

1 [Supplementary material]

2 Delineating the Role of Surface Grafting Density of Organic Coatings on
3 the Colloidal Stability, Transport, and Sorbent Behavior of Engineered
4 Nanoparticles

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19 **Experimental methods**

20 **Column experiments.** Column experiments were carried out in water saturated columns packed
21 with Ottawa sand (U.S. Silica, Berkeley Springs, WV). The column system consisted of syringe
22 pump (NE-1000, New Era Pump Systems Inc., Farmingdale, NY), fraction collector (FRAC-100,
23 Pharmacia, Sweden), and glass column (Spectra/Chrom, TX) with dimensions of 15 cm in length
24 and 1.5 cm in inner diameter. Prior to use, the Ottawa sand was cleaned thoroughly using 1 N of
25 hydrochloric acid (HCl) for 24 h and subsequently rinsed several times by DI water to remove
26 remaining HCl until achieving pH 7 and dried at 200°C. The cleaned sand was sieved for size
27 distribution and the 30-40 mesh size fraction ($d_{50} = 0.5075$ mm) of Ottawa sand was used in this
28 study.

29 For column tests, 1 mM KCl solution was first introduced into the column to completely saturate
30 the sand for 5 pore volume (PV) of column. Following saturation, nonreactive tracer tests were
31 conducted to evaluate hydrodynamic properties of the porous media using 5 PV of 1 mM KBr
32 solution buffered to pH 7 with 0.05 mM of NaHCO_3 . Prior to NPs injection, the packed column
33 was equilibrated and conditioned with 5 PV of 1 mM KCl solution in a background electrolyte
34 with 0.05 mM of NaHCO_3 buffer. The 5 PV of solution containing organic coated Mn_xO_y NPs
35 (1 mg L^{-1} as Mn concentration) with 1 mM of KCl was then introduced to the column for NPs
36 transport experiments. The solutions were introduced into the column at 0.76 m/d of a pore-water
37 velocity, in the range of typical values of real groundwater flow.¹⁻³ The column was rinsed with
38 5 PV of 1 mM KCl solution without NPs (NPs-free solutions) at the same flow rate to investigate
39 the release of attached NPs from sand. Column effluent was collected continuously using fraction
40 collector to the tube and analyzed for the manganese concentrations in the samples using ICP-

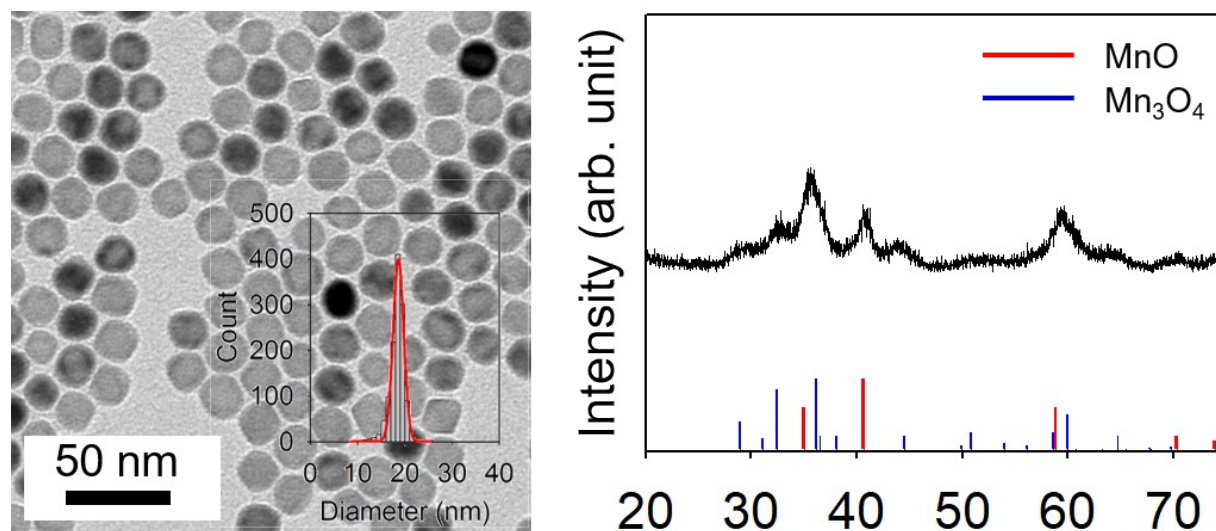
41 OES. After each transport experiment, column were sectioned into 5 segments and retained NPs
42 were extracted from the solid phase using 1 N of HCl solution with sonication. The concentration
43 of NPs in each segment was measured by ICP-OES allowing for retention profiles to be described.

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45 **Soil batch reactor experiments.** Batch reactor experiments were also conducted to evaluate the
46 attachment of NPs to sand media for comparison with column tests. A 5 g of dried Ottawa sand
47 was added to the 10 mL of Mn_xO_y NPs suspensions (1 mg L^{-1} as Mn concentration), prepared in 1
48 mM of KCl as a background electrolyte buffered to pH 7 with 0.05 mM of $NaHCO_3$.⁴ The
49 prepared suspensions were continuously mixed on a rotating rack during 48 h and then let the sand
50 settle for 10 min. The supernatant of solution was collected and the concentration of Mn_xO_y NPs
51 in the supernatant was measured by ICP-OES.

52 References

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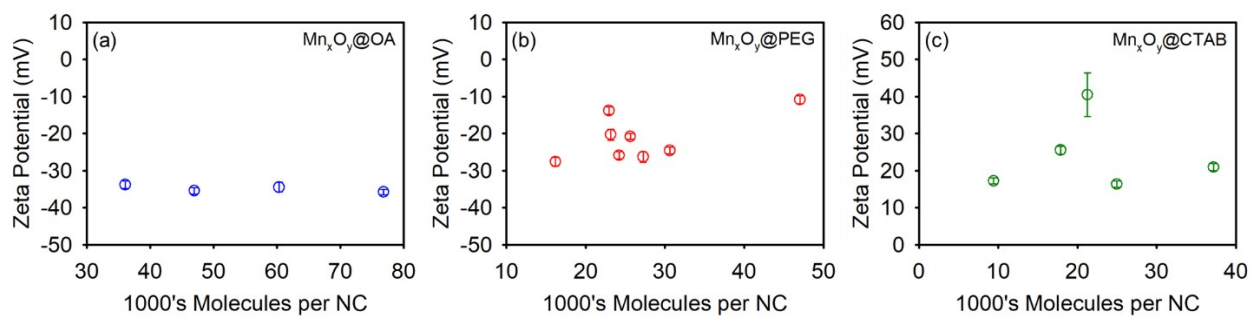
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64 Fig. S1. Highly monodisperse manganese oxide nanoparticles (Mn_xO_y NPs) were precisely
 65 synthesized via manganese oleate (precursor) decomposition in the presence of oleic acid at 320°C.

66 As measured by TEM, the sizes and size distributions of resulting synthesized Mn_xO_y NPs were
 67 18.4 ± 1.5 nm. XRD patterns were matched with MnO (JCPDS Card # 07-0230) and Mn₃O₄

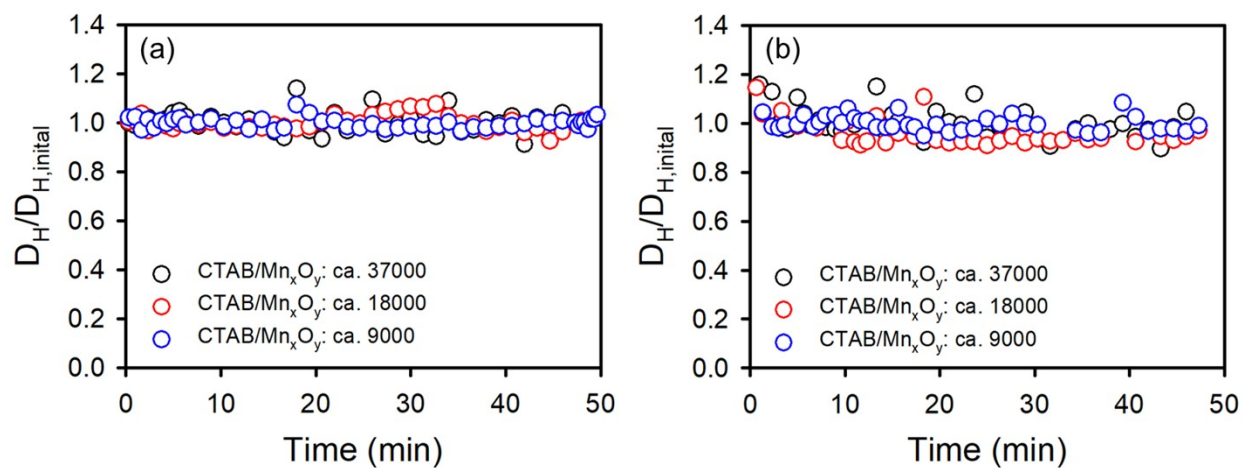
68 (JCPDS Card # 24-0734); the synthesized NPs were well known as MnO core with Mn₃O₄ shell

69 structure.



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71 Fig S2. Zeta potential of the surface functionalized manganese oxide nanoparticles (Mn_xO_y NPs)
 72 with different surface grafting densities at pH 7.0 ± 0.2; (a)Mn_xO_y@OA, (b)Mn_xO_y@PEG, and
 73 (c)Mn_xO_y@CTAB.

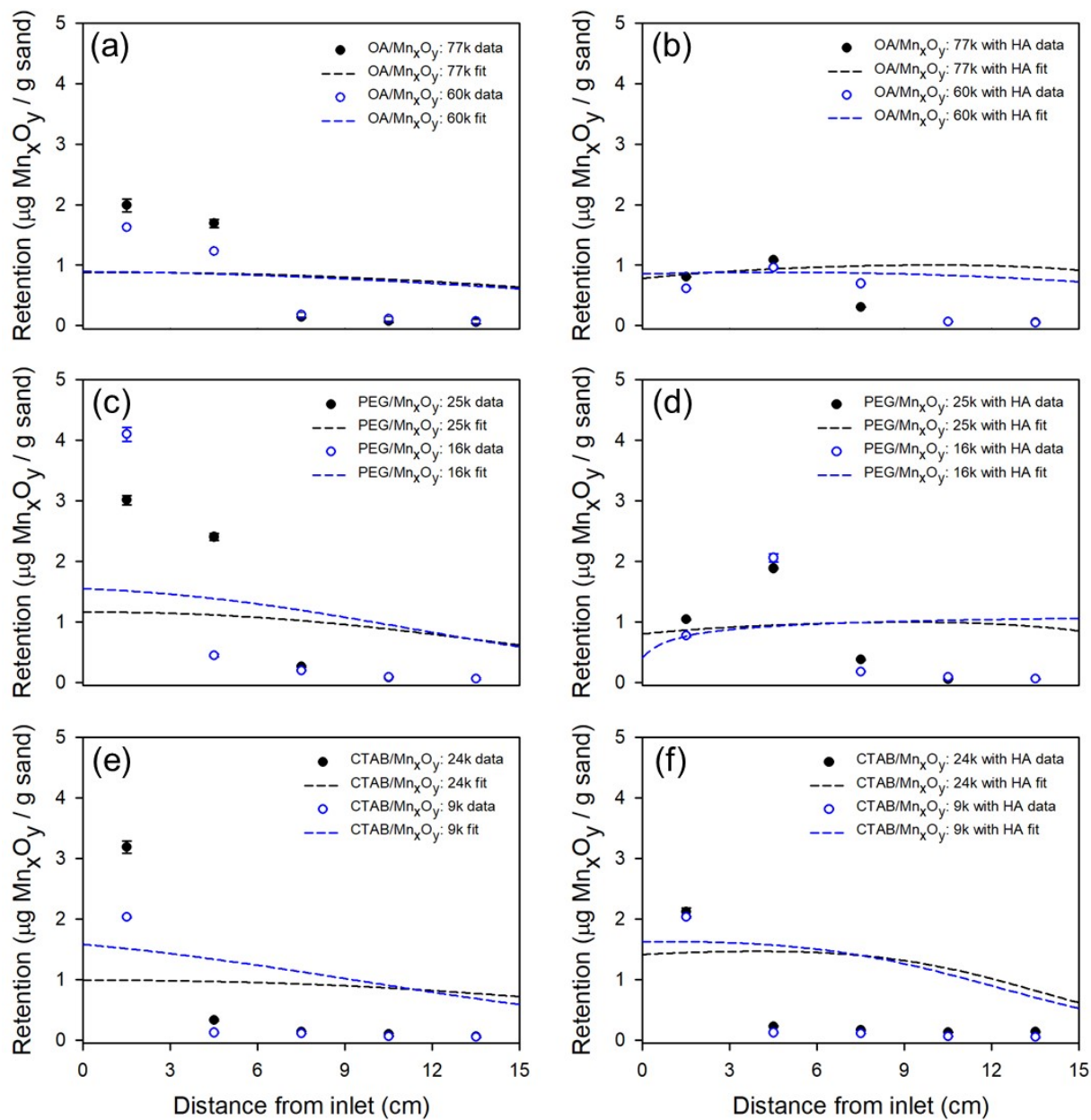


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75 Fig. S3. Aggregation profile of $Mn_xO_y@CTAB$ NPs with various surface grafting densities.

76 Aggregation experiments were performed in the presence of (a) 2000 mM of NaCl and (b) 1000

77 mM of $CaCl_2$ concentrations at pH 7.0 ± 0.2 .



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79 Fig. S4. Experimental and fitted retention profiles of Mn_xO_y NPs with different grafting densities
 80 in water saturated column packed with 30-40 mesh Ottawa sand in the absence and presence of
 81 HA; (a-b) $Mn_xO_y@OA$, (c-d) $Mn_xO_y@PEG$, and (e-f) $Mn_xO_y@CTAB$. All column experiments
 82 were performed at 0.76 m/d of a pore-water velocity and $\text{pH } 7.0 \pm 0.2$.

83 Table S1. Relative concentration (C/C_0) obtained by batch reactor experiments with respect to
84 the surface grafting density of organic coatings. All experiments were conducted at $\text{pH } 7.0 \pm 0.2$.

Surface Stabilizer	Grafting density	Relative Concentration (C/C_0)
OA	77000	0.761 ± 0.01
	60000	0.624 ± 0.01
PEG	25000	0.567 ± 0.01
	16000	0.517 ± 0.01
CTAB	24000	1.088 ± 0.01
	9000	0.851 ± 0.01

85

86 Table S2. Fitted parameters obtained from NP BTC data using MFT model.

Column identifier	k_{att} (h ⁻¹)	k_{det} (h ⁻¹)	S_{max} (μg/g sand)
OA/Mn _x O _y :77k	1.386 ± 0.23	0.016 ± 0.01	1.495 ± 0.26
OA/Mn _x O _y :60k	1.200 ± 0.12	0.015 ± 0.01	1.556 ± 0.32
OA/Mn _x O _y :77k with HA	3.068 ± 0.85	0.027 ± 0.01	1.500 ± 0.16
OA/Mn _x O _y :60k with HA	1.545 ± 0.31	0.019 ± 0.01	1.494 ± 0.24
PEG/Mn _x O _y :25k	1.795 ± 0.29	0.016 ± 0.01	1.970 ± 0.44
PEG/Mn _x O _y :16k	1.820 ± 0.25	0.012 ± 0.00	2.488 ± 0.59
PEG/Mn _x O _y :25k with HA	3.361 ± 1.00	0.024 ± 0.01	1.455 ± 0.18
PEG/Mn _x O _y :16k with HA	30.125 ± 21.49	0.050 ± 0.02	1.230 ± 0.03
CTAB/Mn _x O _y :24k	1.452 ± 0.22	0.011 ± 0.01	1.410 ± 0.16
CTAB/Mn _x O _y :9k	1.426 ± 0.14	0.007 ± 0.00	2.425 ± 0.46
CTAB/Mn _x O _y :24kwith HA	3.317 ± 0.85	0.014 ± 0.01	2.005 ± 0.39
CTAB/Mn _x O _y :9k with HA	2.841 ± 0.55	0.010 ± 0.00	2.160 ± 0.41

87 Values after '±' indicates 95% confidence limits of fitted parameters.