Supporting Information

Tuning the Spin-Crossover Properties of Fe^{II}₄L₆ Cages via the Interplay of Coordination Motif and Linker Modifications

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1 Materials and Methods

Reagents and solvents were purchased from commercial suppliers and used without further purification, unless otherwise specified. Deionised water was used in all cases and for Suzuki reactions, the water was degassed using three freeze-pump-thaw cycles. Tetrahydrofuran was dried using a Pure Solv MD-5 apparatus. Tetrakis(triphenylphosphine)palladium(0) was purchased commercially and stored under a nitrogen atmosphere. CD_3CN for the preparation of the Fe^{II}₄L₆ cages was dried using the following procedure: using dried glassware, 50 mL of CD_3CN was dried over 1 g of CaH₂, distilled and stored over molecular sieves (3-4 Å).

Fluorescence indicator plates (Polygram® SIL G/UV₂₅₄) from Macherey Nagel were used for thin-layer chromatography measurements having a coating thickness of 0.2 mm. Column flash chromatography was performed using an Isolera One from Biotage® with Biotage® SNAP Ultra (10g, 25g) and Biotage® Sfär Silica HC D columns (10g, 25g).

1.1 NMR Spectroscopy

NMR spectra were recorded on Bruker Avance 200, Bruker AvanceNeo 500, or Bruker Avance 600 spectrometers. Chemical shifts for ¹H, ¹³C, and ¹⁹F spectra are expressed in parts per million (ppm) and coupling constants (*J*) are reported in Hertz (Hz). ¹H and ¹³C spectra were referenced to TMS at 0.00 ppm and ¹⁹F spectra were referenced to C₆F₆ at -164.9 ppm. All measurements were carried out at 298 K unless reported otherwise. The following abbreviations are used to describe signal multiplicity for ¹H, ¹³C and ¹⁹F NMR spectra: s: singlet, d: doublet, t: triplet, m: multiplet, b: broad, unres.: unresolved.

The following pulse programs were used for diamagnetic compounds: zg30 (¹H), zgfhigqn (¹⁹F), cosygpmfppqf (COSY), hsqcedetgpsp.3 (HSQC), hmbcgplpndqf (HMBC), zgpg30 (¹³C).

The following pulse programs were used for the paramagnetic complexes: zg30 (¹H), zg (¹H), zgfhigqn (¹⁹F), cosyqf90 (COSY), cosygpqf (COSY), cosyqpmfqf (COSY), hmqcgpqf (HMQC), zg (¹³C).^[1]

1.2 Mass Spectrometry

Electron Ionisation (EI) mass spectrometry was carried out on a Jeol AccuTOF. High resolution electrospray ionisation mass spectrometry (ESI-MS) was carried out on a ThermoFisher Orbitrap (spray voltage 3-4 eV, capillary temperature 40-50 °C) infused from a Harvard syringe pump at a rate of 5-10 μ L per minute.

2 Ligand Synthesis

2.1 Imidazole-Based Ligand

Imidazole-based ligand **14** was synthesized according to Scheme S1 using our recently developed one-pot Sonogashira-type reaction.^[2] Precursor **12** was prepared using a literature-known procedure,^[3] while precursor **13** was obtained after methylation adapting a procedure from Ling and co-workers.^[4]



Scheme S1. Synthesis of imidazole-based ligand 14.

2.1.1 5'-Bromo-2'-(1*H*-imidazol-2-yl)pyridine (**12**)



Adapted from Zhang, Ma and co-workers.^[3a] 5-Bromo-2-cyanopyridine (1.83 g, 10.0 mmol) was dissolved in methanol (10 mL). The reaction mixture was treated with sodium methoxide solution (190 μ L, 1.03 mmol, 5.4 M in methanol) and stirred at 40 °C for 2 h. Afterwards, aminoacetalaldehyde diethyl acetal (1.45 mL, 10 mmol) and glacial acetic acid (1.10 mL) were added at 40 °C, the mixture heated to 70 °C and stirring was continued for 0.5 h. The reaction mixture was cooled to room temperature, methanol (5 mL) and hydrochloric acid (5 mL, 6 M) were added and the mixture was heated at 75 °C for 20 h. After cooling to room temperature, methanol was removed *in vacuo* and the remaining solution cooled to 0 °C. Dropwise addition of potassium carbonate solution (5.00 g in 5.00 mL water) resulted in precipitation of the product, which was filtered and washed with water (400 mL). The product was obtained as a colourless solid (2.00 g, 8.93 mmol, 89%).

Note: After the addition of sodium methoxide, stirring at 40 °C for 2 h is necessary to ensure imidate formation.^[3b]

The analytical data was consistent with literature data.^[3a]

¹**H NMR** (500 MHz, DMSO-*d*₆, 298 K) δ (ppm): 12.9 (br, 1H, *H_a*), 8.71 (d, ${}^{4}J$ = 2.3 Hz, 1H, *H_i*), 8.12 (dd, ${}^{3}J$ = 8.5 Hz, ${}^{4}J$ = 2.3 Hz, 1H, *H_g*), 7.98 (d, ${}^{3}J$ = 8.5 Hz, 1H, *H_f*), 7.19 (s, 2H, *H_{b,c}*).

¹³**C NMR** (151 MHz, DMSO- d_6 , 298 K) δ (ppm): 149.6 (C_i), 147.5 (C_e), 144.5 (C_d), 139.8 (C_g), 125.5 ($C_{b,c}$), 121.01 (C_f), 118.9 (C_h).

EI-MS m/z: 222.97442 (calculated for C₈H₆⁷⁹BrN₃: 222.97451), 224.97246 (calculated for C₈H₆⁸¹BrN₃: 224.97246).



Figure S1. ¹H NMR spectrum (500 MHz, DMSO- d_6 , 298 K) of 5'-bromo-2'-(1*H*-imidazol-2-yl)pyridine (**12**).



Figure S2. ¹³C NMR spectrum (151 MHz, DMSO-*d*₆, 298 K) of 5'-bromo-2'-(1*H*-imidazol-2-yl)pyridine (**12**).

2.1.2 5'-Bromo-2'-(1-methyl-1*H*-imidazol-2-yl)pyridine (13)



Adapted from Ling and co-workers.^[4] 5'-Bromo-2'-(1*H*-imidazol-2-yl)pyridine (**12**) (1.30 g, 5.80 mmol) and potassium carbonate (2.50 g, 18.1 mmol, milled) were dissolved in dimethylformamide (50 mL). After the addition of methyl iodide (500 μ L, 8.04 mmol), the mixture was stirred at room temperature for 24 h. The solution was poured into water (300 mL), the aqueous layer extracted with ethyl acetate (200 mL) and the layers separated. The organic layer was washed with brine (250 mL) and sodium hydroxide solution (200 mL, 10%) and dried over magnesium sulfate. The solvent was removed *in vacuo* and the product obtained as a brown solid (930 mg, 3.91 mmol, 67%).

¹**H NMR** (500 MHz, DMSO-*d*₆, 298 K) δ (ppm): 8.74 (dt, ${}^{4}J = 2.4$ Hz, ${}^{6}J = 0.7$ Hz, 1H, *H_i*), 8.13 (ddd, ${}^{3}J = 8.5$ Hz, ${}^{4}J = 2.4$ Hz, ${}^{6}J = 0.7$ Hz, 1H, *H_g*), 8.03 (dd, ${}^{3}J = 8.5$ Hz, ${}^{5}J = 0.7$ Hz, 1H, *H_i*), 7.35 (s, 1H, *H_b*), 7.05 (s, 1H, *H_c*), 4.03 (d, ${}^{6}J = 0.7$ Hz, 3H, *H_a*).

¹³**C NMR** (125 MHz, DMSO- d_6 , 298 K) δ (ppm): 149.0 (C_e), 148.9 (C_i), 142.9 (C_d), 139.6 (C_g), 128.0 (C_c), 125.6 (C_b), 123.7 (C_f), 118.8 (C_h), 35.9 (C_a).

ESI-MS *m/z*: 237.99731 (calculated for C₉H₉⁷⁹BrN₃: 237.99744).



Figure S3. ¹H NMR spectrum (500 MHz, DMSO- d_6 , 298 K) of 5'-bromo-2'-(1-methyl-1*H*-imidazol-2-yl)pyridine (**13**).



Figure S4. ¹³C NMR spectrum (125 MHz, DMSO- d_6 , 298 K) of 5'-bromo-2'-(1-methyl-1*H*-imidazol-2-yl)pyridine (**13**).



Figure S5. ¹H-¹H COSY NMR spectrum (500 MHz, DMSO-*d*₆, 298 K) of 5'-bromo-2'-(1-methyl-1*H*-imidazol-2-yl)pyridine (**13**).



Figure S6. ¹H-¹³C HSQC NMR spectrum (500 MHz/125 MHz, DMSO- d_6 , 298 K) of 5'-bromo-2'-(1-methyl-1*H*-imidazol-2-yl)pyridine (**13**).



Figure S7. ¹H-¹³C HMBC NMR spectrum (500 MHz/125 MHz, DMSO- d_6 , 298 K) of 5'-bromo-2'-(1-methyl-1*H*-imidazol-2-yl)pyridine (**13**).

2.1.3 1,2-Bis(6"-(1'-methyl-1*H*-imidazol-2'-yl)pyridin-3"-yl)ethyne (14)



Adapted from McConnell and co-workers.^[2] 5'-Bromo-2'-(1-methyl-1*H*-imidazol-2-yl)pyridine (**13**) (172 mg, 722 µmol) and Pd(PPh₃)₄ (41.8 mg, 5 mol%) were added to a pressure tube and the tube was evacuated for 5 min. Tetrabutylammonium fluoride (4.30 mL, 4.30 mmol) and trimethylsilylacetylene (100 µL, 723 µmol) were added, the tube immediately closed and the reaction mixture was heated at 70 °C for 19 h. After cooling to room temperature, water (50 mL) was added and the aqueous layer was extracted with dichloromethane (150 mL). The layers were separated, and the organic layer dried over magnesium sulfate and dried *in vacuo*. The crude product was further purified by flash chromatography (silica gel, 3% methanol/chloroform) and afterwards by an acid extraction where the impure product was dissolved in dichloromethane (10 mL) and the organic layer was extracted with hydrochloric acid (3 x 10 mL, 6 M). The layers were separated and the pH value of the aqueous layer adjusted to 12-14 using ammonia solution (25%). The precipitated product was collected by filtration as a yellow solid (78.5 mg, 231 µmol, 64%).

¹**H NMR** (500 MHz, CDCl₃, 298 K) δ (ppm): 8.74 (dd, ${}^{4}J = 2.1$ Hz, ${}^{5}J = 0.8$ Hz, 2H, H_i), 8.22 (dd, ${}^{3}J = 8.3$ Hz, ${}^{5}J = 0.8$ Hz, 2H, H_f), 7.89 (dd, ${}^{3}J = 8.3$ Hz, ${}^{4}J = 2.1$ Hz, 2H, H_g), 7.16 (d, ${}^{3}J = 1.1$ Hz, 2H, H_c), 7.00 (d, ${}^{3}J = 1.1$ Hz, 2H, H_b), 4.16 (s, 6H, H_a).

¹³**C NMR** (125 MHz, CDCl₃, 298 K) δ (ppm): 150.8 (*C_i*), 149.7 (*C_e*), 144.2 (*C_d*), 139.0 (*C_g*), 128.7 (*C_c*), 125.0 (*C_b*), 121.8 (*C_f*), 118.1 (*C_h*), 90.2 (*C_j*), 36.6 (*C_a*).

ESI-MS *m/z*: 341.15077 (calculated for C₂₀H₁₇N₆: 341.15092).



Figure S8. ¹H NMR spectrum (500 MHz, CDCl₃, 298 K) of 1,2-bis(6"-(1'-methyl-1*H*-imidazol-2'-yl)pyridine-3"-yl)ethyne (**14**).



Figure S9. ¹³C NMR spectrum (125 MHz, CDCl₃, 298 K) of 1,2-bis(6''-(1'-methyl-1*H*-imidazol-2'-yl)pyridine-3''-yl)ethyne (**14**).







Figure S11. ¹H-¹³C HSQC NMR spectrum (500 MHz/125 MHz, CDCl₃, 298 K) of 1,2-bis(6"-(1'-methyl-1*H*-imidazol-2'-yl)pyridine-3"-yl)ethyne (**14**).



Figure S12. ¹H-¹³C HMBC NMR spectrum (500 MHz/125 MHz, CDCl₃, 298 K) of 1,2-bis(6"-(1'-methyl-1*H*-imidazol-2'-yl)pyridine-3"-yl)ethyne (**14**).

2.2 Benzimidazole-Based Ligands with an Alkyne Linker

Benzimidazole-based ligands with an alkyne spacer were prepared according to Scheme S2. The appropriate picolinic acid and amine were reacted according to a procedure from Zhang, Ma and co-workers to obtain the precursors **15** and **20**.^[3a] Methylation or benzylation reactions were adapted from literature-known procedures.^[4-5] Ligands **17**, **19** and **22** were prepared using our previously reported one-pot Sonogashira-type method.^[2]



Scheme S2. Synthesis of benzimidazole-based ligands 17, 19 and 22.

2.2.1 Compounds **15-19**

Compounds **15-19** were synthesised according to a literature procedure and the analytical data was consistent with the literature data.^[2]

2.2.2 2-(5'-Bromo-4'-methylpyridin-2'-yl)-1*H*-benzo[*d*]imidazole (**20**)



Adapted from Zhang, Ma and co-workers.^[3a] Polyphosphoric acid (40 ml) was added to a three-neck flask and heated to 140 °C. *o*-Phenylenediamine (536 mg, 4.96 mmol) and 5bromo-4-methylpyridine-2-carboxylic acid (1.07 g, 4.95 mmol) were added and the reaction was stirred at 180 °C for 23.5 h. After cooling to 140 °C, the reaction mixture was poured into water (300 mL), neutralised with ammonia solution (25%) and the precipitate collected by filtration. After washing the precipitate with water (200 mL), product **20** was obtained as a pink solid (1.03 g, 3.57 mmol, 72%).

¹**H NMR** (600 MHz, DMSO-*d*₆, 298 K) δ (ppm): 13.15 (s, 1H, *H*_a), 8.81 (s, 1H, *H*_m), 8.34 (s, 1H, *H*_j), 7.69 (unres. dt, ³*J* = 8.0 Hz, 1H, *H*_c), 7.53 (unres. dt, ³*J* = 8.0 Hz, 1H, *H*_i), 7.26 (td, ³*J* = 8.0 Hz, ⁴*J* = 1.2 Hz, 1H, *H*_e), 7.22 (td, ³*J* = 8.0 Hz, ⁴*J* = 1.2 Hz, 1H, *H*_d), 2.49 (s, 3H, *H*_n).

¹³**C NMR** (151 MHz, DMSO- d_6 , 298 K) δ (ppm): 150.5 (C_m), 149.9 (C_h), 148.1 (C_i), 147.5 (C_k), 143.8 (C_b), 135.0 (C_g), 123.8 (C_i), 123.6 (C_j), 123.3 (C_e), 122.0 (C_d), 119.3 (C_c), 112.1 (C_i), 21.8 (C_n).

EI-MS m/z: 287.00591 (calculated for C₁₃H₁₀⁷⁹BrN₃: 287.00581), 289.00392 (calculated for C₁₃H₁₀⁸¹BrN₃: 289.00376).



Figure S13. ¹H NMR spectrum (600 MHz, DMSO-*d*₆, 298 K) of 2-(5'-bromo-4'-methylpyridin-2'-yl)-1*H*-benzo[*d*]imidazole (**20**).



Figure S14. ¹³C NMR spectrum (151 MHz, DMSO- d_6 , 298 K) of 2-(5'-bromo-4'-methylpyridin-2'-yl)-1*H*-benzo[*d*]imidazole (**20**).



Figure S15. ¹H-¹H COSY NMR spectrum (600 MHz, DMSO-*d*₆, 298 K) of 2-(5'-bromo-4'-methylpyridin-2'-yl)-1*H*-benzo[*d*]imidazole (**20**).



Figure S16. ¹H-¹³C HSQC NMR spectrum (600 MHz/151 MHz, DMSO- d_6 , 298 K) of 2-(5'-bromo-4'-methylpyridin-2'-yl)-1*H*-benzo[d]imidazole (**20**).



Figure S17. ¹H-¹³C HMBC NMR spectrum (600 MHz/151 MHz, DMSO- d_6 , 298 K) of 2-(5'-bromo-4'-methylpyridin-2'-yl)-1*H*-benzo[d]imidazole (**20**).

2.2.3 2-(5'-Bromo-4'-methylpyridin-2'-yl)-1-methyl-1*H*-benzo[*d*]imidazole (21)



Adapted from Ling and co-workers.^[4] 2-(5'-Bromo-4'-methylpyridin-2'-yl)-1*H*benzo[*d*]imidazole (**21**) (670 mg, 2.33 mmol), methyl iodide (200 μ L, 3.21 mmol) and potassium carbonate (998 mg, 7.22 mmol, milled) were dissolved in dimethylformamide (20 mL) and stirred at room temperature for 17 h. The reaction mixture was poured into water (100 mL) and the aqueous layer was extracted with ethyl acetate (150 mL). The layers were separated and the organic layer was washed with brine (250 mL), sodium hydroxide solution (100 mL, 10%) and dried over magnesium sulfate. The solvent was removed *in vacuo* and the product obtained as a brown solid (547 mg, 1.81 mmol, 78%).

¹**H NMR** (600 MHz, DMSO-*d*₆, 298 K) δ (ppm): 8.82 (s, 1H, *H_m*), 8.32 (s, 1H, *H_j*), 7.71 (dt, ³*J* = 7.7 Hz, ⁴*J* = 1.1 Hz, 1H, *H_t*), 7.65 (dt, ³*J* = 7.7 Hz, ⁴*J* = 1.1 Hz, 1H, *H_c*), 7.34 (unres. ddd, ³*J* = 7.7 Hz, ⁴*J* = 1.1 Hz, 1H, *H_d*), 7.28 (unres. ddd, ³*J* = 7.6 Hz, ⁴*J* = 1.1 Hz, 1H, *H_e*), 4.20 (s, 3H, *H_a*), 2.48 (s, 3H, *H_n*).

¹³**C NMR** (151 MHz, DMSO- d_6 , 298 K) δ (ppm): 150.0 (C_m), 148.9 ($C_{h,i}$), 148.0 (C_k), 142.0 (C_g), 137.1 (C_b), 126.5 (C_j), 123.7 (C_i), 123.3 (C_d), 122.5 (C_e), 119.4 (C_c), 110.9 (C_i), 32.6 (C_a), 21.8 (C_n).

ESI-MS *m/z*: 302.02863 (calculated for C₁₄H₁₃N₃Br: 302.02874).



Figure S18. ¹H NMR spectrum (600 MHz, DMSO-*d*₆, 298 K) of 2-(5'-bromo-4'-methylpyridin-2'-yl)-1-methyl-1*H*-benzo[*d*]imidazole (**21**).



Figure S19. ¹³C NMR spectrum (151 MHz, DMSO- d_6 , 298 K) of 2-(5'-bromo-4'-methylpyridin-2'-yl)-1-methyl-1*H*-benzo[d]imidazole (**21**).



Figure S20. ¹H-¹H COSY NMR spectrum (600 MHz, DMSO- d_6 , 298 K) of 2-(5'-bromo-4'-methylpyridin-2'-yl)-1-methyl-1*H*-benzo[d]imidazole (**21**).



Figure S21. ¹H-¹³C HSQC NMR spectrum (600 MHz/151 MHz, DMSO- d_6 , 298 K) of 2-(5'-bromo-4'-methylpyridin-2'-yl)-1-methyl-1*H*-benzo[*d*]imidazole (**21**).



Figure S22. ¹H-¹³C HMBC NMR spectrum (600 MHz/151 MHz, DMSO- d_6 , 298 K) of 2-(5'-bromo-4'-methylpyridin-2'-yl)-1-methyl-1*H*-benzo[*d*]imidazole (**21**).

2.2.4 1,2-Di(4"-methyl-6"-(1'-methyl-1*H*-benzo[*d*]imidazol-2'-yl)pyridin-3"-yl)ethyne (22)



Adapted from McConnell and co-workers.^[2] 2-(5'-Bromo-4'-methylpyridin-2'-yl)-1-methyl-1*H*benzo[*d*]imidazole (**21**) (218 mg, 712 µmol) and Pd(PPh₃)₄ (41.8 mg, 5 mol%) were added to a pressure tube and the tube was evacuated for 5 min. Tetrabutylammonium fluoride (4.30 mL, 4.30 mmol) and trimethylsilylacetylene (100 µL, 723 µmol) were added, the tube immediately closed and the reaction mixture heated at 70 °C for 21 h. After cooling to room temperature, the precipitate was collected by filtration and washed with cold tetrahydrofuran (250 mL). The product was obtained as a yellow solid (120 mg, 256 µmol, 71%).

¹**H NMR** (600 MHz, TFA-*d*₁, 298 K) δ (ppm): 9.29 (s, 2H, *H_m*), 8.40 (s, 2H, *H_j*), 7.95-7.82 (m, 8H, *H_{c,d,e,f}*), 4.34 (s, 6H, *H_a*), 2.97 (s, 6H, *H_n*).

¹³**C NMR** (151 MHz, TFA- d_1 , 298 K) δ (ppm): 161.7 (C_k), 152.8 (C_m), 144.6 (C_h), 139.1 (C_i), 135.8 (C_b), 132.5 (C_g), 132.0 ($C_{c/d/e/t}$), 131.5 (C_j), 131.4 ($C_{c/d/e/t}$), 127.3 (C_i), 114.3 ($C_{c/d/e/t}$), 112.2 ($C_{c/d/e/t}$), 96.0 (C_o), 35.0 (C_a), 22.7 (C_n).

EI-MS *m/z*: 468.20493 (calculated for C₃₀H₂₄N₆: 468.20624).



Figure S23. ¹H NMR spectrum (600 MHz, TFA- d_1 , 298 K) of 1,2-di(4"-methyl-6"-(1'-methyl-1*H*-benzo[d]imidazol-2'-yl)pyridin-3"-yl)ethyne (**22**).



Figure S24. ¹³C NMR spectrum (151 MHz, TFA-*d*₁, 298 K) of 1,2-di(4''-methyl-6"-(1'-methyl-1*H*-benzo[*d*]imidazol-2'-yl)pyridin-3"-yl)ethyne (**22**).



Figure S25. ¹H-¹H COSY NMR spectrum (600 MHz, TFA- d_1 , 298 K) of 1,2-di(4"-methyl-6"-(1'-methyl-1*H*-benzo[d]imidazol-2'-yl)pyridin-3"-yl)ethyne (**22**).



Figure S26. ¹H-¹³C HSQC NMR spectrum (600 MHz/151 MHz, TFA- d_1 , 298 K) of 1,2-di(4"-methyl-6"-(1'-methyl-1*H*-benzo[d]imidazol-2'-yl)pyridin-3"-yl)ethyne (**22**).



Figure S27. ¹H-¹³C HMBC NMR spectrum (600 MHz/151 MHz, TFA-*d*₁, 298 K) of 1,2-di(4"-methyl-6"-(1'-methyl-1*H*-benzo[*d*]imidazol-2'-yl)pyridin-3"-yl)ethyne (**22**).

2.3 Benzimidazole-based Ligands with no Linker

Benzimidazole-based ligands without a spacer group were synthesised according to Scheme S3. Boronic acid **23** was prepared adapting the synthesis of thiophene-based boronic acids.^[6] In addition, boronic ester **25** was prepared according to Severin and co-workers.^[7]



Scheme S3. Synthesis of ligands 24 and 26.

2.3.1 (6'-(1-Methyl-1*H*-benzo[*d*]imidazol-2-yl)pyridin-3'-yl)boronic acid (23)



Adapted from Phillips and co-workers.^[6] 2-(5'-Bromo-4'-methylpyridin-2'-yl)-1-methyl-1*H*-benzo[*d*]imidazole (**16**) (985 mg, 3.42 mmol) was added to a three-neck flask and the flask was evacuated for 5 min. After the addition of anhydrous tetrahydrofuran (15 mL), the solution was cooled to -78 °C. *n*-Butyllithium (1.50 mL, 3.75 mmol, 2.5 M in hexanes) was added dropwise, the reaction mixture stirred at -78 °C for 2 h before tributyl borate (1.02 mL, 3.78 mmol) was added. Stirring was continued at -78 °C for 2 h. Hydrochloric acid (12 mL, 24 mmol, 2 M) was added and the reaction mixture warmed to room temperature and stirred for 18 h. The precipitate was collected by filtration and washed with cold tetrahydrofuran (3 mL). The product was obtained as a colourless solid (705 mg, 2.79 mmol, 81%).

¹**H NMR** (500 MHz, DMSO-*d*₆, 298 K) δ (ppm): 9.15 (s, 1H, *H_m*), 8.44 (dt, ³*J* = 7.8 Hz, ⁴*J* = 1.9 Hz, 1H, *H_k*), 8.33 (dd, ³*J* = 7.8 Hz, ⁶*J* = 1.3 Hz, 1H, *H_j*), 7.97 (d, ³*J* = 7.9 Hz, 1H, *H_c*), 7.86 (d, ³*J* = 8.1 Hz, 1H, *H_t*), 7.62-7.54 (m, 2H, *H_{d,e}*), 4.32 (d, ⁶*J* = 1.3 Hz, 3H, *H_a*).

¹³**C NMR** (125 MHz, DMSO- d_6 , 298 K) δ (ppm): 154.6 (C_m), 147.6 (C_h), 145.6 (C_i), 143.1 (C_k), 134.7 (C_b), 133.8 (C_g), 131.6 (C_i), 125.6 ($C_{d/e}$), 125.4 ($C_{d/e}$), 124.6 (C_j), 115.9 (C_f), 112.6 (C_c), 33.2 (C_a).

¹¹**B NMR** (160 MHz, DMSO-*d*₆, 298 K) δ (ppm): 28.3 (B_n).

ESI-MS *m/z*: 253.11327 (calculated for C₁₃H₁₃N₃O₂B: 253.11317).



Figure S28. ¹H NMR spectrum (500 MHz, DMSO- d_6 , 298 K) of (6'-(1-methyl-1*H*-benzo[*d*]imidazol-2-yl)pyridin-3'-yl)boronic acid (**23**).



Figure S29. ¹³C NMR spectrum (125 MHz, DMSO- d_6 , 298 K) of (6'-(1-methyl-1*H*-benzo[*d*]imidazol-2-yl)pyridin-3'-yl)boronic acid (**23**).



Figure S30. ¹¹B NMR spectrum (160 MHz, DMSO- d_6 , 298 K) of (6'-(1-methyl-1*H*-benzo[d]imidazol-2-yl)pyridin-3'-yl)boronic acid (**23**).



Figure S31. ¹H-¹H COSY NMR spectrum (500 MHz, DMSO- d_6 , 298 K) of (6'-(1-methyl-1*H*-benzo[d]imidazol-2-yl)pyridin-3'-yl)boronic acid (**23**).



Figure S32. ¹H-¹³C HSQC NMR spectrum (500 MHz/125 MHz, DMSO- d_6 , 298 K) of (6'-(1-methyl-1*H*-benzo[d]imidazol-2-yl)pyridin-3'-yl)boronic acid (**23**).



Figure S33. ¹H-¹³C HMBC NMR spectrum (500 MHz/125 MHz, DMSO- d_6 , 298 K) of (6'-(1-methyl-1*H*-benzo[d]imidazol-2-yl)pyridin-3'-yl)boronic acid (**23**).

2.3.2 6',6"-Di(1-methyl-1*H*-benzo[*d*]imidazol-2-yl)-3',3"-bipyridine (24)



Adapted from Lusby and co-workers.^[8] 2-(5'-Bromopyridin-2'-yl)-1-methyl-1*H*-benzo[*d*]imidazole (**16**) (350 mg, 1.22 mmol), (6'-(1-methyl-1*H*-benzo[*d*]imidazol-2-yl)pyridin-3'-yl)boronic acid (**23**) (309 mg, 1.22 mmol), potassium carbonate (422 mg, 3.05 mmol, milled) and Pd(PPh₃)₄ (70.5 mg, 5 mol%) were added to a three-neck flask and the flask was evacuated for 5 min. Ethanol (4 mL, dry), water (4 mL, degassed) and tetrahydrofuran (9 mL, dry) were added and the reaction mixture was stirred at 80 °C for 19.75 h. The solution was cooled to room temperature and the precipitate was collected by filtration and washed with cold tetrahydrofuran (250 mL). The product was obtained as a colourless solid (374 mg, 898 µmol, 74%)

The analytical data was consistent with literature data.^[8]

¹**H NMR** (600 MHz, CDCl₃, 298 K) δ (ppm): 9.03 (dd, ${}^{4}J = 2.4$ Hz, ${}^{5}J = 0.7$ Hz, 2H, H_m), 8.58 (dd, ${}^{3}J = 8.2$ Hz, ${}^{5}J = 0.7$ Hz, 2H, H_j), 8.15 (dd, ${}^{3}J = 8.2$ Hz, ${}^{4}J = 2.4$ Hz, 2H, H_k), 7.87 (unres. dt, 2H, H_f), 7.48 (unres. dt, 2H, H_c), 7.38 (td, ${}^{3}J = 7.1$ Hz, ${}^{4}J = 1.3$ Hz, 2H, H_d), 7.35 (td, ${}^{3}J = 7.1$ Hz, ${}^{4}J = 1.3$ Hz, 2H, H_d), 7.35 (td, ${}^{3}J = 7.1$ Hz, ${}^{4}J = 1.3$ Hz, 2H, H_d), 7.35 (td, ${}^{3}J = 7.1$ Hz, ${}^{4}J = 1.3$ Hz, 2H, H_e), 4.36 (s, 6H, H_a).

¹³**C NMR** (151 MHz, CDCl₃, 298 K) δ (ppm): 150.5 (*C_i*), 149.7 (*C_h*), 146.8 (*C_m*), 142.7 (*C_g*), 137.5 (*C_b*), 135.0 (*C_k*), 132.7 (*C_i*), 124.9 (*C_j*), 123.6 (*C_d*), 122.9 (*C_e*), 120.2 (*C_i*), 110.0 (*C_c*), 32.9 (*C_a*).

EI-MS *m/z*: 416.17383 (calculated for C₂₆H₂₀N₆: 416.17494).



Figure S34. ¹H NMR spectrum (600 MHz, $CDCI_3$, 298 K) of 6',6"-di(1-methyl-1*H*-benzo[*d*]imidazol-2-yl)-3',3"-bipyridine (**24**).



Figure S35. ¹³C NMR spectrum (151 MHz, CDCl₃, 298 K) of 6',6"-di(1-methyl-1*H*-benzo[d]imidazol-2-yl)-3',3"-bipyridine (**24**).



Figure S36. ¹H-¹H COSY NMR spectrum (600 MHz, CDCl₃, 298 K) of 6',6"-di(1-methyl-1*H*-benzo[*d*]imidazol-2-yl)-3',3"-bipyridine (**24**).



Figure S37. ¹H-¹³C HSQC NMR spectrum (600 MHz/151 MHz, CDCl₃, 298 K) of 6',6"-di(1-methyl-1*H*-benzo[*d*]imidazol-2-yl)-3',3"-bipyridine (**24**).



Figure S38. ¹H-¹³C HMBC NMR spectrum (600 MHz/151 MHz, CDCl₃, 298 K) of 6',6"-di(1-methyl-1*H*-benzo[*d*]imidazol-2-yl)-3',3"-bipyridine (**24**).

2.3.3 1-Benzyl-2-(5'-(4",4",5",5"-tetramethyl-1",3",2"-dioxaborolan-2"-yl)pyridin-2'-yl)-1*H*-benzo[*d*]imidazole (**25**)

Adapted from Severin and co-workers.^[7] 2-(5'-Bromopyridin-2'-yl)-1-benzyl-1*H*-benzo[*d*]imidazole (**18**) (300 mg, 824 µmol), bis(pinacolato)diboron (437 mg, 1.72 mmol), potassium acetate (404 mg, 4.12 mmol) and Pd(dppf)Cl₂·CH₂Cl₂ (33.7 mg, 5 mol%) were added to a three-neck flask and the flask was evacuated for 5 min. Anhydrous 1,4-dioxane (9 mL) was added and the reaction mixture was stirred at 90 °C for 19 h. The solution was cooled to room temperature and the crude reaction mixture was filtered through a silica gel plug (1:1 cyclohexane/ethyl acetate). The solvent was removed *in vacuo*. The impure product was used in the next step without further purification.



¹**H NMR** (500 MHz, DMSO-*d*₆, 298 K) δ (ppm): 8.85 (dd, ${}^{4}J$ = 1.8 Hz, ${}^{5}J$ = 1.0 Hz, 1H, *H*_{*q*}), 8.40 (dd, ${}^{3}J$ = 7.9 Hz, ${}^{5}J$ = 1.0 Hz, 1H, *H*_{*h*}), 8.19 (dd, ${}^{3}J$ = 7.9 Hz, ${}^{4}J$ = 1.8 Hz, 1H, *H*_{*o*}), 7.78-7.75 (m, 1H, *H*_{*g*/*j*}), 7.65-7.62 (m, 1H, *H*_{*g*/*j*}), 7.33-7.26 (m, 2H, *H*_{*h*,*i*}), 7.26-7.21 (m, 2H, *H*_{*b*}), 7.20-7.16 (m, 1H, *H*_{*a*}), 7.14-7.10 (m, 2H, *H*_{*c*}), 6.25 (s, 2H, *H*_{*e*}), 1.33 (s, 12H, *H*_{*t*}).

¹³**C NMR** (125 MHz, DMSO-*d*₆, 298 K) δ (ppm): 153.7 (*C*_q), 152.0 (*C*_m), 148.7 (*C*_i), 143.0 (*C*_o), 142.1 (*C*_k), 137.7 (*C*_d), 136.6 (*C*_f), 128.4 (*C*_b), 127.1 (*C*_a), 126.6 (*C*_c), 123.6 (*C*_{h/l}), 123.5 (*C*_n), 122.7 (*C*_{h/l}), 119.7 (*C*_{g/l}), 111.3 (*C*_{g/l}), 84.3 (*C*_s), 47.9 (*C*_e), 24.6 (*C*_l).

Note: C_{ρ} was not observed.

¹¹**B NMR** (160 MHz, DMSO-*d*₆, 298 K) δ (ppm): Unambiguous assignment of the two signals was not possible due to the presence of a boron-containing impurity.





Figure S39. ¹H NMR spectrum (500 MHz, DMSO-*d*₆, 298 K) of 1-benzyl-2-(5'-(4",4",5",5"-tetramethyl-1",3",2"-dioxaborolan-2"-yl)pyridin-2'-yl)-1*H*-benzo[*d*]imidazole (**25**).



Figure S40. ¹³C NMR spectrum (125 MHz, DMSO-*d*₆, 298 K) of 1-benzyl-2-(5'-(4",4",5",5"-tetramethyl-1",3",2"-dioxaborolan-2"-yl)pyridin-2'-yl)-1*H*-benzo[*d*]imidazole (**25**).



Figure S41. ¹¹B NMR spectrum (160 MHz, DMSO-*d*₆, 298 K) of 1-benzyl-2-(5'-(4",4",5",5"-tetramethyl-1",3",2"-dioxaborolan-2"-yl)pyridin-2'-yl)-1*H*-benzo[*d*]imidazole (**25**).



Figure S42. ¹H-¹H COSY NMR spectrum (500 MHz, DMSO-*d*₆, 298 K) of 1-benzyl-2-(5'-(4",4",5",5"-tetramethyl-1",3",2"-dioxaborolan-2"-yl)pyridin-2'-yl)-1*H*-benzo[*d*]imidazole (**25**).



Figure S43. ¹H-¹³C HSQC NMR spectrum (500 MHz/125 MHz, DMSO-*d*₆, 298 K) of 1-benzyl-2-(5'-(4",4",5",5"-tetramethyl-1",3",2"-dioxaborolan-2"-yl)pyridin-2'-yl)-1*H*-benzo[*d*]imidazole (25).



Figure S44. ¹H-¹³C HMBC NMR spectrum (500 MHz/125 MHz, DMSO-*d*₆, 298 K) of 1-benzyl-2-(5'-(4",4",5",5"-tetramethyl-1",3",2"-dioxaborolan-2"-yl)pyridin-2'-yl)-1*H*-benzo[*d*]imidazole (25).

2.3.4 6',6"-Di(1-benzyl-1*H*-benzo[*d*]imidazol-2-yl)-3',3"-bipyridine (26)



Adapted from Lusby and co-workers.^[8] 2-(5'-Bromopyridin-2'-yl)-1-benzyl-1*H*-benzo[*d*]imidazole (**18**) (79.0 mg, 217 µmol), 1-benzyl-2-(5'-(4",4",5",5"-tetramethyl-1",3",2"-dioxaborolan-2"-yl)pyridin-2'-yl)-1*H*-benzo[*d*]imidazole (**25**) (89.3 mg, 217 µmol), potassium carbonate (150 mg, 1.09 mmol, milled) and Pd(PPh₃)₄ (25.1 mg, 5 mol%) were added to a three-neck flask and the flask was evacuated for 5 min. Anhydrous ethanol (2 mL), water (2 mL, degassed) and anhydrous tetrahydrofuran (5 mL) were added and the reaction mixture was stirred at 80 °C for 23.5 h. The solution was cooled to room temperature, filtered and the precipitate was washed with cold tetrahydrofuran (250 mL). The product was obtained as a colourless solid (59.7 mg, 105 µmol, 49%).

¹**H NMR** (600 MHz, TFA-*d*₁, 298 K) δ (ppm): 9.41 (dd, ${}^{4}J = 2.2$ Hz, ${}^{5}J = 0.7$ Hz, 2H, *H*_{*q*}), 8.64 (dd, ${}^{3}J = 8.2$ Hz, ${}^{4}J = 2.2$ Hz, 2H, *H*_{*o*}), 8.34 (dd, ${}^{3}J = 8.2$ Hz, ${}^{5}J = 0.7$ Hz, 2H, *H*_{*n*}), 7.97 (unres. dt, 2H, *H*_{*j*}), 7.84-7.74 (m, 6H, *H*_{*g*,*h*,*i*}), 7.39-7.32 (m, 6H, *H*_{*a*,*b*}), 7.21-7.16 (m, 4H, *H*_{*c*}), 6.05 (s, 4H, *H*_{*e*}).}

¹³**C NMR** (151 MHz, TFA- d_1 , 298 K) δ (ppm): 150.7 (C_q), 146.7 (C_i), 142.9 (C_m), 141.7 (C_o), 138.4 (C_p), 135.6 (C_i), 134.5 (C_d), 132.6 (C_k), 131.9 (C_b), 131.8 (C_a), 131.6 (C_i), 131.2 (C_h), 129.7 (C_n), 128.5 (C_c), 117.0 (C_j), 115.6 (C_g), 52.8 (C_e).

EI-MS *m/z*: 568.23722 (calculated for C₃₈H₂₈N₆: 568.23754).



Figure S45. ¹H NMR spectrum (600 MHz, TFA- d_1 , 298 K) of 6',6"-di(1-benzyl-1*H*-benzo[d]imidazol-2-yl)-3',3"-bipyridine (**26**).



Figure S46. ¹³C NMR spectrum (151 MHz, TFA- d_1 , 298 K) of 6',6"-di(1-benzyl-1*H*-benzo[*d*]imidazol-2-yl)-3',3"-bipyridine (**26**).



Figure S47. ¹H-¹H COSY NMR spectrum (600 MHz, TFA- d_1 , 298 K) of 6',6"-di(1-benzyl-1*H*-benzo[*d*]imidazol-2-yl)-3',3"-bipyridine (**26**).



Figure S48. ¹H-¹³C HSQC NMR spectrum (600 MHz/151 MHz, TFA-*d*₁, 298 K) of 6',6"-di(1-benzyl-1*H*-benzo[*d*]imidazol-2-yl)-3',3"-bipyridine (**26**).



Figure S49. ¹H-¹³C HMBC NMR spectrum (600 MHz/151 MHz, TFA-*d*₁, 298 K) of 6',6"-di(1-benzyl-1*H*-benzo[*d*]imidazol-2-yl)-3',3"-bipyridine (**26**).

2.4 Benzimidazole-based Ligands with a Phenylene-Based Spacer

Benzimidazole-based ligands with a phenylene-based spacer were synthesised according to Scheme S4. The boronic esters were prepared adapting a literature-known procedure.^[7]



Scheme S4. Synthesis of ligands 27, 28, 30, 32 and 33.

2.4.1 1,4-Di(6"-(1'-methyl-1*H*-benzo[*d*]imidazol-2'-yl)pyridin-3"-yl)benzene (27)



Adapted from Lusby and co-workers.^[8] 2-(5'-Bromopyridin-2'-yl)-1-methyl-1*H*benzo[*d*]imidazole (**16**) (250 mg, 868 µmol), *p*-phenylenediboronic acid (71.9 mg, 434 µmol), potassium carbonate (300 mg, 2.17 mmol, milled) and Pd(PPh₃)₄ (50.2 mg, 10 mol%) were added to a three-neck flask and the flask was evacuated for 5 min. Anhydrous ethanol (3 mL), water (3 mL, degassed) and anhydrous tetrahydrofuran (7 mL) were added and the reaction mixture stirred at 80 °C for 20 h. The solution was cooled to room temperature and the precipitate was collected by filtration and washed with cold tetrahydrofuran (200 mL). The product was obtained as a colourless solid (152 mg, 309 µmol, 71%).

¹**H NMR** (600 MHz, CDCl₃, 298 K) δ (ppm): 9.02 (d, ⁴*J* = 2.3 Hz, 2H, *H_m*), 8.53 (d, ³*J* = 8.2 Hz, 2H, *H_j*), 8.13 (dd, ³*J* = 8.2 Hz, ⁴*J* = 2.3 Hz, 2H, *H_k*), 7.86 (d, ³*J* = 7.7 Hz, 2H, *H_f*), 7.84 (s, 4H, *H_o*), 7.48 (d, ³*J* = 7.6 Hz, 2H, *H_c*), 7.39-7.36 (m, 2H, *H_d*), 7.36-7.33 (m, 2H, *H_e*), 4.36 (s, 6H, *H_a*).

¹³**C NMR** (151 MHz, CDCl₃, 298 K) δ (ppm): 150.0 (*C_h*), 149.7 (*C_i*), 146.9 (*C_m*), 142.7 (*C_g*), 137.5 (*C_b*), 137.4 (*C_n*), 135.6 (*C_i*), 135.0 (*C_k*), 127.9 (*C_o*), 124.8 (*C_j*), 123.5 (*C_d*), 122.7 (*C_e*), 120.0 (*C_t*), 110.0 (*C_c*), 32.9 (*C_a*).

EI-MS m/z: 492.20465 (calculated for C₃₂H₂₄N₆: 492.20624).



Figure S50. ¹H NMR spectrum (600 MHz, $CDCI_3$, 298 K) of 1,4-di(6"-(1'-methyl-1*H*-benzo[*d*]imidazol-2'-yl)pyridin-3"-yl)benzene (**27**).


Figure S51. ¹³C NMR spectrum (151 MHz, CDCl₃, 298 K) of 1,4-di(6"-(1'-methyl-1*H*-benzo[*d*]imidazol-2'-yl)pyridin-3"-yl)benzene (**27**).



Figure S52. ¹H-¹H COSY NMR spectrum (600 MHz, CDCl₃, 298 K) of 1,4-di(6"-(1'-methyl-1*H*-benzo[*d*]imidazol-2'-yl)pyridin-3"-yl)benzene (**27**).



Figure S53. ¹H-¹³C HSQC NMR spectrum (600 MHz/151 MHz, CDCl₃, 298 K) of 1,4-di(6"-(1'-methyl-1*H*-benzo[*d*]imidazol-2'-yl)pyridin-3"-yl)benzene (**27**).



Figure S54. ¹H-¹³C HMBC NMR spectrum (600 MHz/151 MHz, CDCl₃, 298 K) of 1,4-di(6"-(1'-methyl-1*H*-benzo[*d*]imidazol-2'-yl)pyridin-3"-yl)benzene (**27**).

2.4.2 1,4-Di(6"-(1'-benzyl-1*H*-benzo[*d*]imidazol-2'-yl)pyridin-3"-yl)benzene (28)



Adapted from Lusby and co-workers.^[8] 2-(5'-Bromopyridin-2'-yl)-1-benzyl-1*H*-benzo[*d*]imidazole (**18**) (158 mg, 434 µmol), *p*-phenylenediboronic acid (36.0 mg, 217 µmol), potassium carbonate (150 mg, 1.09 mmol, milled) and Pd(PPh₃)₄ (25.1 mg, 10 mol%) were added to a three-neck flask and the flask was evacuated for 5 min. Anhydrous ethanol (1.5 mL), water (1.5 mL, degassed) and anhydrous tetrahydrofuran (3.5 mL) were added and the reaction mixture stirred at 80 °C for 18 h. The solution was cooled to room temperature and the precipitate was collected by filtration and washed with cold tetrahydrofuran (250 mL). The product was obtained as a colourless solid (101 mg, 157 µmol, 72%).

¹**H NMR** (500 MHz, TFA-*d*₁, 298 K) δ (ppm): 9.46 (d, ⁴*J* = 2.1 Hz, 2H, *H*_q), 8.86 (dd, ³*J* = 8.3 Hz, ⁴*J* = 2.1 Hz, 2H, *H*_o), 8.38 (d, ³*J* = 8.3 Hz, 2H, *H*_n), 8.04 (s, 4H, *H*_s), 7.99 (d, ³*J* = 8.3 Hz, 1H, *H*_j), 7.88-7.80 (m, 3H, *H*_{g,h,i}), 7.42-7.34 (m, 3H, *H*_{a,b}), 7.16 (d, ³*J* = 8.0 Hz, 2H, *H*_c), 5.94 (s, 2H, *H*_e).

¹³**C NMR** (151 MHz, TFA- d_1 , 298 K) δ (ppm): 147.6 (C_q), 145.0 ($C_{p/r}$), 144.9 (C_o), 143.3 (C_i), 137.8 ($C_{p/r}$), 136.9 (C_m), 135.3 (C_i), 133.8 (C_d), 132.7 (C_k), 132.0 (C_a), 132.0 (C_i), 131.9 (C_b), 131.7 (C_n), 131.6 (C_h), 131.2 (C_s), 128.6 (C_c), 117.1 (C_j), 115.5 (C_g), 53.0 (C_e).

EI-MS m/z: 644.26771 (calculated for C₄₄H₃₂N₆: 644.26884).



Figure S55. ¹H NMR spectrum (500 MHz, TFA- d_1 , 298 K) of 1,4-di(6"-(1'-benzyl-1*H*-benzo[*d*]imidazol-2'-yl)pyridin-3"-yl)benzene (**28**).



Figure S56. ¹³C NMR spectrum (125 MHz, TFA- d_1 , 298 K) of 1,4-di(6"-(1'-benzyl-1*H*-benzo[*d*]imidazol-2'-yl)pyridin-3"-yl)benzene (**28**).



Figure S57. ¹H-¹H COSY NMR spectrum (500 MHz, TFA- d_1 , 298 K) of 1,4-di(6"-(1'-benzyl-1*H*-benzo[d]imidazol-2'-yl)pyridin-3"-yl)benzene (**28**).



Figure S58. ¹H-¹³C HSQC NMR spectrum (500 MHz/125 MHz, TFA- d_1 , 298 K) of 1,4-di(6"-(1'-benzyl-1*H*-benzo[d]imidazol-2'-yl)pyridin-3"-yl)benzene (**28**).



Figure S59. ¹H-¹³C HMBC NMR spectrum (500 MHz/125 MHz, TFA- d_1 , 298 K) of 1,4-di(6"-(1'-benzyl-1*H*-benzo[d]imidazol-2'-yl)pyridin-3"-yl)benzene (**28**).

2.4.3 2,2'-(2,5-Dimethyl-1,4-phenylene)di(4",4",5",5"-tetramethyl-1",3",2"-dioxaborolane) (29)



Adapted from Severin and co-workers.^[7] 1,4-Dibromo-2,5-dimethylbenzene (500 mg, 1.89 mmol), bis(pinacolato)diboron (1.00 g, 3.94 mmol), potassium acetate (925 mg, 9.42 mmol) and Pd(dppf)Cl₂·CH₂Cl₂ (77.2 mg, 5 mol%) were added to a three-neck flask and the flask was evacuated for 5 min. Anhydrous 1,4-dioxane (20 mL) was added and the mixture was stirred at 90 °C for 23 h. The reaction mixture was cooled to room temperature and the solvent was removed *in vacuo*. Purification by flash chromatography (silica gel, 33% dichloromethane/cyclohexane) gave the product as a colourless solid (196 mg, 547 μ mol, 29%).

The analytical data was consistent with literature data.^[7]

¹**H NMR** (500 MHz, CDCl₃, 298 K) δ (ppm): 7.54 (s, 2H, *H_b*), 2.48 (s, 6H, *H_d*), 1.34 (s, 24H, *H_g*).

¹³**C NMR** (125 MHz, CDCl₃, 298 K) δ (ppm): 140.5 (*C_c*), 136.9 (*C_b*), 83.4 (*C_f*), 24.9 (*C_g*), 21.5 (*C_d*). *C_a* was not observed.

EI-MS m/z: 358.24906 (calculated for C₂₀H₂₈¹¹B₂O₄: 358.24867).



Figure S60. ¹H NMR spectrum (500 MHz, CDCl₃, 298 K) of 2,2'-(2,5-dimethyl-1,4-phenylene)di(4",4",5",5"-tetramethyl-1",3",2"-dioxaborolane) (**29**).



Figure S61. ¹³C NMR spectrum (125 MHz, CDCl₃, 298 K) of 2,2'-(2,5-dimethyl-1,4-phenylene)di(4",4",5",5"-tetramethyl-1",3",2"-dioxaborolane) (**29**).

2.4.4 1,4-Di(6"-(1'-methyl-1*H*-benzo[*d*]imidazol-2'-yl)pyridin-3"-yl)-2,5-dimethylbenzene (**30**)



Adapted from Lusby and co-workers.^[8] 2-(5'-Bromopyridin-2'-yl)-1-benzyl-1*H*benzo[*d*]imidazole (**16**) (80.7 mg, 280 µmol), 2,2'-(2,5-dimethyl-1,4-phenylene)di(4'',4'',5'',5''tetramethyl-1'',3'',2''-dioxaborolane) (**29**) (50.0 mg, 140 µmol), potassium carbonate (96.7 mg, 700 µmol, milled) and Pd(PPh₃)₄ (16.2 mg, 10 mol%) were added to a three-neck flask and the flask was evacuated for 5 min. Anhydrous ethanol (1 mL, dry), water (1 mL, degassed) and anhydrous tetrahydrofuran (3 mL) were added and the reaction mixture was stirred at 80 °C for 23 h. The solution was cooled to room temperature and the precipitate was collected by filtration and washed with cold tetrahydrofuran (250 mL, cooled to -18 °C) to obtain the product as a beige solid (56.0 mg, 108 µmol, 77%).

¹**H NMR** (500 MHz, CDCl₃, 298 K) δ (ppm): 8.75 (dd, ${}^{4}J = 2.3$ Hz, ${}^{5}J = 0.8$ Hz, 2H, H_m), 8.50 (dd, ${}^{3}J = 8.1$ Hz, ${}^{5}J = 0.8$ Hz, 2H, H_{j}), 7.89 (dd, ${}^{3}J = 8.1$ Hz, ${}^{4}J = 2.3$ Hz, 2H, H_{k}), 7.87-7.85 (m, 2H, H_{f}), 7.49-7.47 (m, 2H, H_{c}), 7.37 (td, ${}^{3}J = 7.2$ Hz, ${}^{4}J = 1.3$ Hz, 2H, H_{d}), 7.34 (td, ${}^{3}J = 7.2$ Hz, ${}^{4}J = 1.3$ Hz, 2H, H_{e}), 7.28 (s, 2H, H_{q}), 4.36 (s, 6H, H_{a}), 2.38 (s, 6H, H_{p}).

¹³**C NMR** (125 MHz, CDCl₃, 298 K) δ (ppm): 150.2 (*C_h*), 149.2 (*C_i*), 148.6 (*C_m*), 142.7 (*C_g*), 137.7 (*C_n*), 137.4 (*C_b*), 137.3 (*C_k*), 136.9 (*C_i*), 133.5 (*C_o*), 132.2 (*C_q*), 124.1 (*C_j*), 123.4 (*C_d*), 122.7 (*C_e*), 120.0 (*C_f*), 110.0 (*C_c*), 32.8 (*C_a*), 19.9 (*C_p*).

EI-MS *m/z*: 520.23624 (calculated for C₃₄H₂₈N₆: 520.23754).



Figure S62. ¹H NMR spectrum (500 MHz, CDCl₃, 298 K) of 1,4-di(6"-(1'-methyl-1*H*-benzo[*d*]imidazol-2'-yl)pyridin-3"-yl)-2,5-dimethylbenzene (**30**).



Figure S63. ¹³C NMR spectrum (125 MHz, $CDCl_3$, 298 K) of 1,4-di(6"-(1'-methyl-1*H*-benzo[*d*]imidazol-2'-yl)pyridin-3"-yl)-2,5-dimethylbenzene (**30**).



Figure S64. ¹H-¹H COSY NMR spectrum (500 MHz, CDCl₃, 298 K) of 1,4-di(6''-(1'-methyl-1*H*-benzo[*d*]imidazol-2'-yl)pyridin-3''-yl)-2,5-dimethylbenzene (**30**).



Figure S65. ¹H-¹³C HSQC NMR spectrum (500 MHz/125 MHz, CDCl₃, 298 K) of 1,4-di(6"-(1'-methyl-1*H*-benzo[*d*]imidazol-2'-yl)pyridin-3"-yl)-2,5-dimethylbenzene (**30**).



Figure S66. ¹H-¹³C HMBC NMR spectrum (500 MHz/125 MHz, CDCl₃, 298 K) of 1,4-di(6"-(1'-methyl-1*H*-benzo[*d*]imidazol-2'-yl)pyridin-3"-yl)-2,5-dimethylbenzene (**30**).

2.4.5 2,2'-(2,5-Difluoro-1,4-phenylene)di(4",4",5",5"-tetramethyl-1",3",2"-dioxaborolane) (**31**)



Adapted from Severin and co-workers.^[7] 1,4-Dibromo-2,5-difluorobenzene (514 mg, 1.89 mmol), bis(pinacolato)diboron (1.00 g, 3.94 mmol), potassium acetate (925 mg, 9.42 mmol) and Pd(dppf)Cl₂·CH₂Cl₂ (77.2 mg, 5 mol%) were added to a three-neck flask and the flask was evacuated for 5 min. Anhydrous 1,4-dioxane (20 mL) was added and the mixture was stirred at 90 °C for 23 h. The reaction mixture was cooled to room temperature and the solvent was removed *in vacuo*. Purification by flash chromatography (silica gel, 33% dichloromethane/cyclohexane) gave the product as a colourless solid (143 mg, 391 μ mol, 21%).

¹**H NMR** (500 MHz, CDCl₃, 298 K) δ (ppm): 7.35 (t, ³*J* = 6.5 Hz, 2H, *H*_b), 1.35 (s, 24H, *H*_g).

¹³**C NMR** (125 MHz, CDCl₃, 298 K) δ (ppm): 162.5 (d, ${}^{1}J_{CF}$ = 249.2 Hz, *C_c*), 122.4 (dd, ${}^{2}J_{CF}$ = 20.5 Hz, ${}^{3}J_{CF}$ = 13.7 Hz, *C_b*), 84.3 (*C_t*), 24.8 (*C_g*). A signal for C_a was not observed.

¹¹**B NMR** (160 MHz, CDCl₃, 298 K) δ (ppm): 29.5 (B_e).

¹⁹**F NMR** (470 MHz, CDCl₃, 298 K) δ (ppm): -114.2 (F_d).

EI-MS *m/z*: 366.19828 (calculated for C₁₈H₂₆¹¹B₂¹⁹F₂O₄: 366.19853).



Figure S67. ¹H NMR spectrum (500 MHz, CDCl₃, 298 K) of 2,2'-(2,5-difluoro-1,4-phenylene)di(4",4",5",5"-tetramethyl-1",3",2"-dioxaborolane) (**31**).



Figure S68. ¹³C NMR spectrum (125 MHz, CDCl₃, 298 K) of 2,2'-(2,5-difluoro-1,4-phenylene)di(4",4",5",5"-tetramethyl-1",3",2"-dioxaborolane) (**31**).



Figure S69. ¹¹B NMR spectrum (160 MHz, CDCl₃, 298 K) of 2,2'-(2,5-difluoro-1,4-phenylene)di(4",4",5",5"-tetramethyl-1",3",2"-dioxaborolane) (**31**).



Figure S70. ¹⁹F NMR spectrum (470 MHz, CDCl₃, 298 K) of 2,2'-(2,5-difluoro-1,4-phenylene)di(4",4",5",5"-tetramethyl-1",3",2"-dioxaborolane) (**31**).



Figure S71. ¹H-¹H COSY NMR spectrum (500 MHz, CDCl₃, 298 K) of 2,2'-(2,5-difluoro-1,4-phenylene)di(4",4",5",5"-tetramethyl-1",3",2"-dioxaborolane) (**31**).



Figure S72. ¹H-¹³C HSQC NMR spectrum (500 MHz/125 MHz, CDCl₃, 298 K) of 2,2'-(2,5-difluoro-1,4-phenylene)di(4",4",5",5"-tetramethyl-1",3",2"-dioxaborolane) (**31**).



Figure S73. ¹H-¹³C HMBC NMR spectrum (500 MHz/125 MHz, CDCl₃, 298 K) of 2,2'-(2,5-difluoro-1,4-phenylene)di(4",4",5",5"-tetramethyl-1",3",2"-dioxaborolane) (**31**).

2.4.6 1,4-Di(6"-(1'-methyl-1*H*-benzo[*d*]imidazol-2'-yl)pyridin-3"-yl)-2,5-difluorobenzene (**32**)



Adapted from Lusby and co-workers.^[8] 2-(5'-Bromopyridin-2'-yl)-1-benzyl-1*H*-benzo[*d*]imidazole (**16**) (80.7 mg, 280 µmol), 2,2'-(2,5-difluoro-1,4-phenylene)di(4",4",5",5"-tetramethyl-1",3",2"-dioxaborolane) (**31**) (51.2 mg, 140 µmol), potassium carbonate (96.7 mg, 700 µmol, milled) and Pd(PPh₃)₄ (16.2 mg, 10 mol%) were added to a three-neck flask and the flask was evacuated for 5 min. Anhydrous ethanol (1 mL), water (1 mL, degassed) and anhydrous tetrahydrofuran (3 mL) were added and the reaction mixture was stirred at 80 °C for 23 h. The solution was cooled to room temperature and the precipitate was collected by filtration and washed with cold tetrahydrofuran (200 mL, cooled to -18 °C) to obtain the product as a brown solid (38.7 mg, 73.2 µmol, 52%).

¹**H NMR** (500 MHz, TFA-*d*₁, 298 K) δ (ppm): 9.46 (d, ${}^{4}J$ =2.1 Hz, 2H, *H_m*), 8.88 (dd, ${}^{3}J$ = 8.2 Hz, ${}^{4}J$ = 2.1 Hz, 2H, *H_k*), 8.61 (d, ${}^{3}J$ = 8.2 Hz, 2H, *H_j*), 7.97-7.83 (m, 4H, *H_{c,d,e,f}*), 7.72 (t, ${}^{3}J$ = 8.2 Hz, 2H, *H_q*), 4.37 (s, 6H, *H_a*).

¹³**C NMR** (125 MHz, TFA-*d*₁, 298 K) δ (ppm): 157.7 (d, ${}^{1}J_{CF}$ = 251.6 Hz, *C*_o), 150.3 (*C*_m), 145.3 (*C*_k), 144.5 (*C*_h), 139.1 (*C*_i), 138.6 (*C*_i), 135.6 (*C*_b), 132.4 (*C*_g), 131.8 (*C*_{d/e}), 131.3 (*C*_{d/e}), 130.9 (*C*_j), 127.7 (*C*_n), 120.5 (dd, ${}^{2}J_{CF}$ = 19.4 Hz, ${}^{3}J_{CF}$ = 10.2 Hz, *C*_q), 116.8 (*C*_{c/f}), 114.7 (*C*_{c/f}), 34.8 (*C*_a).

¹⁹**F NMR** (470 MHz, TFA-*d*₁, 298 K) δ (ppm): -120.7 (F_ρ).

EI-MS m/z: 528.18648 (calculated for C₃₂H₂₂¹⁹F₂N₆: 528.18740).



Figure S74. ¹H NMR spectrum (500 MHz, CDCl₃, 298 K) of 1,4-di(6"-(1'-methyl-1*H*-benzo[*d*]imidazol-2'-yl)pyridin-3"-yl)-2,5-difluorobenzene (**32**).



Figure S75. ¹³C NMR spectrum (125 MHz, $CDCl_3$, 298 K) of 1,4-di(6"-(1'-methyl-1*H*-benzo[*d*]imidazol-2'-yl)pyridin-3"-yl)-2,5-difluorobenzene (**32**).



Figure S76. ¹⁹F NMR spectrum (471 MHz, $CDCl_3$, 298 K) of 1,4-di(6"-(1'-methyl-1*H*-benzo[*d*]imidazol-2'-yl)pyridin-3"-yl)-2,5-difluorobenzene (**32**).



Figure S77. ¹H-¹H COSY NMR spectrum (500 MHz, CDCl₃, 298 K) of 1,4-di(6''-(1'-methyl-1*H*-benzo[*d*]imidazol-2'-yl)pyridin-3''-yl)-2,5-difluorobenzene (**32**).



Figure S78. ¹H-¹³C HSQC NMR spectrum (500 MHz/125 MHz, CDCl₃, 298 K) of 1,4-di(6"-(1'-methyl-1*H*-benzo[*d*]imidazol-2'-yl)pyridin-3"-yl)-2,5-difluorobenzene (**32**).



Figure S79. ¹H-¹³C HMBC NMR spectrum (500 MHz/125 MHz, CDCl₃, 298 K) of 1,4-di(6"-(1'- methyl-1*H*-benzo[*d*]imidazol-2'-yl)pyridin-3"-yl)-2,5-difluorobenzene (**32**).

2.4.7 1,4-Bis(4"-methyl-6"-(1'-methyl-1*H*-benzo[*d*]imidazol-2'-yl)pyridin-3"-yl)benzene (**33**)



Adapted from Lusby and co-workers.^[8] 2-(5'-Bromo-4'-methylpyridin-2'-yl)-1-methyl-1*H*-benzo[*d*]imidazole (**21**) (32.2 mg, 107 µmol), 1,4-phenylenediboronic acid (8.84 mg, 53.4 µmol), potassium carbonate (36.9 mg, 267 µmol, milled) and Pd(PPh₃)₄ (5.34 mg, 5 mol%) were added to a three-neck flask and the flask was evacuated for 30 min. Anhydrous ethanol (370 µL), water (370 µL, degassed) and anhydrous tetrahydrofuran (860 µL) were added and the reaction mixture was stirred at 80 °C for 21 h. The solution was cooled to room temperature and the precipitate was collected by filtration and washed with cold (0 °C) tetrahydrofuran (20 mL). The product was obtained as a colourless solid (16.8 mg, 32.3 µmol, 30%).

¹**H NMR** (600 MHz, CDCl₃, 298 K) δ (ppm): 8.62 (s, 2H, H_m), 8.38 (s, 2H, H_j), 7.87 (unres. dd, 2H, H_i), 7.53 (s, 2H, H_p), 7.48 (unres. dd, 2H, H_c), 7.37 (td, ³*J* = 7.2 Hz, ⁴*J* = 1.2 Hz, 2H, H_d), 7.34 (td, ³*J* = 7.8 Hz, ⁴*J* = 1.4 Hz, 2H, H_e), 4.35 (s, 6H, H_a), 2.49 (s, 6H, H_n).

¹³**C NMR** (151 MHz, CDCl₃, 298 K) δ (ppm): 150.3 (*C_h*), 149.4 (*C_i*), 148.9 (*C_m*), 145.6 (*C_k*), 142.6 (*C_g*), 137.4 (*C_b*), 137.3 (*C_o*), 137.1 (*C_i*), 129.5 (*C_p*), 126.1 (*C_j*), 123.3 (*C_d*), 122.7 (*C_e*), 120.0 (*C_t*), 110.0 (*C_c*), 32.8 (*C_a*), 20.0 (*C_n*).

EI-MS *m/z*: 520.23705 (calculated for C₃₄H₂₈N₆: 520.23754).



Figure S80. ¹H NMR spectrum (600 MHz, CDCl₃, 298 K) of 1,4-bis(4"-methyl-6"-(1'-methyl-1*H*-benzo[*d*]imidazol-2'-yl)pyridin-3"-yl)benzene (**33**).



Figure S81. ¹³C NMR spectrum (151 MHz, CDCl₃, 298 K) of 1,4-bis(4"-methyl-6"-(1'-methyl-1*H*-benzo[*d*]imidazol-2'-yl)pyridin-3"-yl)benzene (**33**).



Figure S82. ¹H-¹H COSY NMR spectrum (600 MHz, CDCl₃, 298 K) of 1,4-bis(4"-methyl-6"-(1'-methyl-1*H*-benzo[*d*]imidazol-2'-yl)pyridin-3"-yl)benzene (**33**).



Figure S83. ¹H-¹³C HSQC NMR spectrum (600 MHz/151 MHz, CDCl₃, 298 K) of 1,4-bis(4"-methyl-6"-(1'-methyl-1*H*-benzo[*d*]imidazol-2'-yl)pyridin-3"-yl)benzene (**33**).



Figure S84. ¹H-¹³C HMBC NMR spectrum (600 MHz/151 MHz, CDCl₃, 298 K) of 1,4-bis(4"-methyl-6"-(1'-methyl-1*H*-benzo[*d*]imidazol-2'-yl)pyridin-3"-yl)benzene (**33**).

3 Zn₄L₆ Metal-Organic Cages

Metal-organic cages based on Zn^{II} were self-assembled using six equivalents of the appropriate ligand and four equivalents zinc(II) triflate. The cages were prepared *in situ* and characterisation using NMR spectroscopy and ESI-MS were consistent with the self-assembly of Zn_4L_6 cages. However, a discrete cage did not form with ligand **14** (Section 3.1). Zn_2L_3 structures were also self-assembled with ligands **22**, **27** and **32** (Section 3.7, 3.8 and 3.10).

3.1 Attempted Self-Assembly of Cage 34



Zinc(II) triflate (1.95 mg, 5.36 μ mol) and 1,2-bis(6"-(1'-methyl-1*H*-imidazol-2'-yl)pyridin-3"-yl)ethyne (**14**) (2.74 mg, 8.05 μ mol) were dissolved in CD₃CN (500 μ L) in an attempt to prepare cage **34** *in situ*. However, a discrete cage did not form, even after heating for 117 h at 50 °C, and therefore, assignment of the ¹H NMR spectrum was not possible (Figure S85). In addition, signals consistent with the expected cage were not observed in the ESI mass spectrum.



Figure S85. ¹H NMR spectrum (200 MHz, CD₃CN, 298 K) of the self-assembly for cage 34.



Zinc(II) triflate (2.02 mg, 5.56 μ mol) and 6',6"-di(1-benzyl-1*H*-benzo[*d*]imidazol-2-yl)-3',3"-bipyridine (**26**) (4.70 mg, 8.25 μ mol) were dissolved in CD₃CN (500 μ L) and heated for 24 h at 50 °C to prepare cage **35** *in situ*.

¹**H NMR** (500 MHz, CD₃CN, 298 K) δ (ppm): 8.16 (dd, ${}^{4}J$ = 2.2 Hz, ${}^{5}J$ = 0.7 Hz, 12H, H_{q}), 7.78 (d, ${}^{3}J$ = 8.4 Hz, 12H, H_{n}), 7.60 (dd, ${}^{3}J$ = 8.4 Hz, ${}^{4}J$ = 2.2 Hz, 12H, H_{o}), 7.58 (dt, ${}^{3}J$ = 8.4 Hz, ${}^{4}J$ = 1.0 Hz, 12H, H_{g}), 7.42 (ddd, ${}^{3}J$ = 8.4 Hz, ${}^{3}J$ = 7.3 Hz, ${}^{4}J$ = 1.0 Hz, 12H, H_{h}), 7.37-7.32 (m, 24H, H_{b}), 7.31-7.27 (m, 12H, H_{a}), 7.22 (ddd, ${}^{3}J$ = 8.4 Hz, ${}^{3}J$ = 7.3 Hz, ${}^{4}J$ = 1.0 Hz, 12H, H_{h}), 7.14-7.11 (m, 24H, H_{c}), 6.95 (dt, ${}^{3}J$ = 8.4 Hz, ${}^{4}J$ = 1.0 Hz, 12H, H_{j}), 6.06 (d, ${}^{2}J$ = 18.0 Hz, 12H, H_{e}).

¹³**C NMR** (125 MHz, CD₃CN, 298 K) δ (ppm): 149.7 (*C_q*), 149.5 (*C_l*), 143.0 (*C_m*), 141.9 (*C_o*), 138.8 (*C_k*), 138.5 (*C_l*), 136.7 (*C_p*), 135.6 (*C_d*), 130.3 (*C_b*), 129.3 (*C_a*), 127.4 (*C_h*), 127.1 (*C_l*), 126.8 (*C_c*), 124.4 (*C_n*), 118.1 (*C_j*), 113.7 (*C_g*), 49.7 (*C_e*).

¹⁹F NMR (471 MHz, CD₃CN, 298 K, C₆F₆) δ (ppm): -79.3 (OTf⁻).

HRMS (ESI): m/z = 2283.4282 [**35** + 6OTf]²⁺, 1472.6331 [**35** + 5OTf]³⁺, 1067.2363 [**35** + 4OTf]⁴⁺, 823.7981 [**35** + 3OTf]⁵⁺, 661.5064 [**35** + 2OTf]⁶⁺.

While the signal at m/z 661.5064 is consistent with the 6+ charge for the cage, the theoretical isotopic pattern does not match, possibly due to fragmentation in the gas phase or overlap with other signals.



Figure S86. ¹H NMR spectrum (500 MHz, CD₃CN, 298 K) of cage 35.



Figure S87. ¹³C NMR spectrum (125 MHz, CD₃CN, 298 K) of cage 35.



Figure S88. ¹H-¹H COSY NMR spectrum (500 MHz, CD₃CN, 298 K) of cage 35.



Figure S89. ¹H-¹³C HSQC NMR spectrum (500 MHz/125 MHz, CD₃CN, 298 K) of cage 35.



Figure S90. ¹H-¹³C HMBC NMR spectrum (500 MHz/125 MHz, CD₃CN, 298 K) of cage 35.



Figure S91. ^{19}F NMR spectrum (471 MHz, CD_3CN, 298 K, C_6F_6) of cage 35.





1066.2374

1066.2

1066.6

1067.0

1067.4

1065.9876

1065.7375

1065.8

1065.4877

1065.4

40-

20-

0-

1067.8

1068.2 m/z (Da)

1068.7366

1068.6

1068.9866

1069.0

1069.2366

1069.4

1069.4866 1069.7366

1069.8

1070.2367

1070.2

1070.6

3.3 Cage 36



Zinc(II) triflate (1.96 mg, 5.39 μ mol) and 1,2-di(6"-(1'-benzyl-1*H*-benzo[*d*]imidazole-2'-yl)pyridin-3"-yl)ethyne (**19**) (4.79 mg, 8.08 μ mol) were dissolved in CD₃CN (500 μ L) to prepare cage **36** *in situ*.

¹**H NMR** (600 MHz, CD₃CN, 298 K) δ (ppm): 8.02 (dd, ${}^{3}J$ = 8.5 Hz, ${}^{5}J$ = 0.7 Hz, 12H, $H_{\rm h}$), 7.95 (dd, ${}^{3}J$ = 8.5 Hz, ${}^{4}J$ = 2.0 Hz, 12H, $H_{\rm o}$), 7.84 (dd, ${}^{4}J$ = 2.0 Hz, ${}^{5}J$ = 0.7 Hz, 12H, $H_{\rm q}$), 7.69 (unres. dt, 12H, $H_{\rm g}$), 7.50 (ddd, ${}^{3}J$ = 8.5 Hz, ${}^{3}J$ = 7.4 Hz, ${}^{4}J$ = 0.8 Hz, 12H, $H_{\rm h}$), 7.31 (t, ${}^{3}J$ = 7.7 Hz, 24H, $H_{\rm b}$), 7.25 (ddd, ${}^{3}J$ = 8.3 Hz, ${}^{3}J$ = 7.4 Hz, ${}^{4}J$ = 0.8 Hz, 12H, $H_{\rm h}$), 7.20 (t, ${}^{3}J$ = 7.4 Hz, 12H, $H_{\rm a}$), 7.06 (d, ${}^{3}J$ = 7.6 Hz, 24H, $H_{\rm c}$), 6.99 (dt, ${}^{3}J$ = 8.3 Hz, ${}^{4}J$ = 0.8 Hz, 12H, $H_{\rm h}$), 6.06-5.98 (m, 24H, $H_{\rm e}$).

¹³**C NMR** (151 MHz, CD₃CN, 298 K) δ (ppm): 152.3 (*C*_q), 148.8 (*C*_l), 145.0 (*C*_o), 142.7 (*C*_m), 138.7 (*C*_k), 138.6 (*C*_l), 135.4 (*C*_d), 130.3 (*C*_b), 129.3 (*C*_a), 127.8 (*C*_h), 127.4 (*C*_l), 126.8 (*C*_c), 124.7 (*C*_n), 122.8 (*C*_p), 118.3 (*C*_j), 113.7 (*C*_g), 91.4 (*C*_r), 50.1 (*C*_e).

¹⁹F NMR (471 MHz, CD₃CN, 298 K, C₆F₆) δ (ppm): -79.5 (OTf⁻).

HRMS (ESI): $m/z = 2355.4280 [36 + 6OTf]^{2+}$, 1520.9655 [36 + 5OTf]^{3+}, 1103.4861 [36 + 4OTf]^{4+}, 852.7982 [36 + 3OTf]⁵⁺, 686.1721 [36 + 2OTf]⁶⁺.



Figure S93. ¹H NMR spectrum (600 MHz, CD₃CN, 298 K) of cage 36.



Figure S94. ¹³C NMR spectrum (151 MHz, CD₃CN, 298 K) of cage 36.



Figure S95. ¹H-¹H COSY NMR spectrum (600 MHz, CD₃CN, 298 K) of cage 36.



Figure S96. ¹H-¹³C HSQC NMR spectrum (600 MHz/151 MHz, CD₃CN, 298 K) of cage 36.



Figure S97. ¹H-¹³C HMBC NMR spectrum (600 MHz/151 MHz, CD₃CN, 298 K) of cage 36.



Figure S98. ^{19}F NMR spectrum (471 MHz, CD_3CN, 298 K, C_6F_6) of cage 36.



Figure S99. a) High resolution ESI mass spectrum of cage **36** and b) isotopic patterns of cage **36**: experimental (top) and theoretical (bottom).

S65



Zinc(II) triflate (2.08 mg, 5.72 μ mol) and 1,4-di(6"-(1'-benzyl-1*H*-benzo[*d*]imidazol-2'yl)pyridin-3"-yl)benzene (**28**) (5.53 mg, 8.58 μ mol) were dissolved in CD₃CN (500 μ L) and heated for 74.5 h at 50 °C to prepare cage **37** *in situ*.

¹**H NMR** (500 MHz, CD₃CN, 298 K) δ (ppm): 8.38 (dd, ${}^{3}J$ = 8.5 Hz, ${}^{4}J$ = 2.3 Hz, 12H, H_{0}), 8.18 (dd, ${}^{3}J$ = 8.5 Hz, ${}^{5}J$ = 0.6 Hz, 12H, H_{0}), 8.18 (dd, ${}^{3}J$ = 8.5 Hz, ${}^{5}J$ = 0.6 Hz, 12H, H_{q}), 7.74 (dt, ${}^{3}J$ = 8.5 Hz, ${}^{4}J$ = 0.8 Hz, 12H, H_{g}), 7.51 (ddd, ${}^{3}J$ = 8.5 Hz, ${}^{3}J$ = 7.3 Hz, ${}^{4}J$ = 0.9 Hz, 12H, H_{h}), 7.26 (ddd, ${}^{3}J$ = 8.3 Hz, ${}^{3}J$ = 7.3 Hz, ${}^{4}J$ = 0.8 Hz, 12H, H_{h}), 7.26 (ddd, ${}^{3}J$ = 8.3 Hz, ${}^{3}J$ = 7.3 Hz, ${}^{4}J$ = 0.8 Hz, 12H, H_{h}), 7.09 (s, 24H, H_{s}), 7.02 (dt, ${}^{3}J$ = 8.3 Hz, ${}^{4}J$ = 0.8 Hz, 12H, H_{j}), 6.00 (s, 24H, H_{e}).

¹³**C NMR** (125 MHz, CD₃CN, 298 K) δ (ppm): 149.1 (*C*_{*l*}), 147.8 (*C*_{*q*}), 142.0 (*C*_{*m*}), 139.9 (*C*_{*o*}), 139.1 (*C*_{*p*}), 138.8 (*C*_{*l*}), 138.6 (*C*_{*k*}), 136.0 (*C*_{*l*}), 135.4 (*C*_{*d*}), 130.1 (*C*_{*b*}), 129.0 (*C*_{*a*}), 128.4 (*C*_{*s*}), 127.4 (*C*_{*h*}), 127.0 (*C*_{*i*}), 126.6 (*C*_{*c*}), 124.9 (*C*_{*n*}), 118.2 (*C*_{*j*}), 113.3 (*C*_{*g*}), 49.9 (*C*_{*e*}).

¹⁹F NMR (471 MHz, CD₃CN, 298 K, C₆F₆) δ (ppm): -79.5 (OTf⁻).

HRMS (ESI): m/z = 2512.0241 [**37** + 6OTf]²⁺, 1625.0290 [**37** + 5OTf]³⁺, 1181.2836 [**37** + 4OTf]⁴⁺, 915.2358 [**37** + 3OTf]⁵⁺, 737.8710 [**37** + 2OTf]⁶⁺.



Figure S100. ¹H NMR spectrum (500 MHz, CD₃CN, 298 K) of cage 37.



Figure S101. ¹³C NMR spectrum (125 MHz, CD₃CN, 298 K) of cage 37.



Figure S102. ¹H-¹H COSY NMR spectrum (500 MHz, CD₃CN, 298 K) of cage 37.



Figure S103. ¹H-¹³C HSQC NMR spectrum (500 MHz/125 MHz, CD₃CN, 298 K) of cage 37.



Figure S104. ¹H-¹³C HMBC NMR spectrum (500 MHz/125 MHz, CD₃CN, 298 K) of cage 37.



Figure S105. ¹⁹F NMR spectrum (471 MHz, CD₃CN, 298 K, C₆F₆) of cage 37.



Figure S106. a) High resolution ESI mass spectrum of cage **37** and b) isotopic patterns of cage **37**: experimental (top) and theoretical (bottom).

3.5 Cage 38



Zinc(II) triflate (1.97 mg, 5.42 μ mol) and 6',6"-di(1-methyl-1*H*-benzo[*d*]imidazol-2-yl)-3',3"-bipyridine (**24**) (3.39 mg, 8.14 μ mol) were dissolved in CD₃CN (500 μ L) and heated for 22 h at 50 °C to prepare cage **38** *in situ*.

¹**H NMR** (600 MHz, CD₃CN, 298 K) δ (ppm): 8.34 (dd, ${}^{3}J$ = 8.4 Hz, ${}^{5}J$ = 0.6 Hz, 12H, *H*_j), 8.03 (dd, ${}^{4}J$ = 2.3 Hz, ${}^{5}J$ = 0.6 Hz, 12H, *H*_m), 7.84 (dd, ${}^{3}J$ = 8.4 Hz, ${}^{4}J$ = 2.3 Hz, 12H, *H*_k), 7.77 (unres. ddd, 12H, *H*_c), 7.47 (ddd, ${}^{3}J$ = 8.5 Hz, ${}^{3}J$ = 7.2 Hz, ${}^{4}J$ = 0.8 Hz, 12H, *H*_d), 7.11 (ddd, ${}^{3}J$ = 8.3 Hz, ${}^{3}J$ = 7.2 Hz, ${}^{4}J$ = 0.8 Hz, 12H, *H*_e), 6.92 (unres. ddd, 12H, *H*_i), 4.26 (s, 36H, *H*_a).

¹³**C NMR** (151 MHz, CD₃CN, 298 K) δ (ppm): 149.5 (*C_h*), 149.3 (*C_m*), 143.7 (*C_i*), 141.3 (*C_k*), 138.7 (*C_b*), 138.6 (*C_g*), 136.3 (*C_i*), 126.8 (*C_d*), 126.5 (*C_e*), 125.0 (*C_j*), 118.2 (*C_i*), 113.4 (*C_c*), 34.4 (*C_a*).

¹⁹**F NMR** (471 MHz, CD₃CN, 298 K, C₆F₆) δ (ppm): -79.6 (OTf⁻).

HRMS (ESI): $m/z = 1827.2372 [38 + 60Tf]^{2+}, 1168.5063 [38 + 50Tf]^{3+}, 839.1410 [38 + 40Tf]^{4+}, 641.3225 [38 + 30Tf]^{5+}, 509.4439 [38 + 20Tf]^{6+}.$

While the signal at m/z 509.4439 is consistent with the 6+ charge for the cage, the theoretical isotopic pattern does not match, possibly due to fragmentation in the gas phase or overlap with other signals.



Figure S107. ¹H NMR spectrum (600 MHz, CD₃CN, 298 K) of cage 38.



Figure S108. ¹³C NMR spectrum (151 MHz, CD₃CN, 298 K) of cage 38.



Figure S109. ¹H-¹H COSY NMR spectrum (600 MHz, CD₃CN, 298 K) of cage 38.


Figure S110. ¹H-¹³C HSQC NMR spectrum (600 MHz/151 MHz, CD₃CN, 298 K) of cage 38.



Figure S111. ¹H-¹³C HMBC NMR spectrum (600 MHz/151 MHz, CD₃CN, 298 K) of cage 38.



Figure S112. ¹⁹F NMR spectrum (471 MHz, CD_3CN , 298 K, C_6F_6) of cage 38.



Figure S113. a) High resolution ESI mass spectrum of cage 38 and b) isotopic patterns of cage 38: experimental (top) and theoretical (bottom).



Zinc(II) triflate (2.04 mg, 5.61 μ mol) and 1,2-di(6"-(1'-methyl-1*H*-benzo[*d*]imidazole-2'-yl)pyridin-3"-yl)ethyne (**17**) (3.71 mg, 8.42 μ mol) were dissolved in CD₃CN (500 μ L) to prepare cage **39** *in situ*.

¹**H NMR** (500 MHz, CD₃CN, 298 K) δ (ppm): 8.43 (dd, ${}^{3}J = 8.6$ Hz, ${}^{5}J = 0.8$ Hz, 12H, *H*_j), 8.25 (dd, ${}^{3}J = 8.6$ Hz, ${}^{4}J = 2.0$ Hz, 12H, *H*_k), 7.99 (dd, ${}^{4}J = 2.0$ Hz, ${}^{5}J = 0.8$ Hz, 12H, *H*_m), 7.77 (dt, ${}^{3}J = 8.6$ Hz, ${}^{4}J = 0.9$ Hz, 12H, *H*_c), 7.47 (ddd, ${}^{3}J = 8.6$ Hz, ${}^{3}J = 7.3$ Hz, ${}^{4}J = 0.9$ Hz, 12H, *H*_d), 7.08 (ddd, ${}^{3}J = 8.4$ Hz, ${}^{3}J = 7.3$ Hz, ${}^{4}J = 0.9$ Hz, 12H, *H*_d), 12H, *H*_b), 4.29 (s, 36H, *H*_a).

¹³**C NMR** (125 MHz, CD₃CN, 298 K) δ (ppm): 152.4 (*C_m*), 149.2 (*C_h*), 144.6 (*C_k*), 143.4 (*C_i*), 138.7 (*C_g*), 138.6 (*C_b*), 127.2 (*C_d*), 126.8 (*C_e*), 125.2 (*C_j*), 122.9 (*C_i*), 118.2 (*C_i*), 113.4 (*C_c*), 91.5 (*C_n*), 34.7 (*C_a*).

¹⁹**F NMR** (471 MHz, CD₃CN, 298 K, C₆F₆) δ (ppm): -79.6 (OTf⁻).

HRMS (ESI): $m/z = 1899.7373 [39 + 60Tf]^{2+}, 1216.1738 [39 + 50Tf]^{3+}, 875.1411 [39 + 40Tf]^{4+}, 670.3225 [39 + 30Tf]^{5+}, 533.4437 [39 + 20Tf]^{6+}.$

While the signal at m/z 533.4437 is consistent with the 6+ charge for the cage, the theoretical isotopic pattern does not match, possibly due to fragmentation in the gas phase or overlap with other signals.



Figure S114. ¹H NMR spectrum (500 MHz, CD₃CN, 298 K) of cage 39.



Figure S115. $^{\rm 13}C$ NMR spectrum (125 MHz, CD_3CN, 298 K) of cage 39.



Figure S116. ¹H-¹H COSY NMR spectrum (500 MHz, CD₃CN, 298 K) of cage 39.



Figure S117. ¹H-¹³C HSQC NMR spectrum (500 MHz/125 MHz, CD₃CN, 298 K) of cage **39**.



Figure S118. ¹H-¹³C HMBC NMR spectrum (500 MHz/125 MHz, CD₃CN, 298 K) of cage **39**.



Figure S119. ¹⁹F NMR spectrum (471 MHz, CD₃CN, 298 K, C_6F_6) of cage 39.



Figure S120. a) High resolution ESI mass spectrum of cage **39** and b) isotopic patterns of cage **39**: experimental (top) and theoretical (bottom).

3.7 Cage 40a and Helicate 40b

Zinc(II) triflate (2.03 mg, 5.58 μ mol) and 1,4-di(6"-(1'-methyl-1*H*-benzo[*d*]imidazol-2'yl)pyridin-3"-yl)benzene (**27**) (4.13 mg, 8.38 μ mol) were dissolved in CD₃CN (500 μ L) and heated for 117 h at 50 °C to prepare cage **40a** and helicate **40b** *in situ*.

3.7.1 Cage **40a**



¹**H NMR** (600 MHz, CD₃CN, 298 K) δ (ppm): 8.54 (unres. dd, 12H, H_j), 8.53 (unres. dd, 12H, H_m), 8.24 (unres. dd, 12H, H_k), 7.76 (d, ³*J* = 8.5 Hz, 12H, H_c), 7.48 (s, 24H, H_o), 7.46 (unres. ddd, 12H, H_d), 7.07 (unres. ddd, 12H, H_e), 6.78 (d, ³*J* = 8.4 Hz, 12H, H_f), 4.29 (s, 36H, H_a).

¹³**C NMR** (151 MHz, CD₃CN, 298 K) δ (ppm): 149.6 (*C_h*), 148.2 (*C_k*), 142.9 (*C_i*), 139.8 (*C_j*), 139.1 (*C_n*), 138.8 (*C_b*), 138.7 (*C_g*), 136.5 (*C_i*), 128.9 (*C_o*), 126.7 (*C_d*), 126.4 (*C_e*), 125.3 (*C_m*), 118.1 (*C_t*), 113.3 (*C_c*), 34.7 (*C_a*).

¹⁹**F NMR** (471 MHz, CD₃CN, 298 K, C₆F₆) δ (ppm): -79.6 (OTf⁻).

HRMS (ESI): m/z = 2055.3310 [**40a** + 6OTf]²⁺, 1320.5694 [**40a** + 5OTf]³⁺, 953.1881 [**40a** + 4OTf]⁴⁺, 732.7601 [**40a** + 3OTf]⁵⁺, 585.8077 [**40a** + 2OTf]⁶⁺.

While the signal at m/z 585.8077 is consistent with the 6+ charge for the cage, the theoretical isotopic pattern does not match, possibly due to fragmentation in the gas phase or overlap with other signals.

3.7.2 Helicate 40b



¹**H NMR** (600 MHz, CD₃CN, 298 K) δ (ppm): 8.72 (d, ${}^{3}J$ = 8.3 Hz, 6H, H_{j}), 8.67 (dd, ${}^{3}J$ = 8.3 Hz, ${}^{4}J$ = 1.8 Hz, 6H, H_{k}), 7.87 (d, ${}^{3}J$ = 8.4 Hz, 6H, H_{c}), 7.55 (unres. ddd, 6H, H_{d}), 7.34 (s, 12H, H_{o}),

7.24 (d, ${}^{4}J$ = 1.8 Hz, 6H, H_{m}), 7.11 (unres. ddd, 6H, H_{e}), 6.85 (d, ${}^{3}J$ = 8.2 Hz, 6H, H_{f}), 4.38 (s, 18H, H_{a}).

¹³**C NMR** (151 MHz, CD₃CN, 298 K) δ (ppm): 149.9 (*C_h*), 148.2 (*C_k*), 144.0 (*C_i*), 139.0 (*C_k*), 138.9 (*C_g*), 138.8 (*C_n*), 138.8 (*C_b*), 136.3 (*C_i*), 128.8 (*C_o*), 126.9 (*C_d*), 126.8 (*C_j*), 126.4 (*C_e*), 118.5 (*C_t*), 113.4 (*C_c*), 34.9 (*C_a*).

¹⁹**F NMR** (471 MHz, CD₃CN, 298 K, C₆F₆) δ (ppm): -79.6 (OTf).

HRMS (ESI): m/z = 2054.3328 [**40b** + 3OTf]⁺, 952.6901 [**40b** + 2OTf]²⁺, 585.4759 [**40b** + 1OTf]³⁺, 401.8688 [**40b**]⁴⁺.



Figure S121. ¹H NMR spectrum (600 MHz, CD₃CN, 298 K) of cage **40a** and helicate **40b**. For clarity, only the signals for cage **40a** are labelled.



Figure S122. ¹H DOSY spectrum (600 MHz, CD₃CN, 298 K) of cage **40a** ($D = 5.23 \cdot 10^{-10} \text{ m}^2/\text{s}$) and helicate **40b** ($D = 6.95 \cdot 10^{-10} \text{ m}^2/\text{s}$).



Figure S123. ¹³C NMR spectrum (151 MHz, CD₃CN, 298 K) of cage **40a** and helicate **40b**. For clarity, only the signals for cage **40a** are labelled.



Figure S124. ¹H-¹H COSY NMR spectrum (600 MHz, CD₃CN, 298 K) of cage **40a** and helicate **40b**. For clarity, only the signals for cage **40a** are labelled.



Figure S125. ¹H-¹³C HSQC NMR spectrum (600 MHz/151 MHz, CD₃CN, 298 K) of cage **40a** and helicate **40b**. For clarity, only the signals for cage **40a** are labelled.



Figure S126. ¹H-¹³C HMBC NMR spectrum (600 MHz/151 MHz, CD₃CN, 298 K) of cage **40a** and helicate **40b**. For clarity, only the signals for cage **40a** are labelled.



Figure S127. ¹⁹F NMR spectrum (471 MHz, CD₃CN, 298 K, C_6F_6) of cage 40a and helicate 40b. For clarity, only cage 40a is depicted.



Figure S128. High resolution ESI mass spectrum of cage 40a and helicate 40b.













Figure S129. Isotopic patterns for a) cage 40a and b) helicate 40b: experimental (top) and theoretical (bottom).

3.8 Cage 41a and Helicate 41b

Zinc(II) triflate (2.06 mg, 5.67 μ mol) and 1,4-di(6"-(1'-methyl-1*H*-benzo[*d*]imidazol-2'yl)pyridin-3"-yl)-2,5-difluorobenzene (**32**) (4.36 mg, 8.25 μ mol) were dissolved in CD₃CN (500 μ L) and heated for 24 h at 50 °C to prepare cage **41a** and helicate **41b** *in situ*.

3.8.1 Cage **41a**



¹**H NMR** (600 MHz, CD₃CN, 298 K) δ (ppm): 8.48 (d, ³*J* = 8.5 Hz, 12H, *H*_j), 8.38 (d, ⁴*J* = 2.2 Hz, 12H, *H*_m), 8.33 (dd, ³*J* = 8.5 Hz, ⁴*J* = 2.2 Hz, 12H, *H*_k), 7.75 (dt, ³*J* = 8.4 Hz, ⁴*J* = 0.9 Hz, 12H, *H*_c), 7.46 (ddd, ³*J* = 8.4 Hz, ³*J* = 7.3 Hz, ⁴*J* = 0.9 Hz, 12H, *H*_d), 7.32 (t, ³*J* = 8.7 Hz, 12H, *H*_q), 7.09 (ddd, ³*J* = 8.4 Hz, ³*J* = 7.3 Hz, ⁴*J* = 0.9 Hz, 12H, *H*_e), 6.80 (dt, ³*J* = 8.4 Hz, ⁴*J* = 0.9 Hz, 12H, *H*_l), 4.27 (s, 36H, *H*_a).

¹³**C NMR** (151 MHz, CD₃CN, 298 K) δ (ppm): 157.5 ($C_{n/o}$), 155.9 ($C_{n/o}$), 150.1 (C_m), 149.4 (C_h), 143.3 (C_i), 141.6 (C_k), 138.7 (C_g), 138.6 (C_b), 133.9 (C_i), 126.9 (C_d), 126.5 (C_e), 125.3 (C_j), 118.2 (C_f), 113.4 (C_c), 34.5 (C_a).

¹⁹**F NMR** (471 MHz, CD₃CN, 298 K, C₆F₆) δ (ppm): -79.7 (OTf⁻), -122.9 (F_ρ).

HRMS (ESI): m/z = 2163.7752 [**41a** + 6OTf]²⁺, 1392.5314 [**41a** + 5OTf]³⁺, 1007.1598 [**41a** + 4OTf]⁴⁺, 775.9372 [**41a** + 3OTf]⁵⁺, 621.4557 [**41a** + 2OTf]⁶⁺, 511.8260 [**41a** + OTf]⁷⁺, 428.8537 [**41a**]⁸⁺.

While the signals at m/z 511.8260 and 428.8537 are consistent with the 7+ and 8+ charges, respectively for the cage, the theoretical isotopic pattern do not match, possibly due to fragmentation in the gas phase or overlap with other signals.

3.8.2 Helicate **41b**



Due to the low quantity of helicate present, full characterisation of the helicate by NMR spectroscopy was not possible.

HRMS (ESI): $m/z = 1007.1598 [41b + 20Tf]^{2+}, 621.4557 [41b + OTf]^{3+}, 428.8546 [41b]^{4+}.$



Figure S130. ¹H NMR spectrum (600 MHz, CD₃CN, 298 K) of cage **41a** and helicate **41b**. For clarity, only the signals for cage **41a** are labelled.



Figure S131. ¹H DOSY spectrum (600 MHz, CD₃CN, 298 K) of cage **41a** ($D = 5.22 \cdot 10^{-10} \text{ m}^2/\text{s}$) and helicate **41b** ($D = 6.57 \cdot 10^{-10} \text{ m}^2/\text{s}$).



Figure S132. ¹³C NMR spectrum (151 MHz, CD₃CN, 298 K) of cage **41a** and helicate **41b**. For clarity, only the signals for cage **41a** are labelled.



Figure S133. ¹H-¹H COSY NMR spectrum (600 MHz, CD₃CN, 298 K) of cage **41a** and helicate **41b**. For clarity, only the signals for cage **41a** are labelled.



Figure S134. ¹H-¹³C HSQC NMR spectrum (600 MHz/151 MHz, CD₃CN, 298 K) of cage **41a** and helicate **41b**. For clarity, only the signals for cage **41a** are labelled.



Figure S135. ¹H-¹³C HMBC NMR spectrum (600 MHz/151 MHz, CD₃CN, 298 K) of cage **41a** and helicate **41b**. For clarity, only the signals for cage **41a** are labelled.



Figure S136. ¹⁹F NMR spectrum (471 MHz, CD_3CN , 298 K, C_6F_6) of cage **41a** and helicate **41b**. For clarity, only cage **41a** is depicted.



Figure S137. High resolution ESI mass spectrum of cage 41a and helicate 41b.

















Figure S138. Isotopic patterns for a) cage 41a and b) helicate 41b: experimental (top) and theoretical (bottom).

3.9 Cage 42



Zinc(II) triflate (2.00 mg, 5.50 μ mol) and 1,4-di(6"-(1'-methyl-1*H*-benzo[*d*]imidazol-2'yl)pyridin-3"-yl)-2,5-dimethylbenzene (**30**) (4.30 mg, 8.25 μ mol) were dissolved in CD₃CN (500 μ L) and heated for 24 h at 50 °C to prepare cage **42** *in situ*.

¹**H NMR** (500 MHz, CD₃CN, 298 K) δ (ppm): 8.40 (dd, ${}^{3}J = 8.4$ Hz, ${}^{5}J = 0.7$ Hz, 12H, *H*_j), 8.35 (dd, ${}^{4}J = 2.2$ Hz, ${}^{5}J = 0.7$ Hz, 12H, *H*_m), 8.11 (dd, ${}^{3}J = 8.4$ Hz, ${}^{4}J = 2.2$ Hz, 12H, *H*_k), 7.75 (dt, ${}^{3}J = 8.4$ Hz, ${}^{4}J = 1.0$ Hz, 12H, *H*_c), 7.45 (ddd, ${}^{3}J = 8.4$ Hz, ${}^{3}J = 7.3$ Hz, ${}^{4}J = 1.0$ Hz, 12H, *H*_d), 7.06 (ddd, ${}^{3}J = 8.4$ Hz, ${}^{3}J = 7.3$ Hz, ${}^{4}J = 1.0$ Hz, 12H, *H*_d), 6.63 (dt, ${}^{3}J = 8.4$ Hz, ${}^{4}J = 1.0$ Hz, 12H, *H*_f), 4.23 (s, 36H, *H*_a), 1.92 (s, 36H, *H*_p).

¹³**C NMR** (125 MHz, CD₃CN, 298 K) δ (ppm): 149.8 (*C_m*), 149.5 (*C_h*), 142.7 (*C_i*), 142.5 (*C_k*), 140.8 (*C_i*), 138.7 (*C_b*), 138.6 (*C_g*), 137.0 (*C_n*), 134.4 (*C_c*), 133.4 (*C_q*), 126.7 (*C_d*), 126.5 (*C_e*), 125.2 (*C_j*), 118.1 (*C_t*), 113.3 (*C_c*), 34.6 (*C_a*), 20.0 (*C_p*).

¹⁹**F NMR** (471 MHz, CD₃CN, 298 K, C₆F₆) δ (ppm): -79.6 (OTf⁻).

HRMS (ESI): $m/z = 2139.9267 [42 + 60Tf]^{2+}, 1376.6328 [42 + 50Tf]^{3+}, 995.2357 [42 + 40Tf]^{4+}, 766.3980 [42 + 30Tf]^{5+}, 613.5062 [42 + 20Tf]^{6+}, 504.4410 [42 + 0Tf]^{7+}, 422.8914 [42]^{8+}.$

While the signals at m/z 504.4410 and 422.8914 are consistent with the 7+ and 8+ charges, respectively for the cage, the theoretical isotopic pattern do not match, possibly due to fragmentation in the gas phase or overlap with other signals.



Figure S139. ¹H NMR spectrum (500 MHz, CD₃CN, 298 K) of cage 42.





Figure S141. ¹H-¹H COSY NMR spectrum (500 MHz, CD₃CN, 298 K) of cage 42.



Figure S142. ¹H-¹³C HSQC NMR spectrum (500 MHz/125 MHz, CD₃CN, 298 K) of cage 42.



Figure S143. ¹H-¹³C HMBC NMR spectrum (500 MHz/125 MHz, CD₃CN, 298 K) of cage **42**.



Figure S144. ¹⁹F NMR spectrum (471 MHz, CD₃CN, 298 K, C₆F₆) of cage 42.



993.4 993.6 993.8 994.0 994.2 994.4 994.6 994.8 995.0 995.2 995.4 995.6 995.8 996.0 996.2 996.4 996.6 996.8 997.0 997.2 997.4 997.6 997.8 998.0 998.2 998.4 998.6 998.8 m/z (Da)

Figure S145. a) High resolution ESI mass spectrum of cage 42 and b) isotopic patterns of cage 42: experimental (top) and theoretical (bottom).

3.10 Cage 43a and Helicate 43b

Zinc(II) triflate (1.99 mg, 5.47 μ mol) and 1,2-di(4"-methyl-6"-(1'-methyl-1*H*-benzo[*d*]pyridine-2'-yl)pyridine-3"-yl)ethyne (**22**) (3.85 mg, 8.22 μ mol) were dissolved in CD₃CN (500 μ L) and heated for 74.5 h at 50 °C to prepare cage **43a** and helicate **43b** *in situ*.

3.10.1 Cage 43a



¹**H NMR** (500 MHz, CD₃CN, 298 K) δ (ppm): 8.18 (s, 12H, $H_{\rm J}$), 7.97 (s, 12H, $H_{\rm m}$), 7.77 (dt, ³*J* = 8.6 Hz, ⁴*J* = 0.9 Hz, 12H, $H_{\rm c}$), 7.47 (ddd, ³*J* = 8.6 Hz, ³*J* = 7.3 Hz, ⁴*J* = 0.9 Hz, 12H, $H_{\rm d}$), 7.09 (ddd, ³*J* = 8.4 Hz, ³*J* = 7.3 Hz, ⁴*J* = 0.9 Hz, 12H, $H_{\rm e}$), 6.73 (dt, ³*J* = 8.4 Hz, ⁴*J* = 0.9 Hz, 12H, $H_{\rm f}$), 4.25 (s, 36H, $H_{\rm a}$), 2.48 (s, 36H, $H_{\rm n}$).

¹³**C NMR** (125 MHz, CD₃CN, 298 K) δ (ppm): 156.6 (*C_k*), 152.0 (*C_m*), 149.4 (*C_h*), 143.2 (*C_i*), 138.7 (*C_{b,g}*), 127.1 (*C_d*), 126.7 (*C_e*), 125.6 (*C_j*), 123.2 (*C_i*), 118.2 (*C_i*), 113.5 (*C_c*), 94.0 (*C_o*), 34.7 (*C_a*), 21.3 (*C_n*).

¹⁹**F NMR** (471 MHz, CD₃CN, 298 K, C₆F₆) δ (ppm): -79.6 (OTf⁻).

HRMS (ESI): m/z = 1983.3310 [**43a** + 6OTf]²⁺, 1272.5697 [**43a** + 5OTf]³⁺, 917.1887 [**43a** + 4OTf]⁴⁺, 703.7607 [**43a** + 3OTf]⁵⁺, 561.8079 [**43a** + 2OTf]⁶⁺.

While the signal at m/z 561.8079 is consistent with the 6+ charge for the cage, the theoretical isotopic pattern does not match, possibly due to fragmentation in the gas phase or overlap with other signals.

3.10.2 Helicate 43b



Due to the low quantity of helicate present and overlapping signals with cage **43a**, full characterisation of the helicate was not possible. Therefore, the NMR data below only includes proton and carbon atoms that could be assigned.

¹**H NMR** (500 MHz, CD₃CN, 298 K) δ (ppm): 8.41 (s, 6H, H_j), 7.82 (d, ³*J* = 8.5 Hz, 6H, H_c), 7.52 (unres. ddd, 6H, H_d), 7.42 (s, 6H, H_m), 6.75 (unres. d, 6H, H_f), 4.31 (s, 18H, H_a), 2.65 (s, 18H, H_n).

¹³**C NMR** (125 MHz, CD₃CN, 298 K) δ (ppm): 152.3 (*C_m*), 126.9 (*C_{j,d}*), 113.6 (*C_c*), 35.0 (*C_a*), 21.2 (*C_n*).

¹⁹**F NMR** (471 MHz, CD₃CN, 298 K, C₆F₆) δ (ppm): -79.6 (OTf).

HRMS (ESI): $m/z = 1982.3328 [43b + 3OTf]^+$, 916.1884 [43b + 2OTf]²⁺, 561.1414 [43b + OTf]³⁺, 383.6179 [43b]⁴⁺.



Figure S146. ¹H NMR spectrum (500 MHz, CD₃CN, 298 K) of cage **43a** and helicate **43b**. For clarity, only the signals for cage **43a** are labelled.



Figure S147. ¹H DOSY spectrum (600 MHz, CD₃CN, 298 K) of cage **43a** ($D = 5.33 \cdot 10^{-10} \text{ m}^2/\text{s}$) and helicate **43b** ($D = 6.56 \cdot 10^{-10} \text{ m}^2/\text{s}$).



Figure S148. ¹³C NMR spectrum (125 MHz, CD₃CN, 298 K) of cage **43a** and helicate **43b**. For clarity, only the signals for cage **43a** are labelled.



Figure S149. ¹H-¹H COSY NMR spectrum (500 MHz, CD₃CN, 298 K) of cage **43a** and helicate **43b**. For clarity, only the signals for cage **43a** are labelled.



Figure S150. ¹H-¹³C HSQC NMR spectrum (500 MHz/125 MHz, CD₃CN, 298 K) of cage **43a** and helicate **43b**. For clarity, only the signals for cage **43a** are labelled.



Figure S151. ¹H-¹³C HMBC NMR spectrum (500 MHz/125 MHz, CD₃CN, 298 K) of cage **43a** and helicate **43b**. For clarity, only the signals for cage **43a** are labelled.



Figure S152. ¹⁹F NMR spectrum (471 MHz, CD₃CN, 298 K, C₆F₆) of cage **43a** and helicate **43b**. For clarity, only cage **43a** is depicted.



Figure S153. High resolution ESI mass spectrum of cage 43a and helicate 43b.





a)









Figure S154. Isotopic patterns for a) cage 43a and b) helicate 43b: experimental (top) and theoretical (bottom).

3.11 Cage 44



Zinc(II) triflate (2.00 mg, 5.50 μ mol) and 1,4-bis(4"-methyl-6"-(1'-methyl-1*H*-benzo[*d*]imidazol-2'-yl)pyridin-3"-yl)benzene (**33**) (4.30 mg, 8.26 μ mol) were dissolved in CD₃CN (500 μ L) to prepare cage **44** *in situ*.

¹**H NMR** (500 MHz, CD₃CN, 298 K) δ (ppm): 8.46 (s, 12H, H_j), 7.83 (dt, ³*J* = 8.5 Hz, ⁴*J* = 0.9 Hz, 12H, H_c), 7.70 (s, 12H, H_m), 7.51 (ddd, ³*J* = 8.5 Hz, ³*J* = 7.3 Hz, ⁴*J* = 0.9 Hz, 12H, H_d), 7.08 (s, 24H, H_p), 7.05 (ddd, ³*J* = 8.3 Hz, ³*J* = 7.3 Hz, ⁴*J* = 0.9 Hz, 12H, H_e), 6.63 (dt, ³*J* = 8.3 Hz, ⁴*J* = 0.9 Hz, 12H, H_f), 4.36 (s, 36H, H_a), 2.63 (s, 36H, H_n).

¹³**C NMR** (125 MHz, CD₃CN, 298 K) δ (ppm): 151.9 (*C_m*), 149.9 (*C_h*), 149.7 (*C_i*), 143.1 (*C_k*), 139.8 (*C_l*), 139.0 (*C_b*), 138.8 (*C_g*), 136.8 (*C_n*), 130.4 (*C_o*), 127.8 (*C_q*), 126.8 (*C_d*), 126.3 (*C_e*), 118.4 (*C_j*), 113.3 (*C_i*), 35.0 (*C_c*), 21.0 (*C_a*).

¹⁹**F NMR** (471 MHz, CD₃CN, 298 K, C₆F₆) δ (ppm): -79.7 (OTf⁻).

HRMS (ESI): *m*/*z* = 1375.9668 [44 + 5OTf]³⁺.



Figure S155. ¹H NMR spectrum (500 MHz, CD₃CN, 298 K) of cage 44.



Figure S156. $^{\rm 13}C$ NMR spectrum (125 MHz, CD_3CN, 298 K) of cage 44.



Figure S157. ¹H-¹H COSY NMR spectrum (500 MHz, CD₃CN, 298 K) of cage 44.



Figure S158. ¹H-¹³C HSQC NMR spectrum (500 MHz/125 MHz, CD₃CN, 298 K) of cage 44.



Figure S159. ¹H-¹³C HMBC NMR spectrum (500 MHz/125 MHz, CD₃CN, 298 K) of cage **44**.



Figure S160. ¹⁹F NMR spectrum (471 MHz, CD₃CN, 298 K, C₆F₆) of cage 44.







1379,2991

1379.5

1379.0

1379.9657

1380.0



1377.5 m/z (Da) Figure S161. a) High resolution ESI mass spectrum of cage 44 and b) isotopic patterns of cage 44: experimental (top) and theoretical (bottom).

1378.0

1378.5

1377.0

1374.6340

1375.0

1375.5

1376.0

1376.5

1374.5

1374.3011

20-

0-

1380.5
4 Fe₄L₆ Metal-Organic Cages

Metal-organic cages based on Fe^{II} were self-assembled using six equivalents of the appropriate ligand and four equivalents of iron(II) triflate. The cages were prepared according to the procedures described in Sections 4.1-4.11. Characterisation using NMR spectroscopy and ESI-MS was consistent with the self-assembly of Fe₄L₆ cages. Elemental analysis of the cages was attempted in some cases, but the results were not satisfactory, most likely due to solvent that could not be removed by drying.

General procedure:

The ligands and $Fe(OTf)_2$ were dried *in vacuo* before weighing the appropriate amounts. Before transfer to the glovebox the ligand was stored at 110 °C for 1 h and $Fe(OTf)_2$ was stored under vacuum. The Fe^{II} -based cages were prepared using dried CD_3CN . Cage **2** was precipitated outside of the glovebox using diethyl ether, centrifuged and dried *in vacuo* before transfer back to the glovebox (For further details see Section 4.2).

4.1 Cage 1



Iron(II) triflate (2.32 mg, 5.57 μ mol) and 1,2-bis(6"-(1'-methyl-1*H*-imidazol-2'-yl)pyridin-3"-yl)ethyne (**14**) (2.85 mg, 8.37 μ mol) were dissolved in dry CD₃CN (500 μ L).

¹**H NMR** (600 MHz, CD₃CN, 298 K) δ (ppm): 8.22 (d, ${}^{3}J$ = 8.6 Hz, 12H, *H_f*), 8.00 (d, ${}^{3}J$ = 8.6 Hz, 12H, *H_g*), 7.75 (br, 12H, *H_i*), 7.43 (s, 12H, *H_b*), 6.33 (s, 12H, *H_c*), 4.22 (s, 36H, *H_a*).

¹³**C NMR** (151 MHz, CD₃CN, 298 K) δ (ppm): 158.8 (*C*_i), 152.0 (*C*_h), 147.5 (*C*_e), 140.9 (*C*_g), 131.6 (*C*_c), 130.7 (*C*_b), 122.4 (*C*_f), 120.5 (*C*_d), 90.4 (*C*_j), 37.4 (*C*_a).

¹⁹**F NMR** (471 MHz, CD₃CN, 298 K, C₆F₆) δ (ppm): -78.8 (OTf⁻).

HRMS (ESI): *m*/*z* = 1003.7862 [1 + 50Tf]³⁺, 715.3524 [1 + 40Tf]⁴⁺.

While the signal at m/z 1003.7862 is consistent with the 3+ charge for the cage, the theoretical isotopic pattern does not match, possibly due to fragmentation in the gas phase or overlap with other signals.



S109



Figure S163. ¹³C NMR spectrum (151 MHz, CD₃CN, 298 K) of cage 1.



Figure S164. ¹H-¹H COSY NMR spectrum (600 MHz, CD₃CN, 298 K) of cage 1.



Figure S165. ¹H-¹³C HSQC NMR spectrum (600 MHz/151 MHz, CD₃CN, 298 K) of cage 1.



Figure S166. ¹H-¹³C HMBC NMR spectrum (600 MHz/151 MHz, CD₃CN, 298 K) of cage 1.



Figure S167. ¹⁹F NMR spectrum (471 MHz, CD₃CN, 298 K) of cage 1.



Figure S168. a) High resolution ESI mass spectrum of cage **1** and b) isotopic patterns of cage **1**: experimental (top) and theoretical (bottom).



Iron(II) triflate (11.6 mg, 27.9 μ mol) and 6',6"-di(1-benzyl-1*H*-benzo[*d*]imidazol-2-yl)-3',3"bipyridine (**26**) (23.7 mg, 41.7 μ mol) were dissolved in acetonitrile (3 mL) and the mixture was left to stand at room temperature for 3 h. The solution was poured into diethyl ether (10 mL), centrifuged, and the ether was decanted. Additional diethyl ether (10 mL) was added, and the suspension was centrifuged, decanted, and dried under vacuum (18.0 mg, 3.73 μ mol, 53%).

¹**H NMR** (600 MHz, CD₃CN, 298 K) δ (ppm): 8.61 (d, ${}^{3}J = 8.4$ Hz, 12H, H_{n}), 7.85 (d, ${}^{3}J = 7.9$ Hz, 12H, H_{g}), 7.49 (t, ${}^{3}J = 7.9$ Hz, 12H, H_{h}), 7.44 (dd, ${}^{3}J = 8.4$ Hz, ${}^{4}J = 1.2$ Hz, 12H, H_{o}), 7.29-7.25 (m, 24H, H_{b}), 7.24-7.20 (m, 36H, $H_{a,c}$), 7.18 (s, 12H, H_{q}), 7.09 (t, ${}^{3}J = 7.9$ Hz, 12H, H_{h}), 6.76 (d, ${}^{2}J = 18.0$ Hz, 12H, H_{e}), 6.13 (d, ${}^{3}J = 7.9$ Hz, 12H, H_{f}), 6.07 (d, ${}^{2}J = 18.0$ Hz, 12H, H_{e}).

¹³**C NMR** (151 MHz, CD₃CN, 298 K) δ (ppm): 156.4 (*C_q*), 152.6 (*C_l*), 150.7 (*C_m*), 145.5 (*C_k*), 141.0 (*C_l*), 139.6 (*C_o*), 136.4 (*C_p*), 136.0 (*C_d*), 130.2 (*C_b*), 129.3 (*C_a*), 128.0 (*C_h*), 127.0 (*C_c*), 126.6 (*C_l*), 125.6 (*C_n*), 117.6 (*C_j*), 114.4 (*C_g*), 49.2 (*C_e*).

HRMS (ESI): $m/z = 2264.4426 [2 + 60Tf]^{2+}, 1459.9767 [2 + 50Tf]^{3+}, 1057.7428 [2 + 40Tf]^{4+}.$



Figure S169. ¹H NMR spectrum (600 MHz, CD₃CN, 298 K) of cage 2.



Figure S170. 13 C NMR spectrum (151 MHz, CD₃CN, 298 K) of cage 2.



Figure S171. ¹H-¹H COSY NMR spectrum (600 MHz, CD₃CN, 298 K) of cage 2.



Figure S172. ¹H-¹³C HSQC NMR spectrum (600 MHz/151 MHz, CD₃CN, 298 K) of cage 2.



Figure S173. ¹H-¹³C HMBC NMR spectrum (600 MHz/151 MHz, CD₃CN, 298 K) of cage 2.



Figure S174. ¹⁹F NMR spectrum (471 MHz, CD₃CN, 298 K) of cage 2.



Figure S175. a) High resolution ESI mass spectrum of cage **2** and b) isotopic patterns of cage **2**: experimental (top) and theoretical (bottom).



Iron(II) triflate (21.7 mg, 52.1 μ mol) and 1,2-di(6"-(1'-benzyl-1*H*-benzo[*d*]imidazole-2'yl)pyridin-3"-yl)ethyne (**19**) (46.2 mg, 78.0 μ mol) were dissolved in acetonitrile (5 mL) and the mixture was left to stand at room temperature for 3 h. The solution was poured into diethyl ether (15 mL), centrifuged, and the ether was decanted. The precipitated cage was washed with additional diethyl ether (15 mL), centrifuged, decanted, and dried extensively under vacuum (49.4 mg, 8.21 μ mol, 63%).

¹**H NMR** (600 MHz, CD₃CN, 298 K) δ (ppm): 10.03 (unres. d, 12H, H_n), 8.86 (br, 12H, H_q), 8.54 (d, ${}^{3}J = 8.1$ Hz, 12H, H_g), 7.57 (d, ${}^{3}J = 8.3$ Hz, 12H, H_o), 7.42 (t, ${}^{3}J = 8.1$ Hz, 12H, H_h), 7.38 (t, ${}^{3}J = 7.4$ Hz, 24H, H_b), 7.32 (d, ${}^{3}J = 7.4$ Hz, 24H, H_c), 7.22 (t, ${}^{3}J = 7.4$ Hz, 12H, H_a), 6.84 (t, ${}^{3}J = 6.6$ Hz, 12H, H_i), 6.72 (d, ${}^{2}J = 17.9$ Hz, 12H, H_e), 6.61 (d, ${}^{2}J = 17.9$ Hz, 12H, H_e), 4.91 (unres. d, 12H, H_i).

¹³**C NMR** (151 MHz, CD₃CN, 298 K) δ (ppm): 158.8 (*C_k*), 157.5 (*C_q*), 154.7 (*C_f*), 148.9 (*C_m*), 142.4 (*C_o*), 140.3 (*C_p*), 135.4 (*C_d*), 135.0 (*C_l*), 130.3 (*C_n*), 129.2 (*C_b*), 128.2 (*C_a*), 127.0 (*C_h*), 125.8 (*C_c*), 125.0 (*C_i*), 117.4 (*C_j*), 116.0 (*C_g*), 93.9 (*C_r*), 51.5 (*C_e*).

HRMS (ESI): $m/z = 1508.3126 [3 + 50Tf]^{3+}$, 1093.9949 [3 + 40Tf]⁴⁺, 662.7292 [3 + 30Tf]⁵⁺, 527.4492 [3 + 20Tf]⁶⁺, 430.8211 [3 + 0Tf]⁷⁺.



Figure S176. ¹H NMR spectrum (600 MHz, CD₃CN, 298 K) of cage 3.



Figure S177. ¹³C NMR spectrum (151 MHz, CD₃CN, 298 K) of cage 3.



Figure S178. ¹H-¹H COSY NMR spectrum (600 MHz, CD₃CN, 298 K) of cage 3.



Figure S179. ¹H-¹³C HSQC NMR spectrum (600 MHz/151 MHz, CD₃CN, 298 K) of cage 3.



Figure S180. ¹H-¹³C HMBC NMR spectrum (600 MHz/151 MHz, CD₃CN, 298 K) of cage 3.



Figure S181. ¹⁹F NMR spectrum (471 MHz, CD₃CN, 298 K) of cage 3.







Figure S182. a) High resolution ESI mass spectrum of cage **3** and b) isotopic patterns of cage **3**: experimental (top) and theoretical (bottom).



Iron(II) triflate (2.30 mg, 5.52 μ mol) and 1,4-di(6"-(1'-benzyl-1*H*-benzo[*d*]imidazol-2'yl)pyridin-3"-yl)benzene (**28**) (5.34 mg, 8.28 μ mol) were dissolved in dry CD₃CN (500 μ L).

¹**H NMR** (600 MHz, CD₃CN, 298 K) δ (ppm): 9.89 (s, 12H, H_n), 8.93 (br, 12H, H_q), 8.40 (d, ³*J* = 8.3 Hz, 12H, H_g), 8.09 (d, ³*J* = 8.1 Hz, 12H, H_o), 7.44 (unres. dt, 12H, H_h), 7.28 (d, ³*J* = 7.3 Hz, 24H, H_c), 7.24 (t, ³*J* = 7.3 Hz, 24H, H_b), 7.13 (t, ³*J* = 7.3 Hz, 12H, H_a), 6.91 (t, ³*J* = 7.3 Hz, 12H, H_i), 6.70 (s, 12H, H_s), 6.55 (d, ²*J* = 18.0 Hz, 12H, H_e), 6.49 (d, ²*J* = 18.0 Hz, 12H, H_e), 5.29 (s, 12H, H_i).

¹³**C NMR** (151 MHz, CD₃CN, 298 K) δ (ppm): 157.1 (*C*_k), 153.9 (*C*_m), 152.8 (*C*_f), 150.3 (*C*_i), 147.1 (*C*_p), 143.1 (*C*_q), 138.5 (*C*_o), 137.2 (*C*_r), 136.6 (*C*_d), 130.7 (*C*_n), 130.5 (*C*_b), 129.3 (*C*_a), 128.2 (*C*_s), 127.9 (*C*_h), 127.0 (*C*_c), 126.2 (*C*_i), 118.5 (*C*_j), 116.5 (*C*_g), 52.2 (*C*_e).

HRMS (ESI): *m*/*z* = 2492.5379 [**4** + 6OTf]²⁺, 1612.3752 [**4** + 5OTf]³⁺, 1171.7921 [**4** + 4OTf]⁴⁺, 907.6427 [**4** + 3OTf]⁵⁺, 731.5433 [**4** + 2OTf]⁶⁺, 605.7585 [**4** + OTf]⁷⁺.



Figure S183. ¹H NMR spectrum (600 MHz, CD₃CN, 298 K) of cage 4.



Figure S184. ¹³C NMR spectrum (151 MHz, CD₃CN, 298 K) of cage 4.



Figure S185. ¹H-¹H COSY NMR spectrum (600 MHz, CD₃CN, 298 K) of cage 4.



Figure S186. ¹H-¹³C HSQC NMR spectrum (600 MHz/151 MHz, CD₃CN, 298 K) of cage 4.



Figure S187. ¹H-¹³C HMBC NMR spectrum (600 MHz/151 MHz, CD₃CN, 298 K) of cage 4.



Figure S188. ¹⁹F NMR spectrum (471 MHz, CD₃CN, 298 K) of cage 4.



Figure S189. a) High resolution ESI mass spectrum of cage **4** and b) isotopic patterns of cage **4**: experimental (top) and theoretical (bottom).



Iron(II) triflate (2.31 mg, 5.55 μ mol) and 6',6"-di(1-methyl-1*H*-benzo[*d*]imidazol-2-yl)-3',3"-bipyridine (**24**) (3.46 mg, 8.31 μ mol) were dissolved in dry CD₃CN (500 μ L).

¹**H NMR** (600 MHz, CD₃CN, 298 K) δ (ppm): 11.41 (s, 12H, H_j), 9.72 (br, 12H, H_m), 8.87 (d, ³*J* = 8.4 Hz, 12H, H_c), 7.41 (d, ³*J* = 8.2 Hz, 12H, H_j), 7.30 (dd, ³*J* = 8.4 Hz, ³*J* = 7.4 Hz, 12H, H_d), 6.46 (unres. dd, 12H, H_e), 5.59 (s, 36H, H_a), 4.14 (br, 12H, H_f).

¹³**C NMR** (151 MHz, CD₃CN, 298 K) δ (ppm): 163.6 (*C*_b), 156.5 (*C*_i), 151.4 (*C*_h), 141.0 (*C*_k), 137.5 (*C*_i), 134.3 (*C*_j), 127.0 (*C*_d), 124.9 (*C*_e), 119.1 (*C*_c), 118.6 (*C*_f), 38.3 (*C*_a).

HRMS (ESI): m/z = 1807.7483 [**5**+ 6OTf]²⁺, 1155.8488 [**5** + 5OTf]³⁺, 829.6484 [**5** + 4OTf]⁴⁺, 633.7276 [**5** + 3OTf]⁵⁺.



Figure S190. ¹H NMR spectrum (600 MHz, CD₃CN, 298 K) of cage 5.



Figure S191. ¹³C NMR spectrum (151 MHz, CD₃CN, 298 K) of cage 5.



Figure S192. ¹H-¹H COSY NMR spectrum (600 MHz, CD₃CN, 298 K) of cage 5.



Figure S193. ¹H-¹³C HSQC NMR spectrum (600 MHz/151 MHz, CD₃CN, 298 K) of cage 5.



Figure S194. ¹H-¹³C HMBC NMR spectrum (600 MHz/151 MHz, CD₃CN, 298 K) of cage 5.



Figure S195. ¹⁹F NMR spectrum (471 MHz, CD₃CN, 298 K) of cage 5.



Figure S196. a) High resolution ESI mass spectrum of cage **5** and b) isotopic patterns of cage **5**: experimental (top) and theoretical (bottom).

a)



Iron(II) triflate (2.31 mg, 5.55 μ mol) and 1,2-di(6"-(1'-methyl-1*H*-benzo[*d*]imidazole-2'-yl)pyridin-3"-yl)ethyne (**17**) (3.67 mg, 8.33 μ mol) were dissolved in dry CD₃CN (500 μ L) and heated for 143 h at 50 °C to prepare cage **6** *in situ*.

¹**H NMR** (600 MHz, CD₃CN, 298 K) δ (ppm): 16.3 (s, 12H, *H_j*), 15.1 (br, 12H, *H_m*), 10.8 (s, 12H, *H_c*), 7.19 (s, 48H, *H_{a,k}*), 6.97 (s, 12H, *H_d*), 5.75 (s, 12H, *H_e*), 1.05 (s, 12H, *H_f*).

¹³**C NMR** (151 MHz, CD₃CN, 298 K) δ (ppm): 213.2 (s, C_{quart}), 209.8 (s, C_{quart}), 184.6 (s, C_{quart}), 157.2 (d, ¹*J* = 191 Hz, *C*_m), 152.1 (d, ¹*J* = 164 Hz, *C*_j), 148.1 (d, ¹*J* = 175 Hz, *C*_k), 140.3 (s, C_{quart}), 128.2 (d, ¹*J* = 167 Hz, *C*_c), 126.5 (d, ¹*J* = 165 Hz, *C*_d), 122.2 (d, ¹*J* = 162 Hz, *C*_e), 120.9 (d, ¹*J* = 165 Hz, *C*_f), 108.6 (s, *C*_{quart}), 105.8 (s, *C*_{quart}), 46.0 (d, ¹*J* = 143 Hz, *C*_a).

HRMS (ESI): m/z = 1880.2554 [**6** + 6OTf]²⁺, 1203.8511 [**6** + 5OTf]³⁺, 865.6493 [**6** + 4OTf]⁴⁺, 662.7292 [**6** + 3OTf]⁵⁺, 527.4492 [**6** + 2OTf]⁶⁺, 430.8211 [**6** + OTf]⁷⁺.

While the signal at m/z 430.8211 is consistent with the 7+ charge for the cage, the theoretical isotopic pattern does not match, possibly due to fragmentation in the gas phase or overlap with other signals.



Figure S197. ¹H NMR spectrum (600 MHz, CD₃CN, 298 K) of cage 6.



Figure S198. ¹³C NMR spectrum (151 MHz, CD_3CN , 298 K) of cage 6.



Figure S199. ¹H-¹H COSY NMR spectrum (600 MHz, CD₃CN, 298 K) of cage 6.



Figure S200. ^{1}H - ^{13}C HMQC NMR spectrum (600 MHz/151 MHz, CD₃CN, 298 K) of cage 6.



Figure S201. ¹⁹F NMR spectrum (471 MHz, CD₃CN, 298 K) of cage **6**.









Figure S202. a) High resolution ESI mass spectrum of cage **6** and b) isotopic patterns of cage **6**: experimental (top) and theoretical (bottom).



Iron(II) triflate (2.33 mg, 5.59 μ mol) and 1,4-di(6"-(1'-methyl-1*H*-benzo[*d*]imidazol-2'yl)pyridin-3"-yl)benzene (**27**) (4.14 mg, 8.40 μ mol) were dissolved in dry CD₃CN (500 μ L).

¹**H NMR** (500 MHz, CD₃CN, 298 K) δ (ppm): 13.10 (s, 12H, H_j), 12.2 (br, 12H, H_m), 9.44 (d, ${}^{3}J = 8.1$ Hz, 12H, H_c), 7.86 (d, ${}^{3}J = 8.4$ Hz, 12H, H_k), 7.14 (t, ${}^{3}J = 7.6$ Hz, 12H, H_d), 6.75 (s, 24H, H_o), 6.26 (unres. dd, 12H, H_e), 5.86 (s, 36H, H_a), 3.22 (s, 12H, H_f).

¹³**C NMR** (126 MHz, CD₃CN, 298 K) δ (ppm): 152.5 ($C_{quart.}$), 144.6 ($C_{quart.}$), 141.4 ($C_{quart.}$), 140.2 (C_{o}), 128.7 (C_{s}), 127.2 (C_{d}), 124.3 (C_{e}), 122.5 (C_{c}), 120.0 (C_{f}), 41.7 (C_{a}).

HRMS (ESI): *m*/*z* = 2036.3478 [**7** + 6OTf]²⁺, 1307.9146 [**7** + 5OTf]³⁺, 943.6976 [**7** + 4OTf]⁴⁺, 724.9670 [**7** + 3OTf]⁵⁺, 579.4813 [**7** + 2OTf]⁶⁺.

While the signal at m/z 579.4813 is consistent with the 6+ charge for the cage, the theoretical isotopic pattern does not match, possibly due to fragmentation in the gas phase or overlap with other signals.



Figure S203. ¹H NMR spectrum (500 MHz, CD₃CN, 298 K) of cage 7.



Figure S204. $^{\rm 13}C$ NMR spectrum (126 MHz, CD_3CN, 298 K) of cage 7.



Figure S205. ¹H-¹H COSY NMR spectrum (500 MHz, CD₃CN, 298 K) of cage 7.



Figure S206. ¹H-¹³C HSQC NMR spectrum (500 MHz/126 MHz, CD₃CN, 298 K) of cage 7.



Figure S207. ¹H-¹³C HMBC NMR spectrum (500 MHz/126 MHz, CD₃CN, 298 K) of cage 7.



Figure S208. ¹⁹F NMR spectrum (471 MHz, CD₃CN, 298 K) of cage 7.





2000

1000

_ .

b)



Figure S209. a) High resolution ESI mass spectrum of cage **7** and b) isotopic patterns of cage **7**: experimental (top) and theoretical (bottom).

a)

4.8 Cage 8



Iron(II) triflate (2.31 mg, 5.55 μ mol) and 1,4-di(6"-(1'-methyl-1*H*-benzo[*d*]imidazol-2'-yl)pyridin-3"-yl)-2,5-difluorobenzene (**32**) (4.40 mg, 8.32 μ mol) were dissolved in dry CD₃CN (500 μ L).

¹**H NMR** (600 MHz, CD₃CN, 298 K) δ (ppm): 17.5 (br, 12H, H_m), 17.1 (s, 12H, H_j), 11.0 (s, 12H, H_c), 7.17 (d, ³*J* = 6.5 Hz, 12H, H_k), 7.06 (s, 36H, H_a), 7.00 (t, ³*J* = 7.5 Hz, 12H, H_d), 6.61 (unres. dt, 12H, H_q), 5.97 (unres. d, 12H, H_e), 2.28 (br, 12H, H_f).

¹³**C NMR** (151 MHz, CD₃CN, 298 K) δ (ppm): 156.8 (*C*_{quart.}), 155.2 (*C*_{quart.}), 154.8 (*C*_{quart.}), 146.2 (*C*_k), 136.1 (*C*_{quart.}), 133.8 (*C*_{quart.}), 129.0 (*C*_c), 126.2 (*C*_d), 121.8 (*C*_e), 121.4 (*C*_{quart.}), 46.4 (*C*_a).

HRMS (ESI): m/z = 2144.2912 [**8** + 6OTf]²⁺, 1379.8754 [**8** + 5OTf]³⁺, 997.6674 [**8** + 4OTf]⁴⁺, 768.3437 [**8** + 3OTf]⁵⁺, 615.4611 [**8** + 2OTf]⁶⁺, 506.1155 [**8** + OTf]⁷⁺.



Figure S210. ¹H NMR spectrum (600 MHz, CD₃CN, 298 K) of cage 8.



Figure S211. ¹³C NMR spectrum (151 MHz, CD₃CN, 298 K) of cage 8.



Figure S212. ¹H-¹H COSY NMR spectrum (600 MHz, CD₃CN, 298 K) of cage 8.



Figure S213. ¹H-¹³C HSQC NMR spectrum (600 MHz/151 MHz, CD₃CN, 298 K) of cage 8.



Figure S214. ¹H-¹³C HMBC NMR spectrum (600 MHz/151 MHz, CD₃CN, 298 K) of cage 8.


Figure S215. ¹⁹F NMR spectrum (471 MHz, CD₃CN, 298 K) of cage 8.



Figure S216. a) High resolution ESI mass spectrum of cage 8 (range m/z 500-1000) and b) isotopic patterns of cage 8: experimental (top) and theoretical (bottom).



Figure S217. High resolution ESI mass spectrum (range *m*/*z* 1000-2400) of cage 8.

4.9 Cage 9



Iron(II) triflate (2.36 mg, 5.67 μ mol) and 1,4-di(6"-(1'-methyl-1*H*-benzo[*d*]imidazol-2'yl)pyridin-3"-yl)-2,5-dimethylbenzene (**30**) (4.43 mg, 8.51 μ mol) were dissolved in dry CD₃CN (500 μ L).

¹**H NMR** (600 MHz, CD₃CN, 298 K) δ (ppm): 28.5 (br, 12H, H_m), 25.9 (s, 12H, H_j), 14.6 (s, 12H, H_c), 9.52 (s, 36H, H_a), 6.51 (unres. dt, 12H, H_d), 5.61 (s, 12H, H_k), 5.27 (s, 12H, H_q), 5.23 (s, 12H, H_e), 0.21 (s, 36H, H_p), -0.06 (s, 12H, H_f).

¹³**C NMR** (151 MHz, CD₃CN, 298 K) δ (ppm): 159.7 (C_k), 148.8 (C_c), 134.4 ($C_{quart.}$), 133.4 (C_q), 130.2 ($C_{quart.}$), 128.4 (C_d), 120.0 (C_e), 62.9 (C_a), 18.7 (C_p).

HRMS (ESI): $m/z = 2121.4451 [9 + 6OTf]^{2+}$, 1363.9768 [9 + 5OTf]^{3+}, 985.7433 [9 + 4OTf]^{4+}, 758.8041 [9 + 3OTf]^{5+}, 607.5114 [9 + 2OTf]⁶⁺, 499.4451 [9 + OTf]⁷⁺.



Figure S218. ¹H NMR spectrum (600 MHz, CD₃CN, 298 K) of cage 9.



Figure S219. 13 C NMR spectrum (151 MHz, CD₃CN, 298 K) of cage 9.



Figure S220. ¹H-¹H COSY NMR spectrum (600 MHz, CD₃CN, 298 K) of cage 9.



Figure S221. ¹H-¹³C HSQC NMR spectrum (600 MHz/151 MHz, CD₃CN, 298 K) of cage **9**.



Figure S222. ¹H-¹³C HMBC NMR spectrum (600 MHz/151 MHz, CD₃CN, 298 K) of cage 9.



Figure S223. ¹⁹F NMR spectrum (471 MHz, CD₃CN, 298 K) of cage **9**.







Figure S224. a) High resolution ESI mass spectrum of cage **9** (range m/z 500-1000) and b) isotopic patterns of cage **9**: experimental (top) and theoretical (bottom).



Figure S225. High resolution ESI mass spectrum (range *m*/*z* 1000-2400) of cage 9.

4.10 Cage 10



Iron(II) triflate (2.30 mg, 5.52 μ mol) and 1,2-di(4"-methyl-6"-(1'-methyl-1*H*-benzo[*d*]imidazol-2'-yl)pyridin-3"-yl)ethyne (**22**) (3.88 mg, 8.28 μ mol) were dissolved in dry CD₃CN (500 μ L).

¹**H NMR** (600 MHz, CD₃CN, 298 K) δ (ppm): 19.4 (s, 12H, *H_j*), 19.1 (br, 12H, *H_m*), 12.2 (s, 12H, *H_c*), 8.25 (s, 36H, *H_a*), 6.84 (s, 12H, *H_d*), 5.46 (s, 12H, *H_e*), 2.93 (s, 36H, *H_n*), 0.18 (s, 12H, *H_f*).

¹³**C NMR** (151 MHz, CD₃CN, 298 K) δ (ppm): 163.9 (s, $C_{quart.}$), 163.0 (s, $C_{quart.}$), 160.7 (d, ¹*J* = 182 Hz, $C_{j/m}$), 135.4 (d, ¹*J* = 166 Hz, C_c), 131.2 (s, $C_{quart.}$), 126.7 (d, ¹*J* = 160 Hz, C_d), 123.6 (s, $C_{quart.}$), 122.6 (s, $C_{quart.}$), 121.1 (d, ¹*J* = 161 Hz, C_e), 119.3 (unres. d, C_f), 85.0 (s, C_o), 52.0 (q, ¹*J* = 144 Hz, C_a), 18.5 (q, ¹*J* = 137 Hz, C_n).

HRMS (ESI): $m/z = 1964.3488 [10 + 60Tf]^{2+}, 1259.9138 [10 + 50Tf]^{3+}, 907.6967 [10 + 40Tf]^{4+}, 696.3669 [10 + 30Tf]^{5+}, 555.4804 [10 + 20Tf]^{6+}.$



Figure S226. ¹H NMR spectrum (600 MHz, CD_3CN , 298 K) of cage 10.



Figure S227. 13 C NMR spectrum (151 MHz, CD₃CN, 298 K) of cage 10.



Figure S228. ¹H-¹H COSY NMR spectrum (600 MHz, CD₃CN, 298 K) of cage 10.



Figure S229. ¹H-¹³C HMQC NMR spectrum (600 MHz/151 MHz, CD₃CN, 298 K) of cage 10.



Figure S230. ¹⁹F NMR spectrum (471 MHz, CD₃CN, 298 K) of cage 10.



Figure S231. a) High resolution ESI mass spectrum of cage 10 and b) isotopic patterns of cage 10: experimental (top) and theoretical (bottom).

4.11 Cage 11



Iron(II) triflate (2.31 mg, 5.55 μ mol) and 1,4-bis(4"-methyl-6"-(1'-methyl-1*H*-benzo[*d*]imidazol-2'-yl)pyridin-3"-yl)benzene (**33**) (4.34 mg, 8.34 μ mol) were dissolved in dry CD₃CN (500 μ L).

¹**H NMR** (600 MHz, CD₃CN, 298 K) δ (ppm): 58.3 (s, 12H, H_j), 32.8 (br, 12H, H_m), 24.9 (s, 12H, H_c), 22.1 (s, 36H, H_a), 4.81 (s, 36H, H_n), 2.80 (s, 12H, H_d), -3.32 (s, 12H, H_e), -6.19 (s, 24H, H_p), -29.3 (s, 12H, H_f).

¹³**C NMR** (151 MHz, CD₃CN, 298 K) δ (ppm): 205.0 (*C*_c), 192.0 (*C*_{quart.}), 180.0 (*C*_{quart.}), 123.7 (*C*_p), 122.6 (*C*_d), 112.9 (*C*_a), 104.3 (*C*_e), 51.9 (*C*_{quart.}), 8.6 (*C*_n).

HRMS (ESI): $m/z = 2120.4434 [11 + 60Tf]^{2+}$, 1363.9764 [11 + 50Tf]³⁺, 985.7422 [11 + 40Tf]⁴⁺, 607.5106 [11 + 30Tf]⁵⁺.

While the signals at m/z 985.7422 and 607.5106 are consistent with the 4+ and 5+ charges, respectively, for the cage, the theoretical isotopic pattern does not match, possibly due to fragmentation in the gas phase or overlap with other signals.



Figure S232. ¹H NMR spectrum (600 MHz, CD₃CN, 298 K) of cage 11.



Figure S233. ¹³C NMR spectrum (151 MHz, CD₃CN, 298 K) of cage 11.



Figure S234. ¹H-¹H COSY NMR spectrum (600 MHz, CD₃CN, 298 K) of cage 11.



Figure S235. ¹H-¹³C HMQC NMR spectrum (600 MHz/151 MHz, CD₃CN, 298 K) of cage 11.



Figure S236. ¹⁹F NMR spectrum (471 MHz, CD₃CN, 298 K) of cage 11.









Figure S237. a) High resolution ESI mass spectrum (range m/z 500-1000) of cage **11** and b) isotopic patterns of cage **11**: experimental (top) and theoretical (bottom).



Figure S238. High resolution ESI mass spectrum (range *m/z* 1000-2400) of cage 11.

5 Spin-Crossover Studies

5.1 Evans Method

For the determination of the magnetic susceptibility by the Evans method, a standard is required. Since the standard could potentially bind in the cavity of the cages, *p*-xylene was chosen^[9] over other smaller commonly used standards *t*-BuOH and cyclohexane. However, preliminary studies with cage **6** suggested guest binding based on the chemical shift changes upon the addition of *p*-xylene, thus making it unsuitable as a standard (Figure S239). Since the ideal solution model does not require the use of a standard, the spin-crossover properties of the Fe^{II}₄L₆ cages were determined using this method.



Figure S239. ¹H NMR spectra (500 MHz, CD₃CN, 298 K) of cage **6** with (top) and without (bottom) *p*-xylene.

5.2 Ideal Solution Model

The ideal solution model^[10] can be used to determine the spin-crossover temperature $T_{1/2}$ by fitting δ (the chemical shift of the spin-crossover compound) as a function of temperature (T) to Eq. 1 using Origin (with *R* as 8.31446 J/mol·K) to calculate the values of *C*, ΔH , ΔS and δ_{LS} (the chemical shift of the low-spin state). In initial fits, δ_{LS} was not fixed and the fitting gave large errors or did not converge (Table S1, Sections 5.3-5.13). Therefore, δ_{LS} was fixed in subsequent fittings as the diamagnetic shift of the $Zn^{II}_4L_6$ analogue (Table S1) and a control VT experiment with cage **44** revealed the chemical shift of this proton (j) remained constant within the investigated temperature range (Figure S240).

$$\delta = \delta_{LS} + \frac{C}{T + T \cdot \exp\left(\frac{\Delta H - T\Delta S}{RT}\right)}$$
(1)

For the calculation of $T_{1/2}$ from the obtained ΔH and ΔS values, Eq. 2 describing K_{eq} , the equilibrium constant for the spin transition, is needed. By assuming that [HS] and [LS] (the high spin-state and low spin-state concentrations, respectively) are equal at $T_{1/2}$, Eq. 2 can be simplified to Eq. 3.

$$K_{eq} = \frac{[HS]}{[LS]} = exp\left(\frac{-\Delta G}{RT}\right) = 1$$
(2)

$$1 = \exp\left(\frac{-\Delta G}{RT}\right) \equiv \Delta G = 0 \tag{3}$$

Insertion of Eq. 3 into Eq. 4 gives the spin-transition temperature $T_{1/2}$ as a function of ΔH and ΔS .

$$\Delta G = \Delta H - T \Delta S \tag{4}$$

$$T_{1/2} = \frac{\Delta H}{\Delta S} \tag{5}$$

Furthermore, the spin-state fractions γ_{LS} and γ_{HS} can be calculated for the high-spin and low-spin state, respectively, at any given temperature (Eq. 6).

$$\gamma_{LS} = \frac{[LS]}{[LS] + [HS]} \tag{6}$$

Reorganization of Eq. 2 as Eq. 7 and insertion into Eq. 6 gives Eq. 8. Following simplification, the spin-state fraction γ_{LS} can be calculated using the Gibbs energy (Eq. 9).

$$[HS] = K_{eq} [LS] \tag{7}$$

$$\gamma_{LS} = \frac{[LS]}{[LS] + [LS]K_{eq}} \equiv \frac{[LS]}{[LS]\left(1 + exp\left(\frac{-\Delta G}{RT}\right)\right)}$$
(8)

$$\gamma_{LS} = \frac{1}{1 + exp\left(\frac{-\Delta G}{RT}\right)} \tag{9}$$

Since the sum of the spin-state fractions equals 1 (Eq. 10), the spin-state fraction γ_{HS} can also be obtained (Eq. 11).

$$\gamma_{LS} + \gamma_{HS} = 1 \tag{10}$$

$$\gamma_{HS} = 1 - \frac{1}{1 + \exp\left(\frac{-\Delta G}{RT}\right)}$$
(11)

Table S1. Thermodynamic data from variable temperature NMR experiments (248-348 K)

| | Unfixed δ _{LS} | | Fixed δ _{LS} | | | | |
|------|-------------------------|-----------------------|-----------------------|--------------|-----------------------|------------------|--|
| Cage | ΔH | ΔS | δ_{LS} | ΔH | ΔS | T _{1/2} | |
| _ | [kJ/mol] ^a | [J/molK] ^a | [ppm] ^b | [kJ/mol]ª | [J/molK] ^a | [K] | |
| 1 | С | С | - | С | С | С | |
| 2 | d | d | 7.78 | d | d | d | |
| 3 | d | d | 8.02 | d | d | d | |
| 4 | d | d | 8.18 | d | d | d | |
| 5 | d | d | 8.34 | d | d | d | |
| 6 | 22.88 ± 1.63 | 53.31 ± 8.07 | 8.43 | 24.46 ± 0.58 | 60.94 ± 3.00 | 401 | |
| 7 | 28.46 ± 2.29 | 68.42 ± 11.0 | 8.70 | 27.71 ± 0.91 | 64.48 ± 5.78 | 430 | |
| 8 | 29.98 ± 2.21 | 83.33 ± 7.85 | 8.48 | 27.22 ± 0.93 | 72.61 ± 4.00 | 375 | |
| 9 | 19.55 ± 1.26 | 56.27 ± 4.69 | 8.40 | 21.79 ± 0.48 | 64.32 ± 2.00 | 339 | |
| 10 | 31.14 ± 2.64 | 91.14 ± 8.97 | 8.18 | 27.44 ± 1.12 | 77.65 ± 4.40 | 353 | |
| 11 | d | d | 8.46 | 20.78 ± 0.22 | 85.05 ± 0.93 | 244 | |

^a Determined from fitting the chemical shift change between 248 K and 348 K according to Eq. 1. ^b Chemical shift of the Zn(II) cage analogue. ^c Low spin cage. ^d Could not be determined since fitting did not converge.



Figure S240. a) Chemical shift changes in the range of 248 K to 348 K for diamagnetic cage **44** with b) an expansion of the aromatic region.

5.3 Cage 1

Cage 1 was predominantly low spin between 248 K and 348 K.



Figure S241. Chemical shift changes in the range of 248 K to 348 K for cage 1.



Figure S242. Chemical shift changes from 248 K to 348 K for cage 1.

5.4 Cage 2

Cage **2** was predominantly low spin between 248 K and 348 K. Fitting did not converge with or without fixing δ_{LS} .





Figure S243. Chemical shift changes in the range of 248 K to 348 K for cage 2.



Figure S244. Chemical shift changes in the range of 248 K to 348 K for cage 2.

5.5 Cage 3

Cage **3** was predominantly low spin between 248 K and 348 K. Fitting did not converge with or without fixing δ_{LS} .



Figure S245. Chemical shift changes in the range of 248 K to 348 K for cage 3.



Figure S246. Chemical shift changes in the range of 248 K to 348 K for cage 3.

5.6 Cage 4

Cage **4** was predominantly low spin between 248 K and 348 K. Fitting did not converge with or without fixing δ_{LS} .





Figure S247. Chemical shift changes in the range of 248 K to 348 K for cage 4.



Figure S248. Chemical shift changes in the range of 248 K to 348 K for cage 4.

5.7 Cage 5

Cage **5** was predominantly low spin between 248 K and 348 K. Fitting did not converge with or without fixing δ_{LS} .



Figure S249. Chemical shift changes in the range of 248 K to 348 K for cage 5.



Figure S250. Chemical shift changes in the range of 248 K to 348 K for cage 5.

5.8 Cage 6

While fitting converged with and without fixing δ_{LS} , large errors were obtained for the fit without a fixed δ_{LS} . A spin-crossover temperature of 401 K was obtained from the fitting with δ_{LS} fixed as 8.43 ppm.



Figure S251. Chemical shift changes in the range of 248 K to 348 K for cage 6.



Figure S252. Chemical shift changes in the range of 248 K to 348 K for cage 6.



Figure S253. Fitting of the chemical shift versus temperature data for cage **6** where δ_{LS} was a) fixed as 8.43 ppm b) unfixed.



Figure S254. Spin-state population of cage 6.

5.9 Cage 7

While fitting converged with and without fixing δ_{LS} , large errors were obtained for the fit without fixing δ_{LS} . A spin-crossover temperature of 430 K was obtained from the fitting with δ_{LS} fixed as 8.70 ppm.



Figure S255. Chemical shift changes in the range of 248 K to 348 K for cage **7**. The appearance of additional signals above 318 K is attributed to the formation of other species, possibly due to the lability of the metal-ligand bonds at the elevated temperatures.



Figure S256. Chemical shift changes in the range of 248 K to 348 K for cage 7.



Figure S257. Fitting of the chemical shift versus temperature data for cage **7** where δ_{LS} was a) fixed as 8.70 ppm b) unfixed.



Figure S258. Spin-state population of cage 7.
5.10 Cage 8

While fitting converged with and without fixing δ_{LS} , large errors were obtained for the fit without fixing δ_{LS} . A spin-crossover temperature of 375 K was obtained from the fitting with δ_{LS} fixed as 8.48 ppm.



Figure S259. Chemical shift changes in the range of 248 K to 348 K for cage **8**. The appearance of additional signals above 308 K is attributed to the formation of other species, possibly due to the lability of the metal-ligand bonds at the elevated temperatures.



Figure S260. Chemical shift changes in the range of 248 K to 348 K for cage 8.



Figure S261. Fitting of the chemical shift versus temperature data for cage **8** where δ_{LS} was a) fixed as 8.48 ppm b) unfixed.



Figure S262. Spin-state population of cage 8.

5.11 Cage 9

While fitting converged with and without fixing δ_{LS} , large errors were obtained for the fit without fixing δ_{LS} . A spin-crossover temperature of 339 K was obtained from fitting with δ_{LS} fixed as 8.40 ppm.



Figure S263. Chemical shift changes in the range of 248 K to 348 K for cage **9**. The appearance of additional signals above 308 K is attributed to the formation of other species, possibly due to the lability of the metal-ligand bonds at the elevated temperatures.



Figure S264. Chemical shift changes in the range of 248 K to 348 K for cage 9.



Figure S265. Fitting of the chemical shift versus temperature data for cage **9** (proton j) where δ_{LS} was a) fixed as 8.40 ppm b) unfixed.



Figure S266. Spin-state population of cage 9.



Figure S267. Fitting of the chemical shift versus temperature data for cage **9** (proton m) where δ_{LS} was fixed as 8.35 ppm.



Figure S268. Fitting of the chemical shift versus temperature data for cage **9** (proton c) where δ_{LS} was fixed as 7.75 ppm.



Figure S269. Fitting of the chemical shift versus temperature data for cage **9** (proton a) where δ_{LS} was fixed as 4.23 ppm.



Figure S270. Fitting of the chemical shift versus temperature data for cage **9** (proton d) where δ_{LS} was fixed as 7.45 ppm.



Figure S271. Fitting of the chemical shift versus temperature data for cage **9** (proton k) where δ_{LS} was fixed as 8.11 ppm. The fitting did not converge.



Figure S272. Fitting of the chemical shift versus temperature data for cage **9** (proton q) where δ_{LS} was fixed as 7.06 ppm. The fitting did not converge.



Figure S273. Fitting of the chemical shift versus temperature data for cage **9** (proton e) where δ_{LS} was fixed as 7.06 ppm.



Figure S274. Fitting of the chemical shift versus temperature data for cage **9** (proton p) where δ_{LS} was fixed as 1.92 ppm. The fitting did not converge.



Figure S275. Fitting of the chemical shift versus temperature data for cage **9** (proton f) where δ_{LS} was fixed as 6.63 ppm. The fitting did not converge.

| | Cage 9 | |
|--------|------------------|-----------------|
| Proton | T _{1/2} | δ _{LS} |
| | [K] | [ppm]ª |
| m | 339 | 8.35 |
| j | 339 | 8.40 |
| C | 341 | 7.75 |
| а | 349 | 4.23 |
| d | 226 | 7.45 |
| k | b | 8.11 |
| q | b | 7.06 |
| е | 358 | 7.06 |
| р | b | 1.92 |
| f | b | 6.63 |

Table S2. Spin-crossover temperatures from variable temperature NMR experiments(248-348 K) for cage 9 fitting different protons.

^a Chemical shift of the Zn(II) cage analogue. ^b Could not be determined since fitting did not converge.

5.12 Cage 10

While fitting converged with and without fixing δ_{LS} , large errors were obtained for the fit without fixing δ_{LS} . A spin-crossover temperature of 353 K was obtained from the fitting with δ_{LS} fixed as 8.18 ppm.



Figure S276. Chemical shift changes in the range of 248 K to 348 K for cage **10**. The appearance of additional signals above 318 K is attributed to the formation of other species, possibly due to the lability of the metal-ligand bonds at the elevated temperatures.



Figure S277. Chemical shift changes in the range of 248 K to 348 K for cage 10.



Figure S278. Fitting of the chemical shift versus temperature data for cage **10** where δ_{LS} was a) fixed as 8.18 ppm b) unfixed.



Figure S279. Spin-state population of cage 10.

5.13 Cage 11

Cage **11** was predominantly high spin above 248 K and showed Curie-Weiss behavior above 310 K. Therefore, fitting was performed for data between 248 K and 308 K. While fitting converged with δ_{LS} fixed as 8.48 ppm giving a spin-crossover temperature of 244 K, the fitting did not converge when δ_{LS} was not fixed.



Figure S280. Chemical shift changes in the range of 248 K to 348 K for cage 11.



Figure S281. Chemical shift changes in the range of 248 K to 348 K for cage 11.



Figure S282. Fitting of the chemical shift versus temperature data for cage **11** (proton j) where δ_{LS} was fixed as 8.46 ppm. The fitting without fixing δ_{LS} did not converge.



Figure S283. Spin-state population of cage 11.



Figure S284. Fitting of the chemical shift versus temperature data for cage **11** (proton m) where δ_{LS} was fixed as 7.70 ppm.



Figure S285. Fitting of the chemical shift versus temperature data for cage **11** (proton c) where δ_{LS} was fixed as 7.83 ppm.



Figure S286. Fitting of the chemical shift versus temperature data for cage 11 (proton a) where δ_{LS} was fixed as 4.36 ppm.



Figure S287. Fitting of the chemical shift versus temperature data for cage 11 (proton n) where δ_{LS} was fixed as 2.63 ppm.



Figure S288. Fitting of the chemical shift versus temperature data for cage **11** (proton d) where δ_{LS} was fixed as 7.51 ppm. The fitting did not converge.



Figure S289. Fitting of the chemical shift versus temperature data for cage 11 (proton e) where δ_{LS} was fixed as 7.05 ppm. The fitting did not converge.



Figure S290. Fitting of the chemical shift versus temperature data for cage **11** (proton p) where δ_{LS} was fixed as 7.08 ppm. The fitting did not converge.



Figure S291. Fitting of the chemical shift versus temperature data for cage **11** (proton f) where δ_{LS} was fixed as 6.63 ppm. The fitting did not converge.

| | Cage 11 | |
|--------|------------------|--------------------|
| Proton | T _{1/2} | δ_{LS} |
| | [K] | [ppm] ^a |
| j | 244 | 8.46 |
| m | 286 | 7.70 |
| С | 247 | 7.83 |
| а | 244 | 4.36 |
| n | 268 | 2.63 |
| d | b | 7.51 |
| е | b | 7.05 |
| р | b | 7.08 |
| f | b | 6.63 |

Table S3. Spin-crossover temperatures from variable temperature NMR experiments(248-308 K) for cage **11** for different protons.

^a Chemical shift of the Zn(II) cage analogue. ^b Could not be determined since fitting did not converge.

6 X-Ray Crystallography

Data collection for compound **18**, cages **5** and **7** was performed with a XtaLAB Synergy, Dualflex, HyPix diffractometer with CuK α radiation ($\lambda = 1.54184$). The structures were solved with SHELXT^[11] and refined with SHELXL^[12] using Least Squares minimisation. All non-hydrogen atoms were refined anisotropic. The C-H H atoms were positioned with idealized geometry and were refined isotropic with U_{iso}(H) = 1.2 U_{eq}(C). The crystal of cage **7** was racemically twinned and therefore, a twin refinement was performed leading to a BASF parameter of 0.455(8). In cage **5** six of the eight anions were located, whereas none of them can be located in cage **7**. Therefore, the contribution of the missing anions to the electron density map was removed but they were considered in the calculation of the molecular formula. The anions but also all other atoms have extremely high components of the anisotropic displacement parameters indicating for disorder. Several attempts were made to find a reasonable split model without any success but to reach convergence much of restraints must be used. All these problems can be traced back to the very poor crystal quality. Even on extremely long exposure times, especially cage **7** do not diffract below about 1.2 Å.

A table with selected crystal data and results for the structure refinement can be found in Tables S2 and S3.

CCDC-2207255 (compound **18**), CCDC-2207256 (cage **5**) and CCDC-2207257 (cage **7**) contain the supplementary crystallographic data for this paper. These data can be obtained free charge from the Cambridge Crystallographic Data Centre via http://www.ccdc.cam.ac.uk/data_request/cif.

Preparation of the single crystals

Crystals of compound **18** were obtained from slow evaporation of a solution in acetone. Crystals of cages **5** and **7** were obtained by vapour diffusion of diethyl ether into a CD_3CN solution of the cages.

| Compound | Compound 18 | Cage 5 | Cage 7 |
|---|---------------------|---|---|
| formula | $C_{19}H_{14}BrN_3$ | $C_{162}H_{120}F_{24}Fe_4N_{36}O_{24}S_8$ | $C_{200}H_{144}Fe_4N_{36}O_{24}F_{24}S_8$ |
| MW / g mol ⁻¹ | 364.24 | 3890.81 | 4371.38 |
| crystal system | triclinic | orthorhombic | orthorhombic |
| space group | P-1 | Pccn | Pna2₁ |
| a/Å | 10.6730(1) | 67.4664(11) | 38.3429(5) |
| b/Å | 11.4988(2) | 17.1648(4) | 18.0379(4) |
| c/Å | 12.9614(2) | 32.7981(2) | 37.1508(6) |
| α / ° | 73.861(1) | 3290 | 90 |
| β/° | 85.843(1) | 90 | 90 |
| γl° | 87.851(1) | 90 | 90 |
| V/Å ³ | 1523.75(4) | 37981.7(11) | 25694.5(7) |
| <i>T /</i> K | 100.00(1) | 100.0(1) | 100.0 (1) |
| Ζ | 4 | 8 | 4 |
| $D_{ m calc}$ / g cm $^{-3}$ | 1.588 | 1.361 | 1.130 |
| μ / mm ⁻¹ | 3.685 | 4.041 | 3.040 |
| Crystal size /mm ³ | 0.18×0.06×0.03 | 0.05×0.24×0.25 | 0.2×0.22×0.3 |
| $2\theta_{max}$ / deg | 159.914 | 159.708 | 133.198 |
| measured refl. | 20304 | 115736 | 88529 |
| R _{int} | 0.0169 | 0.0882 | 0.0596 |
| unique refl. | 6503 | 39182 | 36631 |
| refl. <i>F</i> ₀ >4 <i>o</i> (<i>F</i> ₀) | 6385 | 29968 | 21859 |
| parameter | 416 | 2210 | 2091 |
| $R_1 [F_0 > 4\sigma F_0)]$ | 0.0274 | 0.1603 | 0.1165 |
| wR2 [all data] | 0.0764 | 0.4511 | 0.3504 |
| GOF | 1.098 | 1.807 | 1.185 |
| $\varDelta ho_{max/min}$ / e Å ⁻³ | 0.58/-0.52 | 2.62/ -0.1.66 | 0.61/ -0.45 |

Table S4. Selected crystal data and details of the structure refinements for compound 18,cage 5 and cage 7.



Figure S292. Crystal structure of compound 18.

According to Shatruk and co-workers,^[13] the N-N distance from an X-ray crystal structure of a

ligand can be used to predict the spin-state of the corresponding Fe^{II} homoleptic complex. Based on the N-N distance (2.81 Å) in compound **18**, Fe^{II} homoleptic complexes based on this coordination motif should show spin-crossover.



Figure S293. Crystal structure of cage **5**. Hydrogen atoms and non-encapsulated anions are omitted for clarity. The triflate anion was observed to bind in the cavity.



Figure S294. Crystal structure of cage **7**. Hydrogen atoms are omitted for clarity and the anions could not be located during refinement.

7 Anion Binding Studies

Cages **7** and **11** were prepared according to Section 4.7 and 4.11 and the ¹H and ¹⁹F NMR spectra were measured at 298 K and 248 K. Afterwards, 8 equivalents (relative to the cage) of either TBABF₄ (3.65 mg, 11.1 μ mol) or TBANTf₂ (5.80 mg, 11.1 μ mol) were added and the solutions were directly analysed by NMR spectroscopy.





-110 -90 -100 -120 -130 -140 -150 -160 -170 -180 -70 -80 -190 ppm Figure S296. ¹⁹F NMR spectra (471 MHz, CD₃CN, 298 K) of cage 7 before (bottom) and after addition of 8 equivalents TBABF₄ (top).

7.2 Cage 11 + BF₄⁻



Figure S297. ¹H NMR spectra (500 MHz, CD₃CN, 298 K) of cage **11** before (bottom) and after addition of 8 equivalents TBABF₄ (top).



Figure S298. ¹H NMR spectra (500 MHz, CD₃CN, 248 K) of cage **11** before (bottom) and after addition of 8 equivalents TBABF₄ (top).



Figure S299. ¹⁹F NMR spectra (471 MHz, CD₃CN, 298 K) of cage **11** before (bottom) and after addition of 8 equivalents TBABF₄ (top).



Figure S300. ¹⁹F NMR spectra (471 MHz, CD₃CN, 248 K) of cage **11** before (bottom) and after addition of 8 equivalents $TBABF_4$ (top).



Figure S301. ¹H NMR spectra (500 MHz, CD₃CN, 298 K) of cage **7** before (bottom) and after addition of 8 equivalents TBANTf₂ (top).



Figure S302. ¹H NMR spectra (500 MHz, CD₃CN, 248 K) of cage **7** before (bottom) and after addition of 8 equivalents TBANTf₂ (top).





Figure S304. ¹⁹F NMR spectra (471 MHz, CD₃CN, 248 K) of cage **7** before (bottom) and after addition of 8 equivalents TBANTf₂ (top).

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