

SUPPLEMENTARY FILE

Waste-derived Carbon Nanodots for Fluorimetric and Simultaneous Electrochemical Detection of Heavy Metals in Water

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Table S3. Comparison of different fluorescent carbon nanodots derived from green sources for Hg^{2+} and Pb^{2+} photoluminescence (PL) detection

Table S4. Comparison of various carbon nanodots derived from green sources as electrochemical sensors for heavy metal ions

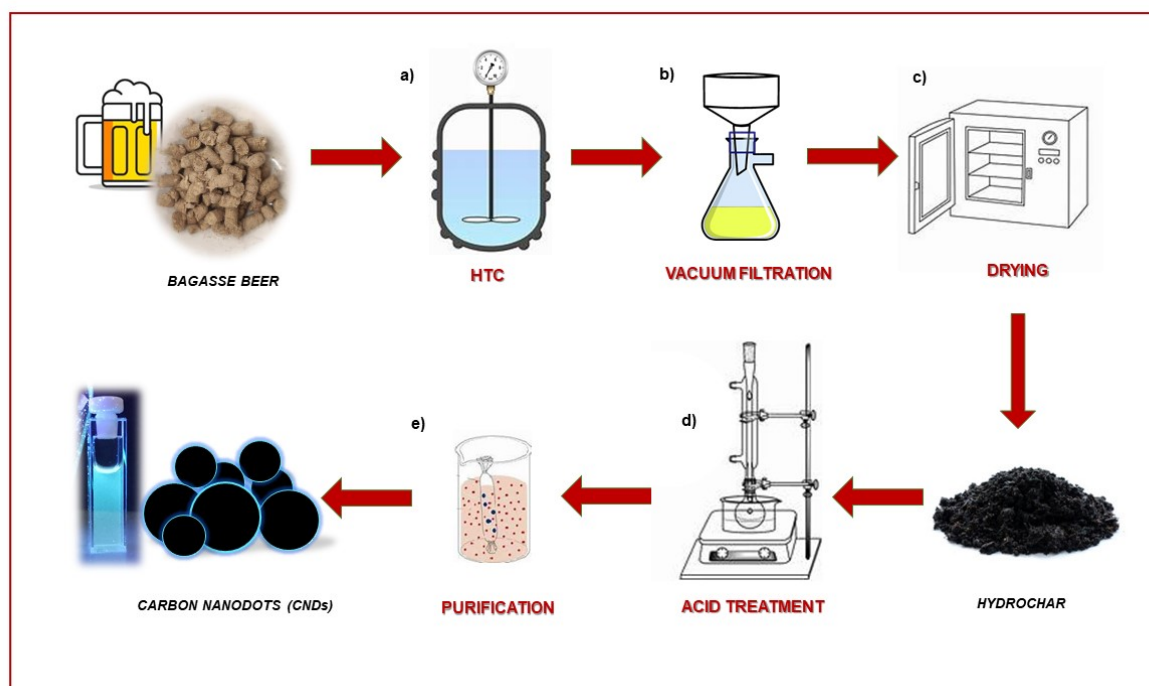
Sample (time-Temperature)

Hydrochar yields* (wt%) \pm sd

| | |
|--------------------------------|--------------|
| Hydrochar _(60–180) | 15.35 ± 0.88 |
| Hydrochar _(60–210) | 18.27 ± 1.11 |
| Hydrochar _(60–240) | 24.27 ± 1.32 |
| Hydrochar _(90–180) | 22.51 ± 0.95 |
| Hydrochar _(90–210) | 31.09 ± 1.81 |
| Hydrochar _(90–240) | 38.55 ± 1.02 |
| Hydrochar _(120–180) | 18.05 ± 2.44 |
| Hydrochar _(120–210) | 23.52 ± 1.53 |
| Hydrochar _(120–240) | 27.10 ± 3.08 |

*hydrochar yield was calculated as follow (1):
$$\text{yield (wt\%)} = \frac{\text{g of product}}{\text{initial g of dry BB}} \times 100$$

Table S1. BB-hydrochar yields



Scheme S1. Pathways involved in the preparation of CNDs from BB

The FTIR spectra of the feedstock and its hydrochar showed absorption peaks characteristic of lignocellulosic biomasses (Figure S1)(2). A strong broad band observed at 3300 cm⁻¹ is attributed to O–H stretching of cellulose hydroxyl groups. The weak absorption peaks between 2900–2850 cm⁻¹ were due to C–H stretching of the methylene groups (3). The peak at 1640 cm⁻¹ suggests the presence of amino groups, and the band between 1500 cm⁻¹ and 1160 cm⁻¹ correspond to the vibrations of C–

O–C and C–N (3). The strong peak observed at 1030 cm^{-1} was attributed due to C–O stretching. The absorption peak detected at 550 cm^{-1} was due to β -glycosidic bond between sugar monomers (2).

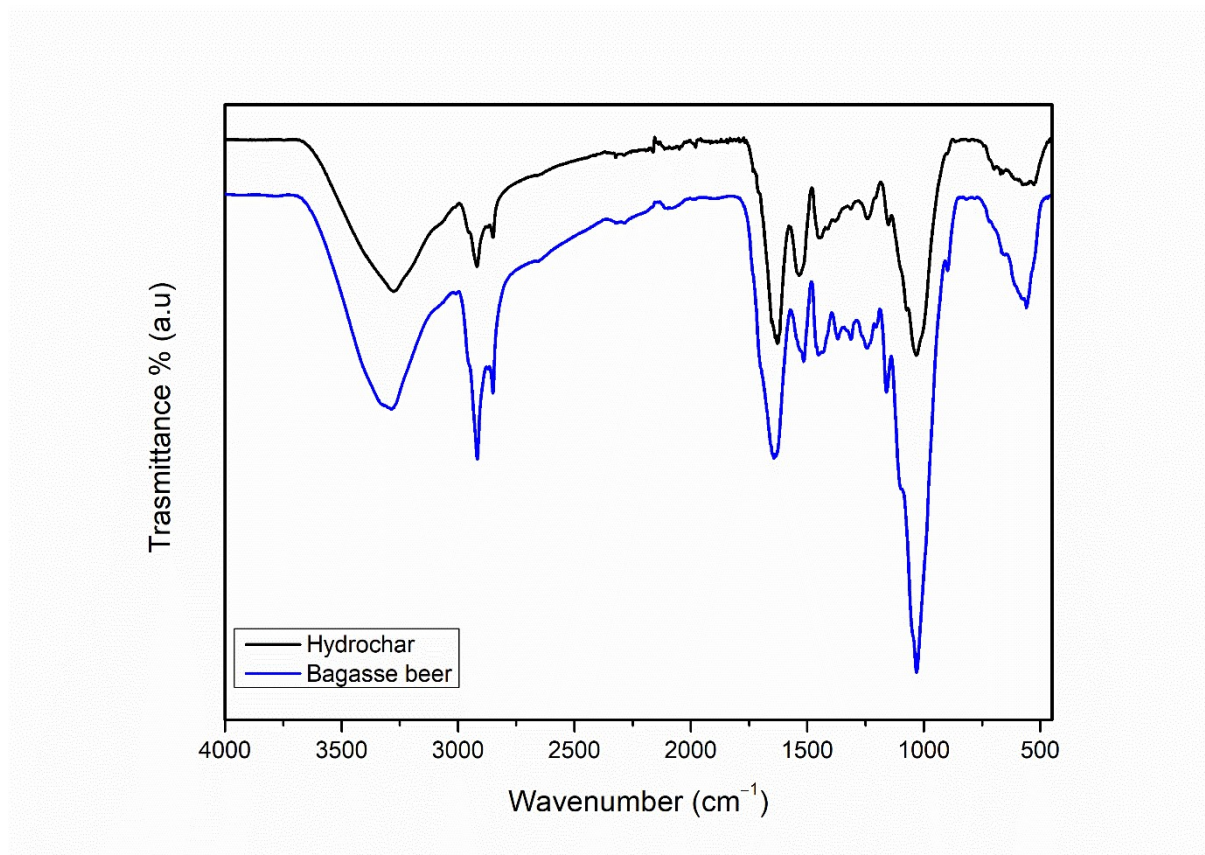


Figure S1. FTIR spectra of Bagasse beer (blue) and Hydrochar (black)

The DLS measurements were carried out in order to verify the size distribution profile and the presence of small particles in the raw materials. The DLS data (Figure S2) showed single size population centred 615 nm for the BB and two populations around 100 nm and 500 nm were observed for the hydrochar: these results suggest that the hydrothermal process helps to obtain nanomaterials but not smaller than 10 nm, therefore, subsequent treatments and purification processes were necessary to reduce the size and minimize the formation of these agglomerated clusters.

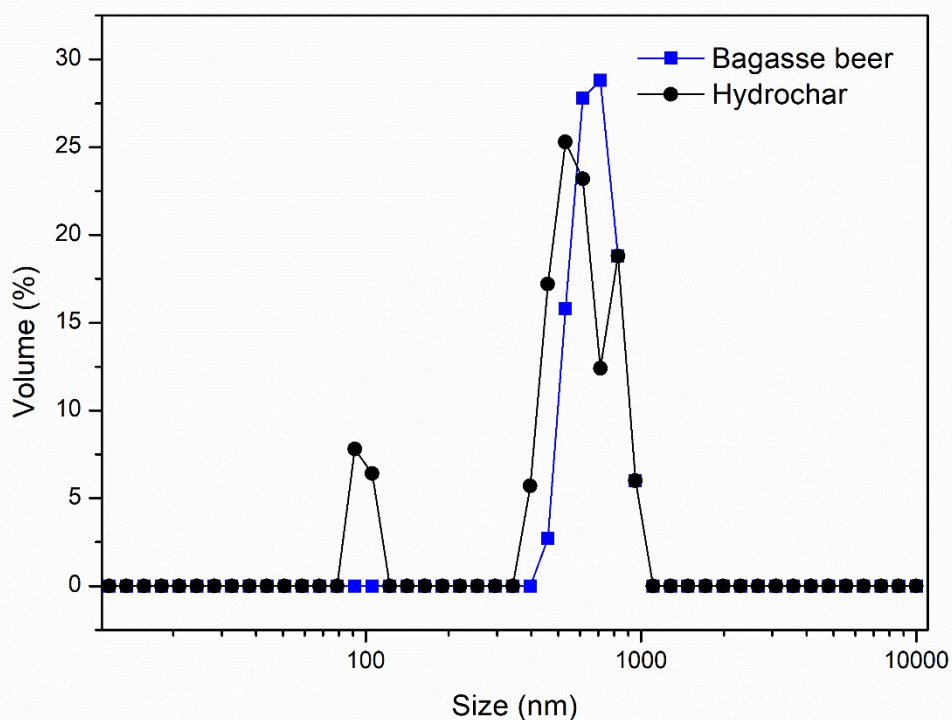


Figure S2. DLS distribution dimensions particles of bagasse beer (blue) and hydrochar (black)

The XRD spectra (Figure S3) show the crystalline cellulose peaks at 16 (101), 22.3 (002), and 35 (040) 2θ degrees, which are more pronounced in the starting material than in the hydrochar. This indicates that the hydrothermal treatment leads to a complete transformation of the "crystalline phase" to the "amorphous phase", considering the disappearance of the peak at 16 2θ (degree) (4). This phenomenon can also be attributed to the well-packed long chains of lignocellulose components in BB, which are characterized by strong hydrogen bonds(4). These hydrogen bonds facilitate the preservation of the sugar rings, which is further promoted by the depolymerisation and hydrolysis reactions occurring during the hydrothermal carbonization process(3).

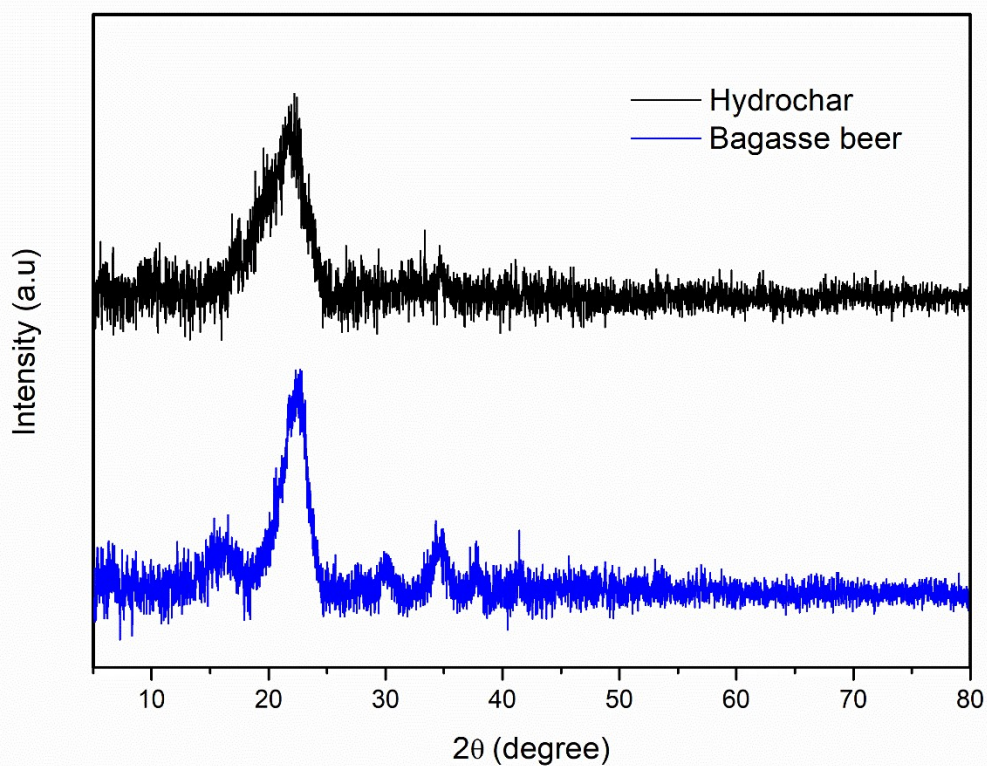


Figure S3. XRD spectra of bagasse beer (blue) and hydrochar (black) after baseline correction

| Sample | C% \pm sd | N% \pm sd | H% \pm sd | S% | O% ^a |
|--------|------------------|-----------------|-----------------|----|-----------------|
| CNDs | 51.33 \pm 1.05 | 6.84 \pm 0.21 | 6.01 \pm 0.25 | 0 | 35.82 |

^acalculated by difference

Table S2. Results of CHNS elemental analysis of CNDs

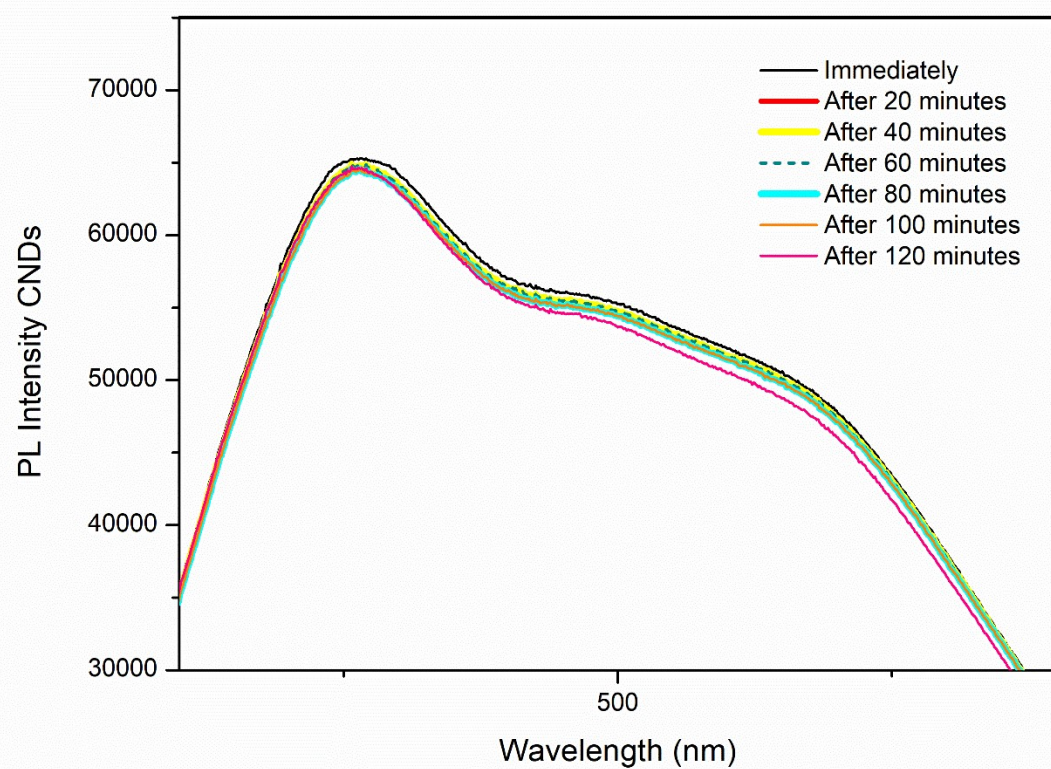


Figure S4. Photostability of CNDs

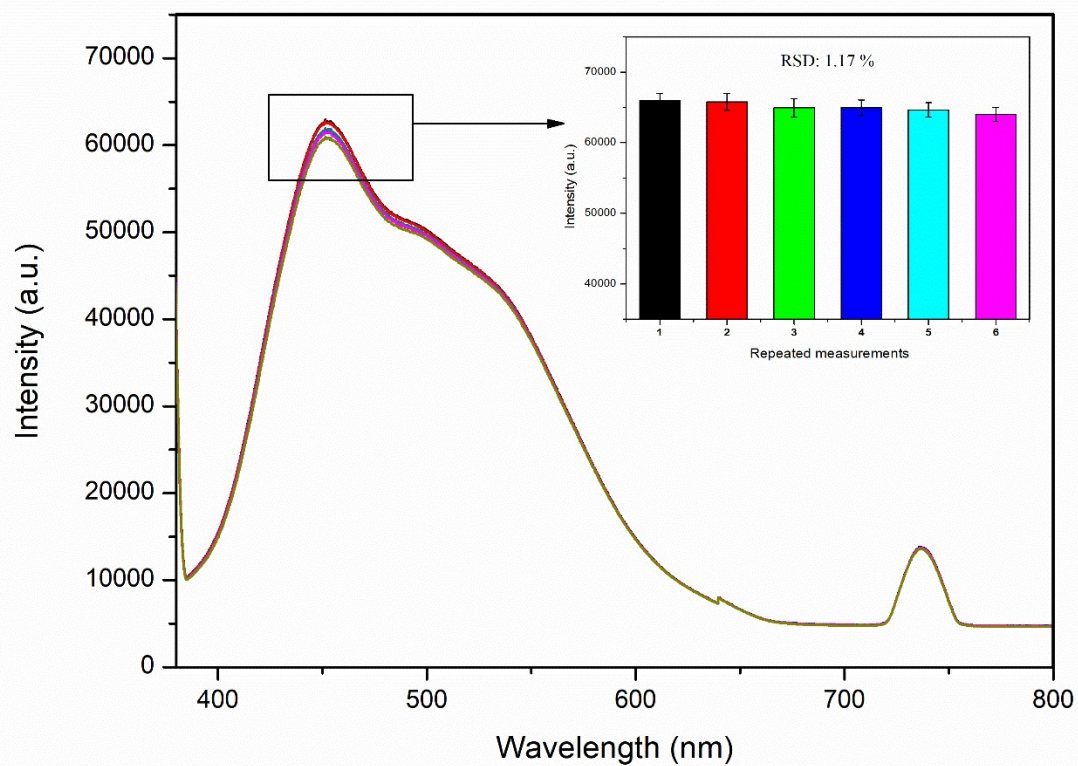


Figure S5. PL Measurement repeatability

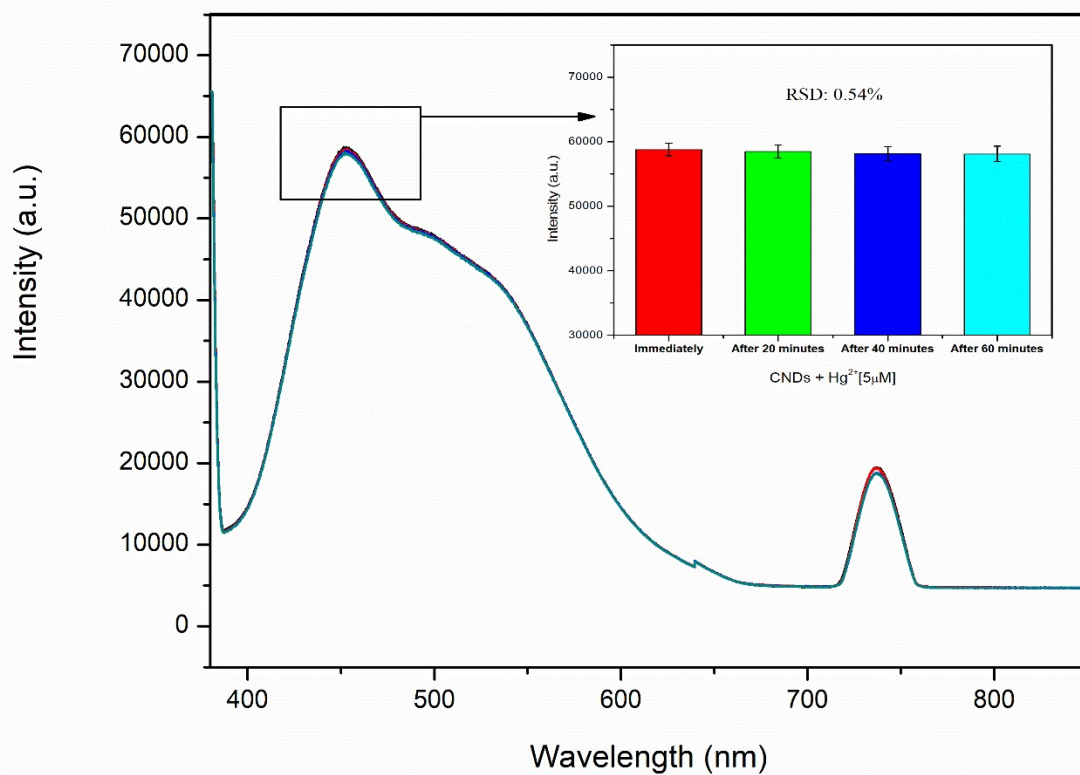


Figure S6. Stability PL response of CNDs with 5 μM of Hg^{2+} monitored after 20 minutes, 40 minutes and 60 minutes

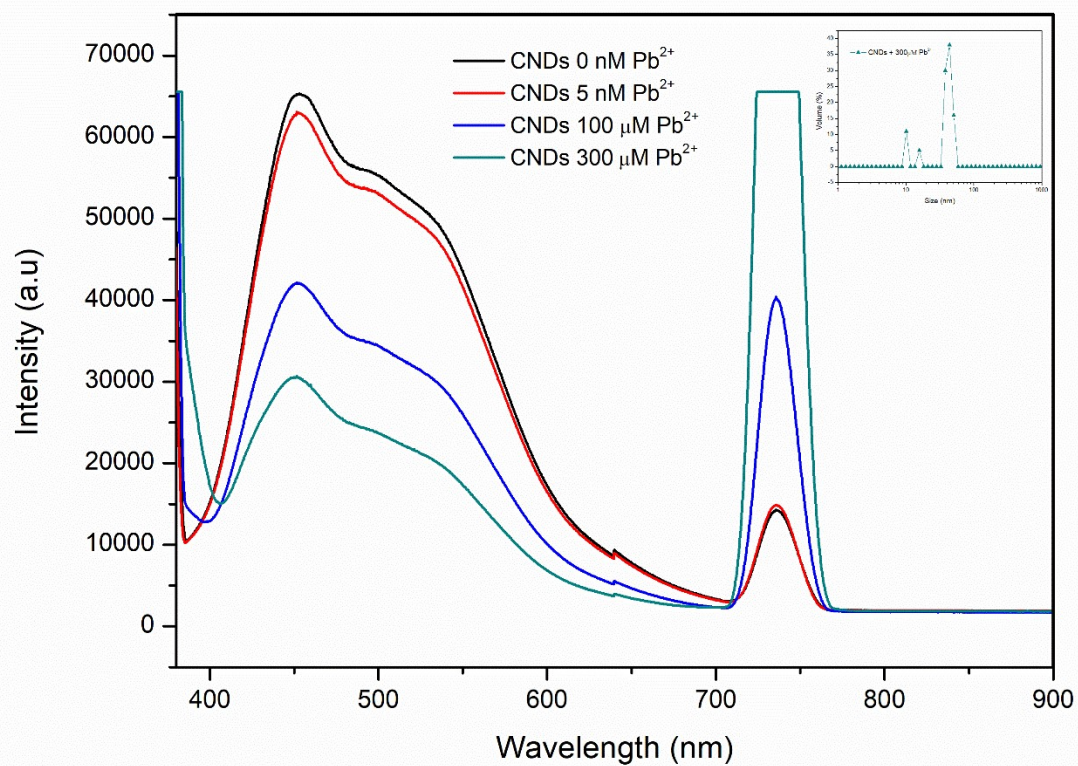


Figure S7. Second harmonic PL CNDs at different addition of Pb^{2+} and (inset) related increasing of CNDs aggregation after 300 μM of Pb^{2+} addition revealed by DLS measurement

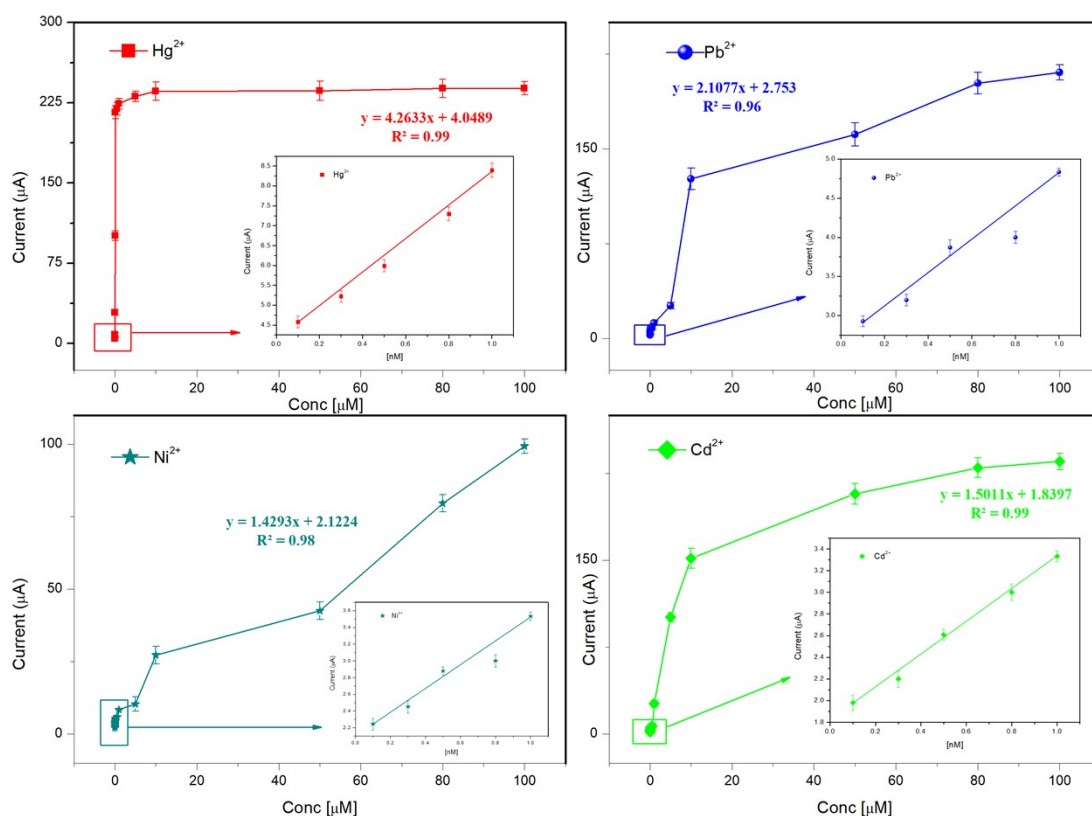


Figure S8. CNDs/SPCE response to Hg^{2+} , Pb^{2+} , Ni^{2+} and Cd^{2+} addition in the studied concentration range [0.1 nM to 100 μM]; insets: calibration graphs for peak current at low concentrations [0.1 to 1 nM] and related fit linear equations ($\text{sd} \leq 3$)

Table S3. Comparison of different fluorescent carbon nanodots derived from green sources for Hg^{2+} and Pb^{2+} photoluminescence (PL) detection

| Samples | Heavy metal ions detected | Detection techniques | LOD (μM) | Sensitivity ($\mu\text{A nM}^{-1} \text{cm}^{-2}$) | Ref. |
|---------------------------------------|---------------------------|----------------------|-----------------------|--|-----------|
| CNDs-bagasse beer derived | Hg^{2+} | PL | 0.011 | 883.44 | This work |
| | Pb^{2+} | PL | 0.079 | 126.64 | This work |
| CNDs-fruit waste derived | Hg^{2+} | PL | 55 | - | (5) |
| CNDs-fruit waste derived | Hg^{2+} | PL | 0.33 | - | (6) |
| CNDs- Pigeon eggs or feathers derived | Hg^{2+} | PL | 0.035 | - | (7) |
| | | PL | 0.039 | - | |
| | | PL | 0.010 | - | |

| | | | | | |
|------------------------------------|------------------|----|--------|---|------|
| CNDs-leaves derived | Pb ²⁺ | PL | 0.0006 | - | (8) |
| CNDs-citric acid derived | Hg ²⁺ | PL | 0.226 | - | (9) |
| CNDs-Microalgae derived | Pb ²⁺ | PL | 0.01 | - | (10) |
| CNDs-glutathione+formamide derived | Hg ²⁺ | PL | 0.039 | - | (11) |
| | Pb ²⁺ | PL | 0.037 | - | |
| CNDs-coconut milk derived | Hg ²⁺ | PL | 0.016 | - | (12) |

Table S4. Comparison of various carbon nanodots derived from green sources as electrochemical sensors for heavy metal ions detection

| Samples | Heavy metal ions detected | Detection techniques | LOD | Sensitivity ($\mu\text{AnM}^{-1}\text{cm}^{-2}$) | Ref. |
|---------------------------------|---------------------------|----------------------|-----------------------|--|-----------|
| CNDs-bagasse beer derived | Hg ²⁺ | SWV | 0.124 $\mu\text{g/L}$ | 34.1 | This work |
| | Pb ²⁺ | SWV | 0.551 $\mu\text{g/L}$ | 21.3 | This work |
| | Cd ²⁺ | SWV | 0.453 $\mu\text{g/L}$ | 32.3 | This work |
| | Ni ²⁺ | SWV | 0.608 $\mu\text{g/L}$ | 11.4 | This work |
| CNDs-activated carbon | Pb ²⁺ | SWASV ^a | 0.7 $\mu\text{g/L}$ | 100.37 | (13) |
| | Cd ²⁺ | SWASV | 0.4 $\mu\text{g/L}$ | 109.45 | |
| CNDs-leaves derived | Pb ²⁺ | DPASV ^b | 0.14 nM | - | (14) |
| GQDs ^c | Pb ²⁺ | SWASV | 8 $\mu\text{g/L}$ | 100.37 | (15) |
| | Cd ²⁺ | SWASV | 11.30 $\mu\text{g/L}$ | 109.45 | |
| CNDs-pomelo peels waste derived | Pb ²⁺ | SWASV | 12 nM | 0.11 | (16) |
| | Cd ²⁺ | SWASV | 13 nM | 0.11 | |

^aSWASV: Square Wave Anodic Stripping Voltammetry; ^bDPASV: Differential Pulse Anodic Stripping Voltammetry; ^cGQDs: Graphene Quantum Dots.

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