Supporting Information

Impact of central atom and halido ligand on structure, antiproliferative activity and selectivity of half-sandwich Ru(II) and Ir(III) complexes with a 1,3,4-thiadiazolebased ligand

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Fig. S1. ESI+ MS spectrum of complex 1 given with a comparison of the experimental and calculated isotopic distributions of the $[Ru(\eta^6-pcym)(L1)Cl]^+$ species (inset).



Fig. S2. ESI+ MS spectrum of complex **2** given with a comparison of the experimental and calculated isotopic distributions of the [Ir(n⁵-Cp*)(L1)Cl]⁺ species (inset).



Fig. S3. ESI+ MS spectrum of complex **3** given with a comparison of the experimental and calculated isotopic distributions of the $[Ru(\eta^6-pcym)(L1)I]^+$ species (inset).



Fig. S4. ESI+ MS spectrum of complex **4** given with a comparison of the experimental and calculated isotopic distributions of the $[Ir(\eta^5-Cp^*)(L1)I]^+$ species (inset).







Fig. S6. ¹H NMR spectrum of complex **1** in DMSO-*d*₆.



Fig. S8. ¹H NMR spectrum of complex **3** in DMSO- d_6 .



Fig. S9. ¹H NMR spectrum of complex 4 in DMSO- d_6 .



Fig. S10. Comparison of ¹H NMR spectra of ligand L1 (*top*) and complexes **2** and **4** (*middle* and *bottom*) in DMSO- d_6 .



Fig. S11. A part of ¹H-¹³C gs-HMQC NMR spectrum of complex **1** in DMSO- d_6 showing the signals of coordinated L1.



Fig. S12. A part of ¹H-¹³C gs-HMQC NMR spectrum of complex **4** in DMSO- d_6 showing the signals of coordinated L1.



Fig. S13. A perspective view on the non-covalent interactions (black dashed lines) involving PF₆⁻ anions (*Left column*). The $V^2 \rho(\mathbf{r})$ contour (*middle column*) and sign λ (*right column*) plots calculated for **1-4**. Blue dots represent (3,-1) critical points, orange dots represent (3,+1) ring critical points and brown dots represents (3,-3) atomic critical points. Brown lines represent bond paths. Atom labelling is provided only in the sign λ plots however, view direction in the $V^2 \rho(\mathbf{r})$ contour plot is identical. The parameters of the depicted non-covalent interactions (in Å) and their interaction energies (*E*_{int} in kcal/mol): **1**, *d*(C10···N2) = 3.300(8), *E*_{int} = 1.49, *d*(C14···F3) = 3.451(9), *E*_{int} = 2.27, *d*(F4···S1) = 3.246(7), *E*_{int} = 1.00, *d*(F5···S1) = 3.422(6), *E*_{int} = 0.69; **2**, *d*(C10···N2) = 3.377(6), *E*_{int} = 1.58, *d*(C14···F2) = 3.688(6), *E*_{int} = 0.78, *d*(C19···F5) = 3.434(6), *E*_{int} = 1.20, *d*(F2···S1) = 3.024(3), *E*_{int} = 1.89, *d*(F5···S1) = 3.156(4), *E*_{int} = 1.31; **3**, *d*(C12···N2) = 3.300(8), *E*_{int} = 2.32, *d*(C4···F2) = 3.314(6), *E*_{int} = 1.32, *d*(C4···F3) = 3.398(5), *E*_{int} = 1.29, *d*(F3···S1) = 3.212(4), *E*_{int} = 1.08, **4**, *d*(C7···N2) = 3.422(8), *E*_{int} = 1.59, *d*(C14···F5) = 3.660(6), *E*_{int} = 0.87, *d*(C19···F3) = 3.459(8), *E*_{int} = 1.33, *d*(F3···S1) = 3.156(8), *E*_{int} = 1.31, *d*(F5···S1) = 3.025(7), *E*_{int} = 1.89.



Fig. S14. A perspective view on the non-covalent interactions (black dashed lines) in **2** (*left*) and **4** (*right*). The selected donor···acceptor distances (in Å): **2**, $d(C21\cdotsO1) = 3.406(7)$, $d(Cl1\cdotsO1) = 3.670(4)$; **4**, $d(C21\cdotsO1) = 3.580(18)$, $d(11\cdotsO1) = 3.668(8)$.



Fig. S15. $\vec{V}^2 \rho(\mathbf{r})$ contour (*left column*), ELF (*middle column*) and sign λ (*right column*) plots calculated for **2** (above) and **4** (below). Blue dots represent (3,-1) critical points, orange dots represent (3,+1) ring critical points and brown dots represents (3,-3) atomic critical points. Brown lines represent bond paths. Atom labelling is provided only in the $\vec{V}^2 \rho(\mathbf{r})$ plots however, view direction in all the depicted plots is identical. Selected energetic and topological parameters: **2**, C21…O1, BCP(3,-1), $H_r = 0.124 \times 10^{-2}$ a.u., $E_{int} = 1.10$ kcal/mol, sign $\lambda = -0.986.10^{-2}$; Cl1…O1, BCP(3,-1), $H_r = 0.589 \times 10^{-3}$ a.u., $E_{int} = 0.40$ kcal/mol, sign $\lambda = 0.290 \times 10^{-2}$, **4**, C21…O1, BCP(3,-1), $H_r = 0.926 \times 10^{-3}$ a.u., $E_{int} = 0.50$ kcal/mol, sign $\lambda = -0.559 \times 10^{-2}$; l1…O1, BCP(3,-1), $H_r = 0.100 \times 10^{-2}$ a.u., $E_{int} = 0.86$ kcal/mol, sign $\lambda = -0.572 \times 10^{-2}$.



Fig. S16. The graphical comparison of relative enthalpies (*left*) and Gibbs energies (*right*) for ground states of *E/Z*-isomers of L1 and **1-4**, and the transition states derived from DFT calculations.

	1	2	3		
Formula	$C_{21}H_{21}CIF_6N_3OPRuS$	$C_{21}H_{22}CIF_6IrN_3OPS$	$C_{21}H_{21}F_6IN_3OPRuS$		
<i>M</i> _r	644.96	737.09	736.41		
Crystal system	Monoclinic <i>, Cc</i>	Monoclinic, P2 ₁ /c	Triclinic, $P\overline{1}$		
a /Å	8.8814(6)	12.5825(5)	9.1768(2)		
<i>b /</i> Å	22.8613(13)	12.3006(3)	10.2321(2)		
<i>c /</i> Å	12.7791(9)	15.4471(6)	15.0893(2)		
α/°	90	90	73.270(2)		
β/°	109.276(5)	90.752(3)	75.493(2)		
γl°	90	90	78.754(2)		
V/ų	2449.2(3)	2390.58(15)	1302.03(5)		
Ζ	4	4	2		
T/K	100.0(2)	100.0(2)	293(2)		
D _c /g cm ⁻³	1.749	2.048	1.878		
μ / mm ⁻¹	8.215 (CuKa)	13.922 (CuKa)	16.092(CuKa)		
F(000)	1288	1424	716		
Data/restraints/para meters	4115/2/320	4541/0/321	4723/0/320		
Goodness-of-fit (GOF) on F ²	0.895	1.106	1.077		
R_1 , w R_2 ($l > 2\sigma(l)$) ^{<i>a, b</i>}	0.0414/0.828	0.0285/0.0687	0.0388/0.1101		
R ₁ , wR ₂ (all data) ^{a, b}	0.0506/0.0842	0.0332/0.0704	0.0399/0.1117		
CCDC number	2266585	2266586	2266587		
^{<i>a</i>} R ₁ = $\sum (F_o - F_c) / \sum F_o $, ^{<i>b</i>} $wR_2 = \{\sum [w(F_o^2 - F_c^2)^2] / \sum [w(F_o^2)^2]\}^{1/2}$					

Table S1. Crystallographic data for the reported compounds 1-4.

	4	
Formula	$C_{21}H_{22}F_{6}IIrN_{3}OPS$	
Mr	828.54	
Crystal system	Monoclinic, P2 ₁ /c	
a /Å	12.5694(2)	
b / Å	12.32870(10)	
c / Å	15.6996(2)	
α/°	90	
β/°	90.5490(10)	
γ/°	90	
V / ų	2432.77(5)	
Ζ	4	
T/K	100.0(2)	
$D_{\rm c}$ / g cm ⁻³	2.262	
μ / mm ⁻¹	22.635 (CuKa)	
F(000)	1568	
Data/restraints/para	80133/0/321	
meters		
Goodness-of-fit (GOF)	1.076	
on F ²		
R_1 , w R_2 ($l > 2\sigma(l)$) ^{<i>a, b</i>}	0.0504/0.1526	
R ₁ , wR ₂ (all data) ^{<i>a, b</i>}	0.0575/0.1569	
CCDC number	2266588	

Table S2. The comparisons of DFT calculated selected bond distances^{*a*} and Mayer bond orders for *E/Z* isomers of **1-4**.^{*b*}

	M-X bond	M-N _{py} bond	M-N _{th} bond
1	2.410/2.410	2.085/2.085	2.037/2.037
[Ru(ŋ ⁶ -pcym)(L1)Cl]⁺	[2.394(2)]	[2.116(10)]	[2.052(10)]
	0.9374/0.9348	0.6050/0.6064	0.5726/0.5697
3	2.738/2.738	2.082/2.082	2.035/2.034
[Ru(ŋ⁰-pcym)(L1)I]⁺	[2.6986(4)]	[2.120(3)]	[2.063(3)]
	0.9336/0.9336	0.6140/0.6158	0.5763/0.5736
2	2.420/2.419	2.112/2.112	2.068/2.069
[Ir(ŋ⁵-Cp*)(L1)Cl]	[2.3857(11)]	[2.114(4)]	[2.063(4)]
	0.7741/0.7733	0.5050/0.5051	0.5257/0.5223
4	2.730/2.731	2.107/2.107	2.063/2.063
[lr(ŋ ⁵ -Cp*)(L1)I]	[2.6445(9)]	[2.093(8)]	[2.054(8)]
	0.6679/0.6683	0.5149/0.5150	0.5389/0.5353

^{*a*} M-X stands for the metal-halogen bond, M-N_{py} stands for the metal-nitrogen bond to the pyridine part of the respective ligand, and M-N_{th} stands for the metal-nitrogen bond to the thiadiazole part of the respective ligand. Bond distances are in Å, and the values in square brackets correspond to the data from X-ray analysis. ^{*b*} Calculated Mayer bond orders are in italics for the respective *E/Z* isomers.



Fig. S17. ¹H NMR spectra of complex 1 in SM1 (30% MeOD- $d_4/70\%$ D₂O), as observed at different time points (t = 0 or 48h).



Fig. S18. ¹H NMR spectra of complex 2 in in SM1 (30% MeOD- $d_4/70\%$ D₂O), as observed at different time points (t = 0 or 48h).



Fig. S19. ¹H NMR spectra of complex 3 in in SM1 (30% MeOD- $d_4/70\%$ D₂O), as observed at different time points (t = 0 or 48h).



Fig. S20. ¹H NMR spectra of complex 4 in in SM1 (30% MeOD- $d_4/70\%$ D₂O), as observed at different time points (t = 0 or 48h).

Fig. S21. ¹H NMR spectra of complex **1** in SM2 (30% MeOD-*d*₄/70% of PBS in D₂O (pH=7.4)), as observed at different time points (t = 0 or 48h).

Fig. S22. ¹H NMR spectra of complex **2** in SM2 (30% MeOD- $d_4/70\%$ of PBS in D₂O (pH=7.4)), as observed at different time points (t = 0 or 48h). Spectra of the dechlorinated species **2**^h (in 30% MeOD- $d_4/70\%$ D₂O) and ligand L1 (in SM2) are depicted as well for comparative purposes.

Fig. S23. ¹H NMR spectra of complex **3** in SM2 (30% MeOD-*d*₄/70% of PBS in D₂O (pH=7.4)), as observed at different time points (t = 0 or 48h).

Fig. S24. ¹H NMR spectra of complex **4** in SM2 (30% MeOD- $d_4/70\%$ of PBS in D₂O (pH=7.4)), as observed at different time points (t = 0 or 48h). Spectra of the dechlorinated species **2**^h (in 30% MeOD- $d_4/70\%$ D₂O) and complex **2** (in SM2) were depicted as well for comparative purposes, with light blue colour denoting the signals of the original iodido complex and with yellow for the chlorido analogue. Other signals are not coloured due to overlap.

Fig. S25. ¹H NMR spectra in the range of 5.7-10 ppm of complex **1** in SM3 (30% MeOD- $d_4/70\%$ of PBS in D₂O (pH=7.4) + 5M equivalents of GSH), as observed at different time points (t = 0 or 48h).

Fig. S26. ¹H NMR spectra in the range of 0.9-4.6 ppm of complex **1** in SM3 (30% MeOD- $d_4/70\%$ of PBS in D₂O (pH=7.4) + 5M equivalents of GSH), as observed at different time points (t = 0 or 48h). Blue arrows indicate β -CH₂ resonances of oxidized GSH (GSSG). For comparative purposes, the ¹H spectra of GSH and GSSG are shown (*top*).

Fig. S27. ¹H NMR spectra in the range of 6.5-9.5 ppm of complex **2** in SM3 (30% MeOD- $d_4/70\%$ of PBS in D₂O (pH=7.4) + 5M equivalents of GSH), as observed at different time points (t = 0 or 48h).

Fig. S28. ¹H NMR spectra in the range of 1.1-4.6 ppm of complex **2** in SM3 (30% MeOD- $d_4/70\%$ of PBS in D₂O (pH=7.4) + 5M equivalents of GSH), as observed at different time points (t = 0 or 48h). For comparative purposes, the ¹H spectra of GSH and GSSG are shown (*top*).

Fig. S29. ¹H NMR spectra in the range of 5.7-10 ppm of complex **3** in SM3 (30% MeOD- $d_4/70\%$ of PBS in D₂O (pH=7.4) + 5M equivalents of GSH), as observed at different time points (t = 0 or 48h).

Fig. S30. ¹H NMR studies in the range of 0.9-4.6 ppm of complex **3** in SM3 (30% MeOD- $d_4/70\%$ of PBS in D₂O (pH=7.4) + 5M equivalents of GSH), as observed at different time points (t = 0 or 48h). For comparative purposes, the ¹H spectra of GSH and GSSG are shown (*top*).

Fig. S31. ¹H NMR spectra in the range of 6.5-9.2 ppm of complex **4** in SM3 (30% MeOD- $d_4/70\%$ of PBS in D₂O (pH=7.4) + 5M equivalents of GSH), as observed at different time points (t = 0 or 48h). A spectrum of **2** in the same medium is depicted for comparative purposes. Orange areas denote discreet signals belonging to chlorido complex, other signals are not highlighted due to overlap.

¹H NMR Chemical shift (ppm)

Fig. S32. ¹H NMR studies in the range of 1.6-4.6 ppm of complex **4** in SM3 (30% MeOD- $d_4/70\%$ of PBS in D₂O (pH=7.4) + 5M equivalents of GSH), as observed at different time points (t = 0 or 48h). Orange area denotes a Cp* signal belonging to chlorido complex. For comparative purposes, the ¹H spectra of GSH and GSSG are shown (*top*).

¹H NMR Chemical shift (ppm)

Fig. S33. ¹H NMR studies of complex 2 in SM4 (30% MeOD-*d*₄/70% of PBS in D₂O (pH=7.4) + 5M equivalents of NADH), as observed at different time points (t = 0 or 48h). Asterisks denote the signals of NAD⁺ originating from oxidation of NADH in the mixture with 2. Blue areas mark the signals of 2. For comparative purposes, the ¹H spectra of NADH and NAD⁺ are shown (*top*).

¹H NMR Chemical shift (ppm)

Fig. S34. ¹H NMR studies of complex 3 in SM4 (30% MeOD-*d*₄/70% of PBS in D₂O (pH=7.4) + 5M equivalents of NADH), as observed at different time points (t = 0 or 48h). Asterisks denote the signals of NAD⁺ originating from oxidation of NADH in the mixture with 3. Blue areas mark the signals of 3. For comparative purposes, the ¹H spectra of NADH and NAD⁺ are shown (*top*).

Fig. S35. ¹H NMR studies of complex **4** in SM4 (30% MeOD- $d_4/70\%$ of PBS in D₂O (pH=7.4) + 5M equivalents of NADH), as observed at different time points (t = 0 or 48h). Asterisks denote the signals of NAD⁺ originating from oxidation of NADH in the mixture with **4**. Blue areas mark the signals of **4**, with blue circles indicating discreet signals of chlorido complex. For comparative purposes, the ¹H spectra of NADH and NAD⁺ are shown (*top*).

Fig. S36 The effect of **1** (*top A, B*), **4** (*middle C, D*) and CDDP (*bottom E, F*) on metabolic activity (MTT assay) of A2780 and A2780cis ovarian carcinoma cell lines. Metabolic activity was analysed 24 and 48h after treatment of cells with selected complexes. The experimental groups were compared with untreated control (* p < 0.05, ** p < 0.01, *** p < 0.001).

Fig. S37. Effect of **1**, **4** and CDDP on changes in viability (A) and mitochondrial membrane potential (B) of A2780 and A2780cis ovarian carcinoma cell lines. TMRE+ cells represent cells with normal mitochondrial membrane potential. Viability and mitochondrial membrane potential were analysed 48h after treatment of cells with selected complexes. The experimental groups were compared with untreated control (*** p < 0.001).

Fig. S39. Control experiments showing the ¹H NMR spectra of NADH in 30% MeOD- $d_4/70\%$ of PBS in D₂O (pH=7.4) at different time points (t = 0 or 48h) measured under the same experimental conditions as for the interaction experiments with **1-4**. No changes are observed. For comparative purposes, the ¹H spectrum of NAD⁺ is shown (*bottom*).