

# Neutral 2-phenylbenzimidazole-based iridium(III) complexes with picolinate ancillary ligand: Tuning the emission properties by handling the substituent on the benzimidazole ring

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## Experimental details: general consideration

### Synthesis

Commercially available reagents were purchased from Sigma-Aldrich, Alfa Aesar, Acros Organics, TCI Chemical, Merck, Strem or Fluorochem and used as received unless otherwise specified. Solvents were obtained from same commercial sources and used without further purification. For moisture sensitive reactions, glassware was oven-dried prior to use. <sup>1</sup>H NMR spectra were recorded on a 400 MHz and on a 500 MHz in deuterated solvent (CDCl<sub>3</sub>, DMSO-d<sub>6</sub> or CD<sub>2</sub>Cl<sub>2</sub>) and data are reported as follows: chemical shift in ppm from tetramethylsilane with the solvent as an internal indicator (CDCl<sub>3</sub> 7.26 ppm, dmsO-d<sub>6</sub> 2.50 ppm, CD<sub>2</sub>Cl<sub>2</sub> 5.32 ppm), multiplicity (s = singlet, d = doublet, t = triplet, q = quartet, p = pentet, hept = heptuplet, m = multiplet or overlap of non-equivalent resonances), *J*-coupling constants (Hz), and proton integration. <sup>13</sup>C{<sup>1</sup>H} NMR spectra were recorded either at 101 MHz or at 126 MHz in suitable deuterated solvent and data are reported as follows: chemical shift in ppm from tetramethylsilane with the solvent as an internal indicator (CDCl<sub>3</sub> 77.16 ppm, DMSO-d<sub>6</sub> 39.52 ppm, CD<sub>2</sub>Cl<sub>2</sub> 53.84 ppm). <sup>19</sup>F NMR spectra were recorded at 376 MHz and at 470 MHz in the suitable deuterated solvent. <sup>31</sup>P NMR spectra were recorded at 202 MHz in the suitable deuterated solvent.

### Electrochemistry

Cyclic voltammograms were recorded with a BioLogic SAS potentiostat. All electrochemical measurements were performed in a standard three-compartment electrochemical cell under an argon atmosphere using tetra-*n*-butylammonium hexafluorophosphate (TBAPF<sub>6</sub>, Aldrich) as supporting electrolyte (0.01 M) in dry CH<sub>3</sub>CN. Silver/silver nitrate (ANE2) in CH<sub>3</sub>CN (Ag/AgNO<sub>3</sub> 10 mM in CH<sub>3</sub>CN) and a platinum coil were used as reference and counter electrodes, respectively). 5 mm diameter vitreous carbon electrodes were used as working electrodes.

## Absorption and emission spectroscopies

Absorption spectra were recorded on a Cary 300 UV–visible spectrophotometer (Varian) and emission spectra (in solution at room temperature and in rigid matrix at 77 K) were recorded on a Horiba Fluoromax 4. Quartz cuvettes with 1 cm optical path were used. Lifetimes were measured using LP900 spectrometer with a Flashlamp pumped by Q-switched Nd:Yag laser operating at 355 nm and with a photomultiplier (PMT) detector, or with a picosecond laser diode operating at 410 nm and using a time-correlated single photon counting detection (TCSPC, PicoHarp 300).

Emission quantum yields in solution were determined in deaerated CH<sub>2</sub>Cl<sub>2</sub> or CH<sub>3</sub>CN solutions at room temperature, and calculated with the following equation.

$$QY_s = QY_{ref} \times \frac{I_s}{I_{ref}} \times \frac{A_{ref}}{A_s} \times \left( \frac{\eta_s}{\eta_{ref}} \right)^2$$

**QY<sub>s</sub>** corresponds to the quantum yield of the sample to be analysed, **QY<sub>ref</sub>** is the quantum yield of the reference compound (in this work is Ir(ppy)<sub>3</sub> in deaerated CH<sub>2</sub>Cl<sub>2</sub> solution with value of 0.46), **I** corresponds to the intensity of the emission (area of the spectrum), **A** is the absorption at the excitation wavelength, **η** corresponds to the refractive index of the solvents.

Solid-state emission spectra were obtained on a Horiba-Jobin-Yvon Fluorolog-3<sup>®</sup> spectrofluorimeter, equipped with a three-slit double-grating excitation and emission monochromator with dispersions of 2.1 nm.mm<sup>-1</sup> (1200 grooves mm<sup>-1</sup> and a R928 photodetector). Samples were introduced into an open capillary quartz tube, which was placed into the cavity of a GMP G8 integration sphere.

The two properties that provide information about the excited state beside the shape of the emission are the **QY (Φ)** and lifetime (τ), represented the following equations.

$$\Phi = \frac{k_r}{k_r + k_{nr}} \quad \tau = \frac{1}{k_r + k_{nr}}$$

The radiative (*k<sub>r</sub>*) and non radiative (*k<sub>nr</sub>*) constants can be obtained from combination and rearrangement of the **QY (Φ)** and τ equations to give equations:

$$k_r = \frac{\Phi}{\tau} \quad k_{nr} = \frac{1}{\tau} - k_r$$

## Crystal Structures Determinations and Refinements.

Single crystals of compounds **IrL<sup>6</sup>pic**, **IrL<sup>9</sup>pic** and **IrL<sup>10</sup>pic**, were picked up from the mother liquor, coated with a parafin mixture, collected with nylon loops and mounted on goniometer heads. Measurements were made at 200 K on a Enraf-Nonius 4 circles kappa goniometer equipped with an Incoatec high brilliance microsource with Montel optics monochromated Mo-

K $\alpha$  radiation ( $\lambda = 0.71073 \text{ \AA}$ ). The detector was a Bruker APEXII and an Oxford Cryosystem cryostream cooler was used. The crystal data and details of the data collections are given in Table S 1. The data were integrated and corrected for Lorentz and polarization effects using Eval14<sup>i</sup>, corrected for absorption using SADABS<sup>ii</sup> and finally merged using Xprep<sup>iii</sup>. Crystallographic structures were solved using direct methods implemented by Superflip.<sup>iv</sup> Refinement was performed using ShelXL-2013<sup>v</sup> run under Olex2<sup>vi</sup>. C, Cl, F, Ir, N, O, and Br atoms were refined anisotropically by the full matrix least-squares method on F<sup>2</sup>. H atoms were set geometrically.

Supplementary data is available on request from the CCDC, 12 Union Road, Cambridge CB2 1EZ, UK, quoting the deposition number CCDC-2246643, -2246644 and -2246645 for **IrL<sup>6</sup>pic**, **IrL<sup>9</sup>pic** and **IrL<sup>10</sup>pic** respectively. These data can be obtained free of charge at [www.ccdc.cam.ac.uk/conts/retrieving.html](http://www.ccdc.cam.ac.uk/conts/retrieving.html) or fax: +44-1223/336-033; E-mail: [deposit@ccdc.cam.ac.uk](mailto:deposit@ccdc.cam.ac.uk)].

Table S 1 : Crystal data and structure refinement.

Compound	<b>IrL<sup>6</sup>pic</b> / CCDC 2246643	<b>IrL<sup>9</sup>pic</b> / CCDC 2246644	<b>IrL<sup>10</sup>pic</b> / CCDC 2246645
Formula	C <sub>42</sub> H <sub>36</sub> F <sub>6</sub> IrN <sub>5</sub> O <sub>4</sub>	C <sub>40</sub> H <sub>32</sub> F <sub>6</sub> IrN <sub>5</sub> O <sub>2</sub> · (CH <sub>2</sub> Cl <sub>2</sub> )	C <sub>42</sub> H <sub>36</sub> F <sub>6</sub> IrN <sub>5</sub> O <sub>4</sub> · 1.54(CH <sub>2</sub> Cl <sub>2</sub> )
$F_w$	980.96	1005.83	1112.05
$T$ [K]	200	200	200
Morphology	needle	block	block
Color	yellow	yellow	orange
Crystal size [mm]	0.10 x 0.17 x 0.40	0.24 x 0.24 x 0.43	0.14 x 0.28 x 0.32
Crystal system	triclinic	monoclinic	monoclinic
Space group	P -1	P 2 <sub>1</sub> /n	P 2 <sub>1</sub> /n
$a$ [Å]	11.392(2)	16.113(3)	15.316(3)
$b$ [Å]	11.878(2)	15.027(3)	15.544(3)
$c$ [Å]	14.728(3)	17.152(3)	19.688(4)
$\alpha$ [°]	85.63(3)	90	90
$\beta$ [°]	88.84(3)	102.83(3)	101.07(3)
$\gamma$ [°]	78.87(3)	90	90
Unit-cell volume [Å <sup>3</sup> ]	1949.7(7)	4049.4(15)	4600.0(17)
$Z$	2	4	4

<sup>i</sup> Duisenberg, A. J. M., Kroon-Batenburg, L. M. J. & Schreurs, A. M. M. (2003). *J. Appl. Cryst.* **36**, 220-229.

<sup>ii</sup> Bruker (2004). *SADABS*. Bruker AXS Inc., Madison, Wisconsin, USA.

<sup>iii</sup> Bruker (2005). *XPREP*. Bruker AXS Inc., Madison, Wisconsin, USA.

<sup>iv</sup> Palatinus, L. & Chapuis, G. (2007). *J. Appl. Cryst.* **40**, 786-790.

<sup>v</sup> Sheldrick, G. M. (2015). *Acta Cryst.* **C71**, 3-8.

<sup>vi</sup> Dolomanov, O.V., Bourhis, L.J., Gildea, R.J, Howard, J.A.K. & Puschmann, H. (2009), *J. Appl. Cryst.* **42**, 339-341

$D_x$ [ $\text{g}\cdot\text{cm}^{-3}$ ]	1.671	1.650	1.606
$\mu$ [ $\text{mm}^{-1}$ ]	3.502	3.499	3.153
Radiation [ $\text{\AA}$ ]	MoK $\alpha$ ( $\lambda = 0.71073$ )	MoK $\alpha$ ( $\lambda = 0.71073$ )	MoK $\alpha$ ( $\lambda = 0.71073$ )
$\Theta$ range for data collection/ $^\circ$	2.296 to 30.000	2.388 to 27.500	2.283 to 27.500
Index ranges	$-16 \leq h \leq 16, -16 \leq k \leq 16,$ $-20 \leq l \leq 20$	$-20 \leq h \leq 20, -19 \leq k \leq 19,$ $-22 \leq l \leq 22$	$-19 \leq h \leq 19, -20 \leq k \leq 19,$ $-23 \leq l \leq 25$
Total reflections	45506	50004	62266
Unique reflections	11358	9220	10403
Used reflections ( $I > 2\sigma(I)$ )	10433	7389	8712
Refined parameters	605	584	650
$R_{int}$ .	0.0305	0.0338	0.0615
$R^a$	0.0197	0.0324	0.0366
$R(w)^a$	0.0442	0.0729	0.0899
Goodness of fit $S$	1.167	1.155	1.132
$\Delta\rho_{min}/\Delta\rho_{max}$ ( $\text{e}\cdot\text{\AA}^{-3}$ )	-0.886/1.247	-2.215/2.185	-1.772/1.950

<sup>a</sup> Refinement based on  $F^2$  where

$w = 1/[\sigma^2(\text{Fo}^2) + (0.0157\text{P})^2 + 1.1400\text{P}]$  where  $\text{P} = (\text{Fo}^2 + 2\text{Fc}^2)/3$  for CCDC-2246643

$w = 1/[\sigma^2(\text{Fo}^2) + (0.0247\text{P})^2 + 17.9483\text{P}]$  where  $\text{P} = (\text{Fo}^2 + 2\text{Fc}^2)/3$  for CCDC-2246644

$w = 1/[\sigma^2(\text{Fo}^2) + (0.0411\text{P})^2 + 12.1613\text{P}]$  where  $\text{P} = (\text{Fo}^2 + 2\text{Fc}^2)/3$  for CCDC-2246645

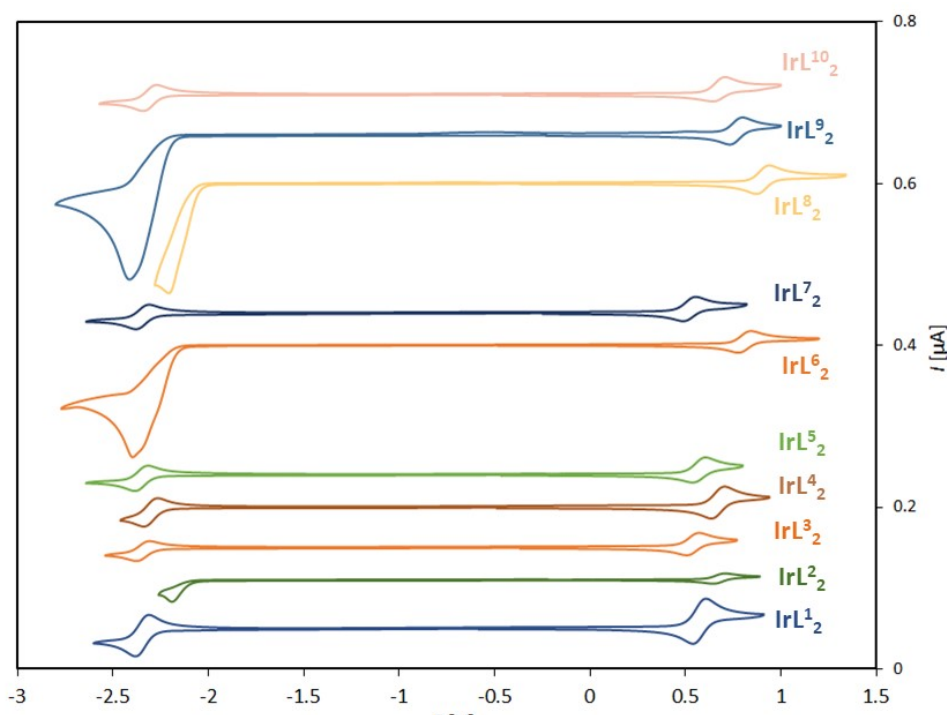


Figure S 1 :Cyclic voltammograms of the complexes ( $10^{-3}$  M to the exception of  $\text{IrL}^3_2$  see below) in deaerated  $\text{CH}_3\text{CN}$ , containing 0.1 M of  $n\text{-Bu}_4\text{PF}_6$ , at a scan rate of  $100 \text{ mV}\cdot\text{s}^{-1}$  using vitreous carbon working electrode (5 mm diameter),  $E(\text{V})$  vs  $\text{Ag}/\text{AgNO}_3$  (0.01 M).  $\text{IrL}^3_2$  displays low solubility in the solvent ( $\text{CH}_3\text{CN}$ ) that leads to lower current during the experiment and the CV was performed in a saturated solution.

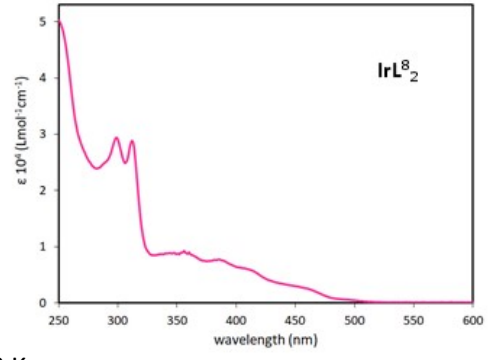
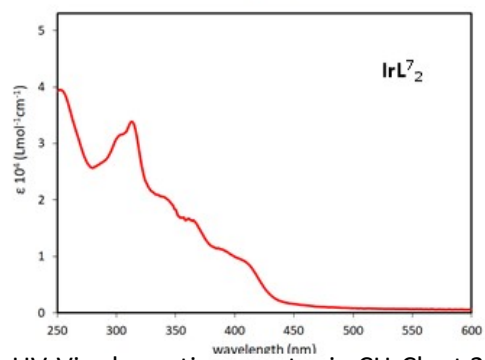
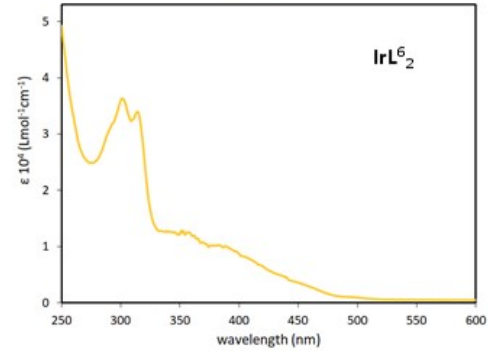
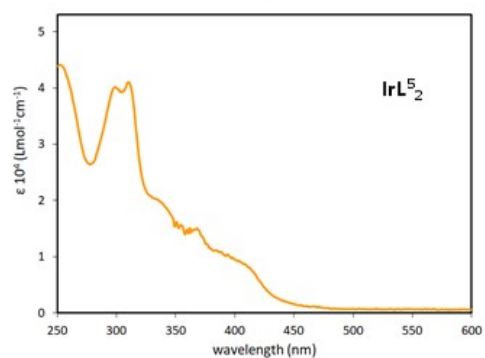
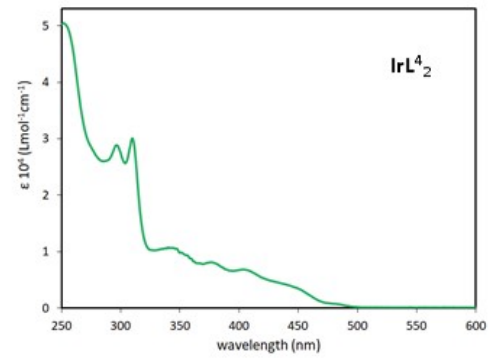
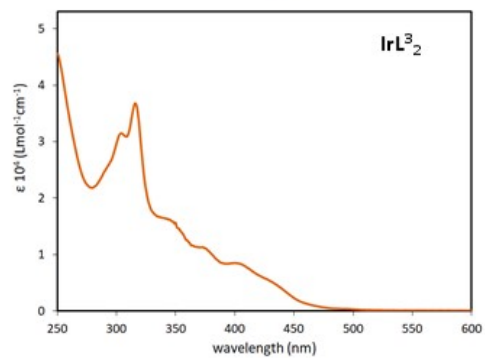
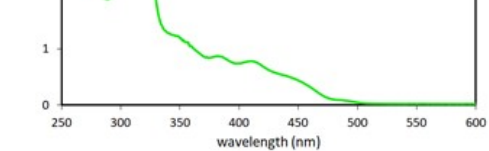
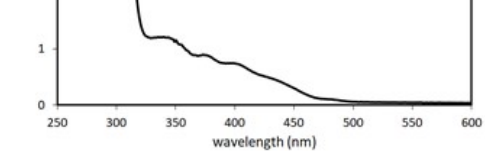
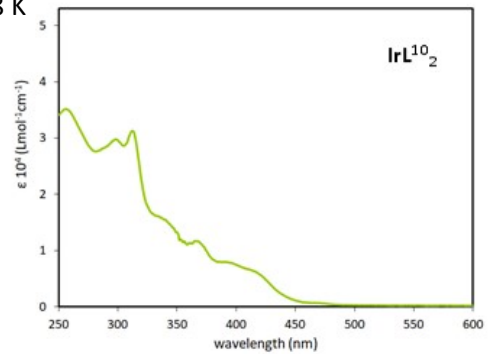
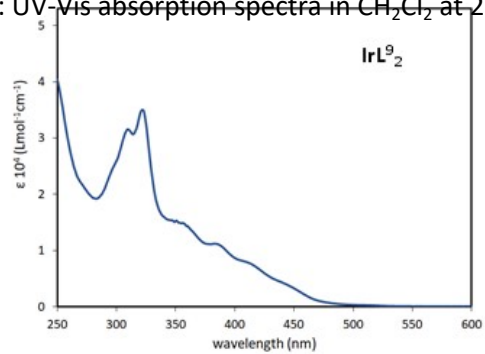


Figure S 2: UV-Vis absorption spectra in CH<sub>2</sub>Cl<sub>2</sub> at 298 K



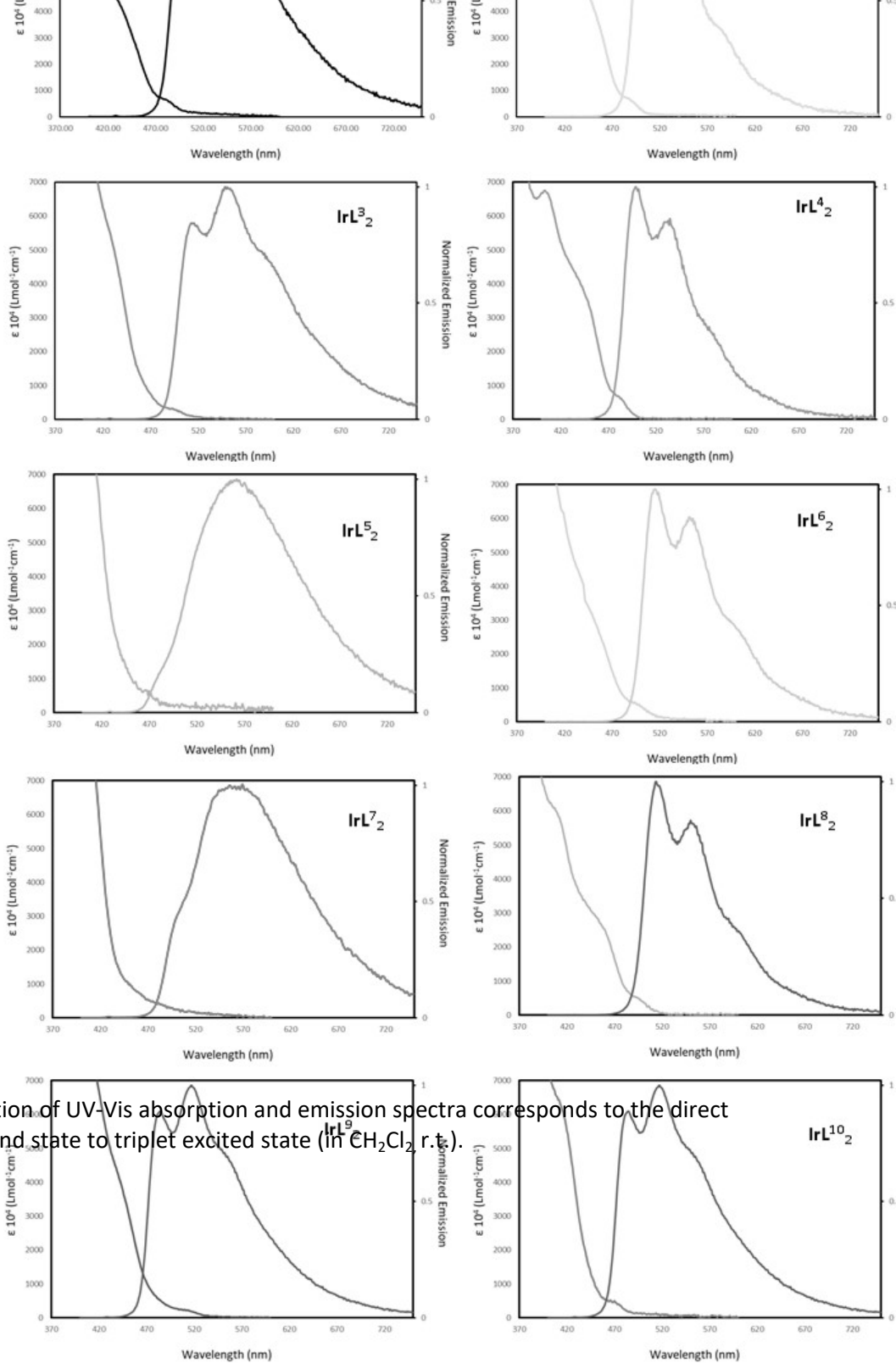


Figure S 3 : The superimposition of UV-Vis absorption and emission spectra corresponds to the direct absorption from singlet ground state to triplet excited state (In CH<sub>2</sub>Cl<sub>2</sub> r.t.).

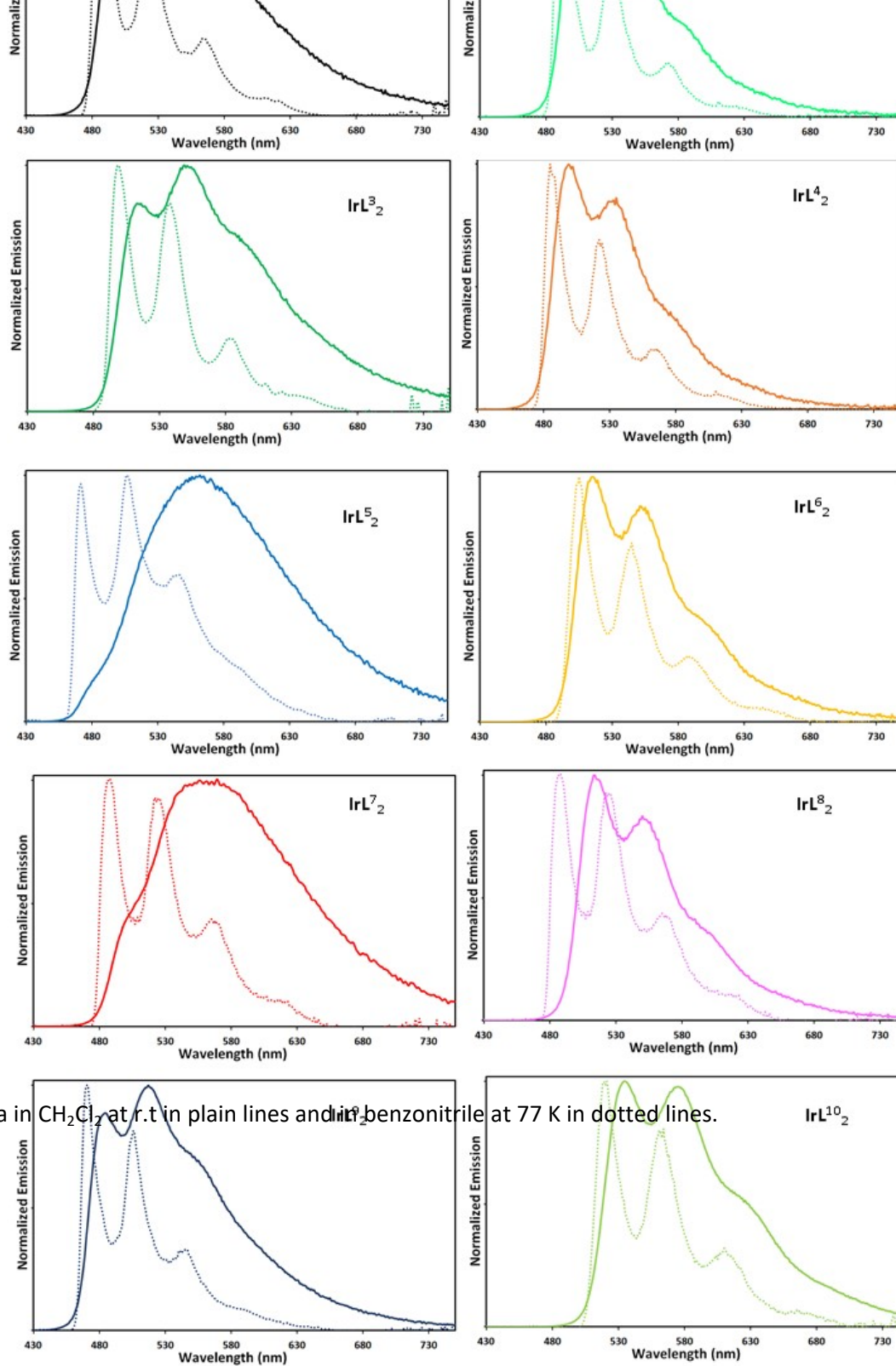


Figure S 4 : Emission spectra in CH<sub>2</sub>Cl<sub>2</sub> at r.t in plain lines and in benzonitrile at 77 K in dotted lines.

# NMR spectra

IrL<sub>2</sub>

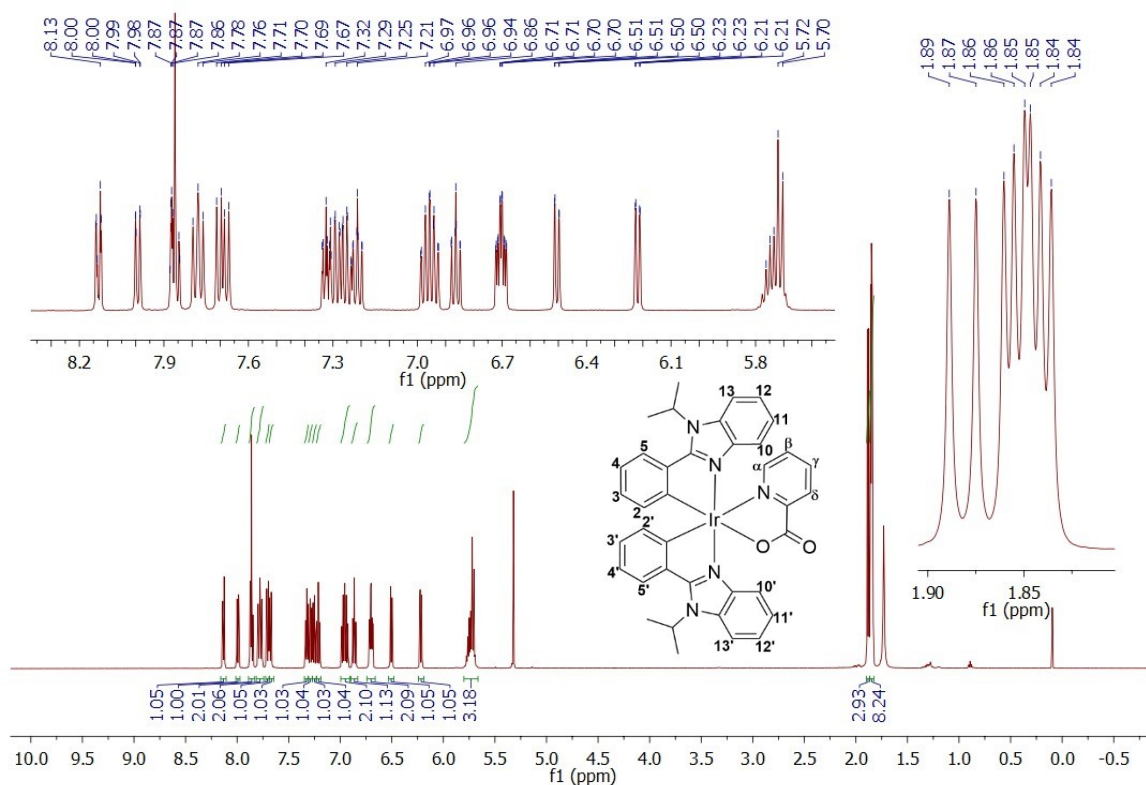


Figure S 5: <sup>1</sup>H NMR spectra (500 MHz, CD<sub>2</sub>Cl<sub>2</sub>)

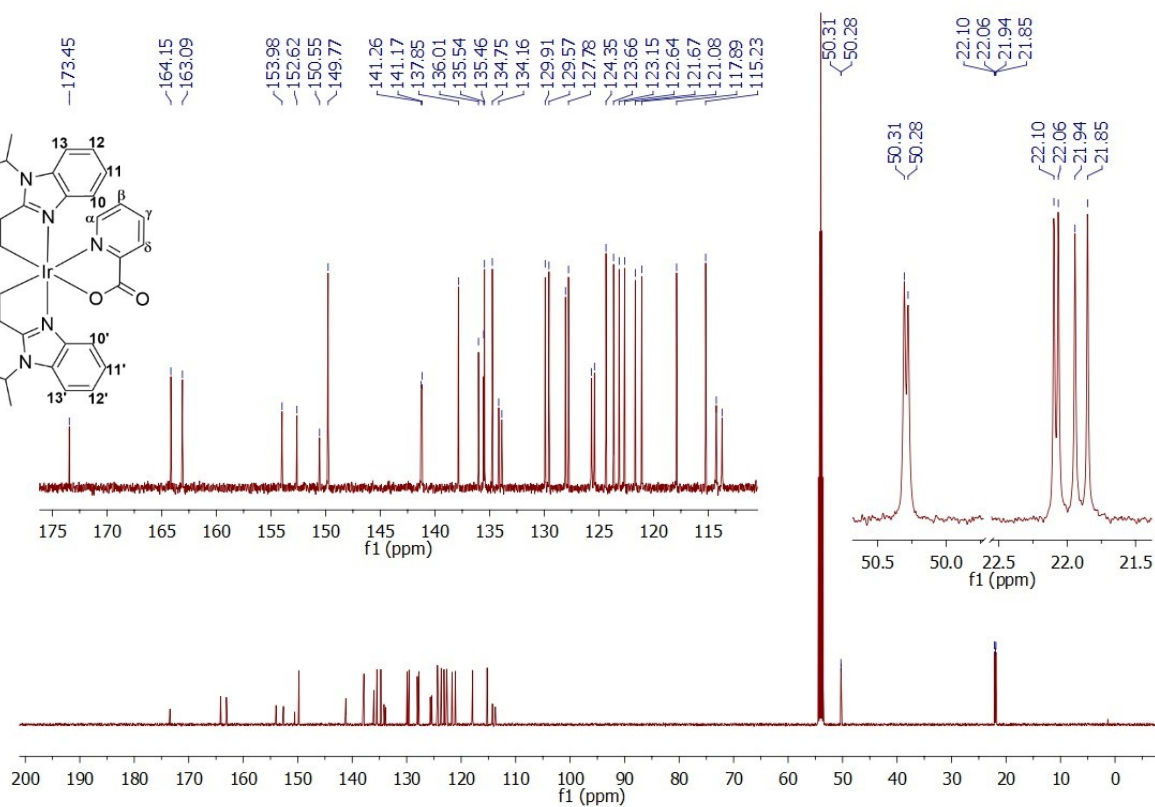


Figure S 6: <sup>13</sup>C NMR spectra (500 MHz, CD<sub>2</sub>Cl<sub>2</sub>)



IrL<sub>2</sub>

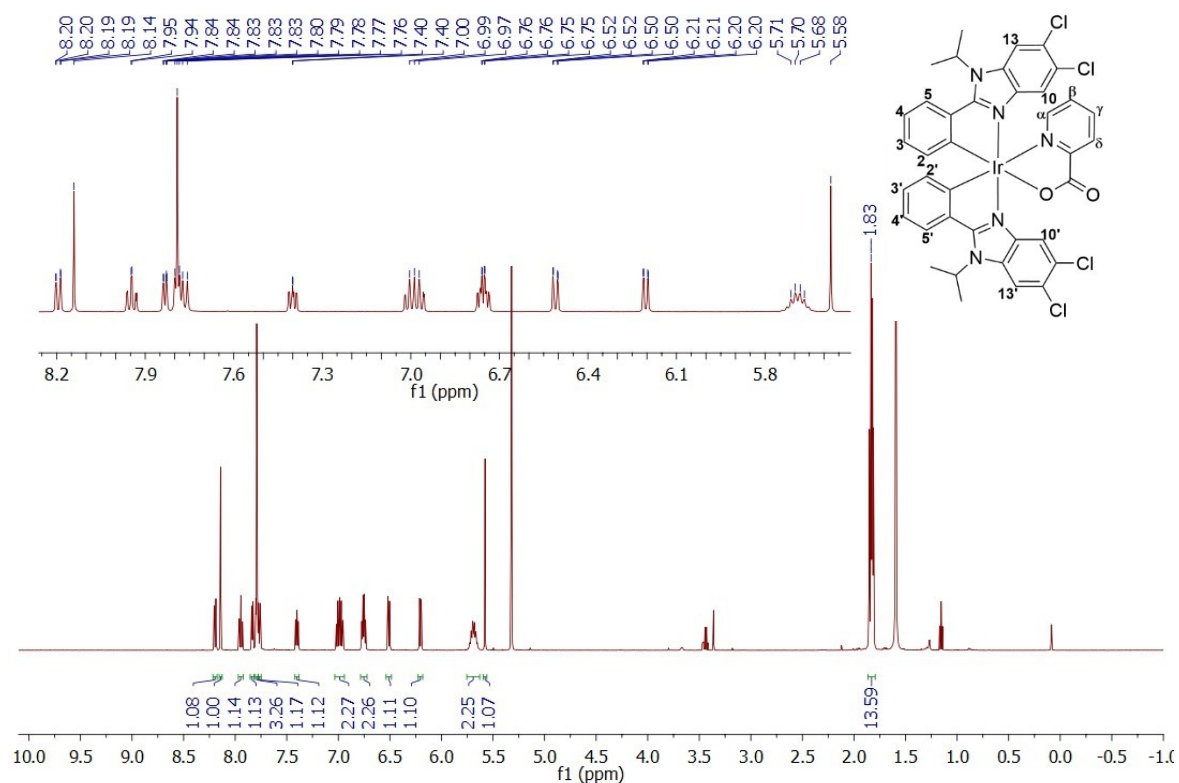


Figure S 7: <sup>1</sup>H NMR spectra (500 MHz, CD<sub>2</sub>Cl<sub>2</sub>)

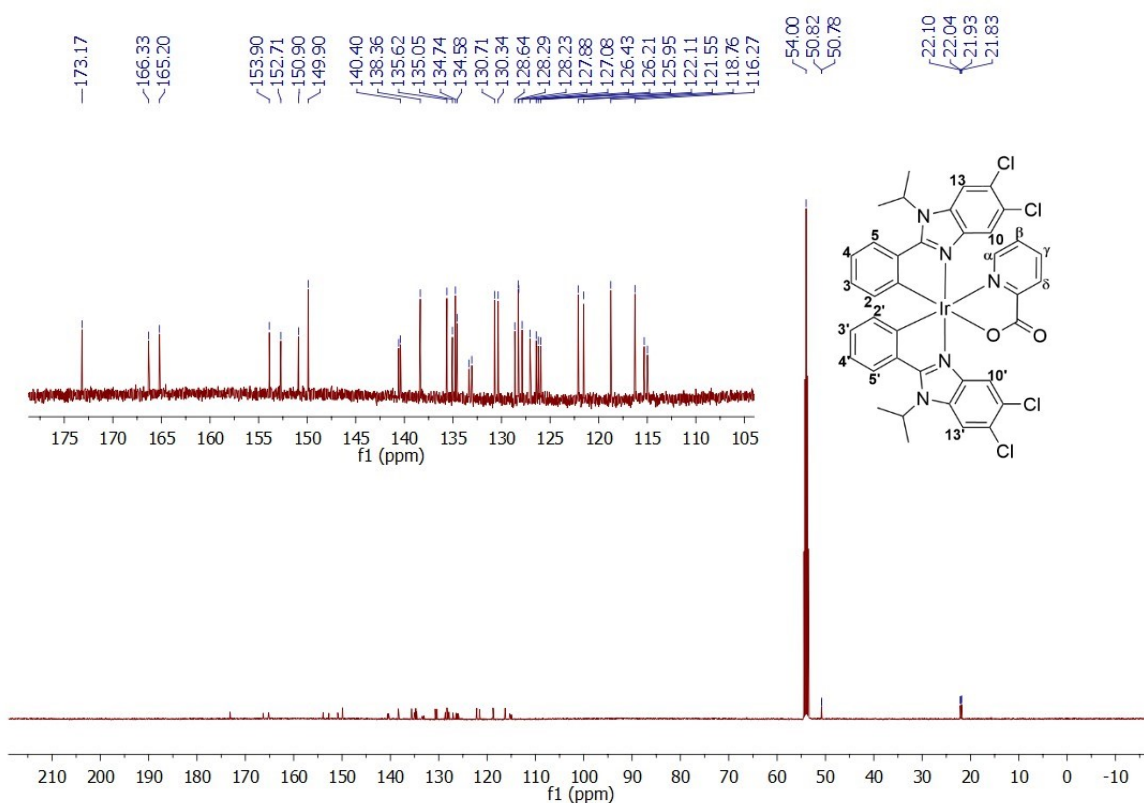


Figure S 8: <sup>13</sup>C NMR spectra (500 MHz, CD<sub>2</sub>Cl<sub>2</sub>)

IrL<sup>3</sup><sub>2</sub>

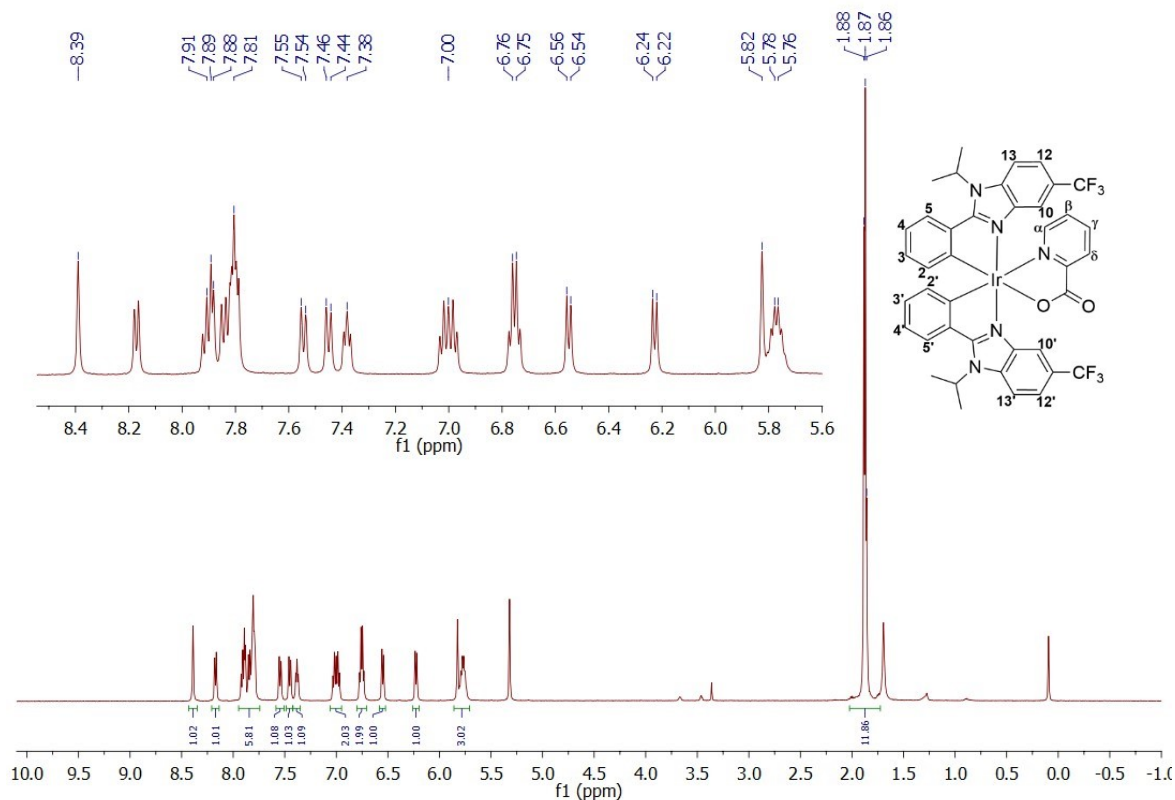


Figure S 9: <sup>1</sup>H NMR spectra (500 MHz, CD<sub>2</sub>Cl<sub>2</sub>)

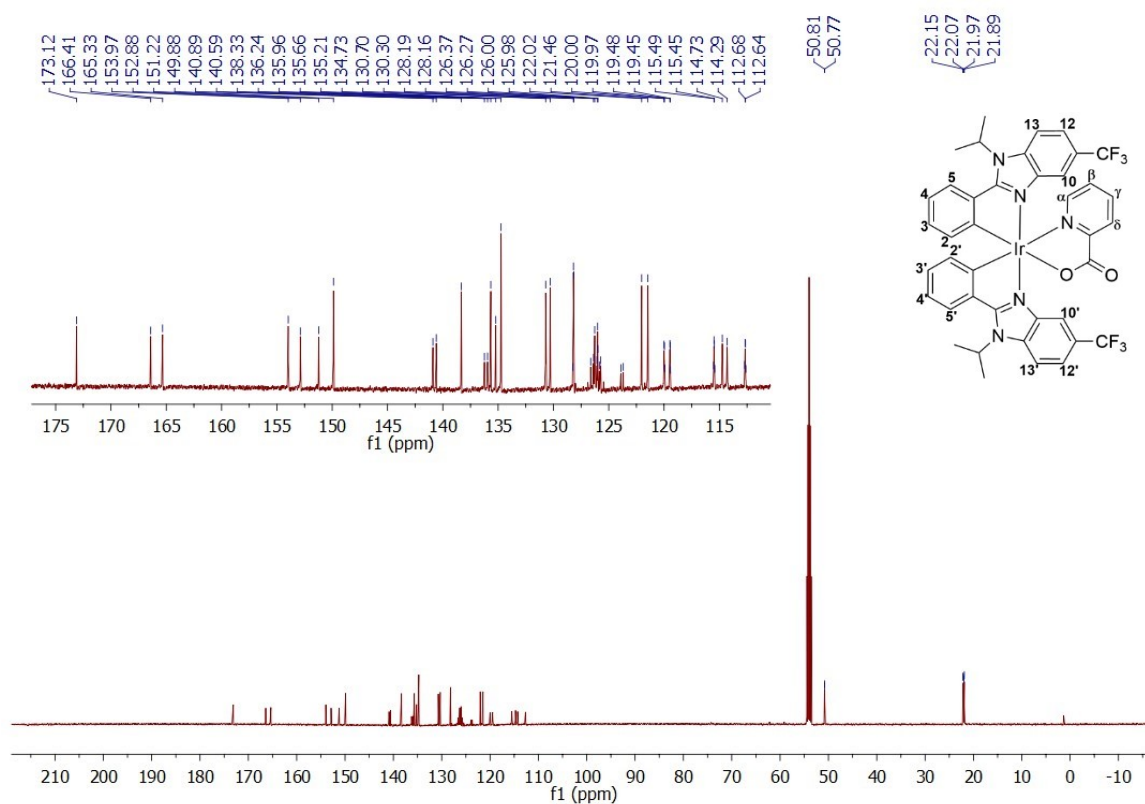


Figure S 10: <sup>13</sup>C NMR spectra (500 MHz, CD<sub>2</sub>Cl<sub>2</sub>)

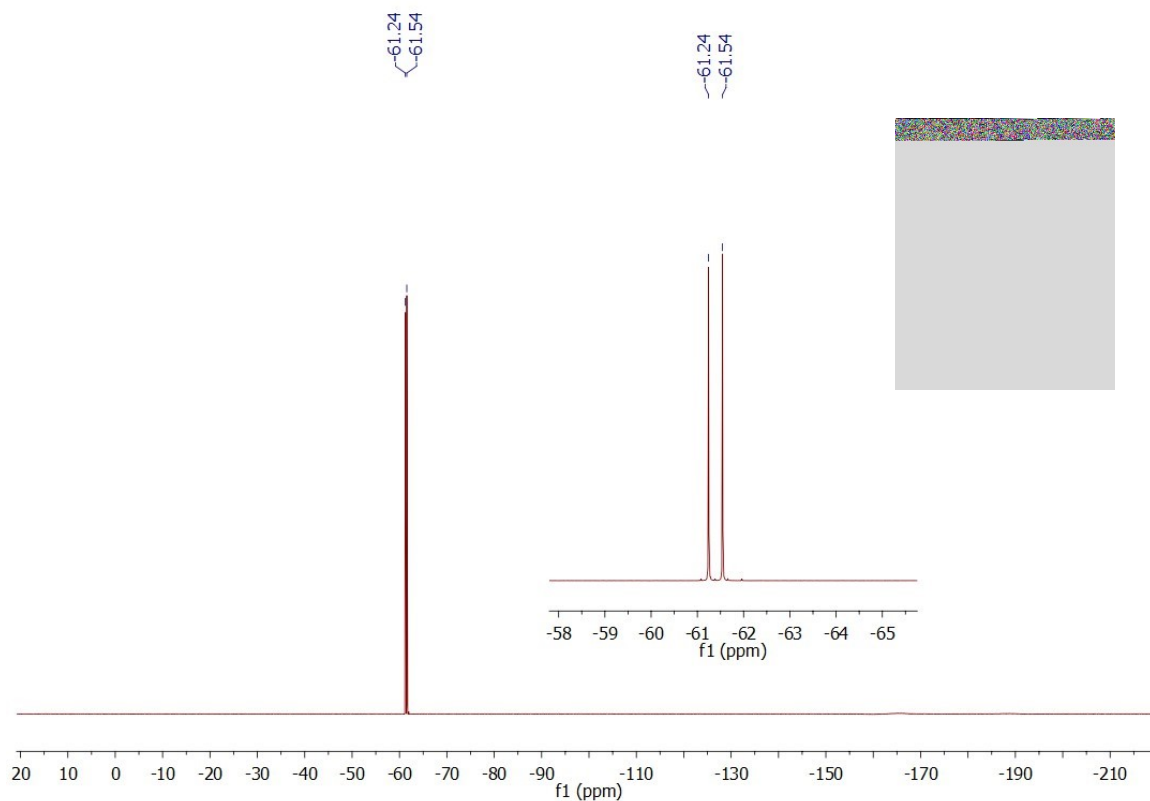


Figure S 11: <sup>19</sup>F NMR spectra (500 MHz, CD<sub>2</sub>Cl<sub>2</sub>)

IrL<sub>2</sub>

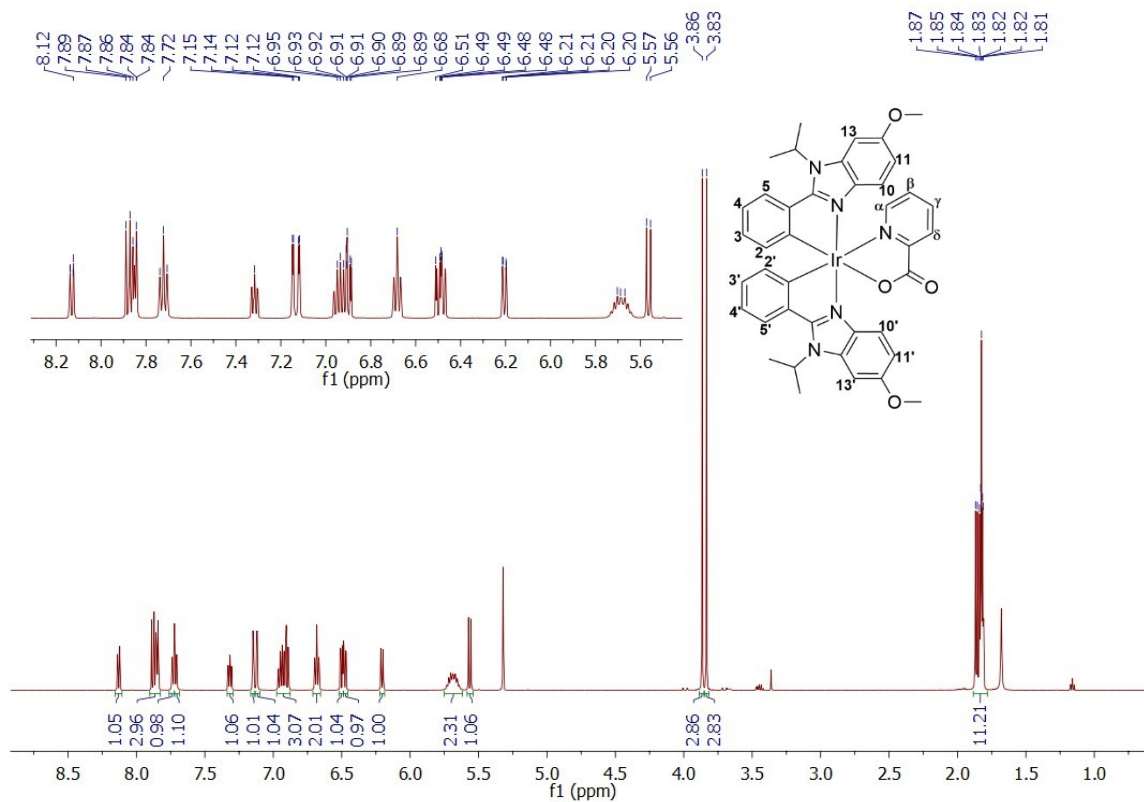


Figure S 12: <sup>1</sup>H NMR spectra (500 MHz, CD<sub>2</sub>Cl<sub>2</sub>)

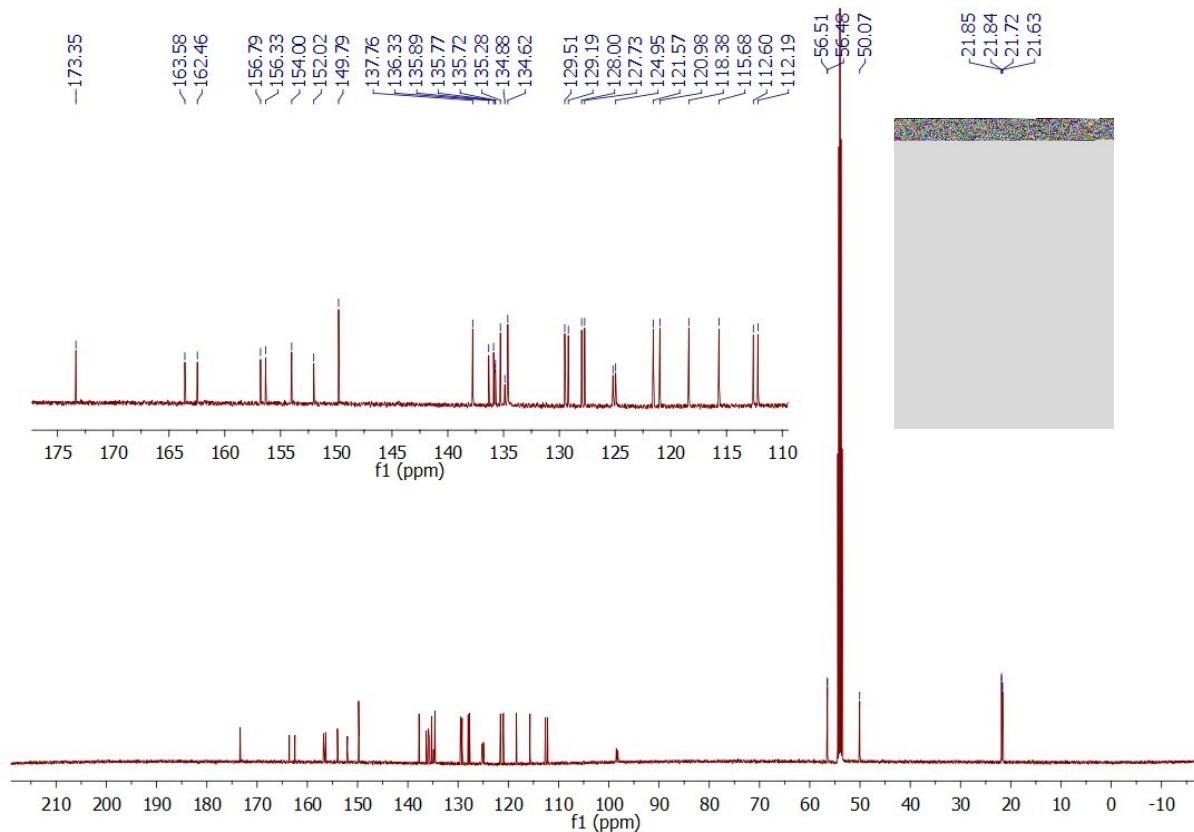


Figure S 13:  $^{13}\text{C}$  NMR spectra (500 MHz,  $\text{CD}_2\text{Cl}_2$ )

$\text{IrL}_5$

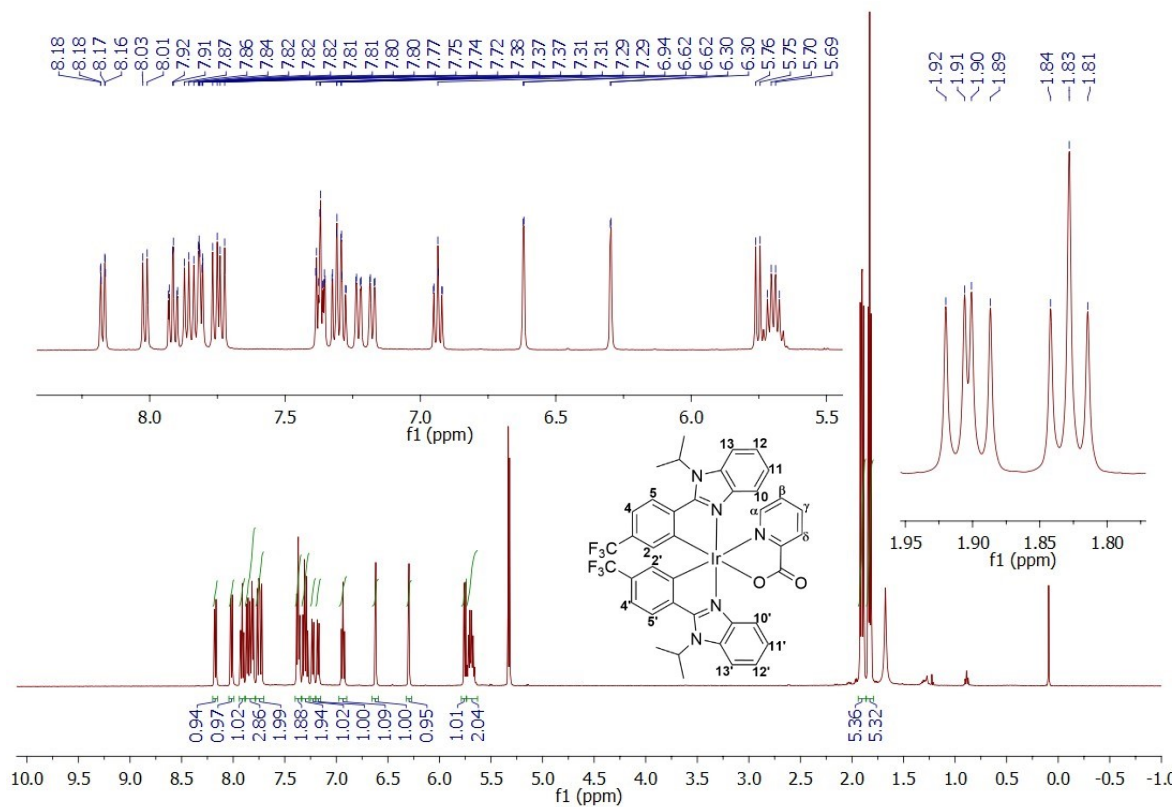


Figure S 14:  $^1\text{H}$  NMR spectra (500 MHz,  $\text{CD}_2\text{Cl}_2$ )

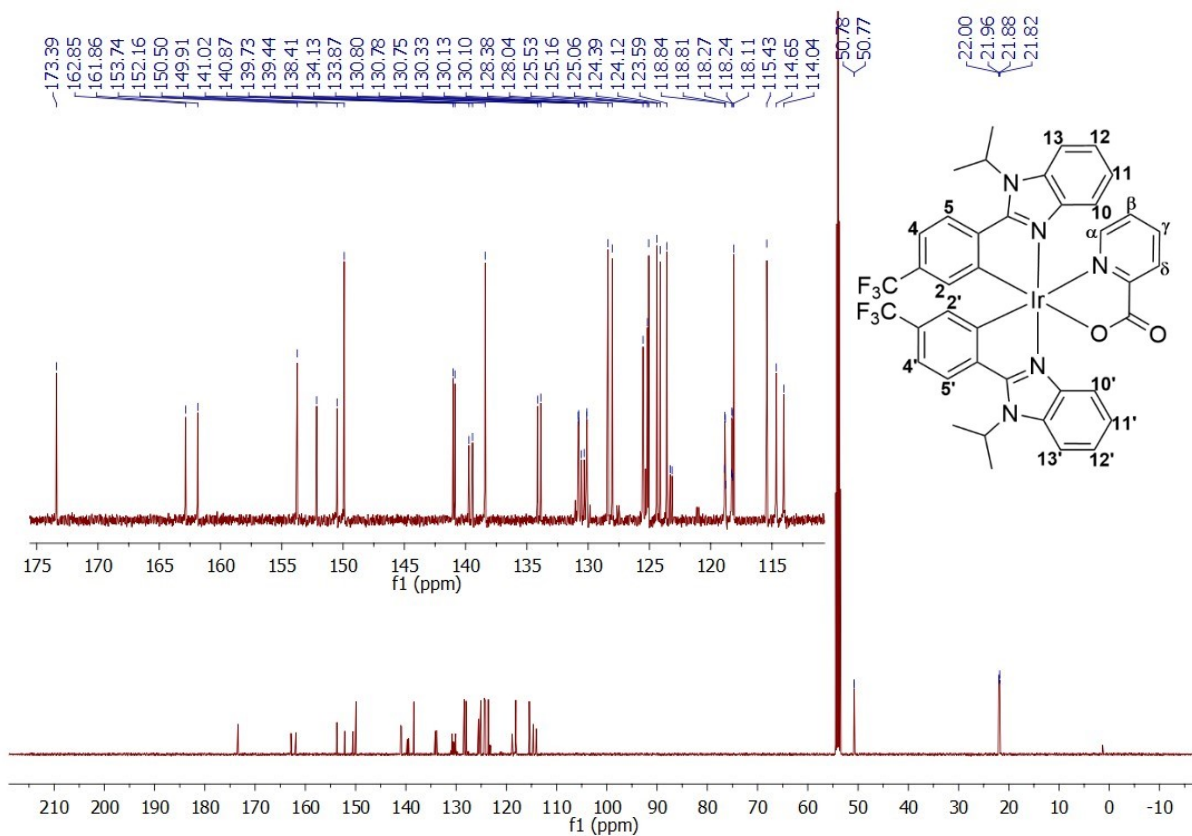


Figure S 15:  $^{13}\text{C}$  NMR spectra (500 MHz,  $\text{CD}_2\text{Cl}_2$ )

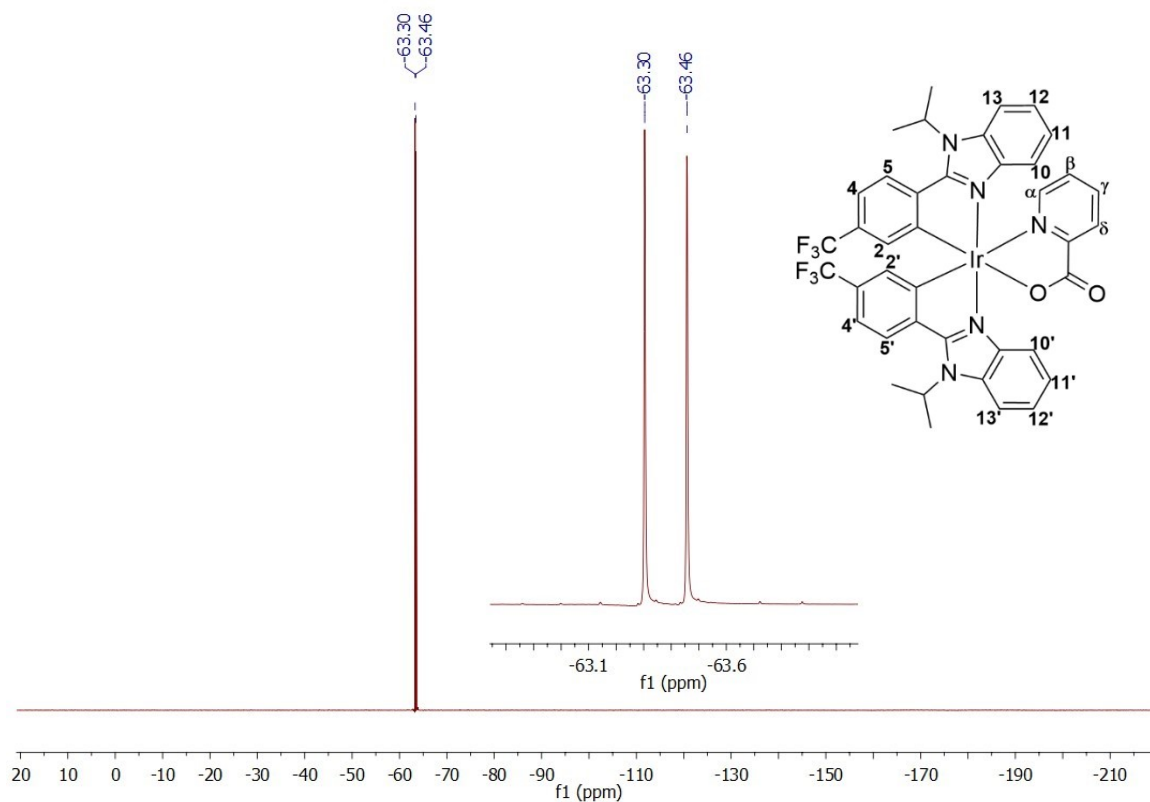


Figure S 16:  $^{19}\text{F}$  NMR spectra (500 MHz,  $\text{CD}_2\text{Cl}_2$ )

IrL<sup>6</sup><sub>2</sub>

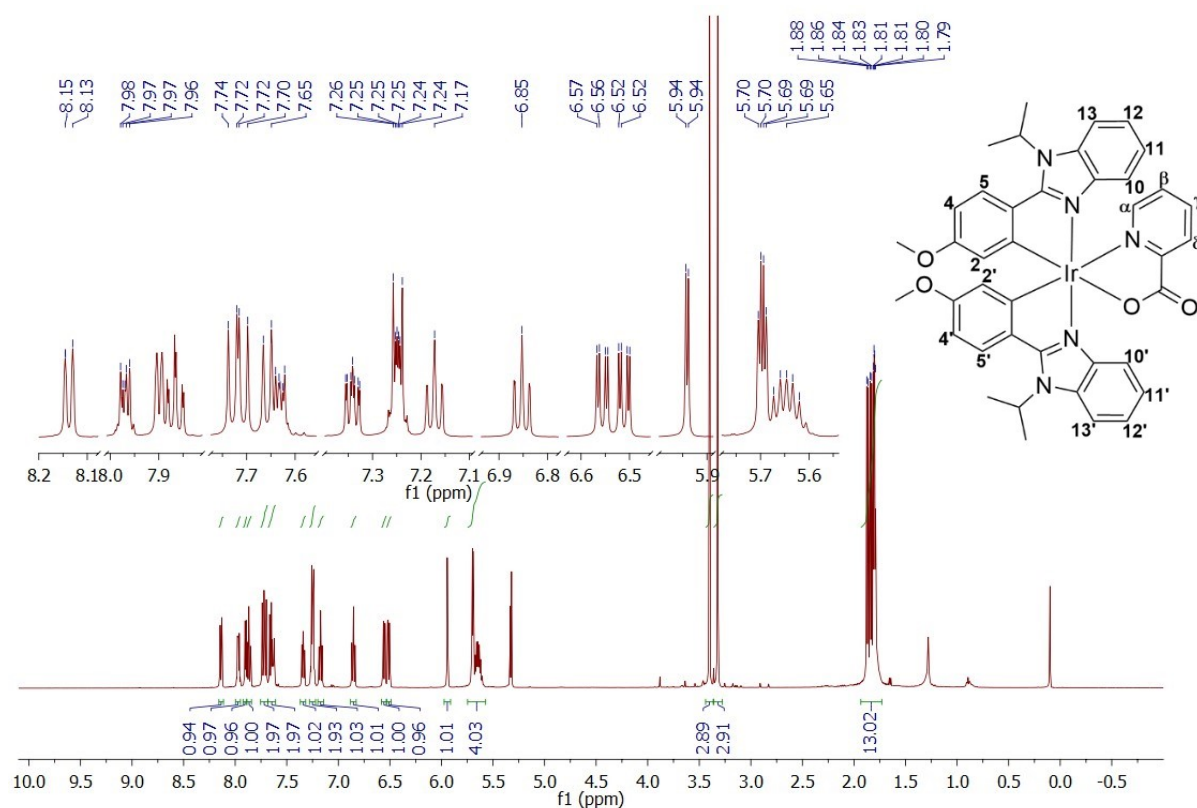


Figure S 17: <sup>1</sup>H NMR spectra (500 MHz, CD<sub>2</sub>Cl<sub>2</sub>)

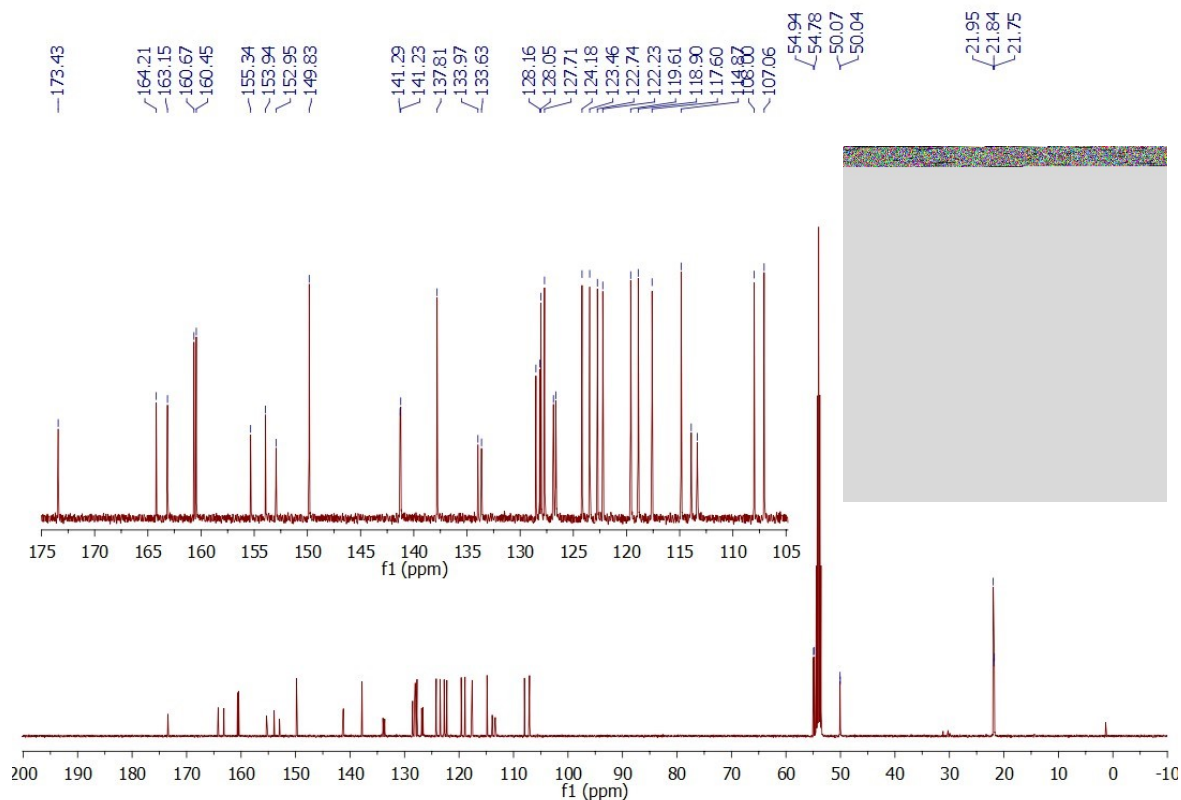


Figure S 18: <sup>13</sup>C NMR spectra (500 MHz, CD<sub>2</sub>Cl<sub>2</sub>)

IrL<sub>2</sub>

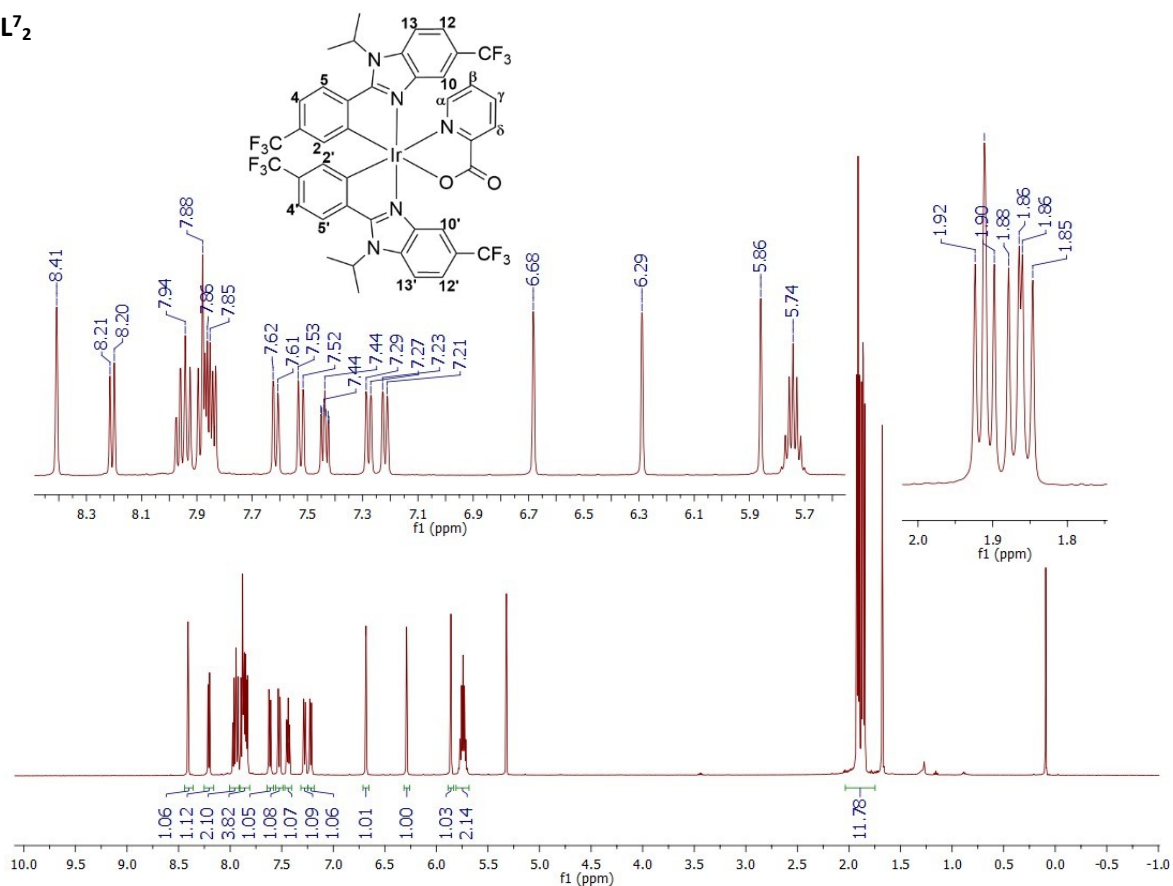


Figure S 19: <sup>1</sup>H NMR spectra (500 MHz, CD<sub>2</sub>Cl<sub>2</sub>)

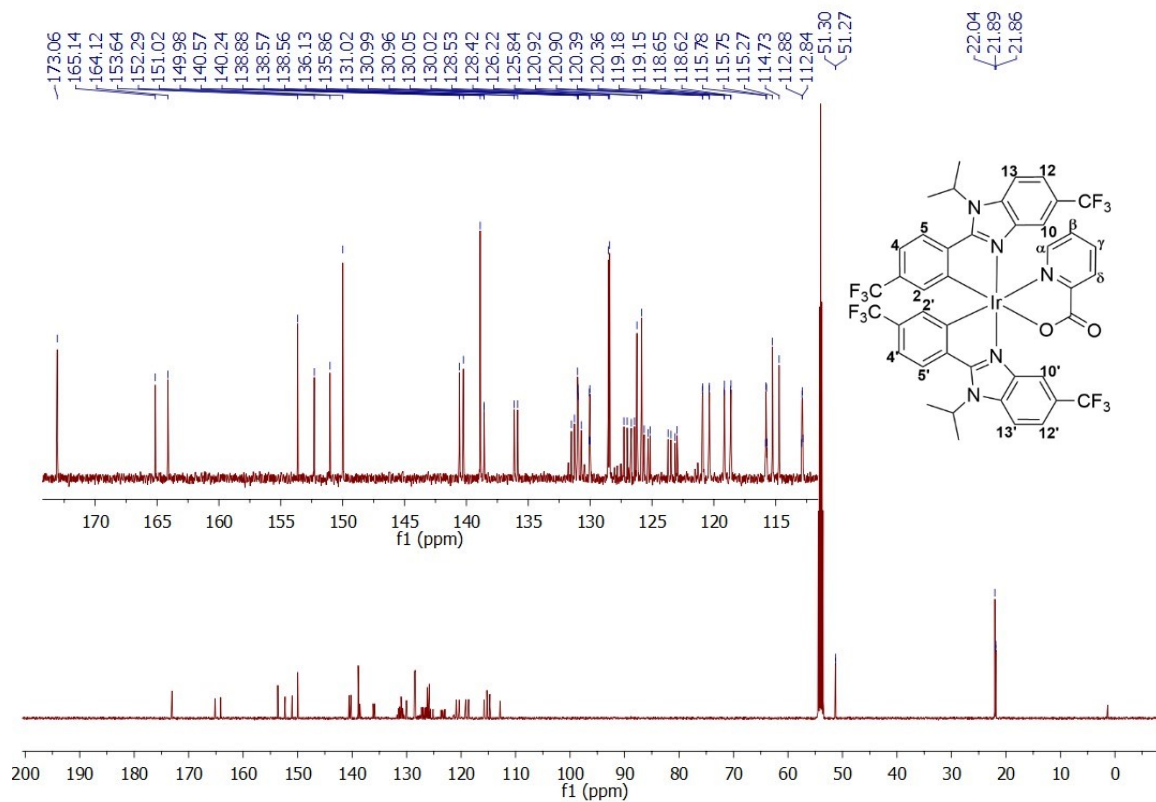


Figure S 20: <sup>13</sup>C NMR spectra (500 MHz, CD<sub>2</sub>Cl<sub>2</sub>)

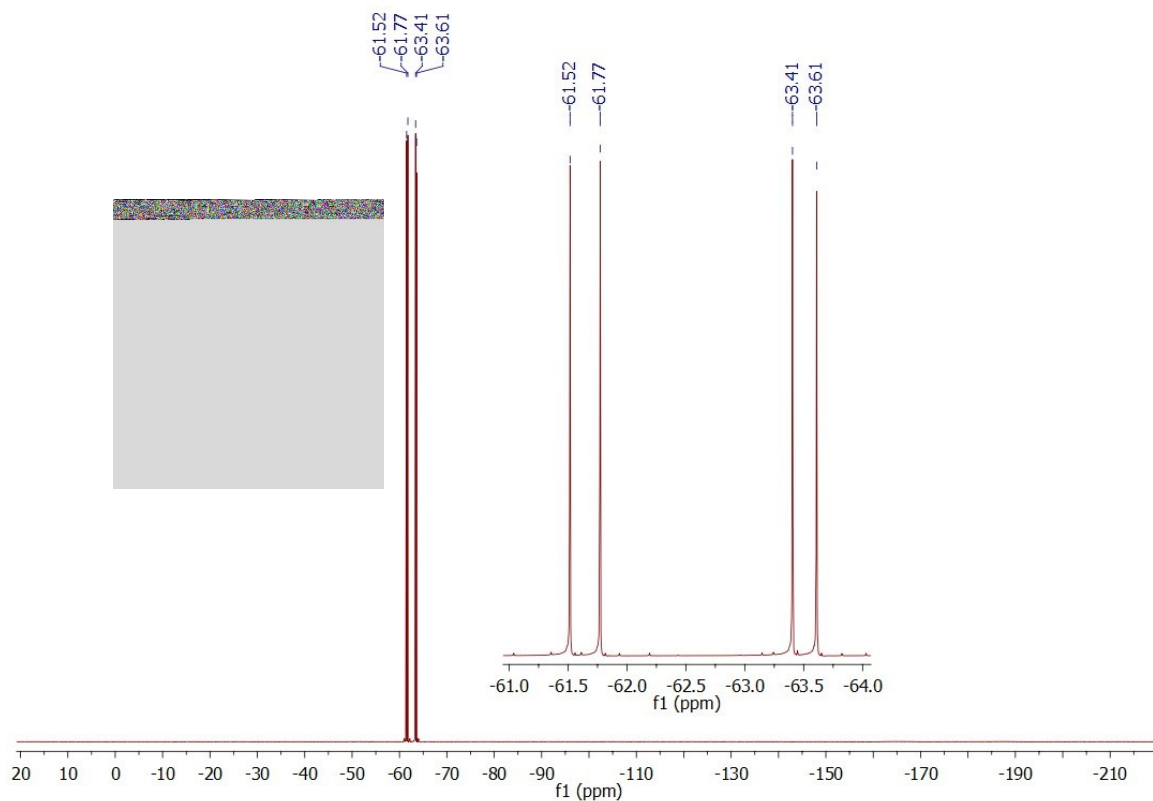


Figure S 21:  $^{19}\text{F}$  NMR spectra (500 MHz,  $\text{CD}_2\text{Cl}_2$ )

$\text{IrL}_2^8$

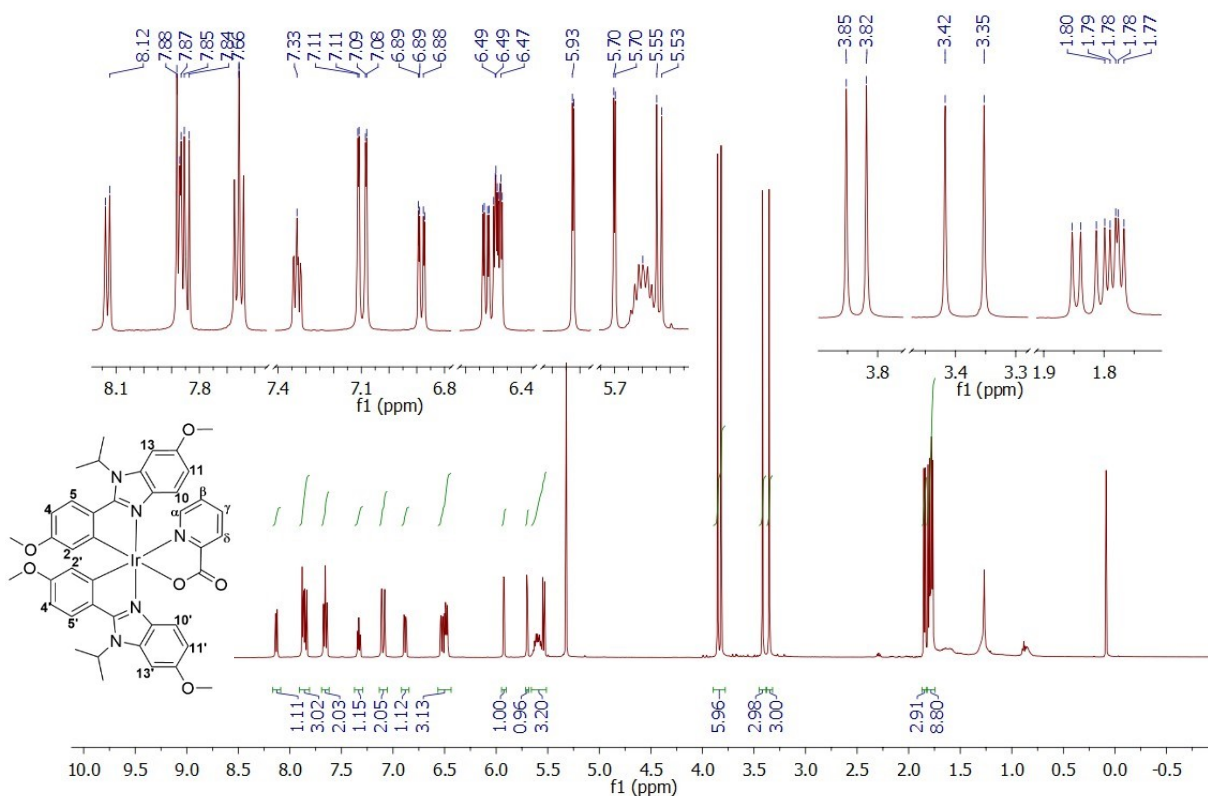


Figure S 22:  $^1\text{H}$  NMR spectra (500 MHz,  $\text{CD}_2\text{Cl}_2$ )



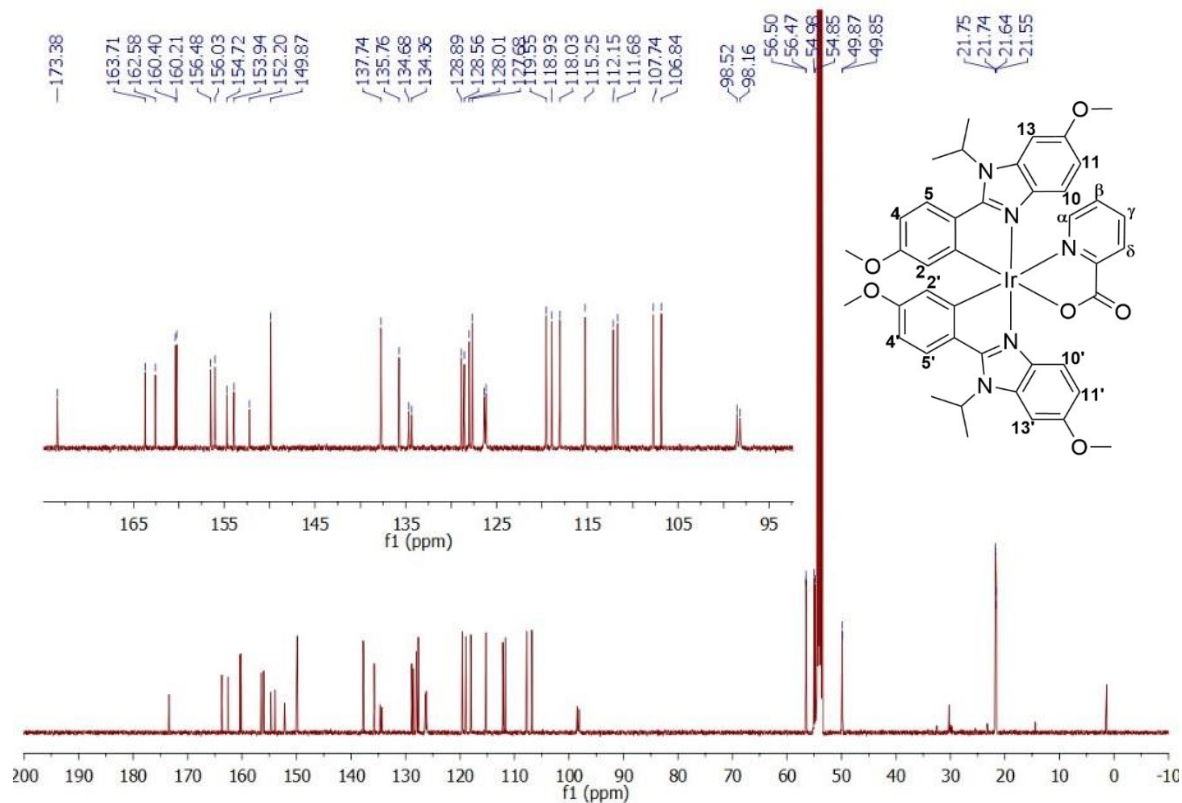


Figure S 23:  $^{13}\text{C}$  NMR spectra (500 MHz,  $\text{CD}_2\text{Cl}_2$ )

$\text{IrL}_2$

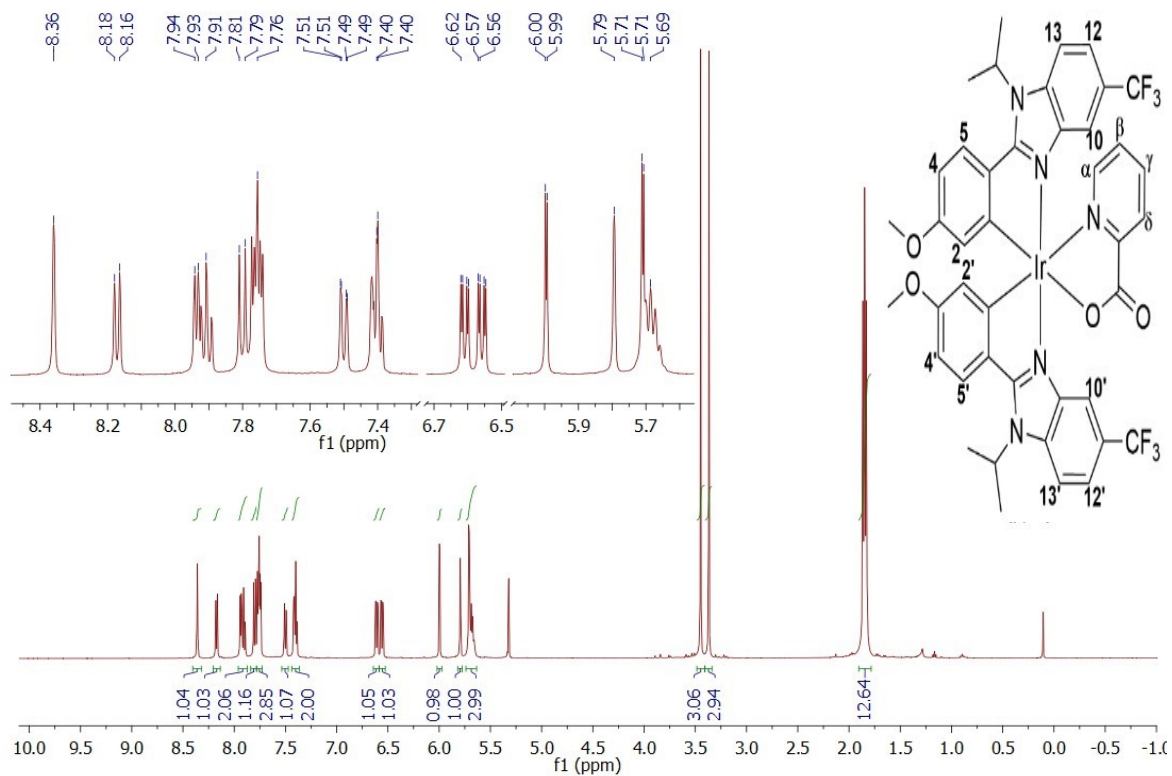


Figure S 24:  $^1\text{H}$  NMR spectra (500 MHz,  $\text{CD}_2\text{Cl}_2$ )

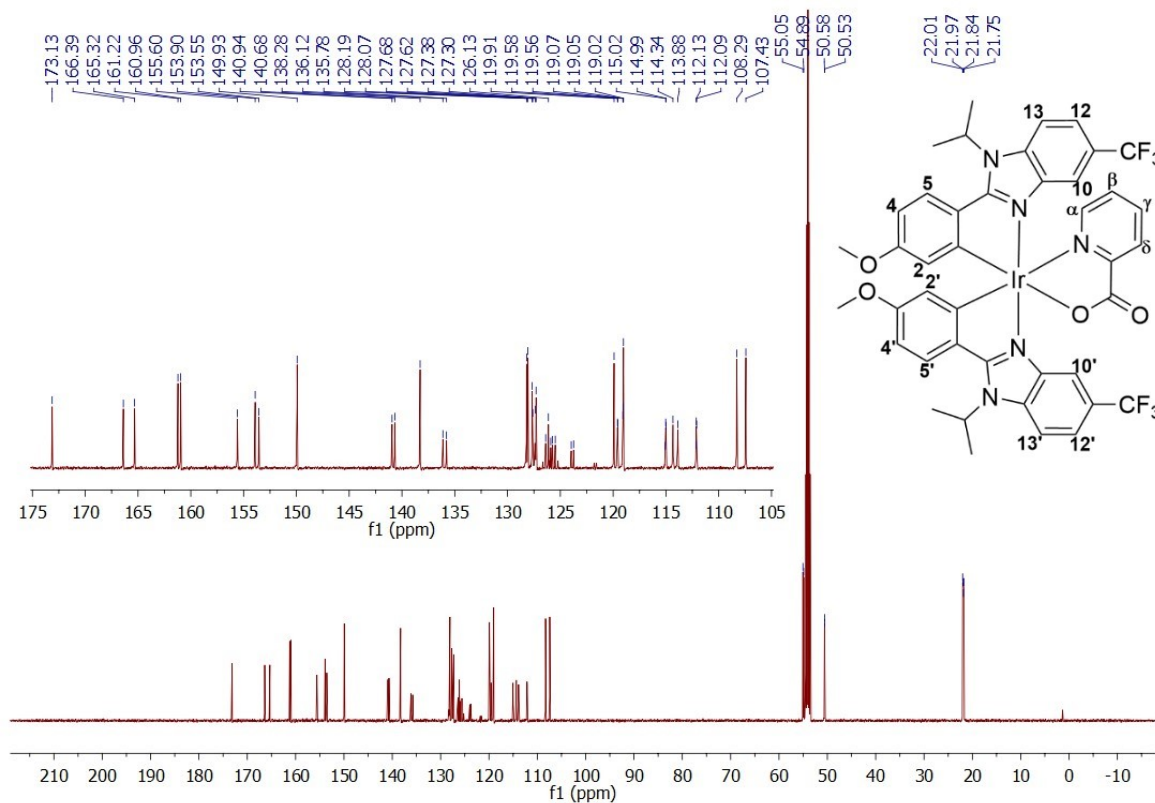


Figure S 25: <sup>13</sup>C NMR spectra (500 MHz, CD<sub>2</sub>Cl<sub>2</sub>)

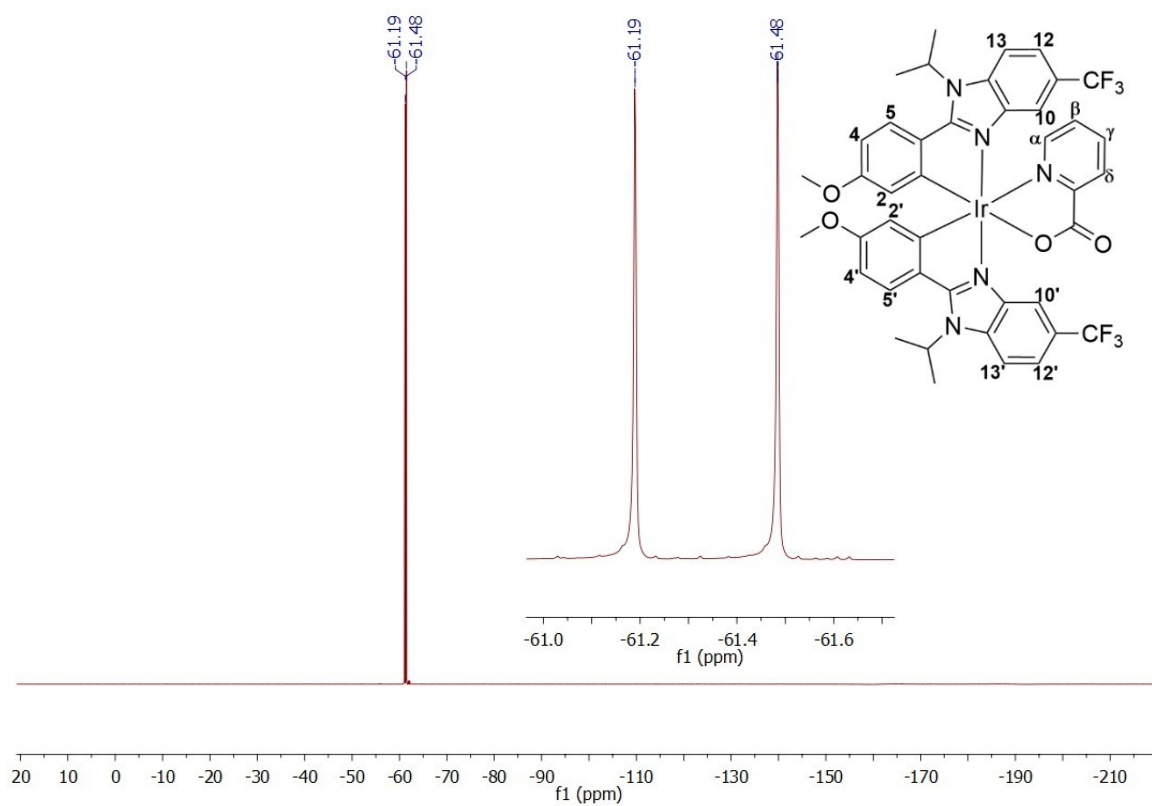


Figure S 26: <sup>19</sup>F NMR spectra (500 MHz, CD<sub>2</sub>Cl<sub>2</sub>)

IrL<sup>10</sup><sub>2</sub>

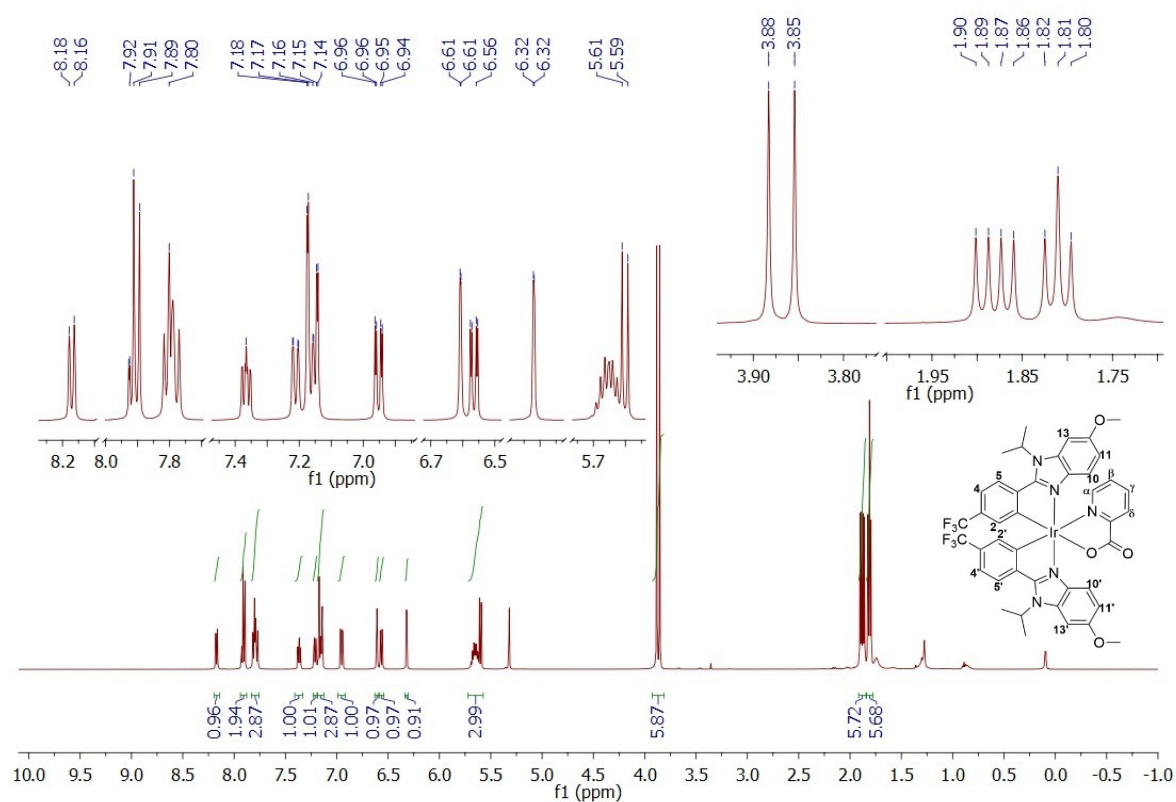


Figure S 27: <sup>1</sup>H NMR spectra (500 MHz, CD<sub>2</sub>Cl<sub>2</sub>)

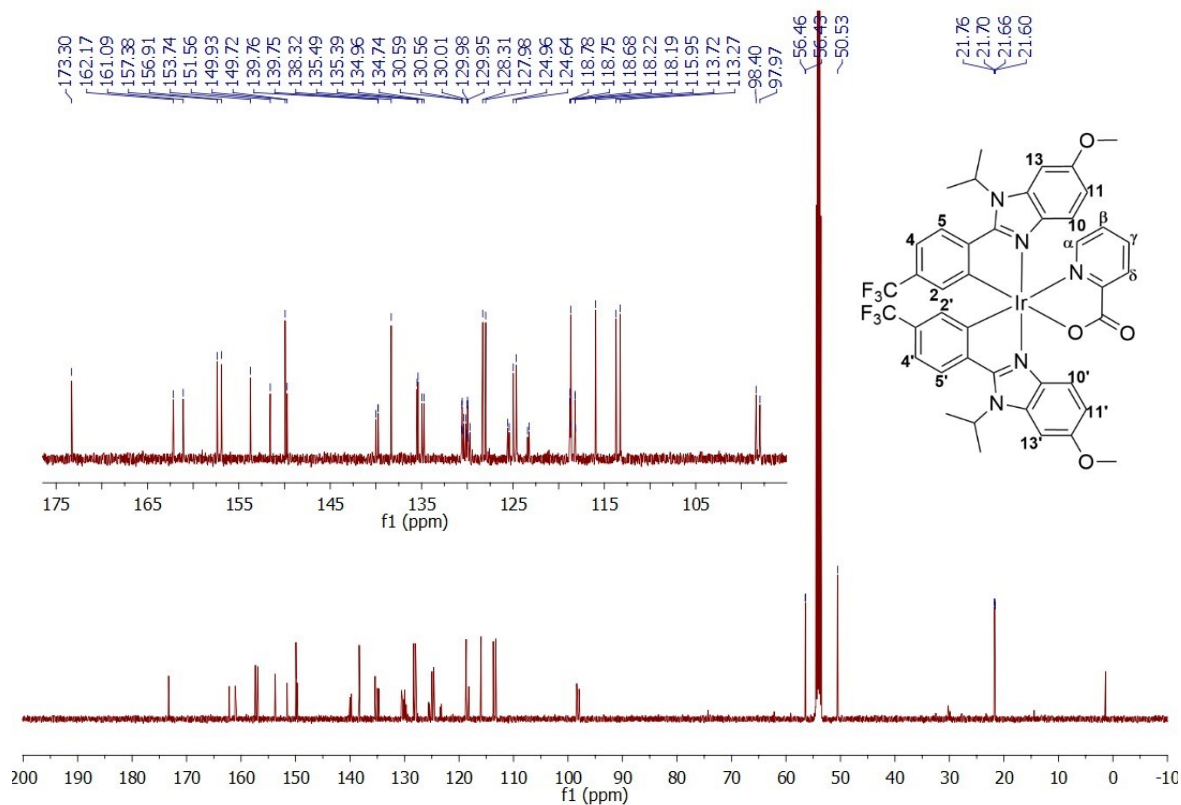


Figure S 28: <sup>13</sup>C NMR spectra (500 MHz, CD<sub>2</sub>Cl<sub>2</sub>)

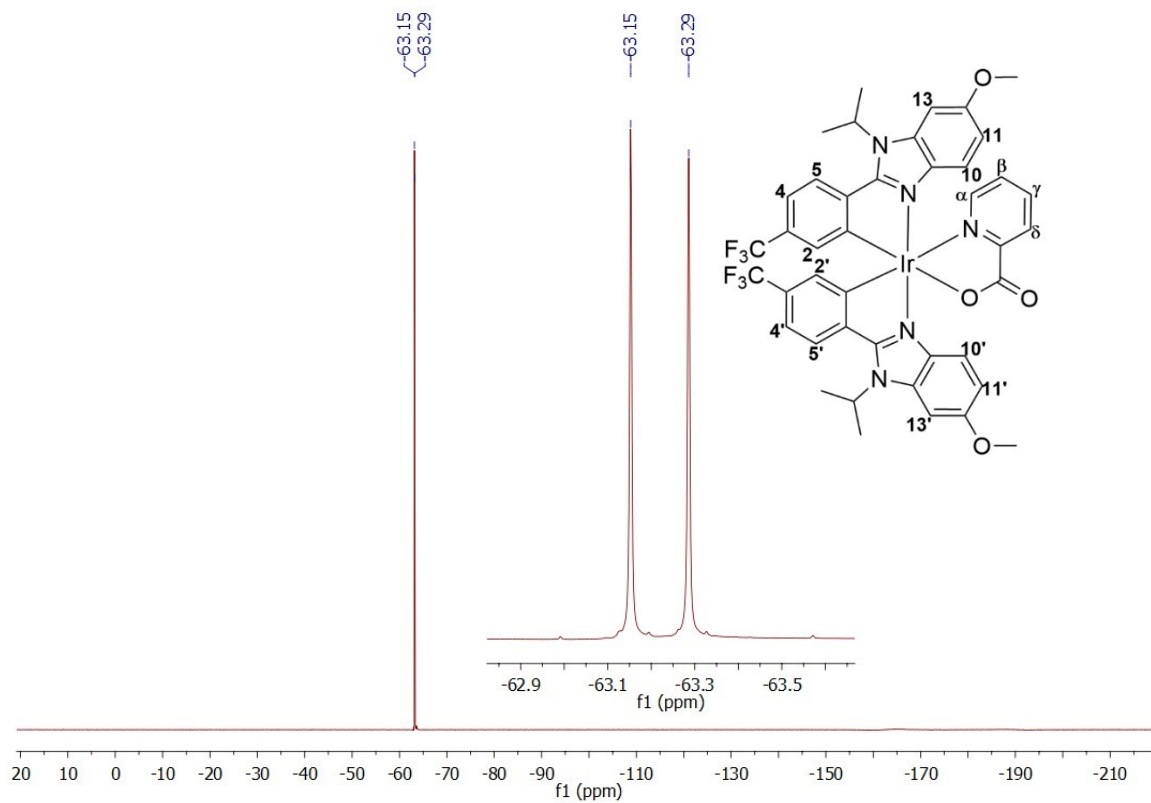
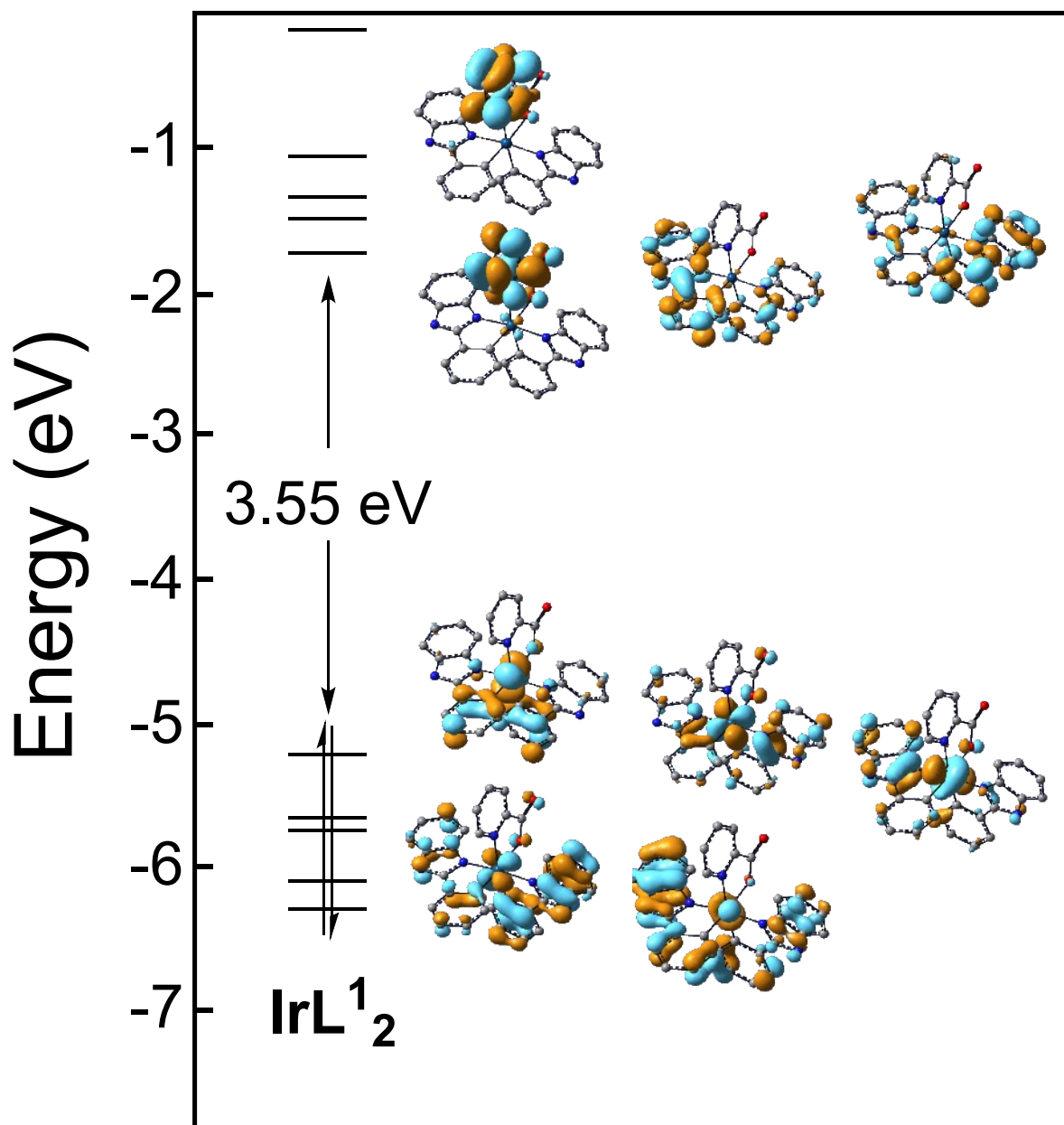
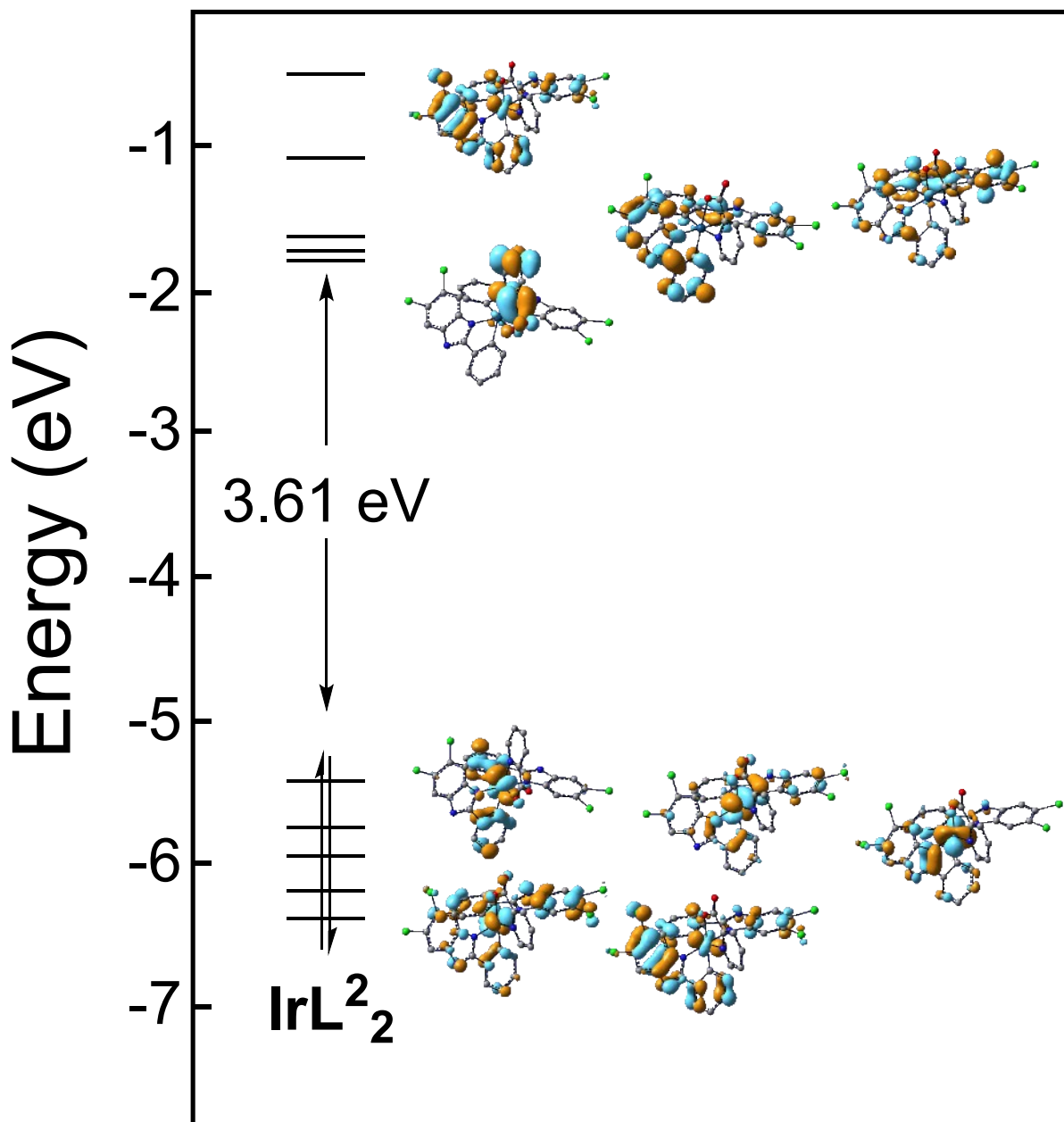
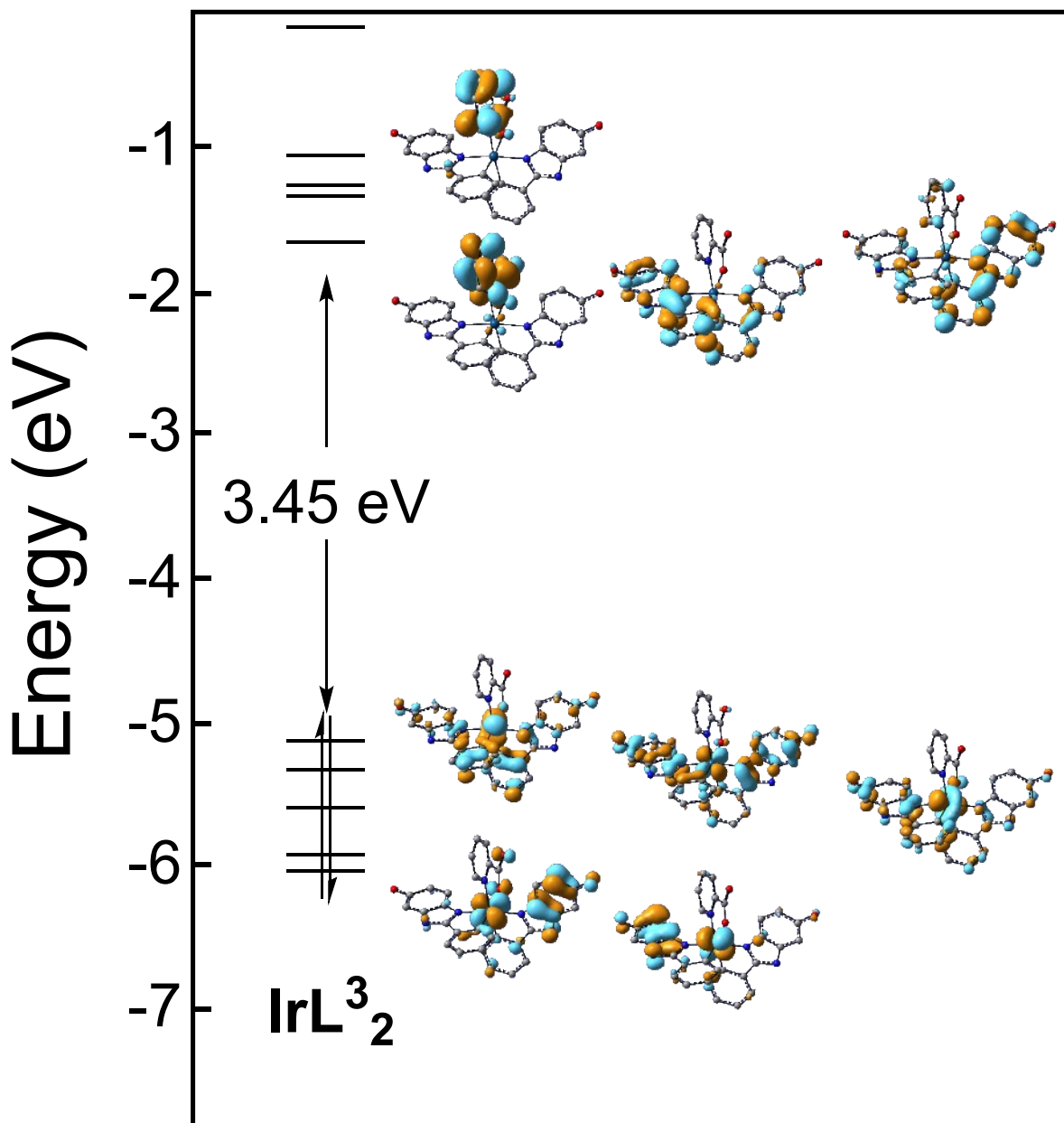


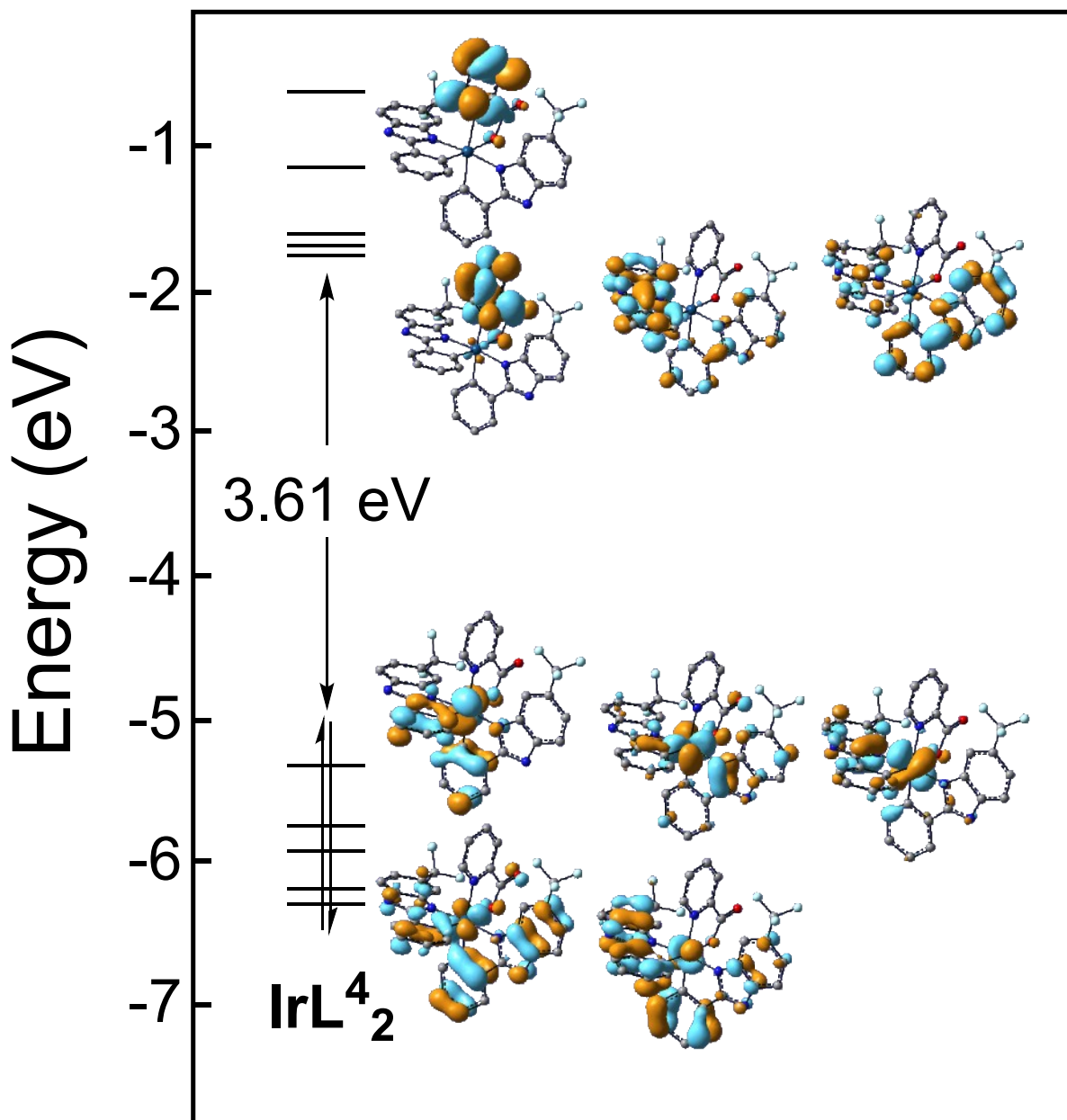
Figure S 29:  $^{19}\text{F}$  NMR spectra (500 MHz,  $\text{CD}_2\text{Cl}_2$ )

Figure S 30. Molecular orbital diagrams of the different species

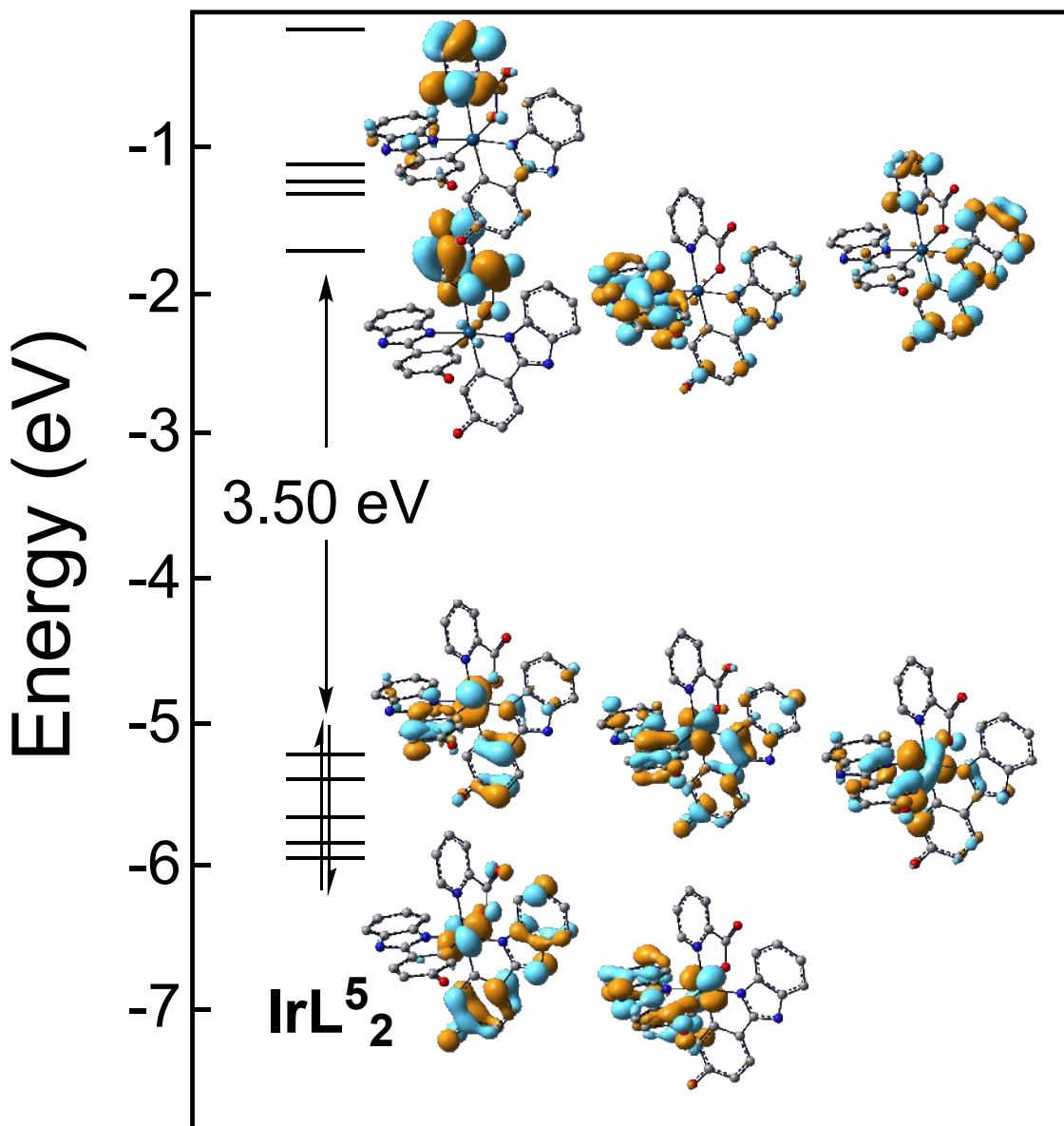


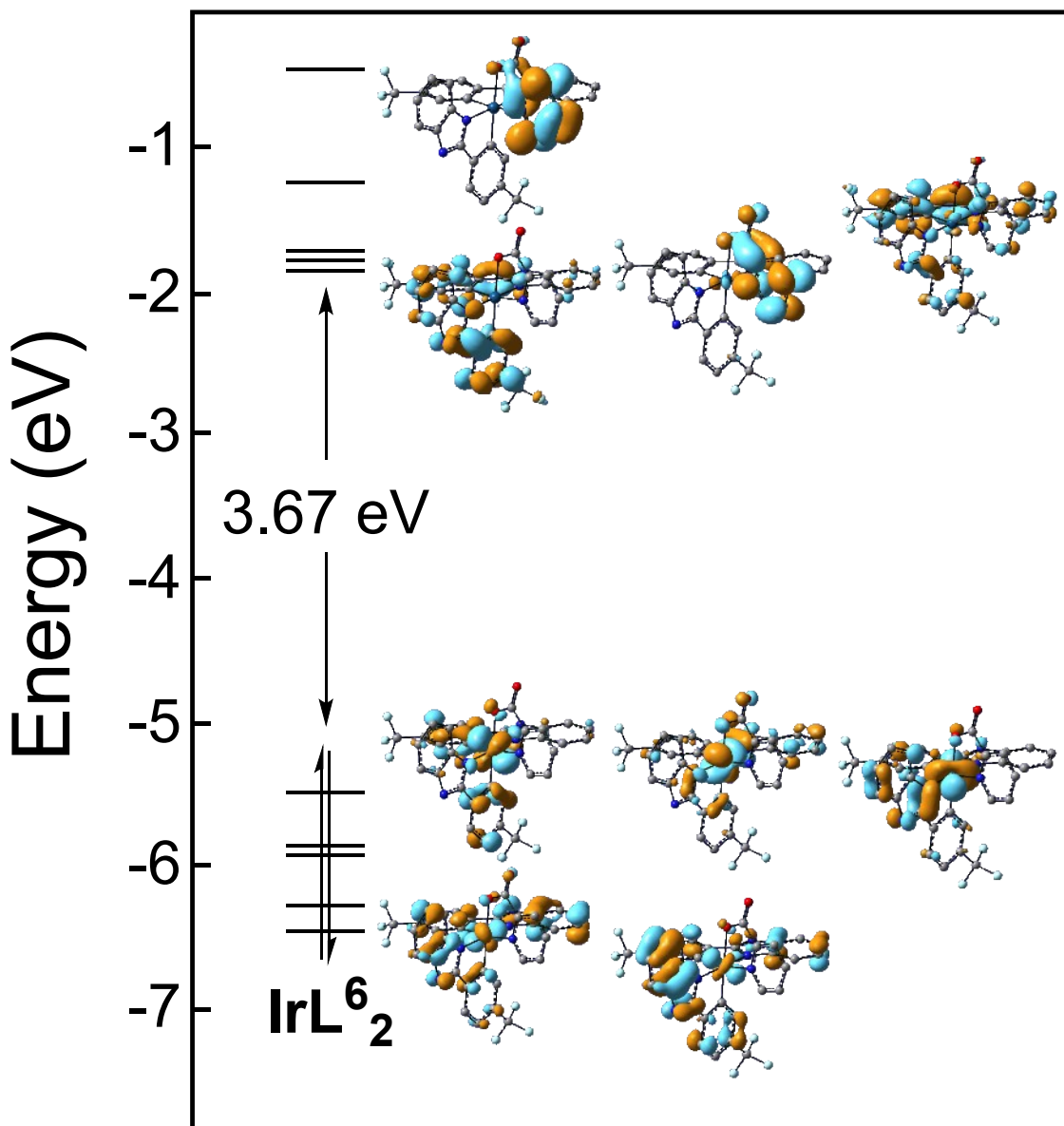


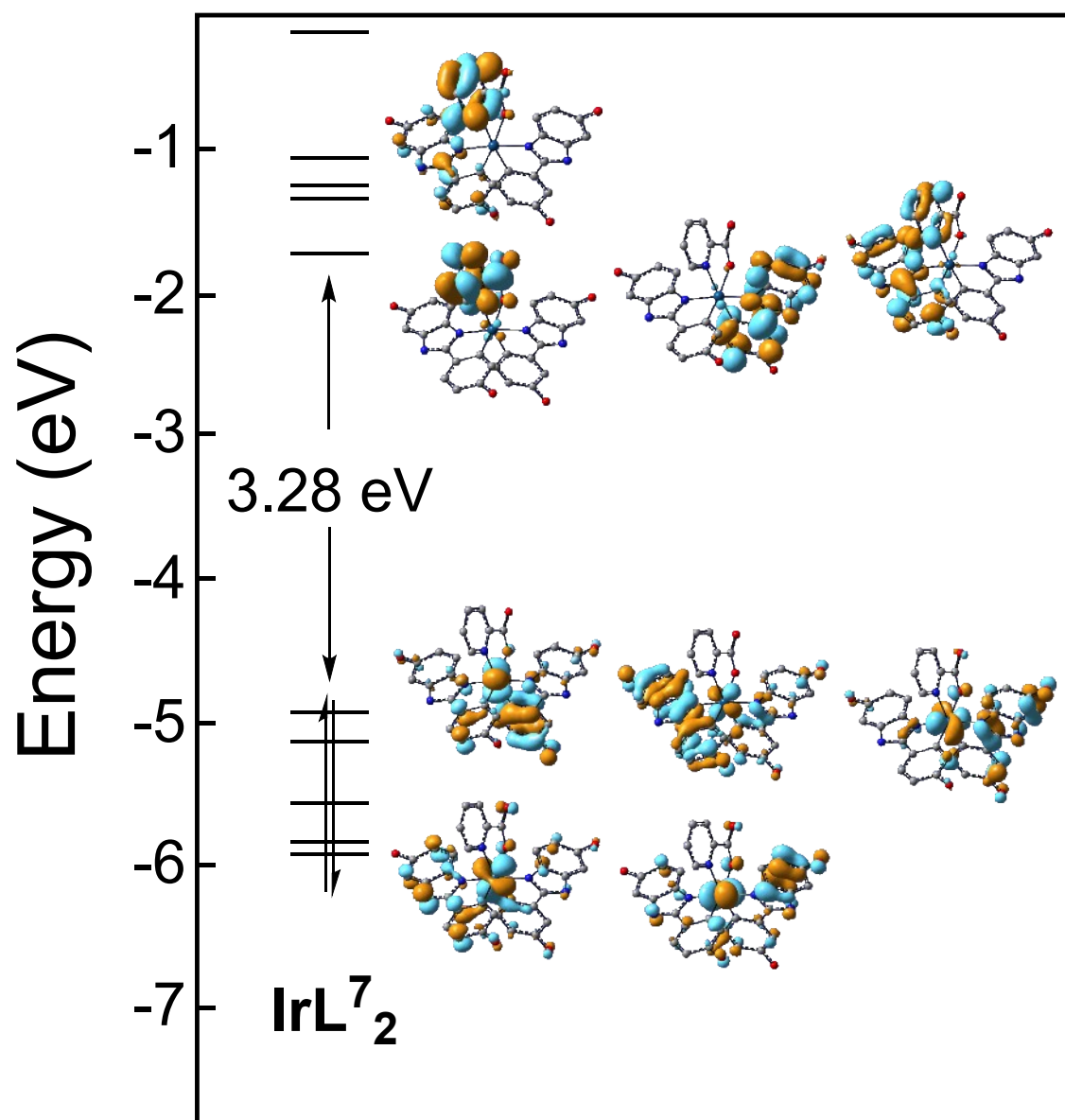


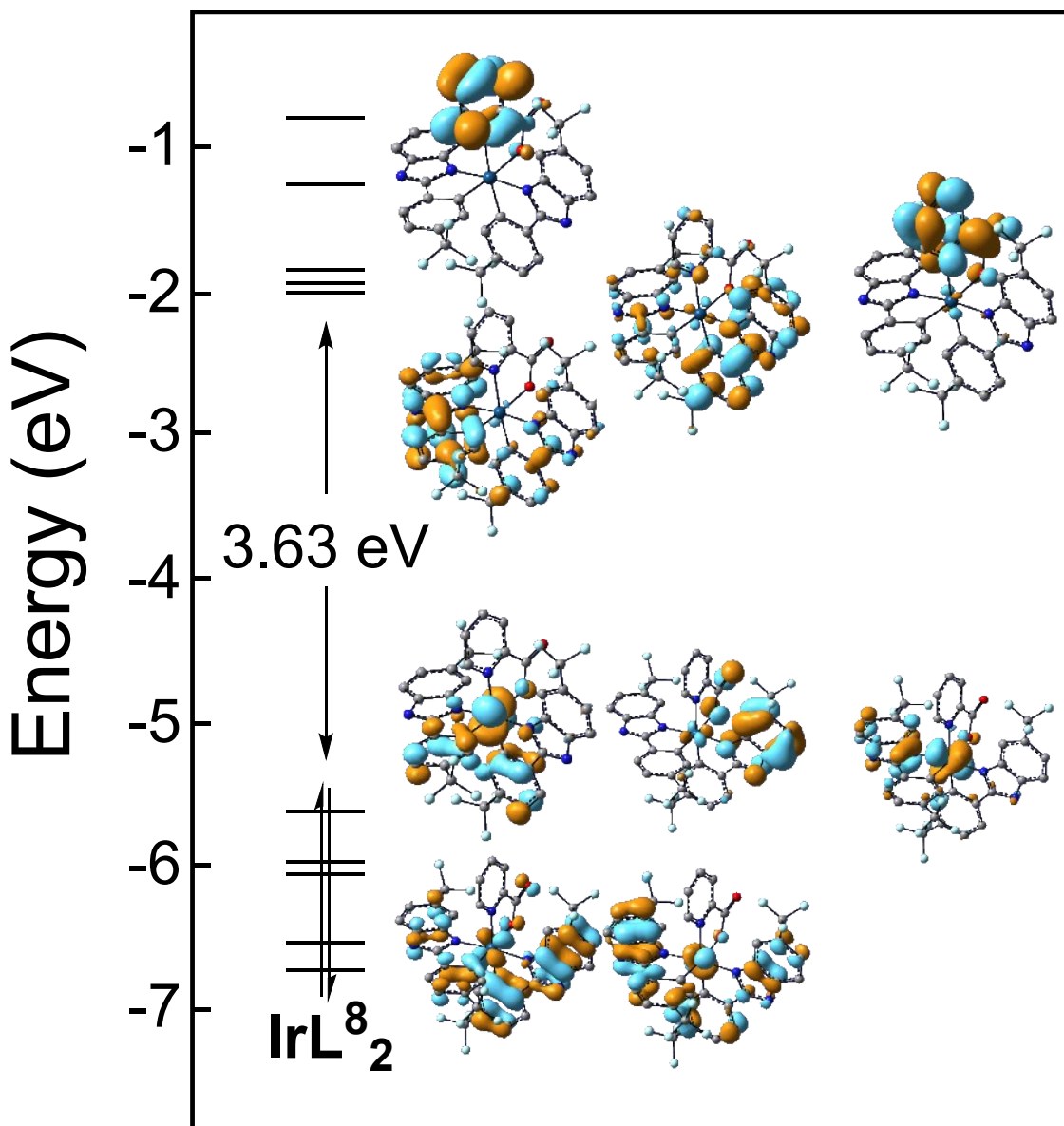


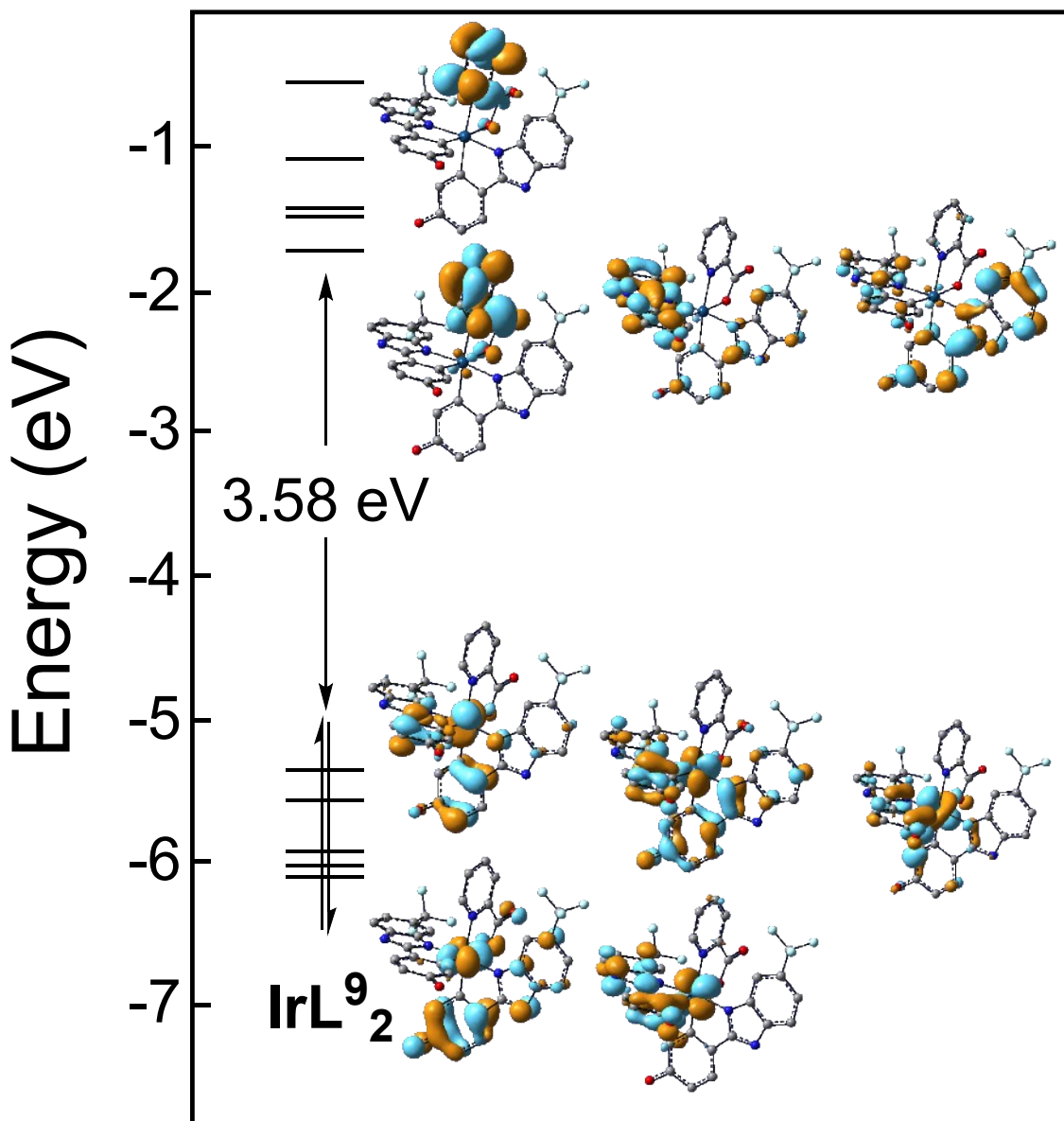


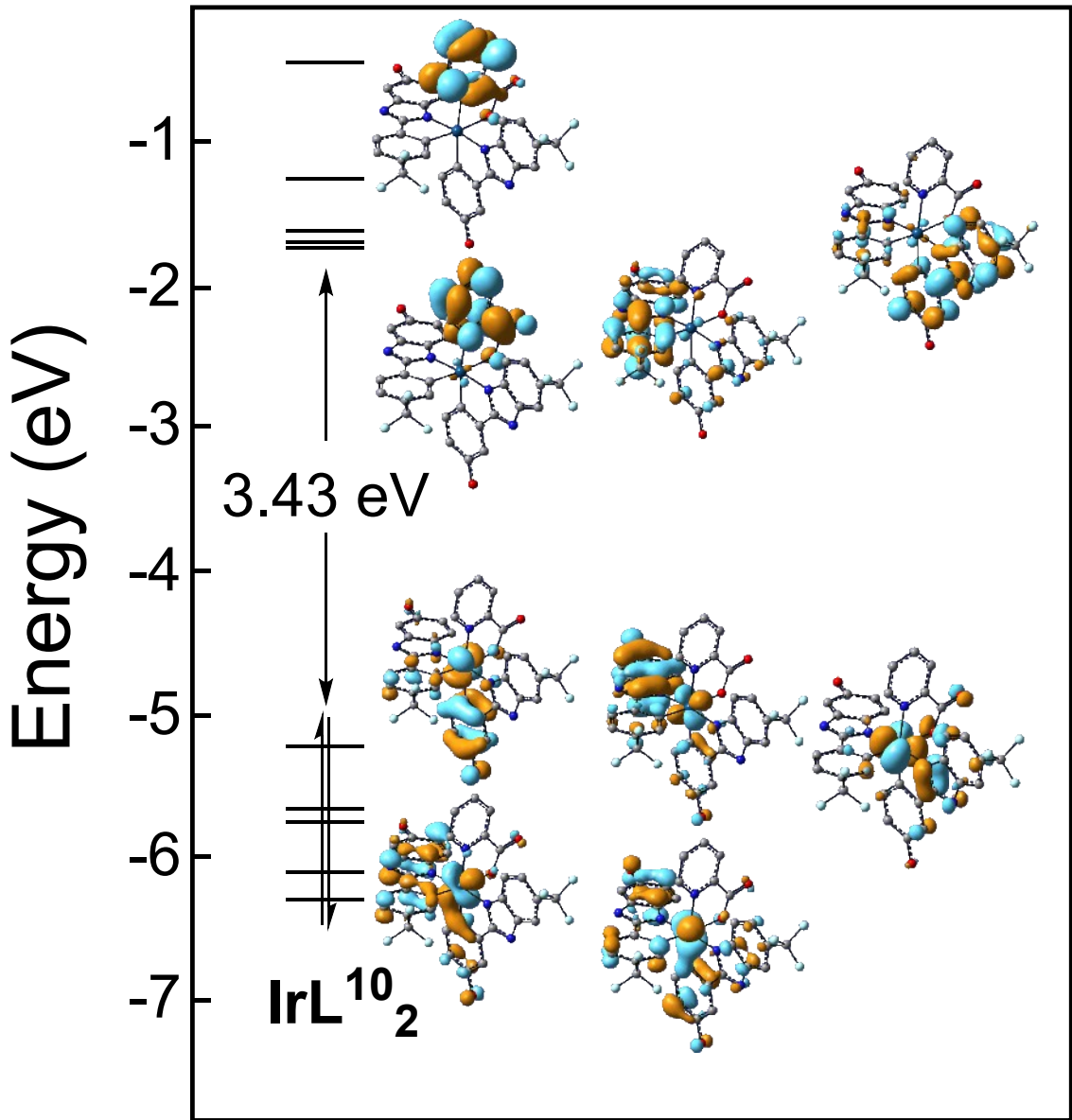












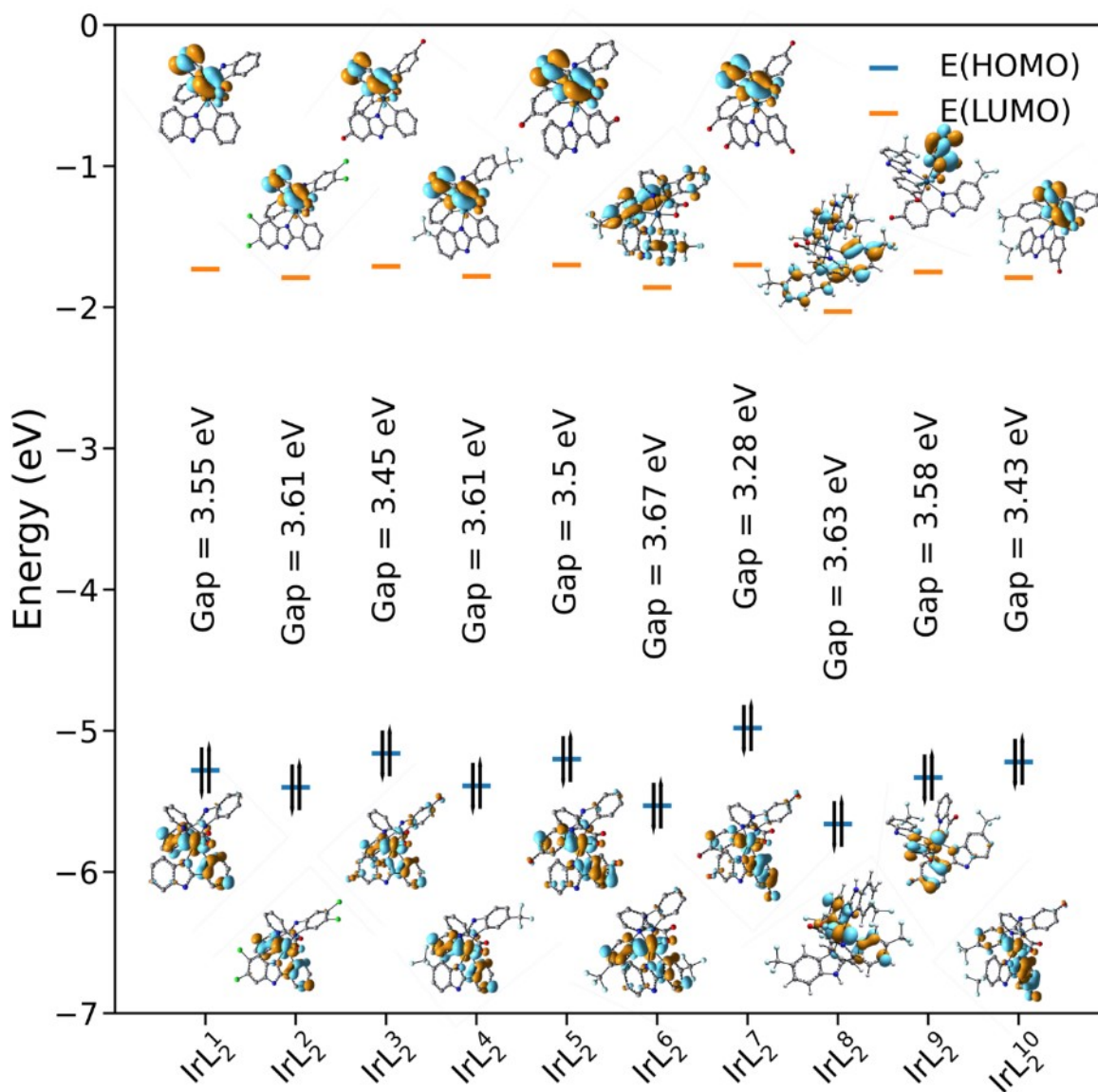


Figure S 31. HOMO-LUMO gap and orbitals for all complexes.

Table S 2. TD-DFT results and absorption band assignments.

Cpd	Transition* (f)	E(eV)	$\lambda$ (nm)	Assignment
IrL <sup>1</sup> <sub>2</sub>	HOMO → LUMO 0.003	2.83	438	ML'CT + LL'CT**
	HOMO → LUMO+1 0.196	3.04	408	<b>MLCT + LLCT</b>
IrL <sup>2</sup> <sub>2</sub>	HOMO → LUMO 0.008	2.90	428	ML'CT + LL'CT
	HOMO → LUMO+1 0.221	2.94	421	<b>MLCT + LLCT</b>
IrL <sup>3</sup> <sub>2</sub>	HOMO → LUMO 0.004	2.76	450	ML'CT + LL'CT
	HOMO → LUMO+1 0.223	3.04	408	<b>MLCT + LLCT</b>
IrL <sup>4</sup> <sub>2</sub>	HOMO → LUMO 0.004	2.90	428	ML'CT + LL'CT
	HOMO → LUMO+1 0.223	2.97	418	<b>MLCT + LLCT</b>
IrL <sup>5</sup> <sub>2</sub>	HOMO → LUMO 0.004	2.81	442	ML'CT + LL'CT
	HOMO-1 → LUMO 0.003	3.09	402	ML'CT + LL'CT
	HOMO → LUMO+1 0.263	3.20	387	<b>MLCT + LLCT</b>
IrL <sup>6</sup> <sub>2</sub>	HOMO → LUMO 0.171	2.94	421	<b>MLCT + LLCT</b>
	HOMO → LUMO+1 0.008	2.99	415	ML'CT + LL'CT
IrL <sup>7</sup> <sub>2</sub>	HOMO → LUMO 0.004	2.61	475	ML'CT + LL'CT
	HOMO → LUMO+1 HOMO-1 → LUMO 0.095	2.95	421	ML'CT + LL'CT + MLCT + LLCT
	HOMO → LUMO+1 HOMO-1 → LUMO 0.152	2.96	419	<b>ML'CT + LL'CT + MLCT + LLCT</b>
IrL <sup>8</sup> <sub>2</sub>	HOMO → LUMO 0.159	2.90	427	<b>MLCT + LLCT</b>
	HOMO → LUMO+1 0.015	3.02	411	ML'CT + LL'CT + MLCT + LLCT



<b>IrL<sup>9</sup><sub>2</sub></b>	HOMO → LUMO 0.004	2.89	430	ML'CT + LL'CT
	HOMO → LUMO+1 0.245	3.14	400	<b>MLCT + LLCT</b>
<b>IrL<sup>10</sup><sub>2</sub></b>	HOMO → LUMO HOMO → LUMO+1 0.018	2.75	450	ML'CT + LL'CT + MLCT + LLCT
	HOMO → LUMO HOMO → LUMO+1 0.165	2.78	447	ML'CT + LL'CT + MLCT + LLCT
	HOMO → LUMO+2 0.152	2.93	423	<b>MLCT + LLCT</b>

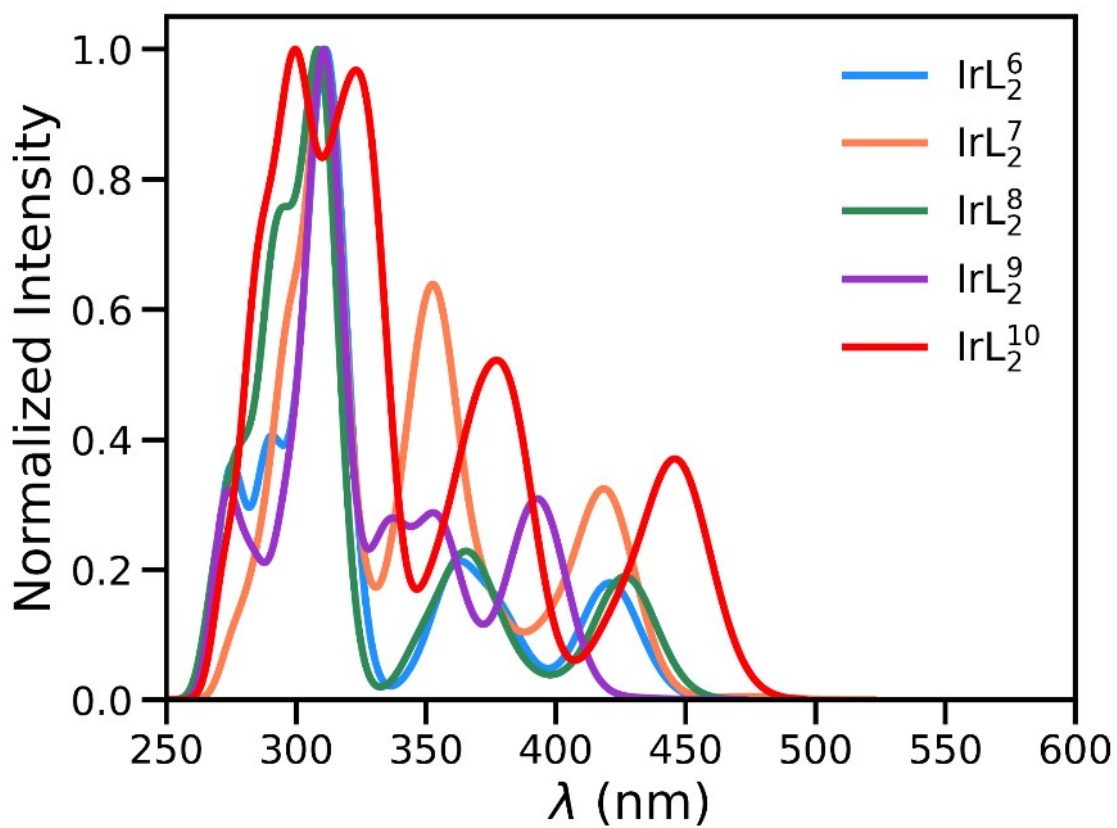
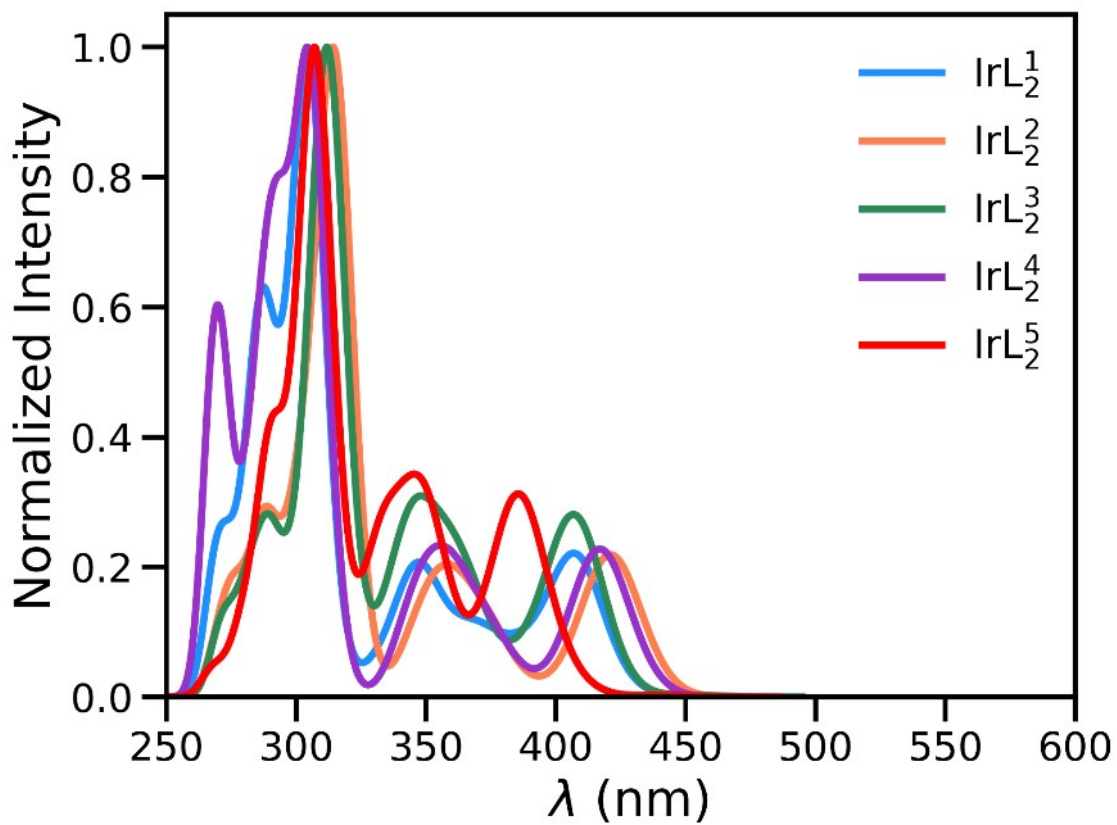
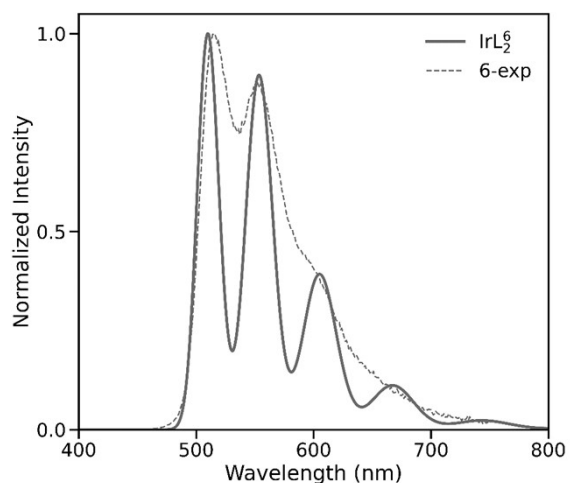
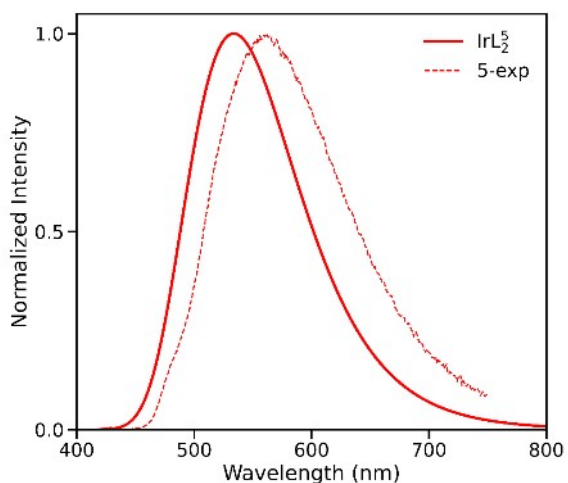
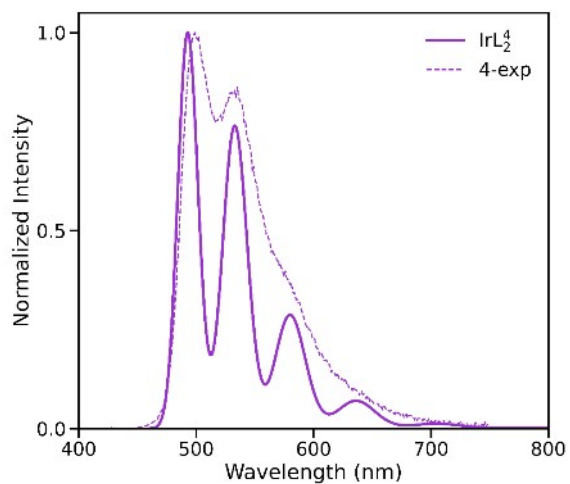
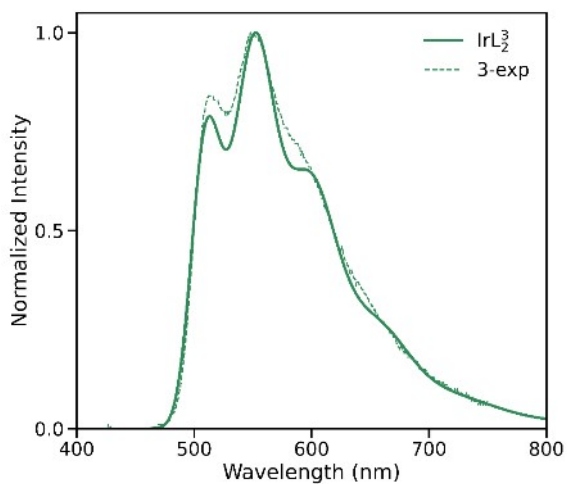
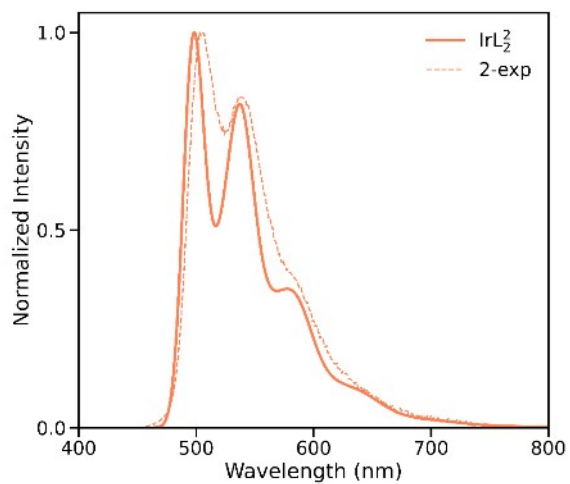
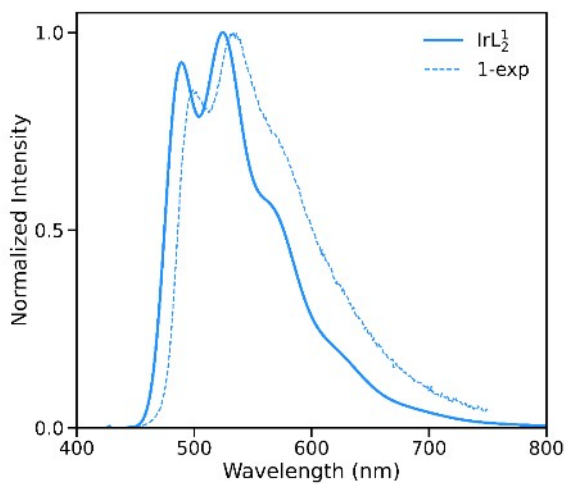


Figure S 32. Simulated absorption spectra using TD-DFT.



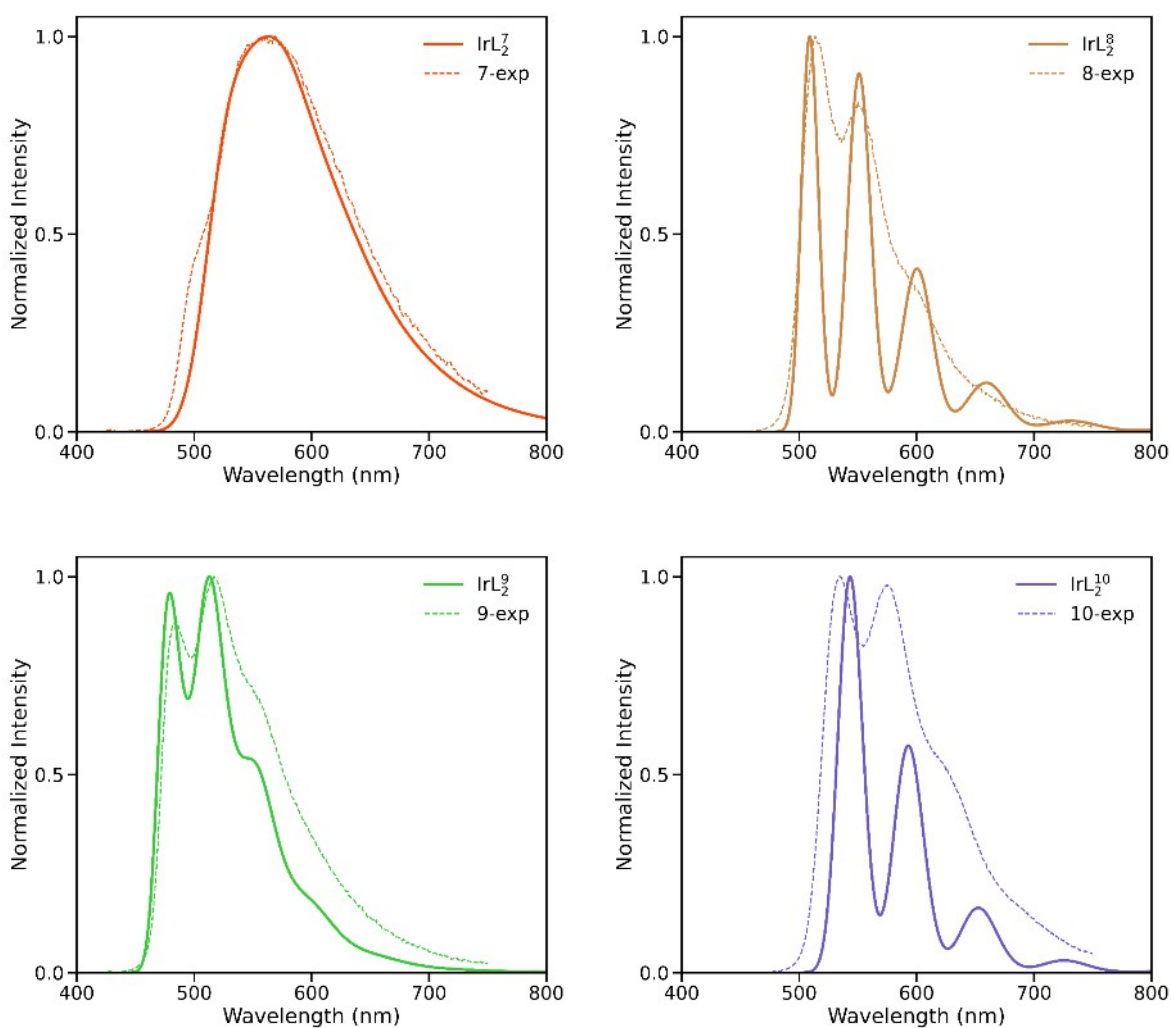


Figure S 33. Recorded (RT) and simulated phosphorescence spectra.

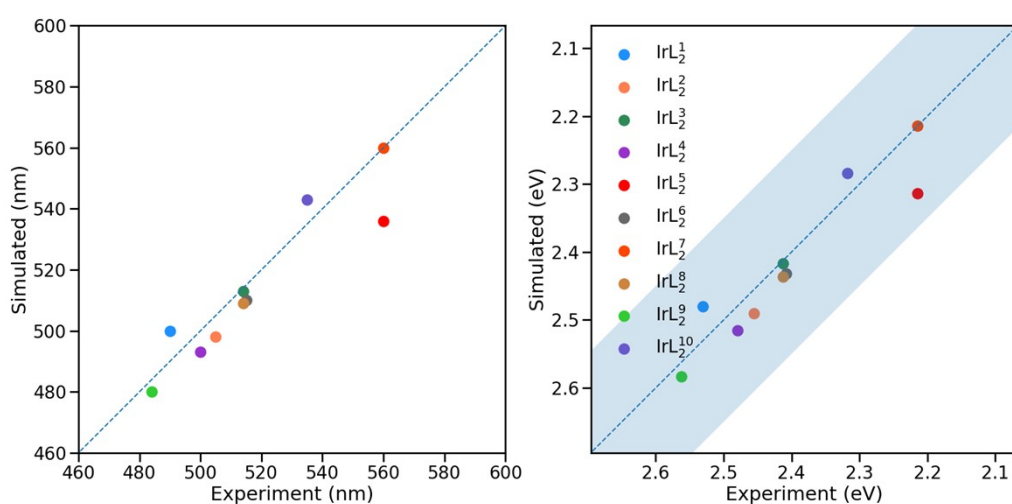


Figure S 34. Correlation between the simulated and recorded luminescence at RT.

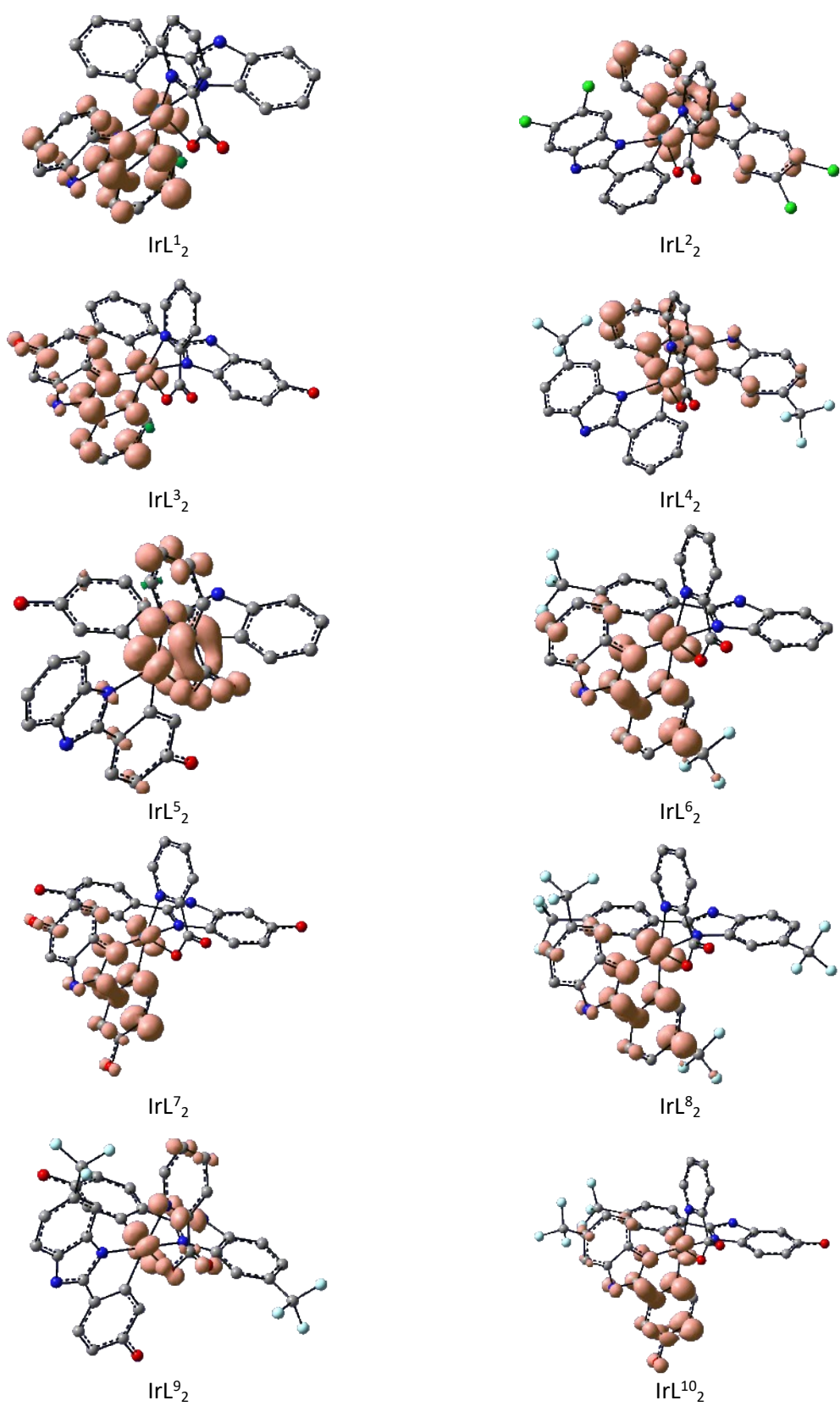


Figure S 35. Spin density localization of each complex in the excited state.

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