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Supporting information

Preparation of silyl-terminated branched polyethylenes catalyzed by Brookhart's nickel diimine complex activated with hydrosilane/B(C₆F₅)₃

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Experimental details

All syntheses and manipulations with air- and moisture-sensitive compounds were performed either under an argon atmosphere using Schlenk techniques or under purified nitrogen in a Labmaster 130 glovebox (mBraun). Solvents (hexane, toluene, dichloromethane) were purified by a solvent purification system (PureSolv MD 7; Innovative Technology, Inc.). Chemicals were used as received from commercial sources (Aldrich, Strem). IR spectra of samples in KBr pellets were measured in an air-protecting cuvette on a Nicolet Avatar FT IR spectrometer in the range 400–4000 cm⁻¹. KBr pellets were prepared under purified nitrogen in the glove box. NMR spectra were measured on a Bruker neo Avance 500 spectrometer at 293 K. Chemical shifts (δ , ppm) are given relative to solvent signal (CD₂Cl₂: $\delta_{\rm H}$ 5.32; toluene- $d_8 \delta_{\rm H}$ 2.08, $\delta_{\rm C}$ 137.48). EPR spectra were recorded on a CW X-band (9.5 GHz) EPR spectrometer MiniScope MS400 (Magnettech) with 100 kHz modulation equipped with microwave frequency counter FC 400 and temperature controller TC H03.

Preparation of 1



BIAN (BIAN = {Ar-N=Ace=N-Ar}, Ace = acenaphthen-1,2-diyl, and Ar = 2,6-(iPr)₂- C_6H_3)) (1.000 g, 2.00 mmol) and 1.98 mmol of NiCl₂DME (where DME = 1,2-dimethoxyethane) (0.435 g, 1.98 mmol), were weighted under nitrogen in the glove box. Flask was connected to an argon/vacuum line and 50 ml of dried CH₂Cl₂ was added. The mixture was stirred (1000 rpm) for 24 h at room temperature. The dark red solution was obtained. All volatiles were evaporated to give brick-red powder. This powder was three times washed with 15 ml of diethyl ether and the solution was filtered off via PTFE capillary. The brick-red powder was dried under a vacuum. The yield was 0.99 g (79 %).

¹H NMR (CDCl₃) δ: 24.42, 23.88, 16.82, 7.26, 5.82, 3.79, 2.84, 1.47, 1.27, 0.89, 0.07, -13.96. ¹H NMR (toluene-*d*₈) δ: 23.52, 16.17, 7.10, 6.98, 4.52, 1.34, 1.29, 1.26, 1.24, -1.66, -13.96. IR (KBr, cm⁻¹): 3061 (m), 2961 (vs), 2867 (s), 1655 (s), 1623 (s), 1598 (s), 1584 (s), 1488 (m), 1464 (s), 1436 (s), 1419 (m), 1384 (s), 1362 (s), 1327 (m), 1290 (s), 1253 (m), 1223 (m), 1202 (m), 1187 (m), 1130 (m), 1112 (m), 1088 (m), 1043 (m), 952 (m), 937 (m), 834 (s), 801 (s), 780 (vs), 758 (vs), 736 (m), 615 (m), 543 (s), 470 (m), 403 (m).

Polymerization

Polymerization procedure for 1/B(C₆F₅)₃/hydrosilane

Ethylene polymerization experiments were performed in a 250 mL Büchi glass double-jacketed laboratory autoclave equipped with a magnetic propeller-like stirrer at constant ethylene pressure. The hot autoclave was three times evacuated and filled with argon, charged successively with a solvent (dichloromethane or toluene), hydrosilane, and $B(C_6F_5)_3$ (toluene stock solution, 3.5×10^{-2} M) and the resulting mixture was stirred at 25 °C under argon for 30 min. The inert gas was vented and the reactor was pressurized with ethylene to the appropriate pressure for 10 min. Then, the ethylene pressure was partially released and the polymerization was started by injecting the desired amount of precatalyst in CH₂Cl₂ under a stream of ethylene and the autoclave was immediately pressurized with ethylene. Final volume of the polymerization solution was 50 mL for all experiments. The autoclave temperature was monitored during the reaction by using an external Pt100 sensor connected to a Julabo F31-C bath. Data collection was performed for temperature changes inside the reactor as well as for ethylene uptake during the polymerization. The consumption of ethylene was followed in each experiment using a calibrated mass flow meter (Bronkhorst, EL-FLOW, 1L/min). After the appropriate time, the reactor was vented and the polymerization was quenched with 80 mL of methanol. The precipitated polymer was stirred for 1 h, filtered off on a glass frit, rinsed repeatedly with methanol and acetone, and dried in a vacuum to constant weight.

Polymerization procedure for 1/B(C₆F₅)₃/binary hydrosilane system

A similar polymerization procedure (vide supra) was used for the binary hydrosilane system. The reactor was charged successively with dichloromethane, triethylhydrosilane, and the solution of borane under the stream of argon, and the resulting mixture was stirred at 25 °C for 30 min. The inert gas was displaced with ethylene and the reactor was pressurized 10 min with ethylene gas at 3×10^5 Pa. After venting the autoclave, the polymerization was started by consecutive loading of diphenylhydrosilane and the desired volume of Ni precatalyst in CH₂Cl₂. The reactor was immediately repressurized with ethylene to 3×10^5 Pa. After the appropriate time, the autoclave was vented and the polymerization was terminated by the addition of 80 mL of methanol. Processing of the precipitated polyethylene was the same as described above.

Polymerization procedure for 1/MMAO system

The polymerization was carried out in the same equipment as above under similar conditions (ethylene pressure 3×10^5 Pa, temperature 25 °C, total volume 50 mL, rpm 800). Dichloromethane was loaded into the reactor and pressurized with ethylene for 10 min. Subsequently, the reactor was vented and polymerization was initiated by successive injection of aluminoxane (solution of MMAO, 7 wt% in toluene, molar ratio MMAO/Ni= 1 000), the appropriate amount of precatalyst (1 µmol, solution in CH₂Cl₂) and immediate repressurization of the autoclave. After 10 min the polymerization mixture was quenched with 80 mL of a 10 % solution of HCl in methanol. The precipitated polymer was stirred for 1 h, filtered off on a glass frit, rinsed repeatedly with methanol and acetone, and dried in a vacuum to constant weight. Activity A= 4 070 kg_{PE} (mol_{Ni} h bar)⁻¹, M_n= 223 kg mol⁻¹, M_w/M_n= 2.1.

Fig. S1 Ethylene uptake during polymerization catalyzed with $1/B(C_6F_5)_3/Et_3SiH$ (red line, entry 1 in Table 1) and Cp₂ZrCl₂/B(C₆F₅)₃/Et₃SiH (blue line). Both polymerizations were conducted at identical conditions. Time period up to 0,2 min corresponds to the pressurization of the autoclave to an appropriate pressure (3 bar).



Fig. S2 Selected ethylene uptake profiles for $1/B(C_6F_5)_3/Et_3SiH$ catalytic system (Table 2), Influence of Si/Ni molar ratio. Time period up to 0,2 min corresponds to the pressurization of the autoclave to an appropriate pressure (3 bar).



Fig. S3. GC-MS analysis of polymerization mixture (entry 2, Table 4) after polymer isolation and volatile species evaporation. No ethylene oligomers nor products of hydrosilane dehydrocoupling were detected. A siloxane $(Et_3Si)_2O$ (at rt 11.45 min) was the only assigned product.



Polymer characterization

Molar masses and dispersities of polyethylene were determined using an HT SEC chromatograph (Polymer char) with infrared and viscometric detectors. NMR of polymers was measured on a Varian Mercury 300 instrument (¹H at 300 MHz, ¹³C at 75 MHz, and 29Si at 59 MHz in toluene- d_8 at 100 °C). The branching content of polymers was determined based on ¹H NMR spectra. FTIR of polymer samples were measured on a Nicolet iS10 FT-IR instrument. Transmission spectra were registered in the region 4000 – 400 cm⁻¹ using 64 scans at a resolution of 4 cm⁻¹. Solid samples of polymers were pressed on a special press at laboratory temperature into thin foils with a thickness optimally in the range of 150-250 µm and directly analyzed in the transmission mode of the spectrometer. The foil thickness was determined from the average of three measurements using a digital micrometer. For silyl-terminated polymers PE-SiH₂Ph and PE-SiHPh₂, the number of silyl groups per 1000 carbons of the polymer chain (N_{Si}= Si/1000C) was calculated from the relation of N_{Si}= 14 A (ρ t ε)⁻¹,

where A is the absorbance of the signal at 2134 cm⁻¹ or 2117 cm⁻¹ (stretching vibrations of SiH₂ and SiH for groups SiH₂Ph and SiHPh₂, respectively), ρ is the density of the polymer (0.95 g m L⁻¹), *t* is the thickness of the PE film (cm) and ϵ denotes the relevant molar absorption coefficient. Coefficients $\epsilon_{2134 \text{ cm}-1} = 165 \text{ Lmol}^{-1}\text{cm}^{-1}$ and $\epsilon_{2117 \text{ cm}-1} = 95 \text{ Lmol}^{-1}\text{cm}^{-1}$ were calculated from a calibration curve by plotting a series of linear silyl-terminated polyethylenes with known silyl content quantified by NMR spectroscopy, against the absorbances of the mentioned IR signals at different thickness of PE foils.

For the quantitative estimation of unsaturated groups in polymer products, the molar absorption coefficients of signals 908 cm⁻¹ for the vinyl group in 1-dodecene ($\epsilon_{908 \text{ cm}-1} = 110 \text{ L}$ mol⁻¹ cm⁻¹), 964 cm⁻¹ for *trans*-vinylene ($\epsilon_{964 \text{ cm}-1} = 168 \text{ L}$ mol⁻¹ cm⁻¹), and 888 cm⁻¹ for vinylidene in 2,3-dimethylbutadiene ($\epsilon_{888 \text{ cm}-1} = 146 \text{ L}$ mol⁻¹ cm⁻¹) were used.¹ In this case, the absorbances of the mentioned alkene signals obtained by subtracting the spectrum of polyethylene after bromination from the original spectrum of the polymer before halogenation were evaluated. Bromination of polymer samples was carried out at laboratory temperature in liquid bromine for 24 hours. After removal of bromine, the sample was thoroughly rinsed with acetone and dried to constant weight in a vacuum oven at a temperature of 120 °C. ATR spectra of polyethylenes were obtained on the same FT-IR spectrometer using an attenuated total reflectance cell equipped with a ZnSe crystal in the range 650-4000 cm⁻¹.

Fig. S4 ¹H NMR spectrum of *br*-PE (entry 6 in Table 2) measured at 100 °C in toluene- d_8 . (*) denote residual proton signals of the solvent. The insert spectrum indicates a trace amount of internal double bonds (multiplet centered at 5.40 ppm) and vinyl chain-ends (multiplets centered at 4.95 and 5.76 ppm) in the polymer.



Fig. S5 ¹H NMR spectrum of *br*-PE-SiPhH₂ (entry 8 in Table 3) measured at 100 °C in toluene- d_8 . (*) denote residual proton signals of the solvent. The insert spectrum indicates a trace amount of internal double bonds in the polymer (multiplet centered at 5.39 ppm).



Fig. S6 FT-IR spectra (PE foils, an expanded region) of polymers obtained from $1/B(C_6F_5)_3/Et_3SiH$ system: red line (entry 1, Table 2), violet line (entry 2, Table 2). For comparison spectra of vinyl-terminated PEs are depicted. The green line shows PE prepared with Cp₂ZrCl₂/B(C₆F₅)₃/Et₃SiH system (Mn = 81 000 g/mol, vinyl content 40 mol%). Turquoise line shows PE prepared with bis (N-(3-tert-butylsalicylidene)anilinato) zirconium(IV)dichloride/B(C-E-)/Et-SiH system

bis {N-(3-tert-butylsalicylidene)anilinato} zirconium(IV)dichloride/B(C_6F_5)₃/Et₃SiH system ($M_n = 5000$ g/mol, vinyl content 98 mol%). Bands at 908 and 990 cm⁻¹ are characteristic of vinyl vibrations, while the band at 964 cm⁻¹ is suggested to be characteristic of internal double bonds.



Fig.S7 ATR-ZnSe spectra (an expanded region) of PEs obtained from binary hydrosilane system activation of 1: $Et_3SiH/Ph_2SiH_2 = 60/1$ (entry 1 in Table 4, bottom), $Et_3SiH/Ph_2SiH_2 = 1/1$ (entry 2 in Table 4, middle) and from activation using sole Ph_2SiH_2 (entry 5 in Table 3, top). For clarity, only vibrations of the terminal silyl group are assigned.



NMR experiments

1/B(C₆F₅)₃/10 eq. Et₃SiH in CD₂Cl₂

In a glovebox $B(C_6F_5)_3$ (20 mg, 0.039 mmol), and 1 (24 mg, 0.039 mmol) were weighed into a flask equipped with a Young valve and then connected to an argon-vacuum line. 1 ml of CD_2Cl_2 was added to dissolve all components. To the orange solution, Et_3SiH (65 µl, 0.407 mmol) was added. The solution was transferred by gastight syringe to an NMR cuvette equipped with a Young valve and sealed off under a vacuum. The evolution of the reaction mixture was followed by multinuclear NMR spectroscopy.

1/B(C₆F₅)₃/20 eq. 1-hexene/10 eq. Et₃SiH in CD₂Cl₂

The initial mixture was prepared as described above from $B(C_6F_5)_3$ (20 mg, 0.039 mmol) and 1 (24 mg, 0.039 mmol). To the orange solution, 1-hexene (120 µl, 0.970 mmol) was added (no color change was observed). Finally, Et₃SiH (65 µl, 0.407 mmol) was added and the solution was transferred by gastight syringe to an NMR cuvette equipped with a Young valve and sealed off under a vacuum. The evolution of the reaction mixture was followed by multinuclear NMR spectroscopy.

1/B(C₆F₅)₃/200 eq. 1-hexene/100 eq. Et₃SiH in tolueneD₈

Stock solutions of **1** (2.4 μ mol, 3 ml of 0.8 mM) and B(C₆F₅)₃ (2.4 μ mol, 3 ml of 0.8 mM) in toluene were transferred into a flask equipped with a Young valve by a gastight syringe, and then connected to an argon-vacuum line. The toluene was evaporated under a vacuum and 1 ml of deuterated toluene was added to dissolve all components. To the orange solution, 1-hexene (60 μ l, 0.480 mmol) was added (no color change was observed). Finally, Et₃SiH (38 μ l, 0.240 mmol) was added and the solution was transferred by gastight syringe to an NMR cuvette equipped with a Young valve and sealed off under a vacuum. The evolution of the reaction mixture was followed by multinuclear NMR spectroscopy.

Fig. S8 ¹H NMR spectra of $1/B(C_6F_5)_3/10$ eq. Et₃SiH/1-hexene in CD₂Cl₂ measured after 15 min, 75 min, 1 day, and 2 days. The inserts showed APT spectrum of the mixture measured 1 day after mixing.



Fig. S9 ²⁹Si{¹H} NMR spectra of $1/B(C_6F_5)_3/10$ eq. Et₃SiH in CD₂Cl₂ measured after 1h, 9h, 2 days, and 7 days.



Fig. S10 ¹H NMR spectra of $1/B(C_6F_5)_3/10$ eq. Et₃SiH in CD₂Cl₂ measured after 35 min, 9h, 2 days, and 7 days. The inserts showed the formation of CD₂HCl (δ_H centered at 2.99 ppm) and CD₂H₂ δ_H centered at 0.19 ppm).





Fig.S11 ²⁹Si{¹H} NMR spectra of $1/B(C_6F_5)_3/100$ eq. Et₃SiH/200 eq. 1-hexene in toluene- d_8 (NS= 20 000) measured 2 months after the mixture preparation.

Fig. S12. ¹³C{¹H) NMR spectrum of mixture $1/B(C_6F_5)_3/100$ eq. Et₃SiH/200 eq. 1-hexene in toluene- d_8 measured 2 months after mixing (a) and ¹³C{¹H) NMR spectrum of poly(1-hexene) isolated thereof mixture (b).



75 70 f1 (ppm) 5 0 140 135 130 125 120 115 110 105 100

EPR experiments

1/B(C₆F₅)₃/hydrosilane

 $B(C_6F_5)_3$ (20 mg, 0.039 mmol) and 1 (24 mg, 0.039 mmol) were weighed into a flask equipped with a Young valve in the glove box and moved to the argon-vacuum line. 5 ml of dichloromethane was added to dissolve all components. The orange solution was then transferred by gastight syringe to an EPR cuvette equipped with a Young valve. All volatiles were evaporated by the vacuum and 6 ml of toluene was added. EPR spectra were collected (no signal was observed in the absence of hydrosilane). An EPR cuvette was then connected to the argon-vacuum line and a hydrosilane (0.405 mmol) was added. In dependence on a hydrosilane nature, the color of the mixture remained yellow (Et₃SiH) or changed steadily/immediately to green/turquoise (Ph₂SiH₂/PhSiH₃). EPR spectra were collected at room and low temperatures (in the range 293-113 K). In addition, the time evolution of Ni(I) signal was monitored for several hours.

1/Et₃SiH /B(C₆F₅)₃ /1-hexene

1 (4 mg, 6 µmol) was weighted in the glovebox into the Schlenk flask. 5.5 ml of toluene was added by syringe to form an orange solution, all solid was dissolved. The solution was transferred to an ampoule equipped with an EPR cuvette. 0.313 mmol of Et₃SiH (0.05 ml, 313 µmol) was added. No change of color was observed. No EPR signal was observed for 4 days. $B(C_6F_5)_3$ (10 mg, 19 µmol) was weighed in the glove box and dissolved in 2 ml of toluene. The colorless solution was transferred into the reaction mixture by syringe. The color of the solution was changed to bright yellow. EPR measurements show signals of Ni(I) and radical (7 lines) at a g-value of 2.002 immediately after mixing. After 24 h only the signal of Ni(I) remains in the EPR spectrum. 1-hexene (10 µl, 80 µmol) were added. EPR spectra without change also at low temperatures were obtained. No color change was observed. After 24 h another 0.5 ml of 1-hexene was added and stirred. After one week of stirring, the ampoule contains a viscous solution (mead consistency). EPR measurement did not show any changes in the EPR spectra of Ni(I) species (measured at low temperatures too). The solution was opened and poured into a mixture of methanol with HCl. The gel was immediately observed together with a color change into pink. The polymer was separated and left stirred in methanol. The yield of polyhexene was 122 mg.

$1/B(C_6F_5)_3/Et_3SiH$

B(C₆F₅)₃ (6 mg, 12 µmol) and **1** (5 mg, 8 µmol) were weighted in the glovebox into the Schlenk flask. 2 ml of dichloromethane were added, and all compounds were dissolved formed a yellow solution. The solution was transferred by syringe to the next ampoule equipped with an EPR cuvette (quartz tube 5 mm). All volatiles were evaporated to vacuum and toluene (6 ml) was added. The yellow solution was obtained and EPR spectra were collected without any signal. After 24 h new small signal (7 lines) was found in the solution at g = 2.002 and its intensity was increased with time. 0.313 mmol of Et₃SiH (0.05 ml, Mw = 116.28) was added and the mixture was stirred, and immediately measured on an EPR spectrometer. Continuous measurement proved the rising signal of Ni(I) and the increased signal of radical too. The next day dark blue-grey solution shows a signal of Ni(I) only. Radical disappeared. All volatiles were evaporated off and measured on GC-MS.

BIAN / $B(C_6F_5)_3$

 $B(C_6F_5)$ (7 mg, 14 µmol) and BIAN (5 mg, 12 µmol) were weighted in the glovebox into the Schlenk flask. 5 ml of toluene was added by syringe. The reaction followed by the color change was observed. All solids were dissolved and the dark yellow solution was transferred to an ampoule equipped with a quartz cuvette for EPR measurement. EPR spectrum shows one signal with splitting (7 lines). Measurements were repeated several times till no change in signal intensity was observed.

Fig. S13 Comparison of X-band EPR spectra of Ni(I) species in the reaction mixture 1, $B(C_6F_5)_3$, Et₃SiH and 1-hexene in toluene solution at room temperature



Discussion concerning the formation of organic ligand (g = 2.001) generated in $1/B(C_6F_5)_3/10$ eq. hydrosilane system.

A second signal detected in the EPR spectrum of mixture $1/B(C_6F_5)_3$ /hydrosilane was the signal of an organic radical with hyperfine splitting. The radical displayed a 7-lined signal at g value 2.002 with hyperfine splitting a = 0.51 mT (Figure 3A). This signal suggests the presence of a radical formed by the interaction of $B(C_6F_5)_3$ with the π -system of BIAN. The splitting of the signal may come from the coupling of the unpaired electron with two ¹⁴N (I=1) and two ¹H (I=1/2) nuclei.



Chart S1

The exact structure of the radical cannot be unequivocally determined from our experiments, however, we tentatively propose a formation of boron species with ligand based radical [(BIAN)B(C₆F₅)₂] depicted in Chart S1. The splitting from ¹¹B and ¹⁰B nuclei was not observed in the experimental spectrum and is suppressed by more significant splitting from two ¹⁴N and two ¹H nuclei, as was reported for [DPP-DAB]⁻BF₂⁺ radical.² The formation of the radical [(BIAN)B(C₆F₅)₂] could be explained as a carboboration of BIAN with B(C₆F₅)₃, followed by C₆F₅ elimination (see Scheme S1), similarly as was described by Stephan for carboboration of dione O₂C₁₄H₈ with B(C₆F₅)₃.^{3.4} Notably, a reaction of free diimine BIAN ligand with B(C₆F₅)₃ generated an identical signal at g 2.002 (see Fig. S14), which supported that its formation did not involve any Ni species derived from **1**.



Scheme S1: Proposed mechanism of the reaction of BIAN with $B(C_6F_5)_3$ leading to the paramagnetic species.

Fig. S14 EPR spectrum of mixture BIAN molecule with $B(C_6F_5)_3$



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