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Table A1 Parameter values used in the present exposure assessment.

Table A2 Genus composition of sampling sites during the three years of the COVID-19.

Table A3 Specific environmental conditions for sampling at each site. Temperature:

TEM(°C), humidity level: HUM (%), light intensity: LI (Lux), Wind speed: WS (m/s). 2019:

A, 2020: B, 2021: C. North Door (ND), West Door (WD), North playground (NP).

Table A1 Parameter values used in the present exposure assessment.

Parameters	Values
Inhalation rate (IR) (m³/d)	19.02 (Adult male), 14.17 (Adult female)
Exposure time (ET) (yr)	24
Exposure frequency (EF) (d/yr)	180
Average body weight (BW) (kg)	66.2 (Adult male), 57.3 (Adult female)
Average time (AT) (d)	73.64×365 (Adult male), 79.43×365 (Adult female)
Exposure skin area (SA) (m ²)	0.215
Skin adherence factor (SL) (kg/(m³/d))	0.07
Dermal absorption factor (ABS)	0.001

Table A2 Genus composition of sampling sites during the three years of the COVID-19.

genus	19ND	19NP	19WD	20ND	20NP	20WD	21ND	21NP	21WD
Pseudomonas	10.41%	9.01%	10.38%	69.38%	49.29%	30.06%	11.42%	13.70%	19.16%
Micrococcaceae	46.51%	19.35%	29.82%	0.86%	1.00%	6.10%	18.38%	11.49%	19.97%
Bacillus	0.12%	0.12%	0.06%	0.72%	2.55%	3.99%	8.31%	26.68%	19.23%
Pantoea	0.01%	0.05%	0.04%	2.15%	5.43%	2.61%	7.54%	7.79%	21.04%
Sphingomonas	7.99%	15.98%	18.93%	0.09%	0.06%	0.07%	0.00%	0.01%	0.00%
Paenarthrobacter	1.47%	20.78%	0.15%	0.29%	1.23%	5.45%	3.72%	2.02%	2.28%
Exiguobacterium	0.02%	0.09%	0.21%	7.06%	6.58%	7.80%	3.39%	0.21%	0.03%
Chitinophagaceae	8.21%	3.73%	7.95%	0.01%	0.00%	0.02%	0.00%	0.00%	0.00%
Stenotrophomonas	3.30%	0.88%	0.03%	0.02%	0.04%	2.03%	10.64%	1.57%	1.10%
Acinetobacter	0.03%	0.02%	0.09%	4.56%	1.21%	6.88%	2.75%	2.70%	0.16%
Others	21.88%	29.97%	32.33%	14.82%	32.58%	34.94%	33.76%	33.77%	16.96%
Unknown	0.04%	0.03%	0.03%	0.04%	0.03%	0.04%	0.09%	0.07%	0.07%

Table A3 Specific environmental conditions for sampling at each site. Temperature: TEM(°C), humidity level: HUM (%), light intensity: LI (Lux), Wind speed: WS (m/s). 2019: A, 2020: B, 2021: C. North Door (ND), West Door (WD), North playground (NP).

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WD Date	Sampling time	TEM	HUM	LI	WS
March 28th	17:18-17:33	16	35	3621	3.6
March 29th	17:27-17:40	19	36	2214	2.8
March 30th	17:20-17:40	16.5	38	1295	2.4
March 31th	17:32-17:49	18.2	40	2935	1.5
April 1st	17:02-17:17	20.5	33	4424	1.2
April 2nd	17:10-17:30	19.7	34	6524	3.2
April 3rd	17:10-17:30	21	48	3023	2.7
April 4th	17:35-17:54	22.4	34	6343	2.4
April 5th	17:34-17:51	26.7	28	2041	3.7
April 6th	17:34-17:51	21	48	1084	4
April 7th	17:31-17:50	24	35	2091	2.1
April 8th	17:32-17:54	14	58	867	1.6
April 9th	17:32-17:51	10	79	935	5
April 10th	17:29-17:50	14	57	676	1.2
April 11th	18:10-18:29	21	65	492	2.6
April 12th	17:26-17:46	19.7	70	2324	2.8
April 13th	17:29-17:47	23	59	321	0
April 14th	17:26-17:44	20.5	40	852	2.4
April 15th	17:08-17:27	21.2	27	1024	4.2
April 16th	17:09-17:28	24	48	4022	5.2
April 17th	18:48-19:04	30	45	109	3.8
April 18th	17:27-17:40	26	44	5935	6.5
April 19th	17:20-17:40	18.2	34	4476	5.2
April 20th	17:32-17:49	24.1	37	9234	2.8
April 21st	17:02-17:17	16.5	89	2324	3.4
April 22nd	17:10-17:30	14.5	90	102	1.2
April 23rd	17:10-17:30	24.5	54	2451	2.5

ND Date	Sampling time	TEM	HUM	LI	WS
March 28th	15:37-15:55	18	35	1037	4.5
March 29th	15:29-15:44	18.4	36	8625	2.4
March 30th	15:32-15:51	15.5	38	1463	2.8
March 31th	15:35-15:56	16.4	40	4212	1.4
April 1st	15:35-15:56	20	33	7035	1.3
April 2nd	15:55-16:16	21	34	1265	3.7
April 3rd	15:40-16:00	20.1	48	1893	2.4
April 4th	15:52-16:14	23.2	34	4212	2.8
April 5th	15:32-15:50	26.8	28	5232	3
April 6th	15:31-15:49	22.1	48	1962	4.2
April 7th	15:33-15:48	22.6	35	1925	2.1
April 8th	15:37-15:55	15	58	1882	1.8
April 9th	15:52-16:07	10	79	532	6.8
April 10th	15:40-16:00	13.6	57	3432	1.4
April 11th	15:32-15:48	20.5	65	7129	2.4
April 12th	15:30-15:44	21.7	70	6243	2.5
April 13th	15:29-15:44	20.4	59	67600	0
April 14th	15:34-15:51	22.5	40	1988	2.3
April 15th	15:32-15:51	20	27	24850	4.3
April 16th	15:35-15:56	22.3	48	16990	5.2
April 17th	15:30-15:49	38.5	45	23212	3.5
April 18th	15:24-15:40	25.1	44	23670	8
April 19th	15:37-15:53	16	34	18580	6,4
April 20th	15:32-15:49	23.1	37	44900	2.4
April 21st	15:32-15:48	16	89	12455	3.1
April 22nd	15:10-15:31	16.4	90	24524	1
April 23rd	15:28-15:45	24	54	42142	2.2

NP Date	Sampling time	TEM	HUM	LI	WS
March 28th	16:12-16:30	15	35	2999	4.2
March 29th	16:07-16:24	18.2	36	2820	2.4
March 30th	15:54-16:09	15.5	38	1037	2
March 31th	15:57-16:12	18	40	4321	1.4
April 1st	16:07-16:24	21	33	4520	1.2
April 2nd	16:02-16:21	20.8	34	6253	3.6
April 3rd	16:03-16:22	22	48	2463	2.2
April 4th	16:04-16:25	24.2	34	5343	2.5
April 5th	16:00-16:21	27.8	28	12241	3.5
April 6th	16:50-17:04	22.6	48	7832	4.6
April 7th	16:00-16:19	25.1	35	35480	2.5
April 8th	16:00-16:16	16.7	58	8140	1.6
April 9th	16:00-16:16	10	79	680	7.5
April 10th	15:40-15:57	14	57	540	1.2
April 11th	15:49-16:06	20.4	65	3975	2.5
April 12th	16:00-16:15	21	70	1079	2.3
April 13th	15:57-16:12	23	59	21220	0
April 14th	16:02-16:21	20.8	40	33570	2.4
April 15th	16:12-16:30	22	27	14270	4.7
April 16th	16:07-16:24	26.1	48	20480	5.2
April 17th	15:54-16:09	30	45	9600	3.7
April 18th	15:57-16:12	26.2	44	52430	9
April 19th	16:00-16:15	16	34	12241	7.2
April 20th	15:57-16:12	25.5	37	42480	2.1
April 21st	16:00-16:20	17	89	35480	3.5
April 22nd	16:12-16:30	19	90	24552	1.5
April 23rd	16:07-16:24	24.6	54	12341	2.5

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WD Date	Sampling time	TEM	HUM	LI	WS
June 24th	18:00-18:25	30	58	2311	0
June 25th	18:10-18:37	32	47	234	1.5
June 26th	19:25-19:53	28.4	74.4	336	0
June 27th	17:35-18:00	30	66.1	3652	0.5
June 28th	19:20-19:37	26.9	38	296	0
June 29th	19:30-19:48	19.6	78	168	3
June 30th	20:15-20:32	27.3	65.4	1	0.6
July 1st	18:55-19:14	28.4	34	1203	0.9
July 2nd	17:17-17:34	29	48	21340	1.8
July 3rd	18:48-19:05	28	34	1864	0.6
July 4th	18:58-19:25	19.1	68	1698	2.5
July 5th	19:02-19:20	29.7	48	2339	0
July 6th	18:54-19:10	32.6	35	2584	1.2
July 7th	18:45-19:03	32.8	58	3212	0.7
July 8th	19:22-19:40	26.4	79	89	0
July 9th	18:48-19:08	29.4	57	2034	0.8
July 10th	18:37-18:55	28.2	65	689	0
July 11th	18:41-19:02	22.1	70	423	2.5
July 12th	18:30-18:50	24.5	59	1864	1.9
July 13th	18:49-19:07	27.1	40	3174	0
July 14th	18:45-19:07	25.4	27	1759	0
July 15th	19:00-19:13	27.6	48	1864	1.6
July 16th	19:13-19:36	27.1	45	545	1.3
July 17th	19:10-19:27	25.6	44	795	0
July 18th	19:03-19:20	25	34	564	0
July 19th	19:25-19:40	26.4	37	419	0
July 20th	19:17-19:34	28.4	89	521	2.6
July 21st	19:07-19:24	19.4	79	685	1.4
July 22nd	18:42-19:00	25.1	54	1464	0

ND Date	Sampling time	TEM	HUM	LI	WS
June 24th	18:32-18:55	20	42.6	270	0.5
June 25th	19:31-19:49	21.7	37.3	580	1
June 26th	20:35-21:00	28	76	3267	0
June 27th	18:36-18:55	31.6	60.5	2249	0
June 28th	20:11-20:23	26.8	71	324	0.6
June 29th	20:01-20:20	19.6	75	29	1.6
June 30th	21:15-21:27	26.4	64.1	22	0
July 1st	19:48-20:05	29	56.5	92	1
July 2nd	18:05-18:23	30.2	61.9	13865	0
July 3rd	19:43-20:03	28.4	63.5	96	0
July 4th	19:23-19:42	19.6	76	103	2.1
July 5th	19:48-20:07	29.6	63.8	86	0
July 6th	19:42-20:00	33.6	47.3	146	0.8
July 7th	19:35-19:52	33.6	51.6	421	3.2
July 8th	20:13-20:30	26.5	86.1	70	0
July 9th	19:40-20:00	29	65.4	126	1.9
July 10th	19:29-19:45	29	68.9	189	1
July 11th	19:46-20:05	18.4	78	201	2.2
July 12th	19:28-19:43	23.8	74.8	523	0
July 13th	19:41-19:56	27.6	62.6	196	0.3
July 14th	19:41-19:57	24.4	86.3	192	0
July 15th	19:45-19:58	28.5	62.4	188	3.2
July 16th	20:08-20:15	28	62.5	31	0
July 17th	20:03-20:12	24.3	77.5	40	0.5
July 18th	19:46-20:00	27.6	67.8	18	0
July 19th	20:10-20:38	24.2	90	12	0
July 20th	20:02-20:20	30.4	55.7	14	2.4
July 21st	20:00-20:19	20.1	69	108	3
July 22nd	19:25-19:40	24.7	82.9	252	0

NP Date	Sampling time	TEM	HUM	LI	WS
June 24th	19:52-20:16	20.4	42.6	670	0.8
June 25th	19:55-20:25	22	37.3	320	1.4
June 26th	20:05-20:30	27.8	75.1	13	0
June 27th	18:05-18:28	31.2	62	6277	1.2
June 28th	19:48-20:02	27.1	69.7	89	0.6
June 29th	18:00-18:20	19.8	70.9	303	2.8
June 30th	20:46-21:10	26.4	66.6	21	0
July 1st	19:25-19:42	29	54.9	786	2.5
July 2nd	17:44-17:56	30.2	60.2	20780	2.6
July 3rd	19:19-19:36	28	64	1223	0
July 4th	19:30-19:46	17.9	88	206	3.3
July 5th	19:28-19:45	30.2	60.9	764	0
July 6th	19:18-19:38	32.7	49.1	1874	1.9
July 7th	19:10-19:28	32.9	53.3	2391	2.3
July 8th	19:50-20:07	26.5	89.1	14	0
July 9th	19:18-19:36	29.7	66.4	1467	1.4
July 10th	19:05-19:24	28.9	70.8	732	0.6
July 11th	19:16-19:33	20.6	71.5	1986	2.5
July 12th	19:01-19:20	24.3	71.8	4345	1.2
July 13th	19:16-19:33	27.7	60.7	1435	0
July 14th	15:10-15:31	25.2	80.8	1261	0
July 15th	19:21-19:37	28.4	62.8	1421	2.2
July 16th	19:40-20:02	28.1	61.9	121	0.9
July 17th	19:36-19:56	24.4	77.7	276	0.4
July 18th	19:25-19:40	26.4	71.7	184	0.7
July 19th	19:49-20:03	24.9	87.8	47	0
July 20th	19:41-19:58	29.9	53.9	87	2.6
July 21st	19:41-19:58	20	88.1	43	2.9
July 22nd	19:08-19:22	24.7	81.3	1159	0

WD I	Date	Sampling time	TEM	HUM	LI	WS
April 1	15th	17:26-18:00	19.3	47.9	19870	0.3
April 1	l 6th	17:27-17:47	21.2	37.6	5935	0.7
April 1	17th	17:18-17:33	20.3	37.6	4476	0.6
April 1	18th	17:27-17:40	22.2	26.7	9492	0.8
April 1	19th	17:20-17:40	18.7	56.3	2324	0.2
April 2	20th	17:32-17:49	25.5	39.5	10810	0.2
April 2	21st	17:02-17:17	19.9	37	2918	0.4
April 2	22nd	17:10-17:30	22.6	29.1	10840	1.5
April 2	23rd	17:10-17:30	24.3	36.9	5249	1.6
April 2	24th	17:35-17:54	25.8	15.3	10610	3.7
April 2	25th	17:34-17:51	24.1	80	525	4
April 2	26th	17:34-17:51	23	26	9410	5.4
April 2	27th	17:31-17:50	21.8	56.8	16720	2.2
April 2	28th	17:32-17:54	19.8	64.2	2389	0.9
April 2	29th	17:32-17:51	23.5	50.7	15120	0.7
April 3	30th	17:29-17:50	27.4	24	27330	1.9
May 1	1st	18:10-18:29	24.9	20.5	3160	2.6
May 2	2nd	17:26-17:46	27.6	13.1	20970	5.7
May 3	3rd	17:29-17:47	23.8	40.2	7865	2.9
May 4	4th	17:26-17:44	22.3	29.5	2847	0.4

ND	Date	Sampling time	TEM	HUM	LI	WS
April	15th	15:32-15:50	20.4	42.6	23670	0.9
April	16th	15:31-15:49	21.7	37.3	18580	1
April	17th	15:33-15:48	20.3	34.8	44900	1.9
April	18th	15:37-15:55	22	23.3	47380	4
April	19th	15:52-16:07	18.3	56.5	9785	0.4
April	20th	15:40-16:00	27.1	34.6	78810	0.4
April	21st	15:32-15:48	20.5	34.1	70510	1.9
April	22nd	15:30-15:44	22.1	28.7	64660	1.2
April	23rd	15:29-15:44	21.9	45.3	34320	2.1
April	24th	15:32-15:51	27.6	14.1	71290	2.4
April	25th	15:35-15:56	22.9	20	6243	1.5
April	26th	15:42-15:59	23.8	24.9	67600	2.4
April	27th	15:34-15:51	21.3	58.4	19880	1.3
April	28th	15:32-15:51	20.6	62.1	24850	1.6
April	29th	15:35-15:56	22	56.1	16990	0.2
April	30th	15:30-15:49	24	29.1	10620	1.2
May	1st	15:24-15:40	26.5	18	108900	2.4
May	2nd	15:37-15:53	28.7	14	114200	5.6
May	3rd	15:32-15:49	29.7	20.2	207300	7.1
May	4th	15:32-15:48	22.7	32.7	64670	2.1

NP]	Date	Sampling time	TEM	HUM	LI	WS
April	15th	16:02-16:20	21.7	39.2	28420	1.5
April	16th	16:00-16:15	25.1	30.9	12700	1.4
April	17th	15:57-16:12	22	30.2	29990	3.2
April	18th	16:00-16:20	24.3	22.7	28110	1.2
April	19th	16:12-16:30	18	57.7	10370	1.1
April 2	20th	16:07-16:24	29	31.6	43680	0.6
April 2	21st	15:54-16:09	21.8	33.2	48200	2.2
April 2	22nd	15:57-16:12	26.3	25.8	62770	2.6
April 2	23rd	15:52-16:09	26	36.2	24630	4
April 2	24th	16:00-16:18	29.9	12	53430	3.5
April 2	25th	16:05-16:23	19.2	22	12241	4.2
April 2	26th	16:07-16:24	23.5	25.2	62480	5.1
April 2	27th	16:02-16:21	21.7	61.5	35480	2.5
April 2	28th	16:03-16:22	20.4	65.3	11840	1.9
April 2	29th	16:04-16:25	23.6	53.4	69680	2.4
April 3	30th	16:00-16:21	26.2	23.6	80240	2.6
May	1st	16:50-17:04	24.4	21.3	39750	6.2
May 2	2nd	16:00-16:19	29.1	13.4	107900	6.4
May :	3rd	16:00-16:16	30.2	19.6	212200	7.6
May	4th	16:00-16:16	23.9	25.4	33570	0.6

A1 bacterial solution sample pre-processing steps.

- 1. Transfer the collected liquid to a centrifuge tube, add 500 μl of Buffer SA, 100 μl of Buffer SC and 0.25 g of grinding beads, mix by vortexing or tissue homogenizer, and then heat lysis at 70°C for 15 min to improve lysis efficiency. 12,000 rpm (~13,400×g) centrifuge for 1 min, transfer the supernatant (about 500 μl) to a new 2 ml centrifuge tube.
- 2. Add 200 μl of Buffer SH, mix well, vortex for 5 sec, and allow to stand at 4°C for 10 min.
- 3. Centrifuge at 12,000 rpm (~13,400 x g) for 3 min, transfer the supernatant to a new 2 ml centrifuge tube, add 500 μl of Buffer GFA (check that isopropanol has been added before use), and mix upside down.
- 4. Add 10 μl of Magnetic Bead Suspension G and mix with shaking for 5 min.
- 5. Place the centrifuge tube on a magnetic rack and let it stand for 30 sec. After the magnetic beads are fully adsorbed, carefully aspirate the liquid.
- Remove the centrifuge tube from the magnetic rack, add 700 µl of Deproteinizing Solution RD (check that anhydrous alcohol has been added before use) and mix for 5 min with shaking.
- 7. Place the centrifuge tube on a magnetic rack and let it stand for 30 sec. After the magnetic beads are fully adsorbed, carefully aspirate the liquid.
- 8. Remove the tube from the magnetic rack, add 700 µl of Rinse Solution PWD (check that anhydrous ethanol has been added before use) and mix for 3 min with shaking.
- 9. Place the centrifuge tube on a magnetic rack and let it stand for 30 sec. After the magnetic beads are fully adsorbed, carefully aspirate the liquid.
- 10. Repeat steps 8 and 9 once.
- 11. Place the tubes on a magnetic rack and allow to air dry at room temperature for 5-10 min. Note: Ethanol residue can inhibit subsequent enzymatic reactions, so make sure that the ethanol evaporates during air drying. However, do not dry the tubes for too long as it may be difficult to elute the DNA.
- 12. Remove the centrifuge tube from the magnetic rack, add 50-100 µl of Elution

- Buffer TB, mix with shaking, place at 56°C, and incubate for 5 min, during which time mix with shaking 3 times, 3-5 times each.
- 13. Place the centrifuge tube on a magnetic rack for 2 min. After the magnetic beads are fully adsorbed, carefully transfer the DNA solution to a new centrifuge tube and store under appropriate conditions.