Supporting Information for

Multiplicity-Driven Photochromism Controls Three-State Fulgimide Photoswitches

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Materials and Methods

Dry solvents were purchased from Acros Organics and Thermo Fisher Scientific, Belgium, and dried over molecular sieves before each reaction. All grade-quality reagents were commercially available compounds used without any further purification and purchased from Merck (Germany), Fluorochem Ltd. (United Kingdom), TCI (Japan) and Penta Chemicals (Czech Republic). Reaction temperatures refer to the temperature of surrounding metal heating block.

Thin-Layer Chromatography (TLC) was performed using silica plates "ALUGRAM SIL G/UV 254" from MACHEREY-NAGEL (Germany). Flash chromatography was performed on an ECOM flash column chromatography system using Silicagel 40–60 μ m (Lach-Ner, Czech Republic). Preparative HPLC was performed on a DeltaChrom system from Watrex (Czech Republic) in reverse phase mode. ¹H and ¹³C NMR spectra were recorded on Bruker Avance IIITM HD 400 MHz and Bruker Avance IIITM HD 400 MHz Prodigy spectrometers (¹H at 401 MHz, ¹³C at 101 MHz, ¹⁹F at 377 MHz) or on a Bruker Avance IIITM HD 600 MHz spectrometer (¹H at 600 MHz, ¹³C at 151 MHz). Deuterated solvents used in the measurements are stated for each compound. Solvent signals were used as a reference for each ¹H and ¹³C measurement, and an internal standard (CFCl₃) was used as reference for ¹⁹F measurements and set to 0 ppm. Chemical shifts (δ) are given in ppm and coupling constants (J) in Hz. Signal multiplicity is specified as follows: singlet (s), broad singlet (bs), doublet (d), doublet of doublets (dd), doublet of doublets (dd), triplet (t), quartet (q), multiplet (m). If product consists of mixture of isomers, hydrogen atom equivalents are rounded to nearest two tenths.

Liquid chromatography–mass spectrometry (LCMS) was performed to monitor the reaction conversion and to confirm the presence and purity of synthesized products on an LC-MS-2020 system from Shimadzu Corporation (Japan) with UV-vis and ESI-MS detectors on a CORTECS C18+ (2.7 μ m, 4.6 × 50 mm) reverse phase-column equipped with a VanGuard (3.9 × 5 mm) pre-column from Waters (USA). All compounds were eluted with a mixture of acetonitrile and water with formic acid (0.1 v/v %) in a gradient mode from 95 to 5% water. LCMS-grade formic acid from VWR (Belgium), HPLC-grade acetonitrile from Fisher Scientific (UK) and Milli-Q water were used for chromatography.

High-resolution electrospray ionization (ESI) mass spectra were recorded on an LTQ Orbitrap XL (Thermo Fisher Scientific, USA) hybrid FT mass spectrometer equipped with a linear ion trap MS and the Orbitrap mass analyser. UV-vis absorption was recorded on Agilent Cary 8454. Fluorescence measurements were recorded on a Horiba Duetta spectrometer. All spectra were measured in cells for fluorescence measurements (3.5 mL volume, 10 mm optical path length, HORIBA).

Kinetic measurements were conducted using custom-designed LED light sources with wavelength maxima at the stated wavelengths (Figure S2-S5). The custom-made cuvette and light-source holder was used in all measurements to ensure that the LEDs had the same position and orientation in relation to the sample, which was perpendicular to the light beam used to measure UV-vis spectrometry. Transient absorption experiments were performed on a commercially available apparatus EOS from Ultrafast Systems. For longer delays, ranging from ns to μ s, the pump laser (EKSPLA NT242) was electronically delayed relative to a sub-nanosecond pulsed probe light source (LEUCOS, a photonic crystal fiber-based supercontinuum laser). Each sample in a quartz cuvette with a 2-mm optical path length was randomly moved at 1 mm/s speed throughout the measurement. The stability of the sample was checked by recording steady-state absorption spectra before and after each measurement.

Supplementary Figures and Schemes

Scheme S1. Stereoselectivity of elimination of lactone 2



The intermediate **i1** can exist as two isomers, *s*-trans and *s*-cis. We do not know their relative energies, but the final cyclization to fulgimide requires *s*-cis conformation. Therefore, if *s*-trans isomer is present throughout the reaction, it must be in equilibrium with the *s*-cis isomer, which is rapidly consumed in the last step, thus changing the equilibrium to yield exclusively the inactive form of the fulgimide.

Scheme S2. Copper-mediated click reaction of 1d with benzyl azide



The increase of active form is contributed to time spent on daylight, solvent evaporation at 40°C and the temperature needed for the reaction.

Figure S1. Quenching of anthracene fluorescence by fulgimide 1b (exponential plot shown in inset).



Even though Stern-Volmer analysis should be linear, our quenching was best fitted by an exponential function. This result may be explained by a combination of static and dynamic quenching, as previously described for anthracene by nitroaromatic compound by Airinei et al.¹

Figure S2. Emission profiles of the 400-nm LED used in this study



Figure S3. Emission profiles of the 626-nm LED used in this study



Figure S4. Emission of the 420-nm light source



Figure S5. Emission of the 385-nm light source



Synthesis of the building blocks

1-(1,2-dimethyl-1*H*-indol-3-yl)-2,2,2-trifluoroethan-1-one² S1

A stirred solution of trifluoroacetic acid anhydride (8.6 g, 5.8 mL, 41.0 mmol, 1.1 equiv.) in dry dichloroethane (40 mL) was cooled to 0°C and equipped with a dropping funnel. Once a solution of 1,2-dimethyl-1*H*-indole (5.42 g, 37.3 mmol) in dry dichloroethane (40 mL) was added dropwise, the reaction mixture was warmed to RT and left to react for 3 hours. The reaction was quenched by addition of saturated solution of NaHCO₃ and extracted to DCM. The combined organic layers were washed with brine, dried over MgSO₄ and evaporated, yielding a yellow solid. Recrystallization from ethyl acetate yielded white needle



Chemical Formula: C₁₂H₁₀F₃NO Molecular Weight: 241.213

yielding a yellow solid. Recrystallization from ethyl acetate yielded white needles of product (7.5 g, 31.1 mmol, 84%).

¹H NMR (400 MHz, CDCl₃) δ 8.08 (m, 1H), 7.40 – 7.31 (m, 3H), 3.78 (s, 3H), 2.82 (s, 3H).

NMR spectra corresponded to the lit. data²

Dimethyl 2-(dicyclopropylmethylene)succinate³ S2

A solution of dimethyl succinate (17 g, 116.3 mmol, 1 equiv.) and dicyclopropylketone (12.8 g, 115.1 mmol, 0.99 equiv.) in *tert*-butanol (25 mL) was added to a refluxing mixture of potasium *tert*-butoxide (13 g, 116.3 mmol, 1 equiv.) in *tert*-butanol (130 mL), and the reaction quickly turned yellow. After 30 minutes of reflux, the reaction was cooled to RT and evaporated to dryness, and the residue was dissolved in 2M NaOH (175 mL) and washed with diethyl ether (175 mL). The water phase was acidified (pH = 1) using conc. HCl and extracted to DCM. Combined

organic layers were washed with brine, dried over $MgSO_4$ and evaporated. Residue was dissolved in methanol (450 mL) and acidified using a mixture of conc. HCl and water (100 mL, 1:1, v/v) and left to react overnight. The reaction mixture was evaporated and basified using a saturated solution of NaHCO₃, the resulting water phase was extracted to diethyl ether, the combined organic layers were washed with brine and dried over MgSO₄, and evaporation yielded slightly yellow oil (8.3 g, 34.8 mmol, 30%), which was used without further purification in the following reaction.

¹H NMR (401 MHz, CDCl₃) δ 3.73 (s, 3H), 3.66 (s, 3H), 3.56 (s, 2H), 1.73 (ttd, *J* = 7.7, 5.9, 1.0 Hz, 1H), 1.34 (ttd, *J* = 8.6, 5.8, 0.9 Hz, 1H), 0.79–0.73 (m, 2H), 0.73–0.67 (m, 2H), 0.66–0.61 (m, 2H), 0.50–0.44 (m, 2H).

NMR spectra corresponded to the lit. data³

(±)-Methyl (2R,3S)-4-(dicyclopropylmethylene)-2-(1,2-dimethyl-1H-indol-3-yl)-5-oxo-2-(trifluoromethyl)tetrahydrofuran-3-carboxylate³ 2

Under an inert atmosphere, **S1** (2.15 g, 8.9 mmol, 1 equiv.) and **S2** (4.25 g, 17.8 mmol, 2 equiv.) were dissolved in dry toluene (125 mL), and a solution of 2 M of LDA in THF/*n*-heptane/ethylbenzene in a 59:28:13 ratio, respectively, (10 mL, 20.1 mmol, 2.25 equiv.) was added over 5 minutes. The reaction mixture was left to react at room temperature for 5 days before being quenched by addition of 1 M HCl, followed by extraction to DCM. The combined organic layers were washed with brine, dried over MgSO₄ and evaporated under reduced pressure. The resulting red oil was purified by flash column chromatography (DCM:CyH 1:1 to 9:1), and the fractions containing the product as a mixture of diastereoisomers were collected and evaporated, yielding a

slightly yellow oil. The oil was dissolved in a small amount of 2-propanol and sonicated; during the sonification, a white powder precipitated. The powder was collected by filtration and dried under vacuo, affording the target diastereoisomer in a 10:1 mixture with an undesired isomer (1.4 g, 3.13 mmol of mixture; 1.27 g, 2.85 mmol, 32% of target diastereoisomer). This mixture was used without further purification in the following reaction.

¹H NMR (401 MHz, $CDCl_3$, target diastereoisomer is, due to blocked rotation, divided to two atropoisomers in 7:3 ratio) δ 8.27 (dt, *J* = 8.1, 1.1 Hz, 0.3H), 7.61–7.55 (m, d, *J* = 8.0 Hz, 0.7H), 7.32–7.03 (m, 3H), 5.08 (s, 0.7H), 4.87 (s, 0.3H), 3.67 (s, 2H), 3.64 (s, 1H), 3.12 (s, 1H), 3.08 (s, 2H), 2.67 (s, 2H), 2.62 (s, 1H), 2.43 (q, *J* = 1.6 Hz, 1H), 1.45–1.26 (m, 1H), 1.07–0.66 (m, 8H).

HRMS (ESI+): calculated for C₂₄H₂₄F₃NO₄Na [M+Na]⁺ 470.1550; found 470.1547.

Synthesis of fulgimides

General procedure for lactone 2 conversion into the inactive Z form of fulgimide

Lactone **2** (50 mg, 0.11 mmol, 1 equiv.) was dissolved in dry DMF (2 mL) and sodium hydride (60% dispersion in paraffin oil, 6.7 mg, 0.17 mmol, 1.4 equiv.) was added in one portion. The reaction was monitored by TLC (DCM, $R_f = 0$). Once the starting material was consumed (typically 1-2 hours), HATU (51 mg, 0.13 mmol, 1.2 equiv.) was added in one portion. After 5 minutes, amine (1.3 equiv.) was added, and the reaction was heated to 50 °C for one hour (general starting conditions, adapted for each amine; conversion assessed by LCMS). Subsequently, the reaction was cooled to RT, and sodium hydride (60%





Chemical Formula: C₂₄H₂₄F₃NO₄ Molecular Weight: 447.454



dispersion in paraffin oil, 9 mg, 0.22 mmol, 2 equiv.) was added in one portion. The reaction was monitored by LCMS. Once the starting material was consumed, the reaction was terminated by adding HATU (25.5 mg, 0.17 mmol, 0.6 equiv.) (typically after 30 mins). The reaction was heated to 50°C for 5 minutes, after which the solvent was evaporated under reduced pressure. The crude product was filtered through a pad of silica (eluted with either DCM or EtOAc, depending on R) and then purified by semi-preparative HPLC (acetonitrile/water, gradient from 5% to pure acetonitrile for 45 mins, then 15 mins pure acetonitrile), sometimes separating into inactive and active isomers (active isomer builds up thermally and under prolonged exposure to day light). If so, it was necessary to keep the inactive isomer in the dark and to evaporate the solvent at RT.

¹H, ¹⁹F and ¹³C NMR spectra of all prepared inactive forms of fulgimides **1a-h** also showed an atypical behavior due to blocked rotation around the methine carbon. To better understand this blocked rotation, cooled NMR studies were conducted on **1d**, revealing both rotamers (for further details on the characteristic signals and on cooling experiments, see section ¹H, ¹⁹F and ¹³C NMR and HRMS Spectra). In some cases, the active and inactive forms were not separated, and their mixture was measured in the given ratios.

¹H NMR

Some ¹H NMR signals were broadened, not displaying sharp splitting. If measured as a mixture, the signals of each form were assigned and distinguished by colors, with the inactive form in purple and the active form in orange.

¹⁹F NMR

As for ¹⁹F NMR spectra, the blocked rotation of the inactive form broadened the trifluoromethyl signal and split into broad singlets in a 2:1 ratio, approximately. Recording the spectrum from a mixture of isomers enabled a clear division of the signals, assigned to each isomer.

$^{13}CNMR$

Because the rotation was blocked and most of carbon atoms were quaternary, most carbon atoms remained invisible. Several carbon atoms gave signal throughout the series of all fulgimides **1a-h** and were thus assigned as usual. Imide substitution was visible in all cases and is highlighted in red.

Fulgimide 1a

Following the general procedure, the target compound was prepared as a yellow solid (32 mg, 0.07 mmol, 61%).

Filtered using DCM. Semi-preparative HPLC yielded 2 fractions: Mixture of the inactive and active forms in a 9:1 ratio and the pure active form.

HRMS (ESI+): calculated for C₂₇H₃₀F₃N₂O₂ [M+H]⁺ 471.2254; found 471.2251.

¹H NMR (401 MHz, CD_2Cl_2 , mixture of inactive and active forms in 9:1 ratio) δ 7.60–6.90 (m, 4H = 3.6H + 0.4H), 3.73 (s, 2.7H), 3.67 (s, 0.3H), 3.62 (td, *J* =



F₃C

Chemical Formula: C₂₇H₂₉F₃N₂O₂ Molecular Weight: 470.536

7.2, 1.5 Hz, 0.2H), 3.46–3.28 (m, 2.7H), 3.00 – 2.85 (m, 0.1H), 2.59 – 2.07 (m, 3H = 2.7H + 0.3H), 1.63 (m, 0.2H), 1.59–1.55 (m, 0.9H), 1.49–1.31 (m, 2H = 1.8H + 0.2H), 1.28–1.00 (m, 7.4H = 7.2H + 0.2H), 0.96 (t, *J* = 7.4 Hz, 0.3H), 0.84 (t, *J* = 7.3 Hz, 2.7H), 0.78–0.43 (m, 1.8H), 0.42–0.19 (m, 0.6H), -0.32–-0.42 (m, 0.1H).

Inactive:

¹³C NMR (101 MHz, CD₂Cl₂, imide part is in red) δ 167.8, 165.9, 137.0, 121.2, 120.1, 109.4, 104.2, 38.0, 30.3, 30.2, 20.5, 17.0, 15.0, 13.8, 11.4, 11.0–8.1 (m).

¹⁹F NMR (377 MHz, CD₂Cl₂) δ -63.38--64.56 (m).

Active:

¹H NMR (401 MHz, CD₂Cl₂) δ 7.34–7.26 (bs, 1H), 7.28 (dt, *J* = 8.2, 0.9 Hz, 1H), 7.16 (ddd, *J* = 8.3, 7.1, 1.2 Hz, 1H), 7.08 (ddd, *J* = 8.1, 7.1, 1.1 Hz, 1H), 3.67 (s, 3H), 3.62 (td, *J* = 7.2, 1.5 Hz, 2H), 2.92 (s, 1H), 2.22 (s, 3H), 1.63 (m, 2H), 1.37 (m, 2H), 0.96 (t, *J* = 7.4 Hz, 3H), 0.72 (m, 2H), 0.28 (m, 6H), -0.37 (s, 1H).

¹⁹F NMR (377 MHz, CD₂Cl₂) δ -56.25.

Fulgimide 1b



Chemical Formula: C₂₉H₂₉F₃N₄O₂ Molecular Weight: 522.572 Following the general procedure, the target compound was prepared as a yellow solid (33 mg, 0.06 mmol, 57%).

Filtered using EtOAc. Semi-preparative HPLC yielded a mixture of the inactive and active forms in a 11:1 ratio.

HRMS (ESI+): calculated for $C_{29}H_{30}F_3N_4O_2$ [M+H]⁺ 523.2315; found 523.2315.

¹H NMR (401 MHz, CD_2Cl_2 , mixture of inactive and active forms in 11:1 ratio) δ 7.65–6.76 (m, 7H = 6.44H + 0.56H), 4.01 (t, *J* = 7.0 Hz, 0.16H), 3.88–3.78 (m, 1.84H), 3.74 (s, 2.76H), 3.67 (s, 0.4H = 0.24H + 0.16H), 3.43 (t, *J* = 6.9 Hz, 1.84H), 3.31 (td, *J* = 8.6, 4.4 Hz, 0.92H), 2.86 (s, 0.08H), 2.59–2.10 (m, 3H = 2.76H + 0.24H), 2.17 (p, *J* = 6.8 Hz, 0.16H), 2.02–1.90 (m, 1.84H), 1.63–1.51 (m, 0.92H), 1.27–0.97 (m, 5.52H), 0.93–0.19 (m, 2.48H = 1.84H + 0.64H), -0.24–0.40 (m, 0.08H).

Active imidazole-CH₂ signal is hidden under a N-CH₃ (3.67 ppm) singlet based on COSY (0.16H)

Inactive:

¹³C NMR (101 MHz, CD₂Cl₂, imide part is in red) δ 167.7, 165.9, 137.5, 137.0, 129.6, 121.4, 120.2, 119.0, 109.5, 104.1, 44.8, 35.5, 30.2, 29.7, 17.2, 15.2, 11.5, 11.1–8.1 (m).

¹⁹F NMR (377 MHz, CD₂Cl₂) δ -63.42--64.73 (m).

Active:

¹⁹F NMR (377 MHz, CD₂Cl₂) δ -56.34.

Fulgimide 1c

Following the general procedure, the target compound was prepared as a yellow solid (35 mg, 0.07 mmol, 65%).

Filtered using EtOAc. Semi-preparative HPLC yielded a mixture of the inactive and active forms in a 10:1 ratio.

HRMS (ESI+): calculated for $C_{27}H_{31}F_3N_3O_2$ [M+H]⁺ 486.2363; found 486.2360.

¹H NMR (401 MHz, CD_2Cl_2 mixture of inactive and active forms in 11:1 ratio) δ 7.60–6.95 (m, 4H = 3.64H + 0.36H), 3.77–3.70 (m, 2.91H = 2.73H + 0.18H),

3.67 (s, 0.27H), 3.55–3.45 (m, 1.82H), 3.41–3.30 (m, 0.91H), 2.97–2.87 (m, 0.09H), 2.54 (t, *J* = 6.8 Hz, 0.18H), 2.51–2.28 (m, 4.55H = 2.73H + 1.82H), 2.28 (s, 0.54H), 2.22 (s, 0.27H), 2.15 (s, 5.46H), 1.62–1.49 (m, 0.91H), 1.23–0.99 (m, 5.46H), 0.86–0.18 (m, 2.54H = 1.82H + 0.72H), -0.28–-0.42 (m, 0.09H).

Active CH_2 is hidden under N-CH₃ (3.73 ppm) singlet based on COSY (0.18H)

Inactive:

¹³C NMR (101 MHz, CD₂Cl₂, imide part is in red) δ 167.8, 165.9, 137.0, 121.2, 120.1, 109.4, 104.2, 56.8, 45.5, 36.1, 30.1, 17.1, 15.0, 11.4, 10.8–8.4 (m).

¹⁹F NMR (377 MHz, CD₂Cl₂) δ -63.18--64.81 (m).

Active:

¹⁹F NMR (377 MHz, CD_2Cl_2) δ -56.38.

Fulgimide 1d

Following the general procedure, the target compound was prepared as a yellow solid (30 mg, 0.06 mmol, 58%).

Filtered using DCM. Semi-preparative HPLC yielded the pure active and inactive forms.

HRMS (ESI+): calculated for $C_{26}H_{24}F_3N_2O_2$ [M+H]⁺ 453.1784; found 453.1782.

Chemical Formula: C₂₆H₂₃F₃N₂O₂ Molecular Weight: 452.477

Inactive:

bemical Formula: CorrHooEoNoO

Chemical Formula: C₂₇H₃₀F₃N₃O₂ Molecular Weight: 485.551





¹H NMR (401 MHz, CD₂Cl₂) δ 7.6–7.22 (bs, 1H), 7.34 (d, *J* = 8.2 Hz, 1H), 7.18 (t, *J* = 7.6 Hz, 1H), 7.06 (t, *J* = 7.6 Hz, 1H), 4.24–4.10 (m, 2H), 3.74 (s, 3H), 3.27 (s, 1H), 2.61–2.21 (m, 3H), 2.17 (t, *J* = 2.6 Hz, 1H), 1.60 (ttd, *J* = 7.9, 6.0, 1.5 Hz, 1H), 1.25–1.00 (m, 6H), 0.96–0.41 (m, 2H).

 13 C NMR (101 MHz, CD₂Cl₂, imide part is in red) δ 166.4, 164.7, 137.0, 121.4, 120.2, 109.5, 104.0, 77.6, 71.0, 30.2, 27.1, 17.2, 15.4, 11.5, 11.4–8.2 (m).

¹⁹F NMR (377 MHz, CD₂Cl₂) δ -63.40--64.62 (m).

Active:

¹H NMR (401 MHz, CD_2Cl_2) δ 7.37–7.21 (bs, 1H), 7.29 (dt, *J* = 8.0, 0.8 Hz, 1H), 7.17 (ddd, *J* = 8.2, 7.0, 1.2 Hz, 1H), 7.08 (ddd, *J* = 8.0, 7.1, 1.1 Hz, 1H), 4.39 (dd, *J* = 2.6, 0.6 Hz, 2H), 3.67 (s, 3H), 2.82 (s, 1H), 2.27 (t, *J* = 2.5 Hz, 1H), 2.23 (s, 3H), 0.83–0.69 (m, 2H), 0.57–0.18 (m, 6H), -0.23–-0.34 (m, 1H).

¹⁹F NMR (377 MHz, CD₂Cl₂) δ -56.53.

Fulgimide 1e

Following the general procedure, the target compound was prepared as a yellow solid (29 mg, 0.06 mmol, 52%).

Filtered using DCM. Semi-preparative HPLC yielded a mixture of the inactive and active forms in a 15:1 ratio.

HRMS (ESI+): calculated for C₂₆H₂₆F₃N₅O₂ [M+Na]⁺ 520.1931; found 520.1931.

¹H NMR (400 MHz, CD₂Cl₂, mixture of inactive and active forms in 15:1 ratio) δ 7.65–6.89 (m, 4H = 3.76H + 0.24H), 3.73 (s, 2.94H = 2.82H + 0.12H), 3.67 (s,

0.18H), 3.49 (t, *J* = 6.9 Hz, 1.88H), 3.40 (t, *J* = 6.6 Hz, 0.12H), 3.37–3.28 (m, 0.94H), 3.27–3.15 (m, 1.88H), 2.90 (s, 0.06H), 2.58–2.07 (m, 3H = 2.82H + 0.18H), 1.94 (dt, *J* = 13.5, 6.7 Hz, 0.12H), 1.74 (p, *J* = 6.8 Hz, 1.88H), 1.61–1.55 (m, 0.94H), 1.23–0.98 (m, 5.64H), 0.88–0.20 (m, 2.36H = 1.88H + 0.48H), -0.32–-0.37 (m, 0.06H).

Active N-CH₂ is hidden under N-CH₃ (3.75 ppm) singlet based on COSY (0.12H)

Inactive:

 13 C NMR (101 MHz, CD₂Cl₂, imide part is in red) δ 167.7, 165.9, 137.0, 121.3, 120.2, 109.5, 104.1, 49.6, 35.7, 30.2, 27.7, 17.1, 15.1, 11.5, 10.4–9.5 (m).

 ^{19}F NMR (376 MHz, CD₂Cl₂) δ -63.31–-64.93 (m).

Active:

 19 F NMR (376 MHz, CD₂Cl₂) δ -56.33.

Fulgimide 1f

Following the general procedure, the target compound was prepared as a yellow solid (34 mg, 0.06 mmol, 52%).

Filtered using DCM. Semi-preparative HPLC yielded a mixture of the inactive and active forms in a 6:1 ratio.

HRMS (ESI+): calculated for $C_{30}H_{27}BrF_3N_2O_2$ [M+H]⁺ 583.1203; found 583.1203.



Chemical Formula: C₃₀H₂₆BrF₃N₂O₂ Molecular Weight: 583.449

¹H NMR (401 MHz, CD_2Cl_2 , mixture of the inactive and active forms in a 6:1 ratio) δ 7.66–6.90 (m, 8H = 6.88H + 1.12H), 4.75 (d, *J* = 1.4 Hz, 0.28H), 4.52 (dd, *J* = 14.8, 14.5 Hz, 1.72H), 3.72 (s, 2.58H), 3.67 (s, 0.42H), 3.36–3.25 (m, 0.86H), 2.90–2.76 (m, 0.14H), 2.60–2.00 (m, 3H = 2.58H + 0.42H), 1.62–1.53 (m, 0.86H), 1.26–0.97 (m, 5.16H), 0.92–0.15 (m, 2.84H = 1.72H + 1.12H), -0.26–0.36 (m, 0.14H).



Chemical Formula: C₂₆H₂₆F₃N₅O₂ Molecular Weight: 497.522

Inactive:

¹³C NMR (101 MHz, CD₂Cl₂, imide part is in red) δ 167.3, 165.6, 137.0, 135.9, 131.9, 130.7, 121.9, 121.3, 120.7, 120.2, 109.5 (d, *J* = 3.6 Hz), 104.1, 41.1, 30.2, 17.2, 15.2, 11.4, 11.1–8.1 (m).

¹⁹F NMR (377 MHz, CD_2Cl_2) δ -63.53--64.60 (m).

Active:

¹⁹F NMR (377 MHz, CD₂Cl₂) δ -56.39.

Fulgimide 1g

The general procedure was followed up to amine addition. Prior to amide coupling, glycine tert-butyl ester hydrochloride (23 mg, 0.13 mmol, 1.2 equiv.) was dissolved in dry DMF (1 mL), and sodium hydride (60% dispersion in paraffin oil, 5.4 mg, 0.13 mmol, 1.2 equiv.) was added; the resulting suspension was then added through a syringe filter to the reaction vessel. The amide coupling reaction mixture was then heated to 60°C for 3 hours. A sodium hydride (60% dispersion in paraffin oil, 9 mg, 0.22 mmol, 2 equiv.) was added, and the reaction mixture of the last step was left standing at RT overnight. After this period, HATU (25.5 mg, 0.17 mmol, 0.6 equiv.) was added, and all subsequent steps followed the general procedure, yielding the target compound as a yellow solid (46 mg, 0.09 mmol, 78%).



Chemical Formula: C₂₉H₃₁F₃N₂O₄ Molecular Weight: 528.572

Filtered using DCM. Semi-preparative HPLC yielded the pure inactive and active forms.

HRMS (ESI+): calculated for C₂₉H₃₂F₃N₂O₄ [M+H]⁺ 529.2309; found 529.2305.

Inactive:

¹H NMR (401 MHz, CD_2Cl_2 , imide part is in red) δ 7.55–7.23 (bs, 1H), 7.32 (d, *J* = 8.1 Hz, 1H), 7.16 (ddd, *J* = 8.1, 7.0, 1.2 Hz, 1H), 7.05 (t, *J* = 7.5 Hz, 1H), 4.10–3.95 (m, 2H), 3.72 (s, 3H), 3.28 (s, 1H), 2.43 (s, 4H), 1.59 (ttd, *J* = 8.0, 6.0, 1.7 Hz, 1H), 1.38 (s, 9H), 1.25–0.99 (m, 6H), 0.94–0.39 (m, 2H).

¹³C NMR (101 MHz, CD₂Cl₂) δ 167.0, 166.3, 165.3, 137.0, 121.3, 120.22, 109.4, 104.1, 82.7, 40.0, 30.2, 28.0, 17.2, 15.3, 11.5, 11.0–9.4 (m).

¹⁹F NMR (377 MHz, CD₂Cl₂) δ -63.38--64.74 (m).

Active:

¹H NMR (401 MHz, CD₂Cl₂) δ 7.35–7.27 (bs, 1H), 7.29 (dt, *J* = 8.2, 0.9 Hz, 1H), 7.17 (ddd, *J* = 8.3, 7.1, 1.2 Hz, 1H), 7.08 (ddd, *J* = 8.0, 7.0, 1.1 Hz, 1H), 4.26 (s, 2H), 3.68 (s, 3H), 2.87 (s, 1H), 2.24 (s, 3H), 1.46 (s, 9H), 0.84–0.67 (m, 2H), 0.60–0.17 (m, 6H), -0.26–0.36 (m, 1H).

¹⁹F NMR (377 MHz, CD₂Cl₂) δ -56.58.

Fulgimide 1h

The general procedure was followed up to amine addition. After this step, the reaction was heated to 100°C for 12h. All subsequent steps followed the general procedure, affording the target compound as a yellow solid (18 mg, 0.03 mmol, 31%).



HRMS (ESI+): calculated for C₃₀H₂₈F₃N₂O₃ [M+H]⁺ 521.2047; found 521.2043.





Inactive:

¹H NMR (401 MHz, CD_2Cl_2) δ 7.70–7.21 (bs, 1H), 7.31 (dt, *J* = 8.2, 0.9 Hz, 1H), 7.17 (t, *J* = 7.6 Hz, 1H), 7.14–7.03 (m, 3H), 6.89 (d, *J* = 8.4 Hz, 2H), 3.78 (s, 3H), 3.71 (s, 3H), 3.35 (s, 1H), 2.66–2.08 (m, 2H), 1.62 (ttd, *J* = 7.9, 6.1, 1.7 Hz, 1H), 1.30–1.02 (m, 6H), 0.96–0.53 (m, 2H).

¹³C NMR (101 MHz, CD₂Cl₂, imide part is in red) δ 167.1, 165.2, 159.7, 137.0, 128.4, 124.7, 121.3, 120.2, 114.4, 109.4, 104.1, 55.8, 30.2, 17.3, 15.3, 11.5, 11.4–8.4 (m).

¹⁹F NMR (377 MHz, CD₂Cl₂) δ -62.51--65.95 (m).

Product of copper-mediated click reaction of 1d and benzyl azide S3

Once **1d** (16 mg, 0.035 mmol, 1 equiv., ratio of inactive and active forms: 9:1) was dissolved in THF/H₂O (1:1, 10 mL), benzyl azide (5.3 mg, 5 uL, 0.04 mmol, 1.1 equiv.) was added to the reaction mixture. The reaction was initiated by adding CuSO₄ \cdot 5 H₂O (4.4 mg, 0.018 mmol, 0.5 equiv.) and sodium ascorbate (7.7 mg, 0.039 mmol, 1.1 equiv.) and heating to 55°C for 15 hours. The solvent was evaporated under reduced pressure, and the crude product was filtered through a pad of silica using EtOAc as solvent. The product was then purified by semi-preparative HPLC, affording 19 mg (0.032 mmol, 92%) of a yellow powder as a mixture of the inactive and active forms in a 85:15 ratio (based on ¹⁹F NMR). The increase of active form is contributed to time spent on daylight, solvent evaporation at 40°C and the temperature needed for reaction.





HRMS (ESI+): calculated for $C_{33}H_{31}F_3N_5O_2$ [M+H]⁺ 586.2424.; found 586.2420.

¹H NMR (401 MHz, CD_2Cl_2 , mixture inactive and active forms in 15:85 ratio) δ 7.61 (s, 0.15H), 7.57–6.89 (m, 9.85H = 8.5H + 1.35H), 5.50 (s, 0.3H), 5.41 (dd, *J* = 14.9, 14.9 Hz, 1.7H), 4.93–4.83 (m, 0.3H), 4.71–4.59 (m, 1.7H), 3.71 (s, 2.55H), 3.66 (s, 0.45H), 3.29 (bs, 0.85H), 2.85 (bs, 0.15H), 2.65–1.80 (m, 2.55H), 2.20 (s, 0.45H), 1.58–1.50 (m, 0.85H), 1.24–0.97 (m, 5.1H), 0.82–0.19 (m, 2.9H = 1.7H + 1.2H), -0.34 (s, 0.15H).

Inactive:

¹³C NMR (101 MHz, CD₂Cl₂, imide part is in red) δ 167.0, 165.5, 143.1, 137.0, 135.3, 129.4, 129.0, 128.4, 128.4, 123.7, 121.3, 120.1, 109.4, 104.0, 33.2, 30.2, 17.2, 15.2, 11.4, 10.9–7.8 (m).

¹⁹F NMR (377 MHz, CD₂Cl₂) δ -63.27--64.82 (m).

Active:

¹⁹F NMR (377 MHz, CD₂Cl₂) δ -56.40.

Photophysical properties and their measurements

Methods

UV/Vis and kinetics. All spectra and kinetics were recorded in HPLC grade acetonitrile. The inactive form of fulgimide (1 mg) was dissolved in acetonitrile (1 mL), and the solution was diluted 10 times and transferred to a fluorescence cuvette equipped with stirring bar, after which the absorbance was adjusted to 0.7. The solution was then subjected to kinetics measurements under stirring and irradiation. Once equilibrium was reached, the ratios of photostationary states were determined via HPLC PDA signals, as described in **Data Evaluation**.

Deactivation. After three cycles of closing and opening, the sample was subjected to deactivation. Sample was transferred from the fluorescence cuvette to a 3-mL vial equipped with a stirring bar. Once 1 equiv. of anthracene was added (5.6 mM solution in acetonitrile), the sample was deoxygenated (15 minutes in sonic bath with balloon filled with nitrogen and outlet), Under stirring, the sample was then simultaneously irradiated with 385- and 626- nm light for 30 minutes. The ratios were then determined via HPLC PDA signals, as described in **Data Evaluation**.

Data Evaluation

Photostationary states (PSS) were determined by HPLC PDA signals (Figure S5). All chromatograms followed the same pattern, only differing in retention times; therefore, only exemplary one of **1b** is shown. The area of the peaks was adjusted by extinction coefficients to calculate the ratio reached in the PSS.

Spectra of pure forms

Inactive forms and active forms. Inactive forms were determined from either the initial spectrum (because only the inactive form was present sometimes) or from a mixture of the active and inactive forms. The active form was primarily produced upon activation, and the wavelength of the isosbestic point was determined. The ratio of HPLC PDA signals measured at the wavelength of the corresponding isosbestic point then directly indicated the molar ratio between isomers in the mixture. As the ratios had in all PSSs less than 5% of inactive form, the contribution of the inactive form was disregarded. Hence, after activation, the spectrum was assigned to pure active forms. The spectrum of the inactive forms was then determined from the initial mixture (molar ratios were determined based on HPLC PDA signals from isosbestic points). According to the Lambert-Beer law, the spectrum of two species is their sum. As such, from the initial spectrum, the spectrum of the active form multiplied by its molar ratio was subtracted, and the resulting spectrum was divided by the molar ratio of the inactive form.

Closed forms were determined from the spectra of PSS. The ratios calculated based on the HPLC PDA signals measured at the wavelength of the respective isosbestic point made it possible to determine the closed form. According to the Lambert-Beer law, the spectrum is a sum of all absorbing species. Thus, the isosbestic point of photoswitching kinetics was used to determine the proportion between E (adjusted by its extinction coefficient at a given wavelength), Z and C. From the spectrum of PSS, the spectra of E and Z multiplied by their molar ratio were subtracted, and the resulting spectrum was divided by the molar ratio of the inactive form.

Figure S6. Chromatogram exported from PDA (absorbance at 473 nm, the corresponding isosbestic point between *E*, *Z* and *C* forms) of LC of **1d** (peaks at 0.5 min correspond to the dead volume of the apparatus), **Red** (1): Mixture after deactivation (**1d** and 1 equivalent of anthracene in degassed acetonitrile irradiated with 385-and 626-nm light for 30 minutes), **Green (2)** PSS in acetonitrile (626 nm) **Blue (3)**: PSS in acetonitrile (420 nm), **Purple (4)**: PSS in acetonitrile (400 nm).



Quantum yields

Quantum yields (Φ) for ring closing and *E*-to-*Z* isomerization were determined using a custom-made LED source with the maximum at 400 nm. Quantum yields for ring opening were determined using a custom-made LED source with the maximum at 626 nm.

Air-saturated solution of fulgimides in acetonitrile ($c \sim 2 \ge 10^{-4}$ M) were irradiated to reach photostationary state at the given wavelength. As only the closed form absorbs at 560 nm and no photo-bleaching was observed, the change in absorbance at 560 nm directly corresponds to the change in closed form concentration.

The quantum yields were calculated from the equations below and averaged from 3 independent measurements $(A_{560 \text{ nm}} = \text{absorbance at 560 nm}, t = \text{time (s)}, V = \text{volume (L)}, \mathcal{E}_{(C, 560 \text{ nm})} = \text{molar extinction coefficient of closed form at 560 nm (M⁻¹cm⁻¹), b = optical path (cm), <math>I_{\text{corr}} = \text{corrected photon flux in acetonitrile (E)}, I_{MeCN} = \text{photon flux in acetonitrile (E)}, c_x = \text{concentration of the observed isomer (M)}, A_{obs} = \text{measured total absorbance at the irradiation wavelength (representing a sum of partial absorbances of each isomer)}, \mathcal{E}_{x,obs} = \text{molar extinction coefficient at the irradiation wavelength (M⁻¹cm⁻¹)}. As absorption spectra of isomers can overlap, it is necessary to recalculate the total photon flux (<math>I_{MeCN}$) to the real photon flux absorbed by the observed isomer (I_{corr}). This is given by the second equation, where the total photon flux was corrected to the photon flux available for the observed isomer at every given time point, as described by K. Börjesson et al.⁴ Next, the quantum yield was calculated from the corrected photon flux by the first equation.

$$\Phi = \frac{\delta A_{560 nm}}{\delta t} \cdot \frac{V}{\varepsilon_{(C, 560 nm,)} \cdot b.I_{corr}}$$

$$I_{corr} = I_{MeCN} \cdot \frac{c_x \cdot \varepsilon_{x,obs}}{A_{obs}} \cdot (1 - 10^{-A_{obs}})$$

$$A_{obs} = c_z \cdot \varepsilon_{z,obs} + c_E \cdot \varepsilon_{E,obs} + c_C \cdot \varepsilon_{C,obs}$$

The photon flux of light sources was determined in toluene, following the work published by Reinfelds et al.² The photon flux was then adjusted from toluene to acetonitrile, based on the difference in their respective refractive indices ($\eta_{MeCN} = 1.3444$, $\eta_{Toluene} = 1.4982$).^{5,6}

$$I_{MeCN} = \frac{(\eta_{MeCN})^2}{(\eta_{toluene})^2} I_{toluene}$$

Z to **E** isomerization. Due to the overlapping absorption spectra of **E** and **Z**, the **Z** to **E** isomerization is directly followed by a much more efficient ring closing (**E** to **C**). Therefore, any active **E** isomer present during the initial stages of the isomerization is immediately transformed to the closed form (**C**). Hence, the increase in the absorbance of **C** at 560 nm directly indicates the decrease in the concentration of the inactive isomer (**Z**). For every given time point, real concentration of inactive isomer (**Z**) was determined and corrected photon flux at 400 nm was calculated. Then, for the initial ten $\delta A_{(560 nm)}/\delta t$ time steps, quantum yield of isomerization was calculated and averaged, which gave the final quantum yield value ($\Phi_{(Z-E, 400 nm)}$).

Ring closing. As this isomerization directly forms the closed form of fulgimide, the increase in the absorbance of *C* at 560 nm directly corresponds to the change in the ratios of the *E* and *C* isomers. For every given time point,

real concentration of active isomer (*E*) was determined and corrected photon flux at 400 nm was calculated. Then, for the initial ten $\delta A_{(560nm)}/\delta t$ time steps, quantum yield of isomerization was calculated and averaged, which gave the final quantum yield value ($\Phi_{(E-C, 400 \text{ nm})}$).

Ring opening. As this isomerization directly leads only to the ring opening from *C* to *E*, the decrease in the absorbance of *C* at 560 nm directly corresponds to the change in the ratios of the isomers. For every given time point, real concentration of closed isomer (*C*) was determined and corrected photon flux at 626 nm was calculated. Then, for the initial ten $\delta A_{(560nm)}/\delta t$ time steps, quantum yield of isomerization was calculated and averaged, which gave the final quantum yield value ($\Phi_{(C-E, 626 \text{ nm})}$).

Absorption spectra

Table S1. Absorption spectra of all three forms of fulgimides **1a-h** and PSS reached after irradiation with 420-nm light in acetonitrile ($c \sim 2 \ge 10^{-4}$ M).



Photoswitching kinetics

All kinetics followed the same pattern, only differing in the time needed to reach PSS as quantum yields differed throughout the series **1a-h**. Therefore, only the photoswitching kinetics of **1b** is shown for illustrative purposes.

Table S2. Kinetics of photoinduced isomerizations of **1b** ($c \sim 2 \ge 10^{-4}$ M) in acetonitrile, **A)** Activation followed by immediate closing (400 nm), **B)** Activation (400 and 626 nm), **C)** Ring closure (400 nm), **D)** Ring opening (626 nm).



Table S3. Kinetics of photoinduced isomerizations of **1b** ($c \sim 1.2 \times 10^{-4}$ M) with anthracene (1 equivalent) in acetonitrile, **A)** Activation followed by immediate closing of the mixture of inactive and active forms (89:11) (400 nm), **B)** Ring closure (400 nm), **C)** Ring opening (626 nm), **D)** Absorption profiles of all forms of fulgimide **1b** with anthracene.



Transient measurements

Figure S7. Exemplary decay traces and the corresponding fit of excited triplet state decay of anthracene ($c \sim 1.7 \times 10^{-4}$ M) in degassed acetonitrile without quencher (black and yellow) and with the active form of fulgimide **1b** ($c \sim 5.6 \times 10^{-4}$ M) as a quencher (red and green).



Γ		А	B1	B2	
Γ	1	Equation	$y = A1^{exp(-x/t1)} + y0$		
	2	уО	0.29611 ± 0.01113	0.20571 ± 0.008	
	3	A1	0.58009 ± 0.011	0.69852 ± 0.01013	
	4	t1	31.9286 ± 1.64266	9.51724 ± 0.43005	
	5	Reduced Chi-Sqr	0.00225	0.00397	
	6	R-Square (COD)	0.94739	0.95031	
	7	Adj. R-Square	0.94697	0.94991	

Figure S8. Linear fit of decays of anthracene as a function of the 1b concentration



	A	B1
1	Equation	y = a + b*x
2	Intercept	22037.08114 ± 5945.67178
3	Slope	1.36823E8 ± 1.79289E7
4	Residual Sum of Squares	1.33016E8
5	Pearson's r	0.9752
6	R-Square (COD)	0.95101
7	Adj. R-Square	0.93468

¹H, ¹⁹F and ¹³C NMR and HRMS Spectra

Characteristic signals of fulgimides



¹**H NMRs**. In its inactive form, free rotation is blocked, broadening hydrogen signals. The signal of the aromatic hydrogen in position 4 is broadened typically between 7.55 and 7.25 ppm. The hydrogens of C-CH₃ are broadened typically between 2.6 and 2.1 ppm. Activation sharpens the peak of hydrogens at position C-CH₃, and the aromatic hydrogens in position 4 are also broadened, albeit to a smaller extent.



Figure S9. ¹H NMR spectrum of the inactive form of compound 1d in CD₂Cl₂

Inactive:

¹H NMR (401 MHz, CD₂Cl₂) δ 7.6 – 7.22 (bs, 1H, Ar-*H*, (4)), 7.34 (d, *J* = 8.2 Hz, 1H, Ar-*H*, (7)), 7.18 (t, *J* = 7.6 Hz, 1H, Ar-*H*, (6)), 7.06 (t, *J* = 7.6 Hz, 1H, Ar-*H*, (5)), 4.24 – 4.10 (m, 2H, N-CH₂), 3.74 (s, 3H, N-CH₃), 3.27 (s, 1H, C-*H*, (16)), 2.61 – 2.21 (m, 3H, C-CH₃), 2.17 (t, *J* = 2.6 Hz, 1H, C-*H*), 1.60 (ttd, *J* = 7.9, 6.0, 1.5 Hz, 1H C-*H*, (17)), 1.25 – 1.00 (m, 6H, 3x CH₂), 0.96 – 0.41 (m, 2H, CH₂).



Figure S10. ¹H NMR spectrum of the active form of compound 1d in CD₂Cl₂

Active:

¹H NMR (401 MHz, CD_2Cl_2) δ 7.37 – 7.21 (bs, 1H, Ar-*H*, (4)), 7.29 (dt, *J* = 8.0, 0.8 Hz, 1H, Ar-*H*, (7)), 7.17 (ddd, *J* = 8.2, 7.0, 1.2 Hz, 1H, Ar-*H*, (6)), 7.08 (ddd, *J* = 8.0, 7.1, 1.1 Hz, 1H, Ar-*H*, (5)), 4.39 (dd, *J* = 2.6, 0.6 Hz, 2H, N-CH₂), 3.67 (s, 3H, N-CH₃), 2.82 (s, 1H, C-*H* (16)), 2.27 (t, *J* = 2.5 Hz, 1H, C-*H*), 2.23 (s, 3H, C-CH₃), 0.83 – 0.69 (m, 2H, CH₂), 0.57 – 0.18 (m, 6H, 3x CH₂), -0.23 – -0.34 (m, 1H, C-*H* (17)).

¹⁹**F NMRs**. Blocked rotation of the inactive form splits the signal of trifluoromethyl group into two broadened singlets in a 2:1 ratio, approximately. Activation restores the sharp singlet (sharp signal around -57 ppm corresponds to the active form, and the broadened singlets around -64 ppm correspond to the inactive form).



Figure S11. Zoomed ¹⁹F NMR spectrum of mixture of inactive and active forms of 1d in CD₂Cl₂

¹³C NMRs. Because the rotation of the inactive form was blocked and most carbon atoms were quaternary, only several carbon atoms were visible in the spectrum. Visible carbon atoms of the fulgide core remained the same through **1a-h**, with minor shifts - two carbonyls (around 167 and 165 ppm), 5 aromatic carbons (around 137 (C8), 121 (C6), 120 (C5), 109 (C7) and 104 (C2) ppm), and signals in aliphatic region (around 30 (C9), 17 (C15), 15 (C16) and 11 (C10) ppm, and a broadened signal around 10 (C18/C19/C20/C21) ppm). The imide substituent was visible in all cases (77.64, 71.00, and 27.11 ppm, for **1d**); in the reported forms, the imide part is indicated in red. Although ¹³C NMRs were measured from mixtures of active and inactive forms, the low concentration of the active form gave almost no signal.



Figure S12. $^{\rm 13}C$ NMR spectrum of the inactive form of 1d in CD_2Cl_2

Inactive:

¹³C NMR (101 MHz, CD₂Cl₂) δ 166.4, 164.7, 137.0, 121.4, 120.2, 109.5, 104.0, 77.6, 71.0, 30.2, 27.1, 17.2, 15.4, 11.5, 11.4 – 8.2 (m).

Cooling experiments

To better understand the system, cooled 1 H, 19 F and 13 C NMR experiments were conducted in CD₃CN on fulgimide **1d**.

¹H NMRs

While cooling down, proton NMRs showed a splitting of both of rotamers in an approximately 2:1 ratio, revealing all signals of the compound with their characteristic splitting. Spectra were recorded at 25 °C and -35°C.



Figure S13. ¹H NMR spectrum of the inactive form (with traces (<5%) of the active form) of **1d** in CD₃CN measured at room temperature



Figure S14. ¹H NMR spectrum of the inactive form (with traces (<5%) of the active form) of **1d** in CD₃CN measured at -35 °C

Major rotamer:

¹H NMR (500 MHz, CD₃CN, 238.15 K) δ 7.36 (d, *J* = 0.8 Hz, 1H, Ar-*H*, (4)), 7.28 (d, *J* = 7.9 Hz, 1H, Ar-*H*, (7)), 7.12 (ddd, *J* = 8.2, 7.1, 1.2 Hz, 1H, Ar-*H*, (6)), 6.98 (ddd, *J* = 8.0, 7.0, 1.0 Hz, 1H, Ar-*H*, (5)), 4.07 (d, *J* = 2.5 Hz, 2H, N-CH₂), 3.73 (s, 3H, N-CH₃), 3.48–3.43 (m, 1H, C-*H* (16)), 2.43 (s, 3H, C-CH₃), 2.42 (t, *J* = 2.5 Hz, 1H, C-*H*), 1.56–1.47 (m, 1H, Δ-CH₂), 1.31–1.18 (m, 2H, Δ-CH₂), 1.11–0.86 (m, 4H, Δ-CH₂), 0.86–0.79 (m, 1H, Δ-CH₂), 0.46–0.38 (m, 1H, C-*H* (17)).

Minor rotamer:

¹H NMR (500 MHz, CD₃CN, 238.15 K) δ 7.50 (d, *J* = 7.8 Hz, 1H, Ar-*H*, (4)), 7.38 (d, *J* = 8.2 Hz, 1H, Ar-*H*, (7)), 7.16 (ddd, *J* = 8.1, 7.0, 1.1 Hz, 1H, Ar-*H*, (6)), 7.07 (ddd, *J* = 7.7, 7.0, 0.9 Hz, 1H, Ar-*H*, (5)), 4.15 (dd, *J* = 22.2, 2.5 Hz, 2H, N-CH₂), 3.69 (s, 3H, N-CH₃), 3.54–3.48 (m, 1H, C-H (16)), 2.46 (t, *J* = 2.5 Hz, 1H, C-*H*), 2.17 (s, 3H, C-CH₃), 1.55–1.46 (m, 1H, Δ-CH₂), 1.30–1.19 (m, 2H, Δ-CH₂), 1.11–0.86 (m, 5H, Δ-CH₂), 0.45–0.38 (m, 1H, C-*H* (17)).

¹⁹F NMRs

Cooling the inactive form split the fluorine signals into two sharp singlets at -15 °C in a 2:1 ratio.







Figure S16. Zoomed-in ¹⁹F NMR spectrum of the inactive form of 1d in CD₃CN measured at -15 °C

Major rotamer:

¹⁹F NMR (470 MHz, CD₃CN, 258.15 K) δ -64.17.

Minor rotamer:

¹⁹F NMR (470 MHz, CD₃CN, 258.15 K) δ -64.75.

¹³C NMRs.

Cooling the inactive form of **1d** split the carbon signals of both rotamers into two sets of carbon signals after a long measurement period. Quaternary carbons 13 and 14 gave no signal, and they were not found by heteronuclear multiple bond correlation (HMBC) analysis either.





Major rotamer:

¹³C NMR (126 MHz, CD₃CN, 238.15 K) δ 167.84 (15), 167.40 (22/23), 164.90 (22/23), 140.58 (1), 137.03 (8), 134.44 (q, J = 2.0 Hz) (11), 127.63 (3), 124.44 (q, J = 275.7 Hz) (12), 121.40 (6), 120.18 (5), 120.02 (d, J = 2.4 Hz) (4), 109.81 (7), 103.76 (2), 78.25 (26), 71.90 (25), 30.17 (9), 27.00 (24), 17.57 (17), 14.34 (q, J = 4.3 Hz) (16), 11.40 (18/19/20/21), 11.30 (10), 11.11 (18/19/20/21), 9.70 (18/19/20/21), 8.24 (18/19/20/21).

Minor rotamer:

¹³C NMR (126 MHz, CD₃CN, 238.15 K) δ 168.14 (15), 167.28 (22/23), 165.15 (22/23), 136.98 (8), 136.72 (1), 135.49 (11), 129.79 (3), 124.13 (q, *J* = 273.8 Hz) (12), 121.29 (6), 120.31 (5), 118.49 (4), 109.94 (7), 103.70 (2), 78.18 (26), 72.00 (25), 30.03 (9), 27.39 (24), 17.74 (17), 14.19 (q, *J* = 3.6 Hz) (16), 11.54 (18/19/20/21), 11.20 (10), 10.94 (18/19/20/21), 9.80 (18/19/20/21), 8.34 (18/19/20/21).

The signal of carbon 4, hidden under CD₃CN, was found by HMBC analysis.

Spectra of fulgimides 1a-h

Fulgimide 1a

Inactive Form (in a 10:1 ratio with the active form):



Figure S18. ¹H NMR spectrum of a mixture of the inactive and active forms in a 10:1 ratio of compound 1a in CD_2Cl_2



Figure S19. ¹⁹F NMR spectrum of a mixture of the inactive and active forms in a 10:1 ratio of compound 1a in CD_2Cl_2 with a zoomed-in inset



Figure S20. $^{\rm 13}C$ NMR spectrum of the inactive form of 1a in CD_2Cl_2

Active Form:



Figure S21. ¹H NMR spectrum of the active form of compound 1a in CD₂Cl₂





Fulgimide 1b

Inactive Form (in a 11:1 ratio with the active form):



Figure S24. ¹H NMR spectrum of a mixture of the inactive and active forms in a 11:1 ratio of compound 1b in CD_2Cl_2



Figure S25. 1 H- 1 H COSY spectrum of a mixture of the inactive and active forms in a 11:1 ratio of compound **1b** in CD₂Cl₂ zoomed in to reveal the presence of a hidden methylene group



Figure S26. ¹⁹F NMR spectrum of a mixture of the inactive and active forms in a 11:1 ratio of compound **1b** in CD_2Cl_2 with a zoomed-in inset



Figure S27. ¹³C NMR spectrum of the inactive form of 1b in CD₂Cl₂



Figure S28. HRMS ESI⁺ spectrum of compound 1b

Fulgimide 1c

Inactive Form (in a 9:1 ratio with the active form):



Figure S29. ¹H NMR spectrum of a mixture of the inactive and active forms in a 9:1 ratio of compound 1c in CD_2Cl_2



Figure S30. 1 H- 1 H COSY spectrum of a mixture of the inactive and active forms in a 9:1 ratio of compound 1c in CD₂Cl₂ zoomed in to reveal the presence of a hidden methylene group



Figure S31. ¹⁹F NMR spectrum of a mixture of inactive and active forms in a 9:1 ratio of compound **1c** in CD₂Cl₂ with a zoomed-in inset



Figure S32. ¹³C NMR spectrum of the inactive form of **1c** in CD₂Cl₂



Figure S33. HRMS ESI⁺ spectrum of compound 1c

Fulgimide 1d

Inactive Form:



Figure S35. 19 F NMR spectrum of the inactive form of compound 1d in CD_2Cl_2 with a zoomed-in inset



Figure S36. ¹³C NMR spectrum of the inactive form of 1d in CD_2Cl_2

Active Form:



Figure S37. ¹H NMR spectrum of the active form of compound 1d in CD₂Cl₂



Figure S38. 19 F NMR spectrum of the active form of compound 1d in CD_2Cl_2 with a zoomed-in inset





Fulgimide 1e

Inactive Form (in a 15:1 ratio with the active form):



Figure S40. ¹H NMR spectrum of a mixture of the inactive and active forms in a 15:1 ratio of compound 1e in CD_2Cl_2



Figure S41. 1 H- 1 H COSY spectrum of a mixture of the inactive and active forms in a 15:1 ratio of compound 1e in CD₂Cl₂ zoomed in to reveal the presence of a hidden methylene group



Figure S42. ¹⁹F NMR spectrum of a mixture of the inactive and active forms in a 15:1 ratio of compound 1e in CD_2Cl_2 with a zoomed-in inset



Figure S43. ¹³C NMR spectrum of the inactive form of 1e in CD₂Cl₂



Figure S44. HRMS ESI⁺ spectrum of compound 1e

Fulgimide 1f

Inactive Form (in a 6:1 ratio with the active form):



Figure S45. $^1\!H$ NMR spectrum of a mixture of the inactive and active forms in a 6:1 ratio of compound 1f in CD_2Cl_2



Figure S46. $^{19}{\rm F}$ NMR spectrum of a mixture of the inactive and active forms in a 6:1 ratio of compound 1f in CD_2Cl_2 with a zoomed-in inset









Fulgimide 1g

Inactive Form:



Figure S49. ^1H NMR spectrum of the inactive form of compound 1g in CD_2Cl_2



Figure S50. ¹⁹F NMR spectrum of the inactive form (with traces of active form) of compound **1g** in CD₂Cl₂ with a zoomed-in inset



Figure S51. ¹³C NMR spectrum of the inactive form of 1g in CD₂Cl₂





Figure S52. ¹H NMR spectrum of the active form of compound 1g in CD₂Cl₂



-56.58



Fulgimide 1h





Figure S56. ¹⁹F NMR spectrum of the inactive form of compound **1h** in CD₂Cl₂ with a zoomed-in inset







Product of copper-mediated click reaction of 1d and benzyl azide S3

Inactive Form (in a 85:15 ratio with the active form):



Figure S59. $^1\!H$ NMR spectrum of a mixture of the inactive and active forms in a 85:15 ratio of compound S3 in CD_2Cl_2



Figure S60. ¹⁹F NMR spectrum of a mixture of the inactive and active forms in a 85:15 ratio of compound S3 in CD_2Cl_2 with a zoomed-in inset







Methyl (2R,3S)-4-(dicyclopropylmethylene)-2-(1,2-dimethyl-1H-indol-3-yl)-5-oxo-2-(trifluoromethyl)tetrahydrofuran-3-carboxylate³ 2

Figure S63. HRMS ESI⁺ spectrum of compound 2

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