

1                    Supporting Information for:  
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4                    Direct non-target analysis of dissolved organic matter and disinfection  
5                    by-products in drinking water with nanoLC-FT-ICR-MS

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15 [Text S1: Drinking water samples](#)

16 The drinking water treatment plants A and B are operating with 38.000 m<sup>3</sup>/d and 25.000 m<sup>3</sup>/d capacity,  
17 respectively. Both DWTPs receive water from a nearby river via bank filtration. The water treatment  
18 process in both plants are, in short: A) bank filtration, mechanical deacidification/aeration, sand filtration,  
19 and disinfection with chlorine (Cl<sub>2</sub>) gas; B) bank filtration, mechanical deacidification/aeration,  
20 flocculation, sand filtration, and disinfection with chlorine dioxide (ClO<sub>2</sub>). After disinfection, both DWTPs  
21 directly supply the drinking water to consumers.

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23 [Table S1](#). Chemical parameters of the drinking water samples used in this study.

DWTP		DOC (mg/L)	Chlorine (mg/L)
DWTP-A	Before disinfection	2.1	ND*
	After disinfection	2.0	0.2
DWTP-B	Before disinfection	2.5	ND*
	After disinfection	2.7	ND*

24 \* ND: Not detected (below detection limit).

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26      Table S2. Chemical properties and retention time (RT) of model compounds on the C18 column used  
 27      in this study.

#	Compound	Concentration (ppm)	Formula	Molecular mass (Da)	Log D (pH3)	RT (min)	Company
1	D-Glucuronic acid	0.2	C <sub>6</sub> H <sub>10</sub> O <sub>7</sub>	194.14	-2.82	7.5-8.0	Sigma-Aldrich
2	2-(4-(2,2-Dicarboxyethyl)-2,5-dimethoxybenzyl)malonic acid	0.04	C <sub>16</sub> H <sub>18</sub> O <sub>10</sub>	370.31	0.01	15.6-16.2	Sigma-Aldrich
3	Isoferulic Acid 3-O- $\beta$ -D-Glucuronide	0.04	C <sub>16</sub> H <sub>18</sub> O <sub>10</sub>	370.31	-0.65	16.9-17.5	Toronto Research Chemicals
4	Vanillic acid	4.7	C <sub>8</sub> H <sub>8</sub> O <sub>4</sub>	168.15	1.14	16.3-16.9	Sigma-Aldrich
5	Fraxin	0.04	C <sub>16</sub> H <sub>18</sub> O <sub>10</sub>	370.31	-1.25	18.2-18.7	Sigma-Aldrich

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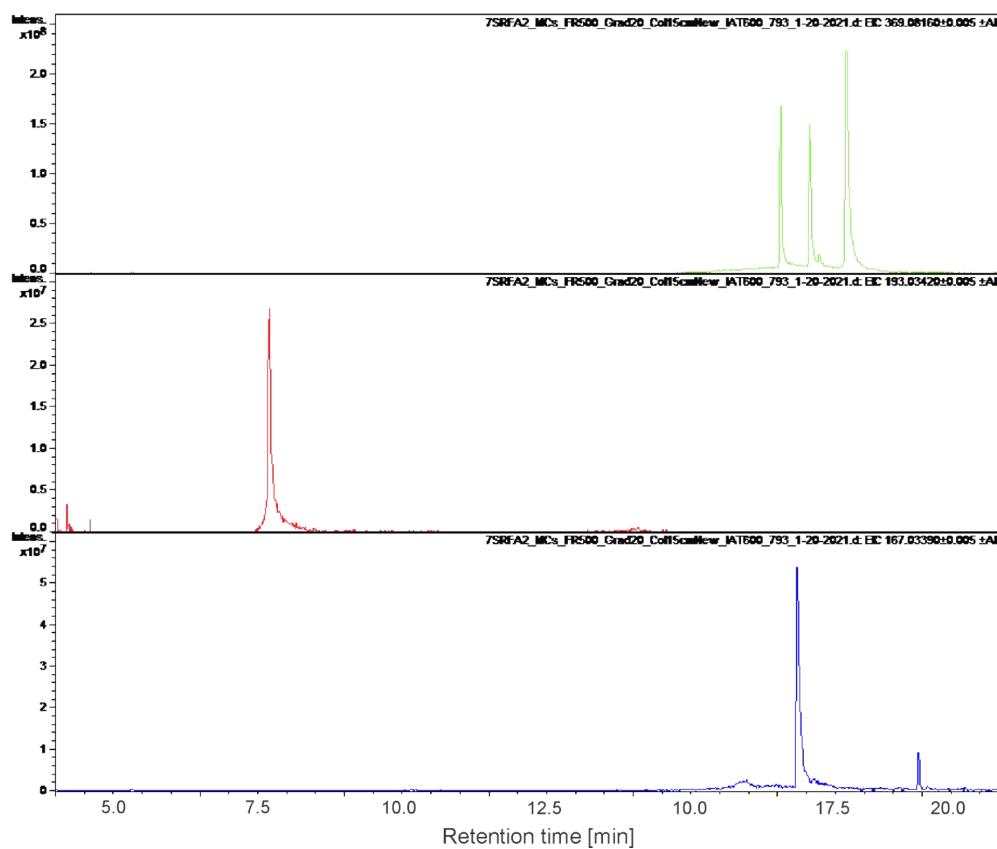
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**Table S3.** The gradient program used in the nanoLC system. The flow rate was set to 500 nL/min.

Time (min)	MeOH (v/v %)	ultrapure water (v/v %)
0	1	99
0.5	1	99
10.5	99	1
30.5	99	1
65	1	99

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35 **Figure S1.** Extracted ion chromatograms (EICs) of model compounds D-Glucuronic acid (red),  
36 Vanillic acid (blue), and 3 isomeric compounds (green: 2-(4-(2,2-Dicarboxy-ethyl)-2,5-  
37 dimethoxy-benzyl)malonic acid, Isoferulic Acid 3-O- $\beta$ -D-Glucuronide, and Fraxin) using the  
38 nanoLC method.

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41      **Table S4.** Comparison of the chromatographic parameters for three isomeric compounds using  
42      HPLC (<sup>1</sup>) and nanoLC (this study) methods.

Compounds	k-value* on nanoLC column <sup>#1</sup>	k-value* on RPLC column <sup>#2</sup>
2-(4-(2,2-Dicarboxy-ethyl)-2,5-dimethoxy-benzyl)malonic acid	1.1	2.3
Isoferulic Acid 3-O- $\beta$ -D-Glucuronide	1.3	2.4
Fraxin	1.5	2.7

43      \* Retention factor (k-values) are calculated by:  $k = (t_R - t_0) / t_0$  with  $t_R$ : retention time of the  
44      analyte and  $t_0$ : column dead time.

45      <sup>#1</sup> Acclaim PepMap, 75  $\mu$ m x 150 mm, 3  $\mu$ m, 100 Å, Thermo Fisher Scientific, USA; flow rate  
46      500 nL/min.

47      <sup>#2</sup> ACQUITY HSS T3, 1.8  $\mu$ m, 100 Å, 150x3mm, Waters, USA; flow rate 0.2 mL/min.

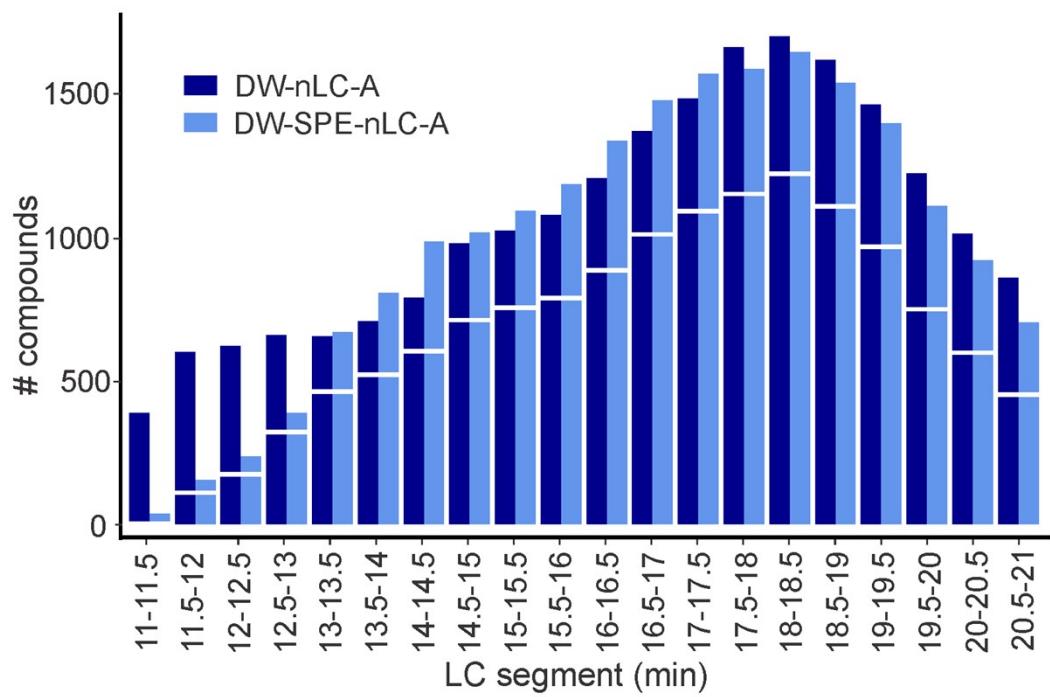
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Table S5. Mass resolving power at m/z 400 in samples DW-nLC-A and DW-SPE-nLC-A in different segments.

	DW-nLC-A	DW-SPE-nLC-A
Segment (min)	Mass resolving power at m/z 400	
11-11.5	278963	NA
11.5-12	287997	359793
12-12.5	253787	288076
12.5-13	270668	273120
13-13.5	291577	261661
13.5-14	288383	272381
14-14.5	278164	265052
14.5-15	268308	263016
15-15.5	267877	278540
15.5-16	275574	280936
16-16.5	263983	268291
16.5-17	266621	279894
17-17.5	275722	276978
17.5-18	273465	277444
18-18.5	270342	266645
18.5-19	270685	260556
19-19.5	268027	275142
19.5-20	256972	274119
20-20.5	262094	260818
20.5-21	274556	271424

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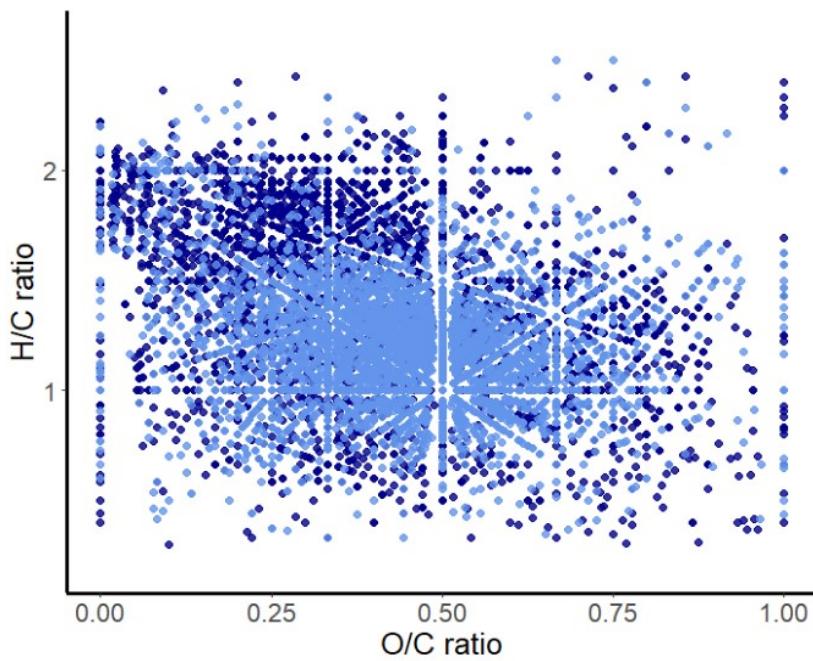


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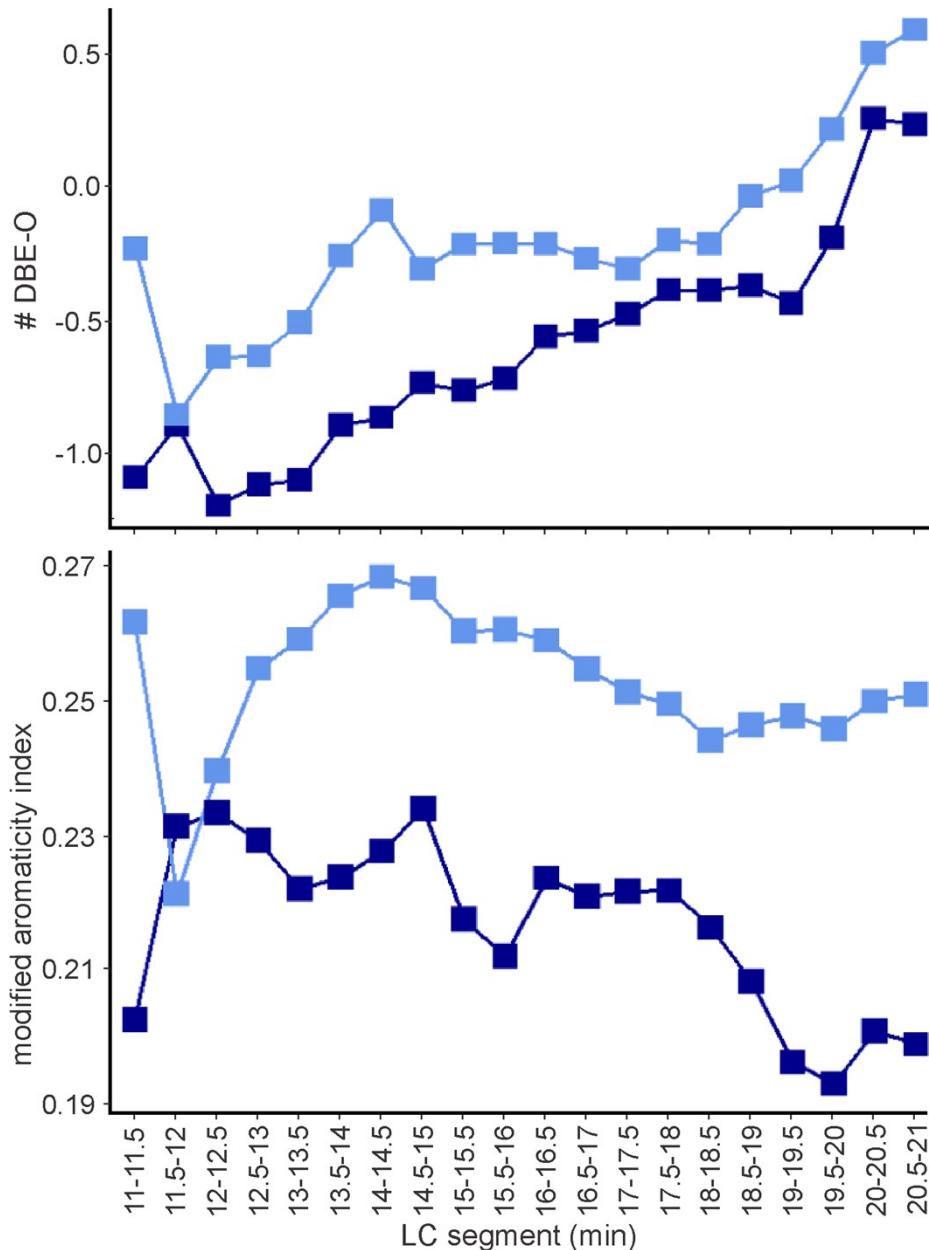
Figure S2. Total number of DOM compounds detected in DW-nLC-A and DW-SPE-nLC-A. The white lines represent the number of shared compounds between both samples in the respective segment.

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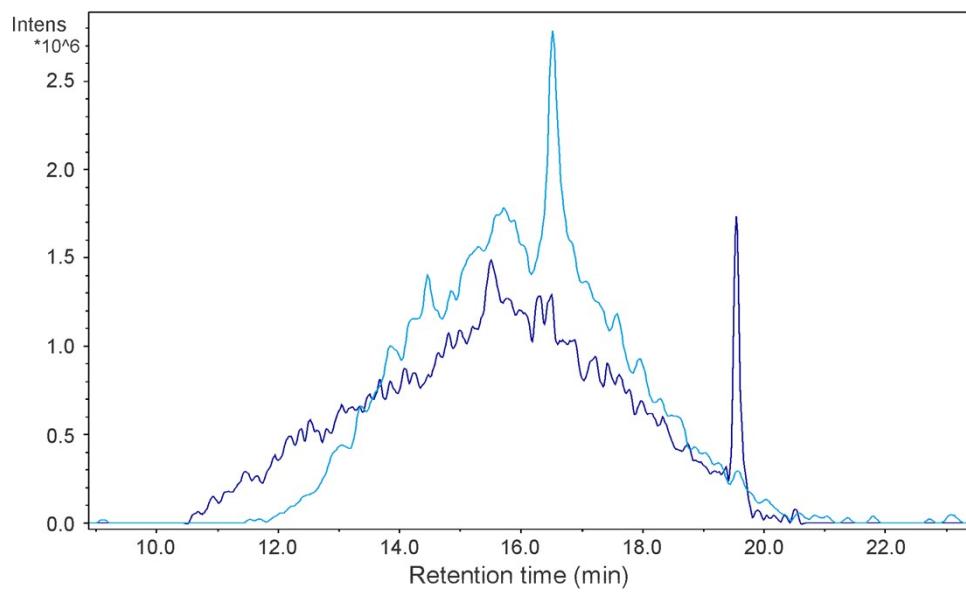
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58      **Figure S3.** All uniquely detected DOM compounds in SPE (DW-SPE-  
nLC-A, blue) and non-SPE (DW-nLC-A, dark blue) processed samples.  
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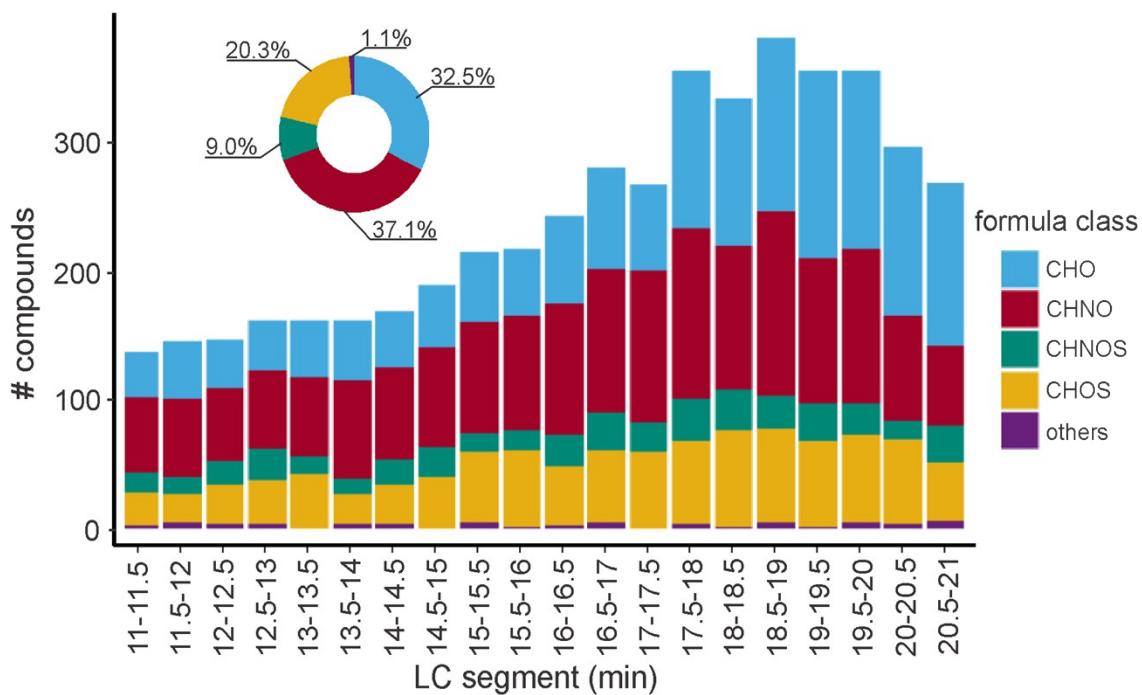
61 **Figure S4.** Intensity weighted average modified aromaticity index ( $Al_{mod}$ ) (a) and DBE-O (b) of DOM  
62 compounds in different segments in SPE (DW-SPE-nLC-A, blue) and non-SPE (DW-nLC-A, dark blue)  
63 processed samples.



66      **Figure S5.** EIC of  $m/z$  313.0929,  $C_{14}H_{18}O_8$  in DW-nLC-A (dark blue, without extraction), DW-SPE-  
67      nLC-A (light blue, extracted with SPE).

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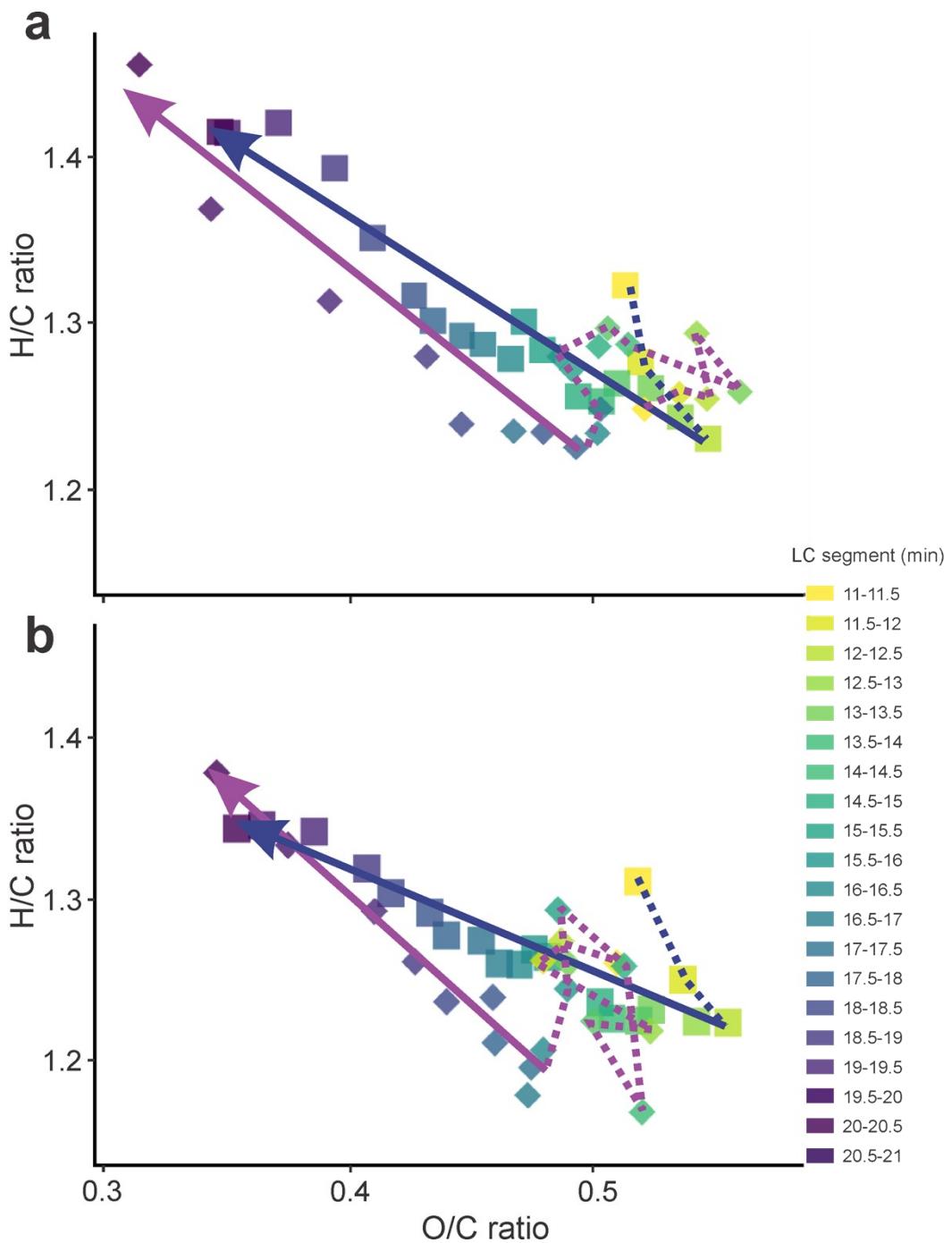
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71 [Figure S6](#). Formula class distribution of non-halogenated DBP compounds in sample DW-nLC-B. The  
 72 bars indicate the total number of non-halogenated DBPs detected in each segment distinguished by  
 73 formula classes (CHO, CHNO, CHOS, CHNOS, and others). The inlay summarizes the percentage of  
 74 formula classes of all non-halogenated DBPs (all segments combined).

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Figure S7. Intensity weighted average hydrogen-to-carbon (H/C) vs oxygen-to-carbon (O/C) ratios of all segments of DOM (square, connected with blue line) and non-halogenated DBPs (diamond, connected with purple line) MFs detected in a) DW-nLC-A and b) DW-nLC-B. Color represents mean retention time (min) of segments. Lines connecting dots are used as visual aid.

83 **Table S6.** Peak mass to charge ratio (*m/z*), signal to noise ratio (S/N), molecular formula (MF)  
 84 MF hydrogen to carbon ratio (H/C), oxygen to carbon ratio (O/C), double bond equivalent (DBE), double  
 85 bond equivalent minus oxygen (DBE-O), and retention time (RT) of corresponding segment of detected  
 86 chlorinated DBPs (Cl-DBPs) in sample DW-nLC-A and DW-nLC-B.

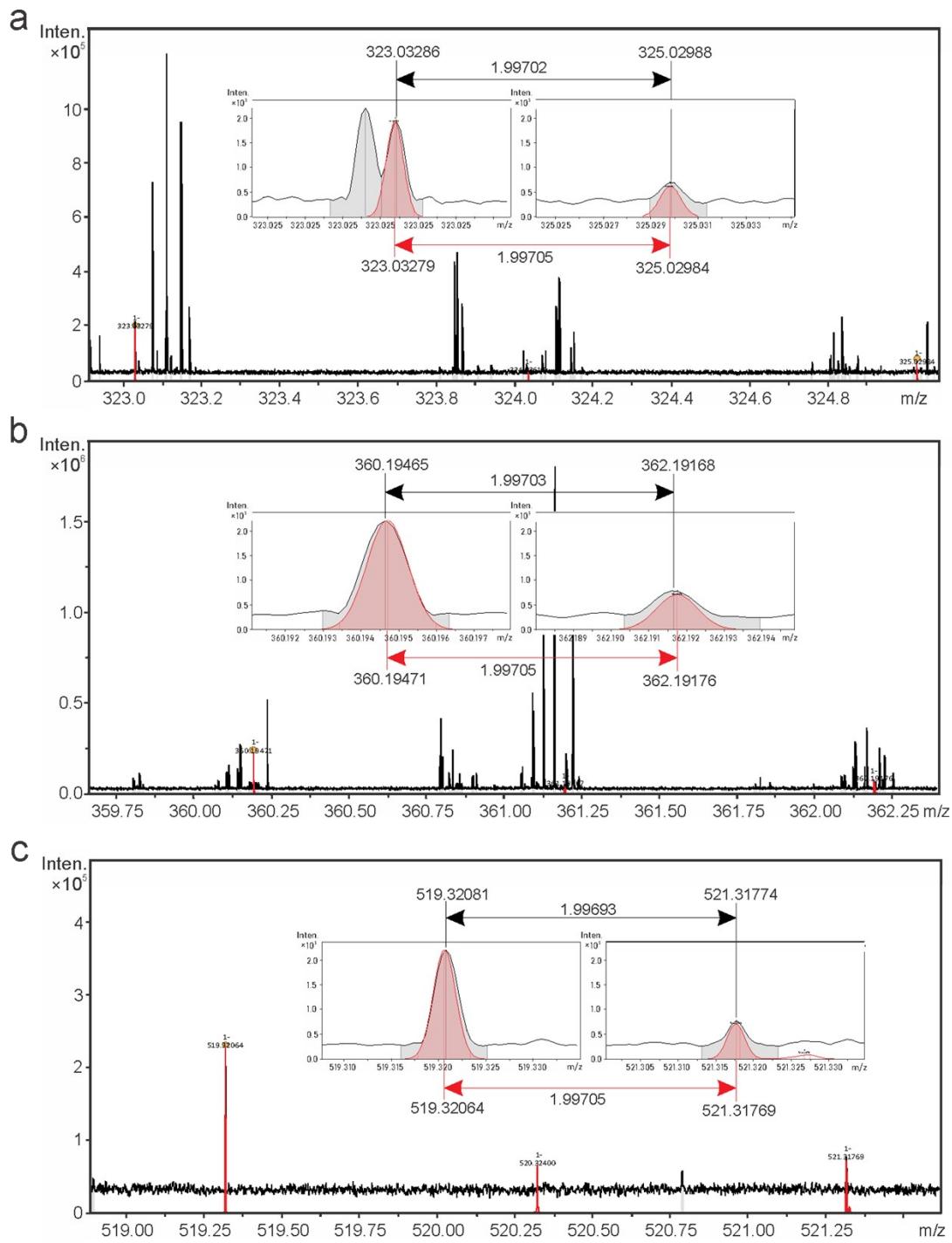
#	peak m/z	peak S/N	Molecular formula	H/C	O/C	DBE	DBE-O	RT (min)	sample	Figure S8
1	323.03286	47	C15 H13 Cl1 O6	0.87	0.40	9	3	16-16.5	DW-nLC-A	a
2	360.19465	52	C18 H32 Cl1 N1 O4	1.78	0.22	3	-1	18-18.5	DW-nLC-A	b
3	519.32081	60	C26 H49 Cl1 N2 O6	1.89	0.23	3	-3	11.5-12	DW-nLC-B	c
4	429.13223	33	C20 H27 Cl1 O8	1.35	0.40	7	-1	16.5-17	DW-nLC-B	d
5	329.06992	48	C17 H15 Cl1 N2 O3	0.89	0.18	11	8	17-17.5	DW-nLC-B	e
6	413.13738	37	C20 H27 Cl1 O7	1.35	0.35	7	0	17-17.5	DW-nLC-B	f
7	378.20526	54	C18 H34 Cl1 N1 O5	1.89	0.28	2	-3	18-18.5	DW-nLC-B	g
8	396.21586	50	C18 H36 Cl1 N1 O6	2.00	0.33	1	-5	18-18.5	DW-nLC-B	h
9	396.21586	59	C18 H36 Cl1 N1 O6	2.00	0.33	1	-5	18.5-19	DW-nLC-B	i
10	280.98919	67	C9 H11 Cl1 O6 S1	1.22	0.67	4	-2	18.5-19	DW-nLC-B	j
11	336.19467	53	C16 H32 Cl1 N1 O4	2.00	0.25	1	-3	18.5-19	DW-nLC-B	k
12	415.14322	38	C22 H25 Cl1 N2 O4	1.14	0.18	11	7	19-19.5	DW-nLC-B	l
13	377.02334	13	C15 H17 Cl3 N2 O3	1.13	0.20	7	4	19.5-20	DW-nLC-B	m
14	382.23658	27	C18 H38 Cl1 N1 O5	2.11	0.28	0	-5	19.5-20	DW-nLC-B	n
15	348.23106	95	C18 H36 Cl1 N1 O3	2.00	0.17	1	-2	20-20.5	DW-nLC-B	o
16	344.19981	101	C18 H32 Cl1 N1 O3	1.78	0.17	3	0	20-20.5	DW-nLC-B	p

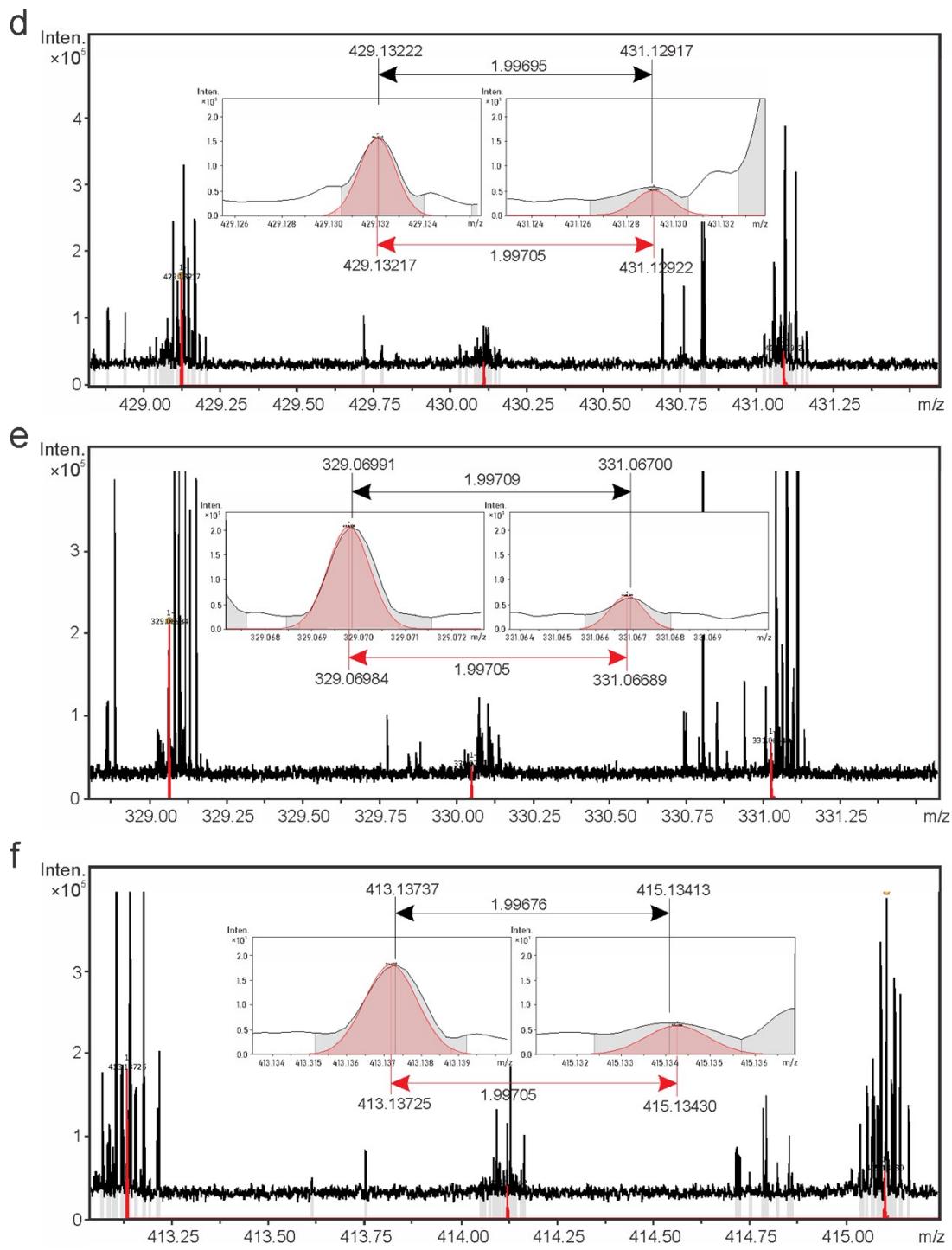
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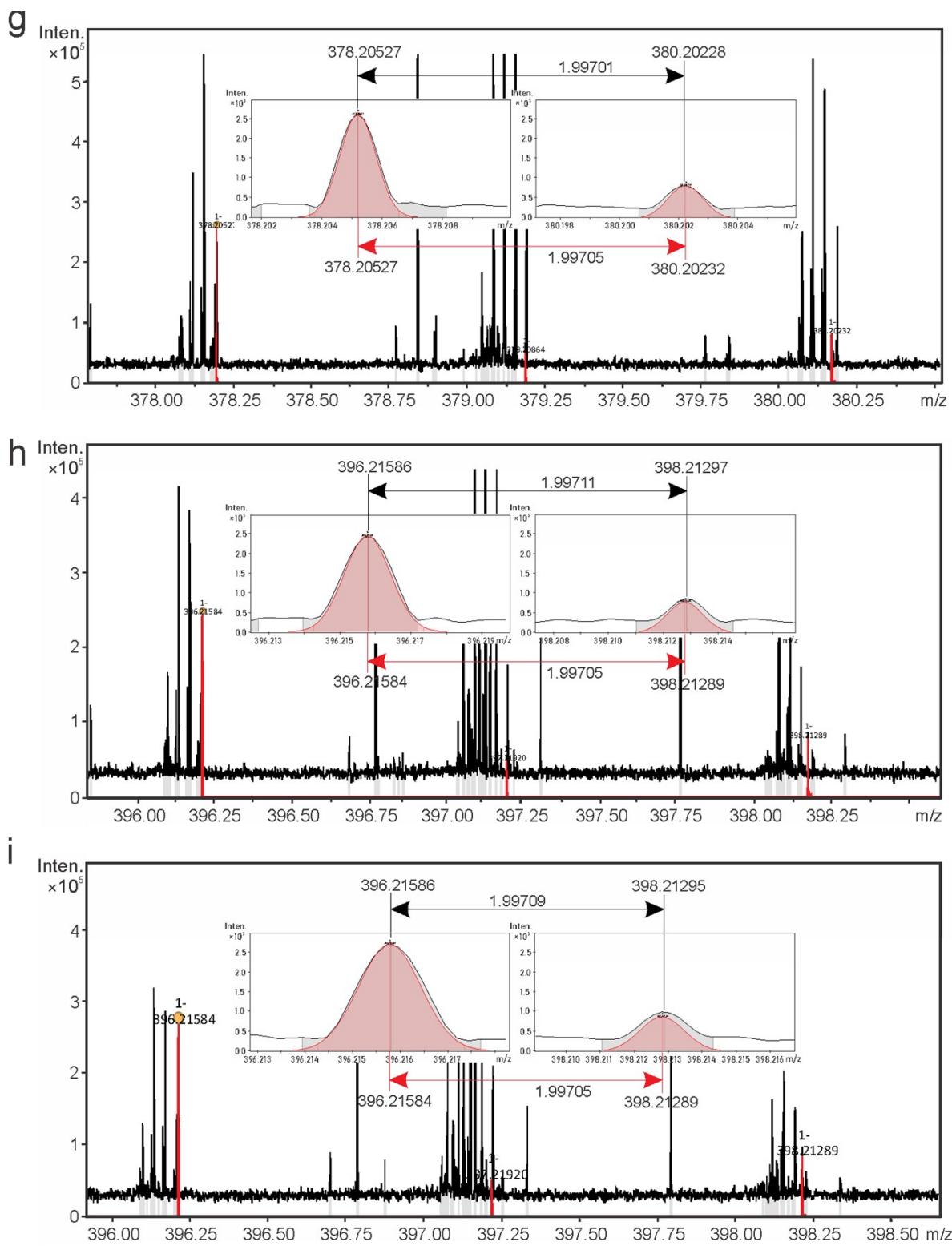
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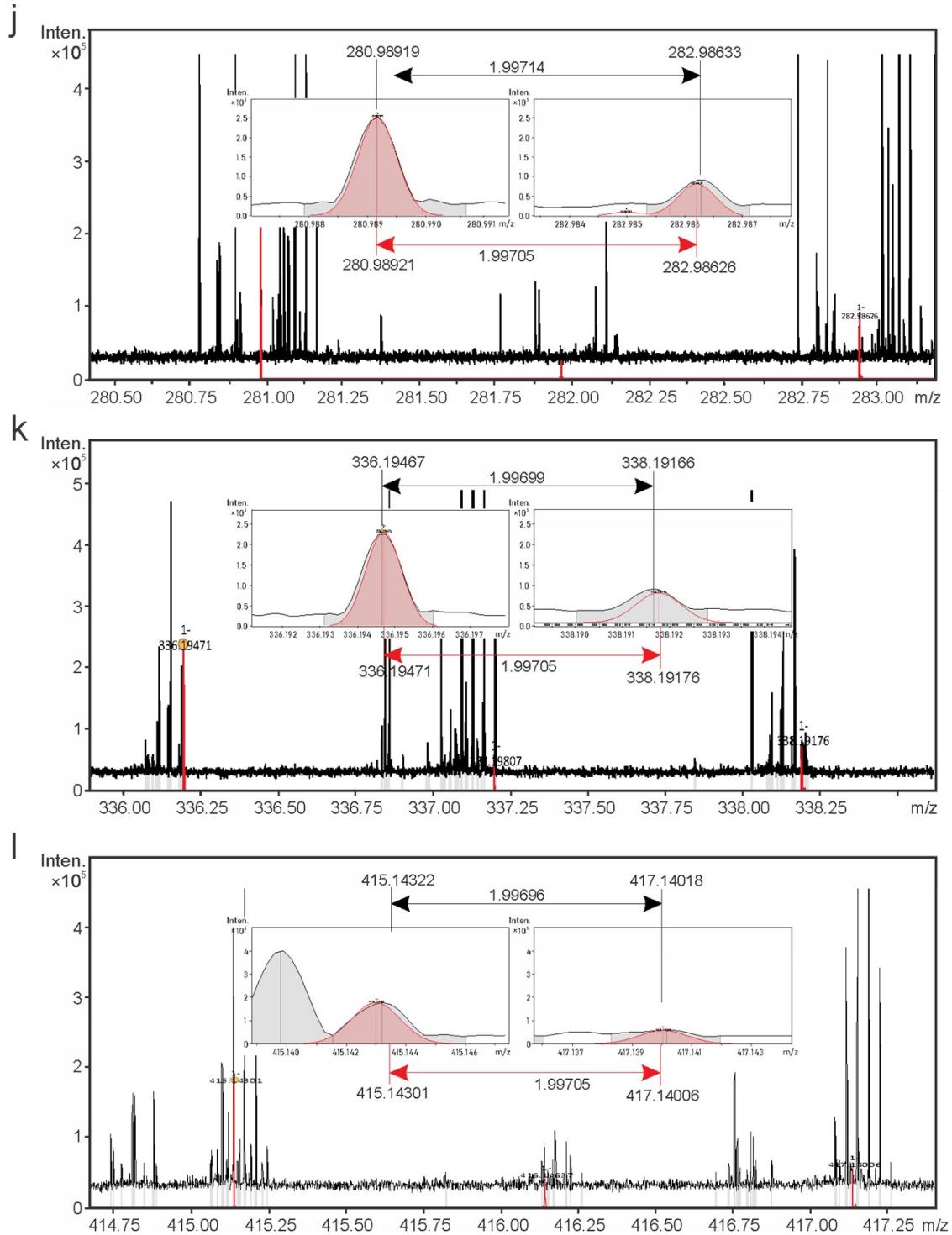
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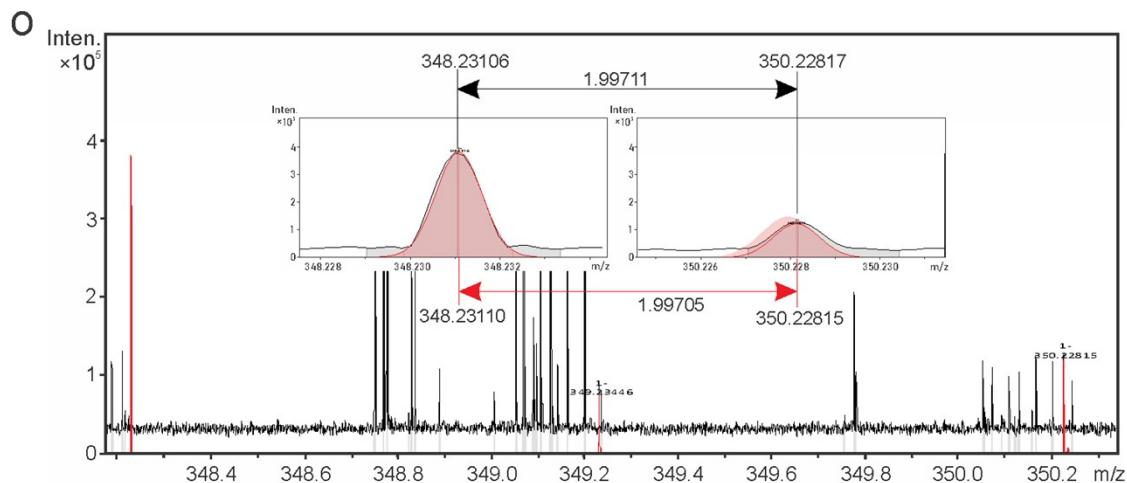
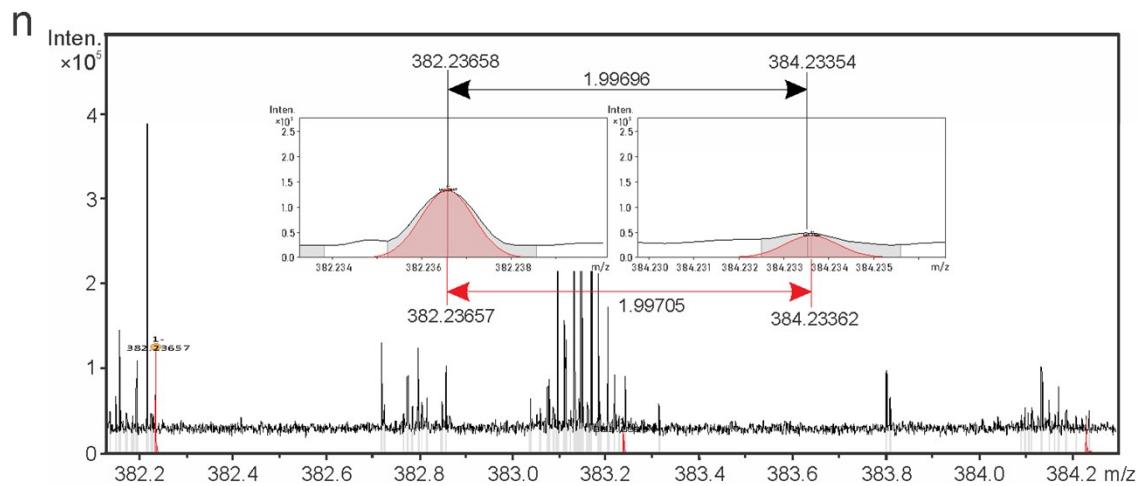
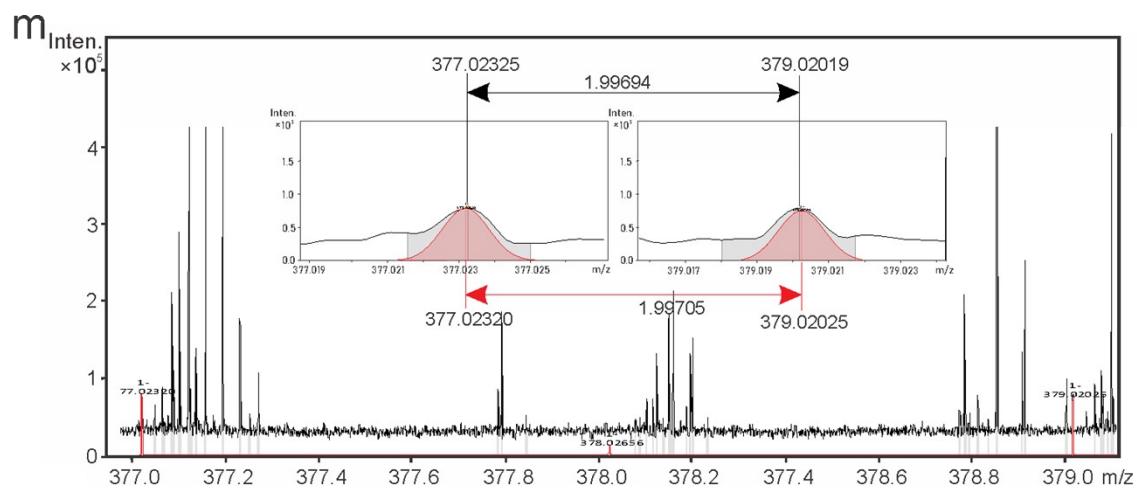
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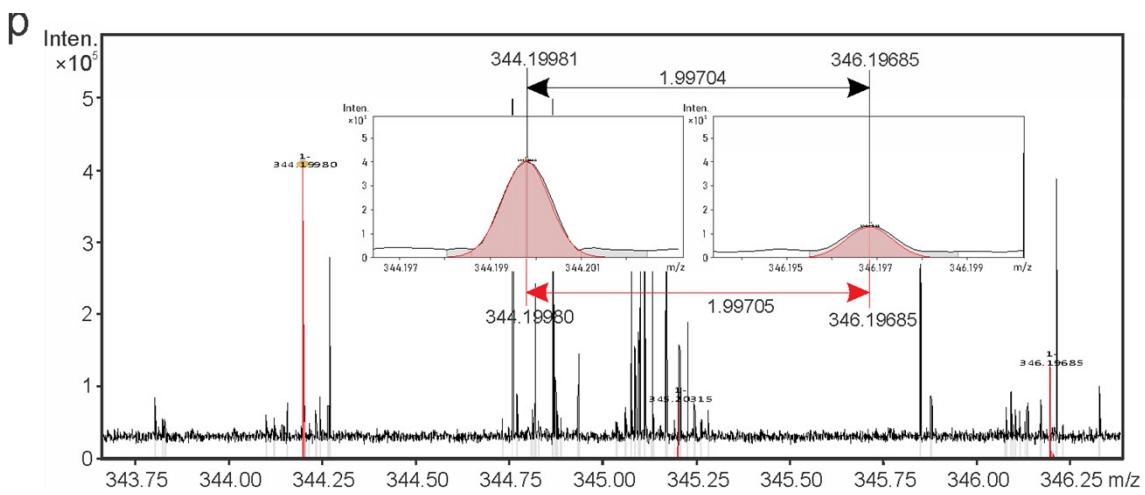






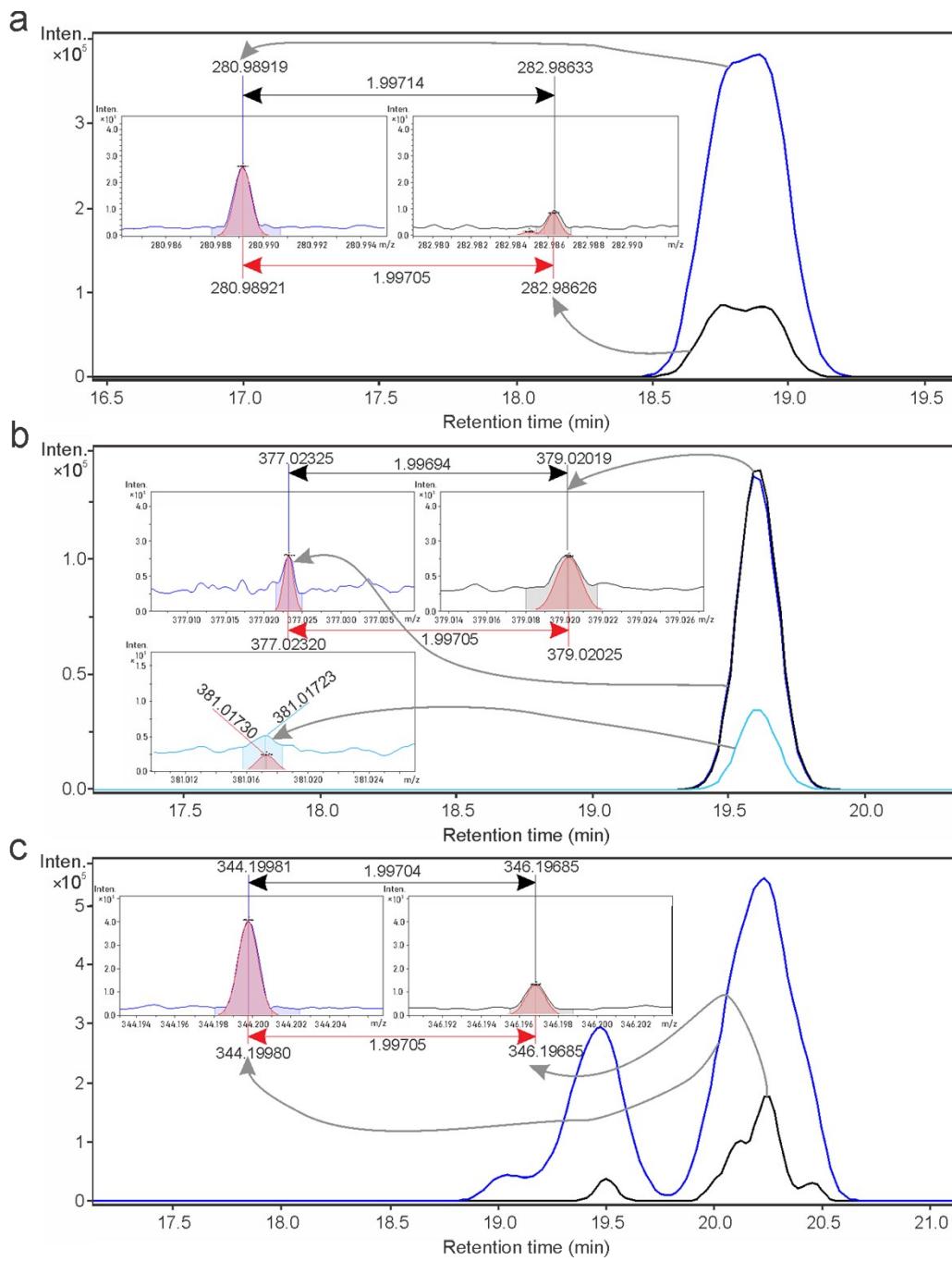






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98 **Figure S8.** Raw spectra (black) and theoretical isotopologue intensity pattern matching for 16  
 99 Cl-DBPs (ref. Table S6) detected in DW-nLC-A (a and b) and DW-nLC-B (c to p). The mass  
 100 differences of isotopologue peaks were indicated with black (raw spectra) and red  
 101 (theoretical isotopologue) arrows.



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103      **Figure S9.** EICs of molecular formula a) C9 H11 Cl1 O6 S1, b) C15 H17 Cl3 N2 O3, and c) C18  
 104      H32 Cl1 N1 O3. For mono-chlorinated MF:  $^{35}\text{Cl}_1$  (blue),  $^{37}\text{Cl}_1$  (black); for tri-chlorinated MF:  
 105       $^{35}\text{Cl}_3$  (blue),  $^{35}\text{Cl}_2\text{ }^{37}\text{Cl}_1$  (black),  $^{35}\text{Cl}_1\text{ }^{37}\text{Cl}_2$  (light-blue). Inside figures: the theoretical isotopologue  
 106      peaks (red) and measured peaks (blue, black, and light-blue) in raw spectra in DW-nLC-B. The  
 107      mass differences of isotopologue peaks were indicated with black (raw spectra) and red  
 108      (theoretical isotopologue) arrows.



110 Additional References:

111 (1) Han, L.; Kaesler, J.; Peng, C.; Reemtsma, T.; Lechtenfeld, O. J. Online Counter Gradient LC-FT-ICR-MS  
112 Enables Detection of Highly Polar Natural Organic Matter Fractions. *Analytical Chemistry* **2021**, *93* (3),  
113 1740-1748. DOI: 10.1021/acs.analchem.0c04426.

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