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The preservation of sarin and *O,O'*-diisopropyl fluorophosphate inside coordination cage hosts

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Cage preparation

For preparation of the H^{PEG} -OH cage, the $[Cd_8(L^{PEG})_{12}](NO_3)_{16}$ complex was dissolved in water and then filtered using a syringe filter (0.22 µm).¹ For preparation of the H^{W} -OH cage, the complex $[Co_8(L^W)_{12}](BF_4)_{16}$ was heated at 80 °C in water for 15 hours.² To obtain the H^{W} -Ol₁₆ and H-Cl₁₆ cages, the complexes $[Co_8(L^W)_{12}](BF_4)_{16}$ and $[Co_8(L)_{12}](BF_4)_{16}$ were separately converted to their chloride form using the ion exchange resin Dowex[®] chloride according to the previously published procedure.³ Residual resin was removed by filtration with use of a syringe filter (0.22 µm).

The concentration of the respective cages were determined using the absorbance at 292 nm (ϵ = 3.088 x 10⁵ M⁻¹ cm⁻¹) in MQ water. Buffered solutions of the cages were prepared using the appropriate quantities of solid boric acid and anhydrous borax. For example, to obtain 2 mL of a 50 mM buffer with a pH value of 8.7, boric acid (50 mM, 6.1 mg) and borax (12.5 mM, 5.0 mg) were added. The resulting suspension was sonicated for 3 minutes and then left to settle for 30 minutes. For the ¹⁹F and ³¹P NMR spectral experiments the final solution was 10% D₂O/90%H₂O (50 mM boric buffer, pH 8.7).



Fig. S1 - Representative hydrolysis curve of DFP (7.5 mM) with H^{PEG}•OH (0.5 mM) over time. Insert: the hydrolysis data fitted to a first order decay model to determine the hydrolysis rate constant and half-life.



Fig. S2 - ¹⁹F NMR spectra: (a) **H**^{PEG}•OH (0.2 mM), (b) **H**^{PEG}•OH (0.2 mM) + NaF (7.5 mM) and (c) NaF (7.5 mM); H₂O/D₂O (90:10 v/v) with boric buffer (pH 8.6, 50 mM).



Fig. S3 - ¹⁹F NMR spectrum of DFP (7.5 mM) and H^{w} •OH (0.5 mM), H₂O/D₂O (90:10 v/v) with boric buffer (pH 8.6, 50 mM).



Fig. S4 - Representative hydrolysis curve of DFP (7.5 mM) with $H^{W\bullet}OH$ (0.5 mM) present over time. Insert: the hydrolysis data (0-9 hours) fitted to a first order decay model to determine the hydrolysis rate constant and half-life.



Fig. S5 - ¹⁹F NMR spectra: (a) DFP (3.0 mM) and $H^{w} \cdot OH$ (0.6 mM); (b) DFP (3.0 mM) and $H^{w} \cdot OH$ (0.6 mM) + cycloundecanone (22 mM); H₂O/D₂O (90:10 v/v) with boric buffer (pH 8.6, 50 mM).



Fig. S6 - Extended packing of **H**•Cl₁₆•DFP viewed down the z-axis showing solvent accessible channels through which the organophosphate guests can access the cages.



Fig. S7 - Crystal structure of H•Cl₁₆•DFP showing DFP resolved in its lower occupancy position (16% occupancy) 2.7 Å further inside the window of the cage than the major position (58% occupancy). H atoms are omitted for clarity, Co – dark blue, C – black, N – blue, O – red, P – purple, F – light green, Cl – dark green.



Fig. S8 - The reaction progress profiles of the hydrolysis of GB; formation of *O*-isopropyl methylphosphonic acid (IMPA) over time as determined by NMR integration; without cage (black) and with H^{PEG} •OH (0.50 mM, purple), measurements in D₂O/H₂O/MeCN (10:85:5 v/v/v) with 50 mM boric buffer, pH 8.7 determined *via* ³¹P NMR spectroscopic analysis.

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