectrum (400 MHz, CDCl ₃) of PLA obtained by $[rac\text{-LA}]/[(R,S)\text{-1}]/[I] = 50/2/1$, solvent = DCM ($P_m = 0.96$) Figure S66. Homonuclear decoupled ¹ H NMR spectrum (400 MHz, CDCl ₃) of PLA obtained by $[rac\text{-LA}]/[(R,S)\text{-1}]/[I] = 50/2/1$, solvent = TOL ($P_m = 0.93$) Figure S67. Homonuclear decoupled ¹ H NMR spectrum (400 MHz, CDCl ₃) of PLA obtained by $[rac\text{-LA}]/[(R,S)\text{-1}]/[I] = 50/2/1$, solvent = CHCl ₃ ($P_m = 0.94$) Figure S68. Homonuclear decoupled ¹ H NMR spectrum (400 MHz, CDCl ₃) of PLA obtained by $[rac\text{-LA}]/[(S,R)\text{-1}]/[I] = 50/2/1$, solvent = DCM ($P_m = 0.96$)	\$65 \$65 \$66 \$67 \$68
Homonuclear decoupled ¹ H NMR spectra of PLAs Table S8. Tetrad probabilities of ESC mechanisms based on non-Bernouillanin Figure S65. Homonuclear decoupled ¹ H NMR spectrum (400 MHz, CDCl ₃) of PLA obtained by $[rac\text{-LA}]/[(R,S)\text{-1}]/[I] = 50/2/1$, solvent = DCM ($P_m = 0.96$) Figure S66. Homonuclear decoupled ¹ H NMR spectrum (400 MHz, CDCl ₃) of PLA obtained by $[rac\text{-LA}]/[(R,S)\text{-1}]/[I] = 50/2/1$, solvent = TOL ($P_m = 0.93$) Figure S67. Homonuclear decoupled ¹ H NMR spectrum (400 MHz, CDCl ₃) of PLA obtained by $[rac\text{-LA}]/[(R,S)\text{-1}]/[I] = 50/2/1$, solvent = CHCl ₃ ($P_m = 0.94$) Figure S68. Homonuclear decoupled ¹ H NMR spectrum (400 MHz, CDCl ₃) of PLA obtained by $[rac\text{-LA}]/[(S,R)\text{-1}]/[I] = 50/2/1$, solvent = DCM ($P_m = 0.96$) Figure S69. Homonuclear decoupled ¹ H NMR spectrum (400 MHz, CDCl ₃) of PLA obtained by $[rac\text{-LA}]/[(R,R)\text{-1}]/[I] = 50/2/1$, solvent = DCM ($P_m = 0.96$)	\$65 \$65 \$66 \$67 \$68
Homonuclear decoupled ¹ H NMR spectra of PLAs Table S8. Tetrad probabilities of ESC mechanisms based on non-Bernouillanin Figure S65. Homonuclear decoupled ¹ H NMR spectrum (400 MHz, CDCl ₃) of PLA obtained by $[rac\text{-LA}]/[(R,S)\text{-1}]/[I] = 50/2/1$, solvent = DCM ($P_m = 0.96$) Figure S66. Homonuclear decoupled ¹ H NMR spectrum (400 MHz, CDCl ₃) of PLA obtained by $[rac\text{-LA}]/[(R,S)\text{-1}]/[I] = 50/2/1$, solvent = TOL ($P_m = 0.93$) Figure S67. Homonuclear decoupled ¹ H NMR spectrum (400 MHz, CDCl ₃) of PLA obtained by $[rac\text{-LA}]/[(R,S)\text{-1}]/[I] = 50/2/1$, solvent = CHCl ₃ ($P_m = 0.94$) Figure S68. Homonuclear decoupled ¹ H NMR spectrum (400 MHz, CDCl ₃) of PLA obtained by $[rac\text{-LA}]/[(S,R)\text{-1}]/[I] = 50/2/1$, solvent = DCM ($P_m = 0.96$) Figure S69. Homonuclear decoupled ¹ H NMR spectrum (400 MHz, CDCl ₃) of PLA obtained by $[rac\text{-LA}]/[(R,R)\text{-1}]/[I] = 50/2/1$, solvent = DCM ($P_m = 0.96$)	\$65 \$65 \$66 \$67 \$68 \$69
Homonuclear decoupled ¹ H NMR spectra of PLAs Table S8. Tetrad probabilities of ESC mechanisms based on non-Bernouillanin Figure S65. Homonuclear decoupled ¹ H NMR spectrum (400 MHz, CDCl ₃) of PLA obtained by [rac-LA]/[(R,S)-1]/[I] = 50/2/1, solvent = DCM (P _m = 0.96) Figure S66. Homonuclear decoupled ¹ H NMR spectrum (400 MHz, CDCl ₃) of PLA obtained by [rac-LA]/[(R,S)-1]/[I] = 50/2/1, solvent = TOL (P _m = 0.93) Figure S67. Homonuclear decoupled ¹ H NMR spectrum (400 MHz, CDCl ₃) of PLA obtained by [rac-LA]/[(R,S)-1]/[I] = 50/2/1, solvent = CHCl ₃ (P _m = 0.94) Figure S68. Homonuclear decoupled ¹ H NMR spectrum (400 MHz, CDCl ₃) of PLA obtained by [rac-LA]/[(S,R)-1]/[I] = 50/2/1, solvent = DCM (P _m = 0.96) Figure S69. Homonuclear decoupled ¹ H NMR spectrum (400 MHz, CDCl ₃) of PLA obtained by [rac-LA]/[(R,R)-1]/[I] = 50/2/1, solvent = DCM (P _m = 0.89) Figure S70. Homonuclear decoupled ¹ H NMR spectrum (400 MHz, CDCl ₃) of PLA	\$65 \$65 \$66 \$67 \$68 \$69
Homonuclear decoupled 1 H NMR spectra of PLAs Table S8. Tetrad probabilities of ESC mechanisms based on non-Bernouillanin Figure S65. Homonuclear decoupled 1 H NMR spectrum (400 MHz, CDCl ₃) of PLA obtained by [rac -LA]/[(R,S) -1]/[I] = 50/2/1, solvent = DCM ($P_{\rm m}$ = 0.96) Figure S66. Homonuclear decoupled 1 H NMR spectrum (400 MHz, CDCl ₃) of PLA obtained by [rac -LA]/[(R,S) -1]/[I] = 50/2/1, solvent = TOL ($P_{\rm m}$ = 0.93) Figure S67. Homonuclear decoupled 1 H NMR spectrum (400 MHz, CDCl ₃) of PLA obtained by [rac -LA]/[(R,S) -1]/[I] = 50/2/1, solvent = CHCl ₃ ($P_{\rm m}$ = 0.94) Figure S68. Homonuclear decoupled 1 H NMR spectrum (400 MHz, CDCl ₃) of PLA obtained by [rac -LA]/[(S,R) -1]/[I] = 50/2/1, solvent = DCM ($P_{\rm m}$ = 0.96) Figure S69. Homonuclear decoupled 1 H NMR spectrum (400 MHz, CDCl ₃) of PLA obtained by [rac -LA]/[(R,R) -1]/[I] = 50/2/1, solvent = DCM ($P_{\rm m}$ = 0.89) Figure S70. Homonuclear decoupled 1 H NMR spectrum (400 MHz, CDCl ₃) of PLA obtained by [rac -LA]/[(S,S) -1]/[I] = 50/2/1, solvent = DCM ($P_{\rm m}$ = 0.87)	\$65 \$65 \$66 \$67 \$68 \$69

obtained by [rac-LA]/[(R,S)-3]/[I] = 50/2/1, solvent = DCM $(P_m = 0.91)$ Figure S73. Homonuclear decoupled ¹H NMR spectrum (400 MHz, CDCl₃) of PLA S73 obtained by [rac-LA]/[(R,S)-6]/[I] = 50/2/1, solvent = DCM $(P_m = 0.90)$ Figure S74. Homonuclear decoupled ¹H NMR spectrum (400 MHz, CDCl₃) of PLA S74 obtained by [rac-LA]/[(R,S)-7]/[I] = 50/2/1, solvent = DCM $(P_m = 0.90)$ Figure S75. Homonuclear decoupled ¹H NMR spectrum (400 MHz, CDCl₃) of PLA S75 obtained by [rac-LA]/[(R,S)-1]/[I] = 200/20/1, solvent = DCM $(P_m = 0.94)$ Figure S76. Homonuclear decoupled ¹H NMR spectrum (400 MHz, CDCl₃) of PLA S76 obtained by [rac-LA]/[(S,R)-1]/[I] = 200/20/1, solvent = DCM $(P_m = 0.93)$ Figure S77. Homonuclear decoupled ¹H NMR spectrum (400 MHz, CDCl₃) of PLA S77 obtained by [rac-LA]/[(R,S)-1]/[I] = 50/2/1, solvent = DCM $(P_m = 0.95)$ Figure S78. Homonuclear decoupled ¹H NMR spectrum (400 MHz, CDCl₃) of PLA S78 obtained by [D-LA]/[(R,S)-1]/[I] = 50/2/1, solvent = CHCl₃ ($P_m = 0.99$) Figure S79. Homonuclear decoupled ¹H NMR spectrum (400 MHz, CDCl₃) of PLA S79 obtained by [rac-LA]/[(S,R)-1]/[I] = 50/2/1, solvent = CHCl₃ ($P_m = 0.95$) Figure S80. Homonuclear decoupled ¹H NMR spectrum (400 MHz, CDCl₃) of PLA S80 obtained by [L-LA]/[(S,R)-1]/[I] = 50/2/1, solvent = CHCl₃ ($P_m = 0.99$) Figure S81. Homonuclear decoupled ¹H NMR spectrum (400 MHz, CDCl₃) of PLA S81 obtained by [D-LA]/[(R,S)-1]/[I] = 50/2/1, solvent = DCM $(P_m = 0.99)$ Figure S82. Homonuclear decoupled ¹H NMR spectrum (400 MHz, CDCl₃) of PLA obtained by [D-LA]/[(R,S)-1]/[I] = 50/2/1, solvent = DCM ($P_m = 0.99$) Figure S83. Homonuclear decoupled ¹H NMR spectrum (400 MHz, CDCl₃) of PLA obtained by [D-LA]/[(R,S)-2]/[I] = 50/2/1, solvent = DCM $(P_m = 0.99)$ References S84

Experimental Details

Reagents, Instruments, and Methods

All synthesis and manipulations of air- and moisture-sensitive materials were carried out in flamed Schlenk-type glassware on a dual-manifold Schlenk line, on a high-vacuum line, or in an inert gas Ar-filled glovebox. High-performance liquid chromatography (HPLC)-grade anhydrous tetrahydrofuran (THF), toluene (TOL), chloroform (CHCl₃), and dichloromethane (DCM) were dried via a Vigor YJC-5 solvent purification system and stored over activated Sigma-Aldrich 4 Å molecular sieves in glovebox. Other commercial reagents were purchased from Energy Chemical and used as received without further purification.

L-Lactide (L-LA), *D*-lactide(D-LA), *rac*-lactide (*rac*-LA) were purchased from Energy Chemical and further purified by recrystallizations from dry toluene and double sublimation under vacuum. The initiators *D*-methyl lactate, *L*-methyl lactate, and benzyl alcohol (BnOH) was purchased from Energy Chemical and used as received without further purification. The initiator 4-methylbenzyl alcohol was purchased from Adamas and purified via sublimation at 55 ℃ under vacuum. Other commercial reagents were purchased from Energy Chemical and used as received.

NMR Spectroscopy

 1 H and 13 C NMR spectra were recorded on an Agilent 400-MR DD2 or a Bruker AV II-400 MHz spectrometer (1 H: 400 MHz, 13 C: 100 MHz). Chemical shifts (δ) for 1 H and 13 C NMR spectra are given in ppm relative to SiMe4. 1 H NMR chemical shifts were referenced as follows: δ 7.26 ppm for chloroform-d (CDCl₃), 13 C NMR chemical shifts were referenced as follows: δ 77.16 ppm for chloroform-d (CDCl₃). The following abbreviations were used to explain the multiplicities: s = singlet, d = doublet, t = triplet, q = quartet, m = multiple. Tacticity of polymers was determined from the methine region of the homodecoupled 1 H NMR spectrum. $P_{\rm m}$, the probability of forming a new isotactic dyad, was calculated utilizing methods established in the literature. 1

High-resolution mass spectra (HRMS)

Analyses were performed on LCMS-IT-TOF (Liquid Chromatograph Mass Spectrometer, shimadzu) that combines QIT (ion trap) and TOF (time-of-flight) technologies.

X-Ray single-crystal diffraction

Data were collected on an Agilent Technologies Gemini plus single crystal diffraction.

Size exclusion chromatography (SEC)

Measurements of polymer number-average molecular weight (M_n) and molecular weight

distributions or polydispersity indices ($D = M_w/M_n$) were performed via SEC. The SEC instrument consisted of an Agilent LC system equipped with one guard column and two PL gel 5 µm mixed-C gel permeation columns and coupled with an Agilent G7162A 1260 Infinity II RI detector; The analysis was performed at 40 °C using THF as the eluent at a flow rate of 1.0 mL/min. The instrument was calibrated with 9 PS standards, and chromatograms were processed with Agilent OpenLab CDS Acquisition 2.5 molecular weight characterization software.

High Performance Liquid Chromatography (HPLC)

HPLC was performed on Agilent 1260 Infinity II Quaternary LC system. Enantiomeric excesses (ee) were determined by chiral HPLC analysis on Daicel chiralpak IA columns. The column employed and the respective solvent mixture are indicated for each experiment. The chromatograms were processed with Agilent OpenLab CDS software.

Differential scanning calorimetry (DSC)

Melting-transition temperature ($T_{\rm m}$) and glass-transition temperature ($T_{\rm g}$) of purified and thoroughly dried polymer samples were measured by differential scanning calorimetry (DSC) on DSC25, TA Instrument. All $T_{\rm m}$, $T_{\rm c}$ and $T_{\rm g}$ values were obtained from a second scan after the thermal history was removed from the first scan. The second heating rate was 10 °C/min and cooling rate was 10 °C/min.

Synthesis of catalysts

Scheme S1. Synthesis of catalysts.

1. Synthesis of (R)-BINAM-Ac-NCS

Literature procedures were modified for the preparation of (R)-BINAM-Ac-NCS.²

To a solution of (*R*)-(+)-1,1'-binaphthyl-2,2'-diamine (2.84 g, 10 mmol) and AcOH (6 mL, 100 mmol) in 70 mL of dried DCM was added acetic anhydride (1.0 mL, 10 mmol) at 0 °C under Ar. The resulting solution was stirred for overnight at room temperature, and 3 M NaOH aqueous solution (about 36 mL) was added to adjust the solution to pH = 10. The reaction mixture was extracted with DCM (3 × 20 mL) and the combined organic phases were washed with saturated brine (70 mL) and dried over anhydrous Na₂SO₄. The solution was filtered and concentrated under reduced pressure. The residue was purified by column chromatography on silica gel (PE/EA = 4/1) to give product (*R*)-BINAM-Ac (2.25 g, 69% yield) as a white solid. ¹H NMR (400 MHz, CDCl₃): δ 8.62 (d, J = 9.0 Hz, 1H), 8.00 (d, J = 9.0 Hz, 1H), 7.91 (d, J = 8.2 Hz, 1H), 7.86 (d, J = 8.9 Hz, 1H), 7.82 (d, J = 7.9 Hz, 1H), 7.43-7.39 (m, 1H), 7.29-7.14 (m, 3H), 7.04 (s, 1H), 6.92 (d, J = 8.3 Hz, 1H), 3.59 (s, 2H), 1.86 (s, 3H). The ¹H NMR spectrum was consistent with literature report.²

(R)-BINAM-Ac (684 mg, 2.1 mmol) was dissolved in DCM (12 mL) and sat. aq. NaHCO₃ (12 mL) was added. The resulting biphasic solution was cooled to 0 °C and thiophosgene (0.2 mL, 2.6 mmol) was then slowly added. The reaction mixture was allowed to warm to room temperature and stir for 12 h. The layers were separated, and the aqueous layer was extracted with DCM ($3 \times 10 \text{ mL}$). The organic layers were collected, washed with brine (30 mL), dried over anhydrous Na₂SO₄. The solution was filtered and concentrated under reduced pressure to give pure product (R)-BINAM-Ac-NCS (667 mg, 86% yield) as a pale-yellow solid, which was used directly in the subsequent reaction without further purification. ^{1}H NMR (400 MHz, CDCl₃): $\delta 8.56 \text{ (d}$, J = 9.1 mes

Hz, 1H), 8.07-7.93 (m, 4H), 7.56-7.35 (m, 4H), 7.30-7.22 (m, 2H), 6.97 (d, J = 8.5 Hz, 1H), 6.64 (s, 1H), 1.83 (s, 3H). The ¹H NMR spectrum was consistent with literature report.²

2. Synthesis of (S)-BINAM-Ac-NCS

Literature procedures were modified for the preparation of (S)-BINAM-Ac-NCS.²

To a solution of (*S*)-(-)-1,1'-binaphthyl-2,2'-diamine (2.84 g, 10 mmol) and AcOH (6 mL, 100 mmol) in 70 mL of dried DCM was added acetic anhydride (1.0 mL, 10 mmol) at 0 °C under Ar. The resulting solution was stirred for overnight at room temperature, and 3 M NaOH aqueous solution (about 36 mL) was added to adjust the solution to pH = 10. The reaction mixture was extracted with DCM (3 × 20 mL) and the combined organic phases were washed with saturated brine (70 mL) and dried over anhydrous Na₂SO₄. The solution was filtered and concentrated under reduced pressure. The residue was purified by column chromatography on silica gel (PE/EA = 4/1) to give product (*S*)-BINAM-Ac (2.58 g, 79% yield) as a white solid. ¹H NMR (400 MHz, CDCl₃): δ 8.63 (d, J = 9.0 Hz, 1H), 8.00 (d, J = 9.0 Hz, 1H), 7.91 (d, J = 8.2 Hz, 1H), 7.86 (d, J = 8.8 Hz, 1H), 7.82 (d, J = 8.0 Hz, 1H), 7.44-7.40 (m, 1H), 7.28-7.14 (m, 3H), 7.04 (s, 1H), 6.92 (d, J = 8.3 Hz, 1H), 3.66 (s, 2H), 1.85 (s, 3H). The ¹H NMR spectrum was consistent with literature report.²

(*S*)-BINAM-Ac (684 mg, 2.1 mmol) was dissolved in DCM (12 mL) and sat. aq. NaHCO₃ (12 mL) was added. The resulting biphasic solution was cooled to 0 °C and thiophosgene (0.2 mL, 2.6 mmol) was then slowly added. The reaction mixture was allowed to warm to room temperature and stir for 12 h. The layers were separated, and the aqueous layer was extracted with DCM (3 × 10 mL). The organic layers were collected, washed with brine (30 mL), dried over anhydrous Na₂SO₄. The solution was filtered and concentrated under reduced pressure to give pure product BINAM-Ac-NCS (741 mg, 96% yield) as a pale-yellow solid, which was used directly in the subsequent reaction without further purification. ¹H NMR (400 MHz, CDCl₃): δ 8.55 (d, J = 9.2 Hz, 1H), 8.07-7.93 (m, 4H), 7.56-7.35 (m, 4H), 7.30-7.22 (m, 2H), 6.97 (d, J = 8.5 Hz, 1H), 6.65 (s, 1H), 1.82 (s, 3H). The ¹H NMR spectrum was consistent with literature report.²

3. Synthesis of (R)-BINAM-TFA-NCS

Literature procedures were modified for the preparation of (R)-BINAM-TFA-NCS.²

To a solution of (R)-(+)-1,1'-binaphthyl-2,2'-diamine (2.84 g, 10 mmol) and trifluoroacetic acid (TFA) (8.5 mL) in 60 mL of dried DCM was added trifluoroacetic anhydride (1.41 mL, 10 mmol) at 0 °C under Ar. The resulting solution was stirred for overnight at room temperature, and 3 M NaOH aqueous solution (about 36 mL) was added to adjust the solution to pH = 10. The reaction mixture was extracted with DCM (3 × 20 mL) and the combined organic phases were washed with saturated brine (70 mL) and dried over anhydrous Na₂SO₄. The solution was filtered and concentrated under reduced pressure. The residue was purified by column chromatography on silica gel (PE/EA = 8/1) to give product (R)-BINAM-TFA (3.27 g, 86% yield) as a white solid. 1 H NMR (400 MHz, CDCl₃): δ 8.56 (d, J = 9.1 Hz, 1H), 8.07 (d, J = 9.0 Hz, 1H), 7.97 (d, J = 8.6 Hz, 1H), 7.93 (s, 1H), 7.89 (d, J = 8.8 Hz, 1H), 7.83 (d, J = 8.1 Hz, 1H), 7.52-7.48 (m, 1H), 7.36-7.14 (m, 4H), 7.16 (d, J = 8.8 Hz, 1H), 6.87-6.84 (m, 1H), 3.68 (s, 2H). The 1 H NMR spectrum was consistent with literature report.²

(*R*)-BINAM-TFA (1.75 g, 4.6 mmol) was dissolved in DCM (21 mL) and sat. aq. NaHCO₃ (21 mL) was added. The resulting biphasic solution was cooled to 0 °C and thiophosgene (0.45 mL, 5.6 mmol) was then slowly added. The reaction mixture was allowed to warm to room temperature and stir for 12 h. The layers were separated, and the aqueous layer was extracted with DCM (3 × 20 mL). The organic layers were collected, washed with brine (60 mL), dried over anhydrous Na₂SO₄. The solution was filtered and concentrated under reduced pressure to give pure product (*R*)-BINAM-TFA-NCS (1.67 g, 86% yield) as a pale-yellow solid, which was used directly in the subsequent reaction without further purification. ¹H NMR (400 MHz, CDCl₃): δ 8.49 (d, *J* = 9.0 Hz, 1H), 8.13 (d, *J* = 9.3 Hz, 1H), 8.06 (d, *J* = 8.7 Hz, 1H), 7.99 (dd, *J* = 8.2, 3.4 Hz, 2H), 7.61-7.48 (m, 3H), 7.44-7.32 (m, 3H), 7.17 (d, *J* = 8.6 Hz, 1H), 7.11 (d, *J* = 8.5 Hz, 1H). The ¹H NMR spectrum was consistent with literature report.²

4. Synthesis of (*R*)-H₈-BINAM-Ac-NCS

(R)-H₈-BINAM-Ac-NCS

Literature procedures were modified for the preparation of (R)-H₈-BINAM-Ac-NCS.³

To a stirred mixture of (R)-(+)-1,1'-binaphthyl-2,2'-diamine (284 mg, 1.0 mmol) and Raney Ni-Al alloy (2.00 g) in isopropanol (100 mL) and water (100 mL) was slowly added 1.0 wt% aqueous NaOH solution (200 mL) over 2 h at 90 °C. After stirring for 30 h, the reaction mixture was cooled to room temperature. The mixture was then filtered through Celite, and the filter cake was washed with ethyl acetate (EA). The filtrates were concentrated in vacuo to remove isopropanol and then exacted with DCM, dried over Na₂SO₄, and concentrated under reduced pressure. The residue was purified by column chromatography on silica gel (PE/EA = 7/1) to give product (R)-H₈-BINAM (243 mg, 83% yield) as a white solid. ¹H NMR (400 MHz, CDCl₃): δ 6.92 (d, J = 8.1 Hz, 2H), 6.62 (d, J = 8.1 Hz, 2H), 2.90 (s, 4 H), 2.79-2.65 (m, 4 H), 2.33-2.12 (m, 4H), 1.78-1.61 (m, 8H).

To a solution of (*R*)-H₈-BINAM (1.21 g, 4.1 mmol) and AcOH (2.4 mL, 41.4 mmol) in 30 mL of dried DCM was added acetic anhydride (0.39 mL, 4.1 mmol) at 0 °C under Ar. The resulting solution was stirred for overnight at room temperature for 24 h, and 3 M NaOH aqueous solution was added to adjust the solution to pH = 10. The reaction mixture was extracted with DCM (3 × 20 mL) and the combined organic phases were washed with saturated brine (70 mL) and dried over anhydrous Na₂SO₄. The solution was filtered and concentrated under reduced pressure. The residue was purified by column chromatography on silica gel (PE/EA = 4/1) to give product (*R*)-H₈-BINAM-Ac (0.98 g, 71% yield) as a pale-yellow solid. ¹H NMR (400 MHz, CDCl₃): δ 8.10 (d, J = 8.4 Hz, 1H), 7.11 (d, J = 8.4 Hz, 1H), 6.96 (d, J = 8.1 Hz, 1H), 6.76 (s, 1H), 6.63 (d, J = 8.2 Hz, 1H), 3.32 (s, 2H), 2.78 (t, J = 6.2 Hz, 2H), 2.72 (t, J = 6.3 Hz, 2H), 2.35-2.12 (m, 2H), 2.09 (t, J = 6.3 Hz, 2H), 1.91 (s, 3H), 1.82-1.56 (m, 8H).

(*R*)-H₈-BINAM-Ac (983 mg, 2.9 mmol) was dissolved in DCM (15 mL) and sat. aq. NaHCO₃ (15 mL) was added. The resulting biphasic solution was cooled to 0 °C and thiophosgene (0.2 mL, 2.6 mmol) was then slowly added. The reaction mixture was allowed to warm to room temperature and stir for 11 h. The layers were separated, and the aqueous layer was extracted with DCM (3 × 10 mL). The organic layers were collected, washed with brine (30 mL), dried over anhydrous Na₂SO₄. The solution was filtered and concentrated under reduced pressure to give pure product (*R*)-H₈-BINAM-Ac-NCS (977 mg, 88% yield) as a pale-yellow solid, which was used directly in the subsequent reaction without further purification. ¹H NMR (400 MHz, CDCl₃): δ 7.99 (d, J = 8.4 Hz, 1H), 7.16 (d, J = 8.4 Hz, 1H), 7.14-7.04 (m, 2H), 6.37 (s, 1H), 3.09-2.62 (m, 4H), 2.33-1.99 (m, 4H), 1.91 (s, 3H),1.82-1.63 (m, 8H).

5. Synthesis of (*R*)-DACH-dimethyl

Literature procedures were modified for the preparation of (R)-DACH-dimethyl.^{4,5}

(1R, 2R)-(-)-1,2-Diaminocyclohexane (9.20 g, 80.6 mmol) was dissolved in 60 mL methanol and the solution was cooled to 0 °C. 37% HCl (6.70 mL, 80.6 mmol) was dissolved in 30 mL methanol and added dropwise to the diamine at 0 °C. The reaction mixture was allowed to warm to room temperature and further stirred at RT for 1 h. Di-*tert*-butyl dicarbonate (17.59 g, 80.6 mmol) was dissolved in 30 mL methanol and added dropwise to the reaction mixture at RT over 40 min. The solution was stirred at RT for 3 h. The solvent was removed under reduced pressure, and the resultant yellow solid was washed with diethyl ether (3 × 50 mL), yielding a white solid. The remaining residue was dissolved in 100 mL chloroform and treated with 40 mL 3.0 M NaOH. The collected organic layer was washed with brine and dried over Na₂SO₄. The solvent was removed under reduced pressure to obtain (*R*)-DACH-Boc (15.58 g, 90% yield) as an off-white solid which was used directly for the next step without further purification. ¹H NMR (400 MHz, CDCl₃): δ 4.45 (s, 1H), 3.12 (d, J = 10.8 Hz, 1H), 2.30 (td, J = 10.3 Hz, 4.0 Hz, 1H), 2.08-1.88 (m, 2H), 1.77-1.63 (m, 2H), 1.44 (s, 9H), 1.36-1.01 (m, 6H).

To a solution of (R)-DACH-Boc (2.14 g, 10.0 mmol) in DCM (100 mL) was added aq. HCHO

(37%, 1.5 mL, 20 mmol) and the mixture was stirred at RT for 15 min. NaBH(OAc)₃ (4.88 g, 23 mmol) was added and stirring was continued for 6 h at RT. Saturated NaHCO₃ (300 mL) was added and stirring was continued for 15 min. The layers were separated, and the aqueous phase was extracted with DCM (3 × 40 mL). The combined organic layer was dried over Na₂SO₄. The solution was filtered and concentrated under reduced pressure. The residue was purified by column chromatography on silica gel (EA/MeOH = 8/1) to give product (R)-DACH-Boc-dimethyl (2.21 g, 91% yield) as a pale-yellow oil. ¹H NMR (400 MHz, CDCl₃): δ 5.30 (s, 1H), 3.32-3.11 (m, 1H), 2.44 (d, J = 12.7 Hz, 1H), 2.22 (s, 6H), 1.98 (s, 1H), 1.84-1.74 (m, 2H), 1.69-1.60 (m, 1H), 1.44 (s, 9H), 1.29-0.98 (m, 4H).

To a solution of (*R*)-DACH-Boc-dimethyl (2.21 g, 9.1 mmol) in 100 mL DCM was added 25 mL trifluoroacetic acid. The reaction mixture was stirred at RT for 12 h. Solvent was removed under reduced pressure, 3.0 M NaOH was added until pH = 10 and the mixture was extracted with DCM (3 × 40 mL). The combined organic phases were washed with brine dried over Na₂SO₄ and concentrated under reduced pressure and the residue was purified by flash column chromatography on silica gel (EA/MeOH = 4/1) to give product (*R*)-DACH-dimethyl (0.39 g, 30% yield) as a yellow oil. ¹H NMR (400 MHz, CDCl₃): δ 2.55 (td, J = 10.2, 4.1 Hz, 1H), 2.21 (s, 6H), 2.05-1.97 (m, 2H), 1.96 (s, 2H), 1.94-1.89 (m, 1H), 1.81-1.60 (m, 3H), 1.26-1.00 (m, 4H). The ¹H NMR spectrum was consistent with literature report.⁵

6. Synthesis of (S)-DACH-dimethyl

Literature procedures were modified for the preparation of (S)-DACH-dimethyl.^{4, 6, 7}

(1*S*, 2*S*)-(+)-1,2-Diaminocyclohexane (9.00 g, 78.8 mmol) was dissolved in 60 mL methanol and the solution was cooled to 0 °C. 37% HCl (6.57 mL, 78.8 mmol) was dissolved in 30 mL methanol and added dropwise to the diamine at 0 °C. The reaction mixture was allowed to warm to room temperature and further stirred at RT for 1 h. Di-*tert*-butyl dicarbonate (17.19 g, 78.7 mmol) was dissolved in 30 mL methanol and added dropwise to the reaction mixture at RT over 40 mins. The solution was stirred at RT for 3 h. The solvent was removed under reduced pressure,

and the resultant yellow solid was washed with diethyl ether (3 \times 50 mL), yielding a white solid. The remaining residue was dissolved in 100 mL chloroform and treated with 40 mL 3.0 M NaOH. The collected organic layer was washed with brine and dried over Na₂SO₄. The solvent was removed under reduced pressure to obtain (*S*)-DACH-Boc (14.57 g, 86% yield) as an off-white solid which was used directly for the next step without further purification. ¹H NMR (400 MHz, CDCl₃): δ 4.44 (s, 1H), 3.12 (d, J = 12.2 Hz, 1H), 2.31 (td, J = 10.3, 4.0 Hz, 1H), 2.04-1.90 (m, 2H), 1.76-1.64 (m, 2H), 1.44 (s, 9H), 1.36-1.01 (m, 6H).

To a solution of (*S*)-DACH-Boc (2.14 g, 10.0 mmol) in DCM (100 mL) was added aq. HCHO (37%, 1.5 mL, 20 mmol) and the mixture was stirred at RT for 15 min. NaBH(OAc)₃ (4.88 g, 23 mmol) was added and stirring was continued for 6 h at RT. Saturated NaHCO₃ (300 mL) was added and stirring was continued for 15 min. The layers were separated, and the aqueous phase was extracted with DCM (3 × 40 mL). The combined organic layer was dried over Na₂SO₄. The solution was filtered and concentrated under reduced pressure. The residue was purified by column chromatography on silica gel (EA/MeOH = 8/1) to give product (*S*)-DACH-Boc-dimethyl (2.00 g, 83% yield) as a pale-yellow oil. ¹H NMR (400 MHz, CDCl₃): δ 5.25 (s, 1H), 3.33-3.06 (m, 1H), 2.45 (d, J = 12.7 Hz, 1H), 2.20 (s, 6H), 2.20-2.13 (m, 1H), 1.87-1.73 (m, 2H), 1.69-1.56 (m, 1H), 1.44 (s, 9H), 1.33-0.90 (m, 4H).

To a solution of (*S*)-DACH-Boc-dimethyl (2.00 g, 8.3 mmol) in 90 mL DCM was added 23 mL trifluoroacetic acid. The reaction mixture was stirred at RT for 12 h. Solvent was removed under reduced pressure, 3.0 M NaOH was added until pH = 10 and the mixture was extracted with DCM (3 × 40 mL). The combined organic phases were washed with brine dried over Na₂SO₄ and concentrated under reduced pressure and the residue was purified by flash column chromatography on silica gel (EA/MeOH = 4/1) to give product (*S*)-DACH-dimethyl (0.87 g, 74% yield) as a yellow oil. ¹H NMR (400 MHz, CDCl₃): δ 2.54 (td, J = 10.2, 4.1 Hz, 1H), 2.20 (s, 6H), 2.09-1.95 (m, 2H), 1.94 (s, 2H), 1.92-1.86 (m, 1H), 1.81-1.59 (m, 3H), 1.28-0.96 (m, 4H). The ¹H NMR spectrum was consistent with literature report.⁷

7. Synthesis of (S)-DACH-pyrrolidine

$$(S)$$
 NHBoc + Br (S) Br (S) $(S$

Literature procedures were modified for the preparation of (S)-DACH-pyrrolidine.⁸ (S)-DACH-Boc (4.28 g, 20 mmol) was dissolved in MeCN (40 mL), and K₂CO₃ (14.10 g,

102 mmol) and 1,4-dibromobutane (6.45 g, 30 mmol) were added. The suspension was stirred at RT for 24 h then at 80 °C for 22 h, and cooled to room temperature. The mixture was filtered, and the solid was washed with MeCN, and the combined filtrates were concentrated in vacuo. The residue was suspended in 6.5 M aqueous HCl (22 mL) with vigorous stirring while the evolution of gas was observed. After 12 h, the mixture was washed with diethyl ether (2 × 20 mL) to remove the excess of 1,4-dibromobutane. 3.0 M NaOH was added to the mixture until pH = 10. The aqueous phase was extracted with DCM (3 × 20 mL) and the combined organic phase was dried over Na₂SO₄. The solution was filtered and concentrated under reduced pressure. The residue was purified by column chromatography on silica gel (EA/MeOH = 4/1) to give product (*S*)-DACH-pyrrolidine (2.34 g, 70% yield) as a colorless solid. ¹H NMR (400 MHz, CDCl₃): δ 2.67-2.61 (m, 2H), 2.56-2.50 (m, 2H), 2.29 (td, J = 10.3, 3.2 Hz, 1H), 2.11 (s, 2H), 2.00-1.89 (m, 1H), 1.88-1.54 (m, 8H), 1.35-1.03 (m, 4H). The ¹H NMR spectrum was consistent with literature report. ⁸

8. Synthesis of (S)-DACH-ⁱPr

Literature procedures were modified for the preparation of (S)-DACH-Pr.9

To a solution of (*S*)-DACH-Boc (1.07 g, 5.0 mmol) in DCM (30 mL) was added acetone (0.88 mL, 12 mmol) and the mixture was stirred at RT for 20 min. NaBH(OAc)₃ (2.54 g, 12 mmol) was added and stirring was continued for 7 h at RT. Saturated NaHCO₃ (40 mL) was added and stirring was continued for 15 min. The layers were separated, and the aqueous phase was extracted with DCM (3 × 20 mL). The combined organic layer was dried over Na₂SO₄. The solution was filtered and concentrated under reduced pressure. The residue was added 15 mL DCM and 5 mL trifluoroacetic acid. The reaction mixture was stirred at RT for 11 h. Solvent was removed under reduced pressure, 3.0 M NaOH was added until pH = 10 and the mixture was extracted with DCM (3 × 20 mL). The combined organic phases were washed with brine dried over Na₂SO₄ and concentrated under reduced pressure and the residue was purified by flash column chromatography on silica gel (EA/MeOH = 4/1) to give product (*S*)-DACH-ⁱPr (0.52 g, 52% yield) as a pale-yellow oil. ¹H NMR (400 MHz, CDCl₃): δ 2.96-2.83 (m, J = 6.2 Hz, 1H), 2.32-2.21 (m, 1H), 2.08-1.97 (m, 2H), 1.93-1.82 (m, 1H), 1.72-1.62 (m, 2H), 1.51 (s, 3H), 1.28-1.17 (m, 2H), 1.14-1.07 (m, 1H), 1.04 (d, *J* = 6.3 Hz, 3H), 0.99 (d, *J* = 6.1 Hz, 3H), 0.95-0.81 (m, 1H). The ¹H NMR spectrum was consistent with literature report.⁹

9. Synthesis of (S)-DACH-ethanol-methyl

Br OH
$$\frac{Ac_2O, DMAP}{DCM}$$
 Br OAC + $\frac{K_2CO_3}{NHBoc}$ $\frac{K_2CO_3}{MeCN}$ $\frac{K_2CO_3}{NHBoc}$ $\frac{K_2CO_$

Literature procedures were modified for the preparation of (S)-DACH-ethanol-methyl. 10

To a solution of 2-bromoethanol (4.00 g, 32.0 mmol) and 4-(N,N-dimethylamino)pyridine (DMAP) (5.20 g, 42.3 mmol) in 40 mL of dried DCM was added acetic anhydride (4.0 mL, 42.3 mmol) at 0 °C under Ar. The reaction mixture was then stirred for 23 h at room temperature and 30 mL H₂O was added to quench the reaction. The layers were separated, and aqueous layer was extracted with DCM (2 × 30 mL). The combined organic phases were dried over Na₂SO₄ and concentrated under reduced pressure to give product 2-bromoethyl acetate (4.71 g, 88% yield) as a yellow oil. 1 H NMR (400 MHz, CDCl₃): δ 4.37 (t, J = 6.1 Hz, 2H), 3.50 (t, J = 6.1 Hz, 2H), 2.09 (s, 3H).

(S)-DACH-Boc (1.71 g, 8 mmol) was dissolved in MeCN (30 mL), and K₂CO₃ (5.53 g, 40 mmol) and 2-bromoethyl acetate (1.34 g, 8 mmol) were added. The suspension was stirred at 80 °C for 24 h and then cooled to room temperature. The mixture was filtered, and the solid was washed with MeCN, and the combined filtrates were concentrated under reduced pressure. The residue was purified by column chromatography on silica gel (EA/MeOH= 10/1) to give product (S)-DACH-Boc-EA (1.36 g, 57% yield) as a pale-yellow oil.

To a solution of (S)-DACH-Boc-EA (2.12 g, 7.1 mmol) in DCM (20 mL) was added aq. HCHO (37%, 0.63 mL, 8.5 mmol) and the mixture was stirred at RT for 20 min. NaBH(OAc)₃ (1.94 g, 9.2 mmol) was added and stirring was continued for 11 h at RT. Saturated NaHCO₃ (30 mL) was added and stirring was continued for 15 min. The layers were separated, and the aqueous phase was extracted with DCM (3×15 mL). The combined organic layer was dried over Na₂SO₄. The solution was filtered and concentrated under reduced pressure. The residue was purified by column chromatography on silica gel (PE/EA = 1/12) to give product (S)-DACH-Boc-EA-methyl (1.06 g, 48% yield) as a pale-yellow oil.

The residue was added 15 mL 4 N HCl and the mixture was stirred at 80 °C for 12 h. After being cooling to ambient temperature, 3.0 M NaOH was added until pH = 10. After extraction with DCM, the combined organic layers were dried over Na₂SO₄, concentrated under reduced pressure

to obtain (*S*)-DACH-ethanol-methyl (0.55 g, 95% yield) as a pale-yellow oil, which was used directly in the subsequent reaction without further purification. ¹H NMR (400 MHz, CDCl₃): δ 3.66-3.50 (m, 2H), 2.76-2.67 (m, 1H), 2.63 (td, J = 10.2, 4.1 Hz, 1H), 2.52-2.41 (m, 1H), 2.27 (s, 5H), 2.23-2.07 (m, 1H), 1.95-1.86 (m, 1H), 1.85-1.72 (m, 2H), 1.70-1.62 (m, 1H), 1.29-0.98 (m, 5H). The ¹H NMR spectrum was consistent with literature report. ¹⁰

10. Synthesis of (S)-DACH-diethyl

Literature procedures were modified for the preparation of (S)-DACH-diethyl.¹¹

To a solution of (S)-DACH-Boc (0.64 g, 3.0 mmol) in DCM (15 mL) was added acetaldehyde (0.32 g, 7.2 mmol) and the mixture was stirred at RT for 15 min. NaBH(OAc)₃ (1.53 g, 7.2 mmol) was added and stirring was continued for 6 h at RT. Saturated NaHCO₃ (40 mL) was added and stirring was continued for 15 min. The layers were separated, and the aqueous phase was extracted with DCM (3 × 15 mL). The combined organic layer was dried over Na₂SO₄. The solution was filtered and concentrated under reduced pressure to obtain a pale-yellow solid, which was used directly in the subsequent reaction without further purification

To a solution of (S)-DACH-Boc-diethyl in 15 mL DCM was added 5 mL trifluoroacetic acid. The reaction mixture was stirred at RT for 11 h. Solvent was removed under reduced pressure, 3.0 M NaOH was added until pH = 10 and the mixture was extracted with DCM (3 × 15 mL). The combined organic phases were washed with brine dried over Na₂SO₄ and concentrated under reduced pressure and the residue was purified by flash column chromatography on silica gel (EA/MeOH = 8/1) to give product (S)-DACH-diethyl (0.37 g, 73% yield) as a dark red oil. 1 H NMR (400 MHz, CDCl₃): δ 2.64-2.49 (m, 3H), 2.38-2.25 (m, 2H), 2.15-2.06 (m, 1H), 2.00-1.96 (m, 3H), 1.79-1.58 (m, 3H), 1.23-1.04 (m, 4H), 1.01 (t, J = 7.1 Hz, 6H). The 1 H NMR spectrum was consistent with literature report. 11

11. Synthesis of catalyst (R,S)-1

$$(R)$$
-BINAM-Ac-NCS (S)-DACH-dimethyl (R,S)-1

To a solution of (*R*)-BINAM-Ac-NCS (1.11 g, 3.0 mmol) in THF (40 mL) was added (*S*)-DACH-dimethyl (0.43 g, 3.0 mmol) and the reaction mixture was stirred for 8 h at room temperature. The reaction mixture was concentrated, and the crude product was purified by flash chromatograph on silica gel (DCM/MeOH/NH₃·H₂O = 100/3/1) to give catalyst (*R*,*S*)-1 (1.29 g, 84% yield) as a pale yellow solid. ¹H NMR (400 MHz, CDCl₃) δ 8.36 (d, J = 9.0 Hz, 1H), 8.07 (s, 1H), 8.04 (d, J = 8.8 Hz, 1H), 7.96 (d, J = 3.8 Hz, 1H), 7.94 (d, J = 2.8 Hz, 1H), 7.89 (d, J = 8.2 Hz, 1H), 7.82 (d, J = 8.7 Hz, 1H), 7.53-7.44 (m, 1H), 7.39 (t, J = 7.5 Hz, 1H), 7.30-7.24 (m, 1H), 7.24-7.10 (m, 2H), 6.98 (d, J = 8.5 Hz, 1H), 6.13 (s, 1H), 2.12 (t, J = 11.1 Hz, 1H), 1.92 (s, 6H), 1.82 (s, 3H), 1.66 (d, J = 11.6 Hz, 2H), 1.48 (s, 1H), 1.10-0.72 (m, 4H). ¹³C NMR (100 MHz, CDCl₃) δ 183.07, 169.49, 136.86, 135.60, 132.99, 132.95, 132.60, 131.08, 129.24, 128.79, 128.34, 128.21, 127.50, 127.14, 126.85, 126.50, 126.10, 125.35, 125.18, 123.02, 67.23, 56.95, 40.29, 32.77, 24.73, 24.43, 24.34, 22.64. HRMS (positive ESI) calculated m/z for C₃₁H₃₅N₄OS⁺ ([M+H]⁺): 511.2532; found m/z: 511.2525.

12. Synthesis of catalyst (R,R)-1

$$(R)$$
-BINAM-Ac-NCS (R) -DACH-dimethyl (R,R) -1

To a solution of (*R*)-BINAM-Ac-NCS (368 mg, 1.0 mmol) in THF (25 mL) was added (*R*)-DACH-dimethyl (171 mg, 1.2 mmol) and the reaction mixture was stirred for 8 h at room temperature. The reaction mixture was concentrated, and the crude product was purified by flash chromatograph on silica gel (DCM/MeOH/NH₃·H₂O = 100/3/1) to give catalyst (*R*,*R*)-1 (449 mg, 88% yield) as a pale yellow solid. ¹H NMR (400 MHz, CDCl₃) δ 11.60 (s, 1H), 8.77 (s, 1H), 8.38 (d, *J* = 9.0 Hz, 1H), 7.98 (d, *J* = 8.8 Hz, 1H), 7.93 (d, *J* = 9.1 Hz, 1H), 7.89 (t, *J* = 8.8 Hz, 2H), 7.67 (d, *J* = 8.8 Hz, 1H), 7.43 (t, *J* = 7.5 Hz, 1H), 7.37 (t, *J* = 7.5 Hz, 1H), 7.18 (t, *J* = 7.7 Hz, 2H), 6.96 (t, *J* = 8.4 Hz, 2H), 6.01 (s, 1H), 3.30 (s, 1H), 2.20-2.08 (m, 1H), 2.05-1.94 (m, 1H), 1.90 (s, 3H), 1.72-1.55 (m, 3H), 1.26 (s, 6H), 1.17-1.06 (m, 2H), 1.04-0.92 (m, 2H). ¹³C NMR (100 MHz,

CDCl₃) δ 170.51, 162.64, 135.96, 133.29, 133.11, 132.37, 131.27, 129.13, 128.78, 128.31, 128.06, 128.00, 127.66, 127.00, 126.71, 126.24, 126.21, 125.74, 125.16, 123.39, 122.81, 68.31, 57.65, 39.57, 36.15, 33.51, 29.82, 24.65, 24.53, 24.34, 22.98. HRMS (positive ESI) calculated m/z for $C_{31}H_{35}N_4OS^+$ ([M+H] $^+$): 511.2532; found m/z: 511.2522.

13. Synthesis of catalyst (S,S)-1

$$(S)$$
-BINAM-Ac-NCS (S) -DACH-dimethyl (S,S) -1

To a solution of (*S*)-BINAM-Ac-NCS (1.00 g, 2.7 mmol) in THF (30 mL) was added (*S*)-DACH-dimethyl (0.43 g, 3.0 mmol) and the reaction mixture was stirred for 8 h at room temperature. The reaction mixture was concentrated, and the crude product was purified by flash chromatograph on silica gel (DCM/MeOH/NH₃·H₂O = 100/3/1) to give catalyst (*S*,*S*)-1 (1.19 g, 86% yield) as a pale yellow solid. ¹H NMR (400 MHz, CDCl₃) δ 11.67 (s, 1H), 8.67 (s, 1H), 8.36 (d, J = 9.0 Hz, 1H), 7.99 (d, J = 8.8 Hz, 1H), 7.94 (d, J = 9.0 Hz, 1H), 7.92-7.86 (m, 2H), 7.68 (d, J = 8.8 Hz, 1H), 7.48-7.42 (m, 1H), 7.37 (t, J = 7.3 Hz, 1H), 7.24-7.15 (m, 2H), 6.99 (d, J = 8.4 Hz, 2H), 6.06 (s, 1H), 3.42 (s, 1H), 2.13 (s, 2H), 1.89 (s, 3H), 1.82-1.58 (m, 4H), 1.51-0.92 (m, 8H). ¹³C NMR (100 MHz, CDCl₃) δ 169.88, 135.93, 133.24, 133.10, 132.40, 131.28, 129.18, 128.82, 128.33, 128.08, 127.52, 127.02, 126.75, 126.27, 126.21, 125.68, 125.17, 123.38, 122.81, 68.19, 57.97, 39.55, 33.45, 29.82, 24.61, 24.51, 24.34, 22.99. HRMS (positive ESI) calculated m/z for C₃₁H₃₅N₄OS⁺ ([M+H]⁺): 511.2532; found m/z: 511.2523.

14. Synthesis of catalyst (S,R)-1

$$(S)$$
-BINAM-Ac-NCS (R)-DACH-dimethyl (S,R)-1

To a solution of (S)-BINAM-Ac-NCS (368.5 mg, 1.0 mmol) in THF (15 mL) was added (R)-DACH-dimethyl (142.2 mg, 1.0 mmol) and the reaction mixture was stirred for 8 h at room temperature. The reaction mixture was concentrated, and the crude product was purified by flash chromatograph on silica gel (DCM/MeOH/NH₃·H₂O = 100/3/1) to give catalyst (S,R)-1 (410.2 mg,

80% yield) as a pale yellow solid. 1 H NMR (400 MHz, CDCl₃) δ 8.36 (d, J = 9.0 Hz, 1H), 8.07 (s, 1H), 8.04 (d, J = 8.8 Hz, 1H), 7.96 (d, J = 3.4 Hz, 1H), 7.94 (d, J = 2.5 Hz, 1H), 7.89 (d, J = 8.1 Hz, 1H), 7.82 (d, J = 8.8 Hz, 1H), 7.53-7.44 (m, 1H), 7.39 (t, J = 7.5 Hz, 1H), 7.30-7.25 (m, 1H), 7.25-7.14 (m, 2H), 6.98 (d, J = 8.4 Hz, 1H), 6.11 (s, 1H), 2.16-2.06 (m, 1H), 1.92 (s, 6H), 1.82 (s, 3H), 1.66 (d, J = 11.8 Hz, 2H), 1.48 (s, 1H), 1.05-0.82 (m, 4H). 13 C NMR (100 MHz, CDCl₃) δ 183.08, 169.48, 136.85, 135.61, 132.97, 132.95, 132.61, 131.09, 129.26, 128.79, 128.35, 128.22, 127.48, 127.16, 126.85, 126.52, 126.11, 125.34, 125.19, 123.05, 67.28, 57.00, 41.04, 32.76, 24.72, 24.42, 24.33, 22.66. HRMS (positive ESI) calculated m/z for C₃₁H₃₅N₄OS⁺ ([M+H]⁺): 511.2532; found m/z: 511.2531.

15. Synthesis of catalyst (*R*,*S*)-2

$$(R)$$
-BINAM-Ac-NCS (S)-DACH-pyrrolidine (R,S)-2

To a solution of (*R*)-BINAM-Ac-NCS (1.19 g, 3.2 mmol) in THF (30 mL) was added (*S*)-DACH-pyrrolidine (0.54 g, 3.2 mmol) and the reaction mixture was stirred for 12 h at room temperature. The reaction mixture was concentrated, and the crude product was purified by flash chromatograph on silica gel (DCM/MeOH/NH₃·H₂O = 100/3/1) to give catalyst (*R*,*S*)-2 (1.65 g, 96% yield) as a pale yellow solid. ¹H NMR (400 MHz, CDCl₃) δ 11.60 (s, 1H), 8.36 (d, J = 9.0 Hz, 1H), 8.06 (d, J = 8.8 Hz, 1H), 8.03 (s, 1H), 7.96 (d, J = 9.0 Hz, 2H), 7.91 (d, J = 8.1 Hz, 1H), 7.77 (d, J = 8.7 Hz, 1H), 7.54-7.45 (m, 1H), 7.41 (t, J = 7.5 Hz, 1H), 7.30 (t, J = 7.5 Hz, 1H), 7.22 (t, J = 7.5 Hz, 1H), 6.97 (d, J = 8.5 Hz, 1H), 5.69 (s, 1H), 3.80-3.69 (m, 3H), 2.54 (s, 2H), 2.09 (s, 3H), 1.87-1.82 (m, 6H), 1.69-1.36 (m, 4H), 1.09-0.75 (s, 4H). ¹³C NMR (100 MHz, CDCl₃) δ 169.43, 136.92, 135.51, 133.20, 132.99, 132.76, 131.10, 129.44, 128.65, 128.30, 128.24, 127.58, 127.18, 126.75, 126.59, 126.20, 125.20, 125.17, 123.18, 68.10, 32.73, 25.75, 24.77, 24.55, 23.70, 23.30. HRMS (positive ESI) calculated m/z for C₃₃H₃₇N₄OS⁺ ([M+H]⁺): 537.2688; found m/z: 537.2685.

16. Synthesis of catalyst (*R*,*S*)-3

$$(R)$$
-BINAM-Ac-NCS (S)-DACH-diethyl (R,S)-3

To a solution of (*R*)-BINAM-Ac-NCS (0.61 g, 1.7 mmol) in THF (15 mL) was added (*S*)-DACH-diethyl (0.37 g, 2.2 mmol) and the reaction mixture was stirred for 15 h at room temperature. The reaction mixture was concentrated, and the crude product was purified by flash chromatograph on silica gel (DCM/MeOH/NH₃·H₂O = 100/3/1) to give catalyst (*R*,*S*)-3 (0.51 g, 57% yield) as a pale yellow solid. ¹H NMR (400 MHz, CDCl₃) δ 8.29 (d, J = 9.0 Hz, 1H), 8.11 (s, 1H), 8.05 (d, J = 8.7 Hz, 1H), 7.95 (d, J = 8.5 Hz, 2H), 7.90 (d, J = 8.2 Hz, 1H), 7.77 (d, J = 8.7 Hz, 1H), 7.49 (t, J = 7.5 Hz, 1H), 7.40 (t, J = 7.5 Hz, 1H), 7.32-7.27 (m, 1H), 7.22 (t, J = 7.3 Hz, 2H), 7.00 (d, J = 8.4 Hz, 1H), 6.16 (s, 1H), 2.50-2.13 (m, 6H), 1.79 (s, 3H), 1.64 (t, J = 15.2 Hz, 3H), 1.48-1.37 (m, 1H), 1.05-0.92 (m, 1H), 0.91-0.65 (m, 9H). ¹³C NMR (100 MHz, CDCl₃) δ 183.08, 169.32, 136.57, 135.65, 133.36, 132.86, 132.65, 131.24, 129.52, 128.83, 128.41, 128.24, 127.29, 127.11, 126.78, 126.59, 126.10, 125.20, 125.06, 123.54, 64.66, 57.02, 42.87, 32.90, 31.72, 25.27, 24.45, 24.36, 22.79, 14.33, 14.25, 13.38. HRMS (positive ESI) calculated m/z for C₃₃H₃₉N₄OS⁺ ([M+H]⁺): 539.2845; found m/z: 539.2841.

17. Synthesis of catalyst (R,S)-4

$$(R)$$
-BINAM-Ac-NCS (S)-DACH- Pr (R,S)-4

To a solution of (*R*)-BINAM-Ac-NCS (0.55 g, 1.5 mmol) in THF (10 mL) was added (*S*)-DACH-^{*i*}Pr (0.36 g, 1.8 mmol) and the reaction mixture was stirred for 14 h at room temperature. The reaction mixture was concentrated, and the crude product was purified by flash chromatograph on silica gel (EA/MeOH = 4/1) to give catalyst (*R*,*S*)-4 (0.45 g, 53% yield) as a pale yellow solid. ¹H NMR (400 MHz, CDCl₃) δ 8.42 (d, *J* = 8.9 Hz, 1H), 8.09 (d, *J* = 8.5 Hz, 1H), 8.00-7.94 (m, 3H), 7.92 (d, *J* = 8.2 Hz, 1H), 7.73 (s, 1H), 7.54-7.48 (m, 1H), 7.44-7.38 (m, 1H), 7.35-7.27 (m, 2H), 7.25-7.20 (m, 1H), 7.02 (d, *J* = 8.5 Hz, 1H), 5.33 (s, 1H), 2.60 (p, *J* = 6.4 Hz, 1H), 1.85 (s, 3H), 1.83-1.74 (m, 1H), 1.68-1.60 (m, 1H), 1.56 (d, *J* = 12.8 Hz, 1H), 1.52-1.36 (m, 3H), 1.01-0.89 (m, 2H), 0.87 (d, *J* = 6.3 Hz, 3H), 0.85-0.79 (m, 1H), 0.78 (d, *J* = 6.2 Hz, 3H), -0.09 (t, *J* =

11.4 Hz, 1H). ¹³C NMR (100 MHz, CDCl₃) δ 184.17, 169.33, 137.20, 135.36, 133.43, 132.78, 132.74, 130.92, 129.50, 129.33, 128.39, 128.26, 128.24, 127.82, 127.08, 126.49, 126.44, 125.98, 125.06, 123.89, 123.34, 62.08, 59.91, 48.28, 33.24, 32.87, 24.75, 24.55, 24.49, 23.31, 22.55. HRMS (positive ESI) calculated m/z for C₃₃H₃₉N₄OS⁺ ([M+H]⁺): 525.2688; found m/z: 525.2684.

18. Synthesis of catalyst (R,S)-5

$$(R)$$
-BINAM-Ac-NCS (S)-DACH-ethanol-methyl (R,S)-5

To a solution of (*R*)-BINAM-Ac-NCS (1.18 g, 3.2 mmol) in THF (30 mL) was added (*S*)-DACH-ethanol-methyl (0.55 g, 3.2 mmol) and the reaction mixture was stirred for 15 h at room temperature. The reaction mixture was concentrated, and the crude product was purified by flash chromatograph on silica gel (DCM/MeOH/NH₃·H₂O = 100/3/1) to give catalyst (*R*,*S*)-5 (1.51 g, 87% yield) as a pale yellow solid. ¹H NMR (400 MHz, CDCl₃) δ 8.31 (d, J = 9.0 Hz, 1H), 8.02 (s, 2H), 7.97-7.91 (m, 2H), 7.89 (d, J = 8.2 Hz, 1H), 7.78 (s, 1H), 7.50-7.44 (m, 1H), 7.43-7.36 (m, 1H), 7.28-7.24 (m, 2H), 7.28-7.24 (m, 2H), 7.24-7.19 (d, J = 8.5 Hz, 1H), 7.06 (d, J = 8.5 Hz, 1H), 7.01 (d, J = 8.5 Hz, 1H), 6.65 (s, 1H), 3.26 (s, 2H), 2.45-2.24 (m, 4H), 2.18 (t, J = 11.2 Hz, 1H), 1.99 (s, 3H), 1.85 (s, 3H), 1.80-1.66 (m, 3H), 1.51 (s, 1H), 1.08 (s, 3H), 0.94-0.78 (m, 1H). ¹³C NMR (100 MHz, CDCl₃) δ 182.15, 169.90, 136.59, 135.33, 132.99, 132.92, 132.30, 131.13, 129.06, 128.96, 128.38, 128.31, 127.12, 126.30, 125.78, 125.42, 125.34, 122.70, 122.57, 66.75, 59.26, 55.76, 55.21, 35.60, 32.85, 25.29, 24.53, 24.49, 23.91, 21.13, 14.23. HRMS (positive ESI) calculated m/z for C₃₂H₃₇N₄O₂S⁺ ([M+H]⁺): 541.2637; found m/z: 541.2630.

19. Synthesis of catalyst (R,S)-6

$$(R)$$
-BINAM-TFA-NCS (S)-DACH-dimethyl (R,S)-6

To a solution of (R)-BINAM-TFA-NCS (0.63 g, 1.5 mmol) in THF (10 mL) was added (S)-DACH-dimethyl (0.28 g, 2.0 mmol) and the reaction mixture was stirred for 18 h at room temperature. The reaction mixture was concentrated, and the crude product was purified by flash

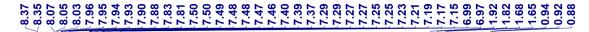
chromatograph on silica gel (DCM/MeOH/NH₃·H₂O = 100/3/1) to give catalyst (R,S)-6 (0.45 g, 53% yield) as a pale yellow solid. ¹H NMR (400 MHz, CDCl₃) δ 10.86 (s, 1H), 9.28 (s, 1H), 8.07 (d, J = 8.7 Hz, 1H), 8.05-8.00 (m, 2H), 7.99-7.91 (m, 2H), 7.60 (d, J = 8.7 Hz, 1H), 7.53-7.46 (m, 2H), 7.36-7.29 (m, 2H), 7.19 (t, J = 9.4 Hz, 2H), 6.07 (s, 1H), 2.16 (s, 1H), 2.10 (s, 6H), 1.69 (t, J = 16.6 Hz, 2H), 1.47 (s, 1H), 1.09-0.66 (m, 4H). ¹³C NMR (100 MHz, CDCl₃) δ 183.50, 155.97, 136.84, 132.74, 132.71, 132.58, 132.51, 132.23, 130.32, 129.17, 128.57, 128.37, 127.61, 127.29, 126.71, 126.44, 126.35, 125.58, 125.34, 123.59, 119.99, 117.12, 114.25, 111.39, 67.64, 56.96, 40.58, 32.62, 24.66, 24.37, 22.87. HRMS (positive ESI) calculated m/z for C₃₁H₃₂F₃N₄OS⁺ ([M+H]⁺): 565.2249; found m/z: 565.2241.

20. Synthesis of catalyst (R,S)-7

$$(R)$$
-H₈-BINAM-Ac-NCS (S)-DACH-dimethyl (R,S)-7

To a solution of (*R*)-H₈-BINAM-Ac-NCS (488.6 mg, 1.3 mmol) in THF (15 mL) was added (*S*)-DACH-dimethyl (227.6 mg, 1.6 mmol) and the reaction mixture was stirred for 6 h at room temperature. The reaction mixture was concentrated, and the crude product was purified by flash chromatograph on silica gel (DCM/MeOH/NH₃·H₂O = 100/3/1) to give catalyst (*R*,*S*)-7 (590.2 mg, 88% yield) as a pale yellow solid. ¹H NMR (400 MHz, CDCl₃) δ 7.97-7.82 (m, 1H), 7.77 (d, *J* = 8.3 Hz, 1H), 7.66-7.45 (m, 1H), 7.35 (d, *J* = 8.2 Hz, 1H), 7.18 (d, *J* = 8.1 Hz, 1H), 7.08 (d, *J* = 8.3 Hz, 1H), 6.45 (s, 1H), 3.21 (s, 1H), 2.92-2.68 (m, 4H), 2.26-2.19 (m, 2H), 2.16 (s, 6H), 2.10 (t, *J* = 6.2 Hz, 3H), 1.90 (s, 3H), 1.83-1.59 (m, 12H), 1.19-1.06 (m, 3H), 1.01-0.88 (m, 1H). ¹³C NMR (100 MHz, CDCl₃) δ 182.28, 168.96, 136.97, 136.88, 134.83, 134.47, 134.39, 134.33, 133.63, 132.27, 129.56, 129.38, 125.41, 121.50, 66.71, 56.55, 40.45, 32.52, 29.92, 29.70, 27.70, 27.53, 27.29, 25.10, 24.69, 24.41, 23.39, 23.10, 23.03, 22.85, 22.53. HRMS (positive ESI) calculated m/z for C₃₁H₄₂N₄OS⁺ ([M+H]⁺): 519.3158; found m/z: 519.3152.

NMR spectra of catalysts



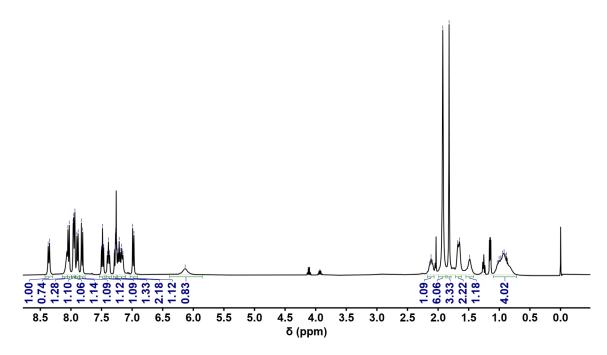


Figure S1. ¹H NMR spectrum (400 MHz, CDCl₃) of (*R*,*S*)-1.

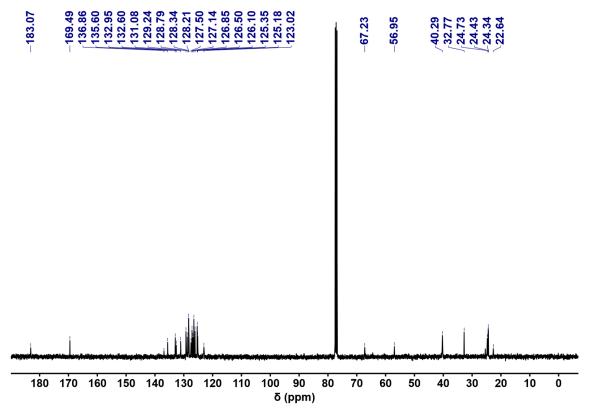


Figure S2. ¹³C NMR spectrum (100 MHz, CDCl₃) of (*R*,*S*)-1.

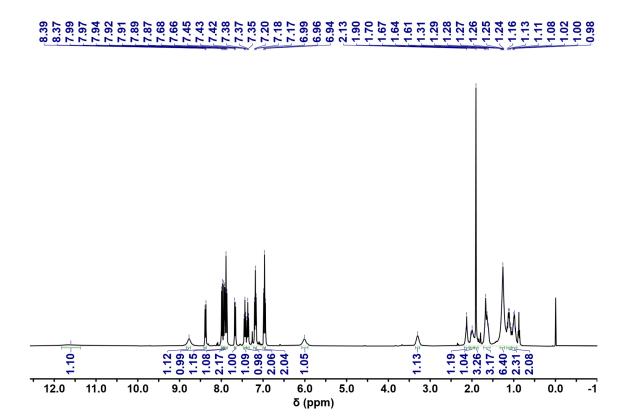


Figure S3. 1 H NMR spectrum (400 MHz, CDCl₃) of (R,R)-1.

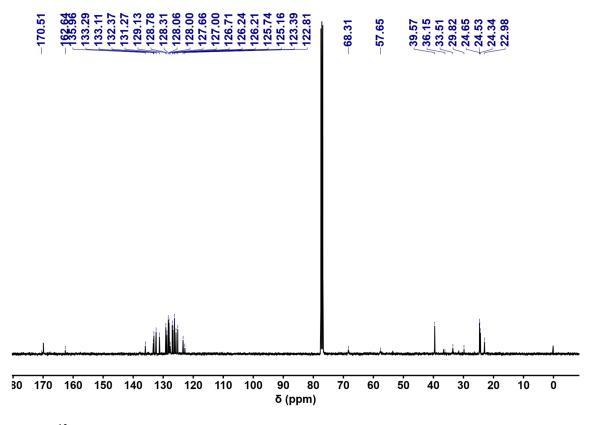


Figure S4. 13 C NMR spectrum (100 MHz, CDCl₃) of (R,R)-1.

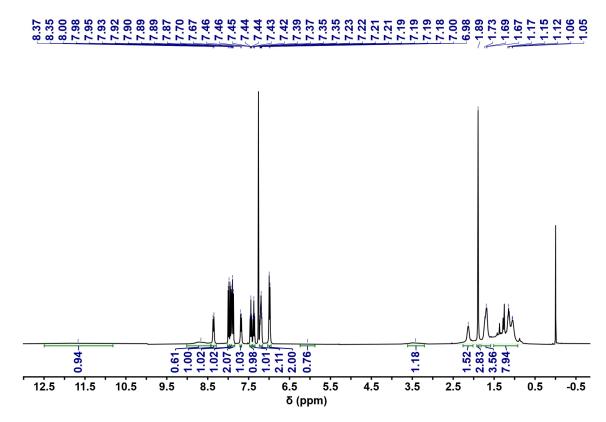


Figure S5. ¹H NMR spectrum (400 MHz, CDCl₃) of (*S*,*S*)-1.

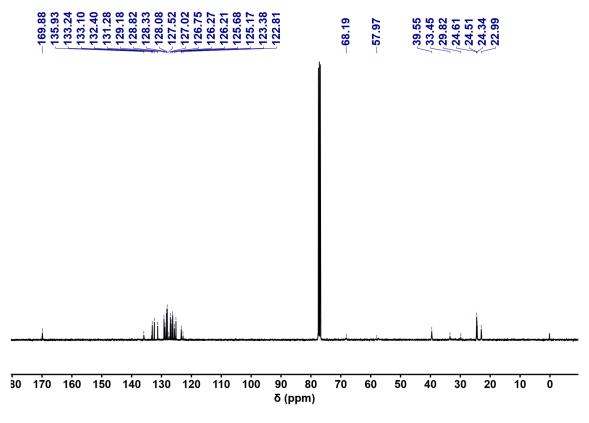


Figure S6. ¹³C NMR spectrum (100 MHz, CDCl₃) of (S,S)-1.

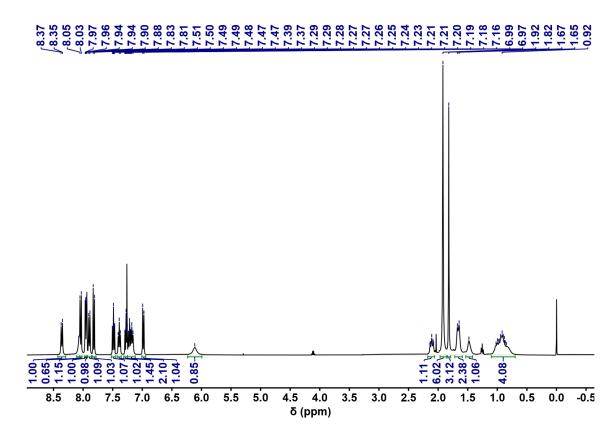


Figure S7. 1 H NMR spectrum (400 MHz, CDCl₃) of (S,R)-1.

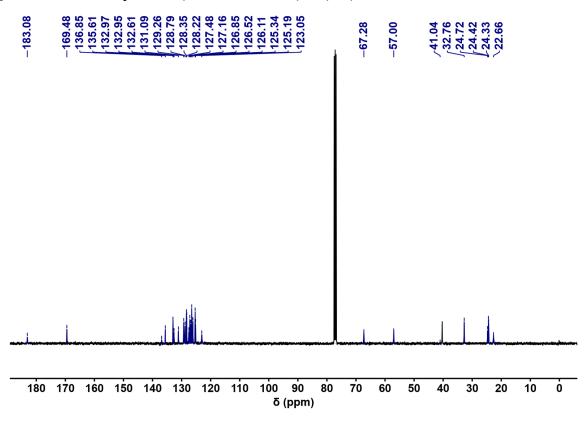


Figure S8. 13 C NMR spectrum (100 MHz, CDCl₃) of (S,R)-1.

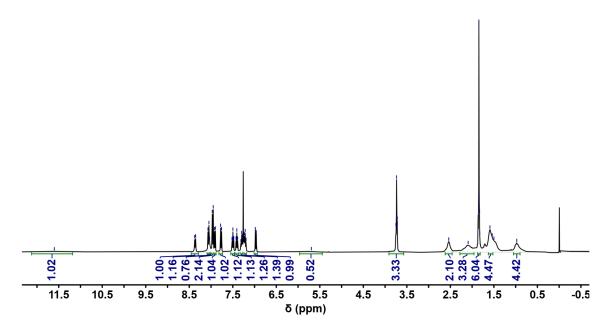


Figure S9. 1 H NMR spectrum (400 MHz, CDCl₃) of (R,S)-2.

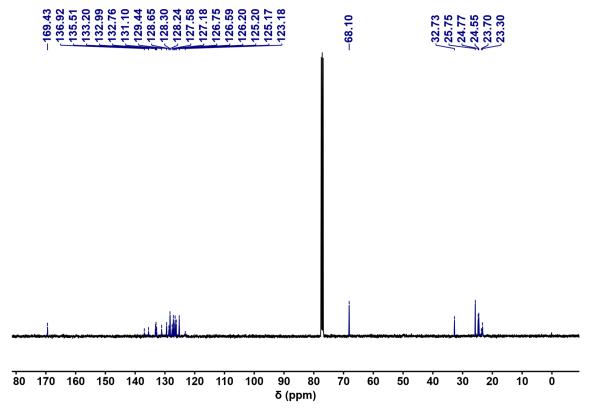


Figure S10. ¹³C NMR spectrum (100 MHz, CDCl₃) of (*R*,*S*)-2.

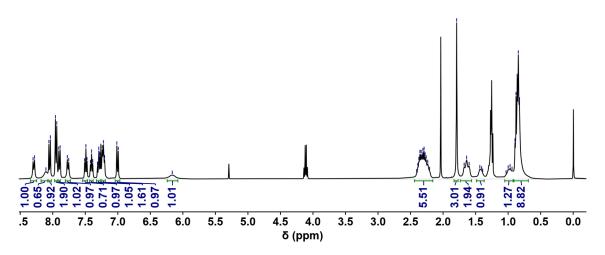


Figure S11. ¹H NMR spectrum (400 MHz, CDCl₃) of (*R*,*S*)-3.

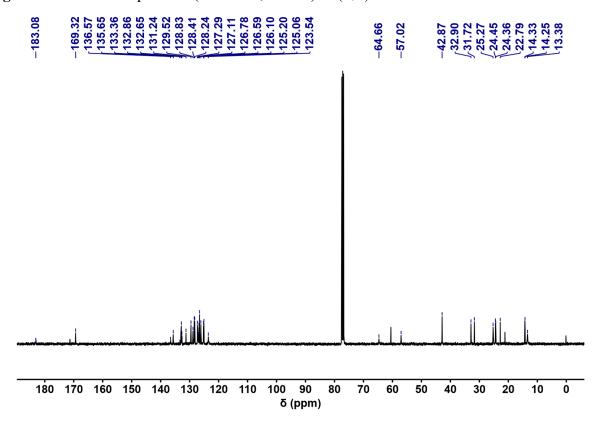


Figure S12. 13 C NMR spectrum (100 MHz, CDCl₃) of (R,S)-3.

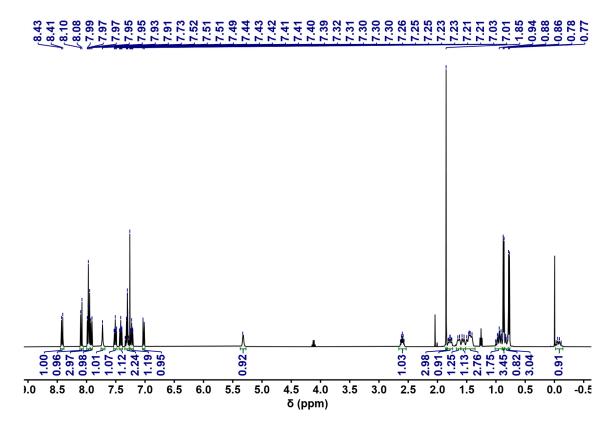


Figure S13. ¹H NMR spectrum (400 MHz, CDCl₃) of (*R*,*S*)-4.

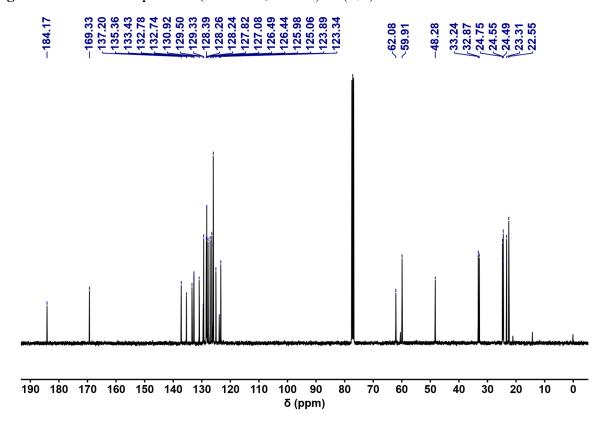


Figure S14. ¹³C NMR spectrum (100 MHz, CDCl₃) of (*R*,*S*)-4.

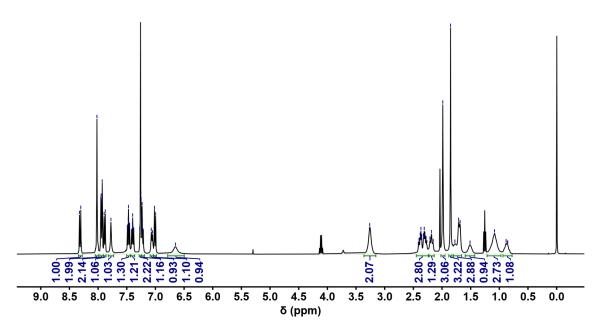


Figure S15. ¹H NMR spectrum (400 MHz, CDCl₃) of (*R*,*S*)-5.

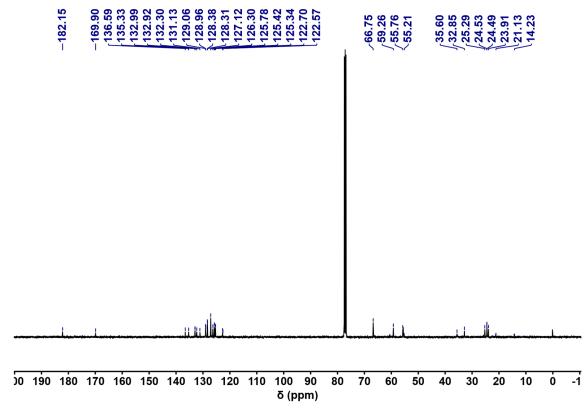


Figure S16. ¹³C NMR spectrum (100 MHz, CDCl₃) of (*R*,*S*)-5.

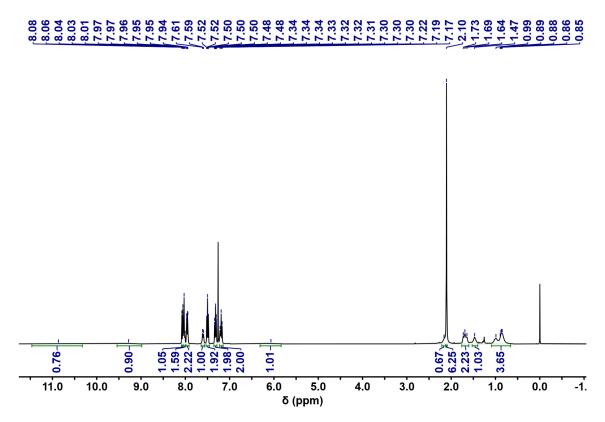


Figure S17. ¹H NMR spectrum (400 MHz, CDCl₃) of (*R*,*S*)-6.

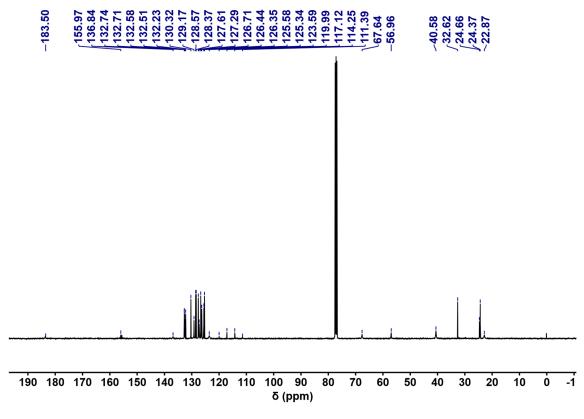


Figure S18. ¹³C NMR spectrum (100 MHz, CDCl₃) of (*R*,*S*)-6.

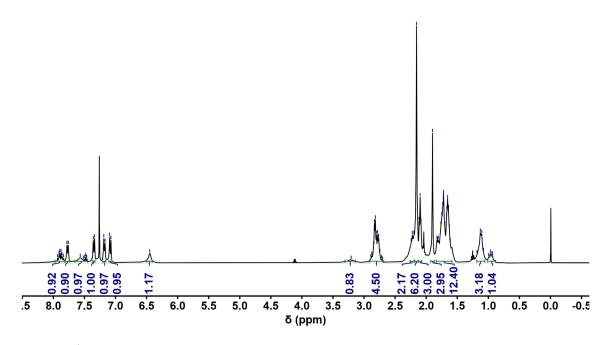


Figure S19. 1 H NMR spectrum (400 MHz, CDCl₃) of (R,S)-7.

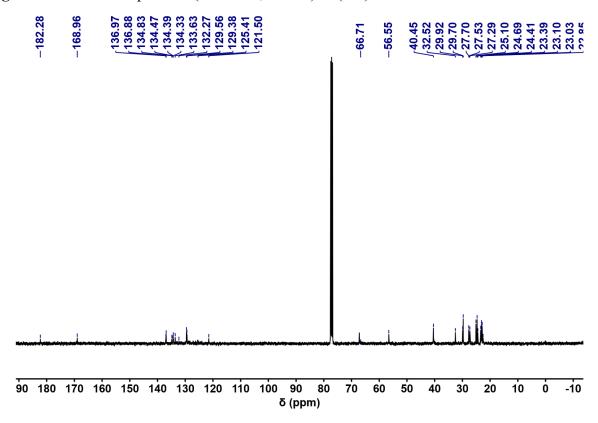


Figure S20. ¹³C NMR spectrum (100 MHz, CDCl₃) of (*R*,*S*)-7.

X-Ray crystallographic analysis

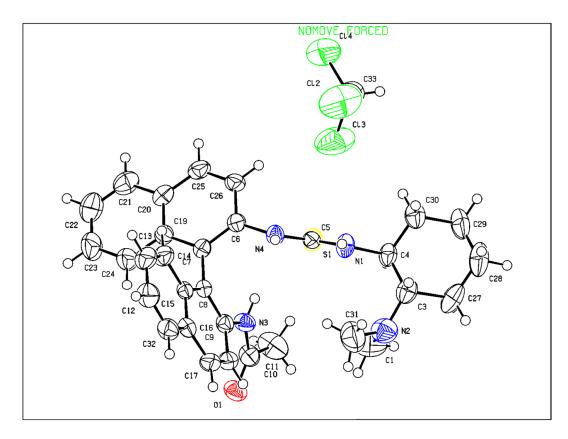


Figure S21. X-ray structure of (R,S)-1.

Table S1. Crystallographic table for (*R*,*S*)-1.

	(R,S)-1	
CCDC Deposition number	2261514	
Empirical formula	C32H35Cl3N4OS	
Formula weight	630.05	
Temperature (K)	290.0	
Crystal system	Monoclinic	
Space group	P2 ₁	
a (Å)	10.4638(10)	
b (Å)	13.6095(10)	
c (Å)	11.5026(10)	
α (°)	90	
β (°)	102.123(4)	
γ (°)	90	
Volume (ų)	1601.5(2)	
Z	2	
ρ_{calc} (g/cm ³)	1.307	
μ (mm ⁻¹)	0.383	
F(000)	660.0	
Crystal size (mm ³)	$0.24 \times 0.13 \times 0.1$	
Radiation	$MoK\alpha (\lambda = 0.71073)$	
Theta range for data collection (°)	3.982 to 55.074	
Index ranges	$\text{-}13 \leqslant h \leqslant 13, \text{-}17 \leqslant k \leqslant 17, \text{-}14 \leqslant l \leqslant 14$	
Reflections collected	18095	
Independent reflections	7340 [$R_{int} = 0.0530$, $R_{sigma} = 0.0659$]	
Data/restraints/parameters	7340/1/373	
Goodness-of-fit on F ²	1.000	
Final R indexes [I>=2σ (I)]	$R_1 = 0.0529, wR_2 = 0.1186$	
Final R indexes [all data]	$R_1 = 0.0842, wR_2 = 0.1383$	
Largest diff. peak/hole (e Å-3)	0.40/-0.36	
Flack parameter	0.07(4)	

Procedure for polymerization

Polymerizations were performed in 4 mL glass reactors inside the glovebox. In a typical polymerization reaction, catalyst (0.056 mmol, 2.0 equiv), initiator (0.028 mmol, 1.0 equiv) and *rac*-LA (200 mg, 1.39 mmol, 50 equiv) were dissolved in 1.19 mL of dichloromethane at 40 °C. At predetermined time intervals, 70 μL aliquots were withdrawn from the polymerization reaction and quickly quenched into 0.5 mL of CDCl₃ acidified with benzoic acid (1.0 wt%). The quenched aliquots were analyzed by ¹H NMR for monomer conversion.

After a desired period of time, the polymerization was quenched by addition of 2.0 mL CHCl₃ acidified with benzoic acid (1.0 wt%). The quenched mixture was precipitated into 50 mL of cold methanol, filtered, and washed with cold methanol to remove any catalyst residue or unreacted monomer and dried in a vacuum oven at 60 °C to a constant weight.

Table S2. Ring-opening polymerization results of LA by different catalysts.^a

Entry	Cat.	M	Solvent	[M]:[Cat]:[I]	T(°C)	<i>t</i> (h)	Conv. (%) ^b	$M_{\rm n}^{c}({\rm kDa})$	\mathcal{D}^{c}	$T_{\mathrm{m}}{}^{d}({}^{\circ}\mathrm{C})$	P_{m}^{e}
1	(R,S)- 1	rac-LA	DCM	50:2:1	40	77	39	5.5	1.09	140	0.95
2^f	(R,S)- 1	rac-LA	CHCl ₃	50:2:1	40	84	48	n.d.	n.d	n.d.	n.d.
3^f	(<i>R</i> , <i>S</i>)-1	D-LA	CHCl ₃	50:2:1	40	84	96	12.4	1.08	163	0.99
4 ^f	(R,S)- 1	_L -LA	CHCl ₃	50:2:1	40	84	16	-	-	-	-
5 ^f	(S,R)- 1	rac-LA	CHCl ₃	50:2:1	40	84	45	6.7	1.08	140	0.95
6 ^f	(S,R)- 1	D-LA	CHCl ₃	50:2:1	40	84	13	-	-	-	-
7 ^f	(S,R)- 1	L-LA	CHCl ₃	50:2:1	40	84	92	11.8	1.07	163	0.99
8 ^g	(R,S)- 1	D-LA	DCM	50:2:1	40	96	91	13.5	1.06	168	0.99
9 g	(R,S)- 1	_L -LA	DCM	50:2:1	40	96	12	-	-	-	-
10 ^g	(R,S)- 2	rac-LA	DCM	50:2:1	40	120	42	7.0	1.07	131	0.91
11 ^g	(R,S)- 2	D-LA	DCM	50:2:1	40	120	83	12.8	1.06	163	0.99
12 ^g	(R,S)- 2	_L -LA	DCM	50:2:1	40	120	14	-	-	-	-
12	(R,S)-1/		TO	50.2.1	70	40	00	6.1	1.10	1.65	0.02
13	(S,R)- 1	rac-LA	TOL	50:2:1	70	48	88	6.1	1.10	165	0.83
1.4	(R,R)- 1 /		TOI	50.2.1	70	40	0.7	<i>C</i> 1	1.00	1	0.70
14	(S,S)- 1	rac-LA	TOL	50:2:1	70	48	87	6.1	1.09	n.d.	0.79

^aReaction conditions: concentration of monomer (M) is 1.0 M, using BnOH as initiator (I), reaction temperature (T), reaction time (T), (T),

Table S3. Ring-opening polymerization results of *rac*-LA by (*R*,*S*)-1 and Base.^a

Entry	Base	Solvent	[M]:[Cat]:[base]:[I]	T(°C)	t (min)	Conv. (%) ^b	$M_{\rm n}^{c}$ (kDa)	\mathcal{D}^{c}	$T_{\mathrm{m}}{}^{c}({}^{\circ}\mathrm{C})$	$P_{\mathrm{m}}{}^{e}$
1 ^f	TBD	CHCl ₃	100:1:1:1	25	60	≥99	20.0	1.49	n.d.	0.67
2^g	MeOK	THF	100:3:1:0	25	5	≥99	24.0	1.54	n.d.	0.65
3^g	MeONa	THF	100:3:1:0	25	1	≥99	18.6	1.46	n.d.	0.71

^aReaction conditions: concentration of monomer (M) is 1.0 M, reaction temperature (T), reaction time (t). ^bMonomer conversion measured by ¹H NMR of the quenched solution. ^cNumber-average molecular weight (M_n) and dispersity index ($D = M_w/M_n$) determined by size exclusion chromatography (SEC) at 40 °C in THF. ^d T_m values were determined by DSC, n.d. = not detected. ^cDetermined by homonuclear decoupled ¹H NMR and calculated based on ESC mechanism. ^fTBD = 1,5,7-[4.4.0]dec-5-ene, BnOH as initiator. ^gmethoxide as initiator.

Procedure for chiral HPLC chromatograms of the unreacted monomer

<u>Control experiment</u>: D-LA (2.0 mg) and L-LA (5.5 mg) were dissolved in 1.5 mL of DCM/hexane = 1/2 solution for standard HPLC test.

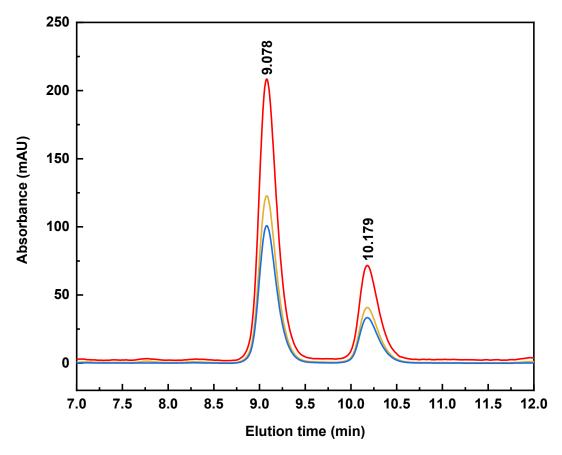


Figure S22. HPLC chromatograms of $_D$ -LA/ $_L$ -LA = 4/11. Column, Chiralpak IA; flow rate, 1.0 mL/min; eluent, hexane/isopropanol = 90/10; detector, UV (230 nm; 210 nm; 202 nm); temperature, 30 °C.

signal	#	Time	Area	Height	Area%	ee%
210	L-LA	9.079	1763.12	122.61	73.38	46.76
210 nm	_D -LA	10.178	639.60	40.58	26.62	
202	L-LA	9.078	2964.36	206.32	73.70	47.39
202 nm	_D -LA	10.179	1058.00	68.91	26.30	
220	L-LA	9.079	1444.22	100.96	73.31	46.61
230 nm	D-LA	10.178	525.91	33.48	26.69	

Procedure for chiral HPLC test of the unreacted reaction mixture: After a desired period of time, the polymerization was quenched by addition of 2.0 mL CHCl₃ acidified with benzoic acid (1.0 wt%). The quenched mixture was precipitated into 50 mL of cold methanol, filtered, and washed with cold methanol to remove any catalyst residue or unreacted monomer. The solution was concentrated under reduced pressure, and the residue was purified by flash chromatograph on silica gel (DCM as eluent) to obtain unreacted monomer. The unreacted monomer was dissolved in a DCM/hexane = 1/2 solution and the ee of the monomer was measured using an Agilent 1260 Series HPLC. The selectivity factor, s, was determined from the equation $s = {ln[(1 - c)(1 - ee)]}/{ln[(1 - c)(1 + ee)]}$, where c is the monomer conversion and ee is the enantiomeric excess of the unreacted monomer.

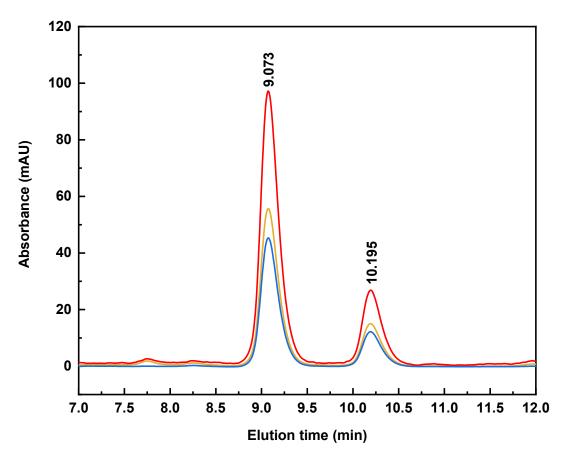


Figure S23. HPLC chromatograms of unreacted monomer obtained by (R,S)-1 (**Table S2** Entry 1). Column, Chiralpak IA; flow rate, 1.0 mL·min⁻¹; eluent, hexane-isopropanol = 90/10; detector, UV (230 nm); temperature, 30 °C.

signal	#	Time	Area	Height	Area%	ee%
210	_L -LA	9.074	801.95	55.57	77.63	55.26
210 nm	_D -LA	10.192	231.11	14.97	22.37	
	L-LA	9.073	1378.22	96.26	77.29	54.59
202 nm	_D -LA	10.195	404.84	26.17	22.71	
230 nm	_L -LA	9.074	648.81	45.50	77.54	55.09
	_D -LA	10.192	187.90	12.26	22.46	

$$ee\% = (54.98 \pm 0.35)\%$$

 $s = k_D/k_L = \{ln[(1-c)(1-ee)]\}/\{ln[(1-c)(1+ee)]\} = 23.0$

Kinetic study of the ROP of rac-LA, D-LA, and L-LA

Table S4. The raw date of kinetic for ROP of rac-LA, D-LA, and L-LA catalyzed by (R,S)-1.^a

Entry	t (h)	Conv. _{rac-LA} (%)	Conv. _{D-LA} (%)	Conv. _{L-LA} (%)
1	12	11	27	2
2	24	19	53	3
3	36	26	72	5
4	48	33	82	8
5	60	39	90	12
6	72	45	93	13
7	84	48	94	16

^aReaction conditions: [M]/[(R,S)-1]/[I] = 50/2/1, concentration of monomer (M) = 1.0 M, initiator (I) = 4-methylbenzyl alcohol, solvent = CHCl₃, reaction temperature = 40 °C.

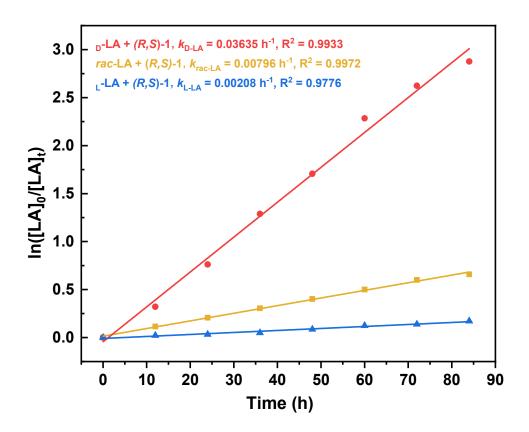


Figure S24. Plots of $\ln([LA]_0/[LA]_t)$ versus time for polymerization catalyzed by (R,S)-1 $(k_{(R,S)-1/D-LA}/k_{(R,S)-1/L-LA} = 17.5)$. Red dot: ROP of D-LA, $k_{(R,S)-1/D-LA} = 0.03635 \text{ h}^{-1}$, $R^2 = 0.9933$. Yellow dot: ROP of rac-LA, $k_{(R,S)-1/rac}$ -LA = 0.00796 h⁻¹, $R^2 = 0.9972$. Blue dot: ROP of L-LA, $k_{(R,S)-1/L-LA} = 0.00208 \text{ h}^{-1}$, $R^2 = 0.9776$.

Table S5. The raw date of kinetic for ROP of rac-LA, D-LA, and L-LA catalyzed by (S,R)-1.^a

Entry	<i>t</i> (h)	Conv. _{rac-LA} (%)	Conv. _{D-LA} (%)	Conv. _{L-LA} (%)
1	12	11	2	27
2	24	19	4	49
3	36	27	5	65
4	48	32	7	77
5	60	38	10	84
6	72	44	12	89
7	84	45	13	92

^aReaction conditions: [M]/[(S,R)-1]/[I] = 50/2/1, concentration of monomer (M) = 1.0 M, initiator (I) = 4-methylbenzyl alcohol, solvent = CHCl₃, reaction temperature = 40 °C.

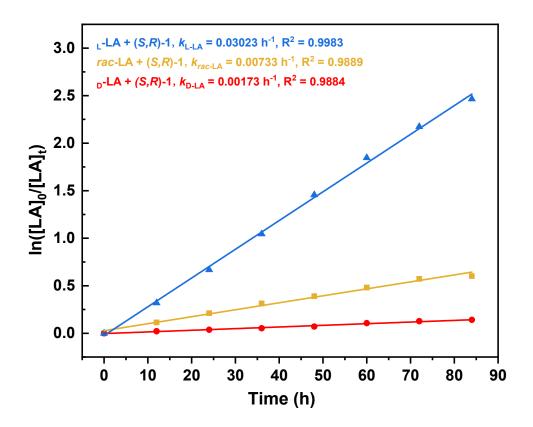


Figure S25. Plots of $\ln([LA]_0/[LA]_t)$ versus time for polymerization catalyzed by (S,R)-1 (k(S,R)-1/L-LA)/k(S,R)-1/D-LA = 17.5). Red dot: ROP of D-LA, k(S,R)-1/D-LA = 0.00173 h⁻¹, R² = 0.9884. Yellow dot: ROP of rac-LA, k(S,R)-1/rac-LA = 0.00733 h⁻¹, R² = 0.9889. Blue dot: ROP of L-LA, k(S,R)-1/L-LA = 0.0302 h⁻¹, R² = 0.9983.

Table S6. The raw date of kinetic for ROP of rac-LA, D-LA, and L-LA catalyzed by (R,S)-1.^a

Entry	<i>t</i> (h)	Conv. _{rac-LA} (%)	Conv. _{D-LA} (%)	Conv. _{L-LA} (%)
1	12	9	20	1
2	24	16	40	2
3	36	22	54	3
4	48	27	67	5
5	60	32	74	7
6	72	36	82	9
7	84	42	87	10
8	96	45	91	12

^aReaction conditions: [M]/[(R,S)-1]/[I] = 50/2/1, concentration of monomer (M) = 1.0 M, initiator (I) = D-methyl lactate, solvent = DCM, reaction temperature = 40 °C.

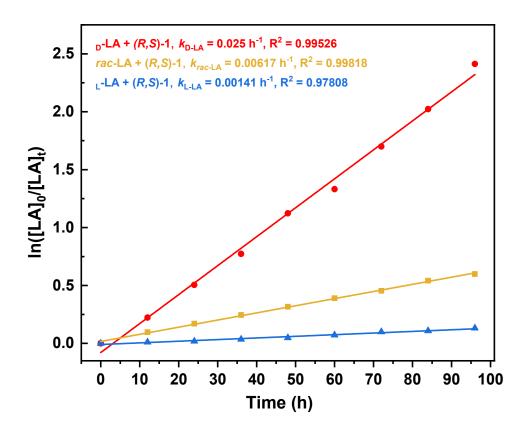


Figure S26. Plots of $\ln([LA]_0/[LA]_t)$ versus time for polymerization catalyzed by (R,S)-1 $(k_{(R,S)-1/D-LA}/k_{(R,S)-1/L-LA} = 17.7)$. Red dot: ROP of D-LA, $k_{(R,S)-1/D-LA} = 0.0250 \, h^{-1}$, $R^2 = 0.9953$. Yellow dot: ROP of rac-LA, $k_{(R,S)-1/rac}$ -LA = 0.00617 h^{-1} , $R^2 = 0.9982$. Blue dot: ROP of L-LA, $k_{(R,S)-1/L-LA} = 0.00141 \, h^{-1}$, $R^2 = 0.9781$.

Table S7. The raw date of kinetic for ROP of rac-LA, D-LA, and L-LA catalyzed by (R,S)-2.

Entry	t (h)	$\operatorname{Conv.}_{rac\text{-}\operatorname{LA}^{a}}\left(\%\right)$	Conv. _{D-LA} b (%)	Conv. _{L-LA} c (%)
1	12	6	16	1
2	24	12	30	2
3	36	18	42	5
4	48	24	55	6
5	60	30	64	8
6	72	35	73	10
7	84	39	78	12
8	96	42	83	14

^aReaction conditions: [M]/[(R,S)-2]/[I] = 50/2/1, concentration of monomer (M) = 1.0 M, initiator (I) = D-methyl lactate, solvent = DCM, reaction temperature = 40 °C.

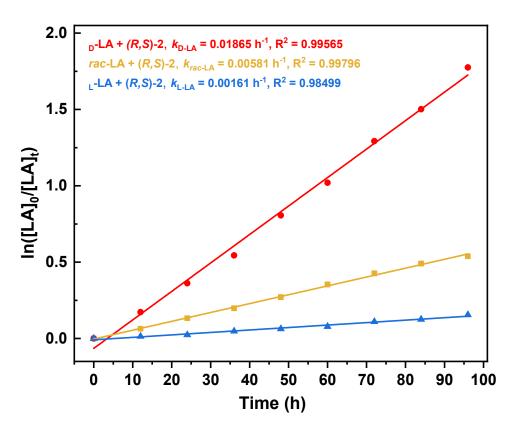


Figure S27. Plots of $\ln([LA]_0/[LA]_t)$ versus time for polymerization catalyzed by (R,S)-2 $(k_{(R,S)-2/D-LA}/k_{(R,S)-2/L-LA} = 11.6)$. Red dot: ROP of D-LA, $k_{(R,S)-2/D-LA} = 0.0187 \, h^{-1}$, $R^2 = 0.9957$. Yellow dot: ROP of rac-LA, $k_{(R,S)-2/rac-LA} = 0.00581 \, h^{-1}$, $R^2 = 0.9980$. Blue dot: ROP of L-LA, $k_{(R,S)-2/L-LA} = 0.00161 \, h^{-1}$, $R^2 = 0.9850$.

Thermal properties of polymers

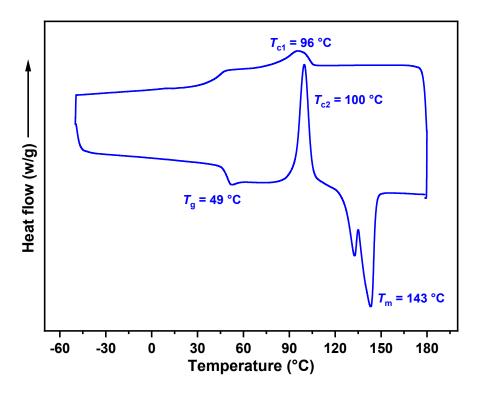


Figure S28. DSC curves of PLA obtained by [rac-LA]/[(R,S)-1]/[I] = 50/2/1, solvent = DCM.

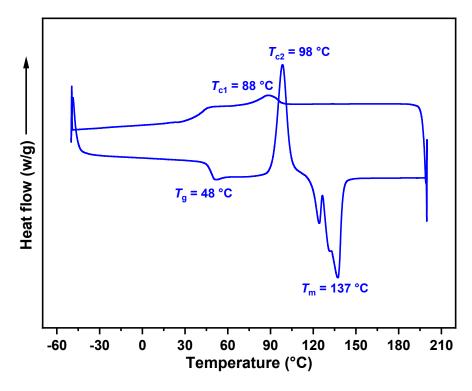


Figure S29. DSC curve of PLA obtained by [rac-LA]/[(R,S)-1]/[I] = 50/2/1, solvent = TOL.

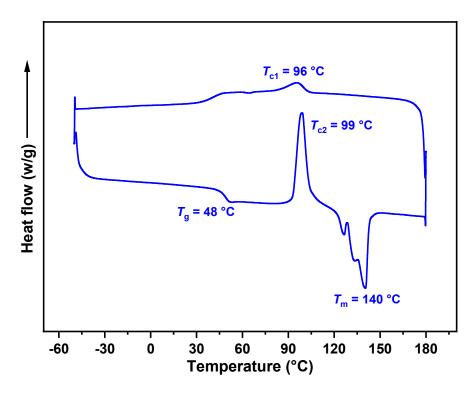


Figure S30. DSC curves of PLA obtained by [rac-LA]/[(R,S)-1]/[I] = 50/2/1, solvent = CHCl₃.

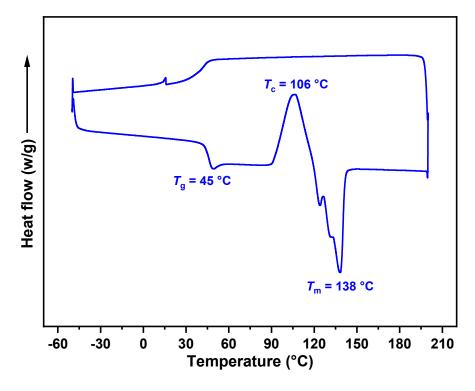


Figure S31. DSC curves of PLA obtained by [rac-LA]/[(S,R)-1]/[I] = 50/2/1, solvent = DCM.

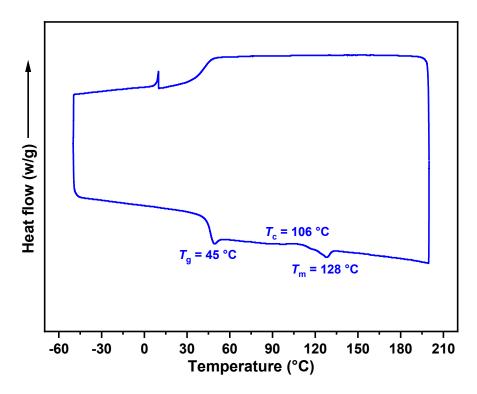


Figure S32. DSC curves of PLA obtained by [rac-LA]/[(R,R)-1]/[I] = 50/2/1, solvent = DCM.

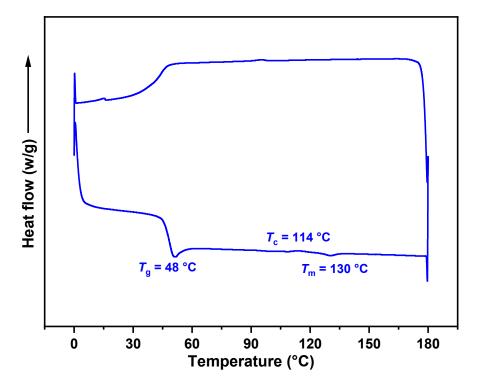


Figure S33. DSC curves of PLA obtained by [rac-LA]/[(S,S)-1]/[I] = 50/2/1, solvent = DCM.

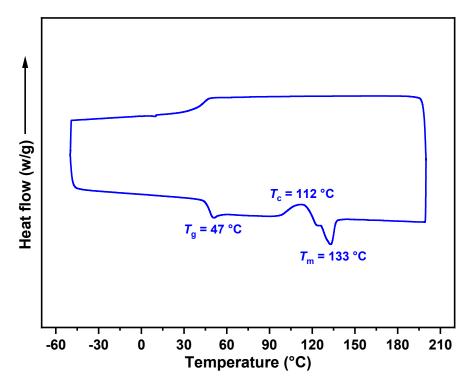


Figure S34. DSC curves of PLA obtained by [rac-LA]/[(R,S)-2]/[I] = 50/2/1, solvent = DCM.

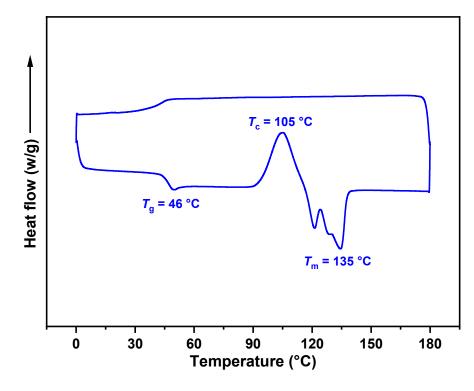


Figure S35. DSC curves of PLA obtained by [rac-LA]/[(R,S)-3]/[I] = 50/2/1, solvent = DCM.

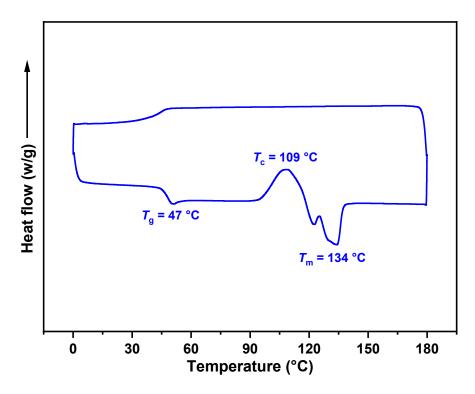


Figure S36. DSC curves of PLA obtained by [rac-LA]/[(R,S)-6]/[I] = 50/2/1, solvent = DCM.

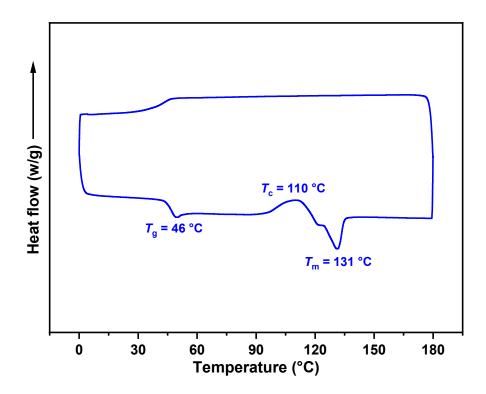


Figure S37. DSC curves of PLA obtained by [rac-LA]/[(R,S)-7]/[I] = 50/2/1, solvent = DCM.

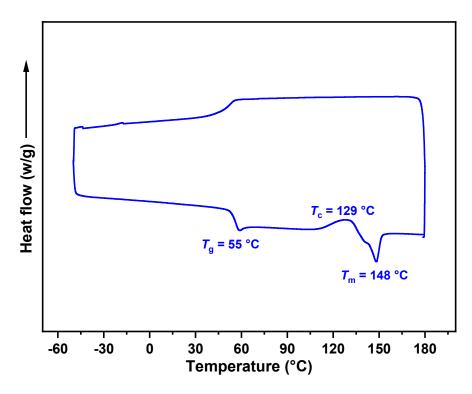


Figure S38. DSC curves of PLA obtained by [rac-LA]/[(R,S)-1]/[I] = 200/20/1, solvent = DCM.

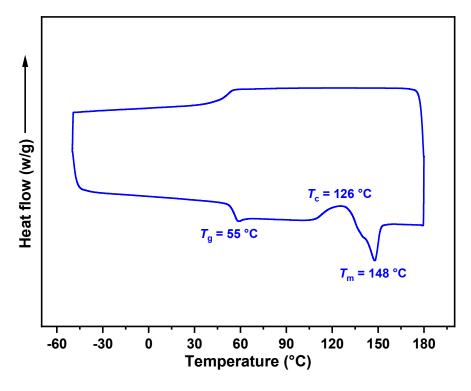


Figure S39. DSC curves of PLA obtained by [rac-LA]/[(S,R)-1]/[I] = 200/20/1, solvent = DCM.

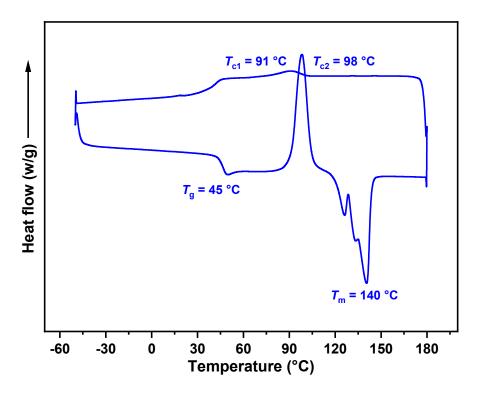


Figure S40. DSC curves of PLA obtained by [rac-LA]/[(R,S)-1]/[I] = 50/2/1, solvent = DCM.

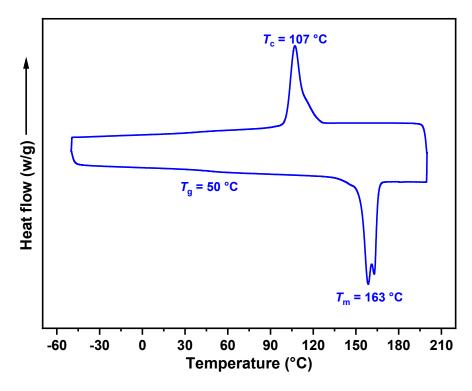


Figure S41. DSC curves of PLA obtained by [D-LA]/[(R,S)-1]/[I] = 50/2/1, solvent = CHCl₃.

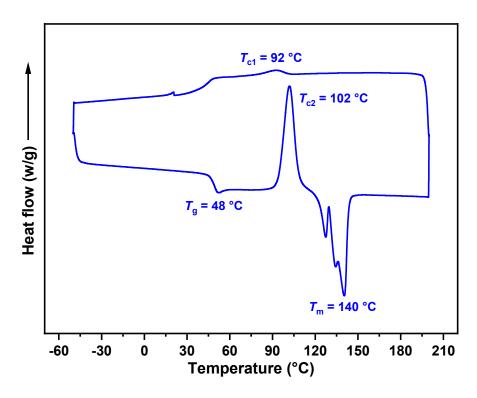


Figure S42. DSC curves of PLA obtained by [rac-LA]/[(S,R)-1]/[I] = 50/2/1, solvent = CHCl₃.

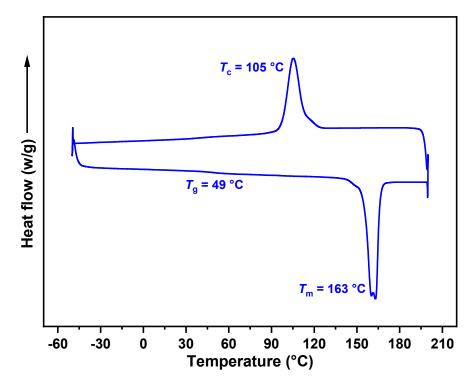


Figure S43. DSC curves of PLA obtained by [L-LA]/[(S,R)-1]/[I] = 50/2/1, solvent = CHCl₃.

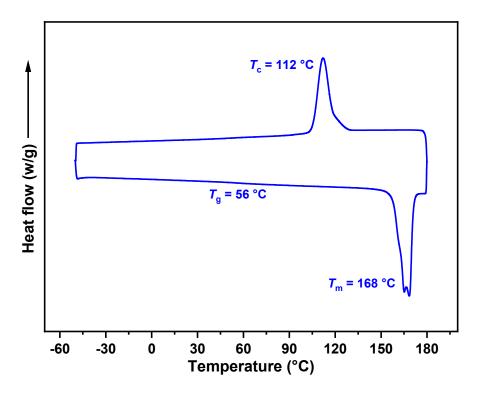


Figure S44. DSC curves of PLA obtained by $[_D-LA]/[(R,S)-1]/[I] = 50/2/1$, solvent = DCM.

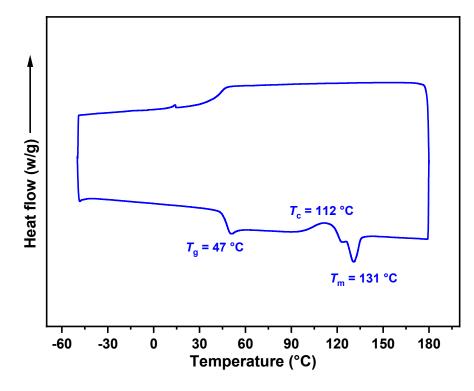


Figure S45. DSC curves of PLA obtained by [rac-LA]/[(R,S)-2]/[I] = 50/2/1, solvent = DCM.

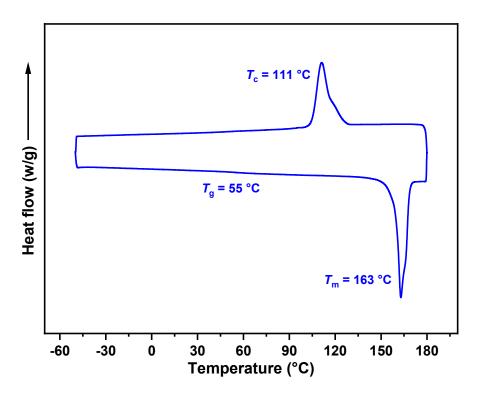


Figure S46. DSC curves of PLA obtained by $[_D$ -LA]/[(R,S)-2]/[I] = 50/2/1, solvent = DCM.

SEC trace of PLAs

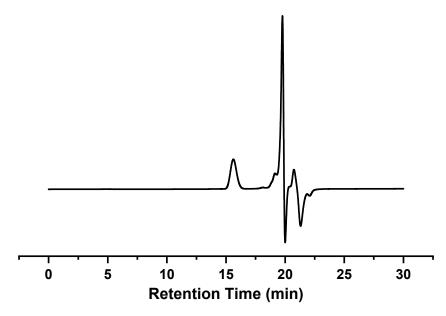


Figure S47. SEC trace of PLA obtained by [rac-LA]/[(R,S)-1]/[I] = 50/2/1, solvent = DCM $(M_n = 6.4 \text{ kg/mol}, D = 1.08)$.

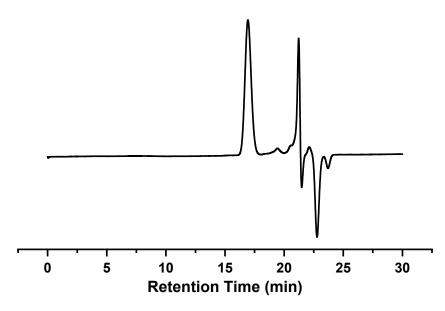


Figure S48. SEC trace of PLA obtained by [rac-LA]/[(R,S)-1]/[I] = 50/2/1, solvent = TOL $(M_n = 5.6 \text{ kg/mol}, D = 1.08)$.

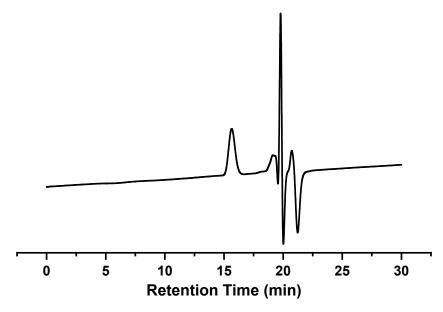


Figure S49. SEC trace of PLA obtained by [rac-LA]/[(R,S)-1]/[I] = 50/2/1, solvent = CHCl₃ ($M_n = 6.2$ kg/mol, D = 1.09).

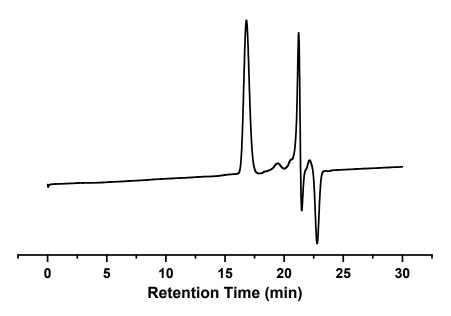


Figure S50. SEC trace of PLA obtained by [rac-LA]/[(R,R)-1]/[I] = 50/2/1, solvent = DCM $(M_n = 6.0 \text{ kg/mol}, D = 1.06)$.

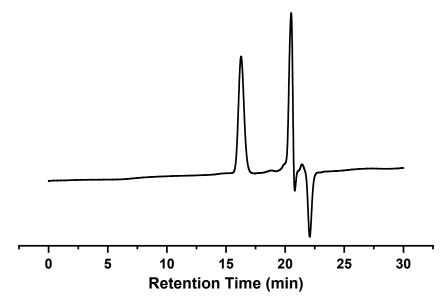


Figure S51. SEC trace of PLA obtained by [rac-LA]/[(S,S)-1]/[I] = 50/2/1, solvent = DCM $(M_n = 6.2 \text{ kg/mol}, D = 1.06)$.

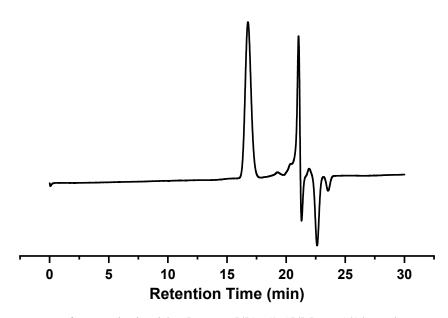


Figure S52. SEC trace of PLA obtained by [rac-LA]/[(R,S)-2]/[I] = 50/2/1, solvent = DCM $(M_n = 5.6 \text{ kg/mol}, D = 1.07)$.

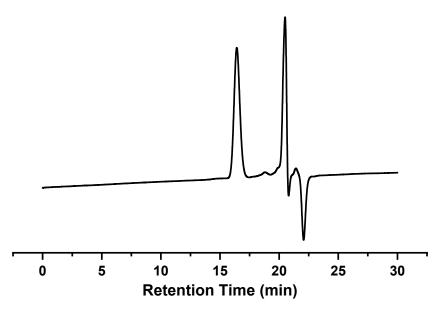


Figure S53. SEC trace of PLA obtained by [rac-LA]/[(R,S)-3]/[I] = 50/2/1, solvent = DCM $(M_n = 5.3 \text{ kg/mol}, D = 1.07)$.

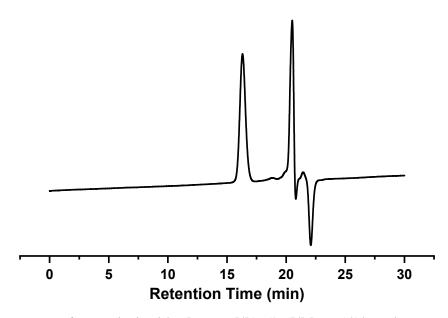


Figure S54. SEC trace of PLA obtained by [rac-LA]/[(R,S)-6]/[I] = 50/2/1, solvent = DCM $(M_n = 6.0 \text{ kg/mol}, D = 1.07)$.

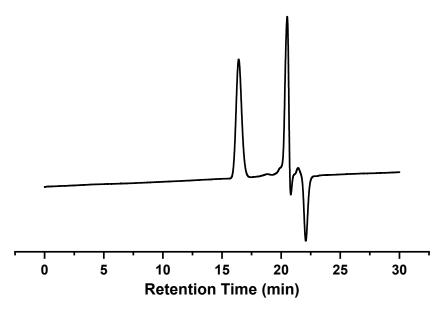


Figure S55. SEC trace of PLA obtained by [rac-LA]/[(R,S)-7]/[I] = 50/2/1, solvent = DCM $(M_n = 5.3 \text{ kg/mol}, D = 1.07)$.

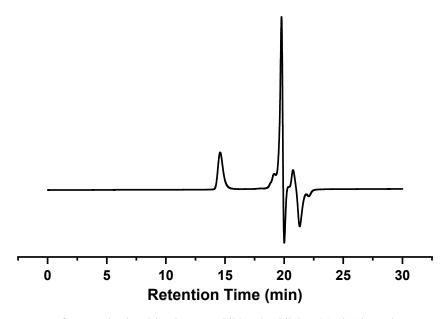


Figure S56. SEC trace of PLA obtained by [rac-LA]/[(R,S)-1]/[I] = 200/20/1, solvent = DCM $(M_n = 19.7 \text{ kg/mol}, D = 1.09)$.

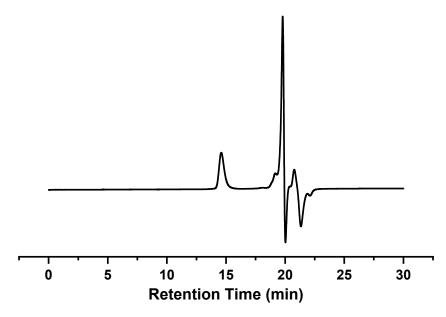


Figure S57. SEC trace of PLA obtained by [rac-LA]/[(S,R)-1]/[I] = 200/20/1, solvent = DCM $(M_n = 19.1 \text{ kg/mol}, D = 1.10)$.

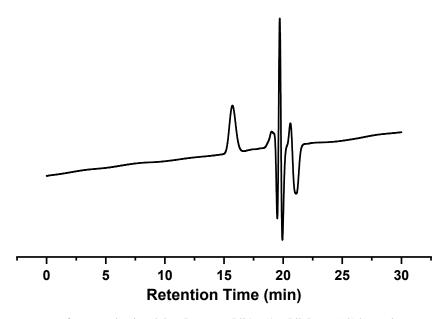


Figure S58. SEC trace of PLA obtained by [rac-LA]/[(R,S)-1]/[I] = 50/2/1, solvent = DCM $(M_n = 5.5 \text{ kg/mol}, D = 1.09)$.

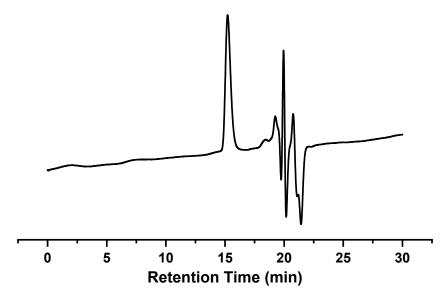


Figure S59. SEC trace of PLA obtained by $[_D$ -LA]/[(R,S)-1]/[I] = 50/2/1, solvent = CHCl₃ ($M_n = 12.4$ kg/mol, D = 1.08).

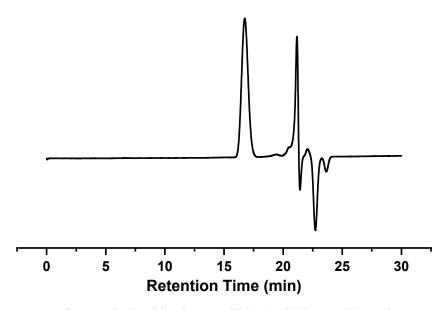


Figure S60. SEC trace of PLA obtained by [rac-LA]/[(S,R)-1]/[I] = 50/2/1, solvent = CHCl₃ ($M_n = 6.7$ kg/mol, D = 1.08).

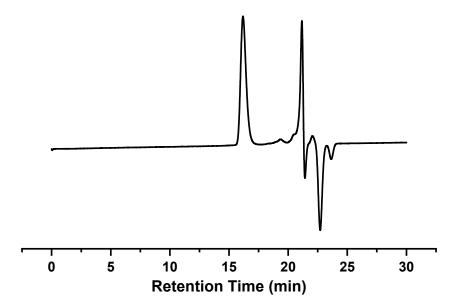


Figure S61. SEC trace of PLA obtained by [L-LA]/[(S,R)-1]/[I] = 50/2/1, solvent = CHCl₃ ($M_n = 11.8$ kg/mol, D = 1.07).

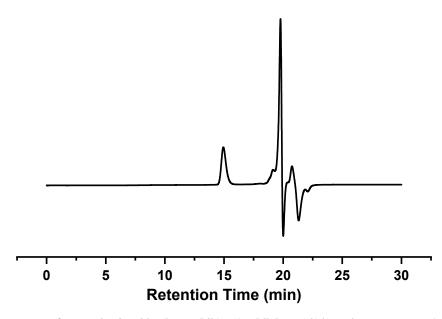


Figure S62. SEC trace of PLA obtained by $[_D$ -LA]/[(R,S)-1]/[I] = 50/2/1, solvent = DCM (M_n = 13.5 kg/mol, D = 1.06).

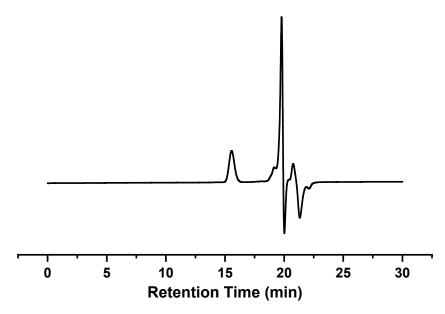


Figure S63. SEC trace of PLA obtained by [rac-LA]/[(R,S)-2]/[I] = 50/2/1, solvent = DCM $(M_n = 7.0 \text{ kg/mol}, D = 1.07)$.

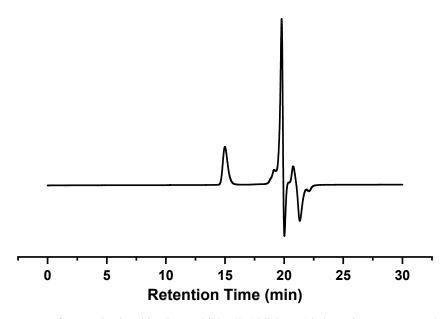


Figure S64. SEC trace of PLA obtained by $[_D$ -LA]/[(R,S)-2]/[I] = 50/2/1, solvent = DCM (M_n = 12.8 kg/mol, D = 1.06).

Homonuclear decoupled ¹H NMR spectra of PLAs

Homodecoupled ${}^{1}H$ NMR spectra were recorded for all the obtained PLA samples. The mechanism of this catalytic system is enantiomorphic site control mechanism (ESC). So the calculation of $P_{\rm m}$ values for each sample followed tetrad probabilities of ESC mechanisms based on non-Bernouillanin in the literature. 1

Table S8. Tetrad probabilities of ESC mechanisms based on non-Bernouillanin.

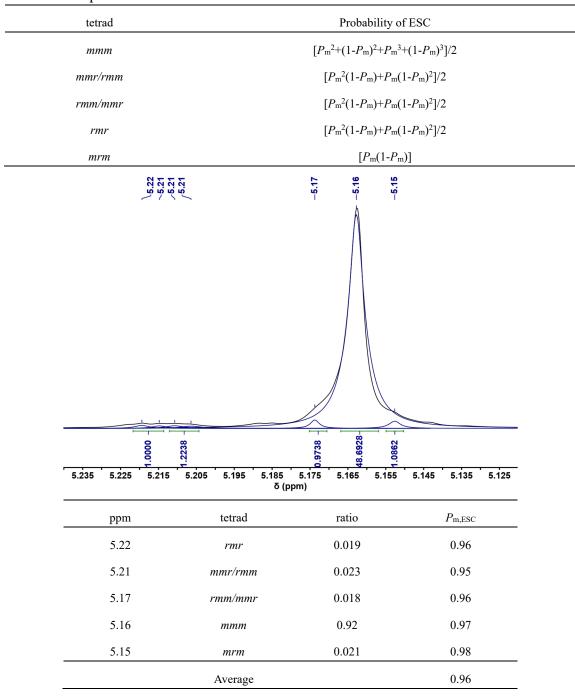


Figure S65. Homonuclear decoupled ¹H NMR spectrum (400 MHz, CDCl₃) of PLA obtained by [*rac*-LA]/[(R,S)-1]/[I] = 50/2/1, solvent = DCM (P_m = 0.96).

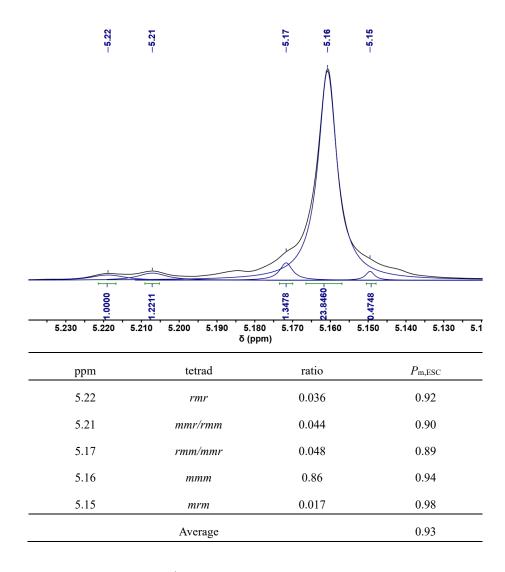


Figure S66. Homonuclear decoupled ¹H NMR spectrum (400 MHz, CDCl₃) of PLA obtained by [rac-LA]/[(R,S)-1]/[I] = 50/2/1, solvent = TOL (P_m = 0.93).

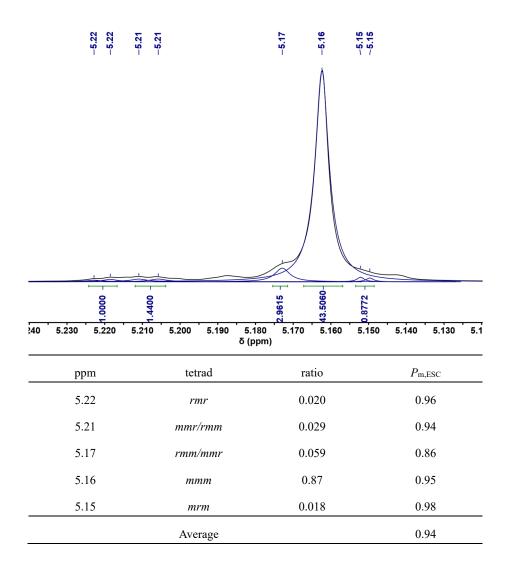


Figure S67. Homonuclear decoupled ¹H NMR spectrum (400 MHz, CDCl₃) of PLA obtained by [rac-LA]/[(R,S)-1]/[I] = 50/2/1, solvent = CHCl₃ (P_m = 0.94).

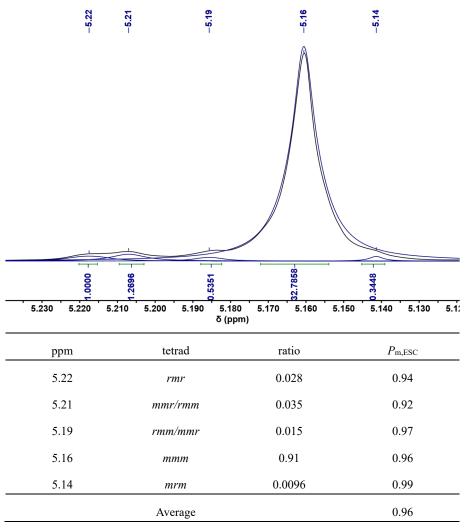


Figure S68. Homonuclear decoupled ¹H NMR spectrum (400 MHz, CDCl₃) of PLA obtained by [rac-LA]/[(S,R)-1]/[I] = 50/2/1, solvent = DCM (P_m = 0.96).

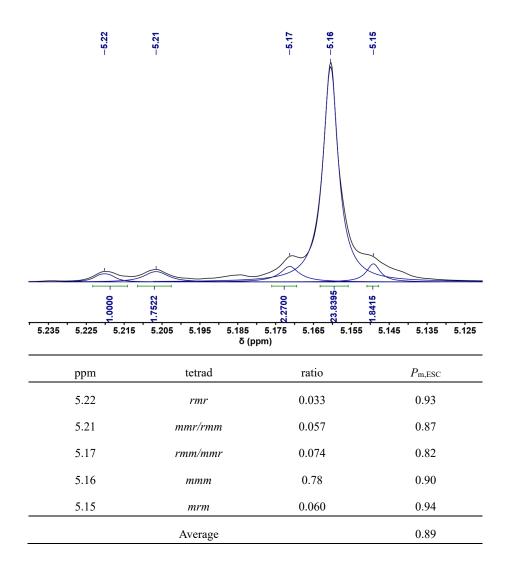


Figure S69. Homonuclear decoupled ¹H NMR spectrum (400 MHz, CDCl₃) of PLA obtained by [rac-LA]/[(R,R)-1]/[I] = 50/2/1, solvent = DCM (P_m = 0.89).

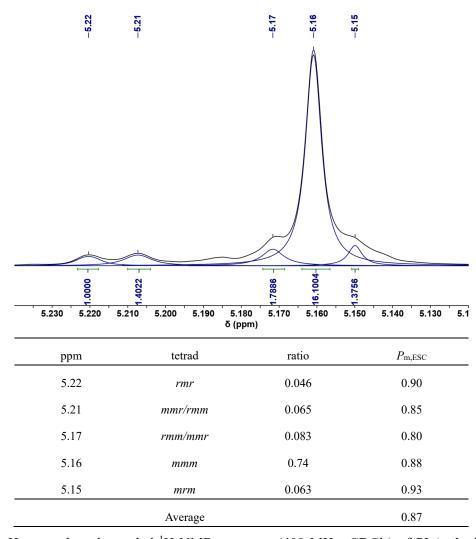


Figure S70. Homonuclear decoupled ¹H NMR spectrum (400 MHz, CDCl₃) of PLA obtained by [rac-LA]/[(S,S)-1]/[I] = 50/2/1, solvent = DCM (P_m = 0.87).

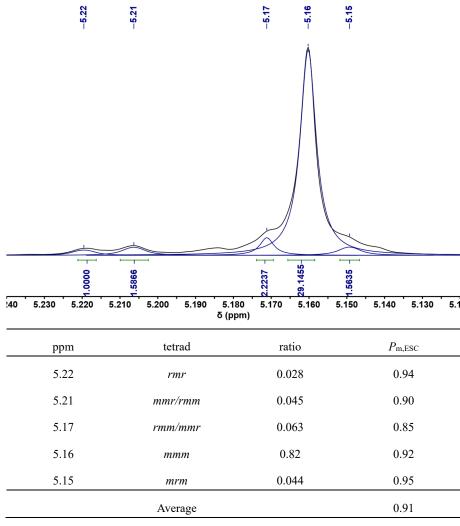


Figure S71. Homonuclear decoupled ¹H NMR spectrum (400 MHz, CDCl₃) of PLA obtained by [rac-LA]/[(R,S)-2]/[I] = 50/2/1, solvent = DCM (P_m = 0.91).

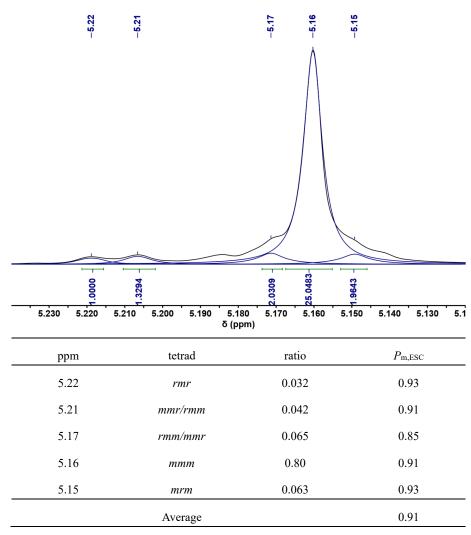


Figure S72. Homonuclear decoupled ¹H NMR spectrum (400 MHz, CDCl₃) of PLA obtained by [rac-LA]/[(R,S)-3]/[I] = 50/2/1, solvent = DCM (P_m = 0.91).

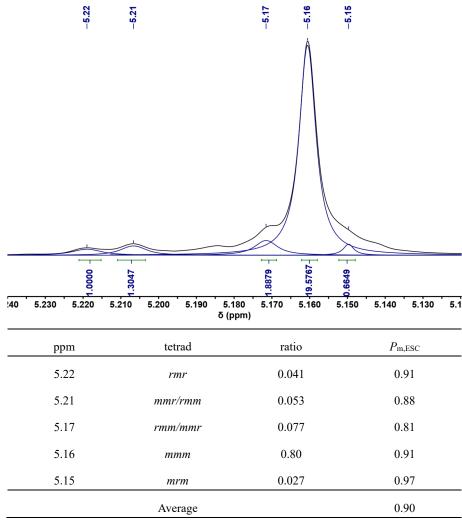


Figure S73. Homonuclear decoupled ¹H NMR spectrum (400 MHz, CDCl₃) of PLA obtained by [rac-LA]/[(R,S)-6]/[I] = 50/2/1, solvent = DCM (P_m = 0.90).

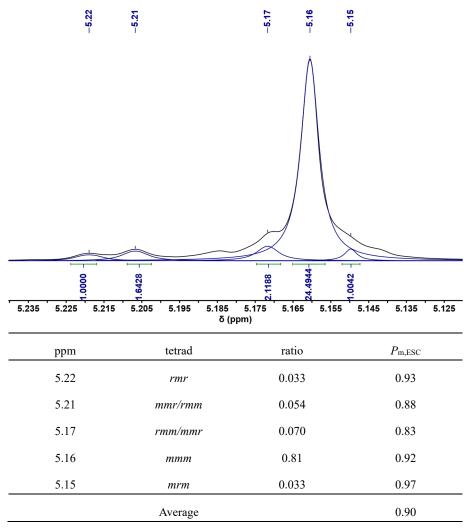


Figure S74. Homonuclear decoupled ¹H NMR spectrum (400 MHz, CDCl₃) of PLA obtained by [rac-LA]/[(R,S)-7]/[I] = 50/2/1, solvent = DCM (P_m = 0.90).

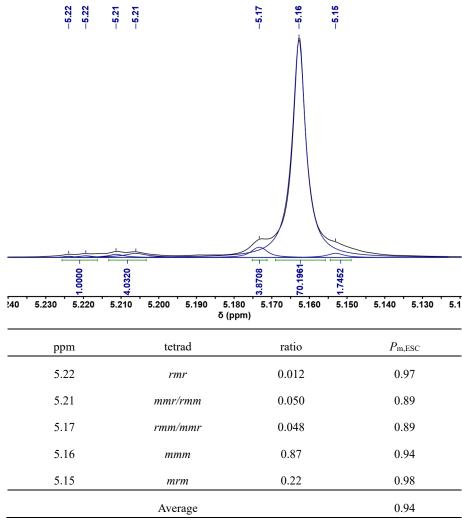


Figure S75. Homonuclear decoupled ¹H NMR spectrum (400 MHz, CDCl₃) of PLA obtained by [rac-LA]/[(R,S)-1]/[I] = 200/20/1, solvent = DCM (P_m = 0.94).

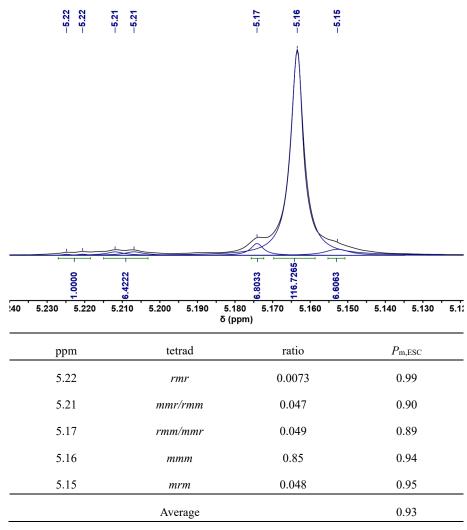


Figure S76. Homonuclear decoupled ¹H NMR spectrum (400 MHz, CDCl₃) of PLA obtained by [rac-LA]/[(S,R)-1]/[I] = 200/20/1, solvent = DCM (P_m = 0.93).

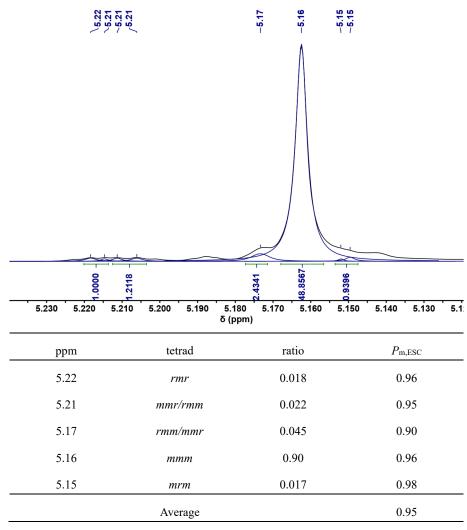


Figure S77. Homonuclear decoupled ¹H NMR spectrum (400 MHz, CDCl₃) of PLA obtained by [rac-LA]/[(R,S)-1]/[I] = 50/2/1, solvent = DCM (P_m = 0.95).

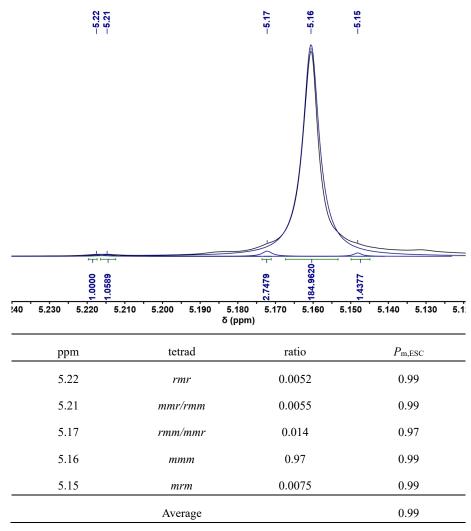


Figure S78. Homonuclear decoupled ¹H NMR spectrum (400 MHz, CDCl₃) of PLA obtained by [D-LA]/[(R,S)-1]/[I] = 50/2/1, solvent = CHCl₃ (P_m = 0.99).

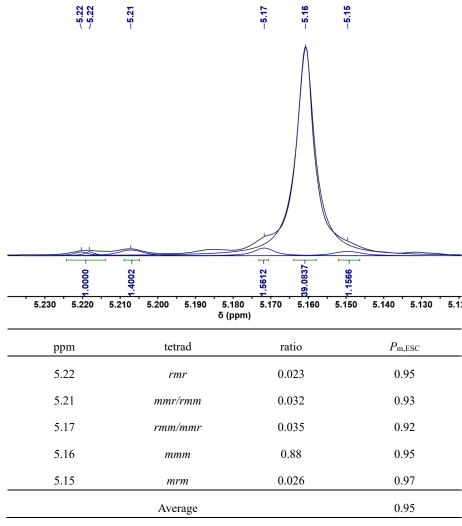


Figure S79. Homonuclear decoupled ¹H NMR spectrum (400 MHz, CDCl₃) of PLA obtained by [rac-LA]/[(S,R)-1]/[I] = 50/2/1, solvent = CHCl₃ (P_m = 0.95).

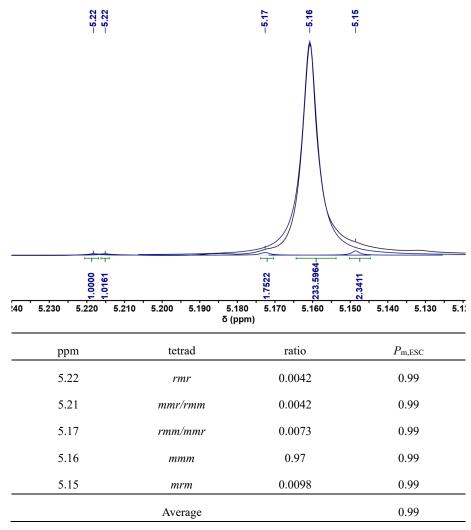


Figure S80. Homonuclear decoupled ¹H NMR spectrum (400 MHz, CDCl₃) of PLA obtained by [L-LA]/[(S,R)-1]/[I] = 50/2/1, solvent = CHCl₃ (P_m = 0.99).

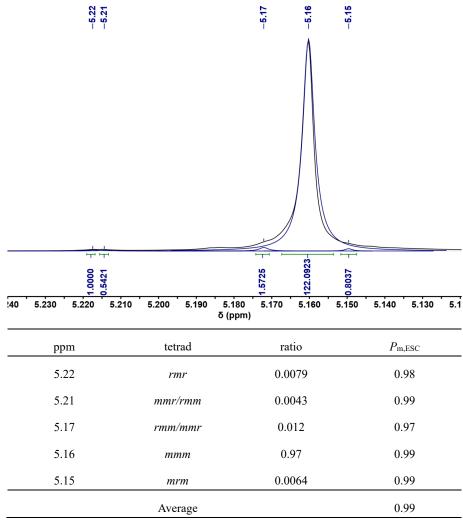


Figure S81. Homonuclear decoupled ¹H NMR spectrum (400 MHz, CDCl₃) of PLA obtained by [D-LA]/[(R,S)-1]/[I] = 50/2/1, solvent = DCM (P_m = 0.99).

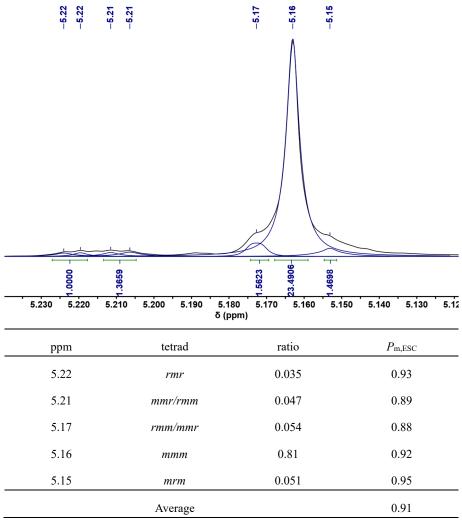


Figure S82. Homonuclear decoupled ¹H NMR spectrum (400 MHz, CDCl₃) of PLA obtained by [rac-LA]/[(R,S)-2]/[I] = 50/2/1, solvent = DCM (P_m = 0.91).

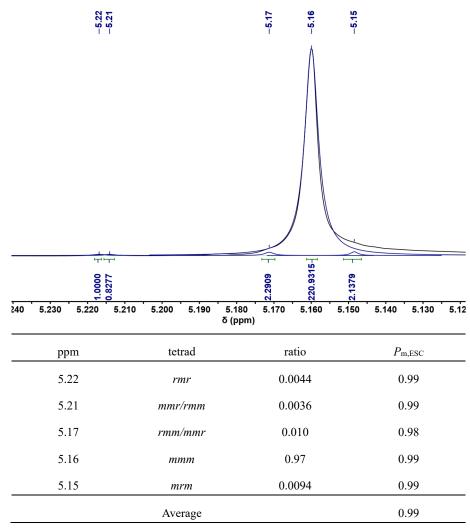


Figure S83. Homonuclear decoupled ¹H NMR spectrum (400 MHz, CDCl₃) of PLA obtained by [D-LA]/[(R,S)-2]/[I] = 50/2/1, solvent = DCM (P_m = 0.99).

References

- 1. Orhan, B.; Tschan, M. J.-L.; Wirotius, A.-L.; Dove, A. P.; Coulembier, O.; Taton, D. Isoselective Ring-Opening Polymerization of *rac*-Lactide from Chiral Takemoto's Organocatalysts: Elucidation of Stereocontrol. *ACS Macro Lett.* **2018**, *7*, 1413-1419.
- 2. Zhu, J.-B.; Chen, E. Y.-X. From *meso*-Lactide to Isotactic Polylactide: Epimerization by B/N Lewis Pairs and Kinetic Resolution by Organic Catalysts. *J. Am. Chem. Soc.* **2015**, *137*, 12506-12509.
- 3. Masson, G.; Bekkaye, M. Synthesis of New Axially Chiral Iodoarenes. *Synthesis* **2015**, *48*, 302-312.
- 4. Kulkarni, C.; Berrocal, J. A.; Lutz, M.; Palmans, A. R. A.; Meijer, E. W. Directing the Solid-State Organization of Racemates via Structural Mutation and Solution-State Assembly Processes. *J. Am. Chem. Soc.* **2019**, *141*, 6302-6309.
- 5. Amarasinghe, N. R.; Turner, P.; Todd, M. H. The First Catalytic, Enantioselective Aza-Henry Reaction of an Unactivated Cyclic Imine. *Adv. Synth. Catal.* **2012**, *354*, 2954-2958.
- 6. Tiffner, M.; Novacek, J.; Busillo, A.; Gratzer, K.; Massa, A.; Waser, M. Design of chiral urea-quaternary ammonium salt hybrid catalysts for asymmetric reactions of glycine Schiff bases. *RSC Adv.* **2015**, *5*, 78941-78949.
- 7. Bulman Page, P.; Farah, M.; Buckley, B.; Chan, Y.; Blacker, A. Preparation of C_2 -Symmetric Biaryl Bisiminium Salts and Their Use as Organocatalysts for Asymmetric Epoxidation. *Synlett* **2015**, *27*, 126-130.
- 8. Yan, L.; Huang, G.; Wang, H.; Xiong, F.; Peng, H.; Chen, F. Squaramide-Linked Chloramphenicol Base Hybrid Catalysts for the Asymmetric Michael Addition of 2,3-Dihydrobenzofuran-2-carboxylates to Nitroolefins. *Eur. J. Org. Chem.* **2018**, 2018, 99-103.
- 9. Liu, F.; Gou, S.; Li, L. Asymmetric Henry reactions of aldehydes with various nitroalkanes catalyzed by copper(II) complexes of novel chiral *N*-monoalkyl cyclohexane-1,2-diamines. *Appl. Organometal. Chem.* **2014**, *28*, 186-193.
- 10. Yamaoka, Y.; Miyabe, H.; Takemoto, Y. Catalytic enantioselective petasis-type reaction of quinolines catalyzed by a newly designed thiourea catalyst. *J. Am. Chem. Soc.* **2007**, *129*, 6686-6687.
- 11. Suez, G.; Bloch, V.; Nisnevich, G.; Gandelman, M. Design and Development of Bioinspired Guanine-Based Organic Catalyst for Asymmetric Catalysis. *Eur. J. Org. Chem.* **2012**, 2012, 2118-2122.