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## **Supporting Information**

# Palladium-catalyzed direct carbonylation of thiophenes and furans under CO/CO<sub>2</sub>-binary conditions leading to carboxylic acids

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	$\left< \begin{array}{c} S \\ \end{array} \right> + CO (20 \text{ atm})  \frac{Pd(OAc)_2}{solvent}$	2 (10 mol%), addtive 3 mL), 100 °C, 20 h	ОН
	<b>1j</b> (3 mmol)	2j	
Entry	Solvent	Additive	Yield <b>2i</b> (%) <sup><i>a</i></sup>
1	Glacial AcOH	-	N. D.
2	Glacial AcOH	NaOAc (1 equiv.)	N. D.
3	Glacial AcOH	H <sub>2</sub> O (100 μL)	N. D.
4	Glacial AcOH	PivOH (30 mol%)	N. D.
5	Glacial AcOH	K <sub>2</sub> CO <sub>3</sub> (30 mol%)	N. D.
6	Glacial AcOH (CO <sub>2</sub> bubbling)	-	8
7	Glacial AcOH ( $N_2 + O_2$ (79 : 21) bubbling)	-	N. D.

Table S1. Reaction Conditions for Pd-mediated Carbonylation of Thiophene 1j

<sup>a</sup>Yields were determined by <sup>1</sup>H NMR spectroscopy based on **1j**.

	Table S2.	Optimization	of Reaction	Conditions	for Pd-catal	yzed	Carbony	lation of	of 11	1
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CI	+ CO (30 atm) + CO <sub>2</sub> (5 atm)	Pd(OAc) <sub>2</sub> (x mol%) <i>p</i> -BQ (1.5 equiv.) AcOH (3 mL), 100 °C, time	CI S O OH
<b>1h</b> (3 mmc	l)		2h
Entry	Pd cat. (mol%)	Time (h)	Yield <b>2e</b> (%) <sup><i>a</i></sup>
1	1	20	45
2	1	72	66
3	5	20	87

"Yields were determined by <sup>1</sup>H NMR spectroscopy.

Table S3. Optimization of Reaction Conditions for Pd-Catalyzed Carbonylation of Furfuryl Acetate  $3a^{a}$ 

	CO(x  atm) +	$CO_{1}$ (5 atm)	Pd(OAc) <sub>2</sub> (y mol%) <i>p</i> -BQ (z equiv.)	
AcO´ \\// +	CO (x atm) +	$OO_2$ (5 atm)	AcOH (3.0 mL), 100 °C, 20 h	AcO OH
<b>3a</b> (3 mmol)				4a

Entry	CO (atm)	CO <sub>2</sub> (atm)	$Pd(OAc)_2 (mol\%)$	<i>p</i> -BQ (equiv.)	Yield <b>4a</b> (%)
1	10	1	10	3.0	41
2	20	1	10	3.0	60
3	50	1	10	3.0	90
4	30	5	1	1.5	55
5	30	5	5	1.5	76

6	30	5	10	1.5	78
7	50	5	5	1.5	40
8	30	5	5	3.0	92
9	30	-	5	3.0	70

"Yields were determined by <sup>1</sup>H NMR spectroscopy.

Table S4. Optimization of Reaction Conditions for Pd-catalyzed Carbonylation of 2-Ethylfuran 3b<sup>a</sup>

	+ CO (30 atm) + CO <sub>2</sub> (5 atm) AcC	Pd(OAc) <sub>2</sub> (x mol%) <i>p</i> -BQ (3 equiv.) DH (3 mL), 100 °C, time	о он
<b>3b</b> (3 mmo	1)		4b
Entry	Pd cat. (mol%)	Time (h)	Yield <b>4b</b> (%)
1	5	20	29
2	15	20	54
3	15	48	(78)
$4^b$	15	48	39

<sup>a</sup>Yields were determined by <sup>1</sup>H NMR spectroscopy (isolated yield). <sup>*b*</sup>The reaction was conducted under CO (30 atm).

## General Remarks

Unless otherwise stated, all starting materials and catalysts were purchased from commercial sources and used without further purification. All solvents were distilled and degassed with nitrogen before use. Thiophene derivatives ( $1e^{S1} 1f^{S2}$ ) and furan derivatives ( $3f^{S3} 3g^{S4}$ ) were prepared according to the previously reported procedures. <sup>1</sup>H NMR spectra were recorded on a JEOL JNM-ECS400 (400 MHz) FT NMR system, a JEOL JNM-ECX400 (400 MHz) FT NMR system, or a Bruker BioSpin Ascend 400 spectrometer (400 MHz) with Me<sub>4</sub>Si as an internal standard. <sup>13</sup>C{<sup>1</sup>H} NMR spectra were recorded on a JEOL JNM-ECX400 (100 MHz) FT NMR, a JEOL JNM-ECS400 (100 MHz) FT NMR system, or a Bruker BioSpin Ascend 400 spectrometer (100 MHz). The <sup>1</sup>H NMR yields of the crude mixture were determined using 1,3,5-trioxane as the internal standard. IR spectra were recorded on a Jasco FT/IR-410, and reported in wavenumbers (cm<sup>-1</sup>).

## Experimental Procedure for Pd-Catalyzed Carbonylation of Thiophenes (Table 2).

Thiophene **1** (3 mmol),  $Pd(OAc)_2$  (1–5 mol%), *p*-benzoquinone (1.5 equiv. 486.4 mg), and AcOH (3.0 mL, bubbled with CO<sub>2</sub> for 30 min) were sequentially added to a 50 mL stainless steel autoclave with a magnetic stirring bar under a N<sub>2</sub> atmosphere. The vessel was purged five times with CO<sub>2</sub> and then charged with CO<sub>2</sub> (5 atm) and CO (30 atm), respectively. The reaction was conducted with magnetic stirring at 100 °C for 20–72 h. The resulting mixture was filtered through silica-gel with CH<sub>2</sub>Cl<sub>2</sub>, and

concentrated under reduced pressure. The residue was basified by sat. NaHCO<sub>3</sub> aq. (pH = 9), and the solvent was removed under reduced pressure. Then, the residue was washed with Et<sub>2</sub>O (30 mL), and acidified with 10% HCl aq. (pH = 1). The solution was concentrated under reduced pressure. The residue was dissolved in acetone (10 mL), and filtered. Finally, the filtrate was concentrated to give the corresponding carboxylic acids **2**.

5-Ethylthiophene-2-carboxylic acid (2a) (CAS no. 23229-72-3)<sup>S5</sup>

Brown solid, 403.0 mg, 86%; <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>):  $\delta$  11.08 (br, 1H), 7.73 (d, J = 3.7 Hz, 1H), 6.84 (d, J = 3.2 Hz, 1H), 2.89 (q, J = 7.5 Hz, 2H), 1.34 (t, J = 7.6 Hz, 3H); <sup>13</sup>C{<sup>1</sup>H} NMR (100 MHz, CDCl<sub>3</sub>):  $\delta$  168.1, 157.7, 135.5, 129.8, 125.1, 24.1, 15.7.

5-Methylthiophene-2-carboxylic acid (2b) (CAS no. 1918-79-2)<sup>S6</sup>



Brown solid, 358.5 mg, 84%; <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>):  $\delta$  10.98 (br, 1H), 7.71 (d, J = 3.6 Hz, 1H), 6.80 (d, J = 3.2 Hz, 1H), 2.54 (s, 3H); <sup>13</sup>C{<sup>1</sup>H} NMR (100 MHz, CDCl<sub>3</sub>)  $\delta$  168.0, 150.1, 135.7, 130.2, 126.9, 16.0.

5-Pentylthiophene-2-carboxylic acid (2c) (CAS no. 63068-75-7)<sup>S7</sup>

<sup>n</sup>Pen S OH

Brown solid, 475.8 mg, 80%; <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>):  $\delta$  9.58 (br, 1H), 7.72 (d, *J* = 3.7 Hz, 1H), 6.82 (d, *J* = 3.7 Hz, 1H), 2.85 (t, *J* = 7.6 Hz, 2H), 1.71 (m, 2H), 1.35 (m, 4H), 0.90 (m, 3H); <sup>13</sup>C{<sup>1</sup>H} NMR (100 MHz, CDCl<sub>3</sub>):  $\delta$  167.7, 156.1, 135.3, 129.8, 125.6, 31.2, 31.1, 30.6, 22.3, 13.9.

5-(2-Ethylhexyl)thiophene-2-carboxylic acid (2d) (CAS no. 1810058-94-6)<sup>S8</sup>



Brown solid, 653.5 mg, 91%; <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>):  $\delta$  10.07 (br, 1H), 7.73 (d, *J* = 3.6 Hz, 1H), 6.80 (d, *J* = 3.6 Hz, 1H), 2.79 (d, *J* = 6.8 Hz, 2H), 1.64-1.61 (m, 1H), 1.38-1.28 (m, 8H), 0.91-0.88 (m, 6H); <sup>13</sup>C{<sup>1</sup>H} NMR (100 MHz, CDCl<sub>3</sub>):  $\delta$  167.8, 155.0, 135.4, 130.0, 126.6, 41.6, 34.6, 32.4, 28.9, 25.6, 23.0, 14.2, 10.9.

5-Benzylthiophene-2-carboxylic acid (2e) (CAS no. 13132-16-6)<sup>S9</sup>

Gray solid, 655.8 mg, 100%; <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>): δ 10.30 (br, 1H), 7.70 (m, 1H), 7.30–7.29 (m, 2H), 7.23 (m, 3H), 6.80 (m, 1H), 4.13 (s, 2H); <sup>13</sup>C{<sup>1</sup>H} NMR (100 MHz, CDCl<sub>3</sub>): δ 167.9, 154.3, 139.0, 135.5, 130.9, 128.8, 128.7, 127.0, 126.5, 36.7.

5-((Benzyloxy)methyl)thiophene-2-carboxylic acid (2f) (CAS no. 1481274-88-7)



Dark brown solid, 618.0 mg, 83%; <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>):  $\delta$  10.36 (br, 1H), 7.77 (d, J = 3.6 Hz, 1H), 7.37-7.36 (m, 4H), 7.33-7.29 (m, 1H), 7.00 (d, J = 3.6 Hz, 1H), 4.71 (s, 2H), 4.59 (s, 2H); <sup>13</sup>C{<sup>1</sup>H} NMR (100 MHz, CDCl<sub>3</sub>)  $\delta$  167.8, 150.9, 137.4, 135.1, 132.4, 128.7, 128.1, 128.0, 126.6, 72.4, 66.7.

5-Phenylthiophene-2-carboxylic acid (2g) (CAS no. 19163-24-7)<sup>S10</sup>



Light brown solid, 540.3 mg, 88%; <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>):  $\delta$  7.87 (d, *J* = 4.1 Hz, 1H), 7.66 (d, *J* = 7.2 Hz, 2H), 7.45-7.38 (m, 3H), 7.33 (d, *J* = 3.6 Hz, 1H); <sup>13</sup>C{<sup>1</sup>H} NMR (100 MHz, CDCl<sub>3</sub>):  $\delta$  167.6, 153.1, 136.1, 133.3, 131.2, 129.25, 129.16, 126.4, 124.0.

5-Chlorothiophene-2-carboxylic acid (2h) (CAS no. 24065-33-6)<sup>S11</sup>

Brown solid, 336.7 mg, 69%; <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>):  $\delta$  9.52 (br, 1H), 7.69 (d, *J* = 4.1 Hz, 1H), 6.97 (d, *J* = 4.1 Hz, 1H); <sup>13</sup>C{<sup>1</sup>H} NMR (100 MHz, CDCl<sub>3</sub>):  $\delta$  167.9, 140.4, 135.9, 132.0, 128.8.

5-Bromothiophene-2-carboxylic acid (2i) (CAS no. 7311-63-9)<sup>S12</sup>



Dark gray solid, 433.5 mg, 70%; <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>):  $\delta$  9.68 (br, 1H), 7.63 (d, *J* = 4.0 Hz, 1H), 7.10 (d, *J* = 4.0 Hz); <sup>13</sup>C{<sup>1</sup>H} NMR (100 MHz, CDCl<sub>3</sub>):  $\delta$  166.5, 136.3, 133.9, 131.3, 122.3.

2-Thiophenylcarboxylic acid (2j) (CAS no. 527-72-0)<sup>S13</sup>



Brown solid, 211.6 mg, 55%; <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>):  $\delta$  7.91 (dd, J = 3.9, 1.1 Hz, 1H), 7.66 (dd, J = 5.0, 1.4 Hz, 1H), 7.15 (dd, J = 5.0, 4.1 Hz, 1H); <sup>13</sup>C{<sup>1</sup>H} NMR (400 MHz, CDCl<sub>3</sub>):  $\delta$  167.7, 135.1, 134.1, 132.9, 128.2.

Thieno[3,2-b]thiophene-2-carboxylic acid (2k) (CAS no. 1723-27-9)<sup>S14</sup>



Light brown solid, 311.3 mg, 56%; <sup>1</sup>H NMR (400 MHz, DMSO-*d*<sub>6</sub>):  $\delta$  8.12 (s, 1H), 7.93 (d, *J* = 5.3 Hz, 1H), 7.52 (d, *J* = 5.3 Hz, 1H); <sup>13</sup>C{<sup>1</sup>H} NMR (100 MHz, DMSO-*d*<sub>6</sub>):  $\delta$  163.9, 143.7, 139.1, 136.1, 133.5, 126.6, 120.8.

3-Methylthiophene-2-carboxylic acid (A) and 4-methylthiophene-2-carboxylic acid (B) (2l)



Brown solid, 322.0 mg, 75%; [compound A] (CAS no. 23806-24-8)<sup>S15</sup>: <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>):  $\delta$  10.1 (br, 1H), 7.48 (d, *J* = 5.0 Hz, 1H), 6.95 (d, *J* = 5.0 Hz, 1H), 2.58 (s, 3H); [compound B] (CAS no. 14282-78-1)<sup>S15</sup>: <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>):  $\delta$  10.1 (br, 1H), 7.69 (d, *J* = 1.3 Hz, 1H), 7.24-7.23 (m, 1H), 2.30 (s, 3H).

## 3-Hexylthiophene-2-carboxylic acid (A) and 4-hexylthiophene-2-carboxylic acid (B) (2m)



Reddish brown solid, 551.9 mg, 87%; [compound A] (CAS no. 214409-28-6)<sup>S16</sup>: <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>):  $\delta$  11.6 (br, 1H), 7.47 (d, J = 5.0 Hz, 1H), 6.98 (d, J = 5.0 Hz, 1H), 3.02 (t, J = 7.8 Hz, 2H), 1.67–1.59 (m, 2H), 1.39–1.26 (m, 4H), 0.91–0.87 (m, 3H); [compound B] (CAS no. 1261539-24-5)<sup>S8</sup>: <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>):  $\delta$  11.6 (br, 1H), 7.72 (d, J = 1.4 Hz, 1H), 7.23 (d, J = 1.4 Hz. 1H), 2.61 (t, J = 7.7 Hz, 2H), 1.67–1.59 (m, 2H), 1.39–1.26 (m, 4H), 0.91–0.87 (m, 3H).

3-Chlorothiophene-2-carboxylic acid (A) and 4-chlorothiophene-2-carboxylic acid (B) (2n)



Light Brown solid, 343.9 mg, 71%; [compound A] (CAS no. 59337-89-2)<sup>S17</sup>: <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>):  $\delta$  7.58 (d, *J* = 5.0 Hz, 1H), 7.07 (d, *J* = 5.4 Hz, 1H); [compound B] (CAS no. 59614-95-8)<sup>S18</sup>: <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>):  $\delta$  7.73-7.73 (m, 1H), 7.43-7.43 (m, 1H)

3-Bromothiophene-2-carboxylic acid (A) and 4-bromothiophene-2-carboxylic acid (B) (20)



Light gray solid, 482.3 mg, 78%; [compound A] (CAS no. 7311-64-0)<sup>S19</sup>: <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>):  $\delta$  7.56 (d, J = 5.2 Hz, 1H), 7.15 (d, J = 5.2 Hz, 1H); [compound B] (CAS no. 16694-18-1)<sup>S20</sup> <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>):  $\delta$  7.79 (d, J = 1.5 Hz, 1H), 7.54 (d, J = 1.5 Hz, 1H).

# Experimental Procedure for Gram-scale Synthesis of 2a via Pd-catalyzed Direct Carbonylation (Table 2).

2-Ethylthiophene **1a** (10 mmol, 1.12 g), Pd(OAc)<sub>2</sub> (1 mol%, 22.5 mg), *p*-benzoquinone (1.5 equiv. 1.62 g), and AcOH (10 mL, bubbled with CO<sub>2</sub> for 30 min) were sequentially added to a 50 mL stainless steel autoclave with a magnetic stirring bar under a N<sub>2</sub> atmosphere. The vessel was purged three times with CO<sub>2</sub> and then charged with CO<sub>2</sub> (5 atm) and CO (30 atm), respectively. The reaction was conducted with magnetic stirring at 100 °C for 20 h. After the reaction was completed, the resulting mixture was filtered through silica-gel with CH<sub>2</sub>Cl<sub>2</sub> and concentrated under reduced pressure. The residue was basified by sat. NaHCO<sub>3</sub> aq. (pH = 9), and the solution was removed in vacuo. The residue was washed with Et<sub>2</sub>O (50 mL), then acidified with 10% HCl aq. (pH = 1). The solution was concentrated under reduced pressure. The residue was dissolved in acetone (20 mL), and filtered. Finally, the filtrate was concentrated to give pure **2a** in 83% isolated yield (brown solid, 1.3 g).

## Experimental Procedure for Pd-Catalyzed Carbonylation of Furans (Table 3).

Furan **3** (3 mmol),  $Pd(OAc)_2$  (5–15 mol%), *p*-benzoquinone (3 equiv. 972.8 mg), and AcOH (3.0 mL, bubbled with CO<sub>2</sub> for 30 min) were sequentially added to a 50 mL stainless steel autoclave with a magnetic stirring bar under a N<sub>2</sub> atmosphere. The vessel was purged three times with CO<sub>2</sub> and then charged with CO<sub>2</sub> (5 atm) and CO (30 atm), respectively. The reaction was conducted with magnetic stirring at 100 °C for 20–48 h. After the reaction was completed, the resulting mixture was filtered

through silica-gel with  $CH_2Cl_2$  and concentrated under reduced pressure. The residue was basified by sat. NaHCO<sub>3</sub> aq. (pH = 9), and the solution was removed in vacuo. The residue was washed with Et<sub>2</sub>O (30 mL), then acidified with 10% HCl aq. (pH = 1). The solution was concentrated under reduced pressure. The residue was disollved in acetone (10 mL), and filtered. Finally, the filtrate was concentrated to give the corresponding carboxylic acids **4**.

5-Ethylfuran-2-carboxylic acid (4b) (CAS no. 56311-37-6)<sup>S21</sup>

Dark brown solid, 326.3 mg, 78%; <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>):  $\delta$  10.1 (br, 1H), 7.26 (d, *J* =3.6 Hz, 1H), 6.18 (d, *J* = 3.4 Hz, 1H), 2.75 (q, *J* = 7.6 Hz, 2H), 1.29 (t, *J* = 7.6 Hz, 3H); <sup>13</sup>C{<sup>1</sup>H} NMR (100 MHz, CDCl<sub>3</sub>):  $\delta$  164.1, 163.7, 142.0, 121.6, 107.4, 21.8, 11.8.

5-Propylfuran-2-carboxylic acid (4c) (CAS no. 14497-25-7)<sup>S22</sup>



Brown solid, 384.7 mg, 83%; <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>):  $\delta$  8.85 (br, 1H), 7.25 (d, *J* = 3.4 Hz, 1H), 6.18 (d, *J* = 3.4 Hz, 1H), 2.69 (t, *J* = 7.5 Hz, 2H), 1.78–1.68 (m, 2H), 0.98 (t, *J* = 7.4 Hz, 3H); <sup>13</sup>C{<sup>1</sup>H} NMR (100 MHz, CDCl<sub>3</sub>):  $\delta$  163.6, 162.8, 142.1, 121.6, 108.2, 30.3, 21.1, 13.7.

5-Butylfuran-2-carboxylic acid (4d) (CAS no. 67238-23-7)<sup>S23</sup>

Brown solid, 400.1 mg, 79%; <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>):  $\delta$  8.91 (br, 1H), 7.25 (d, *J* = 3.4 Hz, 1H), 6.17 (d, *J* = 3.4 Hz, 1H), 2.71 (t, *J* = 7.7 Hz, 2H), 1.68 (m, 2H), 1.38 (m, 2H), 0.93 (t, *J* = 7.4 Hz, 3H); <sup>13</sup>C{<sup>1</sup>H} NMR (100 MHz, CDCl<sub>3</sub>):  $\delta$  163.7, 163.0, 142.0, 121.6, 108.1, 29.7, 28.1, 22.2, 13.7.

## 5-Phenylfuran-2-carboxylic acid (4f) (CAS no. 52938-97-3)<sup>S24</sup>



Brown solid, 203.4 mg, 36%; <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>):  $\delta$  9.98 (br, 1H), 7.80 (d, *J* = 7.5 Hz, 2H), 7.45–7.35 (m, 4H), 6.78 (d, *J* = 2.7 Hz, 1H); <sup>13</sup>C{<sup>1</sup>H} NMR (100 MHz, CDCl<sub>3</sub>) :  $\delta$  163.7, 158.8, 142.8, 129.31, 129.25, 128.9, 125.1, 122.3, 107.3.

5-((Benzyloxy)methyl)furan-2-carboxylic acid (4g) (CAS no. 42890-01-7)<sup>S25</sup>



Light brown solid, 574.4 mg, 82%; <sup>1</sup>H NMR (400 MHz, DMSO-*d*<sub>6</sub>):  $\delta$  7.38–7.28 (m, 5H), 7.18 (d, *J* = 3.3 Hz, 1H), 6.63 (d, *J* = 3.3 Hz, 1H), 4.59 (m, 4H); <sup>13</sup>C{<sup>1</sup>H} NMR (100 MHz, CDCl<sub>3</sub>) :  $\delta$  163.4, 157.6, 143.9, 137.3, 128.9, 128.7, 128.3, 121.7, 112.4, 73.9, 65.7.

5-Bromofuran-2-carboxylic acid (4h) (CAS no. 585-70-6)<sup>S26</sup>



Dark brown solid, 497.2 mg, 87%; <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>): δ 7.26 (s, overlapped with CDCl<sub>3</sub>, 1H), 6.51 (s, 1H); <sup>13</sup>C{<sup>1</sup>H} NMR (100 MHz, DMSO-*d*<sub>6</sub>): δ 158.7, 147.2, 127.2, 120.6, 114.9.

Furan-2-carboxylic acid (4i) (CAS no. 88-14-2)<sup>S27</sup>



Light brown solid, 200.3 mg, 60%; <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>):  $\delta$  8.79 (br, 1H), 7.65 (m, 1H), 7.34 (d, J = 3.5 Hz, 1H), 6.57–6.56 (m, 1H); <sup>13</sup>C{<sup>1</sup>H} NMR (100 MHz, CDCl<sub>3</sub>):  $\delta$  163.5, 147.5, 143.8, 120.2, 112.3.

3-Bromofuran-2-carboxylic acid (4j) (CAS no. 14903-90-3)<sup>S28</sup>



Brown solid, 546.1 mg, 95%; <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>): δ 10.2 (br, 1H), 7.59 (m, 1H), 6.67 (m, 1H); <sup>13</sup>C{<sup>1</sup>H} NMR (100 MHz, CDCl<sub>3</sub>): δ 162.8, 146.9, 140.4, 117.5, 111.1.

## Experimental Procedure for the Structual Charactrization of Palladium Carbonyl Complexes by IR Measurment

[Fig. 1, eq. 1]

 $Pd(OAc)_2$  (0.3 mmol) and AcOH (3.0 mL, bubbled with Ar for 30 min) were sequentially added to a 50 mL stainless steel autoclave with a magnetic stirring bar under a N<sub>2</sub> atmosphere. The vessel was purged five times with CO and then charged with CO (1 atm). The reaction was conducted with

magnetic stirring at 50 °C for 15 min. The resulting mixture was filtered by suction and the residue was washed with dry  $Et_2O$  (15 mL), and the suction was continued for 10 min. The resulting solid was then dried between filter papers and the IR measurment of the resulting powder was conducted by KBr method without any further purification.

## [Fig. 1, eq. 2]

 $Pd(OAc)_2$  (0.3 mmol) and AcOH (3.0 mL, bubbled with Ar for 30 min) were sequentially added to a 50 mL stainless steel autoclave with a magnetic stirring bar under a N<sub>2</sub> atmosphere. The vessel was purged five times with CO and then charged with CO (30 atm). The reaction was conducted with magnetic stirring at 50 °C for 15 min. After removing excess CO from the reaction vessel, the vessel was purged five times with CO<sub>2</sub> and then charged with CO<sub>2</sub> (5 atm). The reaction was conducted with magnetic stirring at 100 °C for 15 min. The resulting mixture was filtered by suction and the residue was washed with dry Et<sub>2</sub>O (15 mL), and the suction was continued for 10 min. The resulting solid was then dried between filter papers and the IR measurment of the resulting powder was conducted by KBr method without any further purification.

## [Fig 2, eq. 6]

 $Pd(OAc)_2$  (0.3 mmol) and AcOH (3.0 mL, bubbled with Ar for 30 min) were sequentially added to a 50 mL stainless steel autoclave with a magnetic stirring bar under a N<sub>2</sub> atmosphere. The vessel was purged five times with CO and then charged with CO (10 atm). The reaction was conducted with magnetic stirring at 100 °C for 15 min. The resulting mixture was filtered by suction and the residue was washed with dry Et<sub>2</sub>O (15 mL), and the suction was continued for 10 min. The resulting solid was then dried between filter papers and the IR measurment of the resulting powder was conducted by KBr method without any further purification.

## [Fig. 2, eq. 7]

 $Pd(OAc)_2$  (0.3 mmol) and AcOH (3.0 mL, bubbled with Ar for 30 min) were sequentially added to a 50 mL stainless steel autoclave with a magnetic stirring bar under a N<sub>2</sub> atmosphere. The vessel was purged five times with CO<sub>2</sub> and then charged with CO<sub>2</sub> (5 atm) and CO (10 atm), respectively. The reaction was conducted with magnetic stirring at 100 °C for 15 min. The resulting mixture was filtered by suction and the residue was washed with dry Et<sub>2</sub>O (15 mL), and the suction was continued for 10 min. The resulting solid was then dried between filter papers and the IR measurment of the resulting powder was conducted by KBr method without any further purification.



Figure S1 Copies of <sup>1</sup>H and <sup>13</sup>C{<sup>1</sup>H} NMR spectra of 2a



Figure S2 Copies of <sup>1</sup>H and <sup>13</sup>C{<sup>1</sup>H} NMR spectra of 2b

# Figure S3 Copies of <sup>1</sup>H and <sup>13</sup>C{<sup>1</sup>H} NMR spectra of 2c





1.3 Q 1.2 ЮH Ξ 1.0 2d (100 MHz, CDCl<sub>3</sub>) 0.9 0.8 0.7 0.6 0.5 0.40.3 0.2 0.1 ibundance Median Contenting of the later of the 0 220.0 210.0 200.0 190.0 180.0 170.0 160.0 150.0 140.0 130.0 120.0 110.0 100.0 90.0 80.0 70.0 60.0 50.0 0 -10.0 -20.0 40.0 30.0 20.0 10.0 1111 135.361 <sup>-</sup> 130.034 <sup>-</sup> 126.643 <sup>-</sup> 77.429 77.103 76.787 167.848 154.972 41.570 34.633 32.411 28.885 25.590 23.022 14.189 10.874 X : parts per Million : Carbon13

Figure S4 Copies of <sup>1</sup>H and <sup>13</sup>C{<sup>1</sup>H} NMR spectra of 2d

Figure S5 Copies of <sup>1</sup>H and <sup>13</sup>C{<sup>1</sup>H} NMR spectra of 2e





Figure S6 Copies of <sup>1</sup>H and <sup>13</sup>C{<sup>1</sup>H} NMR spectra of 2f



Figure S7 Copies of <sup>1</sup>H and <sup>13</sup>C{<sup>1</sup>H} NMR spectra of 2g

Figure S8 Copies of <sup>1</sup>H and <sup>13</sup>C{<sup>1</sup>H} NMR spectra of 2h



Figure S9 Copies of <sup>1</sup>H and <sup>13</sup>C{<sup>1</sup>H} NMR spectra of 2i



Figure S10 Copies of <sup>1</sup>H and <sup>13</sup>C{<sup>1</sup>H} NMR spectra of 2j







Figure S12 The copy of <sup>1</sup>H NMR spectrum of 2l



Figure S14 The copy of <sup>1</sup>H NMR spectrum of 2n



Figure S15 The copy of <sup>1</sup>H NMR spectrum of 20



Figure S16 Copies of <sup>1</sup>H and <sup>13</sup>C{<sup>1</sup>H} NMR spectra of 4b



Figure S17 Copies of <sup>1</sup>H and <sup>13</sup>C{<sup>1</sup>H} NMR spectra of 4c



Figure S18 Copies of  ${}^{1}$ H and  ${}^{13}C{}^{1}$ H} NMR spectra of 4d



Figure S19 Copies of <sup>1</sup>H and <sup>13</sup>C{<sup>1</sup>H} NMR spectra of 4f



# Figure S20 Copies of <sup>1</sup>H and <sup>13</sup>C{<sup>1</sup>H} NMR spectra of 4g



Figure S21 Copies of <sup>1</sup>H and <sup>13</sup>C{<sup>1</sup>H} NMR spectra of 4h





# Figure S22 Copies of ${}^{1}H$ and ${}^{13}C{}^{1}H$ NMR spectra of 4i



# Figure S23 Copies of <sup>1</sup>H and <sup>13</sup>C{<sup>1</sup>H} NMR spectra of 4j





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