

Supporting Information

Rational identification of catalytically promiscuous nitrilase by predicting a unique catalytic triad motif feature through an *in-silico* strategy

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Table S1 Information on sequences used in phylogenetic analysis.

	UniProt ID	Submitted name	Organism	Length
CHT	P32964	Cyanide hydratase	<i>Microdochium sorghi</i>	368
	A9QXE0	Cyanide hydratase	<i>Aspergillus niger</i>	356
	P32963	Cyanide hydratase	<i>Gibberella baccata</i>	355
	C8V9R7	Cyanide hydratase	<i>Emericella nidulans</i>	364
	Q7RVT0	Cyanide hydratase	<i>Neurospora crassa</i>	351
	Q96UG7	Cyanide hydratase	<i>Fusarium solani</i>	363
	P9WEU5	Cyanide hydratase	<i>Stereum hirsutum</i>	371
	E3RV84	Cyanide hydratase	<i>Pyrenophora teres f. teres</i>	366
	G2XQT1	Cyanide hydratase	<i>Botryotinia fuckeliana</i>	371
	I1RP53	Cyanide hydratase	<i>Gibberella zeae</i>	355
	B6HCY4	Cyanide hydratase	<i>Penicillium rubens</i>	358
	A0A2H3E4G0	Cyanide hydratase	<i>Armillaria gallica</i>	356
	Q7Z8M6	Cyanide hydratase	<i>Alternaria brassicicola</i>	363
	Q9P8V3	Cyanide hydratase	<i>Leptosphaeria maculans</i>	356
CP-NLase	V5IPE4	Arylacetonitrilase	<i>Neurospora crassa</i>	327
	C7YS90	Arylacetonitrilase	<i>Fusarium vanettenii</i>	339
	P9WEU6	Arylacetonitrilase	<i>Trametes versicolor</i>	320
	G9AIU0	Aliphatic nitrilase	<i>Rhizobium fredii</i>	308
	Q19A54	Nitrilase	<i>Gibberella moniliformis</i>	320
	G7XTA8	Aromatic nitrilase	<i>Aspergillus kawachii</i>	320
	Q5EG61	NitA	<i>Pseudomonas fluorescens</i>	350
	Q55949	Nitrilase	<i>Synechocystis sp.</i>	341
	C5XY71	Nitrilase	<i>Sorghum bicolor</i>	361
NLase	A2R6M7	Arylacetonitrilase	<i>Aspergillus niger</i>	337
	D4B1Q8	Arylacetonitrilase	<i>Arthroderma benhamiae</i>	335
	K2QXC4	Arylacetonitrilase	<i>Macrophomina phaseolina</i>	344
	G7X8S6	Arylacetonitrilase	<i>Aspergillus kawachii</i>	328
	A0A0P1DJE3	Arylacetonitrilase	<i>Auricularia subglabra</i>	331
	Q2U4D6	Arylacetonitrilase	<i>Aspergillus oryzae</i>	333
	A2RA31	Arylacetonitrilase	<i>Aspergillus niger</i>	328
	G9N4E3	Arylacetonitrilase	<i>Hypocrea virens</i>	329
	Q9UYV8	Nitrilase	<i>Pyrococcus abyssi</i>	262
	Q89GE3	Nitrilase bll6402	<i>Bradyrhizobium diazoefficiens</i>	334
	P20960	Nitrilase, arylacetone-specific	<i>Alcaligenes faecalis</i>	356
	Q500U1	Nitrilase	<i>Pseudomonas syringae</i> pv. <i>Syringae</i>	336
	B6Q5I3	Nitrilase	<i>Talaromyces marneffeii</i>	322
	B6HVR6	Nitrilase	<i>Penicillium rubens</i>	328
	A5DNJ4	Nitrilase	<i>Meyerozyma guilliermondii</i>	317
	Q89PT3	Nitrilase blr3397	<i>Bradyrhizobium diazoefficiens</i>	321
	Q02068	Aliphatic nitrilase	<i>Rhodococcus rhodochrous</i>	383

Table S2 Information on reported crystal structures used in this study.

UniProt ID	PDB code	Organism	Length	Resolution (Å)	Method
Q9UYV8	3ivz	<i>Pyrococcus abyssi</i> GE5	262	1.57	X-RAY DIFFRACTION
P46011	6i00	<i>Arabidopsis thaliana</i>	361	3.4	ELECTRON MICROSCOPY
Q55949	3wuy	<i>Synechocystis</i> sp. PCC 6803 substr. Kazusa	349	3.1	X-RAY DIFFRACTION
Q5EG61	6zby	<i>Pseudomonas</i> <i>fluorescens</i>	350	3.1	ELECTRON MICROSCOPY

Table S3 Information on potential CP-NLase candidates.

UniProt ID	Submitted name	Organism	Length
A0A1W2TCV9	Putative aliphatic nitrilase	<i>Rosellinia necatrix</i>	323
A0A378TBT1	Nitrilase 2	<i>Mycolicibacterium tokaiense</i>	316
A0A182BSR4	Nitrilase	<i>Gordonia terrae</i>	366
A0A653FFS7	Nitrilase	<i>Mycolicibacterium smegmatis</i>	341
A0A385ZDY9	Nitrilase	<i>Streptomyces griseorubiginosus</i>	339
Q1W2L4	Nitrilase	<i>Acidovorax facilis</i>	369
S5PQ64	Nitrilase	<i>Gordonia terrae</i>	366
H9NN54	Nitrilase	<i>Nocardia globerula</i>	366
G1FTR4	Nitrilase	<i>Rhodococcus</i> sp. BX2	366
F6GDZ2	Nitrilase	<i>Lacinutrix</i> sp.	316
F4CZ44	Nitrilase	<i>Pseudonocardia dioxanivorans</i>	349
A0A085F6G6	Nitrilase	<i>Bosea</i> sp. LC85	323
C7CG56	Putative nitrilase	<i>Methylobacterium extorquens</i>	305
B6R8T8	Nitrilase, putative	<i>Pseudovibrio</i> sp. JE062	331
B6R8T7	Nitrilase	<i>Pseudovibrio</i> sp. JE062	332
A0A0U1TDG4	Regioselective nitrilase	<i>Acidovorax facilis</i>	372
A0A7Y9DS80	Nitrilase	<i>Actinomyces sp. corticicola</i>	346
A0A839SAC7	Nitrilase	<i>Mucilaginibacter gotjawali</i>	318
G0LAG6	Nitrilase	<i>Zobellia galactanivorans</i>	316
A0A099KNW3	Nitrilase	<i>Thalassotalea</i> sp. ND16A	324
A0A517WTX7	Nitrilase	<i>Gimesia aquarii</i>	347
A0A7W8A8A8	Nitrilase	<i>Nonomuraea endophytica</i>	306
A0R378	Nitrilase	<i>Mycolicibacterium smegmatis</i>	341
A0A7W0CD04	Nitrilase	<i>Nonomuraea soli</i>	303
F0RCY1	Nitrilase	<i>Cellulophaga lytica</i>	316
Q0AQN1	Nitrilase	<i>Maricaulis maris</i>	310

E6X4J4	Nitrilase	<i>Cellulophaga algicola</i>	316
A0A517W1X2	Nitrilase	<i>Gimesia aquarii</i>	357
A0A2Z4LQF7	Nitrilase	<i>Muricauda aurantiaca</i>	317
L2G5Z2	Arylacetonitrilase	<i>Colletotrichum fructicola gloeosporioides</i>	372
A0A2N6NA38	Arylacetonitrilase	<i>Beauveria bassiana</i>	320
A0A329T4D1	Arylacetonitrilase	<i>Phytophthora cactorum</i>	341
A0A4T0WFD2	Arylacetonitrilase	<i>Colletotrichum higginsianum</i>	371
A0A6A4AXL9	Arylacetonitrilase	<i>Phytophthora rubi</i>	309
A0A6A3GCP7	Arylacetonitrilase	<i>Phytophthora rubi</i>	289
A0A6A3GEF2	Arylacetonitrilase	<i>Phytophthora rubi</i>	309
A0A177DUN2	Arylacetonitrilase	<i>Alternaria alternata</i>	342
A0A4Q4S236	Arylacetonitrilase	<i>Alternaria tenuissima</i>	343
A0A420QY53	Arylacetonitrilase	<i>Fusarium oxysporum</i>	326
A0A420MER3	Arylacetonitrilase	<i>Fusarium oxysporum</i>	326
A0A3L6MVK0	Arylacetonitrilase	<i>Fusarium oxysporum f. sp. Cepae</i>	326
A0A6V8R3E7	Arylacetonitrilase	<i>Trichoderma asperellum</i>	309

Table S4 The sequence similarity of putative CP-arylacetonitrilase candidates.

UniProt ID	1	2	3	4	5	6	7	8	9	10	11	12	13
1 L2G5Z2		91.4	24.73	25	21.77	25	25	27.69	14.78	27.69	27.69	26.88	22.04
2 A0A4T0WFD2	91.4		24.19	24.19	22.04	24.73	24.73	27.69	16.13	27.69	27.69	27.96	23.12
3 A0A4Q4S236	24.73	24.19		97.58	30.91	28.76	29.03	27.42	27.96	27.96	27.96	24.19	27.42
4 A0A177DUN2	25	24.19	97.58		30.91	29.03	29.3	27.69	28.23	28.23	28.23	24.46	27.69
5 A0A329T4D1	21.77	22.04	30.91	30.91		29.57	29.57	26.34	25.27	26.34	26.34	25	42.74
6 A0A6A3GEF2	25	24.73	28.76	29.03	29.57		99.46	34.68	31.99	34.41	34.41	35.75	48.39
7 A0A6A4AXL9	25	24.73	29.03	29.3	29.57	99.46		34.41	31.99	34.14	34.14	35.75	48.12
8 A0A420QY53	27.69	27.69	27.42	27.69	26.34	34.68	34.41		30.65	99.46	99.46	56.72	32.26
9 A0A6A3GCP7	14.78	16.13	27.96	28.23	25.27	31.99	31.99	30.65		30.65	30.65	35.22	31.72
10 A0A3L6MVK0	27.69	27.69	27.96	28.23	26.34	34.41	34.14	99.46	30.65		100	56.45	31.72
11 A0A420MER3	27.69	27.69	27.96	28.23	26.34	34.41	34.14	99.46	30.65	100			31.72
12 A0A6V8R3E7	26.88	27.96	24.19	24.46	25	35.75	35.75	56.72	35.22	56.45	56.45	100	30.91
13 A0A2N6NA38	22.04	23.12	27.42	27.69	42.74	48.39	48.12	32.26	31.72	31.72	31.72	30.91	

Table S5 The enantioselectivity of reported NLase or NHase toward mandelonitrile.

Enzyme	Organisms	ee (%)	References
Nitrile hydratase	<i>Rhodococcus rhodochrous</i> ATCC BAA-87	R, 81	1
Nitrile hydratase	<i>Klebsiella oxytoca</i> 38.1.2	S, 89	2
Nitrile hydratase	<i>Rhodococcus rhodochrous</i> J1	S, 52.6	3
Mutant of nitrile hydratase	<i>Rhodococcus rhodochrous</i> J1	S, 96.8	3

NLase	<i>Neurospora crassa</i> OR74A	S, 51 (amide ratio, 15%)	4
NLase	<i>P. fluorescens</i> EBC191	S, 84 (amide ratio, 19%)	5
NitFO	<i>Fusarium oxysporum</i>	R, 90.3 (amide ratio, 16%)	This study
NitAT	<i>Alternaria tenuissima</i>	S, 96.5 (amide ratio, 28%)	This study

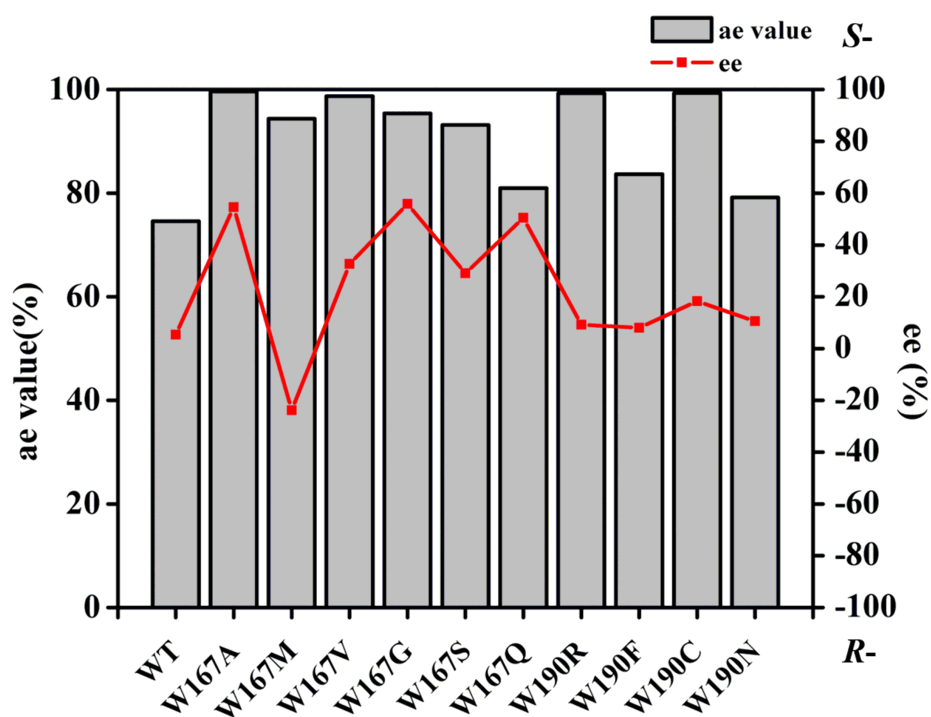


Fig. S1 The enantioselectivity of beneficial mutants.

Table S6 The substrate tolerance of NitPC-W167A toward mandelonitrile.

Substrate (mM)	Reaction time (h)	Yield (%)
50	4	100
100	16	100
150	24	10.2
200	24	4.3

References

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