Phosphorescent Cyclometalated Ir(III) Complex-

Functionalized Molecularly-Imprinted Polymer for Specific

Trimethylamine Detection

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Supplementary Information

Contents

1. Experimental Section

2. Supplementary Figures

Fig. S1	SEM images of PTMA and PTMA-Ir .				
Fig. S2	SEM images of the control polymers without TMA-imprinting cavities (P				
	and P-Ir) prepared according to identical synthetic procedures for PTMA				
	and PTMA-Ir except no imprinting template was used in the synthesis.				
Fig. S3	UV-vis reflectance spectra of PTMA, Ir-2 and PTMA-Ir deposited on a				
	quartz plate with a 1-cm circular recess.				
Fig. S4	FT-IR spectra of PTMA and PTMA-Ir.				
Fig. S5	(a) XPS survey spectrum of PTMA and deconvoluted fittings of the high				
	resolution (b) C 1s, (c) O 1s and (d) Cl 2p signals.				
Fig. S6	(a) XPS survey spectrum of PTMA-Ir and deconvoluted fittings of the high				
	resolution (b) C 1s, (c) O 1s, (d) N 1s and (e) Ir 4f signals.				
Fig. S7	Nanosecond time-resolved emission spectra of PTMA-Ir suspended in H ₂ O				
	$(\lambda_{ex} = 355 \text{ nm}).$				
Fig. S8	Overlaid emission spectra of PTMA-Ir and PTMA suspended in $H_2O(0.2)$				
	mg polymer in 3 mL H ₂ O) ($\lambda_{ex} = 350$ nm)				
Fig. S9	Extracted-ion chromatograms (XIC) of derivatized TMA and TEA				
	monitored at their specific multiple reaction monitoring (MRM) transitions				
	in the study of PTMA-Ir . (a), (b) and (c) are spiked sample replicates, and				
	(d) is the negative control.				
Fig. S10	Extracted-ion chromatograms (XIC) of derivatized TMA and TEA				
	monitored at their specific MRM transitions in the study of P-Ir. (a), (b)				
	and (c) are spiked sample replicates, and (d) is the negative control.				
Fig. S11	Emission spectra of PTMA-Ir suspended in an acidic aqueous solution (pH				
	= 2) after incubation in different concentrations of TMA·HCl for 120 min				
	$(\lambda_{ex} = 350 \text{ nm}).$				

- **Fig. S12** Emission spectra of **Ir-2** in CH₃CN/H₂O (3:2 v/v) after incubation with different concentrations of TMA·HCl ($\lambda_{ex} = 350 \text{ nm}$)
- Fig. S13 Emission spectra of T-Ir upon exposure to different concentrations of TMA vapor under an air atmosphere ($\lambda_{ex} = 350$ nm).
- **Fig. S14** Time-dependent emission spectra of **PTMA-Ir** under an argon atmosphere upon exposure to 634 ppm TMA vapor ($\lambda_{ex} = 350$ nm).
- Fig. S15Emission responses of T-Ir under an air atmosphere upon exposure to 1800ppm of TMA vapor and removal from TMA vapor over five cycles.
- Fig. S16 Emission quenching of T-Ir under an argon atmosphere upon exposure to 3950 ppm vapor of TMA and saturated vapor of other interfering reagents at 20 °C [TEA (~75,000 ppm), DIPEA (~14,100 ppm), acetic acid (~14,800 ppm), DMF (~3,800 ppm), THF (~188,000 ppm) and MeCN (~108,600 ppm)].
- Fig. S17Emission quenching of T-Ir under an air atmosphere upon exposure to 1800ppm vapor of TMA and other interfering reagents.
- **Fig. S18** Emission spectra of **Ir-2** loaded on a filter paper upon exposure to different concentrations of TMA vapor under an argon atmosphere ($\lambda_{ex} = 300$ nm).
- **Fig. S19** Emission spectra of **Ir-2** loaded on a filter paper upon exposure to different concentrations of TMA vapor under an air atmosphere ($\lambda_{ex} = 300$ nm).
- Fig. S20 ¹H NMR spectra of (a) [Ir-1] in $(CD_3)_2CO$ and (b) [Ir-2] in CDCl₃.
- Fig. S21 Positive ESI mass spectrum of Ir-1. The inset shows the expanded ion cluster at m/z = 713.2 and simulated isotope pattern based on the molecular formulation of Ir-1.
- Fig. S22 Positive ESI mass spectrum of Ir-2. The inset shows the expanded ion cluster at m/z = 741.2 and simulated isotope pattern based on the molecular formulation of Ir-2.
- Table S1.
 Comparison of PTMA-Ir sensor with recently reported TMA sensors

3. References

1. Experimental Section

{**Ir(ppy)**₂[**phen(OH)**₂]}**PF**₆ (Ir-1). ¹H NMR (300 MHz, (CD₃)₂CO): δ 8.38 (2H, s, 5-, 6-H's of phenathroline), 8.22 (2H, d, J = 8.1 Hz, two 6-pyridyl H's of ppy), 7.89 (6H, m, two 3-pyridyl H's of ppy, two 4-pyridyl H's of ppy and two 2-phenyl H's of ppy), 7.80 (2H, d, J = 5.7 Hz, 3-, 8- H's of phenathroline), 7.31 (2H, d, J = 6.1 Hz, 2-, 9- H's of phenathroline), 7.05 (4H, m, two 3-phenyl H's of ppy and two 4-phenyl H's of ppy), 6.93 (2H, m, two 5pyridyl H's of ppy), 6.48 (2H, d, J = 7.4 Hz, two 5-phenyl H's of ppy), 3.83 (s, 2H, OH). Positive-ion ESI-MS: m/z 713.2 [M]⁺; IR (KBr disc / cm⁻¹): 3059 br (O-H), 831 (P-F).

{**Ir(ppy)**₂[(**OMe**)₂**phen**]}**PF**₆ (Ir-2). ¹H NMR (300 MHz, CDCl₃): δ 8.36 (2H, s, 5-, 6- H's of phenathroline), 8.07 (2H, d, *J* = 6.0 Hz, two 6-pyridyl H's of ppy), 7.89 (2H, d, *J* = 8.3 Hz, 3-, 8- H's of phenathroline), 7.70 (4H, d, *J* = 7.4 Hz, two 3-pyridyl H's of ppy and two 4-pyridyl H's of ppy), 7.50 (2H, d, *J* = 6.1 Hz, two 2-phenyl H's of ppy), 7.16 (2H, d, *J* = 6.1 Hz, 2-, 9- H's of phenathroline), 7.05 (2H, m, two 5-pyridyl H's of ppy), 6.94 (4H, m, two 3-phenyl H's of ppy and two 4-phenyl H's of ppy), 6.41 (2H, d, *J* = 7.5 Hz, two 5-phenyl H's of ppy), 4.19 (6H, s, CH₃). Positive-ion ESI-MS: m/z 741.2 [M]⁺; IR (KBr disc / cm⁻¹): 846 (P-F).

Physical Measurements and Instrumentation.

Instrumentation for NMR, IR, UV-vis diffuse reflectance, time-resolved emission and Raman spectroscopy, elemental analysis, TGA and ICP-OES are carried out according to our previously reported literature [1,2]. For Raman spectra, excitation beam (309 nm) is produced by a Nd:YAG laser (Spectra-Physics LAB-170). Solvent signals are subtracted and frequencies are calibrated by methanol Raman bands.

The concentrations of TMA·HCl are quantified by LC-MS using Agilent 1200 HPLC equipped with Sciex API 2000 MS/MS system. Ethyl bromoacetate was used to derivatize TMA, to improve ESI-MS/MS detection and separation of TMA. A portion of 125 μ L supernatant was mixed with 425 μ L of 20% concentrated ammonium solution, triethylamine (TEA) as an internal standard in acetonitrile and 100 μ L of 4.0 M ethyl bromoacetate in acetonitrile. The mixture was stand overnight at 25 °C. Thereafter, 250 μ L of MeCN/Milli-Q water/HCOOH (50:50:1 v/v/v) was added to terminate the reaction. The solution was ready for LC-MS/MS analysis. The chromatographic separation was performed using a Water HILIC silica column (3 μ m, 2.1 × 100 mm) with isocratic elution using 10 mM HCOONH4 in water / 0.1% HCOOH in MeCN (v/v 1:9) as eluent (flow rate: 400 μ L/min). The LC-MS calibration curve is shown below:



A: Peak area, IS: internal standard, triethylamine

Emission quenching experiments with different concentrations of TMA·HCl are conducted in a quartz cell, degassed by bubbling argon. The quenching rate constants are calculated through the Stern-Volmer equation:

$$\tau_0/\tau = 1 + k_q \tau_0[Q]$$

 τ_0 and τ are emission lifetimes without and with quencher, [Q] is quencher concentration in mol dm^{-3,} and k_q is the bimolecular quenching rate constant.

TMA-sensing study

The emission titration was performed on an Edinburgh Instruments FLS980 spectrometer. The **PTMA-Ir** suspended in an aqueous solution was incubated with different concentrations of TMA·HCl for 2 hours, and degassed before the emission measurement. To test the amine vapor sensing, **PTMA-Ir** adsorbed on filter paper (**T-Ir**) was prepared by dipping a filter paper strip (23 mm × 13mm) on a **PTMA-Ir**-suspended solution (1 mg **PTMA-Ir** in 10 mL *n*-hexane). The emission of **PTMA-Ir** on **T-Ir** in a closed quartz cuvette with different concentrations of TMA vapor prepared by the injection of saturated TMA vapor was examined. Saturated TMA vapor is prepared by dissolving TMA·HCl (1 g) in NaOH solution (600 mg NaOH in 10 mL milliQ water) in a closed system at 25 °C for 3 hours. The injected TMA contents are verified by FT-IR spectroscopy with a 10-cm infrared gas cell according to the peak area at 1459 cm⁻¹ as shown in the following figures.



(a) FT-IR spectra and (b) calibration curve of TMA vapor (25 °C).

2. Supplementary figures



Fig. S1 SEM images of (a) PTMA and (b) PTMA-Ir.





Fig. S2 SEM images of the control polymers without TMA-imprinting cavities (P and P-Ir) prepared according to identical synthetic procedures for PTMA and PTMA-Ir except no imprinting template was used in the synthesis.



Fig. S3 UV-vis reflectance spectra of **PTMA**, **Ir-2** and **PTMA-Ir** deposited on a quartz plate with a 1-cm circular recess.







Fig. S5 (a) XPS survey spectrum of **PTMA** and deconvoluted fittings of the high resolution (b) C 1s, (c) O 1s and (d) Cl 2p signals.



Fig. S6 (a) XPS survey spectrum of **PTMA-Ir** and deconvoluted fittings of the high resolution (b) C 1s, (c) O 1s, (d) N 1s and (e) Ir 4f signals.



Fig. S7Nanosecond time-resolved emission spectra of PTMA-Ir suspended in H2O
 $(\lambda_{ex} = 355 \text{ nm}).$



Fig. S8 Overlaid emission spectra of **PTMA-Ir** and **PTMA** suspended in H₂O (0.2 mg polymer in 3 mL H₂O) ($\lambda_{ex} = 350$ nm).



Fig. S9 Extracted-ion chromatograms (XIC) of derivatized TMA and TEA monitored at their specific multiple reaction monitoring (MRM) transitions in the study of **PTMA-Ir**. (a), (b) and (c) are spiked sample replicates, and (d) is the negative control.



Fig. S10 Extracted-ion chromatograms (XIC) of derivatized TMA and TEA monitored at their specific MRM transitions in the study of **P-Ir**. (a), (b) and (c) are spiked sample replicates, and (d) is the negative control.



Fig. S11 Emission spectra of PTMA-Ir suspended in an acidic aqueous solution (pH = 2) after incubation in different concentrations of TMA·HCl for 120 min $(\lambda_{ex} = 350 \text{ nm}).$



Fig. S12 Emission spectra of **Ir-2** in CH₃CN/H₂O (3:2 v/v) after incubation with different concentrations of TMA·HCl ($\lambda_{ex} = 350$ nm).



Fig. S13 Emission spectra of **T-Ir** upon exposure to different concentrations of TMA vapor under an air atmosphere ($\lambda_{ex} = 350$ nm).



Fig. S14 Time-dependent emission spectra of **PTMA-Ir** under an argon atmosphere upon exposure to 634 ppm TMA vapor ($\lambda_{ex} = 350$ nm).



Fig. S15Emission responses of T-Ir under an air atmosphere upon exposure to 1800ppm of TMA vapor and removal from TMA vapor over five cycles.



Fig. S16 Emission quenching of T-Ir under an argon atmosphere upon exposure to 3950 ppm vapor of TMA and saturated vapor of other interfering reagents at 20 °C [TEA (~75,000 ppm), DIPEA (~14,100 ppm), acetic acid (~14,800 ppm), DMF (~3,800 ppm), THF (~188,000 ppm) and MeCN (~108,600 ppm)].



Fig. S17Emission quenching of T-Ir under an air atmosphere upon exposure to 1800ppm vapor of TMA and other interfering reagents.



Fig. S18 Emission spectra of **Ir-2** loaded on a filter paper upon exposure to different concentrations of TMA vapor under an argon atmosphere ($\lambda_{ex} = 300$ nm).



Fig. S19 Emission spectra of **Ir-2** loaded on a filter paper upon exposure to different concentrations of TMA vapor under an air atmophere ($\lambda_{ex} = 300$ nm).







Fig. S21 Positive ESI mass spectrum of [Ir-1]. The inset shows the expanded ion cluster at m/z = 713.2.



Fig. S22 Positive ESI mass spectrum of [Ir-2]. The inset shows the expanded ion cluster at m/z = 741.2.

Sensing Material	Sensing Temp. (°C)	Tested Range (ppm)	LOD	Response Time	Ref.
MoO ₃	300	0.1 - 1000	0.01 ppm	8 s	[3]
Ru-SnO ₂	400	1 – 5	1 ppm	/	[4]
Graphene quantum dots / α-Fe2O3	270	0.01 - 1000	0.01 ppm	11 s	[5]
rGO / In ₂ O ₃	RT	100 - 1500	100 ppm	2 s	[6]
α-Fe ₂ O ₃	250	1 - 100	1 ppm	< 1 min	[7]
Au / PdO / WO _{2.72}	240	1 - 100	1 ppm	4 s	[8]
Co ₃ O ₄ / ZnO	250	0.33 - 66	130 ppb	3 s	[9]
Au / WO3	300	0.5 - 25	0.5 ppm	8 s	[10]
PTMA-Ir	25	0 - 1179 0 - 1276	9 ppm 15 ppm	5 s	This work

Table S1.Comparison of **PTMA-Ir** sensor with recently reported TMA sensors.

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