## Synthesis, Structure diversity, and Antimicrobial Studies of Ag(I) Complexes with Quinoline-type Ligands

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**Figure S1**. FTIR spectra of the free ligand (**5NO**<sub>2</sub>**Qu**); (upper) and; the [Ag(5NO<sub>2</sub>Qu)<sub>2</sub>]BF<sub>4</sub> (1) complex (lower).





Figure S2 <sup>1</sup>H NMR (upper) and <sup>13</sup>C NMR (lower) spectra of the free ligand 5NO<sub>2</sub>Qu.



**Figure S3** <sup>1</sup>H NMR (upper) and <sup>13</sup>C NMR (lower) spectra of the [Ag(5NO<sub>2</sub>Qu)<sub>2</sub>]BF<sub>4</sub> (**1**) complex.







Figure S4 <sup>1</sup>H NMR (upper) and <sup>13</sup>C NMR (lower) spectra of the free ligand (Qu3CN).

Figure S5<sup>1</sup>H NMR (upper) and <sup>13</sup>C NMR (lower) spectra of the [Ag(Qu3CN)(H<sub>2</sub>O)]<sub>n</sub>(BF<sub>4</sub>)<sub>n</sub> (2) complex.



Figure S6 FTIR spectra of the free ligand (Qu3CN); (upper) and; the [Ag(Qu3CN)(H<sub>2</sub>O)]<sub>n</sub>(BF<sub>4</sub>)<sub>n</sub> (2) polymer (lower).



CSD analysis of C-NO2 ... Ag interactions

Figure S7 FTIR Heat plot of Ag...O distances and N-O...Ag angles, a clear peak  $120^{\circ}-130^{\circ}$  at distances 2.9 Å -3.1 Å

## Method S1

Antimicrobial activity of the two Ag(I) complexes and the free ligand was screened against Grampositive bacteria ; *Staphylococcus aureus* (ATCC 25923), Methicillin-resistant *Staphylococcus aureus* (MRSA ATCC 43300), MRSA (1) clinical isolates and multidrug resistant (MDR) *Enterococcus fecium* (31) clinical isolates and against Gram-negative bacteria including; *Klebsiella pneumonia* (ATCC 700603), *Pseudomonas aeruginosa* (ATCC 29853), *Escherichia coli* (ATCC 25922), *Acinetobacter baumannii* (ATCC 19606), MDR clinical isolates including two *Klebsiella pneumonia* isolates (50 & R124), one *Pseudomonas aeruginosa* (5) isolate, *Proteus mirabilis* and one *Acinetobacter baumannii* (8) in addition to one fungal isolate; *Candida albicans* for determination of their minimum inhibitory concentration according to the CLSI reference standards. The most active compounds were further tested for determination of their minimum bactericidal concentration.

## Determination of minimum inhibitory concentration (MIC)

Investigation of antimicrobial activity of the studied compounds was performed by microbroth dilution assay for determination of minimum inhibitory concentration (MIC). In summary, 100  $\mu$ L of Muller-Hinton broth (MHB) (Oxoid® Limited, Basingstoke, UK) were disseminated in 96 multi-well microtiter plates, followed by the addition of 100  $\mu$ L of the tested compound into the first row of the microtiter plate. Then, from the first to the twelfth well, serial dilutions was performed. Each well received 7  $\mu$ L of freshly prepared bacterial or fungi suspension (1.5 x10<sup>8</sup> cfu/mL). For each bacterial or fungi strain, positive and negative controls were carried out. Plates were incubated for 18-24 hours at 37 °C, with Amoxicillin 1000  $\mu$ g/mL serving as reference standard antibiotic. The MIC was estimated as the minimum concentration that demonstrated no detectable bacterial growth.

## Determination of minimum bactericidal concentrations (MBC)

The minimum bactericidal concentrations (MBC), was determined by inoculating 10  $\mu$ L from each corresponding wells that showed no apparent growth in multi-well microtiter plates onto Muller-Hinton agar (MHA) and incubating the agar plates at 37°C for 24 hours. MBC was determined as the lowest concentration that demonstrated no bacterial growth on the agar surfaces.