

Origin, exposure routes and xenobiotics impart nanoplastics with toxicity on freshwater bivalves

Adeline Arini¹, Sandra Muller¹, Véronique Coma², Etienne Grau², Olivier Sandre^{2*}, Magalie Baudrimont^{1*}

¹ Univ. Bordeaux, CNRS, Bordeaux INP, EPOC, UMR 5805, F-33600 Pessac, France

² Univ. Bordeaux, CNRS, Bordeaux INP, LCPO, UMR 5629, F-33600 Pessac, France

*Contact e-mails: olivier.sandre@u-bordeaux.fr (OS), magalie.baudrimont@u-bordeaux.fr (MB)

Electronic Supplementary Information

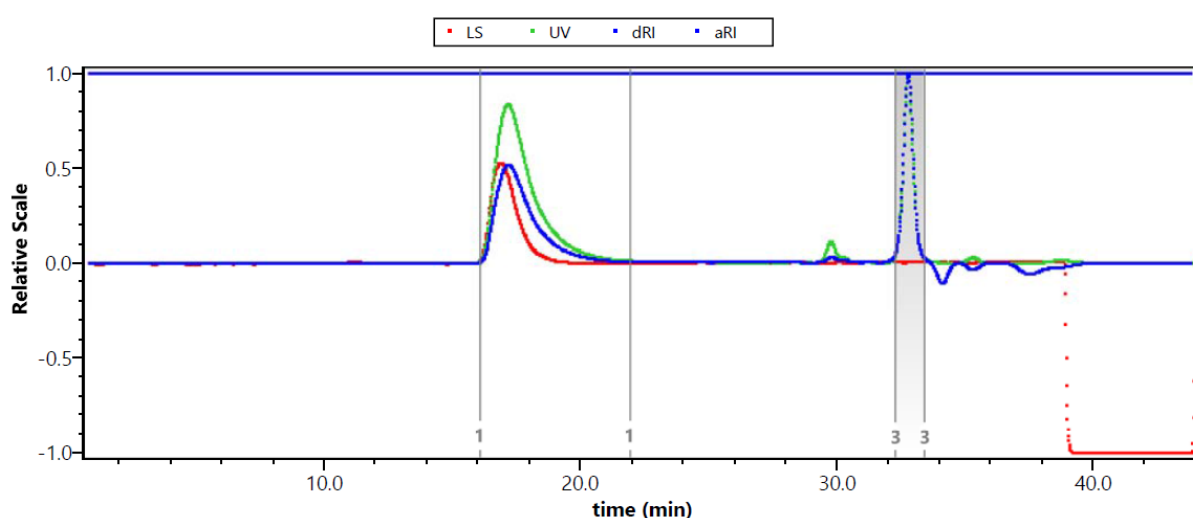


Figure S1: Size exclusion chromatogram of the commercial PS chains used to prepare NP-PS model nanoplastics. The peak 1 on the three detectors (light scattering, UV and differential RI) is assigned to the sample yielding to $M_n=89$ kg/mol, $M_w=196$ kg/mol and $\mathcal{D}=M_w/M_n=2.21$. Peak 3 is the flow marker.

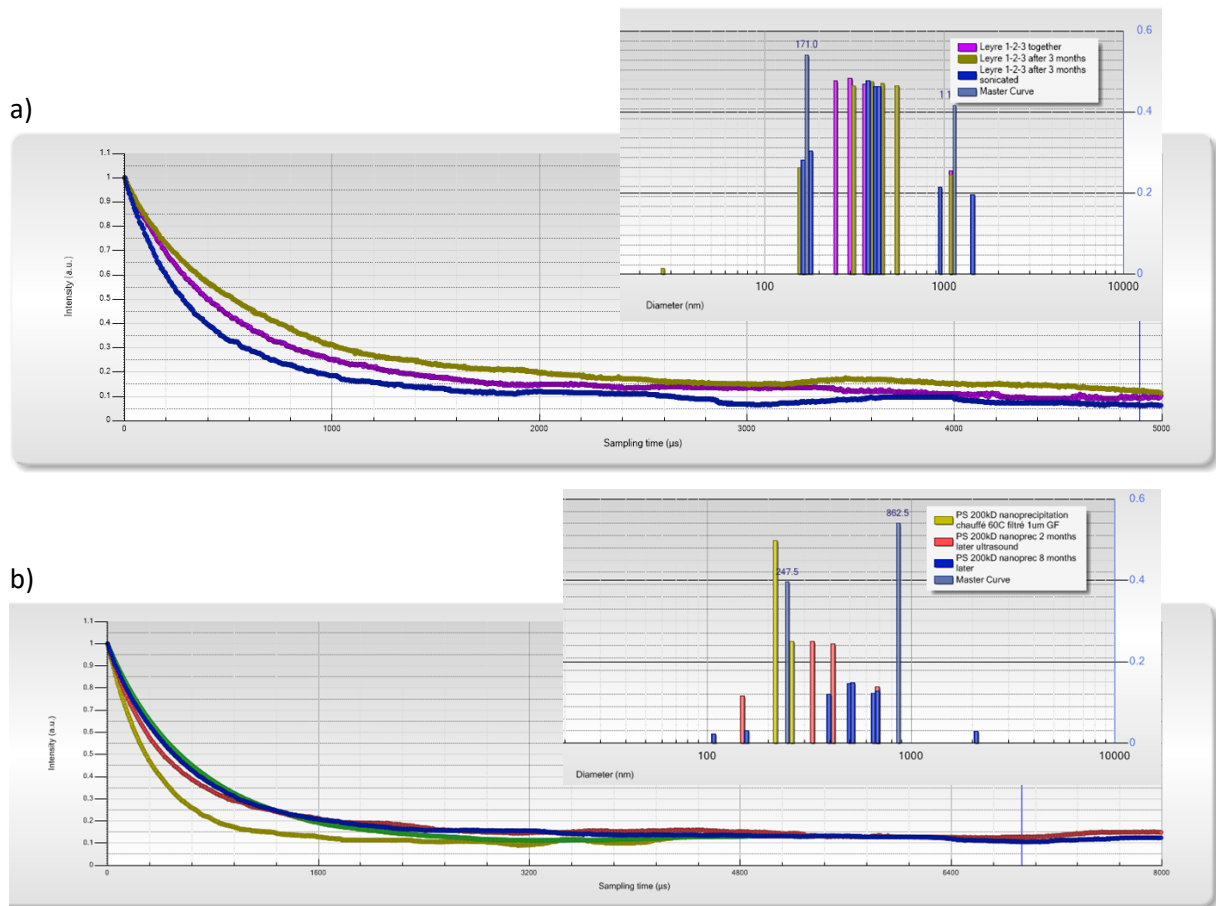
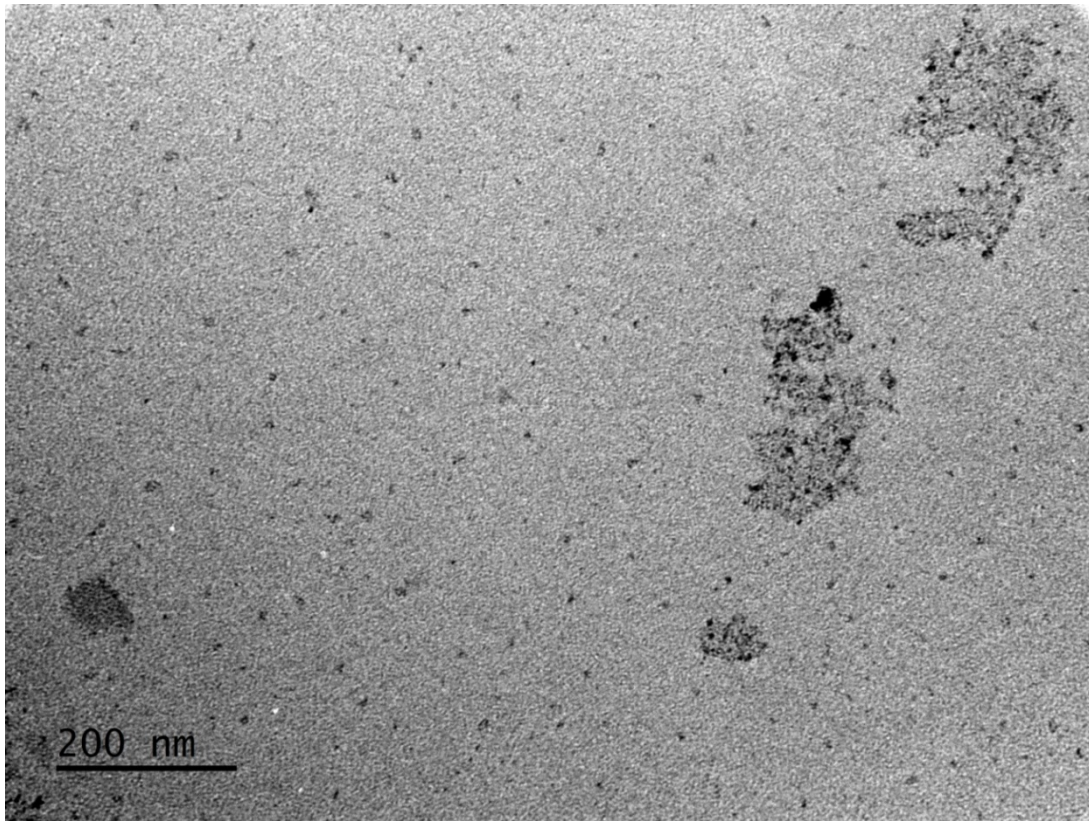
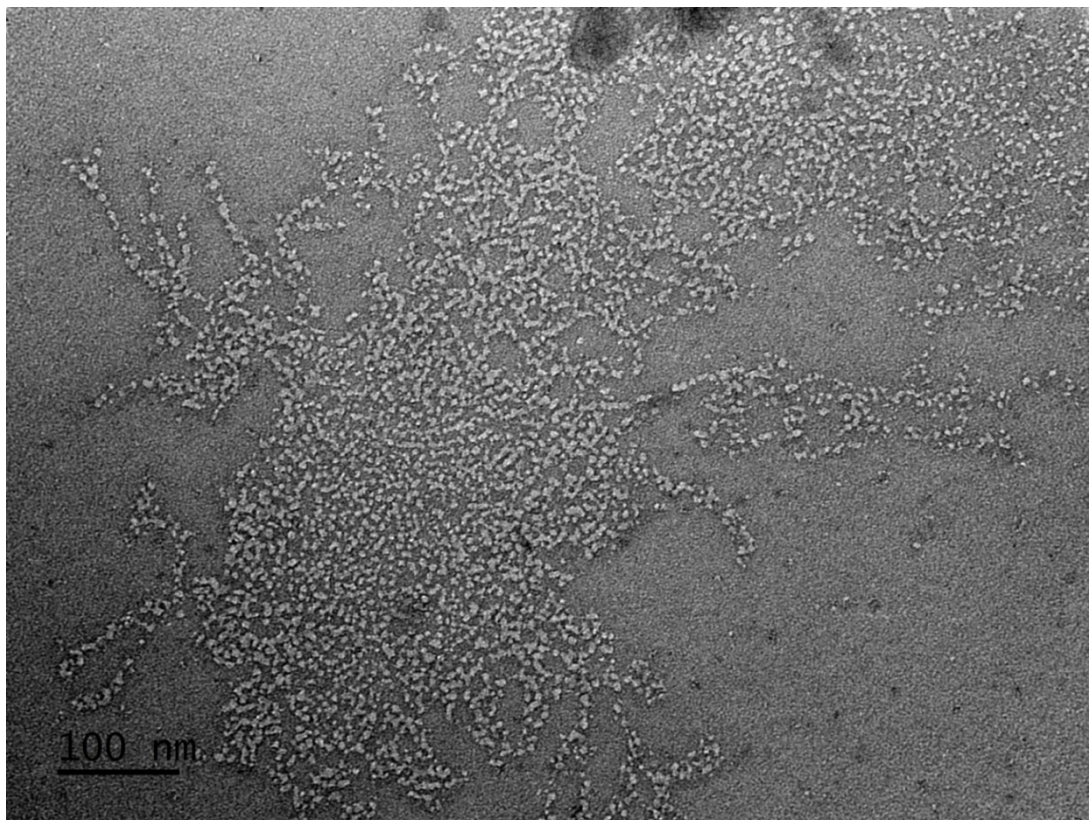


Figure S2: Kinetic study of colloidal stability by DLS (at 165° scattering angle, on a Vasco™ Flex remote-head backscattering DLS instrument, Cordouan Technologies, Pessac, France) of a) NP-L and b) NP-PS during several months after their preparation (when indicated, simple sonication with an ultrasound bath for 5 min was performed, before measurement). Insets show hydrodynamic size histograms obtained by the multimode Pade-Laplace fitting method.



a)



b)

Figure S3: TEM image of a) NP-L and b) NP-PS (with samarium acetate staining specifically hydrophilic regions) showing nanoparticles of undefined morphologies in both cases.

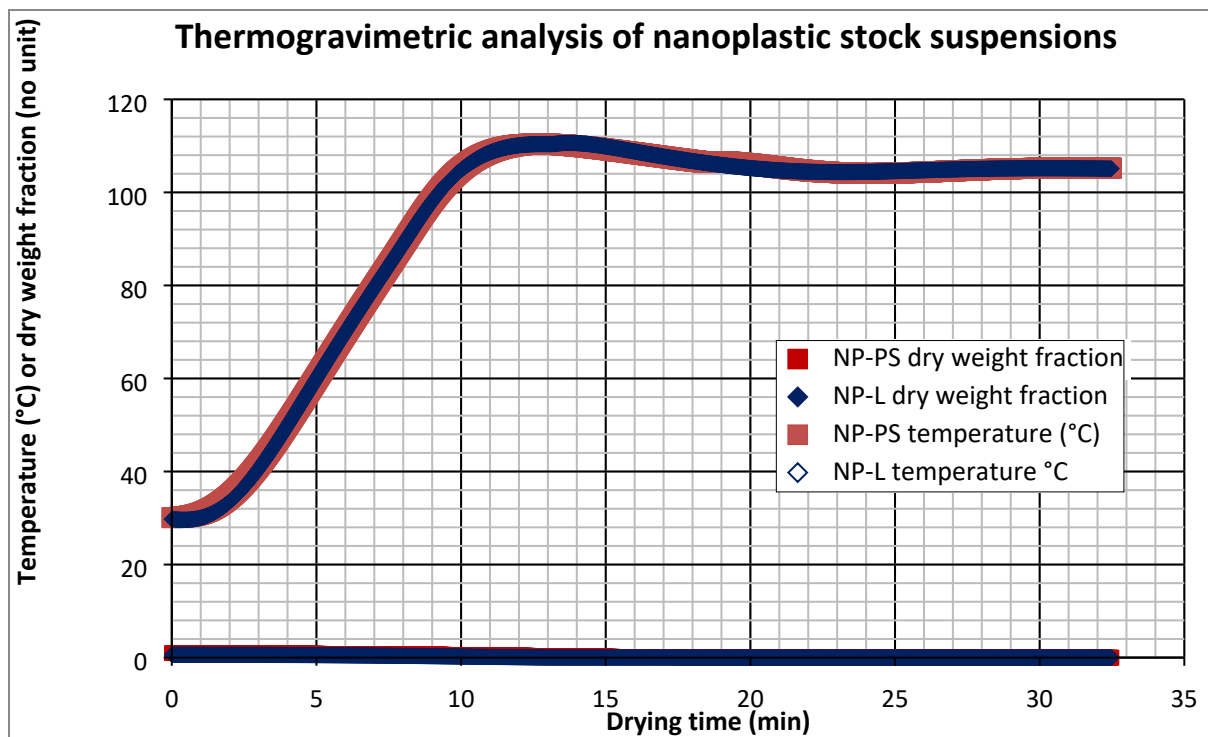


Figure S4: Thermogravimetry analyses (TGA) of a) NP-L and b) NP-PS suspensions yielding the weight concentrations of the stock suspensions: 147 mg/L for NP-L and 420 mg/L for NP-PS.